



August 12th, 2019

Dr. Anja Rammig, Associate Editor
Biogeosciences
The Technical University of Munich
Hans-Carl-v.-Carlowitz-Platz 2
85354 Freising, Germany

Dear Dr. Rammig:

Thank you for considering our manuscript, “Anoxic conditions maintained high phosphorus sorption in humid tropical forest soils” for publication in *Biogeosciences*. We appreciate the comments provided by you and the two reviewers.

We have addressed all the concerns raised by the reviewers. Most importantly, we provided justification for not including blank or low P concentrations in the sorption isotherm experiment. We have checked and improved consistency in the terminology and discussion. We also discussed how P concentration might affect our sorption isotherm results and compared our results to past studies. We also added additional information about our methods as requested by the reviewers and improved the clarity and flow of the text.

We hope you will find our revised manuscript suitable for publication in *Biogeosciences*. Please feel free to contact us if you have any questions.

Sincerely,

A handwritten signature in black ink, appearing to read "Yang Lin".

Yang Lin
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Reviewer #1 (Dr. Eric Roy)

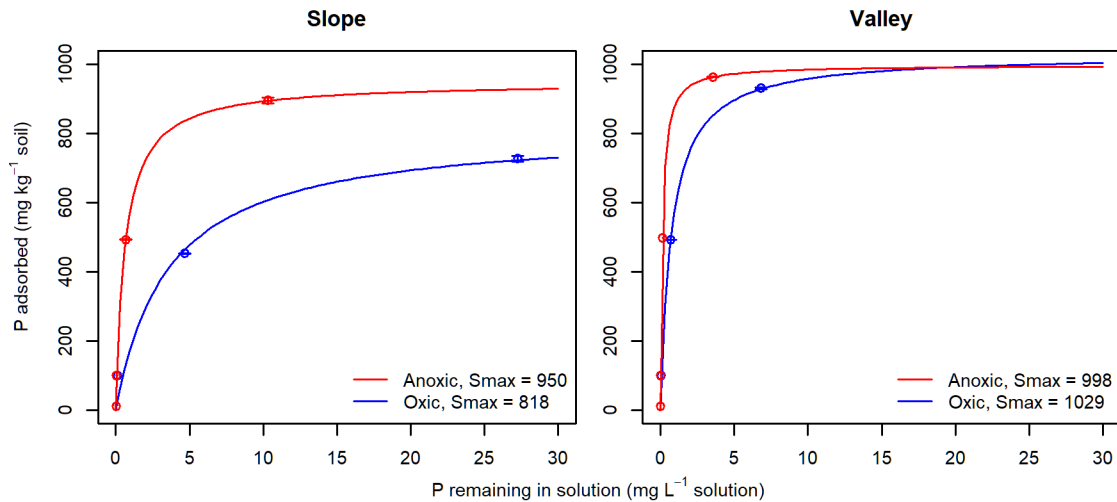
General comments: This manuscript by Lin et al. examines the influence of anoxic conditions on phosphorus sorption in highly weathered, acidic soils in humid tropical forests. It is well known that phosphorus solubility and bioavailability in soils and sediments can be impacted by fluctuations in redox conditions that lead to iron reduction and pH changes. However, to my knowledge (and based on the literature reviewed by Lin et al.), this topic has not been well studied in tropical forest soils (or tropical soils used for agriculture) that are rich in Fe- and Al-minerals and characterized by large P sorption capacity. Therefore, this is a welcome study and the authors have done a nice job generating results that increase understanding of this topic. They perform their experiments using humid tropical forest soils (0-15 cm depth) from two sites in Puerto Rico featuring different parent materials and soil characteristics. Tests include P sorption isotherms, P sorption time curves, soil Fe and Al analyses and P fractions, and a P solubility experiment. The methods used are appropriate for the study questions. I support the publication of this manuscript in Biogeosciences once my comments below have been taken into account.

We appreciate Dr. Roy's interest in our work.

Specific comments: 1) Re: the choice of P concentrations for the sorption isotherm experiments: a. Can the authors provide more justification for their decision to omit the lower concentrations (i.e., 10 and 100 mg P kg⁻¹ soil)? Typically, sorption isotherm experiments have included a 0 mg P kg⁻¹ soil treatment, as well as lower concentrations where most P is sorbed (e.g., Graetz and Nair 2000). Would including these results influence the Langmuir models in this study?

We did not include a P-blank treatment because there was no detectable water-extractable P (or anion exchange resin P, which is higher in concentration) in these soils (McGroddy and Silver 2000 Biotropica). Please see the next section for our response to the comment that our P sorption experiments did not include low P concentration.

b. The range of concentrations used (500 – 10,000 mg P kg⁻¹ soil) is very high. It should be noted that estimates for S_{max} can be dependent on the concentrations used in the P sorption isotherm experiment, particularly when testing a material with high P sorption capacity. For example, Drizo et al. (2002) found that the S_{max} for a steel slag material tested for use in constructed wetlands varied from 0.31 to 3.93 g P kg⁻¹ material depending on the range of P concentrations used. Here, the authors report maximum P sorption capacities ranging from 2526 ± 667 mg P kg⁻¹ to 8256 ± 2517 mg P kg⁻¹ soil. These are high compared to other studies of P-fixing soils that used lower, more conventional ranges in P concentrations (e.g., Brazilian forest soils tested in Roy et al. 2017 using the range of 0 - 1,500 mg P kg⁻¹ soil exhibited S_{max} up to 1167 mg P kg⁻¹ soil). I think the authors should provide some discussion of whether their S_{max} results are method-dependent, and how their results compare to other studies, including those that used more conventional, lower ranges of P concentrations. This discussion can add nuance to their conclusion that S_{max} was “at least one of magnitude above total soil P concentrations.”



As mentioned in the manuscript, our preliminary trials included low P concentrations (10, 100, 500, and 1000 mg P kg⁻¹). As shown in the figure above, the two Icacos soils showed P sorption capacity (S_{max}) extremely close to or higher than the maximum P addition (1000 mg P kg⁻¹). Furthermore, average P concentrations in solution were 0.01 and 0.04 mg P L⁻¹ in samples receiving 10 and 50 mg P kg⁻¹, respectively. Many samples showed no detectable P in solution. Thus, we needed to use higher P concentrations to discern actual sorption potentials. We included the above justification in the methods (Page 4 Line 15-20) and added the figure to the supplement (Figure S1).

We agree with Dr. Roy that concentrations of P would influence the estimation of sorption capacity. We have added the following text in the discussion: “Our estimates of S_{max} were relatively high relative to other humid tropical soils. For example, de Campos et al. (2016) applied up to 8000 mg P kg⁻¹ to a set of strongly weathered Brazilian forest soils and reported a wide range of S_{max} (61-5460 mg P kg⁻¹). Compared to our preliminary trials with maximum P addition of 1000 mg P kg⁻¹ (Fig. S1), S_{max} were higher when more P was added (5000 and 1,0000 mg P kg⁻¹; Table 2), suggesting that S_{max} can be influenced by the concentrations of P used in the experiment. However, even when the maximum P addition level was similar (up to 1500 mg P kg⁻¹ soil), S_{max} from the preliminary trials were also high compared to other strongly weathered soils, including Brazilian forest soils (295-1167 mg P kg⁻¹, Roy et al., 2017) and Thai upland soils (47-1250 mg P kg⁻¹, Wisawapipat et al., 2009).”

2) The authors state that “sorption isotherms of all soils followed Langmuir functions”, however, no model fit diagnostics appear in the manuscript. Can the authors please provide quantitative evidence to justify this statement? The standard errors for S_{max} presented seem somewhat high to me and it would be helpful to know how well the Langmuir model fit the data. Model efficiency is one option (Bolster and Hornberger 2007).

We adopted the Bolster and Hornberger’s method as recommended by the reviewer and updated the estimates of S_{max} and standard errors. Model efficiency ranged between 0.781 to 0.967, indicative of good model fit (Page 7 Line 23-24). Results of model efficiency were included in the supplement (Table S1).

3) Page 6, Line 20 – Can the authors please explain here how they determined that vivianite precipitation was occurring? There is some discussion on subsequent pages that I suggest be moved to the first mention of vivianite precipitation. Furthermore, the calculations made using Visual MINTEQ should be described somewhere in the Methods.

We suspected that vivianite precipitation would contribute to P sorption in anoxic treatments, as the concentrations of P and Fe(II) were high in these samples. We moved some of the discussion from later pages to the results where we first mentioned vivianite precipitation. We also added information on the Visual MINTEQ modeling to the methods (Page 6 Line 13-22).

4) Page 8, Line 10 – The evidence that soils experienced reducing conditions mentioned here should probably come earlier in the paper given its importance for all results presented.

We agreed with this comment and reorganized the results to emphasize the effects of redox treatments (Page 7, Lines 5-21).

References Bolster, C. H., & Hornberger, G. M. (2007). On the use of linearized Langmuir equations. *Soil Science Society of America Journal*, 71(6), 1796-1806. Drizo, A., Comeau, Y., Forget, C., & Chapuis, R. P. (2002). Phosphorus saturation potential: a parameter for estimating the longevity of constructed wetland systems. *Environmental science & technology*, 36(21), 4642-4648. Graetz, D. A., & Nair, V. D. (2000). Phosphorus sorption isotherm determination. In *Methods of phosphorus analysis for soils, sediments, residuals, and waters* (Vol. 369, pp. 35-38). North Carolina State University Raleigh. Roy, E. D., Willig, E., Richards, P. D., Martinelli, L. A., Vazquez, F. F., Pegorini, L., ... & Porder, S. (2017). Soil phosphorus sorption capacity after three decades of intensive fertilization in Mato Grosso, Brazil. *Agriculture, ecosystems & environment*, 249, 206- 214.

Reviewer #2:

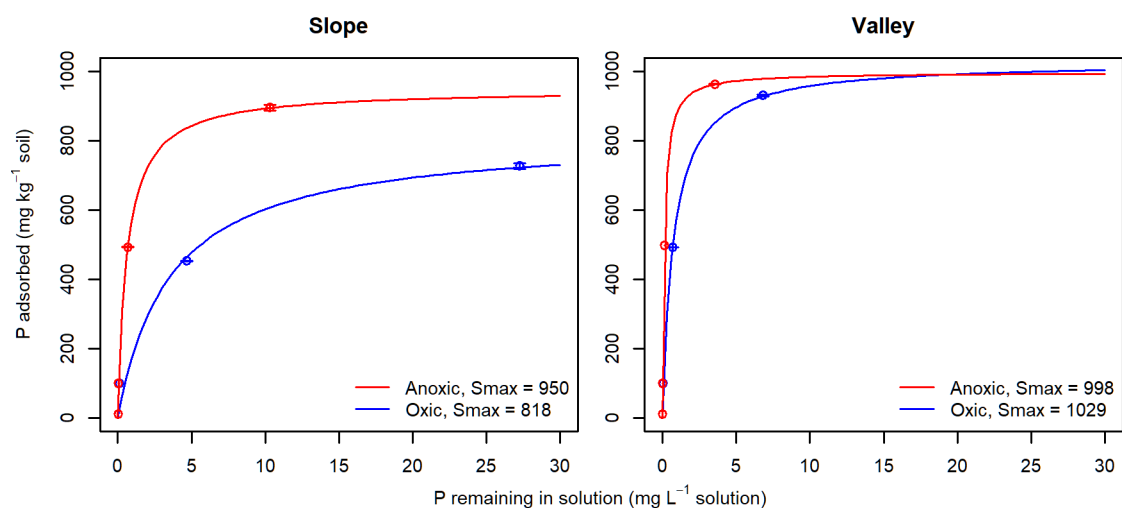
General Comments The manuscript "Anoxic conditions maintained high phosphorus sorption in humid tropical forest soils" describes an experiment using tropical soils from the Luquillo experimental station to assess the effect of anoxic conditions on the adsorption capacity, kinetics and P sorption strength in those soils. Two sets of soils, two topographical units (slope and valley) and two oxic conditions (anoxic and oxic) were used. The results are of great importance for the biogeochemistry community because: (1) few studies on the influence of anoxic conditions to the P sorption capacity/strength in tropical soils are published, (2) because it contradicts the established assumption that reducing conditions increase P availability. While I have no specific grammar corrections, the fluidity of the text should be revisited. Moreover, inconsistencies in the methods, results and discussion sections of the manuscript are visible. I would recommend the publication of the study, after the following major revisions.

We appreciate reviewer's comments. We improved the fluidity of the text during the revision. We also fixed the inconsistencies identified later by the reviewer, including the lack of background soil chemistry data (Table 1) and the inappropriate use of the term 'rate'. We made sure that the rate of P sorption was only used when discussing coefficient k of the P sorption curve (e.g., Page 9 Line 14).

Major comments Although the authors have executed a very interesting work, lack of information on the sample prior to analysis and subsequent analysis during the discussion are not well documented. The authors justify their chosen topographical units as inherent more oxic and less oxic sites yet, no information on the P content, FeOA, AIOA, FeHCl and AlHCl prior incubation is available in Table 1, while the reader is referred to several citations, the addition of this data to Table 1 would be very beneficial. Moreover, while the authors aim to compare the effect on anoxic conditions on the soil capacity to adsorb P no information on the soil specific surface area (SSA) before and after the experiment is available. Moreover, I would recommend the use of SSA for isotherm to analyze the P loading on the minerals (g P/m²) while adsorption occurred.

We added background soil information to Table 1, including concentrations of HCl-extractable Fe(II) and Fe(III) and AO-extractable Fe and Al. We agree with the reviewer that SSA or P loading data would be interesting, but were beyond the scope of this experiment. Given the high surface area of AO-extractable minerals, we consider their concentrations as a surrogate for SSA.

We also question the use of just 4 points in each isotherm, without lower values or zero, the lack of fit statistics (r², among others) and general lack of detail. Later the authors also rely in the lower values of PSI to analyze and compare their sorption curves, while they also refer to the PSI concentration as rate in page 6 line 19. The authors also mention several times the precipitation of vivianine, while no evidence more than the mention of a MINTEQ simulation to the reader. Although the information in in the supplementary section the reader is never referred to it. The details of this MINTEQ simulation are also omitted in the Methods section.



For the justification of no zero or low values, please see the response and additions mentioned above for Reviewer 1. We also included model efficiency as a measure of model fit (Table S1, Page 7 Lines 23-24). We only used four points in the isotherms because 1) our anoxic treatment was limited by the size of the glove box, and we already have a large number of samples (n = 128); 2) including low P concentrations did not significantly affect the shape of isotherm, as our preliminary trials showed that samples receiving 10 and 50 mg P kg⁻¹ had extremely low P concentration in solution (< 0.1 mg P L⁻¹). We also justified our focus on the

PSI values calculated at 1000 mg P kg⁻¹, as this P concentration is comparable to past studies and facilitated comparison with P sorption time curves measured at the same P addition level (Page 6 Lines 29-31).

We avoided the inappropriate terms including ‘PSI rates’ and ‘P addition rates’. We justified the presence of vivianite using literature and also provided information on how MINTEQ simulation was conducted (Page 6 Line 16-22) and introduced the supplementary table in the main text.

At the results, the authors describe the P concentrations added as rates, this is extremely confusing as rates refer to a quantity over time. Which would be the kinetic data. The authors also continue to discuss data from figure 1 (adsorption isotherms) while comparing p-values for the different concentrations of the isotherm while no table or figure is mentioned. The data that this refers is supplementary table 1, where the author refers to the rates as levels, yet in the paragraph no mention to this supplementary table is made.

We no longer refer to P concentrations as rates and abandoned terms such as ‘PSI rates’ and ‘P addition rates’. Rate was only used when discussing coefficient k of the P sorption curve (Page 6 Line 35). We also added a supplementary table (Table S1) to show the P concentrations from the isotherms and indicate statistical significance.

In the discussion, the authors disregarded their kinetic and solubility data and support their discussion on the PSI and its correlation with AIOA and FeOA. The author does not seek to discuss the nuances a faster sorption rate at their simulated anoxic condition in the slope sites. On the other hand, the authors never discuss what minerals/solid phases could be the ones extracted by their HCl(FeII) and HCl(FeII) and how is this related to the higher P sorption and rates in some soils. They base their conclusion on their solubility analysis yet this information is never related to the previous analysis. I would recommend the authors to discussing their results in comparison with the study “Sorption isotherms and kinetics of sediment phosphorus in a tropical reservoir” by Adhityan Appan, and Hong Wang; which is very similar to theirs.

We clarified our discussion of the results mentioned by the reviewer, including the kinetic and solubility data (Page 9 Line 13-30), faster sorption found under anoxic conditions (Page 9 Line 31 to Page 10 Line 12), and the phases of HCl-extractable Fe (Page 9 Line 33-35). We also discussed Adhityan and Appan 2000 in the text (Page 10 Line 24).

Minor revisions

Page 12 line 9: change The to their

The text was revised accordingly.

Page 12 line 10 change “due to the” to “do to the tropical soils”

We changed the text to ‘Due to the high P sorption capacity of tropical soils’.

Anoxic conditions maintained high phosphorus sorption in humid tropical forest soils

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Abstract. The strong phosphorus (P) sorption capacity of iron (Fe) and ~~aluminum~~aluminium (Al) minerals in highly weathered, acidic soils of humid tropical forests is generally assumed to be an important driver of P limitation to plants and microbial activity in these ecosystems. Humid tropical forest soils often experience fluctuating redox conditions that reduce Fe and raise pH. It is commonly thought that Fe reduction generally decreases the capacity and strength of P sorption. Here we examined the effects of 14-day oxic and anoxic incubations on soil P sorption dynamics in humid tropical forest soils from Puerto Rico. Contrary to the conventional belief, soil P sorption capacity did not decrease under anoxic conditions, suggesting that soil minerals remain strong P sinks even under reducing conditions. Sorption of P occurred very rapidly in these soils, with at least 60% of the added P disappearing from the solution within six hours. Estimated P sorption capacities were ~~one~~~~much higher, often by an~~ order of magnitude ~~higher~~, than the soil total P contents. However, the strength of P sorption under reducing conditions was weaker, as indicated by the increased solubility of sorbed P in NaHCO₃ solution. Our results show that highly weathered soil minerals can retain P even under anoxic conditions, where it might otherwise be susceptible to leaching. Anoxic events can also potentially increase P bioavailability by decreasing the strength, rather than the capacity, of P sorption. These results improve our understanding of the redox effects on biogeochemical cycling in tropical forests.

Keywords: Luquillo CZO and LTER, phosphorus availability, nutrient limitation, sorption kinetics, sorption isotherms, sorption rate, anaerobic, aerobic, redox oscillation

1 Introduction

Phosphorus (P) is often thought to limit net primary productivity and organic matter decomposition in humid tropical forests that grow on strongly weathered soils (Vitousek and Sanford Jr., 1986; Cleveland et al., 2011; Camenzind et al., 2017). In these soils, geochemical reactions of adsorption and precipitation, also known as sorption, directly compete with plant roots and microorganisms for phosphate (Thompson and Goyne, 2012). Sorption reactions can immobilize a large ~~of~~ amount of P that can exceed the actual size of the labile soil P pools (de Campos et al., 2016; Roy et al., 2017; Gross et al., 2018) at the scales of seconds to hours (Olander and Vitousek, 2004; McGechan and Lewis, 2002). The sorbed P is generally not readily available for plant and microbial uptake

(Tiessen and Moir, 1993). Thus, mineral sorption plays a key role in constraining the biological availability of P in these ecosystems (Johnson et al., 2003; Reed and Wood, 2016).

How do plants and microbes acquire P in tropical forest soils with high P sorption potential? One of the commonly hypothesized, yet rarely tested, mechanisms is that reducing events can decrease the effectiveness of soil minerals in sorbing P (Chacón et al., 2006; Lin et al., 2018). These soils often contain high concentrations of redox-sensitive poorly crystalline or amorphous Fe minerals (Hall and Silver, 2015; Wilmoth et al., 2018; de Campos et al., 2016). Under anoxic conditions, Fe(III) minerals undergo reductive dissolution and are transformed into Fe(II) phases that are assumed to be less effective at binding P (Quintero et al., 2007; Rakotoson et al., 2014; Shenker et al., 2005).

Reducing conditions increase pH and decrease surface charges that are thought to be central to P sorption (Oh et al., 1999; Willett, 1989). Although Al minerals, which are often enriched in these soils, are not redox-active, redox-induced pH changes can control the speciation of Al minerals and consequently affect P sorption (Haynes, 1982; Gustafsson et al., 2012).

Redox effects on soil P sorption processes have not been well studied in tropical forest soils, even though periodic redox oscillations are well documented in these environments (Barcellos et al., 2018; Schuur et al., 2001; Chen et al., 2018; Keiluweit et al., 2016; Wieder et al., 2011; O'Connell et al., 2018; Silver et al., 1999). Some past studies from tropical ecosystems have reported increases in extractable soil P during anoxic events (Chacón et al., 2006; Peretyazhko and Sposito, 2005; Maranguit et al., 2017), which is consistent with the hypothesis that anoxic conditions weaken P sorption, although the mechanisms remain unclear. To better understand the importance of redox events in regulating P bioavailability in tropical forest soils, it is necessary to examine both the capacity and strength of P sorption. Sorption capacity is usually characterized by the maximum amount of P that soil minerals can sorb and can be measured using the sorption isotherm method (McGechan and Lewis, 2002). The strength of P sorption is related to the potential bioavailability of the sorbed P, as strongly sorbed P is considered to be largely unavailable plant roots and microbes. ~~This~~ can be evaluated by measuring the solubility of sorbed P in various ~~solvents~~extracting solutions such as NaHCO₃ and NaOH ~~solution~~ (Ryden and Syers, 1977).

We determined the effects of redox conditions (oxic vs. anoxic incubations) on P sorption characteristics in soils from humid tropical forests in Puerto Rico. We conducted two types of P sorption experiments: 1) the standard sorption isotherm to evaluate the capacity of P sorption; and 2) the P sorption time curve to evaluate the rate of P sorption over time. These P sorption experiments were conducted using soils from two distinct parent materials with different Fe, Al, and P concentrations and two topographic positions (frequently reduced valleys and more aerated slopes), allowing us to evaluate the effects of soil redox history on P sorption processes. We also measured the relative solubility of sorbed P in NaHCO₃ and NaOH solution to evaluate the strength of P sorption. We hypothesized that anoxic conditions would decrease soil P sorption capacity due to a decrease in Fe(III) concentrations and associated weakening of the Fe-P bond. We also hypothesized that redox-induced decreases in P

sorption capacity would be accompanied by increases in the solubility of sorbed P, indicative of weaker P sorption strength.

2 Material and Methods

5 2.1 Study sites and soil sampling

Soils were collected from humid tropical forests in the Luquillo Experimental Forest (LEF) in Puerto Rico, part of the NSF sponsored Long Term Ecological Research (LTER) and Critical Zone Observatory (CZO) networks, and the DOE sponsored NGEE Tropics program. The mean annual temperature decreased from about 23 °C at 350 m to about 19 °C at 930 m elevations above sea level (Weaver and Murphy, 1990; Brown et al., 1983). Mean annual rainfall increased from about 3,200 mm at 300 m to about 4,800 mm at 1000 m with no clear seasonal pattern (Murphy et al., 2017). Mineral soils (0-15 cm depth) were collected from two sites, El Verde and Rio Icacos, featuring different parent materials and soil characteristics (Table 1). The El Verde site is located at approximately 380 m elevation within the Tabonuco forest zone. The Rio Icacos site is located at approximately 630 m within the Colorado forest zone, and characterized by more frequent cloud condensation and abundant epiphytes. Soils from El Verde were Hapludoxes and derived from volcanoclastic material (Scatena, 1989). Soils from Rio Icacos were Dystrudepts and developed from quartz diorite material, which contained approximately half as much total P compared to volcanoclastic material and lower concentrations of Fe and Al oxides (Mage and Porder, 2012). As a result, soils from El Verde generally have higher total concentrations of P, Fe, and Al minerals (Mage and Porder, 2012; Coward et al., 2017). Topographic locations (ridge, slope, and valley) play a key role in determining soil redox conditions and biogeochemical processes (Scatena, 1989; Silver et al., 1994; O'Connell et al., 2018; Hall and Silver, 2015). In these highly dissected landscapes, catenas typically shift from well-aerated ridges to valley bottoms that experience frequent reducing events. Slopes cover approximately 65% of the landscape, while ridges and valleys make up the rest in approximately equal proportions (Scatena and Lugo, 1995). For the sorption experiments, soils from both sites were collected from valley and slope positions after removing the surface litter layer. For the solubility experiment, we only analyzed the slope and valley soils from the El Verde site due to logistical limitations in accessing the Rio Icacos site after Hurricane Maria. The valley soil sampled for the solubility experiment was significantly drier than that sampled previously (Table 1). Soils were shipped overnight to the University of California, Berkeley, at an ambient temperature and were immediately gently homogenized by hands upon arrival and visible plant debris, rocks, and macro-fauna removed.

30

2.2 Redox treatments

To examine the effects of low redox conditions on soil P sorption processes, soil samples were pre-incubated under anoxic or oxic conditions for 14 days. Soils were subsampled in quart-size, glass jars with gas-tight lids (~ 100 g each, oven-dry equivalent (ODE)). For those in the anoxic treatment, the jar headspace was evacuated and flushed

with N₂ three times before being transferred to an anaerobic glove box (90% N₂, 8% CO₂, and 2% H₂; Coy Laboratory Products, Grass Lake, MI). Jars were sealed inside the glove box and vented every two or three days. Jars in the oxic treatment were sealed under an ambient atmosphere and vented following the same schedule as those from the anoxic treatment. Jars were stored in cardboard boxes in the dark during the incubation. Soils from El Verde and Rio Icacos were analyzed in two separate campaigns following the same experimental approach due to space limitations in the glovebox.

2.3 Phosphorus sorption experiments

After pre-incubation, soil P sorption was evaluated in two ways. We used P sorption isotherm experiments to determine sorption across a range of standard P loadings over 24 hours. We also conducted P sorption time curves that characterize the disappearance of solution P with one level P of addition at multiple time points over 48 hours (Henry and Smith, 2006). For P sorption isotherms, aliquots of 15 g ODE soil were subsampled into pint-size glass jars containing 150 ml of 0.01 M CaCl₂ to reach a soil to solution ratio of 1:10. The CaCl₂ solution had been previously spiked with 100 mg P L⁻¹ KH₂PO₄ stock solution to reach four levels of P concentrations (500, 1000, 5000, and 10,000 mg P kg⁻¹ soil). Preliminary trials showed that P addition at lower concentrations (i.e., 10 and 100 mg P kg⁻¹ soil) resulted in near complete sorption within 24 hours. (Fig. S1), and many samples had no nondetectable P in solution. The estimated maximum sorption capacity was extremely close to the highest P addition level (i.e., 1000 mg P kg⁻¹ soil) used in traditional sorption studies (Roy et al., 2017; Wisawapipat et al., 2009; Gustafsson et al., 2012; Borggaard et al., 1990). Thus, here we used higher levels of P concentrations including 5000 and 10,000 mg P kg⁻¹ soil. Similar levels of P addition have been previously used (de Campos et al., 2016; Zhang et al., 2003). There was a total of 128 samples (4 replicates × 4 levels of P addition × 2 redox treatments × 2 topographic locations × 2 sample sites). Soil slurry was then amended with 1 mL of toluene to inhibit microbial activity before being manually shaken for one minute. Samples from the anoxic treatment were prepared in the anoxic glove box with degassed solutions. The slurry was manually shaken periodically to mix soils with solution. After 24 hours, 5 mL subsamples of soil slurry were extracted from each jar and filtered through 0.45 μm syringe filters into test tubes and then acidified with HCl to a final H⁺ concentration of 0.1 N to prevent the oxidation and precipitation of dissolved Fe(II).

For P sorption time curves, batch experiments were started by subsampling aliquots of 30 g soil (ODE) into quart-size glass jars with 300 ml of 0.01 M CaCl₂ solution with a single pulse of 1000 mg P kg⁻¹ soil and 2 mL of toluene. A total of 32 samples (4 replicates × 2 redox treatments × 2 topographic locations × 2 sample sites) were prepared. Subsamples of suspended soil were taken after 5 minutes, 40 minutes, 2 hours, 6 hours, 12 hours, 24 hours and 48 hours following the previously described method. Solution P concentrations from the sorption experiments were determined colorimetrically following Murphy and Riley (1962).Murphy and Riley (1962).

2.4 Soil Fe and Al analyses and P fractions

Redox-active Fe pools were measured after the pre-incubation. Acid-soluble Fe was extracted by mixing 4 g ODE soil in 40 ml of 0.5 M hydrochloric acid (HCl) solution and shaking for 1 h followed by centrifugation. The HCl-extractable Fe(II) and Fe(III) (HCl-Fe(II) and HCl-Fe(III)) concentrations were determined using a modified ferrozine assay (Viollier et al. 2000). Ammonium oxalate (AO) solution was used to estimate the concentrations of poorly crystalline Fe and Al minerals. A pre-treatment with 0.1 M HCl was used to remove Fe(II) before the AO extraction to avoid catalytic dissolution of crystalline Fe minerals in the presence of Fe(II) and oxalate at reducing conditions (Heiberg et al. 2012). Aliquots of 1 g ODE soil were mixed with 10 ml of 0.1 M HCl for 10 minutes on an end-to-end shaker, followed by centrifugation. Samples were then washed with 40 ml of H₂O, centrifuged, and subsequently used for AO extraction (40 mL) at pH 3.0 in the dark for 2 hours. The AO extract was then filtered through 0.45 µm syringe filter before determination of Fe and Al concentrations (AO-Fe and AO-Al) with inductively coupled plasma optical emission spectrometry (ICP-OES) on three analytical replicates per sample (Perkin Elmer, Optima 5300 DV series, CA, USA).

Soil P fractions were estimated using a modified Hedley scheme after the pre-incubation and before the sorption test (Hedley et al., 1982). Aliquots of 1 g ODE soil were sequentially extracted using 40 mL of 0.5 M sodium bicarbonate (NaHCO₃) and 0.1 M sodium hydroxide (NaOH) solution, each for 16 hours. The NaHCO₃ solution extracts inorganic and organic P that are weakly associated to soil particles and commonly assumed to be biologically available (Tiessen and Moir, 1993). The NaOH solution mobilizes P compounds that are more strongly bound to Fe and Al minerals than those extracted by NaHCO₃ solution, thus having intermediate availability (Tiessen and Moir, 1993). Total P concentration of both solutions was determined following ~~Murphy and Riley (1962) after autoclaving the solution with ammonium persulfate ((NH₄)₂S₂O₈)-Murphy and Riley (1962) after autoclaving the solution with ammonium persulfate ((NH₄)₂S₂O₈).~~

2.5 Phosphorus solubility experiments

We assessed the strength of P sorption by measuring the relative solubility of P for the slope and valley at the El Verde site. Overall, there were 96 samples (4 replicates × 2 extractants × 3 levels of P addition × 2 redox condition × 2 soil types). Approximately 10 g ODE soil was weighed into pint-size glass jars and incubated either under ambient air or in an anaerobic glovebox for 14 days, following the method above. Samples then received 100 ml of 0.01 M CaCl₂ solution containing 0, 100, or 1000 mg P kg⁻¹ soil and 1 mL toluene. Jars were capped and shaken for 24 hours after which 5 mL subsamples of soil slurry were taken to estimate the amount of P remaining in solution and P sorbed by minerals using the methods described above. Microcosms then received 300 ml of either 0.667 M NaHCO₃ or 0.133 M NaOH solution to reach the final concentrations of 0.5 M NaHCO₃ and 0.1 M NaOH, respectively. The two extracting solutions were used because they have different P extraction efficiencies. Solutions were shaken every hour for three hours at both the beginning and the end of a 16-hour, overnight period prior to extraction. The anoxic treatment was conducted in the anoxic glove box with degassed solutions. Determination of

NaHCO₃ and NaOH total P concentrations (NaHCO₃-P_t and NaOH-P_t, respectively) followed the previously described methods. The extracted P came from two potential sources: the P amendment (for the two treatments with P added) and native extractable P. We accounted for the native extractable soil P in the treatment without P addition. We also accounted for the amendment P remaining in solution in order to estimate the amount of sorbed P recovered during extraction. The recovered P was then reported as a percentage of the sorbed P, or the relative P solubility.

2.6 Data analyses

Sorption isotherm data were modelled using the Langmuir equation as in Eq. (1):

$$S = \frac{aS_{max}C}{1+aC}, \quad (1)$$

in which S is the amount of P sorbed during the batch experiment (mg P kg⁻¹ soil), S_{max} corresponds to a predicted maximum sorption capacity (mg P kg⁻¹ soil), C is the concentration of P remaining in solution (mg P L⁻¹), and a represents a coefficient related to the bonding strength of P to soil minerals (L mg⁻¹ P). For each combination of soil type and redox treatment, all data points were used to fit the Langmuir equation using following the nls function in R ver. 3.4.4 approach described by (R Core Team, 2018); Bolster and Hornberger (2007). This approach estimated the means and standard errors of all model parameters, including S_{max} . The goodness of fit was evaluated by model efficiency (E), where $E = 1$ indicates a perfect fit to the data.

It is possible that vivianite precipitation contributed to P sorption in the anoxic treatments, especially at high levels of P addition (5000 and 10,000 mg P kg⁻¹) when Fe(II) concentrations were also high (Borch and Fendorf, 2007; Heiberg et al., 2012). The saturation index of vivianite was calculated using Visual MINTEQ, ver. 3.1 (Gustafsson, 2015). Model inputs included solution pH and concentrations of inorganic P and Fe(II) in solution. We assumed that 10% of HCl-Fe(II) was soluble in solution based on past studies using similar soils from Puerto Rico (Peretyazhko and Sposito, 2005; Wilmoth et al., 2018).

We also calculated a P sorption index (PSI, L kg⁻¹ soil) (Bache and Williams, 1971) using the sorption isotherm results under all four P addition levels in Eq. (2):

$$PSI = \frac{S}{\log_{10} C}, \quad (2)$$

in which S is the amount of P sorbed during the batch experiment (mg P kg⁻¹ soil), and C corresponds to the concentration of P remaining in solution (mg P L⁻¹). High PSI values correspond to high P sorption capacity. Our discussion was focused on PSI values calculated at 1000 mg P kg⁻¹ because this rate level of P addition was similar to the one used by Bache and Williams (1971) (1,500 mg P kg⁻¹) and facilitated comparison with P sorption time curves measured at the same P addition rate. At P addition levels of 5000 and 10,000 mg P kg⁻¹, PSI values were also influenced by vivianite precipitation. Thus, we explored the relationships among PSI at 1000 mg P kg⁻¹, other P sorption indices, and soil Fe and Al concentrations.

Data from P sorption time curves were modelled using a power function as in Eq. (3):

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$$100 - p = at^{-k}, \quad (3)$$

in which p represents the percent of P sorbed (%), t indicates time (h), k corresponds to the rate of P sorption, and a is a coefficient (h^{-1}). Higher values of k indicate faster P sorption. Model fitting was also conducted using the nls function in R, [ver. 3.4.4 \(R Core Team, 2019\)](#). Effects of redox manipulation on PSI values and P solubility were compared using student's T-tests in each soil at $\alpha = 0.05$ level. All analyses were conducted in R.

3 Results

3.1 Redox effects on Fe, Al, and P sorption

In all four types of soils, the anoxic treatment had significantly higher HCl-Fe(II) concentrations relative to the oxic treatment (up to two orders of magnitude greater), providing evidence that soils experienced reducing conditions (Table 2). This effect was particularly strong in the slope soil at El Verde and at Rio Icacos (both $P < 0.001$). Anoxic conditions decreased soil HCl-Fe(III) concentrations relative to oxic conditions in the slope ($P < 0.01$) and valley ($P < 0.001$) soil at El Verde. However, soil HCl-Fe(III) concentrations did not respond significantly to redox treatments in soils from Rio Icacos. Soil HCl-Fe(III) concentrations were significantly higher at El Verde than at Rio Icacos ($P < 0.001$, Table 2).

Anoxic conditions decreased AO-Fe concentrations in the valley soils from Rio Icacos ($P < 0.001$), and increased concentrations of AO-Al in the valley soil at El Verde ($P < 0.05$) (Table 2). However, effects of redox on AO-Fe and AO-Al concentrations were relatively small compared to their differences across sampling sites and topographic positions. Soils at El Verde had higher AO-Fe concentrations ($P < 0.001$) and lower AO-Al concentrations ($P < 0.001$) than those at Rio Icacos. Within the El Verde samples, the valley soil had higher AO-Fe concentrations ($P < 0.01$) and lower AO-Al concentrations ($P < 0.001$) than the slope soil. Within the Rio Icacos site, the valley soil had nearly doubled the AO-Al concentrations compared to the slope soil ($P < 0.001$), while the two topographic zones had similar levels of AO-Fe.

Sorption isotherms of all soils generally followed the Langmuir functions (Fig. 1), as E varied between 0.781 and 0.967 with an average of 0.898 (Table S1). Estimated maximum sorption capacities ranged from $2526 \pm 6672627 \pm 303$ mg P kg^{-1} to 8256 ± 25172519 mg P kg^{-1} (Table 2, mean \pm S.E. unless otherwise noted), which was at least one order of magnitude higher than total soil P concentrations (140–400 mg P kg^{-1} ; Mage and Porder 2012). Effects of redox treatments on P remaining in solution and the P sorbed differed among levels of P addition. Under high P additions (5000–10,000 mg P kg^{-1}), P sorption (vertical axis Fig. 1) was generally greater under the anoxic treatment with lower concentrations of P remaining in solution relative to the oxic treatment (horizontal axis Fig. 1). Phosphorus sorption rates were high at low levels of addition (500 and 1000 mg P kg^{-1} , Fig. 1); Table S1). When 1000 mg P kg^{-1} was added, there was significantly lower P remaining in solution under anoxic conditions at both slope soils ($P < 0.01$ at El Verde and $P < 0.001$ at Rio Icacos) and the valley soils at Rio Icacos ($P < 0.01$). Only the

valley soils at El Verde showed significantly more P remaining in solution under anoxic conditions ($P < 0.01$). Similar trends were also observed at the lowest level of P addition (500 mg P kg^{-1}): there was significantly less P remaining in solution for the two slope soils under anoxic conditions ($P < 0.01$ at El Verde and $P < 0.001$ at Rio Icacos), and the opposite was true for the valley soils at El Verde ($P < 0.01$). ~~However, the~~The valley soils at Rio Icacos showed no significant differences in solution P remaining between the two redox treatments: at the P addition level of 500 mg P kg^{-1} .

The PSI values offered another way to examine the redox treatments on P sorption. Under high P additions ($5000\text{--}10,000 \text{ mg P kg}^{-1}$), anoxic conditions led to greater or similar PSI values as oxic conditions (Table S4S2). When 500 mg P kg^{-1} was added, PSI values became negative in two valley soils under oxic conditions because their average P concentrations remaining in solution were lower than 1 mg L^{-1} , resulting in negative logarithms. Under P addition of $1000 \text{ mg P kg}^{-1}$, anoxic conditions led to lower PSI values than oxic conditions in the El Verde valley soil ($P < 0.05$, Fig. 2), while redox incubations did not significantly affect PSI in the valley soil from Rio Icacos. In contrast, anoxic conditions increased PSI values relative to the oxic treatment in the two slope soils (both $P < 0.01$). The valley soils had $116 \pm 25\%$ and $64 \pm 14\%$ higher PSI values than the slope soils at El Verde and Rio Icacos, respectively (both $P < 0.05$). Averaging between topographic positions, differences in PSI between the two sites were relatively small (El Verde vs. Rio Icacos: $720 \pm 89 \text{ L kg}^{-1}$ soil vs. $890 \pm 7 \text{ L kg}^{-1}$ soil, $P < 0.05$) compared to the effects of redox conditions and topographic positions. For the remainder of the text, we will refer to PSI values as those calculated under P addition of $1000 \text{ mg P kg}^{-1}$ unless noted otherwise.

Rapid P sorption was observed in all soils, as at least 60% of the added P had disappeared from solution within the first six hours of sorption experiment (Fig. 3). In the two slope soils, P sorption occurred more rapidly after anoxic incubation than after oxic incubation, as indicated by the lower concentrations of P remaining in solution under anoxic conditions (all $P < 0.05$ after 12 hours). In valley soils from El Verde, however, P sorption occurred more rapidly after oxic incubation, as $2.6 \pm 0.4\%$ vs. $30.3 \pm 3.3\%$ of the added P remained in solution after the first six hours after oxic and anoxic incubation, respectively ($P < 0.001$). In valley soils from Rio Icacos, more P was sorbed in soils from the oxic treatment than those from anoxic treatment during the first six hours (all $P < 0.05$), while afterwards no effects of redox treatment were found. The rate of P sorption (k) was strongly correlated with PSI values ($r = 0.86$, $P < 0.01$, Table 2).

3.2 Redox effects on Relationships of P sorption to other soil characteristics

~~We measured soil Fe and Al concentrations to determine relationships with P sorption indices. Soil HCl-Fe(III) concentrations were significantly higher at El Verde than at Rio Icacos ($P < 0.001$, Table 2). Anoxic conditions decreased soil HCl-Fe(III) concentrations relative to oxic conditions in the slope ($P < 0.01$) and valley ($P < 0.001$) soil at El Verde. However, soil HCl-Fe(III) concentrations did not respond significantly to redox treatments in two soils from Rio Icacos. In all four types of soils, the anoxic treatment had significantly higher soil HCl-Fe(II)~~

concentrations relative to the oxic treatment (up to two orders of magnitude greater), providing evidence that soils experienced reducing conditions (Table 2). This effect was particularly strong in the slope soil at El Verde and at Rio Icaicos (both $P < 0.001$). We explored the relationships between soil Fe and Al concentrations and P sorption indices. Among all samples, soil HCl-Fe(III) concentrations were weakly positively correlated with PSI values ($r = 0.43$, $P < 0.05$; Table 2), while HCl-Fe(II) values were not. The correlation between HCl-Fe(III) and PSI was strongest at El Verde (Fig. S4S2; $r = 0.89$, $P < 0.001$).

Soils at El Verde had higher AO-Fe concentrations ($P < 0.001$, Table 2) and lower AO-Al concentrations ($P < 0.001$) than those at Rio Icaicos (Table 2). Within the El Verde samples, the valley soil had higher AO-Fe concentrations ($P < 0.01$) and lower AO-Al concentrations ($P < 0.001$) than the slope soil. Within the Rio Icaicos site, the valley soil had nearly doubled the AO-Al concentrations compared to the slope soil ($P < 0.001$), while the two topographic zones had similar levels of AO-Fe. Anoxic conditions decreased AO-Fe concentrations in the valley soils from Rio Icaicos ($P < 0.001$), and increased concentrations of AO-Al in the valley soil at El Verde ($P < 0.05$) (Table 2). Among all samples, concentrations of AO-Al were weakly positively correlated with PSI values ($r = 0.39$, $P < 0.05$), but AO-Fe were not. A positive correlation between AO-Al concentrations and PSI was found at Rio Icaicos (Fig. S4S2; $r = 0.74$, $P = 0.002$), while their correlation was negative at El Verde ($r = -0.71$, $P = 0.002$).

3.3 Phosphorus solubility

We explored the extractability/solubility of P following P additions at the El Verde site. The average relative solubility of P was much lower in NaHCO₃ solution than in NaOH solution ($20.5 \pm 1.3\%$ vs. $69.7 \pm 2.1\%$; $P < 0.001$, Fig. 4). Phosphorus additions of 100 mg P kg⁻¹ yielded higher relative P solubility than 1000 mg P kg⁻¹ additions, in both NaHCO₃ ($P < 0.001$, Fig. 4) and NaOH solutions ($P < 0.001$), as extractants might have reached their limits of dissolving P under higher P addition level. Anoxic conditions generally increased P solubility in NaHCO₃ solution, except in the slope soil with P addition of 100 mg P kg⁻¹. Phosphorus solubility in NaOH solution decreased in the anoxic treatment in the valley soil with the lower level of P addition only ($P < 0.001$). Note that anoxic conditions increased PSI values in both soils under P additions of 1000 mg P kg⁻¹ (Fig. S3). The average relative solubility of P was much lower in NaHCO₃ solution than in NaOH solution ($20.5 \pm 1.3\%$ vs. $69.7 \pm 2.1\%$; $P < 0.001$ (S2)).

4 Discussion

4.1 Anoxic conditions maintained high P sorption

Contrary to our first hypothesis that anoxic conditions would decrease P sorption, anoxic conditions led to similar or greater rates (k) of P sorption as oxic conditions in all but one treatment (P sorption time curve for El Verde valley). This suggests that soils remain strong P sinks even under reducing conditions. Under both oxic and anoxic

conditions, estimated maximum P sorption capacities were much higher than the range of total soil P concentrations (Mage and Porder, 2012), highlighting the significant potential of these soils for retaining P. Our estimates of S_{max} were relatively high relative to other humid tropical soils. For example, de Campos et al. (2016) applied up to 8000 mg P kg⁻¹ to a set of strongly weathered Brazilian forest soils and reported a wide range of S_{max} (61-5460 mg P kg⁻¹).

5 Compared to our preliminary trials with maximum P addition of 1000 mg P kg⁻¹ (Fig. S1), S_{max} were higher when more P was added (5000 and 1,0000 mg P kg⁻¹; Table 2), suggesting that S_{max} can be influenced by the concentrations of P used in the experiment. However, even when the maximum P addition level was similar (up to 1500 mg P kg⁻¹ soil), S_{max} from the preliminary trials were also high compared to other strongly weathered soils, including Brazilian forest soils (295-1167 mg P kg⁻¹, Roy et al., 2017) and Thai upland soils (47-1250 mg P kg⁻¹,

10 Wisawapipat et al., 2009). Sorption of P occurred very rapidly under both redox conditions, with over 60% of the added P removed from solution within six hours in all soils. Results indicate that low redox events are unlikely to induce significant P release to the soil solution in these soils. High P sorption potential is very likely responsible for the extremely low P concentrations of stream water in local watersheds (McDowell, 1998; McDowell and Liptzin, 2014). Our results also suggest that new P entering the ecosystem via atmospheric sources such as dust or smoke

15 (Pett-Ridge, 2009) would likely be rapidly sorbed by soil minerals.

Three mechanisms are likely responsible for the high P sorption capacities under anoxic conditions. First, mixed Fe(III)-Fe(II) or Fe(II) minerals formed during Fe reduction may have a high sorption capacity. Past research in this ecosystem has shown that approximately 90% of the HCl-extractable Fe(II) was in the mineral phase (Peretyazhko and Sposito, 2005; Wilmoth et al., 2018; Chen et al., 2018). These minerals can feature a more amorphous structure compared to Fe(III) phases and thus have a higher surface area available for P sorption (Patrick and Khalid, 1974; Holford and Patrick, 1979; Borch and Fendorf, 2007). Soils from the study site have very large and diverse populations of microbial Fe reducers that facilitate rapid Fe reduction (Dubinsky et al., 2010), contributing to the formation and maintenance of high amorphous Fe minerals. Second, Fe reduction is known to increase soil pH

20 (Lindsay, 1979) that consequently increases the degree of hydroxylation and surface area of Al and organo-Al complexes (Haynes, 1982). These changes in Al speciation have commonly been used to explain the increased P sorption capacities under liming (Haynes and Swift, 1989; Gustafsson et al., 2012). Finally, formation of Fe(II)-P minerals, such as vivianite, can also contribute to high P retention (Heiberg et al., 2012; Walpersdorf et al., 2013). In our experiment, the soil slurry appeared to be supersaturated with respect to vivianite precipitation as calculated by Visual-MINTEQ (Gustafsson, 2011), especially under high P loadings (5000 and 10,000 mg P kg⁻¹, Table S2S3).

25 However, the effect of vivianite formation was likely to be small under low P loadings (e.g., 1000 mg P kg⁻¹), as its precipitation kinetics is slow and depends on P concentration (Borch and Fendorf, 2007; Heiberg et al., 2012).

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4.2 Phosphorus sorption and Fe and Al minerals

Our results identified amorphous Fe(III) and Al minerals as the best predictors of soil PSI values, but not Fe(II) minerals. These results suggest that soil Fe(II) minerals alone were insufficient to explain P sorption capacity compared to Fe(III) or Al minerals, consistent with previous studies (Sallade and Sims, 1997; Quintero et al., 2007; Rakotoson et al., 2014). However, soil Fe(III) and Al minerals were less responsive to redox manipulation compared to Fe(II) minerals. Persistence of amorphous minerals, as well as crystalline Fe and Al minerals, are all likely contributors to the high P sorption capacity under both redox conditions (Gérard, 2016; McGechan and Lewis, 2002). We found that anoxic conditions did not affect HCl-Fe(III) concentrations in soils from Rio Icaos, and that changes in HCl-Fe(III) and AO-Fe were smaller in magnitude compared to the increases in HCl-Fe(II) across all soil types. Together, this indicates that Fe reduction mobilized crystalline or poorly crystalline Fe that was not soluble in HCl or AO solution. [Appan and Wang \(2000\) found no effects of redox on P sorption in tropical reservoir sediments with high Al concentrations.](#) Overall, concentrations of soil Fe(III) and Al minerals influenced soil P sorption behavior across soil types, and their persistence contributed to high P sorption under both redox environments.

Patterns in soil Fe and Al concentrations helped to explain the variability of soil P sorption behavior across the two topographic zones. Valley soils, which are characterized by frequent high magnitude redox fluctuations and low redox events (Silver et al., 1999), had higher concentrations of HCl-Fe(III) than slope soils at both sites, and had similar or higher levels of AO-Fe. The valley soil at Rio Icaos had twice as much AO-Al as the slope soil. The higher levels of amorphous Fe and Al minerals in valley soils likely contributed to higher PSI values relative to slopes in both locations. The differences in amorphous Fe and Al concentrations likely resulted from the long-term changes in redox history and soil transport along the catena (Hall and Silver, 2015; Mage and Porder, 2012). Water and organic matter transport leads to higher soil moisture content in valleys than in slope soils and helps to create more frequent and intensive reducing events (Silver et al., 1999). These conditions facilitate the formation and persistence of amorphous Fe and Al minerals.

Differences in Fe and Al minerals also helped to explain the patterns of P sorption across two sites. Soils at El Verde were enriched in HCl-Fe(III) and AO-Fe but depleted in AO-Al compared to soils at Rio Icaos, and the positive correlation between HCl-Fe(III) and PSI was only significant at El Verde. Soil AO-Al concentrations were positively correlated with PSI at Rio Icaos, but had a negative correlation at El Verde. These results suggest that Fe minerals may play a primary role in sorbing P in the volcanoclastic soils, while Al minerals were more important to P sorption in the dioritic soils. Together our results show that redox history and parent material influenced the patterns of soil P sorption across topographic zones and study site, respectively, at long timescales.

In the valley soil from El Verde, the effects of redox manipulation on P sorption differed greatly among levels of P addition, a pattern not observed in the other three sampling sites. High concentrations of Fe(II), derived from the low-redox legacy of the soil, and soluble P under high P loadings (5000 and 10,000 mg P kg⁻¹) represented favorable conditions for vivianite formation; [\(Table S3\)](#), which likely contributed to the higher P sorption capacity under

anoxic conditions in this soil. Vivianite formation did not appear to play a major role in P sorption at a lower P loading (1000 mg P kg⁻¹), as the anoxic treatment decreased the PSI value and P sorption rate (*k*) in this soil. The strong decline in amorphous Fe(III) concentrations as a result of Fe reduction was potentially responsible for the reduced PSI and P sorption rate under anoxic conditions in these soils. The specific effects of different P-loadings under redox manipulation also led to the lack of significant correlation between S_{max} and PSI values, because S_{max} was mostly driven by data from high P loadings, while PSI was calculated at the P loading of 1000 mg P kg⁻¹.

The PSI values of the valley soil at El Verde showed different responses to anoxic incubation in two separate trials. Soil moisture content measured in the P solubility experiment was significantly lower than its mean value calculated from continuous field observations (O'Connell et al., 2018) and lower than that measured in the initial trial, likely due to natural background variability in rainfall. The PSI value and its response to redox manipulation resembled those of the slope soil. It is possible that the dry period decreased the reactive surface areas of soil minerals potentially by oxidizing reduced species and increasing the crystallinity of secondary Fe and Al minerals. These results suggest that soil P dynamics could be highly sensitive to changes in environmental conditions in tropical forests (O'Connell et al., 2018).

4.3 Implications for P solubility and bioavailability

Our results showed that, regardless of redox conditions, significantly more P was soluble in NaOH than NaHCO₃ and that the relative P solubility was high (> 50% of sorbed P) in NaOH for both the 100 and 1000 mg P kg⁻¹ additions. The NaOH extraction is stronger than the NaHCO₃ solution, and together they are thought to represent a continuum of P bound to Fe and Al minerals (Tiessen and Moir 1993). Anoxic conditions generally increased the solubility of the sorbed P in NaHCO₃ solution, indicating that the strength of P sorption was weaker when soils had more anaerobic microsites. The change in P sorption likely reflected the lower binding strengths of reduced Fe minerals to P than their oxidized forms (Zhang et al., 2003; Holford and Patrick, 1979). Past research has reported increased extractable P-solubility in response to Fe reduction using soils from the tropics (Peretyazhko and Sposito, 2005; Liptzin and Silver, 2009; Chacón et al., 2006; Maranguit et al., 2017; Lin et al., 2018). Our results suggest that Fe-redox dynamics increase P-solubility via decreasing the strength, rather than the capacity of soil P sorption.

Our results have important implications for understanding P bioavailability in tropical forest soils. These results suggest that reducing events can potentially increase P bioavailability by decreasing the P sorption strength of minerals, even though P sorption capacity remained high. An interesting question for future studies is whether plant roots and microbes can take advantage of the increased P solubility during reducing events. Anaerobic conditions can be stressful for plants and microbes; however, studies have reported similar soil respiration rates under aerobic and anaerobic conditions in humid tropical forest soils (DeAngelis et al., 2010; Pett-Ridge, 2005); (DeAngelis et al., 2010; Pett-Ridge, 2005; Bhattacharyya et al., 2018). Soil microbes appear to be well-adapted to dynamic redox

conditions, at least at the scales of days to weeks. Thus, it is possible that microbes can benefit from the increased P solubility under anoxic conditions. At a watershed scale, topography and parent material are important controls of soil sorption behavior and P bioavailability. Although P sorption capacity was high in soils with high Fe and Al concentrations, these soils also responded more strongly to reducing events. Thus, redox dynamic may be particularly important in these soils to facilitate biological uptake and ecosystem retention of P.

5 Conclusions

We found that minerals can retain high P sorption capacity during reducing events in highly weathered tropical forest soils. ~~The~~**Their** high P sorption capacity is expected to contribute to low P concentrations in soil solution and limit the potential for P leaching. Due to the high P sorption capacity **in tropical soils**, current and future increases in precipitation associated with climate change are unlikely to drastically alter P leaching in these environments. Reducing events also decreased the strength of P sorption and potentially increased P bioavailability. Thus, episodic reducing events could serve as ‘hot moments’ for plants and microbes to acquire soil P that would otherwise be tightly bound to minerals. As a result of altered rainfall regimes, more frequent or intensive redox oscillation could increase P bioavailability, if it does not impose a strong O₂ limitation on primary productivity or decomposition.

Author contributions: YL, AG, and WLS conceived the study; YL, AG, and CSO performed the research; YL led the manuscript development and data analysis; all authors contributed to writing.

Competing interests: The authors declare that they have no conflict of interest.

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References

- 30 [Appan, A., and Wang, H.: Sorption isotherms and kinetics of sediment phosphorus in a tropical reservoir, *Journal of Environmental Engineering*, 126, 993-998, 2000.](#)
- Bache, B. W., and Williams, E. G.: A phosphate sorption index for soils, *Journal of Soil Science*, 22, 289-301, 1971.
- Barcellos, D., Cyle, K. T., and Thompson, A.: Faster redox fluctuations can lead to higher iron reduction rates in humid forest soils, *Biogeochemistry*, 10.1007/s10533-018-0427-0, 2018.
- 35 [Bhattacharyya, A., Campbell, A. N., Tfiaily, M. M., Lin, Y., Kukkadapu, R. K., Silver, W., Nico, P. S., and Pett-Ridge, J.: Redox fluctuations control the coupled cycling of iron and carbon in tropical forest soils, *Environmental Science & Technology*, 10.1021/acs.est.8b03408, 2018.](#)

- Bolster, C. H., and Hornberger, G. M.: On the use of linearized Langmuir equations, *Soil Science Society of America Journal*, 71, 1796, 10.2136/sssaj2006.0304, 2007.
- Borch, T., and Fendorf, S.: Chapter 12 Phosphate Interactions with Iron (Hydr)oxides: Mineralization Pathways and Phosphorus Retention upon Bioreduction, *Developments in Earth and Environmental Sciences*, 7, 321-348, 10.1016/S1571-9197(07)07012-7, 2007.
- Borggaard, O. K., Jørgensen, S. S., Møberg, J. P., and Raben - Lange, B.: Influence of organic matter on phosphate adsorption by aluminium and iron oxides in sandy soils, *Journal of Soil Science*, 41, 443-449, 10.1111/j.1365-2389.1990.tb00078.x, 1990.
- Brown, S., Lugo, A. E., Silander, S., and Liegel, L.: Research history and opportunities in the Luquillo Experimental Forest, U.S. Department of Agriculture Forest Service, General Technical Report SO-44, Southern Forest Experiment Station, New Orleans, LA, 128 p, 1983.
- Camenzind, T., Hättenschwiler, S., Treseder, K. K., Lehmann, A., and Rillig, M. C.: Nutrient limitation of soil microbial processes in tropical forests, *Ecological Monographs*, 0, 1-18, 10.1002/ecm.1279, 2017.
- Chacón, N., Silver, W. L., Dubinsky, E. A., and Cusack, D. F.: Iron reduction and soil phosphorus solubilization in humid tropical forests soils: The roles of labile carbon pools and an electron shuttle compound, *Biogeochemistry*, 78, 67-84, 10.1007/s10533-005-2343-3, 2006.
- Chen, C., Meile, C., Wilmoth, J. L., Barcellos, D., and Thompson, A.: Influence of pO₂ on iron redox cycling and anaerobic organic carbon mineralization in a humid tropical forest soil, *Environmental Science & Technology*, 2018.
- Cleveland, C. C., Townsend, A. R., Taylor, P., Alvarez - Clare, S., Bustamante, M., Chuyong, G., Dobrowski, S. Z., Grierson, P., Harms, K. E., and Houlton, B. Z.: Relationships among net primary productivity, nutrients and climate in tropical rain forest: a pan - tropical analysis, *Ecology Letters*, 14, 939-947, 2011.
- Coward, E. K., Thompson, A. T., and Plante, A. F.: Iron-mediated mineralogical control of organic matter accumulation in tropical soils, *Geoderma*, 306, 206-216, 10.1016/j.geoderma.2017.07.026, 2017.
- de Campos, M., Antonangelo, J. A., and Alleoni, L. R. F.: Phosphorus sorption index in humid tropical soils, *Soil and Tillage Research*, 156, 110-118, 10.1016/j.still.2015.09.020, 2016.
- DeAngelis, K. M., Silver, W. L., Thompson, A. W., and Firestone, M. K.: Microbial communities acclimate to recurring changes in soil redox potential status, *Environmental Microbiology*, 12, 3137-3149, 10.1111/j.1462-2920.2010.02286.x, 2010.
- Dubinsky, E. A., Silver, W. L., and Firestone, M. K.: Tropical forest soil microbial communities couple iron and carbon biogeochemistry, *Ecology*, 91, 2604-2612, 10.1890/09-1365.1, 2010.
- Gérard, F.: Clay minerals, iron/aluminum oxides, and their contribution to phosphate sorption in soils - A myth revisited, *Geoderma*, 262, 213-226, 10.1016/j.geoderma.2015.08.036, 2016.
- Gross, A., Pett-Ridge, J., and Silver, W.: Soil Oxygen Limits Microbial Phosphorus Utilization in Humid Tropical Forest Soils, *Soil Systems*, 2, 65, 10.3390/soilsystems2040065, 2018.
- Gustafsson, J. P., Mwamila, L. B., and Kergoat, K.: The pH dependence of phosphate sorption and desorption in Swedish agricultural soils, *Geoderma*, 189-190, 304-311, 10.1016/j.geoderma.2012.05.014, 2012.
- Gustafsson, J. P. *Visual MINTEQ 3.1 user guide. KTH, Department of Land and Water Resources, Stockholm, Sweden: <https://vminiteq.lwr.kth.se/>, access: July 22nd, 2019, 2015.*
- Hall, S. J., and Silver, W. L.: Reducing conditions, reactive metals, and their interactions can explain spatial patterns of surface soil carbon in a humid tropical forest, *Biogeochemistry*, 125, 149-165, 10.1007/s10533-015-0120-5, 2015.
- Haynes, R. J.: Effects of liming on phosphate availability in acid soils, *Plant and Soil*, 68, 289-308, 10.1007/BF02197935, 1982.
- Haynes, R. J., and Swift, R. S.: The effects of pH and drying on adsorption of phosphate by aluminium - organic matter associations, *Journal of Soil Science*, 40, 773-781, 10.1111/j.1365-2389.1989.tb01317.x, 1989.
- Hedley, M. J., Stewart, J. W. B., and Chauhan, B.: Changes in inorganic and organic soil phosphorus fractions induced by cultivation practices and by laboratory incubations, *Soil Science Society of America Journal*, 46, 970-976, 1982.
- Heiberg, L., Koch, C. B., Kjaergaard, C., Jensen, H. S., and Hans Christian, B. H.: Vivianite precipitation and phosphate sorption following iron reduction in anoxic soils, *Journal of Environmental Quality*, 41, 938-949, 10.2134/jeq2011.0067, 2012.
- Henry, P. C., and Smith, M. F.: Two-step approach to determining some useful phosphorus characteristics of South African soils: a review of work done at the ARC-Institute for Soil, Climate and Water, *South African Journal of Plant and Soil*, 23, 64-72, 2006.
- Holford, I. C. R., and Patrick, W. H.: Effects of reduction and pH changes on phosphate sorption and mobility in an acid soil, *Soil Science Society of America Journal*, 43, 292-297, 1979.
- Johnson, A. H., Frizano, J., and Vann, D. R.: Biogeochemical implications of labile phosphorus in forest soils determined by the Hedley fractionation procedure, *Oecologia*, 135, 487-499, 10.1007/s00442-002-1164-5, 2003.
- Keiluweit, M., Nico, P. S., Kleber, M., and Fendorf, S.: Are oxygen limitations under recognized regulators of organic carbon turnover in upland soils?, *Biogeochemistry*, 10.1007/s10533-015-0180-6, 2016.
- Lin, Y., Bhattacharyya, A., Campbell, A. N., Nico, P. S., Pett-Ridge, J., and Silver, W. L.: Phosphorus fractionation responds to dynamic redox conditions in a humid tropical forest soil, *Journal of Geophysical Research: Biogeosciences*, 123, 3016-3027, 10.1029/2018JG004420, 2018.
- Lindsay, W. L.: Iron, in: *Chemical equilibria in soils*, edited by: Lindsay, W. L., John Wiley and Sons Ltd., 129-149, 1979.
- Liptzin, D., and Silver, W. L.: Effects of carbon additions on iron reduction and phosphorus availability in a humid tropical forest soil, *Soil Biology and Biochemistry*, 41, 1696-1702, 10.1016/j.soilbio.2009.05.013, 2009.

- Mage, S. M., and Porder, S.: Parent material and topography determine soil phosphorus status in the Luquillo Mountains of Puerto Rico, *Ecosystems*, 16, 284-294, 10.1007/s10021-012-9612-5, 2012.
- Maranguit, D., Guillaume, T., and Kuzyakov, Y.: Effects of flooding on phosphorus and iron mobilization in highly weathered soils under different land-use types: Short-term effects and mechanisms, *Catena*, 158, 161-170, 10.1016/j.catena.2017.06.023, 2017.
- 5 McDowell, W. H.: Internal nutrient fluxes in a Puerto Rican rain forest, *Journal of Tropical Ecology*, 14, 521-536, 10.1017/S0266467498000376, 1998.
- McDowell, W. H., and Liptzin, D.: Linking soils and streams: Response of soil solution chemistry to simulated hurricane disturbance mirrors stream chemistry following a severe hurricane, *Forest Ecology and Management*, 332, 56-63, 10.1016/j.foreco.2014.06.001, 2014.
- 10 McGechan, M. B., and Lewis, D. R.: Sorption of Phosphorus by Soil, Part I: Principles, Equations and Models, *Biosystems Engineering*, 82, 1-24, 10.1006/bioe.2001.0013, 2002.
- Murphy, J., and Riley, J. P.: A modified single solution method for the determination of phosphate in natural waters, *Analytica Chimica Acta*, 27, 31-36, 1962.
- 15 Murphy, S. F., Stallard, R. F., Scholl, M. A., González, G., and Torres-Sánchez, A. J.: Reassessing rainfall in the Luquillo Mountains, Puerto Rico: Local and global ecohydrological implications, *PLoS ONE*, 12, 10.1371/journal.pone.0180987, 2017.
- O'Connell, C. S., Ruan, L., and Silver, W. L.: Drought drives rapid shifts in tropical rainforest soil biogeochemistry and greenhouse gas emissions, *Nature Communications*, 9, 1348, doi:10.1038/s41467-018-03352-3, 2018.
- 20 Oh, Y. M., Hesterberg, D. L., and Nelson, P. V.: Comparison of phosphate adsorption on clay minerals for soilless root media, *Communications in Soil Science and Plant Analysis*, 30, 747-756, 1999.
- Olander, L. P., and Vitousek, P. M.: Biological and geochemical sinks for phosphorus in soil from a wet tropical forest, *Ecosystems*, 7, 404-419, 10.1007/s10021-004-0264-y, 2004.
- Patrick, W. H., and Khalid, R. A.: Phosphate release and sorption by soils and sediments: effect of aerobic and anaerobic conditions, *Science*, 186, 53-55, 1974.
- 25 Peretyazhko, T., and Sposito, G.: Iron(III) reduction and phosphorous solubilization in humid tropical forest soils, *Geochimica et Cosmochimica Acta*, 69, 3643-3652, 10.1016/j.gca.2005.03.045, 2005.
- Pett-Ridge, J.: Rapidly fluctuating redox regimes frame the ecology of microbial communities and their biogeochemical function in a humid tropical soil, Ph.D., University of California, Berkeley, 2005.
- 30 Pett-Ridge, J. C.: Contributions of dust to phosphorus cycling in tropical forests of the Luquillo Mountains, Puerto Rico, *Biogeochemistry*, 94, 63-80, 10.1007/s10533-009-9308-x, 2009.
- Quintero, C. E., Gutiérrez-Boem, F. H., Befani, M. R., and Boschetti, N. G.: Effects of soil flooding on P transformations in soils of the Mesopotamia region, Argentina, *Journal of Plant Nutrition and Soil Science*, 170, 500-505, 10.1002/jpln.200625015, 2007.
- 35 R Core Team: R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>, 2018.
- Rakotoson, T., Amery, F., Rabeharisoa, L., and Smolders, E.: Soil flooding and rice straw addition can increase isotopic exchangeable phosphorus in P-deficient tropical soils, *Soil Use and Management*, 30, 189-197, 10.1111/sum.12120, 2014.
- 40 Reed, S. C., and Wood, T. E.: Soil phosphorus cycling in tropical soils: An ultisol and oxisol perspective, in: *Soil phosphorus*, Taylor & Francis, 247-283, 2016.
- Roy, E. D., Willig, E., Richards, P., Martinelli, L. A., Vazquez, F. F., Pegorini, L., Spera, S. A., and Porder, S.: Soil phosphorus sorption capacity after three decades of intensive fertilization in Mato Grosso, Brazil, *Agriculture, Ecosystems and Environment*, 249, 206-214, 10.1016/j.agee.2017.08.004, 2017.
- 45 Ryden, J. C., and Syers, J. K.: Origin of the labile phosphate pool in soils, *Soil Science*, 123, 353-361, 1977.
- Sallade, Y. E., and Sims, J. T.: Phosphorus transformations in the sediments of Delaware's agricultural drainageways: II. Effect of reducing conditions on phosphorus release, *Journal of Environmental Quality*, 26, 1579-1588, 1997.
- Scatena, F. N.: An introduction to the physiography and history of the Bisley Experimental Watersheds in the Luquillo Mountains of Puerto Rico, US Department of Agriculture, Forest Service, Southern Forest Experiment Station, 1989.
- 50 Scatena, F. N., and Lugo, A. E.: Geomorphology, disturbance, and the soil and vegetation of two subtropical wet steepland watersheds of Puerto Rico, *Geomorphology*, 13, 199-213, 1995.
- Schuur, E. A. G., Chadwick, O. A., and Matson, P. A.: Carbon cycling and soil carbon storage in mesic to wet Hawaiian montane forests, *Ecology*, 82, 3182-3196, 2001.
- 55 Shenker, M., Seitelbach, S., Brand, S., Haim, A., and Litaor, M. I.: Redox reactions and phosphorus release in re-flooded soils of an altered wetland, *European Journal of Soil Science*, 56, 515-525, 2005.
- Silver, W. L., Scatena, F. N., Johnson, A. H., Siccama, T. G., and Sanchez, M. J.: Nutrient availability in a montane wet tropical forest: spatial patterns and methodological considerations, *Plant and soil*, 164, 129-145, 1994.
- Silver, W. L., Lugo, A. E., and Keller, M.: Soil oxygen availability and biogeochemistry along rainfall and topographic gradients in upland wet tropical forest soils, *Biogeochemistry*, 44, 301-328, 10.1023/A:1006034126698, 1999.
- 60 Thompson, A., and Goynes, K. W.: Introduction to the sorption of chemical constituents in soils, *Nature Education Knowledge*, 4, 7, 2012.

- Vitousek, P. M., and Sanford Jr., R. L.: Nutrient cycling in moist tropical forest, *Annual review of Ecology and Systematics*, 17, 137-167, 1986.
- Walpersdorf, E., Koch, C. B., Heiberg, L., O'Connell, D. W., Kjaergaard, C., and Hansen, H. C. B.: Does vivianite control phosphate solubility in anoxic meadow soils?, *Geoderma*, 193-194, 189-199, 10.1016/j.geoderma.2012.10.003, 2013.
- 5 Weaver, P. L., and Murphy, P. G.: Forest structure and productivity in Puerto Rico's Luquillo Mountains, *Biotropica*, 22, 69-82, 1990.
- Wieder, W. R., Cleveland, C. C., and Townsend, A. R.: Throughfall exclusion and leaf litter addition drive higher rates of soil nitrous oxide emissions from a lowland wet tropical forest, *Global Change Biology*, 17, 3195-3207, 2011.
- 10 Willett, I. R.: Causes and prediction of changes in extractable phosphorus during flooding, *Soil Research*, 27, 45-54, 1989.
- Wilmoth, J. L., Moran, M. A., and Thompson, A.: Transient O₂ pulses direct Fe crystallinity and Fe(III)-reducer gene expression within a soil microbiome, *Microbiome*, 6, 189, 10.1186/s40168-018-0574-5, 2018.
- [Wisawapipat, W., Kheoruenromne, I., Suddhiprakarn, A., and Gilkes, R. J.: Phosphate sorption and desorption by Thai upland soils, *Geoderma*, 153, 408-415, <https://doi.org/10.1016/j.geoderma.2009.09.005>, 2009.](https://doi.org/10.1016/j.geoderma.2009.09.005)
- 15 Zhang, Y. S., Lin, X. Y., and Werner, W.: The effect of soil flooding on the transformation of Fe oxides and the adsorption/desorption behavior of phosphate, *Journal of Plant Nutrition and Soil Science*, 166, 68-75, 10.1002/jpln.200390014, 2003.

Site	Position	Gravimetric soil moisture (%)	pH	Total soil C (%)	Total soil N (%)	HCl-Fe(II) (g Fe kg ⁻¹)	HCl-Fe(III) (g Fe kg ⁻¹)	AO-Fe (g Fe kg ⁻¹)	AO-Al (g Al kg ⁻¹)
El Verde 1	Slope	95.7 ± 2.4	5.27 ± 0.02	7.06 ± 0.37	0.386 ± 0.007	0.054 ± 0.002	1.97 ± 0.11	0.84 ± 0.15	0.37 ± 0.04
	Valley	124.5 ± 1.3	5.70 ± 0.06	5.91 ± 0.16	0.307 ± 0.004	9.42 ± 0.29	1.94 ± 0.05	4.65 ± 0.72	0.45 ± 0.07
Rio Icaeos	Slope	71.6 ± 0.4	4.72 ± 0.01	3.90 ± 0.06	0.196 ± 0.002	0.14 ± 0.04	0.84 ± 0.13	0.35 ± 0.04	1.18 ± 0.07
	Valley	103.7 ± 0.3	5.26 ± 0.06	4.28 ± 0.11	0.237 ± 0.002	0.14 ± 0.01	0.82 ± 0.04	0.37 ± 0.02	2.13 ± 0.12
El Verde 2	Slope	85.1 ± 2.4	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
P solubility	Valley	78.5 ± 0.4	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.

Table 1. Initial soil characteristics (n = 4).

Site	Position	Gravimetric soil moisture (%)	pH	Total soil C (%)	Total soil N (%)
El Verde 1	Slope	95.7 ± 2.4	5.27 ± 0.02	7.06 ± 0.37	0.386 ± 0.007
	Valley	124.5 ± 1.3	5.70 ± 0.06	5.91 ± 0.16	0.307 ± 0.004
Rio Icaeos	Slope	71.6 ± 0.4	4.72 ± 0.01	3.90 ± 0.06	0.196 ± 0.002
	Valley	103.7 ± 0.3	5.26 ± 0.06	4.28 ± 0.11	0.237 ± 0.002
El Verde 2	Slope	85.1 ± 2.4	n.d.	n.d.	n.d.
P solubility	Valley	78.5 ± 0.4	n.d.	n.d.	n.d.

Means and S.E. are shown. n.d., not determined.

Table 2. Maximum P sorption capacities (S_{max}), rates of P sorption (k), and soil Fe and Al fractions in response to redox treatments and their correlations with phosphorus sorption index (PSI) calculated from sorption data under 1000 mg P kg⁻¹ ($n = 4$)

Site	Position	Treat-ment	S_{max} (mg P kg ⁻¹)	k	HCl-Fe(II) (g Fe kg ⁻¹)	HCl-Fe(III) (g Fe kg ⁻¹)	AO-Fe (g Fe kg ⁻¹)	AO-Al (g Al kg ⁻¹)
El Verde	Slope	Anoxic	7451 ± 562 4471-4893 ± 5453	0.275 ± 0.020	3.37 ± 0.12a	0.86 ± 0.09b	0.92 ± 0.06a	0.29 ± 0.03a
		Oxic	5664352 ± 387 7142 ± 5586569 ± 487	0.244 ± 0.016	0.038 ± 0.003b	1.55 ± 0.12a	0.75 ± 0.09a	0.30 ± 0.03a
		Anoxic	7282 ± 1207 6291 ± 44695369 ± 1056	0.288 ± 0.027	13.72 ± 0.22a	1.85 ± 0.08b	1.37 ± 0.03a	0.20 ± 0.004a
		Oxic	2526 ± 6672627 ± 303 8256 ± 2519	0.486 ± 0.049	4.06 ± 0.69b	5.63 ± 0.55a	1.33 ± 0.23a	0.17 ± 0.01b
Rio Icacos	Slope	Anoxic	2290 ± 250 3290 ± 250	0.303 ± 0.015	3.36 ± 0.17a	0.67 ± 0.13a	0.29 ± 0.01a	1.00 ± 0.04a
		Oxic	0.366673 † 0.372068 †	0.194 ± 0.013	0.060 ± 0.002b	0.80 ± 0.05a	0.31 ± 0.03a	0.98 ± 0.07a
	Valley	Anoxic	0.863 † 0.863 †	0.365 ± 0.022	3.39 ± 0.30a	0.90 ± 0.06a	0.29 ± 0.004b	2.01 ± 0.05a
		Oxic	0.092 0.092	0.353 ± 0.025	0.193 ± 0.124b	0.92 ± 0.10a	0.34 ± 0.01a	1.89 ± 0.05a
Correlation with PSI ($n = 32$)		r	0.366673 † 0.372068 †	0.863 †	0.092	0.434	0.001	0.395
		P	0.366673 † 0.372068 †	0.006 †	0.616	0.013	0.998	0.028

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Means and S.E. are shown. Different letters indicate significant effects of redox treatment in each combination of site and topographic position (T-tests). Significant correlation coefficient (r) and P values are in bold. †, $n = 8$.

Figure 1. Effects of anoxic vs. oxic pre-incubation on the P sorption isotherms of slope (left panel) and valley (right panel) soils from El Verde (upper panel) and Rio Icacos (lower panel). Means and standard errors of means are shown. Inserts are the zoomed-in versions of the lower left portion of the respective plots.

- 5 Figure 2. Effects of anoxic vs. oxic pre-incubation on the P sorption index (PSI) of slope and valley soils from El Verde and Rio Icacos. The PSI values were calculated from sorption isotherm data under $1000 \text{ mg P kg}^{-1}$, which better represented P sorption behavior under low levels of P addition than the estimated maximum P sorption capacity. Means and standard errors of means are shown. * indicates significant difference of PSI for the respective combination of site and topographic position.
- 10 Figure 3. Effects of anoxic vs. oxic pre-incubation on the P sorption time curves of slope (left panel) and valley (right panel) soils from El Verde (upper panel) and Rio Icacos (lower panel). Phosphorus was added at the level of $1000 \text{ mg P kg}^{-1}$. Means and standard errors of means are shown.
- 15 Figure 4. Effects of anoxic vs. oxic pre-incubation on the relative solubility of the sorbed P in NaHCO_3 and NaOH solution under P addition of 100 mg P kg^{-1} (P100, left panel) and $1000 \text{ mg P kg}^{-1}$ (P1000, right panel). Soils were collected from slope and valley positions at El Verde. Means and standard errors of means are shown. * indicates significant difference of PSI for the respective combination of site, topographic position, and level of P addition.

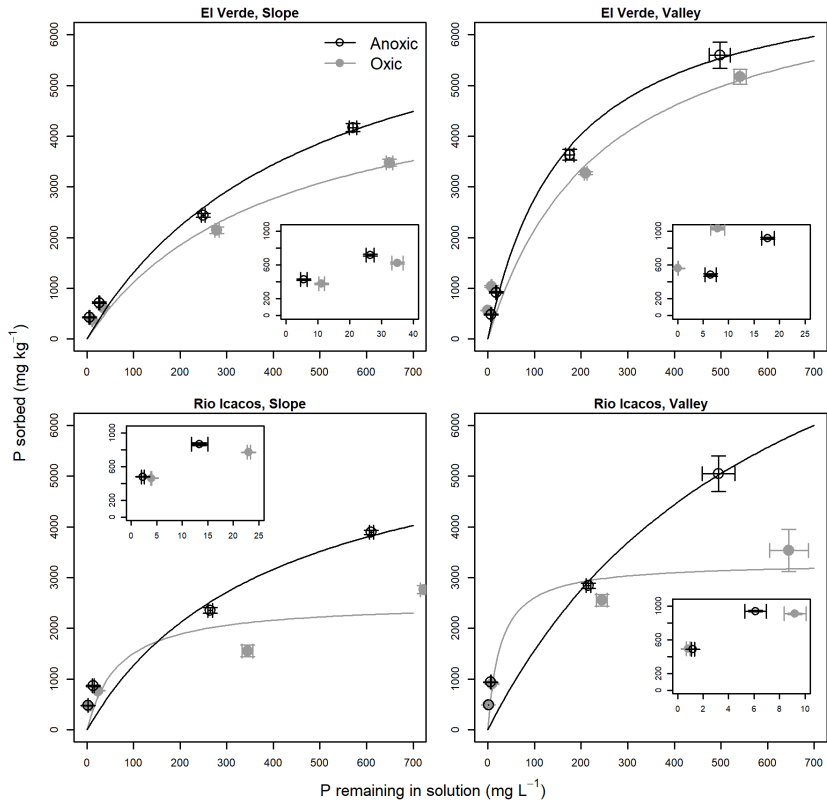


Fig. 1

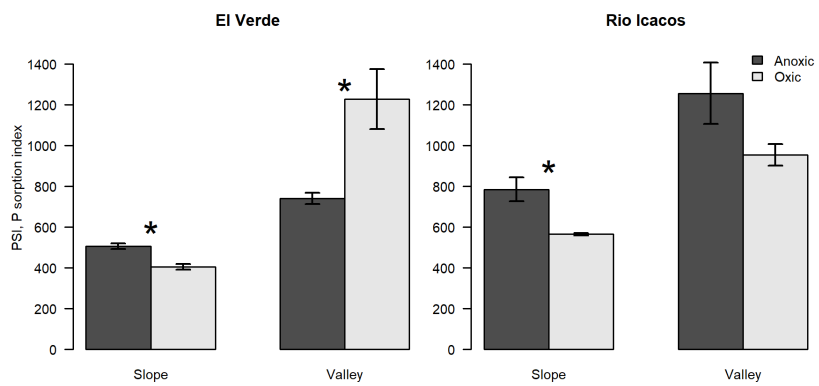


Fig. 2

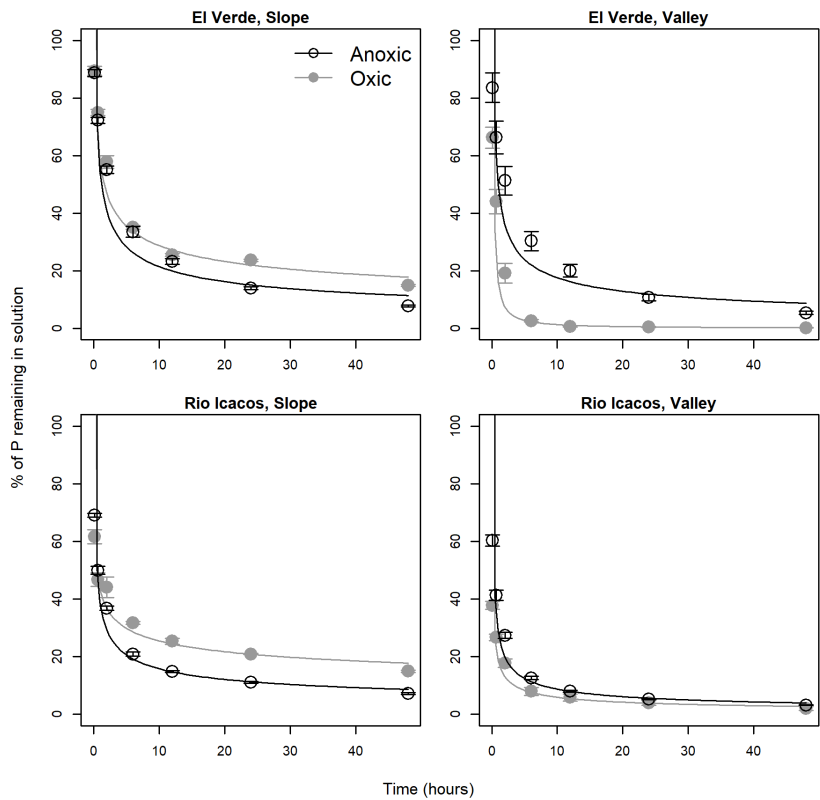


Fig. 3

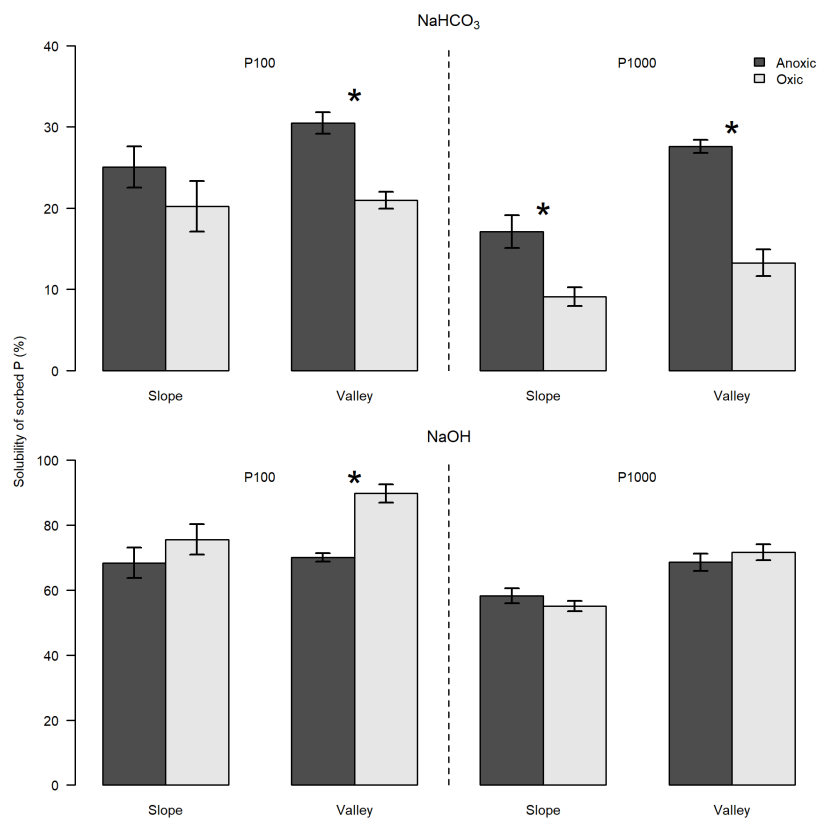


Fig. 4