Authors' response to referee's comments (RC1) on the manuscript, "Organic matter and sediment properties determine in-lake variability of sediment CO2 and CH4 production and emissions of a small and shallow lake" by L.S.E. Praetzel et al.

Dear reviewer,

thank you very much for your comments and suggested improvements on our manuscript "Organic matter and sediment properties determine in-lake variability of sediment CO2 and CH4 production and emissions of a small and shallow lake". We are sure that consideration of your raised points and ideas will considerably improve the manuscript and therefore we will especially focus on revising the discussions part. Please find our responses to each of your comments below, they will be structured as follows:

(1) comments from referee, (2) author's response, (3) author's changes in manuscript.

We hope to be able to answer your open questions and to adequately include your suggestions.

Kind regards, Leandra Praetzel & Co-Authors

Major points:

(1) 1.L.79-81: The Gibbs free energies given in the ms are either not found in the quoted literature (Whiticar 1999) or are different (Conrad 1999). I assume the reason is that they were calculated using energies of formation for gases in dissolved rather than in gaseous state. This would be consistent with the Nernst equations mentioned later (L.250) also probably using gas concentrations rather than partial pressures. However, the authors should clarify the procedures.

(2) Gibb's free energies were calculated using formation energies Gf0 for the elements involved in the reactions in aqueous state. Used Gf0 values are reported by Stumm & Morgan 1996 and Nordstrom & Munoz 1994. In the Nernst equation, dissolved and gaseous concentrations in incubation vials were used for calculations. This will be mentioned in the revised manuscript.

We will include the following explanation on calculation of delta Gr0 in the revised manuscript:

(3) "Values for Δ Gr0 are calculated from standard formation energies Δ Gf0 at 25°C in aqueous state listed in Stumm and Morgan (1995) and Nordstrom and Munoz (1994)."

(1) 2.L.187-200: There are no isotopic data reported, therefore the description of IRMS methodology is not necessary.

(2) delta 13C and delta 15N values are reported in Table 2 and will be discussed in more detail in the results section of the revised manuscript. IRMS was also used to determine mass contents of C and N. The authors therefore prefer to leave the methodology description in the manuscript.

(3) "C and N isotopic values did not vary much between sites and were on average -27.6 % and -0.9 % respectively with only one outlier for δ 15N at site 3.50 (-6.3 %)."

"Neither CO2 nor CH4 production rates were exhibited significant correlation with C content or, C/N ratio, δ 13C or δ 15N,..."

(1) 3. Table 2: L.411-412 mentions strong FTIR absorption features of polysaccharides. However, this compound class is not listed in the Table.

(2) Absorption maxima for polysaccharides will be listed in Table 2 of the revised manuscript.

(1) 4.L.422-425. This is an overview of measured rates. However, the numbers seem to be slightly different from those shown in Fig.2. Although there is probably a reasonable explanation for these differences, I found it confusing. In fact I would be happy just looking at the data in the figure without reading the text. However, one could mention that the rates decreased from the shore to the centre, since this point is later relevant in the Discussion.

(2) We double-checked the accordance of production rates stated in the text and displayed in Figure 2. We confirm that values in the text are the same as in the figure. We note that production rates in the text are mean values, whereas bold lines of boxplots are median values. We suppose that this fact might have caused the perceived discrepancy.

Moreover, we will simplify the figure by removing data from 5-10 cm depth and instead include this data in the supporting information.

We will further add a sentence to emphasize that production rates increased from the center to the shore as follows:

(3) "Overall, production rates decreased from the shore to the center of the lake."

(1) 5.L.477-487: Here applies the same as in point 4. The data in the text seem to slightly different from those seen in Fig. 7.

(2) See also answer to point 4. We double-checked the data and confirm that the values in text and figure are the same. Please note that the stated values in the text are mean values +/- standard deviations of triplicate measurements at the respective sites where minima and maxima were measured whereas boxplots in the figure represent the whole dataset at each sampling date.

(1) 6.L.507: Again the data in the text seem to slightly different from those seen in Fig. 8.

(2) We will adjust the mentioned values in the text according to the displayed flux rates in Fig. 8 as follows:

(3) "CO₂ and CH₄ fluxes measured from intact sediment core incubations ranged from 10.8 \pm 4.4 to 17.9 \pm 2.0 and 0.02 \pm 0.01 to 1.5 \pm 2.6 mmol m⁻² d⁻¹ respectively."

(1) 7. The discussion is too wordy and should be focused to the really novel results. I also recommend a different structure for the Discussion. I think it is not ideal having individual chapters on spatial variability of OM quality, spatial variability of CO2 and CH4 production rates, and influence of OM quality on gas production, since such structure results in too much repetition and also is not very suitable for explaining gas production rates on the basis of OM quality.

(2) We will shorten the discussion and focus on novel results in the revised manuscript. The chapters 4.1.1, 4.1.2 and 4.1.3 will be summarized as one, entitled "4.1.1 Variability of CO2 and CH4

production rates and influence of OM". Like this, we will considerably shorten the paragraph and thus avoid repetitions.

(1) 9. The discussion on temperature effects can be much shorter, since it is rather well reported in the literature.

(2) In the discussion on temperature effects, we emphasize the sensitivity of small lakes to temperature changes compared to large lakes. We will shorten that section in the revised manuscript but prefer to leave that statement in the discussion, as this strong and variable temperature effect on production rates has, to our knowledge, not been shown before for small and shallow lakes.

(3) "In accordance with previous studies (den Heyer and Kalff, 1998; Sobek et al., 2009; Gudasz et al., 2015), we found that with a temperature increase of 10°C, production rates of CO2 doubled and those of CH4 were 2 to 11 times higher. Q10-values for CO2 were thus within the range of earlier reported values by Liikanen et al. (2002) and Berström et al. (2010), whereas Q10-values for CH4 production were slightly higher than values found by Duc et al. (2010). The large observed range of Q10-values, especially for CH4, implies that responses to temperature increases might not be homogeneously distributed within a lake. We point out that sediment CH4 production is more sensitive to increasing temperatures compared to CO2 production and that this leads to a stronger feedback on global warming when considering the higher global warming potential of CH4 compared to CO2 (Marotta et al., 2014). The observed negative correlation between Q10-values and FTIR peak ratios further suggests that sites with more labile OM are more susceptive to increasing temperatures in terms of CH4 production, whereas at sites with more recalcitrant OM, this recalcitrant OM may limit the degradation processes. We therefore assume that sediment greenhouse gas production in small and shallow lakes might in the course of global warming increase to a larger extent than in deeper lakes, as shallow waters, compared to deeper lakes, do not get thermally stratified in summer and therefore shallow sediments warm much faster (Jankowski et al., 2006)."

(1) 10. The discussion of methanogenic pathways (L.648-680) is not really relevant, since the data just show that both methanogenic pathways were exergonic and thus, could well operate. Everything else is speculation and not relevant. The magnitude of the Gibbs free energy does not allow to conclude whether the one pathway is more prevalent than the other. One could however discuss the correlation of the concentrations of H2 and acetate, and the respective Delta G, with sediment OM quality, since correlations were reported in the Results.

(2) We will shorten the discussion of methanogenic pathways and increase the focus on the reported relationships between Gibb's free energies and OM quality in the revised manuscript.

(3) "[...] suggesting that both pathways could contribute to CH4 production during the whole experiment. Still, this approach does not allow to evaluate which of the pathways predominates."

"[...] we would have expected a reverse pattern. Concomitantly, Gibb's free energy of hydrogenotrophic methanogenesis exhibited significant positive correlations with some FTIR peak ratios, although we expected that a high abundance of recalcitrant OM compounds would make hydrogenotrophic methanogenesis more feasible (Miyajima et al., 1997). Acetate and H2 concentrations on the other hand, both exhibited significant negative correlations with some FTIR peak ratios. While this seemed reasonable for acetate concentrations (less acetate available in strongly decomposed OM), this result again proved to be against our expectations in terms of H2 concentrations. One reason for these ambiguous findings might be that the system was not in a

steady state in terms of thermodynamic conditions. Similarly, long time scales of experiments still not reaching such steady state have also been observed earlier for peat columns (Bonaiuti et al., 2017)."

(1) 11. The discussion of alternative electron acceptors (L.682-698) is rather short. The authors only discuss correlations. They miss the chance to discuss stoichiometric relations of reduced EAC with the amounts of CO2 production. Although such mass comparisons apparently have recently been done by other members of the Knorr group (Gao et al. 2019), they would also be interesting for this particular lake. I have the impression that the magnitudes of reducible EACs might explain the CO2 production in the beginning of the incubations, when rates of CO2 production were larger than those of CH4 production, while methanogenic decomposition of OM should result in equal rates. I wonder why this point is not addressed.

(2) In the revised manuscript, we will, besides observed correlations, also discuss the stochiometric relationships between measured EAs and CO2 production: We find that calculated CO2 production from prevalent EAs is lower than the measured CO2 production (see. Fig.). We propose this is due to unknown consumed EAs during the incubation that we did not capture; most likely iron in the solid phase. We indeed measured solid phase iron, but are not able to make statements about its speciation. But starting from the total Fe concentration in the solid phase (2-3%), we suppose that this is high enough to explain the missing EAs to reach a 1:1 ratio of measured CO2 production and calculated EA turnover.



(1) 12. I noticed that lake sediments were anoxically preincubated for either one week (L:178) or 50 days (L.331). Please clarify! Anyway, the preincubation might have depleted most of the reducible iron and sulfur compounds. This may be the explanation for the low values of EACinorg (Fig. 7), but is not discussed.

(2) The incubations were pre-incubated for one week, data from the sediment mesocosms was only used for analysis after 50 days of deployment in the climate chamber, in order the leave the cores adapt to laboratory conditions. We verified steady state conditions via the gas concentration in the sediment profile and observed constant conditions (i.e. no increase of concentrations) after 50 days. We will give a more detailed explanation on the procedure concerning sediment mesocosms in in the revised manuscript:

(3) "For statistical analyses and discussion, we only used measurements that were made >50 days after the deployment of the intact sediment core incubations in the climate chamber. This was done in order to ensure the system had adapted to experimental conditions and had reached a steady state. Steady state conditions were indicated by quasi-constant CO_2 and CH_4 concentrations in the sediment."

(2) We will additionally change the subtitle of the sections to clarify the difference between incubations and sediment mesocosms:

- (3) 2.3 Intact sediment core incubations
 - 2.3.1 CO₂ and CH₄ fluxes
 - 2.3.2 Sediment gas stock change

(2) Further, as suggested by the other reviewer, we will rename the sediment incubations to "slurry incubations" and the sediment mesocosms to "intact sediment core incubations".

(2) The low EACinorg values due to 1 week of preincubation will be discussed in the revised manuscript.

(3) "Nevertheless, both absolute EAC_{inorg} values as well as relative changes were very low, which might have been caused by the one-week preincubation, where most of the reducible inorganic compounds might have already been depleted."

Minor:

(1) 1.L.28: what means 'sufficiently' ? rho=0.65 is sufficient? Would rho=0.6 also be sufficient. Is there an objective criterion for sufficiency?

(2) The word sufficiently will be removed from the text.

(1) 2.L.30-32. I cannot follow the argument of this sentence. I suggest rephrasing.

(2) The sentence will be rephrased in the revised manuscript as follows:

(3) "Our results show that within a small lake, CO2 and CH4 production show significant spatial variability, which is mainly driven by spatial differences in the degradability of the sediment OM."

(1) 3.L.67: cellulose is also a polysaccharide. I suggest rephrasing.

(2) Cellulose will be replaced by natural organic matter in the revised manuscript.

(1) 4.L.83: The Delta G-zero of hydrogenotrophic methanogenesis is more negative than of aceticlastic methanogenesis. Therefore the acetoclastic pathway is less (not more) energetically favorable.

(2) This will be corrected in the revised manuscript.

(1) 5.L.91: EAC has not yet been defined. Please check also for other abbreviations.

(2) This will be corrected in the revised manuscript. We will also check for other abbreviations and make sure that they will be defined when being used the first time and that afterwards, abbreviations will be used consistently throughout the whole manuscript.

(1) 6.L.107-109: The '4' in CH4 as superscript

(2) This will be corrected in the revised manuscript.

(1) 7.L.168: 12 locations; please harmonize with the 13 sampling sites mentioned in the legend of Fig.1.

(2) Samples for incubations were taken from 12 sampling site, samples for sediment mesocosms were taken from 4 sites, whereas one of the mesocosms (S.150) represents the three sites of 150 cm depth from the incubations (1.150, 2.150 and 3.150) so that there are in total 13 sampling sites. This will be explained in more detail in the caption of Fig. 1 as follows:

(3) "Location of the study area and 13 sampling sites within Lake Windsborn. Rhombus: Sampling sites for slurry incubations and intact sediment core incubations, circles: sites for slurry incubations only, asterisk: site for intact sediment core incubation only (reference for 1.150, 2.150 and 3.150)."

(1) 8.L.207: 'relative abundance' compared to what?

(2) FTIR absorption peaks show relative abundances of the corresponding functional moieties in a single sample compared to another moiety. We therefore calculate peak ratios normalized to the polysaccharide peak. The sentence will be changed as follows in the revised manuscript:

(3) "Distinct peaks at specific wavelengths were assigned to functional groups according to Artz et al. (2008) and normalized to the peak intensity at 1031 - 1035 cm-1 (indicative of polysaccharides) in order to obtain inter-comparable peak-ratios of functional moieties in all samples as FTIR spectra only provide information about the relative abundance of certain functional moieties in one sample."

(1) 9.L.268: EAC/EDC: I think you mean EAC & EDC rather than the ratio between both. I found similar possible confusions at many places in the text (e.g., L.293, L.369, 370, 383 and in the labels of Fig. 7. Please check carefully.

(2) Notations will be changed in the revised manuscript as suggested by the reviewer.

(3) (EAC_{OM}, and EDC_{OM})

 $EAC_{OM} (EDC_{OM})$ $\Delta CO_2 = ((c(CO_2)_{end} * V_{seg}) - (c(CO_2)_{start} * V_{seg})) / \Delta t$ $\Delta CH_4 = ((c(CH_4)_{end} * V_{seg}) - (c(CH_4)_{start} * V_{seg})) / \Delta t$ $\Delta CO_2 (\Delta CH_4)$ total C and NFig. 7 label: EAC & EDC (µmol e⁻ gC⁻¹)

(1) 10. L.299. The reference Tamura et al. (1974) only describes the analysis of Fe(II) (albeit in the presence of Fe(III)). How was Fe(III) analyzed?

(2) Fe (III) in the samples was reduced to Fe (II) with 10% ascorbic acid and determined likewise. The procedure will be explained in the revised manuscript as follows:

(3) "Because 1,10-phenanthroline can only detect Fe^{2+} , the Fe^{3+} in the samples was reduced to Fe^{2+} with 10% ascorbic acid. Then, the determined concentration of total Fe was used to calculate the concentration of Fe^{3+} in the samples."

(1) 11. L.477-479: I cannot follow this sentence. Also compare major point 6 above. Please also note, that Fig. 7 is not mentioned in the text, and that Figure number should be exchanged with that of Fig. 6, since Fig.6 is reported later in the text than Fig. 7.

(2) The order of the figures will be changed and mentioning and numbering of figures will be adjusted in the revised manuscript. The sentence in II. 447-449 will be changes as follows:

(3) "EAC_{OM} lay between 218.69 ± 97.15 and 545.71 ± 60.33 μ mol e⁻ gC¹ at t0 and decreased on average by 44.85 μ mol e⁻ gC¹ until t6. Highest values for EAC_{OM} were found at site 3.125 corresponding to lowest measured CH₄ production rates at that site."

(1) 12.Table 3: Showing the time line as t0, t1, t2 etc. is awkward, since one has to consult the explanation in the methods section. I suggest listing the actual time points, i.e. 0, 1, 3 etc. days.

(2) The captions in Table 3 will be changed as suggested by the reviewer.

(1) 13. Table 4: The numbers in the table show too many decimal positions. Please report only those that are significant. In fact, at numerous places in the text numbers seem to show non-significant decimal positions. Please check and correct.

(2) The numbers in Table 4 as well as other number with non-significant decimal positions will be changed as suggested by the reviewer.

		CH₄ flux		CO ₂ flux			
	rho	р	n	rho	р	n	
Clay	0.648	< 0.05	12	0.605	< 0.05	12	
Silt	0.497	n.s.	12	0.302	n.s.	12	
Sand	-0.648	< 0.05	12	-0.605	< 0.05	12	
Fats, waxes, lipids	-0.833	< 0.05	8	-0.333	n.s.	8	
Phenols; humics	-0.833	< 0.05	8	-0.357	n.s.	8	
Aromates	-0.595	n.s.	8	-0.524	n.s.	8	
Lignin	-0.786	< 0.05	8	-0.381	n.s.	8	
C/N	-0.881	< 0.01	8	-0.333	n.s.	8	
C (%)	-0.714	n.s.	8	-0.190	n.s.	8	
CH ₄ sediment stock change	-0.222	n.s.	41	0.05	n.s.	35	
CO ₂ sediment stock change	-0.049	n.s.	41	-0.064	n.s.	35	

(3)

- (1) 14.L.535, 538: Should be Table 4 rather Table 5.
- (2) The numbering will be changed in the revised manuscript.
- (1) 15. References. Some of the references use capital letters for the titles.
- (2) Capitals will be changed according to the reviewer's suggestion.

Authors' response to referee's comments (RC2) on the manuscript, "Organic matter and sediment properties determine in-lake variability of sediment CO2 and CH4 production and emissions of a small and shallow lake" by L.S.E. Praetzel et al.

Dear reviewer,

Thank you very much for your comprehensive comments on our manuscript "Organic matter and sediment properties determine in-lake variability of sediment CO2 and CH4 production and emissions of a small and shallow lake". We appreciate your suggestions both in form and content and believe that they will substantially improve the paper. In the following, we will outline our responses to your comments one by one and thereby hope to clarify open questions and incorporate your suggestions to your satisfaction.

Each answer will be structured as follows:

(1) comments from referee, (2) author's response, (3) author's changes in manuscript.

Kind regards,

Leandra Praetzel & Co-Authors

General

(1) The manuscript needs careful line editing to take care of non-idiomatic English. An example is the frequent usage of wrong tenses (e.g. in line 60: "is mainly depending on" rather than "mainly depends on"). The authors may seek help from a native English speaker for this purpose. I have pointed out a few instances below, but these are by no means exhaustive. I must also concede that I am not a native English speaker!

(2) We asked a native speaker for help and feel certain that her corrections will substantially improve the grammatical style of the paper.

Specific

(1) Lines 14-15: Change "... to the atmosphere, following recent studies this is particularly the case for small and shallow lakes." to "... to the atmosphere; recent studies have shown that this is particularly the case for small and shallow lakes."

(2) The sentence will be rephrased in the revised manuscript as follows:

(3) "Inland waters, particularly small and shallow lakes, are significant sources of carbon dioxide (CO2) and methane (CH4) to the atmosphere."

(1) Line 16: Delete "yet" and "thus".

(2) The terms will be deleted in the revised manuscript.

(1) Lines 21-22: Change "... were significantly negative (p<0.05, rho<-0.6) correlated" to "... exhibited significant negative correlation (p<0.05, rho<-0.6)". Please make similar changes elsewhere.

(2) The sentence will be rephrased according to the reviewer's suggestion. Similar changes will be made elsewhere in the revised manuscript.

(1) Lines 32-34: The last sentence states the obvious. Who has suggested such a "replacement"?

(2) Some studies implicitly equal production and emission rates, e.g.

Grasset et al. 2018: doi: 10.1002/lno.10786

Sollberger et al. 2014: https://doi.org/10.1007/s00027-013-0319-2

We will adjust the statement in the revised manuscript as follows:

(3) "We highlight that studies on production rates and sediment quality need to be interpreted with care in terms of deducing emission rates and patterns as it this neglects physical sediment properties and production and oxidation processes in the water column."

(1) Line 52: Change "has been" to "have been" (here majority is plural), and remove "is".

(2) The term will be changed in the revised manuscript and "is" will be deleted.

(1) Line 56: Remove hyphen between "in" and "lake".

(2) The hyphen will be removed in the revised manuscript.

(1) Lines 58-59: Why is it crucial? Your results show that it is not.

(2) The sentences will be rephrased in the revised manuscript as follows:

(3) "Nevertheless, anoxic sediments are important for whole lake C cycling as the CO2 and CH4 produced there can be released through the water column to the atmosphere. To understand the spatial patterns of CO2 and CH4 emissions, it is therefore of interest to also understand CO2 and CH4 production processes in the sediment as well as their major controls."

(1) Line 64: Also its origin (e.g. lignin).

(2) This will be added in the revised manuscript as follows:

(3) "...and therefore its origin and degree of decomposition..."

(1) Line 74: Remove "being".

(2) The term will be removed in the revised manuscript.

(1) Line 82: As also pointed out by the other referee a more negative deltaG change would make R2 thermodynamically more favourable.

(2) The statement will be corrected in the revised manuscript.

(1) Lines 86-87: Change "are attributed to" to "may arise from".

(2) The phrasing will be changed in the revised manuscript.

(1) Line 90: Change "remain" to "remains".

(2) The term will be corrected in the revised manuscript.

(1) Lines 92-93 and elsewhere: As also pointed out by the other referee please define each abbreviation when you use it the first time and maintain consistency.

(2) All abbreviations will be defined when being used for the first time and only abbreviations will be used afterwards in the revised manuscript.

(1) Line 96: Why "to a small extent"? In such shallow systems wind-driven turbulence could disturb the sediments. Lines 97-98: Add "penetration" after "oxygen" and remove "in our case". What do you mean by "perennial circulation".

(2) The sentence will be rephrased in the revised manuscript as follows:

(3) "...but might in the upper parts of the sediments be influenced by oxygen penetration from the water column due to a well-mixed water body."

(1) Line 99: Please use present indefinite tense, not present continuous.

(2) The tense will be corrected in the revised manuscript.

(1) Line 103: "other" sediment properties?

(2) The term "other" will be added in the revised manuscript.

(1) Line 108: Change "is accountable" by "accounts".

(2) The term will be changed in the revised manuscript.

(1) Lines 110-114: Please rephrase this sentence.

(2) The sentence will be rephrased as follows:

(3) Until now, laboratory incubations of lake sediments were mostly conducted with samples from one or few sites within one lake with a focus on comparing different lakes with each other rather than covering a high in-lake variability of production rates. Further, these studies emphasize temperature effects on production rates (Duc et al., 2010; Gudasz et al., 2010; Gudasz et al., 2015; Fuchs et al., 2016). Unlike peat soils, where a broad range of controls on CO2 and CH4 production has been investigated, to our knowledge, controls such as organic matter (OM) quality, the occurrence of alternative electron acceptors (EAs), thermodynamic processes and sediment grain size have not, or only individually, been systematically surveyed in small lakes.

(1) Line 119: Did you actually investigate "connected productions patterns to OM"?

(2) This sentence might be ambiguous. We will rewrite the sentence as follows:

(3) "...in order to relate observed production patterns to measured OM and sediment characteristics, thermodynamics, and water-atmosphere fluxes."

(1) Line 121 and elsewhere: I am not sure if these experiments can be termed as "mesocosm". These were incubations of cores in the lab.

(2) To clarify the experimental procedure and the differences between the two laboratory experiments, we changed the descriptions throughout the whole manuscript: *Sediment incubations* to *slurry incubations* and *Sediment mesocosms* to *Intact sediment core incubations*.

(1) Line 125: Change "hypothesize" to "hypothesized".

(2) The term will be corrected in the revised manuscript.

(1) Line 137: Change "blast" to "blasted".

(2) The term will be corrected in the revised manuscript.

- (1) Line 138: Change "arose" to "formed".
- (2) The term will be corrected in the revised manuscript.

(1) Figure 1 captions: Technically the depth categories are wrong. For example by <150 cm, you imply depths between 125 and 150 cm, but 20 cm is also <150 cm. This should be clarified (e.g. 125 indicates 100cm<depth_125 cm).

(2) The description of lake depths categories will be clarified in the revised manuscript as follows:

(3) "...numbers 50, 100, 125 and 150 indicate lake depth category (50: <50 cm, 100: 50-100 cm, 125: 100-125 cm, 150: 125-150 cm)."

(1) Table 1, caption: Analytical procedures do not have to be mentioned here; they should be described in methodology section.

(2) The description of analytical procedures will be deleted. These can be found in the methods section 2.4 of the revised manuscript.

(1) Line 167: Change "at three occasions" to "on three occasions".

(2) The term will be changed in the revised manuscript.

(1) Line 169: Why randomly? It should be selectively based on a reason.

(2) We randomly chose the sites in order to avoid detecting differences between sites, water depths or transects due to the sampling date. E.g. if we took all four samples from one transect on the same sampling date, we could not have been sure that potentially observed differences in production rates were because of different site characteristics or because of the sampling date. The same would be true for water depths, so we decided to perform a random sampling at each date.

(1) Line 172: Add "respectively" at the end of the sentence.

(2) The term will be added in the revised manuscript.

- (1) Line 176: Change "added with" to "containing".
- (2) The term will be changed in the revised manuscript.

(1) Line 180: Change "stored" to "maintained".

(2) The term will be changed in the revised manuscript.

(1) Line 192: Referring to a comment by the other reviewer, I note that some isotope data are presented in Table 2 (not for sulphur though), although not at all discussed in the text. It is not clear whether the sample was decalcified. Also what was the reproducibility of measurements? In fact the precision of analysis is not given for any parameter.

(2) Isotopic values of C and N will be mentioned in the revised manuscript in the results sections 3.1.1 and 3.1.2.

Sulphur isotope data was not measured an will therefore be deleted from the methods section.

The samples were not decalcified before analyses. But we analyzed samples for carbonate content which confirmed that carbonate contents were very low (< 0.9 mg/g). We therefore assume that carbonates in the samples only have a minor influence on the results of isotopic data.

During every run of samples, multiple working standards were measured to assure reproducibility of measurements. Precision of standards are: < 1% for C, < 0.1 % for N, < 0.05 ‰ for delta13C, < 0.5 ‰ for delta15N. Information on precision will be added to the methods section 2.2.2 of the revised manuscript.

- (1) Line 201: Change "Therefore" to "For this purpose".
- (2) The term will be changed in the revised manuscript.
- (1) Line 253: Something missing in the sentence.
- (2) The term will be corrected in the revised manuscript.
- (1) Line 267: Change "analyzed for" to "measured".
- (2) The term will be corrected in the revised manuscript.

(1) Lines 274, 291: Change "measured" to "analyzed". (Note samples are analyzed, parameters are measured).

- (1) Line 301: Change "Therefore" to "For this purpose".
- (2) The term will be corrected in the revised manuscript.
- (1) Line 313: These are lab experiments, NOT mesocosms!
- (2) See comment on line 121.

(1) Line 331: Change "conducted" to "made".

(2) Will be changed in the revised manuscript. Please note that the whole sentence should be rephrased in order to the other reviewer's suggestion as follows:

(3) "For statistical analyses and discussion, only measurements that were made > 50 days after the deployment of the sediment mesocosms in the climate chamber were used. This was done in order to ensure the system had adapted to experimental conditions and had reached a steady state. Steady state conditions were indicated by quasi-constant CO2 and CH4 concentrations in the sediment."

(1) Line 361: There is no other way to quantify inputs is ebullition?

(2) The only way to directly quantify ebullition is via inverse funnels that could trap the emitted methane bubbles. We tested this method in our sediment cores, but without success so that we decided to only measure total methane fluxes and separate diffusive and ebullitive fluxes mathematically as this has been suggested by Bastviken et al. 2004 and adapted by many others when measuring in-situ methane fluxes with a floating chamber approach.

(1) Line 408: Change "nor" to "or".

(2) The term will be changed in the revised manuscript.

(1) Fig. 2: Figure difficult to digest. I could not follow "Different letters indicate significant differences between these sites." What do letters "a"-"d" mean? Am I missing anything?

(2) For more clarity, the figure will be split into two and production rates in 5-10 cm depth will be displayed in the supporting information.

a-d denote if there are significant differences between sites. Same letters mean no significant differences. The description will be rephrased in the revised manuscript as follows:

(3) "Identical lowercase letters indicate production rates that were not significantly different (i.e. p > 0.05) from each other."

(1) Line 483: Change "by averagely" to "on a average by".

(1) Lines 490, 493, 499: See earlier comments on Lines 21-22.

(2) The sentences will be rephrased according to the above-noted suggestion.

(1) Line 508 and elsewhere: I believe sediment ebbulition in inferred from k>o. I am not sure. Was there any bioturbation that could increase the emission?

(2) Ebullition was inferred from piston velocity k > 2 as described in the methods section 2.3 (see lines 359-361 of the submitted manuscript).

We did not observe any bioturbation during the experiment.

(1) Table 4: "n.s." presumably means not significant (p<0.05). Is is mentioned somewhere? Significance also depends on the number of values that are not given.

(2) n.s. means not significant. An explanation will be added to the table's description as follows:

(3) "n.s. means that correlations were not significant (p > 0.05)."

We will revise the table including number of values as follows:

	(CH ₄ flux	CO ₂ flux			
	rho	р	n	rho	р	n
Clay	0.648	< 0.05	12	0.605	< 0.05	12
Silt	0.497	<i>n.s.</i>	12	0.302	n.s.	12
Sand	-0.648	< 0.05	12	-0.605	< 0.05	12
Fats, waxes, lipids	-0.833	< 0.05	8	-0.333	n.s.	8
Phenols; humics	-0.833	< 0.05	8	-0.357	n.s.	8
Aromates	-0.595	<i>n.s.</i>	8	-0.524	n.s.	8
Lignin	-0.786	< 0.05	8	-0.381	n.s.	8
C/N	-0.881	< 0.01	8	-0.333	n.s.	8
C (%)	-0.714	<i>n.s.</i>	8	-0.190	n.s.	8
CH ₄ sediment stock change	-0.222	n.s.	41	0.05	n.s.	35
CO ₂ sediment stock change	-0.049	<i>n.s.</i>	41	-0.064	<i>n.s.</i>	35

(2) Numbers of values for other calculated correlations will be given in the Supporting information Tab. S1.

(1) Lines 528: Change "concentration" to "concentrations" and "was" to "were".

(2) The term will be changed in the revised manuscript.

(1) Line 536: Change "were significantly negative correlated" to "showed significant negative correlation"

(2) The term will be changed in the revised manuscript.

(1) Line 542: What do you mean by "narrower"? lower?

(2) Narrower will be changed to lower in the revised manuscript.

(1) Lines 545-556: Authors have emphasized on C/N ratio. They have observed increase in C/N with depth in the inner part of the lake. C/N ratio may not be a very efficient parameter to characterize organic source owing to rapid remobilization of nitrogen as well as reabsorption of ammonium on particulates. The paragraph 405 "The C content in the samples was between 2.15 and 33.16% with lowest values at site 3.50 and highest at site 1.50. C/N ratios ranged from 10.97 at site 1.150 to 19.06 at site 3.100. Neither C content nor C/N ratio showed significant changes with sediment nor lake depth, but C/N ratio was significantly higher in samples taken close to the shore (50) than in samples from the lake center (150) (p < 0.01)." is very confusing. A graph showing distribution of C/N ratio across the horizontal length of the lake would suitable to comprehend the results better.

(2) We will include a figure showing C/N ratios and absorption ratios for fats/polysaccharides to better illustrate our results.



(1) Line 551-552: I do not believe in shallow depths it matters.

(2) We additionally propose the mechanism of resuspension and focusing of small particles, that could alter the degree of decomposition of OM.

(3) "...As this process might not be of the same importance in shallow lakes compared to deeper lakes, we additionally suggest that the more decomposed OM in the lake center might have undergone degradation processes during resuspension and focusing of small particles as a result of wind-induced bed-shearing (Mackay et al., 2011)."

- (1) Line 559: Change "buried" to "getting buried"
- (2) The term will be changed in the revised manuscript.
- (1) Line 571: Change "role for" to "role in"
- (2) The term will be changed in the revised manuscript.
- (1) Line 578: Change "e.g." to "among other things"
- (2) The term will be changed in the revised manuscript.

(1) Line 587: Change "in the following" to "below"

(2) The term will be changed in the revised manuscript.

(1) Line 591: Remove "the" before "in other studies"

(2) The term will be removed in the revised manuscript.

(1) Lines 612-614: Laborious sentence. All you are saying is that such shallow depths do not get thermally stratified in summer.

(2) We will rewrite the sentence as follows:

(3) "...especially regarding the fact that shallow waters, as against deeper lakes, do not get thermally stratified in summer and therefore shallow sediment warm much faster (Jankowski et al., 2006)."

(1) Lines 619-620: Change the tense to present indefinite.

(2) The term will be changed in the revised manuscript.

(1) Line 627-630: What do you mean by "wider" and "narrower"? I do not follow this sentence.

(2) "Wider" and "narrower" will be changed to "higher" and "lower". We will restructure the whole discussions part about OM quality and CO2 and CH4 production rates according to the other reviewer's suggestions. To explain what we mean by this statement: C/N ratios can be interpreted in two ways: a) high C/N ratios = low decomposition state, low C/N ratios = high decomposition state; b) but C/N ratio can also be used to differentiate between OM of terrestrial and aquatic origin (see Meyers 1994) whereas high ratios = terrestrial and low ratios = aquatic origin. It is known that OM of a low decomposition state is easier degradable for microorganisms and therefore leads to higher production rates of CO2 and CH4, but on the other hand, aquatic OM is usually easier degradable for aquatic microorganisms and would therefore lead to higher production rates compared to OM of terrestrial origin (see e.g. Grasset et al. 2018). We therefore conclude that, although there exist two contradicting effects (low vs. high decomposition or aquatic vs. terrestrial origin), the fact that OM closer to the shore is in a lower decomposition state (although it is probably of terrestrial origin) fuels CO2 and CH4 production. The paragraph on C/N ratio and production rates will be revised as follows:

(3) "C/N ratios are frequently used to characterize the degradation state of OM, but we did not find correlations between C/N ratios and CO2 and CH4 production rates in the slurry incubations. Although OM of autochthonous origin was found to fuel higher degradation rates than allochthonous OM (West et al., 2012; Grasset et al., 2018) we found evidence of predominant inputs of allochthonous (terrestrial) material at sites with higher production rates close to the shore (higher C/N ratios), whereas sites with lower production rates in the lake center received mainly autochthonous (aquatic) OM as indicated by lower C/N ratios (Meyers, 1994). On the other hand, high C/N ratios also indicate a lower degradation state and therefore higher degradation potential whereas low C/N ratios are usually typical of highly decomposed OM having a lower CO2 and CH4 production potential (Malmer and Holm, 1984; Kuhry and Vitt, 1996). These two possibilities of interpreting C/N ratios might be the reason for apparently contradicting findings and the missing relationship between C/N ratios and CO2 and CH4 production rates."

(1) Line 637: Change the tense to present indefinite.

(2) The term will be changed in the revised manuscript.

(1) Line 654: But the acetate concentration increased!

(2) Please not the discussion in section 4.1.4 of this observation in lines 660-665 of the submitted manuscript.

- (1) Line 655: Remove "of" before "importance"
- (2) The term will be changed in the revised manuscript.
- (1) Line 671: OM quality is not quantified so instead of low you should perhaps use poor.
- (2) The term will be changed in the revised manuscript.

(1) Line 673: Change "of energy" to "in energy"

(1) Line 675: Change "... acetate, but rather is fermentation" to " ... acetate. Instead fermentation may be rate limiting"

(2) The term will be changed in the revised manuscript.

(1) Line 677: Bring "Further" before "it".

(2) The term will be changed in the revised manuscript.

(1) Line 678: Change "finding emphasizes" to "supports"

(2) The term will be changed in the revised manuscript.

(1) Line 692: If the relationship was insignificant the trend cannot be "clear".

(2) The term will be removed in the revised manuscript.

(1) Line 693: Not at all clear, and so is the following conclusion. I find this whole paragraph speculative.

(2) We will rewrite the whole paragraph 4.1.5 - also following the other reviewer's suggestions – where we will elaborate the relationships between CO2 and CH4 production and alternative EAs more precisely. Instead of discussing relationships between EAC and CH4 production, we emphasize that measured inorganic and organic EAs can explain 40-80% of measured CO2 production. The missing capacity can probably be explained by solid-phase iron, which we found ranging from 2 to 3 %, but whereof we do not have information on its speciation.

We further emphasize, that missing correlations between EAC and CH4 production are due to our experimental set-up: the one-week pre-incubation might have already depleted a large amount of reducible organic and inorganic EAs so that subsequent changes and therefore correlations were low.

(1) Line 702: Change "something" to "somewhat"

(1) Line 706: Change "approaches" to "factors"

(2) The term will be changed in the revised manuscript.

(1) Line 730: Authors attempt to correlate ebullition with grain size. They believe that higher sand content leads to lesser ebullition. Which is highly unlikely since ebullition depends on permeability of sediments and not porosity. Sand always has higher permeability than silt and clay although lesser porosity. You need to elaborate your concept with more clarity

(2) When explaining CH4 ebullition with the concept of grain size distribution/porosity, it is not primarily of importance how permeable the material is, but how effective bubbles can actually accumulate in the sediment (so that they can subsequently be released). Lui et al. 2018 found that the dominant pathway of bubble formation is by displacing the surrounding sediment, and that this is easier in soft, silty sediment compared to sandy sediments. This sediment displacement would lead to more macropores and therefore a higher connectivity creating conduits for bubble release.

We will change the paragraph explaining these mechanisms more precisely as follows:

(3) "We found ebullition supporting significantly to total CH₄ fluxes in two of our four intact sediment core incubations, whereas sites with higher shares of sand exhibited less ebullitive fluxes confirming the findings of Liu et al. (2016) and (2018). The authors explain their findings with the dominant pathway of bubble formation in the sediment, which is by displacing surrounding sediment particles. As this mechanism is more efficient in soft silty sediments compared to sandy material, CH₄ bubbles likely accumulate more easily in silt, creating a network of macropores and therefore conduits for subsequent bubble release. We further found OM quality partly exhibiting significant negative correlations with CH₄ fluxes, but to a lesser extent than with CH₄ production. When preparing slurry incubations, the physical sediment structure is destroyed, so that OM quality becomes the major controlling factor for gas production. These findings suggest that grain size distribution is besides OM quality a main driver of spatial CH₄ flux patterns in intact sediment core incubations and that only a combination of physical characteristics and sediment OM quality could sufficiently explain CH₄ emission patterns from lakes."

(1) Line 747: Change "experiment" to "results"

- (1) Line 749: Remove "especially"
- (2) The term will be changed in the revised manuscript.
- (1) Line 753: Change "vulnerable" to "sensitive"
- (2) The term will be changed in the revised manuscript.
- (1) Line 754: Change "unroll" to "expect"
- (2) The term will be changed in the revised manuscript.
- (1) Line 755: Change "lower water columns" to "shallow depths"
- (2) The term will be changed in the revised manuscript.
- (1) Line 761: Change "refer" to "attribute"
- (2) The term will be changed in the revised manuscript.

(1) Lines 764-765: Then why do you not find strong relationship between methane production and (EACorg)?

(2) Please see also comment on Line 693. The statement will be discussed in more detail in the revised manuscript.

(1) Line 770: Measuring "production rate" does not neglect water column processes, interpretation of these data alone would.

(2) The sentence will be rephrased in the revised manuscript as follows:

(3) "Further, measuring production rates only would neglect the importance of the water column as a sink of sediment generated CH4..."

List of relevant changes made in the manuscript

Overall

- 1. Editing of English language and grammar
- 2. "...were significantly correlated to..." was changed to "...exhibited significant negative correlation..."
- 3. rephrasing of "incubation" and "mesocosms" to "slurry incubations" and intact sediment core incubations"
- 4. correction of non-significant decimal places
- 5. revision of use of abbreviations

Introduction

- L. 105 ff. Correction of standard formation energies
- L. 144 ff. Revision of the whole paragraph describing knowledge gaps

Materials and Methods

Fig. 1 caption: Renaming of lake depth categories

L. 466 ff.: Adding of description of methods for determining lake water parameters

Results

L. 501: Adding of Fig. 2, which displays C/N ratio and fats/polysaccharide ratio at different lake depth categories

Fig. 3: Deletion of production rates from 5-10 cm depth for better clarity

L. 567 ff.: Adding of description of correlations of acetate and hydrogen concentrations with OM quality

L. 610 ff.: Adding of calculation of potential CO2-production from prevalent electron acceptors + Figure

Discussion

L. 669 ff.: The chapters "Spatial variability of OM", "Spatial variability and temperature dependency of CO2 and CH4 production", and "Influence of OM quality on CO2 and CH4 production rates" have been merged and therefore shortened

L. 871 ff.: The chapter "Methanogenic pathways" has been shortened and refocussed on correlations between methanogenic pathways and OM quality

L. 919 ff.: The chapter "Alternative Eas" has been refocussed; the speculative part on CH4 production has been removed and the discussion on CO2 production has been added L. 1009 ff.: The discussion on correlation between CH4 bubble formation and grain size distribution has been described in more detail

Organic matter and sediment properties determine in-lake variability of sediment CO₂ and CH₄ production and emissions of a small and shallow lake

Leandra Stephanie Emilia Praetzel¹, Nora Plenter^{2,3}, Sabrina Schilling¹, Marcel Schmiedeskamp¹, Gabriele Broll², Klaus-Holger Knorr¹

⁵ ¹University of Münster, Institute of Landscape Ecology, Biogeochemistry and Ecohydrology Research Group

²University of Osnabrück, Institute of Geography, Agroecology and Soil Research Group

³University of Applied Sciences Osnabrück, Faculty of Agricultural Sciences and Landscape Architecture

10 *Correspondence to:* Leandra S. E. Praetzel (leandra.praetzel@uni-muenster.de) or Klaus-Holger Knorr (kh.knorr@uni-muenster.de)

Abstract

- Inland waters, particularly small and shallow lakes, are significant sources of <u>carbon dioxide (CO₂)</u> and methane (CH₄) to the atmosphere. , following recent studies this is particularly the case for small and
- shallow lakes. The <u>However, the</u> spatial in-lake heterogeneity of CO_2 and CH_4 production processes and their drivers in the sediment <u>yet</u>-remain poorly studied. We thus measured potential CO_2 and CH_4 production in <u>sediment slurry</u> incubations from 12 sites within the small and shallow crater lake Windsborn in Germany as well as fluxes at the water-atmosphere interface <u>of intact sediment core</u>
- ²⁰ <u>incubations at from</u> four sites. Production rates were highly variable and ranged from 7.2 and to 38.5 μ mol CO₂ gC⁻¹ d⁻¹ and from 5.4 to 33.5 μ mol CH₄ gC⁻¹ d⁻¹. Fluxes <u>lay-ranged between-from</u> 4.5 and to 26.9 mmol CO₂ m⁻² d⁻¹ and <u>between from</u> 0 and to 9.8 mmol CH₄ m⁻² d⁻¹. Both CO₂ and CH₄ production rates and <u>the CH₄ fluxes exhibited a significant and negative correlation (p < 0.05, rho < -0.6) were significantly negative (p<0.05, rho<-0.6) correlated with the prevalence of recalcitrant organic matter</u>
- 25 (OM) compounds in the sediment as identified by Fourier-transformed infrared FTIR-spectroscopy. The C/Ncarbon/nitrogen ratio exhibited a significant negative correlation (p<0.01, rho=-0.88)was significantly (p<0.01, rho=-0.88) correlated with CH₄ fluxes, but neither not with production rates nor CO₂ fluxes. Availability of iInorganic (nitrate, sulfate, ferric iron) and organic (humic acids) electron acceptors failed together could explain differences in CH₄ production rates, assuming a competitive
- 30 suppression, ($R^2=0.22$) whereas but observed non-methanogenic CO₂ production could be explained by up to 91% by prevalent electron acceptors. we-We did not find clear relationships between organic matterOM quality, thermodynamics of methanogenic pathways (acetoclastic vs. hydrogenotrophic) and electron accepting capacity of the organic matterOM. Differences in CH₄ fluxes were also ableinterestingly to a large part be explained by Grain-grain size distribution could sufficiently (p < 0.05,
- 35 rho_=_±0.65) explain differences in CH₄ fluxes. Surprisingly though, sediment gas storage, potential production rates and water-atmosphere fluxes were decoupled from each other and did not show any correlations. Our results show that within a small lake, there exists a significant spatial variability of sediment gas-CO₂ and CH₄ production show significant spatial variability, even within a small lakes which is mainly driven by spatial differences in the degradability of the sediment organic matterOM can
- 40 be explained by the origin and pre-processing, and therefore the degradability of the organic matter. We highlight that studies on production rates and sediment quality need to be interpreted with care, though, in terms of deducing emission rates and patterns, measuring production rates is not a suitable way to replace in situ flux measurements as it that approaches based on production rates only neglects physical sediment properties and production and oxidation processes in the water column as major
- 45 <u>controls on actual emissions</u>.

1 Introduction

Inland waters play an important role in the global carbon (C) cycle and contribute significantly to the natural emissions of the greenhouse gases carbon dioxide (CO₂) and methane (CH₄) (Cole et al., 2007;
Battin et al., 2009; Bastviken et al., 2011; Raymond et al., 2013; Regnier et al., 2013). Lakes and reservoirs are estimated to emit in total 0.32 – 0.39 Pg C of CO₂ and 0.58 Pg C (CO₂ -eq.) of CH₄ year⁻¹ (Cole et al., 2007; Bastviken et al., 2011; Raymond et al., 2013)...), but Especially especially small lakes (< 0.1 km²) have previously been underestimated in the past in regard toregarding their spatial expansion and, therefore, their contribution to global greenhouse gas emissions (Downing, 2010).
Although small lakes account for approx. ~1/3 of the total lake area and cover less than 1% of the global land surface, they contribute 35% of the CO₂ and 72% of the CH₄ emissions from lakes worldwide (Downing et al., 2006; Holgerson and Raymond, 2016). Further, Even even if the highest number of

lakes can bethough most lakes are found in boreal zones, the largest areas occur in lower latitudes around 50° (Verpoorter et al., 2014). In small lakes, Due due to their shallowness, shorter water residence times and a-smaller perimeter perimeter to to-volume ratios, metabolic processes and earbon (C) turnover in small lakes is are much faster compared tothan in larger lakes, making small lakes potentially and they can therefore be expected to be more vulnerable susceptible to environmental and climatic changes than the latter (Wetzel, 1992; Downing, 2010).

Recently, many studies have shown that both CO₂ and CH₄ fluxes are highly variable both on a spatial and temporal scales, but the majority of these measurements has have been taken outperformed on larger lakes and is-concentrated in boreal regions (Schilder et al., 2013; Wik et al., 2013; Bastviken et al., 2015; Natchimuthu et al., 2016; Natchimuthu et al., 2017; Spafford and Risk, 2018). Beyond that, few studies have examined on greenhouse gas (GHG) production processes in the sediments, or have attempted to link and studies linking sediment gas GHG production to emissions both remain scarce. Nevertheless, anoxic sediments play a crucial role in lake C cycling as they are the main producer of CO₂ and CH₄ which is subsequently released through the water column to the atmosphere. Nevertheless, anoxic sediments are important for whole-lake C cycling, as the CO₂ and CH₄ produced there can be released through the water column to the atmosphere. To understand the spatial patterns of CO₂ and CH₄ emissions, it is therefore erucial of interest to also understand CO₂ and CH₄ production processes in the sediment as well as their major controls.

The degradability of OM, and therefore the amount of produced CO₂ and CH₄, produced in lake sediments relates to the degradability of the organic matter (OM) present, which is mainly depending depends primarily on its componentsquality, the microbial biomass and enzyme activities (Updegraff et al., 1995; McLatchey and Reddy, 1998; Fenchel et al., 2012). While C/N ratios can be used to determine the origin and the degradation state of OM can be determined using carbon/nitrogen (C/N) ratios,

80

Fourier-transformed infrared (FTIR) spectroscopy can provide qualitative information about OM components can be determined via Fourier-transformed infrared (FTIR) spectroscopy. The latter

<u>technique</u>, <u>which</u> can, therefore, <u>also</u>further provide information aboutand therefore its <u>origin and grade</u> <u>degree</u> of decomposition (Meyers, 1994; Broder et al., 2012; Biester et al., 2014; Li et al., 2016). The

- 85 prevalence of different OM compounds leads to specific features in FTIR spectra, as functional moieties have wavelength-specific absorption maxima (Niemeyer et al., 1992; Cocozza et al., 2003). Artz et al. (2008) compiled a range of functional moieties that is used to characterize OM in peat soils, but is they are also applicable to OM in general. While polysaccharides and proteins are preferentially degraded by microorganisms, cellulose natural organic matteraliphatic (e.g. waxes) or and aromatic compounds (e.g.
- 90 lignin) are <u>due to their molecular structure</u> more recalcitrant <u>(due to their molecular structure)</u> and, therefore, <u>residually</u> accumulate in the anoxic sediment (Fenchel et al., 2012; Tfaily et al., 2014). The prevalence of different OM compounds leads to specific FTIR spectra, as functional moieties have wavelength-specific absorption maxima (Niemeyer et al., 1992; Cocozza et al., 2003).
- In anoxic sediments, CO_2 and CH_4 are produced during the breakdown of OM over a cascade of 95 microbially induced mediated processes. After the fermentation of cleavage of complex organic polymers, resulting monomers are fermented, which mainly produces hydrogen (H₂) and OM-low molecular weight organic compounds of low molecular weight (e.g. acetate), the latter low molecular weight products compounds are being further oxidized to CO_2 , and H_2 to H_2O , together with a setupon consumption of electron acceptors (EAs: nitrate, sulfate, ferric iron or humic substances). 100 before ThenOnly subsequently, CH₄ production initiates asisas the final step of terminal electron <u>electron-accepting processes</u>, <u>which happensinitiates</u> after <u>depletion of</u> all other, thermodynamically more favorable electron acceptorEAs are depleted (Achtnich et al., 1995; Blodau, 2011). CH₄-in turn is mainly produced via two different pathways, the acetoclastic pathway and the hydrogenotrophic pathway-with, where for which either acetate is the substrate (acetoclastic, R1) or H₂ as-is the substrate 105 and CO₂ as is the electron acceptorEA (hydrogenotrophic, R2) (Conrad, 1999; Whiticar, 1999). Values for ΔG_r^0 for these terminal electron accepting processes and methanogenesis can be are calculated from standard formation energies ΔG_{f}^{0} in aqueous state listed in Stumm and Morgan (1995) and Nordstrom

and Munoz (1994).

$CH_3COOH \rightarrow CO_2 + CH_4$	$\Delta G_{\rm r}^{0} (25^{\circ}{\rm C}) = -50.99 \text{ kJ mol}^{-1}$	R1
------------------------------------	-------------------------------------------------------------------------	----

110	$CO_2 + 4 H_2 \rightarrow 2 H_2O + CH_4$	$\Delta G_{\rm r}^{0} (25^{\circ}{\rm C}) = -193.03 \text{ kJ mol}^{-1}$	R2
-----	------------------------------------------	--------------------------------------------------------------------------	----

Several studies have shown that in anoxic wetland and marine sediments rich in labile organic compounds, the acetoclastic pathway <u>dominates</u>, <u>which is energetically more less</u> favorable for <u>microorganisms</u>, <u>dominates</u>, <u>whereas Thus</u>, <u>wW</u>ith increasing recalcitrance, the hydrogenotrophic pathway becomes more important as acetate as direct precursor of CH₄ gets is depleted (Schoell, 1988; Hornibrook et al., 1997; Miyajima et al., 1997). As suggested by Lojen et al. (1999), the OM quality in lake sediments might thus be responsible for suggested that seasonal changes in the dominant methane CH₄ pathway in a lake sediment are attributed to OM quality.

Whereas <u>many studies have shown that the potential of inorganic electron acceptorEAs to can suppress</u> methanogenic activity is <u>well studied (e.g.</u> Yao et al., 1999; Fenchel et al., 2012), information on the
role of humic substances as organic <u>electron acceptorEAs</u> remains scarce. <u>In one study</u>, Klüpfel et al. (2014) revealed the potential of<u>that</u> humic substances to <u>can</u> be reduced and re-oxidized at oxic-anoxic interfaces in peatlands, sediments or soils underlying water table fluctuations and <u>another study showed</u> it has recently been shown that in peat soils poor in inorganic <u>EAs</u>, the electron accepting capacity (EAC) of availability of EAC in OM controls represents the major control on CO₂ and CH₄ production in peat soils poor in inorganic <u>electron acceptors (EA)s</u> (Gao et al., 2019). As sediments from the lake under studywe investigated here-are also rich in OM, we wanted to verify if whether the electron accepting (EAC) and electron-donating capacities (EDC) of humic substances also play a <u>vital</u> role in explaining the spatial variabilities of CO₂ and CH₄ production in lake sediments. <u>Although that-they</u> are not subjected to water table fluctuations, <u>but-they</u> might, to a small extent in the upper parts of the

130 sediments, be influenced by oxygen <u>penetration</u> from the water column due to the <u>a</u> in our case prevalent perennial circulationwell-mixed water body particularly in shallow lakes (Lau et al., 2016).

135

<u>Overall, The-the</u> amount of CO₂ and CH₄ produced <u>is</u>-therefore either <u>depending-depends</u> on the availability and degradability of the OM itself, the presence of EAs, and the <u>concentration-levels</u> of H₂ and acetate as substrates for methanogenesis (Segers, 1998; Conrad, 1999; Megonigal et al., 2003; Blodau, 2011; Fenchel et al., 2012).

Within lakes, ^Tthe spatial distribution of OM and <u>other</u> sediment properties within lakes has been found to be highly variablevary considerably in terms of their origin (terrestrial vs. aquatic), degradability, elemental geochemistry and grain size (Muri and Wakeham, 2006; Ostrovsky and Tęgowski, 2010; Tolu et al., 2017). It has also been shown thatFurther, sediment grain size is an important factor for the evolution of CH₄ bubbles in sediments (Ostrovsky and Tęgowski, 2010; Liu et al., 2018)...); for example, Liu et al. (2016) for example revealed a decrease inthat CH₄ ebullition <u>decreases</u> with increased increasing shares of sand in lake sediments. Overall though, CH₄ ebullition in turn is accountable is thought to account for a large proportion (75%) of CH₄ emissions from lakes (Bastviken et al., 2011).

Until now, laboratory incubations of lake sediments were mostly conducted with samples from one or
 few sites within one lake with a focus on comparing different lakes with each other rather than covering
 a high in-lake variability of production rates. Further, these studies emphasize temperature effects on
 production rates (Duc et al., 2010; Gudasz et al., 2010; Gudasz et al., 2015; Fuchs et al., 2016). Unlike
 in peat soils, where a broad range of process based controls on CO₂ and CH₄ production has been studie,
 in small lakes, controls such as OM quality, the occurrence of alternative EAs, thermodynamic processes

150 and sediment grain size have not, or have only individually, been systematically surveyed. So far, experimental incubations of lake sediments were only conducted with samples from one or few sites within one lake, rather comparing different lakes with each other than to study within lake variations of CO₂ and CH₄ production and with a focus on temperature effects on production rates (Duc et al., 2010;

Gudasz et al., 2010; Gudasz et al., 2015; Fuchs et al., 2016). Although a broad range of controls on CO₂
 and CH₄ production has been widely investigated for anoxic peatland soils, studies on lake sediment are rare. To our knowledge, controls such as organic matter (OM) quality, the occurrence of alternative electron acceptors (EAs), thermodynamic processes and sediment grain size have not, or only individually, been systematically surveyed.

To close this knowledge gap, we determined the magnitude and spatial variability of sediment CO₂ and
 CH₄ production in a small and shallow temperate lake, in order to relate observed production patterns to and connected productions patterns to measured OM and sediment characteristics, thermodynamics of methanogenesis, and water-atmosphere fluxes. To this end, we conducted slurry and intact sediment mesocosm core incubations with sediment from the crater lake Windsborn in Germany. This site was

chosen as a model system having because it has a high sediment OM content (~30%), a very small
 catchment area and no surficial in- or outflows, meaning minimal in order to keep influences from the surrounding areacatchment as small as possible.

We hypothesized that (I) that there exists a noticeable spatial variability of CO_2 and CH_4 production vary spatially in the sediment, (II) that the variability of in the production rates is reflected in the flux patterns, and that (III) that the variation in production rates, methanogenic pathways and flux patterns can be explained by factors of OM degradability, the occurrence of availability of organic and inorganic EAs, and grain size distribution, or a combination of these factors.

170

2 Materials and Methods

2.1 Study site

175 The studied Lake Windsborn is a polymictic, small-and, shallow crater lake in the Volcanic Eifel, Rhineland Palatinate, south-west Germany and is part of the "Mosenberg" volcano group (see Fig. 1) (LfU, 2013). The climate is temperate with a mean annual temperature of 8.3°C and 931 mm precipitation (multi-annual mean 1981-2010; DWD, 2019). Lake Windsborn is the only genuine crater lake north of the Alpes. It emerged approx. 29,000 years ago after a volcanic eruption when the top of 180 the volcano was-had been blasted away and the newly formed crater was subsequently filled with water (LfU, 2013). The present lake arose formed around 1850, after drainage of the lake and the partial removal of the peat from the lake bottom (Kappes and Sinsch, 2005). The lake is part of a conservation area that was established in 1927 (Kappes and Sinsch, 2005). From 1950 until the 1990s, the lake was used a fishing ground, and was, therefore, stocked up with fish and limed (LfU, 2013). The lake is nearly 185 circular and surrounded by a 20- to 30-30-meter-meter-high rampart which consists of an alternation of red-brown ashes, slag and lapilli from the eruption. Therefore, it has a very small catchment of only about 8 ha compared to the lake surface of 1.41 ha without any, and it has no surficial in- and outflows and is only fed by groundwater and precipitation (LfU, 2013; Meyer, 2013). The maximum lake depth varies between 1.3 and 1.7 meters (Kappes and Sinsch, 2005). The area is underlain by devonic 190 Devonian quartzite (Kampf, 2005).





incubations, circles: sites for slurry incubations only, asterisk: site for intact sediment mesocosm core incubation 195 only (reference for 1.150, 2.150 and 3.150). Depths were interpolated by bivariate linear interpolation. Numbers 1, 2 and 3 refer to the transect number of transect from the lake shore to center, numbers 50, 100, 125 and 150 indicate lake depth category (50: <50 cm, 100: <50-100 cm, 125: <100-125 cm, 150 <: 125-150 cm).

The lake's shoreline is vegetated with *Carex rostrata*, *Comarum palustre* and *Menynathes trifoliata*, all indicating poor nitrogen (N) supply (Ellenberg et al., 2001). At the north-western riparian zone, there is 200 emerges a quaking bog of mainly dominated by Sphagnum spec. whose expansion will slowly lead to the silting up of the lake. Lake Windsborn was previously considered as humic-oligotrophic, but in the early 1990's it has transitioned to an eutrophic lake, and in the early 1990's now it is slowly recovering from human impacts and nutrient input (Kappes et al., 2000). During our measurement campaigns in 2017 and 2018, the lake exhibited partly meso- and eutrophic features as shown in Table 1.

²⁰⁵ Table 1: Selected lake water characteristics measured in Lake Windsborn in 2017 and 2018-: pH, conductivity (cond.), dissolved oxygen (O_2), chlorophyll alpha (Chl. α), dissolved organic carbon (DOC), total dissolved nitrogen (TN), chloride (Cl), calcium (Ca), iron (Fe), potassium (K), magnesium (Mg), sodium (Na), phosphorous (P) and sulfur (S).pH, conductivity and O2 concentration were determined in situ, DOC and TN were determined by catalytic oxidation, chloride was determined by ion chromatography, and other elements by inductively coupled 210 plasma optical emission spectroscopy (ICP-OES).

parameter	рН	cond.	O ₂	Chl <u>.orophyll</u> α	DOC	TN	Cl
unit		µS cm ⁻¹	mg L ⁻¹	μg L-1		mg L ⁻¹	
n	398	387	397	163	419	419	361
average	6.8	19. 35<u>4</u>	9. <mark>_6</mark> 7	27. 6 7	13.74	0.96<u>1.0</u>	2. 86 9
± SD	0.84	1.7 <mark>2</mark>	0.9 <mark>1</mark>	18. 66<u>7</u>	2.3 0	1. <mark>8</mark> 9	3.44
parameter	Ca	Fe	K	Mg	Na	Р	S
unit				mg L ⁻¹			
n	379	378	379	379	379	329	379
average	1. <u>2</u> 17	0.14	0. <u>8</u> 77	0.7 2	4.0 1	0. <u>1</u> 06	0.41
± SD	0. 27<u>3</u>	0. <u>1</u> 06	0.4 2	0.1 3	5.0 1	0. <u>1</u> 06	0.1 <mark>3</mark>

2.2 Sediment Slurry incubations

2.2.1 Sampling and preparation of slurry incubations

215

220

Samples fFor the slurry incubation experiment, samples were taken at on three occasions (in March, April and May 2018) from in total 12 of the in total 13 locations sampling sites within the lake from three transects covering multiple water depths (<50, <100, <125, and <150 cm) (see. Fig. 1). On each sampling date, four of the 12 sampling sites were chosen randomly as it was not possible to set up the experiment with all samples at once. At the sampling dates, Measured-measured air, water and sediment temperatures at the sampling dates were 7.7°C, 5.6°C and 5.2-°C (March), 13.8°C, 15.1°C and 10.1-°C (April) and 23.9°C, 23.6°C and 13.8 °C (May), respectively. Sediment samples were taken in duplicates with a gravity corer (UWITEC, Mondsee, Austria) from a boat with a gravity corer (UWITEC, Mondsee, Austria)-in 60 cm long PVC tubes and transported in an insulated box at ~5°C. The next day, Sediment sediment_cores were cut with a core cutter in the laboratory-the next day in segments of 5 cm thickness (0-5 and 5-10 cm sediment depth). Duplicate samples were homogenized, and then 20 g of each sediment was filled into 120 mL crimp vials added withcontaining 20 mL lake-water, after which the vials were and closed with a butyl-_rubber stopper and aluminum crimp cap. Samples were flushed with nitrogen N2 for 30 minutes in order to remove any remaining oxygen from the water and headspace, and then again flushed with nitrogen-N2 prior to the actual incubation. Slurry Incubations-incubations were set up in triplicates and stored-maintained at 25°C (corresponding to maximum measured in-situ sediment temperatures in summer 2018) in the dark. At-During each run, one set of parallel samples was incubated at 10°C in order to determine a Q10-value for CO2 and CH4 production rates.

The remaining sample material was freeze-dried (Alpha 1-4 <u>LPplusLDplus</u>, Christ, Osterode, Germany), ground with a ball mill (Mixer Mill MM 400, Retsch, Haan, Germany) and used for solid phase analyses, as outlined below.

2.2.2 Sediment organic matterOM quality

235

240

Freeze-dried and ground sediment samples were analyzed for total carbon (C), and nitrogen (N) and sulfur (S) concentrations and <u>C-and N-</u>stable isotopes using isotope-ratio mass spectrometry (IRMS; Eurovector EA3000 coupled with Nu Instruments Nu Horizon, Hekatech, Wegberg, Germany) and for organic matter (OM) components using FTIR-<u>Spectroscopy</u> <u>spectroscopy</u> (Cary 670 FTIR Spectrometer, Agilent, Santa Clara, USA).

For IRMS, 5 mg of sample were was weighed out into a tin cup together with 4 mg of Vanadiumvanadium-Pentoxide-pentoxide (V2O5). The combustion and reduction furnace were set to 1000°C and 650 °C, respectively, and the resultant gaseous compounds were quantified by IRMS.
Results are provided in % by mass for C, and N and S contents (precision < 1% for C and < 0.1% for N) and in ‰ vs. VPDB/AIR/VCDT for C, and N and S isotopic signatures (precision < 0.05‰ for ¹³C and < 0.5‰ for ¹⁵N). For isotope analyses, appropriate certified reference materials were used: IAEA 600 (δ¹³C = -27.774 ‰; δ¹⁵N = 1.0 ‰) and S - 1 (δ³⁴S = -0.30 ‰) for calibration, and BBOT (2.5-Bis-(5-tert.-butyl-2-benzo-oxazol-2-yl)thiophen; Hekatech, Wegberg, Germany), birch leaf, wheat flour and sorghum flour standards (IVA Analysetechnik e. K., Meerbusch, Germany) as working standards covering a range of -27.5‰ to -13.68-‰ for ¹³C, and -0.6‰ to 2.12-‰ for ¹⁵N, and -9.3 to -1.42 ‰ for ³⁴S.

Functional groups of OM compounds were identified by FTIR spectroscopy. ThereforeFor this purpose,
2 mg of freeze-dried sample were-was ground together in a mortar with 200 mg KBr (potassium
bromide) in a mortar, pressed into 13 mm pellets, and analyzed. Each sample was scanned from 599 to
4000 cm⁻¹ with a resolution of 2 cm⁻¹ and baseline corrected. Distinct peaks at specific wavelengths were
assigned to functional groups according to Artz et al. (2008) and normalized to the peak intensity at

1031 - 1035 cm⁻¹ (indicative of polysaccharides) in order to obtain inter-comparable peak-_ratios of functional moieties in all samples, as FTIR spectra only provide information about <u>the</u> relative abundance of certain functional moieties in one sample.

2.2.3 CO₂ and CH₄ production rates from <u>slurry</u> incubations

260

280

Potential CO₂ and CH₄ productions rates were determined by measuring the increase in concentration of CO_2 and CH_4 in the incubation vials over time. Concentrations were obtained from analysis of analyzing the headspace at the beginning of the experiment (t0), and after 1, 3, 8, 11, 14 and 18 days 265 (t1-t6). Samples were taken from the vial with a 10 mL PP-polypropylene syringe equipped with a three3-way-_stopcock and a_0.6 mm needle. Before each sampling, the pressure inside the vial was determined with a pressure sensor (GMH 3110, Greisinger, Regenstauf, Germany), and the syringe was three-times-flushed three-times with nitrogen N_2 , then Then 2 mL of N_2 nitrogen were was left in the syringe before stabbing the needle was stabbed through the vial's stopper, whereby the N2nitrogen was 270 added to the headspace, mixed, and, subsequently 2 mL of the sample were was taken from the vial so that the volume inside the vial remained constant. The gas samples were analyzed for CO₂ and CH₄ concentrations with a gas chromatograph (8610 GC-TCD/FID, SRI Instruments, Torrance, USA) equipped with a fFlame illonisation Ddetector (FID) and Mmethanizer to simultaneously measure CO₂ and CH₄. Before every sampling day, Tthe gas chromatograph was calibrated with standard gas mixtures 275 of known concentrations (CO₂: 385, 5,000 and 50,000 ppmV; CH₄: 5, 1,000 and 50,000 ppmV) before every sampling day.

First, measured concentrations in ppmV were pressure corrected and converted using the ideal gas law:

(1)

$$n = (p * V) / (R * T)$$

where n is the amount of substance in mol, p is the gas partial pressure in atm, V is the headspace volume in L, R is the ideal gas constant (0.082 L atm mol⁻¹ K⁻¹) and T is the laboratory temperature in K.

Total CO₂ and CH₄ concentrations in <u>the</u> gas and water phase in the incubation vials were calculated from headspace concentrations <u>with-using</u> Henry's Llaw:

$$c = K_h * p \tag{2}$$

where c is the concentration in the water phase in mol L⁻¹, K_h is the temperature-dependent Henryconstant (CO₂, 25°C = 0.0339 mol L⁻¹ atm $^{-1}$; CH₄, 25°C = 0.00129 mol L⁻¹ atm $^{-1}$ (Sander, 2015)) and p is the gas partial pressure in atm.

Moreover, CO_2 concentrations were pH-corrected in order to obtain pH-independent values for total CO_2 concentrations using <u>the</u> Henderson-Hasselbalch-equation and equilibrium constants according to Stumm and Morgan (<u>19961995</u>):

290
$$n_{\Sigma CO2} = n_{water} * 10^{\text{pH-6.4}} + (n_{water} * 10^{\text{pH-6.4}}) * 10^{\text{pH-10.25}}$$
 (3)

where $n_{\Sigma CO2}$ is the pH-corrected CO₂ amount in mol, and n_{water} is the calculated amount of CO₂ in the water phase in mol.

Finally, production rates were calculated as by linear regression ($R^2 > 0.8$ for CO_2 and > 0.9 for CH_4) from concentration change in gas and solute phase over time.

To evaluate the effect of temperature on CO_2 and CH_4 production, we calculated Q_{10} -values, describing the relative increase of production rates with an increase in temperature of 10 Kelvin (Fenchel et al., 2012).

$$Q_{10} = (R2/R1)^{[10/(T2-T1)]}$$
(4)

where R2 is the production rate at T2 (25°C), and R1 is the production rate at T1 (10°C).

300 2.2.4 Thermodynamics and methanogenic pathways

In order to calculate the thermodynamic energy yield for hydrogenotrophic and acetoclastic methanogenesis we measured hydrogen (H₂) concentrations at the beginning-(t0), after 8 days (t3) and at the end-(t6) of the experiment, and we measured acetate (H₃COO⁻)-concentrations at the beginning and at the end of the incubation. The thermodynamic energy yield, expressed as Gibb's free energy, was calculated using the Nernst equation and total dissolved and gaseous concentrations of educts and products in the incubation vials as described in Beer and Blodau (2007):

 $\Delta G_{\rm r} = \Delta G_{\rm r}^{0} + R * T \ln \left(\Pi_{\rm i} (\rm products)^{\rm vi} / \Pi_{\rm i} (\rm educts)^{\rm vi} \right)$ (5)

By calculating the Gibb's free energy (Δ G_r), it is possible to evaluate whether these processes are feasible under given conditions. In order for each reaction to occur, Therefore a theoretical threshold of Δ G_r = -20 to -25 kJ mol⁻¹ for hasd to be exceeded (Schink, 1997; Conrad, 1999; Blodau, 2011).

For H₂ concentration measurements, 2 mL of sample were taken from the incubation headspace with a syringe and needle and replaced with the same amount of N2. Samples were analyzed with a <u>r</u>Reduction <u>gGas dDetector</u> (RGD) <u>Hhydrogen and <u>c</u>Carbon <u>m</u>Monoxide <u>Aa</u>nalyzer (ta3000R Gas Analyzer, Ametek, Pittsburgh, USA) that was calibrated with gas standards of 5, 25, and 50 ppmV H₂. Measured H₂ concentrations were corrected for pressure and converted into dissolved concentrations using Henry's <u>Llaw</u> (K_h(H₂, 25°C) = 0.00078 mol L⁻¹ atm⁻¹ (Sander, 2015)) analogous to CO₂ and CH₄.</u>

315

320

305

310

Acetate concentrations were determined by ion chromatography IC-with chemical suppression (883 Basic IC plus, Metrohm, Filderstadt, Germany; A-supp 5 column, Metrohm, Filderstadt, Germany). Aqueous samples were filtered with 0.45 μ m Nnylon + Gglass Mmicro-Ffibre syringe filters (Simplepure, BGB Analytik, Rheinfelden, Germany) and kept frozen at -21°C until analysis.

2.2.5 Alternative Electron AcceptorEAs

To quantify alternative Electron Acceptors (EAs) that could support anaerobic respiration and potentially suppress methanogenesis in anoxic incubations, we analyzed formeasured nitrate (NO_3^-),
sulfate $(SO_4^{2^-})$, <u>ferrousic</u> iron (III)-(Fe³⁺), and <u>the electron acceptingEAC</u> and <u>donating capacityEDC</u> of the <u>organic matterOM</u> (EAC_{OM}, and EDC_{OM}) at the beginning (t0) and the end (t6) of the <u>slurry</u> incubation.

As the analysis of EAC_{OM} and EDC_{OM} is so far_only possible on finely ground materials, thus providing potential capacities rather than true in-situ capacities, we set up a second set of slurry incubations for a sediment depth of 0-5 cm with samples from 10 sites (all except 3.50 and 3.100) was set up withusing
0.4 g of the freeze-dried sediment material and 100 mL of Millipore-Milli-Q water. Incubations-Slurry incubations were set up in six replicates, flushed with N₂, pre-incubated and stored analogous to the first set of slurry incubations. Samples were measured-analyzed at the beginning and at the end of the experiment, whereas since at every sampling occasion, three of the replicates had to be sacrificed (destructive sampling). To this endPrior to analysis, samples were transferred into a glovebox (O₂ < 1 ppm, Innovative Technology, Amesbury, USA) prior to analysis to avoid alteration of the samples'

redox state.

EAC_{OM} and EDC_{OM} were measured chronoamperometrically using chronoamperometry (CHI1000C, CH Instruments, Austin, USA) by mediated electrochemical reduction (MER) and oxidation (MEO) (Aeschbacher et al., 2010; Klüpfel et al., 2014). The cell consisted of a cylindrical glassy carbon-C 340 working electrode, a pplatinum wire counter electrode in a glass-ceramic frit, and an Ag/AgCl reference electrode. Cells were filled with 10 mL of 0.01 M/0.1 M MOPS/KCl-bBuffer to stabilize the pH at 7 and were continuously stirred during measurement. To facilitate electron transfer, organic mediators were added to the buffer : 180 µL DQ (diaquat-dibromide monohydrate, Sigma-Aldrich) for MER, and 180 µL ABTS (2,2'-azino-bis(3-ethylbenzthiazoline-sulfonic acid), Sigma-Aldrich) for MEO at a 345 potential of $E_h = -0.69$ V and $E_h = +0.41$ V, respectively (reported vs. the standard hydrogen <u>H</u>₂ electrode but experimentally measured vs. the Ag/AgCl reference electrode). To determine the electron transfer, 100 µL of suspended samples were added to the buffer solution (Lau et al., 2015), which resulted in an increase of the current, recorded as a peak in the analysis software. After approx. ~30 minutes, when the baseline was reached again, the next sample was added to the cells. Samples were measured analyzed in duplicates. The electron transfer was calculated with the Nernst- equation and normalized to the C 350 content in the samples (Lau et al., 2015).

$$EAC_{OM} \vdash (EDC_{OM}) = peak area / (V * F * C)$$
(6)

with EAC_{OM} <u>/and</u> EDC_{OM} in μ mol e⁻ gC⁻¹, peak area in μ A sec, V = sample volume in μ L, F = Faraday constant 96,485 A sec / mol e⁻, and C = <u>carbon-C</u> content in mg L⁻¹.

EAC_{OM} and EDC_{OM} had to be corrected for <u>either ferric iron (Fe³⁺)</u>, or <u>iron (II)ferrous iron</u> (Fe²⁺) and sulfide (S²⁻) concentrations, respectively, <u>as since</u> with the applied potential <u>also</u>-Fe³⁺ would be reduced and Fe²⁺ and S²⁻ <u>would be</u> oxidized (Lau et al., 2015; Agethen et al., 2018). Fe²⁺, Fe³⁺ and S²⁻ were determined colorimetrically (Gilboa-Garber, 1971; Tamura et al., 1974) with a spectrophotometer (Cary 100 UV-Vis, Agilent, Santa Clara, USA). <u>As the determination withBecause</u> <u>1,10-phenanthroline can only detect Fe²⁺, the Fe³⁺ in the samples was reduced to Fe²⁺ with 10% ascorbic acid. The likewiseThen, theFerric iron was thus determined as the difference of concentration of total Fe and could then be used to calculate the concentration of Fe³⁺ ferrous iron in the samples.</u>

 NO_3^- and SO_4^{2-} concentrations were determined with IC, as described above. Therefore For this purpose, samples were filtered with a 0.45 µm syringe filter, filled-added to in micro-centrifuge tubes, retrieved from the glovebox and stored frozen at -21°C until analysis.

Total electron accepting capacity<u>EAC</u> (EAC_{tot}) was calculated as the sum of EAC_{OM} and EAC_{inorg} (EAC from nitrate, sulfate, <u>ferric</u> iron-(III)) considering the respective amounts of electrons transferred during the main pathways of dissimilatory reduction, i.e., assuming a reduction of NO_3^- to N_2 , of SO_4^{2-} to S^{-2} , and of Fe³⁺ to Fe²⁺ (Konhauser, 2009):

370
$$EAC_{tot} = EAC_{OM} + NO_3^{-*} 5e^{-} + SO_4^{2^-} * 8e^{-} + Fe^{3+} * 1e^{-}$$
 (7)

375

360

365

2.3 Sediment mesocosmsIntact sediment core incubations

2.3.1 :- CO₂ and CH₄ fluxes and sediment gas stock change

To obtain ex-situ CO₂ and CH₄ gas fluxes and estimate changes in sediment CO₂ and CH₄ stocks, intact 380 sediment cores (PVC tubes, 60 cm length, 5.8 cm diameter) were taken in triplicates from four sites out of the twelve 12 sites above in November 2017 (1.50, 2.100, 3.125, S.150; see Fig. 1). S.150 was chosen as one site representing to represent the sites 1.150, 2.150 and 3.150 from the same lake depth category. Sediment cores were transported cooled and deployed in a climate chamber (CLF Plant Master, CLF Plant Climatics GmbH, Wertingen, Germany) at constant conditions (temperature 20°C, humidity 60 385 %). Cores were taken to ensure that each tube contained a sediment layer of on with average thickness of 35 cm-thickness, and was covered with a lake water column of 20 cm; in the lab, we created a headspace of approx. ~150 mL. The cores were equipped with eight sampling ports; -: one in the headspace, one in the water phase and six in the sediment. Of the latterports in the sediment, three were used to sample dissolved gases in the sediment, and three were used for sediment pore water extraction. The gas samplers consisted of a gas-gas-permeable silicon tubes of 5 cm in length and 0.8 cm in diameter 390 equipped with a 3three-way- stopcock, modified after Kammann et al. (2001). This technique allows for sampling of dissolved gases in the sediment by diffusive equilibration through the silicon membrane.

For pore water sampling, a vacuum was applied with a syringe to suction samplers (Rhizon, Eijkelkamp Agrisearch, Giesbeek, Netherlands) of 5 cm in length, 0.25 cm in diameter, and about 0.1 μ m pore size (Seeberg-Elverfeldt et al., 2005). Gas and pore water samplers were deployed in average depths of 5.0 \pm 2.8, 15.3 \pm 2.9, and 23.6 \pm 2.1 cm below the sediment surface.

For statisticaldata analyses-and discussion, we only used measurements that were conductedmade >-50 days after the deployment of the intact sediment core incubations in the climate chamber-were used. This was done in order to ensure the system had adapted to experimental conditions and had reached a steady state. Steady state conditions wereAll measurements were conducted after 50 days of incubation

- of the cores in the climate chamber when the system had reached a steady state as indicated by quasiconstant CO₂ and CH₄ concentrations in the sediment, as identified by repeated monitoring of dissolved gases.
- For the determination of To determine CO₂ fluxes, the cores were closed gas-_tight with a stopper and
 connected to a laser-based, portable greenhouse-_gas analyzer (Los Gatos Research, San Jose, USA),
 which allowed to for measure measuring real-time increase of CO₂, CH₄ and H₂O concentrations in the
 headspace of the cores with a resolution of 1 Hz. As the headspace was too small for the instrument's
 flow rate, a gas bag with a volume of 150 mL was interposed between the headspace and the analyzer.
 The headspace was closed for 10 minutes, and the diffusive CO₂ flux was calculated by linear regression
 (R² > 0.8) of using the increase in concentration over time and by the ideal gas law, corrected for air
- pressure and temperature and related to the water surface area:

$$\mathbf{F} = \Delta \mathbf{c} / \Delta \mathbf{t} * (\mathbf{p} * \mathbf{V}) / (\mathbf{A} * \mathbf{R} * \mathbf{T})$$
(8)

where F is the CO₂ flux in μ mol m⁻² d⁻¹, $\Delta c/\Delta t$ is the slope of the linear regression in ppm d⁻¹, p is the air pressure in atm, V is the sum of headspace and gas bag volume in m³, A is the water surface area in m², R is the ideal gas constant 8.2*10⁻⁵ m³ atm mol⁻¹ K⁻¹, and T is the temperature in K.

CH₄ fluxes were determined by closing the cores with the stopper for 24 hours, and <u>then</u> taking a gas sample right after closing, and <u>then</u> again after 24 hours with a syringe from the headspace. CH₄ fluxes were calculated according to Bastviken et al. (2004):

$$F(CH_4) = k * (C_w - C_{fc})$$

$$\tag{9}$$

420 where F is the CH₄ flux in mmol $m^{-2} d^{-1}$, k is the piston velocity in m d⁻¹, C_w is the measured CH₄ concentration in the water phase in mmol m^{-3} and C_{fc} is the CH₄ equilibrium concentration in the headspace at the given CH₄ water concentration.

The piston velocity k was determined as:

395

400

$$\mathbf{k} = (-\ln((\mathbf{c}_{\text{sat}} - \mathbf{c}_{\text{end}})/(\mathbf{c}_{\text{sat}} - \mathbf{c}_{\text{start}}))/\Delta \mathbf{t} * \mathbf{V}) / (\mathbf{A} * \mathbf{K}_{\text{h}} * \mathbf{R} * \mathbf{T})$$
(10)

425 where c_{sat} is the saturation concentration in the chamber headspace at the measured CH₄ water concentration, c_{end} is the measured CH₄ concentration in the chamber headspace at the end of the flux measurement, <u>and</u> c_{start} is the measured CH₄ concentration in the chamber headspace at the beginning of the flux measurement (all in µatm).

The flux was corrected for the non-linear increase of CH_4 concentration in the headspace over time due to saturation and divided into diffusive and ebullitive proportions based on the piston velocity (k < 2 = diffusion, k > 2 = ebullition).

2.3.2 Sediment gas stock change

435

445

CH₄ and CO₂ concentrations in the sediment were obtained from gas-permeable silicon tubes, determined by gas chromatography as described above (2.2.3) and calculated by Henry's \underline{L} aw using temperature temperature-corrected Henry's constants (see formula equation 4). Measured CO₂ concentrations were corrected for pH (formula equation 5).

The storage change of CO_2 and CH_4 in the sediment was calculated for each depth segment between two sampling ports as the difference between of CO_2 and CH_4 concentrations obtained from silicon gas samples at the beginning and at the end of gas flux measurements:

440
$$\Delta \text{CO}_2/\text{CH}_4 = \left(\left(c(\text{CO}_2/\text{CH}_4)_{\text{end}} * \text{V}_{\text{seg}}\right) - \left(c(\text{CO}_2/\text{CH}_4)_{\text{start}} * \text{V}_{\text{seg}}\right)\right) / \Delta t$$
(11)

$$\Delta CH_4 = \left(\left(c(CH_4)_{end} * V_{seg}\right) - \left(c(CH_4)_{start} * V_{seg}\right)\right) / \Delta t$$
(12)

where Δ -CO₂ (Δ /CH₄-) is the storage change in mmol d⁻¹, c(CO₂)_{end/start}/(c(CH₄)_{end/start}) is the CO₂ <u>or</u>/ CH₄ sediment pore gas concentration at the end/beginning of the flux measurement in mmol m⁻³, and V_{seg} is the volume of the sediment core segment between two samplers in m³.

After completion of flux measurements, <u>intact</u> sediment <u>mesocosms-core incubations</u> were eventually cut into 10 cm slices, freeze-dried and ground for solid phase analyses, as described above.

2.4 Other chemical and physical parameters in water and sediment and lake water samples

- Total phosphorus (P), sulphur-sulfur (S), manganese (Mn) and iron (Fe) in the sediment were determined by wavelength dispersive X-ray fluorescence (WD-XRF; ZSX Primus II, Rigaku, Tokyo, Japan). To this end, 500 mg of freeze-dried and ground sample were pressed into pellets together with 50 mg of wax (Hoechst Wax C, Merck, Darmstadt, Germany) as a pelleting agent. Calibration of the instrument was done using a set of 22 certified reference materials.
- 455 Dissolved organic carbon (DOC) and total nitrogen (TN) concentrations in watery samples and sediment pore water were determined by catalytic oxidation and subsequent NDIR detection with a total

carbon/nitrogen analyzer (TOC-L/TNM-L, Shimadzu, Kyoto, Japan). Calibration was verified at each measurement day with potassium hydrogen phthalate (5 and 25 mg L⁻⁴) and potassium nitrate (1 and 10 mg L⁻⁴) standard solutions.

Grain size distribution was determined after Austrian standards (OENorm B 4412; OENorm L 1050; OENorm L 1061) by the Physio-geographic Lab of the Institute of Geography and Regional Research of the University of Vienna. To this end, the organic substance-matter was removed from the samples with hydrogen peroxide (H₂O₂) prior to analyses and mineral fine soil-sediment was divided into clay (< 2 µm), silt (fine (2-6 µm), medium (6-20 µm), coarse (20-63 µm)) and sand (fine (63-200 µm), medium (200-630 µm), coarse (630-2000 µm)).

Values of pH, conductivity and dissolved O₂ concentration were determined in-situ with a multi-probe (WTW Multi 3420 + IDS sensor, WTW GmbH, Weilheim, Germany). DOC and TN in lake water samples were determined by catalytic oxidation and subsequent NDIR detection with a total C and N analyzer (TOC-L/TNM-L, Shimadzu, Kyoto, Japan). Total Cl, Ca, Fe, K, Mg, Na, P and S concentrations in lake water samples were determined by inductive-coupled plasma optical emission

spectroscopy (ICP-OES, Spectroblue, SPECTRO Analytical Instruments GmbH, Kleve, Germany).

2.5 Statistics

470

All statistical analyses were conducted with R Studio, Version 3.5.-2 (R Core Team, 2018). Data was tested for normal distribution and homoscedasticity with <u>the</u> Shapiro-Wilk and <u>the</u> Levene- <u>T</u><u>t</u>est (Fox and Weisberg, 2011), respectively. For non-normally distributed data, significant differences between groups were identified using <u>the</u> Kruskal-Wallis <u>test</u> and <u>the</u> <u>Ppost-hoc Dunn- t</u><u>T</u>est (Dinno, 2017) for more than two groups and <u>using the</u> Mann-Whitney-<u>T</u><u>t</u>est for comparing two groups. If the condition of homoscedasticity was not fulfilled, groups were compared with Mood's <u>Mm</u>edian <u>T</u><u>t</u>est. Correlations and regressions between production rates and sediment parameters were calculated by Spearman's
<u>Rr</u>ank <u>C</u><u>c</u>orrelation and by linear regression models respectively. All data was tested on a 95% confidence interval and <u>a</u> significance level of α = 0.05.

3 Results

3.1 Sediment Slurry incubations

485 3.1.1 Sediment Organic MatterOM Quality

The C content in the samples was between 2.15-2% and 33.162%, with <u>the</u> lowest values at site 3.50 and <u>the</u> highest at site 1.50. C/N ratios ranged from 10.9711.0 at site 1.150 to 19.06-1 at site 3.100. Neither <u>the</u> C content nor <u>the</u> C/N ratio showed significant changes with sediment nor lake depth, but <u>the</u> C/N ratio was significantly higher in samples taken close to the shore (50) than in samples from the lake center (150) (p < 0.01) (see Fig. 2a). C and N isotopic values did not vary much between sites and

490

505

were on average -27.6 ‰ and -0.6 ‰ respectively.

Organic matter<u>OM</u> quality as identified by FTIR-_analysis was predominated by strong absorption features of polysaccharides, lignin, humic acids, phenolic and aliphatic structures, aromatic compounds, and fats, waxes and lipids (see Fig. S1). Except for lignin, which was not identified at sites 2.50, 2.100,

2.125, 3.50, and 3.100, all components were <u>found-abundant</u> at all sites. Ranges of peak ratios for <u>every</u> <u>each</u> component <u>class</u> can be found in Tab.-le 2. Overall, <u>the</u> lowest <u>FTIR</u> peak ratios were present at site 3.50 and <u>the</u> highest at site 3.125, corresponding to highest and lowest CH₄ production rates <u>(see below)</u>. All peak ratios correlated with each other. They tended to increase <u>(i.e., indicate more decomposed material)</u> with sediment depth and towards the lake center, <u>respectively</u>, but this change was not significant <u>(see Fig. 2b)</u>.



Figure 2: (a) C/N ratio and (b) absorption ratio of fats and polysaccharides at different lake depth categories. Identical lowercase letters indicate C/N ratios that were not significantly different (i.e. p > 0.05) from each other. N.s. means that absorption ratios did not exhibit significant differences between lake depths. n(50, 100) = 5, n(125, 150) = 6

	Min	Max	Average
C (%)	2. 15 2	33. 16 2	2645
N (%)	0. <u>152</u>	2.45	<u>+2</u> .97 <u>0</u>
C/N ratio	1 0.97<u>1.0</u>	19. 06<u>1</u>	13.8 <mark>1</mark>
δ ¹³ C (‰)	-27.84	-27.3	-27.6
δ ¹⁵ N (‰)	-6. 27<u>3</u>	1.3 <mark>3</mark>	-0.9 <mark>3</mark>
P (%)	0.3	0.5 1	0. 37<u>4</u>
S (%)	0.1 3	1.2 2	0.9 3
Mn (%) (ppm)	0.037<u>370</u>	<u>0.121160</u>	0.049<u>490</u>
Fe (%)	2.1 3	7.04	<u>2.953.0</u>
Polysaccharides (1033-1035 cm ⁻¹)	0.58	<u>1.24</u>	0.81
Lignin (1220-1234 cm ⁻¹)	0	0.4 <u>3744</u>	0. 235<u>24</u>
Humic acids (1417-1419 cm ⁻¹)	0. 039<u>04</u>	0.491	0. 328<u>33</u>
Phenols & aliphatics (1456 cm ⁻¹)	0. 018<u>02</u>	0.48 <mark>0</mark>	0.324
Other Aromatics (1623-1646 cm ⁻¹)	0.21 0	0. 825 83	0. 629<u>63</u>
Fats, waxes, lipids (2850-2856 cm ⁻¹)	0.178	0.492	0. 349<u>35</u>

Table 2: C, N, P, S, Mn, and Fe contents, C/N and FTIR peak ratios (peak maximum for polysaccharides) of identified OM compounds and their minimum and maximum values. n = 16-22.

3.1.2 CO₂ and CH₄ production rates

- 510 <u>Overall, production rates decreased from the shore to the center of the lake.</u> Rates for potential CO₂ production in-at_0-5 cm depth ranged from 10.309 \pm 0.003 µmol gC⁻¹ d⁻¹ at site 3.125 to 38.515 \pm 2.48 5 µmol gC⁻¹ d⁻¹ at site 2.100. Potential CH₄ production lay-was between 7.3 \pm 0.14 µmol gC⁻¹ d⁻¹ at site 3.125 and 33.53 \pm 7.151-2 µmol gC⁻¹ d⁻¹ at site 3.50. Production rates in-at_5-10 cm depth were always lower compared to the upper sediment layer and were between 7.189-2 \pm 0.072-1 and 14.714 \pm 0.435 µmol gC⁻¹ d⁻¹ for CO₂ and between 5.361-4 \pm 0.259-6 and 14.264-3 \pm 0.341 gC⁻¹ d⁻¹ for CH₄. <u>Overall,</u> production rates decreased from the shore to the center of the lake.</u>Both CO₂ and CH₄ production rates showed significant differences between sites (Kruskal-Wallis, p < 0.001) and were significantly (Dunn's, p < 0.05) higher at the shore (50+100) than in the center of the lake (125+150) (see Fig. 2-3 + Fig. S2 & S3).
- CO₂ and CH₄ production rates decreased with time. CO₂ production was highest at the beginning, while CH₄ production had its peakwas retarded and highest only after three to eight days of incubation (s. Fig. <u>43</u>, Tab-<u>le</u> 3). The CO₂/ CH₄ amount ratio constantly decreased during the incubation, with a maximum value of 62.1³ ± 58.44 at the beginning of the incubation at site 1.125 and minimum values of 1.17-<u>2</u> ± 0.004 at the end of the incubation at site 2.50, approaching ratios of 1 as expected under strictly methanogenic conditions (s. also Tab-le 3).



Figure 3: CO_2 (a) and CH_4 (b) production rates <u>in-at</u> 0-5 (grey) and 5-10 (white) cm sediment depth. n = 3. Production rates <u>are-were</u> calculated from <u>the</u> linear regression of concentration increase over time. Bold lines are the median, boxes show the 25 and 75 percentile, and whiskers indicate minima and maxima within 1.5 times the interquartile range. Different letters indicate significant differences between these sites Identical lowercase letters indicate production rates that were not significantly different (i.e. p > 0.05) from each other.



Figure 4: $CO_2(a)$ and $CH_4(b)$ production over time at all sites <u>in-at_0-5</u> cm sediment depth. Top: transect 1-: middle: transect 2-: bottom: transect 3. Line<u>s</u> are average values of triplicate measurements \pm SD.

535

540

Table 3: CO_2 and CH_4 production rates and <u>the CO₂/CH₄ amount</u> ratio over time. Rates are in μ mol gC⁻¹ d⁻¹ -: ratio is in μ mol. Values in brackets are SDs.

	sediment	day of incubation						
	depth	ŧO	ŧ1	<u>t23</u>	<u>t38</u>	ŧ4 <u>11</u>	t5<u>14</u>	t6<u>18</u>
CO ₂	0-5 cm		91. <mark>49<u>5</u></mark>	44. 6 7	26. <mark>49<u>5</u></mark>	21.14	6. 79<u>8</u>	13.3 2
production			(40. 29 <u>3</u>)	(16. 77<u>8</u>)	(9. 69<u>7</u>)	(7. <u>6</u> 59)	(3. <u>6</u> 59)	(5.0 <mark>0</mark>)
rate	5-10 cm		39. 86 9	19.6 <mark>3</mark>	11.0 1	11.4 <mark>0</mark>	6.0 <mark>0</mark>	8.9 <mark>1</mark>
			(14. 18<u>2</u>)	(<u>56</u> . <u>950</u>)	(2. <u>697</u>)	(3. 06<u>1</u>)	(2.2 <mark>0</mark>)	(2.7 4)
CH ₄	0-5 cm		14. 37<u>4</u>	22.8 <mark>0</mark>	23. 88 9	16.7 <mark>4</mark>	11.6 <mark>0</mark>	13. 58<u>6</u>
production			(7.4 4)	(11. 26 <u>3</u>)	(11. 47<u>5</u>)	(7. <u>9</u> 86)	(4. 69<u>7</u>)	(578)
rate	5-10 cm		9. 36<u>4</u>	11. 68<u>7</u>	11. 56 6	9. 72 7	7. 89 9	9. 32 3
			(2. <mark>67<u>7</u>)</mark>	(2. <mark>859</mark>)	(3. <mark>13<u>1</u>)</mark>	(2. <mark>34<u>3</u>)</mark>	(2. <u>101</u>)	(2. 72<u>7</u>)
CO ₂ /CH ₄	0-5 cm	14. 55<u>6</u>	8. 70<u>7</u>	3. 84<u>8</u>	2. 33<u>3</u>	2. 11 1	1. 89 9	1. 73<u>7</u>
ratio		(6. <mark>71<u>7</u>)</mark>	(2. 26 <u>3</u>)	(1. 32 <u>3</u>)	(0. <mark>48<u>5</u>)</mark>	(0. <u>455</u>)	(0. <u>374</u>)	(0. 30 <u>3</u>)
	5-10 cm	16. 19 2	5. 61<u>6</u>	2. 92 9	1. 93 9	1. 74<u>7</u>	1. 57<u>6</u>	1.44 <u>4</u>
		(24. <mark>909</mark>)	(1. 60<u>6</u>)	(0. 88 <u>9</u>)	(0. <u>354</u>)	(0. <u>343</u>)	(0. 25 <u>3</u>)	(0. 20 2)

 Q_{10} -values were between $1.\underline{56} \underline{6} \pm 0.\underline{13} \underline{1}$ and $2.\underline{19} \underline{2} \pm 0.\underline{14} \underline{1}$ for CO_2 production rates and between $2.\underline{65}$ $\underline{7} \pm 0.\underline{48} \underline{5}$ and $11.\underline{37} \underline{4} \pm 0.\underline{961} \underline{.0}$ for CH_4 production rates.

Sampling dates did not have any impact on production rates as confirmed by <u>a</u> Kruskal-Wallis-test (CH₄: p = 0.173, CO₂: p = 0.75576) Neither CO₂ nor CH₄ production rates were correlated<u>exhibited significant correlations</u> with C content $\Theta_{\frac{1}{2}}$ the C/N ratio, $\frac{\delta^{13}C}{\delta^{15}N}$, but we found significant negative correlations (p < 0.01, rho < -0.6) between all <u>FTIR</u> peak ratios and CO₂ and CH₄ production rates as well as <u>of FTIR peak ratios</u> with <u>and</u> Q₁₀-values of CH₄ production (p < 0.05, rho = -0.82) (see Fig. 45, test statistics see Table S1, SI).



Figure 5: Correlations between CO₂ and CH₄ production rates and OM quality parameters <u>as determined by</u> <u>elemental analysis or FTIR spectroscopy</u>. <u>Different colors denote different depth categories (see Fig. 4)</u>.

550

3.1.3 Methanogenic pathways

Both H_2 and acetate concentrations increased during the incubation. H_2 concentrations were between 0 and $1587.42 \pm 170.12 - 1$ nmol L⁻¹ and acetate concentrations ranged from $219.74 - 7 \pm 104.25 - 3$ to $1212.71-7 \pm 11.35-4$ µmol L⁻¹ (see Fig. 56). Gibb's free energy for acetoclastic methanogenesis was 555 between - 96.32.3 \pm 0.84.8 and -66.37.4 \pm 0.28.3 kJ mol⁻¹ and for hydrogenotrophic methanogenesis it was between $-62.41 \pm 0.85 - 9$ and -7.41 ± 0.0 kJ mol⁻¹. Energy yields decreased for the acetoclastic pathway and significantly increased (Mood's median, t0-t6: p < 0.05, t3-t6: p < 0.001) increased for the hydrogenotrophic pathway throughout the incubation (see Fig. 56). Energy yields did not differ significantly between lake depths except for the acetoclastic pathway at the end of the incubation (t6, p < 0.001), when the energy yield was highest in samples from the center of the lake. Significant 560 differences between all sites were found for the acetoclastic and hydrogenotrophic pathway at the beginning of the incubation (Kruskal-Wallis, p < 0.01) and at t6 the end of the incubation for acetoclastic methanogenesis only (Kruskal-Wallis, p < 0.01). H₂ concentrations at the end of the experiment exhibited significant positive correlation were significantly correlated with average CO_2 (p < 0.001, rho 565 = +0.51) and CH₄ (p < 0.001, rho = +0.45) production rates (test statistics see Table S2, SI). Further, Gibb's free energy of acetoclastic methanogenesis exhibited a significant positive correlation was positively correlated with C/N ratio (p < 0.05, rho = +0.45) at the end of the experiment (t6). Acetate concentration at the end of the incubation exhibited significant negative correlations with FTIR peak ratios and C/N ratio (p < 0.05, rho < -0.44). Gibb's free energy of hydrogenotrophic methanogenesis exhibited significant positive correlations with FTIR peak ratios of fats, humic acids, phenolics and C content (p < 0.05, rho > 0.17), i.e. a high energy yield was associated with low FTIR peak ratios and C content. Correspondingly, H₂ concentrations exhibited significant negative correlations with all FTIR peak ratios and C content (p < 0.05, rho < -0.15) (test statistics see Table S2, SI).

570





3.1.4 Alternative Organic and Inorganic Electron AcceptorEAs

EAC_{OM} lay was between 218.69-7 ± 97.15-2 and 545.71-7 ± 60.33-3 µmol e⁻ gC⁻¹ at t0-the beginning and decreased by anon average of by 44.85-9 µmol e⁻ gC⁻¹ to 170.76 ± 32.70 to 460.79 ± 51.47 µmol e⁻ gC⁻¹ at until t6the end of the slurry incubation. with hThe highest values for EAC_{OM} were found at site 3.125 corresponding to lowest measured CH₄ production rates at that site. EAC_{inorg} was, with values between 19.82-2.9 ± 10.52-1.0 and 218.01-39.2 ± 24.79-4.4 µmol e⁻ gC⁻¹. EAC_{tot} ranged from 226.95 250.8 ± 83.87-93.9 µmol e⁻ gC⁻¹ at site 3.150 to 614.58-551.4 ± 80.72-60.3 µmol e⁻ gC⁻¹ at site 3.125 and significantly decreased from the beginning to the end of the incubation(t-test, p < 0.001) decreased by averagely by on average 87.25-51.2 µmol e⁻ gC⁻¹ from t0 to t6. EDC_{OM} was between 149.54-5 ± 26.72-7

and $462.69-7 \pm 18.57-6 \,\mu$ mol e⁻ gC⁻¹ at the beginning and between $152.94-9 \pm 53.78-8$ and $370.65-7 \pm 196.22-2 \,\mu$ mol e⁻ gC⁻¹ at the end of the incubation with showing a slight increase decrease by, on average, $31.384 \,\mu$ mol e⁻ gC⁻¹. The lLowest EDC_{OM} at both t0-the beginning and t6-the end of the incubation was found at site 2.100, corresponding to the highest CO₂ production rates there (see Fig. 67).

We further found significant differences for EAC_{OM} and EAC_{tot} (ANOVA, p < 0.001) and EAC_{tot} (ANOVA, p < 0.01) between all sites, with <u>the</u> highest average values at site 3.125 and <u>the</u> lowest at

- 595 sites <u>1.125</u> (t0) and <u>3.150</u> (t6) (see Fig. <u> s_3S_4 </u>). Average CO₂ and CH₄ production rates <u>exhibited a</u> <u>significant negative correlation</u> were significantly (p < 0.05, rho = 0.7) negative correlated with initial EDC_{OM} (p < 0.05, rho = -0.7), whereas we did not find any significant correlation between the production rates and with EAC, electron exchange capacity (EEC, sum of EAC and EDC) nor or the EAC/EDC ratio, although low CH₄ production rates were associated with high EAC. CO₂, CH₄ and acetate 600 concentration were significantly (p < 0.05, < 0.001) negative correlated with EAC_{tot}. Gibb's free energy of hydrogenotrophic methanogenesis was significantly (p < 0.05) positive and acetate concentration significantly (p < 0.01) negative correlated to EAC_{OM}. Acetate concentration exhibited significant negative correlations with EAC_{OM} and EAC_{tot} (p < 0.01, <0.001, rho < -0.38). Gibb's free energy of hydrogenotrophic methanogenesis exhibited significant positive correlations with EAC_{OM} and 605 EAC_{tot} (p < 0.05, rho = 0.43). We did not find any significant correlations between EAC or EDC and OM quality parameters except for EDC_{OM} and the FTIR peak ratio indicative of fats, waxes and lipids and EDC_{OM}-(p < 0.05, rho = -0.76). Of the inorganic EAs, only total sulfur-S content in the sediment exhibited a significant negative correlation was significantly negative correlated with CH4-CO2 production rates (p < 0.05, <u>rho = -0.45</u>) (test statistics see Tables S1, SI).
- 610 Calculated potential CO₂-production from prevalent organic and inorganic EAs was always lower than the measured CO₂-production in slurry incubations and was between 110.5 and 586.4 μmol CO₂-eq. gC⁻¹ (see. Fig. 8) and could explain 38 to 91% of measured CO₂production, whereof 4 to 51% where explained by EAC_{OM} alone.



615 **Figure 67:** Linear regression of CH_4 and CO_2 production rates with <u>the</u> initial <u>electron acceptingEAC</u> and <u>donating-EDC</u>capacities. Points are mean values of triplicate measurements at each site. <u>At site 2.100, highest</u> <u>CO_2-production rate concurred with lowest EDC_{OM}, at site 3.125 lowest CH_4 -production rate concurred with highest EAC_{OM}. Different colors denote different depth categories (see Fig. 4).</u>



*Figure 8: Calculated expected CO*₂-production rate from prevalent EAs vs. measured potential CO₂-production rate in slurry incubations. Dashed line shows the closed budget of expected and measured CO₂-production.



Figure 7: EAC_{OM}(a), EAC_{inorg}(b), EAC_{tor}(c) and EDC_{OM}(d) at the beginning and the end of the incubation at all sites. n = 60-88. Different letters denote significant differences.

3.2 <u>Sediment mesocosmsIntact sediment core incubations</u>: Fluxes, sediment storage change and grain size

CO₂ and CH₄ fluxes measured from <u>intact</u> sediment <u>core mesocosms-incubations</u> ranged from 4.46<u>10.8</u> \pm 4.4 to 26.917. 9 \pm 2.0 and 0-0.02 \pm 0.01 to 9.81.5 \pm 2.6 mmol m⁻² d⁻¹, respectively. CH₄ ebullition played a major role in cores from sites 2.100 and S.150 and accounted for up to 100% of the fluxes there. CO₂ fluxes at site 3.125 were significantly <u>lower</u> (t-test, p < 0.05) lower than at the other sites while total CH₄ fluxes at site 2.100 were significantly <u>higher</u> (Kruskal-Wallis, p < 0.01) higher compared to all other sites. Fluxes were within the same range <u>like-as</u> potential production rates in <u>slurry</u> incubations but <u>did</u>-only partly follow<u>ed</u> the pattern observed in <u>slurry</u> incubations (see Fig. <u>89a+b</u>).



Figure 7: Comparison of fluxes obtained from <u>intact</u> sediment <u>mesocosms_core incubations</u> and potential production rates from the <u>slurry</u> incubations (<u>in at</u> 0-5 cm sediment depth) for $CO_2(a)$ and diffusive and total $CH_4(b)$. Points are averages (n(slurry) incubations) = 3-9, $n(CO_2 fluxes) = 9-14$, $n(CH_4 fluxes) = 11-13$), horizontal and vertical lines are standard deviations. We decided to display production rates from <u>the</u> 0-5 cm sediment depth, as this depth turned out to be <u>the</u> most important for sediment <u>gas_GHG</u> production (see sections 3.1.2 & 4.1.21). (c) Relative grain size distribution in <u>intact</u> sediment <u>mesocosms_core incubations</u>.

Table 4: Spearman's rank correlation coefficients for CH_4 and CO_2 fluxes (means of each <u>intact</u> sediment <u>mesocosmcore incubation except for correlations with sediment stock changes where values from each sampling date were used</u>) and different sediment characteristics. <u>n.s. means that correlations were not significant (p > 0.05)</u>.

	CH ₄ flux		CO₂ flux	
	rho	P	rho	P
Clay	0.6477503 <u>648</u>	<u>0.02275≤0.05</u>	0.6045669 <u>605</u>	<u><0.05</u> 0.03731
Silt	0.4966<u>5</u>085	n.s.	0.3022835	n.s.
Sand	- 0.6477503<u>648</u>	<u><0.05</u> 0.02275	-	<u> </u>
			0.6045669<u>605</u>	
Fats, waxes, lipids	-0.8333333	<u><0.05</u> 0.01538	-0.3333333	n.s.
Phenols; humics	-0.8333333	<u><0.05</u> 0.01538	-0.3571429	n.s.
Aromates	-0.5952381	n.s.	-	n.s.
			0.5238095<u>524</u>	
Lignin	- 0.7863867	<u><0.05</u> 0.02063	-	n.s.
			0.3805097<u>381</u>	
C/N	-0.8809524 <u>881</u>	0.007242 ≤	-0.3333333	n.s.
		<u>0.01</u>		
C (%)	- 0.7142857	n.s.	-0.1904762	n.s.
CH ₄ sediment stock change	-0.2219706 <u>222</u>	n.s.	0.04985994 <u>05</u>	n.s.
CO ₂ -sediment stock change	-	n.s.	-0.06414566	n.s.
	0.04872526<u>049</u>			

	<u>(</u>	CH ₄ flux		<u>CO₂ flux</u>			
	<u>rho</u>	p	<u>n</u>	<u>rho</u>	p	<u>n</u>	
Clay	0.648	< 0.05	<u>12</u>	<u>0.605</u>	< 0.05	<u>12</u>	
Silt	0.497	<u>n.s.</u>	<u>12</u>	0.302	<u>n.s.</u>	<u>12</u>	
Sand	<u>-0.648</u>	< 0.05	<u>12</u>	<u>-0.605</u>	< 0.05	<u>12</u>	
Fats, waxes, lipids	-0.833	< 0.05	<u>8</u>	-0.333	<u>n.s.</u>	<u>8</u>	
Phenols; humics	-0.833	< 0.05	<u>8</u>	-0.357	<u>n.s.</u>	<u>8</u>	
Aromatics	<u>-0.595</u>	<u>n.s.</u>	<u>8</u>	-0.524	<u>n.s.</u>	<u>8</u>	
Lignin	<u>-0.786</u>	<u>< 0.05</u>	<u>8</u>	<u>-0.381</u>	<u>n.s.</u>	<u>8</u>	
<u>C/N</u>	<u>-0.881</u>	<u>< 0.01</u>	<u>8</u>	<u>-0.333</u>	<u>n.s.</u>	<u>8</u>	
<u>C (%)</u>	-0.714	<u>n.s.</u>	<u>8</u>	-0.190	<u>n.s.</u>	<u>8</u>	
CH ₄ sediment stock change	-0.222	<u>n.s.</u>	<u>41</u>	<u>0.05</u>	<u>n.s.</u>	<u>35</u>	
CO ₂ sediment stock change	-0.049	<u>n.s.</u>	<u>41</u>	-0.064	<u>n.s.</u>	<u>35</u>	

Concentrations of dissolved CH₄ and CO₂ (Σ (CO₂, HCO₃⁻, CO₃²⁻)) in the sediment ranged from 3.974.0 to 129.45 mmol m⁻³ (CH₄) and from 322.92 to 3811.36-4 mmol m⁻³ (CO₂), respectively with the lowest values found at site 3.125 and the highest values at S.150. We did not see significant changes in CO₂ and CH₄ concentrations in the depth profile (see Fig. <u>S4S5</u>).

Observed changes of CH_4 and CO_2 concentrations in the sediment of intact cores was were overall very low and <u>lay-were</u> between 0.06-1 and 2.45 (CH₄) and 0.55-6 and 57.11 mmol d⁻¹ (CO₂) in the upper 5 cm of the sediment. It was within These changes were on the same order of magnitude as measured

fluxes but did not correlate with the latterm.

Grain sizes distribution differed between the four sites, with site 3.125 having the highest share of sand (21.6%) and <u>the</u> lowest shares of silt and clay (35.68–7% and 42.73%) whereas the other sites were dominated by finer material and consisted of contained less than 5% of sandy components (see Fig. <u>89c</u>). CH₄ and CO₂ fluxes <u>exhibited significant correlation</u> were significantly correlated with clay and sand content, but not with silt (see Tab-<u>le 4</u>5).

Similar to <u>slurry</u> incubations, CH₄ fluxes <u>exhibited significant negative correlations</u> were significantly negative correlated with some FTIR peak ratios indicative of recalcitrant OM compounds as well as and with <u>the C/N</u> ratio but not with C content. CO_2 fluxes did not show any significant correlation with OM quality parameters at all (see Tab-le 54).

665

655

660

4 Discussion

4.1 Sediment Slurry incubations

4.1.1 <u>Variability of CO₂ and CH₄ production rates and OM quality</u>Spatial variability of OM quality

- 670 Results from the slurry incubation experiment showed that both CO₂ and CH₄ production rates were despite its small size - highly variable within the examined lake. CH₄ production rates were within the range of previously reported values from lakes in central Sweden, a reservoir in Brazil, and in sediments from Lakes Stechlin and Geneva in Germany and Switzerland, respectively (Duc et al., 2010; Fuchs et al., 2016; Grasset et al., 2018). CO₂ production rates were high compared to rates found in Lake Kinneret
- 675 in Israel and in the Pantanal and Amazon regions in Brazil, exceeding reported values by a factor of 7 to 100 (Schwarz et al., 2008; Conrad et al., 2011), supporting the assumption that small lakes generally have higher C turnover rates compared to larger ones.

In all samples, production rates were higher at the beginning of the experiment than toward the end. While CO₂ production rates were highest right after the start and then constantly decreased until reaching

a plateau around day 8, CH₄ production rates peaked after 3 to 8 days and then slowly decreased afterwards approaching a 1:1 CO₂:CH₄ production ratio. This typically observed three-phase pattern (Yao et al., 1999) might among other things be due to thermodynamic constraints, such as end product

accumulation in the sample vials and subsequent inhibition of respiration and methanogenesis (Beer and Blodau, 2007; Blodau et al., 2011; Bonaiuti et al., 2017). The observed delay in notable CH_4 production was expected as prevalent alternative EAs (NO₃⁻, Fe³⁺, SO₄²⁻, humic substances) likely suppressed methanogenic activity (Lovley et al., 1996; Yao et al., 1999; Fenchel et al., 2012).

685

690

700

Both CO_2 and CH_4 production rates were higher in the topmost sediment layer compared to the 5-10 cm sediment depth, which suggests that the first centimeters of the sediment play a major role in GHG production as a consequence not only of temperature but also of microbial community distribution and changes in OM quality (Falz et al., 1999; Sobek et al., 2009; Wilkinson et al., 2015). This finding

corresponded with decreasing C/N ratios and increasing FTIR peak ratios with sediment depth. While sediment age typically increases with depth, the OM quality and, thus, reactivity usually decreases with sediment depth, as recalcitrant compounds are not being completely mineralized but instead residually enrich get buried and preserved in the sediment (Avnimelech et al., 1984; Burdige, 2007; Sobek et al., 2009).

Similar to den Heyer and Kalff (1998), we found higher production rates in samples close to the lake shore in contrast to lower production rates in the center, which suggests either that the input of OM (quantity) in the sediments is higher at the shore sites and/or the OM quality is more labile and, therefore, more easily degradable. As we did not find any correlations between CO_2 or CH_4 production rates and sediment C content as proposed e.g. by Conrad et al. (2011) or Romeijn et al. (2019), we instead suggest that the OM quality rather than quantity might be the determining factor for CO_2 and CH_4 production.

C/N ratios are frequently used to characterize the degradation state of OM, but we did not find correlations between C/N ratios and CO₂ and CH₄ production rates in the slurry incubations. Although OM of autochthonous origin was found to fuel higher degradation rates than allochthonous OM (West et al., 2012; Grasset et al., 2018) we found evidence of predominant inputs of allochthonous (terrestrial) material at sites with higher production rates close to the shore (higher C/N ratios), whereas sites with lower production rates in the lake center received mainly autochthonous (aquatic) OM as indicated by lower C/N ratios (Meyers, 1994). On the other hand, high C/N ratios also indicate a lower degradation state and therefore higher degradation potential whereas low C/N ratios are usually typical of highly

710 decomposed OM having a lower CO₂ and CH₄ production potential (Malmer and Holm, 1984; Kuhry and Vitt, 1996). These two possibilities of interpreting C/N ratios might be the reason for apparently contradicting findings and the missing relationship between C/N ratios and CO₂ and CH₄ production rates.

The abundance of recalcitrant OM compounds (lignin, aromatics, humics, phenols, and lipids) as indicated by FTIR peak ratios, proved to be a more suitable explanation for observed CO₂ and CH₄ production patterns, as indicated by strong significant negative relationships between peak ratios and production rates. All FTIR peak ratios indicative of more refractory components increased toward the lake center, which indicated an increasing predominance of more recalcitrant OM with greater distance from the lake shore and corresponded with decreasing production rates from lake shore to center. At

- 720 first sight this was unexpected as allochthonous material is known to be richer in recalcitrant compounds like lignin or aromatics compared to autochthonous biomass, therefore, is effectively buried in lake sediments (Sobek et al., 2009). However before reaching the sediment, the internally produced material in the lake center must pass through a deeper water column, meanwhile undergoing degradation processes, such that more decomposed OM reaches the sediment (Meyers, 1994; Torres et al., 2011).
- As this process might not be of the same importance in shallow lakes compared to deeper lakes, we additionally suggest that the more decomposed OM in the lake center might have undergone degradation processes during resuspension and focusing of small particles as a result of wind-induced bed-shearing (Mackay et al., 2011). The less decomposed OM close to the shore might further originate from labile aquatic plant substrates growing at the shoreline (Wetzel, 1992; Cole et al., 2007).
- 730 <u>To summarize, the OM degradability does not necessarily depend on its origin, and the OM of terrestrial</u> origin is not necessarily more recalcitrant to degradation by aquatic microorganisms in this study. Sites closer to the shore probably received higher rates of terrestrial OM matter that was less decomposed, whereas autochthonous production in the lake center was overall low, due to the low nutrient status of the lake, and was already undergoing degradation during sedimentation; this likely lead to more
- 735 recalcitrant OM in the sediment. Earlier studies by West et al. (2012) and Grasset et al. (2018), that found allochthonous material to be less decomposable compared to autochthonous matter were conducted in larger lakes, where the influence of the watershed was overall much lower than in our case, because larger lakes have a lower perimeter to volume ratio. We hypothesize that in small and nutrientpoor lakes, the pattern might be reversed as the system adapts to overall low productivity and a simultaneously high input of terrestrially produced OM.
 - In accordance with previous studies (den Heyer and Kalff, 1998; Sobek et al., 2009; Gudasz et al., 2015), we found that with a temperature increase of 10°C, production rates of CO₂ doubled and those of CH₄ were even 2 to 11 times higher. Q₁₀-values for CO₂ were within the range of earlier reported values by Liikanen et al. (2002) and Bergström et al. (2010), whereas Q₁₀-values for CH₄ production were slightly
- ⁷⁴⁵ higher than values found by Duc et al. (2010). The large observed range of Q_{10} -values, especially for CH₄, implies that responses to temperature increases might not be homogeneously distributed within a lake. The observed negative correlation between Q_{10} -values and FTIR peak ratios further suggests that sites with more labile OM are more susceptive to increasing temperatures in terms of CH₄ production, whereas at sites with more recalcitrant OM, the inherent quality of this recalcitrant OM may limit the
- 750 degradation processes. We therefore assume that sediment GHG production particularly CH₄ production - in small and shallow lakes might in the course of global warming increase to a larger extent than in deeper lakes regarding the fact that shallow waters, as against deeper lakes, do not develop a stable stratification in summer and therefore shallow sediments warm much faster (Jankowski et al., 2006).

- The high measured variability in production rates proves (1) that it is necessary to sample lake sediments from different locations in one lake in order to obtain potentials of CO_2 and CH_4 production representative of the whole lake and (2) that upscaling production rates based on single point measurements may be highly biased. Considering only sediment production rates, based on findings from other studies it seems that the observed spatial variability of CO_2 and CH_4 emissions from lakes
- might to a large extent be controlled by variability in sediment GHG production (Schilder et al., 2013; Bastviken et al., 2015; Natchimuthu et al., 2016; Natchimuthu et al., 2017; Spafford and Risk, 2018). But still, as sediment production and emission patterns have so far only been considered separately, other factors driving actual emissions might have been neglected. Therefore, in section 4.2 we further discuss the relationships between CO₂ and CH₄ production and actual emissions, as assessed in the intact sediment core incubation, and relate them to respective sediment properties.

In our study, we found higher C/N ratios in samples close to the lake shore and narrower ratios in the lake center indicating a predominant input of allochthonous (terrestrial) material at the shore and mainly sedimentation of autochthonous (aquatic) OM in the lake center (Meyers, 1994).

- Besides C/N ratios, FTIR peak ratios revealed information about the OM composition and quality in
 Lake Windsborn. We found FTIR peak ratios for refractory components (lignin, aromatics, humics, phenols, and lipids) increasing towards the lake center, indicating increasing predominance of more recalcitrant OM with higher distance from the lake shore. At first sight, this was unexpected as allochthonous material is known to be richer in recalcitrant compounds like lignin or aromates compared to autochthonous biomass and therefore being effectively buried in lake sediments (Sobek et al., 2009).
 On the other hand, the internally produced material in the lake center has to bypass a deeper water column before reaching the sediment, meanwhile undergoing degradation processes and leading to more decomposed OM reaching the sediment (Meyers, 1994; Torres et al., 2011). The less decomposed OM close to the shore might further originate from labile aquatic plant substrates growing at the shoreline (Wetzel, 1992; Cole et al., 2007).
- 780 Not only on a spatial scale, but also with sediment depth, C/N ratios decreased and FTIR peak ratios increased, corresponding with a decrease in CO₂ and CH₄ production rates. While sediment age typically increases, OM quality and thus reactivity usually decreases with sediment depth, as recalcitrant compounds are not being mineralized completely but instead buried and preserved in the sediment (Avnimelech et al., 1984; Burdige, 2007; Sobek et al., 2009).
- 4.1.2 Spatial variability and temperature dependency of CO₂ and CH₄ production rates
 Our incubation experiment showed that both CO₂ and CH₄ production rates were highly variable within one lake of only small spatial extent. CH₄ production rates were within the range of formerly reported values from lakes in central Sweden, a reservoir in Brasil, and in sediments from Lakes Stechlin and Geneva in Germany and Switzerland (Duc et al., 2010; Fuchs et al., 2016; Grasset et al., 2018). CO₂

Amazon regions in Brazil and exceeded reported values by a factor of 7 to 100 (Schwarz et al., 2008; Conrad et al., 2011). Compared to production rates after addition of fresh OM to lake sediment, our CO_2 and CH_4 -production rates remained low (Grasset et al., 2018).

Both CO₂-and CH₄-production rates were higher in the topmost sediment layer compared to 5-10 cm
 role for gas production as a consequence of temperature and microbial community distribution and changes in OM quality (as discussed below) (Falz et al., 1999; Sobek et al., 2009; Wilkinson et al., 2015).

 In all samples, production rates were higher at the beginning of the experiment than towards the end. While CO₂ production rates were highest right after the start and then constantly decreased until reaching
 a plateau around day 8, CH₄ production rates peaked after 3 to 8 days and then slowly decreased afterwards approaching a 1:1 CO₂:CH₄ production ratio. This typically observed 3 phase pattern (Yao et al., 1999) might e.g. be due to thermodynamic constraints, such as end product accumulation in the sample vials and subsequent inhibition of methanogenesis (Beer and Blodau, 2007; Blodau et al., 2011; Bonaiuti et al., 2017). The observed delay in notable CH₄ production can be explained by the prevalent alternative EAs (NO₃⁻, Fe³⁺, SO₄²⁻, humic substances) suppressing methanogenic activity (Lovley et al., 1996; Yao et al., 1999; Fenchel et al., 2012).

Similar to den Heyer and Kalff (1998), we found higher production rates in samples close to the lake shore in contrast to lower production rates in the center, suggesting that either the input rate of OM to the sediment is higher and therefore fueling higher degradation rates or/and that the organic material at these sites is more labile and therefore more easily degradable for microorganisms. More details on that hypothesis will be given in the following.

810

The high measured variability in production rates showed that it is necessary to sample lake sediments from different locations in one lake, in order to understand potentials of CO₂ and CH₄ production representative of the whole lake and that upscaling production rates based on single point measurements may be highly biased. Considering only sediment production rates, it seems that the in other studies observed spatial variability of CO₂ and CH₄ emissions from lakes (Schilder et al., 2013; Bastviken et al., 2015; Natchimuthu et al., 2016; Natchimuthu et al., 2017; Spafford and Risk, 2018) might to a large extent be controlled by sediment gas production. But still, as sediment production and emission patterns have so far only been considered separately, important relations might have been neglected. In section

820 4.2, we therefore further discuss the relationships between CO₂ and CH₄ production and actual emissions and relate them to sediment properties.

Many studies revealed that higher temperatures lead to enhanced mineralization of OM due to higher microbial activity and thus increased production of CH₄ and CO₂ in sediments (den Heyer and Kalff, 1998; Sobek et al., 2009; Gudasz et al., 2015). Accordingly, we found that production rates of CO₂ were

825 2 times and of CH₄-2 to 11 times higher with a temperature increase of 10°C. Q_{10} values for CO₂ were

within the range of earlier reported values by Liikanen et al. (2002) and Berström et al. (2010) whereas Q₁₀-values for CH₄ production were slightly higher than values found by Duc et al. (2010). The high observed range of Q₁₀-values, especially for CH₄, implies that responses of temperature increase might not be homogeneously distributed within one single lake. The observed negative correlation between Q₁₀-values and OM quality indicators suggests that sites with more labile OM are more vulnerable to increasing temperatures in terms of CH₄-production and that at sites with more recalcitrant OM, the latter may limit the degradation processes. Aben et al. (2017) and Kiuru et al. (2018) showed that increasing temperatures caused higher CO₂ and CH₄-emissions from lakes. Assuming that gas production in the sediment and efflux to the atmosphere are coupled, global warming may thus drastically enhance emissions from small lakes like Lake Windsborn, especially regarding the fact that these sediments warm much faster (max. bottom water temperature in summer 2018: 26.65°C, surface water temperature at the same time: 27.14 °C) compared to deeper lakes which have a deeper water column and stable thermal summer stratification as buffer (Jankowski et al., 2006).

4.1.3 Influence of OM quality on CO₂ and CH₄ production rates

845

850

855

840 The amount and the quality of OM usually determines the degradability and therefore the production of end products of anaerobic mineralization processes, CO₂ and CH₄.

Against our expectations and previous findings e.g. by Conrad et al. (2011) and Romeijn et al. (2019), C content was not correlated with CO_2 nor CH_4 production rates. As the production potential is obviously not depending on the amount of C, we suggest that instead the OM quality might be the determining factor for CO_2 and CH_4 production.

Typically, OM of autochthonous origin fuels higher degradation rates than allochthonous OM (West et al., 2012; Grasset et al., 2018) as the latter is rich in compounds like lignin, cellulose and humic acids, being recalcitrant to decomposition and therefore being effectively buried in lake sediments (Sobek et al., 2009). We however measured higher CO₂ and CH₄ production rates at sites close to the shore receiving higher inputs of allochthonous material compared to the center of the lake receiving mainly autochthonous OM as indicated by the C/N ratio. As a wider C/N ratio does not only indicate OM of terrestrial origin, but also implies that the OM is still less decomposed and therefore has a higher decomposition potential whereas a narrower C/N ratio is not only typical of aquatic OM but also indicates a higher degradation state, higher production rates at sites closer to the shore seem to be reasonable (Malmer and Holm, 1984; Meyers, 1994; Kuhry and Vitt, 1996; Gudasz et al., 2015; Grasset et al., 2018). Anyway, the correlation between production rates and C/N ratio was not significant, implying that the C/N ratio might, due to before explained contradictions, not be a suitable proxy for the mineralization potential in our case. Instead, all FTIR peak ratios were significantly negative

correlated with CO₂ and CH₄ production rates, supporting the hypothesis that OM mineralization is 860 highly dependent on the quality of the OM. To summarize, the quality of OM is not necessarily depending on its origin and that OM of terrestrial origin is not necessarily more recalcitrant to degradation by aquatic microorganisms. Sites closer to the shore probably receive higher rates of terrestrial OM matter being less decomposed, whereas autochthonous production in the lake center is overall low, due to the low nutrient status of the lake, and already undergoing degradation processes during sedimentation. This leads to more recalcitrant OM in the sediment. Earlier studies which found that allochthonous material is less decomposable compared to autochthonous matter were conducted in larger lakes, where the influence of the watershed is overall lower due to a lower perimeter to volume ratio. We hypothesize that in small and nutrient poor lakes the pattern might be reversed as the system adapts to overall low productivity and simultaneous high input of terrestrially produced OM.

4.1.4-2 Methanogenic pathways

865

870

875

To make methanogenesis a profitable reaction for microorganisms, theoretical thresholds of -10.6 kJ mol⁻¹ for the hydrogenotrophic <u>pathway</u> and -12.8 kJ mol⁻¹ for the acetoclastic pathway <u>have tomust</u> be exceeded (Hoehler et al., 2001). Except for a few samples at the beginning of the incubation <u>for the hydrogenotrophic pathway</u>, the theoretical thresholds were always exceeded, suggesting that both pathways <u>could potentially</u> contributed to CH₄ production <u>during the whole experiment</u>. <u>Still, this approach does not allow to evaluate which of the pathways actually predominated.</u>

During the incubation, Gibb's free energy increased for the hydrogenotrophic pathway, implying an increasing contribution of that pathway to total CH₄ production, whereas it was opposite for the 880 acetoclastic pathway. This suggests that labile organic polymers as precursors of acetate got depleted during the incubation so that H₂ gained of importance as substrate for methanogenesis. But when considering H₂ and acetate concentrations, the picture gets more complicated. An overall increase of H₂ concentrations was observed during incubation, but average H2 concentrations at t3 had a high standard deviation (see Fig. 5) because some samples showed a peak in H₂ concentrations at t3 instead of a 885 constant increase. These patterns could hint at an imbalance of the system, where fermenting, syntrophic and methanogenic bacteria were not yet equilibrated (Fey and Conrad, 2003). It is also remarkable, that the energy yield for acetoclastic methanogenesis decreased although acetate concentrations increased. We propose two different explanations for this finding: First, acetate is also used to reduce other EAs, which were present in higher concentrations at the beginning of the incubation, and therefore might have kept acetate concentrations low. Second, higher acetate concentrations do not necessarily mean more 890 favorable thermodynamic conditions due to end product accumulation. During degradation of carbohydrates, H₂ and acetate are being produced in a 2:1 ratio. We found acetate concentration exceeding H₂ concentrations by a factor of 10³, suggesting that homoacetogenesis contributed significantly to acetate production in our incubations (Conrad, 1999).

Against our expectations, Gibb's free energy of acetoclastic methanogenesis <u>at the end of the incubation</u> was significantly <u>lowest lower (i.e. higher energy yield)</u> in the center of the lake at t6 and was

additionally ΔG_r exhibited significant positive correlation positively correlated with C/N ratio. Considering that we found the lowest-most decomposed material (following peak ratios from FTIR analyses and low C/N ratios) OM quality-in the lake center and that low C/N ratios in this case indicate 900 OM of high recalcitrance whereas the acetoclastic pathway is attributed thought to predominate with high quality substrates, we would have expected a reverse pattern (Liu et al., 2017; Berberich et al., 2019). Concomitantly, Gibb's free energy of hydrogenotrophic methanogenesis exhibited significant positive correlations with some FTIR peak ratios indicative of higher contributions of recalcitrant OM compounds, although we expected that a high abundance of recalcitrant OM compounds would make hydrogenotrophic methanogenesis more feasible (Liu et al., 2017). Acetate and H₂ concentrations on 905 the other hand, both exhibited significant negative correlations with some FTIR peak ratios. While this seemed reasonable for acetate concentrations (less acetate available in strongly decomposed OM), this result again proved to be against our expectations in terms of H_2 concentrations. This suggests that more recalcitrant material in general slows fermentative processes, delivering both acetate and H_2 , as rate 910 determining step, and all fermentation products are thus kept at low levels.

Also, we found significant differences of energy yields between the sites, but could not relate them to OM quality. This implies that acetate as intermediate in the anaerobic OM mineralization cascade is not the rate determining step, but rather is fermentation and that OM quality only explains a part of the variance in CO₂ and CH₄ production patterns.

915 It further has to be mentioned, that correlations with CO₂ and CH₄-production rates were only observed for H_2 concentrations at the end of the incubation, but not with acetate. This finding emphasizes the hypothesis that the system was not in balance from the beginning of the incubation, and that fermentation seems to be the rate determining step for OM mineralization.

4.1.5-3 Alternative electron acceptorEAs

- 920 AThe average EAC_{OM} measured in our experiment (302.8 \pm 124.6 μ mol e⁻ gC⁻¹) was slightly lower but in the same order of magnitude compared to values found by Lau et al. (2015) and Gao et al. (2019) in other lake sediments or peat soils and other lake sediments, respectively. Given that C contents in the study by Gao et al. (2019) were much higher than in our study (46.2 - 49.4-%, Agethen and Knorr (2018)) and that Lau et al. (2015) fully oxidized their samples prior to analyses (C content 11.0 - 27.4925 %), our measured capacities seem reasonable.

930

We found that both organic and inorganic EAs decreasing decreased during the experiment, indicating that <u>all</u>EAs are being become similarly depleted with time as they are being reduced by microorganisms. Nevertheless, both absolute EAC_{OM} and EAC_{inorg} values as well as relative changes were rather low, which might have been caused by the one-week preincubation, where a majority of the reducible organic and inorganic compounds might have already been depleted. Klüpfel et al. (2014) showed that the EAC of oxidized humic acids strongly declined already after one day of anoxic incubation and did not change

much afterwards. Inorganic EAs were also shown to decrease exponentially during anoxic incubation

(Yao et al., 1999). The high CO_2 production rates (91.5 µmol gC⁻¹ d⁻¹) at the beginning of the slurry incubation are an indication of these processes.

- 935 Sites with higher EAC_{tot} had lower CH₄ production rates and vice versa, as <u>an</u> increased concentration of oxidized humic substances would suppress methanogenic activity <u>on longer time scales</u>, supporting the findings of Agethen et al. (2018) and Gao et al. (2019). Although the relationship between EAC and CH₄ production was <u>not significant</u> probably due to the low number of samples <u>and the sensitivity of the measuring method</u> not significant, a clear trend was observable.
- 940 <u>Non-methanogenic CO₂-production could by 38 to 91% be explained by prevalent organic and inorganic EAs, whereas organic EAs alone were able to explain up to 52% of measured CO₂-production in slurry incubations. Similar contributions were observed in previous studies from freshwater systems and peatlands, where up to 70 and 95% of non-methanogenic CO₂-production was explained by total EAC and 25 to 55% by decreases in EAC_{OM} alone (Lau et al., 2015; Agethen et al., 2018; Gao et al., 2019).</u>
- 945 The authors suggest several possible reasons for the gap between calculated and experimentally measured CO₂-production: Fermentation processes could explain excess CO₂, but in this case reduced equivalents of fermentation products, such as H₂ would be expected to accumulate. Moreover, decarboxylation processes that occur in oxygen-rich, labile OM might occur. Further, the assumption of a nominal oxidation state of C of zero as assumed in our approach might be wrong, so that when assuming a higher oxidation state, measured changes in EAC_{OM} might explain a higher share of CO₂-
- production (Gao et al., 2019). As we neither observed accumulation of fermentation products nor was the sediment under study strongly oxidized, we instead assume these processes are of minor importance and instead suggest that the proportion of unexplained CO₂-production can be explained by unknown consumed inorganic EAs during the slurry incubation. In this case, this was most likely solid phase iron which we found to occur 2-3% (≙ 53-137 mg Fe gC⁻¹) in our samples, but whereof we were not able to make statements about its speciation. We suppose this concentration is high enough to reach a closed
- budget of measured CO_2 -production and calculated EA turnover; if on average 7.8% (4-18.7%) of solid phase iron were present as ferric iron, this would suffice to close the CO_2 budget.
- The large range (4 to 52%) of the contribution of organic EAs to CO₂-production shows that there exists
 a distinct small-scale in-lake variability of terminal electron accepting processes and their contribution
 to CO₂ formation in the sediment of Lake Windsborn. Interestingly, the contribution of EAC_{OM} was
 highest at site 3.125, where we observed lowest CO₂-production rates and highest OM recalcitrance.
 Given that both the absolute C content in the incubation vials and the C:Fe ratio were highest at this site,
 this support findings of Lau et al. (2015), who stated that the contributions of EEC_{OM} to total EEC
 increase with the OC:Fe ratio, indicating more peat-like material.

Additionally, relationships were stronger with EAC_{tot} that with EAC_{inorg} or EAC_{org} only, suggesting that humic substances play a crucial, but variable role as terminal electron acceptors and should not be neglected when estimating the contribution of EAs to methanogenic activity in lake sediments rich in

OM (~30%) (Lau et al., 2016). The spatial variability of measured EAC and C content in the samples shows that substrate composition is critical when evaluating the contribution of different EAs and CH₄ production patterns.

4.2 Comparison of production rates, fluxes, and sediment parameters

970

Comparing the order of magnitude of potential production rates from slurry incubations and fluxes from intact sediment cores, results were in good accordance with each other. While CO₂ production rates 975 were overall something somewhat below measured fluxes, CH₄ production rates were considerably higher than <u>actual</u> CH₄ fluxes. The differences in CO₂ production rates and fluxes can be explained by the influence of the water column: CO_2 is not only produced in the sediment, but also in the water column, e.g. by the degradation of OM or zooplankton respiration (Kling et al., 1992) and can therefore contribute to water-atmosphere fluxes. The differences concerning CH₄ could be explained by several 980 approachesfactors. First, preparing slurry incubations leads to the homogenization of the sediment and higher surface area, and therefore, an even distribution of substrates and microorganisms, which is then bettersuch that the substrates are more readily available for the latter to the microorganisms (Hoehler et al., 2001). Secondly, sediment fluxes from intact sediment cores may be low compared to homogenized slurry incubations, as maximum sediment concentrations there were already reached in shallow 985 depthswe did not observe significant increases in the depth profile of sediment cores, and suggesting that strong changes in sediment gas concentrations which are driving that drive fluxes are likely to occur only in the upper mm millimeters to emcentimeters. Additionally, the preparation of incubations slurries reduces the thermodynamic constraints which that usually exist in intact profiles due to end product accumulation (Blodau et al., 2011; Bonaiuti et al., 2017). Thirdly, not all CH₄ 990 produced in and emitted by the sediment will reach the water-atmosphere interface, because some of the CH4 will be consumed in the oxygenated water column on the way to the surface: of consumption processes in the oxygenated water column that has to be passed. Through through CH_4 -its oxidation in the water column, the amounts of CH₄ emitted from lakes to the atmosphere can be substantially reduced by 51 to 100-% (Bastviken et al., 2002; Bastviken et al., 2008).

Interestingly, spatial patterns of CO₂ and CH₄ emissions could not be well reproduced from sediment slurry incubations. Although highest CO₂ emissions were observed at site S.150, we found lowest CO₂ production rates there. Additionally, we did not see any correlations between sediment OM quality and CO₂ fluxes, suggesting that CO₂ production processes in the sediment overlying water column play a much more important role than in the sediment itself. Moreover, CH₄ production rates at sites 1.50 and 3.125 differed significantly from each other, while this was not the case when comparing fluxes. We assume that the different patterns of CH₄ fluxes and production can be explained by grain size distribution and, thus, physical sediment properties. Sediment grain size determines sediment pore size which is essential for the evolution of methane <u>CH₄</u> bubbles in the sediment (Boudreau et al., 2005). Ebullition is the major component of CH₄ fluxes in shallow waters, as it can efficiently bypass oxidation

- 1005 in the water column (Bastviken et al., 2004; Wik et al., 2013; Natchimuthu et al., 2016). We found that ebullition supporting significantly to total was responsible for most of the CH₄ fluxes in two of our four intact sediment mesocosms core incubations, whereas sites with higher shares of sand exhibited less ebullitive fluxes, confirming the findings of Liu et al. (2016) and (2018). The authors justify their findings with the dominant pathway of bubble formation in the sediment, which is by displacing 1010 surrounding sediment particles. As this mechanism is more efficient in soft silty sediments compared to sandy material, CH₄ bubbles likely accumulate more easily in silt, creating a network of macropores and therefore conduits for subsequent bubble release. We further found that OM quality being-partly exhibited significant negative correlations with significantly correlated to actual CH₄ fluxes, but to a lesser extent than to with sediment CH₄ production. As said, when preparing slurry incubations, the 1015 physical sediment structure is destroyed, so that OM quality likely becomes the major controlling factor for GHG production. These findings suggest that grain size distribution is besides OM quality, grain size distribution is a main driver of spatial CH₄ flux patterns in intact sediment core incubationsmesocosms and that only a combination of the sediment's physical characteristics and its
- On the other hand, when preparing incubations, the physical sediment structure is destroyed, so that OM 1020 quality becomes the major controlling factor for gas production. Although we also found OM quality being significantly correlated to mesocosm fluxes, we suppose that the combination of physical characteristics and sediment OM quality can sufficiently explain CH₄ emission patterns from lakes.

OM quality could sufficiently explain CH₄ emission patterns from lakes.

The missing link between sediment gas storage and both diffusive and ebullitive emissions could can 1025 potentially be explained by our sampling procedure. While fluxes were partly dominated by bubbles, these bubbles cannot be captured in the sediment by silicon samplers. Silicon samplers rely on the slow diffusion of gas between the pore gas and the sampler-, whereas Bbubbles however may form on shorter time-scales and could can quickly being released to the water-column. Thus, concentrations in the bulk sediment stay similar consistent while the released bubbles cause high fluxes. Moreover, the resolution 1030 of our gas samplers was probably not high enough, as they could not covernot covering the uppermost centimeters of the sediment, which we assume to be crucial for diffusive fluxes.

5 Conclusion

1035

Our <u>experiment results</u> showed that there exists a significant spatial variability of CO_2 and CH_4 production in the sediment of a small and shallow lake and, therefore, that it is therefore not possible to upscale sediment production rates from single point measurements. We further proved that especially CH₄ production is strongly depending depends on temperature, and that the extent of temperature dependence might differ within one a lake, especially between littoral and central parts due to different differences in OM quality. Small lakes seem to be very vulnerable sensitive to temperature increases in terms of accelerated C cycling, which implies they might become larger sources of CO₂ and particularly of CH₄ emissions under climate change scenarios. Still, theyAlthough such lakes might not react 1040

homogeneously, but we could would unroll expect that sediments in shallower parts might react more sensitively due to lower water columnsshallow water depths and input of reactive allochthonous OM, which is particularly important for oligotrophic lakes without significant contribution of autochthonous primary production. The strong negative correlations we found between recalcitrant OM compounds 1045 and CO_2 and CH_4 production rates <u>proved provide</u> evidence that OM quality plays a major role in controlling the mineralization of anoxic lake sediment and can be used to explain and predict withinlake patterns of production rates. Both hydrogenotrophic and acetoclastic methanogenesis were feasible during <u>slurry</u> incubations, but we could neither not clearly refer attribute observed patterns to OM quality nor electron exchange capacities <u>EEC</u>, and therefore, we suggest that both H₂ and acetate were not 1050 limited as substrates but that fermentation was the rate-determining step of OM mineralization and subsequent consumption of acetate and H₂ by electron accepting processes or methanogenesis kept the levels of these fermentation products low as expected. We did not find clear patterns between EAC and CH₄ production, but organic and inorganic EAs could explain up to 91% of the observed nonmethanogenic CO₂-production in slurry incubations. More than organic or inorganic EAs alone, EAC_{tot} 1055 could explain observed variabilities in CH4 production, implying that also organic EAs play a major role in constantly anoxic lake sediments with high C contents.

HoweverIn sum, measured production rates were only partly reflected in CO₂ and CH₄ flux patterns obtained from whole-intact sediment core incubations. We suggest suppose that this is was because the physical sediment structure (e.g. pore size) were destroyed in the slurry incubations due to the destruction of physical sediment parameters in the incubations (e.g. pore size), but these parameters are crucial for the evolution of CH₄ bubbles in the sediment, which are crucial for the evolution of CH₄ bubbles in the sediment, which are crucial for the evolution of CH₄ bubbles in the sediment in the slurry incubations, and due to the lack of thermodynamic and gas transport constraints that exist in intact anoxic sediment profiles were removed. Further, measuring the interpretation of production rates only would neglects the importance of the water column as a sink of sediment generated CH₄ through oxidation and source of CO₂ through degradation and respiration processes which might then cause discrepancies between observed production and flux rates.

So far, direct flux measurements of CO_2 and CH_4 between water and the atmosphere seem to be the most accurate way for determining to determine the magnitude and variability of emissions from lakes, but still our study contributes substantially to the understanding of controls on-underlying potential CO_2 and CH_4 production processes in lake sediments and sediment-inherent factors determining emission.

6 Data availability

The underlying research data of this study can be found in the Supplement.

1075 7 Author contribution

LP, MS, SS, NP and KHK designed the study. NP and SS carried out the experiments and sample analyses with the help of LP and MS. LP performed data analyses and wrote the manuscript with the help of NP, SS, KHK and GB.

8 Competing interests

1080 The authors declare that they have no conflict of interest.

9 Acknowledgements

We thank the technicians of the laboratory of the institute of landscape ecology Ulrike Berning-Mader,Madeleine Supper, Viktoria Ratachin and Melanie Tappe for sample measurements. We thank TanjaBroder for ICP-OES and Stephan Glatzel for grain size analyses. We further thank Rebecca Pabst,1085Christina Hackmann, Fabian Lange, Isabelle Rieke, Friederike Ding, Victoria Tietz and AnjaRadermacher for their assistance during laboratory work-and Stephan Glatzel for grain size analyses.We also thank Celeste Brennecka for her improvements on English language and style of themanuscript. This project was funded by the German Research Foundation DFG (BL 563/25-1). Weacknowledge support from the Open Access Publication Fund of the University of MuensterMünster.

10 References

1115

- Aben, R. C. H., Barros, N., van Donk, E., Frenken, T., Hilt, S., Kazanjian, G., Lamers, L. P. M., Peeters, E. T. H. M., Roelofs, J. G. M., Senerpont Domis, L. N. de, Stephan, S., Velthuis, M., van de Waal, D. B., Wik, M., Thornton, B. F., Wilkinson, J., DelSontro, T., and Kosten, S.: Cross continental increase in methane ebullition under climate change, Nature communications, 8, 1682, https://doi.org/10.1038/s41467-017-01535-y, 2017.
 - Achtnich, C., Bak, F., and Conrad, R.: Competition for electron donors among nitrate reducers, ferric iron reducers, sulfate reducers, and methanogens in anoxic paddy soil, Biology and Fertility of Soils, 19, 65-72, https://doi.org/10.1007/BF00336349, 1995.
- 1100 Aeschbacher, M., Vergari, D., Schwarzenbach, R. P., and Sander, M.: Electrochemical analysis of proton and electron transfer equilibria of the reducible moieties in humic acids, Environ. Sci. Technol., 45, 8385–8394, https://doi.org/10.1021/es201981g, 2011.
- Aeschbacher, M., Sander, M., and Schwarzenbach, R. P.: Novel electrochemical approach to assess the redox properties of humic substances, Environmental science & technology, 44, 87–93, https://doi.org/10.1021/es902627p, 2010.
 - Agethen, S. and Knorr, K.-H.: Juncus effusus mono-stands in restored cutover peat bogs Analysis of litter quality, controls of anaerobic decomposition, and the risk of secondary carbon loss, Soil Biology and Biochemistry, 117, 139–152, https://doi.org/10.1016/j.soilbio.2017.11.020, 2018.
- Agethen, S., Sander, M., Waldemer, C., and Knorr, K.-H.: Plant rhizosphere oxidation reduces methane production and emission in rewetted peatlands, Soil Biology and Biochemistry, 125, 125–135, https://doi.org/10.1016/j.soilbio.2018.07.006, 2018.
 - Artz, R. R.E., Chapman, S. J., Jean Robertson, A. H., Potts, J. M., Laggoun-Défarge, F., Gogo, S., Comont, L., Disnar, J.-R., and Francez, A.-J.: FTIR spectroscopy can be used as a screening tool for organic matter quality in regenerating cutover peatlands, Soil Biology and Biochemistry, 40, 515– 527, https://doi.org/10.1016/j.soilbio.2007.09.019, 2008.
- Avnimelech, Y., McHenry, J. R., and Ross, J. D.: Decomposition of organic matter in lake sediments, Environmental science & technology, 18, 5–11, https://doi.org/10.1021/es00119a004, 1984.

Bastviken, D., Sundgren, I., Natchimuthu, S., Reyier, H., and Gålfalk, M.: Technical Note: Costefficient approaches to measure carbon dioxide (CO<sub>2</sub>2) fluxes and

- 1120 concentrations in terrestrial and aquatic environments using mini loggers, Biogeosciences, 12, 3849–3859, https://doi.org/10.5194/bg-12-3849-2015, 2015.
 - Bastviken, D., Tranvik, L. J., Downing, J. A., Crill, P. M., and Enrich-Prast, A.: Freshwater methane emissions offset the continental carbon sink, Science (New York, N.Y.), 331, 50, https://doi.org/10.1126/science.1196808, 2011.
- 1125 Bastviken, D., Cole, J. J., Pace, M. L., and van de Bogert, M. C.: Fates of methane from different lake habitats: Connecting whole-lake budgets and CH 4 emissions, J. Geophys. Res. Biogeosci., 113, n/a-n/a, https://doi.org/10.1029/2007JG000608, 2008.

Bastviken, D., Cole, J., Pace, M., and Tranvik, L.: Methane emissions from lakes: Dependence of lake characteristics, two regional assessments, and a global estimate, Global Biogeochem. Cycles, 18, n/a-n/a, https://doi.org/10.1029/2004GB002238, 2004.

Bastviken, D., Ejlertsson, J., and Tranvik, L.: Measurement of <u>Mm</u>ethane <u>Oxidation_oxidation</u> in <u>Lakeslakes</u>: A <u>Comparison comparison</u> of <u>Methodsmethods</u>, Environ. Sci. Technol., 36, 3354–3361, https://doi.org/10.1021/es010311p, 2002.

1130

1145

- Battin, T. J., Luyssaert, S., Kaplan, L. A., Aufdenkampe, A. K., Richter, A., and Tranvik, L. J.: The boundless carbon cycle, Nature Geosci, 2, 598–600, https://doi.org/10.1038/ngeo618, 2009.
 - Beer, J. and Blodau, C.: Transport and thermodynamics constrain belowground carbon turnover in a northern peatland, Geochimica et Cosmochimica Acta, 71, 2989–3002, https://doi.org/10.1016/j.gca.2007.03.010, 2007.
- Berberich, M., Beaulieu, J. J., Hamilton, T. L., Waldo, S., and Buffam, I.: Spatial variability of sediment
 methane production and methanogen communities within a eutrophic reservoir: Importance of organic matter source and quantity, Limnology and Oceanography, 65, 1336-1358, https://doi.org/10.1002/lno.11392, 2019.
 - Bergström, I., Kortelainen, P., Sarvala, J., and Salonen, K.: Effects of temperature and sediment properties on benthic CO-2 production in an oligotrophic boreal lake, Freshwater Biology, 39, 1794, https://doi.org/10.1111/j.1365-2427.2010.02408.x, 2010.
 - Biester, H., Knorr, K.-H., Schellekens, J., Basler, A., and Hermanns, Y.-M.: Comparison of different methods to determine the degree of peat decomposition in peat bogs, Biogeosciences, 11, 2691– 2707, https://doi.org/10.5194/bg-11-2691-2014, 2014.
 - Blodau, C.: Thermodynamic Control control on Terminal terminal Electron electron Transfer transfer
- 1150 and <u>Methanogenesismethanogenesis</u>, in: Aquatic Redox Chemistry, edited by: Tratnyek, P. G., Grundl, T. J., and Haderlein, S. B., American Chemical Society, Washington, DC, 65–83, https://doi.org/10.1021/bk-2011-1071.ch004, 2011.
 - Blodau, C., Siems, M., and Beer, J.: Experimental burial inhibits methanogenesis and anaerobic decomposition in water-saturated peats, Environmental science & technology, 45, 9984–9989, https://doi.org/10.1021/es201777u, 2011.
 - Bonaiuti, S., Blodau, C., and Knorr, K.-H.: Transport, anoxia and end-product accumulation control carbon dioxide and methane production and release in peat soils, Biogeochemistry, 133, 219–239, https://doi.org/10.1007/s10533-017-0328-7, 2017.
 - Boudreau, B. P., Algar, C., Johnson, B. D., Croudace, I., Reed, A., Furukawa, Y., Dorgan, K. M.,
- Jumars, P. A., Grader, A. S., and Gardiner, B. S.: Bubble growth and rise in soft sediments, Geol, 33, 517, https://doi.org/10.1130/G21259.1, 2005.
 - Broder, T., Blodau, C., Biester, H., and Knorr, K. H.: Peat decomposition records in three pristine ombrotrophic bogs in southern Patagonia, Biogeosciences, 9, 1479–1491, https://doi.org/10.5194/bg-9-1479-2012, 2012.

- 1165 Burdige, D. J.: Preservation of organic matter in marine sediments: Controls, mechanisms, and an imbalance in sediment organic carbon budgets?, Chemical reviews, 107, 467–485, https://doi.org/10.1021/cr050347q, 2007.
 - Cocozza, C., D'Orazio, V., Miano, T. M., and Shotyk, W.: Characterization of solid and aqueous phases of a peat bog profile using molecular fluorescence spectroscopy, ESR and FT-IR, and comparison
- 1170 with physical properties, Organic Geochemistry, 34, 49–60, https://doi.org/10.1016/S0146-6380(02)00208-5, 2003.
- Cole, J. J., Prairie, Y. T., Caraco, N. F., McDowell, W. H., Tranvik, L. J., Striegl, R. G., Duarte, C. M., Kortelainen, P., Downing, J. A., Middelburg, J. J., and Melack, J.: Plumbing the Global-global Carbon-carbon Cyclecycle: Integrating Inland-inland Waters-waters into the Terrestrial-terrestrial
 Carbon-carbon Budgetbudget, Ecosystems, 10, 172–185, https://doi.org/10.1007/s10021-006-9013-8, 2007.
 - Conrad, R.: Contribution of hydrogen to methane production and control of hydrogen concentrations in methanogenic soils and sediments, FEMS Microbiology Ecology, 28, 193–202, https://doi.org/10.1016/S0168-6496(98)00086-5, 1999.
- 1180 Conrad, R., Noll, M., Claus, P., Klose, M., Bastos, W. R., and Enrich-Prast, A.: Stable carbon isotope discrimination and microbiology of methane formation in tropical anoxic lake sediments, Biogeosciences, 8, 795–814, https://doi.org/10.5194/bg-8-795-2011, 2011.
 - den Heyer, C. and Kalff, J.: Organic matter mineralization rates in sediments: A within- and amonglake study, Limnol. Oceanogr., 43, 695–705, https://doi.org/10.4319/lo.1998.43.4.0695, 1998.
- 1185 Deutscher Wetterdienst (DWD): Observations Germany Climate Multi-annual mean 81-10, ftp://ftpcdc.dwd.de/pub/CDC/observations_germany/climate/multi_annual/mean_81-10/, last access: 21 May 2019, 2019.
 - Dinno, A.: dunn.test: Dunn's Test test of Multiple multiple Comparisons comparisons Using using Rank rank Sumssums, 2017.
- Downing, J. A.: Emerging global role of small lakes and ponds: little things mean a lot, Limnetica, 29, 9–23, 2010.
 - Downing, J. A., Prairie, Y. T., Cole, J. J., Duarte, C. M., Tranvik, L. J., Striegl, R. G., McDowell, W. H., Kortelainen, P., Caraco, N. F., Melack, J. M., and Middelburg, J. J.: The global abundance and size distribution of lakes, ponds, and impoundments, Limnol. Oceanogr., 51, 2388–2397,
- 1195 https://doi.org/10.4319/lo.2006.51.5.2388, 2006.
 - Duc, N. T., Crill, P., and Bastviken, D.: Implications of temperature and sediment characteristics on methane formation and oxidation in lake sediments, Biogeochemistry, 100, 185–196, https://doi.org/10.1007/s10533-010-9415-8, 2010.
- Ellenberg, H., Weber, H. E., Düll, R., Wirth, V., and Werner, W.: Zeigerwerte von Pflanzen in Mitteleuropa, 3., durchgesehene Auflage, Scripta geobotanica, Volume 18, Verlag Erich Goltze GmbH & Co KG, Göttingen, 262 pp., 2001.

- Falz, K. Z., Holliger, C., Großkopf, R., Liesack, W., Nozhevnikova, A. N., Müller, B., Wehrli, B., and Hahn, D.: Vertical <u>Distribution_distribution</u> of <u>Methanogens_methanogens</u> in the <u>Anoxic_anoxic</u> <u>Sediment_sediment</u> of Rotsee (Switzerland), Applied and Environmental Microbiology, 65, 2402– 2408, 1999.
- 1205
- Fenchel, T., King, G. M., and Blackburn, T. H.: Bacterial biogeochemistry: The ecophysiology of mineral cycling, 3rd ed., Academic Press/Elsevier, Boston, Mass, 2012.
- Fey, A. and Conrad, R.: Effect of temperature on the rate limiting step in the methanogenic degradation pathway in rice field soil, Soil Biology and Biochemistry, 35, 1–8, https://doi.org/10.1016/S0038-0515/00000155 M 2002
- 1210 0717(02)00175-X, 2003.
 - Fox, J. and Weisberg, S.: An {R} Companion to <u>Applied applied Regression regression</u>, Sage, Thousand Oaks, CA, 2011.
 - Fuchs, A., Lyautey, E., Montuelle, B., and Casper, P.: Effects of increasing temperatures on methane concentrations and methanogenesis during experimental incubation of sediments from oligotrophic
- 1215 and mesotrophic lakes, J. Geophys. Res. Biogeosci., 121, 1394–1406, https://doi.org/10.1002/2016JG003328, 2016.
 - Gao, C., Sander, M., Agethen, S., and Knorr, K.-H.: Electron accepting capacity of dissolved and particulate organic matter control CO2 and CH4 formation in peat soils, Geochimica et Cosmochimica Acta, 245, 266–277, https://doi.org/10.1016/j.gca.2018.11.004, 2019.
- 1220 Gilboa-Garber, N.: Direct spectrophotometric determination of inorganic sulfide in biological materials and in other complex mixtures, Analytical Biochemistry, 43, 129–133, https://doi.org/10.1016/0003-2697(71)90116-3, 1971.
 - Grasset, C., Mendonça, R., Villamor Saucedo, G., Bastviken, D., Roland, F., and Sobek, S.: Large but variable methane production in anoxic freshwater sediment upon addition of allochthonous and
- 1225 autochthonous organic matter, Limnol. Oceanogr., 63, 1488–1501, https://doi.org/10.1002/lno.10786, 2018.
 - Gudasz, C., Sobek, S., Bastviken, D., Koehler, B., and Tranvik, L. J.: Temperature sensitivity of organic carbon mineralization in contrasting lake sediments, J. Geophys. Res. Biogeosci., 120, 1215–1225, https://doi.org/10.1002/2015JG002928, 2015.
- 1230 Gudasz, C., Bastviken, D., Steger, K., Premke, K., Sobek, S., and Tranvik, L. J.: Temperature-controlled organic carbon mineralization in lake sediments, Nature, 466, 478–481, https://doi.org/10.1038/nature09186, 2010.
 - Hoehler, T. M., Alperin, M. J., Albert, D. B., and Martens, C. S.: Apparent minimum free energy requirements for methanogenic Archaea and sulfate-reducing bacteria in an anoxic marine sediment,
- 1235 FEMS Microbiology Ecology, 38, 33–41, https://doi.org/10.1111/j.1574-6941.2001.tb00879.x, 2001.
 - Holgerson, M. A. and Raymond, P. A.: Large contribution to inland water CO2 and CH4 emissions from very small ponds, Nature Geosci, 9, 222–226, https://doi.org/10.1038/NGEO2654, 2016.

Hornibrook, E. R.C., Longstaffe, F. J., and Fyfe, W. S.: Spatial distribution of microbial methane
 production pathways in temperate zone wetland soils: Stable carbon and hydrogen isotope evidence,
 Geochimica et Cosmochimica Acta, 61, 745–753, https://doi.org/10.1016/S0016-7037(96)00368-7,
 1997.

- Jankowski, T., Livingstone, D. M., Bührer, H., Forster, R., and Niederhauser, P.: Consequences of the 2003 European heat wave for lake temperature profiles, thermal stability, and hypolimnetic oxygen
- 1245 depletion: Implications for a warmer world, Limnol. Oceanogr., 51, 815–819, https://doi.org/10.4319/lo.2006.51.2.0815, 2006.
 - Kammann, C., Grünhage, L., and Jäger, H.-J.: A new sampling technique to monitor concentrations of CH 4 N 2 O and CO 2 in air at well-defined depths in soils with varied water potential, European Journal of Soil Science, 52, 297–303, https://doi.org/10.1046/j.1365-2389.2001.00380.x, 2001.
- 1250 Kampf, J.: Grundwasserlandschaften: Hydrologischer Atlas Rheinland-Pfalz, Landesamt für Umwelt, Wasserwirtschaft und Gewerbeaufsicht Rheinland-Pfalz, Mainz, 2005.
 - Kappes, H. and Sinsch, U.: Tolerance of Ceriodaphnia quadrangula and Diaphanosoma brachyurum (Crustacea: Cladocera) to experimental soft water acidification, Hydrobiologia, 109–115, 2005.
- Kappes, H., Mechenich, C., and Sinsch, U.: Long-term dynamics of Asplanchna priodonta in Lake
 Windsborn with comments on the diet, Hydrobiologia, 432, 91–100, https://doi.org/10.1023/A:1004022020346, 2000.
 - Kiuru, P., Ojala, A., Mammarella, I., Heiskanen, J., Kämäräinen, M., Vesala, T., and Huttula, T.: Effects of Climate <u>climate</u> <u>Change change</u> on CO 2 Concentration <u>concentration</u> and <u>Efflux efflux</u> in a <u>Humic <u>humic</u> Boreal <u>boreal</u> <u>Lakelake</u>: A <u>Modeling modeling</u> <u>Studystudy</u>, J. Geophys. Res. <u>Biogeosci., 123, 2212–2233, https://doi.org/10.1029/2018JG004585, 2018.</u>
 </u>

1260

1265

- Kling, G. W., Kipphut, G. W., and Miller, M. C.: The flux of CO2 and CH4 from lakes and rivers in arctic Alaska, Hydrobiologia, 240, 23–36, https://doi.org/10.1007/BF00013449, 1992.
- Klüpfel, L., Piepenbrock, A., Kappler, A., and Sander, M.: Humic substances as fully regenerable electron acceptors in recurrently anoxic environments, Nature Geosci, 7, 195–200, https://doi.org/10.1038/NGEO2084, 2014.

Konhauser, K.: Introduction to geomicrobiology, John Wiley & Sons Ltd, Hoboken, 425 pp., 2009.

Kuhry, P. and Vitt, D. H.: Fossil <u>c</u>Carbon/<u>Nitrogen nitrogen Ratios-ratios</u> as a <u>Measure-measure</u> of <u>Peat</u> <u>peat</u> <u>Decomposition</u> <u>decomposition</u>, Ecology, 77, 271–275, https://doi.org/10.2307/2265676, 1996.

Lau, M. P., Sander, M., Gelbrecht, J., and Hupfer, M.: Spatiotemporal redox dynamics in a freshwater lake sediment under alternating oxygen availabilities: Combined analyses of dissolved and particulate electron accentors. Environ Chem. 12, 826 https://doi.org/10.1071/EN15217.2016

1275 particulate electron acceptors, Environ. Chem., 13, 826, https://doi.org/10.1071/EN15217, 2016.

Landesamt für Umwelt (LfU): Seenatlas: Windsborn Kratersee, https://geodaten-wasser.rlp-1270 umwelt.de/prj-

wwvauskunft/projects/seenatlas/daten/register1.jsp?seeId=125&see_bez=Windsborn+Kratersee&s howHydrologie=true&showChemismus=true, last access 17 July 2019, 2013.

- Lau, M. P., Sander, M., Gelbrecht, J., and Hupfer, M.: Solid phases as important electron acceptors in freshwater organic sediments, Biogeochemistry, 123, 49–61, https://doi.org/10.1007/s10533-014-0052-5, 2015.
- Li, Y., Zhang, H., Tu, C., Fu, C., Xue, Y., and Luo, Y.: Sources and fate of organic carbon and nitrogen from land to ocean: Identified by coupling stable isotopes with C/N ratio, Estuarine, Coastal and Shelf Science, 181, 114–122, https://doi.org/10.1016/j.ecss.2016.08.024, 2016.

1280

1285

- Liikanen, A., Murtoniemi, T., Tanskanen, H., Väisänen, T., and Martikainen, P. J.: Effects of temperature and oxygen availability on greenhouse gas and nutrient dynamics in sediment of a eutrophic and mid-boreal lake, Biogeochemistry, 59, 269–286, https://doi.org/10.1023/A:1016015526712, 2002.
- Liu, L., Wilkinson, J., Koca, K., Buchmann, C., and Lorke, A.: The role of sediment structure in gas bubble storage and release, J. Geophys. Res. Biogeosci., 121, 1992–2005, https://doi.org/10.1002/2016JG003456, 2016.
- Liu, L., Kock, T. de, Wilkinson, J., Cnudde, V., Xiao, S., Buchmann, C., Uteau, D., Peth, S., and Lorke,
- A.: Methane <u>b</u>Bubble G growth and <u>Mm</u>igration in <u>Aa</u>quatic <u>Ss</u>ediments <u>Oo</u>bserved by X-ray μCT,
 Environ. Sci. Technol., 52, 2007–2015, https://doi.org/10.1021/acs.est.7b06061, 2018.
 - Liu, Y., Conrad, R., Yao, T., Gleixner, G., and Claus, P.: Change of methane production pathway with sediment depth in a lake on the Tibetan plateau, Palaeogeography, Palaeoclimatology, Palaeoecology, 474, 279-286, https://doi.org/10.1016/j.palaeo.2016.06.021, 2017.
- 1295 Lojen, S., Ogrinc, N., and Dolenec, T.: Decomposition of sedimentary organic matter and methane formation in the recent sediment of Lake Bled (Slovenia), Chemical Geology, 159, 223–240, https://doi.org/10.1016/S0009-2541(99)00032-7, 1999.
 - Lovley, D. R., Coates, J. D., Blunt-Harris, E. L., Phillips, E. J. P., and Woodward, J. C.: Humic substances as electron acceptors for microbial respiration, Nature, 382, 445–448, https://doi.org/10.1038/382445a0, 1996.
 - Mackay, E. B., Jones, I. D., Folkard, A. M., and Barker, P.: Contribution of sediment focussing to heterogeneity of organic carbon and phosphorus burial in small lakes, Freshwater Biology, 57, 290-304, https://doi.org/10.1111/j.1365-2427.2011.02616.x, 2011.
 - Malmer, N. and Holm, E.: Variation in the C/N-Quotient quotient of Peat-peat in Relation relation to
- 1305Decomposition decomposition Rate rate and aAge Determination determination with 210 Pb, Oikos,
43, 171, https://doi.org/10.2307/3544766, 1984.
 - McLatchey, G. P. and Reddy, K. R.: Regulation of Organic organic Matter matter Decomposition decomposition and Nutrient nutrient Release release in a wWetland sSoil, Journal of Environment Quality, 27, 1268, https://doi.org/10.2134/jeq1998.00472425002700050036x, 1998.
- 1310 Megonigal, J. P., Hines, M. E., and Visscher, P. T.: Anaerobic <u>Metabolismmetabolism</u>: Linkages to <u>Trace-trace Gases-gases</u> and <u>Aerobic-aerobic Processesprocesses</u>, in: Treatise on Geochemistry, Elsevier, 317–424, https://doi.org/10.1016/B0-08-043751-6/08132-9, 2003.

Meyer, W.: Geologie der Eifel, 4th ed., Schweizerbart'sche Verlagsbuchhandlung, Stuttgart, 2013.

- Meyers, P. A.: Preservation of elemental and isotopic source identification of sedimentary organic matter, Chemical Geology, 114, 289–302, https://doi.org/10.1016/0009-2541(94)90059-0, 1994.
- Miyajima, T., Wada, E., Hanba, Y. T., and Vijarnsorn, P.: Anaerobic mineralization of indigenous organic matters and methanogenesis in tropical wetland soils, Geochimica et Cosmochimica Acta, 61, 3739–3751, https://doi.org/10.1016/S0016-7037(97)00189-0, 1997.

1315

- Muri, G. and Wakeham, S. G.: Organic matter and lipids in sediments of Lake Bled (NW Slovenia):
- Source and effect of anoxic and oxic depositional regimes, Organic Geochemistry, 37, 1664–1679, https://doi.org/10.1016/j.orggeochem.2006.07.016, 2006.
 - Natchimuthu, S., Sundgren, I., Gålfalk, M., Klemedtsson, L., and Bastviken, D.: Spatiotemporal variability of lake pCO-2 and CO-2 fluxes in a hemiboreal catchment, J. Geophys. Res. Biogeosci., 122, 30–49, https://doi.org/10.1002/2016JG003449, 2017.
- 1325 Natchimuthu, S., Sundgren, I., Gålfalk, M., Klemedtsson, L., Crill, P., Danielsson, Å., and Bastviken,
 D.: Spatio-temporal variability of lake CH-4 fluxes and its influence on annual whole lake emission estimates, Limnol. Oceanogr., 61, S13-S26, https://doi.org/10.1002/lno.10222, 2016.
- Niemeyer, J., Chen, Y., and Bollag, J.-M.: Characterization of <u>Humic humic a</u>Acids, <u>Compostscomposts</u>, and <u>Peat peat by d</u>Diffuse <u>Rr</u>eflectance Fourier-Transform <u>i</u>Infrared
 <u>Sspectroscopy</u>, Soil Science Society of America Journal, 56, 135, https://doi.org/10.2136/sssaj1992.03615995005600010021x, 1992.
 - Nordstrom, D. K., and Munoz, J. L.: Geochemical thermodynamics. 2nd ed., Blackwell Scientific, Boston, Mass, 1994.
 - OENorm B 4412: 1974 07 01, Erd- und Grundbau; Untersuchung von Bodenproben; Korngrößenverteilung, 1974.
 - OENorm L 1050: 2016 03 15, Boden als Pflanzenstandort Begriffe und Untersuchungsverfahren, 2016. OENorm L 1061-2: 2019 03 01, Physikalische Bodenuntersuchungen - Bestimmung der
 - Korngrößenverteilung des Mineralbodens in land- und forstwirtschaftlich genutzten Böden Teil 2: Feinboden, 2019.
- 1340 Ostrovsky, I. and Tęgowski, J.: Hydroacoustic analysis of spatial and temporal variability of bottom sediment characteristics in Lake Kinneret in relation to water level fluctuation, Geo-Mar Lett, 30, 261–269, https://doi.org/10.1007/s00367-009-0180-4, 2010.
 - R Core Team: R: A language and environment for statistical computing, R Foundation for Statistical Computing, Vienna, Austria, 2018.
- Raymond, P. A., Hartmann, J., Lauerwald, R., Sobek, S., McDonald, C., Hoover, M., Butman, D.,
 Striegl, R., Mayorga, E., Humborg, C., Kortelainen, P., Dürr, H., Meybeck, M., Ciais, P., and Guth,
 P.: Global carbon dioxide emissions from inland waters, Nature, 503, 355–359,
 https://doi.org/10.1038/nature12760, 2013.
Regnier, P., Friedlingstein, P., Ciais, P., Mackenzie, F. T., Gruber, N., Janssens, I. A., Laruelle, G. G.,
Lauerwald, R., Luyssaert, S., Andersson, A. J., Arndt, S., Arnosti, C., Borges, A. V., Dale, A. W.,
Gallego-Sala, A., Goddéris, Y., Goossens, N., Hartmann, J., Heinze, C., Ilyina, T., Joos, F., LaRowe,
D. E., Leifeld, J., Meysman, F. J. R., Munhoven, G., Raymond, P. A., Spahni, R., Suntharalingam,
P., and Thullner, M.: Anthropogenic perturbation of the carbon fluxes from land to ocean, Nature
Geosci, 6, 597–607, https://doi.org/10.1038/NGEO1830, 2013.

- 1355 Romeijn, P., Comer-Warner, S. A., Ullah, S., Hannah, D. M., and Krause, S.: Streambed Ogrganic Matter-matter Ccontrols on Ccarbon Dioxide-dioxide and mMethane Eemissions from Sstreams, Environ. Sci. Technol., 53, 2364–2374, https://doi.org/10.1021/acs.est.8b04243, 2019.
 - Sander, R.: Compilation of Henry's law constants (version 4.0) for water as solvent, Atmos. Chem. Phys., 15, 4399–4981, https://doi.org/10.5194/acp-15-4399-2015, 2015.
- 1360 Schilder, J., Bastviken, D., van Hardenbroek, M., Kankaala, P., Rinta, P., Stötter, T., and Heiri, O.: Spatial heterogeneity and lake morphology affect diffusive greenhouse gas emission estimates of lakes, Geophys. Res. Lett., 40, 5752–5756, https://doi.org/10.1002/2013GL057669, 2013.
 - Schink, B.: Energetics of syntrophic cooperation in methanogenic degradation, Microbiology and molecular biology reviews MMBR, 61, 262–280, 1997.
- 1365 Schoell, M.: Multiple origins of methane in the Earth, Chemical Geology, 71, 1–10, https://doi.org/10.1016/0009-2541(88)90101-5, 1988.
 - Schwarz, J. I. K., Eckert, W., and Conrad, R.: Response of the methanogenic microbial community of a profundal lake sediment (Lake Kinneret, Israel) to algal deposition, Limnol. Oceanogr., 53, 113– 121, https://doi.org/10.4319/lo.2008.53.1.0113, 2008.
- 1370 Seeberg-Elverfeldt, J., Schlüter, M., Feseker, T., and Kölling, M.: Rhizon sampling of porewaters near the sediment-water interface of aquatic systems, Limnol. Oceanogr. Methods, 3, 361–371, https://doi.org/10.4319/lom.2005.3.361, 2005.
 - Segers, R.: Methane production and methane consumption: a review of processes underlying wetland methane fluxes, Biogeochemistry, 41, 23–51, https://doi.org/10.1023/A:1005929032764, 1998.
- 1375 Sobek, S., Durisch-Kaiser, E., Zurbrügg, R., Wongfun, N., Wessels, M., Pasche, N., and Wehrli, B.: Organic carbon burial efficiency in lake sediments controlled by oxygen exposure time and sediment source, Limnol. Oceanogr., 54, 2243–2254, https://doi.org/10.4319/lo.2009.54.6.2243, 2009.

- Spafford, L. and Risk, D.: Spatiotemporal <u>v</u>+ariability in <u>L</u>lake-A<u>a</u>tmosphere <u>n</u>Net CO-2 <u>Ee</u>xchange in the <u>L</u>littoral <u>Z</u><u>z</u>one of an <u>o</u>Oligotrophic <u>l</u>Lake, J. Geophys. Res. Biogeosci., 123, 1260–1276, https://doi.org/10.1002/2017JG004115, 2018.
- Stumm, W. and Morgan, J. J.: Aquatic chemistry: Chemical equilibria and rates in natural waters, Third <u>3rd editioned.</u>, <u>A Wiley Interscience publication</u>, John Wiley & Sons Inc, New York, Chichester, Brisbane, Toronto, Singapore, <u>1022 pp., 19961995</u>.

Tamura, H., Goto, K., Yotsuyanagi, T., and Nagayama, M.: Spectrophotometric determination of iron(II) with 1,10-phenanthroline in the presence of large amounts of iron(III), Talanta, 21, 314– 318, 1974.

1385

1390

- Tfaily, M. M., Cooper, W. T., Kostka, J. E., Chanton, P. R., Schadt, C. W., Hanson, P. J., Iversen, C. M., and Chanton, J. P.: Organic matter transformation in the peat column at Marcell Experimental Forest: Humification and vertical stratification, J. Geophys. Res. Biogeosci., 119, 661–675, https://doi.org/10.1002/2013JG002492, 2014.
- Tolu, J., Rydberg, J., Meyer-Jacob, C., Gerber, L., and Bindler, R.: Spatial variability of organic matter molecular composition and elemental geochemistry in surface sediments of a small boreal Swedish lake, Biogeosciences, 14, 1773–1792, https://doi.org/10.5194/bg-14-1773-2017, 2017.
- Torres, I. C., Inglett, K. S., and Reddy, K. R.: Heterotrophic microbial activity in lake sediments: Effects
 of organic electron donors, Biogeochemistry, 104, 165–181, https://doi.org/10.1007/s10533-010 9494-6, 2011.
 - Updegraff, K., Pastor, J., Bridgham, S. D., and Johnston, C. A.: Environmental and <u>Ssubstrate Ccontrols</u> over <u>Ccarbon and Nnitrogen Mmineralization in Nnorthern Ww</u>etlands, Ecological Applications, 5, 151–163, https://doi.org/10.2307/1942060, 1995.
- Verpoorter, C., Kutser, T., Seekell, D. A., and Tranvik, L. J.: A global inventory of lakes based on high-resolution satellite imagery, Geophys. Res. Lett., 41, 6396–6402, https://doi.org/10.1002/2014GL060641, 2014.
 - Walpen, N., Getzinger, G. J., Schroth, M. H., and Sander, M.: Electron-<u>D</u>donating <u>Pp</u>henolic and <u>Ee</u>lectron-<u>Aa</u>ccepting <u>q</u>Quinone <u>Mm</u>oieties in <u>pPeat d</u>Dissolved <u>Oo</u>rganic <u>Mm</u>atter: Quantities and
- 1405 Rredox Ftransformations in the Ccontext of Ppeat Bbiogeochemistry, Environ. Sci. Technol., 52, 5236–5245, https://doi.org/10.1021/acs.est.8b00594, 2018.
 - Walpen, N., Schroth, M. H., and Sander, M.: Quantification of Pphenolic Aantioxidant <u>Mm</u>oieties in <u>Ddissolved Oorganic Mmatter by Flow-Injection Aanalysis with Eelectrochemical dDetection</u>, Environ. Sci. Technol., 50, 6423–6432, https://doi.org/10.1021/acs.est.6b01120, 2016.
- West, W. E., Coloso, J. J., and Jones, S. E.: Effects of algal and terrestrial carbon on methane production rates and methanogen community structure in a temperate lake sediment, Freshwater Biology, 57, 949–955, https://doi.org/10.1111/j.1365-2427.2012.02755.x, 2012.

Wetzel, R. G.: Gradient-dominated ecosystems: Sources and regulatory functions of dissolved organic matter in freshwater ecosystems, in: Dissolved Organic Matter in Lacustrine Ecosystems: Energy

- 1415 Source and System Regulator, edited by: Salonen, K., Kairesalo, T., and Jones, R. I., Springer Netherlands, Dordrecht, 181–198, https://doi.org/10.1007/978-94-011-2474-4_14, 1992.
 - Wetzel, R. G.: Gradient-dominated ecosystems: sources and regulatory functions of dissolved organic matter in freshwater ecosystems, in: Salonen K., Kairesalo T., Jones R.I. (eds) Dissolved Organic Matter in Lacustrine Ecosystems, 181–198.

- 1420 Whiticar, M. J.: Carbon and hydrogen isotope systematics of bacterial formation and oxidation of methane, Chemical Geology, 161, 291–314, https://doi.org/10.1016/S0009-2541(99)00092-3, 1999.
 - Wik, M., Crill, P. M., Varner, R. K., and Bastviken, D.: Multiyear measurements of ebullitive methane flux from three subarctic lakes, J. Geophys. Res. Biogeosci., 118, 1307–1321, https://doi.org/10.1002/jgrg.20103, 2013.
- Wilkinson, J., Maeck, A., Alshboul, Z., and Lorke, A.: Continuous <u>s</u>Seasonal <u>r</u>River <u>e</u>Ebullition <u>m</u>Measurements <u>l</u>Linked to <u>s</u>Sediment <u>m</u>Methane <u>f</u>Formation, Environ. Sci. Technol., 49, 13121–13129, https://doi.org/10.1021/acs.est.5b01525, 2015.
 - Yao, H., Conrad, R., Wassmann, R., and Neue, H. U.: Effect of soil characteristics on sequential reduction and methane production in sixteen rice paddy soils from China, the Philippines, and Italy,
- 1430 Biogeochemistry, 47, 269–295, https://doi.org/10.1007/BF00992910, 1999.

Supporting Information to: Organic matter and sediment properties determine in-lake variability of sediment CO₂ and CH₄ production and emissions of a small and shallow lake

Leandra Stephanie Emilia Praetzel, Nora Plenter, Sabrina Schilling, Marcel Schmiedeskamp, Gabriele

5 Broll, Klaus-Holger Knorr



Figure S1: FTIR spectra of samples from transect 3.



Figure S2: $CO_2(a)$ and $CH_4(b)$ production rates in 5-10 cm sediment depth. n = 3.



10

Figure S3: $CO_2(a)$ and $CH_4(b)$ production rates in the upper 5 cm of the sediment vs. lake depth. n=12. Different letters denote significant differences between groups.



Figure <u>S3S4</u>: Spatio-temporal variability of $EAC_{OM}(a, b, c)$, $EAC_{tot}(d, e, f)$ and $EDC_{OM}(g, h, i)$ in the incubation experiment at the beginning (a, d, g, n = 3), and the end (b, e, h, n = 3) of the experiment as well as average values for the whole experiment (c, f, i, n = 6).



Figure <u>S4S5</u>: Depth profile of CH_4 (a) and CO_2 (b) concentration in the sediment of the mesocosms. Different symbols denote three replicates at each site. Values are means over sampling period \pm SD. n = 2-10

Table S1: Spearman's rank correlation coefficients and significance levels for $\underline{CO_2 \text{ and } CH_4 \text{ production}, Q_{10}\text{-values and EAC and EDC with all other all measured parameters.}$ Italic numbers denote no significant correlations. means that correlations were not significant (p > 0.05). ACM = acetoclastic methanogenesis, HTM = hydrogenotrophicmethanogenesis.

	CH ₄ production		CO ₂ production		Q ₁₀ (CH ₄)		acetoclastic methanog.		EDC		EACOM		CH ₄ -flux		CO₂ flux	
	P	rho	P	rho	P	rho	P	rho	P	rho	P	rho	P	rho	p	rho
aromatics	0.001	-0.669	0.002	-0.641	0.034	-0.821							0.132	-0.595	0.197	-0.524
fats, waxes, lipids	0.000	-0.700	0.000	-0.736	0.034	-0.821			0.001	0.565			0.015	-0.833	0.428	-0.333
humic acids	0.003	-0.618	0.001	-0.653	0.034	-0.821							0.015	-0.833	0.389	-0.357
lignin	0.003	-0.606	0.003	-0.610	0.034	-0.821							0.021	-0.786	0.352	-0.381
phenols	0.003	-0.606	0.001	-0.667	0.034	-0.821							0.015	-0.833	0.389	-0.357
e	0.194	-0.287	0.327	-0.219									0.058	-0.714	0.665	- 0.190
C/N	0.356	-0.206	0.378	-0.197			0.013	0.447					0.007	-0.881	0.428	-0.333
H ₂ -concentration	0.000	0.450	0.000	0.515							0.157	-0.285				
acetate concentration	0.492	0.248	0.191	0.455							0.002	-0.387				
hvdrogenotrophic											0.031	0.426				
acetoclastic											0.748	0.042				
EAC _{OM}	0.946	0.030	0.470	- 0.261												
EAC inorg	0.204	- 0.442	1.000	0.006												
EACtot	0.191	-0.455	0.470	- 0.261												
EDC	0.031	-0.697	0.039	-0.673												
EAC/EDC	0.247	0.406	0.407	0.297												
EEC	0.166	-0.479	0.054	-0.636												
S (%)	0.019	-0.577	0.087	- 0.441												
clay													0.023	0.648	0.037	0.605
silt													0.101	0.497	0.340	0.302
sand													0.023	-0.648	0.037	-0.605
CH4-stock-change													0.163	- 0.222	0.714	-0.064
CO₂ stock change													0.762	-0.049	0.776	0.050

	<u>CH₄ production</u>		CO₂ production		<u>Q₁₀ (CH₄)</u>		EAC _{OM}		EAC tot		l	EDC	
	<u>p</u>	<u>rho</u> n	<u>p</u>	<u>rho</u> n	<u>p</u>	<u>rho</u> n	<u>p</u>	<u>rho</u> n	<u>p</u>	<u>rho</u>	<u>n p</u>	<u>rho</u> n	
aromatics	<u>< 0.001</u>	<u>-0.669</u> <u>22</u>	<u>< 0.01</u>	<u>-0.641</u> <u>22</u>	<u>< 0.05</u>	<u>-0.821</u> <u>7</u>	<u>n.s.</u>	<u>0.030</u> <u>10</u>	<u>n.s.</u>	<u>0.042</u> <u>1</u>	<u>0 n.s.</u>	<u>0.515</u> <u>10</u>	
<u>fats, waxes, lipids</u>	<u>< 0.001</u>	<u>-0.700</u> <u>22</u>	<u>< 0.001</u>	<u>-0.736</u> <u>22</u>	<u>< 0.05</u>	<u>-0.821</u> <u>7</u>	<u>n.s.</u>	<u>0.321 10</u>	<u>n.s.</u>	<u>0.333</u> <u>1</u>	0 < 0.05	<u>0.758</u> <u>10</u>	
humic acids	<u>< 0.01</u>	<u>-0.618</u> <u>22</u>	<u>< 0.01</u>	<u>-0.653</u> <u>22</u>	<u>< 0.05</u>	<u>-0.821</u> <u>7</u>	<u>n.s.</u>	<u>0.067</u> <u>10</u>	<u>n.s.</u>	<u>0.042</u> <u>1</u>	<u>0 n.s.</u>	<u>0.455</u> <u>10</u>	
<u>lignin</u>	<u>< 0.01</u>	<u>-0.606</u> <u>22</u>	<u>< 0.01</u>	<u>-0.610</u> <u>22</u>	<u>< 0.05</u>	<u>-0.821</u> <u>7</u>	<u>n.s.</u>	<u>0.031</u> <u>10</u>	<u>n.s.</u>	<u>-0.006</u> <u>1</u>	<u>0 n.s.</u>	<u>0.275</u> <u>10</u>	
<u>phenols</u>	<u>< 0.01</u>	<u>-0.606</u> <u>22</u>	<u>< 0.001</u>	<u>-0.667</u> <u>22</u>	<u>< 0.05</u>	<u>-0.821</u> <u>7</u>	<u>n.s.</u>	<u>0.152</u> <u>10</u>	<u>n.s.</u>	<u>0.115</u> <u>1</u>	<u>0 n.s.</u>	<u>0.479 10</u>	
<u>C</u>	<u>n.s.</u>	<u>-0.287</u> <u>22</u>	<u>n.s.</u>	<u>-0.219</u> <u>22</u>	<u>n.s.</u>	<u>-0.393</u> <u>7</u>	<u>n.s.</u>	<u>-0.394</u> <u>10</u>	<u>n.s.</u>	<u>-0.370</u> <u>1</u>	<u>0 n.s.</u>	<u>-0.321 10</u>	
<u>C/N</u>	<u>n.s.</u>	<u>-0.206</u> <u>22</u>	<u>n.s.</u>	<u>-0.197</u> <u>22</u>	<u>n.s.</u>	<u>-0.074</u> <u>7</u>	<u>n.s.</u>	<u>-0.079 10</u>	<u>n.s.</u>	<u>-0.115</u> <u>1</u>	<u>0 n.s.</u>	<u>-0.103</u> <u>10</u>	
$\delta^{13}C$	<u>n.s.</u>	<u>0.134</u> <u>22</u>	<u>n.s.</u>	<u>0.091</u> <u>22</u>	<u>n.s.</u>	<u>-0.321</u> 7	<u>n.s.</u>	<u>0.309</u> <u>10</u>	<u>n.s.</u>	<u>0.224</u> <u>1</u>	<u>0 n.s.</u>	<u>0.164</u> <u>10</u>	
$\delta^{15}N$	<u>n.s.</u>	<u>-0.267</u> <u>22</u>	<u>n.s.</u>	<u>-0.281</u> <u>22</u>	<u>n.s.</u>	<u>-0.071</u> <u>7</u>	<u>n.s.</u>	<u>0.055</u> <u>10</u>	<u>n.s.</u>	<u>-0.006</u> <u>1</u>	<u>0 n.s.</u>	<u>-0.164</u> <u>10</u>	
H ₂ conc.	<u>< 0.001</u>	<u>0.450</u> <u>22</u>	<u>< 0.001</u>	<u>0.515</u> <u>22</u>			<u>n.s.</u>	<u>-0.285</u> <u>26</u>	<u>n.s.</u>	<u>-0.335</u> 2	<u>6 <i>n.s.</i></u>	<u>-0.139</u> <u>26</u>	
<u>acetate conc.</u>	<u>n.s.</u>	<u>0.248</u> <u>10</u>	<u>n.s.</u>	<u>0.455</u> <u>10</u>			<u>< 0.01</u>	<u>-0.387</u> <u>60</u>	<u>< 0.001</u>	<u>-0.418</u> 6	<u>0 n.s.</u>	<u>-0.035</u> <u>60</u>	
HTM	<u>< 0.01</u>	<u>-0.394</u> <u>22</u>	<u>< 0.001</u>	<u>-0.516</u> <u>22</u>			<u>< 0.05</u>	<u>0.426</u> <u>26</u>	<u>< 0.05</u>	<u>0.426</u> <u>2</u>	<u>6 <i>n.s.</i></u>	<u>0.162</u> <u>26</u>	
<u>ACM</u>	<u>n.s.</u>	<u>0.491 10</u>	<u>n.s.</u>	<u>0.297</u> <u>10</u>			<u>n.s.</u>	<u>0.042</u> <u>60</u>	<u>n.s.</u>	<u>0.042</u> 6	<u>0 n.s.</u>	<u>-0.208</u> <u>60</u>	
EAC _{OM}	<u>n.s.</u>	<u>0.030</u> <u>10</u>	<u>n.s.</u>	<u>-0.261</u> <u>10</u>									
EAC inorg	<u>n.s.</u>	<u>-0.261</u> <u>10</u>	<u>n.s.</u>	<u>0.152</u> <u>10</u>									
EAC tot	<u>n.s.</u>	<u>-0.042</u> <u>10</u>	<u>n.s.</u>	<u>-0.285</u> <u>10</u>									
EDC	< 0.05	<u>-0.697</u> <u>10</u>	<u>< 0.05</u>	<u>-0.673</u> <u>10</u>									
EAC/EDC	<u>n.s.</u>	<u>0.406</u> <u>10</u>	<u>n.s.</u>	<u>0.297</u> <u>10</u>									
EEC	<u>n.s.</u>	<u>-0.479</u> <u>10</u>	<u>n.s.</u>	<u>-0.636</u> <u>10</u>									
<u>S</u>	<u>n.s.</u>	<u>-0.411</u> <u>16</u>	<u>< 0.05</u>	<u>-0.446</u> <u>16</u>									
<u>P</u>	<u>n.s.</u>	<u>0.282</u> <u>22</u>	<u>n.s.</u>	<u>0.305</u> <u>22</u>									
<u>Fe</u>	<u>n.s.</u>	<u>0.338</u> <u>22</u>	<u>< 0.05</u>	<u>0.453</u> <u>22</u>									
<u>Mn</u>	<u>n.s.</u>	<u>0.020</u> <u>22</u>	<u>n.s.</u>	<u>0.121</u> <u>22</u>									
CO ₂ conc.							<u>n.s.</u>	<u>-0.161</u> <u>20</u>	<u>n.s.</u>	<u>-0.146</u> 2	<u>0 n.s.</u>	<u>-0.252</u> <u>20</u>	
<u>CH₄ conc.</u>							<u>n.s.</u>	<u>-0.083</u> <u>20</u>	<u>n.s.</u>	<u>-0.080</u> 2	<u>0 n.s.</u>	<u>-0.219</u> <u>20</u>	

Table S2: Spearman's rank correlation coefficients and significance levels for Gibb's free energy of acetoclastic (ACM) and hydrogenotrophic (HTM) methanogenesis, acetate and H_2 concentrations with OM quality parameters. n.s. means that correlations were not significant (p > 0.05).

	AC	<u>M (t6)</u>	<u>acetate</u>	<u>conc. (t6)</u>]	HTM	H	<u>H₂ conc.</u>			
	<u>p</u>	<u>rho</u> <u>n</u>	<u>p</u>	<u>rho</u> n	<u>p</u>	<u>rho</u>	<u>n p</u>	<u>rho</u>	<u>n</u>		
<u>aromatics</u>	<u>n.s.</u>	<u>-0.052</u> <u>30</u>	< 0.001	<u>-0.585</u> <u>30</u>	<u>n.s.</u>	<u>0.136</u> <u>15</u>	<u>54</u> < 0.05	<u>-0.187</u>	<u>174</u>		
<u>fats, waxes, lipids</u>	<u>n.s.</u>	<u>-0.237</u> <u>30</u>	<u>< 0.05</u>	<u>-0.440</u> <u>30</u>	< 0.01	<u>0.255</u> <u>15</u>	<u>54 < 0.001</u>	<u>-0.352</u>	<u>174</u>		
humic acids	<u>n.s.</u>	<u>0.082</u> <u>30</u>	<u>< 0.001</u>	<u>-0.634</u> <u>30</u>	<u>< 0.05</u>	<u>0.183</u> <u>15</u>	<u>54</u> < 0.01	-0.229	<u>174</u>		
<u>lignin</u>	<u>n.s.</u>	<u>0.265</u> <u>30</u>	< 0.01	<u>-0.533</u> <u>30</u>	<u>n.s.</u>	<u>0.140</u> <u>15</u>	<u>54</u> < 0.05	<u>-0.190</u>	<u>174</u>		
<u>phenols</u>	<u>n.s.</u>	<u>0.104</u> <u>30</u>	<u>< 0.001</u>	<u>-0.668</u> <u>30</u>	<u>< 0.05</u>	<u>0.196</u> <u>15</u>	<u>54</u> < 0.01	<u>-0.246</u>	<u>174</u>		
<u>C</u>	<u>n.s.</u>	<u>-0.062</u> <u>30</u>	<u>n.s.</u>	<u>0.074</u> <u>30</u>	<u>< 0.05</u>	<u>0.174 15</u>	<u>54</u> < 0.05	<u>-0.149</u>	<u>174</u>		
<u>C/N</u>	<u>< 0.05</u>	<u>0.447</u> <u>30</u>	<u>< 0.01</u>	<u>-0.538</u> <u>30</u>	<u>n.s.</u>	<u>-0.057</u> <u>15</u>	<u>54</u> <u>n.s.</u>	<u>0.102</u>	<u>174</u>		