Characterising the surface microlayer in the Mediterranean Sea: trace metal concentrations and microbial plankton abundance

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Abstract

The Sea Surface Microlayer (SML) is known to be enriched by trace metals relative to the underlying water and to harbour diverse microbial communities (i.e. neuston). However, the processes linking metals and biota in the SML are not yet fully understood. The metal (Cd, Co, Cu, Fe, Ni, Mo, V, Zn and Pb) concentrations in aerosol samples in the SML (dissolved and total fractions) and in the subsurface waters (SSW; dissolved fraction at ~1 m depth) from the Western Mediterranean Sea were analysed in this study during a cruise in May-June 2017. The composition and abundance of the bacterial community in the SML and SSW, the primary production and Chl-a in the SSW were measured simultaneously at all stations during the cruise. Residence times in the SML of metals derived from aerosol depositions were highly variable and ranged from minutes for Fe (3.6 ± 6.0 min) to a few hours for Cu (5.8 ± 6.2 h). Concentrations of most of the dissolved metals in both the SML and SSW were positively correlated with the salinity gradient and showed the characteristic eastward increase in the surface waters of the Mediterranean Sea (MS). In contrast, the total fraction of some reactive metals in the SML (i.e. Cu, Fe, Pb and Zn) showed a negative correlation with salinity and a positive correlation with microbial abundance, which might be associated with microbial uptake. Our results show a strong negative correlation between the dissolved and total Ni concentration and heterotrophic bacterial abundance in the

SML and SSW, but we cannot ascertain whether this correlation reflects a toxicity effect or is the result of some other process.

1. Introduction

The Mediterranean Sea (MS) is enriched in many trace metals relative to similar nutrient- depleted waters in the open ocean (e.g. Cd, Cr, Co, Cu, Ni, Fe, Zn) (Bonnet et al., 2013; Boyle et al., 1985; Sarthou and Jeandel, 2001; Sherrell and Boyle, 1988). The enrichment of metals in surface water has been associated with different sources including atmospheric deposition, river inflows, groundwater supply, anthropogenic sources and the Atlantic Ocean inflow through the Gibraltar Strait (Boyle et al., 1985; Elbaz-Poulichet et al., 2001; Migon, 2005; Trezzi et al., 2016). The MS has one of the highest rates of aeolian deposition in the world with strong pulses of mineral dust from Africa, in addition to regular anthropogenic aerosol inputs from Europe. Therefore, atmospheric deposition, both dry and wet, is the dominant pathway for the large scale transport of trace metals to the water column and sediments in the MS (Guieu et al., 2002, 2010; Jordi et al., 2012; Ternon et al., 2010; Tovar-Sánchez et al., 2010, 2014). Many of these metals play an important role in biogeochemical processes of this sea. For example, it has been hypothesized that the high Co concentrations in the MS may stimulate "de novo" synthesis of vitamin B_{12} as Co is the central metal ion in the B_{12} molecule (Bonnet et al., 2013). Although present in a higher concentration than in other oceans, Fe may control phytoplankton growth and composition (Sarthou and Jeandel, 2001). Copper from aerosol deposition has been demonstrated to have toxic effects on marine phytoplankton (Jordi et al., 2012; Paytan et al., 2009) while Ni and Zn concentrations in *Posidonia oceanica* are considered good proxy for aerosols deposition (Tovar-Sánchez et al., 2010).

The SML is considered the skin of the ocean as it serves as a boundary layer between the atmosphere and the ocean. With a thickness of 1−1000 μm, it is a prevalent feature of the oceans' surface that shows distinct physical, chemical, and biological properties. This sea-air interface plays a key role in regulating the exchange of gases, solutes and energy between water and atmosphere and is central to a wide range of global biogeochemical and climate regulation processes (Cunliffe et al., 2013). Characterized by a high abundance of microorganisms (called neuston and ranging from bacteria to larger siphonophores (Wurl et al., 2017)), the SML constitutes a particular marine ecosystem. Neuston in the

SML is sustained by recruitment from the underlying plankton, but its composition and activities are different as they are heavily conditioned by meteorological conditions, intensity of UV radiation, organic matter content and/or aerosol impact among others (Cunliffe et al., 2013; Engel et al., 2017).

Impacted by different allochthonous sources (e.g. aerosols, ice, rivers) the SML is enriched in reactive trace metals (e.g. Cu, Fe, Pb) whose metal stoichiometry signature is different from that of the underlying waters (Tovar-Sánchez et al., 2019). For example, in regions under the influence of dust events, such as the North Atlantic Ocean or Mediterranean Sea, concentrations of Cu, Fe or Pb in the total pool of the SML are up to 800, 200 and 150 times higher than in the dissolved metal pool of the underlying water (Tovar-Sánchez et al., 2019). However, despite such enrichment in the concentration of trace metals, little is known about their residence times, their influence on the activity of the microbial community within the SML, or their impact on the biogeochemistry of the underlying waters. Previous studies, using data from both field sampling and laboratory microcosms, have estimated the residence times of dissolved and particulate trace metals (e.g. Al, Fe, Mn, Ni, Cu, Zn, and Pb) in the SML to range from a few minutes to a few hours. This is likely to be long enough to alter the SML chemically and biologically and affect the composition and activity of the neuston community (Ebling and Landing, 2017; Hardy et al., 1985). However, there is still a lack of estimates of residence times that consider the full range of key processes that directly affect the physical, chemical and biological composition of the surface microlayer, such as dry and wet deposition fluxes, wind speeds, and neuston composition.

Here, we investigate the dissolved (<0.22 μm) and total (unfiltered) trace metal composition (Cd, Co, Cu, Fe, Ni, Mo, V, Zn and Pb) of the SML in the central and western MS. Aerosols were sampled and analysed for trace metals at the same stations and residence time of particulate aerosol metals in the SML were calculated. The microbial composition and abundance in the SML and subsurface water (SSW), the primary production and Chl-a concentration in the subsurface water (1-5m) was analysed and their relationships with trace metal concentrations and distribution were examined.

2. Material and Methods

Samples from the SML, the SSW and aerosols were collected during the PEACETIME (ProcEss studies at the Air-sEa Interface after dust deposition in the MEditerranean sea) cruise on board the French

R/V 'Pourquoi Pas?' in the MS, from May 10th to June 11th, 2017 (Guieu et al., 2020). Twelve stations were sampled (Figure 1). Three of these stations were sampled twice (TYR 1-2, ION 1-2) or five times (FAST 1-5) on different days, resulting in a total of 17 sets of samples (Tables 1-2).

2.1. Aerosol sampling and analysis

A PEGASUS container was installed aboard the R/V Pourquoi Pas? This container is a mobile platform equipped with a set of instruments optimized to collect and analyze gaseous compounds and particles in the atmospheric boundary layer (Formenti et al., 2019) in real time. Atmospheric sampling was performed using isokinetic and wind-oriented aerosol multi-samplers with a total sampling flow rate of ca. 400 L min^{-1} per inlet. This inlet was developed to sample particles with an aerodynamic diameter inferior to 40 μm (Rajot et al., 2008). The total flow was subdivided into various transmission lines, which served the majority of the instrumentation. The aerosol size distribution from 10 nm to 30 μm was measured by a combination of standard optical and electrical mobility analysers (SMPS, TSI Scanning Mobility Particle Sizer and GRIMM Inc. optical particle counters - OPC, 1.109). The total mass concentration was obtained by an on-line Tapering Element Oscillating Microbalance (TEOM, model 1400a, Rupprecht and Patashnick Co. R&P).

One of the sampling lines was equipped with a filtration unit to collect the aerosols on 47-mm polycarbonate membranes of 0.4 μm pore size (Whatman Nuclepore ^{TM}). The volume flow rate was set at 20 L.min^{-1}. All filters had been previously cleaned by immersion in ultrapure HCl (2%) during 2 hours and rinsed with ultrapure waters. A sampling strategy was used to avoid contamination by the ship´s fumes. First, when the vessel was at the station, the R/V was systematically positioned so that the inlets were facing the wind (the PEGASUS container and the ship's stack were on the opposite sides of the deck). While steaming, contamination-free sampling was carried out when the relative wind direction was not in the path of chimney exhaust. In total, 36 series of filters were collected, including 17 filters during the work at the station and 5 blanks filters. The sampling locations for each filter are presented in Figure 1. The aerosol filters were first analysed by X-ray fluorescence spectrometry (SFX, spectrometer PW-2404, Panalytical™) to measure the chemical markers of particle origin sources (such as Al and Ca). The filters were then leached with ultrapure water in order to determine the soluble fraction of metals. Finally,

the filters were mineralized using an acid digestion protocol adapted from (Heimburger et al., 2013) in order to quantify the insoluble (particulate) fraction of metals. The dissolved and digested samples were analysed by HR-ICP-MS (Neptune Plus, Thermo Scientific ™) for the trace metals: Cd, Co, Cu, Fe, Mo, Ni, Pb, V, Zn. The total concentration of metals corresponds to the sum of dissolved and particulate fraction of the metals. Rain was also sampled during the cruise with on-line filtration collector (0.2µm, polycarbonate, Nuclepore Whatman™) (Heimburger et al., 2013) and analysed particulate and dissolved metals by HR-ICP-MS.

2.2. Water sampling and analysis

2.2.1. Trace metals

Surface samples, i.e. SML and subsurface water (SSW: ~1 m depth), were collected from a pneumatic boat deployed 0.5 – 1 mile away from the research vessel in order to avoid contamination of the samples by the vessel. SML samples were collected using a glass plate sampler (Stortini et al., 2012; Tovar-Sánchez et al., 2019) which had been previously cleaned with acid overnight and rinsed thoroughly with ultrapure water (MQ-water). Once at the station, the glass plate and the whole sampler were rinsed with seawater several times, and the three first dips (SML samples) were discharged. The 39 x 25 cm silicate glass plate had an effective sampling surface area of 1950 cm^2 considering both sides. In order to check for procedural contamination, we collected SML blanks at some stations on board of the pneumatic boat by rinsing the glass plate with ultra-pure water and collecting 0.5 L using the glass plate system. The sample signal to blank ratio was typically greater than 5:1 for all elements. The surface microlayer thickness (µm) was calculated following the formula (Wurl, 2009):

$$SML_T = V_S \cdot 10^4 / N(A_p \cdot 2)$$

where VS is the volume sampled (mL), N the number of dips, and A_p the surface area of the glass plate (cm^2).

The total fraction of SML (i.e. T-SML) was collected directly from the glass plate system without filtration in acid cleaned 0.5 L LDPE bottles, while the dissolved fraction in the SML (i.e. D-SML) was rapidly filtered on board the pneumatic boat through an acid-cleaned polypropylene cartridge filter

(0.22μm; MSI, Calyx®). SSW (~1 m depth) were collected using acid-washed Teflon tubing connected to a peristaltic pump and directly filtered on the same cartridge to collect the dissolved fraction (D-SSW). All samples were acidified on board to pH< 2 with Ultrapure-grade HCl in a class-100 HEPA laminar flow hood. The metals, (i.e. Cd, Co, Cu, Fe, Ni, Mo, V, Zn and Pb) were stored for at least 1 month prior to analysis. The samples were pre-concentrated using an organic extraction method (Bruland et al., 1979) and quantified by ICP-MS (Perkin Elmer ELAN DRC-e). Prior to pre-concentration and to ensure the breakdown of metal-organic complexes and the removal of organic matter (Achterberg et al., 2001; Milne et al., 2010), total fraction samples (i.e. T-SML) were digested using a UV system consisting of a UV (80 W) mercury lamp that irradiated the samples (contained in quartz bottles) during 30 min. The accuracy of the pre-concentration method and analysis for trace metals was established using Seawater Reference Material for Trace Elements (CASS 6, NRC-CNRC) and compares well with the certified values given in brackets (in µg/L): Cd: 0.02 (0.022); Co: 0.07 (0.067); Cu: 0.44 (0.418); Fe: 1.58 (1.56); Mo: 8.1 (9.15); Ni: 0.41 (0.418); Pb:0.01 (0.011); V: 0.45 (0.49); Zn: 1.14 (1.127).

2.2.2. Ancillary parameters

The temperature and salinity of the surface seawater were measured with the underway thermosalinograph (TSG) system of the R/V Pourquoi Pas?,which was composed of a *Seabird® SBE 21 seaCAT* linked to a *SBE 38* thermometer situated at the seawater inlet. The seawater inlet was located 3 metres under the sea surface. The wind speed at 10 metres was measured with a *Gill Windsonic* ultrasonic anemometer from the on-board BATOS station deployed by the French meteorological agency *Météo France* on the vessel. Temperature, salinity and wind data were binned every 30 seconds by the ship's data management system TECHSAS (*TECHnical Sensor Acquisition System*). The average values of temperature, salinity and wind speed for a time period of 1 hour around the time of SML sampling are shown in Table 1.

2.3. Biological sampling and analysis

2.3.1. Neuston

The microorganisms inhabiting the SML are collectively referred to as the "neuston" (Engel et al., 2017). The microorganisms in the SML were sampled at the same time as the trace metal sample collection, also using a glass plate system (50 x 26 cm silicate glass plate with an effective sampling surface area of 2600 cm^2 considering both sides; (Cunliffe and Wurl, 2014; Harvey, 1966)). The water from the SSW was collected from around a 20 cm depth in acid-clean borosilicate bottles. The bacterial numbers were determined using flow cytometry from a 4 mL sample that was fixed with 200 mL glutaraldehyde (GDA, 1% final concentration). The samples were stored at -20 ºC for 2.5 months at most until analysis and were then stained with SYBR Green I (Molecular Probes) (Marie et al., 1997). The samples were analysed using a flow cytometer equipped with a 488 nm laser (Becton & Dickinson FACScalibur). A plot of side scatter (SSC) vs. green fluorescence (FL1) was used to detect the unique signature of the bacterial cells. The internal standard consisted of yellow-green latex beads (Polysciences, 0.5 μm). The abundance and area of Transparent Exopolymer Particles (TEP) were measured microscopically following a previously described method (Engel, 2009).

2.3.2. Phytoplankton and Primary Production

Chl-a concentration and primary production were measured in the SSW at a depth of 5 m. The primary production was measured with the ^{14}C-uptake technique, following the methods detailed in (Marañón et al., 2000). The seawater samples, collected in Niskin bottles at dawn, were dispensed into four (3 light and 1 dark) polystyrene bottles of 70 mL in volume, which were amended with 15 μCi of NaH^{14}CO$_3$ and incubated for 24 h inside a deck incubator refrigerated with surface seawater from the continuous water supply. The incubator was covered with a neutral density filter that provided an irradiance level of 70% of incident PAR. After incubation, the samples were filtered, using low vacuum pressure, through 0.2-μm polycarbonate filters. The filters were then exposed to HCl fumes overnight to remove non-fixed, inorganic ^{14}C. After adding 5 mL of liquid scintillation cocktail to the filters, the radioactivity of each sample was determined on-board with a liquid scintillation counter. The dark-bottle DPM value was subtracted from the light-bottle DPM value to compute the rates of carbon fixation. A value of 26,663 μgC L^{-1} was used for the concentration of dissolved inorganic carbon (DIC), which corresponds to the mean DIC concentration at 5 m measured during the cruise. Chl-a concentrations were

measured by HPLC (HPLC Agilent Technologies 1200) following the method described by (Ras et al., 2008).

2.4. Statistical analyses

The Spearman rank correlation coefficient (r_s) was used to determine significant relationships ($p<$ 0.05) between the variables measured in the different compartments (air, SML and SSW). Least square linear regressions and determination coefficients (R^2) were calculated to ascertain the relationship between selected variables. Statistical analyses were performed with the aid of the statistical software package SPSS 25.

3. Results and Discussion

3.1. Aerosols depositions

Metal aerosol compositions are shown in Table 1. The range (ng/m^3) of concentrations in our study (Cd: 0 – 0.2 , Co: 0.05 - 0.3 , Cu: 0.4 – 4.8, Fe: 23 – 266, Ni: 1.6 – 7.1, Mo: 0.0 – 2.1, V: 3 – 18, Zn: 2.0 – 14.6 and Pb: 0.2 – 7.0) was of the same order of magnitude of previous measurements collected in the same region and season (Becagli et al., 2012; Calzolai et al., 2015, p.10; Guerzoni et al., 1997; Tovar-Sánchez et al., 2014; Heimbürger et al., 2010), and were therefore consistent with Western Mediterranean background concentrations. However, concentrations of trace metals (i.e. Pb, Cd and Zn) have decreased remarkably over the last two decades, while crustal metals have not shown any evolution (Heimbürger et al., 2010). No clear gradient (North-South or East-West) in the concentrations of atmospheric metals was observed during the cruise. The composition of metal aerosols was mainly influenced by air masses from the North of Europe and Atlantic Ocean (Figure S1), except between June 1st and June 5th (i.e. for the stations St 9 and Fast 1-4) when African air masses were loaded with dust (Figure S1 and S2). During this period, the aerosol mass concentrations were the highest observed during the cruise with a maximum of around 25 µg.m^{-3} (Figure S3), nevertheless these concentrations were typical of a moderate dust event (Pey et al., 2013). The aerosol Fe concentrations during this period, which averaged 245 ng.m^{-3}, were the highest measured during the cruise. The same observation was made for Co. A positive correlation between Ni and V in the aerosols collected throughout the cruise suggest a

common source associated to heavy oil combustion, i.e. marine ship traffic (Becagli et al., 2012). Some rain events occurred during the cruise, one measured when the vessel was at station, June 5th from 2:36 am to 3:04 am (between Fast 3 and Fast 4 samples). However, the whole zone around the Fast station was rainy from the 3rd of June (Figure S4). As the rain composition collected was typical of wet dust deposition with high particulate concentrations of Al, Fe and Ca (Fu et al., in preparation), we suppose the rain-out of dust in the atmospheric column around this station occurred between the 3rd to the 5th of June and hence dusty rain events impacted the concentration of surface waters during Fast stations. The total trace metal concentrations in the dusty rain collected, ranged from 180 pM for Cd to 343 nM for Fe (Cd: 180 pM , Co: 1380 pM , Cu: 18.1 nM, Fe: 343 nM, Ni: 9.9 nM, Mo: 875 pM, V: 26.9 nM, Zn: 345 nM and Pb: 788 pM).

3.2. Biochemical composition and distribution of the surface water

Trace metal concentrations in the surface waters of the MS varied depending on the layer sampled (i.e. SML or SSW) and along a longitudinal gradient (Table 1).

3.2.1. Trace metals in the SML

With the exception of Pb, trace metal concentrations of T-SML (Table 1) were lower, although, of the same order of magnitude, than those measured in a previous study carried out in the MS (Tovar-Sánchez et al., 2014). The lower content of metals in the SML is probably related to the low dust aerosol deposition during our sampling period and to the lack of desert dust aerosols except for the Fast station. T-SML average concentration (± SD) of Pb (663 ± 320 pM) was one order of magnitude lower than in previous studies which investigated pulses of both mineral African dust and anthropogenic aerosols coming from Europe (5596 ± 1589 pM) (Tovar-Sánchez et al., 2014). The highest concentrations of some metals were measured at the FAST-3 station (Co: 773.6 pM; Cu: 20.1 nM; Fe: 1433.3 nM and Pb: 1294.7 pM), probably affected by the dusty rain events on this area.

Dissolved concentrations of Co, Zn, Pb, Cu and Ni showed a decreasing trend from the SML to the SSW, with average concentrations (± SD) 10.4 ± 0.7, 9.3 ± 5.5, 4.2 ± 1.8, 3.1 ± 1.5, and 1.2 ± 0.1 times higher in the SML than in the SSW, respectively. The SML to SSW concentration ratio for V (1.2

± 0.42) and Fe (1.3 ± 1.5) indicated only slight enrichment in the SML, while the ratio for Mo (1.0 ± 0.1) indicated no difference between layers (Table 1). Only Cd concentrations were consistently lower in the SML compared to the underlying water (0.8 ± 0.2 times lower). Such low ratio of dissolved metals (as it is the case of Cd) in the SML compared to the underlying water has been previously observed in areas without significant aerosol inputs (Ebling and Landing, 2015, 2017). Although not fully understood, some mechanisms such as the dominance of removal processes over diffusion, or the higher influence of metal sources from underlaying water have been suggested previously to explain this difference of metal concentrations between compartments (Ebling and Landing, 2017; Hunter, 1980). Given the high exposition to solar radiation and the enrichment in organic matter, photoreaction is likely to play an important role, driving redox processes that alter metal speciation and complexation between inorganic and organic species, and affecting the distribution of species in the SML and their transfer to the SSW. Photoredox processes in the SML are not fully understood; however, the strong dependence of redox seawater chemistry and complexation of elements such as Cu and Fe on solar radiation have been studied (Croot and Heller, 2012; Moffett and Zika, 1988). Thus, for example Fe(III) and Cu(II) could be photochemically reduced in surface seawaters to highly soluble Fe(II) and Cu(I) (Sunda, 2012). Although these reduced states are unstable in oxygenated waters and are re-oxidized and re-chelated by organic ligands on time scales of minutes (Sunda, 2012), the continuous and intense UV radiation in the SML could be intensifying these reactions, and enhancing the transfer between SML and SSW and its effect on phytoplankton.

The spatial distribution of Co and Ni concentrations in the D-SML was well correlated with that observed in the D-SSW (Spearman's correlation coefficient (r_s): 0.87 for Co and 0.91 for Ni; $p<0.01$, Table 3), indicating an efficient diffusive transfer between these two compartments for these elements. The concentration of these elements was also positively correlated with the surface salinity distribution (r_s: 0.62 for Co and 0.93 for Ni; $p<0.01$, Table 3), and presented an eastward trend of increasing concentration, which is consistent with the characteristic distribution of metals on the surface of the MS (see section 3.2.4. below). The variations in concentrations for the rest of the elements (i.e. Cd, Cu, Fe, Pb, V and Zn) in the D-SML were not correlated either with the concentration in the underlying water or with salinity. Multiple physical, chemical and biological processes taking place in the SML could affect

the mobility and diffusion of these elements between compartments. However, the concentrations of Cu, Fe and Zn in the T-SML showed an opposite longitudinal trend to that of salinity (r_s: -0.59 for Cu; -0.69 for Fe, and -0.61 for Zn; $p<0.01$, Table 3). No correlations between T-SLM and D-SLM trace metal concentrations were found except for Cd (0.789 $p<0.01$) and Ni (0.681 $p<0.01$). Since aerosol metal concentrations did not show any longitudinal trend and no other natural or anthropogenic sources were identified in the region, the longitudinal gradient in the concentration of these reactive trace elements in the T-SML must have been influenced by other factors than source inputs, water exchange or dilution with Atlantic waters. The complex matrix of the SML and the particular organic and inorganic speciation of each metal studied in the SML will affect their distribution. Thus, for example, Cd and Zn characterized by an oxidation state number of II can vary from very weak to very strong complexation. Lead in oxygenated seawater is partitioned between chloride and carbonate complexes, while Fe and Cu speciation are strongly influenced by pH (Byrne, 2002).

3.2.2. Residence time of trace metals in the SML

Estimates of the residence times of metals from aerosol deposition in the SML are critical to understanding the biogeochemical processes that affect the fate and distribution of trace metals in the oceans' surface better. The equation proposed by (Ebling and Landing, 2017) was used to estimate the residence time (t) of particulate metals in the SML:

$$t = [TE]_{SML} \times d/J_{aerosol}$$

where $[TE]_{SML}$ is the concentration of the trace element (TE) in the T-SML, d is the thickness of the SML and, and $J_{aerosol}$ is the aerosol trace metal flux measured. $J_{aerosol}$ was estimated by multiplying the metal aerosol concentrations with the sedimentation velocity, which is dependent on aerosol size. During the cruise, Al and Fe atmospheric concentrations were correlated at all the stations and the ratio Fe/Al is typical of a crustal source (Fu et al., in prep.). The atmospheric iron deposition fluxes are associated to mineral dust particles even during the period when the Saharan dust inputs are very low (Desboeufs et al., 2018; Guieu et al., 2010). Conversely, no correlation with Al is observed for the other metals, except during FAST1-3. Thus, we used a mineral dust deposition velocity for Fe of 1 cm.s^{-1} and an average velocity of fine anthropogenic particles for the other metals, i.e. 0.1 cm.s^{-1} (Baker et al., 2010; Duce et

al., 1991). For the calculations of the residence times throughout our different stations we used simultaneous empirical measurements of total metal concentrations in the SML (assuming that the metal in the total fraction is highly influenced by material from aerosol dust) and metal aerosol fluxes. Since Mo and Cd are not enriched in the SML they were not considered in this calculation. Although highly variable among stations, the average residence times (± SD) of Cu (5.8 ± 6.2 h), V (2.2 ± 1.0 h), Pb (1.8 ± 3.4 h), Zn (1.5 ± 1.2 h), Co (1.2 ± 0.7 h), Ni (48 ± 18 min), and Fe (3.6 ± 6 min) were consistent with previous estimates in regions under low aerosol inputs (Ebling and Landing, 2017) (Table 4). The residence time calculation of Fe using the same deposition rate than the rest of the metals (i.e. 0.1 $cm.s^{-1}$) results in a residence time 10 times longer (i.e. average 0.6 h instead 0.06 h), even so, the residence time of Fe remain being the shortest of all studied metals. The residence time of Cu (St 3-4 and Fast 3-4), Fe (St 3 and Fast-3), and Pb (St 4), were an order of magnitude higher than other stations. Our results indicate that Fast 1-4 stations were affected by the dusty rain events, which increased the concentration of some metals in the T-SML and consequently the residence time (Table 4). However, the reasons for the increase at stations St 3 and St 4 are not evident. On average, while the highest residence times obtained for Cu and Pb are in agreement with their strong affinity to particles and therefore with a high probability of retention in the SML, other reactive elements such as Fe presented the shortest residence times. Since such a quick transfer of these metal particles to the underlying water (in the order minutes) is unlikely (mainly due to the high content of organic matter in the SML), and the dissolution is not immediately reflected in an increase in the concentration in the dissolved fraction (i.e. D-SML), other variables (linked to physical processes, photochemistry or biological activity) probably affected the residence time of metals in the SML. Wind speed seems not to have affected the residence time of any metal in the SML (Table 3), which is probably due to the low speed registered during our campaign (9 ± 4.99 knots) (Table 1).

3.2.3. Neuston composition

The abundance of different microbial groups (Bacteria; High nucleic acid-content bacteria: HNA; Low nucleic acid-content bacteria: LNA; pico-phytoplankton) in the SML is shown in Table 2. Bacterial abundance in the SML ranged from $2x10^5$ to $1x10^6$ cell mL^{-1} (average ± SD: $5.1x10^5 ± 2.2x10^5$ cell mL⁻

[1]), which is of the same order of magnitude as the abundance measured in the SML of coastal Mediterranean Sea (Joux et al., 2006; Obernosterer et al., 2005) and of other regions (e.g. along the Peruvian Coast with an average of $8.9x10^5 \pm 4.3x10^5$ cell mL^{-1}) (Agogue et al., 2005; Engel and Galgani, 2016; Joux et al., 2006; Zäncker et al., 2018). The bacterial community was dominated by low nucleic acid-content bacteria (LNA) with an average abundance ($\pm$ SD) of $2.8 x10^5 \pm 1.0x10^5$ cell mL^{-1}. Bacterial abundances did not differ significantly between SML and SSW (Table 2) in the line of previous studies in coastal Mediterranean Sea that report that the SML was only weakly enriched in heterotrophic bacteria (on average 1.1-fold in Joux et al., 2006 and mean 1.3 in Obernosterer et al., 2005), and comparable to ratio in other environments, such as the Pacific, (Obernosterer et al., 2008). Phytoplankton was only slightly, but significantly (t-test, p=0.002, n=12) enriched in the SML with an average enrichment of 1.5 compared to the SSW; such enrichment were also observed in coastal sites of the Mediterranean Sea (Joux et al., 2006).

Microbial abundance decreases from west to east, reflecting the increasing oligotrophy (mostly due to P limitation) of the surface of Mediterranean waters (Pulido-Villena et al., 2012). Microbial abundance in the SML and reactive elements (i.e. Cu, Fe, and Zn) in the T-SML showed the same longitudinal gradients in this study, with a decreasing eastward concentration along the southern coast of the MS. In fact, bacterial abundances were significantly and positively correlated with these bioactive T-SML metals (i.e. r_s: Cu: 0.65 p<0.01; Fe: 0.53 p<0.05; and Zn: 0.49 p<0.05), suggesting that bacterioneuston could affect the concentration and fate of Cu, Fe and Zn in the SML. Bacteria could efficiently assimilate the fraction of Cu, Fe and Zn available, favouring a decrease in the D-SML fraction (Table 1-2). No general relationship between the concentrations of metals and TEPs (high molecular weight polymers released by phytoplankton and bacteria and with a high metal binding capacity (Passow, 2002)) were found in the SML (Table 5). Metal assimilation by microbial communities could explain the higher residence time of Cu and Zn (in the order of hours) in the SML, although information about the metal content in seston would be necessary to corroborate this hypothesis. However, in the case of Fe with an estimated residence time of a few minutes, other processes in addition to wind speed and neuston uptake, should be contributing to facilitating the transfer from the SML to the underlying water. For example, photochemical reactions driven by exposure to intense solar radiation in the SML could play an

important role in the dissolution processes of this metal (Boyd et al., 2010). On the other hand, Ni was strongly and negatively correlated with bacterial abundance in the D-SML (r_s = -0.93, p<0.01; R^2 = 0.74, p<0.01) suggesting, contrarily to Cu, Fe and Zn, a possible inhibiting role on the microbial growth (Table 5 and Figure 2) (see next section for more discussion).

3.2.4. Subsurface water

The D-SSW concentrations of Cd, Co, Cu, Ni, Mo and Zn showed a longitudinal gradient of concentrations increasing from west to east, with significant positive correlations with longitude for Cd (r^2: 0.60; p<0.05), Co (r^2: 0.77; p<0.01) and Ni (r^2: 0.80; p<0.01) (Figure 3). This trend is consistent with previous studies where the increasing eastward trend in concentration along the southern coast of the MS has been suggested to result from several factors such as: more intense Saharan deposition on the eastern MS (Guieu et al., 2002); more rapid exchange of water masses and margin inputs in the western part (Yoon et al., 1999) or, as suggested for Co, the eastward regeneration of biogenic particulates that yields a decrease towards the west of the dissolved Co in surface (Dulaquais et al., 2017). Since surface salinity showed the same eastward increase and was closely correlated with those metals (r_s ranged from 0.51 p<0.05 for Mo to 0.97 p<0.01 for Ni; Table 3), the exchange with the surface Atlantic Ocean waters seems to be the main cause of this gradient of concentrations in our study. Other metals (i.e. Fe, Pb and V) did not show any clear geographical trend and therefore the variations in their surface concentrations could be influenced by factors other than dilution or exchange, such as vertical diffusive fluxes or specific metal sources, as in the case of Fe and Pb, which have been suggested to be more affected by atmospheric inputs (Nicolas et al., 1994; Yoon et al., 1999). In fact, Pb was the only element that showed significant positive correlation with latitude (r_s: 0.88 p<0.01, Table 3) suggesting an influence of the more heavily industrialised northern region of the MS.

D-SSW concentrations of Ni were strongly and negatively correlated with microbial abundance (mainly with heterotrophic bacteria) in the underlying water (r_s: -0.91, p<0.01; R^2 = 0.87, p<0.01) (Figure 2 and Table 5), which could be interpreted as an indication of a potential negative role of this metal on bacteria and small phytoplankton in the top metre of the western surface of the MS, including the SML. The toxicity of Ni at concentrations of ~50 nM has been previously demonstrated in the western MS, with

inhibitions of 10% (EC_{10}) in phycoerythrin and Chl-a signals of a natural population of the picoplankton *Synechococcus* sp. (Debelius et al., 2011). Although that toxicity concentration (tested in picoplankton) is around 13 times higher than the average values (± SD) measured in our samples (T-SML: 4.1 ± 0.5 nM, D-SML: 3.9 ± 0.6 nM and D-SSW: 3.2 ± 0.6 nM; Table 1), deleterious effects on the neuston and microbial communities at lower concentrations might be possible in the top metre of the sea surface. The toxicity to phytoplankton of divalent, cationic trace metals, such as Ni or Cu, is probably controlled by its free metal ion concentration (Donat et al., 1994). Although the Ni interactions with dissolved organic matter have not been studied well in seawater, they are thought to occur partly as stable organic complexes and with slow dissociation rates (Wen et al., 2011). However, intense UV radiation can alter the concentration, structure, reactivity and metal binding capacity of the organic matter, thus increasing the proportion of free metal ions and their bioavailability and/or potential toxicity (Cheloni and Slaveykova, 2018). Even if a general decreasing trend in microbial abundance from west to east due to the increasing oligotrophy was observed, it is pertinent to mention that primary production and Chl-a concentration (measured a 5 m depth) did not show any significant correlations with Ni (Table 5). It is therefore assumed that the potential toxicity of Ni would mainly affecting the bacterial community in the top metre of the oceans' surface. Nickel, like other transition metals, is an essential cofactor of several enzymes; however, it becomes toxic when homeostasis fails. Multiple potential mechanisms of Ni toxicity to aquatic organisms, and in particular to bacteria, have been identified (Macomber and Hausinger, 2016). Among the different possible toxicity mechanisms (including the inhibition of Zn and Fe metalloenzymes and non-metalloenzymes) the toxicity involving reactive oxygen species (ROS) is the most probably in surface seawater. While Ni itself is a poor generator of ROS when compared to other metals like Fe or Cu, its reactivity and ROS production can be enhanced by the displacement of redox-active iron from iron metallocenters (Macomber and Hausinger, 2016) or when chelated by oligopeptides and histidine (Brix et al., 2017), which are abundant in the SML. It appears that nickel-dependent toxicity involving ROS could be a mechanism of oxidative stress in microbial organisms of the surface of oceans.

4. Conclusions

Our results show that the SML in the MS is enriched by trace metals relative to the SSW, even under low aerosol deposition rates. Residence times of particulate metals derived from aerosols deposition were highly variable, with the largest residence times for Cu (5.8 ± 6.2 h) and the lowest for Fe (3.6 ± 6.0 min). A strong negative correlation between the Ni concentration and heterotrophic bacterial abundance in the SML and SSW could be suggestive of an inhibiting role of this element on the microbial growth in the top metre of the surface; however, further research is needed to confirm this finding.

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35

Figure 1. Location of the sampling stations during the PAEACETIME cruise (May 10th to June 11th, 2017).

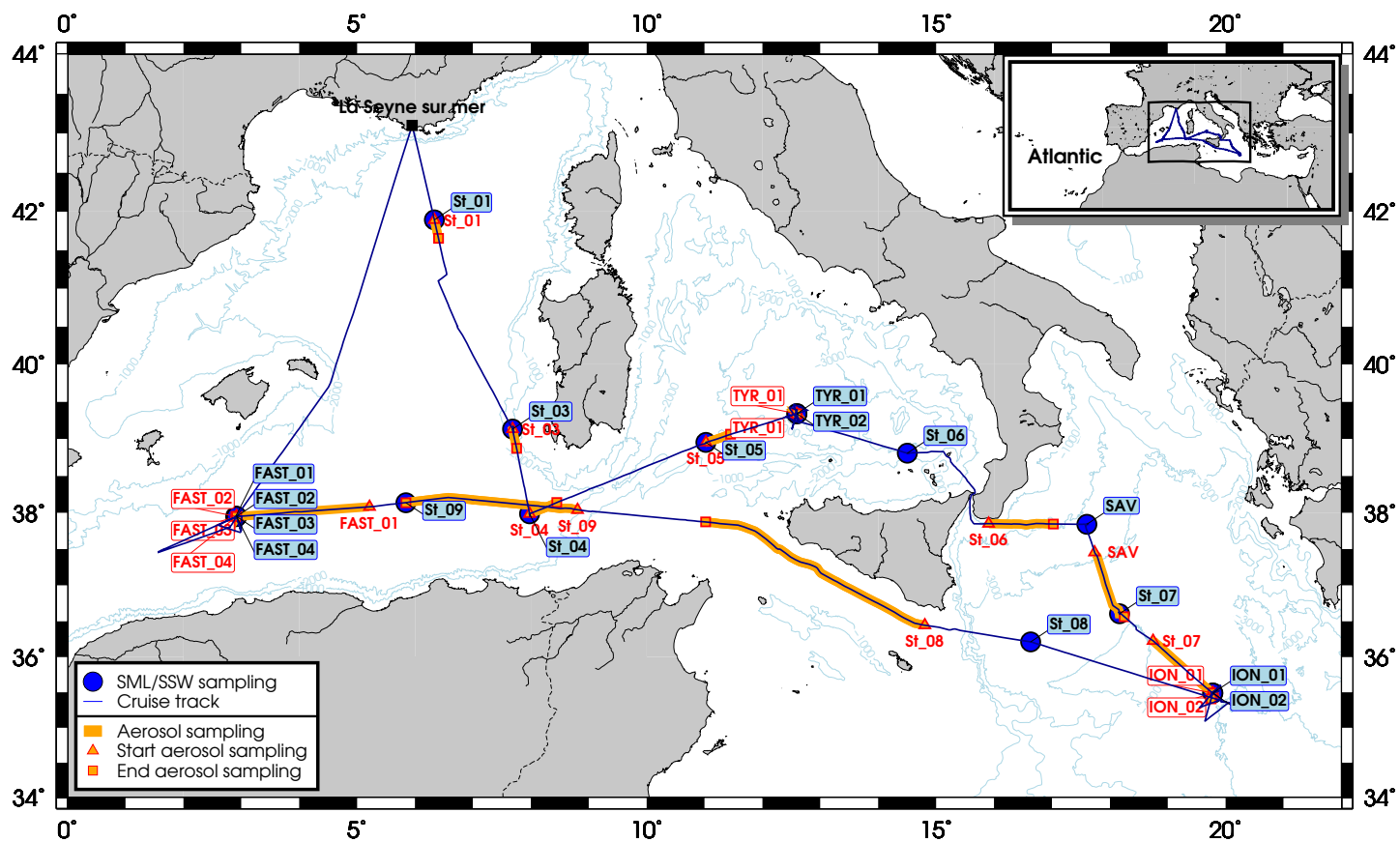
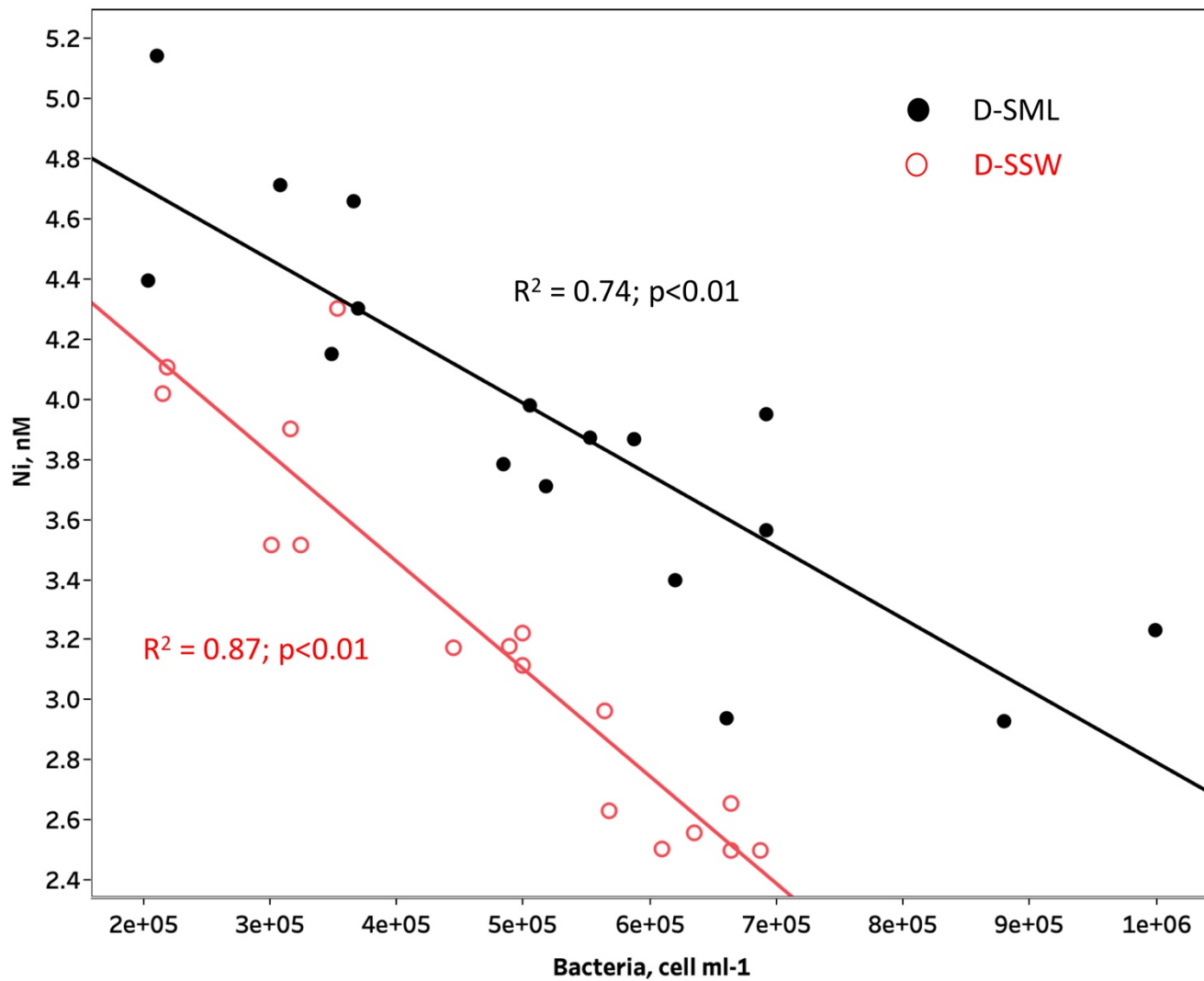



Figure 2. Concentration of dissolved Ni_plotted against bacterial abundance in both the surface microlayer (SML, black dots) and the subsurface water (SSW, red open dots). The lines represent least square linear regressions.

figure

Figure 3. Salinity and concentration of dissolved trace metals in the subsurface waters (SSW) plotted against longitude. The dashed lines represent least square linear regressions.

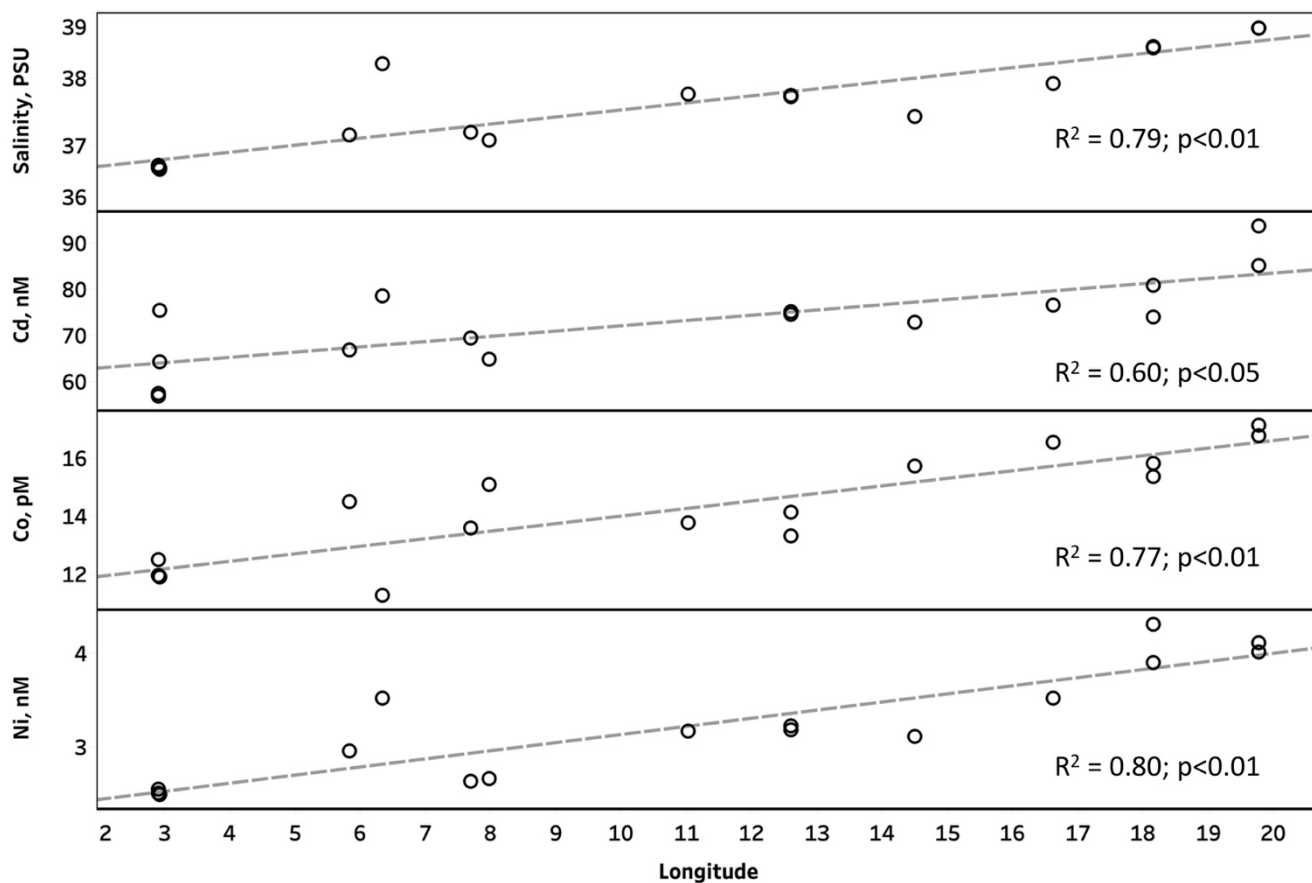



Table 1. Arcillary parameters and metals in water and aerosols measured at all stations. Metals in ng.m^{-3} for aerosols and nM or pM for water compartments.

Station	Date	Time	Lat	Long	Wind Speed	Salinity 5m depth	Temp 5m depth	Sample	Cd	Co	Cu	Fe	Mo	Ni	Pb	V	Zn
		UTC			knots	PSU	ºC		ng·m^{-3} - nM	ng·m^{-3} - pM	ng·m^{-3} - nM	ng·m^{-3} - nM	ng·m^{-3} - nM	ng·m^{-3} - nM	ng·m^{-3} - pM	ng·m^{-3} - nM	ng·m^{-3} - nM
St. 1	12/5/17	13:15	41.8918	6.3333	18.8	38.2	16.4	Aerosol	0.1	0.1	2.0	37.9	0.4	2.7	1.1	8.0	3.6
								D-SML	74.0	109.8	8.0	6.8	125.9	4.1	323.0	14.9	8.0
								T-SML	76.3	112.6	3.6	81.8	121.0	4.5	1217.5	21.7	11.1
								SSW	78.5	11.3	1.6	14.3	130.1	3.5	111.0	23.0	1.9
St. 3	14/5/17	5:30	39.1333	7.6835	5.6	37.2	18.7	Aerosol	0.0	0.1	0.4	22.9	0.0	2.4	0.3	6.5	3.3
								D-SML	62.7	159.6	5.8	3.7	132.5	4.0	766.0	10.6	8.5
								T-SML	72.5	163.1	7.5	72.2	124.9	4.0	858.2	25.4	12.4
								SSW	69.3	13.6	1.2	2.3	124.3	2.6	83.6	23.8	0.6
St. 4	15/5/17	7:30	37.9832	7.9768	6.8	37.1	19.8	Aerosol	0.0	0.1	0.5	27.3	0.1	2.9	0.2	6.8	2.6
								D-SML	57.0	143.3	5.5	1.5	121.7	3.6	601.5	15.2	14.7
								T-SML	59.7	172.0	12.5	32.6	122.4	4.0	881.3	27.1	12.6
								SSW	65.0	15.1	7.2	5.4	125.0	2.7	87.4	18.4	0.7
St. 5	16/5/01	6:00	38.9532	11.023	15.5	37.8	19.9	Aerosol	0.0	0.1	0.6	57.3	0.4	1.9	0.7	3.9	2.6
								D-SML	66.0	143.0	2.6	0.6	122.6	4.0	212.7	35.0	4.7
								T-SML	64.4	152.9	4.8	37.4	128.6	3.9	559.3	31.1	11.1
								SSW	n.m	13.8	1.8	2.0	130.7	3.2	101.2	18.7	1.2
TYR-1	16/5/01	8:00	39.34	12.593	6.6	37.8	20.3	Aerosol	0.1	0.1	4.8	115.3	0.3	3.1	1.9	5.6	7.2
								D-SML	59.9	149.7	4.3	4.4	74.8	3.8	293.4	32.2	6.7
								T-SML	59.1	145.3	11.7	55.2	119.1	3.8	488.3	24.6	7.6
								SSW	75.1	13.3	1.7	2.1	122.9	3.2	90.1	19.4	1.0
TYR-2	18/5/17	8:00	39.3398	12.593	4.9	37.7	21.1	Aerosol	0.1	0.1	2.3	162.3	0.1	2.0	3.9	3.9	8.6
								D-SML	65.7	143.6	7.9	1.1	137.7	3.9	316.3	29.8	7.7
								T-SML	79.9	160.6	5.3	82.5	124.9	3.6	435.2	24.4	7.9
								SSW	74.5	14.1	2.0	2.1	130.0	3.2	90.7	18.6	1.1
St. 6	22/5/17	6:00	38.8077	14.5	10.1	37.4	20.4	Aerosol	0.1	0.1	4.8	189.8	1.2	2.2	2.7	2.9	11.3
								D-SML	55.5	169.7	2.7	13.7	131.7	3.7	281.7	25.2	4.2
								T-SML	56.8	144.2	4.4	42.0	116.6	3.5	572.2	23.2	9.2
								SSW	72.9	15.7	1.7	2.3	131.2	3.1	86.1	18.4	1.0
SAV	23/5/17	10:00	37.8401	17.602	3.0	38.5	20.1	Aerosol	0.1	0.1	2.1	89.9	2.1	7.1	1.4	18.5	4.9
								D-SML	77.5	158.9	2.9	2.2	129.8	4.7	258.1	25.9	6.6
								T-SML	67.4	171.4	5.0	26.7	132.3	4.9	1028.8	27.1	7.9
								SSW	80.8	15.4	1.6	2.1	129.3	4.3	59.1	20.4	1.1
St. 7	24/5/17	4:00	36.6035	18.166	4.8	38.5	20.8	Aerosol	0.0	0.1	2.5	85.6	0.1	4.2	1.3	8.7	6.9
								D-SML	74.1	176.1	6.0	1.6	129.0	4.7	222.0	24.8	9.7
								T-SML	94.1	187.6	8.3	48.6	121.4	4.6	392.3	25.2	10.7
								SSW	74.0	15.8	1.7	2.9	126.2	3.9	60.1	19.7	1.0
ION-1	25/5/17	8:00	35.4892	19.776	8.1	38.8	21.0	Aerosol	0.2	0.1	1.2	88.8	0.2	7.1	1.8	17.9	11.0
								D-SML	53.4	188.8	3.3	5.4	91.7	5.1	284.6	38.2	9.9
								T-SML	69.8	151.5	6.8	24.7	108.4	4.6	527.9	23.9	9.1
								SSW	93.6	17.1	1.9	2.0	128.5	4.1	61.4	18.7	1.2
ION-2	27/5/17	8:00	35.4892	19.777	12.4	38.8	21.1	Aerosol	0.0	0.0	2.9	80.6	0.1	1.7	2.5	3.8	11.3
								D-SML	13.1	154.9	2.8	4.1	130.4	4.4	74.8	19.0	7.1
								T-SML	13.7	168.1	5.9	30.7	127.1	4.7	74.8	28.3	13.6
								SSW	85.2	16.7	2.3	2.1	128.9	4.0	61.9	22.4	1.1
St. 8	30/5/17	5:00	36.2103	16.631	3.7	37.9	21.2	Aerosol	0.0	0.1	2.4	89.8	0.5	4.4	1.1	11.2	12.5
								D-SML	62.9	171.5	4.4	0.2	137.5	4.3	301.1	15.5	4.8
								T-SML	69.6	163.1	3.5	13.9	124.8	3.8	404.0	29.4	4.1
								SSW	76.7	16.5	1.5	3.3	123.8	3.5	62.8	16.3	1.1
St. 9	31/5/17	16:00	38.1347	5.8408	6.2	37.1	22.0	Aerosol	0.1	0.1	1.8	170.7	0.2	2.9	5.7	4.1	10.2
								D-SML	51.0	145.9	5.4	0.9	135.2	3.9	373.6	22.5	21.1
								T-SML	60.7	149.9	7.9	82.5	120.8	3.6	596.5	21.9	11.6
								SSW	66.9	14.5	1.2	2.0	130.5	3.0	91.8	21.2	1.0
FAST-1	2/6/17	17:00	37.946	2.902	13.8	36.7	21.7	Aerosol	0.1	0.1	1.8	226.9	0.5	5.5	7.0	4.7	14.6
								D-SML	46.6	118.5	3.8	0.1	125.4	3.4	359.1	24.0	11.9
								T-SML	45.0	408.4	17.2	207.4	124.7	4.4	880.5	26.0	19.6
								SSW	57.3	12.0	1.1	2.8	114.3	2.5	74.4	17.5	0.7
FAST-3	4/6/17	6:00	37.947	2.9153	4.9	36.7	21.7	Aerosol	0.1	0.2	0.9	245.3	0.2	4.5	3.4	10.2	6.8
								D-SML	47.7	142.8	5.2	4.3	124.2	3.2	454.6	25.4	8.9
								T-SML	58.5	773.6	20.1	1433.3	101.9	4.5	1294.7	26.1	12.1
								SSW	57.0	12.5	1.5	1.6	120.5	2.6	74.7	19.6	1.1
FAST-4	5/6/17	8:00	37.9467	2.9168	8.7	36.6	21.8	Aerosol	0.2	0.3	1.2	266.0	0.2	6.8	4.3	17.1	8.3
								D-SML	53.6	115.9	4.5	3.4	125.4	2.9	247.3	17.5	4.3
								T-SML	57.5	96.8	17.0	64.1	116.9	3.2	635.4	20.5	20.5
								SSW	64.4	11.9	0.8	4.0	113.0	2.5	73.2	18.4	0.6
FAST-5	7/6/01	5:30	37.9465	2.9167	18.2	36.6	21.9	Aerosol	0.0	0.1	0.7	44.9	0.2	1.6	3.2	4.0	2.0
								D-SML	45.0	122.7	4.6	0.0	130.3	2.9	283.3	18.9	8.3
								T-SML	53.3	122.7	13.6	31.1	127.3	3.4	426.1	28.6	17.8
								SSW	75.5	11.9	1.3	1.8	110.8	2.5	70.9	15.4	0.9

Table 2. Biological parameters measured at all stations (Bacteria; High nucleic acid-content bacteria: HNA; Low nucleic acid-content bacteria: LNA; Pico-phytoplankton; Transparent Exopolymer Particles: TEP; Primary Production: PP; and chlorophyll a: Chl-a). n.m: not measured.

Station	Bacteria	HNA	LNA	Pico-phyto	TEP	PP	Chl-a
	cell·ml^{-1} x10^5	cell·ml^{-1} x10^5	cell·ml^{-1} x10^5	cell·ml^{-1} x10^2	particles·l^{-1} x10^6	mg·C·m^{-3}·d^{-1}	µg·l^{-1}
St. 1							
T-SML	3.49	1.62	1.88	84.71	7.67		
SSW	3.02	1.26	1.78	70.74	4.79	5.70	0.187
St. 3							
T-SML	5.06	1.80	3.30	30.76	8.24		
SSW	5.68	2.10	3.62	21.72	1.50	1.65	0.095
St. 4							
T-SML	6.92	3.58	3.37	31.11	5.97		
SSW	6.65	3.59	3.10	24.35	2.66	1.75	0.090
St. 5							
T-SML	4.27	2.05	2.25	16.76	4.55		
SSW	4.45	2.12	2.36	13.74	2.28	1.68	0.060
TYR-1							
T-SML	4.84	1.33	3.46	16.18	3.55		
SSW	4.89	1.43	3.46	12.88	0.68	1.56	0.063
TYR-2							
T-SML	5.53	2.00	3.57	34.60	15.10		
SSW	5.00	1.87	3.16	17.26	1.84	1.77	0.071
St. 6							
T-SML	5.19	2.17	3.04	20.61	8.44		
SSW	4.99	2.03	3.00	12.78	7.47	1.66	n.m
SAV							
T-SML	3.66	1.37	2.30	13.76	19.10	n.m	n.m
SSW	3.53	1.30	2.25	12.83	3.34	n.m	n.m
St. 7							
T-SML	3.08	1.28	1.81	20.38	8.36	n.m	n.m
SSW	3.16	1.32	1.85	15.63	2.96	0.90	0.056
ION-1							
T-SML	2.11	0.81	1.30	17.08	26.55		
SSW	2.19	0.88	1.32	14.29	8.20	1.84	n.m
ION-2							
T-SML	2.04	0.94	1.10	19.14	13.27		
SSW	2.16	0.96	1.21	9.50	7.16	1.79	0.063
St. 8							
T-SML	3.70	1.62	2.10	30.59	26.13		
SSW	3.24	1.48	1.77	11.64	4.65	1.64	0.070
St. 9							
T-SML	5.88	3.01	2.91	33.82	37.07		
SSW	5.65	3.18	2.51	16.70	10.57	2.64	0.072
FAST-1							
T-SML	6.20	3.55	2.68	17.11	6.93		
SSW	6.09	3.76	2.38	14.53	1.27	2.68	0.070
FAST-3							
T-SML	9.99	5.42	4.59	44.31	37.29		
SSW	6.35	3.77	2.62	18.35	2.16	2.44	0.085
FAST-4							
T-SML	8.80	4.01	4.83	44.89	n.m		
SSW	6.87	3.41	3.49	43.40	n.m	2.85	0.081
FAST-5							
T-SML	6.61	3.22	3.41	25.04	3.14		
SSW	6.64	3.57	3.10	23.54	2.88	2.04	0.078

Table 3. Spearman's rank correlation coefficients for selected variables. Significant correlations at p<0.05 and p<0.01 are marked with one asterisk (orange numbers) and two asterisks (red numbers), respectively.

	Variable	Latitude	Longitude	Salinity	Wind Speed	T-SML									D-SML								
						Cd	Co	Cu	Fe	Mo	Ni	Pb	V	Zn	Cd	Co	Cu	Fe	Mo	Ni	Pb	V	Zn
	Latitude	1.00	-0.393	-0.286	0.195	0.221	-0.406	-0.112	.562*	-0.088	-0.422	0.361	-0.362	-0.092	0.318	-0.405	0.373	0.212	-0.005	-0.335	0.394	-0.059	-0.126
	Longitude	-0.393	1.00	.883**	-0.331	0.325	0.105	-.589*	-.692**	0.148	0.434	-.560*	0.136	-.611**	0.349	.802**	-0.315	0.311	0.11	.806**	-.535*	0.325	-0.27
	Salinity	-0.286	.883**	1.00	-0.2	0.455	0.127	-.652**	-.530*	0.161	.670**	-0.362	0.098	-.609**	0.445	.620**	-0.209	0.369	-0.006	.934**	-0.432	0.304	-0.145
D-SSW	Cd	-0.319	.723**	.796**	0.032	0.232	-0.25	-.682**	-.715**	0.347	0.397	-0.435	0.194	-0.494	0.229	0.429	-0.265	0.226	0.124	.721**	-0.491	0.185	-0.3
	Co	-.575*	.857**	.672**	-0.427	0.186	0.338	-0.426	-.630**	0.054	0.39	-.495*	0.228	-0.478	0.086	.870**	-0.346	0.103	0.196	.685**	-0.301	0.277	0.015
	Cu	-0.005	.660**	.568*	0.002	0.152	0.164	-0.35	-0.414	0.093	0.314	-0.287	0.206	-0.368	0.181	0.343	-0.181	0.294	-0.201	0.38	-0.255	0.395	-0.069
	Fe	0.171	0.002	0.007	0.012	0.226	-0.072	-0.184	0.058	-0.08	-0.048	0.128	-0.318	0.055	0.387	-0.112	0.372	0.067	0.088	-0.006	0.135	-.589*	-0.105
	Mo	0.278	0.446	.507*	0.036	0.348	-0.145	-.728**	-0.069	0.086	0.162	-0.002	-0.179	-0.419	0.412	0.277	-0.147	0.282	0.257	.483*	-0.191	0.27	-0.164
	Ni	-0.307	.883**	.970**	-0.341	.486*	0.157	-.661**	-.532*	0.163	.634**	-0.327	0.086	-.699**	.487*	.630**	-0.178	0.303	0.074	.912**	-0.396	0.352	-0.14
	Pb	.884**	-0.435	-0.298	0.364	0.096	-0.365	-0.137	.547*	-0.125	-0.424	0.306	-0.287	0.017	0.115	-.485*	0.275	0.056	-0.029	-0.349	0.407	-0.064	0.012
	V	0.228	0.231	0.459	-0.153	0.359	0.058	-0.221	0.234	0.015	.525*	0.197	-0.248	-0.094	0.262	0.173	0.208	.511*	0.093	.518*	0.047	-0.043	0.131
	Zn	0.007	0.446	.682**	-0.026	0.469	-0.034	-.677**	-0.24	0.139	0.395	-0.098	0.107	-.580*	0.404	0.152	-0.093	0.148	0.044	.621**	-0.24	0.425	-0.137
Residence Time	Co	-.651**	0.233	0.188	-0.405	-0.049	.848**	0.213	-0.179	0.078	.566*	-0.159	.515*	0.066	-0.252	0.382	0.022	-0.162	0.064	0.264	0.137	-0.066	0.441
	Cu	-0.069	-.537*	-.595*	-0.028	-0.132	0.279	.779**	0.218	-0.098	-0.074	0.35	0.145	.551*	-0.292	-0.277	0.206	-0.275	-0.439	-0.457	0.395	-0.14	0.478
	Fe	0.448	-0.456	-0.234	-0.013	0.272	0.374	0.408	.606**	-0.017	0.262	0.459	-0.013	0.251	0.129	-0.356	.650**	0.026	-0.24	-0.161	.592*	-0.191	.560*
	Ni	0.348	0.2	0.095	0.116	0.037	0.034	-0.154	-0.054	0.417	-0.066	-0.348	0.311	0.032	0.01	0.069	0.179	0.005	0.24	0.029	0.012	-0.118	-0.025
	Pb	0.24	0.08	0.18	-0.272	.514*	0.186	-0.24	-0.185	0.071	0.326	.509*	0.175	-0.28	.625**	0.118	0.186	0.232	-0.286	0.245	0.3	-0.148	-0.025
	V	0.359	0.118	-0.048	0.204	-0.228	-0.061	-0.174	0.069	0.297	-0.353	-0.446	0.213	-0.066	-0.15	0.056	-0.147	-0.154	0.311	-0.157	-0.098	0.176	-0.142
	Zn	0.331	-0.386	-0.354	0.218	0.056	-0.125	0.324	0.071	0.301	-0.076	0.304	0.181	0.441	0.132	-0.475	0.338	-0.113	-0.272	-0.311	0.091	-0.309	0.066

Table 4. Number of dips conducted to collect 500mL of SML, thickness (μm) of the SML and calculated residence times (hours) of particulate metals in the SML derived for aerosol depositions.

Station	Dips	Thickness	Co	Cu	Fe	Ni	Pb	V	Zn
St. 1	100	26	0.5	0.8	0.09	0.7	1.6	1.0	1.5
St. 3	60	43	1.2	16.0	0.21	1.2	7.7	2.4	3.0
St. 4	60	43	1.5	19.6	0.08	1.0	13.1	2.4	3.8
St. 5	100	26	0.7	3.4	0.03	0.9	1.2	2.9	2.0
TYR-1	60	43	0.7	1.9	0.03	0.8	0.7	2.7	0.8
TYR-2	60	43	1.0	1.8	0.03	1.3	0.3	3.8	0.7
St. 6	80	32	0.7	0.5	0.01	0.8	0.4	3.6	0.5
SAV	60	43	1.1	1.8	0.02	0.5	1.8	0.9	1.3
St. 7	60	43	1.7	2.5	0.04	0.8	0.7	1.8	1.2
ION-1	60	43	1.2	4.2	0.02	0.4	0.7	0.8	0.6
ION-2	80	32	1.8	1.1	0.02	1.5	0.1	3.4	0.7
St. 8	60	43	1.6	1.1	0.01	0.6	0.9	1.6	0.3
St. 9	80	32	0.9	2.5	0.02	0.7	0.2	2.4	0.7
FAST-1	100	26	1.3	4.4	0.04	0.3	0.2	2.0	0.6
FAST-3	60	43	3.5	16.2	0.39	0.7	1.0	1.5	1.4
FAST-4	60	43	0.2	10.7	0.02	0.3	0.4	0.7	1.9
FAST-5	100	26	1.0	9.3	0.03	0.9	0.2	2.6	4.2
Avergae	72.9	36.8	1.2	5.8	0.06	0.8	1.8	2.2	1.5
S.D.	17.2	7.6	0.7	6.2	0.10	0.3	3.4	1.0	1.2

Table 5. Spearman's rank correlation coefficients for selected parameters. Bacteria, HNA: High nucleic acid-content bacteria; LNA: Low nucleic acid-content bacteria; Pico-Phyto: Pico-Phytoplankton; Transparent Exopolymer Particles: TEP; Primary Production: PP; and chlorophyll a: Chl-a; . Significant correlations at p<0.05 and p<0.01 are marked with one asterisk (orange numbers) and two asterisks (red numbers), respectively.

	Variable	SML					SSW						
		Bacteria	HNA	LNA	Pico-Phyto	TEP	Bacteria	HNA	LNA	Pico-Phyto	TEP	PP	Chl-a
	Longitude	-,815**	-,842**	-,619**	-,485*	0.249	-,785**	-,831**	-,516*	-,667**	0.416	-,630**	-,585*
	Temperature	0.427	0.377	0.265	0.191	0.377	0.341	0.394	0.011	-0.012	0.144	0.42	-0.132
	Salinity	-,958**	-,927**	-,797**	-0.453	0.238	-,957**	-,927**	-,713**	-,565*	0.422	-0.412	-0.475
	Wind Speed	0.049	0.183	-0.109	0.031	-,578*	0.054	0.114	-0.054	0.232	0.078	,521*	0.187
T-SML	Cd	-0.407	-0.43	-0.238	0.161	0.203	-0.377	-0.439	-0.11	0.186	0.044	-0.315	0.024
	Co	-0.012	0.009	-0.174	-0.229	0.285	-0.029	0.176	-0.245	-0.243	-0.326	-0.265	-0.222
	Cu	,654**	,538*	,542*	0.12	-0.182	,684**	,650**	0.453	0.441	-0.468	0.3	0.191
	Fe	,527*	,515*	,488*	,493*	0.024	0.449	,502*	0.449	,493*	-0.421	0.424	0.371
	Ni	-,610**	-,530*	-,669**	-0.347	0.268	-,600*	-0.48	-,667**	-0.248	0.15	-0.044	-0.116
	Mo	-0.238	-0.179	-0.228	-0.353	-0.344	-0.115	-0.088	-0.103	-0.186	-0.191	-0.206	-0.288
	Pb	0.436	,514*	0.319	0.307	0.106	0.407	0.397	0.245	0.478	-0.138	,524*	,754**
	V	-0.096	-0.039	-0.201	-0.45	-0.229	-0.066	0.135	-0.257	-0.387	-0.265	-0.371	-0.349
	Zn	,493*	,579*	0.225	0.34	-0.306	,561*	,569*	0.245	,527*	-0.126	,594*	0.371
D-SML	Cd	-0.385	-0.354	-0.174	-0.085	-0.038	-0.338	-0.397	-0.047	-0.005	-0.082	-0.465	-0.108
	Co	-,583*	-,667**	-0.458	-0.444	0.412	-,539*	-,559*	-0.309	-,627**	0.376	-,732**	-0.495
	Cu	0.221	0.151	0.26	,712**	0.021	0.233	0.123	0.299	,738**	-0.182	0.112	,547*
	Fe	-0.301	-0.327	-0.078	0.08	0.221	-0.346	-0.471	-0.044	-0.047	0.259	-0.029	0.279
	Ni	-,926**	-,888**	-,826**	-0.383	0.356	-,889**	-,853**	-,690**	-0.462	0.453	-0.393	-0.4
	Mo	-0.056	-0.042	-0.034	0.311	0.359	-0.061	-0.088	0.005	-0.137	0.253	-0.079	-0.002
	Pb	0.48	0.41	0.375	0.472	0.121	0.451	0.449	0.385	0.475	-0.315	0.182	,732**
	V	-0.191	-0.265	-0.037	-,574*	0.174	-0.277	-0.208	-0.152	-0.468	-0.076	-0.182	-,670**
	Zn	0.154	0.109	-0.093	0.151	0.129	0.164	0.238	-0.061	0.377	-0.003	0.232	0.275
D-SSW	Cd	-,826**	-,837**	-,638**	-0.421	0.068	-,800**	-,882**	-,579*	-0.429	0.443	-0.211	-0.28
	Co	-,586*	-,591*	-,591*	-0.418	0.491	-,586*	-,532*	-,537*	-,694**	,568*	-,541*	-,538*
	Cu	-0.412	-0.424	-0.284	-0.28	-0.059	-0.439	-0.434	-0.252	-0.324	0.126	-0.376	-0.279
	Fe	-0.022	0.042	-0.052	0.339	-0.235	0.036	-0.064	0.092	0.251	-0.049	-0.026	0.275
	Ni	-,888**	-,876**	-,721**	-0.422	0.365	-,907**	-,899**	-,697**	-,576*	0.462	-0.389	-0.497
	Mo	-0.456	-0.326	-0.429	-0.085	0.226	-,515*	-0.439	-0.326	-0.301	,547*	-0.144	-0.191
	Pb	0.306	0.348	0.321	0.389	-0.285	0.23	0.282	0.38	0.355	-0.176	0.182	0.354
	V	-0.467	-0.443	-0.316	0.04	0.258	-0.448	-0.464	-0.173	0.001	0.163	-0.026	0.143
	Zn	-,602*	-,502*	-,534*	-0.113	0.365	-,726**	-,591*	-,664**	-0.287	0.421	0.024	-0.251