

Dear Dr. Park

On behalf of my coauthors, I would like to thank you for accepting the manuscript. We are grateful to you and anonymous reviewer's comments that help us improve our work scientifically.

I have added the word denitrification in the abstract.

Please find below our responses to the comments below, and also the manuscript with track changes.

With kind regards  
Arezoo Taghizadeh-Toosi  
Researcher  
Aarhus University

Associate Editor Decision: Publish subject to minor revisions (review by editor) (14 Oct 2019) by Ji-Hyung Park

Comments to the Author:

Dear Authors,

Thank you for your efforts to conduct additional statistical analyses and prepare detailed responses to the reviewer comments on your revised manuscript.

I am pleased to let you know that your manuscript can be published after some minor revisions (mostly technical corrections) as detailed below.

- Abstract, lines 20-22: Please provide "ranges" (either XX-XX or from XX to XX).

Response: We have added the ranges and text is read as: *"Equivalent soil gas phase concentrations of N<sub>2</sub>O at grassland sites varied between 0 and 25  $\mu\text{L L}^{-1}$  except for a sampling after slurry application at one of the sites in spring with a maximum of 560  $\mu\text{L L}^{-1}$  at 1 m depth. At the two potato sites the levels of below-ground N<sub>2</sub>O concentrations ranged from 0.4 to 2270  $\mu\text{L L}^{-1}$ , and from 0.1 to 470  $\mu\text{L L}^{-1}$ , respectively, in accordance with the higher soil mineral N availability at grassland sites."*

- Abstract, lines 30-31: This should be a concluding (or at least summarizing) remark. Please specify what "possible pathways of N<sub>2</sub>O production in spring and autumn periods" you would like to highlight.

Response: The summarizing remark has been added as : *"Based on the statistical graphical models and the tentative estimates of reduction capacities, FeS<sub>2</sub> oxidation was unlikely to be important for N<sub>2</sub>O emissions. Instead, archaeal ammonia oxidation in combination with either chemodenitrification or nitrifier denitrification were considered as plausible pathways of N<sub>2</sub>O production in spring, whereas in the autumn heterotrophic may have been more important at arable sites."*

- Conclusions, line 606: for grassland "also" during the autumn?

Response: "also" has been added.

- Table 2: Please describe in the table caption what test was used and what significant differences (between what?) different letters represent.

Response: Table 2 caption has been updated as it requested.

- Fig. 1: To guide readers on the location of your study sites in Demark or Europe, please add more geographical information (e.g., any close city name, cardinal directions,,,) on the main site map

Response: We have added the map of Denmark as well as indicating of the study place. Copenhagen as a capital has been marked in the map as well.

# Regulation of N<sub>2</sub>O emissions from acid organic soil drained for agriculture

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## Abstract

Organic soils drained for crop production or grazing land are agroecosystems with potentially high, but variable emissions of nitrous oxide (N<sub>2</sub>O). The present study investigated the regulation of N<sub>2</sub>O emissions in a raised bog area drained for agriculture, which is classified as potentially acid sulfate soil. We hypothesised that pyrite (FeS<sub>2</sub>) oxidation was a potential driver of N<sub>2</sub>O emissions through microbially mediated reduction of nitrate (NO<sub>3</sub><sup>-</sup>). Two sites with rotational grass, and two sites with a potato crop, were equipped for monitoring of N<sub>2</sub>O emissions and soil N<sub>2</sub>O concentrations at 5, 10, 20, 50 and 100 cm depth during weekly field campaigns in spring and autumn 2015. Further data acquisition included temperature, precipitation, soil moisture, water table (WT) depth, and soil NO<sub>3</sub><sup>-</sup> and ammonium (NH<sub>4</sub><sup>+</sup>) concentrations. At all sites, the soil was acidic with pH ranging from 4.7 to 5.4. Spring and autumn monitoring periods together represented between 152 and 174 d, with cumulative emissions of 4–5 kg N<sub>2</sub>O-N ha<sup>-1</sup> at sites with rotational grass and 20–50 kg N<sub>2</sub>O-N ha<sup>-1</sup> at sites with a potato crop. Equivalent soil gas phase concentrations of N<sub>2</sub>O at grassland sites mostly ranged varied between up to 0.4 and around 1.2–1.5 μL L<sup>-1</sup> at grassland sites except for a sampling after slurry application at one of the sites in spring with a maximum of 560 μL L<sup>-1</sup> at 1 m depth. At the two potato sites the levels of below-ground N<sub>2</sub>O concentrations ranged from 0.4 to 2270 μL L<sup>-1</sup>, and from 0.1 to 470 μL L<sup>-1</sup>, respectively, and up to several hundred μL L<sup>-1</sup> at potato sites, in accordance with the higher soil mineral N availability at grassland sites. Statistical analyses using graphical models showed that soil N<sub>2</sub>O concentration in the capillary fringe (i.e., the soil volume above the water table influenced by tension saturation) was the strongest predictor of N<sub>2</sub>O emissions in spring and, for grassland sites, also in the autumn. For potato sites in autumn, there was evidence that NO<sub>3</sub><sup>-</sup> availability in the top soil and temperature were the main controls on N<sub>2</sub>O emissions. Chemical analyses of intact soil cores from 0–1 m depth, collected at adjacent grassland and potato sites, showed that the total reduction capacity of the peat soil (assessed by cerium (IV) reduction) was much higher than that represented by FeS<sub>2</sub>, and the concentrations of total reactive iron (TRFe) were higher than those of FeS<sub>2</sub>. Based on the statistical graphical models and the tentative estimates of reduction capacities, FeS<sub>2</sub> oxidation was unlikely to be important for N<sub>2</sub>O emissions. Instead, archaeal ammonia oxidation in combination with either chemodenitrification or nitrifier denitrification were

considered as plausible Possible pathways of N<sub>2</sub>O production in spring, whereas in ~~and the~~ and autumn periods heterotrophic denitrification may have been more important, and the potential sources of N<sub>2</sub> are further discussed at arable sites.

**Key words:** Drained peat, potentially acid sulfate soil, rotational grass, potato, nitrous oxide, reactive iron

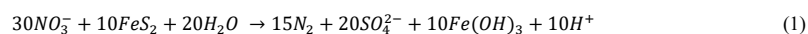
## 1 Introduction

Worldwide, 25.5 million ha of organic soils have been drained for agricultural use, mainly as cropland (Tubiello et al., 2016), and this accelerates decomposition of soil organic matter and net carbon (C) and nitrogen (N) mineralisation above the water table (WT) (Schothorst, 1977). Drained organic soils are significant net sources of greenhouse gas (GHG) emissions as carbon dioxide (CO<sub>2</sub>) and nitrous oxide (N<sub>2</sub>O) (Goldberg et al., 2010; Maljanen et al., 2003). A recent supplement to the 2006 IPCC Guidelines for National Greenhouse Gas Inventories on Wetlands (IPCC, 2014) proposed average annual emission factors of 4.3 and 8.2 kg N<sub>2</sub>O-N ha<sup>-1</sup> yr<sup>-1</sup> for temperate grassland on drained organic soil with low and high nutrient status, respectively, and an emission factor of 13 kg N<sub>2</sub>O-N ha<sup>-1</sup> yr<sup>-1</sup> for cropland. For soil C losses, the emission factors proposed for these three land use categories were between 5.3 and 7.9 Mg CO<sub>2</sub>-C ha<sup>-1</sup> yr<sup>-1</sup> (Hiraishi et al., 2014). Thus, while CO<sub>2</sub> emissions are overall more important, site conditions appear to be more critical for N<sub>2</sub>O.

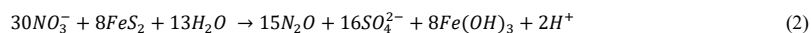
Site conditions are defined by land use, management, inherent soil properties and climate (Mander et al., 2010; Leppelt et al., 2014). Both WT drawdown (Aerts and Ludwig, 1997) and WT rise (Goldberg et al., 2010) may enhance N<sub>2</sub>O emissions, but such effects depend on soil N status (Martikainen et al., 1993; Aerts and Ludwig, 1997). Maljanen et al. (2003) found that WT, CO<sub>2</sub> emissions and temperature at 5 cm depth explained 55% of the observed variability in N<sub>2</sub>O emissions during a two-year field study on a drained organic soil, whereas the response to N fertilisation was limited, and they suggested that N released by soil organic matter mineralisation was the main source of N<sub>2</sub>O. In a study comparing GHG emissions from organic soil with different land uses in three regions of Denmark (in total eight site-years), Petersen et al. (2012) also found that site conditions such as WT, pH and precipitation contributed significantly to explain N<sub>2</sub>O emission dynamics. Among sites with arable crops in three regions, two sites had N<sub>2</sub>O emissions corresponding to, respectively, 38 and 61 kg N ha<sup>-1</sup> yr<sup>-1</sup>; both of these sites showed distinct seasonal patterns with the highest emissions in spring and autumn periods, whereas emissions at the third site were lower (6.4 kg N<sub>2</sub>O-N ha<sup>-1</sup> yr<sup>-1</sup>) and much less variable. Notably, WT depth at the two sites with seasonal patterns of N<sub>2</sub>O emission fluctuated between 10-30 and 10-120 cm depth, whereas WT depth at the third site remained at 90-125 cm depth throughout the 14-month monitoring period.

Several processes can lead to N<sub>2</sub>O formation in acid organic soil: biotic processes include ammonia (NH<sub>3</sub>) oxidation to nitrite (NO<sub>2</sub><sup>-</sup>) by archaea or bacteria (Herrmann et al., 2012; Herold et al., 2012; Stieglmeier et al., 2014), as well as nitrifier denitrification and heterotrophic denitrification by bacteria or fungi (Liu et al., 2014; Maeda et al., 2015; Wrage-Mönnig et al., 2018). The recently discovered process comammox by *Nitrospira* sp. is also a potential, but probably minor, source of N<sub>2</sub>O (Kits et al., 2019; Palomo et al., 2019). Abiotic N<sub>2</sub>O production can occur through

chemodenitrification (Van Cleemput and Samater, 1996; Jones et al., 2015) or abiotic codenitrification (Spott et al., 2011). The two regions showing extreme N<sub>2</sub>O emissions from arable soil had both developed from marine forelands and were categorised as potentially acid sulfate soil, i.e., saturated to poorly drained soil containing pyrite (FeS<sub>2</sub>) that, upon oxidation, may lead to acid production in excess of the soil's neutralising capacity (Madsen and Jensen, 1988). The capillary fringe of organic soils represents an interface between saturated and unsaturated soil conditions, in which the extent of tension saturation depends on pore size distribution (Gillham, 1984). Previously, it has been speculated that oxidation and reduction of iron sulfides could influence N transformations during periods with changing groundwater level (Petersen et al., 2012). Drainage promotes oxidation of FeS<sub>2</sub>, a process which may be linked to microbially mediated nitrate (NO<sub>3</sub><sup>-</sup>) reduction (Jørgensen et al., 2009; Torrento et al., 2010). The complete reduction of NO<sub>3</sub><sup>-</sup> to dinitrogen (N<sub>2</sub>) can proceed as follows:



However, in the capillary fringe residual oxygen (O<sub>2</sub>) or, alternatively, the acidification produced by FeS<sub>2</sub> oxidation, could favour incomplete denitrification with accumulation of the intermediate N<sub>2</sub>O (Torrento et al., 2010):



Nitrate reduction *via* the reaction described in Eq. 2 could potentially have contributed to the very high N<sub>2</sub>O emissions reported previously from two arable sites where groundwater sulfate concentrations were also consistently high (Petersen et al., 2012).

Here, we studied four agricultural sites within one of the regions previously investigated by Petersen et al. (2012), i.e., a raised bog area with acid soil conditions. This study included two sites with rotational grass and two sites with a potato crop, and monitoring took place in spring and autumn periods, where high emissions of N<sub>2</sub>O occurred in previous studies (Petersen et al., 2012; Kandel et al., 2018). We hypothesised that FeS<sub>2</sub> oxidation coupled with NO<sub>3</sub><sup>-</sup> reduction was a possible driver of N<sub>2</sub>O emissions. It was further hypothesised that N<sub>2</sub>O emissions would vary with site conditions affecting denitrification (mineral N availability, rainfall, WT depth and temperature).

## 2 Materials and methods

### 2.1 Study sites

The sites investigated were located in Store Vildmose, which is a 5,000 ha raised bog in northern Jutland, Denmark. The area was, until 150 years ago, the largest raised bog in Denmark, and largely unaffected by human activity. The bog overlies a marine plain formed by the last marine transgression; the sea retreated around 8000 BC, and peat later developed in wet parts of the landscape, attaining a maximum depth of 4.5 to 5.3 m in central parts of the bog (Kristensen, 1945). Between 1880 and 2010, the peat has generally subsided by at least 2 m due to drainage for agriculture or peat excavation (Regina et al., 2016), and today the peat depth is mostly 1-2 m, but in some locations even less (Kandel et al., 2018). The peat and underlying sand is acidic and has been categorised as a potentially acid sulfate soil (Madsen and Jensen, 1988). According to Kandel et al. (2018), the peat at 0-25 cm depth in arable soil in this area has a high degree of humification, which corresponds to H8 on the Von Post scale.

Four sites were selected along an East-West transect (Figure 1a). One arable site (*ARI*) was in a field cropped with second-year potato in 2015, while an adjacent site (*RG1*) in a neighbouring field had second-year rotational grass; these two sites were also represented in the study of Petersen et al. (2012) as sites *N-AR* and *N-RG*, respectively. Land use treatments (i.e., potato and rotational grass) were replicated at sites in other fields and will be referred to as *AR2* and *RG2*. Site *AR2* was located 4.6 km to the west, and site *RG2* was located 1.7 km to the east of the paired *ARI-RG1* sites (Figure 1a and S1).

## 2.2 Experimental design

In January 2015, an area of 10 m × 24 m was selected each site. Sampling positions were georeferenced using a Topcon HiPer SR geopositioning system (Livermore, CA). On 25 February 2015, each site was fenced, and three 10 m × 8 m experimental blocks were defined (Figure 1b). Each site was further divided along its longitudinal axis to establish two 5 m × 24 m fertilisation subplots.

For monitoring of WT depth, piezometer tubes (Rotek A/S, Sdr. Felding, Denmark) were installed to 150 cm depth at the centre of each block. On either side of the piezometers, at 2.7 m distance, collars of white PVC (base area: 55 cm × 55 cm, height: 12 cm [*RG*] or 24 cm [*AR*]) were installed to between 5 and 10 cm depth (Figure 1). The higher collars used at *AR* sites were level with the ridges established around potato rows during the growth period. The collars, which were fixed to the ground by four 40 cm pegs, had a 4 cm wide flange extending outwards 2 cm from the top to support gas flux chambers. To prevent soil disturbance during gas sampling, platforms (60 cm × 100 cm) of perforated PVC were placed in front of each collar to create a boardwalk. The exact headspace of each collar was determined from 16 individual measurements of distance from the upper rim; this procedure was repeated whenever collars had been removed and reinstalled to accommodate field operations.

Sets of five stainless steel diffusion probes for soil gas sampling at 5, 10, 20, 50 and 100 cm depth were installed vertically within 0.5 m of the flux measurement positions in two blocks (Block 2 and 3) at sites *ARI* and *RG1*, while at sites *AR2* and *RG2* diffusion probes were installed only in Block 2. The stainless steel probes were constructed as described in detail by Petersen (2014), with a 10 cm<sup>3</sup> diffusion cell having a 3 mm diameter opening at the sampling depth covered by a silicone membrane, which was connected to the soil surface via two 18G steel tubes with Luer Lock fittings (Figure S1).

A HOBO Pendant Temperature Data Logger (Onset Computer Corp., Bourne, MA) was installed at 5 cm depth in Block 2 at each site. A mobile weather station (Kestrel 4500; Nielsen-Kellerman, Boothwyn, PA) was mounted at 170 cm height at site *RG1* for hourly recording of air temperature, barometric pressure, wind speed and direction, and relative humidity. Daily precipitation was recorded at <10 km distance from the monitoring sites at a meteorological station, from where data to fill a gap in air temperature were also obtained.

## 2.3 Management

135 Management within the fenced experimental sites followed the practices adopted by the respective farmers, e.g., with  
 respect to method of fertiliser application, grass cuts, potato harvest and soil tillage. One exception to this was N  
 fertilisation rate, since N fertiliser was only given to one of the two subplots in each block (Figure 1b). Fertilised  
 subplots of the *RG1* site received 350 kg ha<sup>-1</sup> NS 27-4 fertiliser on 16 April (DOY 106), corresponding to 94.5 kg N ha<sup>-1</sup>.  
 140 Site *RG2* was fertilised with 20-25 Mg ha<sup>-1</sup> acidified cattle slurry (pH 6) on 5 May (DOY 125), and again on 2 July  
 (DOY 183), each time corresponding to 90-110 kg total N ha<sup>-1</sup>. On the day of the second slurry application, *RG2* further  
 received 50 kg N ha<sup>-1</sup> as NS 27-4 fertiliser, which was applied by mistake to both fertilisation subplots. The *AR1* site  
 received 100 kg N ha<sup>-1</sup> as liquid NPS 20-3-3 fertiliser on 21 May (DOY 141), while the *AR2* site received 110 kg N ha<sup>-1</sup>  
 as NS 21-24 pelleted fertiliser on 30 April (DOY 120). The NS fertilisers contained equal amounts of ammonium  
 (NH<sub>4</sub><sup>+</sup>) and nitrate (NO<sub>3</sub><sup>-</sup>), while N in the NPS fertiliser was mainly as NH<sub>4</sub><sup>+</sup>.

145 At the *RG1* site, the grass was cut in late August, while at the *RG2* site the grass was cut in late June and on 9  
 September (DOY 252). Potato harvest at the *AR1* site took place in mid-September (DOY 258), with interruptions due  
 to heavy rainfall. At the *AR2* site, the potato harvest took place on 23 September (DOY 266).

## 2.4 Field campaigns

Based on the patterns of N<sub>2</sub>O emissions observed by Petersen et al. (2012), a monitoring program was conducted  
 150 during spring from 3 March (DOY 63) to 16 June (DOY 169), and during autumn from 3 September (DOY 245) to 10  
 November (DOY 314). Weekly measurement campaigns were conducted at each of the four sites insofar as field  
 operations permitted. Thus, during spring there were 14, 12, 14 and 15 weekly campaigns at the *RG1*, *AR1*, *RG2*, and  
*AR2* sites, respectively. During autumn there were 10, 10, 7 and 10 weekly campaigns at the *RG1*, *AR1*, *RG2*, and *AR2*  
 sites, respectively. Field trips included sampling at two sites, either *AR1* + *RG1* or *AR2* + *RG2*, and thus all four sites  
 155 were visited during two field trips on consecutive days. Campaigns included registration of weather conditions and WT  
 depth, soil sampling, soil gas sampling, and N<sub>2</sub>O flux measurements. With a few exceptions (DOY 169 and 265 at site  
*RG1*, DOY 132 and 314 at site *AR2*), each campaign was initiated between 9:00 and 12:00; the order of sites visited in  
 each trip alternated from week to week.

### 2.4.1 Climatic conditions

160 Air temperature, relative humidity and barometric pressure were logged at the weather station located at *RG1*. During  
 field campaigns, the WT depth was first determined in each of the three piezometers using a Model 101 water level  
 meter (Solinst; Georgetown, Canada). At *AR1* and *AR2*, WT depth in Block 3 was further recorded at 30-minute time  
 resolution for a period during autumn using MaT Level2000 data loggers (MedgeTech; Warner, NH, USA). Soil  
 temperatures at 5, 10 and 30 cm depth were measured in each block using a high precision thermometer (GMH3710,  
 165 Omega Newport, Deckenpfronn, Germany), and in addition continuous measurements of soil temperature at 5 cm depth  
 were collected in block 2 at each site using HOBO Pendant Temperature Data Loggers (Onset Computer Corp., Bourne,  
 MA).

### 2.4.2 Soil sampling

During all field campaigns, soil samples were collected separately from fertilised and unfertilised subplots by random sampling of six 20 mm-diameter cores to 50 cm depth (two per block). Each core was split into 0-25 and 25-50 cm depth, and the six subsamples from each depth were pooled. The pooled samples were transported back to the laboratory in a cooling box and stored at -20°C for later analysis of mineral N and gravimetric water content.

On 23 April (DOY 113), and again on 2 September (DOY 245), undisturbed soil cores (50 mm diameter, 30 cm segments) were collected to 1 m depth within 1 m distance from the positions of flux measurements in Block 3 of sites *RG1* and *AR1* (cf. Figure 1b). A stainless steel corer (04.15 SA/SB liner sampler, Eijkelkamp, Giesbeek, Netherlands) equipped with a transparent plastic sleeve was used. The steel corer's lower end was capped with a 4 cm long cutting head, and hence sampling depths were 0 to 30 cm, 34 to 64 cm and 68 to 98 cm. The intact cores were capped and sealed, and transported in a cooling box to the laboratory, where they were stored at -20°C until analysis (see section 2.4.5).

#### 2.4.3 Soil gas sampling

Soil gas samples were collected in 6 mL pre-evacuated Exetainers (Labco Ltd, Lampeter, UK) as described by Petersen (2014) and demonstrated in Figure S2. In brief, the diffusion probes were flushed *via* the inlet tube with 10 mL N<sub>2</sub> containing 50 µL L<sup>-1</sup> ethylene (AGA, Enköping, Sweden) as a tracer, which displaced the gas in the diffusion cell, though with some mixing of sample and flushing gas. A three-way valve, mounted on the outlet tube, was fitted with a 10 mL glass syringe and an Exetainer. The displaced gas was collected in the glass syringe, and from the glass syringe the 10 mL soil gas sample could be transferred to the Exetainer. After gas sampling, the probe was flushed with 2 × 60 mL N<sub>2</sub> to remove ethylene, and the Luer Lock fittings were capped. Samples of the N<sub>2</sub>/ethylene gas mixture used for sample displacement were also transferred directly to Exetainers for gas chromatographic analysis ( $n = 3$ ) as reference for the calculation of dilution factors (Petersen, 2014). Sampling for soil gas was done in parallel with flux measurements, except when equipment had to be removed during periods with field operations. Due to damage of some probes during spring, it was decided to discontinue soil gas sampling in the unfertilised grassland subplot *RG2-NF*, which had by mistake received fertiliser on DOY 183.

#### 2.4.4 Nitrous oxide flux measurements

Gas fluxes were measured with static chambers (60 cm × 60 cm × 40 cm) constructed from 4-mm white PVC, and equipped with a closed-cell rubber gasket (Emka Type 1011-34; Megatrade, Hvidovre, Denmark) as sealing during chamber deployment. Chambers were further equipped with a 12V fan (RS Components, Copenhagen, Denmark) for headspace mixing that was connected to an external battery (Yuasa Battery Inc.; Laureldale, PA), as well as a vent tube with outlet near the ground to minimise effects of wind (Conen and Smith, 1998; Hutchinson and Mosier, 1981). Also, chambers were equipped with an internal temperature sensor (Conrad Electronic SE; Hirschau, Germany), and a butyl rubber septum on top of each chamber for gas sampling. Handles attached to the top were used for straps fixing the chamber firmly against the collar. Gas samples (10 mL) were taken with a syringe and hypodermic needle immediately after chamber deployment, and then 15, 30, 45 and 60 minutes after closure. Gas samples were transferred to 6 mL Exetainer vials, leaving a 4 mL overpressure.



#### 2.4.5 Soil analyses

205 Soil samples collected during the weekly campaigns were sieved (6 mm) and subsampled for determination of soil  
mineral N and gravimetric water content. Approximately 10 g field moist soil was mixed with 40 mL of 1 M potassium  
chloride (KCl) and shaken for 30 min, and then filtered through 1.6 µm glass microfibre filters. Concentrations of  $\text{NH}_4^+$   
and  $\text{NO}_2^- + \text{NO}_3^-$  in filtered KCl extracts were determined by autoanalyser (Model 3; Bran+Luebbe GmbH, Norderstedt,  
Germany) using standard colorimetric methods (Keeney and Nelson, 1982). Gravimetric soil water content was  
210 determined after drying of soil samples at 80°C for 48 hours.

Additional soil characteristics were determined on the intact soil cores collected in April and September at *AR1* and  
*RG1*. Five cm sections were subsampled from selected depths and analysed for water content, pH, electrical  
conductivity (EC), total soil organic C and N, and  $\text{NO}_2^-$ . Soil pH and EC were measured with a Cyberscan PC300  
(Eutech Instruments; Singapore) in a soil:water solution (1:2.5, w/v). Total soil organic C and total N were measured by  
215 high temperature combustion with subsequent gas analysis using a vario MAX cube CN analyser (Elementar  
Analysensysteme GmbH; Langenselbold, Germany). Soil  $\text{NO}_2^-$ -N concentrations were analysed in soil:water extracts  
(1:5, w/v) using a modified Griess-Ilosvay method (Keeney and Nelson, 1982). Total organic C and total N were further  
determined in bulk soil samples (0-25 cm and 25-50 cm depth) collected at *RG2* and *AR2* in the same weeks as  
sampling of intact cores took place at *AR1* and *RG1*.

220 The concentration of total reactive Fe (TRFe) at selected depth intervals was determined in the samples from both  
April and September samplings of intact soil cores. The analysis of TRFe was done using a dithionite-citrate extraction  
(Carter and Gregorich, 2007; Thamdrup et al., 1994) followed by  $\text{Fe}^{2+}$  analysis with the colorimetric ferrozine method,  
which includes hydroxylamine as reducing agent (Viollier et al., 2000). The extraction dissolves free (ferric) Fe oxides  
(except magnetite,  $\text{Fe}_3\text{O}_4$ ), as well as (ferrous) Fe in FeS, but not  $\text{FeS}_2$ .

225 The intact soil cores from the September sampling were further analysed for acid volatile sulfides (AVS) and  
chromium reducible sulfur (CRS) as indices of FeS and  $\text{FeS}_2$ , respectively. Quantification of AVS and CRS was based  
on passive distillation adapted from Ulrich et al. (1997) and Burton et al. (2008). Briefly, 0.5 g soil and a trap with 4  
mL alkaline Zn-acetate solution (5%) was placed in 120 mL butyl-stoppered (and crimp-sealed) serum bottles, which  
were evacuated (1 kPa) and pressurised with  $\text{N}_2$  (150 kPa) in three cycles to remove  $\text{O}_2$ , eventually leaving the  
230 headspace with  $\text{N}_2$  at atmospheric pressure. Acid volatile sulfide (primarily FeS) was liberated and trapped as ZnS after  
injection of 12 mL anoxic 2 M HCl followed by sonication (0.5 h) and incubation (24 h) on a rotary shaker (20°C).  
Using the same approach with replicate soil samples, combined AVS and CRS (primarily elemental S and  $\text{FeS}_2$ ) was  
trapped after injection of 12 mL 1 M  $\text{Cr}^{2+}$  in 2 M HCl, prepared by reduction of  $\text{CrCl}_3$  (Røy et al., 2014). Trapped  
sulfide (ZnS) in the two traps was measured colorimetrically using diamine reagent (Cline, 1969), and CRS was then  
235 calculated by difference.

Finally, the total reduction capacity of the peat at depths of 27-30 cm, 61-64 cm and 95-98 cm was determined. In  
brief, a suspension (soil:solution, 1:25; w/v) of oven dried (105°C) sieved soil (<2 mm) and 25 mM cerium (IV) sulfate  
reagent,  $\text{Ce}(\text{SO}_4)_2$  in 5% sulfuric acid ( $\text{H}_2\text{SO}_4$ ), was shaken horizontally for 24 h at 275 revolutions per minute (rpm).

240 After centrifugation at 2,000 rpm, residual Ce(IV) was measured by end-point titration using a solution of 5 mM FeSO<sub>4</sub> in 5% H<sub>2</sub>SO<sub>4</sub>. The amount of reduced compounds was calculated and expressed as meq kg<sup>-1</sup>.

#### 2.4.6 Gas analyses

245 Nitrous oxide concentrations were analysed on an Agilent 7890 gas chromatograph (GC) combined with a CTC CombiPal auto-sampler (Agilent, Nærum, Denmark). The instrument had a 2 m back-flushed pre-column with Hayesep P connected to a 2 m main column with Poropak Q. From the main column, gas entered an electron capture detector (ECD). The carrier was N<sub>2</sub> at a flow rate of 45 mL min<sup>-1</sup>, and Ar-CH<sub>4</sub> (95%/5%) at 40 mL min<sup>-1</sup> was used as make-up gas. Temperatures of the injection port, columns and ECD were 80, 80 and 325°C, respectively. Concentrations were quantified with reference to synthetic air and a calibration mixture containing 2013 nL L<sup>-1</sup> N<sub>2</sub>O. Soil profile N<sub>2</sub>O concentrations were frequently at several hundred μL L<sup>-1</sup>; linearity of the EC detector response was ascertained up to 1600 μL L<sup>-1</sup>, but the entire range was not included in analytical runs as a standard practice, and therefore the higher equivalent gas phase concentrations are relatively uncertain.

250 Ethylene concentrations in soil gas samples and flushing gas were analysed following a separate injection with an extended run time. All GC settings were as described above, except that run time was different, and gas from the main column was directed to a flame ionisation detector supplied with 45 mL min<sup>-1</sup> H<sub>2</sub>, 450 mL min<sup>-1</sup> air, and 20 mL min<sup>-1</sup> N<sub>2</sub>; the detector temperature was 200°C.

#### 255 2.5 Data processing and statistical analyses

Equivalent soil gas phase concentrations of N<sub>2</sub>O were calculated assuming full equilibrium (Petersen, 2014) according to Eq. 3:

$$c_s = c_m / \left[ 1 - \frac{(V + d_{out})a_m}{(V - d_{in})a_F} \right] \quad (3)$$

260 where  $c_s$  is the concentration of N<sub>2</sub>O in the diffusion cell and  $c_m$  the observed concentration (μL L<sup>-1</sup>);  $V$ ,  $d_{in}$  and  $d_{out}$  are the volumes of the diffusion cell, inlet tube and outlet tube, respectively (L);  $a_m$  is the concentration of the tracer ethylene in the gas sample analysed (μL L<sup>-1</sup>); and  $a_F$  is the concentration of ethylene in the flushing gas (μL L<sup>-1</sup>).

265 Nitrous oxide mixing ratios were converted to units of mass per volume using the ideal gas law and values of pressure and air temperature recorded by the weather station. Individual N<sub>2</sub>O fluxes were calculated in R (version 3.2.5, R Core Team, 2016) using the package HMR (Pedersen et al., 2010). This program analyses non-linear concentration-time series with a regression-based extension of the model of Hutchinson and Mosier (1981), and linear concentration-time series by linear regression (Pedersen et al., 2010). Statistical data ( $p$  value, 95% confidence limits) are provided by HMR for both categories of fluxes. The choice to use a linear or non-linear flux model was made based on scatter plots and the statistical output.

270 The temporal dynamics of N<sub>2</sub>O fluxes were analysed by season for individual site-crop combinations using a  
 generalised linear mixed model defined with the identity link function, the gamma distribution (see Jørgensen and  
 Labouriau, 2012; McCullagh and Nelder, 1989), and Gaussian random components. The model contained a fixed effect  
 representing the interaction between fertilisation and sampling day, and random effects representing site and sampling  
 position. The model for daily N<sub>2</sub>O emission was used to estimate cumulative emissions by integrating the flux curves  
 over time. Treatment effects were then analysed by specially designed linear contrasts as described in detail by Duan et  
 275 al. (2017), who showed that models with untransformed responses (when using adequate distributions) allow simple  
 statistical inference of the time-integrated N<sub>2</sub>O emissions. WT levels at different time points were compared For each  
 site-crop combination by permutation tests (Good, 2005) with 9999 permutations respecting the block structure.

The dependence structure of variables that were potential drivers of N<sub>2</sub>O fluxes were studied using a class of  
 multivariate models called “graphical models” (Whittaker, 1990, see also Labouriau et al., 2008a,b; and Lamandé et al.,  
 280 2011 for applications in soil science). These models represent the dependence of variables using an undirected graph  
 (not to be confounded with the word “graph” used to refer to a plot), which is a mathematical structure composed of  
*vertices*, represented by points, and *edges* connecting pairs of vertices, represented by lines connecting points,  
 according to the convention explained below. In graphical models, the variables of interest are the vertices of the graph  
 (represented as labelled points). Here the variables used were: soil temperature at 5 cm depth (Temp5); soil temperature  
 285 at 30 cm depth (Temp30); NH<sub>4</sub><sup>+</sup> and NO<sub>3</sub><sup>-</sup> concentrations in the top soil (AmmoniumT and NitrateT); N<sub>2</sub>O  
 concentration of the soil gas diffusion probe closest to, but above the WT, i.e., in the capillary fringe (N<sub>2</sub>OWT); and  
 finally, the N<sub>2</sub>O flux (N<sub>2</sub>O-flux). The dependence structure of these variables was characterised by the conditional  
 covariances between each pair of variables given the other variables. Those conditional covariances were  
 simultaneously estimated using the available data according to a statistical model. The graph representation of the  
 290 model is constructed by connecting the pairs of vertices (i.e., pairs of variables) by an edge when the conditional  
 correlation of the two corresponding variables, given all the other variables, is different from zero. It is possible to show  
 that two variables directly connected in the graph carry information on each other that is not already contained in the  
 other variables (see Whittaker, 1990, Jørgensen and Labouriau, 2012). Moreover, the absence of an edge connecting  
 two vertices indicates that (even a possible) association between the two corresponding variables can be entirely  
 295 explained by the other variables. According to the general theory of graphical models, if two groups of variables, say A  
 and B, are separated in the graph by a third group of variables, say C (i.e., every path connecting an element of A with  
 an element of B necessarily contains an element of C), then A and B are conditionally uncorrelated given C (see  
 Lauritzen, 1999). This property, called the separation principle, was used below to draw non-trivial conclusions on the  
 interrelationship between N<sub>2</sub>O-flux related variables. The graphical models were inferred by finding the model that  
 300 minimised the BIC (Bayesian information criterion, i.e., a penalised version of the likelihood function) as implemented  
 in the R package gRapHD (Abreu et al., 2010). This inference procedure yields an optimal representation of the data in  
 the sense that the probability of correct specification of the model, when using this penalisation, tends to one as the  
 number of observations increases (see Houghton, 1988). The confidence intervals for the conditional correlations were  
 obtained by a non-parametric bootstrap procedure (Davidson and Hinkley, 1997) with 10,000 bootstrap samples.  
 305 Separate analyses were conducted for each combination of site, crop and season.

### 3 Results

#### 3.1 Climatic conditions

In 2015, the annual mean air temperature in the area of this study was 8.7°C, and annual precipitation was 920 mm.

310 This was slightly above the ten-year (2009-2018) average temperature of 8.3°C, and well above the ten-year average annual precipitation of 798 mm. During the spring monitoring period, the daily mean air temperature varied between 1 and 15°C, with an increasing trend over the period, and total rainfall was 220 mm. During the autumn monitoring period, the daily mean air temperature declined from 15 to 5°C, and total rainfall was 148 mm; the most intense daily rain events during spring and autumn were 16.9 and 33.2 mm, respectively.

315 Soil temperature at 5 cm depth showed a clear diurnal pattern (Figure S3), but at all four sites the temperature at the time of chamber deployment was close to the daily mean temperature at this depth. Thus, across the four sites the average deviation ranged from 0.2 to 0.9°C, and the largest deviations on a single day were -2.0 and 2.1°C, respectively.

#### 3.2 Soil characteristics

320 Several soil characteristics were determined by analyses of intact cores collected in late April (DOY 113) 2015 (Table 1). At all sites the soil was acidic, with pH ranging from 4.7 to 5.4. At the paired sites *ARI* and *RGI*, a weak decline in pH was indicated at 40-50 cm depth. Electrical conductivity at *ARI* and *RGI* sites ranged from 0.15 to 0.91 mS cm<sup>-1</sup>, with no obvious trends in the data; the highest value (0.91 mS cm<sup>-1</sup>) occurred at site *ARI* at 93-98 cm in a layer dominated by sand underlying the peat.

325 The organic matter composition of soil profiles at the four sites varied. Total organic C concentrations at sites *ARI* and *RGI* were 34-43% in the upper 0-40 cm, but then dropped to only 0.3-0.6% at c. 1 m depth in the sand. The peat was amorphous and well-decomposed at 0-20 cm depth, while the underlying peat contained intact plant debris. At site *RG2*, the process of peat degradation was evident even at 0-50 cm depth, where TOC concentrations only just met the requirements for being defined as an organic soil; the organic C content was below 20 and 10% at 0-25 and 25-50 cm depth, respectively. Site *AR2* was characterised by a uniform peat layer (33-38% organic C) at 0-50 cm depth. Across 330 all sites, the C:N ratios ranged between 14 and 26 in the organic soil layers.

Two iron sulfide fractions, as well as total reactive iron, were quantified. Acid volatile sulfide ranged from 1.7 to 4.9 µg S g<sup>-1</sup> soil across the four sites and showed no clear relationship with soil depth. This was also the case for CRS, which ranged from 24 to 155 µg S g<sup>-1</sup> dry weight soil. Total reactive Fe (TRFe) concentrations in soil profiles from sites 335 *ARI* and *RGI* ranged from 1.19 to 4.99 mg g<sup>-1</sup> dry weight soil at 0-50 cm depth, and hence concentrations of reactive Fe were up to 1500 times higher than concentrations of Fe in AVS (assuming this was FeS), and 25-120 times higher than Fe in CRS (assuming this was FeS<sub>2</sub>). At sites *ARI* and *RGI*, TRFe declined below 20 cm depth and was close to zero in

the sand below the peat layer (Table 1). The highest concentrations of TRFe at sites *RG1* (Figure 2b) and *ARI* (Figure 2d) occurred at 20 cm depth on 23 April (DOY 113). At site *ARI*, a sink for TRFe at 40-60 cm depth was indicated and, disregarding Depth 6 (93-98 cm), the concentration of TRFe at Depth 5 (47.5-52.5 cm) was significantly lower than concentrations at more shallow depths ( $p < 0.05$ ). Differences in TRFe concentration were observed at site *RG1*, and any differences in the distribution of TRFe between seasons were probably also minor (not tested). There was a strong correlation between TRFe and TOC across all sites ( $r = 0.88$ ,  $n = 16$ ).

The total reduction capacity was determined by a wet oxidation procedure using  $\text{Ce}(\text{SO}_4)_2$ . At both *ARI* and *RG1*, the total reductive capacity of the peat at 27-30 cm depth was outside the range of the analytical method at  $>11,500 \text{ meq kg}^{-1}$ . The reduction capacity dropped to around  $1000 \text{ meq kg}^{-1}$  at 60 to 65 cm depth with a declining organic matter content, and to  $50\text{-}100 \text{ meq kg}^{-1}$  in the sandy layer at 93-98 cm depth.

### 3.3 Soil mineral N dynamics

Soil concentrations of  $\text{NH}_4^+$  and  $\text{NO}_3^-$  at 0-25 and 25-50 cm depth were determined in connection with field campaigns (Tables S1-S4). Since subsamples were pooled for analysis, only a qualitative description of the effects of treatments and temporal dynamics is possible. The residence time for mineral N in the soil solution appeared to be longer at *AR* compared to *RG* sites. At *AR* sites, there was an accumulation of mineral N (Table S2, S4) at both depth intervals during May that also occurred before N fertilisation. Mineral N concentrations were apparently greater at *ARI* compared to *AR2*, and at site *AR2* only  $\text{NO}_3^-$  accumulated. Following fertilisation,  $\text{NH}_4^+\text{-N}$  and  $\text{NO}_3^-\text{-N}$  concentrations were  $100\text{-}200 \mu\text{g g}^{-1}$  dry weight soil at all sites except *RG2* (Table S3), where acidified cattle slurry was applied. Nitrate accumulated at all sites in the weeks after fertilisation, and also there was evidence for some transport to 25-50 cm depth.

Nitrite-N concentrations were determined in soil profiles from the cores sampled at sites *RG1* and *ARI* on 23 April (DOY 113) and 2 September (DOY 245) 2015. At site *RG1*, fertilisation had taken place one week earlier, but a two-sample *t* test did not find evidence for an effect on  $\text{NO}_2^-$  availability ( $p = 0.19$ ), and therefore the results from both *ARI* and *RG1* subplots are presented together. In April, the average concentration of  $\text{NO}_2^-\text{-N}$  at both sites was highest (c.  $10 \mu\text{g g}^{-1}$  dry weight soil) around 40 cm depth and declined towards the surface and deeper layers (Figure 2a,c). However, due to heterogeneity of soil profiles, the differences between depths were not significant ( $p = 0.22$  for *RG1* and  $p = 0.06$  for *ARI*). A decline in  $\text{NO}_2^-\text{-N}$  concentration was indicated at 50 cm depth at site *ARI*, where a depletion of TRFe was indicated. However, there was also less organic matter (cf. TOC in Table 1), which may account for this difference. In September,  $\text{NO}_2^-\text{-N}$  concentrations were  $<1 \mu\text{g g}^{-1}$  dry weight soil at both sites, while the concentrations of TRFe were comparable to those in April.

### 3.4 Groundwater table dynamics

370 Across the four sites, WT changes ranged from 60 to 100 cm. During spring, WT depth at sites *RG1* and *ARI* varied  
between 17 and 81 cm, with a steady decline until the end of April, and by DOY 119 the WT depth was significantly  
(all  $p$  values  $< 0.05$ ) below that of all previous samplings except DOY 112 (Figures 3 and 4). Then followed a period  
with frequent rainfall, where WT fluctuated (no significant changes) around 60-80 cm depth. During the first half of  
September (DOY 246 to 259), rainfall caused the WT to rise from 80 to 40 cm depth ( $p < 0.05$ ; Figures 5 and 6). On  
375 two occasions (DOY 248 and 260) the WT depth rose to 20 cm depth and only gradually declined during the following  
days (data not shown). From mid-September (DOY 258) there followed a period with a gradual WT decline until early  
November (DOY 308), where upon the WT showed an increasing trend from 90 to 45 cm depth during a week with  
intense rainfall. At site *RG2* (Figure 3) the WT initially declined ( $p < 0.05$ ) and then remained mostly at 50-60 cm depth  
during spring, with a temporary rise to 30 cm depth on 3 June (DOY 139;  $p < 0.05$ ). In the autumn, sampling campaigns  
380 were resumed on DOY 245. By this time the WT was close to the surface following intense rainfall, but then declined  
( $p < 0.05$ ) to 80-100 cm in the sandy subsoil (Figure 5). The WT depth at site *AR2* declined initially ( $p < 0.01$ ) and then  
fluctuated between 45 and 60 cm depth during spring except for a transient increase ( $p = 0.05$ ) to 35 cm depth in early  
June (Figure 4). During autumn the WT rose ( $p = 0.05$ ) to the soil surface in September (DOY 260), and then gradually  
withdrew ( $p = 0.05$ ) until early November (DOY 307) when rainfall caused a c. 40 cm increase (Figure 6), as also  
385 observed at sites *RG1* and *ARI*.

### 3.5 Soil N<sub>2</sub>O concentration profiles

Equivalent gas phase concentrations of N<sub>2</sub>O in passive diffusion samplers were determined concurrently with gas  
sampling, and results are presented as contour plots (Figures 3-6; data in Table S5). Concentrations in many cases  
varied by several orders of magnitude between sites and sampling days, and between depths within individual profiles,  
390 and therefore a logarithmic grey scale is used to show gradients. The gaps in Figures 3-6 indicate periods, where  
diffusion probes could not be installed or were temporarily removed due to field operations.

[The concentrations of N<sub>2</sub>O at grassland sites varied between 0 and 25  \$\mu\text{L L}^{-1}\$  with an exception mentioned below.](#)

Under the rotational grass at site *RG1*, soil N<sub>2</sub>O concentrations during spring were mostly between 0.1 and 3  $\mu\text{L L}^{-1}$   
(Figure 3). A higher concentration (15  $\mu\text{L L}^{-1}$ ) was observed at 40-80 cm depth in the fertilised subplot around DOY  
395 139, but only at the lower position (Block 3) of the field plot. At site *RG2*, the concentrations of N<sub>2</sub>O in the soil during  
spring were generally similar to those of *RG1*, although there were more values in the 1-10  $\mu\text{L L}^{-1}$  concentration range  
in the unfertilised plot (Figure 3, Table S5). However, on 3 June (DOY 154) a much higher N<sub>2</sub>O concentration was  
observed in the fertilised part of the plot with a maximum of 560  $\mu\text{L L}^{-1}$  at 100 cm depth (i.e., well below the WT). Soil  
N<sub>2</sub>O concentrations in the unfertilised plot were also elevated around this time, but only to a maximum of 15  $\mu\text{L L}^{-1}$  and  
400 mainly near the soil surface.

The arable site *ARI*, with sampling positions located in a different field, but only 10-20 m from those of site *RG1*,  
had apparently very different soil N<sub>2</sub>O concentration dynamics during spring (Figure 4). There was a consistent  
accumulation of N<sub>2</sub>O at 50 and 100 cm depth where seasonal concentrations averaged 340 and 424  $\mu\text{L L}^{-1}$ , respectively.  
In contrast, at 5, 10 and 20 cm depth the average N<sub>2</sub>O concentrations were 10-30  $\mu\text{L L}^{-1}$ , and there was no clear

405 response to fertilisation on DOY 141 in terms of soil N<sub>2</sub>O accumulation. The soil N<sub>2</sub>O concentrations suggested that there was considerable within-site heterogeneity in soil conditions, as the highest concentrations were often observed in the unfertilised subplot. Maximum concentrations of N<sub>2</sub>O of nearly 1500  $\mu\text{L L}^{-1}$  were recorded at 50 cm depth on DOY 99, and a similar concentration at 100 cm depth on DOY 112, both in the unfertilised subplot. At site *AR2*, the highest soil N<sub>2</sub>O concentrations during early spring were consistently observed at 20 cm depth, but seemed to gradually decline  
410 to reach the background level of 0.3  $\mu\text{L L}^{-1}$  in mid-May (around DOY 130). In the unfertilised subplot, the N<sub>2</sub>O concentration reached 272  $\mu\text{L L}^{-1}$  at 20 cm depth following rainfall and with WT at 35 cm depth. With fertilisation, soil N<sub>2</sub>O concentrations at 10 cm depth reached nearly 400  $\mu\text{L L}^{-1}$  in mid-June. The observed accumulation of N<sub>2</sub>O near the soil surface was accompanied by increasing N<sub>2</sub>O emissions during this period (Figure 4).

During autumn, N<sub>2</sub>O concentrations in the soil profile at the *RG1* and *RG2* sites varied between 0 and 12  $\mu\text{L L}^{-1}$ ,  
415 with a tendency for higher concentrations at 10-20 cm depth (Figure 5). At site *RG1*, where both fertilised and unfertilised subplots could be sampled, this was apparently independent of fertilisation.

September was characterised by heavy rainfall (114 mm in total), and at site *AR1* a substantial rise in the WT from 80 to 40 cm depth was observed (Figure 6). Soil N<sub>2</sub>O concentrations showed a pattern with maxima at 10 and 100 cm depth through to DOY 266 (end of September), and after this time soil N<sub>2</sub>O accumulation rapidly declined concurrently  
420 with WT drawdown. Nitrous oxide concentrations equivalent to several hundred  $\mu\text{L L}^{-1}$  were measured even at 5 cm depth during this period, and the overall highest concentration recorded was 2270  $\mu\text{L L}^{-1}$  at 10 cm depth on DOY 266.  
During late autumn, the N<sub>2</sub>O concentration at 0-50 cm depth varied between 0 and 20  $\mu\text{L L}^{-1}$ , whereas at 100 cm depth it remained high at 100-850  $\mu\text{L L}^{-1}$ . At site *AR2*, the groundwater level was higher than at *AR1* and came close to the soil surface by mid-September (DOY 260). Soil N<sub>2</sub>O accumulated in both fertilised and unfertilised subplots following  
425 saturation of the soil, again the highest concentrations apparently occurred at 20 cm depth. A secondary increase was observed near the soil surface at the last sampling on DOY 314 in November, in response to a period with rainfall and a rapid WT rise.

### 3.6 Nitrous oxide emissions

The weekly sampling campaigns during spring and autumn showed that with few exceptions N<sub>2</sub>O emissions, expressed  
430 as average daily rates, were significantly higher at arable compared to grassland sites (Table 2). One exception was treatment *RG2-F* in spring, where a peak in N<sub>2</sub>O emissions was indicated ( $p = 0.06$ ) on DOY 154, and the flux was still elevated at the next two samplings (Figure 3). This high flux coincided with the accumulation of N<sub>2</sub>O in the soil profile described above, and it was the only time that an effect of fertilisation was observed. The grass in the fertilised subplot showed a clear visual response to fertilisation, which indicated that fertiliser N was effectively taken up by the sward.

435 At site *AR1* the N<sub>2</sub>O fluxes during early spring reached 2000-6000  $\mu\text{g N}_2\text{O m}^{-2} \text{h}^{-1}$  and were higher than in late spring (Figure 4). Since there was no effect of N fertilisation (cf. Table 2), the higher emissions were probably derived from soil N pools and caused by factors other than fertilisation. The potato field at site *AR2* showed a different pattern, with N<sub>2</sub>O fluxes remaining low during early spring, and for several weeks after fertilisation. Independent of

fertilisation, an increasing trend (not significant) was observed in June following a rise in WT, which was at 35 cm  
440 depth on DOY 154.

In the autumn, N<sub>2</sub>O fluxes from site *RG1* were consistently low (Figure 5). The first sampling at site *RG2* was on  
DOY 259 in mid-September, where a high flux of 3000 µg N<sub>2</sub>O m<sup>-2</sup> h<sup>-1</sup> was seen, which dropped to near zero within 1-2  
weeks. Nitrous oxide emissions at site *AR1* were high during September at 4000-10,000 µg N<sub>2</sub>O m<sup>-2</sup> h<sup>-1</sup> across the two  
445 N fertilisation treatments (Table S2), and subsequently declined ( $p < 0.05$ ) to near zero (Figure 6). High fluxes were  
observed on the first day of this monitoring period, DOY 246, at a time where WT depth was still at 40 to 80 cm depth.  
Instead the high N<sub>2</sub>O fluxes may have been triggered by saturation of the top soil after 10 and 22 mm rainfall the  
previous two days. Additional rainfall during the following days then was accompanied by a rise in WT. The  
subsequent decline in N<sub>2</sub>O emissions at *AR* sites coincided with WT draw-down and drainage of the top soil.

Cumulative N<sub>2</sub>O emissions were calculated for the 99-105 d monitoring period in spring, as well as the 47-69 d  
450 period in the autumn. Daily rates were surprisingly consistent in the two periods (Table 2), and therefore the following  
refers to cumulative emissions across both periods. At site *RG1*, the total emission was 4 kg N<sub>2</sub>O-N ha<sup>-1</sup> independent of  
fertilisation, whereas at *RG2* the fertilised and unfertilised subplots were different at 18 and 5 kg N<sub>2</sub>O-N ha<sup>-1</sup>,  
respectively. At *AR* sites with potato, there was no effect of N fertilisation, and the cumulative N<sub>2</sub>O emissions were 18-  
20 kg N<sub>2</sub>O-N ha<sup>-1</sup> at *AR2*, but much higher at 44-52 kg N<sub>2</sub>O-N ha<sup>-1</sup> at site *AR1*.

### 455 3.7 Interrelationships between driving variables of N<sub>2</sub>O production

Graphical models were used to study the dependence structure among selected soil variables and N<sub>2</sub>O fluxes.  
Interestingly, at both *RG* sites, and in both seasons, N<sub>2</sub>O flux consistently depended on N<sub>2</sub>O<sub>WT</sub>, i.e., soil N<sub>2</sub>O  
concentration in the capillary fringe (Figure 7a,b and 8a,b). This was also the case for both *AR* sites in spring (Figure  
7c,d). Where N<sub>2</sub>O<sub>WT</sub> separated N<sub>2</sub>O flux from other variables in the graph, this indicates according to the separation  
460 principle (section 2.5) that information about N<sub>2</sub>O<sub>WT</sub> rendered all the other variables uninformative with respect to N<sub>2</sub>O  
flux. For example, in the analysis of *AR1* in spring (Figure 7b), the variables N<sub>2</sub>O flux and Temp5 were not directly  
connected, and therefore any correlation between Temp5 and N<sub>2</sub>O flux could be completely explained by other  
variables.

The graphical model results were different for *AR* sites in the autumn (Figure 8c,d), where instead soil temperature  
465 at 30 cm depth and (site *AR1* only) NO<sub>3</sub><sup>-</sup>-N concentration in the top soil showed significant correlation with N<sub>2</sub>O flux ( $p$   
< 0.05). All other variables were unrelated to N<sub>2</sub>O flux, or could be accounted for by other variables.

## 4 Discussion

This study investigated seasonal dynamics of N<sub>2</sub>O emissions and soil conditions in an area, which has been designated  
470 as a hotspot for N<sub>2</sub>O emissions (Leppelt et al., 2014). Spring and autumn monitoring periods together covered between



152 and 174 d, and cumulative N<sub>2</sub>O emissions during these periods were in total 4-5 kg N<sub>2</sub>O-N ha<sup>-1</sup> for rotational grass if disregarding the two-week period after slurry application at *RG2-F*, and 20-50 kg N<sub>2</sub>O-N ha<sup>-1</sup> for arable sites with a potato crop. These numbers, representing < 6 month periods, thus confirmed previous results (Petersen et al., 2012) that annual N<sub>2</sub>O emissions from organic soil in this area are comparable to (*RG*), or clearly above (*AR*), the IPCC emission factors for drained organic soil of 8 and 13 kg N<sub>2</sub>O-N ha<sup>-1</sup> yr<sup>-1</sup> for nutrient rich grassland and cropland, respectively (IPCC, 2014). The area has been characterised as potentially acid sulfate soil (Madsen and Jensen, 1988), and a previous study showed groundwater sulfate concentrations in excess of 100 mg L<sup>-1</sup> (Petersen et al., 2012). We therefore hypothesised that NO<sub>3</sub><sup>-</sup> reduction coupled with FeS<sub>2</sub> oxidation could be a pathway of N<sub>2</sub>O formation in this acid organic soil,

Pyrite, measured as CRS, was quantified at selected depths (Table 1). Soil bulk density of the peat varied between 0.15 and 0.3 g cm<sup>-3</sup> (data not shown), and the total amount of CRS at 0 to 50 cm depth could therefore be estimated to 200-350 mmol FeS<sub>2</sub> m<sup>-2</sup>. The N<sub>2</sub>O emissions observed during spring and autumn monitoring periods constituted up to 145 mmol N m<sup>-2</sup> in total (site *ARI*), and it is thus theoretically possible that the process described by Eq. 2 contributed to emissions of N<sub>2</sub>O. However, the FeS<sub>2</sub> concentration (0.4-2.4 mmol kg<sup>-1</sup>) represented a minor part of the total reduction capacity (>11,500 meq kg<sup>-1</sup> at 27-30 cm depth). Also, average concentrations of total reactive Fe were 25-120 times higher than that of FeS<sub>2</sub> (though less in terms of reduction equivalents). It is therefore likely that reducing agents other than FeS<sub>2</sub> were important, a conclusion that was later supported by a laboratory study in which peat amended with FeS<sub>2</sub> did not show enhanced N<sub>2</sub>O production (Taghizadeh-Toosi et al., 2019). Other possible drivers of N<sub>2</sub>O emissions were therefore considered.

#### 4.1 Environmental drivers of N<sub>2</sub>O emissions

A limited number of potential drivers (rainfall, temperature, soil mineral N and soil concentrations of N<sub>2</sub>O) were monitored to help explain N<sub>2</sub>O emission dynamics. Soil N<sub>2</sub>O concentration profiles showed complex patterns where, for example, the highest concentrations were sometimes observed above, and sometimes below the WT depth at both *RG* (Figure 3) and *AR* sites (Figure 4). Fertilisation in spring was associated with higher concentrations of N<sub>2</sub>O below the WT depth at sites *RG2* and *AR2*, indicating which indicated downward transport of fertiliser N, but this was not reflected in elevated N<sub>2</sub>O emissions. ~~In~~ The reason may be that in wet soil, the time required to reach a steady state between N<sub>2</sub>O production and emissions from the soil surface may can be significant and increases with distance (Jury et al., 1982). In accordance with this, Clough et al. (1999) observed a delay of 11 days before <sup>15</sup>N enriched N<sub>2</sub>O produced at 80 cm depth was released from the soil surface at a corresponding rate. ~~The~~ Since the presence of air-filled porosity is critical for the exchange of gases between soil and the atmosphere (Jury et al., 1982), and therefore the soil N<sub>2</sub>O concentration closest to, but above the WT depth (N<sub>2</sub>O<sub>WT</sub>), was taken to represent “subsoil” processes stimulating N<sub>2</sub>O emissions.

The regulation of N<sub>2</sub>O emissions was investigated using a statistical method represented by graphical models. Among the factors considered, the graphical models for individual site-crop combinations consistently identified N<sub>2</sub>O concentration in the capillary fringe as the strongest predictor of N<sub>2</sub>O emissions from both grassland and arable sites in spring (Figure 7a-d), and from grassland sites in the autumn (Figure 8a,b). The implication is that N transformations at

depth in the soil, and not in the top soil, were the main source of N<sub>2</sub>O escaping to the atmosphere in these cases. In accordance with this, there was no apparent effect of N fertilisation on emissions of N<sub>2</sub>O, with emissions at *RG2* after the application of acidified cattle slurry as a notable exception. Other studies also found a limited response to fertilisation (Maljanen et al., 2003; Regina et al., 2004), although Regina et al. (2004) later observed a peak in N<sub>2</sub>O emissions after rainfall. Goldberg et al. (2010) reported that N<sub>2</sub>O emissions from a minerotrophic fen were produced at 30-50 cm depth, in accordance with the observations presented here, where the highest concentrations of N<sub>2</sub>O were mostly observed at 20 or 50 cm depth (Table S5).

Peat decomposing in the capillary fringe during WT drawdown could have been the source of N for N<sub>2</sub>O production. It is well established that N<sub>2</sub>O emissions from organic soil may be enhanced by drainage (Martikainen et al., 1993; Taft et al., 2017), and the response will appear within days, as shown by Aerts and Ludwig (1997) in an incubation study with an oscillating WT. A stimulation of N<sub>2</sub>O emissions by WT drawdown was also observed by Goldberg et al. (2010) when simulating drought under field conditions, although a pulse of N<sub>2</sub>O also occurred after rewetting in that study. In accordance with this effect of rewetting, ~~increasing soil wetness after~~ rain events ~~induring~~ late spring and early autumn often enhanced N<sub>2</sub>O emissions. ~~This was not necessarily a result of WT changes.~~ Despite 32 mm rainfall on DOY 244 and 245, the WT depth at site *ARI* was still at 40 to 80 cm on DOY 246 (Figure 6), but ~~well~~ N<sub>2</sub>O emissions were high. Well-degraded peat will release as little as 10% of its water to drainage (Rezanezhad et al., 2016). ~~It~~, and it is therefore likely that rain water was absorbed by peat above the WT and created conditions suitable for denitrification close to the soil surface, which ~~may explain~~ explains the significant correlation between N<sub>2</sub>O emissions and NO<sub>3</sub><sup>-</sup> (Figure 8c).

The increase in N<sub>2</sub>O emissions during WT cycles reported by Aerts and Ludwig (1997) was observed only with eutrophic peat, whereas a mesotrophic peat showed no effect of WT dynamics on N<sub>2</sub>O emissions, which were consistently low. A similar interaction between nutrient status and WT depth was observed in field studies comparing N<sub>2</sub>O emissions from minerotrophic and ombrotrophic boreal peatlands (Martikainen et al., 1993; Regina et al., 1996). In the present study, soil NH<sub>4</sub><sup>+</sup>-N and NO<sub>3</sub><sup>-</sup>-N concentrations at site *RG1* increased to 133 and 120 µg g<sup>-1</sup> dry weight soil upon fertilisation, respectively, but largely returned to the background level of around 5 and 10 µg g<sup>-1</sup> dry weight soil, respectively, within a week (Table S1). In contrast, at site *ARI* there was substantial accumulation of NH<sub>4</sub><sup>+</sup>-N and NO<sub>3</sub><sup>-</sup>-N even before fertilisation on DOY 141, and soil mineral N remained high for several weeks (Table S2). ~~This accumulation of soil mineral N around the time of potato crop establishment could~~ that may have stimulated N<sub>2</sub>O emissions in the arable soil. Grasslands on organic soil generally show lower emissions of N<sub>2</sub>O compared to arable organic soil (Eickenscheidt et al., 2015), presumably because plants compete successfully with microorganisms for available N. Schothorst (1977) estimated peat decomposition indirectly from the N-content in herbage yield of grassland and concluded that the soil supplied 96 kg N ha<sup>-1</sup> when the drainage depth was 25 cm, but 160 and 224 kg N ha<sup>-1</sup> with the drainage depth at 70 and 80 cm, respectively. Hence, plant uptake of N mineralised from soil organic matter above the WT likely contributed to the much lower N<sub>2</sub>O emissions from rotational grass in this study.

At *RG* sites, soil N<sub>2</sub>O concentrations during spring were generally low and ~~thus~~ did not ~~clearly~~ show evidence for microbial N transformations, which supports the conclusion above that plant uptake was a main sink for the N released during peat decomposition. One exception was the accumulation of N<sub>2</sub>O at ~~0.5-1 m~~ 50-100 cm depth observed at site

RG2 in late May (Figure 3), which could have been caused by leaching of mineral N from the acidified cattle slurry following extensive rain. In contrast, at AR sites N<sub>2</sub>O consistently accumulated in the soil throughout spring irrespective of fertilisation; at site AR1 the highest concentrations occurred at 50 to and 100 cm depth, while at site AR2 with a higher groundwater table the highest concentrations were at 20 cm depth. It indicated that emitted N<sub>2</sub>O was produced mainly in the capillary fringe, consistent with peat decomposition as was a source of mineral N, although N<sub>2</sub>O production and a likely source of N<sub>2</sub>O above, and possibly also in below the saturated zone cannot be ruled out (see next section). Following N fertilisation, the accumulation of N<sub>2</sub>O in the soil profile was mostly associated with precipitation.

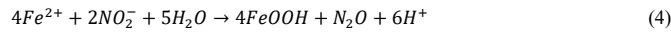
In the autumn, the graphical model models identified NO<sub>3</sub><sup>-</sup> in the top soil (site AR1 only) and soil temperature at 30 cm depth as predictors of N<sub>2</sub>O emissions at arable sites (Figure 7). The accumulation of NO<sub>3</sub><sup>-</sup> was much greater at site AR1 compared to AR2, possibly because AR1 had better drainage of the top soil (e.g., WT at 80 vs. 40 cm depth on DOY 246; Figure 6). It is not clear if the source of N was decomposing potato top residues or accelerated peat decomposition, or both. Rainfall most likely triggered denitrification by increasing soil water-filled pore space and raising WT depth, thereby impeding the O<sub>2</sub> supply to much of the soil profile (Barton et al., 2008). This interpretation is supported by increasing N<sub>2</sub>O concentrations below, as well as above the WT depth depending on site and block, and in fertilised as well as unfertilised subplots (Figure 6). In an annual study, conducted in other parts of the Store Vildmose bog, Kandel et al. (2018) also measured high emissions of N<sub>2</sub>O from a potato crop, i.e., around 2000 µg N<sub>2</sub>O m<sup>-2</sup> h<sup>-1</sup> in October 2014 and 6000 µg N<sub>2</sub>O m<sup>-2</sup> h<sup>-1</sup> in June 2015, which coincided with NO<sub>3</sub><sup>-</sup> accumulation and rainfall. Precipitation was also high during September 2015, and this was accompanied by accumulation of N<sub>2</sub>O in the top soil at all sites. However, N<sub>2</sub>O concentrations reached only around 10 µL L<sup>-1</sup> at RG sites, as opposed to several hundred µL L<sup>-1</sup> at AR sites, confirming that soil mineral N availability was a limiting factor for N<sub>2</sub>O emissions.

#### 4.2 Pathways of N<sub>2</sub>O emissions

Bacterial nitrification, denitrification, and nitrifier-denitrification are all potentially significant pathways of N<sub>2</sub>O formation (Braker and Conrad, 2011). The correlation with NO<sub>3</sub><sup>-</sup> in the top soil at site AR1 in the autumn (Figure 8c) suggested that here denitrification activity controlled N<sub>2</sub>O emissions, but mostly soil mineral N concentrations were low, and ammonia oxidation activity may have limited N<sub>2</sub>O emissions either directly, or indirectly via production of NO<sub>2</sub><sup>-</sup> or NO<sub>3</sub><sup>-</sup>. Ammonia oxidising bacteria (AOB) are scarce in acid peat despite the presence of nitrite oxidising bacteria (NOB) (Regina et al., 1996), and some studies indicate that ammonia oxidising archaea (AOA) predominate in both abundance and activity (Herrmann et al., 2012; Stopnišek et al., 2010). Stieglmeier et al. (2014) isolated an AOA from soil that emitted N<sub>2</sub>O at a rate corresponding to 0.09% of the NO<sub>2</sub><sup>-</sup> produced independent of O<sub>2</sub> availability, but it is not known if this organism is present in acid organic soil, and at this time an indirect control of denitrification activity seems more plausible.

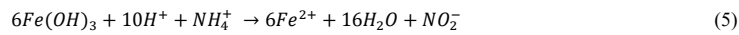
Stopnišek et al. (2010) found that AOA activity was not stimulated by an external source of NH<sub>4</sub><sup>+</sup> and concluded that the activity was associated with N released from decomposing soil organic matter. Well-decomposed peat is dominated by dead-end pores associated with plant cell remains, which are characterised by a slow exchange of solutes with active pore volumes (Hoag and Price, 1997), and hence ammonia oxidation in confined spaces could be important in organic soil. The anaerobic conditions of saturated peat may have limited N mineralisation and hence ammonia

580 oxidation activity during early spring, a constraint which was ~~removed~~alleviated as the WT declined and O<sub>2</sub> entered deeper soil layers. Estop-Aragonés et al. (2012) found that oxic-anoxic interfaces in peat soil were located above the WT depth, and hence the capillary fringe may have been still partly anoxic, which can explain the correlation between N<sub>2</sub>O<sub>WT</sub> and N<sub>2</sub>O emissions in graphical models. In late April (DOY 113), NO<sub>2</sub><sup>-</sup> had accumulated at 20-50 cm depth at both *RG1* and *AR1* sites (Figure 2), ~~which suggests~~suggesting that there was an imbalance between ammonia oxidation and nitrite oxidation activity. Oxygen affinity differs between nitrifiers, with AOA>AOB>NOB (Yin et al., 2018), and hence O<sub>2</sub> limitation could have caused the accumulation of NO<sub>2</sub><sup>-</sup>. In acid soil, this would result in product inhibition by HNO<sub>2</sub> if there were no mechanism to remove NO<sub>2</sub><sup>-</sup>, this would be especially true for *AR* sites, where mineral N accumulation was three to four times higher compared to *RG* sites (Tables S3-S6). Nitrifier-denitrification is a mechanism by which ammonia oxidisers can avoid HNO<sub>2</sub> accumulation, and this process leads to N<sub>2</sub>O formation (Braker and Conrad, 2011). Another potential sink for NO<sub>2</sub><sup>-</sup> is chemodenitrification, an abiotic reaction in which NO<sub>2</sub><sup>-</sup> reacts with Fe<sup>2+</sup> to produce N<sub>2</sub>O (Jones et al., 2015):

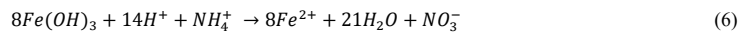


where in Eq. 4 Fe(OH)<sub>3</sub> is shown as anhydrous FeOOH. A possible depletion of TRFe was indicated at 50 cm depth at site *AR1*, which coincided with a similar pattern for NO<sub>2</sub><sup>-</sup> (Figure 2). Nitrifier-denitrification and chemodenitrification are both sinks for NO<sub>2</sub><sup>-</sup>, and therefore both pathways are potential sources of the N<sub>2</sub>O emissions observed during early spring.

At *AR* sites there was often considerable accumulation of N<sub>2</sub>O below the WT, which suggests there was also an anaerobic pathway of N<sub>2</sub>O formation. The fact that TRFe concentrations were much higher than those of AVS or CRS (Table 1) makes it relevant to consider alternative reactions involving iron oxides/hydroxides, which have a potential to produce N<sub>2</sub>O. One such recently described pathway is Feammox, a process whereby ammonia oxidation coupled with ferric iron reduction can produce NO<sub>2</sub><sup>-</sup> below pH 6.5 (Yang et al., 2012):



Nitrate can also be produced under these conditions (Yang et al., 2012; Guan et al., 2018):



605 A shuttle of Fe<sup>2+</sup> between Feammox and chemodenitrification (Eq. 5 and Eq. 4) could account for the accumulation of N<sub>2</sub>O under anoxic conditions in the saturated zone, presumably with the availability of NH<sub>4</sub><sup>+</sup> from peat mineralisation as a limiting factor. The confirmation of pathways will require more detailed investigations that should include molecular analyses targeting microbial communities in the soil profile.

## 610 5 Conclusion

In this one-year study, N<sub>2</sub>O emissions were consistently higher from arable sites compared to rotational grass. There were strong seasonal dynamics in N<sub>2</sub>O emissions, and we present evidence that different pathways were involved.

Concentrations of pyrite were low compared to the total reduction capacity of the peat, and Fe was predominantly in forms other than pyrite. The hypothesis that  $\text{NO}_3^-$  reduction coupled with  $\text{FeS}_2$  oxidation was an important source of  $\text{N}_2\text{O}$  could therefore not be confirmed. Nitrous oxide emissions during spring were independent of fertilisation, since there was mostly no effect of mineral N in the top soil. The significant effect of  $\text{N}_2\text{O}$  concentration in the capillary fringe indicated that emissions during spring, and for grassland also during the autumn, were associated with soil N mineralisation in this environment, as modified by rainfall patterns and WT dynamics. We propose that chemodenitrification (or nitrifier-denitrification) of  $\text{NO}_2^-$  produced in the capillary fringe is a main source of  $\text{N}_2\text{O}$  in acid organic soil during spring, whereas in the autumn heterotrophic denitrification can be a main pathway in arable soil as a result of  $\text{NO}_3^-$  accumulation. Mitigating  $\text{N}_2\text{O}$  emissions from acid organic soil is challenged by this complexity of underlying processes. However, reducing mineral N accumulation by ensuring a vegetation cover outside the main cropping season, and stabilising the WT depth by effective drainage, are potential mitigation strategies.

*Author contributions.* ATT, LEL, TJ and SOP designed the study. ATT, LEL, VE and SOP carried out sampling and analyses. ATT, RL and SOP were responsible for data analyses. ATT and SOP prepared the manuscript with contributions from all co-authors.

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## References

- Abreu, G.C.G., Edwards, D., and Labouriau, R.: High-dimensional graphical model search with the gRapHD R package. *J. Stat. Softw.*, 37, 1-18, doi: 10.18637/jss.v037.i01., 2010
- Aerts, R., and Ludwig, F.: Water-table changes and nutritional status affect trace gas emissions from laboratory columns of peatland soils. *Soil Biol. Biochem.*, 29, 1691-1698, doi: 10.1016/S0038-0717(97)00074-6, 1997
- Barton, L., Kiese, A., Gatter, D., Butterbach-Bahl, K., Buck, R., Hinz, C., and Murphy, D. V.: Nitrous oxide emissions from a cropped soil in a semi-arid climate. *Glob. Change Biol.*, 14, 177-192, doi: 10.1111/j.1365-2486.2007.01474.x, 2008.
- Braker, G., and Conrad, R.: Diversity, structure, and size of  $\text{N}_2\text{O}$ -producing microbial communities in soils—what matters for their functioning? *Adv. Appl. Microbiol.*, 75, 33-70, doi: 10.1016/B978-0-12-387046-9.00002-5, 2011.

- Burton, E.D., Sullivan, L.A., Bush, R.T., Johnston, S.G., and Keene, A.F.: A simple and inexpensive chromium-reducible sulfur method for acid-sulfate soils. *Appl. Geochem.*, 23, 2759-2766, doi: [doi.org/10.1016/j.apgeochem.2008.07.007](https://doi.org/10.1016/j.apgeochem.2008.07.007), 2008.
- 650 Carter, M.R., and Gregorich, E.G., (Eds.): Soil sampling and methods of analysis, Second edition, CRC Press, USA 2007.
- Cline, J.D.: Spectrophotometric determination of hydrogen sulfide in natural waters. *Limnol. Oceanogr.*, 14, 454-458, doi: [doi: 10.4319/lo.1969.14.3.0454](https://doi.org/10.4319/lo.1969.14.3.0454), 1969.
- Clough T.J., Jarvis S.C., Dixon E.R., Stevens R.J., Laughlin R.J., and Hatch D.J.: Carbon induced subsoil  
655 denitrification of <sup>15</sup>N-labelled nitrate in 1 m deep soil columns. *Soil Biol. Biochem.* 31, 31-41, doi: [doi.org/10.1016/S0038-0717\(98\)00097-2](https://doi.org/10.1016/S0038-0717(98)00097-2), 1999.
- Conen, F., and Smith, K.A.: A re-examination of closed flux chamber methods for the measurement of trace gas emissions from soils to the atmosphere. *Eur. J. Soil Sci.*, 49, 701-707, doi: [10.1046/j.1365-2389.1998.4940701.1998](https://doi.org/10.1046/j.1365-2389.1998.4940701.1998), 1998.
- 660 Davidson, A.C., Hinkley, D.V.: Bootstrap Methods and their Application, 1st ed. Cambridge University Press, New York, NY, 1997.
- Duan, Y.F. Kong, X.-W., Schramm, A., Labouriau, R., Eriksen, J., and Petersen, S.O.: Microbial N transformations and N<sub>2</sub>O emission after simulated grassland cultivation: effects of the nitrification inhibitor 3,4-Dimethylpyrazole Phosphate (DMPP). *Appl. Environ. Microbiol.*, 83, e02019-16, doi: [10.1128/AEM.02019-16](https://doi.org/10.1128/AEM.02019-16), 2017.
- 665 Eickenscheidt, T., Heinichen, J., and Drösler, M.: The greenhouse gas balance of a drained fen peatland is mainly controlled by land-use rather than soil organic carbon content. *Biogeosciences*, 12, 5161-5184, doi: <https://doi.org/10.5194/bg-12-5161-2015>, 2015.
- Estop-Aragonés, C., Knorr, K.-H., and Blodau, C.: Controls on *in situ* oxygen and dissolved inorganic carbon dynamics in peats of a temperate fen. *J. Geophys. Res.*, 117, G02002, doi: [10.1029/2011JG001888](https://doi.org/10.1029/2011JG001888), 2012.
- 670 Gillham, R.W.: The capillary fringe and its effect on water-table response. *J. Hydrol.*, 67, 307-324, [doi.org/10.1016/0022-1694\(84\)90248-8](https://doi.org/10.1016/0022-1694(84)90248-8), 1984.
- Guan, Q.S., Cao, W.Z., Wang, G.J., Wu, F.F., Wang, C., Jiang, C., Tao, Y.R., and Gao, Y.: Nitrogen loss through anaerobic ammonium oxidation coupled with iron reduction in a mangrove wetland. *Eur. J. Soil Sci.*, 69, 732-741, doi: [10.1111/ejss.12552](https://doi.org/10.1111/ejss.12552), 2018.
- 675 Goldberg, S.D., Knorr, K.-H., and Gebauer, G.: N<sub>2</sub>O concentration and isotope signature along profiles provide deeper insight into the fate of N<sub>2</sub>O in soils. *Isot. Environ. Health Stud.*, 44, 377-391, doi: [dx.doi.org/10.1080/10256010802507433](https://doi.org/10.1080/10256010802507433), 2008.
- Goldberg, S.D., Knorr, K.-H., Blodau, C., Lischeid, G., and Gebauer, G.: Impact of altering the water table height of an acidic fen on N<sub>2</sub>O and NO fluxes and soil concentrations. *Glob. Change Biol.*, 16, 220-233, doi: [10.1111/j.1365-2486.2009.02015.x](https://doi.org/10.1111/j.1365-2486.2009.02015.x), 2010.
- 680 Good, P.I.: Permutation, Parametric and Bootstrap Tests of Hypotheses. Third Edition. Springer, New York, 2005.
- Haughton, D.M.A.: On the choice of a model to fit data from an exponential family. *Ann. Statist.*, 16, 342-335, doi: [10.1214/aos/1176350709](https://doi.org/10.1214/aos/1176350709), 1988.

Field Code Changed

- Herold, M.B., Baggs, E.M., and Daniell, T.J.: Fungal and bacterial denitrification are differently affected by long-term  
685 pH amendment and cultivation of arable soil. *Soil Biol. Biochem.*, 54, 25-35, doi:  
doi.org/10.1016/j.soilbio.2012.04.031, 2012.
- Herrmann, M., Hädrich, A., and Küsel, K.: Predominance of thaumarchaeal ammonia oxidizer abundance and  
transcriptional activity in an acidic fen. *Env. Microbiol.*, 14, 3013-3025, doi: 10.1111/j.1462-  
2920.2012.02882.x, 2012.
- 690 Hiraishi, T., Krug, T., Tanabe, K., Srivastava, N., Jamsranjav, B., Fukuda, M. & Troxler, T.: Supplement to the 2006  
guidelines for national greenhouse gas inventories: wetlands. Intergovernmental Panel on Climate Change  
(IPCC), Geneva, Switzerland, 354 pp, 2014.
- Hoag, R.S., and Price J.S.: The effects of matrix diffusion on solute transport and retardation in undisturbed peat in  
laboratory columns. *J Contam Hydrol* 28:193-205, 1997.
- 695 Hutchinson, G.L., and Mosier, A.R.: Improved soil cover method for field measurement of nitrous oxide fluxes. *Soil  
Sci. Soc. Am. J.*, 45, 311-316, doi: 10.2136/sssaj1981.03615995004500020017x, 1981.
- IPCC: 2013 Supplement to the 2006 IPCC Guidelines for National Greenhouse Gas Inventories: Wetlands, Hiraishi, T.,  
Krug, T., Tanabe, K., Srivastava, N., Baasansuren, J., Fukuda, M. and Troxler, T.G. (eds). Published: IPCC,  
Switzerland, 2014.
- 700 Jones, L.C., Peters, B., Pacheco, J.S.L., Casciotti, K.L., and Fendorf, S.: Stable isotopes and iron oxide mineral products  
as markers of chemodenitrification. *Environ. Sci. Technol.*, 49, 3444-3452, doi: 10.1021/es504862x, 2015.
- Jury W.A., Letey J., and Collins T.: Analysis of chamber methods used for measuring nitrous oxide production in the  
field. *Soil Sci. Soc. Am. J.* 46, 250–256, doi:10.2136/sssaj1982.03615995004600020007x, 1982.
- Jørgensen, B., and Labouriau, R.: Exponential families and theoretical inference. Springer, Monografias de Matemática,  
705 Rio de Janeiro, Brazil, 2012.
- Jørgensen, C.J., Jacobsen, O.S., Elberling, B., and Aamand, J.: Microbial oxidation of pyrite coupled to nitrate  
reduction in anoxic groundwater sediment. *Environ. Sci. Technol.* 43, 4851–4857, doi: 10.1021/es803417s,  
2009.
- Kandel, T.P., Lærke, P.E., and Elsgaard, L.: Annual emissions of CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O from a temperate peat bog:  
710 Comparison of an undrained and four drained sites under permanent grass and arable crop rotations with  
cereals and potato. *Agric. Forest Meteorol.* 256-257, 470-481, doi: doi.org/10.1016/j.agrformet.2018.03.021,  
2018.
- Keeney, D.R., and Nelson, D.W.: Nitrogen–inorganic forms. In: A.L. Page, T.H. Miller and D.R. Keeney (Eds.),  
Methods of Soil Analysis. Part 2. Agronomy Monographs, 9. American Society of Agronomy and Soil Science  
715 Society of America, Madison, WI, pp. 643-692, 1982.
- Kits, K. D., Jung, M.Y., Vierheilg, J., Pjevac, P., Sedlacek, C.J., Liu, S., Herbold, C., Stein, L.Y., Richter, A., Wissel,  
H., Brüggemann, N., Wagner, M., and Daims, H.: Low yield and abiotic origin of N<sub>2</sub>O formed by the complete  
nitrifier *Nitrospira inopinata*. *Nature Comm.* 10, 1836, doi: //doi.org/10.1038/s41467-019-09790-x, 2019.
- Kristensen, M. K.: Vildmosearbejdet. Det Danske Forlag, Copenhagen, Denmark, 219 pp. (in Danish), 1945.
- 720 Labouriau, R., and Amorim, A.: Comment on ‘An Association Between the Kinship and Fertility of Human Couples’.  
Science, 322 (5908):1634, doi: 10.1126/science.1161907, 2008a.

- Labouriau, R., and Amorim, A.: Human fertility increases with marital radius. *Genetics*, 178, 601-603, doi: 10.1534/genetics.107.072454, 2008b.
- 725 Lamandé, M., Labouriau, R., Holmstrup, M., Torp, S. B., Heckrath, G., Iversen, B. V., and Jacobsen, O.H.: Density of macropores as related to soil and earthworm community parameters in cultivated grasslands. *Geoderma*, 162, 319-326, doi: doi.org/10.1016/j.geoderma.2011.03.004, 2011.
- Lauritzen, S. L.: Causal Inference from Graphical Models. In: *Complex Stochastic Systems*, O. E. Barndorff-Nielsen, D. R. Cox and C. Klüppelberg (Editors), New York, 1999.
- 730 Leppelt, T., Dechow, R., Gebbert, S., Freibauer, A., Lohila, A., Augustin, J., Drösler, M., Fiedler, S., Glatzel, S., Höper, H., Järveoja, J., Lærke, P. E., Maljanen, M., Mander, Ü., Mäkiranta, P., Minkinen, K., Ojanen, P., Regina, K., and Strömgren, M.: Nitrous oxide emission budgets and land-use-driven hotspots for organic soils in Europe. *Biogeosci.*, 11, 6595-6612, doi: doi.org/10.5194/bg-11-6595-2014, 2014.
- Liu, B., Frostegård, Å., and Bakken, L.R.: Impaired reduction of N<sub>2</sub>O to N<sub>2</sub> in acid soils is due to a posttranscriptional interference with the expression of nosZ. *MBio*, 5, e01383-14, doi: 10.1128/mBio.01383-14, 2014.
- 735 Madsen, H.B., and Jensen, N.H.: Potentially acid sulfate soils in relation to landforms and geology. *Catena*, 15, 137-145, doi: doi.org/10.1016/0341-8162(88)90025-2, 1988.
- Maeda, K., Spor, A., Edel-Hermann, V., Heraud, C., Breuil, M. -C., Bizouard, F., Toyoda, S., Yoshida, N., Steinberg, C., and Philippot, L.: N<sub>2</sub>O production, a widespread trait in fungi. *Nature Sci. Rep.*, 5, 9697, doi: 10.1038/srep09697, 2015.
- 740 Maljanen, M., Liikanen, A., Silvola, J., and Martikainen, P.J.: Nitrous oxide emissions from boreal organic soil under different land-use. *Soil Biol. Biochem.*, 35, 1-12, doi: doi.org/10.1016/S0038-0717(03)00085-3, 2003.
- Mander, Ü., Uuemaa, E., Kull, A., Kanal, A., Maddison, M., Soosaar, K., Salm, J.-O., Lesta, M., Hansen, R., Kuller, R., Harding, A., and Augustin, J.: Assessment of methane and nitrous oxide fluxes in rural landscapes. *Landscape Urban Plan.*, 98, 172-181, doi: doi.org/10.1016/j.landurbplan.2010.08.021, 2010.
- 745 Martikainen, P.J., Nykanen, H., Crill, P., and Silvola, J.: Effect of a lowered water-table on nitrous-oxide fluxes from northern peatlands. *Nature*, 366, 51-53, doi: 10.1038/366051a0, 1993.
- McCullagh, P., and Nelder, J.A.: *Generalized Linear Models*, Second Edition. Chapman and Hall/CRC, London, 1989.
- Mu, Z., Huang, A., Ni, J., and Xie, D.: Linking annual N<sub>2</sub>O emission in organic soils to mineral nitrogen input as estimated by heterotrophic respiration and soil C/N ratio. *PLOS ONE*, 9, e96572, doi: doi.org/10.1371/journal.pone.0096572, 2014.
- 750 Palomo, A., Dechesne, A., and Smets, B.F.: Genomic profiling of *Nitrospira* species reveals ecological success of comammox *Nitrospira*. *bioRxiv* doi: <http://dx.doi.org/10.1101/612226>, 2019.
- Pedersen, A.R., Petersen, S.O., and Schelde, K.: A comprehensive approach to soil-atmosphere trace-gas flux estimation with static chambers. *Eur. J. Soil Sci.*, 61, 888-902, doi: 10.1111/j.1365-2389.2010.01291.x, 2010.
- 755 Petersen, S.O.: Diffusion probe for gas sampling in undisturbed soil. *Eur. J. Soil Sci.*, 65, 663-671, doi: 10.1111/ejss.12170, 2014.
- Petersen, S.O., Hoffmann, C. C., Schäfer, C.-M., Blicher-Mathiesen, G., Elsgaard, L., Kristensen, K., Larsen, S.E., Torp, S.B., and Greve, M.H.: Annual emissions of CH<sub>4</sub> and N<sub>2</sub>O, and ecosystem respiration, from eight organic soils in Western Denmark managed by agriculture. *Biogeosci.*, 9, 403-422, doi: doi.org/10.5194/bg-9-403-2012, 2012.
- 760

Field Code Changed



- R Core Team: R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing. Vienna, Austria. <https://www.R-project.org/>, 2016.
- Regina, K., Budiman, A., Greve, M.H., Grönlund, A., Kasimir, Å., Lehtonen, H., Petersen, S.O., Smith, P., and Wösten, H., 2015. GHG mitigation of agricultural peatlands requires coherent policies. *Clim. Policy*, 16, 522–541, doi: [doi.org/10.1080/14693062.2015.1022854](https://doi.org/10.1080/14693062.2015.1022854), 2016.
- Regina, K., Nykanen, H., Silvola, J., and Martikainen, P.J.: Fluxes of nitrous oxide from boreal peatlands as affected by peatland type, water table level and nitrification capacity. *Biogeochem.*, 35, 401–418, doi: [doi.org/10.1007/BF02183033](https://doi.org/10.1007/BF02183033), 1996.
- Regina, K., Syväsalo, E., Hannukkala, A., and Esala, M.: Fluxes of N<sub>2</sub>O from farmed peat soils in Finland. *Eur. J. Soil Sci.*, 55, 591–599, doi: [doi.org/10.1111/j.1365-2389.2004.00622.x](https://doi.org/10.1111/j.1365-2389.2004.00622.x), 2004.
- Rezanezhad, F., Price, J.S., Quinton, W.L., Lennartz, B., Milojevic, T., and Van Cappellen, P.: Structure of peat soils and implications for water storage, flow and solute transport: A review update for geochemists. *Chem. Geol.*, 429, 75–84, [http://dx.doi.org/10.1016/j.chemgeo.2016.03.010](https://doi.org/10.1016/j.chemgeo.2016.03.010), 2016.
- Røy, H., Weber, H.S., Tarpgaard, I.H., Ferdelman, T.G., and Jørgensen, B.B.: Determination of dissimilatory sulfate reduction rates in marine sediment via radioactive <sup>35</sup>S tracer. *Limnol. Oceanogr. Methods*, 12, 196–211, doi: [doi.org/10.4319/lom.2014.12.196](https://doi.org/10.4319/lom.2014.12.196), 2014.
- Schothorst, C.J.: Subsidence of low moor peat soils in the western Netherlands. *Geoderma*, 17, 265–291, doi: [doi.org/10.1016/0016-7061\(77\)90089-1](https://doi.org/10.1016/0016-7061(77)90089-1), 1977.
- Schäfer, C.-M., Elsgaard, L., Hoffmann, C.C. and Petersen, S.O.: Seasonal methane dynamics in three temperate grasslands on peat. *Plant Soil*, 357, 339–353, doi: [doi.org/10.1007/s11104-012-1168-9](https://doi.org/10.1007/s11104-012-1168-9), 2012.
- Spott, O., Russow, R., and Stange, C.F.: Formation of hybrid N<sub>2</sub>O and hybrid N<sub>2</sub> due to codenitrification: First review of a barely considered process of microbially mediated N-nitrosation. *Soil Biol. Biochem.*, 43, 1995–2011, doi: [doi.org/10.1016/j.soilbio.2011.06.014](https://doi.org/10.1016/j.soilbio.2011.06.014), 2011.
- Stieglmeier, M., Mooshammer, M., Kitzler, B., Wanek, W., Zechmeister-Boltenstern, S., Richter, A., and Schleper, C.: Aerobic nitrous oxide production through N-nitrosating hybrid formation in ammonia-oxidizing archaea. *ISME J.*, 8, 1135–1146, doi: [10.1038/ismej.2013.220](https://doi.org/10.1038/ismej.2013.220), 2014.
- Stopnišek, N., Gubry-Rangin, C., Höfferle, S., Nicol, G. W., Mandić-Mulec, I., and Prosser, J.I.: Thaumarchaeal ammonia oxidation in an acidic forest peat soil is not influenced by ammonium amendment. *Appl. Environ. Microb.*, 76, 7626–7634, doi: [10.1128/AEM.00595-10](https://doi.org/10.1128/AEM.00595-10), 2010.
- Taft, H.E., Cross, P.A., Edwards-Jones, G., Moorhouse, E.R., and Jones, D.L.: Greenhouse gas emissions from intensively managed peat soils in an arable production system. *Agr. Ecosyst. Environ.*, 237, 162–172, doi: [doi.org/10.1016/j.agee.2016.11.015](https://doi.org/10.1016/j.agee.2016.11.015), 2017.
- Taghizadeh-Toosi, A., Clough, T., Petersen, S.O., Clough, T., and Elsgaard, L.: Nitrous oxide (N<sub>2</sub>O) turnover dynamics in agricultural peat soil -in response to availability role of nitrate, nitrite, and iron sulfides. *Geomicrobiol. J.* doi: [doi.org/10.1080/01490451.2019.1666192](https://doi.org/10.1080/01490451.2019.1666192), 2019.
- Thamdrup, B., Fossing, H. and Jørgensen, B.B.: Manganese, iron, and sulfur cycling in a coastal marine sediment, Aarhus Bay, Denmark. *Geochim. Cosmochim. Acta*, 58, 5115–5129, doi: [10.1016/0016-7037\(94\)90298-4](https://doi.org/10.1016/0016-7037(94)90298-4), 1994.

Field Code Changed

Field Code Changed

800 Torrento C, Cama J, Urmeneta J, Otero N, and Solar A.: Denitrification of groundwater with pyrite and *Thiobacillus*  
*denitrificans*. Chem. Geol., 278, 80-91, doi: dx.doi.org/10.1016/j.chemgeo.2010.09.003, 2010.  
 Toyoda, S., Yoshida, N., and Koba, K.: Isotopocule analysis of biologically produced nitrous oxide in various  
 environments. Mass Spectrometry Reviews 36, 135-160, doi: doi.org/10.1002/mas.21459, 2017.  
 Tubiello, F.N., Biancalani, R., Salvatore, M., Roissi, S., and Conchedda, G.: A worldwide assessment of greenhouse  
 gas emissions from drained organic soils. Sustainability, 8, 371, 13 pp, doi: 10.3390/su8040371, 2016.  
 805 Ulrich, G.A., Krumholz, L.R., and Suflita, J.M.: A rapid and simple method for estimationg sulfate reduction activity  
 and quantifying inorganic sulfides. Appl. Environ. Microbiol., 63, 1627-1630, 1997.  
 Van Cleemput, O. and Samater, A.H.: Nitrite in soils: Accumulation and role in the formation of gaseous N  
 compounds. Fert. Res., 45, 81-89, doi: doi.org/10.1007/BF00749884, 1996.  
 Viollier, E., Inglett, P.W., Hunter, K., Roychoudhury, A.N., and Van Cappellen, P.: The ferrozine method revisited:  
 810 Fe(II)/Fe(III) determination in natural waters. Appl. Geochem., 15, 785-790, doi: doi.org/10.1016/S0883-  
 2927(99)00097-9, 2000.  
 Whittaker, J.: Graphical models in applied multivariate statistics. John Wiley & Sons, Chichester, UK, 1990.  
 Wrage-Mönnig, N., Horn, M.A., Well, R., Müller, C., Velthof, G., and Oenema, O.: The role of nitrifier denitrification  
 in the production of nitrous oxide revisited. Soil Biol. Biochem. 123, A3-A16,  
 815 doi:doi.org/10.1016/j.soilbio.2018.03.020, 2018.  
 Yang, W.H., Weber, K.A., and Silver, W.L.: Nitrogen loss from soil through anaerobic ammonium oxidation coupled to  
 iron reduction. Nat. Geosci., 5, 538-541, doi: 10.1038/ngeo1530, 2012.  
 Yin, Z., Bi, X., and Xu, C.: Ammonia-oxidizing archaea (AOA) play with ammonia-oxidizing bacteria (AOB) in  
 nitrogen removal from wastewater. Archaea Article ID 8429145, 9 pages, doi: doi.org/10.1155/2018/8429145,  
 820 2018.

**Table 1.** Selected characteristics of soil profiles at the four monitoring sites with rotational grass (*RG1*, *RG2*) and potato crop (*AR1*, *AR2*). All analyses were done in triplicate; results shown represent mean and standard error of two soil profiles for which a complete data set was available. Soils for analyses were collected in late April (DOY 113) except for AVS and CRS (early September). Abbreviations: EC, electrical conductivity; TOC, soil organic carbon; TRFe, total reactive iron; AVS, acid volatile sulfide; CRS, chromium reducible sulfur.

	Depth (cm)	pH	EC	TOC (g 100 g <sup>-1</sup> )	Total N (g 100 g <sup>-1</sup> )	C:N ratio	TRFe (mg Fe g <sup>-1</sup> )	AVS (μg S g <sup>-1</sup> )	CRS (μg S g <sup>-1</sup> )
<b><i>RG1</i></b>									
Depth 1	2.5-7.5	5.1 (0.2)	0.26 (0.10)	37.4 (0.2)	1.75 (0.00)	21.3	3.63 (0.11)	2.51 (0.86)	155 (62)
Depth 2	7.5-12.5	5.3 (0.1)	0.15 (0.02)	38.2 (0.2)	1.79 (0.01)	21.3	4.03 (0.44)	NA	NA
Depth 3	17.5-22.5	5.3 (0.5)	0.37 (0.18)	39.7 (0.3)	1.80 (0.04)	22.1	4.14 (0.32)	NA	NA
Depth 4	36-40	4.8 (0.1)	0.55 (0.02)	43.1 (2.7)	1.85 (0.03)	23.3	3.04 (0.26)	2.60 (0.87)	133 (64)
Depth 5	47.5-52.5	5.1 (0.3)	0.42 (0.13)	31.0 (15.6)	1.47 (0.64)	21.1	2.50 (0.55)	4.86 (1.07)	24 (17)
Depth 6	93-98	5.4 (0.0)	0.51 (0.06)	0.6 (0.3)	0.01 (0.01)	ND	0.14 (0.04)	NA	NA
<b><i>RG2</i></b>									
Depth 1	0-25	5.0	NA	19.8 (3.4)	1.34 (0.13)	14.8	2.29 (0.56)	NA	NA
Depth 2	25-50	5.1	NA	8.9 (3.0)	0.63 (0.23)	14.2	4.48 (NA)	1.71 (0.00)	33 (7.3)
<b><i>AR1</i></b>									
Depth 1	2.5-7.5	5.0 (0.1)	0.45 (0.04)	35.9 (0.1)	1.81 (0.02)	19.9	4.57 (0.09)	1.74 (0.02) §	141 (9)
Depth 2	7.5-12.5	5.2 (0.1)	0.42 (0.06)	34.2 (0.2)	1.76 (0.02)	19.4	4.66 (0.15)	NA	NA
Depth 3	17.5-22.5	5.2 (0.1)	0.34 (0.04)	41.0 (2.2)	1.93 (0.11)	21.3	4.99 (0.43)	NA	NA
Depth 4	36-40	4.7 (0.5)	0.37 (0.05)	41.1 (5.8)	1.84 (0.05)	22.4	3.23 (0.41)	2.17 (0.29)	49 (3)
Depth 5	47.5-52.5	4.7 (0.3)	0.48 (0.08)	5.9 (1.7)	0.37 (0.13)	16.3	1.19 (0.19)	1.98 (0.41)	137 (39)
Depth 6	93-98	5.4 (0.2)	0.91 (0.03)	0.3 (0.1)	0.00 (0.00)	ND	0.18 (0.02)	NA	NA
<b><i>AR2</i></b>									
Depth 1	0-25	5.1	NA	33.4 (1.2)	1.45 (0.03)	23.1	4.11 (0.03)	NA	NA
Depth 2	25-50	5.1	NA	38.4 (0.2)	1.46 (0.02)	26.2	3.78 (0.14)	1.65 (0.02)	45 (8)

ND – Not determined due to TOC and total N concentrations being at the limit of detection.

NA - Not analysed.

§ Taghizadeh-Toosi et al. (2019) presented slightly different AVS and CRS results for *AR1* representing all the six profiles collected at this site.

**Table 2.** Daily emissions of N<sub>2</sub>O during the monitoring periods in spring and autumn [were analysed with a generalised linear mixed model \(see text\)](#). Estimation for each season was performed using the trapezoidal approximation of the integral of the emission curve. Numbers in parentheses indicate 95% confidence intervals, and significant differences [between all eight treatments within each season](#) (at 5% significance level), [with corrected](#)~~ed~~ for multiple testing by the single-step method<sup>‡</sup> are indicated by letters.

			Spring				Autumn			
			DOY	g N <sub>2</sub> O-N ha <sup>-1</sup> d <sup>-1</sup>			DOY	g N <sub>2</sub> O-N ha <sup>-1</sup> d <sup>-1</sup>		
Rotational grass	<i>RG1</i>	<i>NF</i> <sup>#</sup>	62-161	29	(22-36)	a	251-313	20	(16-25)	a
		<i>F</i>		31	(24-37)	a		15	(11-18)	a
	<i>RG2</i>	<i>NF</i>	63-168	23	(17-28)	a	259-306	50	(34-66)	b
		<i>F</i>		146	(94-197)	bc		51	(39-63)	b
Arable	<i>AR1</i>	<i>NF</i>	62-166	303	(229-377)	d	245-307	338	(239-437)	d
		<i>F</i>		234	(180-288)	cd		322	(224-421)	d
	<i>AR2</i>	<i>NF</i>	63-168	102	(84-120)	b	244-313	105	(70-141)	bc
		<i>F</i>		105	(81-128)	b		127	(90-165)	c

<sup>#</sup> *F*, fertilised; *NF*, not fertilised.

## Figure captions

**Figure 1.** A. Location of sites *ARI* and *RGI* (both at 57°13'59.7"N, 9°50'40.3E), *RG2* (57°13'55.9"N, 9°52'20.2E) and *AR2* (57°13'7.6"N, 9°46'26.9E). B. Experimental design at each of the four sites, with three blocks centered around piezometers (●) and two subplots, one of which received N fertiliser at the rate of the surrounding field. Six collars for gas flux measurements (S1-S6) were distributed as indicated, and sets of 5 diffusion probes for soil gas sampling were installed near collars in selected positions (see text).

**Figure 2.** Nitrite-N (a, c) and total reactive iron, TRFe (b, d), in undisturbed soil cores were collected at sites *RG1* and *ARI* on 23 April (DOY 113; white symbols) and 2 September (DOY 245; grey symbols). Results shown are mean and standard error ( $n = 2$ ). At site *RGI* fertilisation had taken place one week earlier, but a two-sample  $t$  test did not find evidence for any effect on  $\text{NO}_2^-$  availability ( $p = 0.19$ ). The dotted lines indicate WT level on the two sampling dates.

**Figure 3.** The top panel shows rainfall, air temperature and management (F – fertilisation) at sites *RGI* (left panels) and *RG2* (right panels) during spring, 3 March (DOY 63) to 16 June (DOY 169). The middle section shows  $\text{N}_2\text{O}$  fluxes (black circles; mean  $\pm$  standard error,  $n = 3$ ) and contour plots of soil  $\text{N}_2\text{O}$  concentrations in fertilised subplots, and the lower section the corresponding results for unfertilised subplots. A logarithmic grey scale was used in order to show trends within both *RG* and *AR* treatments, and between depths. Soil gas sampling positions are indicated in the contour plots; numbers shown are  $\text{N}_2\text{O}$  concentrations ( $\mu\text{L L}^{-1}$ ). Green lines show the WT depth (which varied slightly between blocks). B2 and B3 refer to block number of diffusion probe positions.

**Figure 4.** The top panel shows rainfall, air temperature and management (T – tillage; F – fertilisation) at sites *ARI* (left panels) and *AR2* (right panels) during spring, 3 March (DOY 63) to 16 June (DOY 169). The middle section shows  $\text{N}_2\text{O}$  fluxes (black circles; mean  $\pm$  standard error,  $n = 3$ ) and contour plots of soil  $\text{N}_2\text{O}$  concentrations in fertilised subplots, and the lower section the corresponding results for unfertilised subplots. A logarithmic grey scale was used in order to show trends within both *RG* and *AR* treatments, and between depths. Soil gas sampling positions are indicated in the contour plots; numbers shown are  $\text{N}_2\text{O}$  concentrations ( $\mu\text{L L}^{-1}$ ). Gaps are indicated where soil gas sampling probes were installed late, or removed due to field operations. Green lines show the WT depth (which varied slightly between blocks). B2 and B3 refer to block number of diffusion probe positions.

**Figure 5.** The top panel shows rainfall, air temperature and management (H - harvest) at sites *RGI* (left panels) and *RG2* (right panels) during autumn, 3 September (DOY 245) to 10 November (DOY 314). The middle section shows  $\text{N}_2\text{O}$  fluxes (black circles; mean  $\pm$  standard error,  $n = 3$ ) and contour plots of soil  $\text{N}_2\text{O}$  concentrations in fertilised subplots, and the lower section the corresponding results for unfertilised subplots. A logarithmic grey scale was used in order to show trends within both *RG* and *AR* treatments, and between depths. Soil gas sampling positions are indicated in the contour plots; numbers shown are  $\text{N}_2\text{O}$  concentrations ( $\mu\text{L L}^{-1}$ ); the probes were absent in the unfertilised subplot after harvest. Green lines show the WT depth (which varied slightly between blocks). B2 and B3 refer to block number of diffusion probe positions.

**Figure 6.** The top panel shows rainfall, air temperature and management (H - harvest) at sites *AR1* (left panels) and *AR2* (right panels) during autumn, 3 September (DOY 245) to 10 November (DOY 314). The middle section shows  $N_2O$  fluxes (black circles; mean  $\pm$  standard error,  $n = 3$ ) and contour plots of soil  $N_2O$  concentrations in fertilised subplots, and the lower section the corresponding results for unfertilised subplots. A logarithmic grey scale was used in order to show trends within both *RG* and *AR* treatments, and between depths. Soil gas sampling positions are indicated in the contour plots; numbers shown are  $N_2O$  concentrations ( $\mu L L^{-1}$ ). Green lines show the WT depth (which varied slightly between blocks). B2 and B3 refer to block number of diffusion probe positions.

**Figure 7.** Results of a graphical model for each site crop combination in spring. A. RG1-Spring; B. RG2-Spring; C. AR1-Spring; and D. AR2-Spring. The edges (“lines”) connecting vertices (“points”) indicate significant conditional correlation between the variables given the other variables. The statistical results for direct effects on  $N_2O$  flux are: [1] 0.14 (0.02-0.23,  $p = 0.067$ ); [2] 0.13 (0.06-0.21,  $p = 0.008$ ); [3] 0.18 (0.09-0.26,  $p = 0.001$ ); [4] 0.15 (0.04-0.23,  $p = 0.032$ ); and [5] 0.37 (0.12-0.49,  $p = 0.002$ ). Key to variables: AmmoniumT:  $NH_4^+$ -N at 0-25 cm depth; NitrateT:  $NO_3^-$ -N at 0-25 cm depth;  $N_2O$  WT: equivalent soil gas phase concentration closest to, but above the water table depth; Temp5: soil temperature at 5 cm depth; Temp30: soil temperature at 30 cm depth.

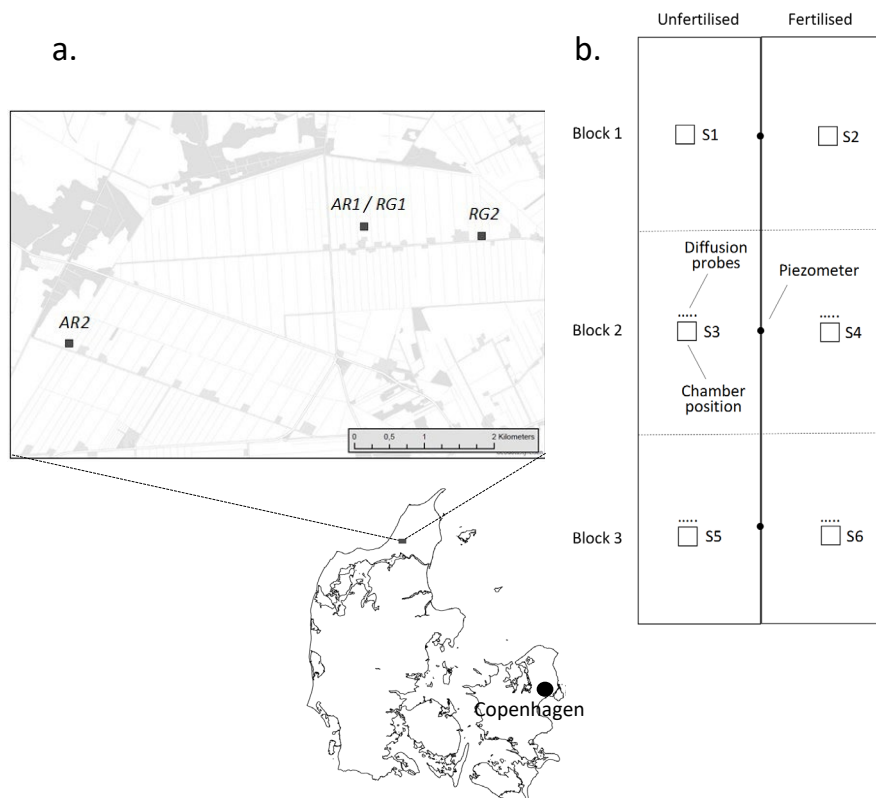
**Figure 8.** Results of a graphical model for each site crop combination in the autumn. A. RG1-Autumn; B. RG2-Autumn; C. AR1- Autumn; and D. AR2- Autumn. The edges (“lines”) connecting vertices (“points”) indicate significant conditional correlation between the variables given the other variables. Statistical results for effects on  $N_2O$  flux are: [1] 0.27 (0.19-0.38,  $p = 0.002$ ); [2] 0.16 (0.007-0.28,  $p = 0.049$ ); [3] 0.19 (0.09-0.28,  $p = 0.021$ ) [4] 0.15 (0.06-0.21,  $p = 0.032$ ); and [5] 0.29 (0.17-0.35,  $p = 0.005$ ). Key to variables: AmmoniumT:  $NH_4^+$ -N at 0-25 cm depth; NitrateT:  $NO_3^-$ -N at 0-25 cm depth;  $N_2O$  WT: equivalent soil gas phase concentration closest to, but above the water table depth; Temp5: soil temperature at 5 cm depth; Temp30: soil temperature at 30 cm depth.



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**Figure 1**



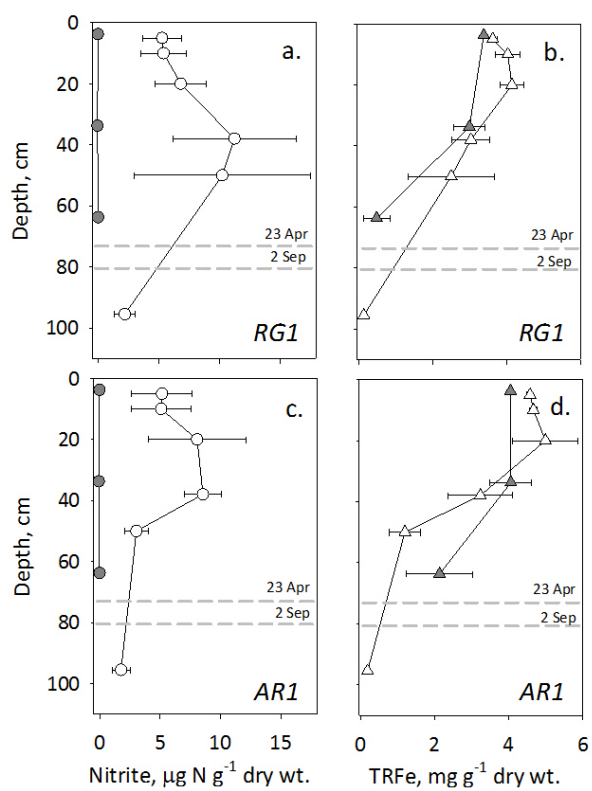
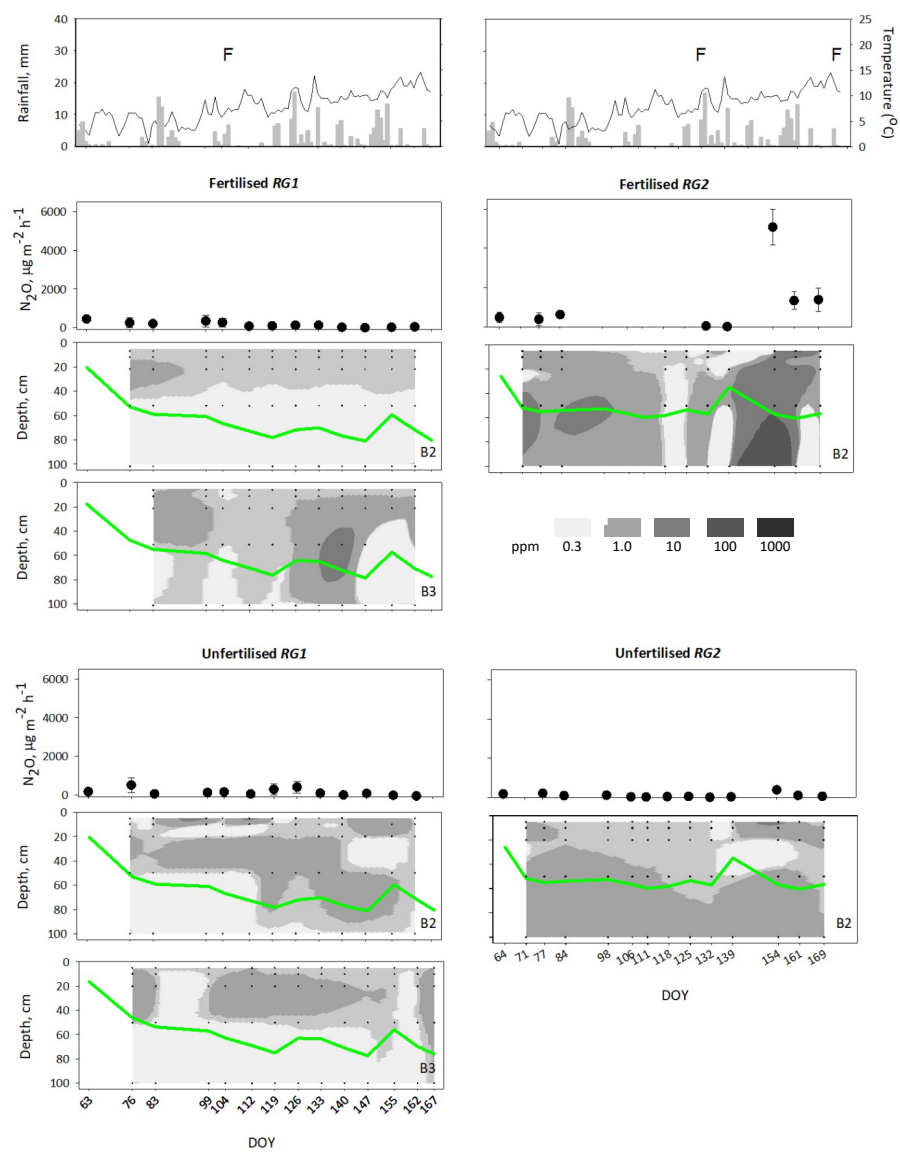


Figure 2.



**Figure 3**

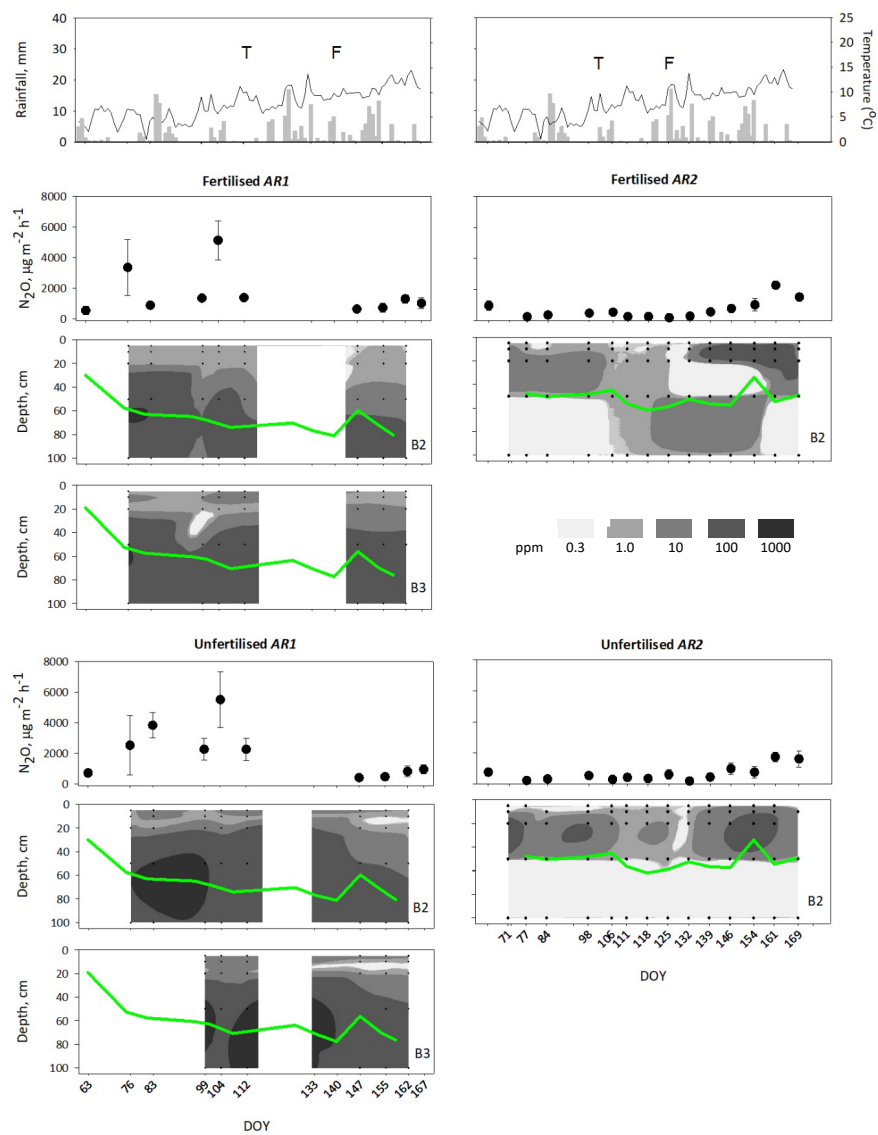


Figure 4

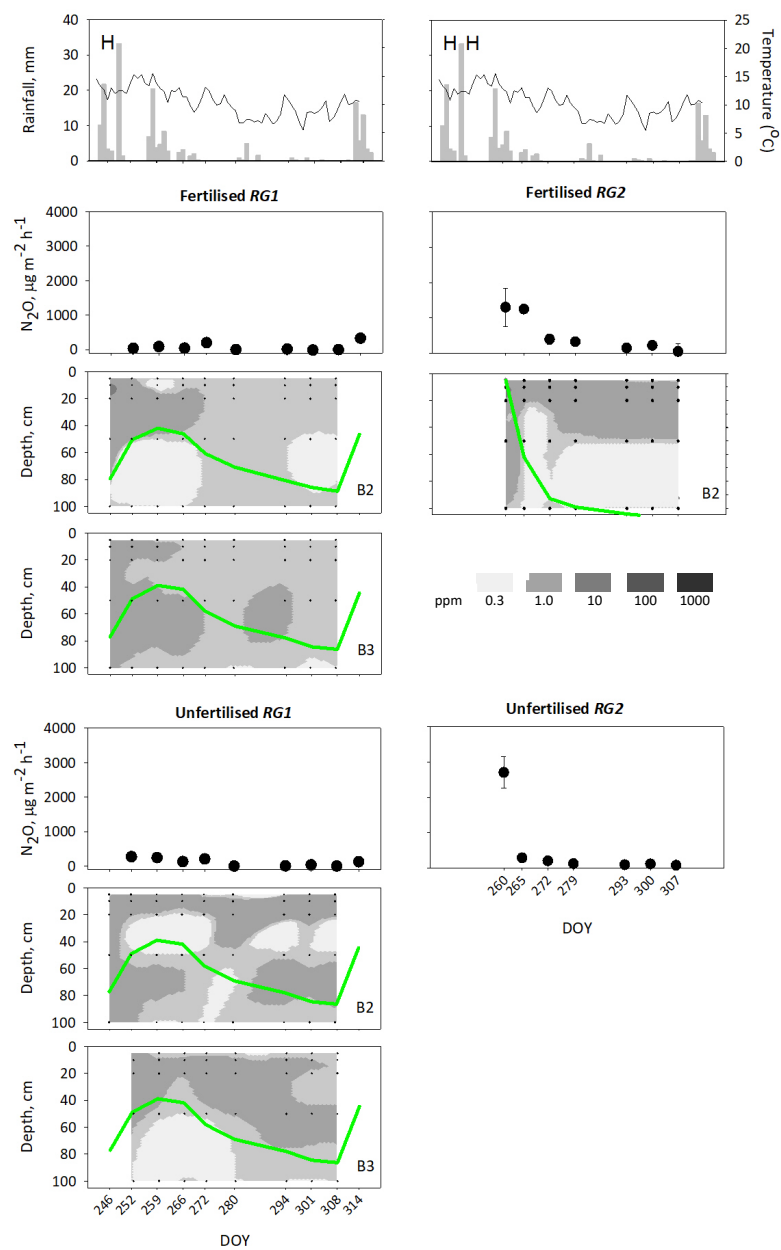


Figure 5

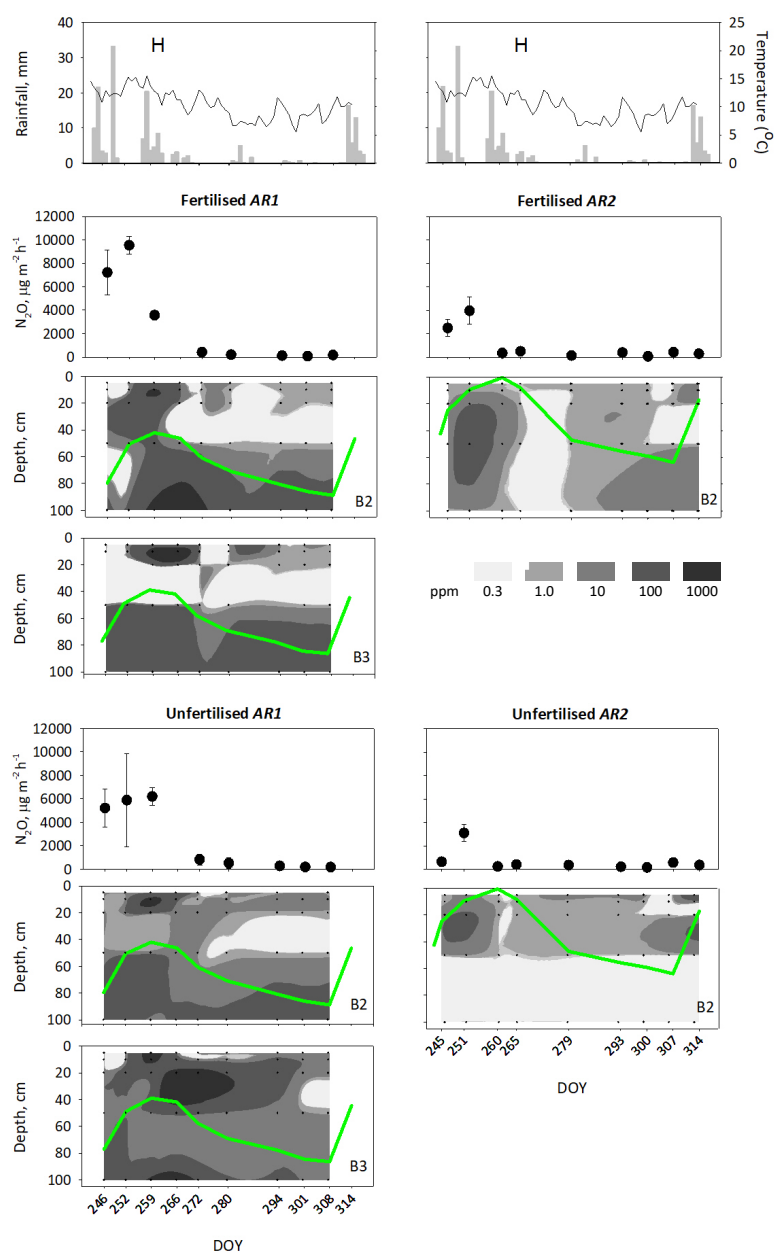


Figure 6

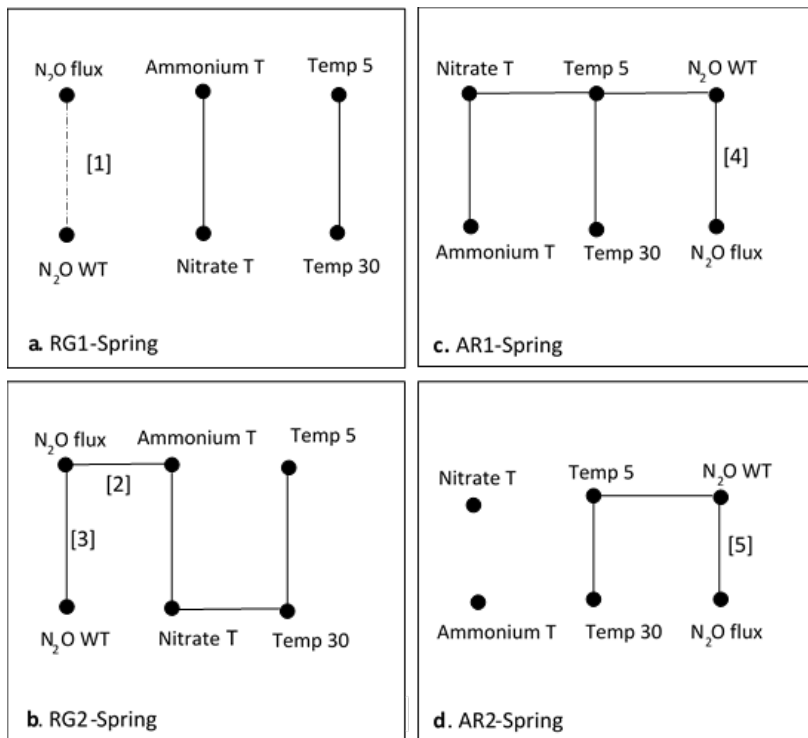
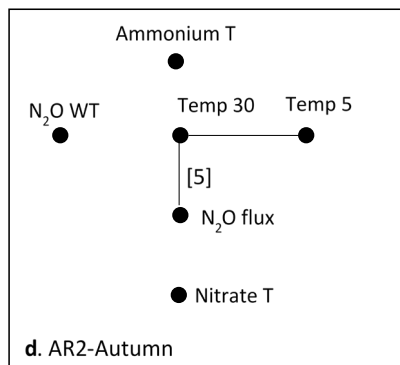
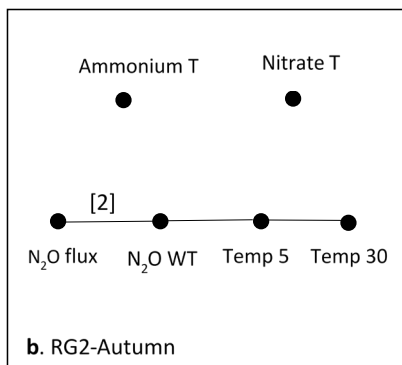
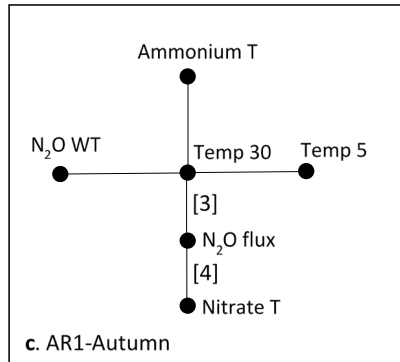
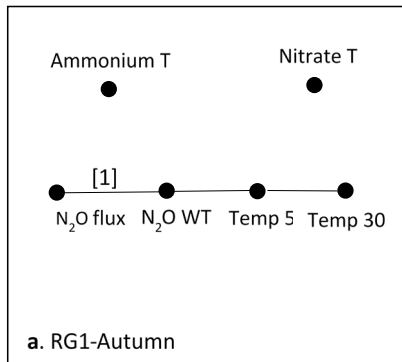


Figure 7



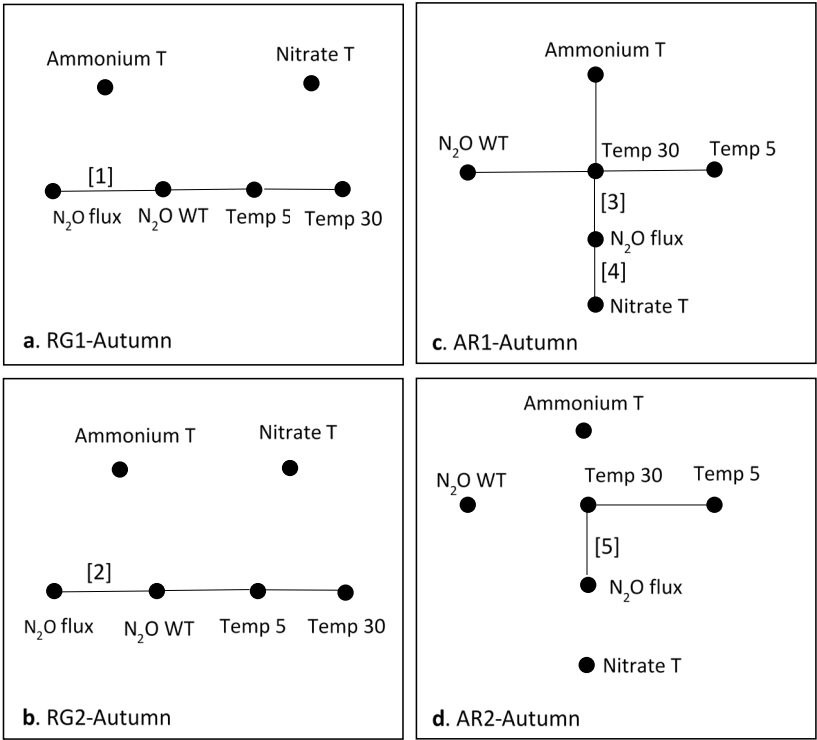


Figure 8



