

Horta, 9 March 2020

Subject: Revision of MS No.: bg-2019-304

Dear Associate editor,

We understand the recurrent issue with the transect lengths vs. the surface estimations, also for comparison purposes with other studies.

Since laser pointers were 6.5 cm apart, these were not visible when altitudes were higher than 2m. We decided to forego the attempts to quantify the actual or exact amount of surface covered and estimated the approximate area covered by using the ROV altitude, time spent at altitude and distance travelled along with predefined image widths (in concordance to Vanreusel et al. (2m at 1m altitude and 4m at 2m altitude, and extrapolated from there)). The methodology for this was added in L126-136. Main results did not change significantly (except for a significant difference between Porifera densities between seamounts and nodule fields L229-230) and main tendencies were withheld.

Figures 3, 6 and 7 and Table A1 were altered accordingly and previous comments regarding figure 3 by this reviewer were re-considered and tests re-done (see below). This has some implications in table and figure numeration which are explained in our detailed comments below.

Sincerely,



Daphne Cuvelier, on behalf of all co-authors

Evaluation of bg-2019-304-manuscript-version3

Methods

R1: Lines 96 ff: I agree that the method for quantification of the samples may be suitable for the comparisons made in this study, provided that the variations in altitude and camera angle were comparable between transects (but I guess that keeping the target altitude on the flat plain was much easier than at the seamounts?). My only concern is the statement that "altitudes >10 m were omitted", meaning that, the other way round, all altitudes between <2 m and 10 m were considered? Since the area correlates with the square of the distance, the area observed could vary by a factor of >25 between sections, which would introduce a substantial bias. Further, although the standardisation to section transect length is perfectly suitable for the comparisons within the study, it would be helpful to include some information on the approximate field of view of the camera, e.g. at the target altitude with a standard pan and tilt setting, in order to get at least some feeling when comparing this study with others.

A: We understand the recurrent issue with the transect lengths vs. the surface estimations, also for comparison purposes, and decided to estimate the approximate surface covered, as explained

above. The bias caused by not taking into account the camera's zoom and pan and tilt is recognised in L137-138

Figures 3, 6 and 7 were altered accordingly and previous comments regarding figure 3 by the reviewer were re-considered and tests re-done (see below).

R1: The authors should also include the information given in their rebuttal, that specimens samples were also used for identifications when possible.

A: This was added in L107-108

Results, Discussion

R1: Line 202: delete "#".

A: # was replaced by "n=". This was done for all other cases in the ms as well.

R1: Lines 252ff (original 220): the modified statement is still not correct. The majority of the ophiuroids were not morphospecies 5, but Ophiuroidea indet.

A: This sentence was rephrased to: "Three Ophiuroidea morphospecies were present at both seamounts and nodule fields (Fig. 2, 3 and 6). Most of the Ophiuroidea observed at the CCZ seamounts that could be identified to morphospecies level, were small and situated on hard substrata (morphospecies 5), while those at nodule fields (including morphospecies 6) were observed on the soft sediments."

R1: Line 333: should read "...showed that an asymptote was reached neither"

A: Change carried out

R1: Line 339: should read "1000 m"

A: Ok

Figures and tables

R1: Fig. 3: I am still not quite happy with this figure, because rare taxa are hardly comparable, despite modifying the axes, and I still suggest to sum up the less abundant morphospecies.

Apart from this, the x-axis labels are incomplete; should read: "Density (ind/100 m)". Negative density values for the seamounts are strange; the minus sign should be deleted. In panel (B), the scaling of the x-axis is not clear; only "-12" is given for the left hand part. I suggest to include a finer scaling here, such as 0.5, 1.0, 1.5 etc. Similar in panel (c). Caption: what does #4 and #5 mean - number of video transects? Rather write "4 transects" or "N=4".

A: Taking these comments into account, we decided to include figure A1 in the ms instead of figure 3 which is now an appended figure. We have added the minimum number of morphospecies to the new figure 3 as well, to facilitate discussing the results. And Table A1 passes now to be Table 2 in the ms. We still believe the back-to-back histogram figure holds valuable information, which is why it is still included in the appendix, though we understand the reviewer's point regarding the rare morphospecies. Figure references have been changed accordingly.

Axes and captions were altered as requested by the reviewer

R1: Fig. 4: Y-axis labels. In panels a and c: "Number of species" (with capital N). And what does "Species" in panels b and c mean? I guess it should also be "Number of species"?

A: change was carried out

R1: Fig. 6: my previous comment is still valid, the y-axis label is incomplete and should comprise quantity and unit; in this case: "Density (ind/100 m)"

A: [Change was carried out](#)

R1: Tab. A1: include sums for higher taxa; e.g., Cnidaria, Anthozoa, Ceriantharia etc.

[Because of the presence of the new figure 3 and the readability and interpretability of the table, we did not include the sums on higher taxa levels in its current lay-out. It would be confusing to use sums and densities across the different taxonomic levels in the table.](#)

1 Are seamounts refuge areas for fauna from polymetallic 2 nodule fields?

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19 Running title: Seamounts as refuge areas for nodule fauna

20 Six keywords: megafauna, seamounts, nodule fields, image analysis, deep sea, mining

21 Abstract

22 Seamounts are abundant and prominent features on the deep-sea floor and intersperse with the
23 nodule fields of the Clarion-Clipperton Fracture Zone (CCZ). There is a particular interest in
24 characterising the fauna inhabiting seamounts in the CCZ because they are the only other ecosystem
25 in the region to provide hard substrata besides the abundant nodules on the soft sediment abyssal
26 plains. It has been hypothesised that seamounts could provide refuge for organisms during deep-sea
27 mining actions or that they could play a role in the (re-)colonisation of the disturbed nodule fields.
28 This hypothesis is tested by analysing video transects in both ecosystems, assessing megafauna
29 composition and abundance.

30 Nine video transects (ROV dives) from two different license areas and one Area of Particular
31 Environmental Interest in the eastern CCZ were analysed. Four of these transects were carried out as
32 exploratory dives on four different seamounts in order to gain first insights in megafauna
33 composition. The five other dives were carried out in the neighbouring nodule fields in the same
34 areas. Variation in community composition observed among and along the video transects was high,
35 with little morphospecies overlap on intra-ecosystem transects. Despite the observation of
36 considerable faunal variations within each ecosystem, differences between seamounts and nodule

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37 fields prevailed, showing significantly different species associations characterising them, thus
38 questioning their use as a possible refuge area.

39 1. Introduction

40 Seamounts are abundant and prominent features on the deep-sea floor (Wessel et al., 2010). They
41 are common in all the world's oceans, occurring in higher abundances around mid-ocean ridges,
42 island-arc convergent areas, and above upwelling mantle plumes (Kitchingman et al., 2007).
43 Seamounts are defined as isolated sub-surface topographic feature, usually of volcanic origin, of
44 significant height above the seafloor (International Seabed Authority (ISA), 2019). They are generally
45 isolated, typically cone shaped undersea mountains rising relatively steeply at least several hundred
46 meters from the deep-sea floor. Seamounts comprise a unique deep-sea environment, characterised
47 by substantially enhanced currents and a fauna that is dominated by suspension feeders, such as
48 corals (Rogers, 2018). They represent hard substrata in the otherwise soft sediment deep sea and
49 can thus be considered habitat islands (Beaulieu, 2001). Given the growing evidence that seamounts
50 differ substantially across a range of spatial scales, the concept of seamounts as a single, relatively
51 well-defined habitat type is outdated (Clark et al., 2012). Depth and substrate type are key elements
52 in determining the composition and distribution of benthic fauna on seamounts, while location is
53 likely the subsequent most important driver of faunal composition and distribution patterns (e.g.
54 Tittensor et al., 2009). Connectivity varies substantially between seamounts, resulting in the
55 presence of taxa with very localised to very wide distributions (Clark et al., 2010).

56
57 The Clarion-Clipperton Fracture Zone (CCZ), in the equatorial eastern Pacific Ocean, is most known
58 for its extensive polymetallic nodule fields that will potentially be mined in the future. In this area,
59 nodules represent the most common hard substrate on the soft-sediment abyssal plains, and many
60 organisms rely on them for survival (Vanreusel et al., 2016). Removal of hard substrate through
61 mining actions will impact all these organisms, which were estimated at about 50% of all megafaunal
62 species in the CCZ (Amon et al., 2016). Nodule fields in the CCZ are interspersed by seamounts
63 (Wedding et al., 2013), the only feature offering hard substrata besides the nodules. Based on this
64 feature/characteristic, it has been hypothesised that seamounts could provide refuge for organisms
65 during deep-sea mining activities or that seamounts could play a role in the (re-)colonisation of the
66 disturbed nodule fields. Whether or not this is true may have important implications for
67 management of the impacts of polymetallic nodule mining in the CCZ. However, knowledge on the
68 biodiversity inhabiting seamounts in this region is currently lacking.

69 The objectives of the current study were twofold: (i) Provide first insights in seamount megafauna
70 within the CCZ, (ii) Compare the benthic fauna inhabiting seamounts and nodule fields in the eastern
71 CCZ. Since this is the first time the seamounts at the eastern CCZ were visited, a separate section is
72 dedicated to describe these first insights.

73 2. Material and Methods

74 2.1. Study site and data

75 During the SO239 ECORESPONSE cruise in 2015 (Martinez Arbizu and Haeckel, 2015), four
76 seamounts were visited for the first time within two different license areas and one area of
77 particular environmental interest (APEI) within the Clarion-Clipperton Fracture zone (CCZ) (Table 1).

78 Nodule fields within the same license areas were visited and sampled as well. Video imagery and
79 faunal samples were collected by a Remotely Operated Vehicle (ROV Kiel 6000 (GEOMAR), equipped
80 with a high definition Kongsberg OE14-500 camera).

81 Seamount transects were carried out uphill, towards the summit resulting in a depth gradient along
82 the transect (Table 1). The four seamount transects were characterised by different depth ranges
83 and lengths and were, due to the vessel's positioning and the predominant South-East surface
84 currents, all carried out downstream, on the north to north-western flanks of the seamounts (Table
85 1 and Fig. 1). The names of the seamounts used here, Rüppel and Senckenberg (BGR, German
86 License area), Heip (GSR, Belgian License area) and Mann Borgese (APEI3), are the ones agreed upon
87 by the scientist during the ECORESPONSE cruise (Martinez Arbizu and Haeckel, 2015), pending
88 incorporation of these names in the GEBCO gazetteer. The seamounts differed in shape and size
89 with Senckenberg and Heip being a sea-mountain range, while Rüppel and Mann Borgese were more
90 isolated, stand-alone seamounts (Fig. 1). Nodule field dives were carried out on relatively flat
91 surfaces (maximum depth range covered during a dive or transect was 30m difference, Table 1) and
92 were referred to by the dive number and license area. The five nodule transects were all located
93 between 4000-5000m depth and the transects differed in length between dives as well (Table 1).
94 Within the same license area, distance between different transects was 16 to 60km, while distance
95 between license areas added up to several hundreds of kilometres (minimum ~700kms BGR – GSR,
96 Fig. 1).

97 Investigated areas were restricted to the eastern part of the CCZ with APEI3 being the most north-
98 and westward bound area.

99 The optical resolution of the camera enabled reliable identification of organisms larger than 3 cm
100 (Martinez Arbizu and Haeckel, 2015). The combination of exploration and opportunistic sampling
101 restricted a systematic image collection. Target ROV travelling altitude was <2m and travelling speed
102 was ~0.2m/s which, along with the camera zoom, were kept constant whenever possible. ~~Due to the~~
103 ~~explorative nature of the dives, the pan and tilt of the ROV camera were not kept constant.~~

104 2.2. Video analysis and statistics

105 All videos were annotated to the lowest taxonomic level possible. The number of morphospecies,
106 defined as morphologically different organisms within the lowest taxonomic group identified, were
107 assessed. Identifications were double checked with scientists working in the same area as well as
108 taxonomic experts and comprise different taxonomic levels (e.g. Genus, Family) and organism
109 samples were used for proper identification whenever possible. Those identifications restricted to
110 higher taxon groups (Family, Class, etc.) and for which it was impossible to attribute a
111 morphospecies, were referred to as taxa and are likely to morphologically differ between transects.
112 Xenophyophores, living on the soft sediment deep-sea floor, were less prominently present at
113 seamounts than at nodule fields and were not quantified. Fish (Actinopterygii), Crustacea
114 (Nematocarcinidae, Aristeidae, Peracarida) and Polychaeta were quantified but left out of the
115 comparing statistical analysis due to their lack of representativity and possible attraction due to ROV
116 lights. The same was done for jellyfish and other doubtful identifications that could not be
117 confidently assigned to a higher taxonomic group (Table A1). A subset of the nodule field transects
118 form BGR, GSR and APEI3 was presented by Vanreusel et al. (2016), corresponding to 44% of what

119 was studied here and limited organism identification to a higher taxonomic level (Order (e.g.
120 Alcyonacea) or Class (e.g. Ophiuroidea)). In our study, the entire transects (100%) were annotated to
121 morphospecies level, allowing a detailed comparison between seamounts and nodule fields.

122 Three categories of substratum types were distinguished: (1) Predominant soft substrata (<40% hard
123 substrata), (2) mix or transition (between 40 and 60% hard substrata) and (3) predominant hard
124 substrata (>60% hard substrata), and were annotated per 10m distance units based on the video
125 footage and tested for correlations with taxonomic abundances.

126 ROV transects on the seamounts were carried out as exploratory dives. Sampling strategy both at
127 seamounts and nodule fields combined video and sampling or specimen collection. Travelling
128 altitude was easier maintained at the relatively flat nodule field transects, where an average of 93%
129 of the time was spent at altitudes <2m. Contrastingly, the uphill seamount transects were more
130 variable in ROV altitude with on average 61% of the time spent at <2m altitudes, and the remaining
131 ~39% spent at higher altitudes, which generally resulted in a higher surface covered at the
132 seamounts. Approximate surface covered (m²) was then estimated by using the ROV altitude, time
133 spent at a predefined altitude, travelled distance, and the image widths at predefined altitudes.
134 Following altitude ranges (and image widths, following Vanreusel et al. 2016 and extrapolated
135 thereon) were taken into account: <1m (2m), 1-2 m (4m), 2-3m (6m), 3-4 m (8m), 4-5m (10m).
136 Ranges from >5m, adding up to 12% for seamount transects and 3% for nodule field transects that
137 were left out since these were the parts where the seafloor was not visualised or organisms could not
138 be quantified. Due to the explorative nature of the dives, the pan and tilt of the ROV camera were
139 not kept constant and thus represents a bias on the surface estimations. Due to varying altitude of
140 the ROV and the use of camera pan, tilt and zoom, it was not possible to use surface coverage as a
141 standardisation measure. We used video transect length instead. For the transect length calculation
142 for each dive, we omitted all parts of the video footage in which the ROV was at an altitude of >10m,
143 or sections where the ROV was not visualising the seafloor (e.g. during transiting or inspecting ROV
144 parts or instruments). Visualisation of ancient disturbance tracks were omitted as well, as these fell
145 out of the scope of the article. Faunal densities were calculated as individuals per square meter
146 (ind./100m²). Faunal densities were calculated as the number of observations per 100m, in order to
147 compensate for time spent collecting samples and differing transect lengths. Statistical testing was
148 carried out in R (R core team, 2018). Non-metric multidimensional scaling analysis (NMDS) was
149 based on Bray-Curtis dissimilarity and carried out with the vegan package (Oksanen et al., 2018). The
150 Kendall's coefficient of concordance (W) was calculated to identify significantly associated groups of
151 species, based on correlations and permutations (Legendre, 2005).

152 3. Results

153 About 80% of all taxa observed across the two adjacent ecosystems, could be identified to a
154 morphospecies level. At a first view, morphospecies revealed to be quite different between
155 seamounts and nodule fields (Fig. 2). While the number of faunal observations at the seamount
156 transects were within similar ranges (344.4-427.6 ind./100m²), those at the nodule transects
157 featured both highest and lowest values (6.3-86730.5-3 ind./100m²) (Table 1). The lowest number of
158 faunal observations were done at the two APE13 nodule transects (ROV13 and 14) and highest at the
159 GSR nodule transect ROV10ROV08. What follows is a first description of eastern CCZ seamount
160 megafauna (section 3.1.) and a detailed comparison with the neighbouring nodule fields (section
161 3.2.)

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162 3.1. Insights in CCZ seamount megafauna

163 The most abundant and diverse (most morphospecies) taxa at the seamount transects comprised
164 Echinodermata (Asteroidea, Crinoidea, Holothuroidea and Ophiuroidea), Anthozoa (Actiniaria,
165 Alcyonacea, Pennatulacea, ~~Scleractinia~~) and Porifera (Hexactinellida) (Table A1, Fig. 3, ~~Fig. A1~~).
166 Keeping in mind the limitation of the video sampling, differences among the benthic seamount
167 community composition are described here. The transect at Mann Borgese (APEI3) was
168 characterised by high densities of Antipatharia, more specifically Antipathidae (~~18.53.5~~ ind./100m²),
169 and solitary Scleractinia (~~7.91.5~~ ind./100m²) (Table A1, Fig. 3). Antipathidae observations were
170 mostly grouped at the end of the video transect, i.e. at the summit. Densities of both Antipatharia
171 and Scleractinia were much lower on the other seamount transects (~~<1.0.2~~ ind./100m²) with
172 Scleractinia being absent from Heip and Senckenberg transects. Alcyonacea corals were observed on
173 all seamount transects. Isididae were found at Senckenberg and Heip transects, and one individual
174 from the Chrysogorgiidae family was observed at the latter as well. Varying numbers of Primnoidae
175 were observed on all transects (Table A1). High abundances of Pennatulacea were observed at
176 Senckenberg (~~3.80.7~~ ind./100m²), representing about ~~2820~~% of sessile fauna annotations for this
177 transect.

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178 Enteropneusta were only observed on Rüppel and Senckenberg transects in the BGR area,
179 represented by two different morphospecies, namely *Yoda* morphospecies (Torquaratoridae) at
180 Rüppel and *Saxipendium* morphospecies (Harrimaniidae) at Senckenberg.

181 Highest Polychaeta densities were observed at Heip transect in the GSR area, which was mainly due
182 to high densities of free-swimming Acroiridae (~~4.20.5~~ ind./100m² vs. ~~0.2~~ ~~0.02-0.03~~ ind./100m² in
183 BGR area Table A1). Aphroditidae polychaetes were only present at the BGR transects (~~0.02~~
184 ind./100m², ~~corresponding to 3 individuals along the transect~~) at Rüppel and ~~0.04 ind./100m~~ ~~(or 1~~
185 individual along the transect) at Senckenberg) (Table A1).

186 Porifera densities were highest at the Heip transect (~~7.50.93~~ ind./100m²), followed by ~~Senckenberg~~
187 ~~Rüppel~~ (~~3.50.38~~ ind./100m²), ~~Senckenberg-Mann Borgese~~ (~~0.36 ind./100m²~~) (~~1.9 ind./100m~~) and
188 lastly ~~Rüppel~~ (~~0.31 ind./100m²~~) ~~Mann Borgese~~ (~~1.8 ind./100m~~) (Table 2, Fig. A1(c)). Six Porifera
189 families were annotated featuring >7 to >10 morphospecies ~~per transect~~ (Fig. 3, Table A12).
190 Cladorhizidae (two individuals) were only observed on Heip transect, and one *Poliopogon* sp.
191 (Pheronematidae) was observed at Mann Borgese transect. Rossellidae gen. sp. nov. was present on
192 three seamount transects, exception being Mann Borgese.

193 Overall Echinodermata densities were highest at Senckenberg seamount (~~17.3.5~~ ind./100m²), ~~adding~~
194 ~~up to 51% of all image annotations for this transect. followed by~~ For comparison, echinoderms at
195 ~~Rüppel-Heip~~ (~~10.1.5~~ ind./100m²) ~~and Rüppel~~ (~~1.4 ind./100m²~~) (Table A1, Fig. 3), ~~both adding up to~~
196 ~~47 were responsible for 37 and 32% of all image annotations for these transects, followed by Mann~~
197 ~~Borgese~~ (~~0.62 ind./100m²~~) or 8.2% of the annotations. The number of morphospecies for all
198 echinoderm taxa (Asteroidea, Echinoidea, Holothuroidea and Crinoidea) was also highest at these 2
199 seamounts in the BGR area (Fig. A1, Table A1 Fig. 3). ~~For comparison, echinoderms at Heip~~ (~~10~~
200 ~~ind./100m~~) ~~and Mann Borgese transects~~ (~~3.3 ind./100m~~) ~~were responsible for 32% and 8.2% of~~
201 ~~observations respectively.~~ Crinoidea ~~and Holothuroidea~~ densities were highest at Senckenberg
202 (~~4.20.9~~ ind./100m² and ~~0.7 ind./100m²~~, respectively), ~~while Holothuroidea were most abundant at~~
203 ~~Rüppel~~ (~~4.4 ind./100m~~). The holothuroid families of Elpidiidae and Laetmogonidae were only

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204 observed at Senckenberg and Rüppel (BGR). Psychropotidae and Synallactidae were observed on all
205 seamounts, represented by different morphospecies. Deimatidae were not observed on Mann
206 Borgese, but were present on the three other seamount transects, again with different
207 morphospecies and densities. Velatid Asteroidea were only observed at Senckenberg and Rüppel
208 (BGR), while Brisingida and Paxillosida were observed on all four seamounts. Aspidodiadematid
209 Echinoidea were absent from the Heip transect and urchinid Echinoidea were absent from the
210 Mann Borgese transect.

211 A species accumulation curve (Fig. 4a) confirmed the limitations of the restricted and exploratory
212 nature of the sampling as no asymptote was reached. The rarefaction curves (Fig. 4b) showed that
213 the transects with the most faunal observations, which corresponded here to the longer transects,
214 were more diverse. However, at smaller sample sizes curves did not cross, thus maintaining the
215 differences observed at higher sample sizes with the Senckenberg transect (ROV04) as most diverse
216 followed by Rüppel (ROV02) (both BGR). The video transect carried out at Mann Borgese (ROV15,
217 APEI3) was the least diverse.

218 A comparison of all morphospecies observed along the 4 transects was presented in a Venn diagram
219 (Fig. 5a). Each seamount transect was characterised by a highest number of unique morphospecies,
220 only observed on the transect in question and not elsewhere. Only three morphospecies were
221 present in all seamount transects, namely *Ceriantharia* msp. 2, a small red galatheid crab and a
222 foliose sponge. Highest number of overlapping morphospecies ($n=16$) was observed between
223 Rüppel and Senckenberg, both in the BGR area (Fig. 5a). Mann Borgese showed the smallest degree
224 of overlap with the other transects (Fig. 5a).

225
226 About 57% of all sessile fauna was associated with predominantly hard substrata, followed by 31%
227 on the mixed substrata. For the mobile taxa, the pattern was less pronounced with 41 and 42%
228 associated with predominantly hard and mixed hard/soft substrata respectively. The amount of
229 predominantly hard and soft substrata was negatively correlated, though not significantly. This was
230 due to the equal amounts (40-60%) of mixed hard/soft substrata. Over all seamount transects
231 pooled together, no taxa were significantly correlated with the amount of hard substrata, nor with
232 soft substrata. When looking at the individual transects, no significant correlations were found
233 between taxa and substrata for ROV02 or ROV04 or ROV09, most likely due to the equal distribution
234 of the amount of hard/soft/mix substrata. In this perspective, ROV15 stood out, as it was dominated
235 by predominantly hard substrata (56%). For this transect, Pennatulacea were significantly
236 negatively correlated with the amount of hard substrata and Zoantharia/Octocorralia were
237 significantly and positively correlated with hard substrata, as were Ophiuroidea, Asteroidea,
238 Crinoidea and Mollusca.

239
240 Due to the limited sample size, the representativity of the observed biological patterns remains to
241 be corroborated by a more elaborate sampling strategy.

242 3.2. Comparison of seamount and nodule field faunal composition and variation

243 The faunal composition and richness (~~number of morphospecies in higher taxonomic groups~~) of the
244 nodule transects can be consulted in Fig. 3, [Fig. A1](#) and Table A1, ~~respectively~~. ~~The only taxon~~
245 ~~showing significant difference in density between seamounts and nodule fields were the Porifera (T-~~
246 ~~test assuming unequal variances, $t=-3.7$, $p<0.05$)~~. In concordance with the seamount transect, the
247 species accumulation curve of the nodule transects did not reach an asymptote either (Fig. 4c). The

248 rarefaction curves showed that the relations among transects were less straightforward for the
 249 nodule transects versus the seamount ones and did cross at smaller sample sizes (<100 individuals,
 250 Fig. 4d). ROV13 and ROV14 transects (both APEI3) were the longest in distance travelled (Table 1)
 251 but featured less faunal observations. At small sample sizes, the richness at ROV13 and 14 was
 252 highest. ROV08 and ROV10 (both GSR) showed parallel curves with ROV08 being more diverse (Fig.
 253 4d).

254 A venn diagram showing the morphospecies overlap among the nodule transects showed a total of 5
 255 species re-occurring on all 5 transects (Fig. 5b). These were: Munnopsidae msp. 1 (Isopoda,
 256 Crustacea), Actiniaria msp.7 (Cnidaria), Ophiuroidea msp. 6 (Echinodermata), *Holascus* sp. and
 257 *Hyalonema* sp. (Hexactinellida, Porifera). There was a high number of unique morphospecies for
 258 each transect, though not as high as for the seamount transects (Fig. 5). ROV13 and 14 (both APEI3)
 259 showed least overlap with the other transects, which is similar to what was observed at the
 260 seamounts.

261 Observations and quantifications of morphospecies confirmed the high degree of dissimilarity
 262 between the two neighbouring ecosystems. Porifera, Ophiuroidea (Echinodermata), Actiniaria and
 263 Alcyonacea (Cnidaria) were more abundant at nodule fields (Fig. 3). These taxonomic groups were
 264 also most diverse on nodule fields (i.e. highest number of morphospecies), exception being the
 265 Alcyonacea which featured more morphospecies on the seamounts (12 to 8 morphospecies for
 266 seamounts and nodule fields respectively) (Fig. 3). Of all Porifera, Cladhorizidae were more diverse
 267 at nodule fields than at seamounts (7 to 1 morphospecies, respectively).

268 There were only 21 morphospecies (10%) that were observed both on seamounts and nodule fields
 269 (Fig. 6). While this subset of morphospecies occurred in both ecosystems, they did so in very
 270 different densities, i.e. very abundant in one ecosystem and very low in abundance in the other:
 271 examples are Galatheididae small red msp. (Decapoda, Crustacea), *Synallactes* white msp.
 272 (Holothuroidea), Ophiuroidea msp. 5 and 6, Comatulida msp. 1 (Crinoidea), *Hyalonema* sp. and
 273 *Hyalostylus* sp. (both Hexactinellida, Porifera) (Fig. 6).

274 Three Ophiuroidea morphospecies were present at both seamounts and nodule fields (Fig. 2, 3 and
 275 6). ~~The majority of the very abundant~~Most of the Ophiuroidea observed at the CCZ seamounts that
 276 could be identified to morphospecies level, were small and situated on hard substrata
 277 (morphospecies 5), while ~~most of the Ophiuroidea~~those at nodule fields (including morphospecies 6)
 278 were observed on the soft sediments. Morphospecies 6 was only rarely observed on the seamounts
 279 (Fig. 3). Another easily recognisable morphospecies was found on Porifera, coral and animal stalks
 280 and was more abundant at seamounts than at nodule fields (morphospecies 4) (Fig. ~~2 and~~3).

281 Crinoidea, Asteroidea (both Echinodermata) and Antipatharia (Cnidaria) were more abundant on the
 282 seamounts (Fig. ~~A13~~). This coincided with a higher diversity for Asteroidea and Antipatharia on the
 283 seamounts as well. Crinoidea diversity was similar (5 to 4 morphospecies comparing seamounts to
 284 nodule fields). Holothuroidea occurred in similar densities in both ecosystems (Fig. ~~A13~~, though they
 285 were characterised by different morphospecies (~~Fig. 3~~Table 2, Fig. ~~A1(b)~~). Overall densities of
 286 Echinoidea were comparable between seamounts and nodule fields, though for the nodule fields
 287 this was mostly due to one very abundant morphospecies, namely Aspidodiadematidae msp 1,
 288 which was absent at the seamounts (~~Fig. 3~~Table 2, Fig. ~~A1(b)~~). Besides this, Echinoidea were more
 289 diverse at seamounts (11 morphospecies vs. 5 at nodule fields).

290 There was no morphospecies overlap for Tunicata, Antipatharia, and Actiniaria. Alcyonacea,
291 Ceriantharia, Corallimorphidae and Crinoidea only shared 1 morphospecies between seamounts and
292 nodule fields, namely *Callozostron* cf. *bayeri*, Ceriantharia msp. 2, *Corallimorphus* msp. 2 and
293 Comatulida msp. 1 respectively (Fig. 6).

294 There were no observations of Enteropneusta, Scleractinia and Zoantharia (Cnidaria), Aphroditidae
295 (Polychaeta) or holothuroid Deimatidae at the nodule fields transects (Table A1, Fig. A1). While
296 Actinopterygii were left out of the analysis, it should be noted that fish observations were more
297 diverse at the seamounts than on the nodule fields.

298 There was quite some faunal variation observed among the video transects of, both seamounts and
299 nodule fields (see Fig. 5 and 7). The (dis)similarities were analysed by a nMDS analysis, which
300 grouped the 9 different video transects based on their taxonomic composition. Despite the large
301 intra-ecosystem variation, they pooled in two distinct groups separating the nodule fields from the
302 seamounts (Fig. 7a). Within each group, BSR and GSR transects were more similar to one another
303 both for seamounts and nodule fields, whilst APEI3 transects stood out more.

304 The Kendall's coefficient of concordance (W, Legendre, 2005) corroborated the existence of two
305 significantly different species associations, whose composition corresponded to the fauna
306 characterising the nodule fields ($W=0.292.03$, $p<0.001$, after 999 permutations) and the seamounts
307 ($W=0.393.04$, $p<0.001$, after 999 permutations).

308 Depth was fitted as a vector on top of the nMDS plot (Fig. 7b) and showed that the discrepancy in
309 faunal composition between the two ecosystems also corresponded to a difference in depth, with
310 the nodule transects all being situated below the 4000m isobath and the seamount transects ranging
311 from 1650 to >3500m (Fig. 7b).

312 4. Discussion

313 4.1. Intra-ecosystem faunal variation

314 Community composition varied markedly at seamounts and nodule fields. The limited sampling (n=9
315 transects), at different locations and additionally, for the seamounts, different depth ranges,
316 precluded any general conclusions on quantifications of biodiversity *per se*. However, taking this into
317 account, it was also the first time seamounts were visited in the area, thus granting first insights in
318 the fauna inhabiting these seamounts and allowing a first comparison with nodule faunal
319 composition.

320 The two BGR seamount transects were most similar in faunal composition, followed by the Heip
321 seamount transect (GSR). These seamount video transects were characterised by more similar depth
322 ranges, and the two BGR transects were also geographically closest to each other. Although for
323 seamounts, distance separating them might be a less determining factor than depth since
324 (mega)faunal communities can be very different even between adjacent seamounts (Schlacher et al.,
325 2014; Boschen et al., 2015). Overall, parameters that vary with depth, such as temperature, oxygen
326 concentration, substratum type, food availability, and pressure are considered major drivers of
327 species composition on seamounts (Clark et al., 2010; McClain et al., 2010). The quantification of the
328 amount of hard and soft substrata was not distinctive enough to explain differences observed here.
329 The difference in depth could also explain the higher dissimilarity with Mann Borgese (APEI3) who
330 featured the shallowest transect and summit, which was dominated by Antipatharia. Antipatharians

331 were previously reported to be more dominant towards peaks as compared to mid-slopes at
332 corresponding depths (Genin et al., 1986). Based on their filter-feeding strategy, Porifera (except
333 carnivorous Cladorhizidae), were also thought to benefit from elevated topography (peaks) or
334 exposed substrata in analogy to corals (Genin et al., 1986; Clark et al., 2010), though no such pattern
335 was apparent here. Porifera are notoriously difficult to identify based on imagery. Although the
336 sampled individuals allowed some identifications to genus or species level (Kersken et al., 2018a and
337 b), identifications remained hard to extrapolate across the different video transects. Generally, as in
338 our study, seamount summits have been more intensively sampled (Stocks, 2009) although the little
339 work done at seamount bases and deep slopes indicated that these areas support distinct
340 assemblages (Baco, 2007).

341 Among the nodule transects a considerable amount of variation in faunal composition was observed
342 (this study, Vanreusel et al., 2016). The two APEI3 nodule transects (ROV13 and 14) stood out in
343 faunal composition, diversity and in low number of faunal observations. They were also the only two
344 transects situated below the 4500m isobaths. But rather than depth, the nodule coverage may be
345 considered an important driving factor, since the density of nodule megafauna was shown to vary
346 with nodule size and density/coverage (Stoyanova, 2012; Vanreusel et al., 2016, Simon-Llédó et al.,
347 2019). Here as well, the APEI3 transects were characterised by a high nodule coverage (~40-88%,
348 Vanreusel et al., 2016), whereas the BGR and GSR nodule transects (ROV3 and ROV 8 + 10,
349 respectively) had a nodule coverage <30% and were also more similar in faunal composition
350 (Vanreusel et al., 2016). Other factors that could be at play are the more oligotrophic surface waters
351 of the northern CCZ which could be the cause of the overall lower faunal densities at APEI3 nodule
352 fields (Vanreusel et al., 2016). Volz et al. (2018) corroborated this, with the location of the APEI3 site
353 in the proximity of the carbon-starved North Pacific gyre being characterised by a reduced POC-flux
354 quantified to being 22-46% lower than the GSR and BGR areas respectively.

355 The species accumulation curves showed that ~~no an~~ asymptote was reached neither at seamounts,
356 nor at nodule fields. Consequently, longer transect lengths might be necessary to representatively
357 quantify and assess megafauna density and diversity (Simon-Llédó et al., 2019). In addition, for a first
358 in-depth description and assessment of seamount fauna composition, one video transect is
359 insufficient to describe the diversity and shifts in faunal assemblages of the surveyed seamounts.
360 Rather, an ampler imaging strategy should be developed, with a minimum transect length exceeding
361 1000 ~~ms~~ (Simon-Llédó et al., 2019) and replicate transects carried out on different faces of the
362 seamount, on slopes with varying degree of exposure to currents and different substrate types.
363 Wider depth ranges should be taken into account as well. Alternatively, across slope transects,
364 following depth contours should be considered as these could provide observation replicates for a
365 given depth. Despite its limitations, this study grants first insights in the seamount inhabiting
366 megafauna of the eastern CCZ and an important first comparison with nodule fauna.

367 4.2. Faunal (dis)similarities between seamounts and nodule fields

368 In other areas, seamounts were shown to share fauna with surrounding habitats (Clark et al., 2010)
369 and could thus potentially serve as source populations for neighbouring environments (McClain et
370 al., 2009). While generally few species seemed restricted to seamounts only (Clark et al., 2010),
371 morphospecies in this study revealed to be quite different between seamounts and nodule fields
372 with little overlap between both. Despite the high degree of variation observed among all the video
373 transects, these grouped into two distinctly separate clusters, separating nodule from seamount

374 transects. The few overlapping morphospecies did occur in different densities in each ecosystem,
375 implying a different role or importance in the ecological community and its functioning.

376 Overall, nodule fields showed higher faunal densities than seamounts. Shifts in density patterns
377 between nodule fields and seamounts were more evident in a number of taxa, where the variety of
378 morphospecies and feeding strategy within each group was likely to be at play. An example of this
379 are the Echinodermata, which include Asteroidea (predators and filter feeders (Brisingida)),
380 Crinoidea (filter feeders), Echinoidea (deposit feeders), Holothuroidea (deposit feeders) and
381 Ophiuroidea (omnivores). Asteroidea were more abundant on seamounts and both Echinoidea and
382 Asteroidea were more diverse in this ecosystem as well. Ophiuroidea were most abundant on the
383 nodule fields (ratio 7 to 1 when compared to seamounts). Same ophiuroid morphospecies were
384 present at seamounts and nodule fields but in very different abundances and they were observed on
385 different substrata types, which indicates different lifestyles, feeding behaviour and corresponding
386 dietary specialisations (Persons and Gage, 1984). Previously it was already demonstrated that
387 Ophiuroidea did not show high levels of richness or endemism on seamounts (O'Hara, 2007). At
388 nodule fields, Ophiuroidea were often observed in association with xenophyophores (Amon et al.,
389 2016, this study) and a similar observation was done at east Pacific seamounts off Mexico (Levin et
390 al., 1986), though no such associations were observed on the seamounts studied here.

391 Holothuroidea densities were thought to possibly decrease when less soft sediment was available
392 since they feed mainly on the upper layers of the soft-bottom sediment (Bluhm and Gebruk, 1999).
393 No significant link was established between holothuroid densities and the amount of hard substrata
394 in this study, but their community composition varied distinctly between nodule fields and
395 seamounts with more families being observed at the latter. Additionally, at the seamounts, many
396 holothurians were observed on top of rocks, possibly reflecting different feeding strategies and
397 explaining the observations of different morphospecies. Geographical variations, different bottom
398 topography, differences in nodule coverages and sizes and/or an uneven distribution of holothurians
399 on the sea floor were thought to play a role in holothuroid community composition (Bluhm and
400 Gebruk, 1999). On the other hand, variability in deep-sea holothuroid abundance was proposed to
401 depend primarily on depth and distance from continents (see Billet, 1991 for a review).

402 Stalked organisms, such as Crinoidea (Echinodermata) and Hexactinellida (except for
403 Amphidiscophora, Porifera) rely on hard substrata for their attachment and are considered being
404 among the most vulnerable organisms when mining is concerned. Crinoidea were more abundant on
405 seamounts, possibly because hard substrata were less limiting than in the nodule fields. Porifera
406 densities (stalked and non-stalked) varied among all analysed transects, revealing no particular
407 trends in abundance. However, the species composition of deep-sea glass sponge communities from
408 seamounts and polymetallic nodule fields was distinctly different. Polymetallic nodule field
409 communities were dominated by widely-distributed genera such as *Caulophacus* and *Hyalonema*,
410 whereas seamount communities seemed to have a rather unique composition represented by
411 genera like *Saccocalyx*.

412 Corals were generally considered to be more abundant on seamounts than adjacent areas, due to
413 their ability to feed on a variety of planktonic or detritus sources suspended in the water column
414 (Rowden et al., 2010). In this study, the Alcyonacea densities were lower on the seamounts than on
415 the nodule transects. The majority of Alcyonacea morphospecies of the seamounts did not occur on

416 the nodule fields and vice versa, with exception of *Callozostron* cf. *bayeri* which was also present at
417 the nodule fields but in very low densities (1/10-8 of those observed at seamounts). The Antipatharia
418 were most abundant at the Mann Borgese seamount (APEI3) compared to all other transects. The
419 depth difference of more than 3000m between this particular seamount and the nodule fields could
420 explain the abundance in Antipatharia which were shown to be more abundant at lower depths
421 (Genin et al., 1986). Additional presence of Pennatulacea at seamounts, a taxon that was virtually
422 absent from the nodule field transects and that appeared more linked to predominant soft substrata
423 at seamounts, resulted in completely distinct coral communities for each ecosystem.

424 Actiniaria were denominated the second most common group at CCZ nodule fields, after the
425 xenophyophores (Kamenskaya et al., 2015) and, in our study, were also more abundant on nodule
426 fields than on seamounts. Depending on the species and feeding strategy, the ratio hard/soft
427 substrata and their preference for either one could play a role. Since morphospecies were distinct
428 between seamounts and nodule fields, their role in the respective communities are likely to differ as
429 well. Combinations of deposit feeding and predatory behaviour in Actiniaria have been observed, as
430 well as burrowing activity, preference for attachment to hard substrata and exposure to currents
431 (Durden et al., 2015a; Lampitt and Paterson, 1987; Riemann-Zürneck, 1998), all factors that could
432 influence the differences in morphospecies observed.

433 Some taxa were only observed on the seamounts in this study, while they occurred on nodule fields
434 elsewhere, be it in low densities. For instance, Enteropneusta, which in this study were found only
435 on seamounts, were observed previously at CCZ nodule fields though observations were rather rare
436 (Tilot, 2006). They appeared more abundant at the nodule fields of the Deep Peru Basin (DISCOL
437 area), though a wide range in abundances was displayed there as well (Bluhm, 2001). The exception
438 were the Scleractinia, which were quite common on seamounts, as also reported in other studies
439 (e.g. Baco, 2007; Rowden et al., 2010), but distinctly absent at nodule fields.

440 Explanation for the discrepancies in faunal composition and the low degree of morphospecies
441 overlap between seamount and nodule fields, as observed here, can be multiple. For one, nodules
442 may not be considered a plain hard substratum, with their metal composition, microbial colonisation
443 and the nodule/sediment interface influencing the epi- and associated megafaunal composition. The
444 possibility of a specific deep-sea faunal community that tolerates or benefits from manganese
445 substrata has been previously proposed (Mullineaux, 1988). The comparison between seamounts
446 and nodule fields as two neighbouring hard-substrata ecosystems also entailed a comparison
447 between depth gradients and possible thresholds (>4000m for nodule fields and 1500>x <4000m for
448 seamounts). Related to this is the steepness of the seamount slope and its current exposure playing
449 a role in the faunal colonisation (Genin et al., 1986; Rappaport et al., 1997). Other studies showed
450 that habitat heterogeneity increased megafaunal diversity at seamounts (Raymore, 1982) and
451 elsewhere, such as abyssal plains (Lapointe and Bourget, 1999; Durden et al., 2015b, Leitner et al.,
452 2017, Simon-Llédó et al., 2019). Within this perspective the smaller-scale substratum heterogeneity
453 transcending the ratio hard/soft substrata or amount of hard substrata could play a role as well.

454 5. Conclusions

455 Based on our current knowledge; seamounts appear inadequate as refuge areas to help maintain
456 nodule biodiversity. In order to conclusively exclude seamount habitats as a refuge for nodule fauna,

a more comprehensive sampling should be carried out. The sampling strategy wielded in this study lacked replicates, uniformity and was limited in sample size. Seamount bases should be taken into consideration as well as they can be characterised by distinctly different assemblages than the summits and they feature depth ranges more similar to nodule fields.

While their role as refuge area for nodule field fauna is currently debatable, the possible uniqueness of the seamount habitat and its inhabiting fauna implies that seamounts need to be included in management plans for the conservation of the biodiversity and ecosystems of the CCZ.

Author Contributions

DC, PAR, SPR, DK analysed the images. DC analysed the data. PMA, PAR, AC conceptualised and carried out the sampling. All authors contributed to the redaction of the manuscript.

Data Availability

Data sets are made available through OSIS-Kiel data portal, BIIGLE and PANGAEA.

Competing interest

The authors declare that they have no conflict of interest

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591 **Tables**
 592 Table 1- Overview table on details of imagery transects analysed in the Clarion-Clipperton license
 593 areas. Video duration includes time spent sampling. **Faunal densities** **Number of observations**
 594 include undetermined organisms. Transect lengths do not include parts visualising ancient
 595 disturbance tracks or parts when the seafloor was not visualised or visible.

Station/Dive	License Area	Seamount (SM) or Nodule field (NF)	Depth (m)	Transect length	Transect lengthApproximate surface	#obs/dive	Faunal densities #

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					covered (m ²)		obs ind. /100m ²
SO239_29_ROV02	BGR	SM: Rüppell	3000-2500	<u>1250m</u>	<u>9458.61</u> 250m	429	<u>4.434.3</u>
SO239_41_ROV03	BGR	NF	4080-4110	<u>1590m</u>	<u>5309.11</u> 590m	932	<u>19.358</u> -6
SO239_54_ROV04	BGR	SM: Senckenberg	3350-2850	<u>2500m</u>	<u>12288.5</u> 2500m	890	<u>6.935.6</u>
SO239_131_ROV08	GSR	NF	4470-4480	<u>710m</u>	<u>1602.57</u> 10m	445	<u>30.362</u> -8
SO239_135_ROV09	GSR	SM: Heip	3900-3550	<u>1000m</u>	<u>6905.41</u> 000m	359	<u>5.335.9</u>
SO239_141_ROV10	GSR	NF	4455-4480	<u>520m</u>	<u>1683.45</u> 20m	351	<u>27.667</u> -5
SO239_189_ROV13	APEI 3	NF	4890-4930	<u>1790m</u>	<u>3580.01</u> 790m	113	<u>3.86.3</u>
SO239_200_ROV14	APEI 3	NF	4650-4670	<u>1490m</u>	<u>2980.01</u> 490m	179	<u>6.212.0</u>
SO239_212_ROV15	APEI 3	SM: Mann Borgese	1850-1650	<u>900m</u>	<u>4805.39</u> 00m	378	<u>7.642.0</u>

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Table 2. Overview of all densities (ind./100m²) observed within each video transect. Higher taxa are in bold. * indicates taxa left out of the statistical analyses due to lack of representativity. Indets were organisms impossible to attribute to a lower taxonomic group. ROV02=Rüppell, ROV04=Senckenberg, ROV09=Heip, ROV15=Mann Borgese

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	SEAMOUNTS				NODULE FIELDS				
	ROV2 Ind./100m ²	ROV4 Ind./100m ²	ROV9 Ind./100m ²	ROV15 Ind./100m ²	ROV3 Ind./100m ²	ROV8 Ind./100m ²	ROV10 Ind./100m ²	ROV13 Ind./100m ²	ROV14 Ind./100m ²
-									
Annelida*									
Polychaeta indet. * (No Serpulidae)	<u>0.018</u>	<u>0.024</u>	-	<u>0.021</u>	<u>0.094</u>	-	<u>0.119</u>	<u>0.028</u>	<u>0.034</u>
<u>Acrocirridae</u>	<u>0.018</u>	<u>0.033</u>	<u>0.515</u>	-	<u>0.170</u>	<u>0.062</u>	<u>0.178</u>	<u>0.894</u>	<u>0.973</u>
<u>Aphroditidae</u>	<u>0.027</u>	<u>0.008</u>	-	-		-		-	-
<u>Echiura msp 1</u>		-	-	-	<u>0.170</u>	<u>0.499</u>	<u>0.356</u>	-	<u>0.101</u>
<u>Polynoidea</u>		-	-	-		-		-	-
<u>Polynoidae msp 2</u>		-	-	-		<u>0.062</u>	<u>0.178</u>	-	-
<u>Polynoidae white msp</u>		-	-	-		<u>0.062</u>		<u>0.028</u>	<u>0.067</u>
-									
Bryozoa									
<u>Bryozoa msp 2</u>		-	<u>0.025</u>	-	-	-	-	-	-
<u>Bryozoa indet.</u>		<u>0.008</u>	-	-	<u>0.132</u>	<u>0.686</u>	<u>0.059</u>	<u>0.056</u>	<u>0.067</u>
-									
Cnidaria									
Anthozoa	-	-	-	-	-	-	-	-	-
Ceriantharia	-	-	-	-	-	-	-	-	-
<u>Ceriantharia msp 1</u>	<u>0.045</u>	<u>0.008</u>	<u>0.049</u>	<u>0.040</u>	-	<u>0.062</u>	-	-	-
<u>Ceriantharia msp 2</u>	-	-	-	<u>0.081</u>	-	<u>0.125</u>	<u>0.059</u>	-	-
<u>Ceriantharia indet.</u>	-	-	<u>0.012</u>	-	-	-	-	-	-
Hexacorralia	-	-	-	-	-	-	-	-	-
Actiniaria	-	-	-	-	-	-	-	-	-
<u>Actinoscyphiidae</u>	-	<u>0.023</u>	-	-	-	-	-	-	-
<u>Actiniidae/Bolocera msp.</u>	<u>0.135</u>	<u>0.039</u>	-	-	-	-	-	-	-
<u>Actiniaria msp 15</u>	<u>0.009</u>	-	-	-	-	-	-	-	-
<u>Actiniaria msp 4</u>	-	<u>0.016</u>	-	<u>0.020</u>	-	-	-	-	-
<u>Actiniaria msp 5</u>	<u>0.009</u>	<u>0.016</u>	-	<u>0.060</u>	-	-	-	-	-

<u>Actiniaria msp 10</u>	-	-	-	-	<u>0.094</u>	-	-	-	<u>0.168</u>
<u>Actiniaria msp 2</u>	-	-	-	-	<u>0.320</u>	<u>0.499</u>	<u>0.059</u>	-	<u>0.134</u>
<u>Actiniaria msp C</u>	-	-	-	-	<u>0.113</u>	<u>0.187</u>	<u>0.119</u>	-	-
<u>Actiniaria msp D</u>	-	-	-	-	<u>0.019</u>	-	-	-	-
<u>Actiniaria msp 7</u>	-	-	-	-	<u>0.188</u>	<u>0.062</u>	<u>0.178</u>	<u>0.028</u>	<u>0.067</u>
<u>Actiniaria msp 8</u>	-	-	-	-	<u>0.038</u>	<u>1.622</u>	<u>0.950</u>	-	<u>0.101</u>
<u>Actiniaria msp 9</u>	-	-	-	-	-	-	<u>0.059</u>	-	-
<u>Actiniaria msp A</u>	-	-	-	-	-	-	<u>0.059</u>	-	<u>0.034</u>
<u>Actiniaria msp B</u>	-	-	-	-	<u>0.075</u>	<u>0.062</u>	<u>0.119</u>	<u>0.028</u>	-
<u>Actiniaria indet.</u>	<u>0.018</u>	<u>0.031</u>	-	-	<u>0.471</u>	<u>0.624</u>	<u>1.366</u>	<u>0.056</u>	<u>0.067</u>
<u>Antipatharia</u>	-	-	-	-	-	-	-	-	-
<u>Antipathidae</u>	-	-	-	-	-	-	-	-	-
<u>Antipathes msp 1</u>	-	-	-	<u>1.591</u>	-	-	-	-	-
<u>Antipathes msp 2</u>	-	-	-	<u>0.020</u>	-	-	-	-	-
<u>Stichopathes msp 1</u>	-	-	-	<u>1.752</u>	-	-	-	-	-
<u>Antipathidae indet.</u>	-	-	-	<u>0.101</u>	-	-	-	-	-
<u>Schizopathidae</u>	-	-	-	-	-	-	-	-	-
<u>Abyssopathes cf. lyra</u>	-	-	-	-	<u>0.151</u>	<u>0.250</u>	<u>0.178</u>	-	-
<u>Bathypathes cf. alternata</u>	-	-	-	-	-	<u>0.062</u>	<u>0.059</u>	-	<u>0.034</u>
<u>Bathypates cf. alternata msp 1</u>	-	-	<u>0.012</u>	-	-	-	-	-	-
<u>Bathypathes cf. alternata msp 2</u>	-	<u>0.023</u>	-	-	-	-	-	-	-
<u>Bathypathes sp.</u>	-	-	-	-	<u>0.057</u>	<u>0.062</u>	-	-	-
<u>Bathypathes msp 1</u>	-	-	<u>0.012</u>	-	-	-	-	-	-
<u>cf. Parantipathes msp 1</u>	-	-	<u>0.016</u>	-	-	-	-	-	-
<u>Umbellapathes aff. bipinnata</u>	-	<u>0.039</u>	<u>0.012</u>	-	-	-	-	-	-
<u>Umbellapathes aff. helioanthes</u>	-	<u>0.117</u>	-	-	-	-	-	-	-
<u>Antipatharia indet.</u>	<u>0.009</u>	<u>0.016</u>	<u>0.012</u>	<u>0.121</u>	<u>0.075</u>	<u>0.125</u>	<u>0.059</u>	-	-
<u>Corallimorpharia/Corallimorphidae</u>	-	-	-	-	-	-	-	-	-
<u>Corallimorphus msp 1</u>	-	<u>0.008</u>	-	-	-	-	-	-	-
<u>Corallimorphus msp 2</u>	-	<u>0.094</u>	<u>0.012</u>	-	<u>0.075</u>	<u>0.125</u>	<u>0.059</u>	-	-
<u>Corallimorpharia msp 3</u>	-	<u>0.008</u>	-	-	-	-	-	-	-

<u>Corallimorpharia msp 4</u>	-	-	<u>0.012</u>	-	-	-	-	-	-	-
<u>Corallimorpharia msp A</u>	-	-	-	-	<u>0.019</u>	-	<u>0.059</u>	-	-	-
<u>Corallimorpharia msp B</u>	-	-	-	-	<u>0.019</u>	-	-	-	-	-
<u>Scleractinia</u>	-	-	-	-	-	-	-	-	-	-
<u>Scleractinia msp 1</u>	<u>0.018</u>	-	-	<u>1.470</u>	-	-	-	-	-	-
<u>Zoantharia</u>	-	-	-	-	-	-	-	-	-	-
<u>Zoantharia msp 2</u>	-	-	-	<u>0.020</u>	-	-	-	-	-	-
<u>Zoantharia indet.</u>	-	<u>0.094</u>	-	<u>0.040</u>	-	-	-	-	-	-
<u>Octocorralia</u>	-	-	-	-	-	-	-	-	-	-
<u>Alcyonacea</u>	-	-	-	-	-	-	-	-	-	-
<u>Alcyoniidae</u>	-	-	-	-	-	-	-	-	-	-
<u>Anthomastus msp 1</u>	<u>0.027</u>	-	-	-	-	-	-	-	-	-
<u>Anthomastus msp 2</u>	<u>0.000</u>	<u>0.031</u>	-	<u>0.020</u>	-	-	-	-	-	-
<u>Coralliidae</u>	-	-	-	-	-	-	-	-	-	-
<u>Corallium sp. nov.</u>	-	-	-	<u>0.020</u>	-	-	-	-	-	-
<u>Chrysogorgiidae</u>	-	-	-	-	-	-	-	-	-	-
<u>Chrysogorgia cf. pinnata</u>	-	-	<u>0.012</u>	-	-	-	-	-	-	-
<u>Isididae</u>	-	-	-	-	-	-	-	-	-	-
<u>Bathygorgia aff. abyssicola 1</u>	-	-	-	-	-	<u>0.062</u>	<u>0.059</u>	-	-	-
<u>Bathygorgia aff. profunda 1</u>	-	<u>0.031</u>	<u>0.012</u>	-	-	-	-	-	-	-
<u>Bathygorgia aff. profunda 2</u>	-	-	<u>0.012</u>	-	-	-	-	-	-	-
<u>Keratoisis aff. flexibilis msp 2</u>	-	-	<u>0.012</u>	-	-	-	-	-	-	-
<u>Isididae msp 1</u>	-	<u>0.008</u>	-	-	-	-	-	-	-	-
<u>Isididae indet.</u>	<u>0.018</u>	-	<u>0.110</u>	<u>0.020</u>	<u>0.038</u>	<u>2.496</u>	<u>0.713</u>	-	-	-
<u>Taiaroidea</u>	-	-	-	-	-	-	-	-	-	-
<u>Taiaroidea msp 1</u>	-	-	-	-	-	-	<u>0.059</u>	-	-	-
<u>Primnoidae</u>	-	-	-	-	-	-	-	-	-	-
<u>Abyssoprinoia cf. gemina</u>	-	-	-	-	-	<u>0.312</u>	<u>0.178</u>	-	-	-
<u>Callozostron cf. bayeri</u>	<u>0.009</u>	<u>0.110</u>	-	-	<u>0.019</u>	-	-	-	-	-
<u>Calyptrophora cf. persephone</u>	-	-	-	-	<u>0.019</u>	-	-	-	-	-
<u>Narella msp 1</u>	-	<u>0.016</u>	-	<u>0.020</u>	-	-	-	-	-	-

<u>Primnoidea indet.</u>	<u>0.081</u>	-	<u>0.025</u>	-	<u>0.810</u>	<u>1.498</u>	<u>0.475</u>	-	-
<u>Alcyonacea msp 1</u>	-	-	-	-	<u>0.038</u>	-	-	<u>0.056</u>	<u>0.067</u>
<u>Alcyonacea indet.</u>	-	<u>0.031</u>	<u>0.209</u>	<u>0.503</u>	<u>2.675</u>	<u>2.933</u>	<u>1.247</u>	-	<u>0.034</u>
<u>Pennatulacea</u>	-	-	-	-	-	-	-	-	-
<u>Umbellulidae</u>	-	-	-	-	-	-	-	-	-
<u>Umbellula msp 1 White</u>	-	-	-	-	-	-	-	-	<u>0.067</u>
<u>Umbellula msp 1 orange</u>	-	<u>0.063</u>	-	<u>0.020</u>	-	-	-	-	-
<u>Umbellula msp 2</u>	-	<u>0.016</u>	-	-	-	-	-	-	-
<u>Umbellulidae indet.</u>	-	<u>0.031</u>	-	-	-	-	-	-	-
<u>Protoptilidae</u>	-	-	-	<u>0.020</u>	-	-	-	-	-
<u>Protoptilum msp 1</u>	-	<u>0.008</u>	-	<u>0.040</u>	-	-	-	-	-
<u>Pennatulacea msp 2</u>	-	<u>0.008</u>	-	-	-	-	-	-	-
<u>Pennatulacea msp 5</u>	-	<u>0.047</u>	-	-	-	-	-	-	-
<u>Pennatulacea msp 6</u>	-	<u>0.023</u>	-	-	-	-	-	-	-
<u>Pennatulacea msp 7</u>	-	<u>0.078</u>	-	-	-	-	-	-	-
<u>Pennatulacea msp 8</u>	-	<u>0.016</u>	-	-	-	-	-	-	-
<u>Pennatulacea indet</u>	<u>0.018</u>	<u>0.423</u>	-	<u>0.020</u>	-	<u>0.062</u>	-	-	-
<u>Octocorallia msp 1</u>	-	-	-	<u>0.040</u>	-	-	-	-	-
<u>Octocorallia msp 2</u>	-	-	-	-	-	-	-	-	-
<u>Anthozoa indet.</u>	<u>0.018</u>	<u>0.023</u>	<u>0.074</u>	<u>0.121</u>	<u>0.038</u>	<u>0.062</u>	<u>0.059</u>	-	-
<u>Hydrozoa</u>	-	-	-	-	-	-	-	-	-
<u>Branchiocerianthus msp</u>	-	<u>0.016</u>	-	-	-	-	-	-	-
<u>Hydrozoa indet.</u>	-	<u>0.016</u>	<u>0.012</u>	<u>0.040</u>	-	<u>0.062</u>	-	-	-
				-	-			-	-
<u>Crustacea*</u>	-	-	-	-	-	-	-	-	-
<u>Decapoda</u>									
<u>Caridea</u>	<u>0.458</u>	<u>0.516</u>	<u>0.466</u>	<u>0.040</u>	<u>0.057</u>	-	<u>0.356</u>	<u>0.056</u>	<u>0.101</u>
<u>Decapoda msp 3</u>	-	<u>0.016</u>	-	-	-	-	-	-	-
<u>Decapoda msp 4</u>	<u>0.009</u>	-	-	-	-	-	-	-	-
<u>Decapoda/Aristeidae</u>	<u>0.009</u>	<u>0.016</u>	-	<u>0.101</u>	<u>0.019</u>	<u>0.250</u>	<u>0.178</u>	<u>0.056</u>	<u>0.034</u>
<u>Decapoda msp 1</u>	-	-	-	-	-	-	-	<u>0.028</u>	-

<u>Galatheidae</u>	-	-	-	-	-	-	-	-	-
<u>Galatheidae small red msp</u>	<u>0.369</u>	<u>0.110</u>	<u>0.025</u>	<u>0.081</u>	<u>0.019</u>	-	-	-	-
<u>Galatheidae small white msp</u>	<u>0.009</u>	<u>0.023</u>	-	-	-	-	-	-	-
<u>Munidopsis spp.</u>	<u>0.108</u>	<u>0.070</u>	<u>0.074</u>	-	-	<u>0.187</u>	-	-	<u>0.101</u>
<u>Galatheidae indet.</u>	<u>0.018</u>	<u>0.031</u>	<u>0.025</u>	<u>0.020</u>	-	-	-	-	-
<u>Parapaguridae</u>	-	-	-	-	-	-	-	-	-
<u>Parapaguridae msp 1/Probebebei sp.</u>	<u>0.072</u>	<u>0.047</u>	-	-	-	-	-	-	-
<u>Peracarida</u>	-	-	-	-	-	-	-	-	-
<u>Amphipoda</u>	-	-	<u>0.012</u>	-	<u>0.019</u>	<u>0.125</u>	-	<u>0.028</u>	<u>0.134</u>
<u>Podoceridae msp 1</u>	-	-	-	-	-	-	-	<u>0.028</u>	-
<u>Amphipoda msp 1</u>	-	<u>0.016</u>	<u>0.025</u>	-	-	-	-	-	-
<u>Isopoda</u>	-	-	-	-	-	-	-	-	-
<u>Munnopsidae msp 1</u>	-	-	-	-	<u>0.170</u>	<u>0.187</u>	<u>0.059</u>	<u>0.084</u>	<u>0.268</u>
<u>Decapoda indet.</u>	-	<u>0.023</u>	<u>0.098</u>	-	-	-	-	-	-
<u>Crustacea indet.</u>	<u>0.009</u>	<u>0.063</u>	<u>0.074</u>	-	-	-	-	-	<u>0.067</u>
<u>Echinodermata</u>	-	-	-	-	-	-	-	-	-
<u>Asteroidea</u>	-	-	-	-	-	-	-	-	-
<u>Brisingida</u>	-	-	-	-	-	-	-	-	-
<u>Brisingida msp 1 (6 arms - orange)</u>	-	<u>0.031</u>	<u>0.074</u>	-	<u>0.075</u>	-	-	-	-
<u>Brisingida msp 1 (8 arms - orange)</u>	<u>0.018</u>	<u>0.078</u>	<u>0.037</u>	-	-	-	-	-	<u>0.034</u>
<u>Brisingida msp 3 (6 arms - white)</u>	-	<u>0.078</u>	<u>0.135</u>	<u>0.040</u>	<u>0.057</u>	<u>0.187</u>	<u>0.059</u>	-	<u>0.067</u>
<u>Brisingida msp 4 (9-10 arms)</u>	<u>0.018</u>	<u>0.078</u>	-	-	-	-	-	-	-
<u>Brisingida indet.</u>	<u>0.036</u>	<u>0.016</u>	-	<u>0.040</u>	-	-	-	-	-
<u>Paxillosida</u>	-	-	-	-	-	-	-	-	-
<u>Solaster msp</u>	-	<u>0.008</u>	-	-	-	-	-	-	-
<u>Paxillosida cf AST_009/AST_007</u>	-	<u>0.102</u>	<u>0.061</u>	-	-	-	-	-	-
<u>Paxillosida msp 1</u>	<u>0.009</u>	-	-	<u>0.020</u>	-	-	-	-	-
<u>Paxillosida msp 2a</u>	-	<u>0.008</u>	-	-	-	-	-	-	-
<u>Paxillosida msp 2b</u>	-	-	<u>0.012</u>	-	-	-	-	-	-
<u>Paxillosida msp 3</u>	-	<u>0.016</u>	<u>0.025</u>	-	-	-	-	-	-

<u>Paxillosida msp 4</u>	-	<u>0.016</u>	-	-	-	-	-	-	-	-
<u>Paxillosida msp 1</u>	-	-	-	-	-	-	-	<u>0.028</u>	-	-
<u>Paxillosida indet.</u>	-	<u>0.133</u>	-	-	-	-	-	-	-	-
<u>Velatida</u>	-	-	-	-	-	-	-	-	-	-
<u>Pterasteridae</u>	-	-	-	-	-	-	-	-	-	-
<u>Hymenaster msp 2</u>	<u>0.009</u>	-	-	-	-	-	-	-	-	-
<u>Pteraster msp</u>	<u>0.027</u>	-	-	-	-	-	-	-	-	-
<u>Velatida cf. AST 014</u>	<u>0.018</u>	<u>0.039</u>	-	-	-	-	-	-	-	-
<u>Velatida msp 2</u>	-	-	-	-	-	-	<u>0.059</u>	-	-	-
<u>Velatida msp 3</u>	-	-	-	-	-	-	-	-	<u>0.034</u>	-
<u>Asteroidea indet.</u>	<u>0.063</u>	<u>0.086</u>	<u>0.160</u>	<u>0.020</u>	<u>0.057</u>	-	-	-	-	-
<u>Crinoidea</u>	-	-	-	-	-	-	-	-	-	-
<u>Comatulida</u>	-	-	-	-	-	-	-	-	-	-
<u>Bourgueticrinina msp 1</u>	-	-	-	-	<u>0.094</u>	-	-	-	<u>0.067</u>	-
<u>Comatulida msp 1</u>	<u>0.261</u>	<u>0.313</u>	-	-	-	-	<u>0.059</u>	-	-	-
<u>Comatulida msp 2</u>	-	-	-	-	-	-	-	-	<u>0.067</u>	-
<u>Hyocrinida</u>	-	-	-	-	-	-	-	-	-	-
<u>Hyocrinidae small msp</u>	-	-	-	-	<u>0.113</u>	<u>0.125</u>	-	-	-	-
<u>Hyocrinidae msp 1</u>	-	<u>0.039</u>	<u>0.012</u>	-	-	-	-	-	-	-
<u>Crinoidea red msp</u>	<u>0.027</u>	<u>0.329</u>	-	<u>0.081</u>	-	-	-	-	-	-
<u>Crinoidea golden msp</u>	<u>0.018</u>	<u>0.078</u>	-	-	-	-	-	-	-	-
<u>Crinoidea msp 1</u>	-	-	-	-	-	-	-	-	<u>0.034</u>	-
<u>Crinoidea indet.</u>	<u>0.009</u>	<u>0.094</u>	<u>0.012</u>	<u>0.020</u>	-	<u>0.062</u>	-	-	-	-
<u>Echinoidea</u>	-	-	-	-	-	-	-	-	-	-
<u>Aspidodiadematidae</u>	-	-	-	-	-	-	-	-	-	-
<u>Aspidodiadematidae msp 1</u>	-	-	-	-	<u>1.187</u>	<u>1.186</u>	<u>0.713</u>	-	-	-
<u>Aspidodiadematidae msp 2</u>	-	<u>0.039</u>	-	-	-	<u>0.062</u>	-	-	-	-
<u>Aspidodiadematidae soft msp</u>	-	-	-	<u>0.020</u>	-	-	-	-	-	-
<u>Aspidodiadematidae spiny msp</u>	<u>0.018</u>	-	-	<u>0.141</u>	-	-	-	-	-	-
<u>Urechinidae</u>	-	-	-	-	-	-	-	-	-	-
<u>Urechinidae msp 1 Nodules</u>	-	-	-	-	-	-	-	-	<u>0.034</u>	-

<u>Urechinidae msp 3</u>	<u>0.027</u>	<u>0.008</u>	<u>0.135</u>	-	-	-	-	-	-	<u>0.034</u>
<u>Urechinidae msp 2 Nodules</u>	-	-	-	-	-	-	-	-	-	-
<u>Urechinidae msp 3 Nodules</u>	-	-	-	-	<u>0.019</u>	-	-	-	-	-
<u>Urechinidae msp 4 Nodules</u>	-	-	-	-	-	-	-	<u>0.028</u>	<u>0.067</u>	-
<u>Urechinidae msp 1</u>	<u>0.027</u>	<u>0.149</u>	<u>0.037</u>	-	-	-	-	-	-	-
<u>Urechinidae msp 2</u>	<u>0.027</u>	<u>0.008</u>	-	-	-	-	-	-	-	-
<u>Urechinidae msp 4</u>	<u>0.063</u>	<u>0.282</u>	<u>0.061</u>	-	-	-	-	-	-	-
<u>Urechinidae msp 5</u>	<u>0.009</u>	-	-	-	-	-	-	-	-	-
<u>Urechinidae msp 6</u>	<u>0.009</u>	-	-	-	-	-	-	-	-	-
<u>Urechinidae msp 7</u>	<u>0.009</u>	-	-	-	-	-	-	-	-	-
<u>Urechinidae indet.</u>	<u>0.018</u>	<u>0.023</u>	<u>0.012</u>	-	-	-	-	-	-	-
<u>Echinoidea indet.</u>	<u>0.009</u>	-	-	-	-	-	-	-	-	-
<u>Holothuroidea</u>	-	-	-	-	-	-	-	-	-	-
<u>Elasipodida</u>	-	-	-	-	-	-	-	-	-	-
<u>Elpidiidae</u>	-	-	-	-	-	-	-	-	-	-
<u>Elpidiidae double-velum msp</u>	-	-	-	-	-	-	<u>0.059</u>	-	-	-
<u>Elpidiidae msp 1</u>	-	-	-	-	-	-	-	<u>0.028</u>	<u>0.034</u>	-
<u>Amperima msp</u>	<u>0.018</u>	-	-	-	-	-	-	-	-	-
<u>Amperima msp 1</u>	-	-	-	-	<u>0.019</u>	-	-	-	-	-
<u>Peniagone "palmata" msp</u>	-	-	-	-	-	<u>0.062</u>	<u>0.119</u>	-	-	-
<u>Peniagone "tulip" msp</u>	-	-	-	-	-	-	<u>0.059</u>	-	-	-
<u>Peniagone cf. leander</u>	-	-	-	-	-	<u>0.062</u>	<u>0.059</u>	-	-	-
<u>Peniagone msp</u>	<u>0.018</u>	<u>0.016</u>	-	-	-	-	-	-	-	-
<u>Peniagone purple msp</u>	-	-	-	-	-	-	-	-	<u>0.034</u>	-
<u>Peniagone white/transparent msp</u>	-	-	-	-	<u>0.019</u>	-	-	<u>0.028</u>	<u>0.034</u>	-
<u>Peniagone indet.</u>	-	-	-	-	<u>0.038</u>	-	-	-	-	-
<u>Laetmogonidae</u>	-	-	-	-	-	-	-	-	-	-
<u>Laetmogonidae msp 1</u>	<u>0.036</u>	<u>0.094</u>	-	-	-	-	-	-	-	-
<u>Laetmogonidae msp 2</u>	<u>0.027</u>	-	-	-	-	-	-	-	-	-
<u>Laetmogonidae msp 3</u>	-	-	-	-	-	-	<u>0.059</u>	-	<u>0.034</u>	-
<u>Pelagothuriidae</u>	-	-	-	-	-	-	-	-	-	-

<u>Enypniastes sp.</u>	-	-	-	-	-	-	-	-	<u>0.034</u>
<u>Psychropotidae</u>	-	-	-	-	-	-	-	-	-
<u>Benthodytes cf. incertae purple msp</u>	-	<u>0.031</u>	<u>0.012</u>	-	-	-	-	-	-
<u>Benthodytes cf. incertae red msp</u>	-	<u>0.086</u>	-	-	-	-	-	-	-
<u>Benthodytes msp</u>	-	<u>0.039</u>	-	-	-	-	-	-	-
<u>Benthodytes msp 1</u>	-	-	-	-	-	-	-	-	<u>0.034</u>
<u>Benthodytes pink msp</u>	-	-	-	<u>0.020</u>	-	-	-	-	-
<u>Benthodytes purple msp</u>	-	-	<u>0.012</u>	-	-	-	-	-	-
<u>Benthodytes red msp</u>	-	<u>0.008</u>	<u>0.012</u>	-	-	-	-	-	-
<u>Psychropotes cf. semperiana</u>	-	-	-	-	-	-	-	<u>0.028</u>	-
<u>Psychropotes longicauda</u>	-	-	-	-	-	-	<u>0.119</u>	-	-
<u>Psychropotes msp 3</u>	-	-	-	-	<u>0.019</u>	-	<u>0.059</u>	-	-
<u>Psychropotes verrucosa</u>	-	-	-	-	<u>0.075</u>	<u>0.062</u>	-	-	-
<u>Psychropotidae msp 1 Nodules</u>	-	-	-	-	<u>0.019</u>	<u>0.062</u>	<u>0.059</u>	-	-
<u>Psychropotidae msp 1</u>	-	<u>0.070</u>	<u>0.012</u>	-	-	-	-	-	-
<u>Psychropotidae msp 2 Nodules</u>	-	-	-	-	-	<u>0.187</u>	-	-	-
<u>Psychropotidae msp 2</u>	-	<u>0.008</u>	-	-	-	-	-	-	-
<u>Psychropotidae msp 3</u>	-	-	-	-	<u>0.038</u>	<u>0.062</u>	-	-	-
<u>Psychropotidae msp 4</u>	-	-	-	-	-	<u>0.062</u>	-	-	-
<u>Psychropotidae red msp</u>	<u>0.018</u>	-	-	-	-	-	-	-	-
<u>Psychropotidae indet.</u>	<u>0.162</u>	<u>0.086</u>	-	<u>0.020</u>	-	-	-	-	-
<u>Holothuriida</u>	-	-	-	-	-	-	-	-	-
<u>Mesothuriidae</u>	-	-	-	-	-	-	-	-	-
<u>Mesothuria msp</u>	<u>0.009</u>	<u>0.023</u>	-	-	-	-	-	-	-
<u>Synallactida</u>	-	-	-	-	-	-	-	-	-
<u>Deimatidae</u>	-	-	-	-	-	-	-	-	-
<u>Deima msp.</u>	-	<u>0.008</u>	-	-	-	-	-	-	-
<u>Deimatidae - irregular papillae length msp</u>	-	<u>0.055</u>	<u>0.012</u>	-	-	-	-	-	-
<u>Oneirophanta msp</u>	<u>0.009</u>	-	<u>0.025</u>	-	-	-	-	-	-
<u>Deimatidae indet.</u>	-	<u>0.008</u>	<u>0.012</u>	-	-	-	-	-	-
<u>Synallactidae</u>	-	-	-	-	-	-	-	-	-

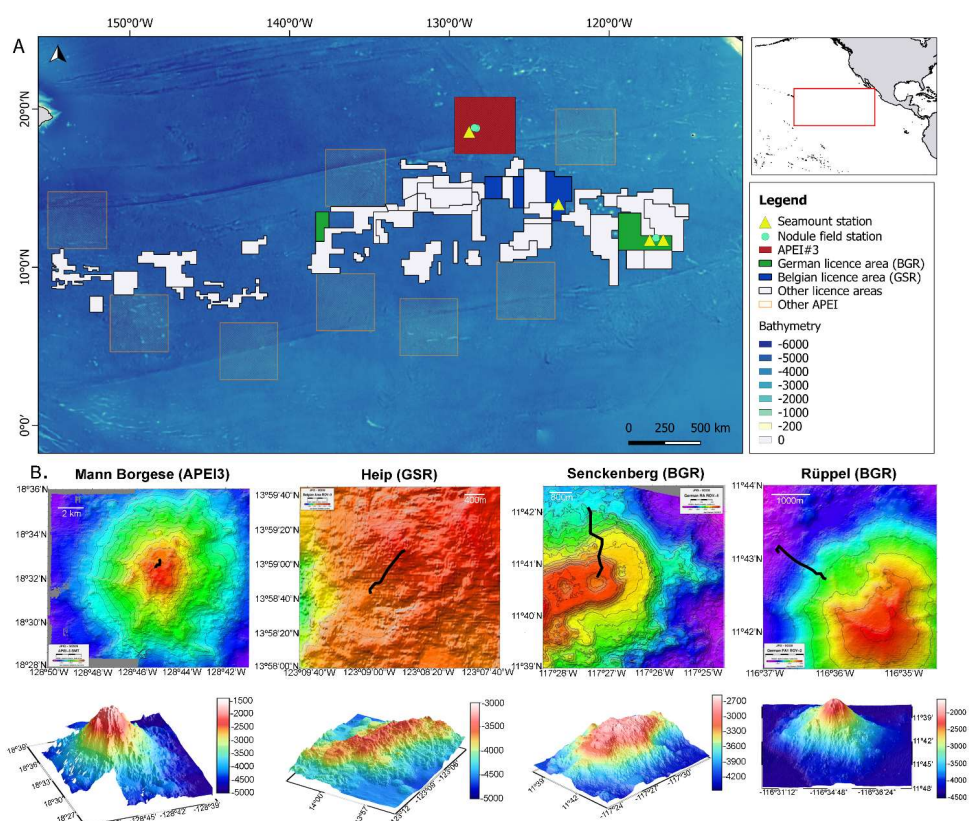
<u>Benthothuria msp</u>	-	-	-	<u>0.081</u>	-	-	-	-	-
<u>Paelopatides "orange" msp</u>	<u>0.009</u>	<u>0.008</u>	-	-	-	-	-	-	-
<u>Synallactes msp 1 (Synallactidae purple msp)</u>	<u>0.009</u>	-	-	-	-	-	-	-	-
<u>Synallactes msp 2</u>	-	<u>0.008</u>	-	-	-	-	-	-	-
<u>Synallactes msp 2 pink</u>	-	-	-	-	<u>0.038</u>	<u>0.250</u>	<u>0.059</u>	-	-
<u>Synallactes msp 2 pink (smooth)</u>	<u>0.027</u>	<u>0.016</u>	-	-	-	<u>0.312</u>	-	-	-
<u>Synallactes sandy-coloured msp</u>	<u>0.018</u>	-	-	-	-	-	-	-	-
<u>Synallactes white msp</u>	<u>0.018</u>	-	-	-	<u>0.697</u>	<u>0.187</u>	<u>0.297</u>	-	<u>0.034</u>
<u>Synallactidae indet.</u>	<u>0.036</u>	-	-	-	-	-	-	-	-
<u>Persiculida</u>	-	-	-	-	-	-	-	-	-
<u>Molpadiodemiae</u>	-	-	-	-	-	-	-	-	-
<u>Molpadiodemas msp</u>	-	<u>0.023</u>	-	-	-	-	-	-	-
<u>Pseudostichopodidae</u>	-	-	-	-	-	-	-	-	-
<u>Pseudostichopus msp</u>	-	-	-	-	-	<u>0.062</u>	-	-	-
<u>Molpadiodemas/Mesothuria</u>	-	-	-	-	<u>0.057</u>	<u>0.125</u>	<u>0.059</u>	-	<u>0.067</u>
<u>Holothuroidea indet.</u>	<u>0.171</u>	<u>0.149</u>	<u>0.037</u>	<u>0.040</u>	<u>0.057</u>	<u>0.062</u>	<u>0.119</u>	-	-
<u>Ophiuroidea</u>	-	-	-	-	-	-	-	-	-
<u>Ophiuroidea msp 1</u>	-	-	-	-	<u>0.019</u>	<u>0.062</u>	<u>0.059</u>	-	-
<u>Ophiuroidea msp 3</u>	-	-	-	-	-	<u>0.125</u>	-	-	-
<u>Ophiuroidea msp 5</u>	<u>0.018</u>	<u>0.391</u>	<u>0.491</u>	-	-	-	-	<u>0.028</u>	<u>0.101</u>
<u>Ophiuroidea msp 6</u>	-	<u>0.031</u>	<u>0.012</u>	-	<u>0.320</u>	<u>1.310</u>	<u>0.653</u>	<u>0.168</u>	<u>0.436</u>
<u>Ophiuroidea msp 4</u>	<u>0.036</u>	<u>0.211</u>	-	-	<u>0.113</u>	-	-	-	-
<u>Ophiuroidea msp7</u>	-	<u>0.008</u>	-	-	-	-	-	-	-
<u>Ophiuroidea indet.</u>	-	<u>0.023</u>	<u>0.037</u>	<u>0.081</u>	<u>5.669</u>	<u>6.677</u>	<u>7.307</u>	-	<u>0.134</u>
-	-	-	-	-	-	-	-	-	-
<u>Enteropneusta</u>	-	-	-	-	-	-	-	-	-
<u>Enteropneusta msp 1 cf. Yoda</u>	-	<u>0.102</u>	-	-	-	-	-	-	-
<u>Enteropneusta msp 2 cf. Saxipendum msp.</u>	<u>0.072</u>	-	-	-	-	-	-	-	-
-	-	-	-	-	-	-	-	-	-
<u>Mollusca</u>	-	-	-	-	-	-	-	-	-
<u>Gastropoda</u>	-	-	-	-	-	-	-	-	-

<u>Limpet</u>	-	-	<u>0.012</u>	-	-	-	-	-	-
<u>Gastropoda msp 1</u>	-	-	<u>0.025</u>	-	-	-	-	-	-
<u>Polyplocophora</u>	<u>0.036</u>	-	-	<u>0.040</u>	-	-	-	-	<u>0.067</u>
<u>Gastropoda indet.</u>	-	-	-	-	-	<u>0.062</u>	-	-	-
<u>Cephalopoda</u>	-	-	-	-	-	-	-	-	-
<u>Octopoda msp 1</u>	<u>0.009</u>	-	-	-	-	-	-	-	-
<u>Pisces*</u>	<u>0.333</u>	<u>0.282</u>	<u>0.074</u>	<u>0.060</u>	<u>0.471</u>	<u>0.187</u>	<u>0.475</u>	<u>0.168</u>	<u>0.168</u>
	-	-	-	-	-	-	-	-	-
<u>Porifera</u>	-	-	-	-	-	-	-	-	-
<u>Demospongiae</u>	-	-	-	-	-	-	-	-	-
<u>Cladorhizidae</u>	-	-	-	-	-	-	-	-	-
<u>Cladorhizidae msp 1</u>	-	-	<u>0.025</u>	-	-	-	-	-	<u>0.067</u>
<u>Cladorhizidae msp 1(soft)</u>	-	-	-	-	-	-	-	-	<u>0.034</u>
<u>Cladorhizidae msp 2</u>	-	-	-	-	-	-	-	<u>0.028</u>	<u>0.034</u>
<u>Cladorhizidae msp 3</u>	-	-	-	-	-	-	-	-	<u>0.034</u>
<u>Cladorhizidae msp 4</u>	-	-	-	-	<u>0.019</u>	-	-	<u>0.056</u>	<u>0.134</u>
<u>Cladorhizidae msp 5</u>	-	-	-	-	-	<u>0.062</u>	-	-	-
<u>Cladorhizidae msp 6</u>	-	-	-	-	-	<u>0.062</u>	-	-	-
<u>Cladorhizidae indet</u>	-	-	-	-	-	-	-	<u>0.028</u>	<u>0.067</u>
<u>Hexactellinida</u>	-	-	-	-	-	-	-	-	-
<u>Euplectellidae</u>	-	-	-	-	-	-	-	-	-
<u>Bathydorus spinosus</u>	<u>0.009</u>	-	-	-	-	-	-	-	-
<u>Bolosoma sp.</u>	-	-	-	<u>0.020</u>	-	-	-	-	-
<u>Corbitella discasterosa</u>	<u>0.009</u>	-	-	-	-	-	-	<u>0.056</u>	-
<u>Docosaccus maculatus</u>	-	-	-	-	-	<u>0.062</u>	-	<u>0.028</u>	-
<u>Docosaccus nidulus</u>	-	-	-	-	-	<u>0.062</u>	-	-	-
<u>Holascusspp</u>	-	-	-	-	<u>0.188</u>	<u>0.125</u>	<u>0.059</u>	<u>0.028</u>	<u>0.034</u>
<u>Hyalostylus schulzei</u>	-	-	-	-	-	-	-	<u>0.028</u>	-
<u>Hyalostylus sp.</u>	-	<u>0.016</u>	<u>0.147</u>	<u>0.020</u>	-	-	-	<u>0.028</u>	<u>0.034</u>
<u>Sacocalyx pedunculatus</u>	-	-	-	<u>0.060</u>	-	-	-	-	-

<u>Sacocalyx sp.</u>	<u>0.036</u>	<u>0.023</u>	<u>0.025</u>	-	-	-	-	-	-
<u>Euretidae</u>	-	-	-	-	-	-	-	-	-
<u>Bathyxiphus subtilis</u>	-	-	-	-	-	-	-	<u>0.028</u>	-
<u>Chonelasma bispinula</u>	-	-	-	-	-	-	<u>0.059</u>	-	-
<u>Chonelasma choanoides</u>	<u>0.009</u>	-	-	-	-	-	-	-	-
<u>Chonelasma sp.</u>	-	-	<u>0.012</u>	-	-	-	-	-	-
<u>Hyalonematidae</u>	-	-	-	-	-	-	-	-	-
<u>Hyalonema spp.</u>	-	<u>0.016</u>	<u>0.098</u>	<u>0.020</u>	<u>0.113</u>	<u>0.312</u>	<u>0.238</u>	<u>0.084</u>	<u>0.235</u>
<u>Rossellidae</u>	-	-	-	-	-	-	-	-	-
<u>Caulophacus sp.</u>	-	<u>0.063</u>	<u>0.074</u>	<u>0.020</u>	<u>0.170</u>	<u>0.062</u>	<u>0.059</u>	-	-
<u>Crateromorpha sp.</u>	-	<u>0.016</u>	-	<u>0.020</u>	-	-	-	-	-
<u>Rossellidae gen. sp.</u>	<u>0.036</u>	<u>0.008</u>	<u>0.025</u>	-	-	-	-	-	-
<u>Pheronematidae</u>	-	-	-	-	-	-	-	-	-
<u>Poliopogon sp.</u>	-	-	-	<u>0.020</u>	-	-	-	-	-
<u>Hexactellinida/foliose sponge msp</u>	<u>0.009</u>	<u>0.023</u>	<u>0.012</u>	<u>0.060</u>	-	-	-	-	-
<u>Hexactellinida - Stalked</u>	-	-	-	-	<u>0.264</u>	<u>0.499</u>	<u>0.535</u>	-	-
<u>Hexactinellida black msp</u>	-	<u>0.008</u>	-	-	-	-	-	-	-
<u>Hexactellinida indet.</u>	<u>0.198</u>	<u>0.203</u>	<u>0.515</u>	<u>0.121</u>	<u>0.979</u>	<u>1.061</u>	<u>1.604</u>	<u>0.447</u>	<u>0.369</u>
<u>Pycnogonida</u>	<u>0.018</u>	-	<u>0.012</u>	-	-	-	-	-	<u>0.034</u>
	-	-	-	-	-	-	-	-	-
<u>Tunicata</u>	-	-	-	-	-	-	-	-	-
<u>Octacnemidae</u>	-	-	-	-	-	-	-	-	-
<u>Megalodicopia msp. 1</u>	<u>0.018</u>	<u>0.008</u>	<u>0.012</u>	-	-	-	-	-	-
<u>Megalodicopia msp. 2</u>	-	-	-	-	-	-	-	-	<u>0.034</u>
<u>Dicopia msp.</u>	<u>0.036</u>	-	-	-	-	-	-	-	-
<u>Pyuridae</u>	-	-	-	-	-	-	-	-	-
<u>Culeolus msp.</u>	-	-	-	-	-	-	-	-	<u>0.034</u>
<u>Tunicata indet.</u>	<u>0.02</u>	<u>0.01</u>	<u>0.01</u>	<u>0.02</u>	-	-	-	-	-
	-	-	-	-	-	-	-	-	-
<u>Paleodictyon nodosum</u>	-	-	-	-	-	-	-	<u>0.03</u>	-

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Figures



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Fig. 1. (A). Location of the Clarion-Clipperton Fracture zone in the equatorial eastern Pacific Ocean featuring the contract areas from the International Seabed Authority (ISA) and the positions of the sampled areas (seamounts and nodule fields). Information on transect length and depth gradients can be found in Table 1. (B). Location of the seamount transects carried out towards the summit on the north–north-western flank and seamount profiles. Rüppel (BGR, ROV02) and Mann Borgese (APEI3, ROV15) are single seamounts, while Senckenberg (BGR, ROV04) and Heip (GSR, ROV09) are sea-mountain ranges.

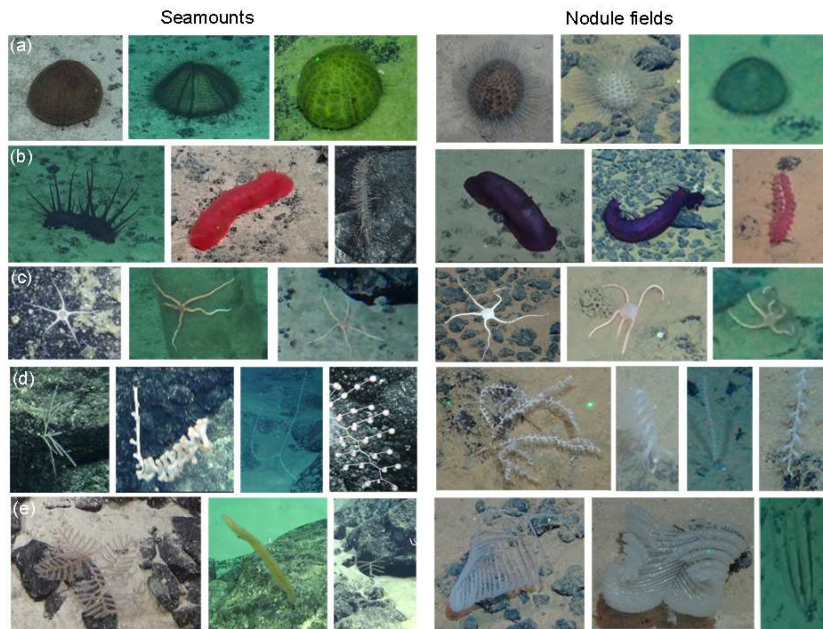
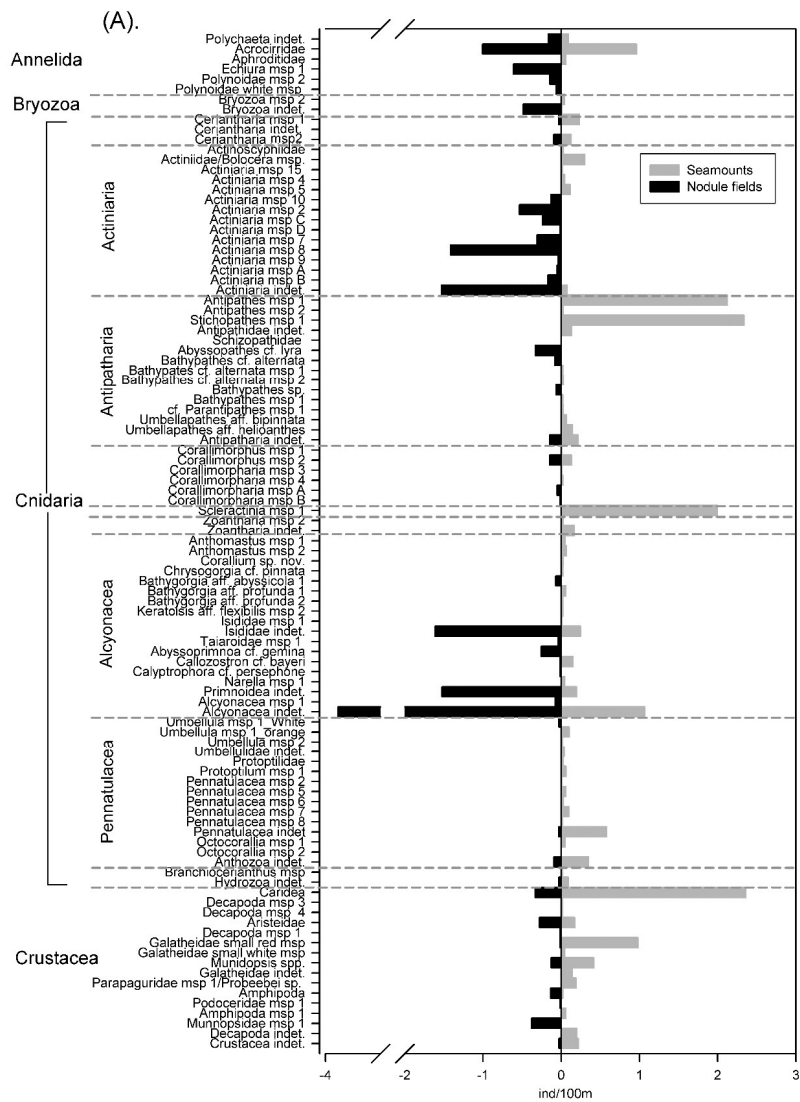
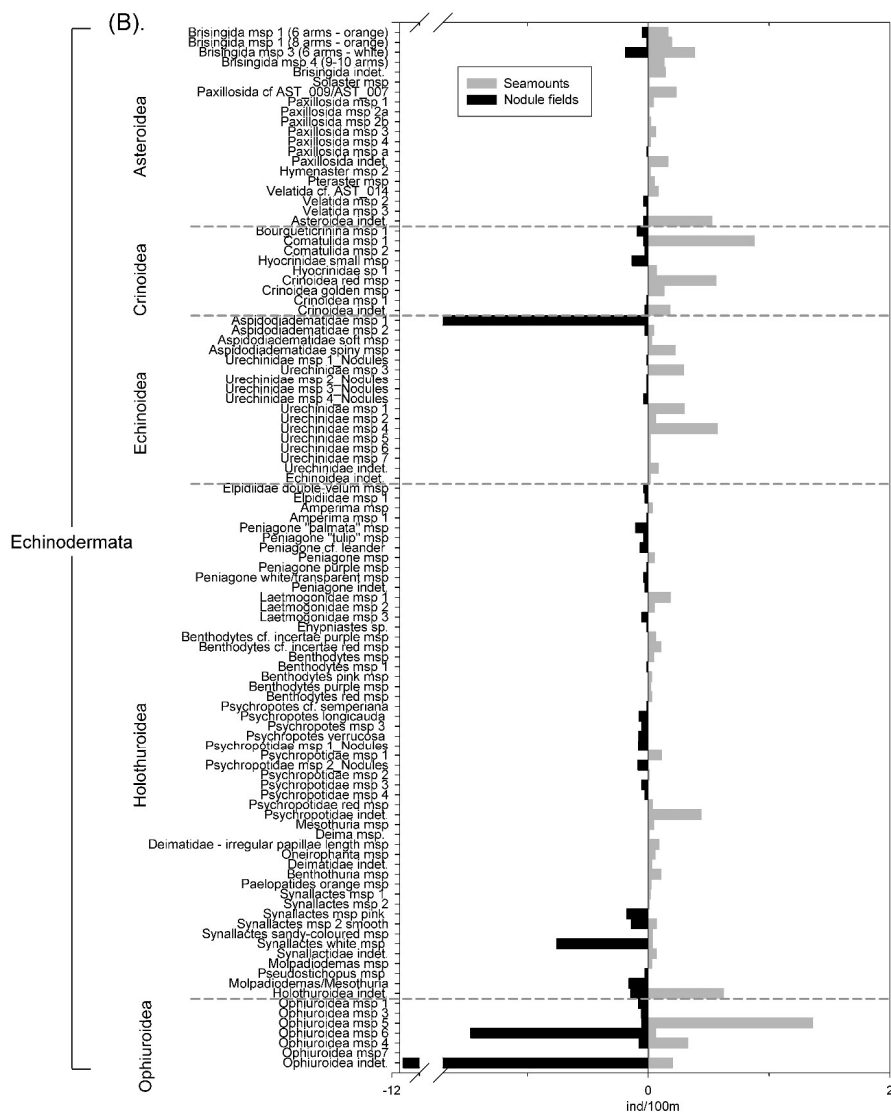
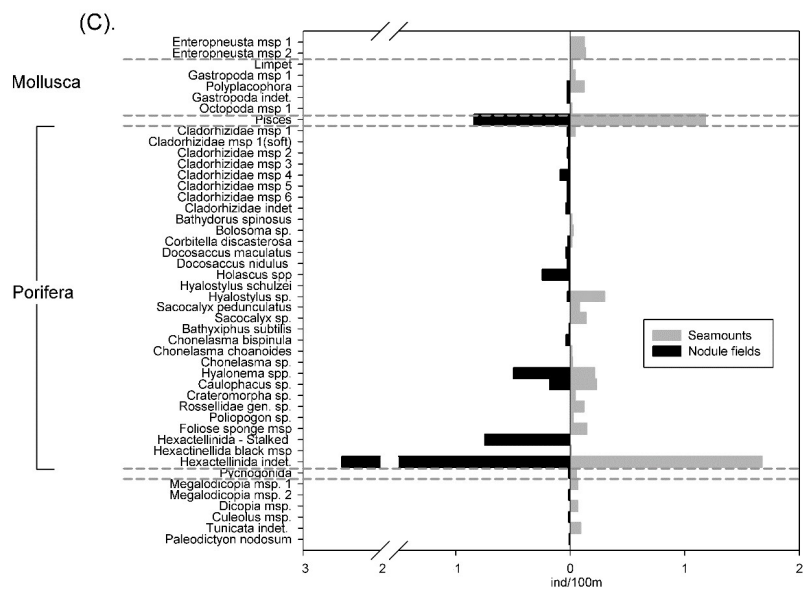


Fig. 2. Some examples of different morphospecies at seamounts and nodule fields in the CCZ. Selected taxa were (a) Echinoidea (from left to right, Urechinidae msp 4 (URC_019), Urechinidae msp 2 (URC_013), Urechinidae msp 3 (URC_009), Urechinidae msp. A (URC_020), Urechinidae msp. B (URC_021), Urechinidae msp. C (URC_005), (b) Holothuroidea (from left to right, Psychropotidae msp 1 (HOL_088), *Benthodytes* red msp. (HOL_101), Deimatidae - irregular papillae msp. (HOL_070), *Psychropotes verrucosa* (HOL_045), Laetmogonidae (HOL_030), *Synallactes* msp 2 pink (HOL_008)(c) Ophiuroidea (from left to right, Ophiuroidea msp. 5 (OPH_003), Ophiuroidea msp. 4 (OPH_005), Ophiuroidea msp. 6 (OPH_006), Ophiuroidea msp. 6 (OPH_006), Ophiuroidea (OPH_012), Ophiuroidea msp. 4 (OPH_005)), (d) Alcyonacea (from left to right, *Callozostron* cf. *bayeri* (ALC_009), *Bathygorgia* aff. *profunda* 2 (ALC_005), *Keratoisis* aff. *flexibilis* msp 2 (ALC_029), *Chrysogorgia* cf. *pinnata*, *Abyssoprinoa* cf. *gemina* (ALC_008), *Bathygorgia* aff. *profunda* 1, *Calyptrophora* cf. *persephone* (ALC_007), *Bathygorgia* aff. *abyssicola* 1 (ALC_003), (e) Antipatharia (*Umbellapathes* aff. *helioanthes* (ANT_018), cf. *Parantipathes* morphotype 1 (ANT_017), *Bathypates* cf. *alternata* msp 1 (ANT_010), *Bathypathes* cf. *alternata* (ANT_006), *Abyssopathes* cf. *lyra* (ANT_022), *Bathypathes* sp. (ANT_003)). Codes refer to an ongoing collaboration in creating one species catalogue for the CCZ and align all morphospecies of different research groups. Copyright: SO239, ROV Kiel 6000, GEOMAR Helmholtz Centre for Ocean Research Kiel







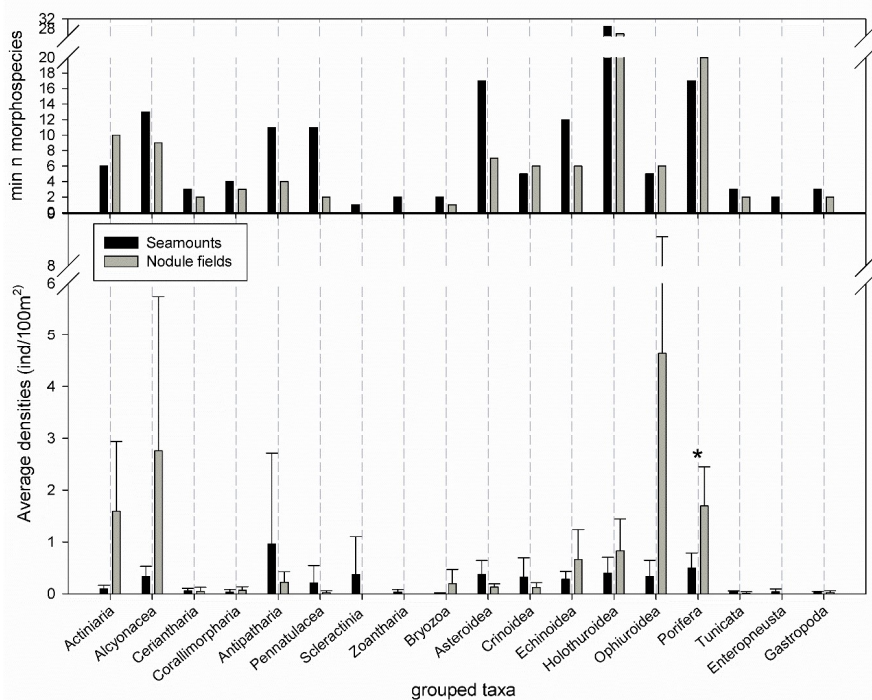
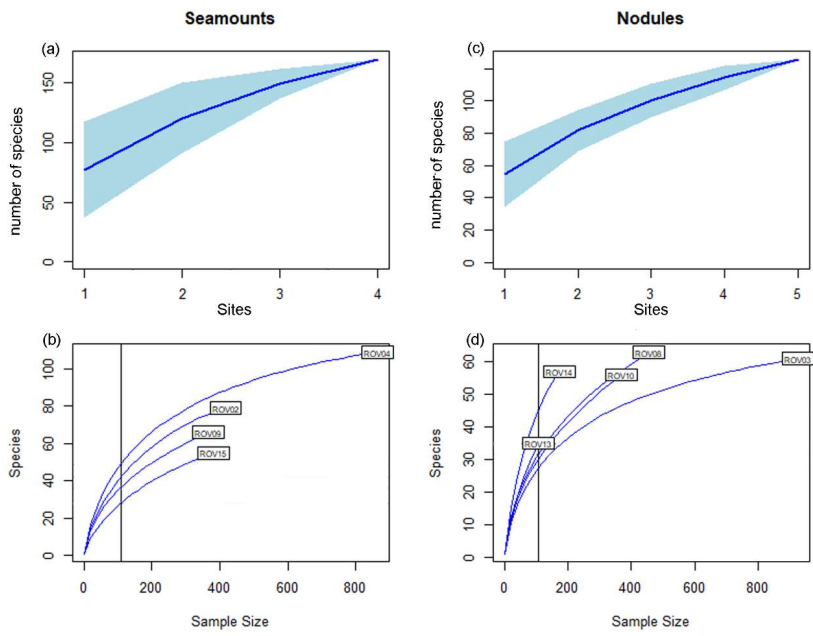


Fig. 3. Average densities at higher taxa level per ecosystem and standard deviation in the lower panel and minimum number of morphospecies per taxon and ecosystem in the upper panel. *= Significant difference in density ($t=-3.7$, $p<0.05$). Back to back histogram comparing average densities of morphospecies and taxa (ind/100m) for seamount (#4) and nodule field (#5) video transects. (a) Annelida, Bryozoa, Cnidaria and Crustacea, (B) Echinodermata and (C) Mollusca, Porifera, Hemichordata and Chordata (Tunicata).

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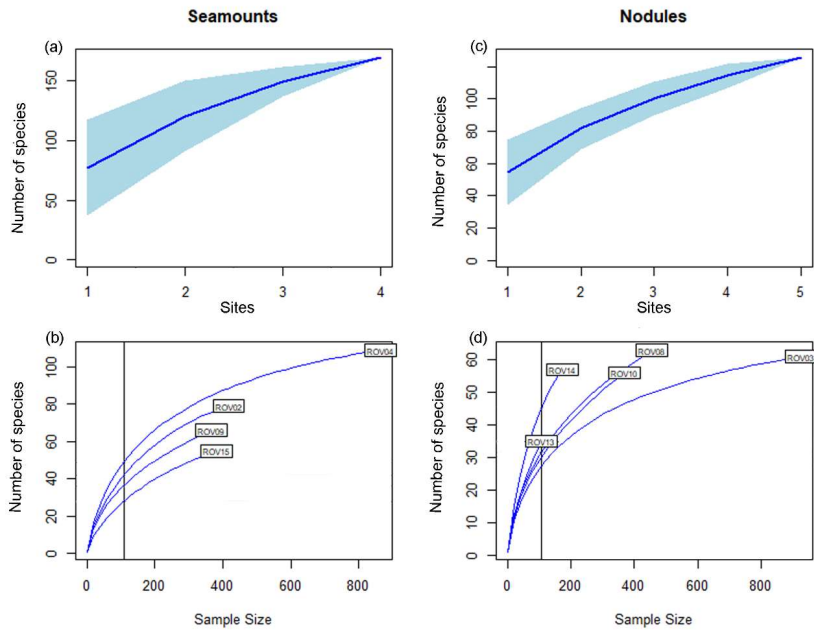
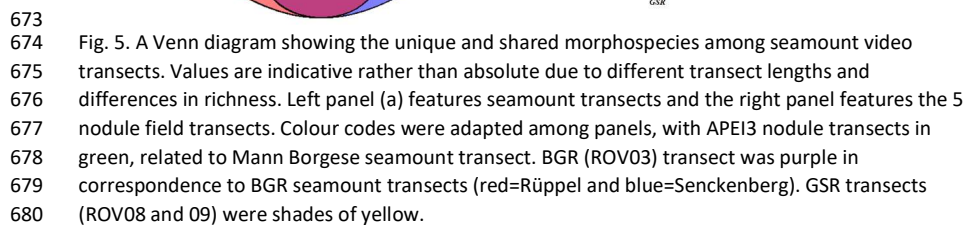
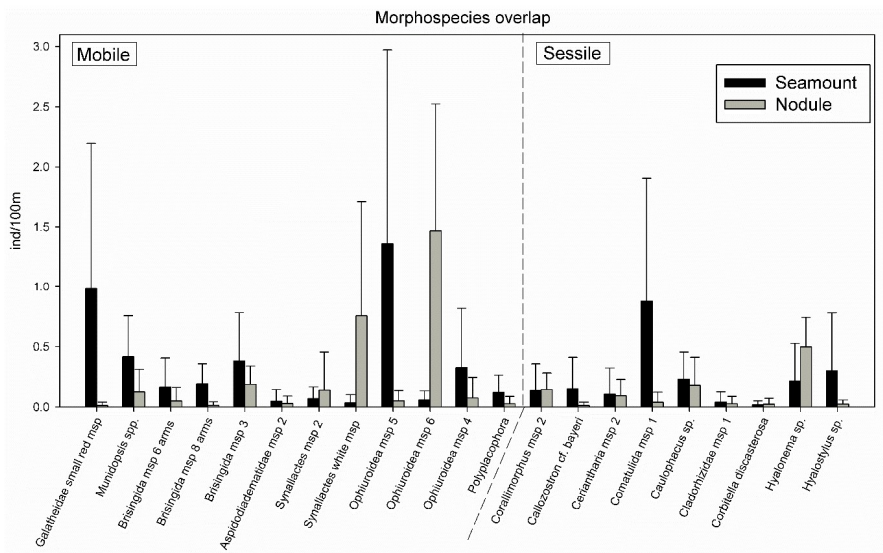


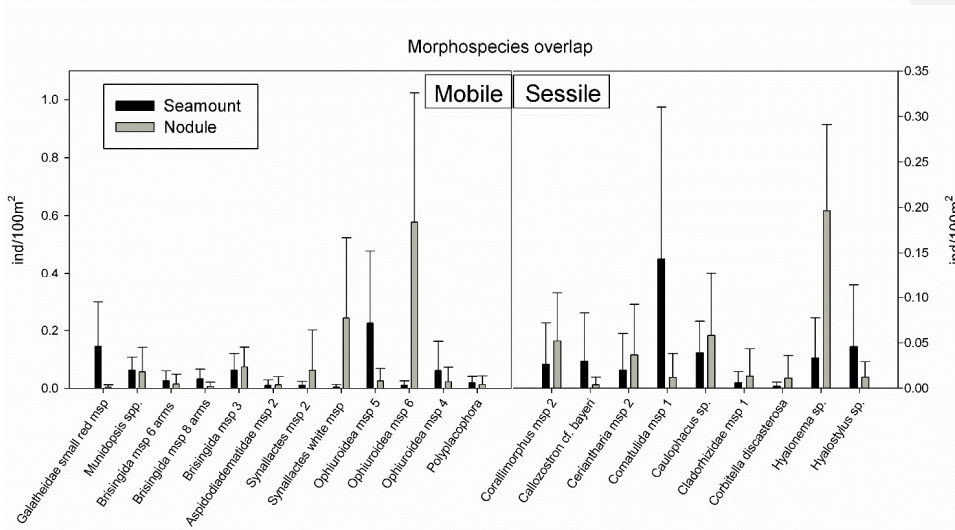
Fig. 4. Species accumulation (upper panel, a and c) and rarefaction curves (lower panel, b and d) for the seamount ($n=4$) and nodule field ($n=5$) transects. Seamount dives: ROV02= Rüppel (BGR), ROV04=Senckenberg (BGR), ROV09=Heip (GSR), ROV15=Mann Borgese (APEI3) in the lower left panel (b). Nodule field dives: ROV03 was carried out in the BGR area, ROV08 and 10 in the GSR area and ROV13 and 14 in the APEI3, presented in the lower right panel (d). Sample size is the number of individuals. Vertical line in the lower panel shows sample size=100.



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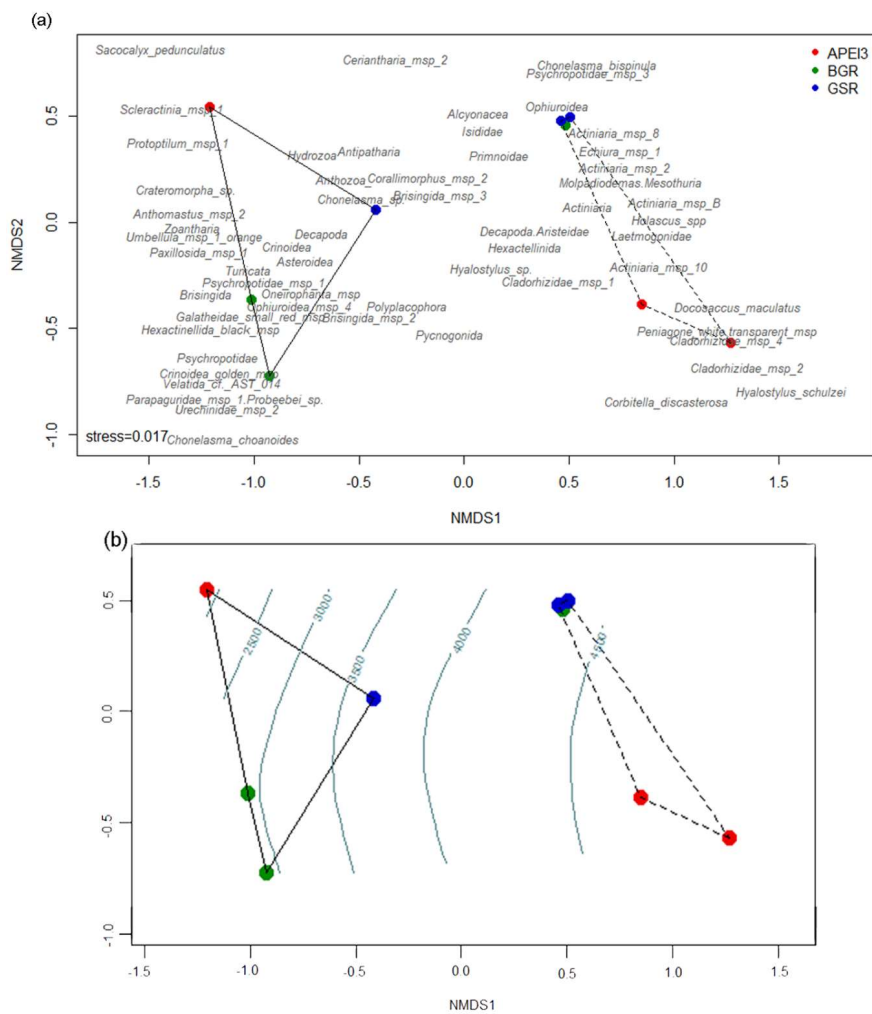
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688 Fig. 6. Morphospecies present in both seamounts and nodule field transects and their average
689 density (ind/100m²) and standard deviation per ecosystem.

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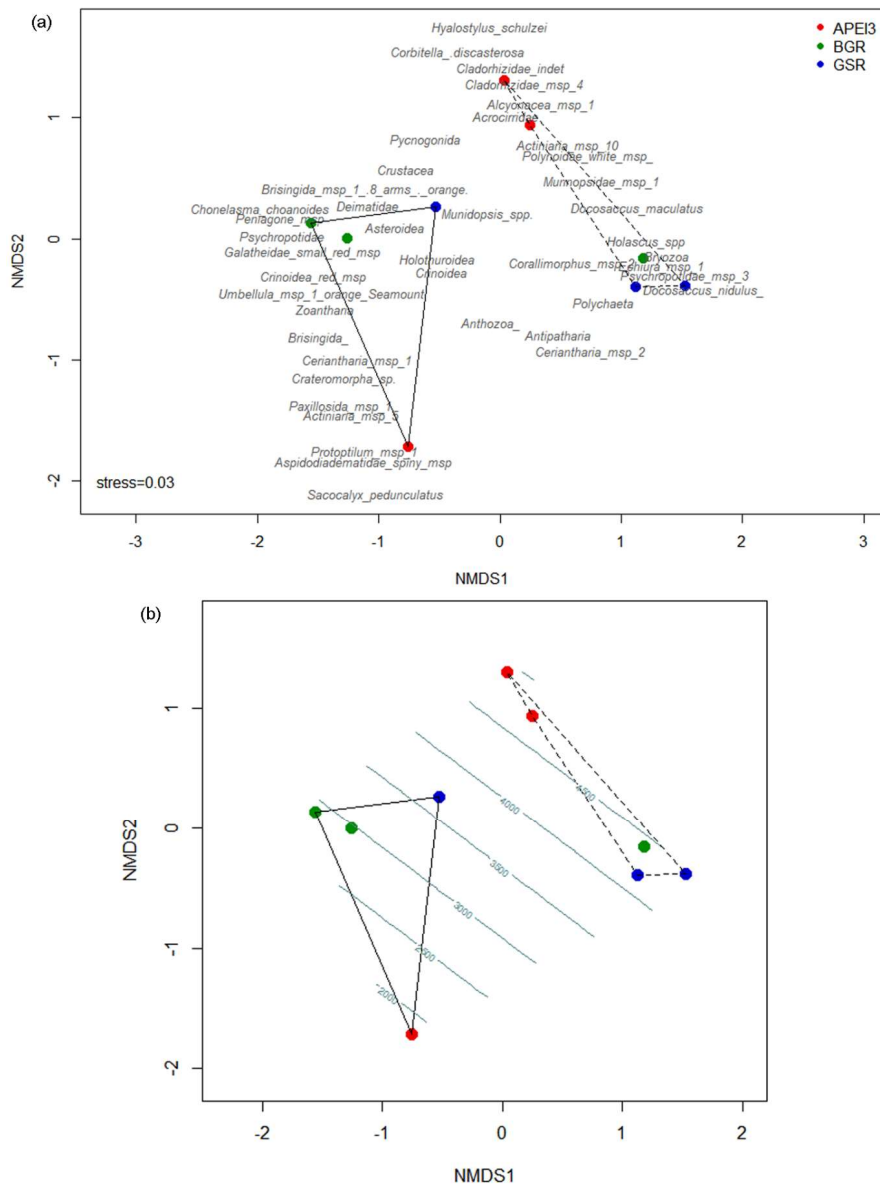


Fig. 7. nMDS-plot with faunal densities and Bray-Curtis distances. Upper panel (a) presents the grouping of the video transects based on their faunal composition and lower panel (b) features the

695 same plot but with depth as a vector fitting. Dotted lines group the nodule transects while the full
696 line groups the seamount transects.

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700 ~~Appendix~~

701 ~~Table A1- Overview of all densities (ind./100m) observed within each video transect. Higher taxa are~~
702 ~~in bold. * indicates taxa left out of the statistical analyses due to lack of representativity. Indets were~~
703 ~~organisms impossible to attribute to a lower taxonomic group. ROV02=Rüppel, ROV04=Senckenberg,~~
704 ~~ROV09=Heip, ROV15=Mann Borgese~~

	SEAMOUNTS				Formatted: Left: 2.54 cm, Right: 2.54 cm, Top: 3.49 cm, Bottom: 2.54 cm, Width: 21 cm, Height: 29.7 cm, Numbering: Continuous			
	ROV2 ind/100m	ROV4 ind/100m	ROV9 ind/100m	ROV15 ind/100m				
-								
Annelida*								
Polychaeta indet. * (No Serpulidae)	0.14	0.12	-	0.11	0.31	-	0.38	0.06
Acrocirridae	0.14	0.16	3.56	-	0.57	0.14	0.58	1.79
Aphroditidae	0.20	0.04	-	-	-	-	-	-
Echiura msp 1	-	-	-	-	0.57	1.13	1.15	-
Polynoidea	-	-	-	-	-	-	-	-
-Polynoidae msp 2	-	-	-	-	-	0.14	0.58	-
-Polynoidae white msp	-	-	-	-	-	0.14	-	0.06
-								
Bryozoa								
Bryozoa msp 2		-	0.17	-	-	-	-	-
Bryozoa indet.		0.038	-	-	0.44	1.55	0.19	0.11
-								
Cnidaria								
Anthozoa								
-Ceriantharia	-	-	-	-	-	-	-	-
-Ceriantharia msp 1	0.34	0.04	0.34	0.22	-	-	-	-
-Ceriantharia msp 2	-	-	-	0.43	-	0.28	0.19	-
-Ceriantharia indet.	-	-	0.08	-	-	-	-	-
-Hexacorallia								
-Actiniaria								
-Actinoscyphiidae	-	0.12	-	-	-	-	-	-
-Actiniidae/Bolocera msp.	1.02	0.19	-	-	-	-	-	-
-Actiniaria msp 15	0.07	-	-	-	-	-	-	-
-Actiniaria msp 4	-	0.08	-	0.11	-	-	-	-
-Actiniaria msp 5	0.07	0.08	-	0.32	-	-	-	-
-Actiniaria msp 10	-	-	-	-	0.31	-	-	-
-Actiniaria msp 2	-	-	-	-	1.07	1.13	0.19	-
-Actiniaria msp C	-	-	-	-	0.38	0.42	0.38	-
-Actiniaria msp D	-	-	-	-	0.06	-	-	-
-Actiniaria msp 7	-	-	-	-	0.63	0.14	0.58	0.06
-Actiniaria msp 8	-	-	-	-	0.13	3.66	3.08	-
-Actiniaria msp 9	-	-	-	-	-	-	0.19	-
-Actiniaria msp A	-	-	-	-	-	-	0.19	-
-Actiniaria msp B	-	-	-	-	0.25	0.14	0.38	0.06
-Actiniaria indet.	0.14	0.15	-	-	1.57	1.41	4.42	0.11
-Antipatharia								
-Antipathidae								
-Antipathes msp 1	-	-	-	8.49	-	-	-	-
-Antipathes msp 2	-	-	-	0.11	-	-	-	-
-Stichopathes msp 1	-	-	-	9.35	-	-	-	-
-Antipathidae indet.	-	-	-	0.54	-	-	-	-
-Schizopathidae								

Abyssopathes cf. lyra	-	-	-	-	0.50	0.56	0.58	-
Bathypathes cf. alternata	-	-	-	-	-	0.14	0.19	-
Bathypates cf. alternata msp 1	-	-	0.08	-	-	-	-	-
Bathypathes cf. alternata msp 2	-	0.12	-	-	-	-	-	-
Bathypathes sp.	-	-	-	-	0.19	0.14	-	-
Bathypathes msp 1	-	-	0.08	-	-	-	-	-
cf. Parantipathes msp 1	-	-	0.11	-	-	-	-	-
Umbellapathes aff. bipinnata	-	0.19	0.08	-	-	-	-	-
Umbellapathes aff. helioanthes	-	0.58	-	-	-	-	-	-
Antipatharia indet.	0.07	0.08	0.08	0.65	0.25	0.28	0.19	-
Corallimorpharia/Corallimorphidae	-	-	-	-	-	-	-	-
Corallimorphus msp 1	-	0.04	0.00	-	-	-	-	-
Corallimorphus msp 2	-	0.46	0.08	-	0.25	0.28	0.19	-
Corallimorpharia msp 3	-	0.04	-	-	-	-	-	-
Corallimorpharia msp 4	-	-	0.08	-	-	-	-	-
Corallimorpharia msp A	-	-	-	-	0.06	-	0.19	-
Corallimorpharia msp B	-	-	-	-	0.06	-	-	-
Scleractinia								
Scleractinia msp 1	0.14	-	-	7.85	-	-	-	-
Zoantharia								
Zoantharia msp 2	-	-	-	0.11	-	-	-	-
Zoantharia indet.	-	0.46	-	0.22	-	-	-	-
Octocorallia								
Alcyonacea								
Alcyoniidae								
Anthomastus msp 1	0.20	-	-	-	-	-	-	-
Anthomastus msp 2	0.00	0.15	-	0.11	-	-	-	-
Coralliidae								
Corallium sp. nov.	-	-	-	0.11	-	-	-	-
Chrysogorgiidae								
Chrysogorgia cf. pinnata	-	-	0.08	-	-	-	-	-
Isididae								
Bathygorgia aff. abyssicola 1	-	-	-	-	-	0.14	0.19	-
Bathygorgia aff. profunda 1	-	0.15	0.08	-	-	-	-	-
Bathygorgia aff. profunda 2	-	-	0.08	-	-	-	-	-
Keratoisis aff. flexibilis msp 2	-	-	0.08	-	-	-	-	-
Isididae msp 1	-	0.04	-	-	-	-	-	-
Isididae indet.	0.14	-	0.76	0.11	0.13	5.63	2.31	-
Taiaroidea								
Taiaroidea msp 1	-	-	-	-	-	-	0.19	-
Primnoidae								
Abyssoprinoa cf. gemina	-	-	-	-	-	0.70	0.58	-
Callozostron cf. bayeri	0.07	0.54	-	-	0.06	-	-	-
Calyptrophora cf. persephone	-	-	-	-	0.06	-	-	-
Narella msp 1	-	0.08	-	0.11	-	-	-	-
Primnoidea indet.	0.61	-	0.17	-	2.70	3.38	1.54	-

-Alcyonacea msp 1	-	-	-	-	0.13	-	-	0.11
-Alcyonacea indet.	-	0.15	1.44	2.69	8.93	6.62	4.04	-
<u>Pennatulacea</u>								
<u>Umbellulidae</u>								
-Umbellula msp 1_White	-	-	-	-	-	-	-	-
-Umbellula msp 1_orange	-	0.31	-	0.11	-	-	-	-
-Umbellula msp 2	-	0.08	-	-	-	-	-	-
-Umbellulidae indet.	-	0.15	-	-	-	-	-	-
-Protoptilidae	-	-	-	0.11	-	-	-	-
-Protoptilum msp 1	-	0.04	-	0.22	-	-	-	-
-Pennatulacea msp 2	-	0.04	-	-	-	-	-	-
-Pennatulacea msp 5	-	0.23	-	-	-	-	-	-
-Pennatulacea msp 6	-	0.12	-	-	-	-	-	-
-Pennatulacea msp 7	-	0.38	-	-	-	-	-	-
-Pennatulacea msp 8	-	0.08	-	-	-	-	-	-
-Pennatulacea indet.	0.14	2.08	-	0.11	-	0.14	-	-
-Octocorallia msp 1	-	-	-	0.22	-	-	-	-
-Octocorallia msp 2	-	-	-	-	-	-	-	-
Anthozoa indet.	0.14	0.12	0.51	0.65	0.13	0.14	0.19	-
Hydrozoa								
-Branchiocerianthus msp	-	0.08	-	-	-	-	-	-
Hydrozoa indet.	-	0.08	0.08	0.22	-	0.14	-	-
Crustacea*								
Decapoda								
-Caridea	3.47	2.54	3.22	0.22	0.19	-	1.15	0.11
-Decapoda msp 3	-	0.08	-	-	-	-	-	-
-Decapoda msp 4	0.07	-	-	-	-	-	-	-
-Decapoda/Aristeidae	0.07	0.08	-	0.54	0.06	0.56	0.58	0.11
-Decapoda msp 1	-	-	-	-	-	-	-	0.06
-Galatheididae								
-Galatheididae-small red msp	2.79	0.54	0.17	0.43	0.06	-	-	-
-Galatheididae-small white msp	0.07	0.12	-	-	-	-	-	-
-Munidopsis spp.	0.82	0.35	0.51	-	-	0.42	-	-
-Galatheididae indet.	0.14	0.15	0.17	0.11	-	-	-	-
-Parapaguridae								
-Parapaguridae msp 1/Probeebei sp.	0.54	0.23	-	-	-	-	-	-
Peracarida								
-Amphipoda	-	-	0.08	-	0.06	0.28	-	0.06
-Podoceridae msp 1	-	-	-	-	-	-	-	0.06
-Amphipoda msp 1	-	0.08	0.17	-	-	-	-	-
-Isopoda								
-Munnopsidae msp 1	-	-	-	-	0.57	0.42	0.19	0.17
Decapoda indet.	-	0.12	0.68	-	-	-	-	-
Crustacea indet.	0.07	0.31	0.51	-	-	-	-	-

Echinodermata									
Asteroidea									
<u>Brisingida</u>									
Brisingida msp 1 (6 arms—orange)	-	0.15	0.51	-	0.25	-	-	-	-
Brisingida msp 1 (8 arms—orange)	0.14	0.38	0.25	-	-	-	-	-	-
Brisingida msp 3 (6 arms—white)	-	0.38	0.93	0.22	0.19	0.42	0.19	-	-
Brisingida msp 4 (9–10 arms)	0.14	0.38	-	-	-	-	-	-	-
Brisingida indet.	0.27	0.08	-	0.22	-	-	-	-	-
<u>Paxillosida</u>									
Solaster msp	-	0.04	-	-	-	-	-	-	-
Paxillosida cf. AST_009/AST_007	-	0.50	0.42	-	-	-	-	-	-
Paxillosida msp 1	0.07	-	-	0.11	-	-	-	-	-
Paxillosida msp 2a	-	0.04	-	-	-	-	-	-	-
Paxillosida msp 2b	-	-	0.08	-	-	-	-	-	-
Paxillosida msp 3	-	0.08	0.17	-	-	-	-	-	-
Paxillosida msp 4	-	0.08	-	-	-	-	-	-	-
Paxillosida msp 1	-	-	-	-	-	-	-	-	0.06
Paxillosida indet.	-	0.65	-	-	-	-	-	-	-
<u>Velatida</u>									
<u>Pterasteridae</u>									
Hymenaster msp 2	0.07	-	-	-	-	-	-	-	-
Pteraster msp	0.20	-	-	-	-	-	-	-	-
Velatida cf. AST_014	0.14	0.19	-	-	-	-	-	-	-
Velatida msp 2	-	-	-	-	-	-	0.19	-	-
Velatida msp 3	-	-	-	-	-	-	-	-	-
Asteroidea indet.	0.48	0.42	1.10	0.11	0.19	-	-	-	-
Crinoidea									
<u>Comatulida</u>									
Bourgueticrinina msp 1	-	-	-	-	0.31	-	-	-	-
Comatulida msp 1	1.97	1.54	-	-	-	-	0.19	-	-
Comatulida msp 2	-	-	-	-	-	-	-	-	-
<u>Hyocrinida</u>									
Hyocrinidae small msp	-	-	-	-	0.38	0.28	-	-	-
Hyocrinidae msp 1	-	0.19	0.08	0.00	-	-	-	-	-
Crinoidea red msp	0.20	1.62	-	0.43	-	-	-	-	-
Crinoidea golden msp	0.14	0.38	-	-	-	-	-	-	-
Crinoidea msp 1	-	-	-	-	-	-	-	-	-
Crinoidea indet.	0.07	0.46	0.08	0.11	-	0.14	-	-	-
Echinoidea									
<u>Aspidodiadematidae</u>									
Aspidodiadematidae msp 1	-	-	-	-	3.96	2.68	2.31	-	-
Aspidodiadematidae msp 2	-	0.19	-	-	-	0.14	-	-	-
Aspidodiadematidae soft msp	-	-	-	0.11	-	-	-	-	-
Aspidodiadematidae spiny msp	0.14	-	-	0.75	-	-	-	-	-
<u>Urechinidae</u>									
Urechinidae msp 1_Nodules	-	-	-	-	-	-	-	-	-

Urechinidae msp 3	0.20	0.04	0.93	-	-	-	-	-
Urechinidae msp 2_Nodules	-	-	-	-	-	-	-	-
Urechinidae msp 3_Nodules	-	-	-	-	0.06	-	-	-
Urechinidae msp 4_Nodules	-	-	-	-	-	-	-	0.06
Urechinidae msp 1	0.20	0.73	0.25	-	-	-	-	-
Urechinidae msp 2	0.20	0.04	-	-	-	-	-	-
Urechinidae msp 4	0.48	1.38	0.42	-	-	-	-	-
Urechinidae msp 5	0.07	-	-	-	-	-	-	-
Urechinidae msp 6	0.07	-	-	-	-	-	-	-
Urechinidae msp 7	0.07	-	-	-	-	-	-	-
Urechinidae indet.	0.14	0.12	0.08	-	-	-	-	-
Echinoidea indet.	0.07	-	-	-	-	-	-	-
<u>Holothuroidea</u>								
<u>Elasipodida</u>								
<u>Elpidiidae</u>								
Elpidiidae double velum msp	-	-	-	-	-	-	0.19	-
Elpidiidae msp 1	-	-	-	-	-	-	-	0.06
Amperima msp	0.14	-	-	-	-	-	-	-
Amperima msp 1	-	-	-	-	0.06	-	-	-
Peniagone "palmata" msp	-	-	-	-	-	0.14	0.38	-
Peniagone "tulip" msp	-	-	-	-	-	-	0.19	-
Peniagone cf. leander	-	-	-	-	-	0.14	0.19	-
Peniagone msp	0.14	0.08	-	-	-	-	-	-
Peniagone purple msp	-	-	-	-	-	-	-	-
Peniagone white/transparent msp	-	-	-	-	0.06	-	-	0.06
Peniagone indet.	-	-	-	-	0.13	-	-	-
<u>Laetmogonidae</u>								
Laetmogonidae msp 1	0.27	0.46	-	-	-	-	-	-
Laetmogonidae msp 2	0.20	-	-	-	-	-	-	-
Laetmogonidae msp 3	-	-	-	-	-	-	0.19	-
<u>Pelagothuriidae</u>								
Enypniastes sp.	-	-	-	-	-	-	-	-
<u>Psychropotidae</u>								
Benthodytes cf. incertae purple msp	-	0.15	0.08	-	-	-	-	-
Benthodytes cf. incertae red msp	-	0.42	-	-	-	-	-	-
Benthodytes msp	-	0.19	-	-	-	-	-	-
Benthodytes msp 1	-	-	-	-	-	-	-	-
Benthodytes pink msp	-	-	-	0.11	-	-	-	-
Benthodytes purple msp	-	-	0.08	-	-	-	-	-
Benthodytes red msp	-	0.04	0.08	-	-	-	-	-
Psychropotes cf. semperiana	-	-	-	-	-	-	-	0.06
Psychropotes longicauda	-	-	-	-	-	-	0.38	-
Psychropotes msp 3	-	-	-	-	0.06	-	0.19	-
Psychropotes verrucosa	-	-	-	-	0.25	0.14	-	-
Psychropotidae msp 1_Nodules	-	-	-	-	0.06	0.14	0.19	-
Psychropotidae msp 1	-	0.35	0.08	-	-	-	-	-

Psychropotidae msp 2_Nodules	-	-	-	-	-	0.42	-	-
Psychropotidae msp 2	-	0.04	-	-	-	-	-	-
Psychropotidae msp 3	-	-	-	-	0.13	0.14	-	-
Psychropotidae msp 4	-	-	-	-	-	0.14	-	-
Psychropotidae red msp	0.14	-	-	-	-	-	-	-
Psychropotidae indet.	1.22	0.42	-	0.11	-	-	-	-
<u>Holothuriida</u>								
Mesothuriidae								
Mesothuria msp	0.07	0.12	-	-	-	-	-	-
<u>Synallactida</u>								
Deimatidae								
Deima msp.	-	0.04	-	-	-	-	-	-
Deimatidae irregular papillae length msp	-	0.27	0.08	-	-	-	-	-
Oneirophanta msp	0.07	-	0.17	-	-	-	-	-
Deimatidae indet.	-	0.04	0.08	-	-	-	-	-
Synallactidae								
Benthothuria msp	-	-	-	0.43	-	-	-	-
Paelopatides "orange" msp	0.07	0.04	-	-	-	-	-	-
Synallactes msp 1 (Synallactidae purple msp)	0.07	-	-	-	-	-	-	-
Synallactes msp 2	-	0.04	-	-	-	-	-	-
Synallactes msp 2 pink	-	-	-	-	0.13	0.56	0.19	-
Synallactes msp 2 pink (smooth)	0.20	0.08	-	-	-	0.70	-	-
Synallactes sandy coloured msp	0.14	-	-	-	-	-	-	-
Synallactes white msp	0.14	-	-	-	2.33	0.42	0.96	-
Synallactidae indet.	0.27	-	-	-	-	-	-	-
<u>Persiculida</u>								
Molpadiodemidae								
Molpadiodemias msp	-	0.12	-	-	-	-	-	-
Pseudostichopodidae								
Pseudostichopus msp	-	-	-	-	-	0.14	-	-
Molpadiodemias/Mesothuria	-	-	-	-	0.19	0.28	0.19	-
Holothuroidea indet.	1.29	0.73	0.25	0.22	0.19	0.14	0.38	-
<u>Ophiuroidea</u>								
Ophiuroidea msp 1	-	-	-	-	0.06	0.14	0.19	-
Ophiuroidea msp 3	-	-	-	-	-	0.28	-	-
Ophiuroidea msp 5	0.14	1.92	3.39	-	-	-	-	0.06
Ophiuroidea msp 6	-	0.15	0.08	-	1.07	2.96	2.12	0.34
Ophiuroidea msp 4	0.27	1.04	-	-	0.38	-	-	-
Ophiuroidea msp 7	-	0.04	-	-	-	-	-	-
Ophiuroidea indet.	-	0.12	0.25	0.43	18.93	15.07	23.65	-
-				-				
<u>Enteropneusta</u>								
Enteropneusta msp 1 cf. Yoda	-	0.50	-	-	-	-	-	-
Enteropneusta msp 2 cf. Saxipendum msp.	0.54	-	-	-	Formatted: English (United Kingdom)			
<u>Mollusca</u>								

Gastropoda									
-Limpet	-	-	0.08	-	-	-	-	-	-
-Gastropoda msp 1	-	-	0.17	-	-	-	-	-	-
-Polyplacophora	0.27	-	-	0.22	-	-	-	-	-
-Gastropoda indet.	-	-	-	-	-	0.14	-	-	-
Cephalopoda									
-Octopoda msp 1	0.07	-	-	-	-	-	-	-	-
Pisces*	2.52	1.38	0.51	0.32	1.57	0.42	1.54	0.34	
Porifera									
Demospongiae									
-Cladorhizidae									
-Cladorhizidae msp 1		-	0.17	-	-	-	-	-	-
-Cladorhizidae msp 1(soft)	-	-	-	-	-	-	-	-	-
-Cladorhizidae msp 2	-	-	-	-	-	-	-	0.06	-
-Cladorhizidae msp 3	-	-	-	-	-	-	-	-	-
-Cladorhizidae msp 4	-	-	-	-	0.06	-	-	-	0.11
-Cladorhizidae msp 5	-	-	-	-	-	0.14	-	-	-
-Cladorhizidae msp 6	-	-	-	-	-	0.14	-	-	-
-Cladorhizidae indet	-	-	-	-	-	-	-	-	0.06
Hexactellinida									
-Euplectellidae									
-Bathydorus spinosus	0.07	-	-	-	-	-	-	-	-
-Bolosoma sp.	-	-	-	0.11	-	-	-	-	-
-Corbitella discasterosa	0.07	-	-	-	-	-	-	-	0.11
-Docosaccus maculatus	-	-	-	-	-	0.14	-	-	0.06
-Docosaccus nidulus	-	-	-	-	-	0.14	-	-	-
-Holascusspp	-	-	-	-	0.63	0.28	0.19	-	0.06
-Hyalostylus schulzei	-	-	-	-	-	-	-	-	0.06
-Hyalostylus sp.	-	0.08	1.02	0.11	-	-	-	-	0.06
-Sacocalyx pedunculatus	-	-	-	0.32	-	-	-	-	-
-Sacocalyx sp.	0.27	0.12	0.17	-	-	-	-	-	-
-Euretidae									
-Bathyxiphus subtilis	-	-	-	-	-	-	-	-	0.06
-Chonelasma bispinula	-	-	-	-	-	-	0.19	-	-
-Chonelasma choanoides	0.07	-	-	-	-	-	-	-	-
-Chonelasma sp.	-	-	0.08	-	-	-	-	-	-
-Hyalonematidae									
-Hyalonema spp.	-	0.08	0.68	0.11	0.38	0.70	0.77	-	0.17
-Rossellidae									
-Caulophacus sp.	-	0.31	0.51	0.11	0.57	0.14	0.19	-	-
-Crateromorpha sp.	-	0.08	-	0.11	-	-	-	-	-
-Rossellidae gen. sp.	0.27	0.04	0.17	-	-	-	-	-	-
-Pheronematidae									
-Poliopogon sp.	-	-	-	0.11	-	-	-	-	-

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Hexactellinida/foiose sponge msp	0.07	0.12	0.08	0.32	-	-	-	-
Hexactellinida - Stalked	-	-	-	-	0.88	1.13	1.73	-
Hexactinellida black msp	-	0.04	-	-	-	-	-	-
Hexactellinida indet.	1.50	1.00	3.56	0.65	3.27	2.39	5.19	0.89
Pycnogonida	0.14	0.00	0.08	0.00				
Tunicata								
Octaenemidae								
-Megalodicopia msp. 1	0.14	0.04	0.08	-	-	-	-	-
-Megalodicopia msp. 2	-	-	-	-	-	-	-	-
-Dicopia msp.	0.27	-	-	-	-	-	-	-
Pyuridae								
-Culeolus msp.	-	-	-	-	-	-	-	-
Tunicata indet.	0.14	0.04	0.08	0.11	-	-	-	-
Paleodictyon nodosum		-	-	-	-	-	-	0.06

Appendix

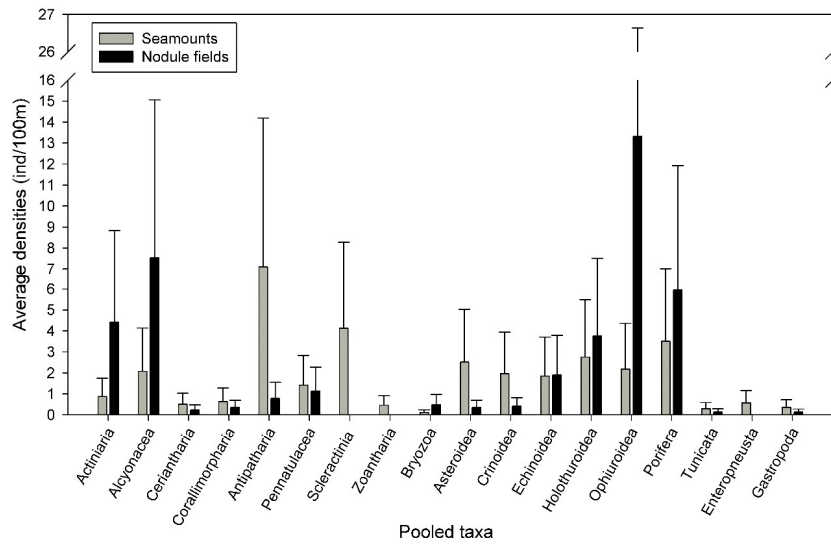
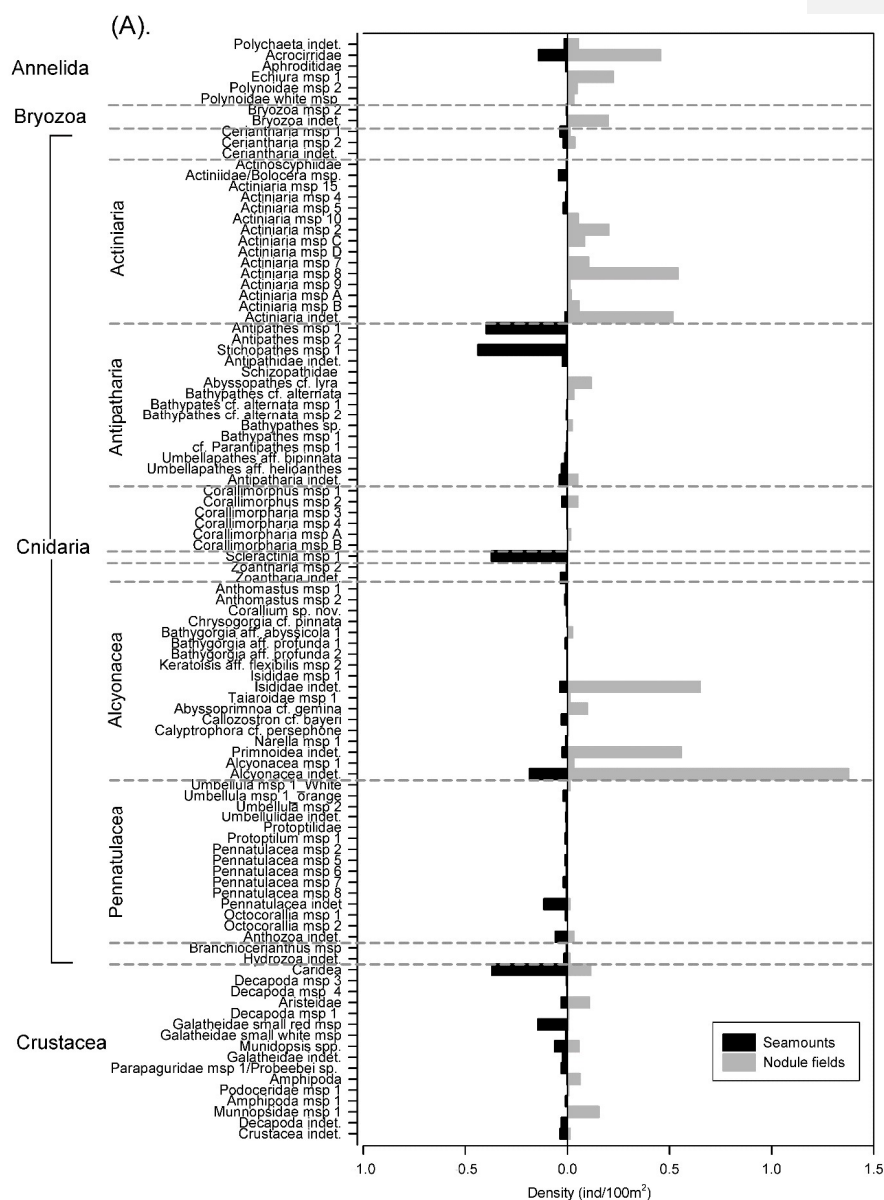
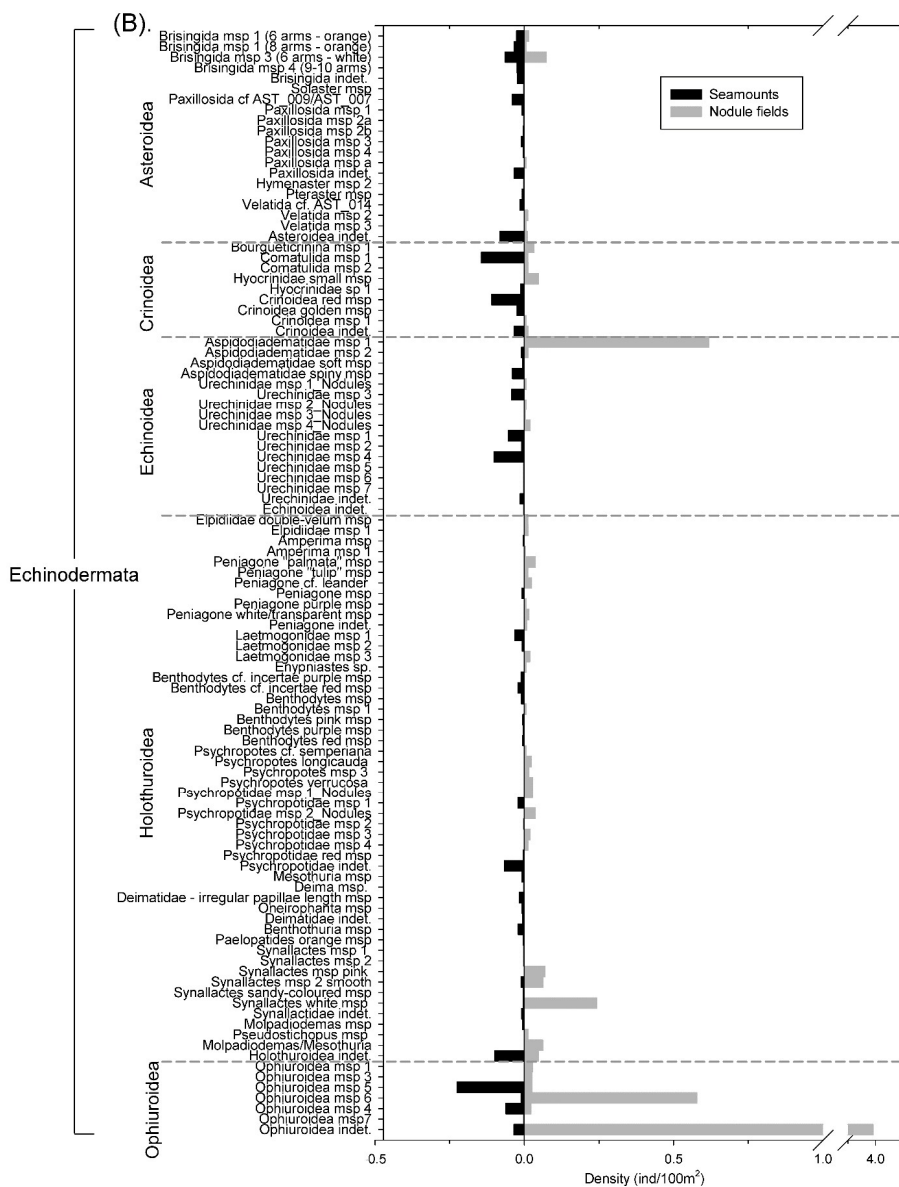


Fig.

A1. Average densities at higher taxa level per ecosystem and standard deviation.





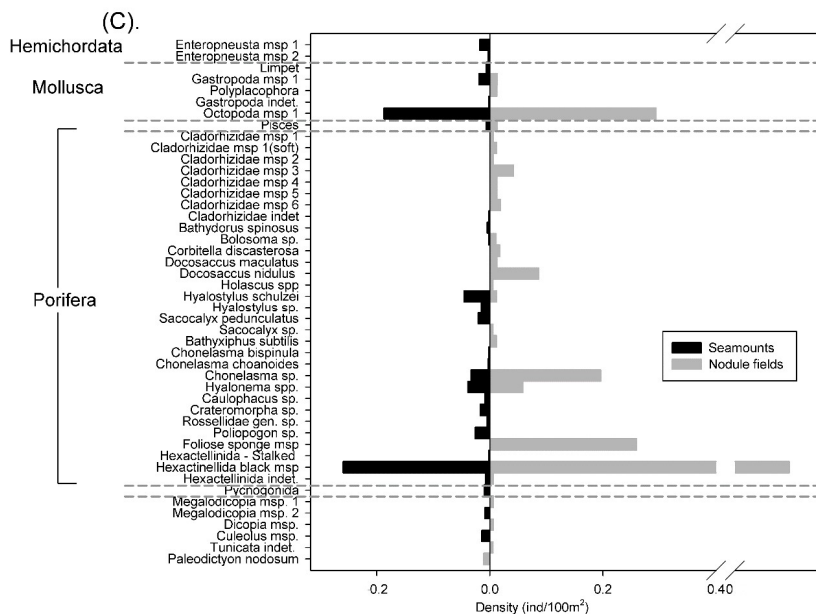


Fig. A1. Back-to-back histogram comparing average densities of morphospecies and taxa (ind/100m) for seamount (n=4) and nodule field (n=5) video transects. (a) Annelida, Bryozoa, Cnidaria and Crustacea, (B) Echinodermata and (C) Mollusca, Porifera, Hemichordata and Chordata (Tunicata).

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