1 Particulate trace metal dynamics in response to increased CO₂ and iron availability in a

2 coastal mesocosm experiment

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14 Abstract. Rising concentrations of atmospheric carbon dioxide are causing ocean acidification and will influence 15 marine processes and trace metal biogeochemistry. In June 2012, in Raunefjord (Bergen, Norway) we performed a 16 mesocosm experiment, comprised of a fully factorial design of ambient and elevated pCO_2 and/or an addition of the 17 siderophore desferrioxamine B (DFB). In addition, the macronutrient concentrations were manipulated to enhance a 18 bloom of the coccolithophore Emiliania huxleyi. We report here the changes in particulate trace metal concentrations 19 during this experiment. Our results show that particulate Ti and Fe were dominated by lithogenic material while 20 particulate Cu, Co, Mn, Zn, Mo and Cd had a strong biogenic component. Furthermore, significant correlations were 21 found between particulate concentrations (mol L⁻¹) of Cu, Co, Zn, Cd, Mn, Mo, and P in seawater and phytoplankton 22 biomass (µgC L⁻¹), supporting a significant influence of the bloom in the distribution of these particulate elements. The 23 concentrations of these biogenic metals (mol L⁻¹) in the *E*. huxlevi bloom were ranked as: $Zn > Cu \approx Mn > Mo > Co >$ 24 Cd. Changes in CO₂ affected total particulate concentrations (mol L⁻¹) and biogenic metal ratios (Me:P) for some 25 metals, while the addition of DFB only affected significantly the concentrations of some particulate metals (mol L 26 ¹). Variations in CO₂ had the most clear, and significant effect on particulate Fe concentrations (mol L⁻¹), decreasing its 27 concentration under high CO2. Indeed, high CO2 and/or DFB promoted the dissolution of particulate Fe, and the 28 presence of this siderophore helped maintaining high dissolved Fe. This shift between particulate and dissolved Fe 29 concentrations, in the presence of DFB, promoted a massive bloom of E. huxleyi in the treatments with ambient CO₂. 30 Furthermore, high CO2 decreased the Me:P ratios of Co, Zn and Mn, while increased the Cu:P ratios. These findings 31 support theoretical predictions that the Me:P ratios of metals whose seawater dissolved speciation is dominated by free 32 ions (e.g. Co, Zn and Mn) will likely decrease or stay constant under ocean acidification. In contrast, high CO2 is 33 predicated to shift the speciation of dissolved metals associated with carbonates, such as Cu, increasing their 34 bioavailability, and resulting in higher Me:P ratios. 35

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Key words: Global change, Fe, CO₂, particulate trace metals, dissolved trace metals, mesocosms, *Emiliania huxleyi*,
 phytoplankton

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40 **1. Introduction**

- 41 Marine phytoplankton contribute half of the world's total primary productivity, sustaining marine food webs and
- 42 driving the biogeochemical cycles of carbon and nutrients (Field et al., 1998). Annually, phytoplankton incorporate
- 43 approximately 45 to 50 billion metric tons of inorganic carbon (Field et al., 1998), removing a quarter of the CO₂
- 44 emitted to the atmosphere by anthropogenic activities (Canadell et al., 2007). Yet, the atmospheric CO₂ concentration
- 45 has increased by 40 % since pre-industrial times as a result of anthropogenic CO₂ emissions, producing rapid changes
- 46 in the global climate system (Stocker et al., 2013). The dissolution of anthropogenic CO₂ in seawater, causes shifts in
- 47 the carbonate chemical speciation, and leads to ocean acidification (OA). Marine ecosystems are sensitive to changes in
- 48 pH because pH strongly affects chemical and physiological reactions (Hoffman et al., 2012). Increased CO₂ in seawater
- 49 may enhance or diminish phytoplankton productivity (Mackey et al., 2015), decrease the CaCO₃ production in most
- 50 planktonic calcifiers (Riebesell and Tortell 2011), and/or inhibit organic nitrogen and phosphorus acquisition (Hutchins
- 51 et al., 2009). Thus, the biogeochemical cycling of nutrients is predicted to be highly affected by OA (Hutchins et al.
- 52 2009), as well as the distribution and speciation of trace metals in the ocean (Millero et al., 2009).
- 53

54 Trace metals, including Fe, Zn, Mn, Cu, Co and Mo, are essential for biological functions (e.g. photosynthesis, 55 respiration and macronutrient assimilation), and Cd can supplement these functions. Trace metals availability can 56 influence phytoplankton growth and community structure (Morel and Price, 2003). In turn, plankton control the 57 distribution, chemical speciation, and cycling of trace metals in the sea (Sunda, 2012), by, for example, releasing 58 organic compounds that dominate the coordination chemistry of metals, internalizing trace elements into the cells, and 59 reducing and/or oxidizing metals at the cell surface. The chemistry of redox speciation of active trace metals is highly 60 dependent on pH. Fe occurs in two main redox states in the environment: oxidized ferric Fe (Fe (III)), which is 61 poorly soluble at circumneutral pH; and reduced ferrous Fe (Fe (II)), which is more soluble in natural seawater, but 62 becomes rapidly oxidized (Millero et al. 1987). Fe speciation and bio- availability are dynamically controlled by the 63 prevalent changing redox conditions. Also, as the ocean becomes more acidic, reduction of Cu (II) will increase, as the 64 ionic form of Cu (II) is reduced to Cu (I) (Millero et al., 2009). The effect of higher concentrations of Cu (I) in surface 65 waters on biological systems is not well known. Therefore, while the effects of OA on inorganic metal speciation will 66 be more pronounced for metals that form strong complexes with carbonates (e.g. copper) or hydroxides (e.g. Fe and 67 aluminium), those that form stable complexes with chlorides (e.g. cadmium) will not be greatly affected. pH mediated 68 changes in concentrations and/or speciation could possibly enhance trace metals limitation and/or toxicity to marine 69 plankton (Millero et al., 2009).

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71 Fe is crucial for phytoplankton growth because it is involved in many essential physiological processes, such as 72 photosynthesis, respiration, and nitrate assimilation (Behrenfeld and Milligan, 2013). The decrease in seawater pH in 73 response to OA may increase Fe solubility (Millero et al., 2009), but it may also result in unchanged or lower Fe 74 the nature of the strong organic Fe ligands (Shi et al., 2010). Consequently, changes in bioavailability, depending on 75 Fe bioavailability due to ocean acidification can affect positively or negatively ocean productivity and CO₂ drawdown. 76 Copper is an essential micronutrient but may be toxic at high concentrations (Semeniuk et al., 2016). An increase in 77 free cupric ion concentrations in coastal areas due to ocean acidification (Millero et al., 2009) could result in negative 78 effects on phytoplankton. Given that trace metals are essential for phytoplankton productivity, and that they are 79 actively internalized during growth, it is important to study the impacts of ocean acidification on the trace metal content

- 80 of ecologically significant plankton species.
- 81
- 82 In a rapidly changing global environment, generated by anthropogenic CO₂ emissions, it is critical to gain adequate

- 83 understanding about ecosystem responses. Due to the complex interactions in aquatic ecosystems, such predictions have
- 84 so far not been possible to do based upon observational data and modelling alone. However, direct empirical studies on
- 85 natural communities offer a robust tool to analyse interactive effects of multiple stressors. Specifically, mesocosm
- 86 experiments allow perturbation studies with a high degree of realism compared to other experimental systems such as in
- 87 the laboratory (high controlled conditions usually far from reality) or *in situ* in the ocean (where not all the interactions
- 88 are contemplated) (Riebesell et al., 2010, Stewart et al., 2013, Riebesell and Gatusso, 2015).
- 89

90 In the present work a bloom of the coccolithophorid *Emiliania huxleyi* was induced in a mesocosm experiment in a

Norwegian fjord, where the speciation of particulate and dissolved trace metals is very dynamic (e.g. Fe; Ozturk et al.
 2002). We aimed to examine and characterize the change of particle trace metals during an E. huxleyi bloom under the

- 93 interactive effects of increased CO₂ and/or dissolved Fe. *Emiliania huxleyi* is the most cosmopolitan and abundant
- 94 coccolithophore in the modern ocean (Paasche, 2002) and its growth and physiology has been studied under this

95 experimental conditions (Segovia et al., 2017, Segovia et al., 2018, Lorenzo et al., 2018). Furthermore, *E. huxleyi* has

96 unique trace metal requirements relative to other abundant phytoplankton taxa (ie. diatoms or dinoflagellates; Ho et al.

97 2003). Coccolithophores play a key role in the global carbon cycle because they produce photosynthetically organic

98 carbon, as well as particulate inorganic carbon through calcification. These two processes foster the sinking of

99 particulate organic carbon—and trace metals—and contribute to deep ocean carbon export (Hutchings, 2011) and

100 ultimately to organic carbon burial in marine sediments (Archer, 1991, Archer and Maier-Reimer., 1994). However,

101 ocean acidification will disproportionally affect the abundance of coccolithophores, as well as their rates of

calcification and organic carbon fixation (Zondervan et al., 2007). The aim of the present study was to characterize the
 changes in particulate trace metal concentrations—in both lithogenic and biogenic particles— during a bloom of
 E. huxlevi under realistic changes in CO₂ and Fe bioavailability expected by 2100.

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107 **2. Materials and methods**

108 2.1 Experimental set-up

109 The experimental work was carried out in June 2012 in the Raunefjord, off Bergen, Norway as described in detail by 110 Segovia et al., (2017). Twelve mesocosms (11 m³ each) were set-up in a fully factorial design with all combinations of 111 ambient and elevated pCO_2 and dFe in three independent replicate mesocosms. The mesocosms were covered by lids

- 112 (both transparent to PAR and UVR) and filled with fjord water from 8 m depth. We achieved two CO₂ levels
- 113 corresponding to present (390 ppm, LC) and those predicted for 2100 (900 ppm, HC) by adding different quantities of
- pure CO₂ gas (Shculz et al., 2009). The specific CO₂ concentration and the CO₂ inlet flows in the mesocosms were
- 115 measured by non-dispersive infrared analysis by using a Li-Cor (LI-820) CO₂ gas analyser (Li-COR, Nebraska, USA).
- 116 CO₂ concentrations in the mesocosms were calculated from pH and total alkalinity measurements using the CO₂ SYS
- 117 software (Robbins et al, 2010). At the beginning of the experiment, nitrate (10 µM final concentration) and phosphate
- 118 (0.3 µM final concentration) were added to induce a bloom of the coccolithophore *Emiliania huxleyi*, according to Egge
- 119 & Heimdal (1994). Following recommendations by Marchetti and Maldonado (2016), t o induce changes in Fe
- 120 availability, and analyse its effects on the plankton community, 70 nM (final concentration) of the siderophore
- 121 desferrioxamine B (DFB) (+DFB and -DFB treatments) (Figure S1b-supplemental material) was added to half of the
- 122 mesocosms on Day 7, when the community was already acclimated to high CO₂. The initial dFe concentration before
- 123 DFB addition was about 4.5 nM. Even though DFB is a strong Fe-binding organic ligand often used to induce Fe
- 124 limitation in phytoplankton (Wells 1999), DFB additions may also in- crease the dissolved Fe pool in environments
- 125 with high concentrations of colloidal and/or particulate Fe, such as fjords (Kuma et al. 1996, Öztürk et al. 2002). By day

- 126 17, dissolved Fe concentrations were significantly higher (by \sim 3-fold) in the high CO₂ and DFB treatments than in the
- 127 control (Segovia et al. 2017). These results support an increase in the solubility of Fe in seawater by either lowering its
- 128 pH (Millero 1998; Millero et al. 2009) and/or the addition of DFB (Chen et al. 2004). The multifactorial experimental
- 129 design consisted of triplicate mesocosms per treatment and the combinations of high and ambient pCO₂ and dFe levels,
- 130 resulted in a total of 12 mesocosms: LC-DFB (control), LC+DFB, HC+DFB and HC-DFB. Water samples from each
- 131 mesocosm were taken from 2 m depth by gentle vacuum pumping of 25 L volume into acid-washed carboys by using
- 132 membrane vacuum pumps (PALL) working at reverse flow. Carboys were quickly transported to the onshore
- 133 laboratory. The biological and chemical variables analysed were phytoplankton abundance and species composition,
- 134 dissolved Fe and Cu concentrations (dFe, dCu), nutrient concentrations (nitrate, phosphate, silicic acid and ammonium)
- 135 and particulate trace metal concentrations.

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137 2.2 Dissolved copper (dCu)

- 138 Low density polyethylene (LDPE) bottles were cleaned with 1% alkaline soap solution for one week, then filled with 6
- 139 M trace metal grade HCl (Seastar, Fisher Chemicals) and submerged in a 2 M HCl bath for one month. For transport, 140
- they were filled with 1 M trace metal grade HCl for one more month and kept double bagged. In between each acid
- 141 treatment, the bottles were rinsed with Milli-Q water (Millipore; hereafter referred to as MQ). Before sampling, the
- 142 bottles were rinsed three times with filtered seawater. Seawater was collected from each mesocosm, filtered through
- 143 AcroPak® capsule filters with 0.2 µm Supor® membrane into the trace metal clean LDPE bottles, and acidified with
- 144 ultra-clean trace metal grade HCl in a Class 100 laminar flow hood. Total dissolved Cu concentrations were measured
- 145 following Zamzow et al., (1998) using a flow injection analysis chemiluminescence detection system (CL-FIA,
- 146 Waterville Analytical). Total dissolved Fe concentrations were measured as described in Segovia et al., (2017) for this
- 147 very experiment. The pH of the 0.2 µm filtered dFe samples was lowered to 1.7 by using SeaStar HCl upon collection.
- 148 Lowering the pH to 1.7, with HCl for more than 24 hours, ensures solubilisation of all the Fe in the sample, as well as 149 the release of all the Fe bound within strong organic complexes (such as Fe-DFB), thus making all dFe available for
- 150 analysis (Johnson et al. 2007). During (FIA-CL, the sample is only buffered to a higher pH immediately before entering 151 the flow cell, right in front of the photomultiplier; so that Fe-DFB complexing kinetics are sufficiently slow to allow 152
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155 2.3 Particulate metals (pMe)

total dFe to be measured.

156 2.3.1 Sampling

157 All equipment and sampling material used during this study was rigorously acid-washed under trace metal clean 158 conditions and protocols according to GEOTRACES . The material was cleaned with Milli-Q water (MQw) with 10 % 159 Extran (Fisher Chemicals) at 60°C for 6h, followed by 3 thorough rinses with MQw at room T. The material was then 160 cleaned with 10 % HPLC grade HCl (Sigma-Aldrich) at 60°C or 12h and then rinsed thoroughly 5 times with MQw at 161 room T. The material was then covered by plastic and transported to the raft. Sampling in the raft was carried out under 162 a mobile plastic cover hood. Filters were precleaned with 10% trace metal grade hydrochloric acid (Seastar, Fisher 163 Chemicals), at 60°C overnight and were rinsed with MQw. Seawater samples (1-3.5 L) were filtered gently onto 0.45 164 µm acid washed Supor ®-450 filters (within a trace metal clean Swinnex filter holder) on days 12, 17 and 21 of the 165 experiment. Four technical replicates were taken from each mesocosm. Two filters were analysed without oxalate-

- 166 EDTA wash and the other two were individually washed with oxalate-EDTA reagent to remove extracellular Fe, as
- 167 well as other metals (Tang and Morel, 2006). Immediately following filtration, the treated filters were soaked with 20

- 168 mL EDTA-oxalate solution, added to the headspace of the Swinnex holders, with an acid-washed polypropylene
- 169 syringe. After 10 min, vacuum was applied to remove the oxalate solution and 10 mL of 0.2 μm filtered chelexed
- 170 synthetic oceanic water (SOW) solution was passed through the filter to rinse off any remaining oxalate solution.
- 171 Replicate filters that were not treated with oxalate solution were transferred directly to 2 mL centrifuge polypropylene
- tubes for storage. The filters with particles were frozen in acid-washed 2 mL polypropylene tubes and then, dried andstored until analysis.
- 174

175 2.3.2 Analytical methods

176 Filters were digested in 7-mL acid-washed Teflon (Teflon, Rochester, NY, USA) vials. Teflon vials were also 177 precleaned using 10% trace metal hydrochloric acid (Fisher, trace metal grade) during two days and then, with nitric 178 acid (Fisher, trace metal grade) at 70 °C during three days. In between each acid treatment, the bottles were rinsed with 179 MQ. Samples were digested in 3 mL of HNO₃ and 0.5 mL of HF (Fisher, trace metal grade) with lids on for 1 h on a 180 hot plate at 200 °C. The lids were then removed to evaporate HF at 200°C. After this, 1.5 mL of HNO3 were added and 181 the samples were heated with lids on overnight at 150 °C. Finally, 2.25 mL of HClO4 (Fisher, Optima grade) were 182 added and the samples were heated for 4 h at 200 °C. After complete digestion, the samples were dried on hot plates at 183 200°C. The dried samples were dissolved in 1% nitric acid with 1 ppb in internal standard. The analysis was performed 184 using a high-resolution inductively coupled plasma-mass spectrometer (ICP-MS, Element XR, Thermo Scientific) and 185 the described instrumental settings (Table S1). Filter blanks were collected and subjected to the same storage, digestion, 186 dilution, and analysis processes, and these blank values were subtracted from sample measurements. Particulate 187 samples for ICP-MS analysis were processed in a trace metal-clean laboratory under a trace metal-clean laminar flow 188 fume hood.

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190 2.3.3 The effect of oxalate-EDTA wash on particulate trace metal concentrations

191 To better estimate the biogenic fraction of the particulate metals, the filters were washed with an oxalate-EDTA 192 solution, which removes extracellular metals and oxyhydroxides (Tovar-Sanchez et al., 2003; Tang and Morel, 2006). 193 In our study, the oxalate wash significantly decreased the concentration of all particulate metals, with the exception of 194 Al and Ti (Tables S2 and S3), as observed by Rauschenberg and Twining (2015). The quantity of metal remaining after 195 the oxalate wash (i.e. biogenic fraction) varied among elements (Tables S2 and S3). In general, the concentrations of Fe 196 and Co in the particles were decreased the least by the oxalate wash by $\sim 25\%$, while Mo and Pb concentrations were 197 decreased the most by ~70%. The concentrations of particulate Cu, Zn, Cd and Mn were reduced by 50% by the oxalate 198 wash. As shown previously (Sanudo-Wilhelmy et al. 2004), the oxalate reagent also removed extracellular P (by ~20%, 199 Table S2 & S3). Compared to Rauschenberg and Twining (2015), the estimates of the biogenic fraction, after the 200 oxalate wash, were in agreement for Co, Cu and P, and lower for Fe, Mn, Zn and Cd concentrations.

201

However, the efficacy of the oxalate wash to dissolve Fe, and other metals, from lithogenic particles is not well

- 203 constrained (Frew et al. 2006, Rauschenberg and Twining., 2015, King et al., 2012). Therefore, the results obtained
- after the oxalate-EDTA wash should be interpreted with caution because we do not know whether the removed metal
- 205 fraction is a) only lithogenic; b) mainly lithogenic but some biogenic fraction is also removed, or c) whether metals
- 206 absorbed onto particles are equally labile to the wash on biogenic and lithogenic particles. Given that many of the
- 207 trends we observed were identical for the oxalate-EDTA washed and non-washed particles [i.e. higher Me
- 208 concentrations in the LC+DFB treatments (Table S2 & S3) and positive correlations between phytoplankton biomass
- and Me concentrations (Lorenzo-Garrido 2016)], below we present and discuss only the non-oxalate wash results.
- 210

211 2.4 Statistical analyses

- 212 Data were checked for normality (by Shapiro-Wilks' test), homoscedasticity (by Levene's test) and sphericity (by
- 213 Mauchly's test). All data met the requirements to perform parametric tests. Statistical significance of treatment effects
- 214 was carried out using Split-Plot ANOVA followed by post-hoc Sidak and Bonferroni tests (considering P < 0.05 as
- 215 significant). All analyses were performed using the General Linear Model (GLM) procedure. The correlation between
- 216 variables was analysed by Pearson's product-moment multiple comparisons (considering P < 0.05 as significant).
- 217 Statistical analyses were carried out using SPSS v22 (IBM statistics) and Sigmaplot 12 (Systat Software, Chicago,
- 218 USA).
- 219

3. Results

221 **3.1 Biological and chemical characteristics during the bloom**

- 222 Plankton community dynamics and their response to the applied treatments in the mesocosms are described in detail by
- 223 Segovia *et al.* (2017). Briefly, at the beginning of the experiment (days 1-10) a bloom of large chain-forming diatoms
- 224 was observed, which declined by day 7 (Figure S1g- supplemental material). This diatom bloom decline was
- associated with a sharp decrease in nitrate and silicic acid concentrations (Figure S2-supplemental material, see Segovia
- et al., 2017 for further details). Picoeukaryotes, dominated the phytoplankton community on day 8 (Figure S1d).
- 227 During the first 10 days of the experiment, there were no significant differences in the chemical variables measured
- between the treatments (Figures S3 and S2 -supplemental material). On day 7, half of the mesocosms were amended by

adding DFB (+DFB treatments). Between day 7 and 17, an increase in dFe was observed in all treatments, except in the

- control (Figure S3). This increase in dFe was sustained for the entire experiment in the DFB treatments (Figure S3).
- 231 Dissolved Cu concentrations were not affected by the different treatments (Figure S3). After day 10, a massive bloom
- of the coccolithophore *E. huxleyi* developed under LC +DFB condition (Figure S1b), out-competing the rest of the
- 233 plankton groups (Figure S1). This bloom was not observed either in the control treatment (LC-DFB) or in the HC
- treatments, although *E. huxleyi* was still the most abundant species in all treatments; with the exception of the HC-DFB
 treatment (Figure S1b).
- 236

237 **3.2** Particulate metal concentrations during the mesocosm experiment

238 The pMe concentrations (nM, mean of all treatments and dates) during the experiment were highest for Al, Fe and Zn, 239 and lowest for Cd, following this trend: Al \approx Fe \approx Zn > Ti > Cu \approx Mn > Mo \approx Pb > Co > Cd (Figure 1, Table S2). 240 Significant changes over time were observed for all particulate trace metal concentrations (Fe, Cu, Co, Zn, Cd, Mn, Mo 241 and Pb), except for Ti and Al (Figure 1, Table 1). The only metal that showed a significant time-dependent decrease in 242 its particulate concentration was Fe (Figure 1, Table 1). In general, the treatments with the highest particulate metals 243 concentrations also exhibited the highest particulate P, except for Al, Ti, Fe, and Pb (Figure 1, Table S2). On days 12 244 and 17, the highest particulate metals concentrations were observed in the LC+DFB treatment, while on day 21, they 245 were observed in both LC treatments (Figure 1, Table S2).

246

247 **3.3** The effects of increased CO₂ and the DFB addition on particulate metal concentrations

248 Increased CO₂ and the DFB addition did not significantly affect the concentrations of particulate Al, Ti, Cu, and Pb

249 (Tables 1 and S2). Similarly, the addition of DFB did not directly influence particulate concentrations of Fe, but high

250 CO₂ had a significant negative impact on particulate Fe (Tables 1 and S2, Figure 1). Particulate Cd concentrations were

- also inversely affected by CO₂, but only in the presence of DFB (CO₂; and CO₂ x DFB effect, Tables 1 and S2, Figure
- 252 1). All other elements (P, Co, Zn, Mn and Mo) exhibited significant effects by CO₂ and by DFB, but there was also a
- significant interaction between these two factors (Table 1, S2). This indicates that, for example, particulate Mn, Zn, Mo,

- 254 Co, and P concentrations were significantly decreased by high CO₂, but only in the +DFB treatments (Figure 1, Table 1,
- S2,). Similarly, the addition of DFB significantly increased pZn and pMn, but only at ambient CO₂ levels (Figure 1,
 Tables 1, S2).
- 257

3.4 Phosphorous-normalized metal ratios in particles collected from the mesocosms and the effects of increased CO₂ and the DFB addition on these ratios

- 260 The P-normalized metal ratios (Figure 2 and means in Table 2) were highest for Al and Fe (mean: 70 ± 38 mmol Al:
- 261 mol P, and 39 ± 34 mmol Fe: mol P), and lowest for Cd and Co (mean 0.02 ± 0.01 mmol Cd: mol P, and 0.07 ± 0.02
- 262 mmol Co: mol P). Fe:P and Ti:P were not significantly affected by increased CO₂ and/or the DFB addition, but showed
- a significant decrease over time (Table 3). The P-normalized Cu, Co and Zn ratios changed significantly over time
- 264 (Table 3). Increased CO₂ significantly decreased Co, Zn and Mn:P ratios, while it increased Cu:P ratios (Figure 2, Table
- 265 3). DFB did not affect the Me:P ratios of any of these bioactive elements (Table 3).
- 266

4. Discussion

268 4.1 The effects of CO₂ and dFe on the plankton community

269 In this experiment we investigated changes in particulate trace metal concentrations, in response to increased CO₂ 270 and/or an addition of the siderophore DFB in a coastal mesocosm experiment. For a better understanding of the 271 processes affecting these stressors, we briefly summarise the mesocosm experiment results originating from Segovia et 272 al. (2017). High CO₂, as well as the DFB addition elevated dFe concentration increasing Fe availability (see Segovia et 273 al. 2027 for further details). The higher dFe concentrations were sustained in the DFB treatments, suggesting that DFB 274 significantly increased the solubility of Fe, as previously shown (Chen et al. 2004). A bloom of the coccolithophore 275 Emiliania huxleyi was observed in the ambient CO₂ treatments, and was especially massive in the presence of DFB 276 Our results suggest that E. huxleyi is able to utilise DFB-bound Fe (Fe-DFB). Indeed, E. huxleyi has (LC+DFB). 277 been shown to produce a wide range of organic compounds with high affinity for Fe (Boye & Van den Berg 2000). 278 Furthermore, E. huxleyi is able to acquire Fe from organic Fe complexes (Hartnett et al. 2012), including Fe-DFB 279 (Shaked & Lis 2012, Lis et al. 2015). Indeed, the biomass of E. huxleyi was negatively affected by increased CO2. 280 However, increased dFe partially mitigated the negative effect of elevated CO₂, indicating that the coccolithophore was 281 able to acclimate better to ocean acidification when Fe availability was high. High dFe also had a positive effect on the 282 cyanobacterium Synechococcus sp, while the rest of the plankton food web did not response to the treatments (Segovia

283 et al. 2017).

4.2 Particulate Fe and Ti are associated with lithogenic sources, while particulate Co, Cu, Zn, Cd, Mo and Mn are associated with biogenic sources

- 286 The particulate trace metal concentrations (nM, mean of all treatments and dates) during the experiment were highest
- for Al, Fe and Zn, and lowest for Co and Cd, following this trend: $Al \approx Fe \approx Zn > Ti > Cu \approx Mn > Mo \approx Pb > Co > Cd$. Lithogenic particles are enriched in Al and low in P (average crustal Al and P content is 2.9 mmol Al and 0.034 mmol
- 289 P g^{-1} dry weight, Taylor 1964), while biogenic particles are enriched in P and low in Al (average plankton Al and P
- 290 content is 0.001 mmol Al and 0.26 mmol P g⁻¹ dry weight, Bruland et al. 1991). Therefore, the distinct high abundance
- of Al and P in lithogenic and biogenic particles, respectively, can be used to evaluate the relative contribution of
- 292 lithogenic and biogenic material in our particulate samples. In order to do this, first, it is important to establish that the
- 293 vast majority of the measured particulate P is associated the biogenic fraction. In this study, the abiotic P was estimated
- using particulate Al concentrations (nM) and the P:Al ratio in crustal material, and was calculated to be negligible (<
- 1% of the total measured particulate P). In addition, a significant correlation (p< 0.003) was found between particulate

200

296 P concentrations and phytoplankton biomass (Table 4). Therefore, we assume a constant trace metal composition in

- 297 biogenic particles (assuming they are rich in phytoplankton) and lithogenic particles (assuming they are rich in crustal
- 298 material). We then calculated the expected metal concentrations in the particulate samples assuming that all the P
- 299 measured in the particles is associated with a biogenic fraction, and that all Al in the particles is associated with the
- 300 lithogenic fraction. Thus, for a given trace metal, its expected particulate trace metal concentration in seawater (mol L⁻
- 301 ¹) can be calculated as the sum of the contribution from biogenic and lithogenic particles, so that:
- 302
- 303 [Me] = a [P] + b [Al]
- 304

305 where [Me] is the total concentration of the metal (mol L^{-1}) expected in the particulate sample; [P] is the P 306 concentration measured in the particles (mol L^{-1}); [A] is the Al concentration measured in the particles (nM L^{-1}); a is 307 the average, well-known metal content in biogenic particles, normalized to P (i.e. mol Me: mol P in marine plankton; 308 Ho 2006) and b is the average, well known metal content in lithogenic particles, normalized to Al (mol Me: mol Al in 309 the Earth crust; Taylor 1964). For example, on day 21 in the HC-DFB treatment, the concentrations of particulate Al 310 and P were 8.22 and 134.8 nM, respectively (Table S2). Assuming a constant 0.0051 mol Fe: mol P in biogenic 311 particles (Ho 2006) and 0.331 mol Fe: mol Al in lithogenic particles (Taylor 1964; Table 2), we calculated an expected 312 particulate Fe concentration of 3.41 nM, where 80% was associated with lithogenic material and 20% with biogenic 313 material. Similar calculations were made for the bioactive metals Mn, Co, Cu, Zn, Cd, and Mo (Table 2). Our 314 calculations indicate that on average, particulate Fe was dominated by the lithogenic component (accounting for an 315 average of 78% of the total expected particulate Fe), while for particulate Co, Cu, Zn, Cd, and Mo the biogenic fraction 316 dominated (accounting for 94, 95, 99, 94 and 98%, respectively, of the total expected concentration; Table 2). 317 Particulate concentrations of Mn were also dominated by the biogenic fraction (65% of the total), but the lithogenic 318 fraction was also significant (35%). Moreover, the expected particulate Mn and Fe concentrations closely matched the 319 particulate Mn and Fe concentration we measured (accounting for an average of $\sim 71\%$ of the measured Mn, and 115% 320 of the measured Fe). For other metals (i.e. Cu, Mo and Zn), the expected particulate concentrations (nM) were lower 321 than measured (23% of the measured pCu, and 8% of measured pZn; Table 2). This suggests that the particles were 322 enriched in Cu, Mo, and Zn relative to what is expected based on natural marine plankton metal quotas (Bruland et al. 323 1991) and crustal ratios (Taylor 1964).

324

To further establish the lithogenic or biogenic source of the pMe in the particles, the particulate metal concentrations were normalized to the concentrations of particulate P and Al (Figure 2, and Table 2). These ratios were then compared

- 327 with well-known molar ratios of metal to Al in the crust (Taylor 1964) and of metal to P ratios in marine plankton
- 328 samples (Ho 2006) and cultures (Ho et al. 2003) (Table 2). The average Fe: Al (506 mmol Fe: mol Al) and Ti:Al ratios
- 329 (119 mmol Ti: mol Al, Table 2) were relatively similar to crustal molar ratios (331 mmol Fe: mol Al and 39 mmol Ti:
- 330 mol Al; Taylor 1964). Additional evidence for the significant lithogenic component in particulate Ti and Fe was
- 331 gathered from Figure 3, where we plotted the molar ratios of the metals relative to P in the collected particles against
- 332 the Al:P ratios measured in those same particles. The slope of these data [(Fe:P)/(Al:P) = mol Me: mol Al] is the ratio
- of Me:Al in the particles and can be compared to well-known Me:Al crustal ratio. Visually, if the data nicely fit the
- 334 Me:Al line for crustal material, these metals are mainly associated with the lithogenic component, as evident for Fe and
- 335 Ti (Figure 3). These combined results suggest that in our experiment, particulate Fe and Ti concentrations were
- aniched by lithogenic material. In support of this finding, we also found no significant correlation between particulate
- 337 Fe and Ti concentrations (nM) and either the total plankton (phytoplankton and microzooplankton) or *E. huxleyi*
- biomass (μ g C L⁻¹; Table 4).

- 339
- 340 In contrast, when the P-normalized metal ratios in the particles collected from the mesoscosms were plotted against the
- 341 Al:P ratios in these particles, there were no correlations for the following metals Co, Cu, Zn, Cd, Mn and Mo (Figure
- 342 3), indicating that these particulate metals were not enriched in lithogenic material. Our measured metal: P ratios were
- 343 comparable to plankton ratios in natural samples and in cultures (Table 2). The concentrations (mol L⁻¹) of these metals
- 344 (i.e. Cu, Co, Zn, Cd, Mn, Mo), as well as P, also showed significant correlations with the biomass (μ gC L⁻¹) of *E*.
- huxleyi and that of total plankton cells (p < 0.05, Table 4), supporting a significant influence of the phytoplankton in the distribution of these particulate elements.
- 347

348 4.3 Particulate metals with a strong biogenic component: their P-normalized ratios

- 349 The concentrations of particulate bioactive metals (mol L^{-1}), with a significant biogenic component (i.e. excluding Fe) 350 in the studied *E. huxleyi* bloom were ranked as: $Zn > Cu \approx Mn > Mo > Co > Cd$ (Figure 1, Table S3), similar to those 351 reported in indigenous phytoplankton populations: Fe \approx Zn > Cu \approx Mn \gg Co \approx Cd, (Twining and Baines, 2013). The 352 only treatment where E. huxleyi did not dominate the community was the HC-DFB; in this treatment the ranking of 353 these biogenic particulate trace metals was the same as that of LC+DFB (with the massive *E. huxleyi* bloom), but their 354 concentrations were lower than those in LC+DFB. At the end of the experiment, the concentrations of these biogenic 355 metals were, in general, comparable in both HC treatments, and lower than those in the LC treatments (Figure 1, Table 356 S3). Therefore, high CO_2 had a tendency to decrease particulate metal concentrations, especially on day 21. Given the 357 strong correlation between concentrations of these particulate bioactive metals and phytoplankton biomass, the lower 358 particulate concentrations in high CO₂ were mainly due to low phytoplankton biomass.
- 359

360 Particulate Zn concentrations were especially high in the LC+DFB treatment (Figure 1), where the highest E. huxleyi 361 biomass was observed. Emiliania huxleyi is well known for its high Zn cellular requirements (~ 1-10 for E. huxleyi vs. 362 1-4 mmol Zn: mol P for other phytoplankton; Sunda and Hunstman 1995, Sunda 2013). But, the Zn: P ratios in the 363 LC+DFB treatment (range 45-69 mmol Zn: mol P; Figure 2, Table S2), as well as in all the other treatment (range 16-364 34 mmol Zn: mol P; Figure 2, Table S2) were significantly higher than these published ratios. This could be explained 365 by, the adsorption of these metals to the outside of the cells, and/or anthropogenic inputs of Zn into the fjord. The Zn:P 366 ratios in the samples washed with the oxalate-EDTA were still high (range 28-57 for LC+DFB and 16-33 mmol Zn: 367 mol P in all other treatments, Table S3), thus adsorption might have not been significant. We hypothesize that 368 anthropogenic aerosols which are rich in anthropogenic particulate metals, such as Zn and Cu (Perry et al. 1999; Narita 369 et al. 1999), and have high percentage of Zn and Cu dissolution (ref.), might be the source of these high Zn

- 370 concentrations and ratios in the particles.
- 371
- 372 Similarly, the Cu:P ratios in the collected particles were relatively elevated $(1.4 \pm 0.8 \text{ mmol Cu: mol P})$ compared to
- 373 those of other phytoplankton, including *E. huxleyi* (Table 2). The dissolved (7.7±0.41 nM Cu, Figure S3) and
- 374 particulate Cu concentrations (0.35±0.25 nM, Table S2) in our experiment were high, and similar to those previously
- 375 measured in this fjord (Muller et al., 2005). Rain events (or wet deposition of anthropogenic aerosols) in this fjord result
- 376 in high dissolved Cu and the active production of strong organic ligands by *Synechococcus*—to lower the free Cu
- 377 concentrations (Muller et al., 2005). Therefore, high Cu might be a general condition in this fjord due to the rainy nature
- 378 of the geographical location, and indigenous plankton might have developed physiological mechanisms to deal with
- 379 high Cu, such as the production of organic ligands to prevent uptake (Vraspir and Butler, 2009), or of heavy-metal-
- 380 binding peptides (phytochelatins) to lower Cu toxicity inside the cell (Ahner and Morel, 1995; Ahner et al., 1995;

- 381 Knauer et al., 1998). Since we measured high particulate Cu, and Cu:P in our experiment, E. huxleyi might have been
- 382 relying mainly on phytochelatins to buffer high intracellular Cu (Ahner et al., 2002).
- 383

384 The Cd:P ratios (average 0.024 ± 0.01 mmol Cd:mol P, Figure 2) were significantly lower than those in phytoplankton

- and *E. huxleyi* (0.36 mmol Cd:mol P, Figure 2). This was surprising, because Cd quotas are normally higher in
- 386 coccolithophores than in diatoms and chlorophytes (Sunda and Huntsman, 2000; Ho et al., 2003). High Cd quotas in
- 387 coccolithophores have been suggested to result from accidental uptake through Ca transporters and channels (Ho et al.,
- 388 2009). The low Cd quotas here may be explained by the antagonistic interaction between Mn and Cd or Zn and Cd
- 389 under high Mn and Zn, respectively (Sunda and Huntsman, 1998, 2000; Cullen and Sherrell, 2005).
- 390 Since high Zn:P ratios were common in this study $(34.02 \pm 18.05 \text{ mmol Zn:mol P}, \text{Figure 2})$, we hypothesize that high 391 Zn levels antagonistically interacted with Cd, resulting in low Cd:P ratios in the particles.
- 392

393 4.4 The effects of increased CO₂ and the DFB addition on particulate metal concentrations and P-normalized 394 ratios

395 Fe enrichment is common in coastal waters, due to sediment resuspension, rivers input, aeolian deposition and mixing 396 or upwelling of deep water. Indeed, Fe was the essential metal with the highest particulate concentrations in our study 397 (Figure 1, Table 3). Furthermore, in this study particulate Fe was characterized by a strong lithogenic component, and 398 was not correlated with phytoplankton biomass. Fe was also unique, in that it was the only trace element whose 399 particulate concentration was significantly and exclusively affected by CO₂ (no interaction between CO₂ and 400 DFB), regardless of the presence or absence of DFB (Table 1) . Furthermore, particulate Fe concentrations (nM) 401 decreased steadily between days 12 and 21, with the exception of the control treatment (LC-DFB; Figure 1, Table 2S). 402 This suggests that the increase in CO₂ and/or the DFB addition reduce the concentration of pFe, despite the 403 phytoplankton bloom. Such a decrease in pFe (range 2.3-fold in LC-DFB, vs. 13.7-fold in HC+DFB; Table S2) might 404 be mediated by the dissolution of particulate Fe by low pH or by the presence of strong organic chelators as observed in 405 this very experiment (Segovia et al. 2017 and references therein) where dFe notably increased in treatments with high 406 CO₂ and/or the addition of DFB (Figure S3). Furthermore, the dissolution of particulate Fe in the treatments with high 407 CO2 and/or the addition of DFB was evident in the Fe partitioning coefficients—the molar ratio between particulate and 408 dissolved concentrations (Figure 4). On day 21, the Fe partitioning coefficients varied by 22-fold between the highest 409 for the control (LC-DFB: 1.039) and lowest for the HC+DFB treatments (HC+DFB: 0.047; Figure 4). Thus, either the 410 DFB addition or high CO₂ promoted the dissolution of pFe. However, at the end of the experiment, high dFe 411 concentrations were only observed in the treatments with the DFB additions, suggesting that the presence of strong 412 organic Fe chelators, such as DFB, mediated the maintenance of high dissolved Fe concentrations, as previously 413 observed (Segovia et al. 2017). Thus, in our future oceans, high CO₂ (low pH) will increase dissolved Fe concentrations 414 in regions rich in particulate Fe, and in strong organic Fe chelators. The deleterious effects of OA on the development 415 of ecologically important species sensitive to increased CO₂ such as *E. Huxleyi*, will be more relevant in high-Fe

- 416 environments than in Fe-limited ones.
- 417 In contrast to the findings for Fe, particulate Cu concentrations a) were not affected by either high CO₂ or the DFB
- 418 addition; b) were dominated by a biogenic component and c) were significantly correlated with phytoplankton biomass
- 419 (Table 4). Furthermore, unique to Cu was a significant increase in Cu:P ratios by day 21 in the high CO₂ treatments,
- 420 especially when no DFB was added. Since the Cu partitioning coefficients only varied by 3.25 fold among treatments
- 421 on day 21 (LC-DFB: 0.065 vs. HC+DFB: 0.047; data not shown), we hypothesize that high CO₂ did not affect the
- 422 partitioning between particulate and dissolved Cu, but instead, it affected the speciation of dissolved Cu, increasing free

- 423 $Cu(Cu^{2+})$ and thus, its bioavailability. This resulted in the highest Cu:P ratios in the high CO₂ treatments, despite the
- 424 low phytoplankton biomass. This increase in bioavailability under lower pH is typical of metals that form strong
- inorganic complexes with carbonates, such as Cu²⁺ (Millero et al., 2009). Thus in our future oceans, high CO₂ (low pH)
 will shift the speciation of dissolved Cu towards higher abundance of free ionic species, increasing its bioavailability
 and likely its toxicity.
- 428

429 Similarly to Cu, particulate Co, Zn and Mn were correlated with biomass and were dominated by the biogenic 430 component. But in contrast to Cu, these metals particulate concentrations were affect by increased CO₂ and/or the DFB 431 addition. However, the effects of high CO2 and/or DFB were very complex because significant interactions between 432 these 2 factors were observed (Table 1); and further studies are required before we are able to discern and conclude a 433 significant trend. Yet, the P-normalized ratios of Co, Zn and Mn were significantly affected by CO₂ (Table 3), 434 exhibiting moderately lower ratios under high CO₂, when phytoplankton biomass was lowest. These results imply that 435 the bioavailability of these metals was not enhanced under acidic conditions. This suggests that under high CO₂ (low 436 pH) the free ionic species of these metals will not significantly increase in the future, as shown for metals that occur

- 437 predominantly as free ionic species in seawater (Millero et al., 2009).
- 438

439 5. Concluding remarks

- 440 The results presented here show that in the fjord where we carried out the present experiment, particulate Fe was 441 dominated by lithogenic material, and was significantly decreased in the treatments with high CO₂ concentrations 442 and/or the DFB addition. Indeed, high CO2 and/or DFB promoted the dissolution of particulate Fe, and the presence 443 of this strong organic complex helped maintaining high dissolved Fe. This shift between particulate and dissolved 444 Fe, in the presence of DFB, promoted a massive bloom of *E. huxleyi* in the treatments with ambient CO_2 , due to 445 increased dissolved Fe bloom of *E. huxleyi*, the concentrations of particulate metals (mol L^{-1}) with During the . 446 a strong biogenic component (Cu, Co, Zn, Cd, Mn, and Mo) were a) highly dynamic, b) positively correlated with 447 plankton biomass, and c) influenced by growth requirements. Furthermore, high CO₂ decreased the Me:P ratios of Co, 448 Zn and Mn , while increased the Cu:P ratios. In contrast DFB had no effects on these ratios. According to our results, 449 high CO₂ may decrease particulate Fe and increase dissolved Fe, but high concentrations of dissolved Fe will only be 450 maintained by the presence of strong organic ligands. Furthermore, ocean acidification will decrease E. huxleyi 451 abundance, and as a result, the sinking of particulate metals enriched in E. huxlevi. Moreover, the Me:P ratios of metals 452 that are predominately present in an ionic free form in seawater (e.g. Co, Zn and Mn) will likely decrease or stay 453 constant. In contrast, high CO₂ is predicated to shift the speciation of dissolved metals associated with carbonates, such 454 as Cu, increasing their bioavailability, and resulting in higher Me:P ratios. We suggest that high Cu might be a 455 common condition in this fjord, and autochthonous plankton might be able to cope with high Cu levels by developing 456 specific physiological mechanisms. Future high CO₂ levels are expected to change the relative concentrations of 457 particulate and dissolved metals, due to the differential effects of high CO₂ on trace metal solubility, speciation, 458 adsorption and toxicity, as well as on the growth of different phytoplankton taxa, and their elemental trace metal 459 composition.
- 460

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469 **Conflict of interest**

- 470 Authors declare no conflict of interest
- 471
- 472 **References**
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Table 1. Statistical analyses (Split-plot ANOVA) of the effects of high CO₂, the addition of DFB, and their interaction, as well as the effect of time, on the concentrations of particulate metals (mmol L^{-1} , data in Table S2, and Figure 3) in particles collected from the different mesocosms treatments. We used all the days for the analyses because the Split-Plot ANOVA integrates fixed factors (Co2 and Fe) and a repeated measures factor (time) by using the post-hoc Bonferroni, thus, time was fully considered during the whole experimental period.

Factor	Al	Ti	Р	Fe	Cu	Со	Zn	Cd	Mn	Мо	Pb
CO ₂	ns	ns	**	*	ns	**	***	***	**	***	ns
DFB	ns	ns	*	ns	ns	*	**	ns	*	*	ns
CO ₂ x DFB	ns	*	**	ns	ns	*	**	*	**	**	ns
Time	ns	ns	ns	***	*	***	***	***	***	***	**

ns: not significant; * p < 0.05; ** p < 0.01; *** p < 0

Table 2. The average metal ratios in the particles collected in this study (without oxalate wash) using the data reported in Table S2. The P-normalized ratios (mmol : mol P, Figure 4) are compared to previous estimates in marine plankton samples and phytoplankton cultures (A). The Al-normalized ratios (mmol : mol Al) are compared to crustal ratios (B).

(mmol : mol P)	Mn:P	Fe:P	Co:P	Cu:P	Zn:P	Cd:P	Mo:P	Al:P	Reference
Phytoplankton Lab	3.8	7.5	0.19	0.38	0.8	0.21	0.03		Ho et al. 2003
Marine Plankton Field	0.68±0.54	5.1±1.6	0.15±0.06	0.41±0.16	2.1±0.88				Но 2006
E. huxleyi _{Lab}	7.1±0.36	3.5±0.07	0.29±0.02	0.07±0.013	0.38±0.002	0.36±0.01	0.022±0.000	13	Ho et al. 2003
This study	1.65±0.41	39.2±34.3	0.07±0.02	1.41±0.55	34.02±18.05	0.02±0.01	0.42±0.12	70±38	
Crust ratio	510	29,738	13	25	32	0.05	0.46	89,972	Taylor 1964
(mmol : mol Al)	Mn:Al	Fe:Al	Co:Al	Cu:Al	Zn:Al	Cd:Al	Mo:Al	Pb:Al	Ti:Al
· · · · ·									
Crustal ratio	5.7	331	0.14	0.27	0.35	0.001	0.005	0.02	39

A)

B)

Factor	Fe:P	Cu:P	Co:P	Zn:P	Cd:P	Mn:P	Mo:P	Pb:P	Ti:P
CO ₂	ns	*	***	**	ns	*	ns	ns	ns
DFB	ns								
CO ₂ x DFB	ns								
Time	***	***	***	***	ns	ns	ns	ns	***

Table 3. Statistical analyses (Split-plot ANOVA) of the effects of CO₂, DFB, and their interaction, as well as the effect of time, on the P-normalized metal quotas (mmol: mol P, data in Figure 4, and Table S2) in particles collected from the different mesocosm treatments.

ns: not significant; * p < 0.05; ** p < 0.01; *** p < 0.001

		Р	Fe	Cu	Со	Zn	Cd	Mn	Мо	Pb	Ti
E. huxleyi	Correlation coefficient	0.622	ns	0.614	0.756	0.747	0.818	0.686	0.825	ns	ns
	P-value	0.003		0.003	7.35.10-5	1.01.10-4	6.02.10-6	5.93.10-4	4.20.10-6		
Total cells	Correlation coefficient	0.641	ns	0.51	0.644	0.889	0.802	0.598	0.53	ns	ns
	P-value	0.002		0.02	1.62.10-3	7.03.10-8	1.23.10-5	4.18.10-3	1.35.10-2		

Table 4. The relationship (Pearson correlations, p < 0.05) between particulate metals concentrations (nmol L⁻¹, no oxalate wash, reported in Table S2) and the biomass (μ gC L⁻¹) of *Emiliania huxleyi* and total cells (phytoplankton and microzooplankton) collected from the different mesocosm treatments.

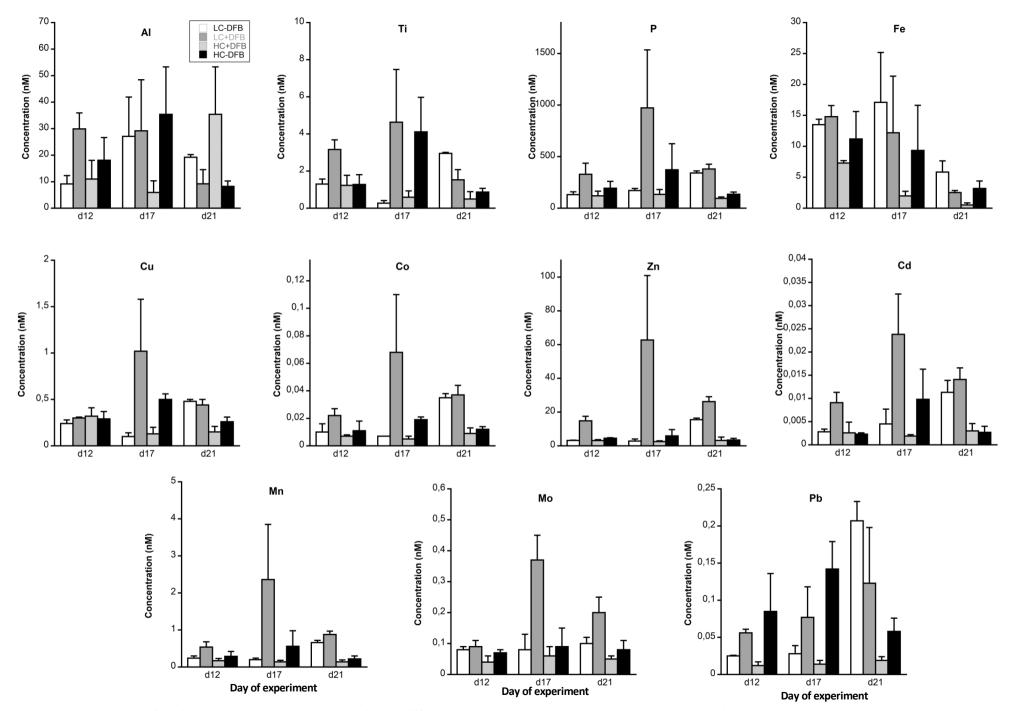


Fig. 1. The concentration of particulate metals in seawater (nM) in the different treatments; LC: ambient CO_2 (390 µatm); HC: increased CO_2 (900 µatm); -DFB (ambient dFe); +DFB (increased dFe) during the development of a bloom of Emiliania huxleyi. Bars are means of measurements in 3 independent mesocosms (n = 3) except for LC–DFB where n = 2. Error bars indicate SD.

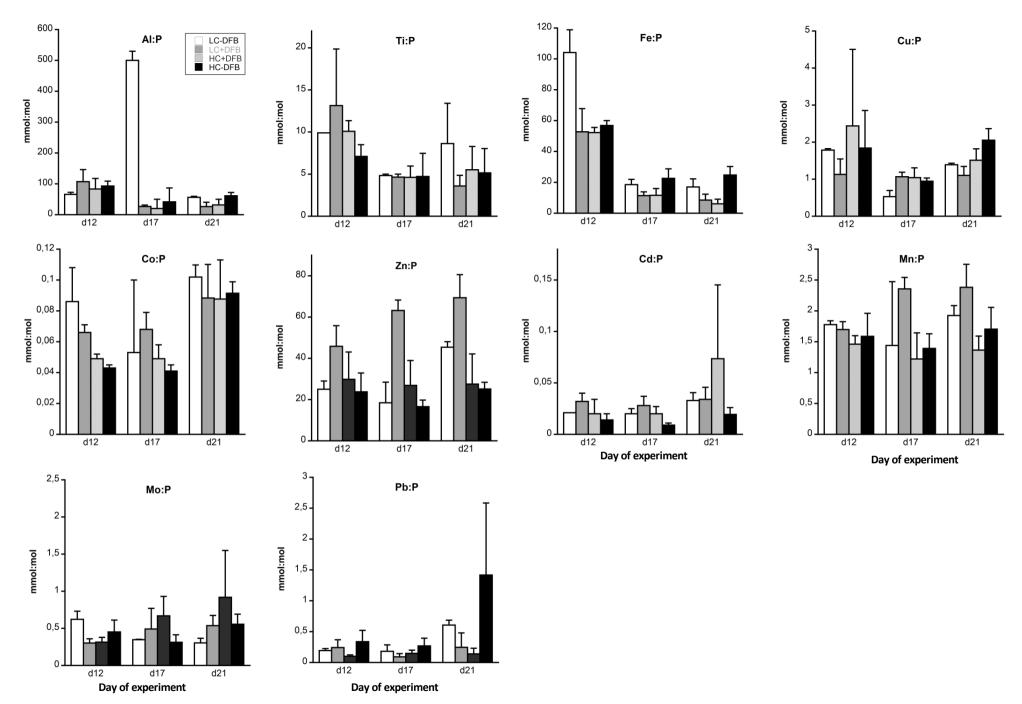


Fig. 2. P-normalized metal quotas (mmol:mol P) of particles from different treatments; LC: ambient CO_2 (390 µatm); HC: increased CO_2 (900 µatm); -DFB (ambient dFe); +DFB (increased dFe) during the development of a bloom of *Emiliania huxleyi*. Bars are means of measurements in 3 independent mesocosms (n = 3) except for LC–DFB where n = 2. Error bars indicate SD.

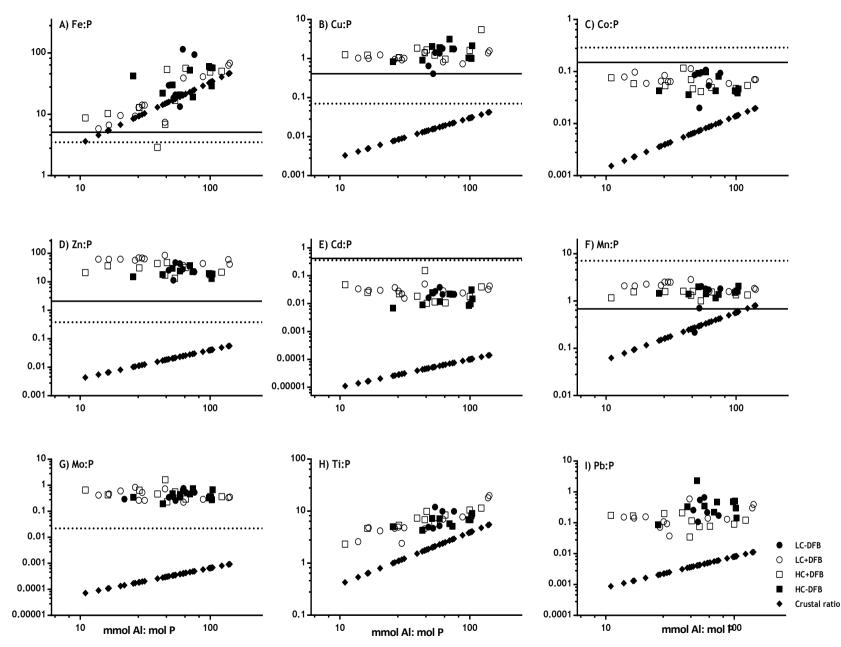


Fig. 3. Comparison of P-normalized metal ratios in particles (mmol:mol P) against mmol Al:mol P ratios in the same particles (without oxalate wash) collected from the different mesocosm treatments (LC: ambient CO_2 ; HC: increased CO_2 (900 µatm); -DFB: no DFB addition; +DFB: with a 70 nM DFB addition) during the development of a bloom on day 12, 17 and 21 (original data reported in Table S2). The x-axis parallel solid and dotted lines represent the average metal quotas obtained from marine plankton assemblages (Ho 2006) and from cultures of *Emiliania huxleyi* (Ho et al. 2003). The slope of the line with the \blacklozenge symbols indicates the average metal : Al (mol:mol) in crustal material (Taylor, 1964). (A) Fe:P, (B) Cu:P, (C) Co:P, (D) Zn:P, (E) Cd:P, (F) Mn:P, (G) Mo:P, (H) Ti:P, (I) Pb:P.

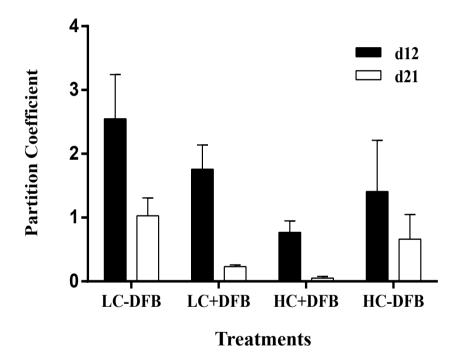


Fig. 4. The Fe partition coefficients (the molar ratio between particulate and dissolved concentrations) in the different mesocosm treatments; LC: ambient CO_2 (390 µatm); HC: increased CO_2 (900 µatm); -DFB: no DFB addition; +DFB: with a 70 nM DFB addition; on day 12 and day 21. . Bars are means of measurements in 3 independent mesocosms (n = 3) except for LC–DFB where n = 2. Error bars indicate SD.