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KINETICS OF CALCITE PRECIPITATION BY UREOLYTIC BACTERIA UNDER AEROBIC AND ANAEROBIC CONDITIONS

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Abstract. The kinetics of urea hydrolysis (ureolysis) and induced calcium carbonate ($CaCO_3$) precipitation for engineering use in the subsurface was investigated under aerobic conditions using *Sporosarcina pasteurii* (ATCC strain 11859) as well as *Bacillus sphaericus* strains 21776 and 21787. All bacterial strains showed ureolytic activity inducing $CaCO_3$ precipitation aerobically. Rate constants not normalized to biomass demonstrated slightly higher rate coefficients for both ureolysis (k_{urea}) and $CaCO_3$ precipitation (k_{precip}) for *B. sphaericus* 21776 (k_{urea} = 0.10 ± 0.03 h^{-1}, k_{precip} = 0.60 ± 0.34 h^{-1}) compared to *S. pasteurii* (k_{urea} = 0.07 ± 0.02 h^{-1}, k_{precip} = 0.25 ± 0.02 h^{-1}). *B. sphaericus* 21787 showed little ureolytic activity but was still capable of inducing some $CaCO_3$ precipitation. Cell growth appeared to be inhibited during the period of $CaCO_3$ precipitation. TEM images suggest this is due to the encasement of cells and was reflected in lower k_{urea} values observed in the presence of dissolved Ca. However, biomass re-growth could be observed after $CaCO_3$ precipitation ceased, which suggests that ureolysis-induced $CaCO_3$ precipitation is not necessarily lethal for the entire population. The kinetics of ureolysis and $CaCO_3$ precipitation with *S. pasteurii* were further analyzed under anaerobic conditions. Rate coefficients obtained in anaerobic environments were comparable to those under aerobic conditions, however no cell growth was observed under anaerobic conditions with NO_3^-, SO_4^{2-} and Fe^{3+} as potential terminal electron acceptors. These data suggest that the initial rates of ureolysis and ureolysis-induced $CaCO_3$ precipitation are not significantly affected by the absence of oxygen but that long-term ureolytic activity might require the addition of suitable electron acceptors. Such variations in the ureolytic capabilities and associated rates of $CaCO_3$ precipitation between strains must be fully considered in subsurface engineering strategies that utilize microbial amendments.

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1. Introduction

Carbonate precipitation is a natural phenomenon which may also be utilized in many subsurface engineering applications (Phillips et al., 2013a) including soil stabilization (van Paassen Leon et al., 2010), immobilization of radionuclides (Mitchell and Ferris, 2005, 2006;Tobler et al., 2012;Warren et al., 2001), and mineral plugging for enhanced oil recovery and carbon sequestration (Dupraz et al., 2009;Ferris et al., 1996;Mitchell et al., 2010;Phillips et al., 2013b). Mineral precipitation can be induced by bacteria as a by-product of common microbial processes, such as urea hydrolysis (ureolysis). In this process, bacteria hydrolyze urea ($CO(NH_2)_2$), an important nitrogen compound found in natural environments) to ammonia (NH_3) and carbonic acid (H_2CO_3) (Equations 1-3). The NH_3 and H_2CO_3 equilibrate in circumneutral aqueous environments to form bicarbonate (HCO_3^-), two ammonium ions (NH_4^+) and one hydroxide ion (OH^-) (Equations 4-5), or at higher pH values to one carbonate ion (CO_3^{2-}) and two NH_4^+ (Equations 4-6). In the presence of dissolved calcium (Ca^{2+}), this increase in carbonate alkalinity shifts the saturation state of the system, allowing for solid calcium carbonate ($CaCO_3$) to form (Equation 7). The overall reaction from the hydrolysis of urea in the presence of Ca^{2+} is summarized by Equation 8.

$$CO(NH_2)_2 + H_2O \rightarrow NH_2COOH + NH_3 \quad (1)$$

$$NH_2COOH + H_2O \rightarrow NH_3 + H_2CO_3 \quad (2)$$

$$CO(NH_2)_2 + 2H_2O \rightarrow 2NH_3 + H_2CO_3 \text{ (Equations 1 + 2 overall)} \quad (3)$$

$$2NH_3 + 2H_2O \leftrightarrow 2NH_4^+ + 2OH^- \quad (4)$$

$$H_2CO_3 \leftrightarrow HCO_3^- + H^+ \quad (5)$$

$$HCO_3^- + H^+ + 2OH^- \leftrightarrow CO_3^{2-} + 2H_2O \quad (6)$$

$$CO_3^{2-} + Ca^{2+} \leftrightarrow CaCO_3 \quad (7)$$

$$CO(NH_2)_2 + 2H_2O + Ca^{2+} \leftrightarrow 2NH_4^+ + CaCO_3 \text{ (Overall process)} \quad (8)$$

The use of ureolytic bacteria in biotechnological applications is appealing for many reasons. Ureolytically active microorganisms are common in a wide variety of soil and aquatic environments, thus, indigenous microorganisms capable of ureolysis can be either stimulated *in situ* or alternatively, they can be used to augment environments lacking ureolytic microorganisms (Fujita et al., 2000;Warren et al., 2001). Urea is a fairly inexpensive substrate and it is often contained in wastewater (Hammes et al., 2003b), so this waste product may be used to stimulate ureolysis in engineering applications (Mitchell et al., 2010). Moreover, the controlled increase of pH and alkalinity in the subsurface by ureolytic bacteria is preferable to the injection of a basic solution (abiotic process), which could lead to instantaneous $CaCO_3$ supersaturation and precipitation at the point of injection limiting the radius of influence of the technology. The injection of urea into the subsurface followed by microbially induced ureolysis would allow for the controlled, gradual ureolysis further away from the injection point, promoting a wider spatial distribution of $CaCO_3$ in the subsurface and avoiding uncontrolled plugging at the point of injection (Cuthbert et al., 2013;Ebigbo et al., 2012;Mitchell and Ferris, 2005;Schultz et al., 2011;Tobler et al., 2012;Tobler et al., 2014).

Among different ureolytic bacteria, *Sporosarcina pasteurii* (formerly known as *Bacillus pasteurii*) has been extensively used as the model urease-producing organism in ureolysis-driven $CaCO_3$ precipitation studies due to its

high ureolytic activity and constitutive production of urease (Phillips et al., 2013). The use of *S. pasteurii* for $CaCO_3$ precipitation is feasible under aerobic conditions and the kinetics of ureolysis under different conditions have been studied. Most studies have reported first order ureolysis rates with respect to urea concentration, with the rate constant (k_{urea}) ranging between 0.002 and 0.090 h^{-1} under aerobic conditions in artificial groundwater without nutrients added (Dupraz et al., 2009;Ferris et al., 2004;Hammes et al., 2003a;Mitchell and Ferris, 2005;Tobler et al., 2012), and 0.35 h^{-1} with the addition of nutrients (Lauchnor et al., 2015). Ureolysis rates have been suggested to be temperature-dependent (Ferris et al., 2004), and it seems to also be affected by cell concentration (inoculum size) (Lauchnor et al., 2015;Tobler et al., 2011).

Although the ureolytic activity of *S. pasteurii* under anoxic conditions has been observed (Martin et al., 2012;Mortensen et al., 2011;Tobler et al., 2012), there is controversy regarding the extent and duration of ureolytic activity that can be achieved in the absence of oxygen. Mortensen et al. (2011) and Tobler et al. (2012) observed extensive ureolytic activity under anoxic conditions, suggesting that the anoxic environment does not inhibit urease activity. Conversely, Martin et al. (2012) observed limited cell growth and poor ureolysis under anoxic conditions and suggested that the ureolytic activity observed was due to the urease already present in the cells.

In this study, the ability of *S. pasteurii* to grow in the absence of oxygen (with or without nitrate (NO_3^-), sulfate (SO_4^{2-}) or ferric ion (Fe^{3+}) as possible electron acceptors) was investigated along with the kinetics of ureolysis and $CaCO_3$ precipitation. Moreover, this study investigates and compares the ureolytic activity of *S. pasteurii* with different strains of *Bacillus sphaericus* under aerobic conditions, which have also been suggested to be capable of ureolysis-induced $CaCO_3$ precipitation (Dick et al., 2006;Hammes et al., 2003a).

2. Materials and methods

2.1. Solutions

Kinetic experiments were carried out using the $CaCO_3$ Mineralizing Medium (CMM) described by Ferris and Stehmeier (1996) (see Supplemental Information [SI], Table SI1.1). Both Ca^{2+} inclusive (CMM+) and Ca^{2+} exclusive (CMM-) versions of this medium were used. Aerobic CMM- was prepared as follows. A double strength solution of nutrient broth was prepared and autoclaved. A nutrient broth was chosen in this experiment to enable cell growth. A separate solution of double strength urea, ammonium chloride, and sodium bicarbonate was prepared and stirred until completely dissolved. These two solutions were combined and adjusted to a pH of 6.0 using concentrated HCl. CMM+ was prepared similarly, but calcium chloride was added after the pH adjustment. Media were filter sterilized into sterile Pyrex bottles using 0.2 µm pore size filters (Nalgene, Rochester, NY). Anaerobic CMM was produced in the same manner. However, all stock solutions were made in an anaerobic chamber using water that had been degassed by stirring overnight in the oxygen-free atmosphere of the chamber (90% N_2, 5% CO_2, 5% H_2). Solutions were filter sterilized into serum bottles and were then capped and sealed inside the chamber. Prior to experiments, the solutions were combined to reach the final concentrations listed in Table SI1.1.

2.2. Bacterial strains and culturing conditions

Three strains of ureolytic bacteria were used: *S. pasteurii* (ATTC 11859), and two isolates from a garden soil and landfill soil, *B. sphaericus* 21776, and *B. sphaericus* 21787 (Belgian Coordinated Collections of Microorganisms, Laboratory of Microbiology, Ghent University) (Hammes et al., 2003b). *Bacillus subtilis* strain 186 (ATCC 23857), a non-ureolytic organism, was used as a control species. Abiotic (i.e. non-inoculated) controls were also set up and run in parallel. Pilot cultures were grown in flasks by adding 100 µL of thawed stock to 100 mL of autoclaved Brain Heart Infusion (BHI) + 2% urea. *S. pasteurii* and *B. sphaericus* were grown on an incubator shaker (New Brunswick Scientific, Edison, NJ) at 30°C and 150 rpm, while *B. subtilis* was grown on an incubator shaker at 37°C and 150 rpm. 100 µL of pilot cultures were transferred at 24 h and 48 h to new flasks containing 100 mL of BHI + 2% urea.

2.3. Aerobic experiments

Once the pilot cultures were ready for inoculation, 40 mL of culture were added to a 50 mL centrifuge tube. This tube was centrifuged at 4303 × *g* using a Sorvall Instruments (Asheville, NC) RC-5C centrifuge for 10 min at 4-6°C. The supernatant was poured off the cell pellet, and it was re-suspended using about 40 mL of CMM-, and again centrifuged for 10 min. This process was repeated once more. After the third run in the centrifuge, the supernatant was poured off and enough CMM- was added to achieve a final optical density reading at 600 nm (OD_{600}) of 0.4 (measured in a 96 well plate using a BioTek Synergy HT plate reader). 1 mL of prepared *S. pasteurii*, *B. sphaericus* strain 21776 or strain 21787, culture was inoculated in 250 mL Pyrex bottles with 150 mL of media (either CMM+ or CMM-) (initial concentration of biomass OD_{600} = ~ 0.015). After inoculation, the systems were statically incubated at 30°C for kinetic experiments.

2.4. Anaerobic experiments

Pilot cultures for anaerobic experiments were limited to the use of *S. pasteurii* and were grown in the same manner as for those used in aerobic experiments. However, cells were transferred into an anaerobic chamber and re-suspended in anaerobically prepared CMM-, then transferred to a serum bottle, sealed and crimped inside the anaerobic chamber. Optical density measurements for time zero were taken after the final suspension, with the same initial density as the aerobic experiments (OD_{600} = ~ 0.015). The first set of experiments investigated cell growth and ureolysis under oxygen-free conditions with a range of potential terminal electron acceptors (TEAs). Experiments were run using a batch system, consisting of 100 mL of CMM- media including 10 mM NO_3^-, SO_4^{2-}, or Fe^{3+} as potential TEAs and inoculated with 1 mL of *S. pasteurii* in 150 mL serum bottles. Concentrated stock solutions of each terminal electron acceptor were made in the anaerobic chamber and filter sterilized: *i)* a concentrated NO_3^- solution was made using 1M $NaNO_3$; *ii)* a concentrated SO_4^{2-} solution was made by combining 1M Na_2SO_4 and 1M Na_2S, Na_2S was added to quench any residual oxygen and make SO_4^{2-} reduction possible; and *iii)* a stock solution of Fe(III) citrate was made using 50 mM Fe(III) citrate as previously described (Gerlach et al., 2011). Appropriate amounts of each stock solution were added to the separate serum bottles containing CMM- and *S. pasteurii*. The growth survey was also conducted in CMM- without the addition of a TEA. After inoculation, the systems were statically incubated at 30°C. Abiotic control experiments, without the inclusion of *S. pasteurii*, were also performed.

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Aliquots were extracted from the systems in the anaerobic chamber and monitored for pH and OD_{600} during the duration of the experiments. Aerobic control experiments were also performed with CMM- media including 10 mM NO_3^-, SO_4^{2-}, or Fe^{3+} and inoculated with 1 mL of *S. pasteurii* in 150 mL serum bottles.

The second set of experiments investigated the detailed kinetics of ureolysis and $CaCO_3$ precipitation with *S. pasteurii* and CMM+ as described above, with NO_3^- as the potential TEA, by monitoring pH, dissolved Ca^{2+} and NH_4^+ concentrations. NO_3^- was chosen as it demonstrated higher growth at early stages and a modest increase in pH compared to the other TEAs from the first set of experiments. Control experiments were also performed with CMM+ without the addition of a TEA and CMM- with NO_3^- as a potential TEA. Here, a stock solution of 10 M $NaNO_3$ was made by mixing and filter sterilizing in the anaerobic chamber, and an appropriate amount was added to the CMM+ or CMM- to reach a final concentration of 1M NO_3^-.

2.5. Experimental sampling and analysis

At different time points, 3 mL of sample were aseptically extracted from the systems and measurements were made of pH, biomass, NH_4^+ and Ca^{2+} concentration. NH_4^+ concentrations were determined using the Nessler Assay (Mitchell and Ferris, 2005). Ca^{2+} concentrations were measured after appropriate dilution in trace-metal grade 5% HNO_3 (Fisher Scientific) using an Agilent 7500ce Inductively Coupled Plasma Mass Spectrometer (ICP-MS). Bacterial biomass was determined using three methods: plate counts, OD_{600} and protein assays. OD_{600} was used as a growth indicator in experiments carried out in the absence of Ca^{2+} (see SI section 1.2). Transmission Electron Microscopy (TEM) images were taken using a LEO 912AB TEM and photographed with a Proscan 2048x2048 CCD camera from a batch culture in CMM+ inoculated with *S. pasteurii*. At the point of crystal formation (after approximately 2.5 h), a mixture of $CaCO_3$ crystals and cells were extracted from the system. Separate samples of *S. pasteurii* grown in the absence of Ca^{2+} were also collected and imaged. Further details are given in the SI section 1.3.

The PHREEQC (version 2) speciation-solubility geochemical model (Parkhust and Appelo, 1999) was used to calculate solution speciation and carbonate mineral saturation. Simulation was performed using calcite -$CaCO_3$- as the only precipitate as it was identified by XRD (data not shown) as the calcium carbonate polymorph present in the systems. The MINTEQ database was used for all calculations and the thermodynamic constants for urea (Stokes, 1967) were added (more information is provided in SI1.4).

2.6. Kinetics of ureolysis and $CaCO_3$ precipitation

The rate coefficient for ureolysis was determined by integrating the following first order differential equation assuming constant biomass concentrations during the period of urea hydrolysis (Ferris et al., 2004; Mitchell and Ferris, 2005):

$$\frac{d[Urea]}{dt} = -k_{urea}[Urea][X] \quad (9)$$

to get:

$$[Urea]_t = [Urea]_o e^{-k_{urea}Xt} \quad (10)$$

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where k_{urea} is the first order rate coefficient for ureolysis, t is the time, and X is the concentration of biomass (SI, section SI2.1). The change in urea concentration was determined according to Equations 3 and 4 (Equation 11).

$$\Delta[Urea] = -0.5 * \Delta[NH_4^+] \quad (11)$$

Ureolysis rates were calculated in two ways. Firstly, it was assumed that the reaction is zero order with respect to biomass ($X = 0$), as performed in other studies of ureolysis kinetics (Cuthbert et al., 2012;Ferris et al., 2004;Mitchell and Ferris, 2005, 2006;Schultz et al., 2011;Tobler et al., 2011), thus k_{urea} rates were not normalized to biomass. Secondly, k_{urea} rates were normalized to the biomass concentration at the onset of precipitation (normalized to the absorbance reading of initial biomass, OD_{600}, and CFU mL^{-1}; SI section 2.2), which was equivalent to the initial biomass in each system ($X = X_0$). This is an appropriate choice of model, since the biomass analysis indicated that the cell density was constant for the duration of $CaCO_3$ precipitation and was equivalent to the initial biomass in the systems, as presented in the results section. The kinetic parameters obtained in this study (k_{urea} normalized to biomass) were compared to other parameters previously published (Ferris et al., 2004;Fujita et al., 2000;Stocks-Fischer et al., 1999;Tobler et al., 2011). The media used in the different studies were similar to those used in this study, all with 25 mM of Ca^{2+}, 333 mM urea and including nutrient broth-based growth media, apart from: *i)* Ferris et al. (2004) who used a dilute artificial groundwater (non-growth medium) with Ca^{2+} and urea concentrations of 1.75 mM and 6 mM, respectively, and *ii)* Tobler et al. (2011) who used different Ca^{2+} concentrations varying from 50 to 500 mM and urea concentrations between 250 and 500 mM.

The precipitation of $CaCO_3$ from the system is dependent on the saturation state of the system, as well as the growth mechanism of $CaCO_3$ (Ferris et al., 2004;Mitchell and Ferris, 2005;Teng et al., 2000). The literature is ambiguous on defining a set rate expression for $CaCO_3$ precipitation, so a non-affinity-based first order rate law was applied to these studies for both its simplicity and the fact that it seems to describe the data well, assuming that for every mole of Ca^{2+} removed from solution one mole of $CaCO_3$ formed (Teng et al., 2000):

$$\frac{d[Ca^{2+}]}{dt} = -k_{precip}[Ca^{2+}] \quad (12)$$

Integration of the above equation yields:

$$[Ca^{2+}]_t = [Ca^{2+}]_o e^{-k_{precip}t} \quad (13)$$

where k_{precip} is the first order rate coefficient for $CaCO_3$ precipitation. Rate constants were found using a non-linear regression method utilizing the Solver function in Microsoft Excel. Due to the significant lag phase, the data used for analysis excluded onset time before ureolysis and $CaCO_3$ precipitation occurred, as previously documented (Tobler et al., 2011).

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3. Results and Discussion

3.1. Aerobic experiments

3.1.1 Solution chemistry

Aerobic experiments with CMM+ medium inoculated with *S. pasteurii*, *B. sphaericus* 21776 and *B. sphaericus* 21787 showed an increase in pH (Table 1) and NH_4^+ (displayed as a stoichiometrically equivalent decrease in urea concentrations, Figure 1) over time. These results support observations from previous studies confirming the ureolytic capabilities of *S. pasteurii* (Ferris et al., 2004;Fujita et al., 2000;Warren et al., 2001) and *B. sphaericus* species (Dick et al., 2006;Hammes et al., 2003a). Differences in the rate of pH change and the amount of urea hydrolyzed between the different bacterial species suggest differences in their ureolytic activity. After 30 h, 58-82% and 72-80% of the available urea was hydrolyzed by *S. pasteurii* and *B. sphaericus* 21776, respectively. *B. sphaericus* 21787 exhibited little utilization of urea (12-15% hydrolyzed) accompanied by only more slight increase in pH values (~ pH 8.7 by 24 ± 3 h) compared to the other bacterial strains [~ pH 9.3 by 24 ± 3 h, consistent with buffering by $NH_4^+ \leftrightarrow NH_3 + H^+$ which has a pK_a value of 9.3 at 30°C (Mitchell and Ferris, 2005)]. Control experiments, inoculated with the non-ureolytic organism *B. subtilis* and sterile controls, did not exhibit significant changes in pH (Table 1) or urea concentrations (data not shown). Geochemical modelling suggested that no $CaCO_3$ precipitation should occur in the absence of ureolysis and that approximately 25.5 mM of urea would have had to be hydrolyzed to achieve supersaturation and for $CaCO_3$ precipitation to commence (see SI1.4).

In all the experiments containing ureolytic bacteria, Ca^{2+} concentration decreased to ~ 5% of the initial Ca^{2+} concentration in the liquid medium after approximately 30 h (Figure 1). The decrease of Ca^{2+} concentrations suggests the precipitation of $CaCO_3$, which was identified by XRD (data not shown) as the polymorph calcite. The onset of $CaCO_3$ precipitation occurred shortly after the start of the experiment (~3-4 h) in the *S. pasteurii* and *B. sphaericus* 21776 experiments, whereas the onset of precipitation was delayed (~ 9 h) for *B. sphaericus* 21787. This supports differences in the rate of urea hydrolysis, and thus the time at which $CaCO_3$ saturation was exceeded (Equations 2, 4, 5 and 7). Differences in ureolysis and $CaCO_3$ precipitation rates can be attributed to differences in the specific ureolytic activities of the organisms or the number of ureolytically active cells. It has been suggested that the inherent variation between different organisms' ability to produce the urease enzyme should affect the rates of ureolysis and associated rates of $CaCO_3$ precipitation (Anbu et al., 2016), as well as the characteristics of the resulting $CaCO_3$ precipitates (Hammes et al., 2003a).

3.1.2 Aerobic bacterial growth and ureolytic activity

Changes in biomass, measured as protein and colony forming units (CFUs), were observed during ureolysis in both aerobic CMM+ and CMM- experiments (Figure 2). CFU and protein concentrations exhibited similar trends for *S. pasteurii* and *B. sphaericus* 21776 where in CMM- experiments, CFUs and protein increased over time asymptotically (Figure 2). In CMM+ experiments, CFUs and protein concentrations seemed to slightly decrease or remain quasi-constant while $CaCO_3$ precipitation occurred (< 10 h), followed by an increase in CFUs and protein once Ca^{2+} had been depleted (Figures 1 and 2). Decrease of biomass growth during $CaCO_3$ precipitation has been suggested to occur

due to the encasement of bacteria within the $CaCO_3$ precipitates (Tobler et al. 2011). Encasement of *S. pasteurii* cells in $CaCO_3$ minerals has been reported (Cuthbert et al., 2012;Ebigbo et al., 2012;Schultz et al., 2011;Stocks-Fischer et al., 1999) and cell indentations in $CaCO_3$ precipitates have been observed (Mitchell and Ferris, 2005). The recovery (*i.e.* re-growth) of biomass after $CaCO_3$ precipitation suggests that ureolysis-induced $CaCO_3$ precipitation does not have to be a lethal event for the population as a whole, and that net cell growth can resume after $CaCO_3$ precipitation ceases (Figure 2). For both *B. sphaericus* strains, in contrast to *S. pasteurii*, CFUs were higher in the CMM+ experiments, despite protein concentrations being lower in the CMM+ experiments. This might suggest that cell mortality of *B. sphaericus* strains is increased in the calcium-free experiments, which may reflect lower tolerance to the higher pH values generally observed in the calcium-free experiments (Table 1, Figure 2).

TEM images and electron energy loss spectroscopy (EELS) of material collected on 0.2 μm pore size filters from the CMM+ *S. pasteurii* systems (Figure 3A-C) confirm that some cells are surrounded by a layer of calcium-containing precipitates. Figure 3D shows *S. pasteurii* grown in CMM- for comparison. The data suggest that cells are removed from suspension and potentially inactivated by $CaCO_3$ encasement, either in large crystals (Mitchell and Ferris, 2005) or by a thin coating (Figure 3A-C).

Quasi-constant biomass concentrations during $CaCO_3$ precipitation (Figure 2) suggest that cell growth might not have to be considered in kinetic descriptions of bacterially induced $CaCO_3$ precipitation. However, it is unclear whether *(i)* $CaCO_3$-encased cells are ureolytically active or *(ii)* $CaCO_3$ precipitates surrounding the cells effectively act as a barrier to urea reaching the cell or to NH_3, OH^-, or NH_4^+ formed by the hydrolysis of urea from diffusing through the $CaCO_3$ to the bulk solution. Therefore, a theoretical analysis of urea diffusion in $CaCO_3$ was performed. The diffusion of oxygen in $CaCO_3$ at high temperatures has been documented (Farver, 1994), but, to the best of our knowledge, information of urea diffusion in $CaCO_3$ at 30°C has not been reported. A number of assumptions were made for the estimates in this study: *(1)* since urea has a lower diffusion coefficient than oxygen in aqueous solutions at 25°C (Stewart, 2003), it was assumed that this will hold true at other temperatures and through other substances, like $CaCO_3$; *(2)* since the diffusion coefficient of oxygen through $CaCO_3$ at 400°C and 100 MPa is 2.66×10^{-22} m^2 s^{-1} (Farver, 1994), and diffusion coefficients generally increase with increasing temperature and pressure, it can be assumed that the diffusion coefficient of urea in $CaCO_3$ at atmospheric pressure and 30°C is smaller than 2.66×10^{-22} m^2 s^{-1}; *(3)* assuming that the geometry of the $CaCO_3$ is a uniformly thick slab with a thickness of approximately 200 nm, as determined from the TEM images (Figure 3B), the time it will take to reach 5% of the bulk urea concentration can be calculated using the relation presented by (Carslaw and Jaeger, 1959):

$$t_5 = 0.1\frac{L^2}{D_e} \quad (13)$$

where L is the slab thickness, D_e is the (estimated) effective diffusion coefficient in $CaCO_3$, and t_5 is the amount of time it will take to reach 5% of the bulk concentration. Using the above assumptions, it would take at least 175 days for 5% of the urea to diffuse through the $CaCO_3$ surrounding the cells. Because $CaCO_3$ precipitation takes place over the course of approximately one day, it can safely be assumed that even if the encased cells are still alive, urea is not able to diffuse through the $CaCO_3$ fast enough for them to hydrolyze significant amounts and contribute to the increase in solution alkalinity. Therefore, it is argued that, at least in the systems described here, cell growth does not have to

be considered in kinetic expressions describing ureolysis during $CaCO_3$ precipitation. Instead, the biomass at the onset of precipitation, which is equivalent to the initial biomass in the system (Figure 2), can be used to normalize the observed ureolysis rates to biomass concentration.

While a physical association between cells and $CaCO_3$ precipitates is evident, precipitation is likely to occur from a combination of *(i)* homogeneous nucleation in the bulk solution, in alkaline microenvironments around bacterial cells (Schultze-Lam et al., 1996;Stocks-Fischer et al., 1999), and *(ii)* heterogeneous nucleation on nascent crystals, bottle walls and the bacterial cell surfaces (Rodriguez-Navarro et al., 2012).

3.1.3 Kinetics of ureolysis and $CaCO_3$ precipitation

Kinetic analyses were performed on the individual CMM+ and CMM- experimental data for all bacterial strains (Figure SI2.1). A summary of the parameters estimated is shown in Table 2 (for detailed results on individual experiments see Table SI2.1). k_{urea} values for the bacterial species varied according to the presence (CMM+) or absence (CMM-) of Ca^{2+} in the medium. *S. pasteurii* and *B. sphaericus* 21776 exhibited similar k_{urea} values in CMM- and CMM+ systems with k_{urea} values being between 1.6 and 2.5 times higher in the absence of Ca^{2+} (*S. pasteurii*: $k_{urea,CMM+}$ = 0.07 ± 0.02 h^{-1}, $k_{urea,CMM-}$ = 0.19 ± 0.10 h^{-1}; *B. sphaericus* 21776: $k_{urea,CMM+}$= 0.10 ± 0.03 h^{-1}, $k_{urea,CMM-}$ = 0.16 ± 0.05 h^{-1}). This is likely due to the encasement of cells by $CaCO_3$ and their inactivation in CMM+ experiments. Some data points were excluded for *S. pasteurii* CMM- k_{urea} calculations because of an estimated increase in urea concentration (based on a decrease in NH_4^+ concentration) likely due to significant volatilization of $NH_4^+ \leftrightarrow NH_3 + H^+$ that can occur at pH > 9 (at 34 h; open marker) (Figure SI2.1B). *B. sphaericus* 21787 exhibited low k_{urea} values in both CMM+ (k_{urea} = 0.02 h^{-1}) and CMM- (k_{urea} = 0.05 h^{-1}). From triplicate experiments, only one experiment showed values that could be used for kinetic analysis (Figure SI2.1), and some outlying data points were not used for the kinetic calculations, hence no standard deviations can be provided (Table 2). Thus, rate coefficients obtained for *B. sphaericus* 21787 are not statistically valid but were estimated for the purpose of comparison to the other studied bacterial strains. The obtained results suggest that *B. sphaericus* 21787 exhibits limited ureolytic capabilities under the experimental conditions used in this study.

A lag time before the onset of ureolysis was observed for all bacterial strains. *S. pasteurii* exhibited a lag time of ~4 h in both CMM+ and CMM-, and *B. sphaericus* 21776 ~5 h in CMM+ and ~3 h in CMM-. Connolly et al. (2013) observed a lag time to the onset of ureolysis of ~15 h for *S. pasteurii*, ~6 h for *Pseudomonas aeruginosa* MJK1, and ~4 h for *Escherichia coli* MJK2, when cultivated with 0.16 mM urea in similarly composed CMM-. In the present study, k_{urea} values normalized to the initial biomass concentration showed similar trends for *S. pasteurii* and *B. sphaericus* 21776, suggesting similar cell specific urease activity between the strains. The standard deviations of k_{urea}, initial biomass and lag time were small between replicate experiments with *S. pasteurii* and *B. sphaericus* 21776, and R^2 values for the fit to Equation 10 were greater than 0.9 (Table SI2.1).

The kinetic parameters obtained in this study were compared to other parameters previously published (Tobler et al., 2011; Ferris et al., 2004; Fujita et al., 2000; Stocks-Fischer et al., 1999). k_{urea} values of *S. pasteurii* obtained in this study as well as in previous publications were standardized to the initial cell concentrations (Table 3). Values of k_{urea} were higher in the present study for both *S. pasteurii* and *B. sphaericus* 21776 (k_{urea} = 0.07 h^{-1} and 0.11 h^{-1},

respectively) than those from *S. pasteurii* in other studies (k_{urea} = 0.008 to 0.028 h^{-1}), except for the results obtained by Tobler et al. (2011) (k_{urea}= 0.13 to 2.29 h^{-1}). This was also apparent once normalized to biomass (this study, *B. sphaericus* 21776 k_{urea} = 7.64 OD_{600}^{-1} h^{-1} and *S. pasteurii* 5.28 OD_{600}^{-1} h^{-1}, compared to k_{urea} = 0.11 to 2.80 OD_{600}^{-1} h^{-1} in previous studies, and 4.33 to 32.71 OD_{600}^{-1} h^{-1} for Tobler et al. 2011). The generally higher k_{urea} values in this study appear to reflect the higher temperature (30°C) used compared to the previous studies, which ranged from 20-25°C. Higher temperatures generally increase reaction rates where chemical reactions are advanced through a transient activated complex (Stumm and Morgan, 1996). In urease, the transitional state involves coordination of urea and water at the active catalytic site of the enzyme (Jabri et al., 1995). Formation of such an activated complex tends to impart a greater temperature dependency on the absolute reaction rate than would be encountered if the reactions were mediated solely by collisions arising from molecular diffusion (Ferris et al., 2004;Mitchell and Ferris, 2005).

The biomass concentration-normalized k_{urea} values from Fujita et al. (2000) are much lower. This could be due to the highest biomass concentrations (OD_{600} = 0.072) used in these studies; very high biomass concentrations could shift the primary kinetic dependency from being catalyst (i.e. enzyme limited) to substrate limited. However, this appears to be opposite to what Tobler et al. (2011) reported indicating k_{urea} increased with increasing inoculum density. The rate constants obtained with the three organisms used in the present study are similar to the range of values measured in deeper vadose zone mineral subsoils which were between 0.00375 h^{-1} to 0.07 h^{-1} (Swensen and Bakken, 1998), suggesting natural levels of ureolytic bacterial activity were reasonably approximated in the aerobic experiments.

On average, *B. sphaericus* 21776 had the highest k_{precip} (0.60 ± 0.34 h^{-1}), although considering its high standard deviation, k_{precip} for *S. pasteurii* is not significantly different (k_{precip} = 0.25 ± 0.02 h^{-1}). R^2 values of the fit to Equation 12 were relatively high (0.84 – 0.93) for *B. sphaericus* 21776 and *S. pasteurii* (Table 2). The lag time for $CaCO_3$ precipitation was 3.3 h for all bacterial strains, which reflects the similar k_{urea} values, and thus the time it took to reach $CaCO_3$ saturation and induce precipitation. Tobler et al. (2011) observed similar lag times until the onset of $CaCO_3$ precipitation (2-3 h) in aerobic experiments (artificial groundwater with no nutrients added, 250-500 mM urea and 50-00 mM Ca) with *S. pasteurii*. First order rate constants for $CaCO_3$ precipitation observed here were also higher (0.21 h^{-1}<k_{precip}<0.60 h^{-1}) compared to other studies (0.01 h^{-1}<k_{precip}<0.11 h^{-1}) (Table 3). This is likely associated with the greater k_{urea} values observed in this study than in previous studies. Temperature is unlikely to account for this variation given the modest decrease in calcite solubility (~ 27 %) that occurs between 20°C and 30°C (Miller, 1952;Stumm and Morgan, 1996). Overall, k_{urea} values are lower than k_{precip} values in this and previous studies, with the exception of Ferris et al. (2004), indicating urea hydrolysis is the rate limiting step during ureolysis-induced $CaCO_3$ precipitation and that $CaCO_3$ precipitation rates are rapid once the critical supersaturation is exceeded (Mitchell and Ferris, 2006).

3.2. Anaerobic experiments

3.2.1 Anaerobic ureolysis and bacterial growth

Given the potential for anaerobic conditions in subsurface environments, screening experiments were performed to assess the capability of *S. pasteurii* to grow and/or increase pH in CMM- in the presence of NO_3^-, SO_4^{2-}, and Fe^{3+} as potential TEAs. There are contradicting reports in the literature regarding *S. pasteurii*'s ability to grow

and hydrolyze urea in the absence of oxygen; some studies suggest that the anoxic environment does not hinder urease activity (Mortensen et al., 2011;Tobler et al., 2011), whereas other studies report limited microbial growth and poor ureolysis (Martin et al., 2012).

In this study, pH increased (pH > 9.0) in all experiments with and without potential TEAs added (Figure 4), suggesting ureolysis by *S. pasteurii* took place in the absence of oxygen. The pH of the abiotic controls did not exceed pH 7, except for the medium containing SO_4^{2-} as the added TEA, which had an initial pH of 7.7, which remained constant throughout the experiments (Figure 4C). Growth in the CMM- anaerobic experiments quantified by OD_{600} absorbance was lower than that observed in aerobic experiments regardless of the presence or absence of TEAs (Figure 4). Although some growth might have occurred during the initial period of the experiments (increased absorbance by ~20 h), the lack of sustained growth over time suggests the inability of *S. pasteurii* to grow under anaerobic conditions. These findings are in agreement with those by Martin et al. (2012), who observed limited cell growth in the absence of oxygen. In the present study, once oxygen was allowed to diffuse into the systems (at 120 h), optical density in all inoculated systems increased, indicating that even though no significant growth was observed in the absence of oxygen, the bacteria were still viable after 120 h of oxygen depletion and can be resuscitated.

Sustained growth of *S. pasteurii* in the absence of oxygen does not appear to be feasible which might limit the potential use of *S. pasteurii* for inducing $CaCO_3$ precipitation in the subsurface to only short-term purposes. However, the potential regrowth of microbes, even after prolonged periods of exposure to oxygen-free conditions, suggests that *S. pasteurii* could be resuscitated and re-stimulated through the injection of oxygenated fluids, which could enable bacterial growth and thus ureolytic activity over longer periods of time.

3.2.2 Kinetics of anaerobic ureolysis and $CaCO_3$ precipitation

After the screening experiments with different TEAs, studies were performed to determine the kinetics of ureolysis and $CaCO_3$ precipitation in the absence of oxygen. For these in-depth studies, only NO_3^- was used as a potential TEA since slight growth and a modest increase in pH were observed in the anaerobic ureolysis and bacterial growth studies (Figure 4A). Average change in urea and Ca^{2+} concentrations for CMM+ and CMM- with NO_3^- (Figure 5), and CMM+ without TEA added are presented (Figure 5). Urea was hydrolyzed under all experimental conditions and $CaCO_3$ precipitation was observed in the presence of Ca^{2+} (Figure 5). However, ureolysis and $CaCO_3$ precipitation did not occur in all replicates for each experiment, which accounts for the high standard deviations.

Rate coefficients were estimated as previously described for aerobic experiments (Figure SI2.2) and a summary of the results is presented in Table 2 (for detailed results for individual experiments see Table SI2.2). Data points which preceded the onset and completion of ureolysis and calcite precipitation were excluded (Figure SI2.2), k_{urea} seems to be lowest in CMM+ with NO_3^- (k_{urea} = 0.04 ± 0.01 h^{-1}), followed by CMM- with NO_3^- (k_{urea} = 0.07 ± 0.01 h^{-1}) and CMM+ without TEA (k_{urea} = 0.08 h^{-1}). The same relative rates were apparent when k_{urea} values were normalized to biomass (Table 2). The presence of NO_3^- appears to have slightly decreased ureolysis rates. While the reasons for this possible decrease in the ureolytic activity due to the presence of NO_3^- are unclear, it could be perceivable that the ureolytic activity is down-regulated in the presence of the alternative nitrogen source NO_3^-. Longer lag times to the onset of ureolysis (ranging from 6.5 to 10 h) were observed in the anaerobic experiments relative to the aerobic ones.

Moreover, comparing the experiments containing NO_3^-, the presence of Ca^{2+} in the medium results in a lower k_{urea} value (Table 2). This could be due to the encasement of cells by $CaCO_3$.

The onset of $CaCO_3$ precipitation occurred after approximately 6 h, which was twice the lag time observed for precipitation under aerobic conditions (Table 2); this was expected as slower ureolysis was observed in the anaerobic experiments (Table 2). Rate constants (k_{precip}) for anaerobic $CaCO_3$ precipitation were not statistically different for CMM+ medium with NO_3^- (0.36 h^{-1} ± 0.22) and CMM+ medium without TEA (0.19 h^{-1} ± 0.05). Tobler et al. (2011) reported a similar k_{urea} value (0.09 h^{-1}) for experiments with *S. pasteurii* under anoxic conditions in natural groundwater containing comparable concentrations of urea and Ca^{2+} (250 mM and 50 mM, respectively); k_{precip} was not reported due to a poor fit of the data with first order kinetics. Experiments containing only indigenous bacteria exhibited far lower rates of ureolysis (k_{urea} = 0.0016 h^{-1}) and $CaCO_3$ precipitation (k_{precip} = 0.009 h^{-1}) than observed in the present study. Differences in ureolysis and $CaCO_3$ precipitation rates between this study and Tobler et al.'s (2011) study is likely due to the low initial biomass of the indigenous ureolytic population in the natural groundwater.

Comparison of kinetics from aerobic and anaerobic experiments in the present study demonstrates that rates are on the same order of magnitude (Table 2). In CMM+, urea hydrolysis rates for *S. pasteurii* under aerobic and anaerobic conditions (with or without TEA) were not significantly different (P_{value}=0.274), even when normalized to initial biomass concentrations (OD_{600} or CFU mL^{-1}). pH increases in the screening experiments suggest anaerobic ureolysis occurred at the same rate as under aerobic conditions (Figure 4). Similarly, k_{precip} in aerobic and anaerobic experiments are comparable (0.19 h^{-1} and 0.25 h^{-1}, respectively). This suggests that oxygen-free environments do not significantly impact the rate of ureolysis or $CaCO_3$ precipitation initially, but that anaerobic growth cannot be conclusively demonstrated under the conditions of this present study. This supports observations by Tobler et al. (2011), who reported similar rates of ureolysis under both oxic and anoxic conditions when amending natural groundwater with *S. pasteurii* (k_{urea} 0.10 h^{-1} in oxic conditions, k_{urea} 0.09 h^{-1} in anoxic conditions with 50 mM Ca^{2+} and 250 mM urea, cf. Table 3). Martin et al. (2012) also observed ureolytic activity by *S. pasteurii* under anoxic conditions but to a lesser extent compared to the extensive activity reported by Tobler et al. (2011), however rate constants were not reported. The current study therefore suggests ureolytic activity observed under anoxic conditions corresponds to the urease already present in the cells as suggested by Martin et al (2012). *S. pasteurii* could therefore potentially be used for $CaCO_3$-induced precipitation in the subsurface in the short-term, and the bacterial growth could be stimulated through multiple injection of bacterial cells or oxygenated medium to re-enable ureolytic activity and thus $CaCO_3$ precipitation. This is supported by our calcite precipitation rate constants under anaerobic conditions, the first to be reported for a ureolytic strain, which are comparable to aerobic rate constants, suggesting anaerobic conditions will not significantly inhibit $CaCO_3$-induced precipitation in the subsurface.

4. Conclusions

All three ureolytic strains studied, *S. pasteurii as well as B. sphaericus* strains 21776 and 21787, were capable of inducing $CaCO_3$ precipitation under aerobic conditions. Data obtained in this study suggest that rates of ureolysis and ureolysis-induced $CaCO_3$ precipitation are affected by differences in the ureolytic species. This information should be considered in subsurface engineering strategies utilizing microbial amendment or stimulation. Specifically, rates

of ureolysis and $CaCO_3$ precipitation were comparable for *S. pasteurii* and *B. sphaericus* 21776. Although *B. sphaericus* 21787 showed poor ureolysis, some $CaCO_3$ precipitation was observed. When rate coefficients were normalized to cell numbers, *B. sphaericus* 21776 had the highest rate of ureolysis and calcite precipitation per cell compared to *S. pasteurii* and *B. sphaericus* 21787, indicating that it may have a higher cell-specific ureolytic activity under the conditions studied here. *B. sphaericus* 21776 may therefore be a candidate species for subsurface augmentation if maximizing rates of ureolysis and precipitation is desirable, although in our experiments we were not able to generate cell concentrations as high as in *S. pasteurii* cultures, so optimization of growth media may be required.

S. pasteurii was capable of ureolysis in anaerobic environments with and without the addition of potential electron acceptors, however, sustained growth of *S. pasteurii* over time in the absence of oxygen did not appear to be possible. Comparison of kinetics from aerobic and anaerobic experiments demonstrates that rates are on the same order of magnitude suggesting that oxygen-free environments do not significantly impact the initial rate of ureolysis or $CaCO_3$ precipitation. Apparent rate coefficients for ureolysis were reduced in CMM+ relative to CMM-. The limited increase in cell biomass during the period of $CaCO_3$ precipitation and TEM images reveal this may be due to the encasement and inactivation of cells. However, populations can recover and re-grow once $CaCO_3$ precipitation has ceased. Therefore, ureolysis-induced $CaCO_3$ precipitation is likely to be efficient in aerobic and anaerobic subsurface systems. However, our data and other recent studies under flow conditions demonstrate that if only one injection of microbes is to occur but longer term ureolysis is desired in subsurface applications, resuscitation and regrowth of microbes, e.g. through the injection of growth media, will be necessary since $CaCO_3$ precipitation greatly inhibits cell growth (Cuthbert et al., 2012;Phillips et al., 2013b).

Author contribution

A.C.M., E.J.E.-O., S.L.B. and R.G. wrote the manuscript. R.G., A.C.M., S.P., A.P. and A.B.C. designed experiments. S.L.B. performed experiments.

Competing interests

The authors declare that they have no conflict of interest.

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Figures

Figure 1. Changes in urea (▲) and dissolved calcium (●) concentrations during ureolysis over time in calcium-inclusive aerobic experiments for (A) *S. pasteurii*, (B) *B. sphaericus* 21776, and (C) *B. sphaericus* 21787. Data points are the averages of triplicate experiments; vertical error bars represent the standard deviations; horizontal error bars indicate standard deviation of the sampling times; error bars are smaller than markers if not visible.

Figure 2. Change in protein concentrations and CFU mL^{-1} over time for calcium inclusive (solid markers) and calcium exclusive (open markers); aerobic medium, (A) *S. pasteurii*, (B) *B. sphaericus* 21776 and (C) *B. sphaericus* 21787. Data points are the average of triplicate experiments; vertical error bars represent the standard deviation of triplicate experiments; horizontal error bars indicate standard deviation of the sampling times.

Figure 3. Transmission electron microscopy images of *S. pasteurii* cells in calcium-inclusive (A-C) and calcium-exclusive (D) media. Arrows indicate a cross-section through a Ca-rich layer, approx. 280 nm thick.

Figure 4. Changes in pH and OD_{600} over time in anaerobic (■) and aerobic (●) calcium-exclusive medium for *S. pasteurii* with (A) NO_3^-, (B) Fe^{3+}, and (C) SO_4^{2-} as terminal electron acceptors; experiments without added terminal electron acceptor are shown in (D). Abiotic controls (▲) are also shown. Open markers indicate values for timepoints after the bottles were opened to the environment allowing oxygen to enter the system.

Figure 5. Changes in urea (triangle markers) and dissolved calcium (cirlce markers) concentrations over time in anaerobic experiments with *S. pasteurii* in calcium inclusive medium with NO_3^- (black solid markers), calcium exclusive medium with NO_3^- (open markers) and calcium inclusive medium without added terminal electron acceptor (grey solid markers). Data points are the averages of triplicate experiments; vertical error bars represent the standard deviations of measurements; horizontal error bars indicate standard deviation of the sampling times; error bars are smaller than markers if not visible.

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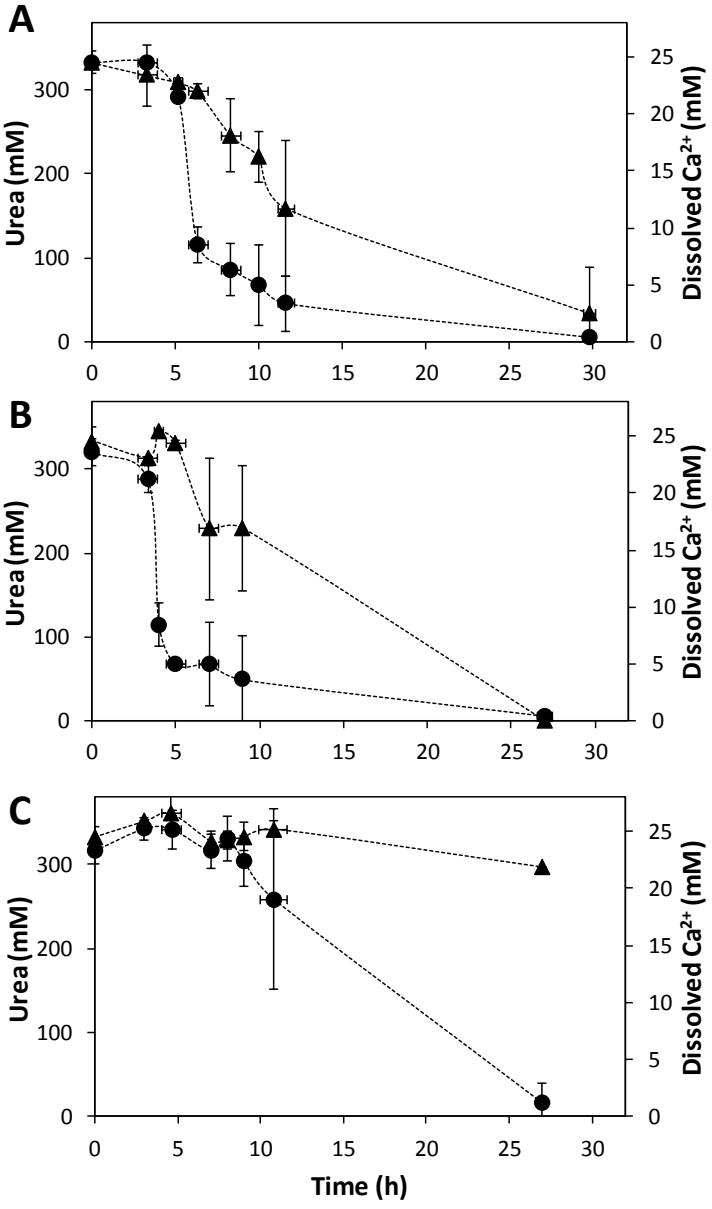

Figure 1.

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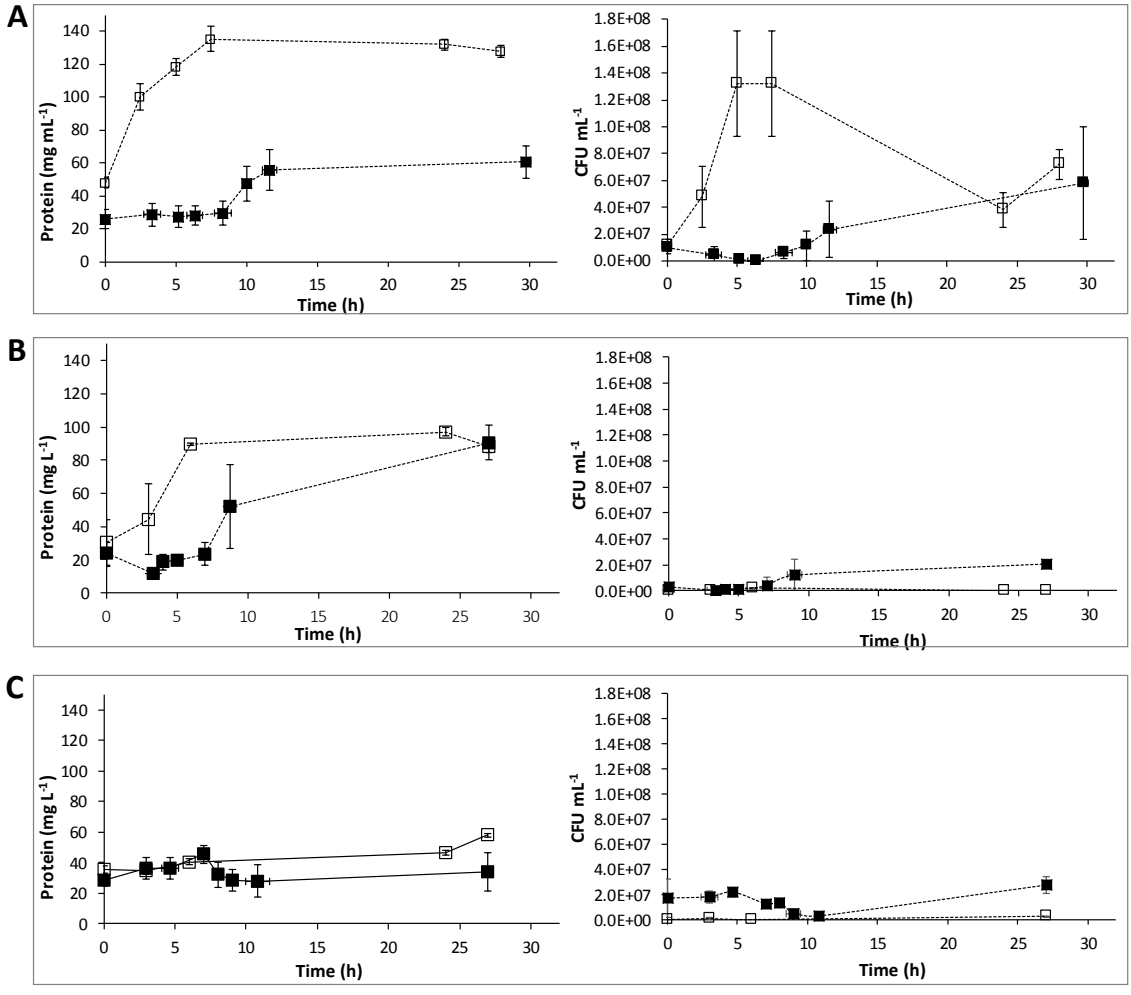

Figure 2.

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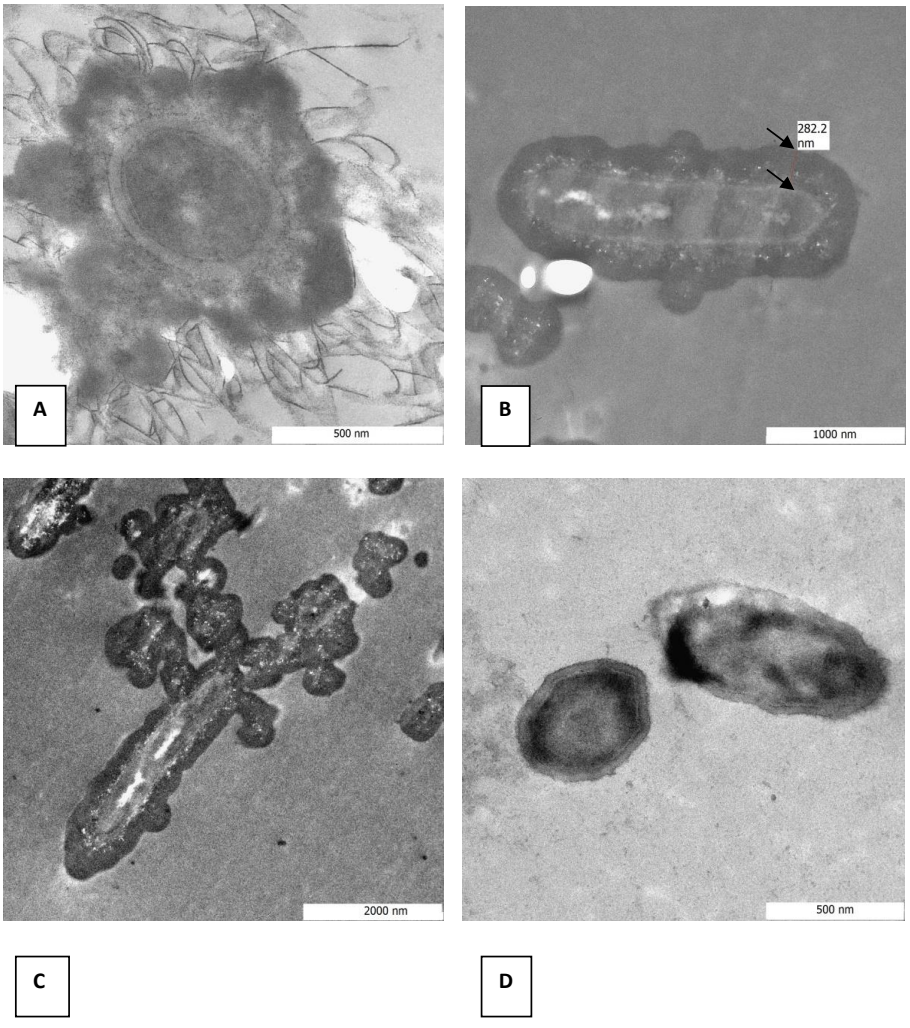

Figure 3.

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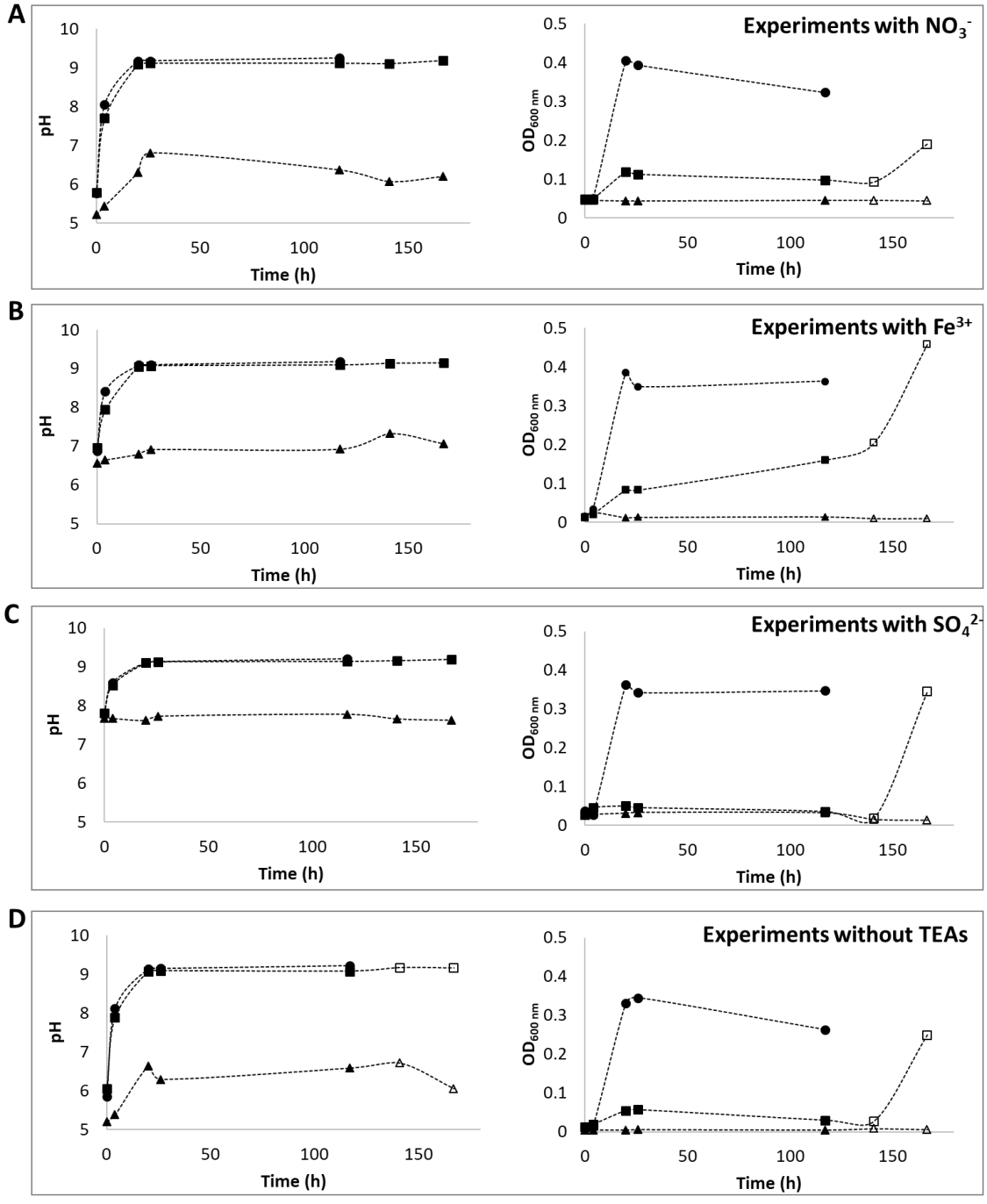

Figure 4.

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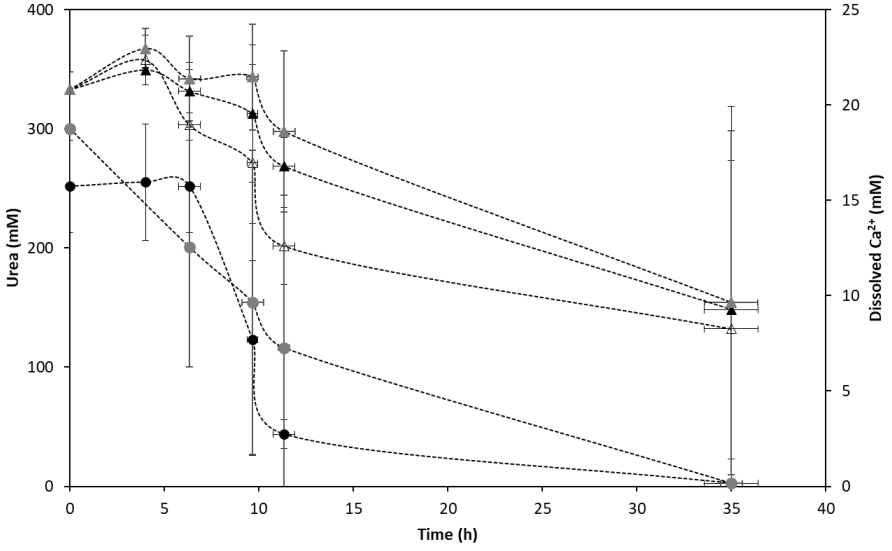

Figure 5.

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Tables

Table 1. Change in pH in aerobic calcium-inclusive (CMM+) and calcium-exclusive experiments (CMM-). Results are averages from triplicate experiments unless stated otherwise. Data for hour 0 was taken immediately after inoculation.

	Species	0 h	10 (±1) h	24 (±3) h
CMM+	*S. pasteurii*	6.66±0.06	8.87±0.08	9.33±0.02*****
	B. sphaericus 21776	7.24±0.30	8.80±0.20	9.23±0.09*****
	B. sphaericus 21787	6.87±0.15	8.06±0.12	8.70±0.26*****
	B. subtilis	6.80§	---	7.50§
	Sterile Control	7.08±0.04	---	7.31±0.05*****
CMM-	*S. pasteurii*	6.91±1.01*****	9.16±0.12*****	9.16§
	B. sphaericus 21776	6.85±0.91*****	9.10§	9.30±0.14*****
	B. sphaericus 21787	7.5§	---	9.00§
	B. subtilis	7.15*****	7.40§	---
	Sterile Control	6.3±0.36	6.56±0.35	6.60±0.28*****

*Data taken from only two experiments
§Data taken from only one experiment
---No data available

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Table 2. Summary of kinetic parameters for urea hydrolysis (k_{urea}) and calcite precipitation (k_{precip}) in aerobic and anaerobic experiments in calcium-inclusive (CMM+) and calcium-exclusive (CMM-) experiments inoculated with *S. pasteurii*, *B. sphaericus* 21776 and *B. sphaericus* 21787. Anaerobic experiments were incubated with or without nitrate as the terminal electron acceptor (TEA). Results are averages from triplicate experiments unless stated otherwise.

	Microorganism	Initial biomass (OD_{600})	k_{urea} (h^{-1})	Lag time (h)	k_{urea} normalized to: OD_{600} (OD_{600}^{-1} h^{-1})	k_{urea} normalized to: CFU (mL CFU^{-1} h^{-1})	k_{precip} (h^{-1})	Lag time (h)
Aerobic								
CMM+	*S. pasteurii*	0.014 ± 0.001	0.074 ± 0.021	5.0 ± 1.0	5.251 ± 1.273	3.22E-08 ± 6.54E-09	0.253 ± 0.021	3.3 ± 0.6
	B. sphaericus 21776	0.014 ± 0.001	0.107 ± 0.038	4.0 ± 0.0	8.020 ± 3.786	5.30E-08 ± 3.23E-08	0.604 ± 0.344	3.3 ± 0.6
	B. sphaericus 21787	0.015 ± (n/a) §	0.023 ± (n/a) §	3 ± (n/a) §	1.526 ± (n/a) §	8.52E-09 ± (n/a) §	0.219 ± (n/a) §	8 ± (n/a) §
CMM-	*S. pasteurii*	0.017 ± 0.000 *	0.192 ± 0.104 *	4.0 ± 0.0 *	11.227 ± 5.988 *	5.88E-08 ± 2.02E-08 *	-	-
	B. sphaericus 21776	0.015 ± 0.001 *	0.168 ± 0.050 *	3.5 ± 0.71 *	10.818 ± 2.797 *	6.20E-08 ± 1.33E-08 *	-	-
	B. sphaericus 21787	0.015 ± (n/a) §	0.067 ± (n/a) §	6.0 ± (n/a) §	3.196 ± (n/a) §	1.75E-08 ± (n/a) §	-	-
Anaerobic								
CMM+	*S. pasteurii*/NO_3^-	0.014 ± 0.002 *	0.048 ± 0.018 *	6.5 ± 0.7 *	3.617 ± (1.092) *	2.34E-08 ± 4.67E-09 *	0.360 ± 0.223	6.5 ± 0.6
	S. pasteurii/no TEA	0.014 ± 0.002	0.082 ± (n/a) §	10.0 ± (n/a) §	6.616 ± (n/a) §	4.68E-08 ± (n/a) §	0.191 ± 0.050	3.3 ± 0.6
CMM-	*S. pasteurii*/NO_3^-	0.014 ± 0.002 *	0.071 ± 0.017 *	8.5 ± 2.1 *	5.278 ± 0.875 *	3.45E-08 ± 2.05E-09 *	-	-

§ One experiment used in analysis
* Two experiments used for kinetic analysis
n/a = No data available

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Table 3. Summary of kinetic coefficients and initial growth conditions for aerobic calcium-inclusive experiments performed in this study and previous studies.

Aerobic conditions	This study	This study	This study	Stocks-Fischer et al. (1999)	Fujita et al. (2000)	Ferris et al. (2003)	Tobler et al. (2011)					
	**B. sphaericus* 21787	*B. sphaericus* 21776	*S. pasteurii* ATCC 11859	*S. pasteurii* ATCC 6453	*S. pasteurii* ATCC 11859	*S. pasteurii* ATCC 11859	*S. pasteurii* ATCC 11859					
Temperature (°C)	30	30	30	25	20	20	20	20	20	20	20	20
Initial pH	6.7	6.7	6.7	8.0	6.5	6.5	6.5	6.5	6.5	6.5	6.5	6.5
$[Ca^{2+}]$ (mM)	25.2	25.2	25.2	25.2	25	1.75	50	200	500	50	200	500
[Urea] (mM)	333	333	333	333	333	6	250	250	500	250	250	500
[cells] (OD_{600})**	0.015	0.014	0.014	0.010	0.072	0.070	0.03	0.03	0.03	0.07	0.07	0.07
k_{urea} (h^{-1})	0.023	0.107	0.074	0.028	0.008	0.038	0.007	0.005	0.005	0.074	0.095	0.044
k_{urea} (OD_{600}^{-1} h^{-1})	1.526	8.020	5.251	2.800	0.111	0.543	0.250	0.180	0.180	1.065	1.363	0.630
k_{urea} (mL CFU^{-1} h^{-1})	8.52E-09	5.30E-08	3.22E-08	3.00E-08	3.73E-10	1.81E-9	9.86E-10	7.13E-10	7.13E-10	3.56E-09	4.56E-09	2.11E-09
k_{precip} (h^{-1})	0.219	0.604	0.253	0.116	0.112	0.014	0.007	0.011	--	--	0.065	--

*Rate coefficients obtained for this strain are not conclusive and should be taken only as a guide for comparison purposes in this study.
**OD_{600} values were converted to a 1 cm path length equivalent where necessary.
--No reported values