Response to Editor

Associate Editor Decision: Publish subject to minor revisions (review by editor) (07 Oct 2019) by Julia Uitz
Comments to the Author:
Dear authors,

My apologies for the long delay in getting back to you regarding the status of your paper.

I believe overall you have satisfactorily addressed the comments and questions raised by both Reviewers. I will be pleased to accept your paper for publication in Biogeosciences, subject to minor changes being made in response to my comments provided below.

Please consider these minor additional comments and include appropriate changes in the revised version of your manuscript. Then upload the final version of your ms onto the BG interface.

Sincerely,
Julia

Response to editor:

Dear Dr. Julia Uitz, Thank you very much for your letter and the additional comments about our paper.

In response to the reviewers' and your comments we have made numerous revisions to our manuscript. We have provided the detailed responses to the comments of the reviewers and you, including a point-by-point reply to the comments.

Additionally, we have modified the first author's affiliations and added Yahui Gao as a co-corresponding author for his contribution on this paper.

We submit here the revised manuscript (with a red color font to mark-up the changes made in the manuscript).

We would like to express our great appreciation again to you for your comments on our paper. Looking forward to hearing from you.

Additional comments

Please consider the comments below, revise the text and correct all typos before you upload the revised version of your manuscript. I've listed several typos below but note that the list is not exhaustive.

The line numbering of the text in red color in the revised text is often wrong, which doesn't help to identify how/if you respond appropriately to the Reviewers' comments.

Unfortunately we neglected to check the line numbers in the response letter against the final revised manuscript. We certainly apologize for this oversight, as we understand it is important for you and the reviewers to be able to locate our revisions quickly and easily. We've also corrected and proofread the manuscript for additional typos, thank you for those that you pointed out.

Responses to RC#1

RC1-Line 210 : Please specify how the equation was modified.

Your response that the wording has been clarified so as to more accurately describe the model in l. 252-255 is confusing and likely refers to the wrong line numbers. The section "Model for population growth…" is in l. 230-239 and includes only minor changes compared to the submitted version. Please make sure you properly accommodate this specific comment by the Reviewer in your revised ms.

Response: We have now expanded this methods text with a much more in-depth description of how the Bernhardt et al. (2018) model works, and why it gives superior estimates under variable thermal regimes compared to older linear models (**lines 230-243**). In addition, we have provided several references for readers who would like to learn the derivation and application of the model for themselves.

RC1-Line 596- : In this section, it might be worth to also expand the discussion…in the community level.

In your response, you mention that changes were made into the text in l. 656-661 but I was not able find any change inhere. Please make sure this important suggestion is accounted for in the revised version of the ms.

Response: We apologize for submitting a revised version with wrong line numbers, we understand this makes it more difficult to review our changes and it shouldn't have happened. Now, we have expanded the existing (but mis-numbered) discussion about effects of thermal variability on potential competition with other phytoplankton taxa such

as the tropical diatoms and cyanobacteria found in the same region as *E. huxleyi*, and point out that this may affect both their relative fitness and community structure. Please check **Line 678-684** in the present revised version.

RC1-Fig 1 : Negative growth rates.
I understand your response. Yet I believe the biological meaning of these negative growth rates should be discussed as other readers may have the same impression as the Reviewers.

Response: We expanded the text about the potential biological meaning and usefulness of the negative growth rates as suggested, and explained our rationale for presenting them (rather than as zero growth rates) in the same fashion that we explained to the reviewer in our response letter. At the end of this paragraph, there is text discussing ecological implications of these negative growth rates for the marine coccolithophore *E. huxleyi* as the Sargasso Sea (where this strain was isolated) continues to warm. **(Lines 458-469)**.

Responses to RC#2
Line 217: The description of the applied statistical tests needs a better description…
In response to this comment, I can see you've simply added the name of the R functions you used to perform the statistical tests. This does not accommodate the Reviewer's comment. I believe she/he is expecting you to indicate the steps in your calculations. By the way, in l. 246 ("two formulas including compare_means() and stat_compare_mean()"),you actually refer to "functions" from the R package, not mathematical formula so please correct the text accordingly.

Response: Thank you for this good suggestion. We have revised the text accordingly, including listing the specific stepwise procedures used in our statistical analyses, as well as the R functions (not formulas, we corrected this) used to calculate them. (**Lines 248-260**)

Typos
l. 139 : please clarify « fluorescent lights were rearranged »
Response: Following your suggestions, we have revised in the manuscript to clarify this. (**Lines 133- 139**)
l. 168 : remove "the" in « very two days the for constant »
Response: We revised the manuscript as suggested. (**Line 167**)
l. 173 : consider adding « at » into the following sentence « always maintained >100 µmol L-1 and >10 µmol L-1 »
Response: We revised the manuscript as suggested. (**Line 172**)
l. 199 : correct « rations »
Response: We revised this mis-spelling. (**Line 196**)
l. 212 : remove « s » at the end of "techniques" in « using a 14C incubation techniques »

or correct sentence appropriately

Response: We followed this suggestion and have revised the manuscript. (**Line 209-210**)

l. 362 : Please define abbreviation or write in full “Cal/Photo ratios”

Response: We followed this suggestion and have revised the manuscript. (**Line 373**)

l. 459: Remove “s” at the end of “rises”

Response: We revised the manuscript as suggested. (**Line 474**)

l. 597: A word must be missing here “of a Southern Hemisphere cultured at”

Response: We added the missing words ‘*E. huxleyi* strain’. (**Line 607**)

l. 676: Please write in full “didn’t address »

We followed this suggestion and have revised the manuscript. (**Line 686**)

Acknowledgement section: Please consider acknowledging the Reviewers for their constructive comments and suggestions.

Response: We agree this is a good idea, and have added acknowledgement of the Reviewers for their constructive comments and suggestions. (**Lines 728-729**)

Response to Anonymous Referee #1

Interactive comment on "How will the key marine calcifier Emiliania huxleyi respond to a warmer and more thermally variable ocean?" by Xinwei Wang et al.

Anonymous Referee #1

Received and published: 1 July 2019

Review on: 'How will the key marine calcifier *Emiliania huxleyi* respond to a warmer and more thermally variable ocean?' by Wang et al. The experiments are well designed and I have only a couple of smaller questions (see specific comments). The manuscript is well written. Overall, I found the discussion not extremely inspiring because I thought it missed a conceptual framework that helps to arrange the numerous datasets. Nevertheless, some of the key conclusions are interesting and the data is valuable. I therefore only have 'minor comments' One major issue, however, is that the authors should deposit their data in a publicly accessible data repository and provide the link within the paper. This is important.

Response: The authors would like to thank the anonymous Reviewers for their constructive comments and suggestions to improve the quality of the paper. Those comments are all valuable and very helpful for revising and improving our paper. We have studied comments carefully and have made correction which we hope meet with approval. Revised portion are marked in red in the paper. The main corrections in the paper and the responds to the reviewer's comments are as flowing:

Response to Reviewer #1 (highlights):

Thank you very much for your helpful comments. Our data from this paper have been submitted to the Biological and Chemical Oceanography Data Management Office (BCO-DMO, bco-dmo.org), as is required by the conditions of our major funding agency (US NSF). The data are currently in the queue to be uploaded, but the data management office is running behind and we have been told that it will be several months more before the data can be quality checked, vetted and formatted, and posted to be made publicly available. When this is finished, the data will be available at our project webpage: www.bco-dmo.org/project/668547. We can provide this link with the paper if the editor agrees, but it will still take some time before the data from this paper are live.

Response to Reviewer #1 (Specific comments):

Line 132: How was light measured and kept identical between treatments? Measuring light in such blocks is challenging and there may be large differences between replicates and treatments. Please provide a detailed description.

Response: We agree that getting the lighting uniform for every replicate within a thermal block is essential but can be difficult, and we went to considerable effort to carefully measure and adjust light levels in each position in the block to be as close to identical as possible. We followed your suggestion, and now provide a detailed section in the Methods on how we measured and adjusted the light intensity in the thermal-blocks. (**Line 133-139**)

Line 135: Was the dilution medium also Aquil? Please clarify.

Response: Yes, the Aquil medium was used as the dilution medium, and have now we clarified this in the manuscript. (**Line 141**)

Line 139: It is unclear to me from this description how negative growth was measured. Wasn't it just the reduction in cell numbers or in your case red fluorescence? Please explain this better.

Response: Yes, the negative growth rate was calculated from the decrease of cell numbers at 28.6 ^{o}C during cultivation. In our preliminary experiments, we repeated this process several times to rigorously verify that cultures were unable to grow at this temperature. We have revised and expanded the description of how negative growth rates were measured in our manuscript. (**Line 146-150**)

Line 146: Please indicate how long it took for the temperature block to reach the new temperature after switching the water bath temperature. Is there a significant time lag? I wonder if this could partially explain the lower response in the one day cycle, as the time lag may have promoted a weaker response.

Response: This is an important point. It took the block about half an hour to re-adjust to the transformed temperature for each growth phase, which shouldn't represent a significant time lag relative to the 24-48 h thermal cycles. The reason for the lower response to the one-day cycle is likely the acclimation characteristics of the coccolithophorid. We have revised and clarified the description of the thermal cycles and their re-adjustment times during transitions in the manuscript. (**Line 158-163**)

Line 154: Weren't the nutrients already in the dilution medium? Or did you adjust to 100 and 10 _mol/L? This is confusing. Please clarify.

Response: Thanks for pointing this out, we agree this text was unclear and confusing. We did adjust the N and P midway through the 4 day cycle (2 day variation treatment) by adding concentrated Aquil stocks at these concentrations to make sure nutrients remained replete throughout the 4 day cycle. We have revised this text in the manuscript to better describe this. (**Line 168-172**)

Line 167: It remains unclear if you always measured both fluorescence and cell number or if this varied between treatments? Please clarify and ideally give the reader an idea how similar the growth rates were when determined with these two measurements.

Response: Following your suggestions, we have revised in the manuscript to clarify this. (**Line 184-186**) Under constant conditions such as in the thermal block and the constant controls of the variation treatment, the cell numbers and the *in vivo* fluorescence are strongly correlated and relatively invariant (as verified by microscopic counts). So, we used the *in vivo* fluorescence to calculate the growth rate. However, the cellular *in vivo* fluorescence (cellular Chl *a* content) changed during temperature fluctuation, so for these treatments we applied cell counts only to calculate the growth rate.

Line 180: Please provide percentage of the HCl acid. Was it 37%? In this case fuming overnight is fairly extreme and may perhaps breakdown POC?

Response: We revised in the manuscript to provide this information (Line **200**)

In our experiment, we used the ~37% saturated HCl for fuming overnight to thoroughly remove the inorganic carbon. We are not aware of any published evidence that ~12h of HCl fuming can degrade organic carbon, but we can consider this possibility if the reviewer knows of any. From our results shown in Fig. 2C, the PIC/POC ratio was extremely low (~0.05), meaning that the POC content was nearly as high as the TPC (PIC+POC) content. This result suggests that the cellular POC is very likely not degraded by our saturated HCl fuming method.

Line 185: Not 100% sure but I assume Fu et al., 2007 did not invent this protocol. Please provide original papers here and also for POC, PON above.

Response: We gave our own references for these methods because in our lab over the years we have made minor modifications to these classic protocols, and this allows readers to look up the exact procedures we used if desired. However, in response to this suggestion we have revised the manuscript by adding the original citations as well for all of these methods (Line **202, 204-206**)

Line 188: See previous comment.

Response: As noted above, we have now added citations to the original protocols preceding the citations of our slightly modified versions of the techniques. (Line **209-210**)

Line 217: The description of the applied statistical tests needs a better description. Perhaps briefly go through the consecutive steps. Just for completeness. Only mentioning which tests were done may raise some eye brows.

Response: We followed this suggestion and have revised in the manuscript to include a better and more in-depth description of the statistical methods. (**Line 248-260**)

Line 227: What is the rationale behind showing the TPC/PON ratio? What meaning does it have and why is it important? I would intuitively say that this dataset could be removed from the results but I am of course interested what the authors think.

Response: We understand that many coccolithophore studies don't present the

TPC/PON ratio, but we feel it is worth presenting as it encompasses all of the C fixed (into both POC and PIC) relative to all of the cellular N quota. We also of course present the more traditional POC:PON and PIC:POC ratios as well.

Line 266: This may indicate a time lag until the high temperature was established so that the warm period was shorter than indicated by assuming an instant change in temperature. Please provide a retention time for how long it lasted until the new temperature was reached within the bottles.

Response: The time for the thermal block to re-equilibrate the experimental bottles after temperatures were switched was only half an hour, which we suggest is too short to significantly affect the overall growth rates in either the one day or two day thermal variation treatments. We have revised in the manuscript with new text to point this out. (Line **309-311**)

Line 267: This comment basically addresses all quota measurements and ratios. When you look at e.g. PIC/POC and do this for a one day period in the cycled experiments. To what extent is the response you measure and report here 'diluted' by the PIC/POC that manifested during the previous temperature that prevailed before? Is there a carry-over to the next day that needs to be accounted for?

Response: We have considered this phenomenon during our experiment, so during dilutions we replaced a large proportion of the culture with fresh medium (up to 80-90%) to avoid significant carry-over from the old growth phase. The ideal condition of course would be to switch from cool phase to warm phase and then cycle without dilution. However, this is impossible as dilution with fresh medium is necessary to avoid nutrient limitation setting in and confounding our results. In addition, volume removed for sampling needs to be replaced with fresh medium in our relatively small volume experimental thermal block setup.

Line 380: The abbreviation TPCs is not ideal because it can be confused with total particulate carbon. I would suggest to use no abbreviation here.

Response: We followed this suggestion and have revised the manuscript to avoid using the abbreviation TPC here, as indeed we had already used to stand for total particulate carbon. Instead, here we now write out the words 'temperature performance curve' (**Line 424-432, 457, 675, 1050-1052**).

Line 406: A particularly comprehensive assessment was done by Zhang et al., 2014 from the Reusch group. This should definitely be considered here.

Response: We followed this suggestion and have revised in the manuscript to include the Zhang et al. reference. (**Line: 452**)

Line 409: The Zhang et al., paper seems an overlooked but important paper here.

Response: As noted above, we have revised in the manuscript to include consideration of the Zhang et al. 2014 study. (Line: **453-456**)

Line 417: Schlueter et al., 2014 (also Reusch group) have shown that Ehux can quickly adapt to warming. Should be mentioned here, perhaps.

Response: We talk about rapid adaptation to warming in the following section, and we have already cited the Schlueter et al. 2014 study in this context. (**Line: 691**)

Line 476: I don't understand how this trend can suggest these things. Isn't the damage of biochemical mechanisms simply your interpretation of what may have happened. Should be rephrased.

Response: We agree that we should be more specific and support our suggestion with evidence from the literature. Accordingly, we have revised in the manuscript to point out that energetic and material investments in cellular repair machinery such as heat shock proteins are needed to deal with stressfully high temperatures, and supported this statement with a new reference (O'Donnell et al. 2018). (Line: **532-536**)

Line 607: 'ectothermic' refers to animals or also plants/microbes? Please specify.

Response: We have now stated that we are specifically referring to ectothermic animals at this point in the manuscript. We agree that even though plants and microbes can't control their body temperature either, the term ectotherm is usually reserved for animals. (Line: **667**)

Fig. 2 shows that the plasticity in PIC/POC is much larger than in the other ratios in this figure. I find this very interesting. Maybe it would be worth discussing this issue.

Response: This is an insightful comment, and so we have added new text to the Discussion to point out the large plasticity in PIC:POC ratios with temperature changes, and to discuss this observation in terms of a prior study by Krumhardt et al. (2017), as well as pointing out potential implications for ballasting of sinking particles. (**Line 499-505**)

Fig 6B: y-axis incomplete.

Response: The Y-axis scale in Fig 6 has now been extended to 1.0 in order to encompass all of the data points, thanks for pointing this out.

I hope my suggestions help the authors to improve their manuscript.

We do appreciate the constructive comments of the reviewer, and they have indeed improved the paper.

Response to Anonymous Referee #2

Interactive comment on "How will the key marine calcifier *Emiliania huxleyi* respond to a warmer and more thermally variable ocean?" by
Xinwei Wang et al.
Anonymous Referee #2
Received and published: 27 August 2019

Temperature is an important driver regulating phytoplankton physiology. Previous laboratory and field investigations suggest that the trend of global warming may strongly affect future phytoplankton communities and the consequent marine biogeochemistry. Most previous studies of warming effects on phytoplankton were mainly conducted under relatively constant temperature regimes. However, under future climate change scenario, in addition to warming (i.e. increasing mean temperature), the magnitude of temperature fluctuation will also be changed. The response pattern of marine phytoplankton to thermal variations/fluctuations is still largely unknown. The present study investigated the physiological response of a well-studied marine coccolithophore species *Emiliania huxleyi* to not only a broad range of temperature regime, but also two different frequencies (one-day and two-day) of thermal variation. The examined physiological parameters include growth, photosynthetic and calcification rates, and elemental compositions. The results suggest that higher thermal variation frequency (one-day) was less inhibitory on *E. huxleyi* physiological processes than two-day variations especially under high temperature, indicating that the frequency of temperature fluctuation may be of importance in regulating the impacts of extreme high temperature events on key phytoplankton groups. The conclusions are valuable and help to predict the relevant marine biogeochemistry under a more realistic condition of a complex and changing marine environment. In general, the manuscript is well written and organized; the results are also well explored and discussed. I would suggest the manuscript to be accepted with minor revisions. My detailed comments and suggestions are listed below.

Response: We appreciate the reviewer's thoughtful comments and enthusiasm for our study, and have described our revisions and responses to their helpful comments below.

Line 140: How often were these cultures diluted? Does this mean that steady-state growth was not observed for 28.6_C treatment?

Response: The cultures were diluted every two days the for constant and one-day variation treatments, and every four days for two-day variation treatments. (Methods, **Line 166-168**). The reviewer is correct, since a negative growth rate was calculated from the decrease of cell numbers at 28.6 ^{o}C during cultivation, the coccolithophore was unable to survive at this temperature, and growth was not at steady state- this treatment could not be diluted due to the declining biomass, and thus represents a batch culture rather than a semi-continuous one. To be certain that 28.6 ^{o}C exceeded the upper thermal limit, we repeated the experiment at this temperature several times. We have discussed this with new text on **Line 146- 150**

Lines 144-148: For the different fluctuation cycles (one-day and two-day), how was the temperature adjusted? Was temperature changed gradually during a one-day or two-day period or the cultures experienced abrupt temperature changes? Was there any lag phase for temperature changes? It would be better to provide the details of temperature fluctuation patterns in different treatments in order to better explain the observed different effects of fluctuation frequencies on *Emiliania huxleyi* physiology.

Response: The temperature setting of the thermal block setup was switched over fully (not gradually) at each transition between fluctuation cycles, but took about ½ hour to equilibrate to the new temperature after being changed, thus allowing some time for the cells to acclimate to the temperature shift. We did not observe any significant growth rate lag following the thermal shifts, just a rapid transition to a new growth rate. We have provided a detailed description in the manuscript. (**Line 158-163**)

Lines 152-155: What was the nutrient condition in the culture medium used for dilution? What do you mean by "100 _mol L-1 nitrate and 10 _mol L-1 phosphate was added every two days"? Please clarify.

Response: We adjusted the N and P midway through the 4 day cycle (2 day variation treatment) by adding concentrated Aquil stocks at these final concentrations to make sure nutrients were replete. We have revised this text in the manuscript to better describe this. (**Line 168-172**)

Line 170: Please delete "GFC"

Response: We have revised in the manuscript as suggested. (**Line 189**)

Lines 174-176: "Total Particulate Carbon" and "Particulate Organic Carbon/Nitrogen" should all be lowercased.

Response: We have made this change. (**Line 193-195**)

Line 206: I found the abbreviation of "TPC" a bit confusing here, since it refers to "total particulate carbon" in the earlier text.

Response: The abbreviation TPC for 'thermal performance curve' has been removed here, since the reviewer is correct, it was used earlier in the paper for 'total particulate carbon'. Thermal performance curve is now written out. (**Line 424-432, 457, 675, 1050-1052**).

Line 209: misspelling of *Emiliania huxleyi*

Response: We revised this mis-spelling. (**Line 225, 230**)

Line 210: Please specify how the equation was modified.

Response: Our approach used for predicting thermal response curves under variable thermal conditions (as opposed to the constant temperatures used in the classic Eppley study) was first published by Bernhardt et al. (2018). It is a non-linear averaging model that incorporates the principle of Jensen's inequality, and so is based on Eppley's equation but with these modifications to deal with fluctuating temperatures. It has been applied in published thermal variation studies by Qu et al (2019) and Kling et al. (in press), both cited here. The full derivation of this thermal variation model is too lengthy to give here, but can be obtained by interested readers from the Bernhardt paper. We changed the original confusing wording to more accurately describe this model on **Line 230-243.**

Line 251: Please rephrase the text to "The growth rates during the cool phase of the one-day variation cycle were lower than those…"

Response: We followed this suggestion and have revised the manuscript. (**Line 293-294**)

Line 419: should be revised to " can be influenced…".

Response: We revised the manuscript as suggested. (**Line 471**)

Line 596 - : In this section, it might be worth to also expand the discussion on how thermal variation would affect the competition advantage of coccolithophores over other phytoplankton functional groups (such as diatoms) in the community level.

Response: Thank you for this good suggestion. We have revised the discussion text accordingly. (**Line 678-684**)

Fig. 1. The growth rates presented in the figure were supposed to be measured during steady growth phase. However, according the context, the cultures were not able to survive at 28.6_C. I assume the negative growth rate was calculated based on the decreased in-vivo fluorescence values over the consecutive sampling days. I'd suggest using the value 0

instead of negative value for fitting at this data point.

Response: As noted above, the negative growth rate was calculated from the decrease of cell numbers at 28.6 ^{o}C during cultivation during a batch culture, an experiment which we repeated several times to robustly verify this result. The magnitude of the negative growth rate here is an expression of the degree of stress the culture experienced at this temperature, and may be useful to some readers for comparison with the other positive growth rate values in the variation experiments. We appreciate the comment, but with the editor's permission would like to keep the negative value here.

How will the key marine calcifier *Emiliania huxleyi* respond to a warmer and more thermally variable ocean?

Xinwei Wang[1], Feixue Fu[2], Pingping Qu[2], Joshua D. Kling [2], Haibo Jiang[3], Yahui Gao[1,4*], David A. Hutchins[2*]

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Key words: thermal variation, *Emiliania huxleyi*, coccolithophore, calcification, growth rate, elemental composition, global warming

Abstract

Global warming will be combined with predicted increases in thermal variability in the future surface ocean, but how temperature dynamics will affect phytoplankton biology and biogeochemistry is largely unknown. Here, we examine the responses of the globally important marine coccolithophore *Emiliania huxleyi* to thermal variations at two frequencies (one-day and two-day) at low (18.5 ^{o}C) and high (25.5 ^{o}C) mean temperatures. Elevated temperature and thermal variation decreased growth, calcification and physiological rates, both individually and interactively. One-day thermal variation frequencies were less inhibitory than two-day variations under high temperature, indicating that high frequency thermal fluctuations may reduce heat-induced mortality and mitigate some impacts of extreme high temperature events. Cellular elemental composition and calcification was significantly affected by both thermal variation treatments relative to each other, and to the constant temperature controls. The negative effects of thermal variation on *E. huxleyi* growth rate and physiology are especially pronounced at high temperatures. These responses of the key marine calcifier *E. huxleyi* to warmer, more variable temperature regimes have potentially large implications for ocean productivity and marine biogeochemical cycles under a future changing climate.

Introduction

Climate-driven changes such as ocean warming alter the productivity and composition of marine phytoplankton communities, thereby influencing global biogeochemical cycles (Boyd et al., 2018; Hutchins & Fu, 2017; Thomas, et al., 2012). Increasing sea surface temperatures have been linked to global declines in phytoplankton concentration (Boyce et al., 2010), changes in spring bloom timing (Friedland et al., 2018), and biogeographic shifts in harmful algal blooms (Fu et al. 2012; Gobler et al., 2017). Warming and acidification may drive shifts away from dinoflagellate or diatom dominance, and towards nanophytoplankton (Hare et al., 2007; Keys et al., 2018). Similarly, Morán et al. (2010) predicted that a gradual shift will occur towards smaller primary producers in a warmer ocean.

Effects of temperature increases on phytoplankton diversity are uncertain. Warming and phytoplankton biodiversity were found to be inversely correlated in a coastal California diatom assemblage, at least on short timescales (Tatters et al., 2018). In contrast, a five-year long mesocosm experiment found that elevated temperature can modulate species coexistence, thus increasing phytoplankton species richness and productivity (Yvon-Durocher et al. 2015). Globally, rising temperatures may result in losses of phytoplankton biodiversity in the tropics, but gains in the polar regions (Thomas et al., 2012). It is thought that ocean warming will lead to a poleward range expansion of warm-water species at the expense of cold-water species (Boyd et al., 2010; Gao et al., 2018; Hallegraeff, 2010; Hutchins & Fu, 2017; Thomas et al., 2012). It is evident that rising ocean temperatures will benefit some groups, while having

detrimental consequences for others (Boyd et al., 2010, 2015, 2018; Feng, et al., 2017; Fu et al., 2014). For example, recent decades of satellite observations show a striking poleward shift in the distribution of blooms of the coccolithophore *Emiliania huxleyi*, a species that was previously virtually absent in polar waters (Boyd et al., 2010; Neukermans et al., 2018).

Coccolithophores are the most successful calcifying phytoplankton in the ocean, and contribute almost half of global marine calcium carbonate production. They play crucial biogeochemical roles by performing both photosynthesis and calcification, and facilitate carbon export to the deep ocean through the ballasting effects of their calcium carbonate shells (Klaas & Archer, 2002; Krumhardt et al., 2017; Monteiro et al., 2016). *E. huxleyi* (Lohm.) is the most abundant and cosmopolitan coccolithophore, forming prolific blooms in many regions (Holligan, et al., 1983; 1993; Iglesias-Rodríguez et al., 2002; Westbroek et al., 1993).

The responses of *E. huxleyi* to global change factors have been intensively investigated. Many *E. huxleyi* strains are sensitive to ocean acidification, which negatively affects their growth rates and calcification (Feng et al., 2018; Hoppe et al., 2011). However, among the many currently changing environmental drivers, temperature may be among the most important in regulating coccolithophore physiology (Boyd et al., 2010). Feng et al. (2008) reported that the growth rate of *E. huxleyi* was improved by elevated temperature at low irradiance. Furthermore, temperature was the most important driver controlling both cellular particulate organic and inorganic carbon content of a Southern Hemisphere *E. huxleyi* strain (Feng et al.,

84 2018).

85 Most research about the effects of global warming on *E. huxleyi* and
86 phytoplankton in general has focused on predicted increases in mean temperatures.
87 However, in the natural environment, seawater temperatures fluctuate over timescales
88 ranging from hours, to days, to months (Bozinovic et al., 2011; Jiang et al., 2017).
89 Future climate models predict not only in an increase in mean temperature, but also an
90 increase in temperature variability (frequency and intensity), as well as a higher
91 probability of extreme events (IPCC 2013).

92 The impacts of climatic variability and extremes have been best studied in
93 metazoans, where they may sometimes have a larger effect than increases in climatic
94 averages alone (Vázquez et al., 2017; Vasseur et al., 2014; Zander et al., 2017).
95 Variability can promote greater zooplankton species richness, compared with long-term
96 average conditions (Cáceres 1997; Shurin et al. 2010). In corals, temperature variability
97 could buffer warming stress, elevate thermal tolerance and reduce the risk of bleaching
98 (Oliver & Palumbi, 2011; Safaie et al., 2018).

99 In comparison, we still lack a thorough understanding of how thermal variation
100 affects phytoplankton growth and physiology. Unlike zooplankton, the few available
101 studies suggest increasing thermal variation may decrease phytoplankton biomass and
102 biodiversity, and shift the community towards small phytoplankton (Burgmer &
103 Hillebrand, 2011; Rasconi et al., 2017). Two studies have shown that plastic responses
104 play a key role in acclimation and adaptation to thermal fluctuations in algae (Kremer
105 et al., 2018; Schaum & Collins, 2014). Population growth rates of phytoplankton in

fluctuating thermal environments have been quantitatively modeled based on data from thermal response curves obtained under constant temperatures (Bernhardt et al., 2018).

In view of this relative lack of information on the effects of non-steady state temperatures on biogeochemically important phytoplankton, we carried out a thermal variability study using the Sargasso Sea *E. huxleyi* isolate CCMP371. Our experiments combined ocean warming with thermal variations, with a focus on the increasing frequency of temperature variations under global climate change. We examined growth rates, photosynthesis, calcification and elemental composition under constant, one-day and two-day temperature variations. This study is intended to provide insights into how different frequencies of thermal variation may influence the physiology and biogeochemistry of this important marine calcifying phytoplankton species under both current and future sea surface temperatures.

Materials and methods

The marine coccolithophore *E. huxleyi* (Lohm.) Hay and Mohler stain CCMP371 (isolated from the Sargasso Sea) was maintained in the laboratory as stock batch cultures in Aquil medium (100 μmol L^{-1} NO_3^- , 10 μmol L^{-1} PO_4^{3-}) made with 0.2 μM-filtered coastal seawater collected from the California region (Sunda et al., 2005). Cells were grown at 22 ºC under 120 μmol photons m^{-2} s^{-1} cool white fluorescent light with a 12 h/12 h light/dark cycle.

Experimental set-up

An aluminum thermal gradient block with a range of 13 temperatures was used to perform the thermal response curve and temperature variation experiments. For the

thermal curve experiment, the extreme temperatures of the thermal-block were set to 8.5 °C and 28.6 °C, with intermediate temperatures of 10.5 °C, 12 °C, 13.5 °C, 15.5 °C, 17.5 °C, 18.5 °C, 21.3 °C, 22.6 °C, 24.5 °C, 26.6 °C, and 27.6 °C. The *E. huxleyi* cells were transferred from the stock cultures into triplicate 120 ml acid washed polycarbonate bottles in the thermal block under a 12 h light /12h dark cycle at 180 µmol photons m^{-2} s^{-1}. For the light intensity measurement, irradiance was measured individually at each position in the thermal block using a light meter with a small detector bulb to fit into the round holes drilled to fit the experimental bottles (LI-250A light meter, LI-COR). During measurements the detector bulb was positioned identically in each position, and if necessary, the positions of the fluorescent lights were adjusted nearer or farther until the light intensity was between 175-185 µmol photons m^{-2} s^{-1} for every experimental replicate.

Semi-continuous culturing methods were used for all experiments. Cultures were diluted with Aquil medium every two days to keep them in exponential growth stage while acclimating to the treatment temperatures for two weeks before starting the variation experiment. Dilution volumes were calculated to match growth rates of each individual replicate, as measured using *in vivo* chlorophyll a (Chl *a*) fluorescence. Once steady-state growth rates were recorded for 3–5 consecutive transfers, the cultures were sampled (Zhu et al., 2017). Due to the decrease of cell numbers during cultivation at 28.6 °C (from our preliminary experiment), these cultures were diluted from 22 °C stock cultures. They were then sampled as a batch culture (without dilution) after 4-6 days to estimate the negative growth rates and elemental stoichiometry at this

upper limit temperature point.

Six treatments were used to determine the responses of *E. huxleyi* growth, photosynthesis and calcification to different frequencies of temperature fluctuation. Temperature fluctuation treatments included: 1) Low temperature, constant (18.5 ^{o}C). 2) Low temperature, one-day fluctuation cycle (16-21^{o}C, mean = 18.5^{o}C). 3) Low temperature, two-day fluctuation cycle (16-21^{o}C, mean =18.5^{o}C). 4) High temperature, constant (25.5 ^{o}C). 5) High temperature, one-day fluctuation cycle (23-28^{o}C, mean = 25.5^{o}C). 6) High temperature, two-day fluctuation cycle (23-28^{o}C, mean = 25.5^{o}C). For the variation treatment cycles, cultures were incubated at the cool phase (16 ^{o}C and 23 ^{o}C for low and high temperatures, respectively) for either one or two days. They were then switched to the warm phase (21 ^{o}C and 28 ^{o}C for low and high temperature, respectively) for the same amount of time. It took about 1/2 hour to re-adjust the thermal block to the transformed temperature at the beginning of each new treatment cycle. The experimental *E. huxleyi* cultures were grown in triplicate in 120 ml acid washed polycarbonate bottles using the thermal-block under a 12 h light /12h dark cycle at 180 µmol photons m^{-2} s^{-1}.

For the variable temperature experiment, cultures were diluted semi-continuously with Aquil medium every two days for constant and one-day variation treatments, and every four days for two-day variation treatments. To ensure nutrient-replete conditions in the two-day variation treatments, Aquil nitrate and phosphate stocks were added at the two-day midpoint of every four days thermal cycle to make sure that the final nitrate and phosphate concentrations were not depleted and were always maintained

at >100 μmol L^{-1} and >10 μmol L^{-1}, respectively. Cultures were grown for at least eight dilutions (~16 days for constant and one day variation treatments; ~32 days for two-day variation treatments) to acclimate to the different experimental conditions before final sampling. All variation treatments were sampled twice across the thermal variation cycle, once during the cool phase and once during the warm phase.

Growth rates

In vivo fluorescence was measured daily for the one-day variation treatment and every two days for the constant and two-day variation treatments using a Turner 10-AU fluorometer (Turner Designs, CA). *In vivo*-derived growth rates were subsequently verified using cell samples counted with a nanoplankton counting chamber on an Olympus BX51 microscope. Specific growth rates (d^{-1}) were calculated using the *in vivo* fluorescence and cell count data as: $\mu=\ln[N(T_2)/N(T_1)]/(T_2-T_1)$, in which $N(T_1)$ and $N(T_2)$ are the *in vivo* fluorescence values (for thermal curve experiments and constant treatments) or cell counts (for variation treatments, because of potential changes in cellular *in vivo* fluorescence during fluctuation) at T_1 and T_2.

Chl *a* analysis

Twenty ml culture samples were filtered onto GF/F glass fiber filters (Whatman, Maidstone, UK) for Chl *a* analysis. In vitro Chl *a* was extracted with 90% aqueous acetone for 24 hours at -20 ^{o}C, and then measured using a Turner 10-AU fluorometer (Turner Design, USA). (Fu et al., 2007).

Elemental analysis

Elemental composition sampling included total particulate carbon (TPC),

particulate organic carbon (POC), particulate organic nitrogen (PON), particulate inorganic carbon (PIC) and particulate organic phosphorus (POP), allowing calculation of cellular elemental stoichiometry and calcite/organic carbon ratios (PIC/POC) (Feng et al.; 2008). Culture samples for TPC, POC and PON, were collected onto pre-combusted GF/F glass fiber filters (Whatman) and dried in a 60 ^{o}C oven overnight. For POC analysis, filters were fumed for 24 hours with saturated HCl (~37%) to remove all inorganic carbon prior to analysis. TPC, PON and POC were then measured by a 440 Elemental Analyzer (Costech Inc, CA) according to previous studies (Hutchins et al., 1998; Feng et al., 2008). PIC was calculated as the difference between TPC and POC. For POP measurement, culture samples were filtered onto pre-combusted GF/F filters (Whatman) and analyzed using a molybdate colorimetric method (Solórzano and Sharp, 1980), with minor modifications as in Fu et al. (2007).

Total carbon fixation, photosynthetic and calcification rates & ratios

Total carbon fixation, photosynthetic carbon fixation and calcification rates were measured using the ^{14}C incubation technique (Platt et al., 1980) with slight modifications as in Feng et al. (2008). Sixty mL culture samples from each treatment were spiked with 0.2 μCi $NaH^{14}CO_3$ and then incubated for 4 h under their respective experimental conditions. After incubation, samples were filtered on two Whatman GF/F filters (30mL each) for total carbon fixation and photosynthetic rate separately. The filters for photosynthetic rate measurement were fumed with saturated HCl (~37%) before adding scintillation fluid. Thirty mL from each treatment (10 mL from each

replicate bottle) was filtered immediately, after adding equal amounts of $NaH^{14}CO_3$ for procedural filter blanks. Filters were then placed in 7 mL scintillation vials with 4 mL scintillation fluid overnight in the dark. To determine the total radioactivity (TA), 0.2 µCi $NaH^{14}CO_3$ together with 100 µL phenylalanine was placed in scintillation vials with the addition of 4 mL scintillation solution. All samples were counted on a Perkin Elmer Liquid Scintillation Counter to measure the radioactivity. Total carbon fixation and photosynthetic rate were calculated from TA, final radioactivity and total dissolved inorganic carbon (DIC) values. Calcification rate was then calculated as the difference between total carbon fixation and photosynthetic rate for each sample.

Model for population growth of *E. huxleyi*

Growth rates measured under constant temperatures in the thermal block were fitted to the Eppley thermal performance curve (Eppley, 1972; Norberg, 2004; Thomas et al., 2012). This function quantifies parameters of growth temperature effects, including the temperature optimum for growth (T_{opt}), and high and low temperature limits (T_{max} and T_{min} respectively) in our strain of *E. huxleyi*. To predict growth rates under variable thermal regimes from our constant temperature thermal curves, we applied a recently developed model based on the Eppley curve, but incorporating non-linear averaging in conjunction with consideration of Jensen's inequality (Bernhardt et al., 2018). This new thermal fluctuation model takes into account the amount of time that the cells spend at each portion of their thermal performance curve, as well as incorporating the observation that growth rates usually increase slowly with temperature at the cooler end of the curve, but then drop off very quickly at the upper, warm end of the curve

(Jensen's inequality). This model is more realistic and skillful under variable temperatures than previous work assuming a linear relationship between temperature and growth rates, as non-linear averaging allows much more accurate predictions when dealing with skewed thermal curves. Several recent studies of phytoplankton thermal variability responses have successfully applied the Bernhardt et al. (2018) model (Qu et al. 2019, Kling et al. in press).

Statistical analysis

The mean values of most parameters measured under the variation treatments were calculated by averaging the values from the cool and warm phases, including all the elemental content and ratios, photosynthetic and calcification rates and ratios. Statistical analyses were performed using R (version 3.5.0). For the response of *E. huxleyi* to warming, the mean growth rate or elemental ratios of three replicates at 12 temperature points were used to fit the growth rate or elemental ratios curves. A one-way ANOVA was applied to analyze the difference between the average value for the entire temperature range and the value at each individual temperature for the elemental ratios. For the response of *E. huxleyi* to thermal variation, a one-way ANOVA was performed to test the statistical significance in growth rate, elemental stoichiometry, photosynthetic and calcification rates and ratios among different frequencies (constant, one-day and two-day) of temperature variabilities at cool/warm phases under high/low temperatures. *p*-values were calculated based on Student's t-test via two functions including compare_means() and stat_compare_mean() in the ggpubr package, and the figures were generated via the ggplot package in open source statistical software R

version 3.5.0 (R Foundation).

Results

Responses of *E. huxleyi* to warming

The growth rates of *E. huxleyi* at constant temperature increased significantly with warming from 0.09±0.01 d^{-1} at 8.5 ºC to a maximum value of 0.90±0.02 d^{-1} at 21.3 ºC. Growth was optimal up to 24.5 ºC, and then decreased rapidly to -0.46±0.05 d^{-1} at 28.6 ºC ($p<0.05$, Fig. 1).

The elemental ratios of the cells in the different temperature treatments were compared to the average elemental ratios across the entire temperature range (Fig. 2). The thermal trends of TPC/PON ratios were generally similar with those of growth rates, in that ratios increased from 8.5 to 17.5 ºC, and then decreased from 24.5 to 27.6 ºC. The TPC/PON ratios at 8.5, 10.5 and 27.6 ºC were significantly lower than the average level of all the temperature points ($p<0.05$, Fig 2A). The POC/PON ratios of most temperature points were very close to the mean value of 6.3, except at 27.6 ºC (7.1) and 28.6 ºC (7.4), which were significantly higher than the average ($p<0.05$, Fig 2B). The highest PIC/POC ratio was 0.49±0.07 at 22.6 ºC, and the lowest PIC/POC ratio was 0.05±0.04 at 27.6 ºC, a value that was almost 90% less than the highest value. The PIC/POC ratios at the lowest temperature tested (10.5 ºC) and at the high end of the temperature range (26.6 and 27.6 ºC) were significantly lower than the average level (Fig. 2C). Chl *a*/POC ratios were significant lower at 8.5, 10.5 and 27.6 ºC than the mean, and at 17.5, 21.3, 22.6 and 24.5 ºC were significantly higher than the average ($p<0.05$, Fig. 2C). The trends of PIC/POC and Chl *a*/POC ratio were similar, in that

282 they gradually increased from low temperature to the highest value at 22.6 ^{o}C, and
283 then dropped rapidly as temperature increased further. (Fig. 2C, D).

284 **Responses of *E. huxleyi* to temperature variations**

285 **Growth rate**

286 In low temperature experiments, both one-day and two-day temperature variations
287 had a negative effect on growth rate. The mean growth rates of the one-day (0.71±0.01
288 d^{-1}) and two-day (0.72±0.01 d^{-1}) variation treatments were not significantly different
289 from each other ($p>0.05$), but both were lower than that of the constant 18.5 ^{o}C
290 treatment (0.76±0.01, $p < 0.05$) (Fig. 3A). Growth rates were low during the cool phase
291 (16 ^{o}C) of the experiment (~0.5-0.6 d^{-1}), but those of the two-day variation cycle were
292 not significantly different from the constant control at this temperature ($p>0.05$).
293 However, the growth rates during the cool phase of the one-day variation cycle were
294 lower than those of the constant 16 ^{o}C treatment ($p<0.05$). During the warm phase of
295 the thermal cycle (21^{o}C), there were no significant differences in the elevated growth
296 rates (~0.85-0.9 d^{-1}) of the constant control and those of either variable treatment
297 ($p>0.05$, Fig. 3A).

298 In the high temperature experiments, as in the low temperature experiments, both
299 temperature variation frequencies had a negative effect on mean growth rates. The
300 growth rates in the two-day variation treatment were (0.20±0.02 d^{-1}), a decrease of
301 ~74% compared with the constant 25.5 ^{o}C ($p<0.05$), and ~62% of the one-day
302 variation treatment value ($p<0.05$, Fig. 3B). During the cool phase (23 ^{o}C), the growth
303 rate of the one-day variation treatment was slightly lower ($p<0.05$) than the constant

304 23 °C, but there were no significant changes between two-day variations and the
305 constant 23 °C treatment ($p > 0.05$, Fig. 3B). During the warm phase (28 °C), the
306 constant 28 °C and two-day variation treatment both had negative growth rates of -
307 0.45±0.05 d^{-1} and -0.45±0.04 d^{-1}, respectively. However, the one-day variation
308 treatment had a low but positive warm phase growth rate at 0.25±0.02 d^{-1} (Fig. 3B).
309 There was a time lag of ~1/2 hour to switch to the transformed temperature for each
310 new growth phase, which should thus have had only minimal effects on overall growth
311 rates across the one-day and two-day thermal variations.

312 **Cellular PIC and POC contents and ratios**

313 In low temperature experiments, the cellular PIC content of the constant 18.5 °C
314 treatment was 3.5±0.3 pg/cell, and there were no significant differences with
315 temperature variation treatments ($p> 0.05$, Table 1). However, the cellular POC
316 content of the constant 18.5 °C treatment was 8.0±0.6 pg/cell, which was lower than
317 in the two-day variation treatment, but significantly higher than in the one-day
318 variation treatment ($p<0.05$).

319 Like POC, the PIC/POC ratio was significantly affected by temperature variations
320 (Fig. 4A). The lowest PIC/POC ratio was found in the one-day variation treatment
321 (0.38 ±0.07), which was significantly lower than the two-day variation treatment value
322 ($p < 0.05$), but close to that in the constant 18.5 °C ($p > 0.05$). A similar trend was
323 found in both the cool (16 °C) and warm phases (21 °C) of the two variation treatments,
324 in that the PIC/POC ratio of the one-day variation treatment was lower than of the
325 two-day variation treatment ($p < 0.05$, Fig. 4A). Both variation treatments had lower

PIC/POC ratios during the warm phase than during the cool phase, although these differences were not significant ($p>0.05$).

High temperature experiments showed particulate carbon trends that were contrary to those of the low temperature treatments. The PIC content and PIC/POC ratios were significantly decreased by temperature variation. The cellular PIC content of the constant treatment (25.5 ^{o}C) was 5.5±0.3 pg/cell, which was ~ 200% higher than that of the one-day variation and ~ 160% higher than in the two-day variation treatments ($p<0.05$, Table 1). The same trend was found for PIC/POC ratios in one-day variation and two-day variation treatments, which decreased ~ 67% and 33% compared with the constant 25.5 ^{o}C treatment, respectively ($p<0.05$, Fig. 4B). However, the POC content of one-day and two-day variation treatments was higher than in the constant 25.5 ^{o}C treatment ($p < 0.05$,Table 1). During the cool phase (23 ^{o}C), the PIC content and PIC/POC ratio of the one-day variation treatment was significantly lower than in the two-day variation treatment, but contrary to PIC content, the POC content of the one-day variation treatment was significantly higher than that in the two-day variation treatment. During the warm phase (28 ^{o}C), there were no significant differences of PIC content, POC content, or PIC/POC ratio between the one-day and two-day variation treatments (Fig. 4B, Table 1).

Photosynthetic and calcification rates and ratios

In low temperature treatments, there were no differences between total carbon fixation rates (photosynthesis plus calcification) for the two variable treatments relative to the constant control (Fig. 5A). However, during the cool phase total

carbon fixation rates were higher in the one-day variation than in the two-day variation ($p<0.05$, Fig 5A), while this rate was the same in both variation treatments during the warm phase ($p > 0.05$, Fig. 5A). In high temperature experiments, the total carbon fixation rates of the one-day and two-day variation treatments were significantly decreased by about ~20% and ~18% respectively, compared with the constant 25.5 ^{o}C treatment ($p<0.05$, Fig. 5 B).

The photosynthetic and calcification rates of the constant 18.5 ^{o}C treatment were 0.04±0.00 pmol C cell^{-1} hr^{-1} and 0.02±0.00 pmol C cell^{-1} hr^{-1}, respectively, which were not significantly different from both of the temperature variation treatments ($p > 0.05$, Fig. 5 C,E). Photosynthetic rates changed within the thermal cycle for both one-day and two-day variation treatments, with a decrease of 22% and 28% from the warm phase to the cool phase, respectively (Fig. 5C). However, there were no significant changes in calcification rates under either variation frequency treatment between the cool and warm phases of the thermal cycles ($p > 0.05$).

In the mean 25.5 ^{o}C experiment, photosynthetic rates were not significantly different between the one-day variation and constant treatments ($p > 0.05$), while the photosynthetic rate of the two-day variation was slightly higher than that of the constant 25.5 ^{o}C treatment ($p<0.05$, Fig. 5D). In contrast, calcification rates of one-day and two-day variation treatments at a mean temperature of 25.5 ^{o} were significantly decreased by about ~46% and ~51%, respectively, relative to the constant control ($p<0.05$, Fig. 5F). There were no significant differences in total carbon fixation, photosynthetic and calcification rates between the one-day variation and two-day

variation treatments during both the cool (23 ^{o}C) and warm (28 ^{o}C) phases ($p>0.05$, Fig. 5 B,D,F).

In the low temperature treatments, there were no significant differences in calcification to photosynthesis (Cal/Photo) ratios between the constant and the two variable treatments ($p > 0.05$, Fig 6A). In contrast, in the high temperature experiments, the Cal/Photo ratio of the one-day variation and two-day variation treatments were decreased by ~40% and 49%, respectively, compared with the constant 25.5 ^{o}C treatment ($p<0.05$, Fig. 6B). For both low and high temperature experiments, there were no significant differences between the one-day and two-day variation treatments in either the cool or warm phases of the thermal cycle ($p > 0.05$, Fig. 6B). However, in both temperature treatments the lower photosynthetic rates during the cool phase (Fig. 5C,D) resulted in an increase in the Cal/Photo ratio during the cool phase for both the one-day and two-day variation treatments ($p<0.05$ Fig. 6A,B).

Elemental content and stoichiometry

In the low temperature experiments, the one-day variation and two-day thermal variations had different effects on cellular elemental contents and ratios, relative to the constant 18.5 ^{o}C treatment. One-day variation increased most of the cellular elemental and biochemical contents (TPC, PON, and Chl *a*) but with no significant difference ($p>0.05$), except for POP content ($p<0.05$), compared with the constant 18.5 ^{o}C treatment (Table 1). In contrast, the two-day variation treatment decreased all the measured cellular elemental and biochemical contents (TPC, PON, POP and Chl *a*,

392 $p<0.05$) in relation to the constant 18.5 ^{o}C treatment (Table 1). However, the
393 TPC/PON and Chl *a*/POC ratios of the two-day variation treatment were higher than
394 those of the one-day variation and constant 18.5 ^{o}C treatments ($p<0.05$, Fig. 7A,E),
395 while the PON/POP ratio was lower than in the one-day variation and constant 18.5
396 ^{o}C treatments ($p<0.05$, Fig. 7C). There were no significant differences in TPC/PON,
397 PON/POP and Chl *a*/POC ratios between the constant 18.5 ^{o}C and the one-day
398 variation treatments ($p > 0.05$, Fig. 7A).

399 In high temperature experiments, the highest cellular TPC, PON and POP contents
400 were all obtained under the one-day variation treatment, which was significantly
401 higher than under constant 25.5 ^{o}C conditions ($p<0.05$, Table 1). However, there were
402 no significant differences in cellular Chl *a* content between the constant 25.5 ^{o}C and
403 both variation treatments ($p > 0.05$, Table 1). The TPC/PON ratio of the constant 25.5
404 ^{o}C treatment was ~22% and ~35% higher than that of the two-day variation and one-
405 day variation treatments, respectively ($p<0.05$, Fig. 7B), while the PON/POP ratio was
406 highest in the one-day variation, followed by the two-day variation and finally by the
407 constant control (Fig. 7D). The Chl *a*/POC ratio of the one-day variation treatment
408 was significantly lower than that of the constant 25.5 ^{o}C and two-day variation
409 treatments ($p<0.05$), but there were no significant differences between the constant
410 25.5 ^{o}C and two-day variation treatments ($p > 0.05$, Fig. 7F).

411 During the cool phase of the high temperature experiments (23 ^{o}C), the cellular
412 TPC, PON, POP and Chl *a* content of two-day variation were all significantly lower
413 than in the one-day variation treatment ($p<0.05$). Similar decreasing trends during the

414 cool phase were observed for the TPC/PON ratios (Fig. 7B), but not the Chl *a*/POC
415 ratio, which was ~32% higher than in the one-day variation treatment ($p<0.05$, Fig.
416 7F). During the warm phase (28 ^{o}C), there were no significant differences of cellular
417 TPC, PON and POP contents between one-day and two-day variation treatments ($p >$
418 0.05, Table 1) as well as the TPC/PON ratio (Fig 7B). However, the Chl *a* content of
419 the one-day variation treatment was ~20% lower than that of the two-day variation
420 treatment ($p<0.05$). The Chl *a*/POC ratio was not significantly different between the
421 one-day and two-day variation treatments at the warm phase ($p > 0.05$, Table 1, Fig.
422 7F).

423 **Experimental constant temperature performance curves and measured and**
424 **modeled fluctuating temperature performance curves**

425 The experimentally-determined constant condition temperature performance
426 curves and the predicted fluctuating temperature condition temperature performance
427 curves based on the Bernhardt et al. (2018) non-linear averaging model are shown in
428 Fig. 8 for *E. huxleyi*. Compared with the measured temperature performance curve
429 under constant thermal conditions, the modeled curve of the fluctuating temperature
430 condition showed a leftward shift towards lower temperatures at optimum
431 temperatures and above. The maximum and optimal temperature of the modeled
432 fluctuating temperature performance curve were all lower than those of the measured
433 constant condition curve. In particular, the optimal temperature for growth decreased
434 from 22^{o}C in constant conditions to 21 ^{o}C under fluctuating temperature conditions.
435 At the same time, the maximum growth rate (μ_{max}) of the fluctuating temperature

condition was 0.8 d^{-1}, which was lower than the constant condition value of 0. 9 d^{-1}. The measured growth rates of experimental one-day (0.71 d^{-1}) and two-day (0.72 d^{-1}) variation treatments at the relatively low mean temperature of 18.5 ^{o}C closely matched the model-predicted fluctuating temperature growth rate at this temperature (0.74^{-1}, Fig. 8). However, measured and predicted growth rates did not match as well at the higher mean temperature. At 25.5 ^{o}C, the measured growth rate of the one-day variation was 0.52 d^{-1}, 30% higher than the predicted fluctuating temperature growth rate of 0.40 d^{-1}. In contrast, the measured growth rate of the experimental two-day variation treatment was 0.20 d^{-1}, a decrease of 50% compared to the model-predicted fluctuating temperature growth rate of 0.40 d^{-1} at this temperature (Fig. 8).

Discussion

Effects of warming on *Emiliania huxleyi* growth rates and elemental ratios

Thermal response curves and optimum growth temperatures describe the importance of temperature as a control on the distribution of *E. huxleyi* strains in the ocean (Buitenhuis et al., 2008; Paasche, 2001). The optimal temperature range of 21.3-24.5 ^{o}C found in our study is similar to that of some other *E. huxleyi* strains (De Bodt et al., 2010; Feng et al., 2017; Rosas-Navarro et al., 2016; Zhang et al., 2014). Most studies have focused on the lower part of the temperature curve where growth rates increase with rising temperatures (Feng et al., 2017; Matson et al., 2016), with relatively few examining stressfully warm temperatures where growth is inhibited (Zhang et al., 2014).

In our study, the descending portion of the upper temperature performance curve

458 ranged from 24.5 °C to 28.6 °C, at which point growth rates became negative. We
459 repeated the upper thermal limit part of the curve several times to rigorously verify that
460 cultures were unable to grow at this temperature. The magnitude of the negative growth
461 rate is presented here as it represents an expression of the degree of stress the culture
462 experienced at this temperature, and so makes a useful comparison with the other
463 positive growth rate values in the variation experiments. This *E. huxleyi* strain was
464 isolated from the Sargasso Sea where the sea surface temperature can reach 29 °C in
465 the summer, and will undoubtedly be higher in the future with global warming
466 (https://seatemperature.info/sargasso-sea-water-temperature.html). This suggests that
467 this strain may be currently living at or very near its upper thermal limit for part of the
468 year, as are many other tropical and subtropical phytoplankton (Thomas et al. 2012),
469 and that it may therefore be vulnerable to further warming.

470 Calcification is the key biogeochemical functional trait of this species, and the
471 PIC/POC ratio of *E. huxleyi* can be influenced by factors that include CO_2 concentration,
472 nutrient status, irradiance and temperature (Feng et al., 2008, 2017; Raven & Crawfurd,
473 2012). The cellular PIC/POC of *E. huxleyi* has been reported to decrease as irradiance
474 and CO_2 concentration rise, but to increase under nitrate and phosphate limitation (Feng
475 et al., 2017; Paasche, 1999; Riegman et al., 2000). The effect of temperature on *E.*
476 *huxleyi* cellular PIC/POC ratio is however more complex. De Bodt et al. (2010) and
477 Gerecht et al. (2014) observed that higher cellular PIC/POC ratios were obtained at
478 lower temperatures for both *E. huxleyi* and *Coccolithus pelagicus*. Sett et al. (2014),
479 however, found an opposite trend, whereby the PIC/POC ratio increased with

480 temperature in another strain of *E. huxleyi*. Feng et al. (2017) reported that the cellular
481 PIC/POC of *E. huxleyi* was increased as the temperature rose from 4 ^{o}C to 11 ^{o}C, but
482 decreased with warming from 11 ^{o}C to 15 ^{o}C and remained steady afterwards.

483 In our study, the cellular PIC/POC ratio of *E. huxleyi* was positively correlated to
484 growth rate (R^2=0.73), and increased with warming from 8.5 ^{o}C to a maximum at 22.6
485 ^{o}C, and then decreased with further warming to 27.6 ^{o}C. In a meta-analysis of studies
486 using different coccolithophore subgroups, Krumhardt et al. (2017) found that the
487 highest PIC/POC ratios were observed between 15 °C and 20 °C, in the same thermal
488 range where the highest growth rates of *E. huxleyi* are found, as seen here and in Sett
489 et al. (2014). In contrast, Rosas-Navarro et al. (2016) reported that the cellular PIC/POC
490 ratio showed a minimum at optimal growth temperature (between 20 and 25 °C) for
491 three strains of *E. huxleyi*. However, the *E. huxleyi* strain used here was isolated from
492 a warmer area (the Sargasso Sea) compared with isolates from coastal Japan and New
493 Zealand in previous studies (Rosas-Navarro et al. 2016; Feng et al. 2017). The growth
494 temperature for our stock cultures was 22-24^{o}C, higher than that of the other two *E.*
495 *huxleyi* strains. Feng et al. (2017) also found that the optimal temperature for
496 calcification was close to the stock culture maintenance temperature in their study.

497 Our results also support suggestions that stressful high temperatures may lead to
498 decreases in cellular PIC/POC ratios and calcification (De Bodt et al., 2010; Feng et al.,
499 2017; Gerecht et al., 2014; Krumhardt et al., 2017). The cellular PIC/POC ratio of *E.*
500 *huxleyi* was much more plastic than the other ratios we measured, including TPC/PON
501 and POC/PON. Indeed, PIC/POC ratios may change dramatically (>2-fold) with

temperature for some coccolithophore subgroups (Krumhardt et al., 2017). The plasticity in PIC/POC ratios of *E. huxleyi* during temperature changes in our study may have implications for shifts in the ballasting of coccolith-containing particles during sinking, thus affecting the ocean carbon cycle.

The cellular Chl *a*/POC ratio of *E. huxleyi* showed a similar pattern with the PIC/POC ratio, as it was also positively correlated to growth rate. Zhu et al. (2017) reported the cellular Chl *a*/POC ratio of a Southern California diatom was also correlated to growth rate across a very similar temperature range. In contrast, Feng et al. (2017) found that the cellular Chl *a*/POC ratio of *E. huxleyi* dramatically decreased with warming. However, in our experiments, the cellular Chl *a*/POC ratio was lower at 27.6 ^{o}C than at 28.6 ^{o}C, likely due to the negative growth rates and consequent lack of acclimation of the cultures maintained at the highest temperature. Traits such as PIC/POC ratios, Chl *a*/POC ratios and TPC/PON ratios also showed some evidence for possible carryover from the stock cultures (22-24 ^{o}C) in this 28.6 ^{o}C treatment, as we were forced to sample before the cells died completely, after only 2-3 cycles of dilution.

Effect of thermal variation on *Emiliania huxleyi* growth and physiology

Constant vs variable temperature

Thermal variability in the surface ocean is becoming an increasingly relevant topic as global warming proceeds. In our study, we found that the growth rates of a subtropical *E. huxleyi* strain were quite sensitive to temperature variation under both low (18.5 ^{o}C, “winter”) and high (25.5 ^{o}C, “summer”) mean temperatures. In both low and high temperature experiments, growth rates always decreased under temperature

524 variation, compared with the constant mean temperature. This result agrees with
525 previous studies showing that temperature variation slowed the growth rates of the
526 fresh water green alga *Chlorella pyrenoidosa* and the marine diatom *Cyclotella*
527 *meneghiniana*, as observed in laboratory work but also during long-term field
528 observations (Zhang et al., 2016).

529 This growth rate inhibition under temperature variation was more pronounced at
530 high temperature than at low temperature, indicating that variability at the warm range
531 boundary will have a stronger negative effect on population growth rate than
532 variability near the lower thermal limits (Bernhardt et al., 2018). This trend suggests
533 that acclimation to high temperature (whether constant or variable) may require greater
534 investment in cellular repair machinery, such as heat shock proteins, thus potentially
535 diverting nutrient and energy supplies and thereby reducing growth rates (O'Donnell
536 et al., 2018). However, following Jensen's inequality model to predict the thermal
537 performance curve, there should be an inflection point where the transfer between
538 positive and negative effects of temperature variability will occur compared with the
539 constant thermal curve. Conversely, for phytoplankton living in regions of suboptimal
540 temperatures, thermal variation can enhance growth (Bernhardt et al., 2018). Thus, for
541 some polar phytoplankton or for temperate species extending their ranges poleward,
542 such as *E. huxleyi* (Neukermans et al., 2018), not only warming but also thermal
543 variability may need to be taken into consideration in order to understand changes in
544 high latitude microbial communities and biogeochemistry cycles.

545 Temperature variation affected the physiology of *E. huxleyi* differently compared

546 with constant temperature. Physiological traits that were affected by thermal
547 fluctuations also differed at low temperature ("winter") and high temperature
548 ("summer"), suggesting different response mechanisms. Under low temperature
549 variations (16-21 ^{o}C), photosynthesis and calcification were correlated with
550 temperature, leading to rates similar to those observed with constant temperature.
551 However, elemental contents and ratios under thermal variations differed from
552 constant temperature. For instance, the cellular POC, PON, POP and Chl *a* contents
553 increased during one-day variations but decreased during two-day variations,
554 compared with constant temperature.

555 These cellular quota changes were reflected in elemental ratio differences
556 (PIC/POC, Chl a/POC and TPC/POC) between the thermal variation treatments and
557 constant temperature. However, the changes between thermal variation and constant
558 treatments were not significant under low temperature ("winter"), indicating that the
559 thermal variation wouldn't significantly influence biogeochemical cycles under these
560 conditions. Unlike constant temperature treatments where selection may favor a higher
561 growth rate, the trade-off for the thermal variation treatments may involve sacrificing
562 increased growth rate in order to adjust cellular stoichiometry to adapt to the
563 fluctuating environment.

564 In contrast, photosynthetic and calcification rates under high temperature thermal
565 variations (23-28 ^{o}C) were significantly different from those seen under constant
566 temperature (25 ^{o}C), especially the calcification rate. Thermal variation treatments
567 transiently but repeatedly experienced the extreme high temperature point (28 ^{o}C),

568 leading to extremely low calcification rates and PIC contents, and thus relatively low
569 PIC/POC and Cal/Photo ratios. Previous *E. huxleyi* studies agree that high temperature
570 decreases PIC content, PIC/POC ratios and Cal/Photo ratios (Feng et al., 2017; 2018;
571 Gerecht et al., 2014). The two different patterns of responses to thermal variation we
572 observed under low and high temperatures imply a seasonal pattern in the ways that
573 thermal variations will affect the elemental stoichiometry of *E. huxleyi* .

574 Under other stresses such as nutrient limitation, trade-offs between growth rates
575 and resource affinities may be necessary to adapt to thermal changes. For instance,
576 nitrate affinity declines in cultures of the large centric diatom *Coscinodiscus*
577 acclimated to warmer temperatures (Qu et al. 2018), while warming decreases cellular
578 requirements for iron in the nitrogen-fixing cyanobacterium *Trichodesmium* (Jiang et
579 al. 2018). In nitrogen-limited cultures of the marine diatom *Thalassiosira pseudonana*,
580 long-term thermal adaptation acted most strongly on systems other than those involved
581 in nitrate uptake and utilization (O'Donnell et al., 2018). Thus, it is possible that our
582 thermal response results with *E. huxleyi* might have differed under nutrient-limited
583 growth conditions.

584 ***One-day vs two-day thermal variation***

585 As temperature fluctuations in the surface ocean increase along with climate
586 change, phytoplankton will be influenced by the frequencies and intensities of these
587 thermal excursions. We found that the responses of *E. huxleyi* to one-day versus two-
588 day temperature variations were different at both low and high temperature. For
589 instance, under low temperature the transition from the warm phase to the cool phase

590 during the thermal variation could be treated as a low temperature stress leading to a
591 lag phase in growth. The growth rate of the one-day variation treatment at the cool
592 phase was lower than that of the two-day variation, suggesting that physiological
593 acclimation is not rapid enough to accommodate to the shorter variation treatment,
594 while the two-day variation allows enough time for growth to recover. However, at
595 the warm phase (21 ^{o}C) there was no difference in growth rates between the one-day
596 and two-day variations compared with the constant 21-degree treatment. These results
597 imply that there was a shorter lag phase after transfer at the optimal temperature point
598 (21 ^{o}C at the warm phase) than during low temperature stress (16 ^{o}C at the cool phase).
599 There was no significant difference in photosynthetic rates between the one-day
600 and two-day variation during the warm phase (21 ^{o}C), but both were higher than during
601 the cool phase, indicating the photosynthetic rate was correlated to the thermal
602 variation cycle. However, for the calcification rate there was no significant difference
603 between one-day and two-day variations during either the cool or warm phases. These
604 results suggested that photosynthesis was more responsive to temperature variations
605 than calcification, and so ultimately determined the growth rate in both cool and warm
606 phases. Feng et al. (2017) reported a similar relationship between growth and
607 photosynthetic rates of a Southern Hemisphere *E. huxleyi* strain cultured at different
608 temperatures.
609 Temperature variation frequencies also strongly influenced elemental
610 composition. In low temperature experiments, the cellular contents of PON, POP and
611 POC in the one-day variation treatment were all higher than under two-day variations.

612 A notable exception to this trend was the cellular PIC content, which was not
613 significantly different between one-day and two-day variation treatments. The PIC
614 content was positively correlated to calcification and relatively stable, indicating that
615 coccolith production and storage of *E. huxleyi* was relatively independent of the
616 frequency of thermal variation.

617 Unlike the photosynthetic rate, the cellular elemental content of one-day and two-
618 day variations were significantly different, but were not changed during temperature
619 variation when transitioning from the warm phase to the cool phase or vice versa.
620 The temperature dependent photosynthetic enzyme activity likely determined the
621 similar photosynthetic rate of one-day and two-day variation treatments at both cool
622 and warm phase in our short-term experiment, but the divergent responses of cellular
623 stoichiometry in one-day and two-day thermal variations indicated different
624 mechanisms of rapid acclimation to different thermal fluctuation frequencies. Our
625 results imply that the responses of *E. huxleyi* to one-day and two-day thermal
626 variations have different patterns, but both reach stable states during extended periods
627 of temperature fluctuation. Due to decreasing POC content, the PIC/POC ratio
628 increased in the two-day variation compared with the one-day variation, suggesting
629 that more rapid thermal fluctuations might lead to a decrease in calcite ballasting of
630 sinking organic carbon.

631 Under the high temperature scenario, thermal variation forces the microalgae to
632 intermittently deal with a lethal high temperature during the warm phase (28 ^{o}C), with
633 potentially irreversible damage to the cells. In the “summer” experiments, the mean

growth rate of the two-day variation was much lower than that of the one-day variation. This mainly resulted from the negative growth rate of two-day variation cultures during the warm phase (28 ^{o}C), whereas the growth rate of the one-day variation was >0.20 d^{-1}. This result demonstrates that high frequency temperature variations (one-day) can partly mitigate growth inhibition by high temperatures in *E. huxleyi*, and so allow tolerance to extreme thermal events relative to longer exposures. This observation agrees with previous studies of other marine organisms such as corals (Oliver & Palumbi, 2011; Safaie et al., 2018). In the case of our experiments, the lag phase and metabolic inertia would help to maintain the microalgae during short exposures (one-day) to high temperature when transitioning from the cool phase (23 ^{o}C) to the warm phase (28 ^{o}C).

Likewise, the particulate organic element contents (PON, POP and POC) of *E. huxleyi* were more stable in one-day than in two-day temperature variation treatments. The relatively steady status of cellular particulate organic matter content in the high frequency temperature variation treatment may conserve energy, compared to the energy-intensive redistribution of major cellular components under lower frequency temperature variations. This differential energetic cost may help to explain the differences in growth rates between the two treatments. Adaptation to high temperature may also require higher investment in repair machinery, such as heat shock proteins, leading to an increased demand for nitrogen and other nutrients, thus increasing cellular POC, PON and POP contents (O'Donnell et al., 2018).

Prediction and modelling of *E. huxleyi* responses to thermal variation

Mathematical curves based on population growth rates from laboratory studies have been used to predict future population abundance, persistence or fitness in a changing world (Bernhardt et al., 2018; Deutsch et al., 2008; Jiang et al. 2017). We applied a modified version of the Eppley thermal performance curve model with the addition of non-linear averaging (Bernhardt et al., 2018) to predict the influence of thermal variation on the growth rate of *E. huxleyi* (Fig. 8). *E. huxleyi* growth rates were predicted to be much lower at warmer temperatures under variable conditions compared to constant conditions, but there were no significant differences at cooler temperatures. Thus, the effect of thermal variation on population growth at the upper thermal limit was predicted to be stronger than that in the lower portion of the thermal range (Bernhardt et al., 2018; Sunday et al., 2012). This phenomenon has been widely observed in ectothermic animal taxa (Dell et al., 2011), but this model for the effect of thermal variation on population growth rate may lack the ability to predict species responses at the extreme edges of their ranges (Bernhardt et al., 2018).

Our results showed that the measured effects of a variable thermal regime on *E. huxleyi* growth rate fitted well with model-predicted values at a relatively low temperature (mean=18.5 ^{o}C), but differed considerably at high temperature (mean=25.5 ^{o}C). This was especially evident under the two-day variation conditions at a mean of 25.5 ^{o}C, where the growth rate was sharply lower than predicted from the constant temperature performance curve-based model. This result suggests that transient heat waves may erode thermal tolerances of *E. huxleyi* populations already growing near their upper thermal limits, and that the frequency and duration of such extreme events

678 is critically important in determining the magnitude of this stress. Qu et al. (2019)
679 reported that the tropical cyanobacterium *Trichodesmium erythraeum* only showed a
680 slight decrease of growth rate with thermal variation treatments at high temperature
681 (average 30^{o}C), compared with constant 30^{o}C treatments. In contrast, the sensitivity of
682 this *E. huxleyi* isolate to increasing thermal variability may reduce its fitness and its
683 ability to compete with other taxa such as diatoms and cyanobacteria, with implications
684 for community structure in the future sub-tropical ocean.

685 Although thermal variation at high temperature negatively impacted the growth
686 rate of *E. huxleyi* in our experiment, our relatively short-term study did not address the
687 potential for *E. huxleyi* to evolve under selection by frequent extreme heat events.
688 Evolutionary change in the thermal optimum and the maximum growth temperature in
689 response to ocean warming may reduce heat-induced mortality, and mitigate some
690 ecological impacts of global warming (O'Donnell et al., 2018, Thomas et al., 2012). For
691 example, Schlüter et al. (2014) found that after one year of experimental adaptation to
692 warming (26.3°C), the marine coccolithophore *E. huxleyi* evolved a higher growth rate
693 when assayed at the upper thermal tolerance limit. Similar results were reported for the
694 marine diatom *Thalassiosira pseudonana* in recent studies (O'Donnell et al., 2018;
695 Schaum et al., 2018). Schaum et al. (2018) also found that the evolution of thermal
696 tolerance in marine diatoms can be particularly rapid in fluctuating environments.
697 Furthermore, populations originating from more variable environments are generally
698 more plastic (Schaum & Collins, 2014; Schaum et al., 2013). Long-term evolutionary
699 experiments with *E. huxleyi* will be necessary to determine how the thermal

performance curve of this important marine calcifier may diverge under selection by different frequencies and durations of extreme thermal variation events.

Understanding the combination of ocean warming and magnified thermal variability may be a prerequisite to accurately predicting the effects of climate change on the growth and physiology of the key marine calcifier *E. huxleyi.* This information will help to inform biogeochemical models of the marine and global carbon cycles, and ecological models of phytoplankton distributions and primary productivity. How changing thermal variation frequencies and heat wave events will affect marine phytoplankton remains a relatively under-explored topic, but one that is likely to become increasingly important in the future changing ocean.

Data availability

The data are available by request from the corresponding author (DAH), and at www.bco-dmo.org/project/668547.

Author contributions

XW, F-XF and DAH contributed to conceiving and planning the experiments. XW, F-XF, PQ, JDK, and H-BJ performed the lab experiments. XW, F-XF, Y-HG and DAH contributed to the data analysis and to writing the paper. All of the authors contributed comments, revisions and editing.

Competing interests

The authors declare that they have no conflict of interest.

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Figure legends:

Fig. 1 Thermal performance curve showing cell-specific growth rates (d^{-1}) of *Emiliania huxleyi* CCMP371 across a temperature range from 8.5 to 28.6 ℃. Symbols represent means and error bars are the standard deviations of three replicates at each temperature, but in many cases the errors bars are smaller than the symbols.

Fig. 2 Changes in *Emiliania huxleyi* TPC/PON ratios (**A**), POC/PON ratios (**B**), PIC/POC ratios (**C**) and Cha/POC ratios (**D**) across a temperature range from 8.5 to 28.6 ℃. Dashed lines represent the average ratios for the entire temperature range. Bars represent means and error bars are the standard deviations of three replicates at each temperature. Symbols * represent the significant difference ($p<0.05$) between average ratios and the ratio at each temperature.

Fig. 3 *Emiliania huxleyi* growth rate responses to constant temperatures, and during the warm and cool phases of the two thermal variation frequencies (one-day and two-day), under low (**A**) and high (**B**) mean temperatures. The thick black line in the boxplots represent median values for each experimental treatment; whiskers on boxplots indicate 1.5 × interquartile range. Listed p-values with their respective brackets are the statistical significance between two treatments.

Fig. 4 Responses of *Emiliania huxleyi* PIC/POC ratios to constant temperatures, and during the warm and cool phases of two thermal variation frequencies (one-day and two-day), under low (**A**) and high (**B**) mean temperatures. LT: Low temperature; HT: High temperature. The thick black line in the boxplots represent median values for each experimental treatment; whiskers on boxplots indicate 1.5 × interquartile range. Listed

1023 p-values with their respective brackets denote the statistical significance between two
1024 treatments.

1025 **Fig. 5** Responses of *Emiliania huxleyi* photosynthetic carbon fixation and calcification
1026 at constant temperatures and during the warm and cool phases of two thermal variation
1027 frequencies (one-day and two-day), including: total carbon fixation (photosynthesis +
1028 calcification) at low (**A**) and high (**B**) temperatures; photosynthetic carbon fixation at
1029 low (**C**) and high (**D**) temperatures; and calcification rates at low (**E**) and high (**F**)
1030 temperatures. LT: Low temperature; HT: High temperature. The thick black line in
1031 the boxplots represent median values for each experimental treatment; whiskers on
1032 boxplots indicate 1.5 × interquartile range. Listed p-values with their respective
1033 brackets denote the statistical significance between two treatments.

1034 **Fig. 6** Responses of *Emiliania huxleyi* calcification to photosynthesis ratios (cal/photo)
1035 to constant temperatures, and during the warm and cool phases of two thermal variation
1036 frequencies (1 day and 2 day), under low (**A**) and high (**B**) mean temperatures. LT: Low
1037 temperature; HT: High temperature. The thick black line in the boxplots represent
1038 median values for each experimental treatment; whiskers on boxplots indicate 1.5 ×
1039 interquartile range. Listed p-values with their respective brackets denote the statistical
1040 significance between two treatments.

1041 **Fig. 7** Responses of *Emiliania huxleyi* elemental ratios in two thermal variation
1042 frequency treatments (1 day and 2 day) compared to constant temperatures, for:
1043 TPC/PON (**A**, cool phase and **B**, warm phase), PON/POP (**C**, cool phase and **D**, warm
1044 phase) and Chl *a*/POC ratios (**E**, cool phase and **F**, warm phase). LT: Low temperature;

1045 HT: High temperature. The thick black line in the boxplots represent median values for
1046 each experimental treatment; whiskers on boxplots indicate 1.5 × interquartile range.
1047 Listed p-values with their respective brackets denote the statistical significance between
1048 two treatments.

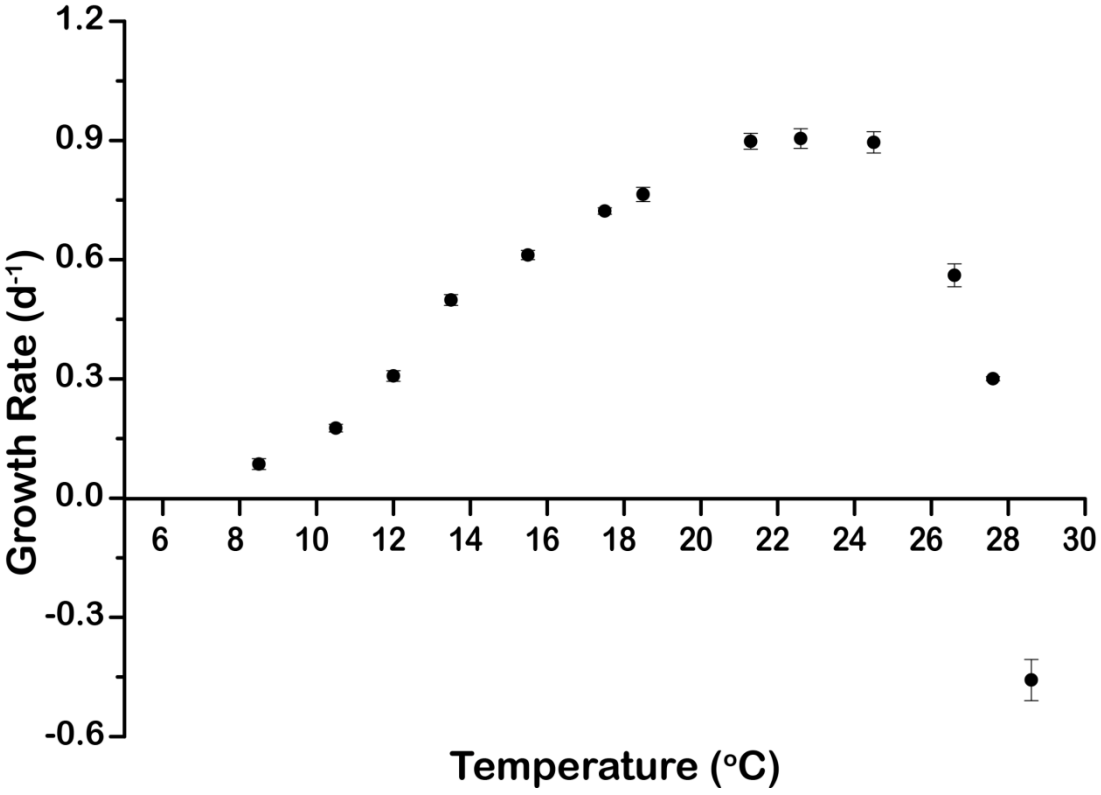
1049 **Fig. 8** Thermal performance curves based on specific growth rates (d^{-1}) of *Emiliania*
1050 *huxleyi*, including our experimentally determined constant condition temperature
1051 performance curve (black symbols and solid line) and a predicted fluctuating condition
1052 temperature performance curve (dashed line) according to the model of Bernhardt et al.
1053 (2018). Measured growth rates from the two low and high temperature experiments
1054 are shown for constant thermal conditions (red symbols), one-day (green symbols) and
1055 two-day (blue symbols) variation treatments.

1056

Table 1 The effect of temperature variation under low and high temperature on total carbon (pg/cell), cellular POC (pg/cell), cellular PIC (pg/cell), cellular PON (pg/cell), cellular POP (pg/cell) and cellular Chl *a* (pg/cell) of *Emiliania huxleyi.*

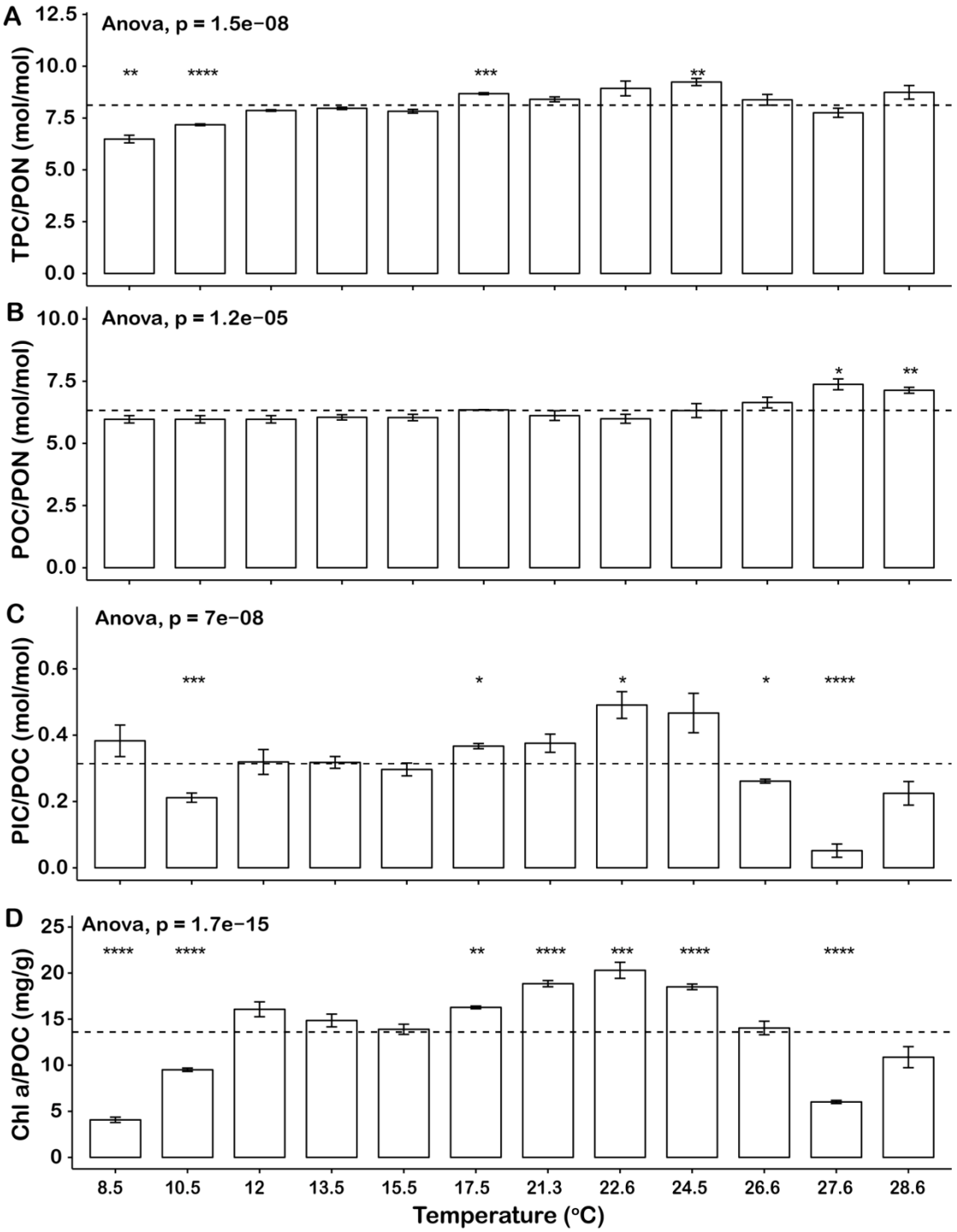
Treatment		Total Carbon	Cellular PON	Cellular POP	Cellular POC	Cellular PIC	Cellar Chl *a*
Low temperature	18.5 °C	11.5±0.4	1.8±0.2	0.17±0.00	8.0±0.6	3.5±0.3	0.14±0.00
	One-day cool point (16)	13.0±0.5	2.2±0.3	0.18±0.00	8.9±0.3	4.1±0.3	0.15±0.01
	One-day warm point (21)	12.0±0.7	2.1±0.3	0.19±0.00	9.3±0.9	2.7±0.9	0.19±0.00
	Two-day cool point (16)	10.1±0.7	1.3±0.2	0.16±0.01	6.0±0.9	4.0±0.3	0.12±0.01
	Two-day warm point (21)	10.4±0.5	1.5±0.2	0.17±0.01	6.6±0.5	3.8±0.3	0.15±0.01
High temperature	25.5 °C	15.0±0.7	2.0±0.1	0.21±0.01	9.5±0.3	5.5±0.7	0.18±0.02
	One-day cool point (23)	16.1±1.4	3.0±0.2	0.21±0.00	12.9±1.5	3.2±0.2	0.15±0.01
	One-day warm point (28)	19.1±0.8	4.4±0.3	0.24±0.01	17.0±0.6	2.1±0.2	0.20±0.02
	Two-day cool point (23)	12.4±1.0	1.9±0.2	0.19±0.01	7.5±1.0	4.8±0.3	0.13±0.01
	Two-day warm point (28)	19.4±2.0	3.9±0.8	0.25±0.03	18.3±3.7	2.1±0.9	0.25±0.02

1061 **Fig. 1**



1062

1063 **Fig. 2**



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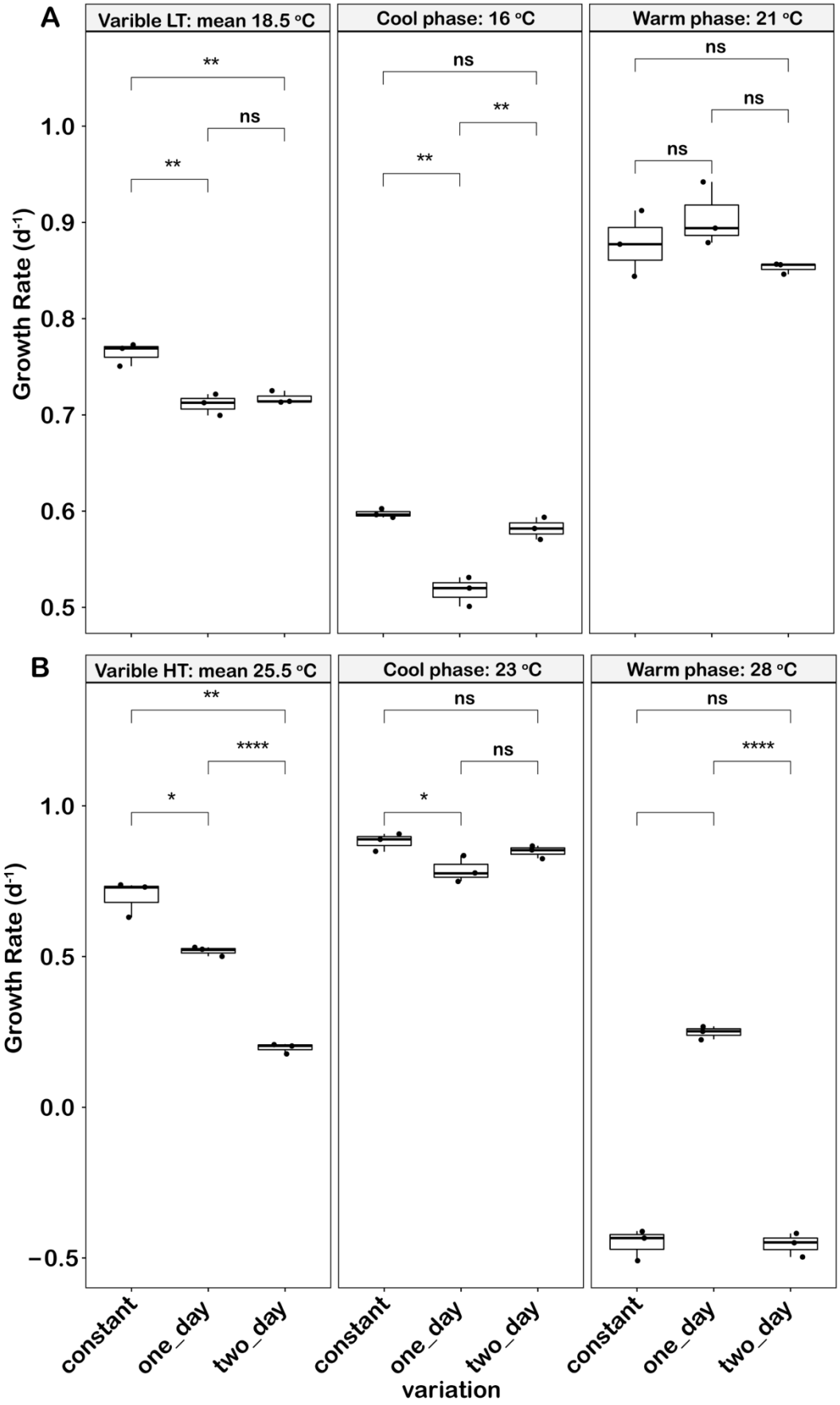
1066 **Fig. 3**



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1068 **Fig. 4**

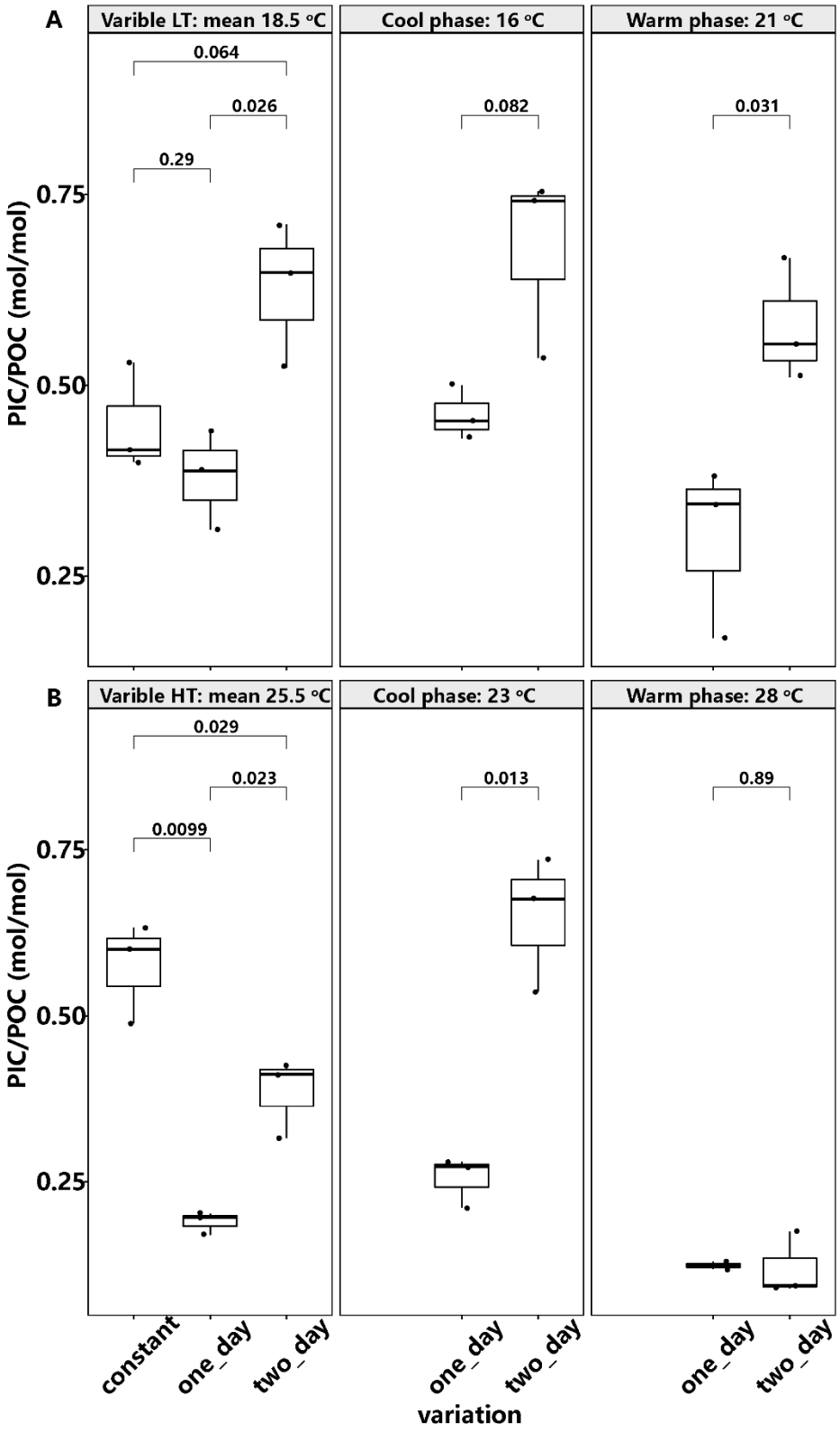

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Fig. 5

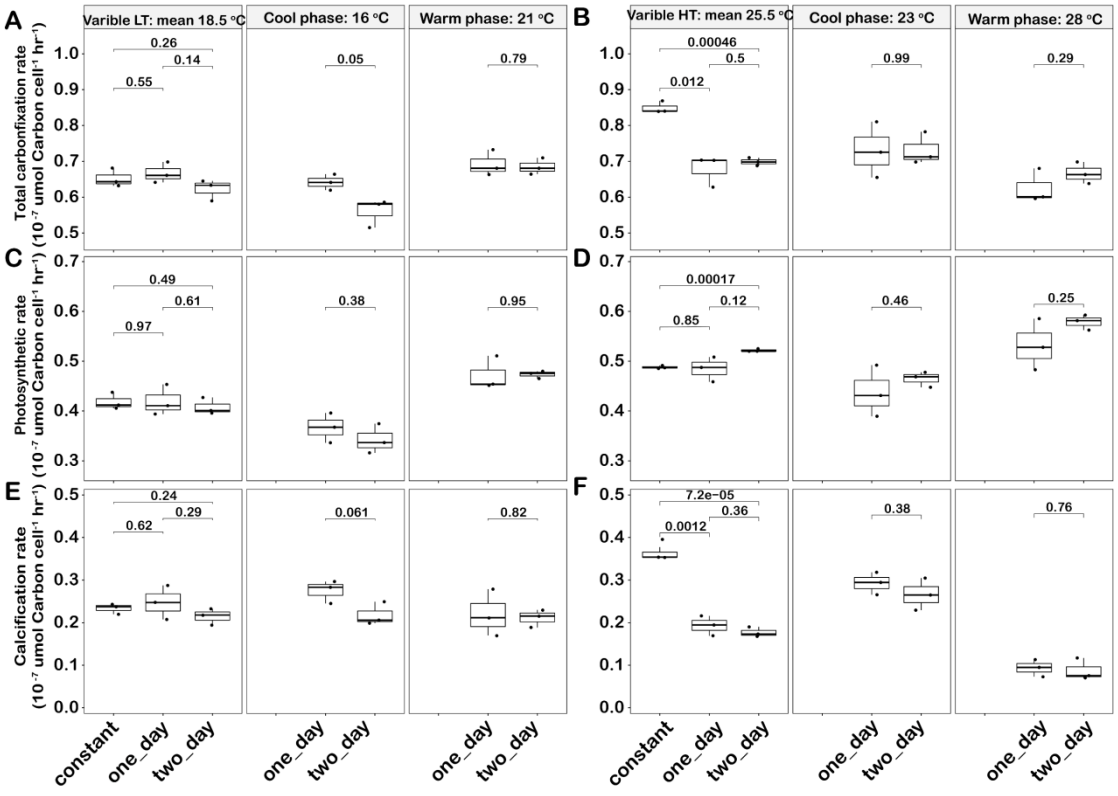

A
Varible LT: mean 18.5 °C
Cool phase: 16 °C
Warm phase: 21 °C
B
Varible HT: mean 25.5 °C
Cool phase: 23 °C
Warm phase: 28 °C
Total carbonfixation rate (10⁻⁷ umol Carbon cell⁻¹ hr⁻¹)
C
D
Photosynthetic rate (10⁻⁷ umol Carbon cell⁻¹ hr⁻¹)
E
F
Calcification rate (10⁻⁷ umol Carbon cell⁻¹ hr⁻¹)
0.26
0.14
0.55
0.05
0.79
0.00046
0.5
0.012
0.99
0.29
0.49
0.61
0.97
0.38
0.95
0.00017
0.12
0.85
0.46
0.25
0.24
0.29
0.62
0.061
0.82
7.2e−05
0.36
0.0012
0.38
0.76
constant
one_day
two_day

Fig. 6

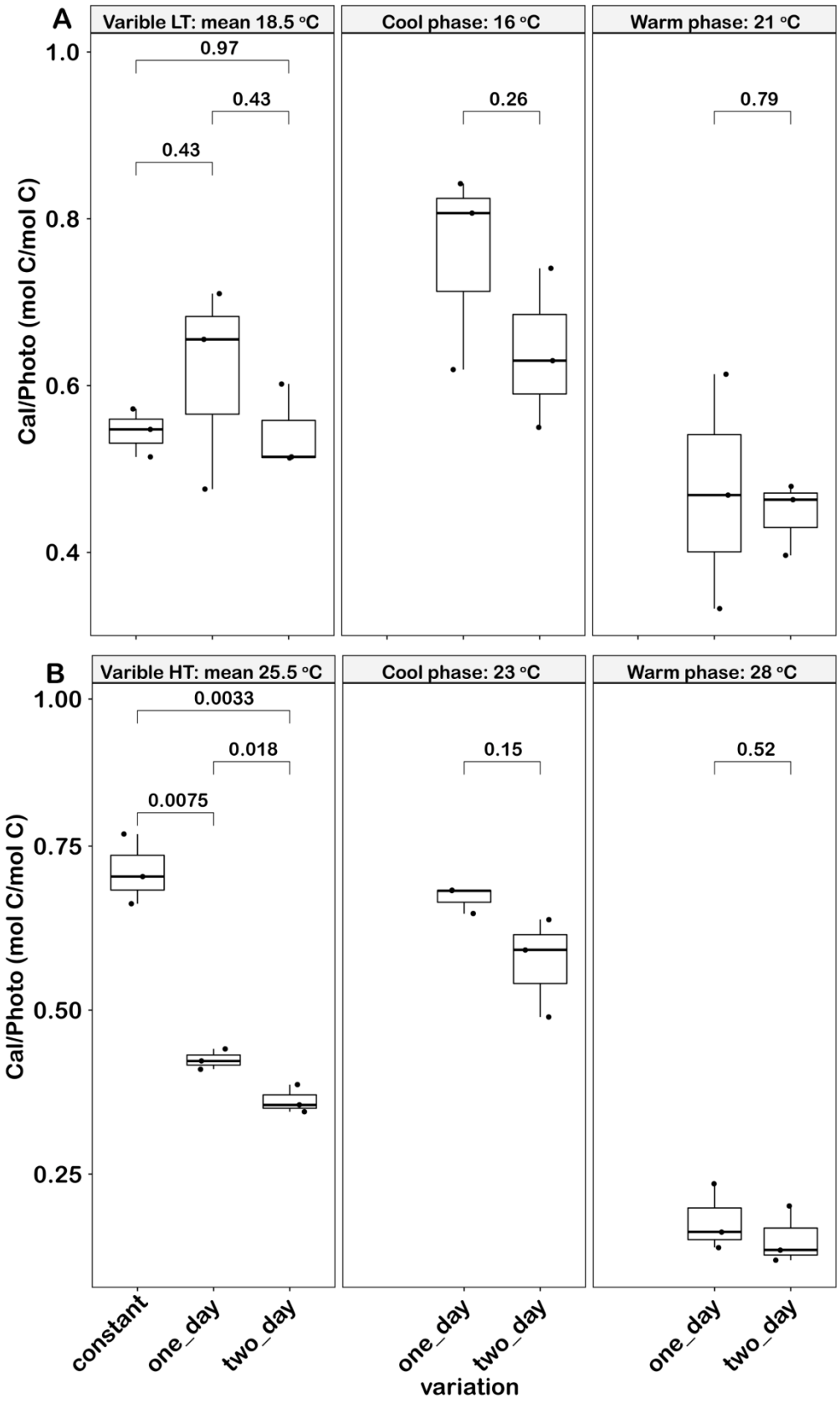
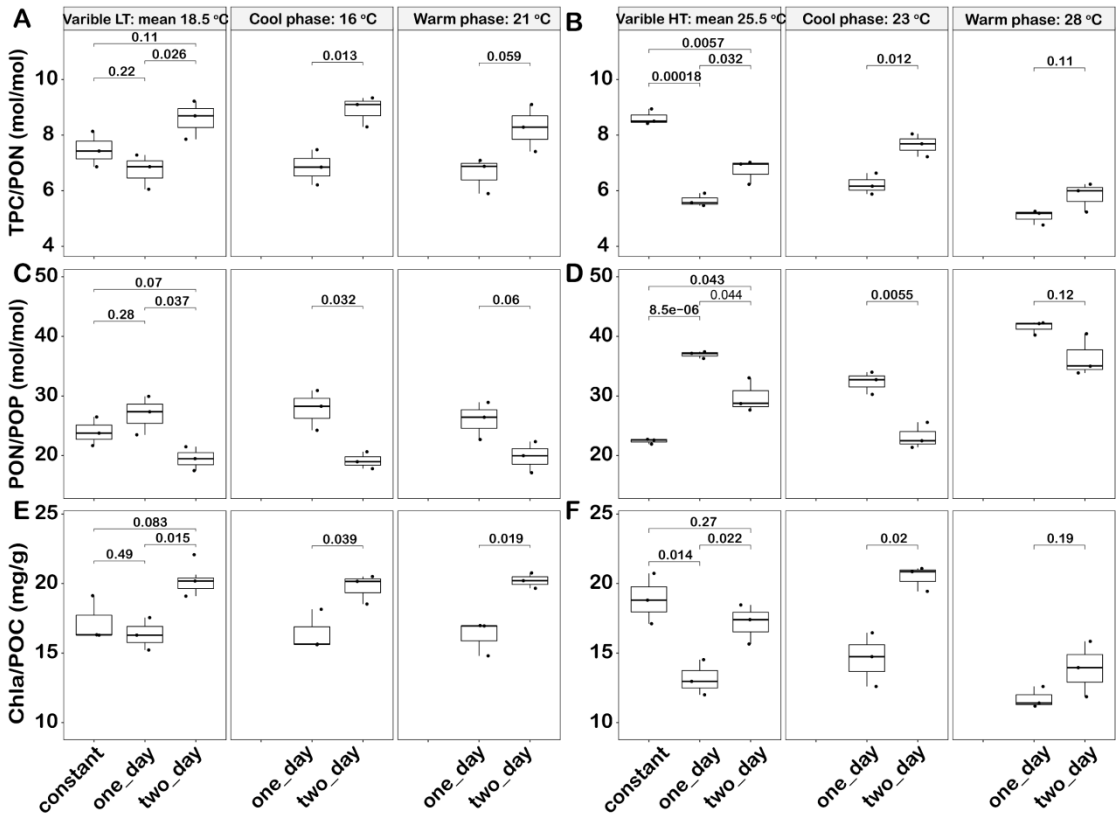
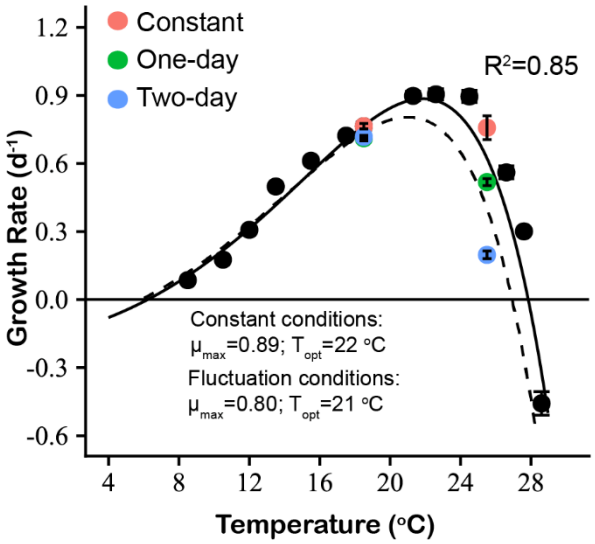

A
Varible LT: mean 18.5 °C
Cool phase: 16 °C
Warm phase: 21 °C
0.97
0.43
0.43
0.26
0.79
Cal/Photo (mol C/mol C)
1.0
0.8
0.6
0.4
B
Varible HT: mean 25.5 °C
Cool phase: 23 °C
Warm phase: 28 °C
0.0033
0.018
0.0075
0.15
0.52
Cal/Photo (mol C/mol C)
1.00
0.75
0.50
0.25
constant
one_day
two_day
one_day
two_day
one_day
two_day
variation

Fig. 7


A
Varible LT: mean 18.5 °C
Cool phase: 16 °C
Warm phase: 21 °C
B
Varible HT: mean 25.5 °C
Cool phase: 23 °C
Warm phase: 28 °C
TPC/PON (mol/mol)
PON/POP (mol/mol)
Chla/POC (mg/g)
C
D
E
F
0.11
0.22
0.026
0.013
0.059
0.0057
0.00018
0.032
0.012
0.11
0.07
0.28
0.037
0.032
0.06
0.043
8.5e−06
0.044
0.0055
0.12
0.083
0.49
0.015
0.039
0.019
0.27
0.014
0.022
0.02
0.19
constant
one_day
two_day

1081 **Fig. 8**



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