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Bremen, August 7th 2019

Dear Editor(s),

Please find attached the revised manuscript initially entitled "**Calcification and distribution of extant coccolithophores across the Drake Passage during late austral summer 2016**" by Mariem Saavedra-Pellitero* (corresponding author), Karl-Heinz Baumann, Miguel Angel Fuertes, Hartmut Schulz, Yann Marcon, Nele Manon Vollmar, Jose Abel Flores and Frank Lamy.

We addressed all the comments and suggestions by the two referees and other members of the scientific community and we thank you for the 3 weeks extension you gave us.

If possible, we would like to take the chance to change the title to "**Calcification and latitudinal distribution of extant coccolithophores across the Drake Passage during late austral summer 2016**", because we think it describes better this piece of research.

All the best,



Mariem Saavedra-Pellitero

Comments:

We thank all the reviewers for their comments and suggestions which definitely helped us to improve this manuscript.

The reply to the reviewers is structured as follows:

Comments from the reviewers in black

Our reply to reviewers in red.

Note that line numbers refer to the 1st submitted version, not the revised one.

Francisco Díaz-Rosas

Dear Saavedra-Pellitero and co-authors, I have read your paper in BGS discussion, and I have a short comment to make: the description of the central area of the heavy calcified morphotype-R as "Grill" is not aligned with recently observation in the Eastern South Pacific, in which, it's reported as completely or nearly completely covered (see Fig. 3b in Beaufort et al., 2011 and Fig. 2d in von Dassow et al., 2018).

We thank Francisco Rias-Rosas for pointing out this. We absolutely agree with him, and we accordingly modified Table 1. The morphology of the central area is now described as "covered or nearly covered" and we also added the reference "von Dassow et al. (2018)" in the last column (comparable morphotypes in the literature)

Anonymous Referee #1

Summary:

Saavedra-Pellitero and co-authors present coccolithophore measurements from along the western side of the Drake Passage. They quantify coccolithophore species counts and coccolith mass. They present some oceanographic measurements but also use global databases to retrieve nutrients and carbonate chemistry quantities. They find a poleward decrease in both coccolithophore diversity and calcification. They group *E. huxleyi* morphotypes into two major groups (A and B) and are able to show that the southward decrease in calcification is related to a shift from A morphotypes along the Chilean margin to B morphotypes in the subantarctic and polar front zones. Coccolithophore calcification is inversely related to alkalinity, dissolved inorganic carbon, and pH. A Principle component analysis reveals three distinct clusters: Chilean coastal, SAZ, and PFZ. Temperature seems to be an important factor in controlling the distribution of coccolithophore species, as well as overall coccolithophore abundance and calcification.

General comments:

Referee #1 (R#1): This manuscript is well written and of high quality. The authors present a valuable dataset with respect to observed coccolithophores. They present fantastic detailed plots of coccolithophore species in this Southern Ocean transect. This transect is slightly westward of

the transects presented Charalampopoulou et al. (2016), which is a similar study. This manuscript offers more information on depth variations in coccolithophore abundances than previous studies in this region, which is great! This study reaches much of the same overall conclusions as previous transects observing coccolithophores in the Southern Ocean, so it is not groundbreaking, but adds to a solid overall conclusion of coccolithophores transitioning from more calcified species/morphotypes in the subtropics to less calcified ones in the ACC region. The conclusion that temperature is a controlling factor on coccolithophore abundance agrees with previous studies (e.g., Charalampopoulou et al., 2016). I think this manuscript is in great shape and only needs minor revisions.

Mariem Saavedra-Pellitero et al. (MSP): We thank reviewer #1 for his/her insightful comments and we agree with most of them and made changes accordingly.

R#1: One piece that is missing is a bit more specific speculation about how coccolithophore abundance/calcification could change with climate change. The authors say that coccolithophores will be strongly influenced, but not how they will be influenced. I think it's important to hypothesize the direction of change, given current observations and relationships with environmental variables presented in the study. I also think that the positive relationship between temperature and coccolith mass needs to be emphasized a bit more.

MSP: This point has been made also by reviewer #2.

We included more information regarding the missing information at the end of section 4.3.

R#1: It is a bit of a shame that nutrients and carbonate chemistry parameters were not measured in situ, but I do not think that having these measurements would have changed the conclusions (it would have just added more strength to them).

MSP: We absolutely agree with reviewer#1 and we are aware that it is a limitation we have face in this study.

R#1: I also think that the depth variations between the three different oceanic region clusters could be more emphasized (especially because this was not as well presented in previous studies, so I find it to be new information): maximum depth of coccolithophores decreases poleward.

MSP: We added more details regarding depth variations for each of the clusters/zones in section 4.2.

Additionally, we mentioned it now in the abstract, conclusions and also briefly in section 4.1.

Specific comments:

R#1: Page 1, Abstract: maybe add in something about the decreasing depth of coccolithophores as you go poleward (as shown in Figure 3a)

MSP: We wrote: "We find that coccolithophore abundance, diversity and maximum depth habitat decrease southwards marking different oceanographic fronts as ecological boundaries" in the abstract.

R#1: Page 2, Line 6: extra "substantial".

MSP: The extra "substantial" was deleted.

R#1: Page 2, Lines 11-13: This sentence is awkward and a bit hard to understand. Maybe it would be best rewritten like this: “Coccolithophores produce up to 40% of open ocean calcium carbonate (Poulton et al., 2013) and are responsible for 20% of global net marine primary production (Malone et al., 2017). Therefore, how coccolithophores respond to changing oceanic conditions is of utmost importance for marine ecology and carbon cycling.”

MSP: We rewrote this sentence (literally) using the suggestion from reviewer #1.

R#1: Page 2, Line 32: I think that it’s important to include that the Beaufort et al (2011) study includes both modern samples and paleodata from the last 40000 years. Maybe just add “over long timescales”: “A known positive correlation exists over long timescales between surface-ocean: :.”

MSP: We agree and therefore we added “over long timescales” to the text.

R#1: Page 3, Line 6: replace “actually” with “recently”

MSP: We replaced “actually” with “recently”.

R#1: Page 3, Line 15: Perhaps replace “species levels” with “overall coccolithophore calcification” since Beaufort et al. (2011) and Freeman and Lovenduski (2015) both have drawn conclusions based on overall coccolithophore calcification. While the Beaufort study has some species level information, the Freeman and Lovenduski study does not.

MSP: Following the suggestion of reviewer#1 we made this sentence simpler, and wrote: “Even with a temperature-driven range expansion of coccolithophores in the SO, surface ocean carbonate chemistry is now capable of exerting a first-order control on the composition of coccolithophore assemblages as well as on overall coccolithophore calcification (Cubillos et al., 2007; Mohan et al., 2008; Beaufort et al., 2011; Freeman and Lovenduski, 2015)”.

R#1: Page 3, Line 23: Break this sentence up into two sentences for clarity: “Accordingly, we calculated extant coccolithophore species numbers at different stations between 10 and 150 m of the water column and evaluated the coccolith mass variations of *E. huxleyi*. We compared these observations with in situ conductivity–temperature– depth (CTD) measurements, carbonate chemistry parameters, as well as to previously published Southern Ocean coccolithophore and calcification datasets.”

MSP: We split the sentence into two shorter ones.

R#1: Page 3, Line 28: no need to capitalize “stations”

MSP: We changed it to “stations”.

R#1: Page 6, Line 7: instead of “a taxon” say “one taxon”

MSP: We use “one” instead of “a”.

R#1: Page 7: Line 22: Add references to Figures 4 and 5: “: : , grouped into A (Figure 4) and B (Figure 5) according to Young et al. (2019). Also, by “Young et al., 2019” do you mean Nanotax3 website? It is unclear what reference this is referring to in the bibliography.

MSP: We added the references to Figures 4 and 5 as suggested.

Yes, as pointed out by reviewer#1, we unclearly referenced Nanotax3 website in the previous version. To ensure that we cite it correctly this time, we double checked in <http://www.mikrotax.org/Nannotax3/pages/ntax-citation.html> and referenced accordingly.

R#1: Page 7, Line 30: Type A overcalcified and Type R seem very similar to me. How are they different exactly?

MSP: They are indeed similar and showed similar distribution, but they still show slightly different morphologies, as shown in Plate 1.

In *E. huxleyi* type R the slits between distal shield elements are almost or totally closed, and the tube is usually thick (Plate 1 a) giving a *Reticulofenestra*-like appearance, while type A overcalcified shows just a closed or nearly closed central area (Plate 1 b, c), but not almost closed slits.

Since this information can be found in the original sources mentioned in the section 2.1, we simply added the references to the specific pictures of the different morphotypes displayed Plate 1, e.g., “Type R (Plate 1 a)...”

R#1: Page 8, Line 23: When you say that *Syracosphaera* dominates in the SAZ, do you mean that it dominates among the rare coccolithophore assemblage or among coccolithophores overall? Please modify to be more specific.

MSP: We specify now that it dominates among the rare coccolithophore assemblage.

R#1: Page 9, Line 7: Take out the extraneous “the” before 77.4%.

MSP: “The” was deleted.

R#1: Page 10, Lines 25-29: Could silica be becoming more limiting north of the PF, opening a niche for coccolithophores? Perhaps competition among phytoplankton is another possibility of coccolithophores increasing in abundance at the PF.

MSP: This is a likely possibility, as far as we know from other papers in the Atlantic sector of the Southern Ocean (e.g., Smith et al., 2017). However, we do not have in situ silica measurements or diatoms counts, so we decided to avoid speculation and did not to include this suggestion in the new version of the manuscript.

Future research could usefully address the interesting shift in dominance from coccolithophores to diatoms across the PF (also mentioned by reviewer #2) and assess the interrelationship between both groups, even at a sub-species level.

R#1: Page 11, Line 10: rather than saying “up to 61.7_S”, maybe it’s more appropriate to say “down to 61.7_S”

MSP: We changed it to “down to”.

R#1: Page 11, Line 21: misspelling of the word “coastal”; and instead of saying “: : : increasing towards oceanic regions” maybe say “: : : increasing towards open ocean regions”

MSP: We made both changes

Page 12, Line 12: change “communities” to “community”

MSP: We could not find the word “communities” in page 12 line 12, so we assumed that reviewer#1 was referring to page 12 line 3. We changed “communities” to “community” in that sentence.

R#1: Page 12, Line 19: The widely used *E. huxleyi* strain NZEH (morphotype R) and *E. huxleyi* strain RCC1216 (morphotype R) were both isolated from around New Zealand so I believe that would count as “observing” it there too.

For example, see Methods in Iglesias-Rodriguez et al. (2017) and Langer et al., (2009): Iglesias-Rodriguez, Maria Debora, Bethan M. Jones, Sonia Blanco-Ameijeiras, Mervyn Greaves, Maria Huete-Ortega, and Mario Lebrato. "Physiological responses of coccolithophores to abrupt exposure of naturally low pH deep seawater." *PloS one* 12, no. 7 (2017): e0181713.

Langer, Gerald, Gernot Nehrke, Ian Probert, J. Ly, and Patrizia Ziveri. "Strain-specific responses of *Emiliania huxleyi* to changing seawater carbonate chemistry." *Biogeosciences* 6, no. 11 (2009): 2637-2646.

MSP: We thank reviewer#1 for those interesting papers. They are cited now in the new version.

R#1: Page 12, Line 30: change “up to ca. 6_C” to “down to ca. 6_C”

MSP: We changed it to “down to ca. 6_C”.

R#1: Page 13, Line 20: change “decreases” to “decrease”

MSP: We changed it to “decrease”.

R#1: Page 14, section 4.3 in general: I think that the temperature as a controlling factor needs to be discussed more. It’s in the abstract (Page 1, Line 28/29) as a greater limiting factor than carbonate chemistry, which I totally agree with, but I think it needs more discussion in the paper.

MSP: We included more information regarding this point at the end of section 4.3.

R#1: Are colder temperatures in the poleward direction selecting for lightly calcified species/morphotypes? Or could it be a physiological change induced by colder temperatures?

MSP: These are in fact very interesting questions that we will keep in mind for future work.

However, we believe that we would need more data in order to properly answer them.

In the new version of the manuscript, we highlighted the relevant role of temperature, and mentioned the degree of adaptive potential of coccolithophores (last sentence in section 4.3), but we did not want to speculate more.

R#1: I think a bit more speculation (perhaps bringing in some laboratory experiments) would be nice in this section.

MSP: We added more speculation (and more references citing culture experiments) in section 4.3.

R#1: There’s a summary of the effects of temperature on coccolithophore calcification in Krumhardt et al. (2017): Krumhardt, Kristen M., Nicole S. Lovenduski, M. Debora Iglesias-Rodriguez, and Joan A. Kleypas. "Coccolithophore growth and calcification in a changing ocean." *Progress in oceanography* 159 (2017): 276-295.

MSP: We thank reviewer#1 for suggesting this paper. We cited it in the new version in section 4.3.

R#1: Page 14, Line 20: misspelled the word “mass”

MSP: We corrected it.

R#1: Page 14, Lines 20 – 25: It’s good that you pointed out the fact that the carbonate chemistry parameters have been estimated, rather than measured. However, the latitudinal gradients in carbonate chemistry parameters are pretty well established and I don’t think it would affect the relationships you’re seeing.

MSP: We thank reviewer#1 for this positive comment.

R#1: Page 15, Line 3: misspelled the word “Striking”

MSP: We corrected it.

R#1: Page 15, Line 10. Here is where it would be good to speculate on the direction of change in coccolithophore abundance/calcification (or latitudinal species/morphotype shifts) with ongoing climate change. You could bring up the positive correlation with temperature shown in Table 4 and the PC analysis.

MSP: We speculated about possible future scenarios in section 4.3.

R#1: Figure 13: I like that you included this comparison to the Charalampopoulou et al. (2016) paper. Is the direction arrow on the right hand side of the figure supposed to say “East” rather than “West”? I thought the Charalampopoulou transects were to the east of the present study: :

MSP: We thank reviewer#1 for spotting this mistake. We modified Figure 13 and wrote “East”, because those transects are east of our study.

R#1: Table 1 and Plate 1: I like that you grouped the E hux morphotypes into 2 main groups. There seems to be a fluidity between all these morphotypes and grouping into only 2 groups makes the information much more digestible.

MSP: We are glad that reviewer#1 appreciate our approach.

R#1: Table 4: misspelling on the line HCO₃- CO₂Sys

MSP: We corrected it.

Anonymous Referee #2

Referee #2 (R#2): The paper presents well documented distributional data of coccolithophores and in particular E. huxleyi morphotypes across the oceanographic fronts in the area of the Drake Passage, an important zone for monitoring the path of the Antarctic Circumpolar Current. The presentation of the methods and data is clear and the discussion is well supported, showing a consistent latitudinal trend of decreasing coccolith mass along with temperature decrease and a gradient in carbonate chemistry parameters. Overall the manuscript represent a substantial

contribution in the field of coccolithophore studies, adding new information and providing accurate measurements of both Ehux types and coccolith mass. The data are well presented, with figures and plates are of excellent quality.

Mariem Saavedra-Pellitero et al. (MSP): We thank reviewer#2 for his/her positive feedback and insightful comments. We made modifications in the text based on his/her suggestions.

R#2: Some points deserve further discussion, in particular: section 4.3 is well organized and points to significant changes in the coccolith mass of *E. huxleyi* across the different fronts and zones of the ACC.

However, while the degree of calcification is considered as the main driver of coccolith mass variation (but the assessment of degree of calcification and mass is done with different techniques, so there can be no direct attribution), there is no discussion about the influence of coccolith size on coccolith mass, e.g. type C is smaller than B/C which is smaller than type B, by definition.

MSP: This was one of the main challenges of this study: to choose the best way to characterize the coccolithophore assemblage while minimizing the error in the coccolith mass estimates. For this reason we choose to combine SEM analyses and C-calcita LM measurements.

Using just LM techniques would not have allowed us to distinguish the different *E. huxleyi* morphotypes (i.e., O type from B/C), and using only SEM techniques, would have implied to calculate the coccolith mass with the equations of Young and Ziveri (2000), being forced to assume some of the values (e.g., the shape-dependent constant K_s). That is the reason behind using different techniques for this research.

Regarding the influence of coccolith size on coccolith mass, we are aware that (by definition) there is size variation (Table 1) and certain overlap in size among different *E. huxleyi* morphotypes, which makes a direct comparison complicated. That is why we provided (as a first approach) Figure 11, but we decided to stick just to the mass variations in the manuscript. Biometric work would be required for the discussion that reviewer#2 is asking here, but we think that it is a topic for future research.

R#2: Carbonate chemistry parameters.

The discussion of the relation between coccolithophore calcification and the carbonate chemistry of the water column should be considered even more carefully, given the fact that the data are not measured in the same samples and the pattern of pH variation is not so clear – a different trend appears if the different calculations are considered, e.g. fig. 12.

MSP: Yes, we agree with reviewer#2, but as pointed out by reviewer#1, we made it very clear throughout the manuscript that those are not in situ measurements. We believe that it has been well specified, and it is depicted even in the conclusions.

R#2: However, the correlation with omega calcite seems meaningful, looking at the graphs, but this parameter is not considered in the discussion.

MSP: We added more discussion in the section 4.3 regarding the weak correlation between calcite saturation and coccolith mass.

R#2: The last sentence of the conclusion is however not supported and does not explain how climate change will affect the calcification mode of coccolithophores, given that no clear

relationship between the degree of calcification and the carbonate chemistry of sea water are established yet, but rather different correlations seem to exist in different areas of the world oceans and under different oceanographic conditions, so the question remains open.

MSP: This point has somehow been made also by reviewer #1 and it has already been addressed, highlighting the temperature as a controlling factor.

R#2: Page 11 line 12: also Malinverno et al., 2016 show the shift in dominance from coccolithophores to diatoms in water samples across the PF / sACCF, so this could be cited.

MSP: We thank reviewer#2 for suggesting this paper. We included this reference and reworded the sentence as follows: “In any case the southernmost extent of coccolithophores is also influenced by the clear dominance of diatoms south of the PF, as suggested by the high diatom concentration (valves/g dry sediment) and biogenic opal content recorded in surface sediment samples from the AZ of the Drake Passage (Cárdenas et al., 2018) and from Pacific Southern Ocean extant plankton studies (e.g., Saavedra-Pellitero et al., 2014; Malinverno et al., 2016).”

R#2: Typos: Page 1, line 22: classified -> identified

MSP: We changed it to “identified”.

R#2: Page 2, line 6: delete “substantial” which is repeated twice

MSP: The extra “substantial” was deleted, as suggested also by reviewer#1.

R#2: Page 2, line 7: dissolved carbon » dissolved inorganic carbon

MSP: We changed it to “dissolved inorganic carbon”.

R#2: Page2, line 16: phosphate is mis-spelled

MSP: We corrected it.

R#2: Page 2, line 28: the future -> in the future

MSP: We changed it.

R#2: Page 4, line 6: (2004) is repeated

MSP: We deleted one of the “(2004)”.

R#2: Page 7 line 20: this taxa -> this taxon

MSP: We changed it to “taxon”.

R#2: Page 8 line 1: later -> latter

MSP: We corrected it.

R#2: Page 13 line 5: established -> established by

MSP: We added “by”.

Additional changes:

Title: Calcification and **latitudinal** distribution of extant coccolithophores across the Drake Passage during late austral summer 2016

We would like to add the word “latitudinal” to the title, because it makes it more precise.

Page 1, L8: includes...a unicellular

Page 1, L14: To avoid repetition we changed “expected” to “predicted”

Page 1, L27: succession

Page 3, L9: *E. huxleyi* (because the whole name was previously mentioned)

Page 3, L31: a instead of an

Page 4, L14: missing space

Page 4, L33: we deleted the dash in “derived- variables”

Page 5, L2: The comma should not superscript

Page 6, L5: We added “H=” to equation (1)

Page 6, L27: sp.

Page 8, L20: we deleted an extra space

Page 8, L26: *Papposphaera* sp.

Page 9, L18: environment

Page 9, L22: we changed “maximum” to “maximal”

Page 10, L3: pg instead of picrograms

Page 10, L10: portray

Page 10, L26: occurring....Antarctic Circumpolar Current

Page 11, L31: mesoscale

Page 12: We changed the title of section 4.2, because it seemed random/preliminary title.

Page 12, L 22: has already been

Page 13, L7: might instead of may

Page 13, L13: spp.

Page 13, L31: sp.,

Page 13, L31: lightly calcified instead of lightly-calcified

Page 14, L10: observed

Page 14, L10: estimated masses instead of estimated mass values

Page 14, L27: succession

Page 15, L21: sp.

Page 16: We changed the title to just “Conclusions”

Page 16: We added the link for the Pangaea data repository:

<https://doi.org/10.1594/PANGAEA.901294>.

References: We added the missing reference for Schlitzer (2015)

Fig. 13 caption: we deleted some extra commas

Additionally we added/deleted some missing spaces/commas throughout the manuscript.

In the conclusions we capitalized some of the terms (e.g., Polar Front), just for consistency.

Calcification and latitudinal distribution of extant coccolithophores across the Drake Passage during late austral summer 2016

Mariem Saavedra-Pellitero^{1,2}, Karl-Heinz Baumann^{1,3}, Miguel Ángel ~~Fuertes~~³Fuertes⁴, Hartmut ~~Schulz~~⁴Schulz⁵, Yann Marcon^{1,5,6}, Nele Manon Vollmar¹, José-Abel ~~Flores~~³Flores⁴, Frank Lamy⁶

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Abstract:

Coccolithophores are globally distributed microscopic marine algae that exert a major influence on the global carbon cycle through calcification and primary productivity. There is recent interest in coccolithophore polar communities, however field observations regarding their biogeographic distribution are scarce for the Southern Ocean. This study documents the latitudinal, as well as in depth, variability in the coccolithophore assemblage composition and the coccolith mass variation of the ecologically dominant *Emiliana huxleyi* across the Drake Passage. Ninety-six water samples were taken between 10 and 150 m water depth from 18 stations during POLARSTERN Expedition PS97 (February-April, 2016). A minimum of 200 coccospheres per sample were ~~classified-identified~~ in scanning electron microscope and coccolith mass was estimated with light microscopy. We find that coccolithophore abundance, ~~-and~~ diversity and maximum depth habitat decrease southwards marking different oceanographic fronts as ecological boundaries. We characterize three zones: (1) the Chilean margin, where *E. huxleyi* type A (normal and overcalcified) and type R are present; (2) the Subantarctic Zone (SAZ), where *E. huxleyi* reaches maximum values of $212.5 \cdot 10^3$ cells/L and types B/C, C, O are dominant. (3) The Polar Front Zone (PFZ), where *E. huxleyi* types B/C and C dominate. We link the decreasing trend in *E. huxleyi* coccolith mass to the poleward latitudinal succession from type A to type B group. Remarkably, we find that coccolith mass is strongly anticorrelated to total alkalinity, total CO₂, bicarbonate ion and pH. We speculate that low temperatures are a greater limiting factor than carbonate chemistry in the Southern Ocean. However, further in situ oceanographic data is needed to verify the proposed relationships. We hypothesize that assemblage composition and calcification modes of *E. huxleyi* in the Drake Passage will be strongly influenced by the ongoing climate change.

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1. Introduction

5 The carbon chemistry of the ocean has a fundamental impact on marine life. The current influx of anthropogenic CO₂ into the surface ocean is causing a substantial ~~substantial~~-perturbation to marine chemistry, as exhibited by variations in alkalinity, carbonate ion, saturation state or pH (e.g., Gattuso et al., 2011). Many organisms use dissolved inorganic carbon for photosynthesis and/or the production of calcium carbonate biominerals. Open ocean phytoplankton includes coccolithophore algae, ~~an~~ unicellular organism belonging to the phylum Haptophyta (Young and Bown, 1997; Young et al., 2003). Within a single coccolithophore cell there are dual pathways of carbon utilization – for photosynthesis and for biomineralization of calcium carbonate platelets, called coccoliths. ~~As primary producers of Coccolithophores produce~~ up to ~40% of open ocean calcium carbonate (Poulton et al., 2013) and ~~are responsible for~~ ~20% of global net marine primary production (~~Malone et al., 2017~~), (~~Malone et al., 2017~~). ~~Therefore, how~~ coccolithophore responds to changing oceanic ~~chemistry conditions~~ is ~~therefore key of utmost importance~~ for marine ecology and carbon cycling. The ratio between particulate organic carbon formed during photosynthesis and particulate inorganic carbon produced via calcification varies depending on species or even morphotypes within species (Blanco-Ameijeiras et al., 2016), but can also be highly influenced by environmental conditions, such as seawater CO₂ concentration, total alkalinity, and phosphate concentration (Findlay et al., 2011). As climate varies, it is expected that these key conditions will change, and it is ~~expected predicted~~ that upper oceans may experience increased stratification and decreased nutrient availability in the upper photic zone (Cabré et al., 2015). How exactly coccolithophores will respond to these changes is subject to debate.

20 Ocean acidification combined with the increase in sea-surface temperature due to global warming –are major concerns in polar and subpolar regions (e.g., Wassmann et al., 2011; Post et al., 2013; Freeman and Lovenduski, 2015), triggering an increasing interest in coccolithophore ecology at high latitudes (e.g., Harada et al., 2012; Dylmer et al., ~~2013~~2015; Balch et al., 2016; Charalampopoulou et al., 2016; Giraudeau et al., 2016; Saruwatari et al., 2016; Nissen et al., 2018; Rigual Hernández et al., 2018; Krumhardt et al., 2019). Questions remain about how coccolithophore populations will adapt to predicted changes in their environment, if at all. There is growing concern that increasing levels of CO₂ in the atmosphere and the subsequent acidification of the ocean may disrupt the production of coccoliths. As more of the water column becomes undersaturated in CaCO₃ in the future (Fabry et al., 2009), carbonate dissolution will be favored over precipitation and coccolithophores may be less successful in exporting carbon to the deep ocean (Fabry et al., 2008). Additionally, any change in the global distribution and abundance of coccolithophore species relative to non-calcifying groups of phytoplankton (e.g., naked Haptophyceae cells, diatoms, etc) will have important effects on the biogeochemical cycling of carbon and climatic feedbacks. A known positive correlation exists over long timescales between surface-ocean carbonate ion concentrations [CO₃²⁻] and the mean coccolith mass of the associated Noëlaerhabdaceae assemblage, a family of coccolithophores, which includes the extant species *Emiliania huxleyi* (Beaufort et al., 2011). This correlation is driven by the replacement of more- by less-heavily calcified morphotypes or species with declining [CO₃²⁻]. Although the physiological driver for this strong ecological selective pressure is not known (Beaufort et al., 2011), it may determine Noëlaerhabdaceae biogeography, particularly in high latitudes, both in the past and future (Cubillos et al., 2007).

Geographical shifts in the occurrence or abundance of coccolithophores and assemblage compositions have been ~~actually-recently~~ observed (e.g., Rivero-Calle et al., 2015; Krumhardt et al., 2016). Repeated sampling in the Australian sector of the Southern Ocean (SO) over the past four decades has shown a dramatic range expansion of *Emiliania-E. huxleyi* south of 60°S (Cubillos et al., 2007), where any ocean acidification effect appears outweighed by surface-ocean warming. Other authors also recorded a southward expansion of the habitat of *E. huxleyi* in the SO during the last two decades (Winter et al., 2014), although the actual cause of this latitudinal expanse is still under debate (e.g., Patil et al., 2014; Malinverno et al., 2015). Even with a temperature-driven range expansion of coccolithophores in the SO, surface ocean carbonate chemistry is now capable of exerting a first-order control on the composition of coccolithophore assemblages as well as on ~~overall coccolithophore calcification calcification by coccolithophores from sub-specific morphotypes to species levels~~ (Cubillos et al., 2007; Mohan et al., 2008; Beaufort et al., 2011; Freeman and Lovenduski, 2015).

With significant changes in marine species distributions already occurring, it is crucial to understand the ecosystem structure as well as the potential impact of environmental change on the provision of essential ecosystem services (O'Brien et al., 2016). In this work, we assess the potential relationship between environmental parameters and the community composition, biogeography and calcification mode of modern high latitude coccolithophore communities across the Drake Passage. Accordingly, we calculated extant coccolithophore species numbers at different stations between 10 and 150 m of the water column, evaluated the coccolith mass variations of *E. huxleyi*. ~~We-and~~ compared these observations with in situ conductivity-temperature-depth (CTD) measurements, carbonate chemistry parameters, as well as to previously published Southern Ocean coccolithophore and calcification datasets.

2. Material and methods

2.1. Sample preparation for scanning electron microscope analyses and coccolithophore taxonomical considerations

Ninety-six water samples were taken at 18 ~~s~~ Stations located in the southern Chilean continental margin and across the western end of the Drake Passage (Fig. 1) from February to April 2016 during Expedition PS97 (Lamy, 2016). Seawater samples were obtained at different depths using a rosette sampler with 24 × 12 L Niskin bottles (Ocean Test Equipment Inc.) attached to a CTD Seabird SBE911 plus device (Lamy, 2016). The bottles were fired by ~~an~~ SBE32 carousel. For the study of coccolithophore assemblages, 4 to 7 samples per station, between 10 and 150 m water depth, were chosen. Two litres of water were filtered onto 0.45 µm pore size Polycarbonate Track-Etch Membrane, air-dried and stored over silica gel. A small part of the filter was cut out, fixed on an aluminum Scanning Electron Microscope (SEM) stub and sputtered with gold/palladium. A specific area of the center of the filter was analysed with Zeiss DSM 940A SEM at the University of Bremen, to determine quantitative cell counts for all morphotypes, species and total coccolithophore abundance at magnifications of 1000x, 2000x and 5000x when required. A minimum of 200 whole coccospheres per sample were counted and classified following Young et al. (2003), the revised classification of Jordan et al. ~~(2004)~~(2004) and the electronic guide to the biodiversity and taxonomy of coccolithophores Nannotax 3 (ina.tmsoc.org/Nannotax3/index.html) by Young et al. (2019).

Initially, seven different morphotypes of *Emiliania huxleyi* were distinguished in the study area belonging to two main groups, types A and B (for further details see Nannotax). These are type A (*huxleyi*), type A overcalcified, type B (*pujosiae*), type B/C, type C (*kleijneae*), type R and type O (which included specimens with an opened central area and specimens with the central area covered by a thin plate) (Table 1, Plate 1). Additionally, the degree of calcification was visually assessed while counting, that is

why the terms “normal”, “calcified” and “heavily calcified” are used in this work to denote some of the most robust *E. huxleyi* placoliths regardless the morphotype (see Plate 1). Semi-quantitative estimates of preservation were based on SEM observations on the coccolithophore assemblage. “Good” preservation implied little or no evidence of carbonate dissolution. Coccoliths with the main morphological characteristics partially altered but still identifiable at species level were tagged as “moderate” (e.g., T-elements within the taxon *E. huxleyi* were present). Specimens affected by strong dissolution or high fragmentation were regarded as “poor” (e.g., T-elements within the taxon *E. huxleyi* were dissolved).

2.2. Oceanographic data

The CTD-rosette hydrocasts during Expedition PS97 (Lamy, 2016) provided vertical water column profiles of in situ sea surface temperature (SST), salinity (SSS), density, oxygen and fluorescence (reflecting chlorophyll-a concentrations) (Fig. 2). According to the criteria specified by Orsi et al. (1995), different oceanographic fronts were crossed in this latitudinal transect, the Subantarctic Front (SAF) between stations PS97/036-1 and PS97/037-1 and the Antartic Polar Front (PF), at PS97/040-1 (Fig. 1). The areas between these fronts are referred to as Subantarctic Zone (SAZ), north of the SAF, Polar Front Zone (PFZ), between the SAF and PF and Antarctic Zone (AZ), south of the PF.

Phosphate, nitrate and silicate contents were retrieved from the World Ocean Atlas 2013 (WOA13) 1°x1° grid austral summer collection (December to February) (Garcia et al., 2014). Total carbon dioxide (T_{CO_2}) and total alkalinity (T_{ALK}) values were obtained from the Ocean Data View (ODV) global alkalinity and total dissolved carbon 1°x1° grid collection from the uppermost 150 m of the water column during austral summer (December to February) (Goyet et al., 2000). Since carbonate chemistry parameters were not measured in situ and data availability in the Drake Passage/Southern Ocean is limited, bicarbonate ion (HCO_3^-), carbonate ion (CO_3^{2-}), saturation state (Ω_{Ca}) and pH were calculated (as derived-variables) with ODV software version 4.6.3 (Schlitzer, 2015) using the uppermost 150 m of the GLODAPv2 collection (Key et al., 2015) and excluding measurements done before 1980. HCO_3^- , CO_3^{2-} , Ω_{Ca} and pH were also calculated (for comparison purposes) using the CO2SYS.XLS program for 10-20 m depth (Pierrot et al., 2006) and considering the interpolated values of T_{ALK} , T_{CO_2} and nutrients as well as the in situ measurement of temperature, salinity and pressure.

The values of each of these oceanographic parameters were estimated at the location of every CTD station by interpolating the available data points. The interpolation used a triangulation-based linear method for CTD stations located within the boundaries of the available data, and a nearest neighbour extrapolation method for any CTD stations located outside of the data boundaries. Latitude/longitude coordinates were projected to Universal Transverse Mercator (Zone 19E, World Geodetic System 1984) before interpolation in order to minimize the distance distortion inherent in geographic coordinates. The calculations were done in MATLAB™ using a custom function that is provided as supplementary material.

2.3. Sample preparation and coccolith calcite estimates in light microscope

Fifteen samples were selected in a latitudinal transect for coccolith calcite estimates, between 10 and 20 m water depth. Sample preparation was designed and carried out at the University of Salamanca (Spain). A part of the filter (ca. 1/4 of the original Polycarbonate Track-Etch Membrane) was cut out and carefully placed into small plastic bags. Buffered water (pH = 9) was prepared with 0.075g/L of Na_2CO_3 , 0.1 g/L and 0.04g/L of unflavored gelatin (“Gold Gelatin”). Six hundred μ L of buffered water were added, and the bags were sealed consistently in a triangular shape. After 30 minutes, the bags were shaken in a lab vortex for

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3 minutes to ensure that coccoliths were fully detached from the filter and re-suspended. Holding each bag on top of a cover slide placed on a hot plate (ca. 60°C), an incision was made with a scalpel in one of the bag angles, allowing the solution to slowly drop onto the cover slide. Once the water was evaporated, the cover slip was mounted with mixture of 50% Canada balsam and 50% Xylene and left in an oven (40° C) for at least 24h. This technique ensured that coccoliths were in the same plane of focus for polarized light microscopy. The dried filter was checked for presence/absence of coccoliths later on in the SEM of the University of Bremen (see supplementary material).

In this study, a Nikon Eclipse LV100 POL polarized light microscope with a 100x H/N2 objective set-up with circular polarization was used at the University of Salamanca. In order to determine coccolith mass and thickness, between 20 and 53 random fields of view were imaged using the Nikon DS-Fi1 digital 8-bit colour camera and the NisElements software, keeping the light level of the microscope and aperture settings constant. The images were processed with the C-Calcita software (for further details see Fuertes et al., 2014). Calibration was done with images of a well preserved and in-focus calcareous spine from the sample PS97/033-1 at 10 m (see supplementary material). In total, 796 coccoliths were analysed, with a minimum of 34 coccoliths (up to 94) measured per sample.

2.4. Diversity indices

The Shannon index (H), the Simpson diversity index ($1-D$) and the Fisher's alpha index were calculated with Paleontological Statistics (PAST™) software version 3.22 (Hammer et al., 2001) using the raw coccolithophore counts. H was determined with the following equation (1):

$$H = -\sum_i \frac{n_i}{n} \ln \frac{n_i}{n} \quad (1)$$

where n_i is the number of individuals in taxon i , and n is the total number of all individuals. This index takes into account the number of individuals as well as the number of taxa. H ranges from 0 for communities with just a-one taxon to higher values for communities with many taxa, each with few individuals (Harper, 1999). Dominance, D , was determined with the equation (2):

$$D = \sum_i \left(\frac{n_i}{n}\right)^2 \quad (2)$$

The Simpson index $1-D$ varies from 0 to 1 and measures evenness of the community. Fisher's alpha was calculated with the following equation (3):

$$S = a * \ln(1 + n/a) \quad (3)$$

where S is number of taxa, n is number of individuals and a is the Fisher's alpha.

2.5. Statistics

A principal component analysis (PCA) was performed on the coccolithophore relative abundance data using PAST™ software version 3.22 (Hammer et al., 2001). The objective of the PCA is to find hypothetical variables, called components, that capture the maximum proportion of the variance in the multivariate dataset as possible (Davis, 1986; Harper, 1999). These new variables are linear combinations of the original variables (Hammer et al., 2001). The first principal component has the largest variance possible, and each subsequent component explains the next greatest variance possible. Samples in which less than 50 coccospheres were counted were excluded from the original database, therefore just 74 samples were considered for the PCA. In order to avoid skewness, the relative abundances of the different coccolithophore taxa (x) were log-transformed prior to the PCA using the

formula: $y=\log(x+1)$. This transformation enhanced the importance of rare taxa, and minimized the dominance of few abundant taxa (Mix et al., 1999), in this case of *E. huxleyi* types B/C and C.

Seventeen coccolithophore taxa were considered for the PCA (see the taxonomical groups in Table 2). Due to the similar ecological preferences observed, *Papposphaera* sp. and *Pappomonas* spp. were lumped together in one taxonomical group, in the same way as holococcolithophores, the *Syracosphaera* and the *Ophiaster* species (Table 2). In contrast, *E. huxleyi* morphotypes were regarded as different groups. A correlation matrix between the principal component scores and the environmental variables (i.e., SST, SSS, fluorescence, oxygen and density measured in situ, as well as nitrate, phosphate and silicate contents interpolated from the WOA13) was performed in order to identify potential relationships between the environmental parameters and the coccolithophore components.

3. Results

3.1. Coccolithophore distribution

The analysis of 96 water samples shows in general higher cell concentrations in the uppermost 100 m of the water column in the SAZ and at shallower depths (ca. 60 m) in the PFZ (Fig. 3 a). The highest coccolithophore number, 214.6×10^3 cells/L, is reached at station PS97/034-2 (60 m) in SAZ, but high cell concentrations are generally observed north of the SAF (Fig. 3 a). The uppermost 150 m average of coccolithophore concentrations drastically drop at the oceanographic fronts, from 119.9×10^3 to 56.4×10^3 cells/L (PS97/036-1, /037-1) at the SAF, and from 32.2×10^3 to 0.1×10^3 cells/L (PS97/040-1, /041-1) at the PF. Stations south of the PF show very low cell numbers or are devoid of coccolithophores, but detached coccoliths are occasionally observed at the southernmost locations, even at station PS97/50-2. Coccolithophore preservation is in general moderate to good north of the PF, with optimum values at 10-20 m, but becomes notably poorer in the AZ (Fig. 3 c).

3.2. Coccolithophore community composition

Twenty-three different coccolithophore taxa (including morphotypes) are observed in this transect across the Drake Passage (Table 2). The most dominant species is *E. huxleyi*, although less abundant taxa also dwell in these (sub-) polar waters. In the following lines, we will comment on the main species composing the coccolithophore community, from the dominant taxa to the rare ones.

3.2.1. *Emiliana huxleyi*

Emiliana huxleyi dominates the coccolithophore assemblage, reaching values up to 212.5×10^3 cells/L at PS97/034-2 (60 m). This taxon is present in all the stations, except at coccolithophore-barren samples south of the PF, where coccospheres are occasionally recorded in low numbers, always below 3.0×10^3 cells/L. Seven well established morphotypes of *E. huxleyi* are observed; these are types A, A overcalcified, B, B/C, C, R and O (Plate 1), grouped into A (Fig. 4) and B (Fig. 5) according to Young et al. (2019). Neither malformed *E. huxleyi*, morphotype D, nor var. *corona* are present in the studied samples. However, a variation in the degree of coccolithophore calcification is observed; i.e., heavily calcified specimens as well as weakly calcified specimens are present in this transect (Table 1, Plate 1).

Emiliana huxleyi A group is less abundant than group B and includes type A, type A overcalcified and type R, all of them present in coastal waters (Fig. 4 a). Type R (Plate 1 a) is the most uncommon *E. huxleyi* morphotype, and it just dwells offshore Chile and

at station PS97/038-1 (20 m), where it reaches a maximum of 0.9×10^3 cells/L (Fig. 4 c). A similar distribution pattern is shown by *E. huxleyi* type A overcalcified (Fig. 4 d, [Plate 1 b, c](#)), with concentrations up to 1.8×10^3 cells/L at PS97/018-1 (100 m). *Emiliania huxleyi* type A (normal form or moderately calcified, [Plate 1 d-f](#)) is restricted to the continental margin, where it records numbers of up to 1.8×10^3 cells/L (Fig. 4 b).

Emiliania huxleyi B group specimens dwell north of the PF, showing a broader distribution than A group (Fig. 5 a); it includes types B, B/C, C and O. Type B ([Plate 1 g-i](#)) is the most unusual *E. huxleyi* morphotype within group B, with maximum numbers of 7.4×10^3 cells/L at PS97/029-1 (10 m). *Emiliania huxleyi* type B is abundant in a narrow deep band (0-150 m) at the Chilean margin/open ocean transition in the SAZ, and it is occasionally present in shallow waters of the PFZ (Fig. 5 b). Types B/C and C ([Plate 1 j-l](#)) show a broader latitudinal distribution and they are the most abundant *E. huxleyi* taxa with maxima of 74.0×10^3 cells/L at PS97/030-1 (150 m) and 103.2×10^3 cells/L at PS97/034-2 (60 m), respectively (Table 2, Fig. 5 c, d). Type B/C reaches higher concentrations at shallower depths (10-20 m), when compared to type C (20-100 m). Type O, with an “opened” central area ([Plate 1 m](#)), displays a similar distribution pattern to type B/C and reaches the highest numbers (8.0×10^3 cells/L at PS97/032-1, 10 m) mainly at shallow depths (10-20 m) (Fig. 5 e). On the contrary, type O with a central area covered by a thin “lamella”, which is highly variable in size, is notably more abundant and dwells at deeper depths (ca. 0-100 m), with maximum abundances of 72.0×10^3 cells/L at PS97/036-1, 60 m (Fig. 5 f). *Emiliania huxleyi* detached coccoliths were not counted but their presence or absence was assessed during the SEM analyses. Free detached coccoliths of *E. huxleyi* types B/C-C show a broader distribution than coccospheres and are rarely recorded up to 61.7°S at station PS97/050-2.

3.2.2. Other taxa

On top of *E. huxleyi*, less abundant taxa are observed. *Ophiaster* spp., including *O. hydroideus* (Plate 2) and sporadically *O. reductus*, is present primarily at the Chilean margin to a maximum 150 m water depth, but unexpectedly reaches up to 10.1×10^3 cells/L at PS97/038-1 at a depth of -10 m north of the PF (Fig. 6 a). *Calciopappus caudatus* is found in the uppermost 60 m of the SAZ and PFZ with maximum abundances of 5.3×10^3 cells/L also at PS97/038-1, 10 m (Fig. 6 b). Four species belonging to the genus *Syracosphaera* are recorded in the Drake Passage (i.e., *S. dilatata*, *S. corolla*, *S. marginaporata* and *S. pulchra*). *Syracosphaera* spp. dominates [among the rare coccolithophore assemblage](#) in the SAZ, except at coastal stations, and its highest numbers (up to 5.3×10^3 cells/L PS97/038-1) are recorded between 10 and 60 m (Fig. 6 c). In contrast, *Calcidiscus* s.l., mainly *Calcidiscus leptopus*, displays moderate numbers at the coastal stations offshore Chile (from 10 to 150 m), but reaches higher numbers (up to 1.4×10^3 cells/L at PS97/038-1, 10 m) southwards in a rather patchy, but shallow distribution (Fig. 6 d). *Papposphaera* sp. and *Pappomonas* spp. (including sp. 1 and sp. 5) are observed mainly in the PFZ or at shallow depths in the SAZ, with a maximum concentration of 1.4×10^3 cells/L at PS97/038-1, 10 m (Fig. 6 e). Holococcolithophores, mainly *Syracosphaera strigilis* HOL (Plate 2), are restricted to the uppermost 60 m in the SAZ with maximum values of 1.0×10^3 cells/L (PS97/038-1) (Fig. 6 f).

Rare coccolithophore taxa (maximum below 0.8×10^3 cells/L) are also recorded in the Drake Passage. *Gephyrocapsa muelleriae* is restricted to the northernmost stations offshore of Chile, while *Chrysolita* sp. is occasionally observed in the SAZ and *Calciosolenia murrayi* in the PFZ (Fig. 7 c-e). *Acanthoica quattrosipina* and *Wigwamma antarctica* display a broader distribution, even south of the PF, in the case of the latter (Fig. 7 a, b). Additionally, non-coccolithophores haptophytes belonging to the genera *Petasaria* and *Chrysochromulina* are generally present in the SAZ, PFZ and AZ, reaching maximum concentrations of 7.8×10^3 cells/L at PS97/037-1 at 60 m water depth (Fig. 7 f).

In order to investigate the relationship between the coccolithophore composition and the environmental variables in the study area, a PCA, was performed using the data from 74 sampling points (Fig. 8). Based on the broken stick method, the PCA indicates the existence of three main principal components (PC) explaining ~~the~~ 77.4% of the total variance (see supplementary material). PC1 explains 45.3% of the variance and it is positively related to the abundance of *E. huxleyi* type O and negatively related to *E. huxleyi* type C. PC2, connected to *E. huxleyi* type B/C, in a lesser extent to type B, and negatively correlated to *E. huxleyi* type A, accounts for 17.3% of the variance. PC1 seems associated to a marked temperature gradient. Positive PC1-values are linked to warm/moderately warm water taxa, which dwell north of the SAF, and negative PC1-values are connected to taxa that live in colder waters at higher latitudes. PC1 is correlated to SST and anticorrelated to density, oxygen content, macronutrients (phosphate, nitrate) and silicate (Table 3). PC2 is related to salinity variations (Table 3). Negative values of PC2 are related to extant coccolithophore taxa observed in the low salinity waters offshore Chile or to taxa living in the PFZ. On the contrary, positive values of PC2 are linked to taxa dwelling in the SAZ, where the SSS values are the highest of the studied transect. The coccolithophore assemblage composition separates out three different clusters in the PCA (Fig. 8) corresponding to three different oceanographic areas. The SAZ is characterized by positive values PC1 and PC2, the PFZ by negative PC1 and PC2 values, and the Chilean coastal ~~environment~~environment (Chl) by positive PC1 and negative PC2.

3.3. Diversity

Coccolithophore diversity indices (Shannon index *H*, Simpson index 1-D, Fisher's alpha) as well as the number of taxa are highest offshore Chile (0-150 m) and they became restricted to the uppermost 60 m in the PFZ (Fig. 9). The number of taxa is maximal ~~at~~ at station PS97/038-1 (10 m) and drastically drops south of the PF. The Shannon and Simpson 1-D indices display a similar pattern (Fig. 9 b, d), are highly correlated ($r=0.95$, supplementary material) and show that coccolithophore diversity decreases southwards. The number of taxa and diversity indices are strongly related to latitude (Fig. 9 c, f), with station PS97/038-1 (10-20 m) being an outlier due to the high diversity estimates recorded there.

3.4. Coccolith calcification –*E. huxleyi*–

Emiliana huxleyi specimens were classified into non-standardized sub-categories (e.g., regarding level of coccolith calcification) while counting in the SEM. Specimens of type A calcified/heavily calcified and overcalcified (Fig. 10 a) are present at the same locations as the common *E. huxleyi* type A (Fig. 4). Calcification in the group B implied in most of the cases a thicker central tube in the coccolith central area and thicker T-elements (Plate 1). Although type B calcified is recorded with rather low numbers, it shows a similar distribution to type B/C calcified (Fig. 10 b, c); both are restricted to the uppermost 150 m of the water column at stations PS97/029-1, /030-1 and /031-1. Type C calcified is occasionally recorded where types B and B/C are present, but shows a much broader and patchy distribution north of the PF (Fig. 10 d).

Coccolith mass measured with the software C-Calcita ranges from 19.8 to 0.8 ~~picograms~~ picograms, and median values (per station) vary from 7.3 to 2.4 pg. Relatively high *E. huxleyi* masses are recorded in the SAZ (Fig. 11), but not at the stations with the highest cell concentrations (such as PS97/034-2). Measurements of the coccolith mass allowed us to compare to the identified morphotypes. In general, coccolith mass decreases southwards across the Drake Passage (Fig. 11). While high coccolith masses are reached offshore Chile (i.e., stations PS97/017-1, /018-1) where *E. huxleyi* types R and A (overcalcified) are present, low coccolith masses are reached in the PFZ where types B/C and C dominate (i.e., stations PS97/037-1, /040-1). The gradual latitudinal mass decrease is occasionally interrupted by sudden drops in the mass estimates. These decreases in mass estimates appeared to be controlled by the predominance of a specific morphotype, for instance the low *E. huxleyi* mass values recorded at PS97/016-1 coincide with a sudden increase in the relative abundance of *E. huxleyi* type C (Fig. 11).

4. Discussion

4.1. Latitudinal variations in the coccolithophore abundance, distribution and diversity.

The observed maximum coccolithophore abundance recorded (up to 214.6×10^3 cells/L) is in agreement with previous studies carried out in different sectors of the Southern Ocean, which estimated maximum numbers between 130 and 640×10^3 cells/L (e.g., Eynaud et al., 1999; Findlay and Giraudeau, 2000; Cubillos et al., 2007; Gravalosa et al., 2008; Mohan et al., 2008; Hinz et al., 2012; Saavedra-Pellitero et al., 2014; Malinverno et al., 2015; Balch et al., 2016; Charalampopoulou et al., 2016). The coccolithophore abundance, ~~and diversity~~ and maximum depth habitat drastically drop from North to South and portrays the oceanographic fronts as ecological boundaries. Marked shifts in the coccolithophore numbers, community composition and diversity occurring at the SAF and PF observed here, were also previously noted by other authors in different sectors of the Southern Ocean (e.g., Eynaud et al., 1999; Gravalosa et al., 2008; Saavedra-Pellitero et al., 2014; Malinverno et al., 2015; Balch et al., 2016; Charalampopoulou et al., 2016). Although the aforementioned studies reported increases in the abundance of coccolithophores at the SAF and PF, we only observe an increase in the number of cells/L in the PF at shallow depths (< 60 m, Fig. 3), which is not so evident in the SAF. The increase in coccolithophore abundance recorded in the PF could be linked to the high biological productivity occurring at the Antarctic Circumpolar Current fronts (e.g., Murphy, 1995; Pollard et al., 2002; Patil et al., 2013) due to the frontal dynamics itself (e.g., Laubscher et al., 1993) or to the physical accumulation of particulate matter and nutrients at these convergence zones (e.g., Franks, 1992; Eynaud et al., 1999; Gravalosa et al., 2008; Balch et al., 2016).

The southernmost extent of *E. huxleyi* has been extensively discussed (e.g., Winter et al., 2014; Malinverno et al., 2015). The PF constitutes a natural sharp barrier which marks a drop in coccolithophore diversity and number of coccospheres (Saavedra-Pellitero et al., 2014; Saavedra-Pellitero and Baumann, 2015). Several studies observed the absence of *E. huxleyi* south of the PF (e.g., Verbeek, 1989; Charalampopoulou et al., 2016). However, specimens of *E. huxleyi* and *W. antarctica* are sporadically recorded south of the PF in the studied transect with numbers of $< 3 \times 10^3$ cell/L in the uppermost 80 m of the water column at stations PS97/043-3 and /047-1. *Emiliania huxleyi* is observed in low numbers at temperatures between 1.7 and -0.7°C (Fig. 2), below the 2°C isotherm limit that McIntyre and Bé (1967) originally established for the Atlantic Southern Ocean. Although it is unusual, few authors occasionally found *E. huxleyi* also dwelling in cold waters $< 2^\circ\text{C}$ (see Table 1 in Holligan et al., 2010). Monospecific assemblages of *E. huxleyi* have been also recorded south of the PF by other authors in the Pacific sector (e.g., Gravalosa et al., 2008; Saavedra-Pellitero et al., 2014), Australian sector (e.g., Nishida, 1986; Findlay and Giraudeau, 2000; Cubillos et al., 2007; Malinverno et al., 2015), in the Atlantic sector (e.g., Eynaud et al., 1999; Holligan et al., 2010) and Indian sector (e.g., Mohan et al., 2008; Patil et al., 2014). We speculate that the free detached coccoliths of *E. huxleyi* observed in our study area, up-down to 61.7°S , and showing a broader distribution than coccospheres, are not in situ and could have been transported. In any case the southernmost extent of coccolithophores is also influenced by the clear dominance of diatoms south of the PF, as suggested by the high diatom concentration (valves/g dry sediment) and biogenic opal content recorded in surface sediment samples from the same study area from the AZ of the Drake Passage in the AZ (Cárdenas et al., 2018) and from Pacific Southern Ocean extant plankton studies (e.g., Saavedra-Pellitero et al., 2014; Malinverno et al., 2016).

The number of taxa and coccolithophore diversity decreases southwards (Fig. 9) in agreement with other studies performed in the Drake Passage, the Australian and Pacific sectors of the Southern Ocean (e.g., Findlay and Giraudeau, 2000; Gravalosa et al., 2008;

Saavedra-Pellitero et al., 2014; Charalampopoulou et al., 2016). Coccolithophore diversity is related to the temperature gradient, as shown by the correlation between Shannon index and SST ($r=0.8$, see supplementary material). Contrary to Saavedra-Pellitero et al. (2014), the highest coccolithophore diversity values do not always occur at stations that showed the highest coccolithophore abundances (Figs. 3, 9). Few studies offshore Chile (ca. 33°S, 36°S) and in the Drake Passage showed low coastal coccolithophore diversity increasing towards open oceanic regions (Charalampopoulou et al., 2016; Menschel et al., 2016; von Dassow et al., 2018) which contrasts with the high number of taxa recorded in this work at uppermost 100 m of the water column at the Chilean margin (Fig. 9). Amongst different environmental factors, temperature could be one of the main variables favouring high coccolithophore diversity at the coastal stations.

The unexpected high diversity and number of taxa recorded at 10-20 m at station PS97/038-1 (previously labelled as “outlier”), coincident with relatively high density of coccospheres, does not seem to have been promoted by high SSTs, but rather by an occasional variation in the nutrient availability. So far, there are no nutrient measurements in situ available for this transect, but the interpolated values from the WOA13 austral summer suggest that nutrients are generally available this part of the SAZ, and shallow (10, 20 m) nitrate or phosphate concentrations (Fig. 12 h, i) do not abruptly change at PS97/038-1. Therefore this could be due to mesoscale eddies which could have advected nutrients (Frenger et al., 2018). We speculate that an increase in the fluorescence values (Fig. 2), reflecting higher chlorophyll-*a* concentrations, could be attributed to a higher availability of nutrients, which could have favored coccolithophores. The available fluorescence data (Fig. 2) seem to primarily reflect diatom concentration, south of the PF, followed by the non-coccolithophore haptophytes (*Petasaria* and *Chrysochromulina* spp.) in the SAZ and PFZ (up to 100 m water depth) superimposed to the coccolithophore distribution. Future quantitative analyses of extant diatoms and nutrients performed at the same stations and depths; are envisaged and will be required for better understanding of the phytoplankton community interactions and ecological patterns across the Drake Passage.

4. 2. Coccolithophore assemblages/Community composition in the study area/across the Drake Passage.

Based on the PCA (Fig. 8) it was possible to distinguish three main different oceanographic areas, characterized by different coccolithophore assemblages in the study area.

(1) The Chilean margin. Coccolithophores dwell mostly up to 400-150 m water depth in this zone (Fig. 3). *Emiliania huxleyi* type A is present in the stations closest to the Chilean coast (i.e., PS97/018-1 and /017-1), which recorded the highest SST and lowest SSS in the study area. This morphotype of *E. huxleyi* has been also observed in low abundances north of the SAF in different Pacific/Australian Southern Ocean (e.g., Cubillos et al., 2007; Saavedra-Pellitero et al., 2014; Malinverno et al., 2015), although was not observed by others authors (e.g., Gravalosa et al., 2008; Charalampopoulou et al., 2016), probably due to the high latitudes of those transects. Specimens of type A with different degree of calcification were present ranging from normal to overcalcified (Plate 1), the latter being more abundant. In the Northern Hemisphere, *E. huxleyi* type A dominates the coccolithophore assemblage in the North Atlantic and in Norwegian coastal waters (e.g., van Bleijswijk et al., 1991; Holligan et al., 1993), but not in the Southern Ocean (e.g., Cook et al., 2011; Hagino et al., 2011). *Emiliania huxleyi* type R was observed for first time in the Drake Passage, although it has been previously just observed off New Zealand (Langer et al., 2009; Iglesias-Rodriguez et al., 2017) and in the Eastern South Pacific (Beaufort et al., 2008; Beaufort et al., 2011; von Dassow et al., 2018).

Other minor taxa present in the Chilean margin are *Calcidiscus* s.l., *Ophiaster* spp and *G. muellerae*. The first has already been found by other authors in the SAZ (e.g., Saavedra-Pellitero et al., 2014; Malinverno et al., 2015), but its patchy distribution north of the PF is in agreement with observations by Charalampopoulou et al. (2016). *Ophiaster* spp. shows uneven distribution in the SAZ and PFZ, but it is also present in the Chilean continental margin. *Ophiaster* spp. is recorded offshore Chile with low number living up to 100 m water depth, which contrast the high numbers observed in the SAZ offshore New Zealand always above 60 m (Saavedra-Pellitero et al., 2014). Occasional low numbers of extant *G. muellerae* have also been observed in the Southern Ocean by Saavedra-Pellitero et al. (2014) and Findlay and Giraudeau (2000).

(2) The SAZ. This oceanographic zone is bounded in the south by the SAF. Salinity values are relatively constant at about 34 psu, but SST gradually decreases, ~~up-down~~ to ca. 6° C, while the nitrate and phosphate contents progressively increase (Figs. 2, 12). The SAZ is characterized by the dominance of *E. huxleyi* types C, B/C, O and B. ~~The maximum coccolithophore depth habitat progressively decreases southwards within the SAZ, from 150 m, in the transitional zone south of the chilean margin to up-to 100 m in the open ocean (Fig. 3).~~

The shift in occurrence from type A group to type B group has been recorded by some authors at the STF in the Australian sector (e.g., Hiramatsu and De Deckker, 1996; Findlay and Giraudeau, 2000; Malinverno et al., 2015). *Emiliana huxleyi* type B appears in very low abundance, in agreement with Saavedra-Pellitero et al. (2014), restricted to a transitional narrow deep band (0-150 m) between the Chilean coastal margin and more open conditions of the SAZ. It has been observed also north of the STF in the Indian Southern Ocean (Patil et al., 2013). This morphotype was otherwise only found in the Northern Hemisphere (Cook et al., 2011).

Emiliana huxleyi type O was established by Hagino et al. (2011), who observed this morphotype extensively distributed in the Southern Ocean. However, so far only Malinverno et al. (2015) and this study described *E. huxleyi* type O. This would mean that the reported geographic distributions of types B/C and C ~~might~~ be biased by inclusion of Type O (Hagino et al., 2011). *Emiliana huxleyi* type O (including the opened and lamella forms) is abundant in the SAZ, which is in agreement with Malinverno et al. (2015). *Emiliana huxleyi* type B/C and C (see Table 1) are the dominant taxa in the SAZ of the Southern Ocean (e.g., Findlay and Giraudeau, 2000; Cubillos et al., 2007; Gravalosa et al., 2008; Mohan et al., 2008; Saavedra-Pellitero et al., 2014; Saavedra-Pellitero and Baumann, 2015).

Among minor taxa, *Syracosphaera* spp., *Calcidiscus* sp., *A. quattrosipina* as well as holococcolithophores are found in the SAZ, in agreement with the assemblage observed by Charalampopoulou et al., (2016) north of the PF. Four species belonging to the genus *Syracosphaera* (plus *S. strigilis* HOL) are recorded in the study area, as previously observed (Gravalosa et al., 2008; Charalampopoulou et al., 2016). *Acanthoica quattrosipina*, a species tolerant to low salinity (Supraha et al., 2014) is present in southern high latitudes (Eynaud et al., 1999; Findlay and Giraudeau, 2000; Malinverno et al., 2015), but has not been recorded in other polar transects (e.g., Mohan et al., 2008).

(3) The PFZ. This oceanographic zone is bounded by the SAF and the PF. Salinity and SST gradually decreases with respect to the SAZ, and nutrient contents continue to progressively increase poleward (Figs. 2, 12). The maximum coccolithophore depth habitat in this zone is restricted to 60 m. *Emiliana huxleyi* types B/C and C dominate the PFZ and reach relatively high numbers north of the PF, although *E. huxleyi* type O is still present in low abundance (Fig. 11), as also observed by Malinverno et al. (2015). The high coccolithophore numbers observed in the SAZ and at the shallowest depths of the PFZ are part of the Great Calcite Belt, a region of high surface reflectance in the Southern Ocean due to the increased seasonal concentrations of coccolithophore and

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particulate inorganic carbon (Balch et al., 2011; Balch et al., 2016). We suggest that the uneven distribution of some of the coccolithophore taxa is driven by the physical processes of the ACC; i.e., it is primarily linked to the positions of frontal boundaries but also affected by the dynamics of mesoscale eddies, as mentioned by Holligan et al. (2010).

Minor taxa present in the PFZ, and broadly north of the PF include species of the family Papposphaeraceae (i.e., *Papposphaera* sp., *Pappomonas* spp. and *W. Antarctica*). They have small-sized and lightly calcified coccoliths, which makes them easily overlooked even under SEM. Specimens from the genera *Papposphaera* and *Pappomonas* have been observed at polar waters in the North Hemisphere (Thomsen, 1981; Samtleben and Schröder, 1992; Charalampopoulou et al., 2011; Thomsen and Østergaard, 2014) and also in the Southern Ocean (e.g., Gravalosa et al., 2008; Saavedra-Pellitero et al., 2014; Charalampopoulou et al., 2016). Apart from the fact that the lightly calcified polar coccolithophores are non-photosynthetic heterotrophs, which gives them a strong competitive advantage to dwell in the darkness for months every year, very little is known about them (Thomsen and Østergaard, 2013). However, because they are weakly calcified, they will be one of the first polar taxa to be threatened by ocean acidification (Thomsen and Østergaard, 2013).

4.3. *Emiliania huxleyi* mass variations across the Drake Passage.

In this study, the coccolith mass of *E. huxleyi* was measured across the Drake Passage up to the PF at depths ranging between 10 and 20 m water depth. The general southwards decreasing trend in *E. huxleyi* mass (Fig. 11) is in agreement with trends observed by Charalampopoulou et al. (2016) across the Southern Ocean (Fig. 13). Differences in the estimated mass values can be attributed to the distinct taxonomical considerations, to the methodologies used in both studies, and mainly to the different oceanographic conditions during the sampling periods (2009, 2016). The mean coccolith mass is related to strong latitudinal gradients in temperature ($r=0.75$), also observed by Charalampopoulou et al. (2016), total alkalinity ($r=-0.89$), total CO_2 ($r=-0.86$), HCO_3^- ($r=-0.81$ or -0.68 depending on the method used to calculate it) in agreement with Beaufort et al., (2011) and nutrient content (nitrate: $r=-0.75$, phosphate -0.71) noted by Charalampopoulou et al., (2016) (Table 4, Fig. 12). In contrast, the coccolith mass relationship to salinity, fluorescence, silicate content, carbonate ion, calcite saturation is not significant (Table 4). Still it could be argued that the poleward decrease in coccolith mass decrease roughly coincides with a reduction in Ω_{Ca} (Fig. 12) and CO_3^{2-} (Table 4); in agreement with different studies that found depressed coccolith calcification at low Ω_{Ca} and CO_3^{2-} values (e.g., Riebesell et al., 2000; Beaufort et al., 2011). Although the pH variation is rather reduced, the anticorrelation between coccolith mass and pH becomes significant ($r=-0.7$) depending on the method used (Table 4, Fig. 12), in agreement with the biogeochemistry and optics South Pacific experiment (BIO SOPE) data from Beaufort et al (2011) ($r=-0.52$). The negative correlation of the present data contrasts with the global and well established relationship between coccolith mass and pH ($r=0.75$) (Beaufort et al., 2011). However, the relationship between coccolith mass and the carbonate chemistry parameters should be considered carefully. T_{ALK} , T_{CO_2} , Ω_{Ca} , pH and HCO_3^- have been calculated from the GLODAP-v2 database (in which the majority of the datapoints are scattered and samples were measured just in August 2005 and in February 2009) (Key et al., 2015) or from the global alkalinity and total dissolved carbon collection (Goyet et al., 2000) which shows an austral summer average using the CO2SYS.XLS program (Pierrot et al., 2006).

The observed decreasing trend of the coccolith mass can be linked to the latitudinal succession from type A group to type B group (Fig 12 b), in agreement with other authors who observed a latitudinal trend from *E. huxleyi* more calcified to weakly calcified

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morphotypes (e.g., Cubillos et al., 2007; Mohan et al., 2008). In the Chilean margin, the highest coccolith masses recorded are related to the presence of *E. huxleyi* type A (including normal, calcified and overcalcified specimens) and type R (Fig. 11), observed also by other authors at lower latitudes offshore Chile (Beaufort et al., 2008; Beaufort et al., 2011; von Dassow et al., 2018). Although it is uncommon, heavily calcified *E. huxleyi* morphotypes have been recorded in reduced pH and Ω_{Ca} conditions in other parts of the globe (e.g., Smith et al., 2012; Triantaphyllou et al., 2018). The presence of calcified specimens of *E. huxleyi* type B/C, and in a lesser extent of type C, in the transitional zone from the Chilean margin to the open ocean reflects an increase in the coccolith masses (Figs. 10, 11). In contrast, a higher relative abundance of *E. huxleyi* type C corresponds to smaller coccolith masses in the SAZ. Striking are the relatively low coccolith mass values in the open ocean of the SAZ that coincide with maxima in the density of coccolithophores (Fig. 12).

The dataset presented here constitutes an important contribution to the coccolithophore ecology sparsely studied at high latitudes. This work is also relevant for future climate and ocean model simulations in the context of global warming and ocean acidification threatening calcifying plankton. Taking into account the existing relationships between the physico-chemical parameters and the coccolithophore components, changes in the composition and calcification modes of *E. huxleyi* morphotypes are expected to occur in the Drake Passage with the ongoing climate change. However, our study does not provide enough evidence to infer how coccolithophores will cope with a forthcoming changing ocean. We speculate that future sea surface warming and stratification (Boyd et al., 2008), concomitant with a southward migration of the Antarctic Circumpolar fronts, will lead to a increase in the numbers of *E. huxleyi* at higher latitudes and to a potential higher calcification poleward in agreement with the model from Krumhardt et al. (2017). At the same time, pH, CO_3^{2-} and Ω_{Ca} are predicted to decrease (e.g., Hauri et al., 2015), which will also affect coccolithophores. Based on our limited data, conflicting conclusions can be drawn from the carbonate parameters. We could hypothesize that *E. huxleyi* will calcify more in a future Southern Ocean scenario at lower pH, consistent with few culture experiments (e.g., Iglesias-Rodriguez et al., 2008) but also that this species will reduce its calcification rate at lower CO_3^{2-} , Ω_{Ca} and higher HCO_3^- in agreement with the negative effects of ocean acidification suggested by several authors (e.g., Riebesell et al., 2000). Bearing in mind that culture experiments in different strains of *E. huxleyi* have already shown different responses to changing carbonate chemistry (Langer et al., 2009), it seems necessary to consider also the degree of adaptive potential of coccolithophores in future studies to predict their upcoming performance in the Polar realm.

5. Conclusions/Summary

This study documents the latitudinal and the depth variability in the coccolithophore assemblage composition and calcification of *Emiliana huxleyi*, the dominant species, across the Drake Passage, driven by physical, chemical and biological parameters in the surface ocean. Coccolithophore abundance, and diversity and maximum depth habitat decrease southwards portraying the oceanographic fronts as ecological boundaries. Marked shifts in the coccolithophore numbers, community composition and diversity occur at the Subantarctic Front (SAF) and Polar Front (PF). Three main different oceanographic areas are characterized, based on the coccolithophore composition:

(1) The Chilean margin. *Emiliana huxleyi* type A (normal and overcalcified) and type R are present in the stations closest to the Chilean coast, which record the highest SST and lowest SSS in the study area. Rare taxa present offshore Chile are *Calcidiscus* s.l., *Ophiaster* spp. and *Gephyrocapsa muelleriae*.

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(2) The Subantarctic **Z**one (SAZ). This zone is bounded by the SAF in the south. Salinity values are relatively constant and SST gradually decreases while nutrient content increases. The SAZ is characterized by the dominance of *E. huxleyi* types C, B/C, O and B. *Emiliana huxleyi* reaches maximum values of $212.5 \cdot 10^3$ cells/L north in the SAZ. Minor taxa include *Syracosphaera* spp. (including holococcolithophores from this genus), *Calcidiscus* sp. and *Acanthoica quattrosolina*.

(3) The **P**olar **F**ront **Z**one (PFZ). It is bounded by the SAF in the north and the PF in the south. Salinity and SST progressively decreases with respect to the SAZ, and nutrient contents continue to increase poleward. *Emiliana huxleyi* types B/C and C dominate the PFZ and reach relatively high numbers north of the PF, although *E. huxleyi* type O is still present.

Minor taxa present in the PFZ, and broadly north of the PF include species of the family Papposphaeraceae (i.e., *Papposphaera* sp., *Pappomonas* spp. and *Wigwamma antarctica*). Specimens of *E. huxleyi* and *W. antarctica* are sporadically recorded south of the PF with numbers of $< 3 \cdot 10^3$ cell/L and dwelling at temperatures $< 2^\circ$ C.

The general decreasing trend in *E. huxleyi* coccolith mass can be linked to the latitudinal succession from type A group (in the Chilean margin) to type B group (in the PFZ). Coccolith mass and coccolithophore diversity are related to the strong latitudinal gradient in temperature. Coccolith mass also shows anticorrelation to total alkalinity, total CO₂, bicarbonate ion (HCO₃⁻), pH and nutrient content, which contrasts with the global and well established positive relationship between coccolith mass and pH as well as total alkalinity. However, the relationship between coccolith mass and the carbonate chemistry parameters should be considered carefully, since in situ measurements are not available. The existing relationships between the physico-chemical parameters and the coccolithophore components in the Drake Passage suggest that assemblage composition and calcification modes of *E. huxleyi* will be strongly affected by the ongoing climate change.

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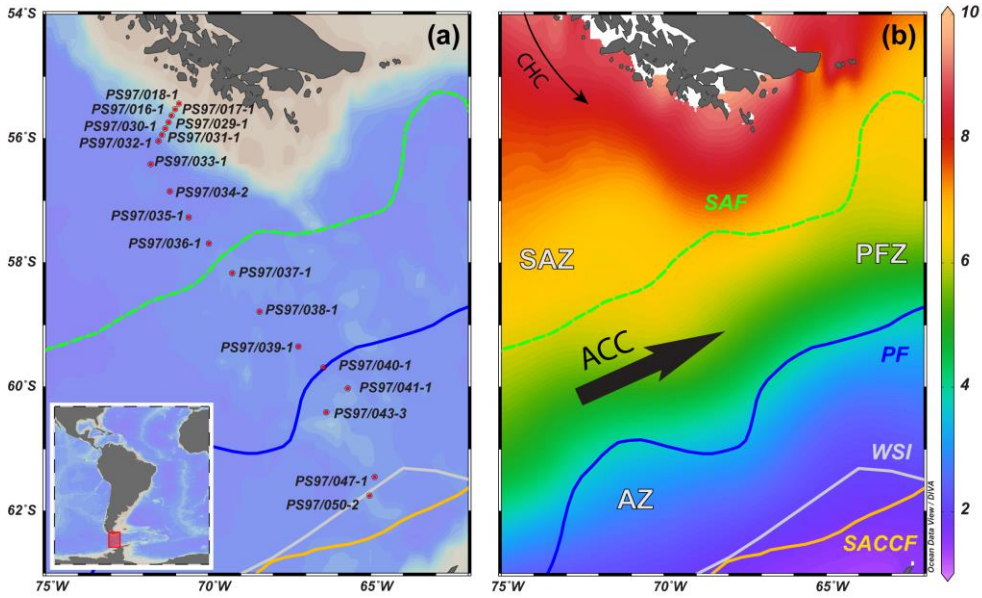
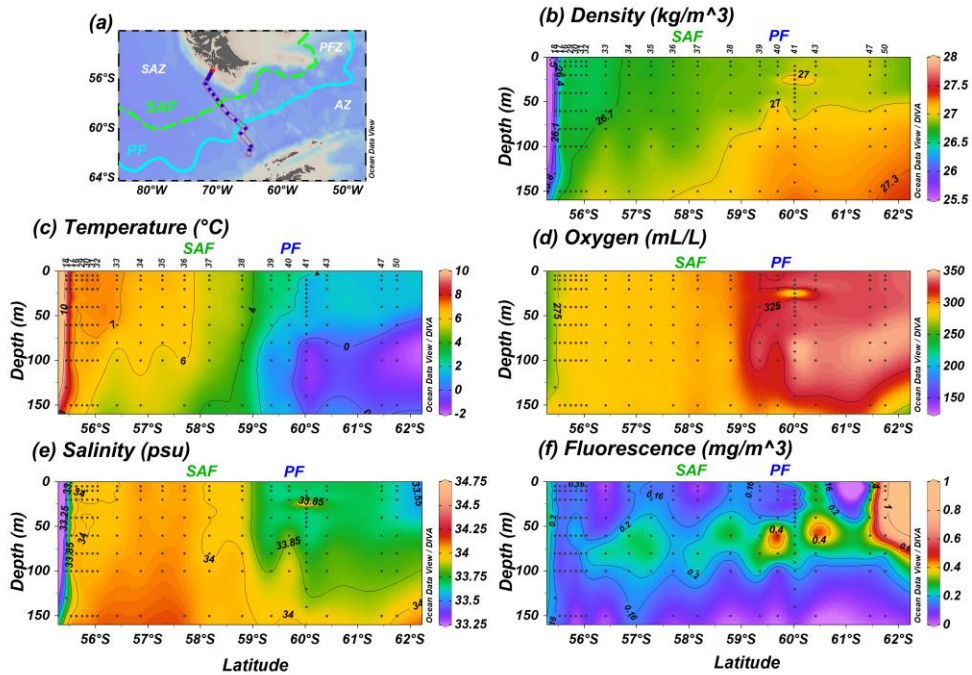


Fig. 1. (a) Overview map showing the bathymetry of the Drake Passage area and the CTD stations studied; (b) sea surface temperature (°C) seasonal average (January, February, March) at 0 m water depth from the World Ocean Atlas 2013, 0.25° grid (Locarnini et al., 2013) plotted with Ocean Data View (ODV) software version 4.6.3 (Schlitzer, 2015). The different oceanographic fronts are indicated as follows: Subantarctic Front (SAF) with a green dashed line, Antarctic Polar Front (PF) with a blue line and southern Antarctic Circumpolar Current Front (SACCF) with an orange line (Orsi et al., 1995). The areas between the fronts are referred to as Subantarctic Zone (SAZ), Polar Front Zone (PFZ) and Antarctic Zone (AZ). CHC: Cape Horn Current, ACC: Antarctic Circumpolar Current, WSI: winter sea ice extent (Comiso, 2003).

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Fig. 2. (a) Location of the PS97 CTD stations studied, (b) water density (kg/m^3), (c) sea surface temperature ($^{\circ}\text{C}$), (d) dissolved oxygen (mL/L) (e) sea surface salinity (psu) and (f) fluorescence (mg/m^3) profiles from 55.4°S to 61.7°S up to a depth of 150 m. Black dots indicate sampling points. The different oceanographic fronts (according to Orsi et al., 1995) are indicated as SAF—green dashed line, PF—blue line.

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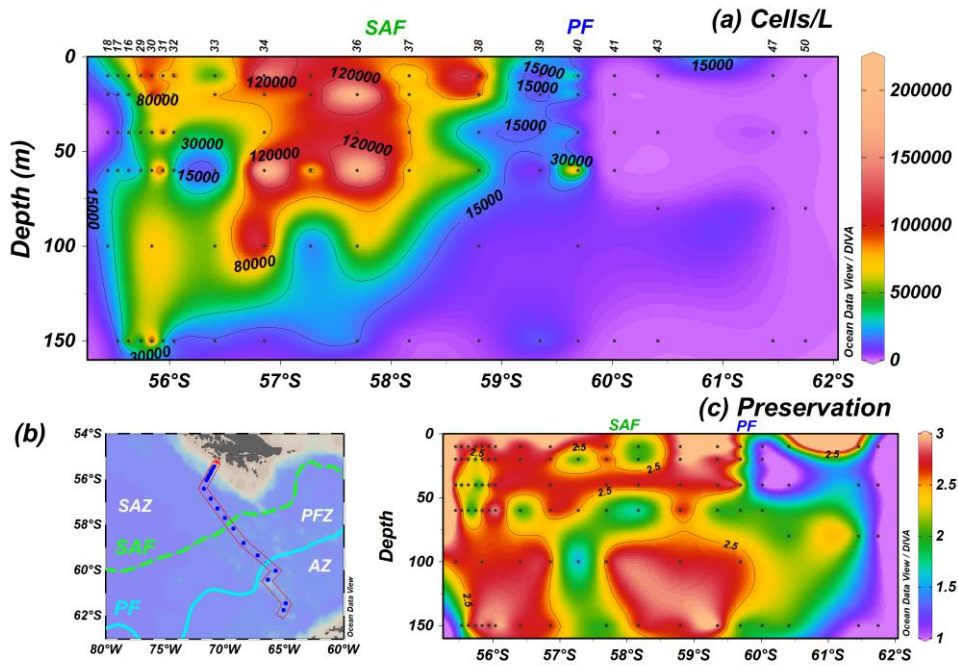
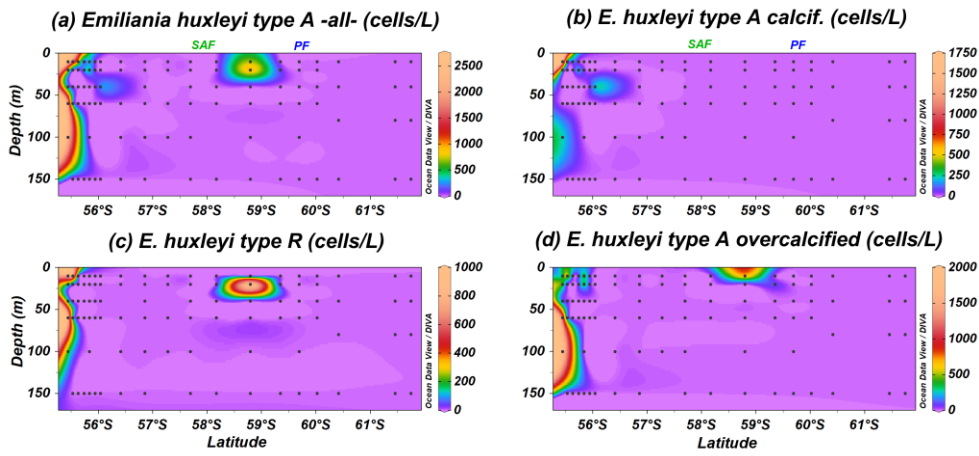


Fig. 3. PS97 CTD stations showing (a) the number of coccospheres/l across a (b) latitudinal transect in the Drake Passage and (c) semiquantitative estimates of coccolith preservation for the uppermost 150 m of the water column (1=poor, 2=moderate, 3=good). Black dots indicate sampling points and the different oceanographic fronts according to Orsi et al., (1995) are indicated as SAF—green dashed line, PF—blue line.



10 Fig. 4. Location of the PS97 CTD stations studied showing the number of coccospheres/L of (a) *Emiliana huxleyi* type A group, (b) *E. huxleyi* type A moderately calcified, (c) *E. huxleyi* type R and (d) *E. huxleyi* overcalcified. The Subantarctic Front (SAF) and the Polar Front (PF) according to Orsi et al. (1995) are indicated.

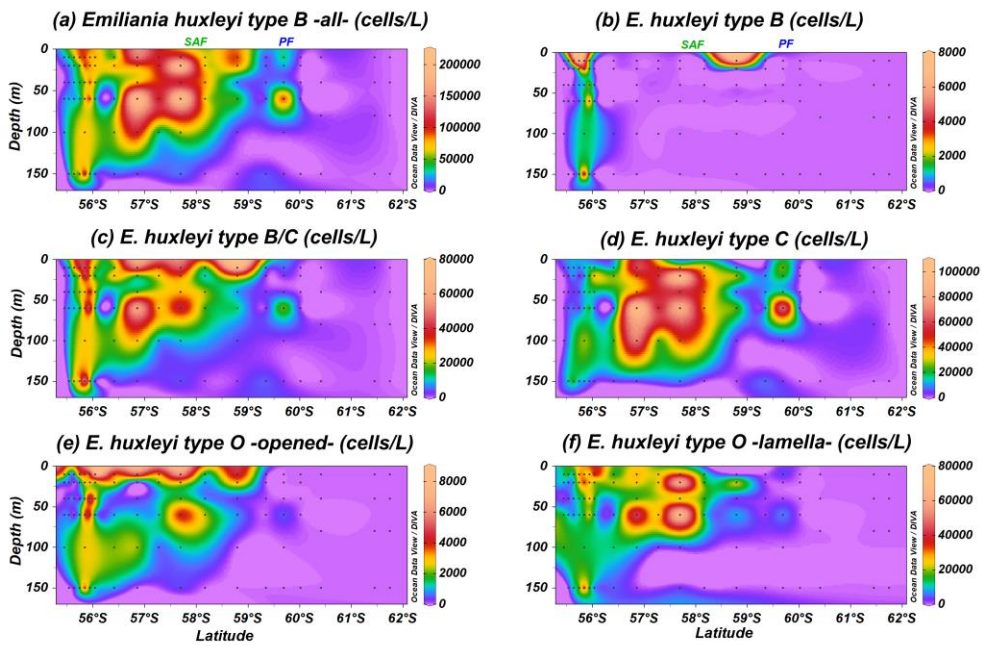


Fig. 5. Location of the PS97 CTD stations studied showing the number of coccospheres/L of (a) *Emiliana huxleyi* type B group, (b) *E. huxleyi* type B, (c) *E. huxleyi* type B/C, (d) *E. huxleyi* type C, (e) *E. huxleyi* type O (with opened central area) and (f) *E. huxleyi* type O (with lamella). The Subantarctic Front (SAF) and the Polar Front (PF) according to Orsi et al. (1995) are indicated.

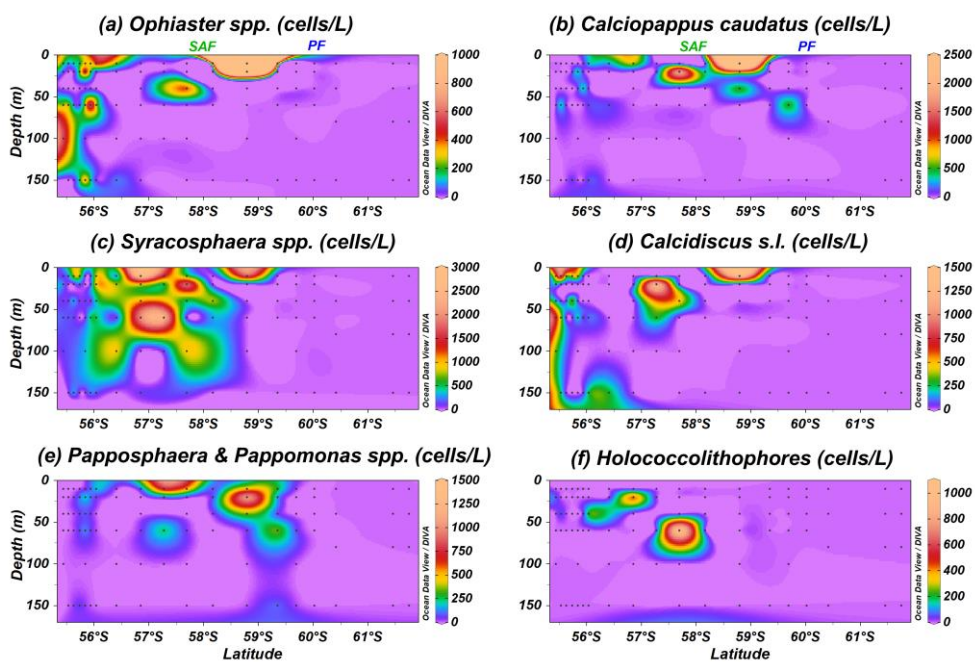


Fig. 6. Location of the PS97 CTD stations studied showing the number of coccospheres/L of (a) *Ophiaster* spp., (b) *Calciopappus caudatus*, (c) *Syracosphaera* spp., (d) *Calcidiscus* s.l., (e) *Papposphaera* & *Pappomonas* spp., and (f) holococcolithophores. The Subantarctic Front (SAF) and the Polar Front (PF) according to Orsi et al. (1995) are indicated.

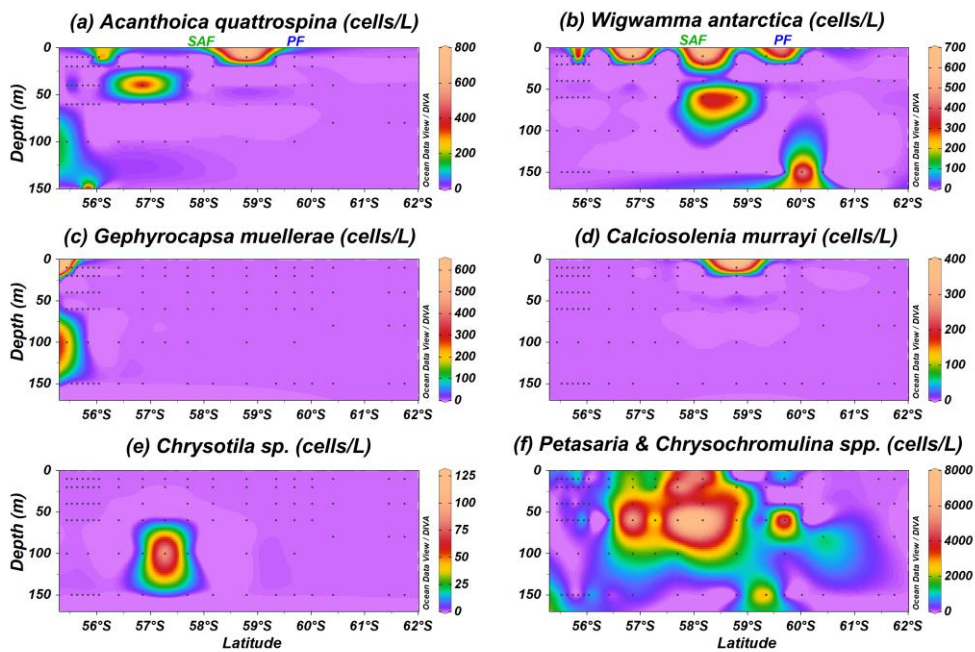


Fig. 7. Location of the PS97 CTD stations studied showing the number of coccospheres/L of (a) *Acanthoica quattropsina*, (b) *Wigwamma antarctica*, (c) *Gephyrocapsa muelleriae*, (d) *Calciosolenia murrayi*, (e) *Chrysofila* sp., (f) *Petasaria* and *Chrysochromulina* spp. The Subantarctic Front (SAF) and the Polar Front (PF) according to Orsi et al. (1995) are indicated.

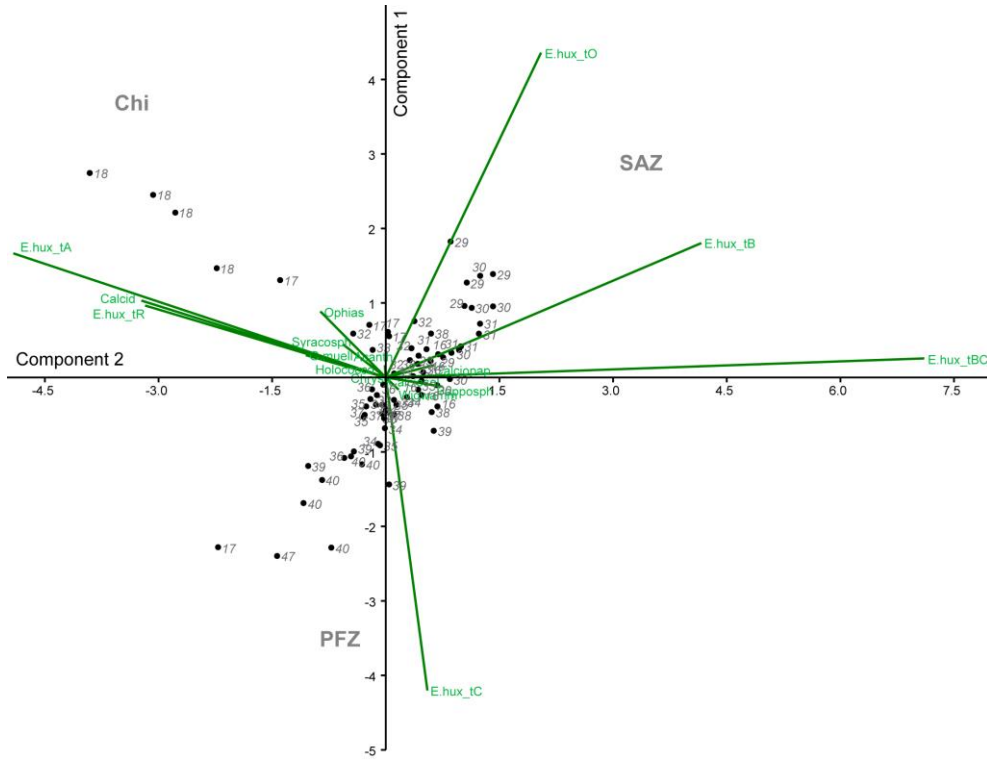


Fig. 8. Principal Component Analysis (PCA) biplot performed on the coccolithophore dataset. The two main components, PC1 (y axis) and PC2 (x axis), coccolithophore taxa, station numbers and the different clusters are indicated. The following abbreviations are used in the figure: E.hux_tA (*E. huxleyi* type A), E.hux_tB (*E. huxleyi* type B), E.hux_tBC (*E. huxleyi* type BC), E.hux_tC (*E. huxleyi* type C), E.hux_tO (*E. huxleyi* type O), E.hux_tR (*E. huxleyi* type R), Calcid (*Calcidiscus* s.l.), Acanth (*Acanthoica quattrosolina*), Calcipap (*Calcipapirus caudatus*), Calciosol (*Calciosolenia murrayi*), Chryso (*Chrysotila* sp.), G.mueller (*G. muelleriae*), Holococco (*Holococcolithophores*), Ophias (*Ophiaster* spp.), Papposph (*Papposphaera* sp and *Pappomonas* spp.), Syracosph (*Syracosphaera* spp.), Wigwamm (*Wigwamma antarctica*), SAZ (subantarctic zone), PFZ (polar front zone) and Chi (Chilean coastal zone).

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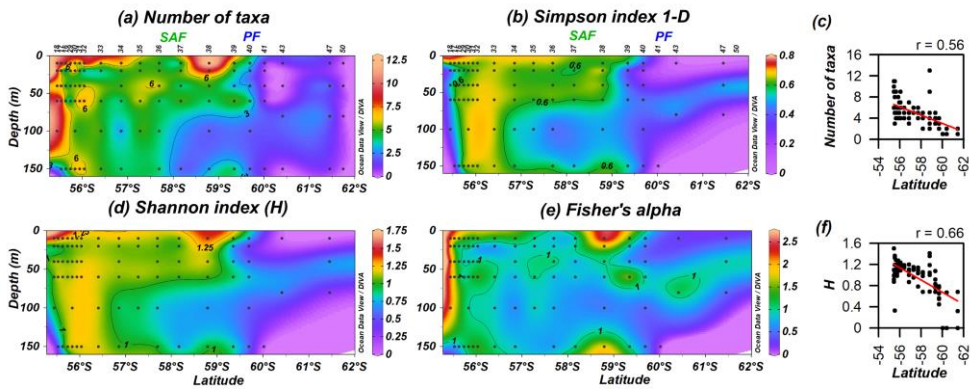


Fig. 9. PS97 latitudinal transects showing coccolithophore diversity: (a) number of taxa, (b) Simpson index 1-D, (c) number of taxa vs latitude, (d) Shannon index (H), (e) Fisher's alpha and (f) H vs latitude. The Subantarctic Front (SAF) and the Polar Front (PF) according to Orsi et al. (1995) are indicated.

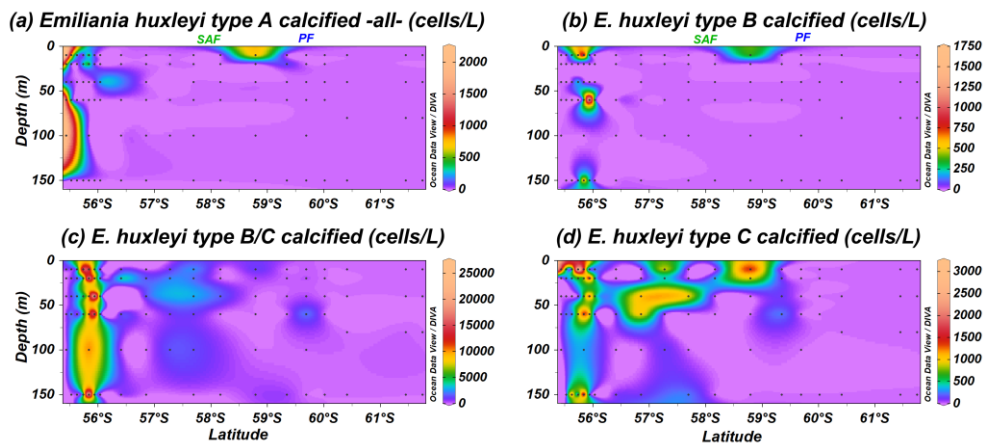


Fig. 10. Location of the PS97 CTD stations studied showing the number of coccospheres/L of *Emiliana huxleyi* calcified and overcalcified (a) type A group, (b) type B, (c) type B/C and (d) type C. The Subantarctic Front (SAF) and the Polar Front (PF) according to Orsi et al. (1995) are indicated.

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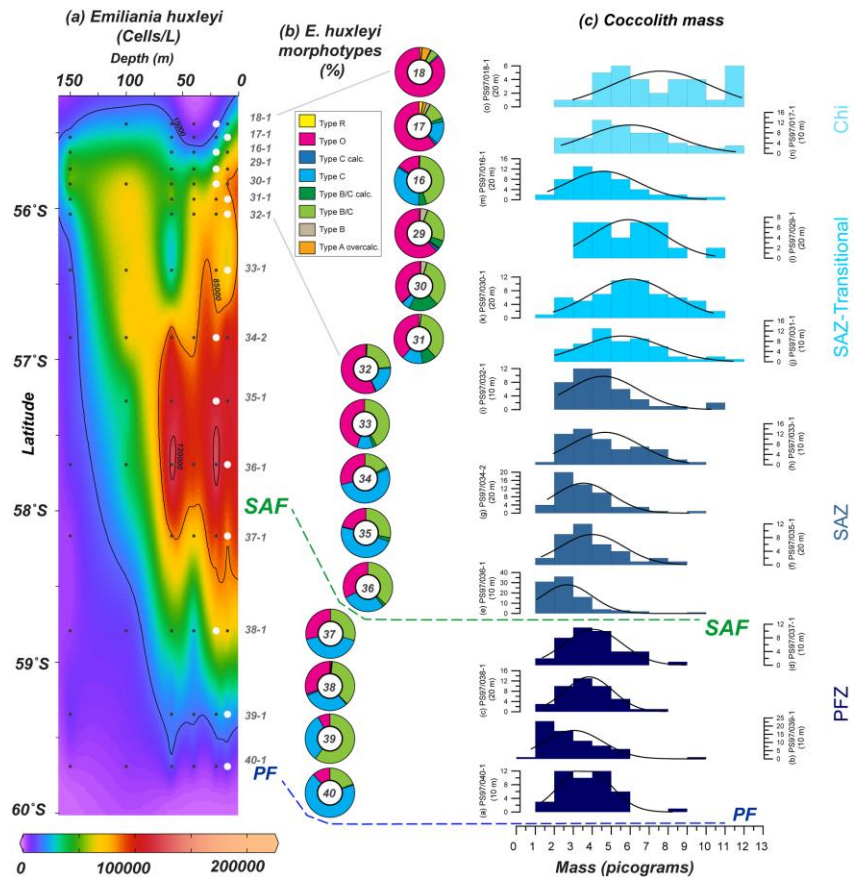


Fig. 11. (a) Location of the PS97 CTD stations between 55.4 and 60°S showing the number of cells/L of *Emiliana huxleyi* from 0 to 150 m. (b) Relative abundance of *E. huxleyi* morphotypes (including the different degree of calcification observed in SEM) in a latitudinal transect (10-20 m, indicated with white dots in (a)), and (c) coccolith mass histograms at those specific locations. Bin size in (c) is 1 picogram and normal distribution fitting line is shown. Station's number, different zones: Chi (Chilean coastal zone), SAZ-transitional (from Chi to open ocean), SAZ (subantarctic zone), PFZ (polar front zone), and the main oceanographic fronts: Subantarctic (SAF) and Polar Fronts (PF) according to Orsi et al. (1995) are indicated.

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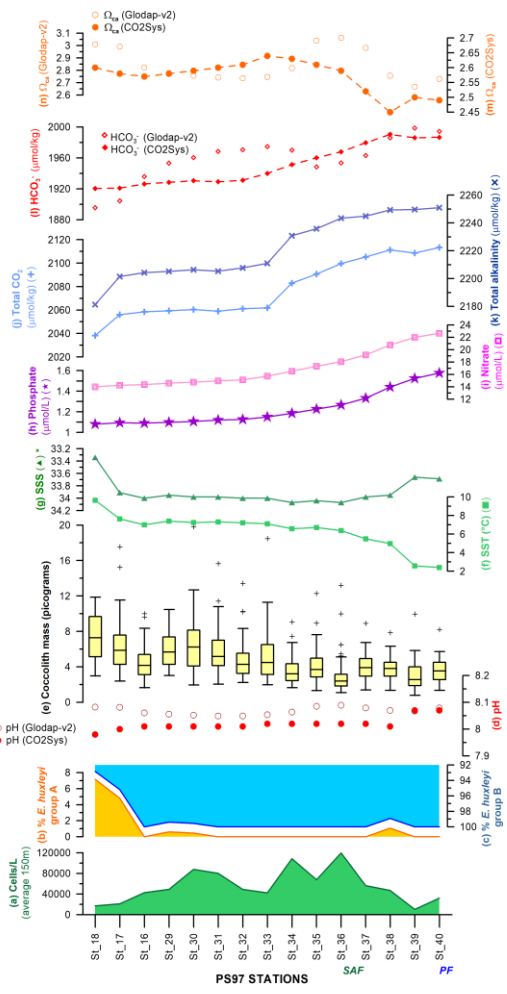
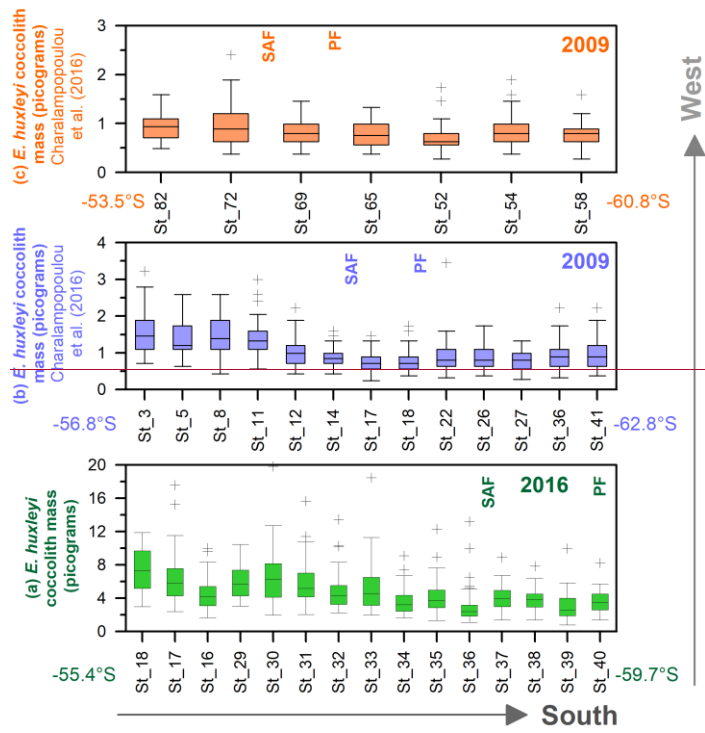


Fig. 12. Latitudinal transect showing on the left hand side: (a) the uppermost 150 m average of cells/L, relative abundance of *E. huxleyi* morphotypes belonging to (b) group A and (c) to group B, (d) pH calculated with different approaches (Pierrot et al., 2006; Key et al., 2015), (e) coccolith mass boxplot (outliers have been indicated with “+”), (f) sea surface temperature (SST, °C) and (g) salinity (° note the inverted scale) measured in situ (Lamy, 2016). Interpolated (h) phosphate (µmol/L) and (i) nitrate contents (µmol/kg) (Garcia et al., 2014), (j) total alkalinity (µmol/kg) and (k) total CO₂ (µmol/kg) (Goyet et al., 2000), (l) bicarbonate ion (HCO₃⁻, µmol/kg) and (m) calcite saturation state (Ω_{Ca}) calculated with different approaches (Pierrot et al., 2006; Key et al., 2015). The subantarctic (SAF) and Polar Fronts (PF) according to Orsi et al. (1995) are indicated.

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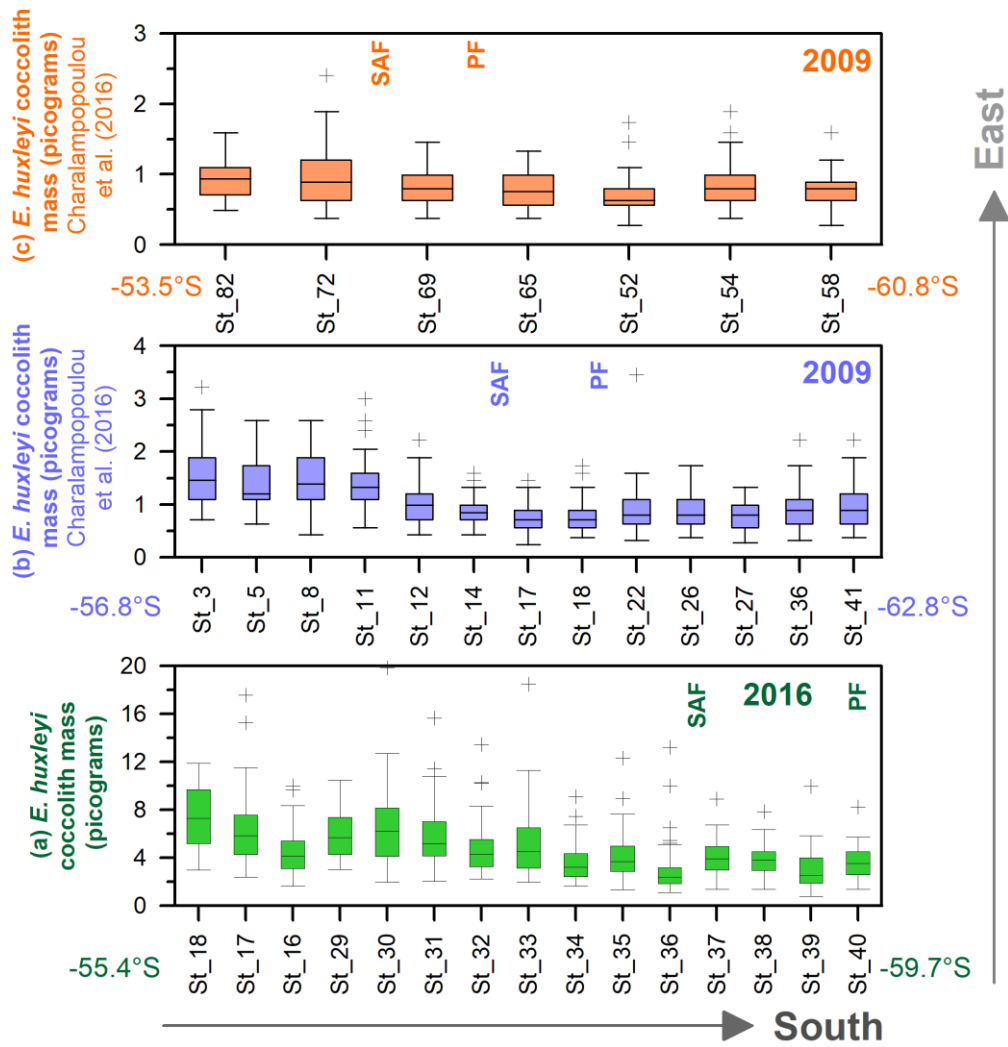


Fig. 13. Drake Passage latitudinal transects from East to West showing boxplot coccolith mass estimates (in picograms): (a) this study, (b) transect at around 68°W Charalampopoulou et al., (2016), (c) transect at around 55-58°W Charalampopoulou et al., (2016). Note that (b, c) have been calculated from Charalampopoulou et al., (2016). Outliers have been indicated with “+” and numbers in the x axes refer to the original station numbers. The approximate location of the Subantarctic (SAF) and Polar Fronts (PF) are shown as well as the year of the sampling.

Table 1. Classification scheme of *Emiliana huxleyi* morphotypes observed in the present study (modified from Hagino et al. (2011) regarding Young and Westbroek (1991), Cook et al. (2011) and Young et al. (2019)).

<i>E. huxleyi</i> morphotype	Morphology of the distal shield	Morphology of the central area	Distal shield length	Comparable morphotypes in the literature	
A group	Type A	Moderate-heavily calcified elements	Grill	<4 μm	Warm type (McIntyre and Bé, 1967), var. <i>huxleyi</i> (Medlin et al., 1996)
	Type A overcalcified	Moderate-heavily calcified elements	closed or nearly closed	<4 μm	Type A overcalcified (Young et al., 2003)
	Type R	<i>Reticulofenestra</i> -like heavily calcified distal shield elements	Grill Covered or nearly covered	<4 μm	Type R (Young et al., 2003), (von Dassow et al. 2018)
B group	Type B	Lightly calcified elements	solid plate	≥4 μm	Type B (Young et al., 2003), var. <i>pujosae</i> (Verbeek, 1990) Medlin & Green in Medlin et al. (1996)
	Type B/C	Lightly calcified elements	solid plate	<4 μm	Type B/C (Young et al., 2003), var. <i>aurorae</i> (Cook, 2011)
	Type C	Lightly calcified elements	solid plate	<3.5 μm	Cold type (McIntyre and Bé, 1967), Type C (Young et al., 2003), var. <i>kleijniae</i> (Medlin et al., 1996)
	Type O	Lightly calcified elements	opened or lamella	variable in size	Subarctic type (Okada and Honjo, 1973), Type B (Hagino et al., 2005)

Table 2. Summary of the coccolithophore taxa/groups in the studied plankton samples. The presence of each species in the Subantarctic Zone (SAZ), in the Polar Front Zone (PFZ) or in the Antarctic Zone (AZ) is indicated with “+” and with “-” if it is occasional. The station and water depth where the maximum number of cells/L is recorded has been also indicated.

Coccolithophore taxa / groups	SAZ	PFZ	AZ	Max. (cells/L)	Station	Depth (m)
<i>Acanthoica quattrosolina</i>	+	-		0.7*10 ³	PS97/038-1	10
<i>Calcidiscus</i> s.l.	+	-		1.4*10 ³	PS97/038-1	10
<i>Chrysotila</i> sp.	-			0.1*10 ³	PS97/035-1	100
<i>Calciosolenia murrayi</i>	+	-		0.3*10 ³	PS97/038-1	10
<i>Calcioppapus caudatus</i>	+	-		5.3*10 ³	PS97/038-1	10
<i>Emiliana huxleyi</i> type A (incl. overcalcified)	+	-		2.1*10 ³	PS97/018-1	100
<i>Emiliana huxleyi</i> type R	+	-		0.9*10 ³	PS97/038-1	20
<i>Emiliana huxleyi</i> type B	+	-		7.4*10 ³	PS97/029-1	10
<i>Emiliana huxleyi</i> type B/C	+	+	-	74.0*10 ³	PS97/030-1	150
<i>Emiliana huxleyi</i> type C	+	+	-	103.2*10 ³	PS97/034-2	60
<i>Emiliana huxleyi</i> Type O (incl. lamella)	+	+	-	76.2*10 ³	PS97/036-1	60
<i>Gephyrocapsa muelleriae</i>	+			0.6*10 ³	PS97/018-1	10
Holococcolithophores	+			1.0*10 ³	PS97/036-1	60
<i>Ophiaster</i> spp. (incl. <i>hydroideus</i> & <i>reductus</i>)	+	-		10.2*10 ³	PS97/038-1	10
<i>Papposphaera</i> sp.	-			0.2*10 ³	PS97/035-1	60
<i>Pappomonas</i> spp. (incl. sp. 1 & 5)	+	+		1.4*10 ³	PS97/038-1	20
<i>Syracosphaera</i> spp. (incl. <i>dilatata</i> , <i>corolla</i> , <i>marginaporata</i> , <i>pulchra</i>)	+	-		2.8*10 ³	PS97/034-2	10
<i>Wigwamma antarctica</i>	-	+	-	0.7*10 ³	PS97/034-2	10

Table 3. Correlation matrix between Principal component (PC) scores and the environmental variables. Significant Pearson correlation coefficients are indicated in bold ($p < 0.05$)

Variables	PC 1	PC 2
SST	0.77	-0.01
SSS	-0.23	0.66
Density	-0.77	0.41
Oxygen	-0.70	-0.02
Fluorometer	-0.13	-0.13
Phosphate	-0.70	-0.21
Nitrate	-0.70	-0.20
Silicate	-0.61	-0.22

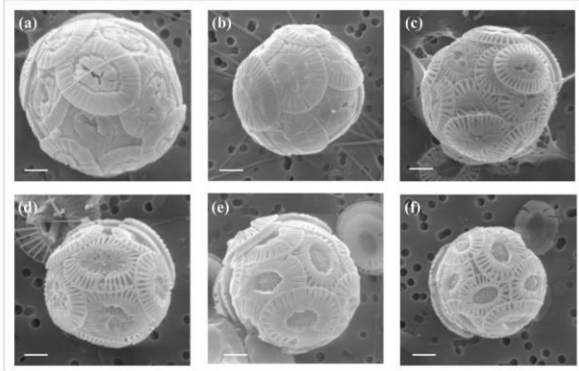
Table 4. Correlation matrix between mean / median coccolith values and the environmental variables. Significant Pearson correlation coefficients are indicated in bold ($p < 0.05$). Measurements in situ and calculated values are indicated.

Environmental variables		Mean coccolith mass (pg)	Median coccolith mass (pg)
In situ	Temperature	0.75	0.71
	Salinity	-0.35	-0.37
	Fluorescence	0.10	0.13
Calculated	Phosphate	-0.71	-0.66
	Nitrate	-0.75	-0.70
	Silicate	-0.35	-0.31
	Total CO ₂	-0.82	-0.77
	Total Alkalinity	-0.85	-0.81
	HCO ₃ ⁻ Glodap-v2	-0.68	-0.66
	HCO ₃ ⁻ CO2Sys st	-0.81	-0.75
	pH Glodap-v2	-0.25	-0.22
	pH CO2Sys	-0.70	-0.69
	CO ₃ ²⁻ Glodap-v2	0.09	0.08
	CO ₃ ²⁻ CO2Sys	0.40	0.32
	Ωca Glodap-v2	0.10	0.10
	Ωca CO2Sys	0.39	0.31

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Emiliana huxleyi type A group



Emiliana huxleyi type B group

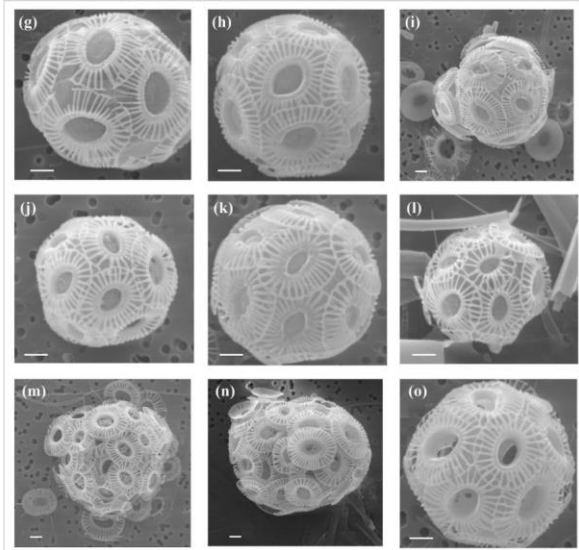


Plate 1. Specimens of *Emiliana huxleyi*. Group A includes (a) type R, sample PS97/018-1 at 10 m water depth; (b) type A overcalcified, PS97/018-1 at 60 m; (c) type A overcalcified, PS97/018-1 at 100 m; (d) type A calcified, PS97/018-1 at 60 m; (e) type A calcified PS97/018-1 at 20 m; (f) type A slightly calcified, PS97/018-1 at 60 m.

Emiliana huxleyi group B includes: (g) type B PS97/016-1 at 5 m; (h) type B PS97/029-1 at 60 m; (i) type B calcified, PS97/29-1 at 150 m; (j) type B/C PS97/016-1 at 10 m; (k) type B/C slightly calcified PS97/030-1 at 150 m; (l) type C PS97/043-1, at 10 m; (m) type O -opened-, PS97/032-1 at 10 m; (n) type O -lamella-, PS97/038-1 at 10 m; (o) type O -lamella- PS97/017-1 at 60 m. The scale bar indicates 1 μ m.

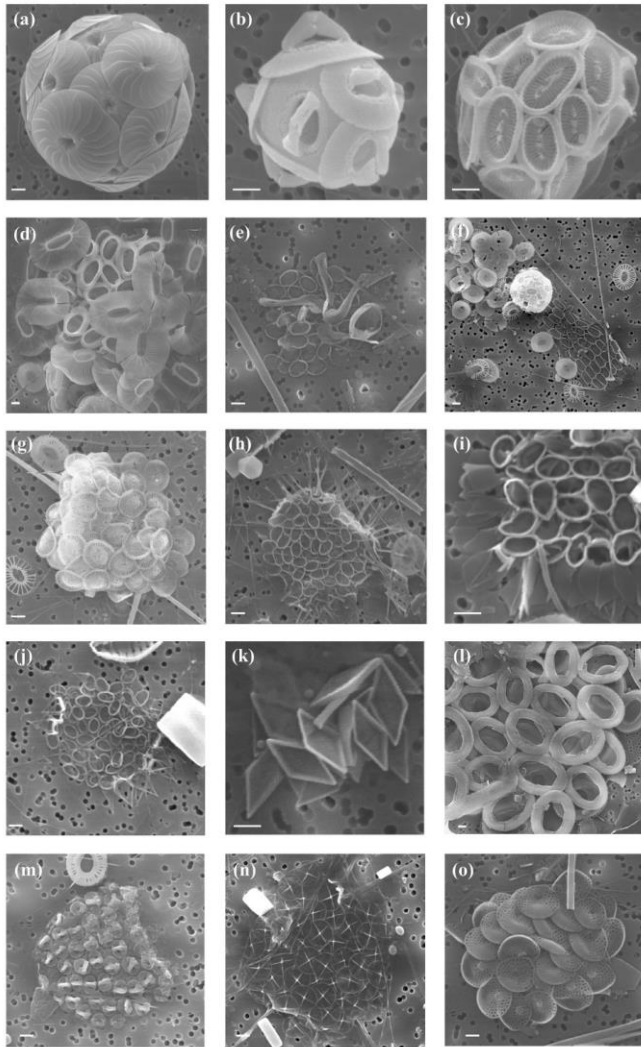


Plate 2. Other taxa present in the study area. Specimens of (a) *Calcidiscus leptoporus* s.s., sample PS97/016-1 at 5 m water depth; (b) *Gephyrocapsa muelleriae*, PS97/018-1 at 10 m; (c) *Syracosphaera dilatata*, PS97/32-1 at 10 m; (d) *Syracosphaera corolla*, PS97/38-1 at 10 m; (e) *Ophiaster hydroideus*, PS97/38-1 at 10 m; (f) *Calciopappus caudatus* and *Emiliana huxleyi*, PS97/34-2 at 10 m; (g) *Acanthoica quattrosipina*, PS97/32-1 at 10 m; (h) *Papposphaera sagittifera* sp., PS97/38-1 at 20 m; (i) *Papposphaera* cf. *heldalii*, PS97/30-1 at 40 m; (j) *Wigwamma antarctica*, PS97/39-1 at 10 m; (k) *Calciosolenia* sp., PS97/38-1 at 10 m; (l) *Chrysotila* sp., PS97/35-1 at 100 m; (m) *Syracosphaera strigilis* HOL, PS97/17-1 at 40 m; (n) *Chrysochromulina* sp.; (o) *Petasaria heterolepis* sp., PS97/38-1 at 40 m. The scale bar indicates 1 μ m.