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# Physical constraints for respiration in microbial hotspots in soil and their importance for denitrification

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10 **Abstract** Soil denitrification is the most important terrestrial process returning reactive nitrogen to the atmosphere, but remains poorly understood. In upland soils, denitrification occurs in

hotspots of enhanced microbial activity, even under well-aerated conditions, and causes harmful

emissions of nitric (NO) and nitrous oxide (N2O). Timing and magnitude of such emissions are

difficult to predict due to the delicate balance of oxygen (O2) consumption and diffusion in soil.

15 To study how spatial distribution of hotspots affects O<sub>2</sub> exchange and denitrification, we

embedded porous glass beads inoculated with either Agrobacterium tumefaciens (a denitrifier

lacking N<sub>2</sub>O reductase) or *Paracoccus denitrificans* (a "complete" denitrifier) in different

architectures (random vs. layered) in sterile sand adjusted to different water saturations (30%,

60%, 90%) and measured gas kinetics (O2, CO2, NO, N2O and N2) at high temporal resolution.

Air connectivity, air distance and air tortuosity were determined by X-ray tomography after the

experiment. The hotspot architecture exerted strong control on microbial growth and timing of

denitrification at low and intermediate saturations, because the separation distance between the

microbial hotspots governed local oxygen supply. Electron flow diverted to denitrification in

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anoxic hotspot centers was low (2-7%) but increased markedly (17-27%) at high water saturation.

X-ray analysis revealed that the air phase around most of the hotspots remained connected to the

headspace even at 90% saturation, suggesting that the threshold response of denitrification to soil

moisture could be ascribed solely to increasing tortuosity of air-filled pores. Our findings suggest

that denitrification and its gaseous product stoichiometry do not only depend on the amount of

microbial hotspots in aerated soil, but also on their spatial distribution. We demonstrate that

combining measurements of microbial activity with quantitative analysis of diffusion lengths

using X-ray tomography provides unprecedented insights into physical constraints regulating soil

microbial respiration in general and denitrification in particular. This opens new avenues to use

observable soil structural attributes to predict denitrification and to parameterize models. Further

experiments with natural soil structure, carbon substrates and microbial communities are required

35 to demonstrate this under realistic conditions.

1. Introduction

Soil carbon and nitrogen turnover is governed by soil heterogeneity at the microscale. Much of

the turnover is concentrated in microsites, providing favorable conditions (pO2, temperature, pH)

and substrates (carbon, nutrients) for soil microbial activity. The partitioning of aerobic and

anaerobic respiration in microsites is largely controlled by the water content in the soil matrix

which defines the scale across which O2 diffuses towards microsites of high O2-consuming

activity. Aqueous diffusion lengths range from distances across thin water films in well-aerated

soils, to individual soil aggregates of different radii at field capacity, up to the distance to the soil

surface when the soil is saturated (Smith et al., 2003; Elberling et al., 2011; Ball, 2013; Parkin,

1987). Aerobic respiration is less affected by soil moisture than anaerobic respiration and

typically peaks around water saturations of 20-60% in forest, grass and cropland soils (Schaufler

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et al., 2010; Ruser et al., 2006; Moyano et al., 2012). Bulk soil respiration starts to decline at

higher saturations due to the development of anoxic microsites with lower redox potential,

supporting carbon mineralization at typically only a tenth of the rates observed under oxic

conditions (Keiluweit et al., 2017). Denitrification, i.e. the dissimilatory respiration of N

oxyanions instead of oxygen, is commonly observed at water saturations above 60-70% and

peaks beyond 90% (Ruser et al., 2006; Linn and Doran, 1984). The occurrence of anaerobic

microsites is governed by the balance between saturation-dependent diffusion and microbial

consumption of O2, which in turn depends on the quantity, quality and distribution of soil organic

matter in the soil matrix terms and environmental factors like temperature and pH, which control

microbial activity (Tecon and Or, 2017; Nunan, 2017; Smith et al., 2003). In fact, water films

around decaying plant material may suffice to induce anaerobic respiration, if microbial

respiration exceeds O<sub>2</sub> diffusion through that minute barrier (Parkin, 1987; Kravchenko et al.,

2017).

60 The interplay between physical constraints and biological activity in soil controls microbial

respiration at microscopic scales and complicates the prediction of denitrification and N-gas

fluxes at larger scales. For instance, nitrous oxide (N<sub>2</sub>O) emissions show notoriously large spatial

variability, which has been attributed to heterogeneous distribution of anoxic microsites in the

soil (Mathieu et al., 2006; Röver et al., 1999; Parkin, 1987; Parry et al., 1999). Together with the

often observed high temporal variability of microbial respiration and its fluctuations under

transient conditions, this has led to the notion of "hotspots" and "hot moments" for microbial

activity and emissions (Groffman et al., 2009; Kuzyakov and Blagodatskaya, 2015). "Hotspots"

of denitrification have traditionally been linked to diffusion constraints in soil aggregates. Cell

numbers and O2 concentration have been shown to decline exponentially towards aggregate

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70 centers (Sexstone et al., 1985; Horn et al., 1994; Zausig et al., 1993; Højberg et al., 1994) and the

critical aggregate radius for the development of anoxic centers and is typically >1 mm (Sierra and

Renault, 1996; Højberg et al., 1994; Schlüter et al., 2018), However, anoxic microsites have also

been reported for smaller aggregates (equivalent diameter of 0.03-0.13 mm) in well-aerated,

repacked soils (Keiluweit et al., 2018).

75 An important, but often neglected aspect of physical diffusion constraints on microbial

respiration is the spatial distribution of microbial hotspots within the soil matrix. Incubation

experiments were either designed to control the aggregate size in repacked soil (Mangalassery et

al., 2013; Miller et al., 2009) or the volume fraction of sieved soil mixed evenly into sterile quartz

sand (Keiluweit et al., 2018). Some incubation studies were carried out with undisturbed soil and

investigated diffusion constraints within the pore network (Rabot et al., 2015). However, these

studies did not address the location of hotspots nor the diffusion lengths towards air-filled pores.

The vast majority of incubations studies merely reports bulk soil properties like carbon and

nitrogen content, bulk density and water saturation. Notable exceptions are Kravchenko et al.

(2017) who controlled the position of microbial hotspots by placing decaying plant leaf material

into repacked soils with different aggregate sizes and water saturations and Ebrahimi and Or

(2018), who placed several layers of remolded aggregates as artificial hotspots into a sand matrix

and controlled the volume fraction of anaerobic and aerobic respiration by adjusting the water

table in the sand column. Such systematic studies with simplified soil analogues, yet fully

accounting for transport processes from and towards hotspots, including interactions between

hotspots, are needed to improve our understanding about how physical constraints on microbial

respiration control the anaerobic soil volume and transient denitrification activity.

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The objective of the present study was to study the interplay between microbial activity and

physical diffusion in controlling aerobic and anaerobic respiration for different spatial

distributions of hotspots. We embedded uniform artificial hotspots inoculated with denitrifying

pure cultures (Schlüter et al., 2018) in sterile sand, which was adjusted to different water

saturations. We hypothesized that the competition for oxygen would depend on the separation

distance between the hotspots, which in turn would control microbial cell growth and O2

consumption and thus affect the timing of the aerobe-anaerobe transition in respiration, i.e. the

onset of denitrification. Further, by placing hotspots inoculated with complete (P. denitrificans)

and truncated (A. tumefaciens) denitrifiers in distinct horizontal layers, we expected to see

interactions with respect to overall N2O turnover. To capture the highly dynamic respiration

kinetics, we monitored O2, CO2, NO, N2O and N2 exchange between the headspace and the sand-

hotspot matrix at high temporal resolution. The morphology of the air-filled pore space in terms

of air connectivity, air tortuosity and air distance was determined by X-ray computed tomography

after the experiment.

2. Material and methods

2.1. Microbial hotspots

Two facultative anaerobic bacteria were used in this study. Paracoccus denitrificans expresses all

denitrification enzymes necessary to reduce NO<sub>3</sub><sup>-</sup> to N<sub>2</sub>, whereas Agrobacterium tumefaciens

lacks the gene nosZ encoding nitrous oxide reductase (N2OR), which makes N2O the final

denitrification product. Moreover, the two strains differ in their regulatory phenotypes with

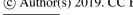
respect to inducing denitrification in response to oxygen depletion, which leads to characteristic

patterns of product accumulation (Bergaust et al., 2011). P. denitrificans induces NO and N2O

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reductase early during O<sub>2</sub> depletion (Bergaust et al., 2010), thus releasing little N<sub>2</sub>O. By contrast,

A. tumefaciens is known to be less stringent in controlling intermediates which may result in the

release of large amounts of NO, up to cell-toxic, milli-molar concentrations (Bergaust et al.,

2008). Both strains were grown in Sistrom's medium (Sistrom, 1960) as described in a previous

study (Schlüter et al., 2018), but at double strength to provide enough substrate for depleting O<sub>2</sub>

during aerobic growth. The medium was amended with 10 mM NH<sub>4</sub>NO<sub>3</sub> and 5 mM KNO<sub>3</sub> for

anaerobic growth. To produce microbial hotspots, porous borosilicate glass beads (VitraPOR

P100, ROBU Glasfilter Geräte GmbH) with a diameter of 7 mm, a porosity of 32% and a

medium pore diameter of 60  $\mu$ m were saturated with freshly inoculated growth medium ( $\approx 10^8$ 

cells ml<sup>-1</sup>) by submersion into one of the two cultures. In the following, the inoculated porous

glass beads are referred to as At- (A. tumefaciens) and Pd- (P. denitrificans) hotspots. Detailed

information about the culture conditions and the inoculation procedure can be found in Schlüter

et al. (2018).

2.2. Repacked sand

Fifty At and Pd hotspots each were placed into 120 ml of washed, sterile quartz sand (0.2-0.5 mm

grain size) yielding a volume fraction of 14% (20 ml; Fig. S1a). The sand was packed into 240

ml glass jars (Ball Corporation, Bloomfield, CA) in portions of 10 ml layers and adjusted to

target saturation by adding sterile water with a spray can. The packing procedure resulted in some

minor changes in porosity between layers and some larger gaps around the hotspots (Fig. S3)

which affected air distribution in the sand (Fig. S2a). Three saturations were used, corresponding

to water-filled pore spaces (WFPS) of 30, 60 and 90%. The fully saturated hotspots were placed

into the sand at three different architectures (Figure 1). For the "random" distribution, the

hotspots were placed in five equidistant (~9.8 mm, center to center) horizontal layers with a

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random distribution of ten At and ten Pd hotspots per layer. For the "layered At/Pd" and "layered

Pd/At" distributions, all fifty hotspots of each strain were placed into one of two horizontal layers

spaced 21 mm from each other (center to center) at an average headspace distance of 18.2 and

39.2 mm, respectively, where the order represents top/bottom. Care was taken to keep the

hotspots cool (on crushed ice) during the packing procedure. The pore size distribution of the

porous hotspots and the sand in the bulk soil and in hotspot vicinity are reported in Figure S3.

2.3. **Incubation** 

To establish aerobic and anaerobic growth patterns and denitrification kinetics for both bacterial

strains when growing inside the glass beads, a pre-experiment was conducted without sand. Fifty

Pd or At hotspots were placed in empty 120 ml serum bottles (Fig. S1b) and incubated at 15°C

under either oxic (He/O<sub>2</sub> 80/20%) or anoxic (He 100%) conditions in two replicates per

treatment. Headspace concentrations of O2, CO2, NO, N2O and N2 were measured every 4 h by

piercing the septum with a hypodermic needle mounted to the robotic arm of an autosampler

(GC-PAL, CTC Analytics, Switzerland). The autosampler was connected to a gas chromatograph

(Agilent Model 7890A, Santa Clara, CA, USA) and a NO analyzer (Teledyne 200. San Diego,

CA, USA) via a peristaltic pump. Detailed information about the robotized incubation system and

the experimental setup can be retrieved elsewhere (Molstad et al., 2007; Schlüter et al., 2018).

In the main experiment, freshly inoculated glass beads were packed into incubation vessels as

described above, three replicates for each of the nine combinations of saturation and hotspot

distribution. Jars with 30% and 60% WFPS were flushed with He/O<sub>2</sub> for 40 min, using ten cycles

of vacuum (3 min) and purging (1 min). Jars with 90% WFPS were flushed using 180 cycles of

mild vacuum (~ 600 mbar) and O<sub>2</sub>/He purging to avoid structural changes of the packed columns

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due to bubbling of trapped gas. The jars were then placed into a water bath kept at 15°C and after temperature equilibration  $O_2$ /He overpressure was released. Gas concentrations in the headspace were analyzed as described above. Gas production and consumption kinetics were used to calculate the fraction of electrons diverted to  $O_2$  or N oxyanions and thus to estimate the contribution of denitrification to total respiration ( $e_{denit}^{-}/e_{total}^{-}$ ) (Schlüter et al., 2018; Bergaust et al., 2011). The NO/(NO+  $N_2$ O +  $N_2$ ) and  $N_2$ O/(NO+  $N_2$ O +  $N_2$ ) product ratios were estimated from the cumulative release of gaseous denitrification products (NO,  $N_2$ O,  $N_2$ ), after subtracting precursors from products (NO from  $N_2$ O +  $N_2$  and NO +  $N_2$ O from  $N_2$ ). The rationale behind the latter was to mimic an open system, in which N-gases released to the atmosphere are not available any longer as electron acceptors for denitrification. Details about the calculation of denitrification product ratios can be found in the Supporting Information (SI 1.2).

## 2.4. X-ray tomography and image analysis

After the incubation experiment, the glass jars were scanned with X-ray micro-tomography (X-tek XCT 225, Nikon Metrology) with a beam energy of 145 kV, a beam current of 280  $\mu$ A, an exposure time of 708 ms per frame, a 0.5 mm copper filter for reducing beam hardening artefacts and a total of 3000 projection for a full scan. Individual hotspots were also scanned (100 kV, 90 $\mu$ A, 1000ms per frame, no filter) to analyze the internal pore morphology. The 2D projections were reconstructed into a 3D image with a resolution of 35  $\mu$ m using a filtered-back projection algorithm in the X-tek CT Pro 3D software. Image processing from raw gray-scale data (**Figure 1a**) to segmented data including sand grains, air and water (**Figure 1b-c**) was carried out according to well-established protocols for multi-phase segmentation (Schlüter et al., 2014). The porous glass beads were assigned to At or Pd hotspots according to the orientation of the flat end in the random architecture or by the vertical position in the layered architecture (**Figure 1b-c**).

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The segmented images were analyzed with respect to three different spatial attributes of the air-

filled pore spaces deemed important for oxygen supply. 1. Air connectivity by distinguishing

isolated air-filled pores and air-filled pores with a continuous path to the headspace (yellow and

red in Figure 1d). Air connectivity is then defined as the ratio of connected air-filled pore space

and total air-filled pore space 2. Air tortuosity as derived from the geodesic length of connected

air-filled pores. The geodesic length is the distance of any connected air voxel to the headspace

along curved paths around obstacles like solid particles and water-blocked pores (Fig. 1e). Air

tortuosity is the ratio between geodesic and vertical Euclidean distance to the headspace averaged

over all connected, air-filled voxels. It is a proxy for the diffusive transport of gaseous oxygen in

air-filled pores 3. Air distances of water-filled pores as defined by the average geodesic distance

from any water voxel to the closest air-filled pore with headspace connection (white in Fig. 1f).

Air distance is a proxy for the slow diffusive transport of dissolved oxygen. All image processing

steps were carried out with Fiji/ImageJ (Schindelin et al., 2012) and associated plugins (Legland

et al., 2016; Doube et al., 2010) or with VG Studio Max 2.1 (Volume Graphics). Each image

processing and analysis step is explained in detail in the supporting information (SI 1.3).

[Figure 1]

### 3. Results

# 3.1. Aerobic respiration and denitrification in unconstrained

# hotspots without sand

At grew faster than Pd at 15°C in the experiment with loosely placed porous glass beads as

indicated by faster O<sub>2</sub> consumption and CO<sub>2</sub> accumulation in the oxic treatment (Figure 2a,b).

Also under fully anoxic conditions, At accumulated CO<sub>2</sub> faster than Pd (Figure 2b). N-gas

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kinetics clearly reflected the distinct regulatory phenotypes of the two bacterial denitrifiers.

Anoxic At instantly accumulated a large amounts of NO (Figure 2c) which persisted until all

 $NO_3$  was reduced to  $N_2O$  (as judged from the stable plateau in  $N_2O$ , **Figure 2d**). Due to slower

growth and  $O_2$  consumption, Pd induced denitrification much later than At, but accumulated less

intermediates (NO, N2O) than At. Oxically incubated Pd accumulated no detectable NO,

indicating efficient regulation of denitrification when switched slowly to anaerobic conditions in

hotspots. Also, NO may have been reduced to N<sub>2</sub>O when diffusing from the anoxic center to the

boundary of the hotspot. In the initially oxic treatments, denitrification contributed 7% to the total

electron flow in At hotspots and 13% in Pd hotspots, reflecting the fact that (i) Pd has one more

reduction step in the denitrification sequence and that (ii) At used less nitrate for anaerobic

respiration in anoxic hotspots centers and more oxygen for aerobic respiration in oxic hotspots

215 margins than Pd.

[Figure 2]

3.2. Effects of hotspot distribution in sand

The distribution of microbial hotspots within the sand strongly impacted bulk respiration. This is

evident for treatments with medium saturation (60% WFPS) for the first 210 h of incubation

(Figure 3) and with other saturations for the entire incubation period (300 h; Figures S4-6). The

random distribution of hotspots allowed for much faster aerobic growth than the layered

architectures, leading to complete consumption of O<sub>2</sub> from the jars after 70 h (**Figure 3a**). Given

the slow growth of Pd (Figure 2a), initial  $O_2$  consumption was dominated by the activity of At

hotspots turning them partly anoxic. Hence, the pronounced NO peak in the random treatment,

coinciding with complete  $O_2$  exhaustion from the headspace (**Figure 3c**), was due to At activity,

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similar to what was seen in the unconstrained At hotspots under anoxic conditions (Figure 2c).

N<sub>2</sub>O production was observed long before O<sub>2</sub> was depleted from the headspace (Figure 3d) and

is attributed entirely to At denitrification. Pd denitrification did not start before all O2 was

depleted and manifested itself in a transient increase in N<sub>2</sub>O production at ~70 h together with an

exponential increase in N<sub>2</sub> production (Figure 3e) which was also observed with unconstrained

Pd hotspots (Figure 2e). Note that the apparent net consumption of CO<sub>2</sub> (Figure 3b) upon O<sub>2</sub>

depletion was due to internal alkalization driven by accelerating denitrification, once all hotspots

turned anoxic.

[Figure 3]

235 In the layered architectures, O2 consumption was slower and complete anoxia was not reached

before 120 h into the incubation. In contrast to the random architecture, less O<sub>2</sub> was available for

each individual hotspot in the densely packed hotspot layers, allowing for less aerobic growth per

unit time. As a consequence, there was more time for fully denitrifying At hotspots to interact

with Pd hotspots which induced denitrification gradually between 80 and 120 h. Indeed, less  $N_2O$ 

accumulated in the headspace than in the random treatment (Figure 3d, S6d) and the onset of N<sub>2</sub>

accumulation appeared long before complete O<sub>2</sub> depletion from the headspace (Figure 3a,e). In

other words, Pd hotspots consumed N2O produced in At hotspots. Upon O2 depletion in the

headspace, a burst of NO production occurred (Figure 3c) as seen previously with At hotspots

(Figure 2c). However, since Pd denitrification was now fully developed, the NO peak was much

more short-lived than with the random distribution, because Pd hotspots reduced NO produced

by At hotspots all the way to  $N_2$ .

The effect of vertical order in the layered hotspot architecture was small, but consistent among all

denitrification products. The distribution with Pd hotspots on top (layered Pd/At) consumed the

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NO and  $N_2O$  produced in At hotspots much quicker than the At/Pd architecture (**Figure 3c-d**) and

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accumulated N<sub>2</sub> faster after complete O<sub>2</sub> depletion (Figure 3e). Both observations highlight the

effect of shorter diffusion distances between the headspace and the Pd hotspot layer in the

layered *Pd/At* architecture.

3.3. Effects of matrix saturation

Differences in water saturation resulted in different absolute amounts of oxygen initially present

in the jars (Figure 4a) but did not affect the O2 concentration in the sand matrix. Oxygen was

depleted slightly faster at 60% than at 90% saturation even though there was absolutely more O2

initially present at 60% WFPS. This illustrates the paramount role of oxic growth for the oxic-

anoxic transition in the hotspots: the more O2 available initially, the stronger the aerobic growth

and the faster the oxic-anoxic transition.

Increasing saturation from 60 to 90% in the randomly distributed hotspots had a strong effect on

the timing and accumulation of denitrification products. The expected NO burst upon O<sub>2</sub>

depletion was damped by two orders of magnitude (Figure 4c), because the oxic-anoxic

transition proceeded more smoothly in the 90% treatment and NO was reduced further to  $N_2O$ 

before it could escape to the headspace. On the other hand, N<sub>2</sub>O and N<sub>2</sub> production commenced

earlier in the 90% than in the 60% treatment (Figure 4d-e), indicating that O<sub>2</sub> availability was a

priori smaller irrespective of metabolic activity (which was larger in the 60% treatment). The

switch from net  $N_2O$  production to net  $N_2O$  consumption indicates the moment when microbial

activity in Pd hotspots caught up with At hotspots.

[Figure 4]

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270 Surprisingly, O<sub>2</sub> consumption in the 30% treatments was slow despite having the largest amount of O<sub>2</sub> in the jar. This was caused by unintended substrate limitation. Due to overlapping pore size distribution between porous hotspots and sand (Fig. S3c), medium was sucked by capillary force from the hotspot into the surrounding sand, as could be seen in a parallel experiment with brilliant blue dye (Fig. S7). This separated cells, which were likely immobilized in the pore space 275 of the hotspots, temporarily from a considerable fraction of the carbon and NO<sub>3</sub> supplied with the medium, before the dissolved substrate would diffuse back into the hotspots due to the evolving gradient induced by consumption in the hotspots. Decreasing the saturation from 60% to 30% also resulted in different timing and accumulation of denitrification products. The slow oxic growth of both At and Pd hotspots due to the substrate diffusion limitation at 30% WFPS 280 provided more time for Pd hotspots to interact with At hotspots than in the 60% WFPS treatment. Indeed, the NO burst from At hotspots after complete O2 exhaustion in the random architecture was 50% higher at 30% WFPS indicating higher At cell numbers due to prolonged oxic growth (Figure 4c, Figure 5c), whereas the N<sub>2</sub>O peak was 50% lower, due to concomitant N<sub>2</sub>O reduction in Pt hotspots (Figure 4d, Figure 5d).

#### 3.4. Mass balances

By the end of the incubation, oxygen was exhausted in all treatments. Likewise, NO<sub>3</sub> was consumed by all treatments, except for the layered hotspots at 30% and 60% WFPS. This means that respiration was electron acceptor limited and that the cumulated recovery of denitrification products can be compared with the amount of NO<sub>3</sub> initially present (Figure S8). The balance between aerobic and anaerobic respiration,  $e_{denit}^{-}/e_{total}^{-}$  (Bergaust et al., 2011), is given by the electron flow to nitrogenous electron acceptors relative to the total electron flow, including O<sub>2</sub> respiration (Figure 5). When seen over all three water saturations, early stage denitrification

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under oxic headspace conditions (Figure 5a) showed a threshold response to increasing moisture

with disproportionally higher  $e^-_{denit}/e^-_{total}$  ratios at 90% WFPS (17-27%) than at 60% or 30%.

The proportions of electrons diverted to denitrification at low and medium saturations were small

(2-7%) and even smaller than those observed in unconstrained hotspots (7-13%). Differences

between saturations were less pronounced when the entire incubation period is considered

(Figure 5b), since fully anoxic conditions during late stage incubation overrode saturation

effects. Overall, the effect of hotspot architecture on  $e_{denit}^{-}/e_{total}^{-}$  ratios was smaller than the

effect of saturation.

This stands in stark contrast to the pronounced effect of hotspot architecture on denitrification

product ratios (Figure 5c, d). Hotspot architecture governed growth rates through local

competition for  $O_2$  and therewith the number of active cells involved in net production sites (At

hotspots) and net consumption sites (Pd hotspots) of NO once O2 was exhausted. In layered

hotspot architectures there was hardly any net-release of NO to the headspace irrespective of

saturation (Figure 5c). With random hotspot architecture, there was substantial NO release, the

magnitude of which, however, decreased linearly with saturation. This pattern in NO

stoichiometry clearly reflects the number of At cells at the moment of complete O<sub>2</sub> depletion, as

affected by oxic growth which lasted longer with lower saturation. The N2O product ratio

(Figure 5d) was influenced by both saturation and hotspot architecture. In layered architectures,

the N<sub>2</sub>O ratio increased exponentially with increasing saturation similar to what was observed for

relative electron flow to denitrification (Figure 5a). In random architectures, the N<sub>2</sub>O product

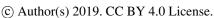
ratio was consistently higher than in layered architectures irrespective of saturation, yet the

highest ratio was reached at 60% WFPS, due to the most vigorous growth, and hence fastest oxic-

anoxic transition at intermediate saturation.

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[Figure 5]

# 3.5. Pore space properties

At the lowest saturation (30% WFPS), the entire air-filled pore space was connected to the headspace (Figure 6a) and tortuosity was close to unity, i.e. the diffusion lengths in air only depended on the vertical distance to the headspace (Figure 6b). The diffusion distances in waterfilled pores (Figure 6c) corresponded to the size of small, evenly distributed water clusters. At medium saturation (60% WFPS), the amount of disconnected air was still negligible and tortuosity only slightly increased. The increase in air distance was due to a few large water pockets, which were caused by the step-wise addition of water to the repacked sand. Only at 90% saturation a considerable air volume of 5-20% became disconnected from the headspace. The path along which the remaining air was connected to the headspace became more tortuous with increasing saturation and average diffusion distances in water to the connected air cluster increased to 1 mm. This is still surprisingly short as compared to the size of the hotspots (7 mm). Independent tests showed that the high air connectivity at this low air content was facilitated by vacuum application during He/O<sub>2</sub>-purging prior to the incubation. Directly after packing, the continuous air cluster only reached 10-15 mm into the sand (data not shown), whereas bubbling due to vacuum application formed continuous air channels that reached deep into the sand matrix connecting even the deepest hotspots with the headspace. Moreover, some larger gaps remained around hotspots during packing which tended to be air-filled after wetting. This is reflected in the consistently higher air-connectivity, lower air tortuosity and lower air distance, when only pores in the direct vicinity of hotspots are analyzed (Figure 6a-c). More than 90% of hotspot surfaces still had a direct air-filled connection with the headspace at 90% WFPS (Figure 6a). Depth profiles of these pore space attributes are reported in Fig. S2.

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[Figure 6]

### **4. Discussion**

# 4.1. Physical constraints on denitrification kinetics

The experimental setup in this incubation study was designed to investigate physical constraints on microbial respiration in hotspots as affected by the interplay between gaseous diffusion through a sterile matrix and local competition for oxygen. For this, we compared different combinations of water saturation in the matrix and spatial distributions of hotspots. The setup is a coarse simplification of soil in which metabolic activity in hotspots not only depends on oxygen supply, but also on diffusion of substrates from the matrix to the hotspots. As such, our experiment does not allow to draw direct conclusions about the functioning of hotspots in real soils with respect to denitrification and its product stoichiometry. However, by placing denitrifiers and their substrates into hotspots, we considerably reduced the level of complexity and created a system that is amenable to studying the dynamic interrelations between denitrifier growth, oxygen consumption and induction of denitrification by gas kinetics. Soil N<sub>2</sub>O emissions are known to be highly variable in time and a unifying concept incorporating dynamic changes in denitrification activity and product stoichiometry in response to changing environmental conditions is still missing. Our model system provides a first data set for validating mathematical process models that are explicit for structural distribution of hotspots and dynamic changes in boundary conditions (here mimicked by different hotspot architectures and declining oxygen concentrations in the headspace of batch incubations, respectively). The development of such models is a core activity of the DASIM project (http://www.dasim.net/). By combining metabolic

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360 measurements with advanced structural imaging and computation, we also provide a link to

parameterizing such models with real soil data in future research.

Inoculating growing denitrifiers into porous glass beads and embedding them in sterile sand

resulted in a highly dynamic system with respect to oxygen consumption and induction of

denitrification. This was intended for the sake of experimental depth, but it must be noted that

oxic-anoxic transitions are likely slower, i.e. less dynamic in real-soil hotspots. In real soils, even

highly organic hotspots contain a fair amount of recalcitrant organic C that limits microbial

growth and oxygen consumption. Also with respect to denitrification stoichiometry, real soils

may be expected to be less dynamic as multiple denitrifying phenotypes contained in the natural

soil microbiome (Roco et al., 2017) utilize denitrification intermediates mutually.

Notwithstanding, soil NO and N<sub>2</sub>O emissions are known to be episodic in nature. Large,

denitrification driven emission pulses occur upon abrupt changes in O2 availability, caused by

external factors like heavy rainfalls or soil freezing (Flessa et al., 1995), O<sub>2</sub> consumption by

nitrification after ammoniacal fertilization (Huang et al., 2014) or incorporation of easily

degradable organic matter (Flessa et al., 1995) which cannot be captured satisfactorily by

common steady-state models for soil respiration and N<sub>2</sub>O emission (Parton et al., 2001; Li et al.,

1992). Even though the concept of hotspots is central in the understanding of denitrification

dynamics in upland soils, common soil denitrification models do not account for the dynamics of

spatially explicit hotspots in the soil matrix but rather scale bulk denitrification with a generic

anoxic volume fraction (Li et al., 2000; Blagodatsky et al., 2011). To advance soil denitrification

models, it is obvious that microbial respiration dynamics in hotspots have to be targeted, both

conceptually (Wang et al., 2019) and experimentally (Kravchenko et al., 2017; Ebrahimi and Or,

2018). Our study is a first step in this direction.

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One of the main findings of this study is that soil microbial respiration and the propensity to

develop denitrifying anoxic hotspots does depend on their distribution in space. The onset of

denitrification and its kinetics was linked to the spatial and temporal extent of anoxia developing

in hotspot centers, which was governed by the interplay between denitrifier growth and diffusion

constraints and hotspot architecture had a strong impact on this interplay. When distributed

randomly, microbial activity was most disperse relative to available oxygen, resulting in more

growth, faster O<sub>2</sub> draw down and earlier anoxia than when packed densely in layers (Figure 3).

Rapid oxic-anoxic transition led to higher release rates of denitrification intermediates increasing

the product ratios of NO and  $N_2O$  (Figure 5c-d). This effect was most pronounced at low and

intermediate saturations but was dampened at 90%WPFS because oxygen supply was impeded

by bulk diffusion irrespective of hotspot placement. Thus, our results highlight the significance of

hotspot distribution at low soil moistures and exemplifies why N<sub>2</sub>O emissions are notoriously

395 difficult to predict under these conditions.

Even though we failed to fully synchronize At and Pd growth in time, our experiment

demonstrates that contrasting denitrification phenotypes may interact in modulating N<sub>2</sub>O flux to

the atmosphere. Pd hotspots reduced N<sub>2</sub>O released from At hotspots irrespective of the layers'

orientation (Figure 3d), which can be attributed to the high degree of air connectivity in the sand

column (**Figure 1d**). We had expected more  $N_2O$  reduction with Pd on top (layered Pd/At), but

since At grew faster than Pd, partial anoxia and NO and  $N_2O$  formation was induced in At, long

before N<sub>2</sub>O consuming activity was induced in Pd hotspots. Future experiments with artificial

hotspots should therefore carefully consider potential growth rates and air connectivity in packed

soil.

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4.2. Physical constraints on cumulative denitrification

The cumulative release of gaseous denitrification products, as described by electron flow ratios, depended less on hotspot architecture than on soil moisture. Electron flows to denitrification ranged from <5% of total respiratory flow at low to medium saturations (30, 60% WFPS) to almost 23% at 90% WFPS (Figure 5a). We attribute this low denitrification electron flow to the small active volume relative to the sterile sand matrix (the total volume fraction of hotspots was 14%, less of which was actually anoxic) and the large amount of oxygen initially present in the incubation jars. Yet, we found a typical, non-linear denitrification response to soil moisture (Figure 5a). This threshold behavior is well known (Weier et al., 1993) and has been attributed to a disproportional contribution of small pores to the anoxic volume at higher saturation (Schurgers et al., 2006). In our system, consisting of coarse sand with a relatively homogenous pore size distribution, we attribute the non-linear response to an increase in tortuosity of air-filled pores that was pronounced enough to impair the supply of hotspots with oxygen. Air connectivity and distance to the next continuous, air-filled pore also increased non-linearly, but did not reach a critical value (Figure 6), ruling out that differences in NO and N2O release at different saturations were due to gas entrapment but rather due to elongated diffusion pathways in airfilled pore networks, leading to longer residence times of denitrification intermediates and stronger reduction of intermediates in hotspots along the way to the headspace. Saturationdependent threshold behavior for denitrification is a well-studied phenomenon in soils (Linn and Doran, 1984; Ruser et al., 2006; Paul et al., 2003), but for a lack of pore scale measurements often attributed to reduced bulk soil diffusivity. In undisturbed soil, the relative importance of air connectivity and distances between air-filled and water-filled pores might be more relevant for impairing oxygen supply and inducing denitrification. Air connectivity to the headspace was shown to affect N<sub>2</sub>O emissions in terms of intensity and speed in repeated wetting/drying cycles

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in an intact soil column (Rabot et al., 2015). In agricultural soil with different crop rotations, N<sub>2</sub>O emissions were shown to correlate positively with the volume fraction of soil with macropore

distances larger than 180 µm, used as an ad-hoc definition for poorly aerated soil (Kravchenko et

al., 2018). In a mesocosm study on microstructural drivers for local redox conditions, none of the

investigated soil pore metrics derived from X-ray CT data (excluding those examined here)

correlated with redox kinetics during a wetting/drying cycle (Wanzek et al., 2018). Hence,

combining metabolic monitoring by high-resolution gas kinetics with direct assessment of

diffusion lengths of gaseous and dissolved oxygen and denitrification products via X-ray

microtomography emerges as a promising tool to study physical constraints for aerobic and

anaerobic respiration in soil. However, meaningful metrics derived from X-ray data relevant for

denitrification are yet to be developed and will require additional experiments with both artificial

and real soils. Improved understanding of factors and mechanisms controlling denitrification and

N gas emission on a three-dimensional micro-scale may help to design and test soil management

strategies that mediate the return of excess nitrogen to the atmosphere in a controlled way, i.e.

with as little as possible NO and N<sub>2</sub>O release, be it by crop residue (Kravchenko et al., 2017), pH

(Russenes et al., 2016) or irrigation (Bergstermann et al., 2011) management. At the same time,

our experiments call for the implementation of spatially explicit reaction-diffusion algorithms

(Hron et al., 2015; Ebrahimi and Or, 2016) in soil process models. For instance, diffusion lengths

between hotspots and air-filled pores connected to the headspace may serve as useful measure to

parametrize model concepts like the anaerobic soil volume fraction in larger-scale continuum

models (Li et al., 2000; Schurgers et al., 2006; Blagodatsky et al., 2011).

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5. Conclusions

Using a highly simplified model system, we demonstrate that the factorial combination of water

saturation and hotspot architecture creates a wealth of denitrification kinetics in response to

declining oxygen concentrations with highly variable NO and N<sub>2</sub>O release rates. Even though our

experiment was conducted in a closed system, with growing denitrifier strains and a limited

amount of substrate, the results are relevant for real soils in that they give a worst-case scenario

of population dynamics and metabolic activity in hotspots. Hotspot architecture played a more

pronounced role for denitrification kinetics at lower soil moisture (30 and 60% WFPS). Hence,

denitrification and its gaseous product stoichiometry do not only depend on the amount of

microbial hotspots in aerated soil, but also on their spatial distribution. The total amount of

denitrification measured as cumulative electron flow, in turn, depended more on water saturation

which is in line with the well-known saturation-dependent threshold behavior in denitrification

also found in natural soil. For the case of artificial soil used in our study, we found that this

threshold behavior was best explained by increased air tortuosity at high saturations. Future

experiments with artificial and natural soils are needed to fully capture the regulation of

465 denitrification at the micro-scale.

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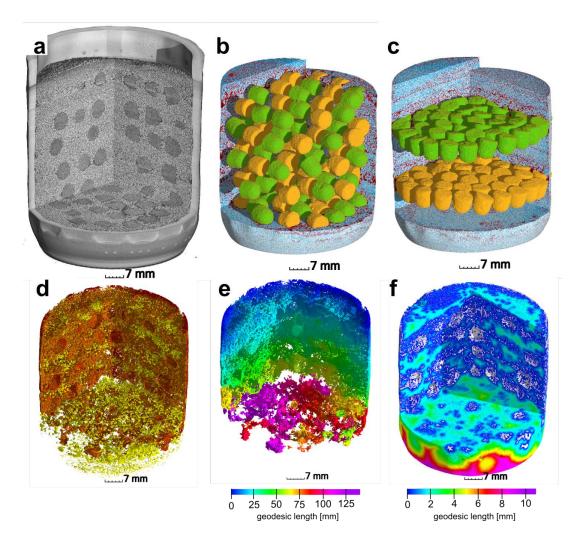


Figure 1. Upper panel: (a) X-ray CT scan of an incubation jar with random hotspot architecture and medium saturation (60% WFPS). (b) Image segmentation of the same jar into air (red), water (blue), sand (transparent), A. tumefaciens hotspots (orange) and P. denitrificans hotspots (green). (c) A different jar at medium saturation (60% WFPS) with layered Pd/At hotspot architecture. Lower panel: a jar with random distribution at high saturation (90% WFPS). (d) Air connectivity, determined as the volume fraction of air connected to the headspace (red, disconnected air shown in yellow). (e) Air tortuosity as derived from the geodesic length to the headspace within the connected air cluster. (f) Diffusion lengths determined as the geodesic length to the closest connected air cluster (white) within water-filled pores.





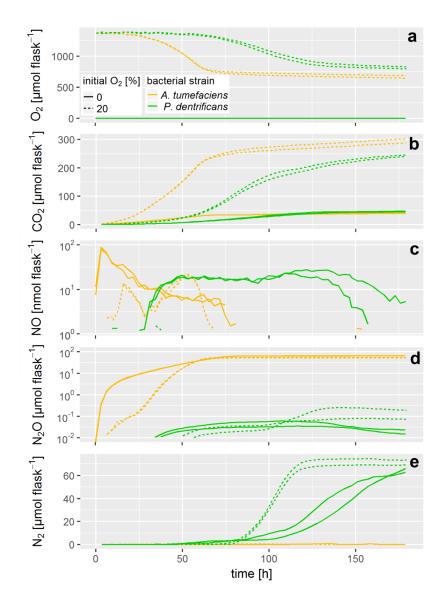


Figure 2: Gas kinetics of individual sets of hotspots inoculated with two different bacterial strains, under oxic and anoxic conditions: (a) O<sub>2</sub>, (b) CO<sub>2</sub>, (c) NO, (d) N<sub>2</sub>O, (e) N<sub>2</sub>. Note the logarithmic ordinate in (c) and (d).





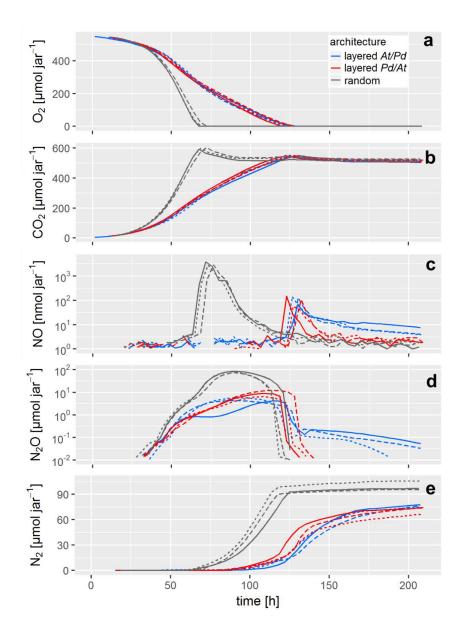


Figure 3: Gas kinetics in all treatments at medium saturation (60% WFPS) for three different hotspot architectures: (a)  $O_2$ , (b)  $CO_2$ , (c) NO, (d)  $N_2O$ , (e)  $N_2$ . Note the logarithmic ordinate in (c) and (d). Different lines styles represent replicates.





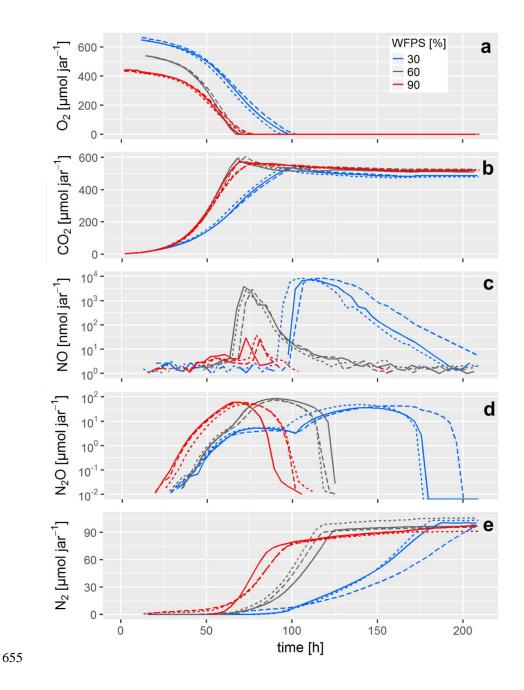


Figure 4: Gas kinetics of randomly placed hotspots at three different saturations: (a)  $O_2$ , (b)  $CO_2$ , (c) NO, (d)  $N_2O$ , (e)  $N_2$ . Note the logarithmic ordinate in (c) and (d). Different lines styles represent replicates.





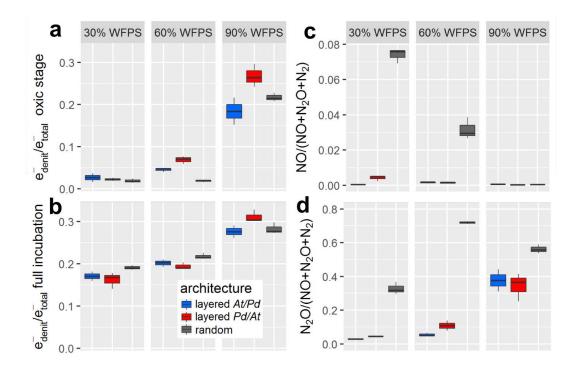


Figure 5: The proportion of denitrification in total respiration expressed as relative electron flow for all architectures and saturations. Values are reported for (a) the initial, oxic to hypoxic stage (O<sub>2</sub> present in headspace) and (b) for the full incubation period of 300 h. The product ratios for NO (c) and N<sub>2</sub>O (d) consider the full incubation period and are corrected for the release of precursor gases. Data shown as box-whisker plots; Whiskers- min-max, middle lines - median.

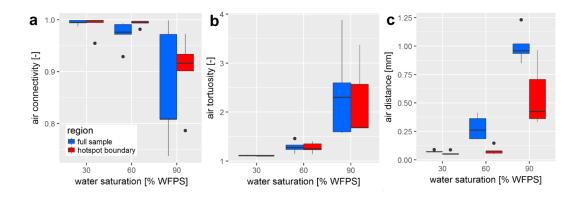


Figure 6: Morphological properties of air-filled pores at different saturations averaged over different hotspots architectures (n=5). These properties are reported separately for the entire pore space within the region of interest (full sample) and for the pore space in direct vicinity to the porous glass beads (hotspot boundary): (a) air connectivity represents the volume fraction of air with direct connection to the headspace. (b) Air tortuosity represents the ratio between geodesic length to the headspace and Euclidean distance for any voxel within the connected air-cluster. (c) Air distance represents the geodesic distance to the connected air cluster within the water-filled pores. Data shown as boxwhisker plots: Whiskers- min-max, middle lines – median, dots: outliers.