

Flux variability of phyto- and zooplankton communities in the Mauritanian coastal upwelling between 2003 and 2008

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Abstract

Continuous multiyear records of sediment trap-gained microorganism fluxes are scarce. Such studies are important to identify and to understand the main forcings behind seasonal and multiannual evolution of microorganism flux dynamics. Here, we assess the long-term flux variations and population dynamics of diatoms, coccolithophores, calcareous and organic

dinoflagellate cysts, foraminifera and pteropods in the Eastern Boundary Upwelling Ecosystem of the Canary Current. A multiannual, continuous sediment trap experiment was conducted at the mooring site CB_{eu} (*Cape Blanc eutrophic*, ~20°N, 18°W; trap depth = ca. 1,300 m) off Mauritania (northwest Africa), between June 2003 and March 2008. Throughout the study, the reasonably consistent good match of fluxes of microorganisms and bulk mass reflects the seasonal occurrence of the main upwelling season and relaxation, and the contribution of microorganisms to mass flux off Mauritania. A clear successional pattern of microorganisms, *i.e.* primary producers followed by secondary producers, is not observed. High fluxes of diatoms, coccolithophores, organic dinoflagellate cysts, and planktonic foraminifera occur simultaneously. Peaks of calcareous dinoflagellate cysts and pteropods mostly occurred during intervals of upwelling relaxation. A striking feature of the temporal variability of populations' occurrence is the persistent pattern of seasonal groups' contribution. Species of planktonic foraminifera, diatoms, and organic dinoflagellate cysts typical of coastal upwelling as well as cooler water planktonic foraminifera and the coccolithophore *Gephyrocapsa oceanica* are abundant at times of intense upwelling (late winter through early summer). Planktonic foraminifera and calcareous dinoflagellate cysts are dominant in warm pelagic surface waters and all pteropod taxa are more abundant in fall and winter, when the water column stratifies. Similarly, coccolithophores of the upper- and lower photic zone, together with *Emiliania huxleyi*, and organic dinoflagellate cysts dominate the assemblage during phases of upwelling relaxation and deeper layer mixing.

A significant shift in the 'regular' seasonal pattern of taxa relative contribution is observed between 2004 and 2006. Benthic diatoms strongly increased after fall 2005 and dominated the diatom assemblage during main upwelling season. Additional evidence for a change in population dynamics are the short dominance of the coccolithophore *Umbilicosphaera annulus*, the occurrence of the pteropod *Limacina bulimoides*, and the strong increase in the flux of calcareous dinoflagellate cysts, abundant in tropical, warm oligotrophic waters south of the study area after fall 2005. Altogether, this suggests that pulses of southern waters were transported to the sampling site via the northward Mauritania Current. Our multiannual trap experiment provides a unique opportunity to characterize temporal patterns of variability that can be extrapolated to other EBUEs, which are experiencing or might experience similar future changes in their plankton

community.

1. Introduction

A way to obtain insights into the impact of climate variability on marine ecosystems is monitoring multiannual evolution and changes covering key species or groups of species representing different trophic levels. To date, multiyear continuous, *in situ* monitoring records of marine communities are scarce. Information about open ocean ecosystems is even more rare (e.g., IOC-UNESCO TS129 IGMETS 2017; Bringué et al., 2019). Furthermore, records providing information about organism groups of different trophic levels are practically unknown or cover only a few species (e.g., Schlüter et al., 2012; Rembauville et al., 2016).

Eastern Boundary Upwelling Ecosystems (EBUEs) are among the most important marine ecosystems, both ecologically and economically (Cropper et al., 2014). Despite the fact that they cover only 10% of the global surface ocean area, they provide about 25% of the global fish catch (Pauly and Christensen, 1995) and build extraordinary marine hotspots of high primary production and biodiversity (Aristegui et al., 2009). In doing so, they play a key role in the marine biological pump, as together with other continental margins may be responsible for more than 40% of the CO₂ ocean sequestration (Muller-Karger et al., 2005). As EBUEs are highly dynamic with large seasonal and interannual variability (e.g., Chavez and Messié, 2009; Fischer et al., 2016), gaining information on their long-term variability is essential to understand their potential response to current global climate change.

One of the EBUEs that have been thoroughly studied over the past three decades is the coastal ecosystem off Mauritania (northwest Africa), which is part of the Canary Current (CC) EBUE (Cropper et al., 2014). The Mauritanian system is characterized by intense offshore Ekman transport and strong mesoscale heterogeneity, which facilitate the exchange of neritic and pelagic water masses (Mittelstaedt, 1983; Zenk et al., 1991; Van Camp et al., 1991; Aristegui et al., 2009; Chavez and Messié, 2009; Meunier et al., 2012; Cropper et al., 2014). In addition, regional factors such as nutrient trapping efficiency (Aristegui et al., 2009), the giant chlorophyll filament (Gabric et al., 1993; Barton, 1998; Lange et al., 1998; Helmke et al., 2005); dust deposition (Fischer et al., 2016, 2019) and/or the shelf width (Hagen, 2001; Cropper et al., 2014) strongly affect the temporal

88 dynamics of primary and secondary producers communities in surface waters along the
 Mauritanian coast. In this ecosystem, several long-term, continuous, sediment trap-based
 90 monitoring records are available since the late 1980s. Until now, studies monitoring variability of
 this seasonally dynamic ecosystem mostly focused on the variability of bulk fluxes (Fischer et al.,
 92 1996, 2009, 2016, 2019; Bory et al., 2001; Marcello et al., 2011; Skonieczny et al., 2013), particular
 groups of microorganisms (Lange et al. 1998; Romero et al., 1999, 2002, 2003; Köbrich and
 94 Baumann, 2008; Romero and Armand, 2010; Zonneveld et al., 2010; Köbrich et al., 2016; Romero
 and Fischer, 2017; Guerreiro et al., 2019) or sea-surface temperature (Müller and Fischer, 2001;
 96 Mollenhauer et al., 2015). However, the simultaneous comparison of the seasonal and interannual
 dynamics of several phyto- and zooplankton communities by means of multiyear sediment trap
 98 experiments has not been performed in this region and is rare in other EBUEs or other ocean
 areas as well (Bringué et al., 2019).

100 This paper builds on the long-term studies of the variability of sea-surface temperature and
 fluxes (SST) at the site CBeu published by Mollenhauer et al. (2015), Romero and Fischer (2017),
 102 and Fischer et al. (2019). We describe the seasonal and interannual variability of fluxes of several
 primary and secondary producers in the Mauritanian coastal upwelling over a continuous trap
 104 experiment extending almost 1,900 days between June 2003 and March 2008 (Table 1). In
 addition to the previously reported diatom fluxes and reconstructed SST values, we present here
 106 new data of fluxes of coccolithophores, calcareous and organic-walled dinoflagellate cysts,
 planktonic foraminifera and pteropods as well as the species-specific composition of the
 108 assemblages that have been collected at the mooring site CBeu (*Cape Blanc eutrophic*), located
 around 80 nm west of the Mauritanian coastline below a giant chlorophyll filament (Fig. 1). The
 110 microorganisms producing the calcareous, opaline or organic remains have different water column
 habitats, life strategies, and use different nutrient sources (Romero et al., 1999, 2002; Baumann et
 112 al., 2005; Romero and Armand, 2010; Zonneveld et al., 2013), and are widely used as proxies in
 paleostudies carried on Mauritanian sediments (Jordan et al., 1996; Romero et al., 2008; McKay et
 114 al., 2014) and similar paleoenvironments (e.g., Baumann and Freitag, 2004; Bouimetarhan et al.,
 2009; Romero et al., 2015; Weiser et al., 2016; Hardy et al., 2018). The emphasis of our
 116 multiannual trap experiment is on the comparison of temporal dynamics and the species-specific

composition of the primary and secondary producer plankton community off Mauritania. The simultaneous assessment of fluxes of several microorganism groups collected over long intervals provides substantial information about potential changes in the coastal upwelling community. Results are discussed in the context of varying environmental conditions of the low-latitude Northeastern Atlantic. To our knowledge, this is the first multiyear trap-based record of major primary and secondary producers that provides multiyear information on the dynamics of major components of the plankton community in a highly productive EBUE.

2. Oceanographic setting of the study area

The CC-EBUE is located in the eastern part of the North Atlantic Subtropical Gyre (Fig. 1; Arístegui et al., 2009; Chavez and Messié, 2009; Cropper et al., 2014). Both the temporal occurrence and the intensity of the upwelling along northwestern Africa depend on the shelf width, the seafloor topography, and wind direction and strength (Mittelstaedt, 1983; Hagen, 2001). The Mauritanian shelf is wider than the shelf northward and southward and gently slopes from the coastline into water depths below 200 m (Fig. 1b; Hagen, 2001). The shelf break zone with its steep continental slope extends over a distance of approximately 100 km (Hagen, 2001). As a result of the coastal and shelf and slope topography, the ocean currents and wind systems, the coastal region off Mauritania is characterized by almost permanent upwelling. Its intensity varies throughout the year (Lathuilière et al., 2008; Cropper et al., 2014). Our trap site CBeu is located at the southern boundary of this permanent upwelling zone (Fig. 1; Table 1).

The surface hydrography is strongly influenced by two wind-driven surface currents: the southwestward-flowing CC and the poleward-flowing coastal countercurrent or Mauritania Current (MC) (Fig. 1). The eastern branch of the subtropical gyre, the surficial CC detaches from the continental slope between 25° and 21°N and supplies waters to the North Equatorial Current. The CC water is relatively cool because it entrains upwelled water from the coast as it moves southward (Mittelstaedt, 1991). The MC gradually flows northward along the coast up to about 20°N (Mittelstaedt, 1991), bringing warmer surface waters from the equatorial realm into the study area. Towards late autumn, the MC is gradually replaced by a southward flow associated with upwelling water due to the increasing influence of trade winds south of 20°N (Zenk et al., 1991), and becomes a narrow strip of less than 100 km width in winter (Mittelstaedt, 1983). The MC

146 advances onto the shelf during summer and is enhanced by the relatively strong Equatorial
Countercurrent and the southerly monsoon (Mittelstaedt, 1983). The presence of strong coastal
148 currents during the upwelling season causes substantial horizontal shear within the surface layer,
where currents tend to converge (Mittelstaedt, 1983). This convergence favors the formation of the
150 Cape Verde Frontal Zone (CVFZ, Fig. 1; Zenk et al., 1991).

A coastal countercurrent, the Poleward Undercurrent (PUC; Fig. 1) occurs mainly due to wind-
152 driven offshore divergence (Pelegri et al., 2017). North of Cape Blanc (ca. 21°N), the intense
northeasterly winds cause the coastal upwelling to move further offshore and the upper slope is
154 filled with upwelled waters. South of Cape Blanc, northerly winds dominate throughout a year, but
surface waters remain stratified and the PUC occurs as a subsurface current. South of Cape
156 Timiris (ca. 19°30'N), the PUC intensifies during summer-fall and remains at the subsurface during
winter-spring (Pelegri et al., 2017). The encountering of the northward flowing MC-PUC system
158 with the southward flowing currents in the Canary Basin leads to flow confluence at the CVFZ
(Zenk et al., 1991) and the offshore water export visible as the giant Mauritanian chlorophyll
160 filament (Gabric, 1993; Pelegri et al., 2006; Pelegri et al., 2017). This filament extends over 300
km offshore (e.g., Van Camp et al., 1991; Aristegui et al., 2009; Cropper et al., 2014) and carries
162 South Atlantic Central Water (SACW) offshore through an intense jet-like flow (Meunier et al.,
2012; Fig. 1). Intense offshore transport forms an important mechanism for the export of cool,
164 nutrient-rich shelf and upper slope waters offshore Mauritania. Based on satellite imagery and *in*
situ data, it has been estimated that the giant Mauritanian filament could export about 50% of the
166 particulate coastal new production offshore toward the open ocean during intervals of most intense
upwelling, while coastal phytoplankton at the surface might be transported as far as 400 km
168 offshore (Gabric et al., 1993; Barton, 1998; Lange et al., 1998; Helmke et al., 2005). The transport
effect could extend to even more distant regions in the deep ocean, since sinking particles are
170 strongly advected by lateral transport (Fischer and Karakaş, 2009; Karakaş et al., 2006; Fischer et
al., 2009).

172 The nutrient concentration of the upwelled waters off Mauritania varies depending on their origin
(Fütterer, 1983; Mittelstaedt, 1991; Zenk et al., 1991). The source of upwelling waters off
174 Mauritania are either North Atlantic Central Water (NACW), north of about 23°N, or SACW (south

of 21°N, Fig. 1). Both water masses are mixed in the filament area off Cape Blanc. The SACW occurs in layers between 100 and 400 m depth off Cape Blanc and the Banc d'Arguin. The hydrographic properties of the upwelling waters on the shelf suggest that they ascend from depths between 100 and 200 m south off the Banc d'Arguin (Mittelstaedt, 1983). North of it, the SACW merges gradually into deeper layers (200-400 m) below the CC (Mittelstaedt, 1983). During intense upwelling, the stratification of the shelf waters weakens, and so is the stratification further offshore, usually within the upper 100 m (Mittelstaedt, 1991). The biological response is drastically accelerated in the upwelling waters when the SACW of the upper part of the undercurrent feeds the onshore transport of intermediate layers to form mixed-water types on the shelf (Zenk et al., 1991).

3. Material and Methods

3.1. Moorings, sediment traps and bulk flux assessment

Sediment trap moorings were deployed at site CBeu off Mauritania in the CC-EBUE (Fig. 1). Details on trap location and depth, sample amount, water depth and sampling intervals as well as the sample identification (cruise and GeoB numbers) are presented in Table 1. Large-aperture time-series sediment traps of the Kiel type were used with 20 to 40 cups (depending on ship-time availability) and 0.5 m² openings, equipped with a honeycomb baffle (Kremling et al., 1996). Traps were moored in intermediate waters (1,256 m – 1,296 m) and sampling intervals varied between 6.5 and 23 days (Table 1). Uncertainties with the trapping efficiency due to strong currents (e.g. undersampling) and/or due to the migration and activity of zooplankton migrators ('swimmer problem') are assumed to be minimal in the depth range of our trap experiments (Buesseler et al., 2007; Fischer et al., 2019).

Prior to each deployment, sampling cups were poisoned with 1 ml of concentrated HgCl₂ per 100 ml of filtered seawater. Pure NaCl was used to increase the density in the sampling cups up to 40 ‰. Upon recovery, samples were stored at 4°C and wet split in the MARUM sediment trap laboratory (University Bremen, Bremen) using a rotating McLANE wet splitter system. Larger swimmers, such as crustaceans, were handpicked with forceps and removed by carefully filtering through a 1-mm sieve. All flux data hereafter refer to the size fraction of <1 mm. Detailed

information about sampling and laboratory analysis is given in Mollenhauer et al. (2015), Romero and Fischer (2017) and Fischer et al. (2019). These papers present the bulk fluxes for the deployments CBeu 1-5 (Table 1). Alkenone-derived sea surface temperature (SST) for the CBeu deployments 1-4 were provided by Mollenhauer et al. (2015).

Using $\frac{1}{4}$ or $\frac{1}{5}$ wet splits, analysis of the <1 mm fraction was carried out (Fischer and Wefer, 1991; Fischer et al., 2016). Samples were freeze-dried and homogenized before being analyzed for bulk (total mass), organic carbon (OC), calcium carbonate (CaCO_3) and biogenic silica (BSi, opal). Total organic carbon (TOC) and CaCO_3 were measured by combustion with a CHN-Analyzer (HERAEUS, Department of Geosciences, University of Bremen). TOC was measured after removal of carbonate with 2 N HCl. Overall analytical precision based on internal lab standards was better than 0.1% ($\pm 1\sigma$). Carbonate was determined by subtracting OC from total carbon, the latter being measured by combustion without pre-treatment with 2N HCl. Organic matter was estimated by multiplying the content of total organic carbon by a factor of two as about 50-60% of marine organic matter is constituted by OC (Hedges et al., 2002).

BSi was determined with a sequential leaching technique with 1M NaOH at 85°C (Müller and Schneider, 1993). The precision of the overall method based on replicate analyses is mostly between ± 0.2 and $\pm 0.4\%$, depending on the material analyzed. The lithogenic fluxes were estimated by subtracting the masses of CaCO_3 , BSi, and 2 x OC from the total mass flux.

3.2. Assessment of organism fluxes and species identification

3.2.1. Diatoms

Diatom data shown here were previously published in Romero and Fischer (2017). For this study $\frac{1}{25}$ and $\frac{1}{125}$ splits of the original samples were used. Samples were prepared for diatom studies following the method proposed by Schrader and Gersonde (1978). A total of 185 sediment trap samples were processed. Each split was treated with potassium permanganate, hydrogen peroxide, and concentrated hydrochloric acid following previously used methodology (Romero et al., 2002, 2009a, b, 2016, 2017). Identification and count of the species assemblage were done on permanent slides (*Mountex*® mounting medium) at x1000 magnification using a Zeiss® Axioscop with phase-contrast illumination (MARUM, Bremen). The counting procedure and definition of counting units follows Schrader and Gersonde (1978). Depending on valve abundances in each

sample, several traverses across each slide were examined. Total amount of counted valves per
slide ranged between 300 and 800. At least two cover slips per sample were scanned in this way.
Valve counts of replicate slides indicate that the analytical error of concentration estimates is ca.
10 %. The resulting counts yielded abundance of individual diatom taxa (absolute and relative) as
well as daily fluxes of diatoms as valves per $\text{m}^{-2} \text{d}^{-1}$, calculated according to Sancetta and Calvert
(1988).

3.2.2. Coccolithophores

Aliquots of each sample were 1/125. Depending on the total flux, samples were further split
down to 1/625 to 1/2500 of the original sample volume and were filtered afterward onto
polycarbonate membrane filters (Schleicher and Schuell™ 47mm diameter, 0.45 μm pore size). A
piece of the filter was cut and mounted on a Scanning Electron Microscopy stub. Qualitative and
quantitative analyses of the trapped assemblages were performed using a Zeiss® DSM 940A at
10kV accelerating voltage (Department of Geosciences, University of Bremen, Bremen). In an
arbitrarily chosen transect, coccoliths were counted until a total of at least 500 specimens were
reached. The taxonomic classification of identified species was based on Young et al. (2003) as
well as on Nannotax 3 (Young et al., 2019).

3.2.3. Calcareous and organic-walled dinoflagellate cysts

1/125 splits of the original trap material were ultrasonically treated and sieved with tap water
through a high precision metal sieve (Storck-Veco®) with a 20 μm pore size. The residue was
transferred to Eppendorff cups and concentrated to 1 ml of suspension. After homogenization of
the material, a known aliquot was transferred to a microscope slide where it was embedded in
glycerin-gelatine, covered with a cover slip and sealed with wax to prevent oxidation of the
organic material. After counting, cyst fluxes were calculated by multiplying the cyst counts with
the aliquot fraction and the split size (1/125) and dividing through the amount of days during
which the trap material was sampled as well as the trap-capture surface. No chemicals were
used to prevent dissolution of calcite and silicate. Cyst assemblages were determined by light
microscopy (Axiovert, x400 magnification). The taxonomy of calcareous dinoflagellate cysts
follows Vink et al. (2002) and Elbrächter et al. (2008), while that of organic walled dinoflagellate
cysts is according to Zonneveld and Pospelova (2015),

3.2.4. Planktonic foraminifera and pteropods

Depending on the absolute magnitude of the total mass flux, a 1/5 or a 1/25 split of the wet solution (fraction <1mm) was used to pick planktonic foraminifera and pteropods (pelagic mollusks). Specimens of both groups of calcareous microorganisms were rinsed three times by using tap water, dried at 50°C in an oven overnight and then separated from each other. Identification and count of shells were done by using a stereomicroscope Zeiss® Stemi 2000 (MARUM, Bremen). The foraminifera fluxes (all size fractions) were determined in mg per m² and day with a Sartorius BP 211D analytical balance. Additionally, the total amount of specimens/sample of foraminifera and pteropods (>150 µm) were manually counted. Planktonic foraminifera were identified and classified according to Hemleben et al. (1989) and Schiebel and Hemleben (2017). Out of 15 species of planktonic foraminifera identified, only six species were used as environmental indicators.

3.2.5. Alkenones

1/5 wet splits of the <1 mm fraction were used for alkenone analysis. Briefly, freeze-dried CBeu 1-4 samples were solvent extracted. The resulting total lipid extracts (TLEs) saponified and the alkenone fractions were obtained by means of column chromatography of the neutral lipid fractions from the saponification. Details are given in Mollenhauer et al. (2015).

A slightly different, miniaturized analysis procedure has been applied for the CBeu trap 5 samples. 1/5 wet splits of the freeze-dried <1mm fraction were weighted in 10 ml Pyrex tubes and a known amount of an internal standard (*n*-Nonadecan-2-one) was added. Samples were then 3x ultrasonically extracted with a mixture of 3 ml dichloromethane/methanol (9:1 vol./vol.), centrifuged and the supernatant solvent combined as total lipid extract (TLE). TLEs were evaporated to dryness and saponified in a 0.1M potassium hydroxide solution in methanol/water (9:1 vol./vol.) for two hours at 80°C. Neutral lipids, recovered with hexane, were afterwards separated into fractions of different polarity by silica gel chromatography and elution with hexane, dichloromethane/hexane (1:1 vol./vol.) and dichloromethane/methanol (9:1 vol./vol.), respectively. The second fraction containing the alkenones was dried, re-dissolved in 20µl hexane and analyzed on a 7890A gas chromatograph (GC, Agilent Technologies) equipped with a cold on-column injection system, a DB-5MS fused silica capillary column (60 m, ID 250 µm, 0.25 µm film coupled to a 5 m, ID 530 µm

deactivated fused silica precolumn) and a flame ionization detector (FID). Helium was used as carrier gas (constant flow, 1.5 mL/min) and the GC oven was heated using the following temperature program: 60°C for 1 min, 20°C/min to 150°C, 6°C/min to 320°C and a final hold time of 35 min. Alkenones were identified by comparison of the retention times with a reference sample composed of known compounds. Peak areas were determined by integrating the respective peaks.

The $U_{37}^{K'}$ index was calculated using the following equation (Prahl and Wakeham, 1987)

$$U_{37}^{K'} = \frac{C_{37:2}}{C_{37:2} + C_{37:3}}$$

and converted to SST using the global surface water calibration from Conte et al. (2006):

$$SST = \frac{U_{37}^{K'} - 0.0709}{0.0322}$$

3.3. Environmental physical parameters

SST, Sea Surface Temperature Anomaly (SSTA), mixed layer depth (MLD) and upper ocean chlorophyll-a concentration data are based on satellite-derived data achieved from the NASA supported Giovanni project (<https://giovanni.gsfc.nasa.gov/>). SST is the mean of daily surface ocean temperature and MLD values of the sampling interval in a 4 km² area around the trap position (Table 1). In the study area, SST at the mooring location is influenced by seasonal air temperature changes as well as the presence of upheld water surfacing at the trap position.

To compensate for seasonal air temperature changes the SSTA is calculated by subtracting the above-calculated SST at the trap position from mean SST values of simultaneous sampling intervals in a 4 km² block 200 nm west of the trap position. Both SSTA and MLD are parameters reflecting active upwelling in the study area. Upper ocean chlorophyll a data and MLD represent monthly mean values in a 9 km² block around the trap position. Wind speed and wind directions are provided by Nouadhibou airport (20°56'N, 17°2'W) (Institut Mauretanien de Recherches Océanographiques et des Pêches, Nouadhibou, Mauritania). For statistical analyses, the means of daily values during the trap sampling intervals were calculated.

3.4. Multivariate analyses

To determine if a linear or unimodal based ordination method should be applied on the data we performed a Detrended Correspondence Analysis previous to the statistical analysis. Results of

318 this analysis revealed a total length of gradient of 1.2 sd, which indicates a linear species respond
on environmental gradients. The ordination techniques Principal Component (PCA) and
320 Redundancy Analysis (RDA) have been accordingly performed. Significance of the environmental
variables has been tested with a Monte-Carlo permutation test with unrestricted permutations.

322 PCA and RDA have been performed with the software Package Canoco 5 (ter Braak and
Smilauer, 2012; Smilauer and Leps, 2014). To obtain insights into the temporal relationship
324 between fluxes of organism groups (diatoms, coccolithophores, calcareous and organic-walled
dinoflagellate cysts, planktonic foraminifera and pteropods) and bulk components as well as the
326 environmental conditions in surface waters and low atmosphere, a RDA has been performed. RDA
compares the total flux of organism groups with environmental parameters and TOC, BSi, CaCO₃
328 and lithogenic fluxes (Table 2). Since the fluxes of the individual groups differ by several orders of
magnitudes, it is essential to normalize their flux values prior to the statistical analysis in order to
330 be able to determine temporal relationships of flux variability. As a consequence, the total flux of
the organism groups has been normalized to values between 0 and 1000 previous to the analyses
332 according to formula 1:

$$nFl_i = (FL_{i/y} / FL_i \text{ max}) \times 1000n$$

334 Fl_i = normalized flux of species group i

$Fl_{i/y}$ = flux of species group i in sample y

336 $FL_i \text{ max}$ = maximal flux observed in species group i

To better understand the relationship within the individual organism groups, a PCA has been
338 performed (Table 2). For these analyses, the total flux of the organisms/species groups have been
normalized to values between 0 and 1000 according to formula 2:

340 $nFl_j = (FL_{ij/y} / FL_i \text{ max}) \times 1000n$

Fl_j = normalized accumulation rate of ecological entity j in species group i

342 $Fl_{j/y}$ = accumulation rate of ecological entity j in sample y $FL_j \text{ max}$ = maximal accumulation
rate observed in species group i

344 Within coccolithophores, *Umbilicosphaera anulus* had exceptionally large fluxes in one sample
only. This flux exceeded the maximal flux of the other species by a factor of three. This value has
346 been excluded from the analysis and the $FL_j \text{ max}$ in this group is determined by excluding this

outlier.

4. Results

4.1. Bulk fluxes and fluxes of organism groups

Independent of the year, the total mass flux of particulates at site CBeu is mainly composed of biogenic components (calcium carbonate, BSi and TOC, Table 3; Fischer et al., 2019). Calcium carbonate is the main bulk biogenic component. The daily average flux of CaCO_3 –calculated for each full calendar year sampled between 2004 and 2007- varies between 121.1 and 150.9 $\text{mg m}^{-2} \text{d}^{-1}$ (Table 3). BSi is the second most important biogenic bulk component (35.8-54.61 $\text{mg m}^{-2} \text{d}^{-1}$, Table 3), followed by TOC (19.00-29.40 $\text{mg m}^{-2} \text{d}^{-1}$). Lithogenics is the main non-biogenic contributor to the total mass flux (84.6-134.2 $\text{mg m}^{-2} \text{d}^{-1}$, Table 3). Coccolithophores, planktonic foraminifera, calcareous dinoflagellates and pteropods are main contributors to the CaCO_3 flux, while diatoms dominate the siliceous community.

The fluxes of total mass, CaCO_3 , TOC, BSi and lithogenics show major peaks in winter and spring (Fig. 2). Secondary maxima were found during late summer/fall, mainly in 2003, and less clearly in 2005, 2006 and 2007 (Fig. 2). However, the individual components reveal different flux amplitudes and point to some interannual variability (Table 3). CaCO_3 fluxes were exceptionally high in early winter 2005 compared to the other years. Fluxes of BSi and TOC match well the flux pattern of total mass and show less interannual variability (Fig. 2c, d). The flux of the lithogenic fraction has the highest amplitudes in spring 2006 and 2007 (Fig. 2e).

Fluxes of microorganisms are dominated by diatoms and coccoliths (Fig. 3a, b). These exceeded the fluxes of organic- and calcareous walled dinoflagellate cysts, planktonic foraminifera and pteropods by a factor of four to five. Highest coccolithophore and diatom fluxes reach 4.2×10^9 coccoliths $\text{m}^{-2} \text{d}^{-1}$, and 1.2×10^8 valves $\text{m}^{-2} \text{d}^{-1}$, respectively. Maximal fluxes of organic-walled dinoflagellates reach up to 7.1×10^4 cysts $\text{m}^{-2} \text{d}^{-1}$, of planktonic foraminifera 0.9×10^4 shells $\text{m}^{-2} \text{d}^{-1}$, and of pteropods 1.1×10^4 shells $\text{m}^{-2} \text{d}^{-1}$.

Each group of microorganisms shows large seasonal and interannual variability (Table 5).

Diatoms had their maximal flux in fall/winter 2005 and spring/summer 2006 (Fig. 3a).

Coccolithophores had their highest fluxes mostly in winter/spring throughout the sampling interval

and exceptionally in July/August 2003 and 2007 and in fall 2005. On the long-term, low coccolithophore fluxes were observed fall and winter 2007/2008 (Fig. 3b). Calcareous dinoflagellate cysts were practically absent until fall 2005 (Fig. 3c). After September 2005, calcareous dinoflagellate cysts showed maximal export fluxes in fall/winter 2005/2006 and fall/winter 2006/2007 (Fig. 3c). Fluxes decreased again after spring 2007. Organic-walled dinoflagellate cysts had their highest export fluxes in summer 2003, spring/summer 2006 and summer 2007 (Fig. 3d). Planktonic foraminifera showed maximal fluxes in summer 2003, winter/spring 2004, 2005, 2007 and spring/summer 2006 (Fig. 3e). Pteropods had their maximal fluxes in summer 2003, fall/winter 2003/2004, 2004/2005 and 2006/2007 as well as summer 2005 and 2007 (Fig. 3f).

4.2. Species- and group-specific composition of assemblages

The studied plankton community at the CBeu site is highly diverse and is composed by at least 220 identified species. Table 5 presents the species-specific composition of groups depicted in Fig. 4.

Out of 170 marine *diatom* species, the 70 most abundant diatom taxa (average relative contribution >0.75% for the entire studied interval) were attributed to four groups, according to the main ecological conditions they represent: (1) benthic, (2) coastal upwelling, (3) coastal planktonic, and (4) open-ocean waters (see also Romero and Fischer, 2017). The diatom groups show a clear seasonal pattern and strong interannual variability (Fig. 4a, Table 4). Benthic diatoms have higher relative contributions during spring and summer, whereas the coastal upwelling group mainly occurred between late spring and early fall. Open-ocean diatoms were more abundant from fall to early spring while the coastal planktonic taxa tended to be more abundant during fall and winter. Most noticeably, a drastic shift in the relative contribution of the benthic diatoms occurred in spring-summer 2006 when the abundance of benthic diatoms strongly increased from 2006 onward, compared to 2003–2005 (Fig. 4a, Table 4). In spite of the increased relative contribution of benthic diatoms after 2005, the seasonal pattern of the predominantly high spring-summer total diatom flux remained unaltered (Fig. 3a).

Coccolithophores are consistently dominated by *Emiliana huxleyi* and *Gephyrocapsa oceanica*, whose combined contribution always represents > 50% of the community throughout the

sampling period (Fig. 4b, Table 4). Oligotrophic upper photic zone (UPZ, e.g., *Umbellicosphaera*
 406 *tenuis*, *U. irregularis*) and lower photic zone species (LPZ, e.g., *Florisphaera profunda*, *Gladiolithus*
flabellatus) make up the majority of the remaining species. Whereas *E. huxleyi* showed a less clear
 408 seasonal pattern, *G. oceanica* tends to be more abundant during late spring and early fall (Fig. 4b).
 In contrast, UPZ and LPZ taxa have higher relative contributions during winter and spring. The
 410 appearance of *Umbilicosphaera anulus* (present in consistently low relative abundances of 5-10%
 until the summer of 2006) accounts for up to 65% of the community in winter 2005/06. Other
 412 common taxa with an average relative contribution >0.75% for the entire studied interval are listed
 in Table 5.

414 **Calcareous dinoflagellate cysts** can be attributed to five groups based according to the
 main ecological conditions they represent (Siggelkow et al., 2002; Richter et al., 2007; Kohn and
 416 Zonneveld, 2010); (1) upwelling, (2) warm waters, (3) terrestrial mineral input, (4) cosmopolitan
 and (5) other species (Table 5). Until fall 2005 abundances are very low such that the
 418 recognition of a seasonal pattern is hampered (Fig. 4c). After fall 2005, their occurrence shows a
 clearer seasonal pattern. In spring-summer of 2006 upwelling species dominate the association.
 420 After fall 2006, the community is composed by the interplay of cosmopolitan species, warm
 water taxa and upwelling-dependent species, where warm water taxa dominate. Whereas
 422 upwelling species are most abundant in spring and fall, warm water and mineral indicators are
 more abundant in fall/winter (Fig. 4).

424 **Organic dinoflagellate cysts** can be attributed to five groups based on the relationship
 between their geographic distribution in surface sediments from the Cape Blanc area and the
 426 environmental conditions in surface and subsurface waters as well as long-term surveys of their
 seasonal cyst production (Susek et al., 2005; Holzwarth et al., 2010; Smayda, 2010; Smayda
 428 and Trainer, 2010; Trainer et al., 2010; Zonneveld et al., 2012, 2013): (1) upwelling, (2)
 upwelling relaxation, (3) potential toxic, (4) cosmopolitan, and (5) other species. Throughout the
 430 investigated time interval upwelling species are abundant in spring and fall/winter whereas
 upwelling relaxation species have higher relative abundances in fall (Fig. 4d). Potential toxic
 432 species are abundant in fall/winter 2004/2005 and 2007/2008. Organic-walled dinoflagellate

cysts do not show a clear change in their composition between 2005 and early 2006 (Table 4)

434 as observed for many other groups.

The distribution and abundance of **planktonic foraminifera** species is linked to surface-water
 436 properties. We use prominent species as tracers of surface water properties: *Globigerina bulloides*,
 usually thriving in temperate to subpolar waters during intervals of enhanced primary productivity
 438 due to high nutrient availability (Schiebel and Hemleben, 2017), is generally most abundant
 between summer and fall (fig. 4e). Transitional and subpolar species *Globorotalia inflata* and
 440 *Neogloboquadrina incompta* are usually present throughout a year, only decreasing in abundance
 in fall and winter when warm water taxa peaked (*Globigerinoides ruber* pink, *G. ruber* white and *G.*
 442 *sacculifer*; Kucera, 2007, Schiebel and Hemleben, 2017) (Fig. 4e). The only exception is in fall and
 winter 2004/2005, when warm-water taxa are almost absent.

444 As large secondary carbonate producers off Mauritania, **pteropods** are important contributors
 to the carbonate flux in the CC-EBUE (Fischer et al., 2016). The community is composed of
 446 relatively few taxa. *Heliconoides inflatus* dominates the assemblage throughout most of the studied
 interval (Fig. 4f). It is often the only species found in the assemblage until winter 2005/2006, when
 448 a sudden and drastic shift in the relative contribution occurred. *Limacina bulimoides* appears for
 the first time in winter to spring 2006 - and again in fall and winter 2006/2007 - and dominates the
 450 assemblage together with a group of unspecified uncoiled pteropods. However, another
 occurrence of *L. bulimoides* is missing in winter 2007/2008.

452 4.3. Statistical analyses

Comparison of the fluxes of the microorganisms, bulk components and the environmental
 454 conditions in surface waters and the lower atmosphere (MLD, average wind speed, wind direction,
 chlorophyll-a concentration (Chl-a), SST and SSTA) resulted in a significant relationship within the
 456 first and second RDA axes that correspond to 34 % and 11% of the variance within the dataset,
 respectively (Table 2).

458 All microorganism groups are ordinated on the positive part of the first axis showing a positive
 relationship with all bulk parameters (Fig. 5). This implies that the fluxes of total mass, TOC, BSi
 460 and CaCO₃ increased with increasing fluxes of all studied microorganism groups (Fig. 5, Tables 3
 and 4). Fluxes of planktonic foraminifera, diatoms and –to a lesser extent– coccolithophores and

organic dinoflagellate cysts are ordinated on the negative side of SST and, with the exception of organic dinoflagellate cysts, ordinated on the positive side of MLD (Fig. 5). This implies that their fluxes are enhanced whenever SST is low and MLD is deep, *i.e.* under a well-mixed uppermost water column. Diatoms, coccolithophores, organic dinoflagellate cysts and planktonic foraminifera also show a positive correlation with SSTA, implying that enhanced fluxes of these microorganisms occur when temperature anomalies between waters overlying site CBeu and the offshore pelagial is large. The flux of pteropods and calcareous dinoflagellate cysts are positively related to the average wind direction, and negatively to MLD and average wind speed (Fig. 5).

Interestingly, the satellite-derived Chl-a data do not show a significant correlation with fluxes of major microorganism groups studied at site CBeu. This possibly indicates that (1) a large portion of satellite-measured chlorophyll concentration is delivered by microorganisms, which did not reach the CBeu trap, and/or (2) due to the strong ballasting effect, part of the microorganisms' remains reach the trap cups independent of intervals of highest satellite chlorophyll values. An alternative explanation is that (3) satellites measure the chlorophyll concentration in the uppermost centimeters of the water column while microorganisms collected with the CBeu traps thrive mostly in waters deeper than those reached by satellite sensors.

To better understand the correlation of the fluxes of the species groups within the microorganism groups, PCA has been performed (Fig. 6, Table 2). The first two axes correspond to 26.3 % and 16.2% of the variance within the dataset respectively. Based on their ordination on the first and second axis, three groups are recognized (Fig. 6):

- Groups 1 and 2 are ordinated at the negative side of the second axis. Group 1 (in blue, Fig. 6) is built by planktonic foraminifera characteristic of cooler or high nutrient water masses (For-cold, For-hinut); benthic and upwelling-related diatoms (Dia-bent and Dia-upw); organic dinoflagellates characteristic for upwelling regions (OD-upw), and the coccolithophore *Gephyrocapsa oceanica* (Co-Gocean). Group 2 (in brown, Fig. 6) consists of upwelling-related and other calcareous dinoflagellates cysts (CD-upw and CD-other), other coccolithophores (Co-other), and coastal planktonic and open-ocean diatoms (Dia-coast and Dia-ocean).
- Ordinated on the positive side of the second axis and central part of the first axis, group 3

assembles planktonic foraminifera mainly thriving in warm waters (For-warm), calcareous
 492 dinoflagellates characteristic of warm water conditions and those responding to mineral
 input (CD-warm, CD-min), and all pteropods taxa (in black, Fig. 6).

- 494 - Group 4 is ordinated at the central/positive part of the second axis and positive site of the
 first axis. Species assigned to group 4 are: organic walled dinoflagellate cysts typical of the
 496 upwelling relaxation (OD-upw relax); UPZ and LPZ coccolithophores (Co-up phot and Co-
 low phot); *E. huxleyi* (Co-*Ehux*), other coccolithophores (Co-other), *U. anulus* (Co-*Uanu*),
 498 and cosmopolitan calcareous dinoflagellate cysts (CD-cosm) (in red, Fig. 6).

500 5. Discussion

502 5.1. Relationship between microorganisms' fluxes at site CBeu and the physical and biogeochemical settings off Mauritania

Both the visual examination of the flux variability and the performed statistical analysis evidence
 504 that the seasonality of most microorganism groups at the CBeu site closely follows the temporal
 pattern of changes in upper water oceanographic conditions off Mauritania between June 2003
 506 and March 2008. Fluxes of diatoms, coccolithophores, organic-walled dinoflagellate cysts and
 planktonic foraminifera increase whenever the uppermost water column is well mixed (Fig. 7e),
 508 SSTs are low (Fig. 7d), and SSTA are high (Figs. 2, 3, 5). This strong match supports the scenario
 of simultaneous occurrence of intense upwelling off Mauritania and high microorganisms' fluxes at
 510 site CBeu. Several previous studies have separately documented enhanced production of diatoms,
 coccolithophores, organic-walled dinoflagellate cysts and planktonic foraminifera occurring when
 512 the nutrient concentration in the uppermost water column off Mauritania increases (Baumann et al.,
 2005; Zonneveld et al., 2012; Guerreiro et al., 2017; Romero and Fischer, 2017; Pospelova et al.,
 514 2018).

The atmospheric, hydrographic and biochemical conditions deliver the physical parameters and
 516 nutrient conditions that determine the temporal pattern of population dynamics as recorded by the
 CBeu trap. Wind and upper water conditions off Mauritania show a clear seasonal pattern of
 518 variability (Fig. 7a-e). The highly stratified uppermost water column (above 40 m water depth)
 overlying site CBeu is an effect of winds blowing mainly from the N-NE between late winter and

520 early summer (Fig. 7a, b, e). The stratification off Mauritania breaks down mostly in early to middle
 winter, when the predominant winds turn from N-NE into S-SE (Fig. 7a). Following this setting,
 522 upwelling reaches its highest intensity between late winter/early spring and early summer
 (Mittelstaedt, 1991; Meunier et al., 2012; Cropper et al., 2014). The SST record (Fig. 7d) matches
 524 well the seasonal alternation of stratification and mixing conditions: lowest temperatures occur
 mostly between winter and early spring (increasing SSTA in late winter and throughout spring).
 526 Throughout the period investigated, this SST cyclicity remains fairly constant.

Fluxes of total mass and biogenic bulk components (CaCO_3 , BSi, TOC) are clearly seasonal in
 528 nature (Fig. 3 a-d; Fischer et al., 2019) and reflect the temporal productivity pattern of the
 Mauritanian upwelling region (Meunier et al., 2012; Cropper et al., 2014). The good temporal
 530 match between flux maxima of most of the studied microorganism groups and biogenic bulk
 components unambiguously evidences the contribution of primary and secondary producers to the
 532 total mass/biogenic mass fluxes off Mauritania (Figs. 2, 3). Higher absolute values of CaCO_3 over
 BSi (Fig. 2b, d, Table 3) support the scenario of calcareous primary and secondary producers
 534 (coccolithophores, planktonic foraminifera and pteropods) dominating the plankton community in
 the Mauritanian upwelling system (Fischer et al., 2019). Diatoms are the main contributors to the
 536 BSi flux (Fig. 3a, 2d; Romero et al., 2002; Romero and Fischer, 2017).

A strong match among fluxes of diatoms, coccolithophores, organic-walled dinoflagellate cysts
 538 and planktonic foraminifera with lithogenic fluxes at times of intense upwelling is observed (Figs.
 2e, 3 a-c, e). The RDA supports this correlation (Fig. 5). The good correlation between lithogenic
 540 and microorganisms' fluxes demonstrates that winds –responsible for the water column mixing off
 Mauritania (Mittelstaedt, 1983; Meunier et al., 2012)– might additionally enrich surface waters
 542 overlying site CBeu with land-derived nutrients. Primary and secondary producers may remarkably
 benefit from this eolian-transported pool of nutrients. Lithogenic material is brought into
 544 Mauritanian ocean waters in the form of dust that it is transported from the Sahara and the Sahel
 (Romero et al., 2003; Friese et al., 2017). Numerous studies have thoroughly documented that the
 546 particle flux off Mauritania predominantly occurs in the form of aggregates, often rich in lithogenic
 particles (e.g., Karakaş et al., 2009; Iversen et al., 2010; Iversen and Ploug, 2010; Nowald et al.,
 548 2015; Fischer et al., 2016; van der Jagt et al., 2018). Recent experiments have also shown that

aggregates' abundance and sinking velocities increase toward deeper waters when aggregates
 550 are ballasted with lithogenic particles, whereas aggregates are not able to scavenge lithogenic
 material from deeper waters (van der Jagt et al., 2018). Further evidence of a possible ballasting
 552 effect on the flux of bulk components was recently presented by Fischer et al. (2019). These
 authors observed that individual high BSi maxima at site CBeu revealed a peak-to-peak correlation
 554 with the dust fluxes. This was interpreted to indicate a strong coupling between dust deposition
 (lithogenic flux) and the efficiency of the biological pump under dry depositional conditions in winter
 556 off Mauritania. Based on these observations, Fischer et al. (2019) proposed that the ballasted,
 organic-rich aggregates built in surface waters immediately react to any additional dust supply with
 558 aggregation followed by rapid sedimentation. Experimental studies on aggregate ballasting and
 scavenging off Mauritania (van der Jagt et al., 2018) support this view as well.

560 A remarkable finding of our multiannual trap experiment is that flux maxima of diatoms,
 coccolithophores, organic-walled cysts producing dinoflagellates (all primary producers) and
 562 planktonic foraminifera (secondary producers) seem to occur fairly simultaneously (Figs. 3, 5). We
 propose three possible interpretations: (i) no clear short-term succession of the microorganism
 564 groups occurred (no temporal turnover in phytoplankton composition within a few days, Roelke and
 Spatharis, 2015), (ii) the succession is not properly captured due to low temporal resolution of
 566 some sediment trap intervals (Table 1), and/or (iii) the microorganisms –originally produced in
 surface and subsurface waters by different communities– sink with different velocities through the
 568 water column toward the ocean bottom and get 'mixed' during their sinking, mainly due to
 dissimilar weights and sizes of their remains.

570 Although it is widely believed that the supply of resources regulates the marine community
 structure (Roelke and Spatharis, 2015), experimental data show that the competition of resources
 572 *per se* does not lead to succession of phytoplankton populations. For instance, Barber and Hiscock
 (2006) observed that marine picoplankton is not replaced by diatoms when chemical transient
 574 conditions (e.g., added iron) abruptly provide a more favorable growth setting. The enormous
 proportional increase in diatom biomass has fostered the misconception that diatoms replace the
 576 non-diatom taxa by succession as the bloom matures. Additional evidence is provided by
 observational studies. Bringué et al. (2018) observed that autotrophic dinoflagellates in the Cariaco

578 Basin do not appear to compete with diatoms for resources as both groups respond positively to
 upwelling dynamics. Similarly, Anabalón et al. (2014) observed equal and simultaneous
 580 contributions of diatoms and pigmented dinoflagellates to total autotrophic biomass off Cape Ghir
 located in the northern CC-EBUEs. Contrary to conventional wisdom, Anabalón et al. (2014)
 582 demonstrate that groups of phytoplankton increase in growth rates and absolute abundance, but
 the biomass increase of the ambient non-diatom assemblage is modest, especially compared to
 584 the order of magnitude or more increase of diatom biomass.

The first three CBeu trap experiments had long, low-resolution sampling intervals (>18
 586 days/sample, June 2003 through September 2006, Table 1). However, the high-resolution intervals
 of CBeu deployments 4 and 5 (up to 7.5 days per sample, Oct 2006-March 2008, Table 1) should
 588 have captured a possible short-term succession of major groups (e.g., diatoms quickly reacting to
 increasing nutrient availability, whereas photosynthetic dinoflagellates becoming more abundant
 590 during upwelling relaxation, Margalef, 1963; Jiménez-Quiroz et al., 2019). Although we do not
 dismiss the possibly occurrence of succession within studied populations at site CBeu, at this
 592 stage we do not disregard either its occurrence. It should also be kept in mind that the deployed
 traps capture those microorganism remains that reach the trap cups at around 1,300 m water
 594 depth, while they hardly capture green algae or cyanobacteria thriving in surface waters. CBeu
 traps at ca. 1,300 m water depth capture a mixed signal of sinking particles from a surface
 596 catchment area of at least ca. 100 km² (Siegel and Deuser, 1997, Fischer et al., 2016,) due to (i)
 differential settling velocities of particles (Fischer and Karakaş, 2009; Iversen et al., 2010; van der
 598 Jagt et al., 2018), and (ii) highly heterogeneous and dynamic surface water conditions due to
 filament and eddy activity off Mauritania (Mittelstaedt, 1991; Gabric et al., 1993; Meunier et al.,
 600 2012; Cropper et al., 2014). Additionally, the trapped signal is always affected by dissolution of
 particular species and/groups of organisms sinking through the water column into deeper waters
 602 (e.g., Romero et al., 1999, 2002).

5.2. Temporal variations of the species-specific composition of the plankton community

604 We are aware that 1,900 days of continuous sampling cannot deliver a definite picture of all
 temporal changes affecting the composition of the plankton community in the very dynamic
 606 Mauritanian upwelling. However, the overall temporal pattern observed led us to propose a general

sequence of seasonal variability. Most of the major microorganisms' groups occur simultaneously and clear successional trends are not quite distinguishable (Fig. 3). Despite strong interannual variability (Table 4), a consistent seasonal pattern in the occurrence of species or groups of species can still be recognized. Figure 4 shows the seasonal evolution of populations responding to the temporal dynamics of nutrient availability, e.g. following short-period dust events (Fig. 2e) and/or vertical mixing events associated with stronger winds (Fig. 7a, e). Based on the visual data examination and the statistical analysis, four groups of species are recognized (Figs. 3, 6). Populations of group 1 (Dia-bent, Dia-upw, Co-Gocean, OD-upw, For-cold = in blue in Fig. 6) have higher relative contribution during the most intense phase of the upwelling season (mainly between late winter/early spring and early summer; Mittelstaedt, 1983, Cropper et al., 2014). Group 1 quickly responds to intense mixing and lowered SST at the CBeu site (Fig. 7d, e) and represents the typical upwelling-related association off Mauritania. This observation confirms the ecological characterization of the species groups that has been separately presented in previous biogeographical/ecological studies (Romero et al., 2002; Kucera, 2007; Köbrich et al., 2008, 2016; Zonneveld et al., 2013; Romero and Fischer, 2017).

Diatoms of coastal regions (Dia-coast, non-upwelling related) and those thriving in open ocean waters (Dia-ocean) together with other calcareous dinoflagellates (CD-other), cosmopolitan organic dinoflagellate cysts (OD-cosm) and 'other coccolithophores' (Co-other) are assigned to group 2 (in brown in Fig. 6). Except for the cosmopolitan organic dinoflagellate cysts, all components of group 2 are primary producers and occur more abundantly between early fall and late winter (Fig. 4), at times of deepening of the ML and upwelling relaxation (Fig. 7e). Group 2 represents a primary producer signal typical of meso- to oligotrophic waters conditions off Mauritania, occurring under weakened upwelling, when winds predominantly blow from the N-NE, SST start decreasing after their summer peak, and the uppermost water column stratifies (Fig. 7a, d, e).

Except for warm waters (CD-warm) and dust input-sensitive (CD-min) calcareous dinoflagellate cysts, group 3 is mainly composed by secondary producers: warm-water planktonic foraminifera and all pteropods (Fig. 4e, f). As such, this group represents the calcareous fraction of zooplankton feeding on (primary) phytoplankton, occurring mainly during phases of predominantly warmer

636 SSTs (Fig. 7d), N-NE-originated winds (Fig. 7a) and stratified uppermost water column (Fig. 7e).
 SST appears to be the most important factor controlling assemblage composition of planktonic
 638 foraminifera (Kucera, 2007). Large, symbiont-bearing specialists like *G. ruber* and *G. sacculifer* are
 adapted to more oligotrophic and warmer waters. They show their maximum abundance in warm
 640 waters with a deeper mixed-layer depth (Fig. 7e, f).

The seasonal dynamics of group 4 is similar to that of group 3 (intervals of weakened upwelling
 642 conditions), but they differ in their composition: group 4 is mainly made of calcareous primary
 producers. These populations dominate the plankton community during intervals of weakened
 644 upwelling, shallow MLD and predominantly oligotrophic water conditions. Similar to group 3, group
 4 consists mainly of coccolithophores (the dominant *E. huxleyi*, accompanied by UPZ and LPZ, *U.*
 646 *anulus*, Figs. 4b, 6), as well as organic dinoflagellate cysts characteristic for upwelling relaxation
 phases (CD-upw relax). The contribution of *E. huxleyi* and accompanying coccolithophore taxa,
 648 and upwelling-relaxation organic dinoflagellate cysts shows highest relative values from early fall
 through early spring and decreases into the most intense upwelling season (when *G. oceanica*
 650 increases, Fig. 4b). As such, this group also bears some resemblance to group 2, though coastal
 and open-ocean water diatoms are component of the latter, while diatoms are absent in group 4.
 652 This difference possibly reflects the distinct nutrient and water depth conditions in which *E. huxleyi*
 and other coccolithophores (group 4) and diatoms (group 2) typically thrive.

654 **5.3. Shifts in the species-specific composition of assemblages between 2004 and 2006**

The persistent seasonal pattern of the groups' and species occurrence experiences occasional
 656 shifts. Several events, which altered the 'regular' pattern of temporal occurrence of species or
 group species at site CBeu, were observed between late 2004 and late 2006 (Fig. 7f-j). We identify
 658 three main shift stages in the species-specific composition of assemblages:

1. Stage 1 (2004): (i) low total biogenic production (summer–fall 2004, Fig. 2b-d), and (ii)
 660 absence of warm-water foraminifera (Fig. 7f). These changes in production/flux were
 accompanied by (iii) a significant decrease in SST as reconstructed with $U_{37}^{K'}$ (Fig. 7d).
2. Stage 2 (late 2005/early 2006): (i) extraordinarily high relative contribution of the
 662 coccolithophore *U. anulus* (Fig. 7i); as well as (ii) the first high occurrence of *L. bulimoides*
 664 and uncoiled pteropods (Fig. 7h).

3. Stage 3 (after fall 2006): (i) strong increase of the relative contribution of benthic diatoms (Fig. 7i) and warm-water calcareous dinoflagellates (Fig. 7j), and (ii) highest longest occurrence and highest relative abundance of *L. bulimoides* (fall 2006/winter 2007, Fig. 7h).

A certain degree of interannual variability of the physical setting (Mittelstaedt, 1983, 1991; Cropper et al., 2014) might explain the shifts in the species-specific composition of the assemblages. The near-disappearance of warm-water planktonic foraminifera in 2004 (Fig. 7f) was most probable the response to lower-than-usual water temperatures (Fig. 7d). However, the SST decrease is not recorded by satellite imagery. The overall climate evolution indicates a longer warm and dry period from 2001-2004 in the Sahel and Sahara (east of site CBeu) and anomalously warm temperatures in the Eastern Atlantic (Zeeberg et al., 2008; Alheit et al., 2014). 2004 is the only year of our study with noticeable temporal discrepancies between satellite and $U_{37}^{K'}$ -based temperature (Fig. 7d). This temperature gap suggests a certain decoupling between the temperature signal of the uppermost centimeters of the water column (satellite) and subsurface waters where the alkenone-forming coccolithophores dwell (*E. huxleyi* and *G. oceanica*; Conte et al., 1995). As planktonic foraminifera mainly react to SST variability (Kucera, 2007), cooler than usual subsurface waters between middle winter and early fall 2004 (Fig. 7d) might have been responsible for the strong decrease of the warm-water planktonic foraminifera contribution (Fig. 7f). Additionally, all other plankton groups show lowest fluxes toward late summer. Neither the seasonal pattern nor the MLD show any significant change nor unusually high lithogenic fluxes occurred (Fig. 2e, 7e). Exceptionally, the winter season 2004/2005 is characterized by a high total mass flux (Fig. 2a); this extraordinarily high seasonal value matches well with the highest fluxes of TOC and $CaCO_3$ for the studied interval.

The extraordinary high relative abundance of *U. anulus* in fall 2005 has not yet been observed in similar or other settings, although it is often listed in studies of large-scale distribution patterns of coccolithophores (e.g., Böckel and Baumann, 2008; Estrada et al., 2016; Poulton et al., 2017). So far only Steinmetz (1991) has found *U. anulus* (described as *U. calvata* and *U. scituloma*) in 'frequent' abundances in sediment traps deployed in the equatorial Atlantic, central Pacific, and in the Panama Basin, but without adding detailed information such as fluxes, the timing of its

694 occurrence or its ecological significance. In most of earlier trap studies, *U. anulus* has been
 grouped together with other umbilicosphaerids coccolithophores, since it did not reach high
 696 abundances (e.g., Köbrich et al., 2016; Guerreiro et al., 2017). Nevertheless, umbilicosphaerids
 seem to favor warm and more oligotrophic conditions (Baumann et al., 2016), so that the increased
 698 input of tropical surface waters transported northward via the MC (Mittelstaedt, 1991) can be
 possibly responsible for the advection of *U. anulus* upon the site CBeu.

700 The shift in the pteropod composition from dominating *H. inflatus* towards the appearance of *L.*
bulimoides between winter 2005 and spring 2006 –and again in fall and winter 2006/2007 (Fig.
 702 7h)– can be also explained by the increased influence of warmer surface waters of southern origin.
Heliconoides inflatus is known as a rather cosmopolitan species, occurring across a wide range of
 704 oceanic provinces (Bé and Gilmer, 1977; BurrIDGE et al., 2017), whereas *L. bulimoides* seems to
 prefer waters of subtropical gyres (although it was also present in low numbers in the equatorial
 706 region, BurrIDGE et al., 2017). A stronger transport of the MC from the south may have led to the
 deterioration of the adequate environmental conditions for *H. inflatus*, as can be seen by the
 708 extremely low total pteropods flux during winter 2005 to spring 2006 (Fig. 3e), and, thus, to the
 relative enrichment of *L. bulimoides*. The fact that the latter species is absent again in winter 2008
 710 (Fig. 7h) represents a gradual return to previous ('regular') winter conditions. 'Regular'
 temperatures from early 2005 on allowed the reappearance of warm-water planktonic foraminifera
 712 in fall 2005 (Fig. 7f).

The exceptional increase in the contribution of the benthic diatoms in spring-summer 2006 (Fig.
 714 7i) might have been possibly related to the intensification of lateral advection upon the
 intermediate-waters deployed CBeu trap (Romero and Fischer, 2017). Observational and model
 716 experiments show that the transport of particles from the Mauritanian shelf and the uppermost
 slope via nepheloid layers significantly contributes to the deposition upon the lowermost slope and
 718 beyond than the direct vertical settling of particles from the surface layer (Nowald et al., 2014;
 Karakaş et al., 2006; Fischer et al., 2009; Zonneveld et al., 2018). The relevance of advective
 720 processes within nepheloid layers has been already proposed for similar settings (Puig and
 Palanques, 1998; Inthorn et al., 2006). We speculate that the longer predominance of N-NE winds
 722 between 2005 and 2007 (Fig. 7a) might have possibly intensified the transport of benthic diatoms

from the shallow coastal area into the hemipelagic CBeu trap via the MC (Fig. 1). Enhanced lateral
 724 transport has important environmental implications for the final burial of organic matter in EBUEs.
 As the organic matter can be effectively displaced from the area of production (Inthorn et al.,
 726 2006), carbon depocenters generally occur at the continental slopes between 500 and 2,000 m. In
 the CC-EBUE around Cape Blanc, the depocenter with up to 3% of organic carbon has a depth
 728 range between 1,000 and 2,000m (Fischer et al., 2019).

Most of the populations affected by and responding to shifting environmental conditions off
 730 Mauritania between 2004 and 2006 returned to their 'regular' seasonal pattern of occurrence after
 2006 (Fig. 4). However, some shifts persisted after summer 2006. *Limacina bulimoides* still
 732 dominated the pteropod assemblage (Fig. 7h), the total pteropod flux showed the highest maxima
 for the entire studied interval (might be due to the large food supply and organic matter as
 734 represented by high total fluxes of diatoms, Fig. 3a, e), and warm-water calcareous dinoflagellate
 cysts increased during late fall 2006 (Fig. 7j). An exception to this pattern is the high relative
 736 contribution of benthic diatoms (Figs. 4a, 7i; Romero and Fischer, 2017). At this stage, we cannot
 fully disregarded that the shift in the species-specific composition of the diatom community (also
 738 present after 2008; Romero and Fischer, 2017; Romero, unpublished observations) might be due
 to the natural long-term variability due to external forcings (e.g., North Atlantic Oscillation) or due to
 740 on-going climate change.

Our multiannual trap experiment provides a unique opportunity to study the long-term evolution
 742 of the plankton community in an ecologically important EBUE. Rapid shifts in the population
 contribution at site CBeu demonstrate that calcareous, siliceous and organic plankton
 744 microorganisms rapidly react to environmental changes in the CC-EBUE off Mauritania. Time-
 series trap experiments continuously conducted over many years –as those currently in the CC-
 746 EBUE (Fischer et al., 2016, 2019; Romero et al., 2002, 2016, 2017)– deliver a reliable
 observational basis on the occurrence of long-lasting variations of populations in response to key
 748 environmental forcings. Among others, our multiannual observations are useful for future model
 experiments on plankton dynamics and evolution in low- and mid-latitude EBUEs and how
 750 organisms influencing the global carbon cycle might react to global and ocean warming.

752 **6. Conclusions**

- 754 - The seasonal amplitude of the flux variations of primary and secondary producers in the upper
 water column off Mauritania was well recorded in our continuous trap experiment that lasted 1,900
 days. The repeated yearly pattern of higher fluxes of diatoms, coccolithophores, organic-walled
 756 dinoflagellate cysts and planktonic foraminifera between early spring and early/middle summer
 matched well the temporal occurrence of the most intense upwelling interval in waters overlying
 758 site CBeu. Conversely, fluxes of calcareous dinoflagellate cysts and pteropods were higher during
 intervals of upwelling relaxation (late summer through late fall).
- 760 - The good temporal match between maxima of (most of) studied microorganism groups and
 biogenic bulk components unambiguously evidences the contribution of primary and secondary
 762 producers to the total mass/biogenic mass fluxes. The notorious coupling between fluxes of
 lithogenics and major microorganism groups provides compelling evidence for the biological pump
 764 off Mauritania to be strongly dependent on the dust input from the Sahara/Sahel and the eolian-
 transported nutrient deposition.
- 766 - 1,900 days of continuous trap record of microorganism fluxes allowed for the recognition of a
 general sequence of seasonal variations of the main plankton populations thriving in coastal
 768 waters off Mauritania. The temporal turnover (succession) was better shown by the temporal
 variations of particular species or groups of species.
- 770 - A significant shift in the 'regular' seasonal pattern of populations' occurrence was recognized in
 species relative contributions between 2004 and 2006. Several events altering the regular
 772 seasonal pattern were observed and occurred in three main stages: summer–fall 2004, late
 2005/early 2006, and after fall 2006. Although most of the populations return 'to normal' after fall
 774 2006, a few did not.
- 776 - Our multiannual trap experiment emphasizes the significance of long-term records on
 evaluating the impact of changing environmental conditions on living populations. Time-series trap
 experiments conducted over many years –as those currently conducted in the CC-EBUE by
 778 MARUM– deliver a broad observational basis on the occurrence of persistent seasonal and
 interannual patterns. They also help in the interpretation of long-lasting variations of population

780 changes in response to key environmental forcings, such as nutrient input, water masses
variability, lateral transport and/or climate change. The applicability of the flux dynamics of primary
782 and secondary producers here presented is not limited to the Mauritanian upwelling system, and it
might comparable to other EBUEs.

784

Code and Data Availability

786 Data are available at <https://doi.pangaea.de/10.1594/PANGAEA.904390>

Author Contributions

788 All authors collected the data. Oscar E. Romero wrote the manuscript. All authors contributed to
790 results interpretation and discussion.

Competing Interests

792 The authors declare that they have no conflict of interest.

794

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Figures

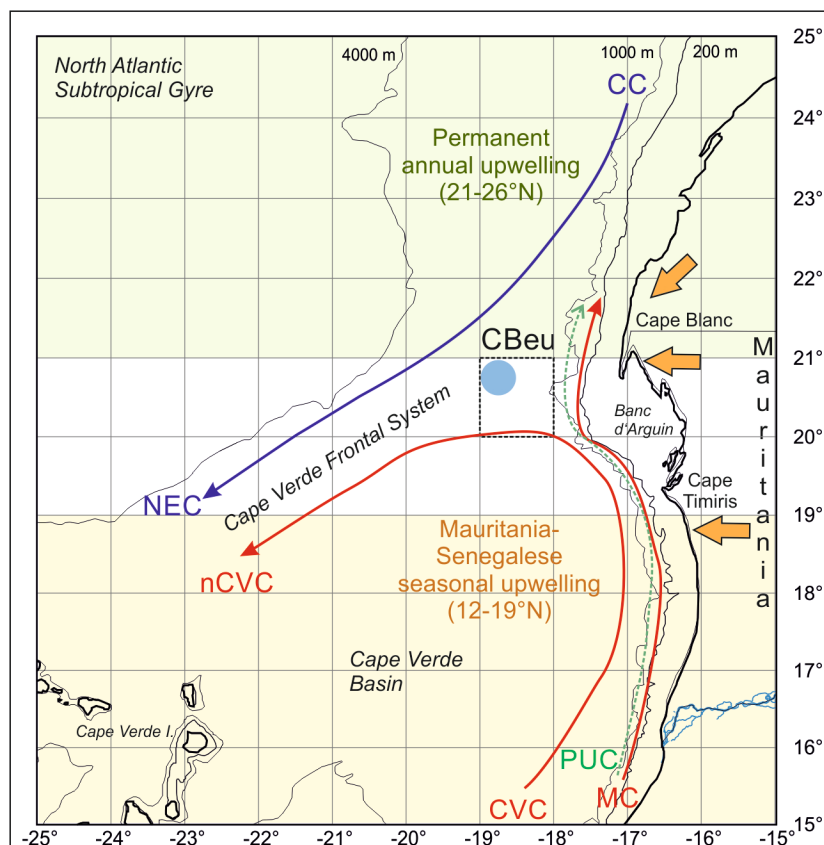


Figure 1. Map of the study area showing the location of trap site CBeu (full light blue dot), surface currents, and main wind system. Surface currents (Canary Current, CC, violet line; North Equatorial Counter Current, blue arrow; Mauritanian Current; red arrow), North Equatorial Current (NEC), Cape Verde Current (CVC), north Cape Verde Current (nCVC), PUC are depicted after Mittelstaedt (1983, 1991) and Zenk et al. (1991). The Cape Verde Frontal Zone (CVFZ) builds at the confluence of the NACW and the SACW (Zenk et al., 1991). Trade winds and Saharan Air Layer are represented by orange arrows (Nicholson, 2013). The upwelling zones are depicted after Cropper et al. (2014).

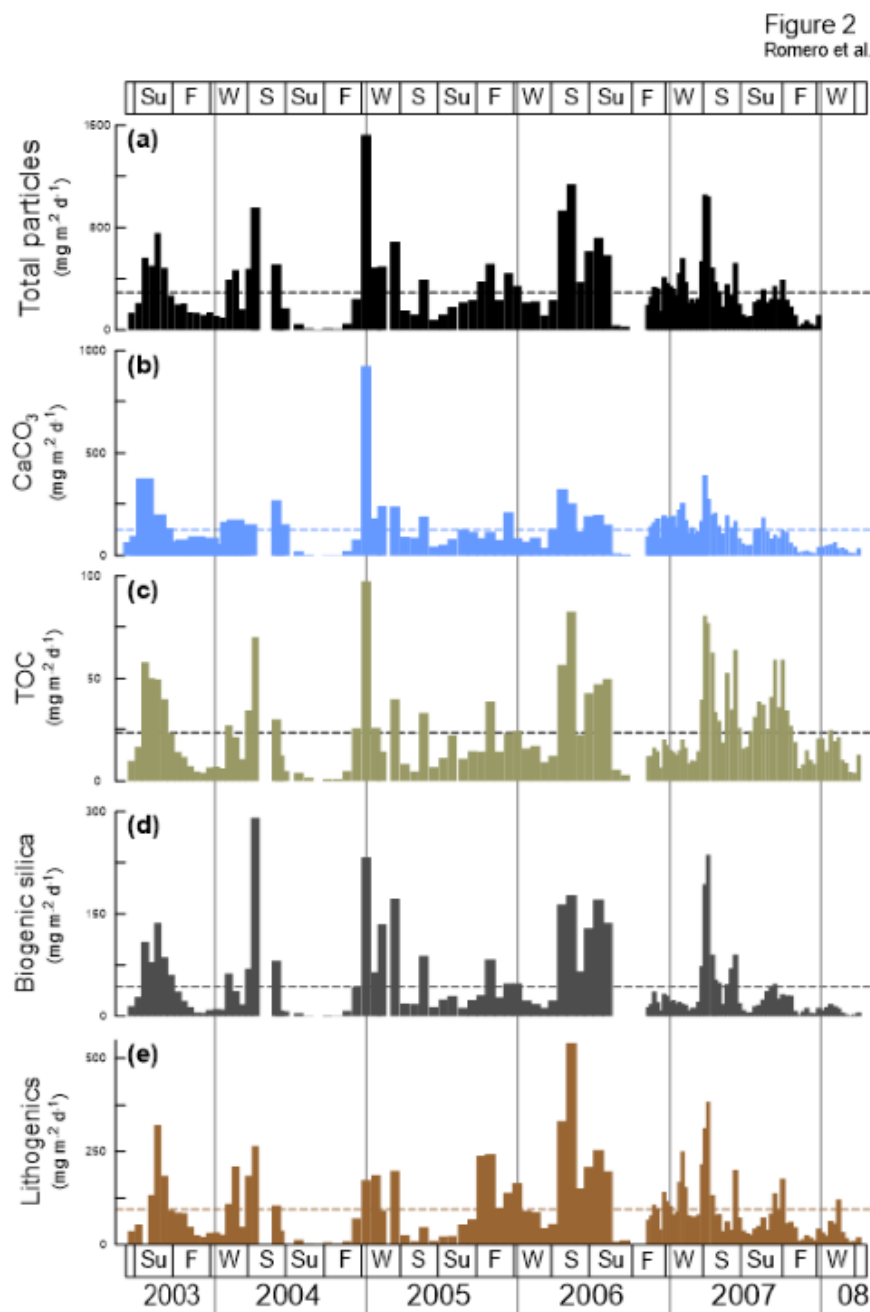


Figure 2. Total mass particle and bulk fluxes at the trap site CBeu between June 2003 and March 2008. From top to bottom: (a) total mass particle ($\text{mg m}^{-2} \text{d}^{-1}$, black bars), (b) calcium carbonate (CaCO_3 , $\text{mg m}^{-2} \text{d}^{-1}$, light blue bars), (c) total organic carbon (TOC, $\text{mg m}^{-2} \text{d}^{-1}$, olive bars), (d) biogenic silica (BSi, opal, $\text{mg m}^{-2} \text{d}^{-1}$, dark grey bars), and (e) lithogenics ($\text{mg m}^{-2} \text{d}^{-1}$, brown bars). The horizontal striped line for each parameter represents the average flux for the whole studied interval (Table 1). The boxes in the upper and lower panels represent seasons (Su=summer, F=fall, W=winter, S=spring). The vertical background gray lines indicate calendar year separation. For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.

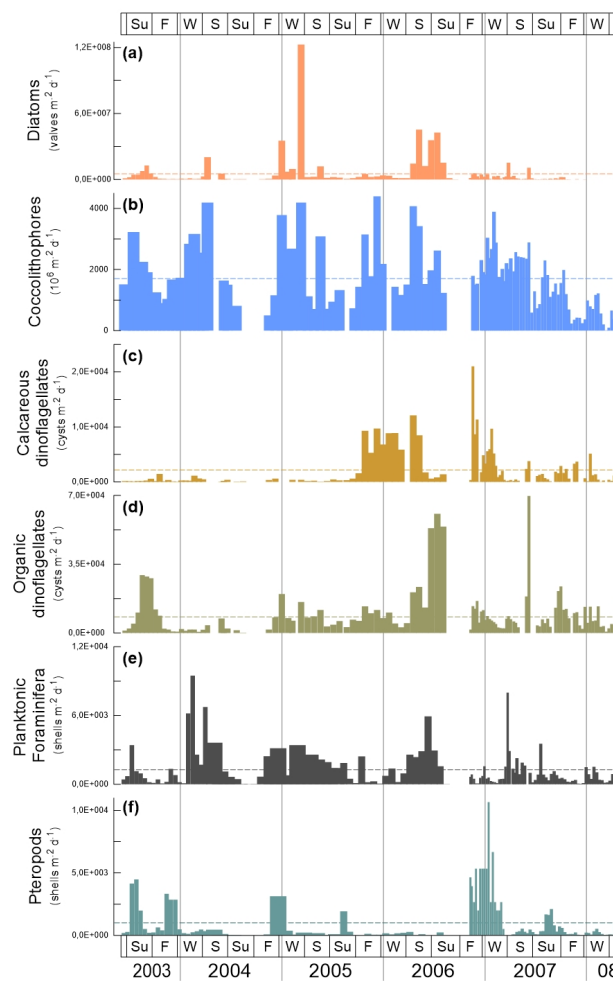
Figure 3
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Figure 3. Fluxes of microorganisms at the trap site CBeu between June 2003 and March 2008.

From top to bottom: (a) diatoms (valves $\text{m}^{-2} \text{d}^{-1}$, peach bars; note that ten samples corresponding to CBeu 5 –12/13/2007 through 03/17/2008– were not available for diatom analysis); (b) coccolithophores (coccoliths $\text{m}^{-2} \text{d}^{-1}$, light blue bars); (c) calcareous dinoflagellates (cysts $\text{m}^{-2} \text{d}^{-1}$; gold bars); (d) organic dinoflagellates (cysts $\text{m}^{-2} \text{d}^{-1}$; khaki bars); (f) planktonic foraminifera (shells $\text{m}^{-2} \text{d}^{-1}$; grey bars), and (f) pteropods (shells $\text{m}^{-2} \text{d}^{-1}$; ocean green bars). The horizontal striped line for each group of organisms represents the average flux for the whole study interval. The boxes in the upper and lower panels represent seasons (Su=summer, F=fall, W=winter, S=spring). The vertical background gray lines indicate calendar year separation. For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.

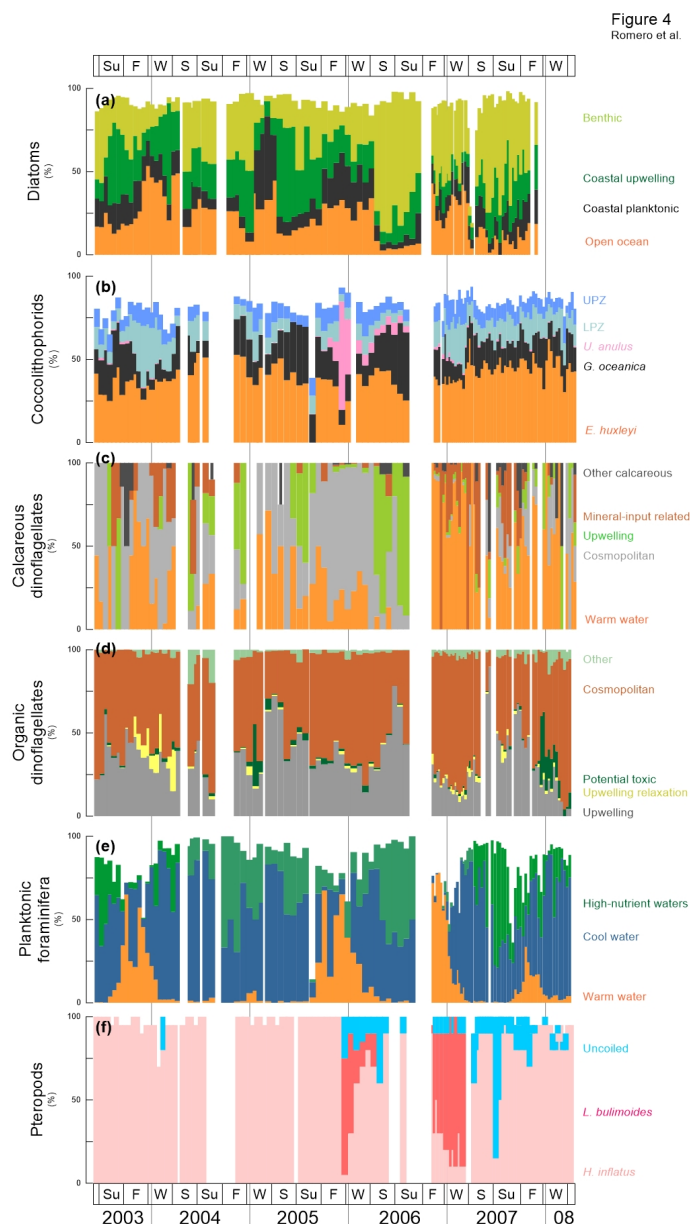


Figure 4. Cumulative relative abundance (%) of main species or group of species of diatoms, coccolithophores, dinoflagellates, planktonic foraminifera and pteropods at the trap site CBeu between June 2003 and March 2008 (Table 1). From top to bottom: (a) diatoms - benthic, light green bars; coastal upwelling, dark green bars; coastal planktonic, black bars; and open-ocean (%, orange bars; note that ten samples corresponding to CBeu 5 –12/13/2007 through 03/17/2008– were not available for diatom analysis); (b) coccolithophores –upper photic zone, blue bars; lower photic zone, moss green; *Umbilicosphaera anulus*, pink bars; *Gephyrocapsa oceanica*, black bars; *Emiliana huxleyi*, orange bars; (c) calcareous dinoflagellates – other calcareous, dark grey bars; mineral-input related, brown bars; upwelling, light green bars; cosmopolitan, light grey; warm water, orange bars; (d) organic dinoflagellates – upwelling

species (grey bars); upwelling relaxation species (light yellow bars); potential toxic (dark green bars); cosmopolitan, red brown bars; other, faded green bars; (e) planktic foraminifera – high nutrient water, green bars; cool water, blue bars; warm water, orange bars; and (f) pteropods – uncoiled species, light blue bars; *Limacina bulimoides*, red bars; *Heliconoides inflatus*, pink bars. Note that the relative contribution of diatoms, coccolithophores and planktonic foraminifera does not add up to 100% since the ecology of some taxa of these groups is not well known constrained. The species-specific composition of groups is presented in Table 5. The boxes in the upper and lower panels represent seasons (Su=summer, F=fall, W=winter, S=spring). The vertical background gray lines indicate calendar year separation. For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.

Figure 5
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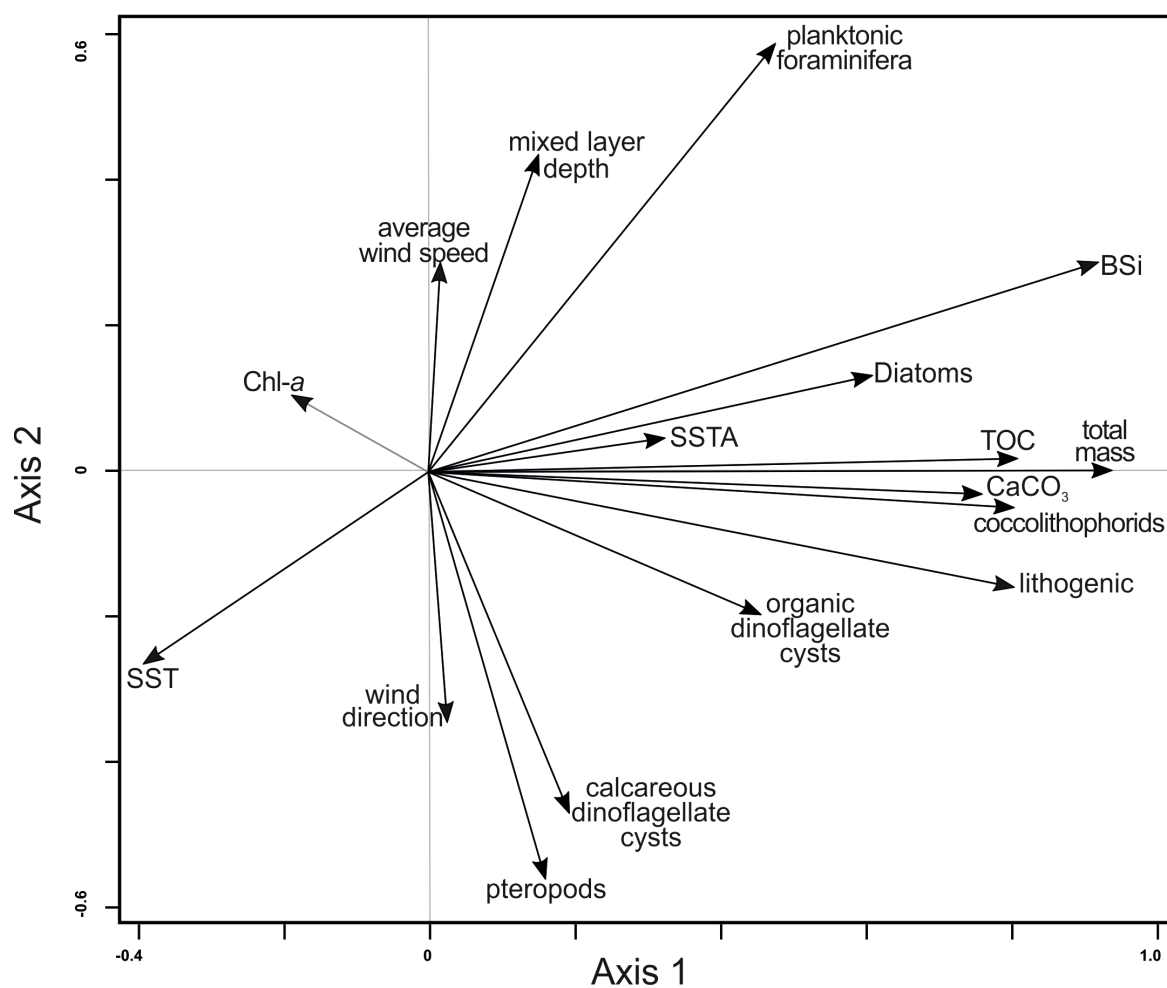


Figure 5. RDA ordination diagram depicting the relationship between the accumulation rates of organism groups and bulk fluxes and environmental conditions in upper waters. References: Av. wind speed=average wind speed; Chl-*a*=chlorophyll *a*; TOC=total organic carbon; CaCO₃=calcium carbonate; MLD=mixed layer depth; SST=sea surface temperature; SSTA=sea surface temperature anomalies. For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.

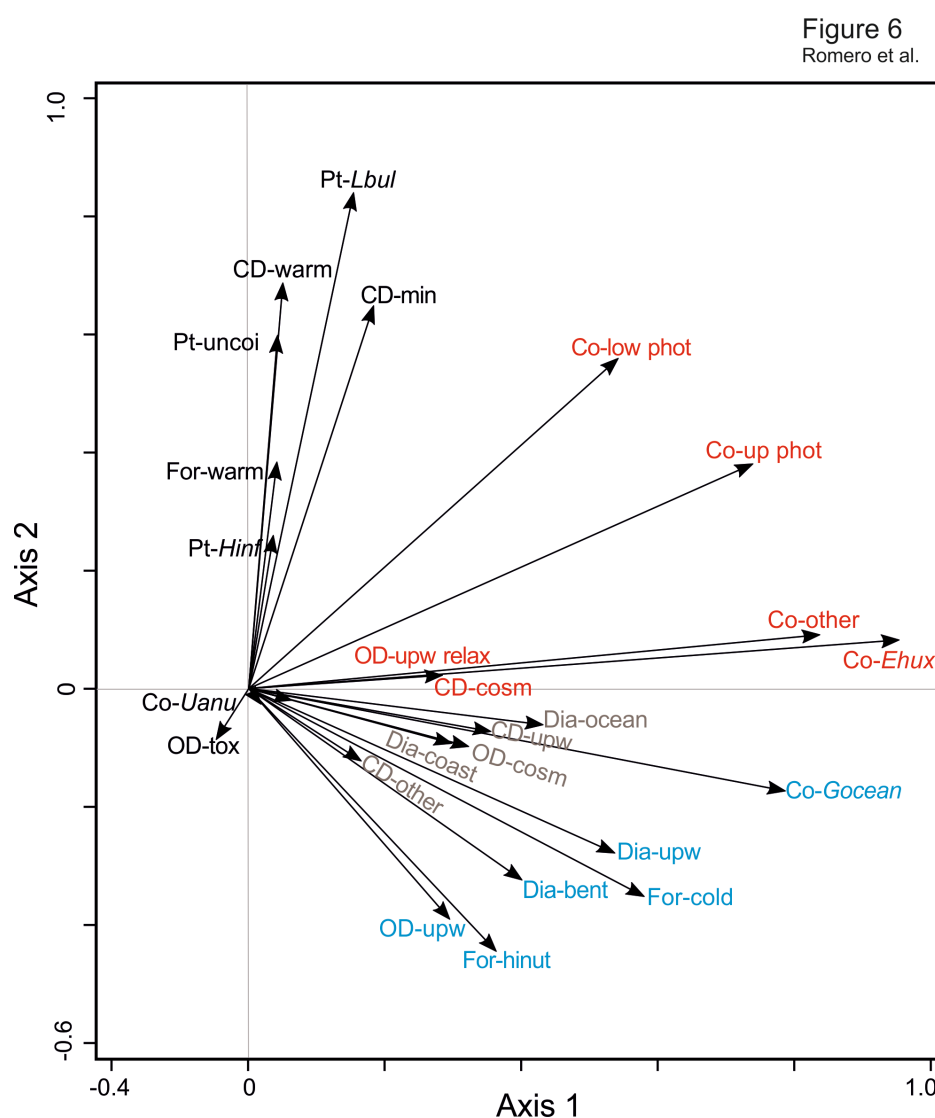


Figure 6. Results of a PCA analysis of ecological groups of the organism groups at the CBeu trap site between June 2003 and March 2008. References: Dia (diatoms): -bent = benthic, -coast = coastal planktonic, -ocean = open ocean, -upw = upwelling; Co (coccolithophores): -*Ehux* = *Emiliana huxleyi*, -*Gocean* = *Gephyrocapsa oceanica*, -low phot: low photic zone, -other: other coccolithophroids, -*Uanu* = *Umbilicosphaera anulus*, -up phot: upper photic zone; CD (calcareous

dinoflagellate cysts): -cosm = cosmopolitan group, -min = terrestrial mineral group, -other =
species that do not fit in one of the other ecological groups, -upw = upwelling, warm: warm
waters; OD (organic-walled dinoflagellate cysts): -cosm = cosmopolitan group, -other = species
that do not fit in one of the other ecological groups, -tox = potential toxic group, -upw = upwelling,
-upw relax = upwelling relaxation; For (planctonic foraminifera): -cold: cold water group, -hinut =
high nutrient waters group; -warm = warm water group; and Pt (pteropods): -*Hinf* = *Heliconoides inflatus*, -*Lbul* = *Limacina bulimoides*, -uncoi: uncoiled. Groups of microorganisms are identified
by colors (light blue, group 1; brown, group 2; black, group 3; and red, group 4). The species-
specific composition of groups is presented in Table 5. For interpretation of the references to
color in this figure legend, the reader is referred to the web version of this article.

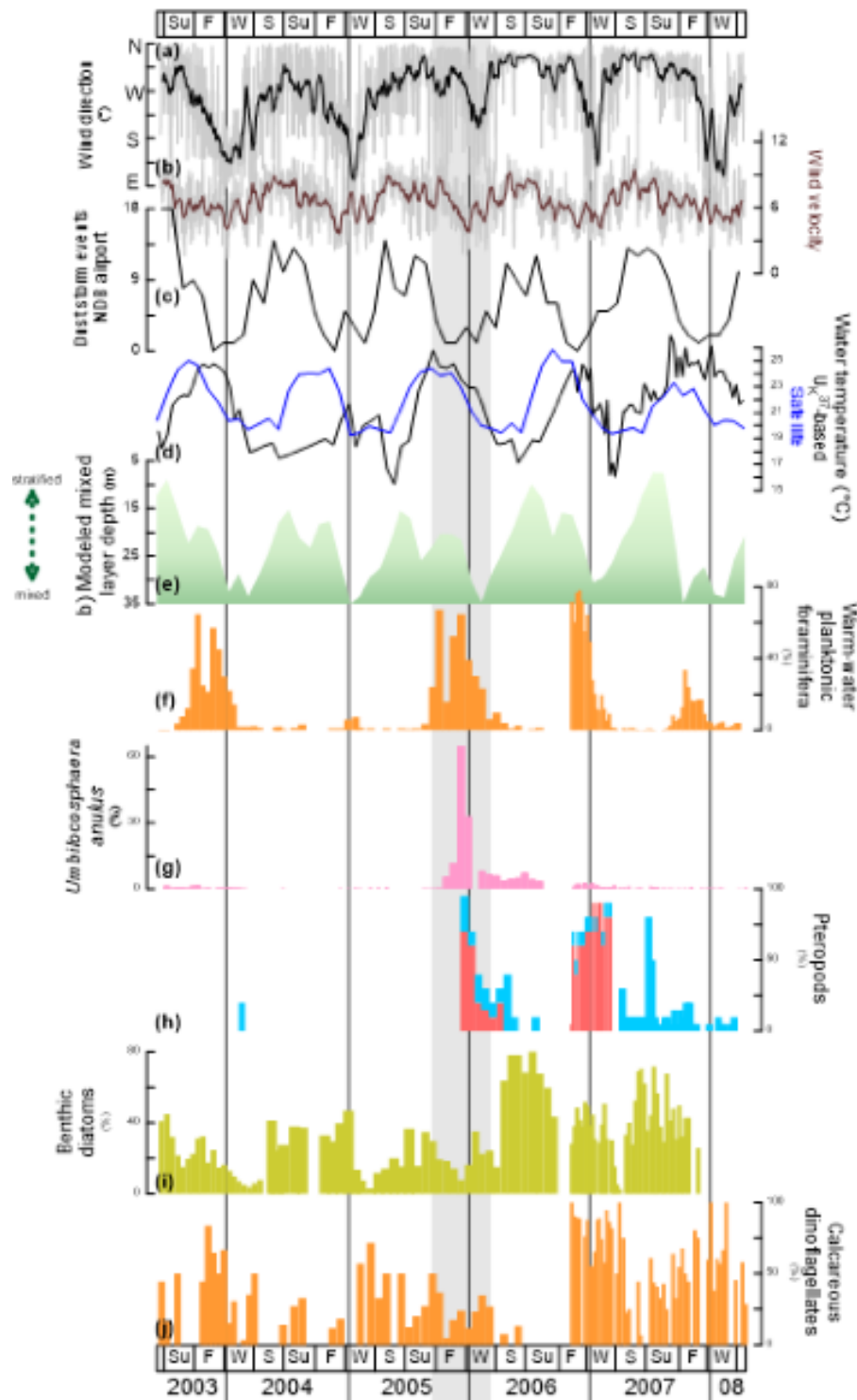
Figure 7
Romero et al.

Figure 7. Comparison physical data (a-e) and relative abundance of selected species or group/s of species (f-j) at site CBeu between June 2003 and March 2008. Physical data: (a) origin of daily wind direction (the grey line are daily data, the thicker black line represent the 17-running point average); (b) daily wind velocity (m s^{-1} , the grey line are daily data, the thicker dark brown line represent the 17-running point average), (c) dust storm event (number of events) recorded at the

meteorological station of the Airport of Nouadhibou (ca. 20°57'N, 17°02'W, Mauritania); (d) seawater temperature (°C): $U_{37}^{K'}$ -based reconstruction (black line) and satellite-imagery generated data (blue line); (e) mixed layer depth (m, grey line, <https://modis.gsfc.nasa.gov> for the area between 19°-18°W and 20°-21°N). Relative contribution (%) of (f) warm-water planktonic foraminifera (orange bars); (g) the coccolithophore *A. anulus* (pink bars); (h) pteropods *L. bulimoides* (red bars) and uncoiled species (light blue bars); (i) benthic diatoms (banana yellow bars); and (j) calcareous dinoflagellates (light orange bars). The species-specific composition of groups is presented in Table 5. The vertical gray lines indicate years separation. The light grey shading in the background highlights the interval of main shift in fluxes values and/or the relative contribution of particular species or group of species (see discussion in 5.3.). For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.

Tables

Table 1 – Romero et al.

Mooring CB _{eu}	Coordinates	GeoB-#/cruise	Trap depth (m)	Ocean bottom depth (m)	Sample amount	Capture duration (sample/days)	Sampling interval
1	20°45'N 18°42'W	- POS 310	1,296	2,714	20	1 = 10.5, 2-20 = 15.5	06/05/2003 – 04/05/2004
2	20°45'N 18°42'W	9630-2 M 65-2	1,296	2,714	20	1-20 = 22, 2-19 = 23	04/18/2004 – 07/20/2005
3	20°45.5'N 18°41.9'W	11404-3 POS 344-	1,277	2,693	20	21.5	07/25/05 – 09/28/2006
4	20°45.7'N 18°42.4'W	11835-2 MSM 04b	1,256	2,705	20	1 = 3.5, 2-20 = 7.5	10/28/2006 – 03/23/2007
5	20°44.9'N 18°42.7'W	12910-2 POS 365-	1,263	2,709	38	1, 2 = 6.5, 3-38 = 9.5	03/28/2007 – 03/17/2008

Table 1: Data deployment at site CBeu (Cape Blanc eutrophic, off Mauritania): coordinates, GeoB

location and cruise, trap depth, ocean bottom depth, sample amount, capture duration of each

sample and sampling interval. Two gaps in the sampling intervals occurred: 04/05/2004–

04/18/2004, and 09/28/2006–10/28/2006.

Table 2

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Analysis	Method	Analysed Parameters	Length of gradient	Eigenvalue Axis 1 (%)	Eigenvalue Axis 2 (%)	Eigenvalue Axis 3 (%)	Eigenvalue Axis 4 (%)	P-value
1	RDA	Fluxes of microorganism s and bulk parameters, environmental parameters	1.8	34.5	10.7	4.7	2.1	0.002
2	PCA	microorganisms	1.4	26.3	16.2	9.8	6.9	

References: RDA, Redundancy Analysis; PCA, Principal Component Analysis.

Table 2: Main result values of the ordination techniques Redundancy (RDA) and Principal

Component (PCA) analyses performed with the software Package Canoco 5 (ter Braak and Smilauer, 2012; Smilauer and Leps, 2014).

Table 3

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Year	Total mass (mg m ⁻² d ⁻¹)	CaCO ₃ (mg m ⁻² d ⁻¹)	(%)	TOC (mg m ⁻² d ⁻¹)	(%)	Biogenic silica (mg m ⁻² d ⁻¹)	(%)	Lithogenics (mg m ⁻² d ⁻¹)	(%)
2004	309.5+403.9	150.9+360.2	48.75	21.93+39.7	7.15	54.61+85.6	17.57	84.6+66.5	27.33
2005	317.5+176.6	125.5+65.3	39.45	19.00+11.00	6.00	52.4+46.2	16.50	101.7+82.5	32.03
2006	377.9+281.9	142.8+79.1	37.79	23.3+20.6	6.19	54.07+60.2	14.31	134.2+122.6	35.51
2007	282.8+225.1	121.1+80.2	42.18	29.4+19.6	10.38	35.8+46.9	12.66	92.9+83.4	32.85

Table 3: Average daily fluxes of total mass, calcium carbonate (CaCO₃), total organic carbon

(TOC), biogenic silica and lithogenics (mg m⁻² d⁻¹) calculated at site CBeu for full calendar years 2004, 2005, 2006 and 2007.

Table 4

Year	Diatoms					Coccolithophores					Calcareous dinoflagellate cysts					Organic diatoms			
	Flux	Benthic	Coastal upwelling	Coastal planktonic	Open-ocean	Flux	<i>E. huxleyi</i>	<i>G. oceanica</i>	Low photic zone	Upper photic zone	<i>A. anulus</i>	Flux	Other calcareous	Mineral input	Upwelling	Cosmopolitan	Warm waters	Flux	Upwelling
(valves m ⁻² d ⁻¹)	Relative abundance (%)					(coccoliths ×10 ⁶ m ⁻² d ⁻¹)	Relative abundance (%)					(cysts m ⁻² d ⁻¹)	Relative abundance (%)					(cysts m ⁻² d ⁻¹)	
2004	4770323±14831598	23.96±15.54	27.56±10.95	27.56±10.95	27.56±10.95	2094±1159	43.73±6.70	14.77±7.98	13.86±2.21	6.72±1.92	0.14±0.18	389±312	2.73±2.50	14.47±14.71	20.24±29.22	32.25±28.00	14.93±16.85	3823±5164	30.59±10.23
2005	11189496±29887484	17.66±9.31	27.26±17.40	27.26±17.40	27.26±17.40	2014±1224	36.95±9.23	20.19±6.21	9.87±3.09	9.45±3.14	7.41±17.53	2228±3457	1.68±6.23	1.23±3.07	10.17±18.84	51.47±29.53	29.20±21.69	8128±3626	40.42±15.41
2006	9699612±13964939	48.40±19.03	12.76±6.88	12.76±6.88	12.76±6.88	2056±833	37.29±5.22	21.63±9.70	10.45±5.11	10.16±3.85	3.93±2.22	5556±5490	1.85±2.92	11.83±24.40	21.16±32.24	25.73±33.54	39.42±38.65	17160±17780	32.21±16.86
2007	1785459±3114341	40.05±17.03	17.76±9.12	17.76±9.12	17.76±9.12	1616±872	44.72±6.53	15.35±5.17	14.61±5.75	9.64±2.63	0.53±0.44	1851±2104	4.27±10.39	20.57±23.73	6.26±8.67	9.57±13.36	53.61±28.29	9600±11757	33.00±20.06

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nofflagellate cysts			Planktonic Foraminifera				Pteropods			
Upwelling relaxation	Potential toxic	Cosmopolitan	Flux	Upwelling	Cool waters	Warm waters	Flux	Uncoiled	<i>L. bulimoides</i>	<i>H. inflatus</i>
Relative abundance (%)			(shells m ⁻² d ⁻¹)		Relative abundance (%)		(shells m ⁻² d ⁻¹)	Relative abundance (%)		
5.46±8.48	1.61±1.55	56.38±8.26	2286±2801	22.76±20.69	63.77±26.21	2.29±3.30	326±741	1.18±4.85	0.00±0.00	65.88±44.88
0.73±0.62	4.92±8.90	52.04±14.46	1228±1129	19.00±13.57	49.42±28.58	17.53±24.59	221±838	1.56±6.25	4.38±17.50	86.88±33.01
1.26±1.57	0.83±1.07	63.85±15.69	1287±1417	20.87±23.64	28.54±25.30	31.83±30.75	1962±2342	10.25±8.66	31.00±29.23	43.50±30.83
2.12±2.07	4.13±9.75	57.33±18.81	996±1373	22.32±20.49	52.20±21.22	7.01±8.18	1034±1981	10.49±14.48	15.61±32.41	64.63±36.75

Table 4: Average daily fluxes diatoms (valves m⁻² d⁻¹), coccolithophores (coccoliths m⁻² d⁻¹),

calcareous and organic dinoflagellate cysts (cysts m⁻² d⁻¹), planktonic foraminifera and pteropods

(shells m⁻² d⁻¹) and average relative contribution of groups of taxa for each of the above-

mentioned microorganisms calculated at site CBeu for full calendar years 2004, 2005, 2006 and

2007.

Table 5

Romero et al.

Diatoms	References
1) Benthic	
<i>Actinoptvchus</i> spp.	Round et al. (1990)
<i>Amphora</i> spp.	
<i>Cocconeis</i> spp.	
<i>Cymatosira belgica</i>	
<i>Delphineis surirella</i>	
<i>Grammatophora marina</i>	
<i>Licmophora</i> sp.	
<i>Odontella mobiliensis</i>	
<i>Psammodyction panduriformis</i>	
<i>Tabularia</i> spp.	
2) Coastal upwelling	
Resting spores of:	Hasle and Svendsen (1996)
<i>Chaetoceros affinis</i>	
<i>Chaetoceros cinctus</i>	
<i>Chaetoceros compressus</i>	
<i>Chaetoceros constrictus</i>	
<i>Chaetoceros coronatus</i>	
<i>Chaetoceros debilis</i>	
<i>Chaetoceros diadema</i>	
<i>Chaetoceros radicans</i>	
3) Coastal planktonic	
<i>Actinocyclus curvatulus</i>	Crosta et al. (2012), Romero et
<i>Actinocyclus octonarius</i>	al. (2009, 2012, 2016, 2017),
<i>Actinocyclus subtilis</i>	—
<i>Chaetoceros concavicomis</i> (vegetative cell,	
<i>Chaetoceros lorenzianus</i> (VC)	
<i>Chaetoceros pseudobrevis</i> (VC)	
<i>Coscinodiscus argus</i>	
<i>Coscinodiscus decrescens</i>	
<i>Coscinodiscus radiatus</i>	
<i>Cyclotella litoralis</i>	
<i>Skeletonema costatum</i>	
<i>Thalassionema nitzschioides</i> var. <i>capitulata</i>	
<i>Thalassiosira angulata</i>	
<i>Thalassiosira conferta</i>	
<i>Thalassiosira oestrupii</i> var. <i>venrickae</i>	
<i>Thalassiosira poro-irregularis</i>	
4) Open-ocean	
<i>Asteromphalus flabellatus</i>	Hasle and Svendsen (1996),
<i>Asteromphalus sarcophagus</i>	Romero et al. (2005), Crosta et
<i>Azpetia neocrenulata</i>	al. (2012)
<i>Azpetia nodulifera</i>	
<i>Azpetia tabularis</i>	
<i>Detonula pumila</i>	
<i>Dytilum brightwellii</i>	
<i>Fragilariaopsis doliolus</i>	
<i>Hemiaulus hauckii</i>	
<i>Hemidiscus membranaceus</i>	
<i>Leptocylindrus mediterraneus</i>	
<i>Neodelphineis denticula</i>	
<i>Nitzschia bicapitata</i>	
<i>Nitzschia capuluspalae</i>	
<i>Nitzschia interruptestrata</i>	
<i>Nitzschia sicula</i>	
<i>Planktoniella sol</i>	
<i>Pseudo-nitzschia inflata</i> var. <i>capitata</i>	
<i>Pseudo-nitzschia pungens</i>	
<i>Pseudo-nitzschia subfraudulenta</i>	
<i>Pseudosolenia calcar-avis</i>	
<i>Pseudotraceratium punctatum</i>	
<i>Rhizosolenia acuminata</i>	
<i>Rhizosolenia bergonii</i>	
<i>Rhizosolenia imbricatae</i>	
<i>Rhizosolenia setigera</i>	
<i>Roperia tessellata</i>	
<i>Stellarima stellaris</i>	
<i>Thalassionema bacillare</i>	
<i>Thalassionema frauenfeldii</i>	
<i>Thalassionema nitzschioides</i> var. <i>capitulata</i>	
<i>Thalassionema nitzschioides</i> var. <i>inflata</i>	
<i>Thalassionema nitzschioides</i> var. <i>parva</i>	
<i>Thalassiosira eccentrica</i>	
<i>Thalassiosira endoseriata</i>	
<i>Thalassiosira ferelineata</i>	
<i>Thalassiosira lineata</i>	
<i>Thalassiosira nanolineata</i>	
<i>Thalassiosira oestrupii</i> var. <i>oestrupii</i>	
<i>Thalassiosira sackettii</i> var. <i>sacketii</i>	
<i>Thalassiosira sackettii</i> var. <i>plana</i>	
<i>Thalassiosira subtilis</i>	
<i>Thalassiosira symmetrica</i>	
Coccolithophores	
1) Cosmopolitan	Boeckel and Baumann (2008),
<i>Emiliana huxleyi</i>	Baumann and Boeckel (2013),
<i>Gephyrocapsa oceanica</i>	Poulton et al. (2017), Young et
2) Lower photic zone	al. (2019)
<i>Algirosphaera robusta</i>	
<i>Calciosolenia murrayi</i>	
<i>Florisphaera profunda</i>	
<i>Gladiolithus flabellatus</i>	
<i>Hayaster perplexus</i>	
3) Warm oligotrophic surface waters	
<i>Discosphaera tubifera</i>	
<i>Helicosphaera carteri</i>	
<i>Rhabdosphaera xiphos</i>	
<i>Umbellosphaera irregularis</i>	
<i>Umbellosphaera tenuis</i>	
<i>Umbilicosphaera anulus</i>	
<i>Umbilicosphaera sibogae</i>	

4) Other miscellaneous species	
	<i>Acanthoica quattropsina</i>
	<i>Calcidiscus leptoporus</i>
	<i>Calcidiscus leptoporus</i> small
	<i>Calcidiscus quadriperforatus</i>
	<i>Gephyrocapsa ericsonii</i>
	<i>Gephyrocapsa muelleriae</i>
	<i>Ophiaster hydroideus</i>
	<i>Ophiaster hydroideus</i>
	<i>Rhabdosphaera stylifer</i>
	<i>Syracosphaera anthos</i>
	<i>Syracosphaera pulchra</i>
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Calcareous dinoflagellates cysts	
1) Upwelling	Siggelkow et al. (2002); Richter et al. (2007); Kohn and Zonneveld (2010)
	<i>Calciadinellum onemusum</i>
	<i>Scrinosiella trochoidea</i>
2) Warm waters	
	<i>Calciadinellum albatmsianum</i>
3) Mineral input	
	<i>Leonella aranifera</i>
4) Cosmopolitan	
	<i>Thoracosphaera heimii</i>
5) Others	
	<i>Calciadinellum levantinum</i>
	<i>Melndemuncula herlinensis</i>
	<i>Pemambucia tuberosa</i>
	<i>Scrinosiella lacrimosa</i>
	<i>Scrinosiella renalis</i>
	<i>Scrinosiella trifida</i>
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Organic dinoflagellates cysts	
1) Upwelling	Susek et al. (2005); Holzwarth et al. (2010); Trainer et al. (2010); Smayda (2010); Smayda and Trainer (2010); Zonneveld et al. (2010; 2013)
	<i>Echinidinium aculeatum</i>
	<i>Echinidinium arnulatum</i>
	<i>Echinidinium transparantum</i>
	<i>Echinidinium zonneveldiae</i>
	<i>Echinidinium</i> spp.
	cvst of <i>Protoperidinium americanum</i>
	cvst of <i>Protoperidinium monospinum</i>
	<i>Stelladinium stellatum</i>
2) Upwelling relaxation	
	<i>Lincolodinium machaerophorum</i>
	cvst of <i>Polvkrikos schwartzii</i>
	cvst of <i>Polvkrikos kofoidii</i>
3) Potential toxic	
	cvsts of <i>Gymnodinium</i> spp.
	cvsts of <i>Alexandrium</i> spp.
4) Cosmopolitan	
	<i>Braiantedinium</i> spp.
	<i>Spiniferites</i> species
	<i>Immagidinium</i> species
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Planktonic Foraminifera	
1) Warmer waters	Hemleben et al. (1989), Schiebel and Hemleben (2017)
	<i>Globaerinoidea ruber</i> (pink and white)
	<i>Globaerinoidea sacculifer</i>
2) Cooler waters	
	<i>Globorotalia inflata</i>
	<i>Neogloboquadrina incompta</i>
3) Upwelling	
	<i>Globaerina bulloides</i>
4) Additional secondary species	
	<i>Beella digitata</i> (Brady 1879)
	<i>Globigerinella calida</i> (Parker 1962)
	<i>Globaerinaella siphonifera</i> (d'Orbigny 1839)
	<i>Globorotalia crassaformis</i> (Galloway and Wissler 1927)
	<i>Globorotalia menardii</i> (Parker, Jones and Brady 1865)
	<i>Globorotalia scitula</i> (Brady 1882)
	<i>Globorotalia truncatulinoides</i> (d'Orbigny 1839)
	<i>Neogloboquadrina dutertrei</i> (d'Orbigny 1839)
	<i>Orbulina universa</i> (d'Orbigny 1839)
	<i>Pulleniatina obliquiloculata</i> (Parker and Jones 1865)
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Pteropods	
	<i>Heliconoides inflatus</i> (d'Orbigny 1835, formerly known as <i>Limacina inflata</i>)
	<i>Limacina bulimoides</i> (d'Orbigny 1835)

Table 5: Species composition of the assemblage of (a) diatoms, (b) coccolithophores, (c)

calcareous and (d) organic dinoflagellate cysts, (e) planktonic foraminifera and (f) pteropods at site CBeu between June 2003 and March 2008.