

Isotopic evidence for alteration of nitrous oxide emissions and producing pathways contribution under nitrifying conditions

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Abstract. Nitrous oxide (N₂O) emissions by a nitrifying biofilm reactor were investigated with N₂O isotopocules. The nitrogen isotopomer site preference of N₂O (¹⁵N-SP) indicated the contribution of producing and consuming pathways in response to changes in oxygenation level (from 0 to 21 % O₂ in the gas mix), temperature (from 13.5 to 22.3 °C), and ammonium concentrations (from 6.2 to 62.1 mg N L⁻¹). Nitrite reduction, either nitrifier-denitrification or heterotrophic denitrification, was the main N₂O producing pathway under the tested conditions. Difference between oxidative and reductive rates of nitrite consumption was discussed in relation to NO₂⁻ concentrations and N₂O emissions. Hence, nitrite oxidation rates seem to decrease as compared to ammonium oxidation rates at temperatures above 20 °C and under oxygen-depleted atmosphere, increasing N₂O production by the nitrite reduction pathway. Below 20 °C, a difference in temperature sensitivity between hydroxylamine and ammonium oxidation rates is most likely responsible for an increase in the N₂O production via the hydroxylamine oxidation pathway (nitrification). A negative correlation between the reaction kinetics and the apparent isotope fractionation was additionally shown from the variations of δ¹⁵N and δ¹⁸O values of N₂O produced from ammonium. The approach and results obtained here, for a nitrifying biofilm reactor under variable environmental conditions, should allow application and extrapolation on N₂O emissions from other systems such as lakes, soils and sediments.

1 Introduction

Nitrogen (N) cycling lies on numerous biological processes exploited and altered by anthropic activities (Bothe et al., 2007). One of the major issues related to N cycle alteration is the production of nitrous oxide (N₂O) a potent ozone-depleting and greenhouse gas whose emissions exponentially increased during the industrial era (Crutzen et al., 1979; IPCC, 2014; Ravishankara et al., 2009). Wastewater resource recovery facilities (WRRFs) contribute to about 3 % of annual global anthropogenic N₂O sources (ca. 6.7 ± 1.3 Tg N-N₂O in 2011; IPCC, 2014); with 0 to 25 % of the influent nitrogen loads

emitted as N₂O (Law et al., 2012b). The challenges in mitigation of these emissions rely on the understanding of the N₂O producing processes and their controls.

Two microbial processes are responsible for the production of N₂O (nitrification and heterotrophic denitrification), with only one of these capable of consuming it (denitrification; Fig. 1a; Kampschreur et al., 2009). Nitrification is the oxidation of ammonium to nitrite (NO₂⁻) via the intermediate hydroxylamine (NH₂OH) conducted by ammonia oxidizers and the subsequent oxidation of NO₂⁻ to nitrate (NO₃⁻) by nitrite oxidizers. During nitrification, N₂O can be produced as reaction side-product from hydroxylamine oxidation by biotic, abiotic or hybrid processes (Caranto et al., 2016; Heil et al., 2015; Terada et al., 2017). Heterotrophic denitrification and nitrifier-denitrification produce N₂O from nitrite reduction conducted by denitrifiers and ammonium oxidizers, respectively.

Temperature, electron donor and acceptor concentrations have been identified to control the N₂O emission from WRRFs (Bollon et al., 2016; Kampschreur et al., 2009; Tumendelger et al., 2014, 2016; Wunderlin et al., 2012). These variables may induce N₂O accumulation due to inhibition or disturbance of enzyme activity (Betlach and Tiedje, 1981; Kim et al., 2008; Otte et al., 1996). In addition to this, the different N₂O producing processes, nitrification, nitrifier-denitrification or heterotrophic denitrification, are rarely observed independently from each other in heterogeneous environments like wastewater, natural waters, soils or sediments. However, the understanding of the influence that environmental conditions have on the balance between these processes and the N₂O producing pathways remain to a large extent unexplored.

In order to decipher N₂O producing/consuming pathways, the analysis of N₂O isotopocules, molecules that only differ in either the number or position of isotopic substitutions, has been applied (Koba et al., 2009; Sutka et al., 2006) (Figs. 1b-d). The isotope composition of substrates and fractionation mechanisms influence both nitrogen and oxygen isotope ratios of N₂O (reported as δ¹⁵N and δ¹⁸O, respectively, Fig. 1b). Basically, the oxygen atom in the N₂O molecule produced by hydroxylamine oxidation originates from atmospheric dissolved oxygen with δ¹⁸O of 23.5 ‰ (Andersson and Hooper, 1983; Hollocher et al., 1981; Kroopnick and Craig, 1972), while the oxygen atom in N₂O produced by nitrite reduction originates from nitrite that has undergone oxygen-exchange with water (Kool et al., 2007; Snider et al., 2012). Nonetheless, the δ¹⁸O-N₂O resulting from the nitrite reduction conducted by the nitrifiers ranges from 13 to 35 ‰ (Snider et al., 2012). In contrast, the N₂O produced by the heterotrophic denitrifiers through the nitrite reduction pathway has δ¹⁸O over 35 ‰ (Snider et al., 2013). However, the O-exchange between the N₂O precursors and water can decrease it to values below 35 ‰ (Snider et al., 2015). Therefore, the δ¹⁸O alone does not enable differentiation between the N₂O producing pathways.

In combination with δ¹⁸O, the δ¹⁵N-N₂O allows to identify the N₂O producing pathways (Fig. 1b). However, the isotope fractionations (or isotope effects) largely influence the δ¹⁵N-N₂O due to wide variations between and within the reactions involved in the nitrogen cycle (Denk et al., 2017). The isotopic fractionation results from the difference in equilibrium constant or reaction rate observed between the heavier and lighter isotopes in both abiotic and biotic processes. The net isotope effects (Δ) approximated from the difference between δ¹⁵N of product and substrate characterize the production of compounds resulting from sequential or branched reactions and have been recently reviewed (Denk et al., 2017; Toyoda et al., 2017). So

65 far, only two estimates of the net isotope effect of N₂O production by ammonium oxidation via hydroxylamine of -46.5 and -32.9 ‰ have been proposed (Sutka et al., 2006; Yamazaki et al., 2014). These values are imbricated between -52.8 and -6 ‰, the range of net isotope effects related to the N₂O production through nitrite reduction performed by nitrifiers or heterotrophic denitrifiers (Lewicka-Szczebak et al., 2014; Sutka et al., 2008).

Similarly to isotope ratios, the nitrogen isotopomer site preference (¹⁵N-SP), the difference between the relative abundances
70 of N₂O molecules enriched in ¹⁵N at central (N^α) position and terminal (N^β) position, differ according to N₂O producing pathway (Figs. 1c and d). During heterotrophic or nitrifier-denitrification the ¹⁵N-SP of N₂O produced from nitrate or nitrite ranges from -10.7 to 0.1 ‰, while ranging from 13.1 to 36.6 ‰ when N₂O results from hydroxylamine oxidation (Frame and Casciotti, 2010; Jung et al., 2014; Sutka et al., 2006; Yamazaki et al., 2014). Finally, N₂O reduction to N₂ by heterotrophic denitrifiers increases the values of δ¹⁵N, δ¹⁸O and ¹⁵N-SP of residual N₂O with specific pairwise ratios (Jinuntuya-Nortman et
75 al., 2008; Webster and Hopkins, 1996; Yamagishi et al., 2007).

Nitrogen and oxygen isotope ratios of N₂O have lower potential for N₂O source identification as compared to ¹⁵N-SP. However, we believe that the use of both isotope approaches should strengthen the conclusions from ¹⁵N-SP and reveal additional isotope effects (Fig. 1).

The aim of the current study is to improve our understanding regarding the effects of key environmental variables (oxygenation, temperature, NH₄⁺ concentrations) on N₂O production and emission rates. More specifically using nitrogen and oxygen isotope ratios as well as ¹⁵N-SP of N₂O should allow deciphering the different producing and consuming pathways under these different conditions. In order to achieve this, the nitrifying biomass of a submerged fixed-bed biofilm reactor was investigated. Among the wastewater treatment systems, the biofilm systems are adapted to large urban areas owing to their compactness, flexibility and reliability. An increase of their development is expected in response to the additional 2.5 billion humans predicted in urban
85 areas by 2050 (United Nations Population Division, 2018). However, the biofilm systems have received much less attention than the suspended biomass systems and the relations between the N₂O producing/consuming pathways and controls remain largely unknown (Sabba et al., 2018; Todt and Dörsch, 2016). Although applied here to the nitrifying biomass of a WRRF, the research questions addressed regard a diversity of environments including natural waters, soils and sediments: i) Does the nitrifying biomass emit N₂O and what are the producing pathways at play?; ii) Do oxygenation, temperature, and NH₄⁺
90 concentration alter the N₂O emissions, and what are the involved processes? We hypothesize that the isotope signature of N₂O allows identifying the N₂O origins and the assessment of pathway contribution to N₂O emissions. The results of this study should improve the mechanistic understanding as well as improved prediction of N₂O emissions from WRRFs, currently suffering from high uncertainty.

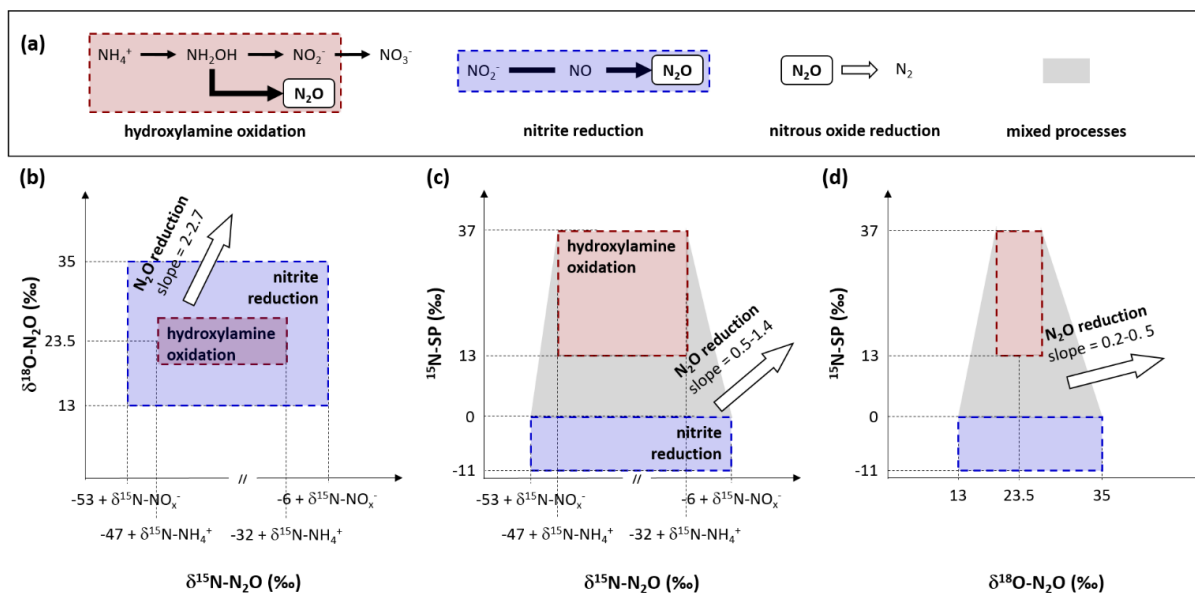


Figure 1: N₂O producing and consuming pathways at play during nitrification and heterotrophic denitrification. Substrate isotope composition, isotope effects and ¹⁵N-SP values from the literature were used to propose the ranges of ¹⁵N (Lewicka-Szczębała et al., 2014; Sutka et al., 2006, 2008; Yamazaki et al., 2014), ¹⁸O (Andersson and Hooper, 1983; Hollocher et al., 1981; Kool et al., 2007; Kroopnick and Craig, 1972; Snider et al., 2012), and ¹⁵N-SP (Frame and Casciotti, 2010; Jung et al., 2014; Sutka et al., 2006; Yamazaki et al., 2014), as well as the slopes relating them with each other during N₂O reduction to N₂ (Jinuntuya-Nortman et al., 2008; Webster and Hopkins, 1996; Yamagishi et al., 2007). The assumptions made and the calculations performed are detailed in the text.

2 Material and methods

95 2.1 Experimental setup for nitrifying experiments

Experiments were carried out with colonized polystyrene beads (diameter 4 mm) sampled from the nitrification biologically active filters (BAF) of a domestic WRRF (Seine Centre, France). In this WRRF, wastewater (240,000 m³ d⁻¹) passes through a pre-treatment stage, followed by a physicochemical decantation, and tertiary biological treatment. The latter is composed of three biofiltration processes; (i) carbon elimination (24 Biofor®), (ii) nitrification (29 Biostyr®), and (iii) denitrification (12 Biofor®). Nitrifying Biostyrs® are submerged fixed-bed biofilm reactors with a unitary section of 111 m² and a filter bed of 3 m high. This unit is operated to receive a nominal load of 0.7 kg NH₄⁺-N m⁻³ d⁻¹.

A lab-scale reactor with a working volume of 9.9 L (colonized Biostyrene® beads and interstitial volume) and a headspace of 1.4 L was operated in continuous down-flow counter-current mode for seven weeks (i.e. solution was down-flowing, while air was up-flowing; Fig. S1). Mass flow meters (F-201CV, Bronkhorst, France) sustained the inflow gas rate at 0.5 L min⁻¹. A peristaltic pump (R3425H12B, Sirem, France) pumped feeding solution from a feeding tank into the reactor at 0.2 L min⁻¹, in order to maintain a hydraulic retention time (HRT) of 27.8 ± 0.6 min. A water jacket monitored by a cryogenic regulator (WK

500, Lauda, Germany) controlled the reactor temperature. The feeding solution consisted of ammonium chloride (NH₄Cl) as substrate, monobasic potassium phosphate (KH₂PO₄) as phosphorus source for bacterial growth, and sodium hydrogen carbonate (NaHCO₃) as pH buffer and inorganic carbon source in 100 or 150 L of tap water (average 0.2 ± 0.4, 2.4 ± 1.1, and 110 2.5 ± 1.3 mg N L⁻¹ of NO₂⁻, NO₃⁻ and sum of both NO_x⁻ molecules, respectively).

The influence of environmental conditions on the ammonium oxidation rates and the N₂O emissions from various combinations of oxygenation levels, temperatures and ammonium concentrations were tested in twenty-four experiments (Table 1). Note that two of them were used twice; as oxygenation and concentration tests. The oxygenation tests were carried out by mixing compressed air and pure nitrogen gas to reach 0 to 21 % O₂ in the gas mixture (Fig. S2a). The tests were performed at five 115 substrate concentrations and at a temperature between 19.2 and 20.6 °C. The temperature tests were carried out by cooling the feeding solution directly in the feeding tank (22.3 to 13.5 °C), with an inflow ammonium concentration close to the nominal load that received the nitrifying biomass; i.e. 20.3-21.1 mg NH₄⁺-N L⁻¹. The ammonium concentration tests were run at an increase (6.2, 28.6 and 62.1 mg NH₄⁺-N L⁻¹) and a decrease (56.1, 42.9, 42.7 and 20.2 mg NH₄⁺-N L⁻¹) in the NH₄⁺ concentrations in the feeding solution, at temperatures ranging from 19.0 to 19.8 °C. Atmospheric oxygenation level (i.e. 21 120 % O₂ in the gas mixture) was imposed for both tests (Figs. S2b and c). This gas mixture using compressed air with 21 % O₂ was considered hereafter as optimal as compared to the oxygen-depleted atmosphere used during the oxygenation tests. Noticeably, the atmospheric oxygenation level is the condition that represents the most optimal conditions of oxygenation applied in nitrification BAF of domestic WRRF.

Table 1. Detailed average conditions (± standard deviation) of oxygenation, temperature and concentration tests.

inflow [NH₄⁺] <i>mg N L⁻¹</i>	inflow gas rate <i>L min⁻¹</i>	O₂ in gas mix <i>%</i>	temperature <i>°C</i>
<i>oxygenation tests</i>			
25.1 ±0.5	0.4	0	19.2 ±0.1
23.8 ±0.6	0.53	4.2	19.9 ±0.1
25.1 ±0.5	0.53	4.2	19.2 ±0.1
37.3 ±0.6	0.5	4.2	20.5 ±0.1
23.8 ±0.6	0.51	10.5	20.2 ±0.1
25.1 ±0.5	0.51	10.5	19.2 ±0.1
37.3 ±0.6	0.5	10.5	20.6 ±0.1
23.8 ±0.6	0.5	16.8	20.1 ±0.1
25.1 ±0.5	0.5	16.8	19.3 ±0.1
37.3 ±0.6	0.5	16.8	20.6 ±0.1
20.2 ±0.5	0.5	21	19.5 ±0.1
25.1 ±0.5	0.57	21	19.6 ±0.5

28.6 ±0.5	0.5	21	19.6 ±0.1
<i>temperature tests</i>			
20.3 ±0.3	0.5	21	13.5 ±0.2
21.1 ±n.a.	0.5	21	15.5 ±0.1
21.1 ±n.a.	0.5	21	16.2 ±0.1
20.3 ±0.3	0.5	21	18.2 ±0.1
21.1 ±n.a.	0.5	21	20.3 ±0.1
20.3 ±0.3	0.5	21	22.3 ±0.1
<i>NH₄⁺ concentration tests</i>			
6.2 ±0.1	0.5	21	19.6 ±0.0
20.2 ±0.5	0.5	21	19.5 ±0.1
28.6 ±0.5	0.5	21	19.6 ±0.1
42.7 ±1.0	0.5	21	19.3 ±0.0
42.9	0.5	21	19.0 ±0.0
56.1 ±0.3	0.5	21	19.0 ±0.1
62.1 ±0.4	0.5	21	19.8 ±0.0

Note that two experiments tested both oxygenation and ammonium concentration.

125 2.2 Reactor monitoring, sampling and concentrations analysis

Dissolved oxygen, temperature (Visiferm DO Arc 120, Hamilton, Switzerland) and pH (H8481 HD, SI Analytics, France) were continuously measured at the top of the reactor and data were recorded at 10 second intervals. The N₂O concentration was continuously analyzed by an infrared photometer (Rosemount™ X-STREAM X2GP, Emerson, Germany) in outflow reactor gas after drying through a condenser and a hydrophobic gas filter (0.2 μm). Minute averages are used for monitored data hereafter. Gas samples were taken for N₂O isotopic signature determination by outlet gas pipe derivation into a sealed glass vial of 20 ml. The vial was first flushed with the sampling gas for > 45 sec prior to 1-5 min sampling. Gas samples were then stored in the dark at room temperature until analysis. Note that gas sampling was lacking for 5 of the 13 oxygenation tests.

The feeding solutions were characterized from 1 to 5 replicate samples collected in the feeding tank. For each tested condition, the outflow was characterized within 5 days from 1 to 14 replicate samples immediately filtered through a 0.2 μm syringe filter and stored at 4 °C. Outflow sampling started after at least one hydraulic retention time (28 ±1 min). Ammonium was analyzed using the Nessler colorimetric method, according to AFNOR NF T90-015 (DR 2800, Hach, Germany). Nitrite and nitrate were measured by ionic chromatography (IC25, Dionex, USA).

2.3 Stable isotope measurements

140 Atmospheric N₂ and Vienna Standard Mean Ocean Water (VSMOW) are the references used for the nitrogen and oxygen
isotopes ratios, respectively, expressed in the conventional δ -notation, in per-mil (‰). Nitrogen and oxygen isotope ratios of
nitrate and nitrite were determined separately following a modified protocol of McIlvin and Altabet (McIlvin and Altabet,
2005; Semaoune et al., 2012). Nitrogen isotope ratios of ammonium were determined following the protocol of Zhang et al.
(2007). These methods consist in the conversion of the substrate (ammonium or nitrite or nitrate) into dissolved N₂O. The $\delta^{15}\text{N}$
145 and $\delta^{18}\text{O}$ for ammonium, nitrite, and nitrate were hence determined from a calibration curve created with a combination of
nitrate or ammonium standards that have undergone the same chemical conversion as the samples (USGS-32, $\delta^{15}\text{N-NO}_3^- = 180$ ‰, $\delta^{18}\text{O-NO}_3^- = 25.7$ ‰; USGS-34, $\delta^{15}\text{N-NO}_3^- = -1.8$ ‰, $\delta^{18}\text{O-NO}_3^- = -27.9$ ‰ and USGS-35 $\delta^{15}\text{N-NO}_3^- = 2.7$ ‰, $\delta^{18}\text{O-NO}_3^- = 57.5$ ‰; or IAEA-N1, $\delta^{15}\text{N-NH}_4^+ = 0.4$ ‰, IAEA-305A, $\delta^{15}\text{N-NH}_4^+ = 39.8$ ‰, USGS-25, $\delta^{15}\text{N-NH}_4^+ = -30.4$ ‰). The
quality of calibration was controlled with additional international standards (IAEA-NO-3, $\delta^{15}\text{N-NO}_3^- = 4.7$ ‰, $\delta^{18}\text{O-NO}_3^- =$
150 25.6 ‰; or IAEA-N2, $\delta^{15}\text{N-NH}_4^+ = 20.3$ ‰). Basically, an analytical sequence was comprised of triplicate standards for
calibration and quality controls and duplicate samples. The average of the analytical replicates was then used for calibration,
quality control and as result.

Since no international standards were available for N₂O isotopes, these were determined the same day as nitrate or ammonium
standard analysis insuring correct functioning of the method and analysis. In addition to this, the internal N₂O standards were
155 previously calibrated by exchange with the laboratory of Naohiro Yoshida and Sakae Toyoda at the Tokyo Institute of
Technology. All isotope measurements were determined using an isotope ratio mass spectrometer (IRMS, DeltaVplus; Thermo
Scientific) in continuous-flow with a purge and trap system coupled with a Finnigan GasBench II system (Thermo Scientific).
The precision was 0.8 ‰, 1.5 ‰ and 2.5 ‰ for $\delta^{15}\text{N}$, $\delta^{18}\text{O}$, and $^{15}\text{N-SP}$, respectively.

2.4 Data processing and statistics

160 The effects of environmental conditions on nitrification were assessed from 4 indices. The ammonium oxidation rate (AOR)
was estimated in each experiment for time ≥ 1 HRT from the difference between influent and effluent NH₄⁺ concentrations
multiplied by the liquid flow rate (kg NH₄⁺-N d⁻¹). The nitrification efficiency was defined as the ratio between AOR and
influent ammonium load. The N₂O emission rate (N₂O-ER) was calculated by multiplying the measured N₂O concentration by
the gas flow rate (mg N₂O-N min⁻¹). The N₂O emission factor (N₂O-EF) was defined as the ratio between N₂O-ER and AOR
165 (% of oxidized NH₄⁺-N). The measurements related to liquid or gas samples were averaged by experiment; i.e. average of data
obtained from the samples collected after one hydraulic retention time.

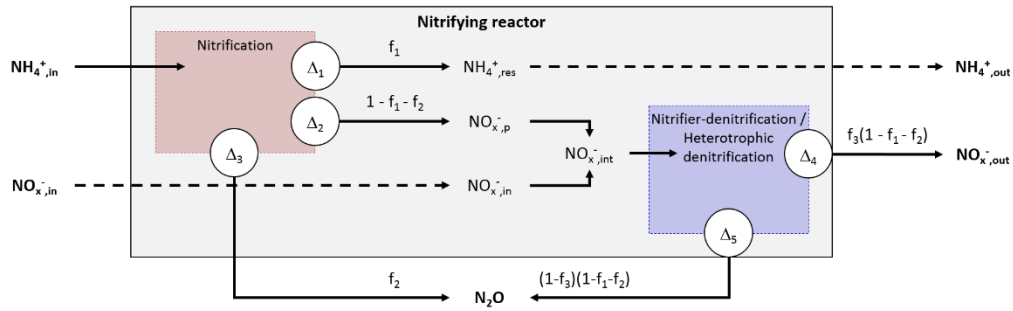
Statistical analysis were performed using the R software (R Core Team, 2014). The value of 0.05 was used as significance
level for spearman correlations (*cor.test* function) and linear regressions (*lm* function). *Adjusted r*² was provided as *r*² for the
latter.

170 **2.5 Estimation of ranges of nitrogen isotope ratio in biologically produced N₂O**

As shown in Fig. 1, the pairwise relationships between $\delta^{15}\text{N}$, $\delta^{18}\text{O}$ and ^{15}N -SP assist the determination of the producing and consuming pathways at play. The N atoms that compose the N₂O molecule originate from NH₄⁺ molecules when produced by hydroxylamine oxidation, while originating from the N atoms of NO₃⁻ or NO₂⁻ molecules when produced by nitrite reduction (NO_x⁻ molecules). However, the nitrogen isotope ratio of N₂O does not equal those of its substrates as it depends on isotope effects associated to each reaction step of N₂O producing process. The isotope effect of reaction step can be determined from the isotope composition of substrates or products. Although being performed on a few tests here, the obtained value can only be applied to a limited number of environmental conditions. The use of estimates from the literature seems therefore suitable. Several equations enable to approximate the isotope effect and its effect on the isotope ratios of substrate and product pools involved in a reaction. These equations vary according to the assumptions made on the system boundaries (Denk et al., 2017).

175 The nitrifying reactor used in this study can be described as an open-system continuously supplied by an infinite substrate pool with constant isotopic composition (NH₄⁺_{,in}). A small amount of the infinite substrate pool is transformed into a product pool (NO_x⁻_{,p}) or a residual substrate pool (NH₄⁺_{,res}) when flowing through the system. The equations describing the input, output and processes considered here are presented in Fig. 2 after Fry (2006). Note that the definitions of f and Δ are inverse to the cited literature and that Δ_1 and Δ_4 are null because no fractionation alter the residual substrate exiting the reaction (Fry, 2006).

180



Equations:

Nitrification:	Nitrifier-denitrification / Heterotrophic denitrification:
$\delta^{15}\text{N}-\text{NH}_4^+_{,\text{res}} \approx \delta^{15}\text{N}-\text{NH}_4^+_{,\text{in}} - \Delta_2(1 - f_1) + (\Delta_3 - \Delta_2)f_2$	$\delta^{15}\text{N}-\text{NO}_x^-_{,\text{out}} \approx \delta^{15}\text{N}-\text{NO}_x^-_{,\text{int}}(1 - f_1 - f_2)^{-1} - \Delta_5(1 - f_3)$
$\delta^{15}\text{N}-\text{NO}_x^-_{,\text{p}} \approx \delta^{15}\text{N}-\text{NH}_4^+_{,\text{in}} + \Delta_2f_1 + (\Delta_3 - \Delta_2)f_2$	$\delta^{15}\text{N}-\text{N}_2\text{O} \approx \delta^{15}\text{N}-\text{NO}_x^-_{,\text{int}}(1 - f_1 - f_2)^{-1} + f_3\Delta_5$
$\delta^{15}\text{N}-\text{NO}_x^-_{,\text{int}} \approx \frac{(\delta^{15}\text{N}-\text{NO}_x^-_{,\text{in}} \times ([\text{NO}_3^-]_{\text{in}} + [\text{NO}_2^-]_{\text{in}}) + \delta^{15}\text{N}-\text{NO}_x^-_{,\text{p}} \times (1 - f_1 - f_2) \times [\text{NH}_4^+]_{\text{in}})}{([\text{NO}_3^-]_{\text{in}} + [\text{NO}_2^-]_{\text{in}} + (1 - f_1 - f_2) \times [\text{NH}_4^+]_{\text{in}})}$	
$\delta^{15}\text{N}-\text{N}_2\text{O} \approx \delta^{15}\text{N}-\text{NH}_4^+_{,\text{res}} - \Delta_2(1 - f_1 + f_2) + \Delta_3(1 + f_2)$	

Figure 2. Diagram and equations of the nitrifying reactor after Fry (2006). It is considered as a sequence of two reactor boxes. (i) The nitrification of inflow ammonium ($\text{NH}_4^+_{,\text{in}}$) to a pool of nitrite and nitrate ($\text{NO}_x^-_{,\text{p}}$), residual ammonium ($\text{NH}_4^+_{,\text{res}}$), and nitrous oxide (N_2O) through the hydroxylamine oxidation pathway. (ii) The subsequent reduction of intermediate $\text{NO}_x^-_{,\text{int}}$; mixing of inflow $\text{NO}_x^-_{,\text{in}}$ and formed $\text{NO}_x^-_{,\text{p}}$ to nitrous oxide (N_2O) through the nitrite reduction pathway, and residual NO_x^- that exits the reactor ($\text{NO}_x^-_{,\text{out}}$). Note that residual substrates and formed products exit the reactor without further isotope fractionation (Δ_1 and Δ_4 are null). See text for details.

185 The balance between input and output of each reactional step allows to propose equations for calculation of the nitrogen isotope ratio of compounds in the inflow and outflow of the system (Denk et al., 2017; Fry, 2006). These equations can be simplified under the assumption that limited amount of N compounds are transformed into N_2O ; i.e. f_2 close to 0 and f_3 close to 1. Therefore, the N isotope ratios of the residual substrate pool can be approximated from Eq. (1).

$$\delta^{15}\text{N}-\text{NH}_4^+_{,\text{res}} \approx \delta^{15}\text{N}-\text{NH}_4^+_{,\text{in}} - \Delta_2(1 - f_1), \quad (1)$$

190 Where f_1 is the remaining substrate fraction leaving the reactor (i.e. remaining fraction of ammonium), ranging from 0 to 1 (0-100 in %), and Δ_2 is the N isotope enrichment factor associated with ammonium oxidation. In their review, Denk et al. (2017) reported a mean value of -29.6 ± 4.9 ‰ for Δ_2 . Therefore, $\delta^{15}\text{N}$ is higher for residual than the initial substrate pool ($\delta^{15}\text{N}-\text{NH}_4^+_{,\text{in}} < \delta^{15}\text{N}-\text{NH}_4^+_{,\text{res}}$). Consequently, the pool of product is depleted in heavier isotope (i.e. nitrite and nitrate hereafter defined as NO_x^- pool; $\delta^{15}\text{N}-\text{NO}_x^-_{,\text{in}} > \delta^{15}\text{N}-\text{NO}_x^-_{,\text{int}}$). It is estimated from Eqs. (2-4):

$$195 \quad \delta^{15}\text{N}-\text{NO}_x^-_{,\text{p}} \approx \delta^{15}\text{N}-\text{NH}_4^+_{,\text{in}} + \Delta_2f_1, \quad (2)$$

Where $\delta^{15}\text{N}-\text{NO}_x^-_{,\text{p}}$ is the nitrogen isotope ratio of the product pool produced by nitrification. The nitrogen isotope ratio of the overall intermediate NO_x^- exiting this process results of mixing between initial and produced NO_x^- pools ($\delta^{15}\text{N}-\text{NO}_x^-_{,\text{int}}$) and can be estimated from Eqs. (3) and (4):

$$\delta^{15}\text{N-NO}_x^-,_{\text{in}} = \frac{(\delta^{15}\text{N-NO}_2^-,_{\text{in}} \times [\text{NO}_2^-]_{\text{in}} + \delta^{15}\text{N-NO}_3^-,_{\text{in}} \times [\text{NO}_3^-]_{\text{in}})}{([\text{NO}_3^-]_{\text{in}} + [\text{NO}_2^-]_{\text{in}})}, \quad (3)$$

$$200 \quad \delta^{15}\text{N-NO}_x^-,_{\text{int}} \approx \frac{(\delta^{15}\text{N-NO}_x^-,_{\text{in}} \times ([\text{NO}_3^-]_{\text{in}} + [\text{NO}_2^-]_{\text{in}}) + \delta^{15}\text{N-NO}_x^-,_{\text{p}} \times (1-f_1) \times [\text{NH}_4^+]_{\text{in}})}{([\text{NO}_3^-]_{\text{in}} + [\text{NO}_2^-]_{\text{in}} + (1-f_1) \times [\text{NH}_4^+]_{\text{in}})}, \quad (4)$$

Note that $\delta^{15}\text{N-NO}_x^-,_{\text{out}}$ equals $\delta^{15}\text{N-NO}_x^-,_{\text{int}}$ when f_3 is close to 1 which means that nitrifier-denitrification and heterotrophic denitrification are negligible. Finally, two options must be considered to approximate the nitrogen isotope ratio of N_2O that exits the reactor. On the one hand, $\delta^{15}\text{N-N}_2\text{O}$ can be estimated from Eq. (5), when the hydroxylamine oxidation is the producing process of N_2O :

$$205 \quad \delta^{15}\text{N-N}_2\text{O} \approx \delta^{15}\text{N-NH}_4^+,_{\text{res}} - \Delta_2(1 - f_1) + \Delta_3, \quad (5)$$

In addition to the influence of the nitrogen isotope composition of the substrate, the $\delta^{15}\text{N-N}_2\text{O}$ depends therefore on the difference between the isotope effects related to the oxidation of NH_4^+ to NO_2^- and the oxidation of NH_2OH to N_2O for complete nitrification ($f_1=0$), while depending only on the latter for limited nitrification ($f_1=1$). On the other hand, $\delta^{15}\text{N-N}_2\text{O}$ can be estimated from Eq. (6), when the nitrite reduction is the producing process of N_2O :

$$210 \quad \delta^{15}\text{N-N}_2\text{O} \approx \delta^{15}\text{N-NO}_x^-,_{\text{int}}(1 - f_1)^{-1} + \Delta_5, \quad (6)$$

In addition to the influence of the nitrogen isotope composition of the substrate, when negligible amounts of N_2O are produced by nitrite reduction during nitrifier-denitrification or heterotrophic denitrification its nitrogen isotope ratio depends on isotope effect related to this process (Δ_5).

3 Results and Discussion

215 Changes in pH, ammonium, nitrite and nitrate concentrations confirmed nitrifying activity in the reactor system (Table S1, Fig. S3). During the ammonium concentration tests, decreases in ammonium concentrations ($[\text{NH}_4^+]$), increases in nitrite and nitrate concentrations ($[\text{NO}_2^-]$ and $[\text{NO}_3^-]$, respectively) were observed, while pH remaining below 8 prevented any relevant loss of ammonium by volatilization. For example, $[\text{NH}_4^+]$ decreased from 6.2 to 1.1, from 28.6 to 17 and from 62.1 to 49.1 mg N L^{-1} by flowing through the nitrifying biomass. At the same time, $[\text{NO}_2^-]$ and $[\text{NO}_3^-]$ increased from 0 to 0.2-0.3 mg N L^{-1} and from 1.4-1.8 to 5-10 mg N L^{-1} , respectively. Over the range of tested conditions, the ratio between ammonium oxidation rate and influent ammonium load ranged from 10 to 82 %, never exceeding 40 % for suboptimal nitrifying conditions imposed during oxygenation and temperature tests (i.e. oxygenation levels < 21 % O_2 , and temperatures < 20 °C). The ammonium concentration, oxygenation level and temperature affected the ammonium oxidation rates, as well N_2O emission rates and factors.

225 3.1 Isotope composition ranges of N_2O produced by hydroxylamine oxidation and nitrite reduction

Ranges of $\delta^{15}\text{N}$ for N_2O produced by different processes were hypothesized from Eqs. (1-5) for pairwise relationships with reviewed data of $\delta^{18}\text{O}$ and $^{15}\text{N-SP}$. To this aim, measurements of isotope ratios of the different nitrogen species were required. The $\delta^{15}\text{N}$ of inflow ammonium, nitrite and nitrate were $-3 \pm 0.1 \text{ ‰}$ ($n=3$), $-15 \pm 0.1 \text{ ‰}$ ($n=2$), $6.9 \pm 0.3 \text{ ‰}$ ($n=3$), respectively

230 during ammonium concentration experiments (Fig. S3 and Table S2). The $\delta^{15}\text{N}$ of residual NH_4^+ and intermediate NO_x^- were estimated from Eqs. (1-4) with $f_1 = 0.1$ or 0.9 (Figs. S2d-f), $\Delta_2 = -30$ ‰, the highest $[\text{NH}_4^+]_{\text{in}}$ (62.1 mg N L^{-1}) and the lowest $[\text{NO}_x^-]_{\text{in}}$ (1.4 mg N L^{-1}). They ranged from -3 to 27 ‰ and from -32 to 7 ‰, respectively, which encompasses a few isotope compositions measured in the outflow during ammonium concentration tests (Fig. S3 and Table S2).

Prior to pairwise comparisons with $\delta^{18}\text{O}$ and ^{15}N -SP, ranges of $\delta^{15}\text{N}$ values for N_2O produced by the hydroxylamine oxidation and nitrite reduction pathways were estimated from Eq. (5). The net isotope effect of N_2O production by ammonium oxidation via hydroxylamine can be estimated by combining the isotope effects of ammonium oxidation and hydroxylamine oxidation to N_2O . The net isotope effect associated to the ammonium oxidation to nitrite ranges from -38.2 to -14.2 ‰ (Casciotti et al., 2003) and can approximate the nitrogen isotope ratio of hydroxylamine transitory produced. The isotope effect related to hydroxylamine oxidation to N_2O ranging from -26.0 to 5.7 ‰ from data in Sutka et al. (2003, 2004, 2006), the net isotope effect of N_2O production by ammonium oxidation via hydroxylamine (Δ_3) can range from -64.2 ‰ ($-26.0 + (-38.2)$) to -8.5 ‰ (235 $5.7 + (-14.2)$). Considering the range of nitrogen isotope ratio of residual ammonium, this method provided a broad range of $\delta^{15}\text{N}$ values, from -65 ‰ ($\delta^{15}\text{N-NH}_4^+_{\text{res}} = -3$ ‰, $\Delta_2 = -30$ ‰, $f_1 = 0.9$ and $\Delta_3 = -64.2$ ‰) to 46 ‰ ($\delta^{15}\text{N-NH}_4^+_{\text{res}} = 27$ ‰, $\Delta_2 = -30$ ‰, $f_1 = 0.1$ and $\Delta_3 = -8.5$ ‰), for N_2O produced from ammonium by hydroxylamine oxidation, according to Eq. (5). These values encompassed the values proposed by others (-46.5 and -32.9 ‰; Sutka et al., 2006; Yamazaki et al., 2014).

A higher range of net nitrogen isotope effect for nitrite reduction than hydroxylamine oxidation pathway was estimated for 245 N_2O production (Figs. 3a and b). Prior to being reduced to N_2O through the nitrite reduction pathway, NO_x^- was mainly derived from ammonium oxidation in the nitrifying system (Eqs. (1-4)); the resulting intermediate $\delta^{15}\text{N-NO}_x^-$ ranging from -32 to 7 ‰. In addition to this, the net isotope effects related to the N_2O production through nitrite reduction performed by nitrifiers or heterotrophic denitrifiers (Δ_5) ranges from -52.8 to -6 ‰ (Lewicka-Szczebak et al., 2014; Sutka et al., 2008). Consequently, the $\delta^{15}\text{N}$ of N_2O produced by nitrite reduction ranged from -89 ‰ ($\delta^{15}\text{N-NO}_x^-_{\text{int}} = -32$ ‰, $f_1 = 0.1$ and $\Delta_5 = -52.8$ ‰) to 64 ‰ (250 $\delta^{15}\text{N-NO}_x^-_{\text{int}} = 7$ ‰, $f_1 = 0.9$ and $\Delta_5 = -6$ ‰), according to Eq. (6). This is consistent with previous findings reporting $\delta^{15}\text{N-N}_2\text{O}$ between -112 and -48 ‰ for nitrifier-denitrifying systems (Mandernack et al., 2009; Pérez et al., 2006; Yamazaki et al., 2014; Yoshida, 1988). However, a similar range of nitrite-derived $\delta^{15}\text{N-N}_2\text{O}$ is suggested for nitrifiers and heterotrophic denitrifiers, because ammonium oxidation influences both processes in the system used in this study where there is low initial amount of NO_2^- and NO_3^- .

255 Pairwise comparisons of $\delta^{15}\text{N}$, $\delta^{18}\text{O}$ and ^{15}N -SP estimates of the different experiments are presented in Fig. 3. These comparisons provided ranges of plausible isotope compositions for N_2O produced by nitrifying or heterotrophic denitrifying bacteria through the hydroxylamine oxidation and nitrite reduction pathways (red and blue boxes, respectively). The measured N_2O isotope compositions were compared to these estimates to identify the N_2O producing and consuming pathways likely at play in oxygenation, temperature, and ammonium concentration tests.

260 This approach suggests that the nitrite reduction pathway was the main contributor to the N_2O emissions. Heterotrophic denitrification likely influenced the N_2O emissions, as shown by oxygen isotope ratios higher than 35 ‰ (Snider et al., 2013)

(Figs. 3a and c). However, this conclusion depends highly on $\delta^{18}\text{O}-\text{N}_2\text{O}$ ranges. Furthermore, the application of atmospheric oxygen $\delta^{18}\text{O}$ (23.5 ‰; Kroopnick and Craig, 1972) to estimate the oxygen isotope ratio of N_2O produced by hydroxylamine oxidation remains uncertain since respiratory activity and air stripping might drive isotopic fractionations and increase the $\delta^{18}\text{O}$ of residual dissolved oxygen (Nakayama et al., 2007). To date, the oxygen isotope fractionation related to air stripping is lacking. Note that this estimate relies on the assumption that there is no accumulation of NH_2OH and that its oxidation to N_2O occurs before or independently of its oxidation to NO_2^- .

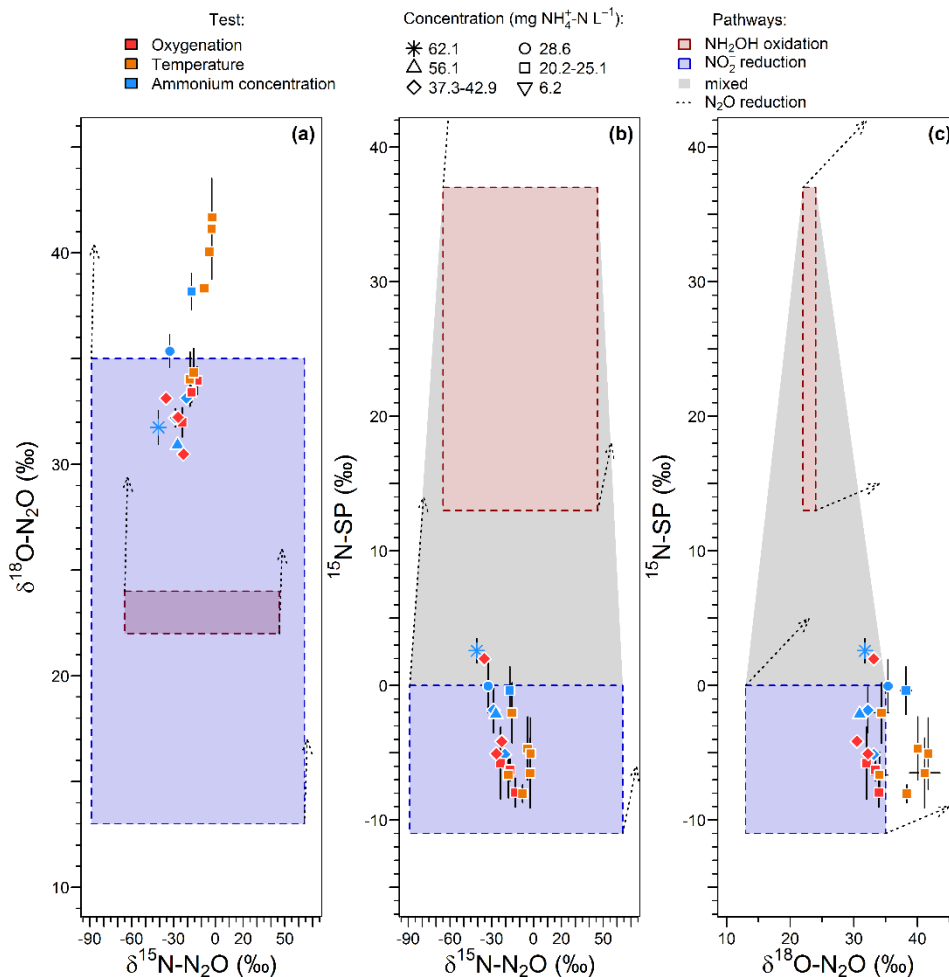


Figure 2: Interpretation maps of the isotope signature of N_2O . Schematic maps of (a) $\delta^{15}\text{N}-\delta^{18}\text{O}$, (b) $\delta^{15}\text{N}-^{15}\text{N-SP}$, and (c) $\delta^{18}\text{O}-^{15}\text{N-SP}$. The red and blue squares show the range of the data for N_2O produced by ‘hydroxylamine oxidation’ and ‘nitrite reduction’, respectively. The shaded area represents mixing of N_2O produced by these pathways. The N_2O reduction increases $\delta^{15}\text{N}$, $\delta^{18}\text{O}$, and $^{15}\text{N-SP}$ with slopes characterizing the pairwise relationships.

3.2 The effect of oxygen limitation on the N₂O producing pathways

Ammonium concentrations decreased from 20.2-37.3 to 11.4-31.1 mg N L⁻¹; with 45 to 89 % of the inflow ammonium remaining in the outflow during the oxygenation tests (Fig. S2d). When measured, the cumulated concentrations of NO₂⁻ and NO₃⁻ ([NO_x⁻]) increased from 2.4-4.1 to 4.7-11 mg N L⁻¹ between inflow and outflow and were composed by at least 74 and 82 % of NO₃⁻, respectively. The mass balance between N compounds that enter and exit the reactor evidenced a default of up to 5 mg N and impacted each test. No significant amount of NO was detected during any tests (data not shown) whereas neither NH₂OH, N₂, nor N mineralization/assimilation in the biofilm were quantified. The accumulation of such amounts of NH₂OH is unlikely. Heterotrophic denitrification, i.e. the reduction of NO_x⁻ and more particularly of N₂O to N₂, may explain the incomplete N mass balance. However, the measurement of small N₂ variations in the gas mixture that exiting the reactor and comprising at least 79 % N₂ was not measured.

The oxygenation level had contrasting effects on ammonium oxidation rates, and N₂O emission rates and factors (Figs. 4a-c). Between an oxygenation of 0 to 10.5 % O₂ in the gas mixture, no clear trend in ammonium oxidation rates was observed although being rather low (1.1 ± 0.5 mg NH₄⁺-N min⁻¹). In the same oxygenation levels interval, the N₂O emission rate increased for two of three inflow [NH₄⁺] tested. It increased from 0.35×10^{-3} to 0.73×10^{-3} mg N min⁻¹ between 0 and 10.5 % O₂ at 25.3 mg NH₄⁺-N L⁻¹, and from 1.34×10^{-3} to 1.4×10^{-3} mg N min⁻¹ between 4.2 and 10.5 % O₂ at 23.8 mg NH₄⁺-N L⁻¹; while it decreased from 2.86×10^{-3} to 2.04×10^{-3} mg N min⁻¹ between 4.2 and 10.5 % O₂ at 37.3 mg NH₄⁺-N L⁻¹. Finally, the N₂O emission factor globally increased from 0.05 to 0.16 % in the 0-10.5 % O₂ interval. At oxygenation levels from 10.5 to 21 % O₂, the ammonium oxidation rates increased from 0.9 ± 0.2 to 2.1 ± 0.4 mg N min⁻¹, with N₂O emission rates remaining stable at $1.2 \times 10^{-3} \pm 0.6 \times 10^{-3}$ mg N min⁻¹ and the emission factors decreasing from 0.15 ± 0.03 to 0.06 ± 0.03 %.

The ¹⁵N-SP varied between -9 to 2 ‰ over the range of imposed oxygenation levels, with a marked increase when oxygenation increased from 16.8 to 21 % O₂ (Fig. 4d). A similar marked change in nitrogen and oxygen isotope ratios of N₂O (decrease and increase, respectively) was observed when oxygenation increased from 16.8 to 21 % O₂ (Figs. 4e and f). Note that to observe the latter variations the effect of ammonium concentration was not included. One way to do so is to compare the isotope composition average at 21% O₂ with the isotope composition measured for 23.8 NH₄⁺-N L⁻¹ at 16.8 % O₂. The ¹⁵N-SP values were close to the range of -11 to 0 ‰ reported for N₂O produced by nitrifying or denitrifying bacteria through nitrifier-denitrification and heterotrophic denitrification (Toyoda et al., 2017; Yamazaki et al., 2014). Additional suggestions can be made from the ¹⁵N-SP dynamics between and variations within the oxygenation levels. If an increase in the hydroxylamine oxidation contribution to the N₂O emission might explain the higher ¹⁵N-SP observed at 21 % O₂ as compared to lower oxygenation levels, an additional mechanism can explain the variations observed for the experiments with oxygen-depleted atmosphere. The ¹⁵N-SP dynamics suggest a higher amount of N₂O was reduced to N₂ at 4.2 than 16.8 % O₂. The reduction of N₂O to N₂ can increase the ¹⁵N-SP of residual N₂O (Mothet et al., 2013). In heterotrophic denitrifying bacteria however, the nitrous oxide reductase involved in this reaction is highly sensitive to inhibition by oxygen (Betlach and Tiedje, 1981; Otte et al., 1996). This might explain the decrease in ¹⁵N-SP from -3.8 ± 4.4 ‰ to -7.2 ± 1.7 ‰ when O₂ increased from 4.2 to 16.8

305 % . This is also consistent with a possible onset of anoxic microsites within the reactor biomass more likely at 4.2 than 16.8 % O₂. The dissolved oxygen (DO) concentration never decreased below 1.5 mg O₂ L⁻¹ in the bulk solution at the top of the reactor (Fig. S2). However, DO decreased from the bulk reactor solution toward the deeper layers of biofilm due to the activity of ammonium oxidizers (Sabba et al., 2018). This is further exacerbated by heterogeneous and varying distribution of air circulation within the static bed. Therefore, oxygen depletion can be assumed within the biofilm. Finally, the N₂O reduction to N₂ likely explains the overall decrease in N₂O emission between 16.8 and 0 % O₂ (Fig. 4b).

310 In general the N₂O reduction to N₂ is accompanied by an increase in nitrogen and oxygen isotope ratios of N₂O (Ostrom et al., 2007; Vieten et al., 2007). However, our results show a decrease in δ¹⁵N-N₂O and the δ¹⁸O-N₂O remained stable between 30.5 and 34.7 ‰ when the N₂O reduction is thought to increasingly constraint the N₂O isotopocules with decreasing O₂ from 16.8 to 4.2 % (Figs. 4e and f). The independence of samples taken during the oxygenation test can explain this. The N₂O sampled at 4.2 % O₂ is not a residual fraction of N₂O produced at 16.8 % O₂ that would have undergone a partial reduction. The oxygenation level can alter the isotope fractionation factors through the control of reaction rates, as evidenced for the reduction of N₂O to N₂ by Vieten et al. (2007). These authors reported lower reaction rates and increased isotope fractionation factors with increasing oxygenation levels. In our case, a similar phenomenon might have influenced both oxidative and reductive processes leading to the production of N₂O and occurring before its ultimate reduction to N₂. However, knowledge regarding the controls such as the oxygenation level have on the net isotope effect related to a sequence of non-exclusive oxidative and reductive processes is still lacking and require further investigations. Additionally, with δ¹⁸O below 35 ‰ for all but one experiment the oxygenation tests did not provide evidence for heterotrophic denitrifier contribution to N₂O emissions, likely due to oxygen-exchange with water (Snider et al., 2015, 2012, 2013).

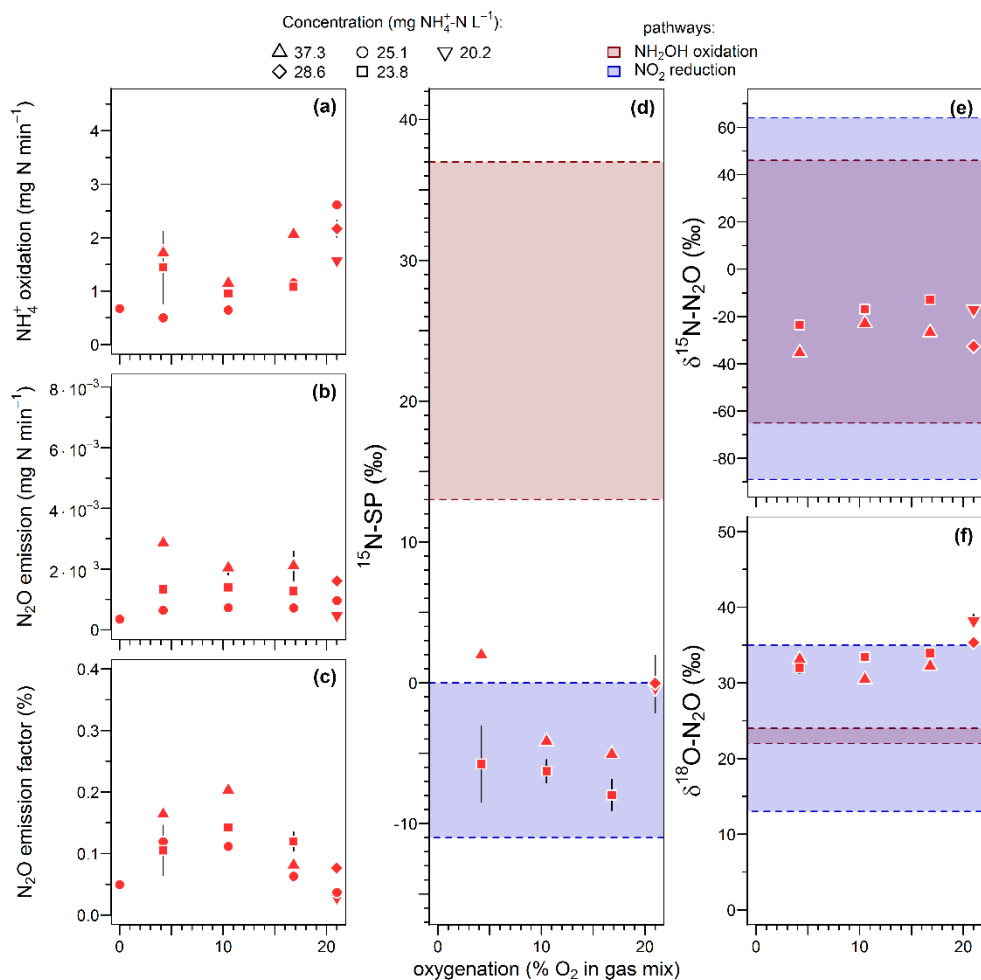


Figure 3: Effect of oxygenation level on (a) the ammonium oxidation rate, (b) the nitrous oxide emission rate, (c) the N₂O emission factor, and (d) the nitrogen isotopomer site preference, (e) the nitrogen isotope ratio, and (f) the oxygen isotope ratio of N₂O. Average and standard deviation (error bars) are calculated for the steady-state conditions. Note that gas sampling for isotope analysis was lacking for 5 of the 13 oxygenation tests.

320 3.3 Difference in temperature dependency of hydroxylamine and ammonium oxidizers as driver of hydroxylamine oxidation contribution to N₂O emissions

Ammonium concentrations decreased from 6.2-62.1 to 0.9-54.1 mg N L⁻¹; from 18 to 79 % of the inflow ammonium remaining in the outflow during the temperature and ammonium concentration tests (Figs. S2e and f). This remaining fraction was positively correlated to ammonium concentrations ($r = 0.96$), and negatively correlated to temperature within a lower range of values (61-67 %; $r = -0.94$). In the ammonium tests, the cumulated concentrations of NO₂⁻ and NO₃⁻ ([NO_x⁻]) increased from 1.4-6.1 to 5.1-19.6 mg N L⁻¹ between inflow and outflow and were composed by at least 74 and 91 % of NO₃⁻, respectively. Noticeably, the nitrite concentrations in the outflow linearly increased with temperature ($r^2 = 0.95$; Fig. S2h).

325

An increase in temperature and inflow ammonium concentrations both positively influenced the rates of NH_4^+ oxidation and N_2O emissions and the emission factor (Fig. 5). The NH_4^+ oxidation rate linearly increased from 1.3 to 1.5 $\text{mg NH}_4^+\text{-N min}^{-1}$ with temperature ($r = 0.89$; Fig. 5a) and increased from 0.97 to 3.49 $\text{mg NH}_4^+\text{-N min}^{-1}$ with a tenfold increase in the inflow ammonium concentration ($r = 0.82$; Fig. 5b). These positive correlations are well known in the temperature range investigated here and are likely due to enhanced enzymatic activity and Michaelis-Menten kinetics, respectively (Groeneweg et al., 1994; Kim et al., 2008; Raimonet et al., 2017). Similarly, the N_2O emission rates increased from 80.4×10^{-6} to 2.5×10^{-3} $\text{mg N}_2\text{O-N min}^{-1}$, and from 83.6×10^{-6} to 6.2×10^{-3} $\text{mg N}_2\text{O-N min}^{-1}$ upon changes in temperature and ammonium concentrations, respectively. These results are in agreement with positive correlations between N_2O emissions with temperature and ammonium concentration observed from modelling and experimental studies on partial nitrification and activated sludge systems (Guo and Vanrolleghem, 2014; Law et al., 2012a; Reino et al., 2017). Altogether this confirms a correlation between the N_2O emission rates and the ammonium oxidation rates. Interestingly, the increase in N_2O emission factor indicates a stronger effect of temperature and ammonium concentration on the N_2O emission rate than on NH_4^+ oxidation. The N_2O emission factors increased from 0.07 to 0.16 %, and from 0.01 to 0.29 % with temperature and inflow ammonium concentration, respectively ($r > 0.94$; Figs. 5e and f). Both experiments suggest that the increase in N_2O emissions results from the increasing production of N_2O by hydroxylamine oxidation or nitrite reduction in combination with a slow rate of or absence of N_2O reduction to N_2 . Furthermore, no nitrite accumulation was observed with increasing ammonium oxidation rate (Fig. S2i). Therefore, if N_2O emission results mainly from the nitrite reduction pathway, this suggests that the nitrite reduction pathway is more responsive to the increasing ammonium oxidation rate than the nitrite oxidation pathway; the latter remaining the main pathway of nitrite consumption.

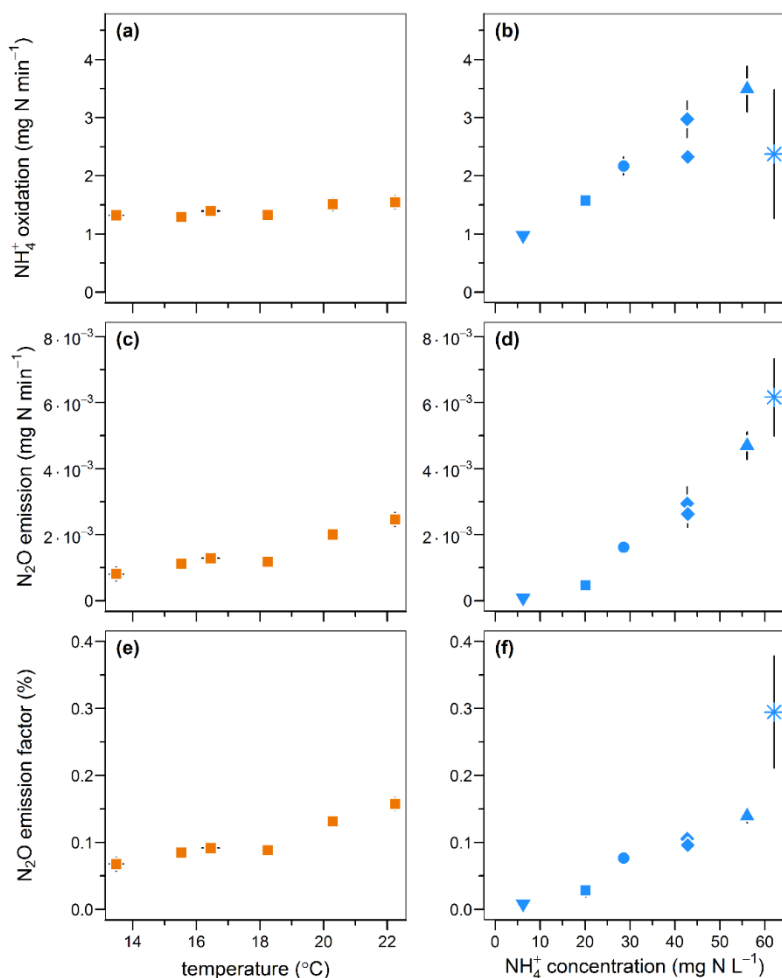


Figure 4: Effect of temperature and inflow ammonium concentration on (a, b) the ammonium oxidation rate, (c, d) the nitrous oxide emission rate, and (e, f) the N₂O emission factor.

The range of nitrogen isotopomer site preference observed during the temperature and concentration tests (from -8 to 2.6 ‰) was similar to those measured during the oxygenation tests; confirming the high contribution of the nitrite reduction pathway to N₂O emissions (Fig. 6a). This is consistent with previous findings based on the ¹⁵N-SP of N₂O emitted from aerobic activated sludge (Toyoda et al., 2011; Tumendelger et al., 2016; Wunderlin et al., 2013), although authors reported ¹⁵N-SP as high as 10 ‰. This can suggest a higher oxygen limitation being favourable to the contribution of the nitrite reduction to N₂O production in the nitrifying reactor studied here. The hydroxylamine oxidation can even be the main N₂O producing pathway, as evidenced by Tumendelger et al. (2014) in some aerated tank.

Furthermore, the ¹⁵N-SP increased with temperature between 13.5 and 19.8 °C. Our data suggest that temperature was the main control on the change in N₂O producing pathways within this temperature range (Fig. 6a). This could explain higher SP obtained with 28.6 than 42.8 mg N L⁻¹ inflow ammonium concentration. The temperature control seems to mitigate here the

effect that ammonium concentration can have on the N₂O producing pathways evidenced elsewhere. Wunderlin et al. (2012, 2013) observed an increase in ¹⁵N-SP from -1.2 to 1.1 ‰ when inflow [NH₄⁺] increased from 9 to 15 mg N L⁻¹. They also observed 3-6 ‰ decreases in ¹⁵N-SP over the course of ammonium oxidation experiments and suggested that NH₂OH oxidation contribution to N₂O production increased when conditions of NH₄⁺ excess, low NO₂⁻ concentrations and high nitrogen oxidation rate occur simultaneously. Our findings are consistent with the observation of Groeneweg et al. (1994) showing that temperature rather than ammonium concentration influenced the ammonium oxidation rate.

The ¹⁵N-SP increased from -6.5 to 2.6 ‰ with increasing temperature from 13.5 to 19.8 °C (Fig. 6a). This ¹⁵N-SP increase may either result from an increase in the N₂O production by the hydroxylamine oxidation pathway or the N₂O reduction to N₂. Since an optimal oxygenation level was imposed and increased emissions were observed, the increasing ¹⁵N-SP is more likely due to N₂O production by the hydroxylamine oxidation pathway. Reino et al. (2017) also observed an increase of N₂O emissions for temperature above 15 °C in a granular sludge airlift reactor performing partial nitrification. The authors suggested two hypothesis to explain their results: (i) the difference in the kinetic dependency with temperature of enzymes involved in ammonium and hydroxylamine oxidation; (ii) the temperature dependency of the acid-base equilibrium ammonium-ammonia. The changes in ¹⁵N-SP observed here are consistent with the former hypotheses. Hydroxylamine oxidation likely becomes the limiting step at a temperature above 15 °C, while being faster than ammonium oxidation at lower temperature (Fig. 7). At temperature above 15 °C, hydroxylamine therefore accumulates and leads to a higher contribution of the hydroxylamine oxidation pathway to N₂O emissions. It would thus be interesting to determine the temperature dependency of the hydroxylamine oxidase.

The change in nitrous oxide producing and consuming pathways had contrasted effects on the nitrogen and oxygen isotope ratios of nitrous oxide (Figs. 6b and c). The δ¹⁵N-N₂O decreased from -2.5 to -40.9 ‰ with an increasing contribution of hydroxylamine oxidation to the N₂O emissions; i.e. when temperature increased from 13.5 to 19.8 °C. This is in contrast with the expected net lower isotope effect for N₂O produced by hydroxylamine oxidation than nitrite reduction and points out that further investigations are needed (Snider et al., 2015; Yamazaki et al., 2014). The changes in δ¹⁸O-N₂O were less straightforward; likely influenced by changes in the reaction rates in addition to changes in the contribution of N₂O producing pathways. The values decreased from 41.1 to 34.3 ‰ with an increasing contribution of hydroxylamine oxidation to the N₂O emissions; when temperature increased from 13.5 to 18.2 °C. It decreased linearly from 38.2 to 31.8 ‰ with increasing reaction rate, when inflow ammonium concentration increased from 20.2 to 62.1 mg NH₄⁺-N L⁻¹ (*r*² = 0.83).

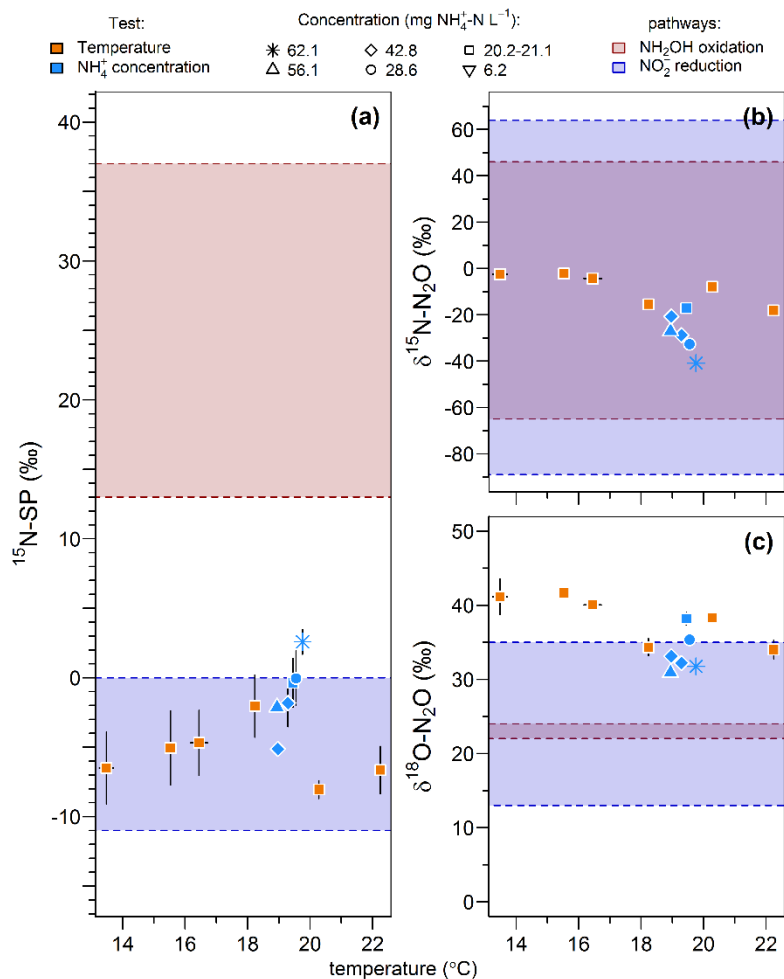


Figure 5: Effect of temperature (orange symbols) and inflow ammonium concentration (blue symbols) on (a) the nitrogen isotopomer site preference, (b) the nitrogen isotope ratio, and (c) the oxygen isotope ratio of N_2O . Average and standard deviation (error bars) are calculated for the samples taken after one hydraulic retention time. Note that the isotopic measurements of gas samples taken at inflow ammonium concentration of 42.7 and 42.9 mg N L^{-1} were both recorded as 42.8 mg N L^{-1} in the legend.

3.4 Difference in oxidation and reduction rates of nitrite as driver of nitrite reduction contribution to N_2O emissions

385 The oxygenation, temperature and ammonium concentration tests revealed a strong control of nitrite oxidizing activity and the contribution of the nitrite reduction pathway to N_2O production. No relationship was observed between NO_2^- concentrations and oxygenation (Fig. S2g). In addition to this, higher $^{15}\text{N-SP}$ at 21 % compared to the 10.5-16.8 % O_2 was observed while temperature remained below 20 °C (Fig. 4d). This is most likely due to higher nitrite oxidation than nitrite reduction rates in response to increasing oxygenation levels to 21 % O_2 , which is consistent with the nitrite oxidation step sensitivity to oxygen limitation (Pollice et al., 2002; Tanaka and Dunn, 1982). Additionally, the $^{15}\text{N-SP}$ close to 0 ‰ observed at the highest

390 oxygenation level indicates a decreasing contribution to N_2O production of nitrite reduction over hydroxylamine oxidation

pathway. The highest oxygenation level thus limits the reduction pathways (i.e. NO_2^- reduction to N_2O and N_2O reduction to N_2) while favoring the ammonium and nitrite oxidation pathways.

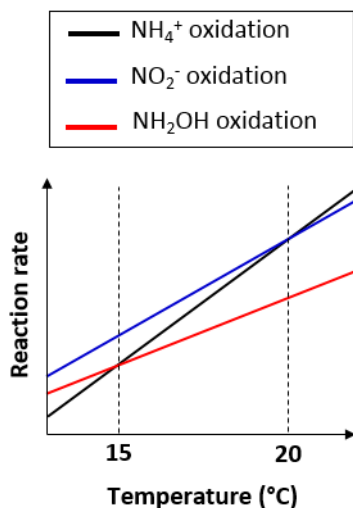


Figure 6: Scheme of the difference in temperature dependency of the reactions involved in nitrification.

During the temperature and ammonium concentration tests, the contribution of the hydroxylamine oxidation pathway to N_2O emissions increased with temperature between 13.5 and 19.8 °C (Sect. 3.3) and decreased in favor of the nitrite reduction pathway when temperature exceeded 20 °C (Fig. 6a). The ^{15}N -SP was low when temperature exceeded 20 °C ($-7.3 \pm 1 \text{‰}$), while being higher than -5‰ ($-1.3 \pm 2.4 \text{‰}$) when temperature ranged from 18.2 to 19.8 °C. At temperatures above 20 °C, ammonium oxidation rates exceed nitrite oxidation rates (Fig. 7; Kim et al., 2008; Raimonet et al., 2017). This explains most likely the increased contribution of the nitrite reduction pathway to N_2O emission, as more nitrite becomes available for nitrifier-denitrification and/or heterotrophic denitrification. As little nitrite accumulated (Fig. S2h), lower rates of nitrite consuming than producing processes can be inferred (nitrite reduction and oxidation vs. ammonium oxidation). Additionally, values of $\delta^{18}\text{O} > 35 \text{‰}$ measured during these tests suggest a significant contribution of heterotrophic denitrifiers to N_2O emissions (Snider et al., 2013). This seems to occur at the lowest hydroxylamine oxidation contribution to N_2O production; below 18°C and at 20.3 °C. Furthermore, the denitrifiers were impacted to a larger extent by temperature than ammonium concentration.

4 Conclusion

Our results demonstrated that whatever the imposed conditions, the nitrifying biomass produced N_2O and nitrite reduction remained the main N_2O producing pathway. The N_2O emissions were sensitive to oxygenation, temperature, and NH_4^+ concentration likely due to control of enzymatic activities. The use of N_2O isotopocules confirmed the processes that control N_2O emissions under oxygenation constrain and improved the knowledge of processes that control N_2O under temperature

constrain. Among the environmental variables tested, temperature appears to be the main control on N₂O producing pathways under nitrifying conditions, due to its dissimilar effects on ammonium and nitrite oxidizing activities. Ranges of optimal temperature for nitrification and limited N₂O emissions can be recommended. The combination of low N₂O emissions and high nitrification rates would occur close to 15 °C. From 15 to 20 °C, increasing nitrification rate increase the N₂O emissions by the hydroxylamine oxidation pathway. Above 20 °C, an increasing nitrification rate increases the N₂O emissions via the nitrite reduction pathway.

We studied the impact of environmental variables on N₂O producing pathways based on the isotope analysis of a limited sample number of dissolved N compounds. The approach and conclusions based on the impact of these variables on N₂O emissions most likely applies to nitrification and denitrification in soils, sediments, lake and marine systems. These systems are subject to dynamic environmental conditions, among which ammonium concentrations, oxygenation and temperature. The comparison of the N₂O isotopocules measured and those hypothesized from the literature provides a useful tool to discuss the N₂O producing and consuming process, as well the underlying control mechanisms at play. Ultimately, this can result in mitigation solutions of N₂O emissions by constraining trough space and time the contribution of N₂O producing and consuming pathways. However, additional efforts seem still needed to reduce, if possible, the ranges of N₂O isotope signatures related to each producing and consuming processes.

Supplementary material

Additional information about the nitrifying activity of the biomass, the experimental conditions, and the time series of ammonium oxidation experiments.

Author contribution

JF, AF and MSp designed the experiments with contributions from GH, MSe and AML. GH, JF and LL carried the experiments out. GH performed the stable isotope measurements with contribution from VV and interpreted them with contribution from MSe. GH and JF processed the data. GH, JF and AML prepared the manuscript with contributions from all co-authors.

Competing interests

The authors declare that they have no conflict of interest.

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440 **References**

- Andersson, K. K. and Hooper, A. B.: O₂ and H₂O are each the source of one O in NO₂⁻ produced from NH₃ by Nitrosomonas: ¹⁵N-NMR evidence, *FEBS Lett.*, 164(2), 236–240, doi:10.1016/0014-5793(83)80292-0, 1983.
- Betlach, M. R. and Tiedje, J. M.: Kinetic explanation for accumulation of nitrite, nitric oxide, and nitrous oxide during bacterial denitrification., *Appl. Environ. Microbiol.*, 42(6), 1074–84, 1981.
- 445 Bollon, J., Filali, A., Fayolle, Y., Guerin, S., Rocher, V. and Gillot, S.: N₂O emissions from full-scale nitrifying biofilters, *Water Res.*, 102, 41–51, doi:10.1016/j.watres.2016.05.091, 2016.
- Bothe, H., Ferguson, S. J. and Newton, W. E.: *Biology of the nitrogen cycle.*, 2007.
- Caranto, J. D., Vilbert, A. C. and Lancaster, K. M.: Nitrosomonas europaea cytochrome P460 is a direct link between nitrification and nitrous oxide emission., *Proc. Natl. Acad. Sci. U. S. A.*, 113(51), 14704–14709, doi:10.1073/pnas.1611051113, 2016.
- 450 Casciotti, K. L., Sigman, D. M. and Ward, B. B.: Linking diversity and stable isotope fractionation in ammonia-oxidizing bacteria, *Geomicrobiol. J.*, 20(4), 335–353, doi:10.1080/01490450303895, 2003.
- Crutzen, P. J., Heidt, L. E., Krasnec, J. P., Pollock, W. H. and Seiler, W.: Biomass burning as a source of atmospheric gases CO, H₂, N₂O, NO, CH₃Cl and COS, *Nature*, 282(5736), 253–256, doi:10.1038/282253a0, 1979.
- 455 Denk, T. R. A., Mohn, J., Decock, C., Lewicka-Szczebak, D., Harris, E., Butterbach-Bahl, K., Kiese, R. and Wolf, B.: The nitrogen cycle: A review of isotope effects and isotope modeling approaches, *Soil Biol. Biochem.*, 105, 121–137, doi:10.1016/j.soilbio.2016.11.015, 2017.
- Frame, C. H. and Casciotti, K. L.: Biogeochemical controls and isotopic signatures of nitrous oxide production by a marine ammonia-oxidizing bacterium, *Biogeosciences*, 7(9), 2695–2709, doi:10.5194/bg-7-2695-2010, 2010.
- 460 Fry, B.: *Stable Isotope Ecology*, Springer-Verlag New York., 2006.
- Groeneweg, J., Sellner, B. and Tappe, W.: Ammonia oxidation in nitrosomonas at NH₃ concentrations near km: Effects of pH and temperature, *Water Res.*, 28(12), 2561–2566, doi:10.1016/0043-1354(94)90074-4, 1994.
- Guo, L. and Vanrolleghem, P. A.: Calibration and validation of an activated sludge model for greenhouse gases no. 1 (ASMG1): prediction of temperature-dependent N₂O emission dynamics, *Bioprocess Biosyst. Eng.*, 37(2), 151–163, doi:10.1007/s00449-013-0978-3, 2014.
- 465 Heil, J., Liu, S., Vereecken, H. and Brüggemann, N.: Abiotic nitrous oxide production from hydroxylamine in soils and their dependence on soil properties, *Soil Biol. Biochem.*, 84, 107–115, doi:10.1016/J.SOILBIO.2015.02.022, 2015.
- Hollocher, T. C., Tate, M. E. and Nicholas, D. J.: Oxidation of ammonia by Nitrosomonas europaea. Definite ¹⁸O-tracer evidence that hydroxylamine formation involves a monooxygenase., *J. Biol. Chem.*, 256(21), 10834–10836, 1981.
- 470 IPCC: *Climate Change 2014: Impacts, Adaptation, and Vulnerability*, *Organ. Environ.*, 24(March), 1–44, 2014.
- Jinuntuya-Nortman, M., Sutka, R. L., Ostrom, P. H., Gandhi, H. and Ostrom, N. E.: Isotopologue fractionation during microbial reduction of N₂O within soil mesocosms as a function of water-filled pore space, *Soil Biol. Biochem.*, 40(9), 2273–2280, doi:10.1016/j.soilbio.2008.05.016, 2008.
- Jung, M.-Y., Well, R., Min, D., Giesemann, A., Park, S.-J., Kim, J.-G., Kim, S.-J. and Rhee, S.-K.: Isotopic signatures of N₂O produced by ammonia-oxidizing archaea from soils, *ISME J.*, 8(5), 1115–25, doi:10.1038/ismej.2013.205, 2014.
- 475 Kampschreur, M. J., Temmink, H., Kleerebezem, R., Jetten, M. S. M. and van Loosdrecht, M. C. M.: Nitrous oxide emission during wastewater treatment, *Water Res.*, 43(17), 4093–4103, doi:10.1016/j.watres.2009.03.001, 2009.
- Kim, J.-H., Guo, X. and Park, H.-S.: Comparison study of the effects of temperature and free ammonia concentration on nitrification and nitrite accumulation, *Process Biochem.*, 43(2), 154–160, doi:10.1016/J.PROCBIO.2007.11.005, 2008.
- 480 Koba, K., Osaka, K., Tobari, Y., Toyoda, S., Ohte, N., Katsuyama, M., Suzuki, N., Itoh, M., Yamagishi, H., Kawasaki, M., Kim, S. J., Yoshida, N. and Nakajima, T.: Biogeochemistry of nitrous oxide in groundwater in a forested ecosystem elucidated by nitrous oxide isotopomer measurements, *Geochim. Cosmochim. Acta*, 73(11), 3115–3133, doi:10.1016/j.gca.2009.03.022, 2009.

- 485 Kool, D. M., Wrage, N., Oenema, O., Dolfig, J. and Van Groenigen, J. W.: Oxygen exchange between (de)nitrification intermediates and H₂O and its implications for source determination of NO₃⁻ and N₂O: a review, *Rapid Commun. Mass Spectrom.*, 21(22), 3569–3578, doi:10.1002/rcm.3249, 2007.
- Kroopnick, P. and Craig, H.: Atmospheric oxygen: isotopic composition and solubility fractionation., *Science*, 175(4017), 54–5, doi:10.1126/science.175.4017.54, 1972.
- 490 Law, Y., Ni, B.-J., Lant, P. and Yuan, Z.: N₂O production rate of an enriched ammonia-oxidising bacteria culture exponentially correlates to its ammonia oxidation rate, *Water Res.*, 46(10), 3409–3419, doi:10.1016/j.watres.2012.03.043, 2012a.
- Law, Y., Ye, L., Pan, Y. and Yuan, Z.: Nitrous oxide emissions from wastewater treatment processes., *Philos. Trans. R. Soc. Lond. B. Biol. Sci.*, 367(1593), 1265–1277, doi:10.1098/rstb.2011.0317, 2012b.
- 495 Lewicka-Szczebak, D., Well, R., Köster, J. R., Fuß, R., Senbayram, M., Dittert, K. and Flessa, H.: Experimental determinations of isotopic fractionation factors associated with N₂O production and reduction during denitrification in soils, *Geochim. Cosmochim. Acta*, 134, 55–73, doi:10.1016/j.gca.2014.03.010, 2014.
- Mandernack, K. W., Mills, C. T., Johnson, C. A., Rahn, T. and Kinney, C.: The δ¹⁵N and δ¹⁸O values of N₂O produced during the co-oxidation of ammonia by methanotrophic bacteria, *Chem. Geol.*, 267(1–2), 96–107, doi:10.1016/J.CHEMGEO.2009.06.008, 2009.
- 500 McIlvin, M. R. and Altabet, M. A.: Chemical conversion of nitrate and nitrite to nitrous oxide for nitrogen and oxygen isotopic analysis in freshwater and seawater, *Anal. Chem.*, 77(17), 5589–5595, doi:10.1021/ac050528s, 2005.
- Mothet, A., Sebilo, M., Laverman, A. M., Vaury, V. and Mariotti, A.: Is site preference of N₂O a tool to identify benthic denitrifier N₂O?, *Environ. Chem.*, 10(4), 281–284, doi:10.1071/EN13021, 2013.
- Nakayama, N., Obata, H. and Gamo, T.: Consumption of dissolved oxygen in the deep Japan Sea, giving a precise isotopic fractionation factor, *Geophys. Res. Lett.*, 34(20), doi:10.1029/2007GL029917, 2007.
- 505 Ostrom, N. E., Pitt, A., Sutka, R., Ostrom, P. H., Grandy, A. S., Huizinga, K. M. and Robertson, G. P.: Isotopologue effects during N₂O reduction in soils and in pure cultures of denitrifiers, *J. Geophys. Res.*, 112(G2), doi:10.1029/2006JG000287, 2007.
- Otte, S., Grobden, N. G., Robertson, L. A., Jetten, M. S. and Kuenen, J. G.: Nitrous oxide production by *Alcaligenes faecalis* under transient and dynamic aerobic and anaerobic conditions., *Appl. Environ. Microbiol.*, 62(7), 2421–6, 1996.
- 510 Pérez, T., Garcia-Montiel, D., Trumbore, S., Tyler, S., Camargo, P. de, Moreira, M., Piccolo, M. and Cerri, C.: Nitrous oxide nitrification and denitrification ¹⁵N enrichment factors from Amazon forest soils, *Ecol. Appl.*, 16(6), 2153–2167, doi:10.1890/1051-0761(2006)016[2153:nonadn]2.0.co;2, 2006.
- Pollice, A., Tandoi, V. and Lestingi, C.: Influence of aeration and sludge retention time on ammonium oxidation to nitrite and nitrate, *Water Res.*, 36(10), 2541–2546, doi:10.1016/S0043-1354(01)00468-7, 2002.
- 515 R Core Team: R: A language and environment for statistical computing, 2014.
- Raimonet, M., Cazier, T., Rocher, V. and Laverman, A. M.: Nitrifying kinetics and the persistence of nitrite in the Seine river, France, *J. Environ. Qual.*, 46(3), 585–595, doi:10.2134/jeq2016.06.0242, 2017.
- Ravishankara, A. R., Daniel, J. S. and Portmann, R. W.: Nitrous oxide (N₂O): the dominant ozone-depleting substance emitted in the 21st century., *Science*, 326(5949), 123–125, doi:10.1126/science.1176985, 2009.
- 520 Reino, C., van Loosdrecht, M. C. M., Carrera, J. and Pérez, J.: Effect of temperature on N₂O emissions from a highly enriched nitrifying granular sludge performing partial nitrification of a low-strength wastewater, *Chemosphere*, 185, 336–343, doi:10.1016/j.chemosphere.2017.07.017, 2017.
- Sabba, F., Terada, A., Wells, G., Smets, B. F. and Nerenberg, R.: Nitrous oxide emissions from biofilm processes for wastewater treatment, *Appl. Microbiol. Biotechnol.*, 102(22), 9815–9829, doi:10.1007/s00253-018-9332-7, 2018.
- 525 Semaoune, P., Sebilo, M., Templier, J. and Derenne, S.: Is there any isotopic fractionation of nitrate associated with diffusion and advection?, *Environ. Chem.*, 9(2), 158–162, doi:10.1071/EN11143, 2012.
- Snider, D., Thompson, K., Wagner-Riddle, C., Spoelstra, J. and Dunfield, K.: Molecular techniques and stable isotope ratios at natural abundance give complementary inferences about N₂O production pathways in an agricultural soil following a rainfall event, *Soil Biol. Biochem.*, 88, 197–213, doi:10.1016/j.soilbio.2015.05.021, 2015.
- 530 Snider, D. M., Venkiteswaran, J. J., Schiff, S. L. and Spoelstra, J.: Deciphering the oxygen isotope composition of nitrous oxide produced by nitrification, *Glob. Chang. Biol.*, 18(1), 356–370, doi:10.1111/j.1365-2486.2011.02547.x, 2012.
- Snider, D. M., Venkiteswaran, J. J., Schiff, S. L. and Spoelstra, J.: A new mechanistic model of δ¹⁸O-N₂O formation by denitrification, *Geochim. Cosmochim. Acta*, 112, 102–115, doi:10.1016/j.gca.2013.03.003, 2013.

- 535 Sutka, R. L., Ostrom, N. E., Ostrom, P. H., Gandhi, H. and Breznak, J. A.: Nitrogen isotopomer site preference of N₂O produced by *Nitrosomonas europaea* and *Methylococcus capsulatus* Bath, *Rapid Commun. Mass Spectrom.*, 17(7), 738–745, doi:10.1002/rcm.968, 2003.
- Sutka, R. L., Ostrom, N. E., Ostrom, P. H., Gandhi, H. and Breznak, J. A.: Nitrogen isotopomer site preference of N₂O produced by *Nitrosomonas europaea* and *Methylococcus capsulatus* Bath, *Rapid Commun. Mass Spectrom.*, 18(12), 1411–1412, doi:10.1002/rcm.1482, 2004.
- 540 Sutka, R. L., Ostrom, N. E., Ostrom, P. H., Breznak, J. A., Pitt, A. J., Li, F. and Gandhi, H.: Distinguishing nitrous oxide production from nitrification and denitrification on the basis of isotopomer abundances, *Appl. Environ. Microbiol.*, 72(1), 638–644, doi:10.1128/AEM.72.1.638, 2006.
- Sutka, R. L., Adams, G. C., Ostrom, N. E. and Ostrom, P. H.: Isotopologue fractionation during N₂O production by fungal denitrification, *Rapid Commun. Mass Spectrom.*, 22(24), 3989–3996, doi:10.1002/rcm.3820, 2008.
- 545 Tanaka, H. and Dunn, I. J.: Kinetics of biofilm nitrification, *Biotechnol. Bioeng.*, 24(3), 669–689, doi:10.1002/bit.260240311, 1982.
- Terada, A., Sugawara, S., Hojo, K., Takeuchi, Y., Riya, S., Harper, W. F., Yamamoto, T., Kuroiwa, M., Isobe, K., Katsuyama, C., Suwa, Y., Koba, K. and Hosomi, M.: Hybrid nitrous oxide production from a partial nitrifying bioreactor: hydroxylamine interactions with nitrite, *Environ. Sci. Technol.*, 51(5), 2748–2756, doi:10.1021/acs.est.6b05521, 2017.
- 550 Todt, D. and Dörsch, P.: Mechanism leading to N₂O production in wastewater treating biofilm systems, *Rev. Environ. Sci. Biotechnol.*, 15(3), 355–378, doi:10.1007/s11157-016-9401-2, 2016.
- Toyoda, S., Suzuki, Y., Hattori, S., Yamada, K., Fujii, A., Yoshida, N., Kouno, R., Murayama, K. and Shiomi, H.: Isotopomer analysis of production and consumption mechanisms of N₂O and CH₄ in an advanced wastewater treatment system, *Environ. Sci. Technol.*, 45(3), 917–922, doi:10.1021/es102985u, 2011.
- 555 Toyoda, S., Yoshida, N. and Koba, K.: Isotopocule analysis of biologically produced nitrous oxide in various environments, *Mass Spectrom. Rev.*, 36(2), 135–160, doi:10.1002/mas.21459, 2017.
- Tumendelger, A., Toyoda, S. and Yoshida, N.: Isotopic analysis of N₂O produced in a conventional wastewater treatment system operated under different aeration conditions, *Rapid Commun. Mass Spectrom.*, 28(17), 1883–1892, doi:10.1002/rcm.6973, 2014.
- 560 Tumendelger, A., Toyoda, S., Yoshida, N., Shiomi, H. and Kouno, R.: Isotopocule characterization of N₂O dynamics during simulated wastewater treatment under oxic and anoxic conditions, *Geochem. J.*, 50(2), 105–121, doi:10.2343/geochemj.2.0390, 2016.
- United Nations Population Division: World urbanization prospects 2018, [online] Available from: <https://population.un.org/wup/Publications/Files/WUP2018-KeyFacts.pdf>, 2018.
- 565 Vieten, B., Blunier, T., Neftel, A., Alewell, C. and Conen, F.: Fractionation factors for stable isotopes of N and O during N₂O reduction in soil depend on reaction rate constant, *Rapid Commun. Mass Spectrom.*, 21(6), 846–850, doi:10.1002/rcm.2915, 2007.
- Webster, E. A. and Hopkins, D. W.: Nitrogen and oxygen isotope ratios of nitrous oxide emitted from soil and produced by nitrifying and denitrifying bacteria, *Biol. Fertil. Soils*, 22(4), 326–330, doi:10.1007/BF00334577, 1996.
- 570 Wunderlin, P., Mohn, J., Joss, A., Emmenegger, L. and Siegrist, H.: Mechanisms of N₂O production in biological wastewater treatment under nitrifying and denitrifying conditions, *Water Res.*, 46(4), 1027–1037, doi:10.1016/j.watres.2011.11.080, 2012.
- Wunderlin, P., Lehmann, M. F., Siegrist, H., Tuzson, B., Joss, A., Emmenegger, L. and Mohn, J.: Isotope signatures of N₂O in a mixed microbial population system: Constraints on N₂O producing pathways in wastewater treatment, *Environ. Sci. Technol.*, 47(3), 1339–1348, doi:10.1021/es303174x, 2013.
- 575 Yamagishi, H., Westley, M. B., Popp, B. N., Toyoda, S., Yoshida, N., Watanabe, S., Koba, K. and Yamanaka, Y.: Role of nitrification and denitrification on the nitrous oxide cycle in the eastern tropical North Pacific and Gulf of California, *J. Geophys. Res.*, 112(G2), doi:10.1029/2006JG000227, 2007.
- Yamazaki, T., Hozuki, T., Arai, K., Toyoda, S., Koba, K., Fujiwara, T. and Yoshida, N.: Isotopomeric characterization of nitrous oxide produced by reaction of enzymes extracted from nitrifying and denitrifying bacteria, *Biogeosciences*, 11(10), 2679–2689, doi:10.5194/bg-11-2679-2014, 2014.
- 580 Yoshida, N.: ¹⁵N-depleted N₂O as a product of nitrification, *Nature*, 335(6190), 528–529, doi:10.1038/335528a0, 1988.
- Zhang, L., Altabet, M. A., Wu, T. X. and Hadas, O.: Sensitive measurement of NH₄⁺ ¹⁵N/¹⁴N (δ¹⁵NH₄⁺) at natural abundance levels in fresh and saltwaters, *Anal Chem*, 79(14), 5297–5303, doi:10.1021/AC070106d, 2007.

