

Response letter

...However, the paper does have some grammatical issues and would benefit from having a professional editor to smooth over the language by improving sentence structure and grammatical flow of the manuscript.

Response: Thank you for your suggestion. We have read over the manuscript and corrected these mistakes.

One place that I see a need for improvement is at the end of the introduction-- I suggest adding a statement there about why the work needs to be done. This is stated in the abstract but not in the introduction.

Response: We added a paragraph at the end of the introduction, from line 66 to line 72.

Further, when referring to nitrogen fixation please use "N₂" instead of N-- I marked all of the places this needs to be fixed.

Response: Corrected.

Modelling biological nitrogen fixation in global natural terrestrial ecosystems

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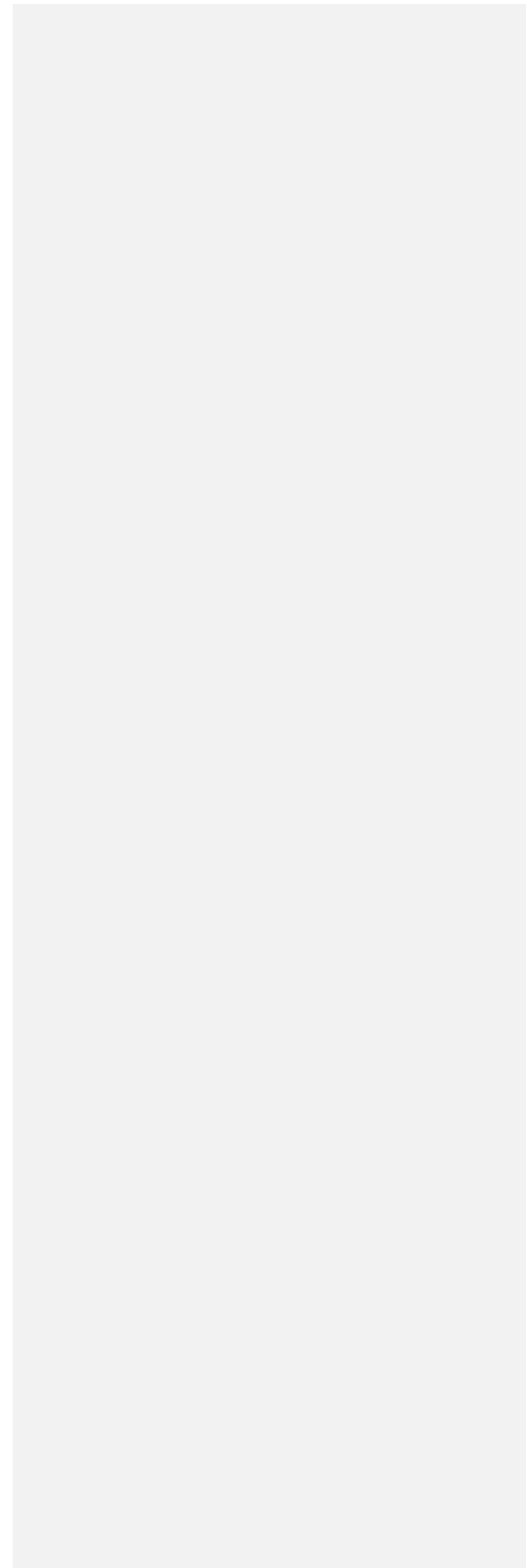
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Abstract. Biological nitrogen fixation plays an important role in the global nitrogen cycle. However, the fixation rate has been usually measured or estimated at a particular observational site. To quantify the fixation amount at the global scale, process-based models are needed. This study develops a biological nitrogen fixation model to quantitatively estimate nitrogen fixation rate by plants in natural environment. The revised nitrogen module better simulates the nitrogen cycle in comparison with our previous model that has not considered the fixation effects. The new model estimates that tropical forests have the highest fixation rate among all ecosystem types, which decreases from the equator to the polar region. The estimated nitrogen fixation in global terrestrial ecosystems is 61.5 Tg N yr⁻¹ with a range of 19.8 - 107.9 Tg N yr⁻¹ in the 1990s. Our estimates are relatively low compared to some early estimates using empirical approaches, but comparable to more recent estimates that involve more detailed processes in their modelling. Furthermore, the contribution of nitrogen made by biological nitrogen fixation depends on ecosystem type and climatic conditions. This study highlights that there are relatively large effects of biological nitrogen fixation on ecosystem nitrogen cycling and the large uncertainty of the estimation calls for more comprehensive understanding of biological nitrogen fixation. More direct observational data for different ecosystems to improve future quantification of fixation and its impacts.

1. Introduction

In most terrestrial ecosystems, nitrogen (N) available for plants is generally limited although it is the most abundant element in the atmosphere (LeBauer and Tresder, 2008). Nitrogen usually enters terrestrial ecosystems through processes of nitrogen deposition and from biological N fixation (BNF). Nitrogen deposition is a physical process, representing the direct input of reactive nitrogen including organic N, ammonia, and nitrogen oxides (NO_y) including nitric oxide (NO), nitrogen dioxide (NO₂), nitric acid (HNO₃) and organic nitrates from the atmosphere to biosphere. BNF, a biochemical process that converts nonreactive nitrogen (N₂) to reactive nitrogen, provides a liaison between the atmosphere and biological systems. Lightning is also a way to convert N₂, adding 3-5 Tg N yr⁻¹ to terrestrial ecosystems (Levy and Moxim, 1996). Nitrogen input via rock weathering is another important source for terrestrial ecosystems, adding 3-10 kg N ha⁻¹ yr⁻¹ (Morford et al., 2011; Houlton et al., 2018). BNF is significantly greater than lightning induced N fixation (Galloway et al., 1995). On a global scale, anthropogenic nitrogen to the environment could be more than 160 Tg N yr⁻¹ (Gruber and Galloway, 2008), which is even greater than terrestrial N fixation (~110 Tg N yr⁻¹). However, taken together, natural N fixation is the primary source in the absence of human activities to global terrestrial ecosystems. For natural terrestrial ecosystems, the amount of N added is approximately balanced by the nitrogen converted back to the atmosphere (Stedman and Shetter, 1983) and lost into ocean and other aquatic systems.

Once entering terrestrial ecosystems, N can be taken up by plants and microbes, and converted into other oxidized forms through mineralization, nitrification and denitrification. In terrestrial ecosystems, N₂N fixation generally affects the nitrogen cycle and nutrient level to constrain plant productivity. Any change of nitrogen input to terrestrial ecosystems will influence their soil nitrogen content.

In the process of BNF, N₂ is converted to ammonia by certain soil microorganisms which can then be utilized by and incorporated into plants. In natural environment, N₂N fixation is conducted by two types of microorganisms:

60 asymbiotic organisms ~~and symbiotic organisms.~~ The former ~~including~~ includes blue-green algae, lichens and free-
living soil bacteria (Belnap, 2002; Granhall & Lid-Torsvik, 1975), ~~and and the later includes symbiotic organisms,~~
~~such as~~ fungi and nodule forming Rhizobium species. Among them, the most dominant fixers are leguminous plants
and their N fixation mechanisms are also ~~the~~ best known (Sullivan et al., 2014; Vitousek et al., 2013). A symbiotic
65 relationship exists between legume plants and bacteria, in which legume plants provide the bacteria energy through
photosynthesis and the bacteria around the rhizobia supplies the legume N in the form of ammonia. To date, the
amount of N fixation by legumes is estimated in the range of 11.3-33.9 kg N ha⁻¹ yr⁻¹ (2.8-8.4 g m⁻² yr⁻¹) in natural
terrestrial ecosystems. ~~A symbiotic relationship exists between legume plants and bacteria. In the process, legume~~
~~plants provide the bacteria energy through photosynthesis, meanwhile, the bacteria around the rhizobia supply the~~
~~legume N in the form of ammonia.~~

70 The biological N₂ fixation rate has been usually measured or estimated at a particular observational site. To
quantify the fixation amount at the global scale, process-based models and sufficient observational data are needed.
This study ~~develops a first models the~~ BNF ~~model considering from~~ the symbiotic relationship between legume
plants and bacteria. ~~The model is extensively calibrated with site-leveled observational data. The model is, then~~
75 ~~extrapolated to the global terrestrial ecosystems to quantify the fixation rate in the 1990s. The analyzes~~ factors
influencing the fixation rate ~~are also analyzed for from~~ different terrestrial ecosystems ~~soils, including the~~
~~distribution of legume plants, soil temperature, and soil properties and types.~~

2. Methods

2.1 Overview

80 We first develop a BNF model and then couple the model with an earlier version of biogeochemistry model
quantifying soil carbon and nitrogen dynamics (Yu and Zhuang, 2019). The revised model is then used to quantify
the BNF at regional and global scales in natural terrestrial ecosystems. The BNF rate estimates consider the effects
of environmental conditions including temperature, soil moisture, soil mineral nitrogen content and soil carbon
content. The modified model is calibrated and evaluated with observed ~~N₂N~~ fixation rate data from published studies
for various natural terrestrial ecosystems from the Arctic to tropical ecosystems. The model sensitivity to model
85 input is analyzed. The model is then extrapolated to the global terrestrial ecosystems at a monthly step and a spatial
resolution of 0.5° by 0.5° for the final decade of the 20th century. The effects of physical conditions on BNF are then
analyzed.

2.2 Model description

90 The Terrestrial Ecosystem Model (TEM) is a process-based model that simulates carbon and nitrogen
dynamics, hydrological and thermal processes for terrestrial ecosystems. Although many efforts were made to
incorporate more details of the N cycle, the N input from the atmosphere to ecosystems has not fully been
incorporated to date, especially the BNF as input. Here we improve the N dynamics within TEM by considering

N_2N fixation by legumes. The model schematic and other calculations including carbon cycle and nitrogen cycle are inherited from an earlier version of TEM (Zhuang et al., 2003; Yu and Zhuang, 2019).

95 BNF is the most significant process in either symbiotic or non-symbiotic forms converting stable molecular N_2 into N chemical compounds that are available to plants. For most terrestrial ecosystems, N_2N fixers could be in many forms, such as free-living bacteria, lichens, and blue algae. But among them, symbiotic BNF is a dominant process to provide biologically accessible N, and most systematical BNF is regulated by legume plants, especially in croplands and semi-natural environments (Mus et al., 2016). In natural environments, contributions from legumes
100 can be significant but with large uncertainties, which is greatly determined by various environmental conditions (Lindemann and Glover, 1996). In this study, the N_2N fixation via legume plants is modeled considering (1) the accessible N concentration in soils, (2) the limitation of temperature, (3) soil water status, (4) the carbon demand for N_2N fixation, and (5) the percentage of N_2N fixing plants for each ecosystem type as:

$$N_{fix} = N_{fixpot} f_t f_w f_N f_c f_{plant} \quad (1)$$

105 where N_{fix} is the nitrogen fixation rate, N_{fixpot} is the potential N_2N fixation rate ($g N day^{-1}$), f_t is the influence function of soil temperature, f_w is the soil water function, f_N is the function of root substrate N concentration, f_c is the function of plant carbon availability, and f_{plant} is the function of legume plant coverage. Please refer to Table 4 for value range of related parameters.

110 The potential N_2N fixation is highly related to the total N demand of plants and the available nitrogen in soils. Theoretically, the definition of potential N_2N fixation rate should be the difference between the demand and supply of N. Both of them vary with plant types, stages of growth and soil conditions. For large spatial-scale simulations for various ecosystem types, it is impossible to derive potential N_2N fixation because of data availability. N_{fixpot} can be estimated based on dry matter of root, nodule or plant dry matter (Voisin et al, 2003, 2007). However, root biomass is also difficult to measure directly. In most published studies, the potential nitrogen
115 fixation rate was measured using an acetylene reduction array (ARA) method (Hardy et al, 1968, 1973), and some used ^{15}N methods (Shearer and Kohl, 1986). In our simulation, N_{fixpot} is assumed to be a constant for each ecosystem type. The N_{fixpot} range is determined from literature and specific values for various ecosystem types are obtained through model parameterization.

120 Soil temperature is a controlling factor for both microbial activities and plant growth. A large number of studies show that different plants have slightly different preferences for temperature (Montanez et al, 1995; Breitbarth et al., 2007; Gundale et al., 2012). For soybean, 20-35 °C is optimal (Boote et al., 2008), and for white clover the optimal temperature can be 13-26 °C (Wu and McGechan, 1999). The activity of microbes responds slightly differently to temperature among species. For most of them, the optimum temperature is 20-25 °C, and at 12-35 °C the activity is not limited. Generally, the relation between the factor and temperature is not exactly a
125 Gaussian distribution. BNF increases as the temperature rises from minimum temperature (0-5 °C) for N fixation to

optimal temperature, maximum rate occurs within an optimal range (15-25°C), and decreases from optimal to maximum temperature above which BNF will stop at 35-40 °C:

$$f_t = \begin{cases} 0 & \text{when } (t < t_{min} \text{ or } t > t_{max}) \\ \frac{t-t_{min}}{t_{optL}-t_{min}} & \text{when } (t_{min} \leq t < t_{optL}) \\ 1 & \text{when } (t_{optL} \leq t \leq t_{optH}) \\ \frac{t_{max}-t}{t_{max}-t_{optH}} & \text{when } (t_{optH} < t \leq t_{max}) \end{cases} \quad (2)$$

130 where the upper limit (t_{max}) is set to 45 °C. There is no lower limit, but when t is low enough, f_t will be close to zero (Wu and McGechan, 1999; Boote et al., 2008; Holzworth et al., 2014) (Table 1). For the convenience in computing, a lower limit is set in our model. When the temperature goes beyond its upper or lower limit, f_t is assumed to be 0.

Water stress has a direct effect on nitrogen fixing system (Sprent, 1972). With proper temperature, soil moisture condition is the major factor controlling nitrogen fixation rate (Srivastava and Ambasht, 1994). Soil water deficit and flood dramatically inhibits N_2N fixation because of drought stress and oxygen deficit, respectively
135 (Omari et al., 2004; Mario et al., 2007). In our model, the water factor is linearly related with soil water content (Williams, 1990; Wu and McGachan, 1999):

$$f_w = \begin{cases} 0 & \text{when } (W_f \leq W_a) \\ \varphi_1 + \varphi_2 & \text{when } (W_a < W_f < W_b) \\ 1 & \text{when } (W_f \geq W_b) \end{cases} \quad (3)$$

where W_f ($J \text{ kg}^{-1}$) is the available soil water, which is defined as the ratio of water content to that at the field
140 capacity. In soils, water potential generally includes osmotic and matrix potentials, ranging from -0.1 to -0.3 bar for typical soils, which has little effects on the N fixation. But when the soil gets very dry, the potential can be up to -100 to -200 bar and increases rapidly. W_a is the bottom threshold below which N_2N fixation is totally restricted by soil moisture. W_b is the upper threshold above which nitrogen fixation is not limited by soil moisture. φ_1 and φ_2 are
145 parameters representing the linear relationship between soil water content and its effect on N_2N fixation, respectively (Table 1).

It is generally thought that more substrate N in soils will slow down the N_2N fixation, because plants can uptake N directly from soil with less energy (Vitousek and Field, 1999). By comparison, N_2N fixation needs more energy and consumes more carbon than plant N uptake does. Thus, the N_2N fixation is only considered to occur when the direct N uptake from soil cannot meet the plant N demand. In our model, the inhibition effect of N is
150 defined as (Wu and McGehan, 1999):

$$f_N = \begin{cases} 1 - f_{Nup} \ln(1000 - N_s) & \text{when } (N_s \geq 0.001) \\ 1 & \text{when } (N_s < 0.001) \end{cases} \quad (4)$$

Where f_{Nup} is a parameter related to legume biological N_2N fixation and soil N. N_s is the soil mineral N ($g \text{ N m}^{-2}$).
155 f_N shows a natural logarithmic relation with the soil mineral N.

N_2N fixers get photosynthetic carbohydrate support from plants. Because the product of every unit of nitrogen fixed consumes a certain amount of carbon, the lack of carbon supply will inhibit the N_2N fixation. The carbon cost for per unit of fixed N_2N varies widely depending on environmental conditions and ecosystem types. For example, the consumption of carbon is only 1.54 times of fixed N_2N for cowpea (Layzell et al., 1979), and it can be 6.3 to 6.8 times for soybeans (Ryle et al., 1979). It is also related to the life cycle of plants. The carbon effect is modeled following a Michaelis-Menten equation (Boote et al., 1998):

$$f_c = \frac{1}{1 + K_c / C_r} \quad (5)$$

where C_r is the soil carbon content ($g\ C\ m^{-2}$) to represent carbon availability from plants to N_2N fixers. K_c is the Michaelis-Menten constant, which is plant species dependent.

2.3 Data

The classification of land cover and leguminous biomes were derived from the combination of the International Geosphere and Biosphere (IGP) land-cover classification system and the study of Schrire et al (2005). The experimental N_2 fixation data for model calibration were collected for 7 major ecosystem types. Nitrogen fixation rates were determined with acetylene reduction assay (ARA) method in most published studies (Table 2, data were from Cleveland et al. (1999)), expressed in $kg\ N\ m^{-2}\ yr^{-1}$. Some of them were measured with the ^{15}N natural abundance technique.

The parameters for N_2 fixation module were initialized with a priori values (Table 2). Ecosystem-specific and microbe guild-specific parameters were inherited from previous TEM model (Zhuang et al., 2003; Yu and Zhuang, 2019). The global simulations were conducted at a spatial resolution of 0.5 by 0.5 degree and at a monthly time step. Historical climate data including temperature, precipitation, cloudiness and water vapor pressure were derived from the Climate Research Unit (CRU) (Mitchell and Jones, 2005). Soil texture data were from Melillo et al. (1993) and Zhuang et al. (2003). Other initial conditions including vegetation properties, soil carbon content and soil nitrogen contents were from Chen and Zhuang (2013) and Zhuang et al. (2012).

For regional simulations, the total amount of fixed N_2N was also influenced by legume coverage. For each ecosystem type, we estimated the coverage according to the distribution of legume plants and field studies (Table 3, the coverage data are compiled from Cleveland et al. (1999)), where the minimum and maximum values were derived from the abundance of N_2N -fixers.

2.4 Model calibration and site-level validation

Most model parameters are legume-specific or vegetation-specific and are adjusted based on value ranges from previous studies (Table 1). Model is parameterized for 7 representative natural terrestrial ecosystems (Table 2). Root mean square error (RMSE) and coefficient of determination ($0 \leq R^2 \leq 1$) were used for model calibration. RMSE was

calculated to show the mean difference between simulated data and observational values. The model is iterated with changing parameters until the RMSE reached a certain value for each site. Most parameters in the model driving nitrogen cycle in the soil have been defined and calibrated in previous studies (Yu and Zhuang, 2019). The calibrated model is evaluated at the site level and then extrapolated to the global terrestrial ecosystems.

2.5 Model sensitivity and uncertainty Analysis

The response of N_2N fixation of different biomes to input data and variation of parameters was analyzed using sensitivity testing. Four major input variables were selected, including air temperature, precipitation, soil nitrogen content and soil organic carbon content. The monthly average input variables were changed by $\pm 10\%$ of the original level for each site and each grid. The variables were changed at 6 levels, respectively, and the rest of input variables were kept at their original values. The sensitivity was calculated by comparing the simulated annual nitrogen fixation to the simulations with the original input values.

3. Results

3.1 Model evaluation

To evaluate the model, thirty-five observational sites were selected for 7 major ecosystem types across the globe, representing different climate and soil conditions. The experimental data of N_2N fixation have a mean value of $12.9 \text{ kg N ha}^{-1} \text{ yr}^{-1}$, with a standard deviation of $17.7 \text{ kg N ha}^{-1} \text{ yr}^{-1}$. The maximum observed fixation occurred in temperate forest in New Zealand, while the minimum rate was also for temperate forest in [the state of Idaho](#) [State of](#) [in](#) the US. Our simulations are comparable with the observed data for all major ecosystem types with the coefficient of determination (R^2) of 0.44 and with a slope of 0.46 (Figure 2). The regression results are mainly influenced by some observed data greater than $30 \text{ kg N ha}^{-1} \text{ yr}^{-1}$. By removing the outliers of observational data, the slope of regression increases to 0.72. Observational data for temperate forests show the greatest variation among all major ecosystem types, with a maximum value reaching 800 times of the minimum one. Simulations are closer to the observations across sites in temperate forests with R^2 of 0.26 and slope of 0.42. Our model underestimated nitrogen fixation rate in temperate forests. The large variation in observations may be due to the distribution of legume plants, different sampling time periods (e.g., growing and non-growing seasons), and varying climate conditions. For tropical forests, our model estimates of N_2N fixation are higher than observations with the slope of 0.75 and R^2 of 0.44.

3.2 Model sensitivity analysis

The model sensitivity analysis quantifies the impact of changes in forcing data on nitrogen fixation rate. Climate conditions including air temperature and precipitation, and soil characteristics of nitrogen content and carbon content varied at 3 levels to examine the sensitivity. The response of nitrogen fixation rate emissions is quantified for each ecosystem type. The sensitivity test was conducted for all observational sites (Table 2). Temperature is the most sensitive variable (Figure 1). Nitrogen fixation is more sensitive to the change of all forcing conditions. Increasing soil nitrogen results in a lower N_2N fixation. Abundant soil nitrogen content inhibits BNF activity, but stimulates nitrification and denitrification processes.

3.3 Biological nitrogen fixation in global terrestrial ecosystems

Tropical forests in South America, Central Africa and South Asia show a wide range of N_2N fixation rates between 1 and 200 kg N ha⁻¹ yr⁻¹ (Bruijnzeel et al, 1991). Here all plants in tropical rainforest are assumed to fix nitrogen and one set of parameters are applied for all tropical forests. The coverage for tropical forests in the landscape was assumed to be 15% (Cleveland et al., 1999), ranging from 5% to 25%. The N_2 fixation rate was estimated to be 18.2 kg N ha⁻¹ yr⁻¹, which is the highest among all vegetation types. Our simulations show that the total fixed nitrogen ranges from 10.8 Tg N yr⁻¹ to 54 Tg N yr⁻¹, with the average value of 32.5 Tg N yr⁻¹(Table 3). Nitrogen fixation in tropical forests is almost half of the global total amount and a principal contributor of BNF in natural ecosystems. Tropical forests have the largest potential to fix nitrogen given that the optimal temperature and soil moisture for BNF is relatively easy to have under tropical climatic conditions.

Temperate forests cover the largest land area from 30°N to 60°N, including temperate coniferous forest, temperate deciduous forest and temperate evergreen forest. Temperate areas have the majority of legumes and many temperate ecosystems are considered to be N limited. Comparing to other ecosystem types in temperate regions, conifers are likely to limit the reproduction of legumes (Wheatley et al, 2010). In general, plant species carrying nitrogen fixers are only distributed in a small percentage of natural temperate forests, like clear-felled areas and pastures (Boring and Swank, 1984). Cleveland et al. (1999) indicated that the legume coverage ranges from 1% to 10% of the land area only. Consequently, our simulations indicate that N_2N fixation by temperate forests was 12.7 kg N ha⁻¹ yr⁻¹. The estimates of the total nitrogen fixation were between 1.9 and 19.14 Tg N yr⁻¹ (Table 3). Nitrogen fixation in temperate areas contributes 12.5% of the global total amount.

Savanna covers over a half of African continent, Australia and large areas of South America. It is an important biome in the Southern Hemisphere. There is a great variation in native legume species. Only in humid savanna, legumes may significantly contribute to the increase of soil nitrogen (Cech et al., 2008). On average, 15% of the vegetation in savanna is considered as legume grass and biological nitrogen fixation occurs when precipitation is greater than 10 mm per month. Generally, nitrogen fixation in savanna is restricted by soil moisture, while temperate grassland is limited by both temperature and soil moisture (Bustamante et al, 1970). Nitrogen fixers are not abundant for these biomes (Woodmansee et al., 1981). The coverage of nitrogen fixers was assumed to be from 5% to 25%, (Cleveland et al., 1999). Our simulation assumed that nitrogen fixers cover 15% of the land, resulting in 1.9 kg N

255 ha⁻¹ yr⁻¹ fixation, representing a much smaller fraction compared to forest ecosystems. Total fixed nitrogen in grasslands appeared to range from 0.62 to 3.1 Tg N yr⁻¹, with an average of 1.86 Tg N yr⁻¹. For savanna, the total contribution was less due to its relatively small area. The minimum, average and maximum values were estimated to be 0.45, 1.34 and 2.23 Tg N yr⁻¹, respectively.

260 In tundra and boreal forest regions, both host plants and their rhizobia are adapted to the environment with low temperature. Nitrogen fixation rate is extremely variable for boreal ecosystems. For tundra, the coverage was assumed to be 3-15%, and for boreal forest, the coverage was 4-18%. But in general, the low temperature and permafrost conditions limit the activity of nitrogen fixers (Alexander, 1981). We estimated that tundra ecosystems fix nitrogen at 3.2 kg N ha⁻¹ yr⁻¹. Their total BNF was between 0.51 to 2.55 Tg N yr⁻¹ with average of 1.54 Tg N yr⁻¹. In boreal forests, the fixation rate was much lower (2.1 kg N ha⁻¹ yr⁻¹) compared to temperate forests.

~~The fixation could be neglected in deserts because of the extremely dry conditions. Only few legumes may exist in deserts and their growth is highly depended on precipitation events. Even in semi-arid areas, the N₂ fixation rate is much lower than that in tropical and temperate forests (5.7 kg N ha⁻¹ yr⁻¹).~~

265 ~~In deserts, although some legumes may exist in extremely dry conditions, and some species may grow rapidly after rainfall, their fixation could be neglected. However, in semi-arid areas, legumes are common plants with several species, their N fixation is lower than tropical and temperate forests (5.7 kg N ha⁻¹ yr⁻¹).~~

270 Mediterranean ecosystems such as in southern California and some areas in southern Australia are characterized with mild rainy winter and hot dry summer, containing both evergreen and deciduous shrublands, in which nodulated legumes are prominent (Sprent et al., 2017). These legumes are more active in comparatively wet season than in dry season (Sánchez-Díaz, 2001). The ability to fix nitrogen is considered to be one of the most important features that enable legumes and plants to survive under severe environments (Crisp et al., 2004). We estimated that the N₂N fixation rate of these legume species is similar to that in grasslands (2.7 kg N ha⁻¹ yr⁻¹).

275 Spatially, the highest rate of N₂N fixation occurred in the tropical and sub-tropical areas, as a result of proper climate and soil characteristics for fixers (Figure 3). N fixation from tropical forests and xeric shrubland contributes to nearly half of the global terrestrial amount (Table 3). A lower N₂N fixation rate was in high latitudes of eastern China, North America and Europe, which were mainly covered with temperate forests. Compared to tropical areas, N₂N fixation in temperate regions shows a larger variability depending on vegetation types. The spatial variation could be attributed to the distribution of legume plants, in addition to the difference of humidity and temperature conditions. N₂N fixation in temperate regions accounts for 35% of the total fixed N₂N.

280 Our model estimated that high BNF rates in the growing season is consistent with other regional and global estimates (Cleveland et al., 1999, 2013; Lee and Son, 2005; Lett and Michelsen, 2014). The energetic cost for active N uptake becomes lowest when soil temperature is around 25°C (Fisher et al. 2010). Similarly, our estimates of high BNF rates also occur at similar temperature conditions in spring and summer. The global soil nitrogen mineralization rate was estimated to be 696 Tg N yr⁻¹ while 15% of plant N demand was provided by BNF (Cleveland et al., 2013). Our estimates of BNF were lower than the estimates by Cleveland et al. (2013) and fell within 10% of the total soil mineralization rate. This result also indicates that about 10% of the mineralized N was induced by BNF.

290 During 1990-2000, our simulations show that BNF in natural terrestrial ecosystems is 61.5 Tg N yr⁻¹, but anthropogenic N_2N fixation was much higher at 140 Tg N yr⁻¹ (Galloway et al., 2002). This large amount of anthropogenic N input to terrestrial ecosystems is expected to inhibit the natural BNF and might lead to less BNF in the future.

4. Discussion

4.1 Comparison with other estimates of biological nitrogen fixation (BNF)

295 There is a large uncertainty in estimating the N input into terrestrial ecosystems, especially from BNF (Sutton et al., 2014) (Table 3). In our study, a calibrated process-based model was applied to estimate site-level and global BNF in natural terrestrial ecosystems. Empirical models provide reasonable estimation based on relationships between N_2 fixation rates and environmental factors (e.g. evapotranspiration) (Cleveland et al, 1999), while process-based approaches consider processes in BNF affected by multiple controlling factors (Fisher et al., 2010; Gerber et al., 2008; Meyerholt et al., 2016). Our estimated BNF in the global terrestrial ecosystems is 61.5 Tg N yr⁻¹ with an uncertainty ranging from 19.8 to 107.9 Tg N yr⁻¹, which is lower than most existing studies. Cleveland et al. (1999) provided a central value of 195 Tg N yr⁻¹ by scaling up field-based experimental data, with a range of 100 - 289 Tg N yr⁻¹. This range represents potential distribution of nitrogen fixation. In reality, N_2N fixation is also affected by climate and soil conditions, making the actual terrestrial BNF smaller than the potential one. In a more recent study of Cleveland et al. (2013), a total of 127.5 Tg N yr⁻¹ was estimated to be related to BNF, based on the relationship between BNF and evapotranspiration (ET). Galloway et al (2002b) also provided several estimates for global BNF. Galloway et al. (2004) further suggested a range of 100 - 290 Tg N yr⁻¹ and implied that the true rate of BNF would be at the low end of this range without large-scale human disturbance. In an earlier study (Galloway et al., 2002b), the mean annual global BNF was estimated to be 89-100 Tg N yr⁻¹. By assuming a steady state between N input to and loss from ecosystems, Vitousek et al. (2013) estimated the BNF to be 58 Tg N yr⁻¹ with a plausible range of 40 - 100 Tg N yr⁻¹, which is similar to our estimates. However, Xu-Ri and Prentice (2017) estimated that the N_2N fixation was about 340 Tg N yr⁻¹ which is almost 5 times larger than our estimates. In their study, BNF was determined by plant N requirement across all biome types.

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320 In our estimation, tropical forests significantly contribute to the total BNF, which is up to 18 kg N ha⁻¹yr⁻¹. This result is highly related to the density of leguminous plants, and the physical conditions in tropical areas (Crews, 1999). Our simulated results are comparable to the estimates of symbiotic N_2 fixation from tropical evergreen (5.5-16 kg N ha⁻¹ yr⁻¹) and deciduous forests (7.5-30 kg N ha⁻¹ yr⁻¹) (Reed et al., 2011). Barron et al. (2010) directly measured N_2 -fixing root nodules across lowland tropical forests and their observations also showed a large variation among individual trees. For a mature forest matrix, the average value was around 10 kg N ha⁻¹ yr⁻¹, but it could be as high as 200 kg N ha⁻¹ yr⁻¹ for some areas. Cleveland (2013) provided a similar estimate to ours (around 12 kg N ha⁻¹ yr⁻¹), but higher values (20-30 kg N ha⁻¹ yr⁻¹) in their earlier studies (Cleveland et al., 1999). Sullivan et al (2014) analyzed human's impact on tropical N fixation and found, depending on forest ages, fixation was 5.7 kg N ha⁻¹ yr⁻¹ with a range from 1.2 to 14.4 kg N ha⁻¹ yr⁻¹, which is lower than our estimates.

325 For temperate and boreal forests, we estimated that BNF fixation is 2.1-18 kg N ha⁻¹ yr⁻¹. The existing BNF
estimates from literature also show a large uncertainty for those forest ecosystems. For instance, LM3V-N model
(Gerber et al., 2009) suggested that the N input to forests to be less than 5 kg N ha⁻¹ yr⁻¹. But their model also
estimated that, in moist forests, the uptake of N could be 30-80 kg N ha⁻¹ yr⁻¹. Deluca et al. (2002) reported that
cyanobacterium and feather moss could act as a supplement to N_2N fixation in boreal forests (0.5 kg N ha⁻¹ yr⁻¹)
330 while the organic N accumulation could be 3 kg N ha⁻¹ yr⁻¹. For the forests in northwest Rocky Mountain, N_2N
fixation amount is on average between 0.5 and 2 kg N ha⁻¹ yr⁻¹ (Clayton and Kennedy, 1985; Fahey et al., 1988)
while Kou-Giesbrecht and Menge's model (2019) estimated the N_2N fixation rate to be 0 -10 kg N ha⁻¹ yr⁻¹ for
temperate forests, and 0 to 6 kg N ha⁻¹ yr⁻¹ for boreal forests.

335 There could be a number of reasons for our comparatively lower estimates. The most important one is that there
is a considerable uncertainty in estimating the coverage of N_2N fixing plants. High diversity in the distribution of
legume plants highly influences the estimation of total plant coverage, because our estimation was based on site-
level experimental data. In order to improve our understanding, more investigation on legume plant distribution and
associated data for N_2N fixers is needed, especially in the Middle Asia, South America and Africa.

340 Large variations of BNF rates exist across terrestrial ecosystems spatially (Figure 3). The global BNF spatial
pattern is similar to other estimates (Cleveland et al., 1999; Xu-Ri and Prentice, 2017). The highest N_2N fixation rate
in tropical regions (more than 50% of the global terrestrial N_2N fixation) is primarily due to their warm and moist
soil conditions. Further, N_2N fixed by human activities became increasingly influential in the past century
(Galloway et al., 2002), especially in temperate regions due to their large human population. The anthropogenic N
deposition contributed more to soil N than BNF did. As a result, soils became N rich, inhibiting BNF in temperate
345 soils. This could explain why the potential N_2N fixation rate was high in temperate ecosystems, but only contributed
to 20% of the total fixation.

4.2 Major controls on biological nitrogen fixation

350 In our simulations, the N_2N fixation was primarily influenced by soil temperature, moisture and soil nitrogen
content. The highest N_2N fixation rate in tropical ecosystems is consistent with our sensitivity analysis for
temperature and soil moisture. The sensitivity analysis indicated that a 1-3°C increase of temperature led to 7%
increase in N_2N fixation rate. Nitrogen cycle responds differently between different biomes and legume types. But in
general, increasing temperature will accelerate processes in the N cycle. Soil moisture correlates with BNF in a
similar way with temperature. A slightly increase of precipitation (10%) increased the nitrogenase activity.
However, the response of N_2N fixation to soil water stress is not as sensitive as that to the change in temperature.
Xeric shrubland and savanna in dry tropical areas still contribute greatly to the global N_2N fixation, while the
355 contribution of boreal forests, with low temperature, is much lower.

BNF is highly regulated by soil nitrogen content. N-deficiency conditions usually favor BNF activities, for
example, in xeric shrubland and savanna. Enhancing soil N content will decrease the N_2N fixation rate, which is also
consistent with our sensitivity analysis. It costs less energy for plants to take up N directly from soils rather than

360 biologically fixing it from the atmosphere (Cannell and Thornley, 2000). However, there is an exception for some
areas in tropical ecosystems. Many tropical soils are comparatively rich in nitrogen, but N_2 -fixing plants are still
active to compensate the nitrogen depletion due to the rapid N cycling (Pons et al., 2007). This explains why N
fertilization inhibits the BNF in temperate ecosystems, but BNF is still active in N-rich soils in tropical ecosystems.
In areas where the energetic cost exceeds the demand of N, the BNF rate will be comparatively lower. Sullivan et
al. (2014) suggested that there were lower rates of BNF in undisturbed mature forests and higher rate in secondary
365 forests, depending on the balance between N-demand and energy consumption.

4.3 Model limitation and future work

The incorporation of BNF into TEM allows us to more adequately simulate nitrogen cycle from natural
terrestrial ecosystems. However, there are several limitations in this study.

370 First, the current model ignores the effect of free-living BNF. Although symbiotic BNF is critical for most
natural and semi-natural ecosystems, asymbiotic organisms play an important role in extreme environments such as
waterlogged soils and deserts. The importance of symbiotic BNF or fixation by leguminous plants may not be as
significant as previously thought. Elbert et al. (2012) suggested that cryptogam contributed nearly half BNF in
terrestrial ecosystems, which was up to 49 Tg N yr⁻¹. In some tropical areas, the spatial N input from free-living
bacteria even exceeds symbiotic input (Sullivan et al., 2014). In addition, legumes are not the only source of
375 symbiotic BNF. Some fungi species have the ability to actively fix atmospheric nitrogen. But in most existing
models, fungi or mycorrhizae symbioses are not considered due to the limited knowledge about their mechanisms of
fixing N (Fisher et al., 2010). A more comprehensive model that covers various types of nitrogen fixation is needed.

380 Second, the BNF process in our model is calibrated with a limited amount of data, imposing a general set of
parameters to all plant species and soil conditions within an ecosystem type. More observational data from natural
terrestrial ecosystems is desirable to improve our model.

385 Third, it is difficult to isolate the N addition via natural processes from human activities. In the US, 20-35% of
annual N input into terrestrial ecosystems are human-related (Sobata et al., 2013). As a result, the quality of
observational data varies from site to site, some BNF data are only semi-natural. The observational data are
imperfect, which might have also biased our estimates through model parameterization process.

5. Conclusions

390 This study developed a process-based biological nitrogen fixation model and coupled it with an extant
biogeochemistry model. The model was evaluated with observed data for N_2 fixation. The model was then
extrapolated to the global natural terrestrial ecosystems. Our model estimates that biological nitrogen fixation in
natural terrestrial ecosystems was 61.5 Tg N yr⁻¹ during the last decade of the 20th century and the greatest fixation
rate occurred in tropical regions. Soil temperature, rather than soil moisture and nutrient content, is the most
dominant control on N_2 fixation. Lacking the knowledge about the distribution of N_2 fixing plants and their
physiological features might have biased our estimates of ~~both~~ biological nitrogen fixation at the global scale.

Data availability

395 Climate data including monthly cloudiness, precipitation, temperature, and water vapor pressure are from the
Climate Research Unit (CRU) <http://www.cru.uea.ac.uk/data> (last access: May 2017). Global vegetation data and
soil data are available in Zhuang et al. (2003) and McGuire et al. (2001). The explicit spatial data on soil water pH
from the ORDL gridded soil properties product (https://daac.ornl.gov/cgi-bin/dsviewer.pl?ds_id=546, last access:
400 ~~March-May, 2017~~2020) are based on the World Inventory of Soil Emission Potentials (WISE) database (Batjes,
2000). The global average carbon dioxide concentration is observed at NOAA's Mauna Loa Observatory. N deposit
data are from NADP monitor and CASTNET. The initial values of soil microbial carbon and nitrogen, and the ratio
of C / V / N at the global scale, were from a compilation of global soil microbial biomass carbon, nitrogen, and
phosphorus data (<https://doi.org/10.3334/ORNLDAAC/1264>, last access: May, 2017). *The data presented in this
paper can be accessed through our research website (<http://www.eaps.purdue.edu/ebdl/>).*

Author contribution

405 Q. Zhuang and T. Yu designed the research. T. Yu performed model simulations and data analysis. Both
authors contributed to the paper writing.

Competing interests

410 The authors declare that they have no conflict of interest.

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420 Reference:

Adams M A, Attiwill P M. Role of Acacia spp. in nutrient balance and cycling in regenerating Eucalyptus regnans F. Muell. forests. I. Temporal changes in biomass and nutrient content. *Aust. J. Bot.*, 32(2): 205-215, 1984.

Alexander V, Billington M M. Nitrogen fixation in the Alaskan taiga, *Forest ecosystems in the Alaskan taiga*. Springer, New York, NY, 112-120, 1986.

425 Baker T G, Oliver G R, Hodgkiss P D. Distribution and cycling of nutrients in *Pinus radiata* as affected by past lupin growth and fertiliser. *Forest Ecol. Manag.*, 17(2-3): 169-187, 1986.

Barron A R, Purves D W, Hedin L O. Facultative nitrogen fixation by canopy legumes in a lowland tropical forest. *Oecologia*, 165(2): 511-520, 2011.

430 [Bate, G. C., and C. Gunton. Nitrogen in the Burkea savanna, in Ecology of Tropical Savannas, edited by B. J. Huntley and B. H. Walker, pp. 498-513, Springer-Verlag, New York, 1982.](#)

Battle M, Bender M, Sowers T, et al. Atmospheric gas concentrations over the past century measured in air from firm at the South Pole. *Nature*, 383(6597): 231, 1996.

Beck, Douglas, and Luis A. Materon, eds. Nitrogen Fixation by Legumes in Mediterranean Agriculture: Proceedings of a Workshop on Biological Nitrogen Fixation on Mediterranean-type Agriculture, ICARDA, Syria, April 14–17, 1986. Vol. 32. Springer Science & Business Media, 2012.

435 Blundon D J, Dale M R T. Dinitrogen fixation (acetylene reduction) in primary succession near Mount Robson, British Columbia, Canada[J]. *Arctic. Alpine. Res.*, 22(3): 255-263, 1990.

Boote K J, Jones J W, Hoogenboom G, et al. Simulation of crop growth: CROPGRO model. *Agricultural systems modeling and simulation*, 18: 651-692, 1998.

440 [Boote K.J., Hoogenboom G., Jones J.W., Ingram K.T. Modeling nitrogen fixation and its relationship to nitrogen uptake in the in the CROPGRO model, in: Ma L., Ahuja L.R., Bruulsema T.W. \(Eds.\), Quantifying and Understanding Plant Nitrogen Uptake for Systems Modeling. CRC Press, Florence, USA, pp. 13–46, 2008](#)

Boring L R, Swank W T, Waide J B, et al. Sources, fates, and impacts of nitrogen inputs to terrestrial ecosystems: review and synthesis. *Biogeochemistry*, 6(2): 119-159, 1988.

445 Boring L R, Swank W T. The role of black locust (*Robinia pseudo-acacia*) in forest succession. *J. Ecol.*, 749-766, 1984.

[Bouniols, A., M. Cabelguenne, C.A. Jones, A. Chalamet, J.L. Charpentau, and J.R. Marty. Simulation of Soybean Nitrogen Nutrition for a Silty Clay Soil in Southern France. Field Crops Res. 26\(1\): 19-34, 1991](#)

450 Bowman W D, Schardt J C, Schmidt S K. Symbiotic N₂-fixation in alpine tundra: ecosystem input and variation in fixation rates among communities. *Oecologia*, 108(2): 345-350, 1996.

Breitbarth E, Oschlies A, LaRoche J. Physiological constraints on the global distribution of *Trichodesmium*? effect of temperature on diazotrophy[J]. *Biogeosciences*, 4(1): 53-61, 2007.

455 Bruijnzeel L A. Nutrient input–output budgets of tropical forest ecosystems: a review. *J. Trop. Ecol.*, 7(1): 1-24, 1991.

Bustamante, M. et al. Nitrogen cycling in tropical and temperate savannas. 10.1007/978-1-4020-5517-1_10, 1970.

Cabelguenne, M., P. Debaeke, and A. Bouniols. EPICphase, A Version of the EPIC Model Simulating the Effects of Water and Nitrogen Stress on Biomass and Yield, Taking Account of Developmental Stages: Validation on Maize, Sunflower, Sorghum, Soybean, and Winter Wheat. *Agric. Syst.* 60(3): 175-96, 1999.

460 Cannell M G R, Thornley J H M. Modelling the components of plant respiration: some guiding principles. *Ann. Bot. London*, 85(1): 45-54, 2000.

Cech P G, Kuster T, Edwards P J, et al. Effects of herbivory, fire and N₂-fixation on nutrient limitation in a humid African savanna. *Ecosystems*, 11(6): 991, 2008.

465 Chapin D, Bliss I C, Bledsoe I J. Environmental regulation of nitrogen fixation in a high arctic lowland ecosystem. *Can J. Botany*. 69. 2744-2755. 1991.

Chen M I N, Zhuang Q. Modelling temperature acclimation effects on the carbon dynamics of forest ecosystems in the conterminous United States. *Tellus B.*, 65(1): 1915, 2013.

470 Christie P. Nitrogen in two contrasting Antarctic bryophyte communities. *J Ecol* 75:73-93. 1987

Clayton J L, Kennedy D A. Nutrient Losses from Timber Harvest in the Idaho Batholith I. *Soil. Sci. Soc. Am. J.*, 49(4): 1041-1049, 1985.

475 Cleveland C C, Townsend A R, Schimel D S, et al. Global patterns of terrestrial biological nitrogen (N₂) fixation in natural ecosystems. *Global biogeochem. Cy.*, 13(2): 623-645, 1999.

Cleveland C C, Houlton B Z, Smith W K, et al. Patterns of new versus recycled primary production in the terrestrial biosphere. *P. Natl. A. Sci.*, 110(31): 12733-12737, 2013.

480 Corre-Hellou G, Brisson N, Launay M, et al. Effect of root depth penetration on soil nitrogen competitive interactions and dry matter production in pea-barley intercrops given different soil nitrogen supplies. *Field Crop. Res.*, 103(1): 76-85, 2007.

Corre-Hellou G, Faure M, Launay M, et al. Adaptation of the STICS intercrop model to simulate crop growth and N accumulation in pea-barley intercrops. *Field Crop. Res.*, 113(1): 72-81, 2009.

485 Crews T E. The presence of nitrogen fixing legumes in terrestrial communities: Evolutionary vs ecological considerations, *New Perspectives on Nitrogen Cycling in the Temperate and Tropical Americas*. Springer, Dordrecht, 233-246, 1999.

Crisp M, Cook L, Steane D. Radiation of the Australian flora: what can comparisons of molecular phylogenies across multiple taxa tell us about the evolution of diversity in present-day communities?. *Philos. T. Roy. Soc. B.*, 359(1450): 1551-1571, 2004.

490 Davidson E A, Reis de Carvalho C J, Vieira I C G, et al. Nitrogen and phosphorus limitation of biomass growth in a tropical secondary forest. *Ecol. Appl.*, 14(sp4): 150-163, 2004.

Dentener F, Drevet J, Lamarque J F, et al. Nitrogen and sulfur deposition on regional and global scales: A multimodel evaluation. *Global Biogeochem Cycles*~~*Global biogeochemical cycles*~~, 20(4), 2006.

- DuBois J D, Kapustka L A. Biological nitrogen influx in an Ohio relict prairie. [Am J Bot-American Journal of Botany](#), 70(1): 8-16, 1983.
- 495 Eckersten, H., Geijerstam, L. A., & Torssell, B. Modelling nitrogen fixation of pea (*Pisum sativum* L.). *Acta. Agr. Scand. B-S P*, 56(2), 129-137, 2006.
- Eisele, L., Schimel, D. S., Kapustka, L. A., & Parton, W. J. Effects of available P and N: P ratios on non-symbiotic dinitrogen fixation in tallgrass prairie soils. *Oecologia*, 79(4), 471-474, 1989.
- Elbert, W., Weber, B., Burrows, S., Steinkamp, J., Büdel, B., Andreae, M. O., & Pöschl, U. Contribution of cryptogamic covers to the global cycles of carbon and nitrogen. *Nat. Geosci.*, 5(7), 459, 2012.
- 500 Erickson H, Davidson E A, Keller M. Former land-use and tree species affect nitrogen oxide emissions from a tropical dry forest. *Oecologia*, 130(2): 297-308, 2002.
- Erickson H, Keller M, Davidson E A. Nitrogen oxide fluxes and nitrogen cycling during postagricultural succession and forest fertilization in the humid tropics. *Ecosystems*, 4(1): 67-84, 2001.
- 505 Fahey T J, Yavitt J B, Pearson J A, et al. The nitrogen cycle in lodgepole pine forests, southeastern Wyoming. *Biogeochemistry*, 1(3): 257-275, 1985.
- Fisher J B, Sitch S, Malhi Y, et al. Carbon cost of plant nitrogen acquisition: A mechanistic, globally applicable model of plant nitrogen uptake, retranslocation, and fixation. *Global Biogeochem. Cy.*, 24(1), 2010.
- Galloway J N, Schlesinger W H, Levy H, et al. Nitrogen fixation: Anthropogenic enhancement-environmental response. *Global Biogeochem. Cy.*, 9(2): 235-252, 1995.
- 510 Galloway J N, Cowling E B, Seitzinger S P, et al. Reactive nitrogen: too much of a good thing?. *AMBIO: A Journal of the Human Environment*, 31(2): 60-64, 2002.
- Galloway, James N., and Ellis B. Cowling. "Reactive nitrogen and the world: 200 years of change." *AMBIO: A Journal of the Human Environment* 31.2 (2002): 64-71.
- 515 Galloway J N, Dentener F J, Capone D G, et al. Nitrogen cycles: past, present, and future. *Biogeochemistry*, 70(2): 153-226, 2004.
- Gerber S, Hedin L O, Oppenheimer M, et al. Nitrogen cycling and feedbacks in a global dynamic land model. *Global Biochemical. Cy.* 24(1), 2010.
- [Goosem S, and Lamb D. Measurements of phylloshpere nitrogen fixation in a tropical and two sub-tropical rainforests. *Journal of Tropical Ecology*, 2, 373-376. 1986.](#)
- 520 Gregory J, Stouffer R J, Molina M, et al. *Climate change 2007: the physical science basis*. 2007.
- Grove T S, Malajczuk N. Nodule production and nitrogen fixation (acetylene reduction) by an understorey legume (*Bossiaea laidlawiana*) in Eucalyptus forest. *J. Ecol.*, 303-314, 1992.
- 525 Gruber, N. and Galloway, J.N. (2008) An Earth System Perspective of the Global Nitrogen Cycle. *Nature*, 451, 293-296.
- Gundale M J, Nilsson M, Bansal S, et al. The interactive effects of temperature and light on biological nitrogen fixation in boreal forests. *New Phytol*, 194(2): 453-463, 2012.

- 530 Hardy R W F, Holsten R D, Jackson E K, et al. The acetylene-ethylene assay for N₂ fixation: laboratory and field evaluation. *Plant physiology*, 1968, 43(8): 1185-1207, 1968.
- Hardy R W F, Burns R C, Holsten R D. Applications of the acetylene-ethylene assay for measurement of nitrogen fixation. *Soil Biol. Biochem.*, 5(1): 47-81, 1973.
- Harvey A E, Larsen M J, Jurgensen M F, et al. Nitrogenase activity associated with decayed wood of living northern Idaho conifers. *Mycologia*, 81(5): 765-771, 1989.
- 535 He X H, Critchley C, Bledsoe C. Nitrogen transfer within and between plants through common mycorrhizal networks (CMNs). *Crit Rev Plant. Sci*, 22(6): 531-567, 2003.
- Heath, B., Sollins, P., Perry, D. A., & Cromack Jr, K. Asymbiotic nitrogen fixation in litter from Pacific Northwest forests. *Can J. Forest Res.*, 18(1), 68-74, 1988.
- 540 [Hendrickson O Q, Chatarpaul L, Burgess D. Nutrient cycling following whole-tree and conventional harvest in a northern mixed forest. *Can J For Res* 18: 1427-1436, 1989.](#)
- Hendrickson, O. Q. Asymbiotic nitrogen fixation and soil metabolism in three Ontario forests. *Soil Biol. Biochem.*, 22(7), 967-971, 1990.
- 545 Holzworth, D. P., Huth, N. I., deVoil, P. G., Zurcher, E. J., Herrmann, N. I., McLean, G., ... & Moore, A. D. APSIM—evolution towards a new generation of agricultural systems simulation. *Environ. Modell. Softw*, 62, 327-350, 2014.
- Houlton, B. Z., Morford, S. L., & Dahlgren, R. A. Convergent evidence for widespread rock nitrogen sources in Earth's surface environment. *Science*, 360(6384), 58-62, 2018.
- 550 Huss-Danell, K. Nitrogen fixation by *Stereocaulon paschale* under field conditions. *Can J. Botany*, 55(5), 585-592, 1977.
- Jarrell, W. M., & Virginia, R. A. Soil cation accumulation in a mesquite woodland: sustained production and long-term estimates of water use and nitrogen fixation. *J. Arid Environ.* 18(1), 51-58, 1990.
- Johnson, H. B., & Mayeux, H. S. *Prosopis glandulosa* and the nitrogen balance of rangelands: extent and occurrence of nodulation. *Oecologia*, 84(2), 176-185, 1990.
- 555 Kapustka, L. A., & DuBois, J. D. Dinitrogen fixation by cyanobacteria and associative rhizosphere bacteria in the Arapaho Prairie in the Sand Hills of Nebraska. *Am. J. Bot.*, 74(1), 107-113, 1987.
- Lambers, H., Raven, J. A., Shaver, G. R., & Smith, S. E. Plant nutrient-acquisition strategies change with soil age. *Trends Ecol. .Evol*, 23(2), 95-103, 2008.
- 560 LeBauer, D. S., & Treseder, K. K. Nitrogen limitation of net primary productivity in terrestrial ecosystems is globally distributed. *Ecology*, 89(2), 371-379, 2008.
- Lee, Y. Y., & Son, Y. Diurnal and seasonal patterns of nitrogen fixation in *alnus hirsuta* plantation of central Korea. *J. Plant Biol.*, 48(3), 332-337, 2005.
- Lepper, M. G., & Fleschner, M. Nitrogen fixation by *Cercocarpus ledifolius* (Rosaceae) in pioneer habitats. *Oecologia*, 27(4), 333-338, 1977.

- 565 Lett, S., & Michelsen, A. Seasonal variation in nitrogen fixation and effects of climate change in a subarctic heath. *Plant Soil*, 379(1-2), 193-204, 2014.
- Levy, H., Moxim, W. J., & Kasibhatla, P. S. A global three-dimensional time-dependent lightning source of tropospheric NO_x. *J Geophys. Res-Atmos.*, 101(D17), 22911-22922, 1996.
- 570 [Ley, R. E., D'Antonio, C.M. Exotic grass invasion alters potential rates of N fixation in Hawaiian woodlands. *Oecologia* 113:179-87. 1998.](#)
- Lindemann, William C., and C. R. Glover. Nitrogen fixation by legumes, 2003.
- [Luken, J. O. and R. W. Fonda. Nitrogen accumulation in a chronosequence of red alder communities along the Hoh River, Olympic National Park, Washington. *Can. J. For. Res.* 13:1228-1237, 1983.](#)
- 575 Maheswaran, J., & Gunatilleke, I. A. U. N. Nitrogenase activity in soil and litter of a tropical lowland rain forest and an adjacent fernland in Sri Lanka. *J Trop Ecol*, 6(3), 281-289, 1990.
- Marino, D., Frendo, P., Ladrera, R., Zabalza, A., Puppo, A., Arrese-Igor, C., & González, E. M. Nitrogen fixation control under drought stress. Localized or systemic?. *Plant Physiol.*, 143(4), 1968-1974, 2007.
- 580 May, D.E., and Webber, P.J., Spatial and temporal variation of vegetation and its productivity on Niwot Ridge, Colorado, in *Ecological Studies in the Colorado Alpine*, a Festschrift for John W. Mart, edited by H. Halfpenny, pp. 35-62, Institute for Arctic and Alpine Research, Univ. of Colorado, Boulder, Colo., 1982.
- Mitchell, T. D., & Jones, P. D. An improved method of constructing a database of monthly climate observations and associated high-resolution grids. *Int J Climatol. - A Journal of the Royal Meteorological Society*, 25(6), 693-712, 2005.
- 585 Montanez, A., Danso, S. K. A., & Hardarson, G. (1995). The effect of temperature on nodulation and nitrogen fixation by five *Bradyrhizobium japonicum* strains. *Appl. Soil Ecol.*, 2(3), 165-174.
- Morford, S. L., Houlton, B. Z., & Dahlgren, R. A. Increased forest ecosystem carbon and nitrogen storage from nitrogen rich bedrock. *Nature*, 477(7362), 78, 2011.
- 590 Mosier, A. R., Delgado, J. A., Cochran, V. L., Valentine, D. W., & Parton, W. J. Impact of agriculture on soil consumption of atmospheric CH₄ and a comparison of CH₄ and N₂O flux in subarctic, temperate and tropical grasslands. *Nutr. Cycl. Agroecosys.*, 49(1-3), 71-83, 1997.
- Mus, F., Crook, M. B., Garcia, K., Costas, A. G., Geddes, B. A., Kouri, E. D., ... & Udvardi, M. K. Symbiotic nitrogen fixation and the challenges to its extension to nonlegumes. *Appl. Environ. Microbiol.*, 82(13), 3698-3710, 2016.
- 595 [Nohrstedt HÖ. Biological activity in soil from forest stands in Central Sweden, as related to site properties. *Microb Ecol* :11\(3\):259-266. 1985.](#)
- O'Connell, A. M., & Grove, T. S. Seasonal variation in C₂H₂ reduction (N₂-fixation) in the litter layer of eucalypt forests of south-western Australia. *Soil Biol Biochem.*, 19(2), 135-142, 1987.
- 600 Omari, K., Mubyana, T., Matsheka, M. I., Bonyongo, M. C., Veenendaal, E., & Musil, C. F. Flooding and its influence on diazotroph populations and soil nitrogen levels in the Okavango Delta. *S Afr. J. Bot.*, 70(5), 734-740, 2004.

- Permar, T. A., & Fisher, R. F. Nitrogen fixation and accretion by wax myrtle (*Myrica cerifera*) in slash pine (*Pinus elliottii*) plantations. *Forest Ecol Manag*, 5(1), 39-46, 1983.
- 605 Pons, T. L., Perreijn, K., Van Kessel, C., & Werger, M. J. Symbiotic nitrogen fixation in a tropical rainforest: 15N natural abundance measurements supported by experimental isotopic enrichment. *New Phytol.*, 173(1), 154-167, 2007.
- Prentice, I. C. Modelling the demand for new nitrogen fixation by terrestrial ecosystems. *Biogeosciences*, 14(7), 2003-2017, 2017.
- 610 Reed, S. C., Cleveland, C. C., & Townsend, A. R. Functional ecology of free-living nitrogen fixation: a contemporary perspective. *Annu Rev Ecol. Evol. S.*, 42, 489-512, 2011.
- Rosswall T, Granhall U. Nitrogen cycling in a subarctic ombrotrophic mire. *Ecol. Bull.*: 209-234, 1980.
- Rundel, P. W., Nilsen, E. T., Sharifi, M. R., Virginia, R. A., Jarrell, W. M., Kohl, D. H., & Shearer, G. B.. Seasonal dynamics of nitrogen cycling for a *Prosopis* woodland in the Sonoran Desert. In *Nitrogen Cycling in Ecosystems of Latin America and the Caribbean* (pp. 343-353), 1982. Springer, Dordrecht..
- 615 Sánchez-Díaz, M. Adaptation of legumes to multiple stresses in Mediterranean-type environments. *Options Méditerranéennes*, 45, 145-151, 2001.
- Schlesinger, W. H., Gray, J. T., Gill, D. S., & Mahall, B. E. *Ceanothus megacarpus* chaparral: a synthesis of ecosystem processes during development and annual growth. *Bot. Rev.*, 48(1), 71-117, 1982.
- 620 Schlesinger, W. H. *Biogeochemical Cycles*. (Book Reviews: *Biogeochemistry. An Analysis of Global Change.*). *Science*, 253, 686-687, 1991.
- Schlesinger, W. H. On the fate of anthropogenic nitrogen. *P Natl. Acad.*, 106(1), 203-208, 2009.
- Schrire, B. D., G. P. Lewis, and M. Lavin. Biogeography of the Leguminosae. *Legumes of the world*: 21-54, 2005..
- Schwintzer, C. R. Nonsymbiotic and symbiotic nitrogen fixation in a weakly minerotrophic peatland. *Am. J. Bot.*, 70(7), 1071-1078., 1983.
- 625 Shearer, G., & Kohl, D. H. N₂-fixation in field settings: estimations based on natural 15N abundance. *Funct. Plant Biol.*, 13(6), 699-756, 1986.
- Sheridan, R. P. Nitrogenase activity by *Hapalosiphon flexuosus* associated with *Sphagnum erythrocalyx* mats in the cloud forest on the volcano La Soufriere, Guadeloupe, French West Indies. *Biotropica*, 134-140, 1991.
- 630 Skujinš, J., Tann, C. C., & Börjesson, I. Dinitrogen fixation in a montane forest sere determined by 15N₂ assimilation and in situ acetylene-reduction methods. *Soil Biol. Biochem.*, 19(4), 465-471, 1987.
- Sobota, D. J., Compton, J. E., & Harrison, J. A. Reactive nitrogen inputs to US lands and waterways: how certain are we about sources and fluxes?. *Front. Ecol. Environ.*, 11(2), 82-90, 2013.
- Sonesson, M., Jonsson, S., Rosswall, T., & Rydén, B. E. The Swedish IBP/PT Tundra Biome Project Objectives-Planning-Site. *Ecol. Bull.*, 7-25, 1980.
- 635 Sprent, J. I. The effects of water stress on nitrogen-fixing root nodules. *New Phytol.*, 71(3), 443-450, 1972.
- Sprent, J. I., Ardley, J., & James, E. K. Biogeography of nodulated legumes and their nitrogen-fixing symbionts. *New Phytol.*, 215(1), 40-56, 2017.

- Srivastava, A. K., & Ambasht, R. S. Soil moisture control of nitrogen fixation activity in dry tropical Casuarina plantation forest. *J. Environ. Manage.*, 42(1), 49-54, 1994.
- 640 Stedman, D. H., and R. Shetter, The global budget of atmosphere nitrogen species, in *Trace Atmospheric Constituent: Properties, Transformations and Fates*, edited by S.S. Schwartz, pp. 411-454, John Wiley, New York, 1983.
- 645 [Stewart, W. D. P., Sampaio, M. J., Isichei, A. O., and Sylvester-Bradley, R. Nitrogen fixation by soil algae of temperate and tropical soils. In: Dobreiner J, R H Burris and A hollaender \(eds\) Limitation and potentials for biological nitrogen fixation in the tropics. Basic Life Sciences, vol 10. Plenum Press, New York. Pp 41-63, 1978.](#)
- Sutton, Mark A., et al., eds. *Nitrogen deposition, critical loads and biodiversity*. Springer Science & Business Media, 2014.
- Sullivan, B. W., Smith, W. K., Townsend, A. R., Nasto, M. K., Reed, S. C., Chazdon, R. L., & Cleveland, C. C. (2014). Spatially robust estimates of biological nitrogen (N) fixation imply substantial human alteration of the tropical N cycle. *P. Natl. A. Sci.*, 111(22), 8101-8106, 2014.
- [Sylvester-Bradley, R. Proc. Int. Lupin Conf., 1st pp. 157-173, 1980.](#)
- Thornley, J. H. M. Simulating grass-legume dynamics: a phenomenological submodel. *Ann Bot-London*, 88(5), 905-913, 2001.
- 655 Vitousek, P. M. Potential nitrogen fixation during primary succession in Hawaii Volcanoes National Park. *Biotropica*, 234-240, 1994.
- Vitousek, P. M., Menge, D. N., Reed, S. C., & Cleveland, C. C. Biological nitrogen fixation: rates, patterns and ecological controls in terrestrial ecosystems. *Philos. T. R. Soc. B.*, 368(1621), 20130119, 2013.
- 660 Voisin, A. S., Salon, C., Jeudy, C., & Warembourg, F. R. Symbiotic N₂ fixation activity in relation to C economy of *Pisum sativum* L. as a function of plant phenology. *J. Exp. Bot.*, 54(393), 2733-2744, 2003.
- Voisin, A. S., Bourion, V., Duc, G., & Salon, C. Using an ecophysiological analysis to dissect genetic variability and to propose an ideotype for nitrogen nutrition in pea. *Ann Bot-London*, 100(7), 1525-1536, 2007.
- Wheatley, Ron E., and J. I. Sprent. *Legume Nodulation: A Global Perspective*. *Exp. Agr.* 46.4: 568, 2010.
- 665 Whitehead, D.C., *Grass Nitrogen*, 397pp., CAB Int., Wallingford, UK
- Williams, J. R. Sharply A N. EPIC-Erosion Productivity Impact Calculator | . Model Documentation. US Department of Agriculture Technical Bulletin, (1990), 1768.
- Woodmansee, R. G., & Wallach, L. S. Effects of fire regimes on biogeochemical cycles. *Terrestrial Nitrogen Cycles. Ecol. Bull.(Stockholm)*, 33, 649-669, 1981.
- 670 Wu, L., & McGechan, M. B. Simulation of nitrogen uptake, fixation and leaching in a grass/white clover mixture. *Grass Forage Sci.*, 54(1), 30-41, 1999.
- Yu, T. and Q. Zhuang. Quantifying global N₂O emissions from natural ecosystem soils using trait-based biogeochemistry models. *Biogeosciences* 16(2): 207-222, 2019.

- 675 Zhuang, Q., Romanovsky, V. E., & McGuire, A. D. Incorporation of a permafrost model into a large-scale ecosystem model: Evaluation of temporal and spatial scaling issues in simulating soil thermal dynamics. *J. Geophys. Res-Atmos.*, 106(D24), 33649-33670, 2001.
- Zhuang, Q., McGuire, A. D., O'Neill, K. P., Harden, J. W., Romanovsky, V. E., & Yarie, J. Modeling soil thermal and carbon dynamics of a fire chronosequence in interior Alaska. *J. Geophys. Res-Atmos.*, 107(D1), FFR-3, 2002.
- 680 Zhuang, Q., McGuire, A. D., Melillo, J. M., Clein, J. S., Dargaville, R. J., Kicklighter, D. W., ... & Hobbie, J. E. Carbon cycling in extratropical terrestrial ecosystems of the Northern Hemisphere during the 20th century: a modeling analysis of the influences of soil thermal dynamics. *Tellus B.*, 55(3), 751-776, 2011.
- Zhuang, Q., Lu, Y., & Chen, M. An inventory of global N₂O emissions from the soils of natural terrestrial ecosystems. *Atmos. Environ.*, 47, 66-75, 2012.
- 685 Zhuang, Q., Chen, M., Xu, K., Tang, J., Saikawa, E., Lu, Y., ... & McGuire, A. D. (2013). Response of global soil consumption of atmospheric methane to changes in atmospheric climate and nitrogen deposition. *Global Biogeochem. Cy.*, 27(3), 650-663, 2013.

Table 1. Description of parameters used in the model

Parameters	Description	Unit	Reference Value	reference
N_fix	nitrogen fixation rate	g N m ⁻² day ⁻¹		
N_fixpot	potential nitrogen fixation rate	g N m ⁻² day ⁻²	0.01-1×10-3	Thornley (2001); Eckertsten et al.(2006); Corre-Hellou et al. (2007); Corre-Hellou et al. (2009);
ft	soil temperature factor	°C		
t_min	the minimum temperature for the start of N fixation	°C	0.5~5	Boote et al. (2008)
t_max	the maximum temperature for the stop of N fixation	°C	40~45	Boote et al. (2008)
t_optL	lower threshold of optimal temperature	°C	10~20	Boote et al. (2008)
t_optH	upper threshold of optimal temperature	°C	25~35	Boote et al. (2008)
fw	soil water factor			
φ1	coefficient for soil moisture		0	
φ2	coefficient for soil moisture		2	APSIM, EPIC (Sharpley and Williams, 1990; Bouniols et al., 1991; Cabelguenne et al., 1999); SOILN (Wu and McGechan, 1999)
Wa	lower threshold of water content below which N fixation is totally restrict by the deficit of soil water		0	APSIM, EPIC (Sharpley and Williams, 1990; Bouniols et al., 1991; Cabelguenne et al., 1999); SOILN (Wu and McGechan, 1999)
Wb	upper threshold of water content above which N fixation is not limited by the deficit of soil water		0.5	APSIM, EPIC (Sharpley and Williams, 1990; Bouniols et al., 1991; Cabelguenne et al., 1999); SOILN (Wu and McGechan, 1999)
Wf	available soil water content to that at field capacity			
f_Nup	parameter relating legume biological nitrogen fixation and soil nitrogen content		0.01~0.1	SOILN model (Wu and McGechan, 1999)
Ns	Soil mineral nitrogen content	g N m ⁻²		
f_N	Soil mineral N effect			
f_C	Soil carbon effect			
Cr	Carbon concentration in the soil	g C g ⁻¹ soil		
Kc	Michaelis-Menten Constant for carbon	g C m ⁻²	0.001~0.01	Thornley (2001); Eckertsten et al. (2006)

Table 2. Calibration Sites of Biological Nitrogen Fixation Rate for Representative Ecosystems

Site name	Ecosystem Type	Lon.	Lat.	Experimental method	Reference	N Fixation Rate kg N ha ⁻¹ yr ⁻¹	Simulation, kg N ha ⁻¹ yr ⁻¹
Stordalen, Sweden	Tundra	18	68	ARA*	Christie (1987); Sonesson et al. (1980)	2	2.5
Truelove Lowland, Canada	Tundra	-84.5	75.5	ARA	Chapin et al. (1990)	3	2.8
Niwot Ridge, Colorado, US	Tundra	-105.5	40	¹⁵ N	Bowman et al. (1996)	4.9	5.1
Central Sweden	Boreal Forest	18	60	ARA	Nohrstedt (1985)	0.93	0.9
PNFI, Ontario, Canada	Boreal Forest	-77	45.5	ARA	Hendrickson (1990)	0.25	1.2
Southern British Columbia, Canada	Boreal Forest	-119	49	ARA	Hendrickson and Burgess (1989)	2.8	2.2
Robson moraines, British Columbia	Boreal Forest	-119	53	ARA	Blundon and Dale (1990)	1.1	1.2
Umea, Sweden	Boreal Forest	19.5	64	ARA	Huss-Danell (1976)	1	1.5
Coweeta Basin,	Temperate Forest	-83	35	N accumulation	Boring and Swank (1984)	48	19.5
Hoh River, Washington, US	Temperate Forest	-123.5	48	ARA	Luken and Fonda (1983)	40	13
Tom Swamp, Massachusetts, US	Temperate Forest	-75	42.5	unspecified	Schwinzer (1983)	35	25.7
Big Creek Basin, Melbourne, Australia	Temperate Forest	145.5	38	ARA	Adams and Attiwill (1984)	24	23.2
Jebo Creek, Utah, US	Temperate Forest	-112	42	¹⁵ N	Skujins et al. (1987)	10.2	12.5
Karri Forest, south-western Australia	Temperate Forest	116	-34.5	ARA	Grove and Malajczuk (1992)	7.93	8.5
Woodhill Forest, New Zealand	Temperate Forest	174.5	-37	N accumulation	Baker et al. (1986)	80	23.5
Gainesville, Florida, US	Temperate Forest	-82	30	N accumulation	Permar and Fisher (1983)	10.6	12.8
Fox park, Wyoming, US	Temperate Forest	-106	41	ARA	Fahey et al. (1985)	13	12.5
Mount Robson, Canada	Temperate Forest	-119	53.1	ARA	Blurdon and Dale (1990)	1.65	3.2
Dwellingup, South-western Australia	Temperate Forest	116	33	ARA	O'Connel and Grove (1987)	2.5	3.1
Adair, Oregon, US	Temperate Forest	-123	44.6	ARA	Heath et al (1988)	0.74	2.4

Priest River Experimental Forestry, Idaho, US	Temperate Forest	-116	48	ARA	Harvey et al (1989)	0.1	1.7
Arapaho Prarie, Nebraska, US	Grassland	-100	42	unspecified	Kaputsa and DuBois (1987)	0.2	0.7
Lynx Prairie Preserve, Ohio, US	Grassland	-83.5	39	ARA	DuBois and Kaputsa(1983)	8.2	1.9
Konza Prarie Research Natural Area, Kansas, US	Grassland	-96	39.5	nitrogenase activity	Eisele et al (1989)	21	3.3
Buso, Papua New Guinea	Tropical Forest	147	-7.5	ARA	Goosem and Lamb (1986)	0.5	5.2
Reserve Ducke, Manaus, Brazil	Tropical Forest	-59	-3	ARA	Sylvester-Bradley et al. (1980)	2.45	3.5
Sinharaja Man and Biosphere reserve	Tropical Forest	80.5	6.5	ARA	Maheswaran and Gunatilleke (1990)	8	8.5
Amazon Territory of Venezuela	Tropical Forest	-67	2	ARA	Jordan et al (1983)	32	20.3
Kilauea, Hawaii, US	Tropical Forest	-155	19	ARA	Vitousek (1994)	2.8	18.5
Volcano La Soufriere, Guadeloupe	Tropical Forest	-61.5	16	ARA	Sheridan (1991)	4.02	7.2
Hawaii Volcanoes National Park, US	Tropical Forest	-155	19.5	ARA	Ley and D'Antonio (1998)	4.9	9.3
Santa Ynez Mountain, California, US	Mediterranean Shrubland	-120	34.5	ARA	Schlesinger et al.(1982)	1	2.4
San Bernardino Mountains, California, US	Mediterranean Shrubland	-116.5	34	ARA	Lepper and Fleschner (1977)	6.9	3.7
Harpers Well, California, US	Xeric Shrubland	-116	33.5	N accumulation	Rundel et al.(1982)	30	18.5
Sonoran Desert, Arizona, US	Xeric Shrubland	-112.5	33	cation accumulation	Jarrell and Virginia (1990)	40	23.5

*ARA denotes the acetylene reduction assay method in determining biological N₂ fixation rates.

Table 3. Model estimated biological nitrogen fixation in global natural terrestrial ecosystems

Ecosystem	Average coverage of N_2 -fixing plants	Coverage range	Reference	N Fixation Rate (kg N_2 -N ha ⁻¹ yr ⁻¹)	Total_Min (Tg N yr ⁻¹)	Total_Max (Tg N yr ⁻¹)	Total_Avg (Tg N yr ⁻¹)	Area (10 ⁸ ha)
wet tundra	9%	3%~15%	May and Webber (1982)	3.2	0.51	2.55	1.54	5.37
alpine tundra & wet tundra	9%	3%~15%	May and Webber (1982)	3.2	0.51	2.55	1.54	5.36
boreal forest	9%	4%~18%	Alexander and Billington (1986); weber and Van Cleve (1981)	2.1	2.01	9.06	4.53	19.3
temperate coniferous forest	5%	1%~10%	Cleveland et al (1999)	12.7	0.71	7.15	3.5	5.51
temperate deciduous forest	5%	1%~10%	Cleveland et al (1999)	12.7	0.76	7.65	3.75	5.89
temperate evergreen forest	5%	1%~10%	Cleveland et al (1999)	12.7	0.43	4.34	2.13	3.35
grassland	15%	5%~25%	Woodmansee et al (1981); Robertson and Rosswall (1986)	1.9	0.61	3.1	1.86	8.4
tropical forest	15%	5%~25%	Cleveland et al (2001)	18.2	10.8	54	32.6	17.8
xeric shrubland	15%	10%~20%	Johnson and Mayeux (1990)	5.7	2.92	14.6	8.35	14.8
Mediterranean shrubland	15%	10%~20%	Johnson and Mayeux (1990)	2.7	0.13	0.66	0.4	1.47
savanna	15%	5%~25%	Stewart et al (1978); Bate and Gunton (1982)	1.9	0.45	2.23	1.34	7.05
Total					19.84	107.89	61.54	94.3

Table 4. Model parameters for various natural terrestrial ecosystems

	N_pot (g N₂ N fixed day ⁻¹)	t_optL (°C)	t_optH (°C)	W_upH (J kg ⁻¹)	fNup	Kc (g C m ⁻²)
1 wet tundra	0.028	10	25	0.8	65	0.002
2 alpine tundra & wet tundra	0.028	10	25	0.8	65	0.002
3 boreal forest	0.032	12	25	0.8	70	0.008
4 temperate coniferous forest	0.55	16	35	0.6	80	0.01
5 temperate deciduous forest	0.55	18	35	0.6	80	0.01
6 temperate evergreen forest	0.55	18	35	0.6	80	0.01
7 grassland	0.05	18	35	0.5	60	0.012
8 tropical forest	0.8	20	35	0.8	100	0.005
9 xeric shrubland	0.7	15	35	0.4	65	0.016
10 Mediterranean shrubland	0.08	19	35	0.5	65	0.016
11 savanna	0.05	20	35	0.5	60	0.012

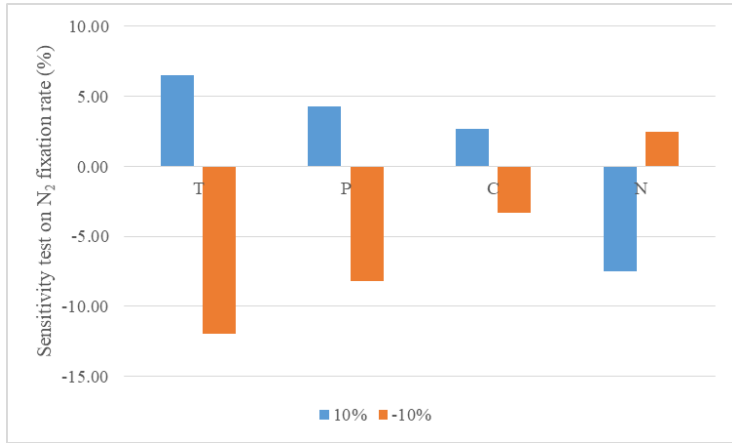
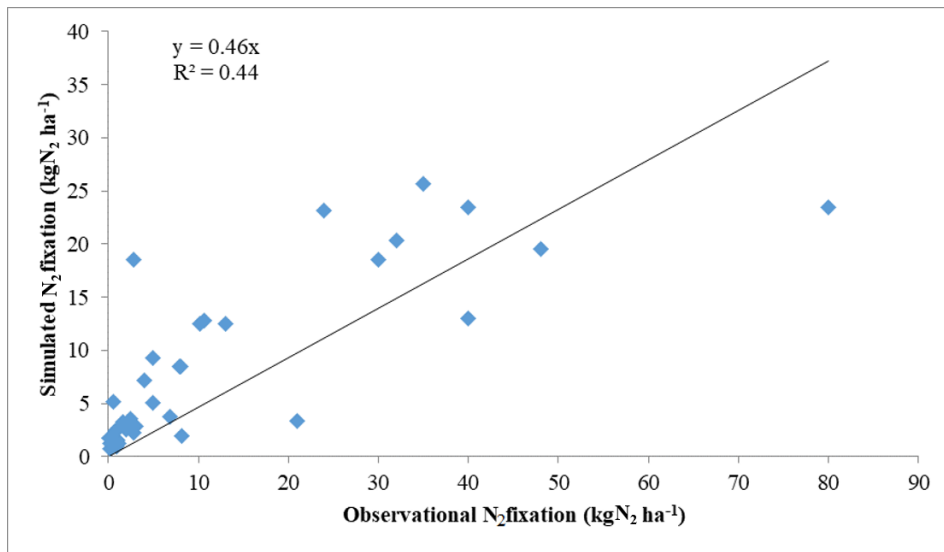
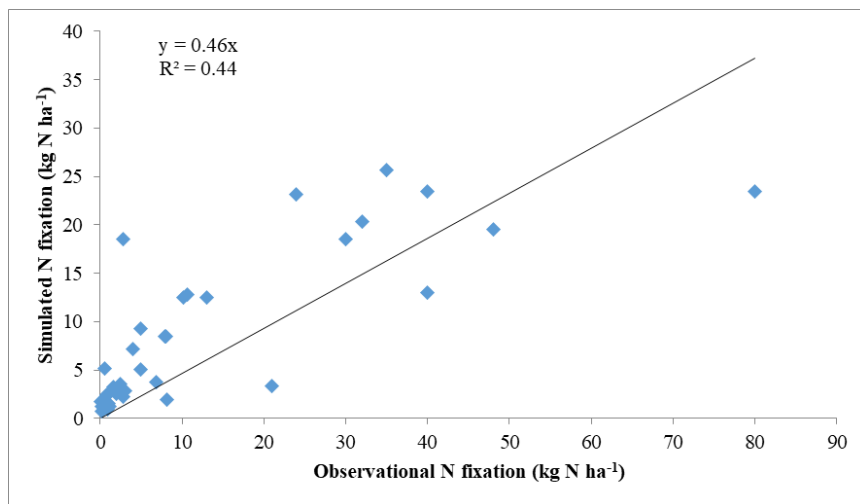


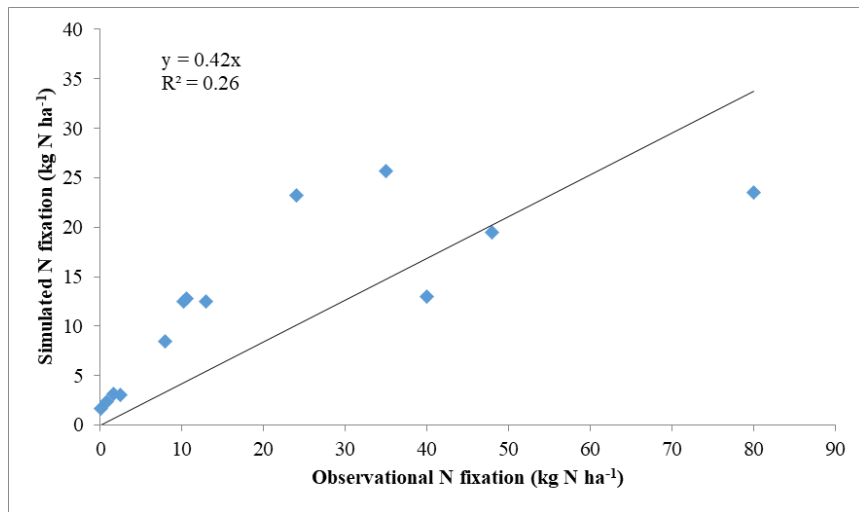
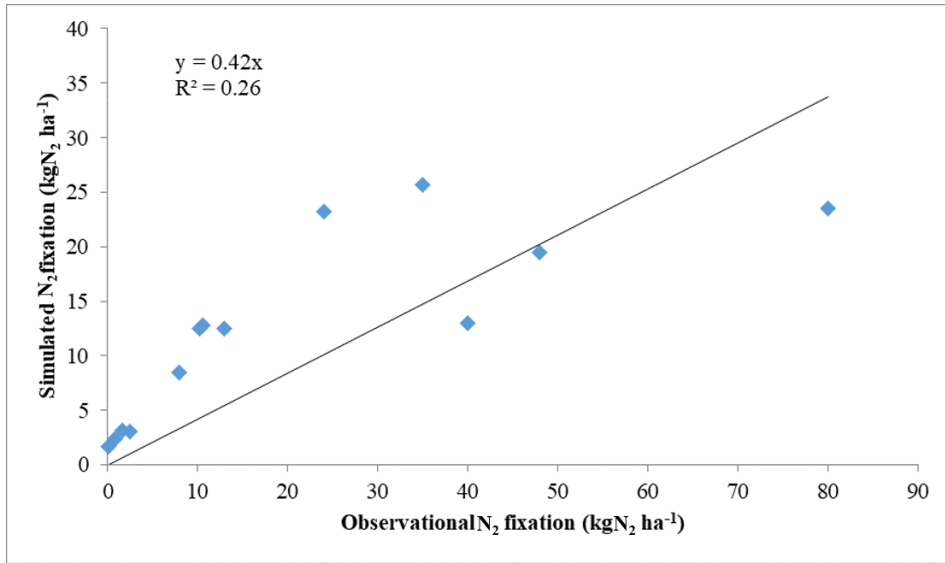
Figure 1. Model sensitivity of N_2 fixation in natural terrestrial ecosystems to changing model input data: Increasing or decreasing each variable by 10% for air temperature (T), precipitation (P), soil carbon content (C), soil nitrogen content (N) for N_2 fixation rate.

(a)



(b)





(c)

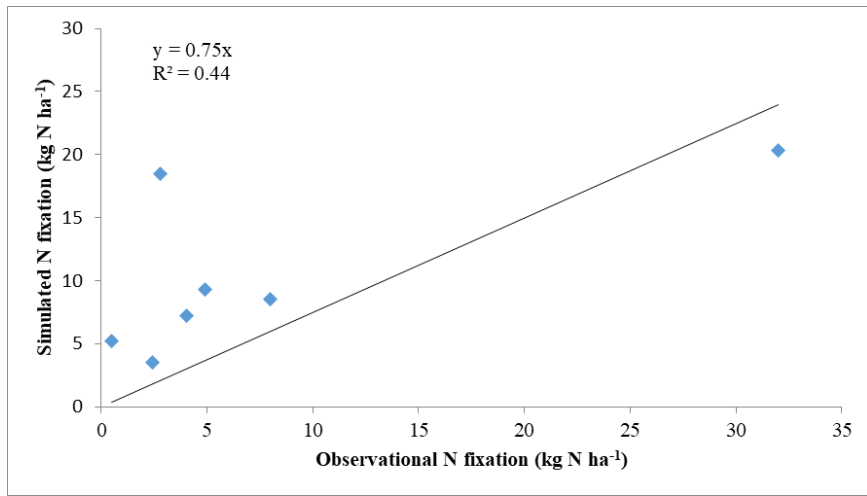
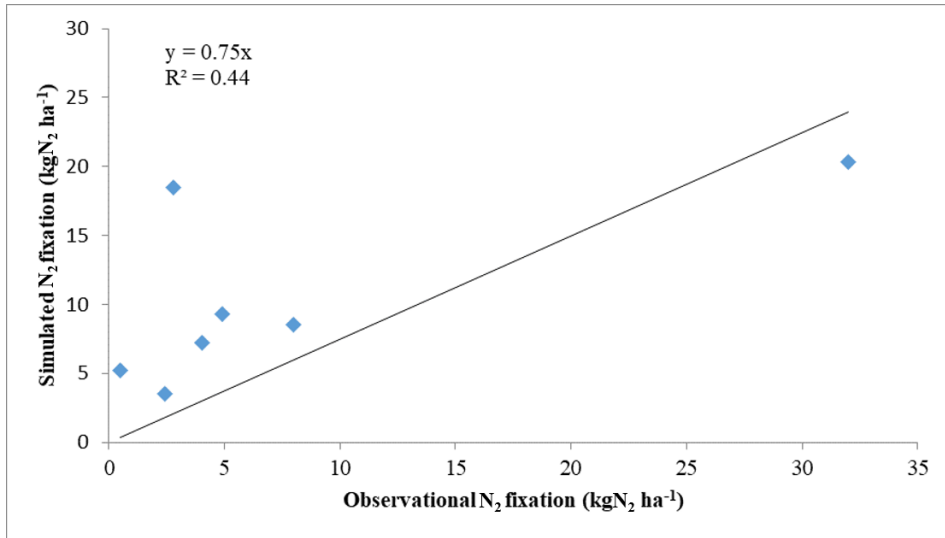


Figure 2. Comparison between Modeled and Observed Nitrogen Fixation Rate at site level: (a) All sites, (b) Temperate Forest, (c) Tropical Forest (data listed in Table 2). Y is simulated N₂ fixation while X represents the observational N₂ fixation.

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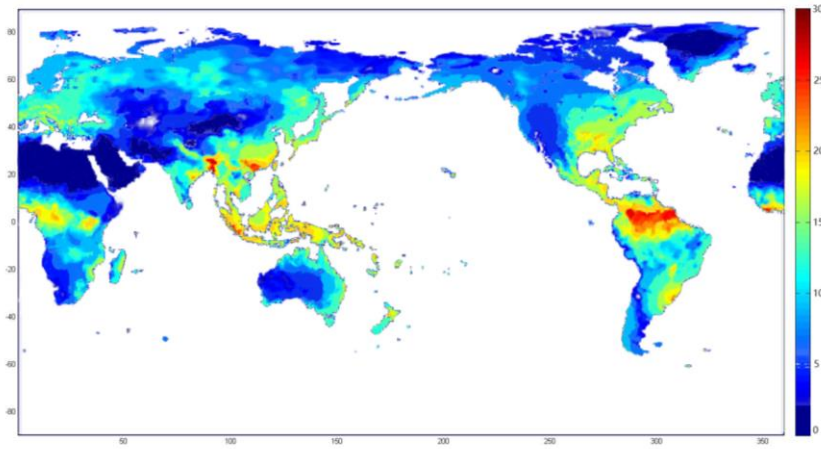


Figure 3. Simulated spatial distribution of BNF rates ($\text{kg N}_2 \text{ ha}^{-1} \text{ yr}^{-1}$) in natural terrestrial ecosystems during 1990-2000 by considering the BNF effects.