

1 **A refinement of coccolith separation methods: Measuring the sinking**  
2 **characters of coccoliths**

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8 **Abstract.** Quantification sinking velocities of individual coccoliths will contribute to optimizing  
9 laboratory methods for separating coccoliths of different sizes and species for geochemical analysis.  
10 The repeat settling/decanting method was the earliest method proposed to separate coccoliths from  
11 sediments, and is still widely used. However, in the absence of estimates of settling velocity for non-  
12 spherical coccoliths, previous implementations have depended mainly on time consuming empirical  
13 method development by trial and error. In this study, the sinking velocities of coccoliths belonging  
14 to different species were carefully measured in a series of settling experiments for the first time.  
15 Settling velocities of modern coccoliths range from 0.154 to 10.67 cm h<sup>-1</sup>. We found that a quadratic  
16 relationship between coccolith length and sinking velocity fits well and coccolith sinking velocity  
17 can be estimated by measuring the coccolith length and using the length-velocity factor,  $k_v$ . We  
18 found a negligible difference in sinking velocities measured in different vessels. However, an  
19 appropriate choice of vessel must be made to avoid ‘hindered settling’ in coccolith separations. The  
20 experimental data and theoretical calculations presented here support and improve the repeat  
21 settling/decanting method.

## 22 **1. Introduction**

23 Coccolithophores are some of the most important phytoplankton in the ocean. They can secrete  
24 calcareous plates called coccoliths, which contribute significantly to discrete particulate inorganic  
25 carbon in the euphotic zone and to CaCO<sub>3</sub> fluxes to the deep ocean (e.g., Young and Ziveri, 2000;  
26 Sprengel et al., 2002). Coccolith morphology, geochemistry and fossil assemblage composition  
27 can reflect paleoenvironmental changes (e.g., Beaufort et al., 1997; Stoll et al., 2002; Zhang et al.,  
28 2016). However, the use of coccolith geochemical analyses in paleoenvironmental reconstructions  
29 was so far hindered by the difficulty of isolating coccolith compared with foraminifera. Two main  
30 methods have been developed to concentrate near-monospecific assemblages of coccoliths from  
31 bulk sediments: one is the method based on a decanting technique (Paull and Thierstein, 1987; Stoll  
32 and Ziveri, 2002) and the other is that based on microfiltration (Minoletti et al., 2009). The  
33 improvement of separation techniques offered a new perspective to study the Earth's history (e.g.  
34 Stoll, 2005; Beltran et al., 2007; Bolton and Stoll, 2013; Rousselle et al., 2013). Moreover, the  
35 development of coccolith oxygen and carbon isotope studies in culture in recent years (e.g. Ziveri  
36 et al., 2003; Rickaby et al., 2010; Hermoso et al., 2016; McClelland et al., 2017) has provided an  
37 improved mechanistic understanding of coccolith isotope data and therefore stimulated the need for  
38 more purified coccolith fraction samples from the fossil record.

39 Both decanting and microfiltering are widely used methods for coccolith separation. The  
40 microfiltering method separates coccoliths with polycarbonate micro-filter membrane (with pore  
41 sizes of 2µm, 3µm, 5µm, 8µm, 10µm and 12µm). This method is highly effective in the larger size  
42 ranges, but is very time consuming in sediments with a high proportion of small (<5µm) coccoliths  
43 (which tends to be the case in natural populations). It is also impossible to separate coccoliths with  
44 similar lengths by microfiltration, such as *Florisphaera profunda* and *Emiliana huxleyi* (Hermoso  
45 et al., 2015). Decanting, on the other hand, is highly effective for the small-sized coccoliths, because  
46 their slow settling times permit a greater ability to separate different sizes. Consequently, in some  
47 studies, a combination of the micro filtering and sinking or centrifugation method were applied for  
48 coccolith separation (Stoll, 2005; Bolton et al., 2012; Hermoso et al., 2015). The repeated  
49 sinking/decanting method, first employed by (Edwards, 1963; Paull and Thierstein, 1987) follows  
50 the simple principle formalized by Stokes' Law for spherical particles: particles of larger size settle

51 more quickly because they have a higher ratio of volume and mass (accelerating sinking) to sectional  
52 area (resistance retarding sinking). However, the sinking velocities of coccoliths with complex  
53 shape are difficult to calculate and have not been quantified in previous studies. Consequently, the  
54 repeated decanting method has generally used settling times based on empirical trial and error.  
55 In the current study, we present a novel and rigorous estimation of sinking velocity for 16 species  
56 of modern and Cenozoic coccoliths, carefully measured in 0.2% ammonia at 20°C. With this new  
57 dataset, we explore how to estimate the sinking velocity of coccoliths based on their shape and  
58 length, which allows our estimations to be generalized for other species, and for situations where  
59 the mean length of coccoliths of a given species was different from that of our study. These  
60 generalizations, together with our results on sinking velocities of one coccolith species  
61 (*Gephyrocapsa oceanica*) in different vessels, should allow a significant improvement in efficiency  
62 of future protocols for separation of coccoliths by repeated decanting.

## 63 **2. Materials and methods**

### 64 **2.1 Sample selections**

65 We measured the sinking velocity of 16 different species of coccoliths, isolated from eight deep-sea  
66 sediment samples from the Pacific and Atlantic Oceans (Figure 1, Table A1). Sample were  
67 principally of Quaternary age but include two Neogene/Paleogene samples. In general, numbers of  
68 small coccoliths, including *E. huxleyi*, *Gephyrocapsa* spp and *Reticulofenestra* spp. are about an  
69 order of magnitude greater than that of larger coccoliths. However, the larger coccoliths'  
70 contributions to carbonate can be as high as 50% (Baumann, 2004; Jin et al., 2016). Moreover, both  
71 small coccoliths and large coccoliths are useful in geochemical analyses (Ziveri et al., 2003; Rickaby  
72 et al., 2010; Candelier et al., 2013; Bolton et al., 2012, 2016; Bolton and Stoll, 2013). Therefore,  
73 both small and large coccoliths were studied in this research. (B). Pictures of the studied coccolith  
74 are shown in Appendix B, and all classifications follow Nannotax3 except *Reticulofenestra* spp.  
75 (Figure C2 in Appendix C).

### 76 **2.2 Experiment designs**

#### 77 **2.2.1 Sample pretreatments**

78 The sinking velocity measurement depends on absolute abundance estimation (more details in 2.2.2).

79 However, on microscope slides, larger coccoliths and foraminifer fragments may cover smaller  
80 coccoliths, reducing the accuracy of coccolith absolute numbers. Thus, before sinking experiments  
81 were carried out, raw sediments were pretreated to purify the target coccoliths to reduce errors in  
82 coccolith counting. The raw sediments were disaggregated in 0.2% ammonia and sieved through a  
83 63 µm sieve and then treated by sinking method or filtering method (Bolton et al., 2012; Minoletti  
84 et al., 2009) to concentrate the target species up to at least more than 50% of the total assemblage  
85 (for Noëlaerhabdaceae coccoliths, a percentage more than 90% can be easily achieved). In one  
86 sample with aggregation (ODP 807), we did a rapid settling (30 min, 2 cm) to eliminate aggregates.  
87 Most of the species were measured individually in settling experiments, except for *Pseudoemiliana*  
88 *lacunosa* and *Umbilicosphaera sibogae*, which were measured together.

### 89 **2.2.2 Measuring the sinking speeds of coccoliths**

90 We are not aware of any prior direct determination of the sinking velocity of individual coccoliths,  
91 although the sinking velocities of live coccolithophores and other marine algal cells have been  
92 successfully measured by the ‘FlowCAM’ method (Bach et al., 2012) or a similar photography  
93 technique (e.g. Miklasz and Denny, 2010). Here we introduce a simple method to measure the  
94 particle sinking speeds without special equipment.

- 95 1. After pretreatment, the coccolith suspensions were gently shaken and then moved into  
96 comparison tubes which were vertically mounted on tube shelves. We set the timer going  
97 and let the suspension settle for a specified period of time, marked as sinking time or  
98 settling duration (T);
- 99 2. Thereafter, we removed the upper 15 ml supernatant into a 50 ml centrifuge tube with a 10  
100 ml pipette. This operation was performed slowly and gently to avoid drawing lower  
101 suspensions upward. The absolute counting of coccolith was achieved by using the ‘drop  
102 technique’ to make quantitative microscope slides (Koch and Young, 2007; Bordiga et al.,  
103 2015). 0.3 ml mixed suspension was extracted and pipettes onto a glass cover and dry the  
104 slider on a hotplate;
- 105 3. The lower suspension was then homogenized and another slider was prepared as described  
106 above;
- 107 4. The number of coccoliths in the upper and lower suspensions were carefully counted on

108 microscope at  $\times 1250$  magnification and the number of coccoliths and fields of view (FOV)  
109 were recorded for further calculations. More than 300 specimens were counted for most of  
110 the measurements. For the *Helicosphaera carteri* measurements, more than 100 FOV were  
111 checked and about 100 specimens were counted.

112 To calculate the sinking velocities of coccoliths, we define a parameter named the separation ratio  
113 (R), which represents the percentage of removed coccoliths in one separation by pumping out the  
114 upper suspension. This parameter is important and will be repeatedly mentioned in the following  
115 part. R was measured using the following equation (more details about derivation can be found in  
116 Appendix D):

$$117 \quad R = \frac{\frac{N_1}{n_1} \times V_1}{\frac{N_1}{n_1} \times V_1 + \frac{N_2}{n_2} \times V_2} \quad (2-1)$$

118 where  $N_1$  and  $N_2$  are numbers of coccoliths counted in upper and lower suspension slides,  
119 respectively;  $n_1$  and  $n_2$  are the number of FOV counted.  $V_1$  and  $V_2$  are the volume of the settling  
120 vessel defined by the settling distance, as shown in Figure 2.

121 The separation ratio, R, also has a relationship with sinking time, T (Appendix D):

$$122 \quad R = \frac{V_1 - \frac{V_1}{D} \times v \times T}{V_1 + V_2} \quad (2-2)$$

123 where  $V_1$ ,  $V_2$  and  $D$  are shape parameters shown in Figure 2; and  $v$  is the average sinking velocity  
124 of measured coccoliths. If we plot R against T, the slope of line has a relationship with  $v$ . Then liner  
125 regressions between R and T were processed with MATLAB to calculate the  $v$  (details about error  
126 analyses can be found in Appendix E).

127 There are still two issues to be explained. Firstly, to eliminate the shape differences among vessels,  
128 all separation ratios have been transferred to calibrated separation ratios ( $R_{cal}$ ), which means the  
129 separation ratio measured in a standard vessel with  $V_1=15$  ml,  $V_2=10$  ml and  $D=6$  cm (more details  
130 about transformation from R to  $R_{cal}$  can be found in Appendix D). Secondly, we treated the average  
131 sinking velocities as the sinking velocities of the coccoliths with the average length. This  
132 approximation has been proved reasonable in Appendix D.

### 133 **2.2.3 Detecting the potential influence of vessels**

134 Seven commonly used vessels were selected to detect the potential influence of vessels (Figure 3).  
135 Two of them are made of plastics (No.2 and No.3 in Figure 3) and all others are pyrex glass vessels.  
136 About 500 mg of sediment from core KX21-2 were pretreated as described in 2.2.1 and suspended

137 in about 500 ml ammonia. After that, settling experiments were performed as described in 2.2.2  
138 using different vessels. In these experiments, only the dominant species, *G. oceanica*, was measured.

### 139 **2.2.4 Other factors influencing the sinking velocity**

140 Temperature can change the density and viscosity of liquid. Generally speaking, the higher the  
141 temperature is, the lower the density and viscosity will become and the faster pellets will sink. Take  
142 water for instance, if the temperature increases from 15 to 30°C, the particle sinking velocity will  
143 increase by ~43% (Table 1). All sinking velocities measured or discussed in the following sections  
144 were velocities at 20°C to minimize the influence of temperature.

145 The calibration of sinking velocity in high concentration suspension has been calculated by  
146 Richardson and Zaki (1954)

$$147 \quad v = v_0(1 - \alpha_s)^{2.7} \quad (2-3)$$

148 where the  $\alpha_s$  is the solids volume fraction. Based on equation 2-3, the higher the suspension  
149 concentration is, the slower the sinking velocity will be. That is so called ‘hindered settling’. When  
150 the  $\alpha_s=0.2\%$ , the reduction of sinking velocity owing to hindered settling is negligible ( $v/v_0$  equals  
151 99.46%). Hence, in this study all suspensions have solid volume fractions lower than 0.2% to avoid  
152 notable reductions of coccolith sinking velocities.

## 153 **3. Results and Discussions**

### 154 **3.1 Influence of vessels**

155 The sinking velocities of *G. oceanica* in the core KX21-2 in 0.2% ammonia at 20°C measured in  
156 different vessels vary from 0.99 to 1.23 cm h<sup>-1</sup>. The lowest value occurred in the 100 ml centrifuge  
157 tube and the highest sinking velocity was measured in the 50 ml centrifuge tube experiments. The  
158 correlations between sinking velocities and different vessel parameters are quite low:  $r=0.13$  for the  
159 vessel inner diameter,  $r=0.0005$  for the sinking distance and  $r=0.051$  for the upper volume and total  
160 volume ratio ( $V_1/(V_1+V_2)$ ). The dissipation of energy by friction between the moving fluid and the  
161 walls can cause a reduction of sinking speed (wall effect). A significant wall effect will be detected  
162 when a particle is settling in a vessel with a diameter that is smaller than the particle size by two  
163 orders of magnitude (Barnea and Mizarchi, 1973). The length of coccoliths is on the micron scales,  
164 so the diameters of vessel used in laboratory are more than four orders of magnitude larger than

165 coccoliths. Moreover, our results show that the difference between vessel materials, glass and  
166 plastics, can also be ignored (Figure 4). Hence, we suggest that vessel type almost has no significant  
167 influence on sinking velocity of coccoliths.

168 However, our experiments were premised on the basis that the concentration of suspension was  
169 equal among different vessels. This means that large vessels can treat more sediment at one time but  
170 if we choose a larger vessel, more suspensions should be pumped and it often costs more time in  
171 sinking (often due to longer sinking distance). Assuming that the sediment is composed of 50%  
172 calcite (with density of  $2.7 \text{ g cm}^{-3}$ ) and 50% clay (about  $1.7 \text{ g cm}^{-3}$ ), the largest amount of sediment  
173 that can be used without significant reduction of the sinking velocity (5%) is about 400 mg in 100  
174 ml suspension (this calculation is based on equation 2-3). However, because sediments accumulate  
175 in the lower suspension, the particle concentration can be more than 4 times higher than in the initial  
176 homogenous concentration. This phenomenon will be more significant for a vessel with a narrow  
177 bottom, such as centrifuge tubes. To avoid this, we recommend using about 100 mg dry sediment  
178 suspended in at least 100 ml suspension to avoid ‘hindered settling’. If more sediment is necessary  
179 for geochemistry analyses, then a larger vessel should be selected to separate enough sample at one  
180 time.

### 181 **3.2 Sinking velocities at 20°C in 0.2% ammonia**

182 We measured the separation ratios of different coccoliths in comparison tubes at 20°C in 0.2%  
183 ammonia (Figure 5). The sinking velocities of coccoliths were then calculated by linear fitting of  
184 separation ratios and settling durations. The sinking velocities of studied coccoliths vary by two  
185 orders of magnitude from  $0.154 \text{ cm h}^{-1}$  to  $10.67 \text{ cm h}^{-1}$  (Table 2). The highest sinking velocity was  
186 found in the measurement of *Coccolithus pelagicus* and the lowest velocity was found for *F.*  
187 *profunda*. The average sinking speed of coccoliths is about 10-50% of the terminal sinking velocities  
188 of calcite spheres calculated by Stokes’ Law (Figure 6c). These ratios are comparable to the oval  
189 objects (e.g. seeds) data from Xie and Zhang (2001) and smaller than steel ellipsoids data from  
190 McNown and Malaika (1950). The sinking velocities of coccoliths measured in our experiment are  
191 about 2-3 orders of magnitude smaller than values from sediment traps of  $143\text{-}243 \text{ m d}^{-1}$  ( $595\text{-}1012$   
192  $\text{cm h}^{-1}$ ) in the North Atlantic (Ziveri et al., 2000 and Stoll et al., 2007), suggesting that the coccoliths  
193 sinking out of the euphotic layer are mainly in the form of sinking aggregates rather than individual

194 coccoliths.

### 195 **3.3 Estimating the sinking velocities**

196 Generally speaking, the sinking velocities of coccoliths increase with distal shield length (Figure  
197 5a), as expected from the increase in volume to sectional area for a given geometry as length  
198 increases. Our data implies that the sinking velocity has a power function relationship with distal  
199 shield length.

200 We propose that the sinking velocity of coccoliths might have a quadratic relationship with distal  
201 shield length as described by Stokes' Law (Figure 6a). If we use data for all species except *H. carteri*  
202 (the reason can be found in the following discussion), the sinking velocities can be described by the  
203 following equation:

$$204 \quad v = 0.0982 (\pm 0.001) * \phi^2 \quad (3-1)$$

205 Based on this quadratic regression, we derive a shape-velocity factor ( $k_v$ ) that relates settling  
206 velocity to coccolith length.

$$207 \quad v = k_v * \phi^2 \quad (3-2)$$

208 Furthermore, this factor is analogous to the shape-mass factor, ' $k_s$ ' used to relate coccolith mass to  
209 coccolith length (Young and Ziveri, 2000). The length and shape-velocity factor of coccoliths can  
210 be used to predict most of the sinking velocity variations, however, variations may also arise due to  
211 changes in coccolith mass and thickness, for a given length, and due to the hydrodynamics of  
212 particular shapes. We noticed that the smaller coccolith *G. caribbeanica* has a greater sinking  
213 velocity than the larger coccolith, *G. oceanica*. We suggest that this was caused by greater mass per  
214 length (or greater average thickness) in the case of *G. caribbeanica* and this may be due to the closed  
215 central area while *G. oceanica* has an open central area. Another example is *H. carteri*, which lower  
216 sinking velocity of which can be explained by the unique structure of *H. carteri* coccolith. Firstly,  
217 the broad edge of *H. carteri* can increase the drag force significantly. Moreover, most of the  
218 measured coccoliths have a ellipticity (major axis length and minor axis length ratio) larger than 0.8,  
219 while the ellipticity of *H. carteri* is around 0.6, which means the mass of *H. carteri* is smaller than  
220 other species of coccoliths with similar lengths (Figure 6d and Figure C3). That is also the reason  
221 *H. carteri* was excluded from the general regression in equation 3-1. In the case of partial dissolution,  
222 the well-preserved *Cyclicargolithus floridanus* may have higher mass than dissolved (or



223 disarticulated) *Cy. floridanus*, and therefore a slightly higher shape-velocity factor.

#### 224 **4. Suggestions for coccolith velocity estimations and separations**

225 To improve coccolith separation by settling methods, we measured sinking velocities of different  
226 coccoliths by gravity. Sinking velocities in this study varied from 0.154 to 10.61 cm h<sup>-1</sup>, about 10%  
227 to 50% of those of calcite spheres with same diameter. The shape of different vessels had little  
228 impact on the sinking velocity. But we should consider the volume of vessels to avoid ‘hindered  
229 settling’. The sinking velocities are mainly controlled by the shape of coccolith, including the distal  
230 shield length, the size of central area, and the ellipticity of coccoliths. Besides the shape of coccoliths,  
231 temperature is also crucial to the coccolith separations because of the dependence of sinking  
232 velocities on temperature. Length-velocity factors were proposed to estimate coccoliths sinking  
233 velocities, so coccolith separation can be achieved by following steps:

- 234 1. Measure the length of coccoliths in your target assemblage under the microscope and  
235 regress the length distribution by the assumption of normal distribution (details are in  
236 Appendix C);
- 237 2. Estimate sinking velocities for each important species. For species which sinking speed  
238 has been directly measured, we can use the length-velocity factor directly ( $v=k_v \cdot \phi^2$ ).  
239 For unmeasured species, we can choose the length-velocity factor of coccoliths with  
240 similar morphology in this study or use the general length-velocity formula  
241 ( $v=0.098(\pm 0.001) \cdot \phi^2$ );
- 242 3. Calculate the separation time for main species. For example, in KX21-2 there are three  
243 main coccoliths, *F. profunda*, *G. oceanica* and *Ca. leptoporus* and we wish to separate  
244 *G. oceanica* out from the bulk sediment. Calculate each coccoliths’ sinking velocity  
245 distributions as described in Step 2 above. As shown in Figure 7, a sinking velocity  
246 intermediate between *F. profunda* (with a length  $2\sigma$  larger than average, marked as  $+2\sigma$ )  
247 and *G. oceanica* (with a length  $2\sigma$  smaller than average, marked as  $-2\sigma$ ) optimal to  
248 separate them, would be 0.6 cm h<sup>-1</sup>. Similarly, we can chose speed thresholds 1.85 cm  
249 h<sup>-1</sup> to separate *G. oceanica* from *Ca. leptoporus*. If we settle in a 50 ml centrifuge tube  
250 with a sinking distance, D, equal to 5.84 cm, the sinking time for separating *F. profunda*  
251 should be  $T=5.84/0.6=9.73$  h. Similarly, we can calculate the time for separating *G.*

252 *oceania* by  $T=5.84/1.85=3.16$  h;

253 4. Homogenize the sediment suspension and let coccoliths settling as the period  
254 calculated in Step 3. After that, pump out the upper part of suspension. In the upper  
255 part, we have exclusively the smaller of the main coccoliths. However, column will  
256 still contain some smaller ones. So this step (settling and pumping) should be repeated  
257 until the lower part no longer has significant contribution from the smaller coccoliths.  
258 This step has been well described in pervious studies and more details can be found in  
259 Stoll and Ziveri (2002) and Bolton et al. (2012).

260 We find, if we use the general formula, a closed central area coccolith will sink faster than prediction  
261 (for *G. caribbeanica* and small *Ca. leptoporus* will settle ~40% faster) and coccoliths with greater  
262 ellipticity can settle much slower (for *H. carteri* will settle as 30% of the predicted sinking velocity  
263 for coccolith with similar length). Moreover, the sinking method cannot separate every species of  
264 coccoliths perfectly. As mentioned in Section 2.2.1, *P. lacunosa* and *U. sibogae* cannot easily be  
265 separated from each other because they have similar sinking velocities. Nevertheless, this study  
266 provides the first direct estimation of coccolith settling velocities, which should simplify  
267 implementation of future methods to separate coccoliths by settling time.

268

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272 Foundation and participating countries under management of the IODP Management International,  
273 Inc (IODP-MI).

274 **Table 1.** The influence of temperature on sinking velocity. Density data is from Kell (1975) and  
 275 viscosity data is from Joseph et al. (1978).

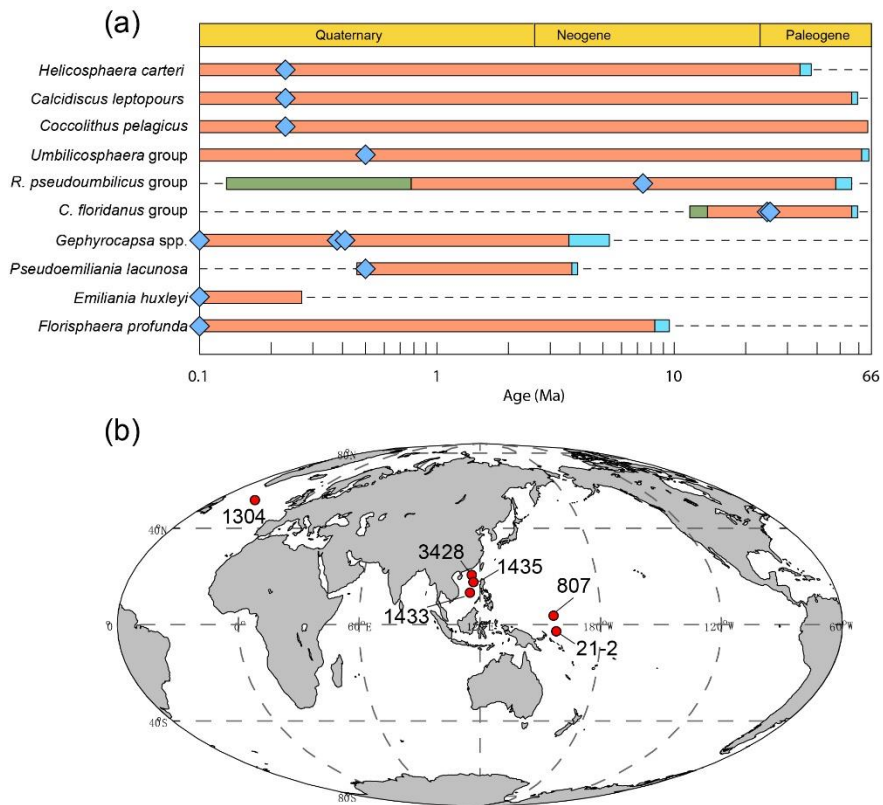
T (°C)	$\rho$ (g cm <sup>-3</sup> )	$\eta$ (mPa s)	$v_T : v_{T=20}$
15	0.9991	1.1447	0.8804
20	0.9982	1.0087	1
25	0.9970	0.8949	1.1279
30	0.9956	0.8000	1.2627

276 **Table 2.** The sinking velocity and shape-velocity factor of different coccolith species:  $\phi$  means the  
 277 distal shield length of coccolith and  $St \phi$  is the standard deviation of distal shield length;  $sv$  represents  
 278 the sinking velocity;  $v$  (95%-) and  $v$  (95%+) represent the lower and higher limit of 95% confidence  
 279 level, respectively. ' $k_v$ ' represents the length-sinking velocity factor. The short name of coccolith can be  
 280 found in the caption of Figure 4. The details of coccoliths length distribution are in Appendix C.

Species	abb.	$\phi$ ( $\mu\text{m}$ )	$St \phi$ ( $\mu\text{m}$ )	sinking velocity ( $\text{cm h}^{-1}$ )	$v$ (95% -)	$v$ (95% +)	$k_v$
<i>F. profunda</i>	Fp-WP	1.508	0.557	0.158	0.010	0.011	0.070
<i>F. profunda</i>	Fp-SCS	1.786	0.641	0.154	0.051	0.052	0.048
small <i>Reticulofenestra</i>	Ret (<4 $\mu\text{m}$ )	2.454	0.509	0.848	0.354	0.416	0.141
<i>E. huxleyi</i>	Emi	2.512	0.469	0.853	0.054	0.064	0.135
<i>Gephyocapsa</i> spp.	G spp	2.755	0.502	0.752	0.125	0.147	0.099
<i>G. caribbeanica</i>	Gcar	3.312	0.352	1.873	0.174	0.192	0.171
<i>U. sibogae</i>	Umb	4.060	0.500	1.268	0.416	0.441	0.077
<i>G. oceanica</i>	Geo	4.187	0.517	1.170	0.155	0.178	0.067
<i>P. lacunosa</i>	Pla	4.350	0.617	1.171	0.337	0.338	0.062
Small <i>Ca. leptoporus</i>	Cal small	4.605	0.629	3.351	0.172	0.199	0.158
large <i>Reticulofenestra</i>	Ret(>4 $\mu\text{m}$ )	4.988	0.605	2.379	0.534	0.641	0.096
<i>Cy. floridanus</i>	Cyf	5.805	0.963	4.174	0.320	0.336	0.124
(dissolved) <i>Cy. floridanus</i>	Cyf -d	6.134	0.727	4.508	0.352	0.417	0.120
Large <i>Ca. leptoporus</i>	Cal large	6.370	0.931	3.737	1.053	1.336	0.092
<i>H. carteri</i>	Hel	8.936	0.994	2.541	1.740	2.440	0.032
<i>Co. pelagicus</i>	Cpl	10.640	1.175	10.610	0.950	1.235	0.094

281

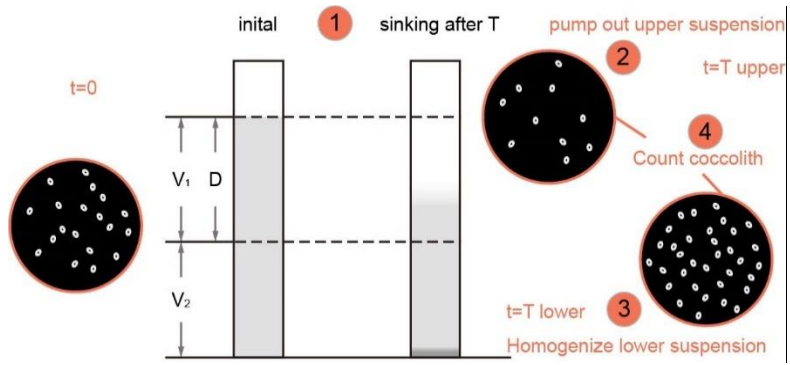
282 **Figure 1.** Temporal and spatial distribution of samples. (a) The evolution of studied coccoliths: first  
 283 occurrence and last occurrence data are from Nannotax3  
 284 (<http://www.mikrotax.org/Nannotax3/index.html>). The blue bars represent ranges of first occurrence  
 285 and the green bars represent ranges of last occurrence. The blue diamonds represent samples used in  
 286 this study. (b) Spatial distribution of samples. 1304 means IODP U1304, 3428 means MD12-3428cq,  
 287 1433 and 1435 means IODP U1433 and U1435, respectively. 807 means ODP 807 and 21-2 means  
 288 KX21-2.



289

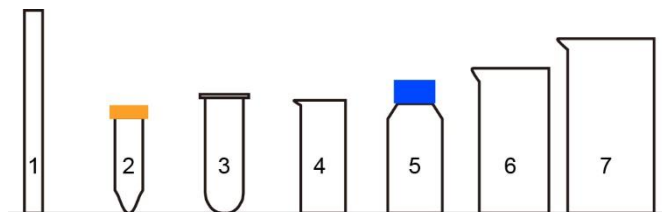
290

291 **Figure 2.** Schematic of settling experiments.  $V_1$  and  $V_2$  are the volumes of the upper and lower  
292 cylinders,  $D$  is the settled distance. The numbers in circles are same as the number of Steps described in  
293 Section 2.2.1.



294

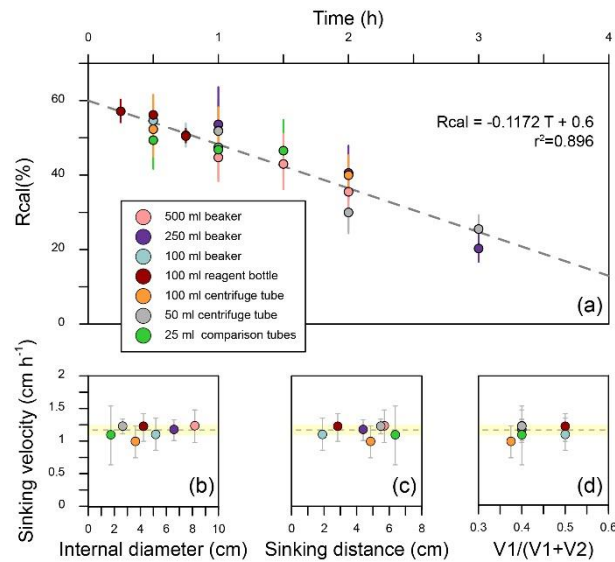
295 **Figure 3.** The shape parameters of vessels.  $V_1$  and  $V_2$  means the volume of upper suspension and lower  
 296 suspension, respectively.  $D$  means sinking distance.  $\Phi$  means average inner diameter which is  
 297 calculated by  $2*(V_1/\pi D)^2$ .



No.	Name	V1	V2	D (cm)	$\Phi$ (cm)
1	25 ml comparison tube	15	10	6.376	1.73
2	50 ml centrifuge tube	30	20	5.480	2.64
3	100 ml centrifuge tube	50	30	4.854	3.62
4	100 ml beaker	40	40	2.834	4.24
5	100 ml reagent bottle	40	40	1.900	5.18
6	250 ml beaker	150	100	4.400	6.59
7	500 ml beaker	300	200	5.700	8.19

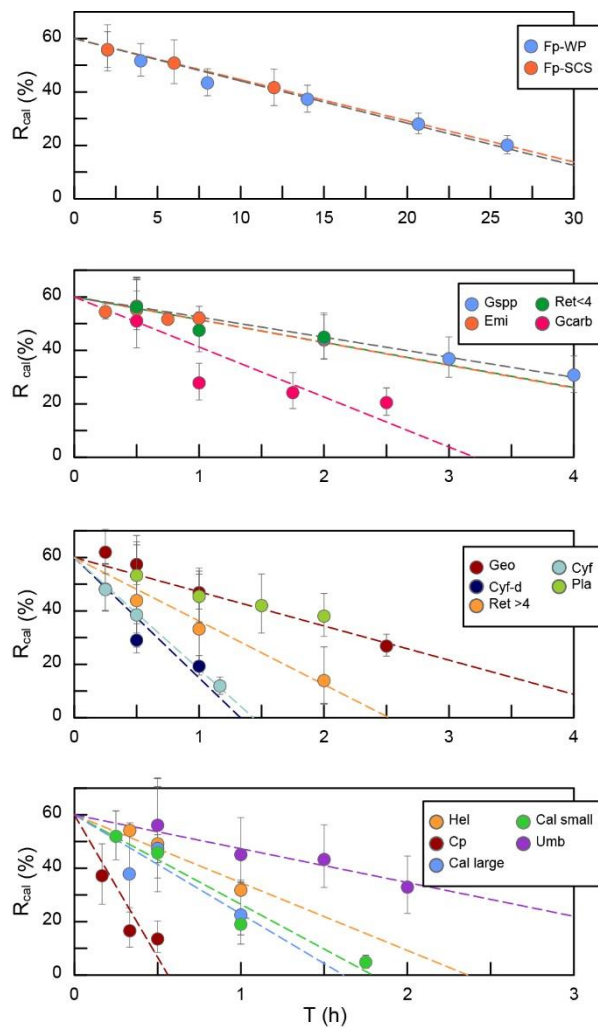
298  
 299

300 **Figure 4.** Sinking velocities of *G. oceanica* in the core KX-21-2 measured in different vessels. (a) The  
 301 calibrated separation ratios measured in different vessels. Error bars show 95% confidence level of  
 302 calibrated separation ratio. (b-d) The relationship between sinking velocity and different vessel shape  
 303 parameters. Error bars represent 95% confidence level of sinking velocity in each vessel and the shade  
 304 area represents 95% confidence level of sinking velocity considering all data points.



305

306 **Figure 5.** The calculated separation ratio ( $R_{cal}$ ) vs sinking duration. Fp-WP means *F. profunda* in the  
 307 West Pacific. Fp-SCS means *F. profunda* in the South China Sea. Emi means *E. huxleyi*. Gspg means  
 308 small *Geophyocapsa*. Geo means *G. oceanica*. Gcarb means *G. caribbeanica*. Ret<4 means small  
 309 *Reticulofenestra*. Ret>4 means large *Reticulofenestra*. Cyf means *Cyclicargolithus floridanus*. Cy-d  
 310 means dissolved *Cy. floridanus*. Umb means *U. sibogae*. Pla means *Pseudoemiliana lacunosa*. Hel  
 311 means *H. carteri*. Cal large means larger *Calcidiscus leptoporus*. Cal small means small *Ca.*  
 312 *leptoporus*. Cpl means *Co. pelagicus*.



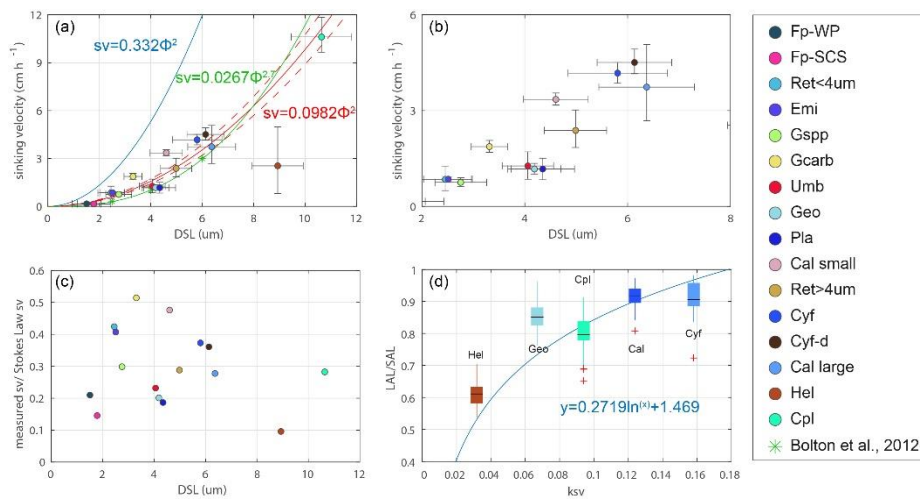
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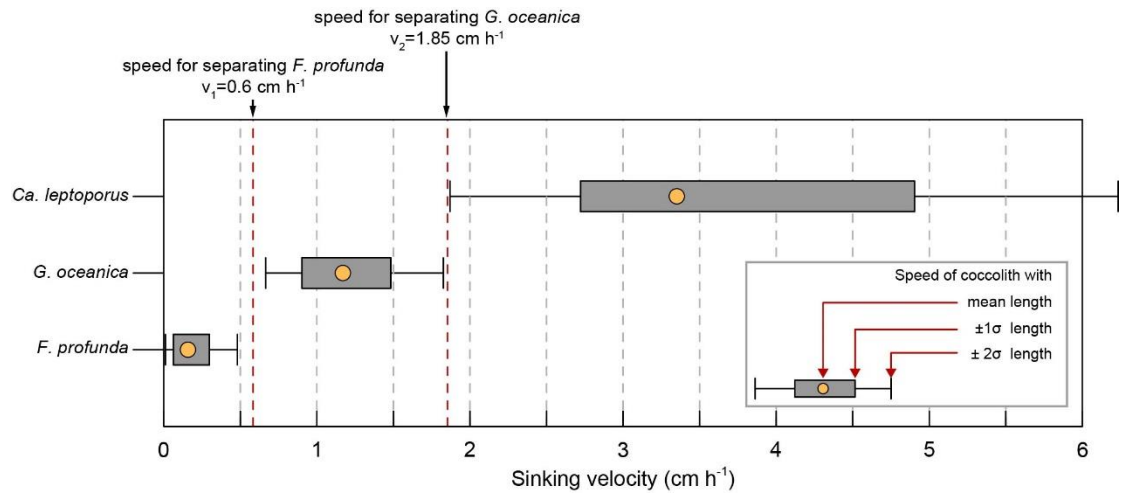
316 **Figure 6.** Coccolith sinking velocities and coccolith shape factors. (a-b) Sinking velocities and mean  
317 distal shield length. The horizontal error bars represent one standard deviation of coccolith length and  
318 the vertical ones represent 95% confidence level of measured sinking velocities. The blue, green and  
319 red lines represent sinking velocity of calcite sphere objects, coccolith sinking velocities estimated by  
320 Bolton et al. (2012) and this study, respectively. (c) The ratio of measured speed and speed calculated  
321 by Stokes' Law. (d) Coccolith short axis length (SAL) and long axis length (LAL) ratio against shape-  
322 velocity factor  $k_v$ . Box shows median value and upper/lower quartiles, whiskers show maximum and  
323 minimum values, outliers larger than 1.5 of the interquartile range are shown as red crosses. The SAL  
324 against LAL plot was shown in Figure C3. The short names of coccoliths can be found in Table 2.



325

326

327 **Figure 7.** The selection of separation velocities: the sinking velocities of three main coccolith species  
 328 in sample from core KX21-2 were calculated by the length distribution and velocity factors in Table 2.  
 329 The yellow dots represent sinking velocities of coccoliths with mean length. The edge of boxes show  
 330 the sinking velocities of coccolith within one standard deviation of length ( $\pm 1\sigma$ ) and the whiskers  
 331 mark the sinking velocities of coccolith within two standard deviation of length ( $\pm 2\sigma$ ).



332

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438 **Appendix A. Sample selections**

439 **Table A1.** Sample selections

Measured coccolith	abb.	Region	Core	Section	Epoch	Age model ref.
<i>F. profunda</i>	Fp-SCS	SCS	MD12-3428	0-1 cm	Holocene	Zhang et al., 2016
<i>F. profunda</i>	Fp-WP	W.P.	KX21-2	2-4 cm	Holocene	Liang et al., 2016
<i>E. huxleyi</i>	Emi	SCS	MD12-3428	0-1 cm	Holocene	Zhang et al., 2016
<i>Gephyocapsa</i> spp.	Gspp	W.P.	ODP 807A	1H 5W 102-104	Pleistocene	Jin et al., 2010
<i>G. oceanica</i>	Geo	W.P.	KX21-2	2-4 cm	Holocene	Liang et al., 2016
<i>G. caribbeanica</i>	Gcarb	N.A.	IODP 1304B	7H 5W 69-70	Pleistocene	Channell et al., 2010
small <i>Reticulofenestra</i>	Ret<4	SCS	IODP 1433B	28R 2W 30-34	Miocene	Li et al., 2013
large <i>Reticulofenestra</i>	Ret>4	SCS	IODP 1433B	28R 2W 30-34	Miocene	Li et al., 2013
<i>Cyclicargolithus floridanus</i>	Cyf	SCS	IODP 1435A	6R 3W 25-29	Oligocene	Li et al., 2013
<i>Cyclicargolithus floridanus</i>	Cyf-d	SCS	IODP 1435A	8R 1W 27-31	Oligocene	Li et al., 2013
<i>Umbilicosphaera sibogae</i>	Umb	W.P.	ODP 807A	3H 5W 92-94	Pleistocene	Jin et al., 2010
<i>Pseudoemiliana lacunosa</i>	Pla	W.P.	ODP 807A	3H 5W 92-94	Pleistocene	Jin et al., 2010
<i>Helicosphaera carteri</i>	Hel	W.P.	ODP 807A	3H 5W 92-94	Pleistocene	Jin et al., 2010
large <i>Calcidiscus leptoporus</i>	Cal large	W.P.	ODP 807A	3H 5W 92-94	Pleistocene	Jin et al., 2010
small <i>Calcidiscus leptoporus</i>	Cal small	N.A.	IODP 1304B	7H 5W 69-70	Pleistocene	Channell et al., 2010
<i>Coccolithus pelagicus</i>	Cpl	N.A.	IODP 1304B	7H 5W 69-70	Pleistocene	Channell et al., 2010

440

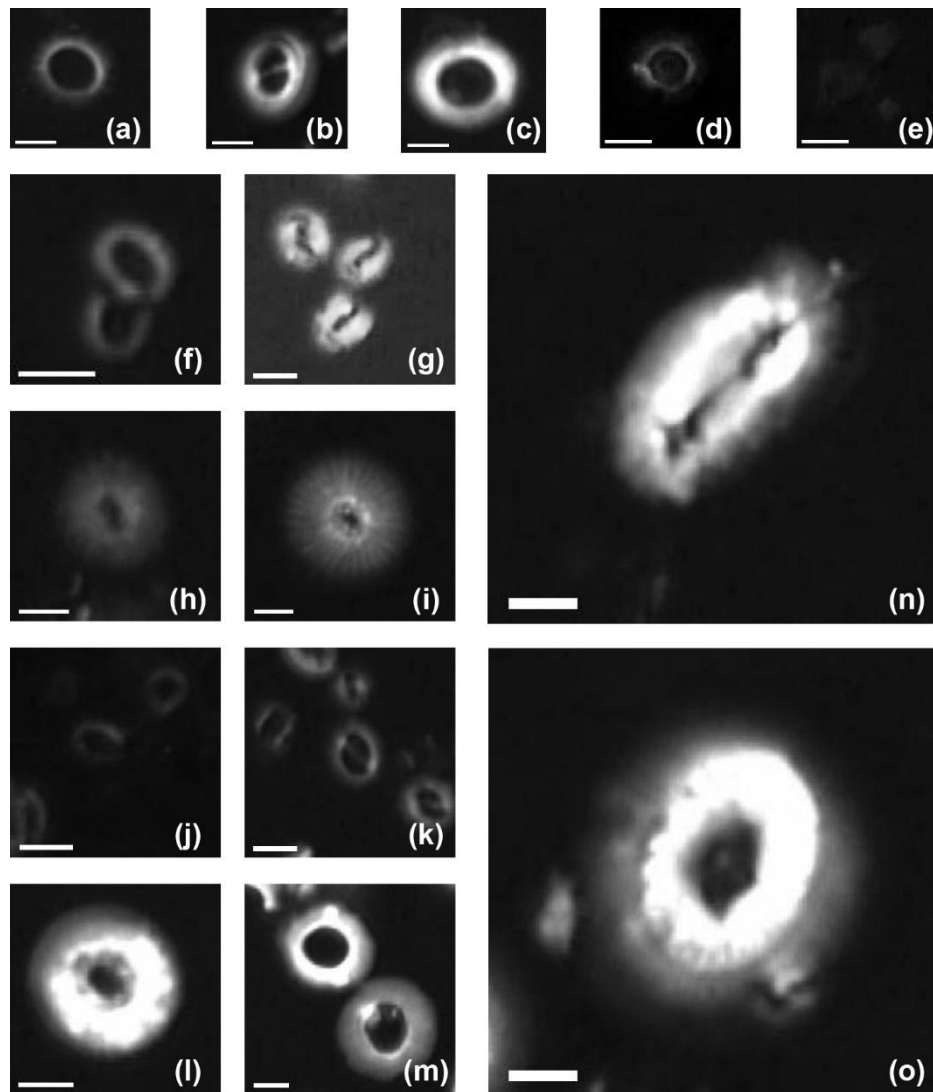
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## Appendix B. Coccolith images under circular polarized light

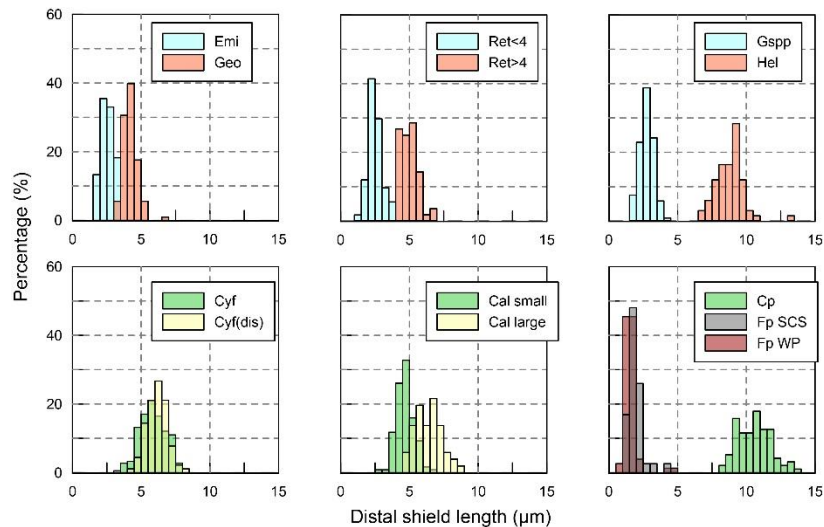


458

459 **Plate B1.** Imaged of measured coccolith in this study: (a) *Pseudoemiliania lacinosa* in the core ODP  
 460 807; (b) *Gephyrocapsa oceanica* in the core KX21-2; (c) *Reticulofenestra* spp. (large) in the core  
 461 IODP U1433B; (d) *Umbilicosphaera sibogae* in the core ODP 807; (e) *Florisphaera profunda* in  
 462 the core KX21-2; (f) *Reticulofenestra* spp. (small) in the core IODP U1433B; (g) *Gephyrocapsa*  
 463 *caribbeanica* in the core IODP U1304B; (h) small *Calcidiscus leptoporus* in the core IODP U1304B;  
 464 (i) large *Calcidiscus leptoporus* in the core ODP 807A; (j) *Emiliana huxleyi* in the surface sediment  
 465 in the South China Sea; (k) *Gephyrocapsa* spp. in the core ODP 807; (l) *Cyclicargolithus floridanus*  
 466 in the core IODP U1435A and (m) dissolved *Cyclicargolithus floridanus* in the same core; (n)  
 467 *Helicosphaera carteri* in the core ODP 807A; (o) *Coccolithus pelagicus* in the core IODP U1304B.  
 468 White bars represent a length of 2  $\mu\text{m}$ .

469 **Appendix C. The length distribution of coccoliths**

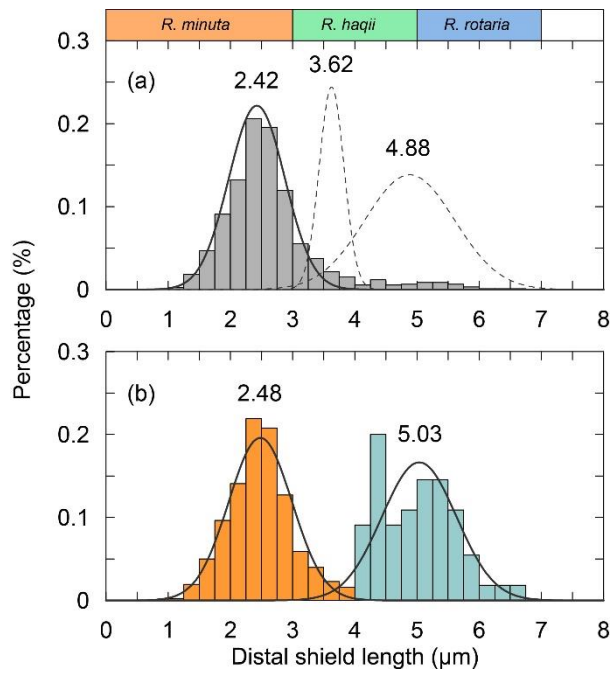
470 To measure the distal shield length of coccoliths, pictures were taken at a magnification of 1250x  
471 under circular polarized light. The coccolith lengths were measured by using the image analysis  
472 software, ImageJ. More than 5 pictures were taken and more than 50 (usually more than 100)  
473 coccolith specimens were measured. The length distributions of coccoliths measured in our  
474 experiments were shown in the Figure C1.



475

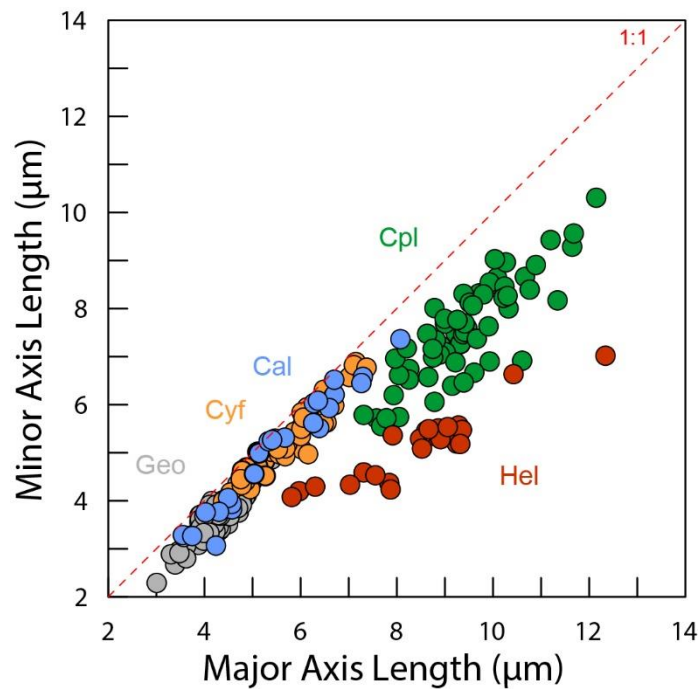
476 **Figure C1.** Size distribution of coccolith measured in the present study. The shorten names of coccolith  
477 follow Table A1.

478 The classification of coccoliths by length was supported by mixture analysis in PAST (Hammer et  
479 al., 2001), such as *Reticulofenestra* spp. and *Gephyrocapsa* spp. *Reticulofenestra* spp. in the  
480 Miocene were classified into two groups, Ret. (<4 µm) and Ret. (>4 µm). The traditional  
481 classification of *Reticulofenestra* spp. is <3 µm, 3-5 µm and 5-7 µm didn't pass the normal  
482 distribution test. Hence, in this study the *Reticulofenestra* spp. are divided at 4 µm (Figure C2).  
483 *Gephyrocapsa* spp. were classified by the shape of coccoliths into small *Gephyrocapsa* (central area  
484 opening and length <3.5 µm), *G. oceanica* (central area opening and length >3.5µm) and *G.*  
485 *caribbeanica* (closed central area) by the length and central area.



486

487 **Figure C2.** The classical classification of *Reticulofenestra* spp. (a) and the classification used in our  
 488 study (b). The curves represent the normal distribution fits of different coccolith groups and the dish  
 489 curve marks that the goodness of fit is below 0.2.



490

491 **Figure C3.** The short axis and long axis length distribution of coccoliths in Figure 6d.

492 **Reference.**

493 Hammer, Ø., Harper, D., Ryan, P., 2001. Paleontological Statistics Software: Package for  
494 Education and Data Analysis. *Palaeontologia Electronica*.

495 **Appendix D. Coccolith movement in gravity settling**

496 In this part, the derivation of equation will be explained in detail including proofs of several  
497 assumptions mentioned in the methods part.

498 When the well mixed sediment begins to sink, the decrease of coccoliths number in the upper  
499 suspension ( $N_u$ ) can be described as following equation:

500 
$$\frac{dN_u}{dT} = -\frac{N_{u(t=0)}}{D} \times v \quad (D-1)$$

501 where the  $D$  is the length of upper suspension and  $N_{u(t=0)}/D$  is the initial number of coccolith in  
502 cross-section with a unit thickness,  $v$  is the sinking velocity of coccolith.

503 Do integration for the equation D-1, we can get the variation of coccolith number in the upper  
504 column over time:

505 
$$N_u = N_{u(t=0)} - \frac{N_{u(t=0)}}{D} \times v \times T \quad (D-2)$$

506 where  $T$  is settling time. After a period of time ( $T$ ), we pump out the upper suspension. Here we  
507 define the number of coccoliths in the upper supernatant dividing the total coccoliths number in the  
508 tube ( $N_t$ ) as separation ratio ( $R$ ), which represents the percentage of total coccoliths removed in one  
509 separation.  $R$  can be expressed by

510 
$$R = \frac{N_u}{N_t} \quad (D-3)$$

511 Assuming all coccoliths are uniformly distributed in the suspension at the beginning of settling,  
512  $N_{u(t=0)}$  has relationship with  $N_t$  as follow:

513 
$$\frac{N_{u(t=0)}}{N_t} = \frac{V_1}{V_1+V_2} \quad (D-4)$$

514 where  $V_1$  is the volume of upper suspensions and  $V_2$  is the volume of lower suspensions.

515 Combining the equation D-1, D-2, D-3 and D-4, we obtain the relationship between separation ratio,  
516  $R$ , and sinking velocity,  $v$ , as follow:

517 
$$R = \frac{N_u}{N_t} = \frac{N_{u(t=0)} - \frac{N_{u(t=0)}}{D} \times v \times T}{N_t} = \frac{V_1 - \frac{V_1}{D} \times v \times T}{V_1+V_2} \quad (D-5)$$

518 If we plot the  $R$  and  $T$  on a figure, the slope of the line is a function of  $V_1$ ,  $V_2$ ,  $D$  and  $v$ . Since the  
519  $V_1$ ,  $V_2$ ,  $D$  are known parameters, we say the slope of  $R$ - $T$  is a function of  $v$ , which is exactly what  
520 we want.

521 Comparison tubes used in our experiments have the same  $V_1$  and  $V_2$  but different  $D$ . Other vessels  
522 used in other experiments have different  $V_1$ ,  $V_2$  and  $D$ . So we should adjust the raw separation ratio

523 to calibrated separation ratio ( $R_{cal}$ ), which represents the separation ratio made in a standard vessel  
 524 with  $V_{1std}=15$  ml,  $V_{2std}=10$  ml and  $D_{std}=6$  cm. This step can be described by equation D-6:

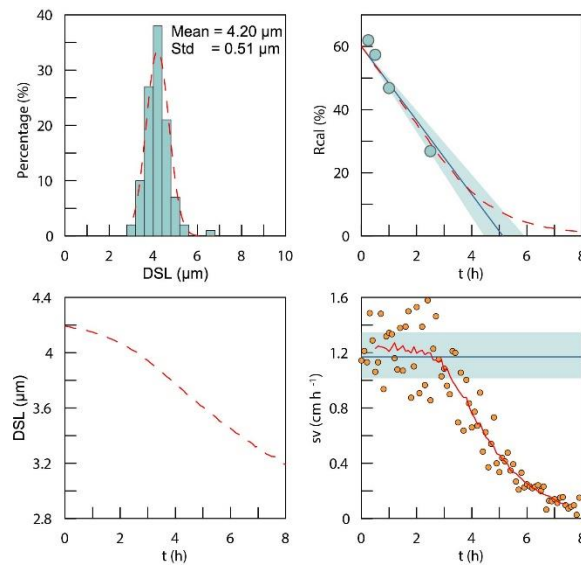
$$525 \quad R_{cal} = \frac{[R \times (V_1 + V_2) - V_1] \times D \times V_{1std}}{(D_{std} \times V_1 + V_{1std}) \times (V_{1std} + V_{2std})} \quad (D-6)$$

526 After calibrated, the slope of  $R_{cal}$ -T (k) has relationship with v as following equation:

$$527 \quad v = -\frac{D_{std} \times (V_{1std} + V_{2std})}{V_{1std}} \times k = -10 \times k \quad (D-7)$$

528 where k is the slope of  $R_{cal}$  against T from regression and other parameters are as described above.  
 529 Hence, the sinking velocity of different coccoliths can be achieved by measuring the variations of  
 530  $R_{cal}$  over time.

531 The coccoliths' lengths in the sediment have some variations. So what we measured is actually the  
 532 bulk settling velocity of whole coccolith population. We also offer a test for the assumption that the  
 533 average sinking velocity of all coccoliths can be treated as the sinking velocity of coccoliths with  
 534 the average length. Here we used the data of *G. oceanica*. A normal distribution was fitted to the  
 535 measured length distribution (Figure D1-a). We generated 100000 coccolith following the normal  
 536 distribution and let these coccolith evenly distributing in the comparison tube at the initial and then  
 537 set them sinking without collisions with each other. The sinking velocities of different size  
 538 coccoliths were calculated by the velocity-shape parameter 'k<sub>v</sub>' as described in discussion part. We  
 539 modeled the coccoliths sinking process and computed the separation ratio (red dash line in Figure  
 540 D1-b), coccolith length (red dash line in Figure D1-c) and instant sinking velocities (orange dots in  
 541 Figure D1-d) at different time sections.



542

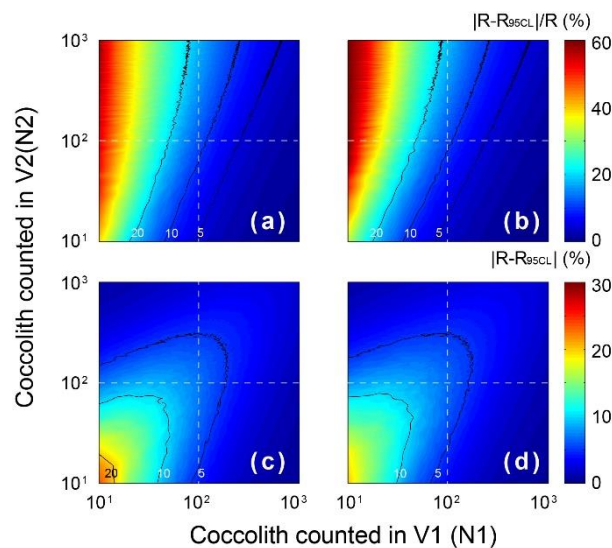
543

544 **Figure D1.** The simulations of coccoliths settling with different lengths: (a) the length distribution of  
545 coccoliths. The green bars represent measured data and red dash line represents the best fit for normal  
546 distribution. (b) The calibrated separation ratio: the green dots are measured data in our settling  
547 experiments, the blue line and shade area represent the calculated sinking velocity based on  $R_{cal}$   
548 measurement and the red dash line represents results obtained from simulations. (c) The average length  
549 of removed coccolith in simulations; (d) the modeling sinking velocities of coccoliths: the orange dots  
550 are instant sinking velocity calculated from derivation of  $R_{cal}$ , the red dash line is weighted average for  
551 the instant sinking velocity. Blue line represents the average sinking velocity we measured and the  
552 green shade area represents 95% confidence level of the measured velocity.

553 For *G. oceanica* experiments, the instant sinking velocity would not change significantly until  
554 settling for more 3 hours. That means for all  $R_{cal}$  larger than 15% are safe for liner regressions. The  
555 minimum safe number of  $R_{cal}$  will descend with the drop of dispersion degree of coccolith length  
556 distribution. Hence our assumption for average sinking velocity and the use of liner regression are  
557 proved to be reasonable.

558 **Appendix E. Statistical and error analyses**

559 The errors of measured separation ratio (R) and calculated sinking velocity (v) are mainly caused  
 560 by counting coccolith, the error of which follows the Poisson distribution. To detect the influence of  
 561 counting number on the result error, the error of separation ratio was simulated by 5000 times Monte  
 562 Carlo calculations with assumptions that ' $V_1:V_2=15:10$ ' and ' $n_1=n_2$ ' (Figure E1). The result shows  
 563 that the number of coccolith counted in the upper column draws more influence on the relative error  
 564 ( $|R-R_{95CL}|/R$ ). That means more coccolith in the upper suspension should be counted to make results  
 565 more accurate. The slope of  $R_{cal}-T$  was calculated by liner fitting with the intercept fixed on  
 566  $V_1/(V_1+V_2)$ . The input  $R_{cal}$  were generated from measured values considering the error of coccolith  
 567 counting. The regressions of  $R_{cal}-T$  were repeated by 5000 times regressions in the software Matlab  
 568 and the error of sinking velocity, v, was source from the distribution slope of  $R_{cal}-T$  in Monte Carlo  
 569 process.



570

571 **Figure E1.** The error distribution with different  $N_1$  and  $N_2$  (ranging from 1 to 1000) simulated 5000  
 572 times by the Matlab with assumptions that the error distributions of  $N_1$  and  $N_2$  follow Poisson  
 573 distribution. The calculation of R follows equation 2-5, and here we assume numbers of FOV are equal  
 574 ( $n_1=n_2$ ). Counter lines mark values equal to 5, 10 and 20. (a) and (c) represent the lower 95%  
 575 confidence level and (b) and (d) represent upper 95% confidence level. (a) and (b) the relative error of  
 576 R and (c) and (d) represent the absolute error of R.