Structure and functioning of epipelagic mesozooplankton and response to dust deposition events during the spring PEACETIME cruise in the Mediterranean Sea

Guillermo Feliú¹, Marc Pagano¹, Pamela Hidalgo², François Carlotti¹

¹ Aix Marseille Univ., Universite de Toulon, CNRS, IRD, MIO UM 110, 13288, Marseille, France

Correspondence to: François Carlotti (francois.carlotti@mio.osupytheas.fr)

(Revised version: new or modified text is underlined in yellow)

Abstract. The PEACETIME cruise (May-June 2017) was a basin scale survey dedicated to the investigation of different planktonic trophic regimes in the Algerian, Tyrrhenian and Ionian basins and, in particular, focusing on areas impacted by Saharan dust deposition. This paper presents the structural and functioning patterns of the zooplankton component during this survey, including their responses to two mature that the Algerian and Tyrrhenian basins. The mesozooplankon was sampled at 12 stations by combining nets with 2 mesh sizes (100 and 200 µm) mounted on a Bongo frame for vertical hauls within the upper 300 meter layer. In this general post-bloom situation, total mesozooplankton showed reduced variations in abundance and biomass over the whole area, with a noticeable contribution of the small size fraction (< 500 µm) of up to 50 % in abundance and 25 % in biomass. The taxonomic structure was dominated by compods, mainly cyclopoids and calanoids, and completed by appendicularians, ostracods and chaetognaths. Distinct complankton taxa assemblages in the three main areas were in agreement with recently proposed regional patterns for the Mediterranean Basin, although the assemblages found in the western Ionian stations presented a closer analogy with those of the Tyrrhenian basin than with those of the Ionian basin, probably due to Atlantic water influence.

Zooplankton carbon demand, grazing pressure, respiration and excretion rates were estimated using allometric relationships to the mesozooplankton size spectrum. On average, the daily zooplankton consumption potentially represents 15 % of the phytoplankton stock, almost the whole of the primary production, with a narrow range of variations, and its excretion contributes roughly one quarter of the N and P requirements of phytoplankton production. The small-size fractions make a significant contribution to these mesozooplankton estimated fluxes. Whereas in the Algerian basin (long-duration station FAST), the initial impact on the pelagic ecosystem of a tracked dust deposition was studied, the survey of the southern Tyrrhenian basin occurred almost a week after another dust event. The changes in mesozooplankton taxonomic structure appear to be a relevant indicator to study this response, with an initial phase with no real dominance of taxa, then a disturbed state of the community with strong dominance of certain filter-feeding taxa and the appearance of carnivorous species, and finally a recovery state towards a more stable system with diversification of the community. To our knowledge,

² Department of Oceanography and Millennium Institute of Oceanography, Faculty of Natural Science and Oceanography, University of Concepcion, P.O. Box 160 C, Concepción, Chile

PEACETIME is the first in situ study allowing observation of mesozooplankton responses before and soon after natural Saharan dust depositions. The changes in rank-frequency diagrams of the zooplankton taxonomic structure are an interesting index to highlight short-term responses of zooplankton to episodic dust deposition events.

35 1 Introduction

The Mediterranean Sea is a semi-enclosed basin connected to the Atlantic Ocean and the Black Sea. It is composed of two major sub-basins, the Eastern and Western Mediterranean, connected by the Sicily strait (Skliris, 2014). The Mediterranean Sea can be considered as a model of the world's oceans (Bethoux et al., 1999; Lejeusne et al., 2010) because of its characteristics, such as the unique thermohaline circulation pattern and the deep water formation process. In addition, it is considered to be oligotrophic with an excess of carbon, a deficiency of phosphorus relative to nitrogen (MERMEX Group, 2011) and a decreasing west-east gradient in chlorophyll-*a* (i.e. Siokou-Frangou et al., 2010).

For the last two hundred years, numerous investigations have documented the pelagic zooplankton community inhabiting the Mediterranean Sea (Saiz et al., 2014), including long-term time series (i.e. Fernández de Puelles et al., 2003; Mazzocchi et al., 2007; Molinero et al., 2008; García-Comas et al., 2011; Berline et al., 2012) and a succession of oceanographic surveys covering wide transects at different time periods of the year (Kimor and Wood, 1975; Nowaczyk et al., 2011; Donoso et al., 2017; Siokou et al., 2019). The regular monitoring of the zooplankton community is essential when considering the high sensitivity of the Mediterranean Sea to anthropogenic and climate disturbance (Sazzini et al., 2014). Some of those disturbances may alter the structure and functioning of the pelagic ecosystem, and this is critical considering that marine ecosystems are being altered by anthropogenic climate change at an unprecedented rate (Chust et al., 2017).

Dust deposition is a major source of micro- and macro-nutrients (Wagener et al., 2010) that can stimulate primary production (Ridame et al., 2014), accelerate carbon sedimentation and possibly aggregation of marine particles (i.e. Neuer et al., 2004; Ternon et al., 2010; Bressac et al., 2014). Large amounts of Saharan dust can be transported in the atmosphere throughout the western and eastern Mediterranean Sea and then deposited on the sea surface by wet or dry deposition. The PEACETIME oceanographic cruise, carried out between May 10 and June 11 of 2017, was designed to study *in situ* the processes occurring in the Mediterranean Sea after atmospheric dust deposition and their impact on marine nutrient budget and fluxes, and on the biogeochemical functioning of the pelagic ecosystem. Thus, the survey strategy was designed to be flexible in order to be able to change the sampling area depending on atmospheric events (Guieu et al., accepted). Consequently, the survey sampling program realized consisted in 14 oceanographic stations in the central and western parts of the Mediterranean Sea.

The aims of the present contribution to the PEACETIME project are 1) to document the zooplankton abundance, biomass and size distribution along the survey transect, with special attention to small-sized zooplankton; 2) to analyze the relationship between zooplankton structure and environmental variability, including dust deposition; 3) to estimate the

bottom-up (nutrient regeneration) and the top-down (grazing) impact of zooplankton on phytoplankton stock and production by estimating its ingestion, respiration, ammonium and phosphate excretions using allometric models.

These objectives will serve to test the following hypotheses: whether the Saharan dust events impact the zooplankton community structure following deposition (H1), and if so, whether the effect would be immediately observable or after a lag time (H2). Finally, whether changes in zooplankton structure driven by dust deposition exceed regional differences under oligothropic conditions (H3).

2 Material and methods

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2.1 Study area and environmental variables

The PEACETIME cruise survey was conducted in May/June 2017 in the Western Mediterranean Sea (Figure 1) on board R.V. *Pourquoi pas?*. Among the 12 stations studied, 10 were sampled once for zooplankton (the short-duration stations ST1 to ST9, and the long-duration station TYR), whereas two long-duration stations ION and FAST, lasting 3 and 5 days respectively, were sampled three times. The station positions along the transect were planned before the cruise in order to sample the principal ecoregions (see Figure 4 in Guieu et al., accepted), with the exception of FAST, an opportunistic station to monitor a wet dust deposition event which occurred on June 5 a few hours after the first sampling date (Table 1). A large area including the southern Tyrrhenian Sea starting on May 10 which could have impacted the samples at ST5, ST6 and TYR which were sampled on May 16, 19 and 22, respectively (pers. comm. C. Guieu).

Hydrological variables (temperature, density, salinity) were measured on vertical profiles using a CTD. Dissolved oxygen was measured using a SBE43 sensor and chlorophyll-a concentration was determined from Niskin bottle samples by HPLC following the protocol of Ras et al. (2008), and with a Fluorescence sensor coupled with the CTD. Primary production was measured with the 14 C-uptake technique, following the methods detailed in (Marañón et al., 2000). The depth of the mixed layer (MLD) was computed using the density difference criterion $\Delta \sigma_{\theta} = 0.03 \ kgr^{-3}$ defined in de Boyer Montégut et al. (2004).

2.2 Ancillary data on dust deposition events occurring during the PEACETIME survey

Guieu et al. oductory paper) detailed how they used three regional dust transport models to identify major dust events during the PEACETIME cruise. Two major wet dust events occurred during the period (Table 2). The first concerned the whole southern Tyrrhenian basin, with predicted flux > 1g m⁻² (Desboeufs et al. in prep.), and started on May 10, several days before the arrival of vessel in this area. The dust event was confirmed by aluminium, iron and lithogenic Si measured in sediment traps at TYR with a lithogenic flux between 200 and 1000m of 150-200 mg m⁻² (Bressac et al., in

prep.). The second was located in the area between the Balearics and the Algerian coast and occurred from 3 to 5 June, with predicted flux of 0.5 g m⁻² (Guieu et al., accepted) after the arrival of the vessel in this area (station FAST). The dust event was confirmed by on-board atmospheric dust deposition samples (Desboeufs, in preparation this special issue), water column observations (nutrients, trace metals) (Tovar-Sánchez et al. 2020) and tracers of dust deposition in sediment traps comm. C. Guieu). The highest aerosol mass concentrations (around 25 μg m⁻³) with the highest iron content (245 ng m⁻³) were measured at FAST between 1 and 5 June, and subsequently the highest trace metal concentrations in the surface microlayer were measured on 4 June (Co: 773.6 pM; Cu: 20.1 nM; Fe: 1433.3 nM; and Pb: 1294.7 pM) (Tovar-Sánchez et al 2020). The chemical composition of rain samples at FAST confirmed wet deposition of dust reaching a total particulate flux of 0.012 g m⁻² (Fu et al., in prep.). The Ionian basin was the only southern area not impacted by dust deposition during the PEACETIME cruise, and results obtained at the long-duration station ION will be use or comparison.

2.3 Zooplankton sampling and sample processing

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A total of 16 zooplankton samples were collected at 12 stations (Table 1) using a Bongo frame (double net ring of 60 cm mouth diameter) equipped with 100 μ m and 200 μ m mesh size nets (noted N_{100} and N_{200} below) mounted with filtering codends. At all sampling stations, the Bongo frame was vertically towed from 300 m depth to the surface at a constant speed of 1ms^{-1} . Sample volume was estimated based on the ring diameter and the towed cable length. The sampling was mostly performed during the morning, except for ST7, ST9 and TYR, and night tows were also performed for the long-duration stations FAST and ION. The samples were preserved in 4% borax-buffered formalin immediately after the net was hauled back onto the deck.

The samples were processed using FlowCAM (Fluid Imaging Technologies Inc. Series VS-IV, Benchtop model) and ZOOSCAN (Gorsky et al., 2010). One of the goals of this study was to achieve determination of the complete size structure of the zooplankton community by combining different plankton mesh size nets and analysis techniques (FlowCAM and ZOOSCAN) in order to optimize the observed size spectrum. The formalin preserved samples were rinsed with tap water to remove the formalin. For net N_{100} , the sample was then split into 3 size fractions: < 200 μ m (noted below $N_{100}F_{<200}$), 200 μ m – 1000 μ m (noted below $N_{100}F_{>1000}$), and > 1000 μ m (noted below $N_{100}F_{>1000}$). For net N_{200} , the sample was split into two size fractions: < 1000 μ m (noted below $N_{200}F_{<1000}$) and > 1000 μ m (noted below $N_{200}F_{>1000}$).

To determine the complete size spectrum, different combinations of size fractions from the two nets and analytical techniques were tested using two-way ANOVA. Taking into account the two mesh sizes, (N_{100}, N_{200}) , the limits of the size spectrum were defined from the fraction $N_{100}F_{<200}$ for the lower limit and from the fraction $N_{200}F_{>1000}$ for the upper limit. Considering that our FlowCAM does not detect particles larger than 1200 μ m of ESD and our ZOOSCAN does not detect particles smaller than 300 μ m of ESD, $N_{100}F_{<200}$ was analyzed by FlowCAM and $N_{200}F_{>1000}$ by ZOOSCAN. The intermediate size fractions $N_{100}F_{200/1000}$ and $N_{200}F_{<1000}$ were both analyzed with ZOOSCAN and FlowCAM. These analyses delivered abundance and biomass values for successive ESD size classes: <200 μ m (noted $C_{<200}$); 200-300 μ m ($C_{200-300}$); 300-500 μ m

 $(C_{300-500})$; 500-1000 μm $(C_{500-1000})$; 1000-2000 μm $(C_{1000-2000})$; > 2000 μm $(C_{200-300})$. The challenge was to choose the best net-analysis technique combination for the intermediate size fractions $(C_{200-300}, C_{300-500} \text{ and } C_{500-1000})$. The abundance of each class for the two nets and the two treatments was statistically compared. Parts of the spectrum corresponding to fractions $C_{200-300}$ and $C_{300-500}$ from N_{100} measured with FlowCAM, and to the fractions $C_{500-1000}$ from N_{200} measured with the ZOOSCAN have significantly higher abundances than other net-analysis technique combinations (P<0.000). Consequently, we combined data for $N_{100}F_{<200}$ and $N_{100}F_{200-1000}$ measured with FlowCAM to compute ESD size classes <500 um (Figure 2A) and data for $N_{200}F_{<1000}$ and $N_{200}F_{>1000}$ measured with ZOOSCAN to compute ESD size classes >500 um (see Figure 2B). The combination of these data enabled us to compute the final size spectrum (Figure 2C), that was used to estimate abundance, biomass and metabolic rates for each ESD size class, and then for the whole sample (sum of all the size classes) and for the total mesozooplankton (sum of the size classes $C_{200-300}$, $C_{300-500}$, $C_{500-1000}$ and $C_{1000-2000}$).

For the FlowCAM analyses, the sample was concentrated in a given water volume. Then, an aliquot of each sample was analyzed using FlowCAM in auto-image mode. For the fraction $N_{100}F_{<200}$, a 4X magnification and 300 μ m FOV flow cell were used and the analysis was carried out up to 3000 counted particles. For the fraction $N_{100}F_{200-1000}$ a 2X magnification and 800 μ m FOV flow cell were used and the analysis was carried out up to 1500 counted particles.

The digitalized images were analyzed using the VisualSpeadsheet® software and classified manually into taxonomic categories. Considered living organism groups for the FlowCAM were copepods, nauplii, crustaceans, appendicularians, gelatinous, chaetognaths and other diverse zooplankton groups (polychaeta, ostracods etc.). Non-organism particles were classified as detritus. Duplicates and bubbles were deleted.

To calculate the number of particles in the sample, the following equation was used.

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$$A = \frac{p_a \times V_c}{V_a \times V_s}$$

where A is the abundance (ind m^{-3}); P_a is the number of particles in the analyzed aliquot; V_c is the given volume in the concentrated sample and V_a is the volume of the analyzed aliquot and V_s is the volume of sea water sampled by the zooplankton net (m^3).

For the ZOOSCAN analyses, the sample was homogenized and split using a Motoda box until a minimum of 1000 particles were obtained. Then, for the digitalization, the subsample was placed on the glass slide of the ZOOSCAN and the organisms were manually separated using a wooden spike to avoid overlapping. After scanning, the images were processed with ZooProcess (version 7.32) using the image analysis software Image J (Grosjean et al.,2004; Gorsky et al., 2010). Particles were classified automatically into taxonomic categories using the Plankton Identifier software (http://www.obs-vlfr.fr/~gaspari/Plankton_Identifier/index.php, last access: November 2019). Then the classification was manually verified to ensure that every vignette is in the correct category. Considered living groups of organisms for the ZOOSCAN were copepods, nauplii, crustaceans, appendicularians, gelatinous, chaetognaths and diverse zooplankton (polychaeta, ostracods etc.). Non-organism particles were classified as detritus. Blurs and bubbles were deleted.

2.4 Normalized biomass size spectrum

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The size spectra were computed for each station using combined FlowCAM and ZOOSCAN data, following Suthers et al. (2006). Firstly, the data were classified in size categories of 0.1 mm of ESD from 0.2 to 2.0 mm. Zooplankton biovolume (mm³) was estimated for each category following the equation:

$$Biovolume = \frac{1}{6} \times \pi \times (ESD)^3$$

with ESD expressed in mm. The X-axis of the normalized biomass size spectrum (NBSS) was calculated by dividing the biovolume by the abundance of each category and transformed into Log10. For the Y-axis, the biovolume of each category was divided by the difference in biovolume between two consecutive categories and transformed into Log10. NBSS slope and intercept were determined using linear regression model. The slope of the NBSS reflects the balance between small and large individuals, a steeper slope corresponding to a higher proportion of small individuals (bottom-up control) and a flatter slope corresponding to a higher proportion of large individuals (top down control) (Donoso et al., 2017; Naito et al., 2019).

2.5 Zooplankton carbon demand, respiration and excretion rates

170 The zooplankton carbon demand (ZCD in mg C m⁻³ d⁻¹) was computed based on estimates of biomass from ZOOSCAN and FlowCAM samples and on estimates of growth rate:

$$ZCD = Ration \times B_{zoo}$$

where B_{zoo} is the biomass of zooplankton in mgC m⁻³, calculated using the area-weight relationships from Lehette and Hernández-León (2009) and converted to carbon assuming that carbon represent 40% of the total body dry weight (Omori and Ikeda, 1984). Ration (d⁻¹) is defined as the amount of food consumed per unit of biomass per day calculated as:

$$Ration = g_z + \frac{r}{A}$$

where g_z is the growth rate, r is the weight specific respiration and A is assimilation efficiency. g_z was calculated following Zhou et al. (2010):

$$g_z(w, T, Ca) = 0.033 \left(\frac{Ca}{Ca + 205e^{-0.125T}} \right) e^{0.09T} w^{-0.06}$$

as a function of sea water temperature (T, °C), food availability (Ca, mgC m⁻³), estimated from Chl-a, and weight of individuals (w, mg C). We consider here that food is phytoplankton following Calbet et al. (1996). Following Alcaraz et al. (2007) and Nival et al. (1975), values of r and A were 0.16 d⁻¹ and 0.7 respectively. ZCD was compared to the phytoplankton stock, converted to carbon assuming a C:Chl/a ratio of 50:1, and to primary production to estimate the potential clearance of phytoplankton by zooplankton.

Ammonium and phosphorus excretion and oxygen consumption rates were estimated using the multiple regression model by Ikeda et al. (1985) with carbon body weight and temperature as independent variables

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Where lnY represent the ammonium excretion, phosphorus excretion or oxygen consumption. a_0 , a_1 and a_2 are constant (see Ikeda et al. 1985), X_1 is the body mass (dry weight, carbon, nitrogen or phosphorus weight) and X_2 is the habitat temperature (°C).

Contribution to nutrient regeneration by zooplankton was estimated using the values of primary production and converted to nitrogen and phosphorus requirement using Redfield ratio. Respiration was converted to respiratory carbon lost assuming a respiratory quotient for zooplankton of 0.97 following Ikeda et al. (2000) and used as carbon requirement for zooplankton metabolism.

2.6 Data analysis

Spatial patterns of the environmental variables were explored using a principal component analysis (PCA). We considered temperature, salinity, dissolved oxygen and Chl-a values from a fluorescence sensor coupled with CTD, using mean values of the 0-300 m layer depth, plus the estimated MLD. The data were normalized prior to analyses performed using PRIMER v7 software (Anderson et al., 2008).

Differences in zooplankton abundance and biomass between size classes and areas were tested using two-way ANOVA. One-way ANOVA with Scheffé post-hoc analysis was applied to compare mean values between areas for total zooplankton and within each size class. Prior analyses data were log-transformed and tested for homogeneity. Dunnett's test was used in case of non-homogeneity. Potential association between univariate zooplankton and environmental data were tested using Spearman's rank-correlations. These analyses were performed with Statistica 7 Software. The 100 µm sample of station TYR was discarded from these data analyses due to poor state of preservation of the sample.

For studying the spatial patterns of zooplankton communities, a taxonomic group-station matrix with the abundance values was created and then square-root transformed to estimate station similarity using Bray Curtis similarity. The similarity matrix was then ordinated using Nonmetric Multidimenstional Scaling (NMDS). The contributions of significant taxa to the similarity or dissimilarity between stations and areas was tested using SIMPER. Then the BIOENV algorithm was used to select the environmental variables best explaining the spatial pattern observed for the zooplankton communities. PERMANOVA was used to test the differences between areas based on environmental or zooplankton multivariate data. All these analyses were performed using PRIMER v7 software (Anderson et al., 2008).

The relationships between the biological and the environmental variables were also studied by coupling multivariate analyses of two datasets. The first dataset featured the abundances of all the zooplankton taxa identified from the 200µm net samples, and the second recorded environmental variables (the same as for the PCA analysis). A factorial correspondence analysis (FCA) and a principal component analysis (PCA) were performed on these two data sets, respectively. Then the results of the two analyses were associated through a co-inertia analysis (Doledec and Chessel, 1994) performed using ADE-

4software (Thioulouse et al., 1997). Prior to the analyses, the data were log-transformed to tend towards the normality of the distributions.

Rank frequency diagrams (RFD) were created using the data from N_{200} to see differences in taxonomic composition between the samples. In order to improve the interpretation of the RFDs, first we used a method derived from Saeedghalati et al. (2017) based on the ordination of normalized rank abundance distribution. Rank-abundance matrix was created with the data standardized by the total abundance. Resemblance was measured with Bray-Curtis similarity and a cluster was created using the complete linkage criterion. Secondly, a rank abundance distribution index was estimated following Mouillot and Lepretre (2000). The RFD for each station was separated into three portions: first the ranks with relative abundance <0.5 % were discarded (rare taxa, between 0 and 30% of the taxa according to all stations; by taking <1% we would discard between 18 and 49% of the taxa) and then the two parts were fitted with a linear regressions. One part with 4 highest ranks (see Mouillot and Lepretre for the justification) and the remaining portion with the following ranks (between 15 and 23 taxa, depending on the station). The slope for both upper and lower RFD portion was calculated (p1 and p2 respectively), then the p1/p2 ratios were estimated to quantify the differences between the RFDs of all the stations.

230 **3 Results**

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3.1 Spatial patterns of environmental variables

The Principle Component Analysis (PCA) on environmental data explains 90.3 % of the total variance in the first two axes and delivers three clusters of oceanographic areas plus two distinct stations (Figure 3). The first axis (62 % of the variance) is mostly influenced by temperature and dissolved oxygen, as shown by their high correlations with the scores of the sampling points on this axis (r= 0.95 with p=0.000 and r=0.92 with p=0.000, respectively), whereas the second axis (28.3 %) is mostly influenced by MLD (r=-0.75, p=0.01), salinity (r=-0.75, p=0.001) and Chl-*a* (r=-0.57, p=0.022).

The cluster of western stations in the Algerian Basin (AB) includes ST3, ST4, ST9 and FAST which are characterized by low temperature, salinity and MLD values. The cluster located in the Tyrrhenian Basin (TB) comprises (stations ST5, ST6 and TYR) and is very close to the first group, but with lower chlorophyll-a concentrations and higher values of temperature and salinity. Eastern stations (stations ST7, ST8 and ION) located in the Ionian Basin (IB) are characterized by the highest temperature and salinity values and the lowest dissolved oxygen concentrations found during the survey. Stations 1 and 2 on the north-south transect (NS) do not cluster with any of the other stations due to deeper MLD and higher chlorophyll-a concentrations.

245 **3.2 Spatial patterns of zooplankton structure**

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Zooplankton abundance (Figure 4A) during the PEACETIME cruise ranges between 265 and 583 x 10³ ind m⁻², with an average of 372 x $10^3 \pm 84$ x 10^3 ind m⁻², and biomass (Figure 4b) from 1160 to 2170 mgDW m⁻², with an average of 1707 \pm 333 mgDW m⁻². The highest abundances are found in PB and AB, and the highest biomass in AB. The averaged total biomass in PB is lower than in AB, due to the very low contribution $C_{>2000}$ and $C_{>2000}$, but size classes from $C_{<200}$ to $C_{500-1000}$ present higher biomass values than in AB. In TB, total biomass values decrease between ST4 and ST6, 250 the latter presenting the lowest biomass value of the whole survey. Note that the biomass of TYR is obtained only for the size classes above 500 um ESD, and the corresponding abundance is comparable to those obtained in ST5 and ST6 for these larger size classes. In IB, total biomass and abundance are lower than in AB and with low variability between stations. Detritus estimated for all analyzed classes by FlowCAM and ZOOSCAN represents between 14.6 to 39.1% of the total 255 biomass. The C₂₀₀₋₃₀₀ ESD size class has the highest averaged contribution (42.9 %) to the total zooplankton abundance, followed by $C_{300-500}$ (28.5%), $C_{<200}$ (17.8 %), $C_{500-1000}$ (8,9 %), $C_{1000-2000}$ (1.7 %) and finally $C_{>2000}$ (0.22 %). In terms of biomass, $C_{500-1000}$ has the highest averaged contribution (25.3%), followed by $C_{1000-2000}$ (23.8%), $C_{300-500}$ (21.3%), $C_{>2000}$ (15.5%), $C_{200-300}$ (11.9%), and finally $C_{<200}$ µm fraction (2.1%). There is no correlation between total zooplankton abundance or biomass and integrated Chl-a, but $C_{300-500}$ biomass is negatively correlated with Chl-a (r=-0.52, p= 0.044). 260 Total abundance is negatively correlated with temperature (r=-0.67, p= 0.006) (Table 3).

Copepods are the most abundant taxonomic group at all stations (Figure 5), representing 40 to 79 % of the abundance and 32 to 85 % of the total biomass. Abundance of zooplankton smaller than 300 µm is dominated by cyclopoid and calanoid copepodites. In N₂₀₀, 51 taxonomic groups are found of which 34 are copepod genus. The adult stages of the copepod community are dominated by the genus *Para/Clausocalanus* spp. (28.7 %), *Oithona* spp. (13.7 %), *Corycaeus* spp. (6.2 %), *Oncaea* spp. (4.1 %) and undefined calanoid copepods (7.0 %). The most abundant non-copepod groups are appendicularians (5.1 %), ostracods (4.8 %) and chaetognaths (3.6 %). The highest contributions of copepods to abundance and biomass are found in PB, and then this proportion tends to decrease southwards where the abundance and biomass of the other groups such as chaetognaths and gelatinous zooplankton increase. The ratio between copepods with length smaller than 1 mm and larger than 1mm (Figure 5) ranges from 2.8 to 8.3 (5.1 on average), with maximum mean values found in TB and minimum in IB.

The two-way ANOVA shows that the PB basin is characterized by significantly lower abundance and biomass in the upper size classes (1000-2000μm and >2000μm) compared to the other areas (p<0.05). One-way ANOVA results show that both total zooplankton and mesozooplankton present significantly higher abundance in PB than in IB, whereas their total biomass was not significantly different between the areas (p>0.05). Significant differences in abundance and biomass between areas were found in the size classes C300-500, C1000-2000 and C>2000 and the biomass of C<200 (P<0.05) (Table 4).

NBSS is calculated for each station as shown in Figure 6 taking ION1 as an example. During the PEACETIME survey, the NBSS slopes (Figure 7) range between -0.60 and 1.27, with an average value of -0.80. The most negative slopes are found in

PB, whereas the IB area has the fewest negative slopes. At the long-duration stations FAST and ION, strong variations in slope values appear depending on the sampling time, with steeper slopes in the samples collected during the daytime indicating higher contributions of small zooplankton compared to large ones, and potentially linked to daily migration of larger forms deeper than 300 m.

The NMDS analysis (Figure 8) on the mesozooplanktonic taxa abundances based on N₂₀₀ delivers a distribution pattern for the stations rather similar to that of the PCA on environmental variables. ST1 and ST2 on PB are the most dissimilar stations due to the higher abundance of copepods, especially *Para/Clausocalanus* at ST1, which is twice as high as at ST2, and between 5 to 13 times higher than the rest of the transect (Figures 8A and 5). Similarly, *Centropages* spp. abundance is 10 times higher at ST1 and ST2 than at other stations of the survey. In contrast, abundances of *Oithona* spp. and *Corycaeus* spp., are respectively 6 and 10 times lower at ST1 and ST2 than at other stations. The zooplankton community in AB is slightly different from those in TB and IB due to appendicularians and unidentified calanoid copepods being more abundant in AB and to *Haloptilus* spp. being more abundant in TB and IB. Within TB and IB, the three sampling dates (ION1, ION2, ION3) at station ION form a unique cluster, whereas, ST7 and 8 are grouped with station TB in another cluster. This differentiation of ST7 and 8 from the ION sampling dates in the NMDS analysis is mainly due to differences in relative abundance of Mesocalanus spp. (more abundant), ostracoda (less abundant), Clytemnestra spp. (absent in ION) and Pontellidae spp. (absent at ST7 and 8).

The SIMPER analysis shows that the lower average similarity between the stations is in PB (64.79 %) mainly due to Para/Clausocalanus spp. The rest of the basins share a higher internal similarity 78.43 %, 79.79 % and 78.03 % for AB, TB and IB respectively. Another interesting point highlighted in the SIMPER analysis is the lower average dissimilarity between TB and ST7 and ST8 from (20.25 %), this dissimilarity increases when the comparison is made between TB and the rest of the stations included in IB (29.04 %); this is in agreement with the NMDS analysis (Figure 8) that related ST7 and ST8 with TB rather than with the stations in their basin.

300 3.3 Relationship between environmental variables and zooplankton community

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Results of the PERMANOVA analysis on the environmental variables and on diversity on taxa are summarized on the following in Table 5. Interestingly, based on the zooplankton diversity of TB and IB, their difference is more significant when ST7 and ST8 are removed from IB and placed on TB (based on the NMDS cluster, Figure 8), whereas it is not the case when considering environmental variables (see Table below). This suggests that the similarity between st7 and st8 and the TB stations is not linked to the environmental context.

The BIOENV results show that salinity and chlorophyll were the environmental variables best explaining the overall spatial distribution of zooplankton community (BIOENV; Rs = 0.657).

The first factorial plane of the Co-inertia analysis (Figure 9) explained 96% of the total variance, with 79 % due to the first axis. On both spaces ('Environment' and 'Zooplankton'), the first axis opposes the IB stations associated with high temperature and salinity values and several zooplankton taxa (namely Echinoderm larvae and some copepod taxa, ie

Pontellidae, Rhincalanus spp., Haloptilus spp. and Phaena spp.) to the PB and AB stations characterized by higher chlorophyll concentrations and by some copepod taxa (mainly Pseudodiaptomus spp., Tortanus spp. and Pleuromama spp.). On this axis, TB stations have an intermediate position, close to the coordinate zero. The second axis opposes northern (st1 and 2 of PB) and southern (AB) stations sampled in the Western Mediterranean basin. On this axis, PB stations are characterized by higher chlorophyll and salinity and deeper MLD, compared to AB and by the association with Pseudodiaptomus spp., whereas southern AB stations are associated with the copepods Heterorhabdus spp., Labidocera spp. and Euterpina spp. As in the preceding multivariate analyses, we note that St 8 and 9 from the IB tend to be closer to the TB stations than to the Ion station on the first factorial plane, particularly in the 'Zooplankton system'. The association between the environmental context and the zooplankton community is high with good correlation between the normalized scores of the stations (R2=0.844 and R2=0.820 for X1 and X2 axes, respectively), and by the positions of the plots of these stations close to the equality lines (i.e. X1 zooplankton = X1 Environment or X2 zooplankton = X2 Environment).

3.4 Zooplankton community changes linked to dust deposition events during the PEACETIME survey

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The zooplankton community changes were analyzed using the variations of RFD between samplings. The RFDs for stations TYR, ST5, ST6, ION and FAST are presented separately in Figures 10A to 10D, and grouped in Figures 10E and 10F. As only one sample was done at station TYR, nine days after a large dust deposition event in the Southern Tyrrhenian Sea, RFDs of ST5 and ST6 also sampled in TB (six and twelve days after the dust event, respectively) are added for comparison (Figures 10A and 10B). At all three TB stations, RFDs are characterized by high dominance of filter-feeding zooplankton Para/Clausocalanus spp. and Oithona spp. in 1st and 2nd position with a strong drop in abundance for the following ranked taxa (undefined calanoid copepods or Corycaeus spp.). Appendicularians drop from the 4th position at ST5 and TYR to the 10th position at ST6. The shapes of RFDs change more between ST5 and TYR than between TYR and ST6. At station ION that was not impacted by dust deposition, RFD shapes are similar at both sampling dates (ION1 and ION3) with the community dominated by Para/Clausocalanus spp. (Figure 10C). Corycaeus spp. changes from the 2nd position to the 4th. calanoid copepods from 3rd to 6th and Oithona spp. from 4th to 2nd. Appendicularians occupy a very similar position in both RFDs (6th and 7th rank at ION1 and ION3 respectively). At station FAST, the taxonomic composition is dominated by copepods (Figure 10D), but the rank order of the most dominant species changes between the two sampling dates (FAST1 and FAST3). Oithona spp. and Para/Clausocalanus spp. have the 1^{rst} and 2nd ranks during FAST1, but this order is reversed at FAST 3. The 3rd place on both days are occupied by calanoid copepods. Appendicularians present one of the most significant changes, with their rank dropping from 4th to 14th between the two dates. It is remarkable that the RFDs change from a convex shape at FAST1 to a more concave one at FAST2, influenced by the high dominance of Para/Clausocalanus at the first rank (Figure 10D). The comparison of the standardized RFDs for all the stations (Figure 10E) highlights that the greatest change in shape is visible at FAST, whereas it stays moderate at ION and negligible at TB. Figure 10F is similar to Figure 10E, but without ION, to visualize changes in zooplankton community composition at different time lags after a dust event, and will be commented on in more detail in the Discussion section.

3.4 Estimated zooplankton carbon demand, grazing pressure, respiration and excretion rates

Zooplankton carbon demand ZCD (Table 6) varies between 145.9 and 280.1 mgC m⁻² d⁻¹at ST6 and FAST1, respectively. Assuming phytoplankton as the major food source, zooplankton consumption potentially represents 15 % of the phytoplankton stock on average per day and 97 % of the primary production (see Table 6). ZCD follows the zooplankton biomass pattern with higher values in AB and lower values in TB, and does not increase with primary production (r= -0.18, p>0.05). The average respiration (mean: 83.1 mgC m⁻² d⁻¹ and range between 62.9 and 112.2 mgC m⁻² d⁻¹) corresponds to 36.4 % of the integrated primary production. Almost half of this zooplankton respiration is due to organisms smaller than 500 μm of ESD. Mean ammonium excretion is 12.3 mg NH4 m⁻² d⁻¹ (range between 9.1 and 17.7 mg NH4 m⁻² d⁻¹), and mean phosphate excretion 1.7 mg PO4 m⁻² d⁻¹ (range between 1.3 to 2.3 PO4 m⁻² d⁻¹). The potential contributions of excreted nitrogen and phosphorus to primary production are respectively 31.5 % (range between 19.9 to 42.6 %) and 26.3% (range between 19.9 to 42.6%). Zooplankton size classes smaller than 500 μm of ESD contribute 45 % and 47 % of the total ammonium and phosphate excretion respectively. The details of the rates for each size class is given as supplementary material (Table S1).

4 Discussion

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4.1 Methodological concerns and the importance of the small zooplankton fraction

This methodology combining two nets (N_{100} and N_{200}) and two sample treatments (FlowCAM and ZOOSCAN) enables us to deliver a more accurate mesozooplankton community size spectrum (200-2000 µm), whereas size classes $C_{<200}$ and $C_{>2000}$ at the edges of the spectrum range remain under-sampled and require other equipment for proper sampling (respectively bottles and larger mesh size net). The length:width ratio of mesozooplankton organisms is quite variable, from 1 for the nearly round-shaped organisms such as nauplii or cladoceran, to more than 10 for long organisms such as chaetognaths (Pearre, 1982) or some copepods such as *Macrosetella gracilis* (Böttger-Schnack, 1989), with an average value between 3 and 4 for copepods (Mauchline, 1998). If we consider that organisms with a length:width ratio of 6 caught by the 200 µm mesh size will present an ESD of at least 490 µm, it is consistent that this net quite correctly samples organisms having an ESD above 500 µm ESD. For these organisms (> 500 µm ESD), ZOOSCAN is the most appropriate tool to deliver the size spectrum. Similarly, the 100 µm mesh size net allows small organisms of width just below 100 µm to pass through, but most of them might have an ESD up to 200 µm because for these smaller sizes, the length:width ratio is mostly below 4 (Mauchline, 1998). Due to the threshold of ZOOSCAN at 300 µm ESD, FlowCAM is the best tool to process organisms in the fraction below 500 µm.

Several authors have already highlighted the limitation of the 200 µm mesh size to catch small zooplankton individuals. Comparisons of different zooplankton mesh size nets comprised between 60 and 330 µm have systematically shown a decrease in abundance with increasing mesh size (Turner, 2004; Pasternak et al., 2008; Riccardi, 2010; Makabe et al., 2012; Altukhov et al., 2015). When the goal of the study is to achieve a full understanding of the complete mesozooplankton

community structure and functioning, the size selectivity of the sampling nets is an important issue: clearly, a large fraction of organisms of ESD between 200 and 500 μ m is undersampled using a single 200 μ m mesh size net. Pasternak et al. (2008) reported that a 220 μ m mesh can lose up to 98% of the abundance of *Oithona* spp. and 80% of copepodite stages of *Calanus* spp. Riccardi (2010) found that a classical 200 μ m net catches only 11% of the abundance and 54 % of the biomass compared to a 80 μ m mesh size, leading also to differences in observed species composition in the Venice lagoon. During the PEACETIME survey, the small size classes ($C_{200-300}$ and $C_{300-500}$) of mesozooplankton have been optimally sampled using a 100 μ m mesh size net (N_{100}). Consequently, these size classes represent very large percentages of the total abundance (respectively 52.3 and 34.8 %) and a significant contribution to the total biomass (respectively 14.5 and 25.9 %). These reliable estimations have direct consequences for the estimated fluxes (see below).

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4.2 Differences in abundance, biomass and zooplankton community structure in relation to regional environmental characteristics

A review of the most relevant information available on zooplankton biomass and abundance in different regions of the Dediterranean Sea (Table 7) shows a wide range of variation that can be attributed to location, sampling seasons and/or sampling methods (net mesh size, depth of tow, etc), and in general, the values during the PEACETIME survey are of the same order of magnitude, although most of other studies were performed with a 200 µm mesh size net and often over a shallower surface layer. However, during this post-bloom period, no clear regional patterns in abundance and biomass were found, unlike other descriptions showing a north-south and west-east decrease in zooplankton stocks (Dolan et al., 2002, Siokou-Frangou, 2004). In PB, Donoso et al. (2017) and Nival et al. (1975) highlighted a strong variability which is consistent with the strong gradient found between ST1 and ST2 during PEACETIME (see Figure 4). In AB, abundance and biomass values obtained during the survey are similar to those recorded in late spring by Nowaczyk et al. (2011), whereas Riandey et al. (2005) found lower abundance and higher biomass values. However, the latter study focused on high resolution of a mesoscale eddy highlighting an important fine-scale variability of abundance and biomass values. For TB, the data are difficult to compare due to different sampling conditions (net mesh size, depth of tow and sampling season). In IB, all biomass values presented in Table 7 are of the same order, but abundances found by Mazzocchi et al. (2003, 2014) are three times lower than those observed during PEACETIME, probably due to a high contribution of $C_{<200}$ and $C_{200-300}$ obtained with N₁₀₀ (see Figure 4). In general, the better sampling of small size classes with N₁₀₀ should lead to higher abundance values. However, the comparison of data in Table 7 shows that regional and temporal variability of these values partially masks this benefits.

In PEACETIME, clear regional differences are found both in terms of environmental variables and zooplankton taxonomic composition. ST1 and ST2 are clearly differentiated from all the others with deeper MLD, higher chlorophyll-a concentrations and a zooplankton community dominated by typical herbivorous copepods of PB (*Centropages*, *Para/Clausocalanus*, *Acartia*, etc), as mentioned by Gaudy et al.(2003) and Donoso et al. (2017), and characterized by a scarcity of thaliaceans which normally occurs in ephemeral and aperiodical patches (Deibel and Paffenhöfer, 2009). AB and

TB are very closely related to each other in terms of hydrological features and chlorophyll-a, but slightly differentiated in salinity and zooplankton taxonomy, probably because they are both strongly influenced by the Modified Atlantic Water (MAW) and its associated mesoscale features (Millot and Taupier-Letage, 2005). In AB, 17 days separated the sampling of ST3 and ST4 with that of ST9 and FAST, but despite this time gap, they are very close in terms of hydrological features, chlorophyll-a level and zooplankton community structure. IB is clearly differentiated from these groups in terms of environmental parameters (see Figure 3) due to higher salinity and lower chlorophyll-a, but in terms of zooplankton community the western Ionian stations (ST7 and ST8) present more analogy with TB than with the ION station (see Figure 8). During PEACETIME, the station ION appears clearly separated from ST7 and ST8 located further westwards by a northsouth jet (ADCP and MVP observations, Berline et al., in preparation), which might correspond to the Mid-Mediterranean Jet (Malanotte-Rizzoli et al., 2014, their Figure 5). The location of ST7 and ST8 within anticyclonic structures of the portion of the Modified Atlantic Water (MAW) flowing through the Sicily Channel could explain their similarity to TB stations in terms of zooplankton assemblages, as TB is directly influenced by the main part of the MAW flowing through the Sardinia Channel. Ayata et al. (2018) also classified the Tyrrhenian Sea as heterogeneous due to complex circulation patterns including transient hydrodynamic structures in the south, which could also explain the similarity of ST7 and ST8 to TB stations in terms of zooplankton assemblages during PEACETIME. This visited area of the IB during PEACETIME certainly represents a transition area between the eastern and western Mediterranean basins (Siokou-Frangou et al., 2010; Mazzocchi et al., 2003).

These regional differences highlighted both in terms of environmental characteristics and zooplankton taxa assemblages are in agreement with the regionalization of the Mediterranean basin by Ayata et al. (2018) based on historical biogeochemical, biological and physical data of the epipelagic zone. For example, ST1 of PEACETIME characterized by high Chl-a, high zooplankton abundance and dominance of small copepods is clearly located in the 'consensual Ligurian Sea Region' sensu Ayata et al. (2018), identified as the most productive of the Mediterranean due to intense deep convection events. Among AB stations, stations 3, 4 and 9 are clearly in the 'consensual Algerian region' (Ayata et al., 2018), whereas station FAST corresponds to the 'western Algerian heterogeneous region'. Among the IB stations, the separation of stations 7 and 8 from the ION stations in terms of zooplankton communities and, to a lesser extent, of environmental variables, also corresponds to the distinction between the 'consensual North Ionian' region and the western part of the 'Ionian Sea region', considered as a heterogeneous region (Ayata et al., 2018).

4.3 Estimated zooplankton-mediated fluxes during the PEACETIME survey

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By using allometric relationships relating zooplankton grazing and metabolic rates to size structure, zooplankton impacts (top-down vs. bottom-up) on primary production have been investigated. We are aware that using constant conversion factors may limit the analysis of the spatial variation, since these factors may display temporal and geographical variations (Minutoli and Guglielmo, 2009). However, our sampling strategy based on a limited number of stations sampled did not

enable us to consider temporal and spatial variations accurately, and our main goal was to have rough estimations of the epipelagic zooplankton mediated fluxes at the scale of the PEACETIME cruise.

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ZCD estimations show that zooplankton required 15 % of the daily phytoplankton stock, with narrow variations over the whole area (between 9.5 to 19.3), which are twice lower than the values estimated by Donoso et al. (2017) during the spring bloom in the North-Western Mediterranean Sea. However, estimated grazing rates are of the order of the estimated primary production, which corresponds to the highest range of the values summarized by Siokou-Frangou et al. (2010) for the whole Mediterranean Sea (from 14 to 100 %), Just estimating ZCD on the basis of mesozooplankton alone certainly leads to overestimation of its top-down impact on phytoplankton. In the Mediterranean Sea, the primary production is consumed by a "multivorous web" including microbial and zooplankton components (Siokou-Frangou et al., 2010). Mesozooplankton simultaneously grazes on phytoplankton and heterotrophic prey, such as heterotrophic dinoflagellates (Sherr and Sherr, 2007) or ciliates (Dolan et al., 2002), and might be quite flexible in its feeding strategy depending on the composition and size of prey as well as on environmental variables such as turbulence (Kleppel, 1993; Yang en al., 2010). On one hand, a large part of the primary production can be consumed by ciliates (Dolan and Marrasé et al., 1995), but on the other hand mesozooplankton can consume almost the entire ciliate production (Pitta et al., 2001; Pérez et al., 1997; Zervoudaki et al., 2007), potentially explaining the wide variations of standing stock of ciliates over the Mediterranean Sea (Dolan et al., 1999; Pitta et al., 2001; Dolan et al., 2002). The extensively described east-west pattern of decreasing grazing impact (Siokou-Frangou et al., 2010) could not be observed during this study as only one station (ION station) was typical of the Eastern Mediterranean Sea.

Estimated NH₃ and PO₄ excretion rates by mesozooplankton during PEACETIME are consistent with the few observations collected in the Mediterranean Sea (Alcaraz, 1988; Alcaraz et al., 1994; Gaudy et al., 2003) and with those obtained at similar latitudes (see review in Hernández-León at al., 2008). From our estimation, zooplankton excretion would contribute respectively to 21 - 44 % and 17 - 38 % of the N and P requirements for phytoplankton production. In the NWMS, Alcaraz et al. (1994) estimated a zooplankton nitrogen excretion contribution to primary production > 40%, whereas Gaudy et al. (2003) reported 31-32 % and 10-100 % N and P contributions. This impact on phytoplankton production can be even greater in proximity to the DCM where zooplankton tends to aggregate fuelling regenerated production (Saiz and Alcaraz, 1990) and enhancing bacterial production (Christaki et al., 1998). Zooplankton grazing impact and nutrient contribution to primary production are higher in the western basin than in the Ionian Sea, mainly linked to variations of zooplankton biomass.

Mean carbon released through zooplankton respiration represents 36 % of the primary production during PEACETIME, which is higher than previous measurements in NWMS (by Alcaraz, 1988 and Gaudy et al., 2003) from onboard incubation experiments on zooplankton collected with a 200 µm mesh size net.

Metabolic estimations clearly show that the size fractions $< 500 \mu m$ (optimally captured with the 100 μm mesh size net) make a significant contribution to the whole mesozooplankton estimated fluxes: 14.9 % of the ZCD is due to organisms $< 300 \mu m$, and this size class contributes 21 % and 20 % of the total ammonium and phosphate excretion, respectively.

4.4 Impact of dust deposition on the zooplankton community

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475 In the past years, responses to Saharan dust inputs in marine systems have been mostly studied in microcosm and mesocosm experiments, but more rarely observed in situ. Most studied responses to dust are focused on the microbial biota and are generally marked by an increase in metabolic rates rather than by standing stock changes (probably due to trophic transfer along the food-web) (Ternon et al., 2011; Guieu et al., 2014; Ridame et al., 2014; Herut et al., 2016). In mesocosms, changes in zooplankton stocks are strongly dependent on the initial conditions, and cannot really reflect what could occur in natural 480 waters within the Mediterranean "multivorous planktonic food-web" (Siokou-Frangou et al., 2010). Pitta et al. (2017) found an increase in mesozooplankton biomass 9 days after the beginning of a mesocosm experiment, probably as a result of an earlier increase of prey (flagellates, ciliates and dinoflagellates). Tsagaraki et al. (2017) described an increase in productivity after an artificial dust deposition that was transferred to higher trophic levels by the classical food web, resulting in an increase of copepod egg production 5 days after the beginning of the experiment. Very few in situ studies have documented 485 mesozooplankton responses to Saharan dust. Abundance increase was observed by Thingstad et al. (2005) in the Eastern Mediterranean Sea, and by Hernández-León et al. (2004) in Atlantic waters close to the Canary Islands one week after the deposition. In this latter area, Franchy et al. (2013) detected increases of zooplankton grazing and zooplankton biomass after another event. Thus, the PEACETIME survey dedicated to the tracking of such events was an opportunity to observe real in situ responses.

At station FAST (an opportunistic station after a Saharan dust deposition event), an increase in nitrate (from 50 nM to 120 nM) and phosphate concentrations (from 8 nM to 16 nM) occurred in the mixed layer (pers. comm. C. Guieu), which led to an increase in primary production from FAST1 to FAST3, but with no visible changes in phytoplankton biom see Table 2). For zooplankton, the total abundance slightly decreases but the community composition presents obvious changes, mainly a decrease of appendicularians and an increase of *Para/Clausocalanus* spp. and of carnivorous taxa (*Candacia* spp., chaetognaths, siphonophores) (see Figure 10D). The sharp decrease of appendicularian abundance (four-fold decrease) and rank position (see Figure 10D) could potentially be linked either to food limitation or to predation. Size and species composition of the phytoplankton community in FAST suggest a change toward larger cells (Table 2) poorly edible by appendicularians and inducing filter clogging. There were also potential increases in food competition with Para/Clausocalanus spp. (Lombard et al., 2010) and/or in predation by chaetognaths and siphonophores (Purcell et al., 2005). Although total zooplankton biomass remains relatively stable at FAST, the contribution of the size classes $C_{500-1000}$ and C₁₀₀₀₋₂₀₀₀ increase relative to the smaller size classes (see Figure 4B) inducing variations on the NBSS slope from -0.76 to -0.63 (see Figure 6). This 15% increase in biomass is mainly due to large migrating taxa such as copepods *Eucalanus* spp., Rhincalanus spp. and Candacia spp., chaetognaths and siphonophores. The daily observation of sediment traps at 200 and 500 meters over five days between FAST1 and FAST ers.comm. C. Guieu) shows a relative increase of swimmers collected at 500 m versus those collected at 200 m, also suggesting increasing numbers of migrants. An obvious planktonic transition occurred during this period but it is difficult to conclude which of the bottom-up (changes in primary producers) or top-down (increase of carnivorous migrants) effects was dominant. The change in the RFDs (Figure 10D), from a convex shape at FAST1, indicating a more stable system with no dominance of the first taxonomic groups, to a more concave shape at FAST3 influenced by the high dominance of *Para/Clausocalanus* at the first rank, could reflect a disturbance effect (*sensu* Pinca and Dallot, 1997) of the dust deposition on the zooplankton community.

A synoptic analysis of the RFDs linked to the dust events observed in the Tyrrhenian basin and at station FAST offers a basis for proposing a conceptual model of a virtual time series of zooplankton community responses after a dust deposition event (Figure 10F): the first sampling is carried out before the event (FAST1), and several other samplings are done with a time-lag of five days (FAST3), six days (ST5), nine days (TYR) and twelve days (ST6) after the event. FAST1 represents an initial steady state (state 1) with no dominance in the first taxa ranks, while FAST3 and ST5 represent a disturbed state of the community (state 2) with strong dominance of the first taxa and the collapse of the following ones. TYR and ST6 represent the beginning of recovery towards a stable system (state 3) with the move up of the second rank. State 1 before the dust event is characterized by oligothropic conditions with low nutrients, low phytoplankton concentration dominated by small-size cells and their typical zooplankton grazers (e.g. appendicularians and thaliaceans), leading to a convex RFD shape (like FAST1, Figure 10F) reflecting a mature community (sensu Frontier, 1976). State 2 is characterized by a nutrient input linked to the dust event stimulating larger phytoplankton cells and their herbivorous grazers (copepods) and attracting carnivorous migrants leading to a more concave RFD shape (like FAST3, ST5 and TYR, Figure 10F) typical of a disturbed community (sensu Frontier, 1976). State 3 is characterized by the diversification of herbivorous taxa leading to changes in RFD towards a convex shape (like ST6, Figure 10F).

The cluster analysis on the RFDs (Figure 11A) is in agreement with this succession of the time series (Figure 10F) by grouping the stations according to impact level of the wet dust deposition. It separates the initial condition (FAST1) from the most disturbed state (stations FAST3 and ST6) and identifies a transition phase before (FAST2) and after (TYR and ST6) the peak disturbance. The changing trends in p1/p2 ratios (Figure 11B) show an interesting development, with a sharp increase until day 5 after the dust deposition and a progressive decrease towards the end of the virtual time series. The linear regression suggests that the community structure will deliver a p1/p2 ratio value similar to the initial value of the time series after 22 days. Is interesting to note that this delay corresponds to an average generation time of zooplankton organisms for this region. Interestingly, in the Co-inertia analysis the stations impacted by the dust (FAST and TB stations) are grouped on the left side of the relationship between X2 axis of environment and zooplankton. In addition, their succession in this graph is consistent with the sequence observed in the virtual time series of RFD (with FAST1 as the initial station before the dust deposition and TYR and St6 corresponding to day 9 and 12 after the dust event) showing the coupled impact of dust on both environment and zooplankton.

5 Conclusion

540 before and soon after natural Saharan dust deposition events and to highlight *in situ* zooplankton responses in terms of community composition and size structure. Our study suggests that a complete understanding of the mesozooplankton community response to a single massive dust event would require continuous observation over two to three weeks, from an initial state just before the event to a complete process of zooplankton community succession after the event. To identify such a succession, the rank-frequency diagrams of the zooplankton taxonomic structure appear to be a more practical and sensitive index than observable changes in stock (abundance and biomass) or in metabolic rates, and should be further tested. Particularly the changes of the p1/p2 ratio might characterize the response of the zooplankton community to a pulse of dust (or any massive disturbance) and its resilience capacity after the forcing event.

This approach requires a complete overview of mesozooplankton size spectrum and community composition which was achieved in our study by combining data from two mesh size nets (100 and 200 μ m) and two analytical techniques (FlowCAM and ZOOSCAN). In our study, this strategy also enabled us to show the importance of small forms (< 500 μ m of ESD) both in terms of stocks and fluxes.

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Data availability

All data and metadata will be made available at the French INSU/CNRS LEFE CYBER database (scientific coordinator: Hervé Claustre; data manager, webmaster: Catherine Schmechtig). INSU/CNRSLEFE CYBER (2020)

570 Authors contribution

GF, MP and FC wrote the paper with contributions by PH. GF participated in the sample treatment. GF, FC, MP and PH participated in the data analysis

Competing interests

The authors declare that they have no conflict of interest.

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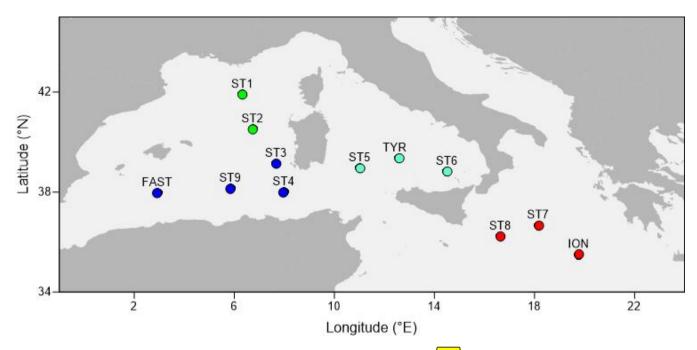


Figure 1: Map with the sampling points during PEACETIME cruise 2017. The considered in the course of the study. Green dots: Provencal basin (PB); Dark brue dots: Algerian Basin (AB); Light blue dots: Tyrrhenian Sea (TB); Red dots: Ionians Basin (IB).

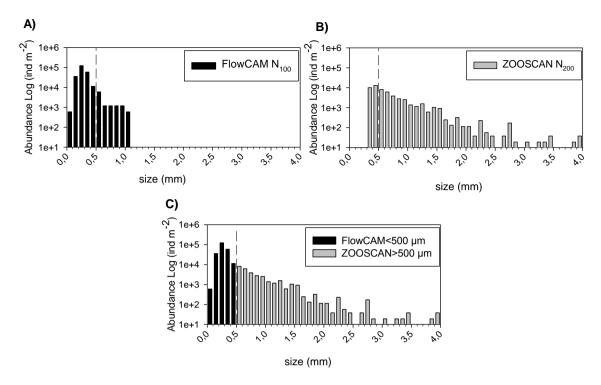


Figure 2: Size spectrum of ST ION1 as an example obtained by: A) FlowCAM (N_{100}) , B) ZOOSCAN (N_{200}) and C) combination of FlowCAM $(N_{100}$ counting only zooplankton smaller than 500 μ m of ESD) and ZOOSCAN (N_{200}) counting only zooplankton bigger than 500 μ m of ESD)

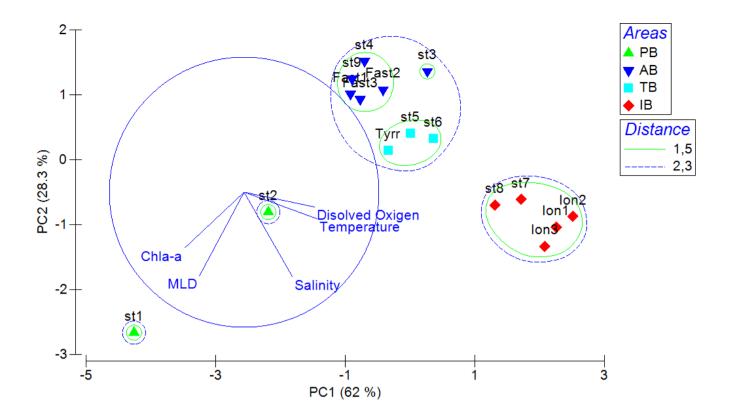
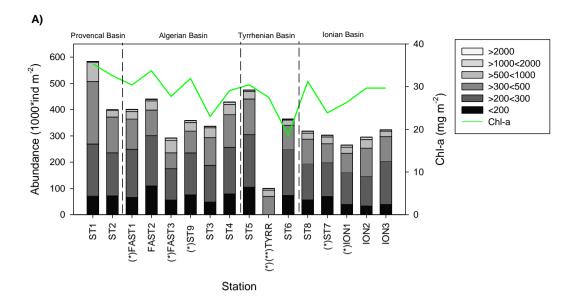


Figure 3: Principal Component Analysis (PCA) ordination of five environmental indicators: Mixed/layer Depth (MLD), Integrated values of Chl-a concentration, mean values on the upper 0/300 m of temperature, salinity and dissolved oxygen. AB: Algerian Basin, PB: Provencal Basin, TB: Tyrrhenian Basin, IB: Ionian Basin.



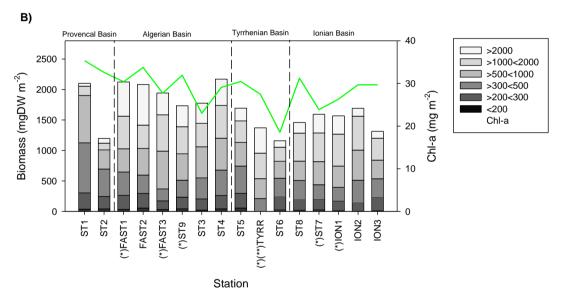


Figure 4: Values of zooplankton abundance (A) and biomass (B) cumulated by ESD size classes across different stations of the PEACETIME cruise. Integrated Chl-a concentrations (green line). (*)Stations sampled during the night. (**) At station TYR, only the abundance and biomass values above 300 µm are presented.

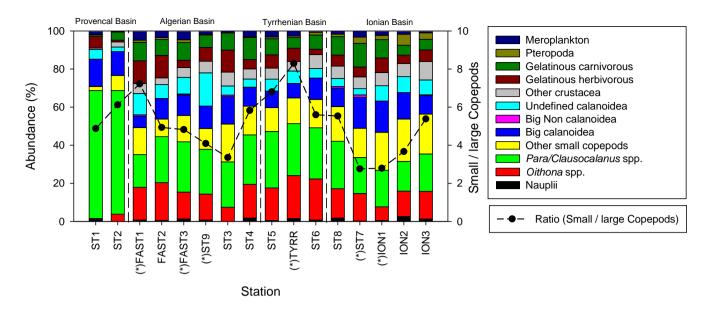


Figure 5: spatial variation of taxonomic groups (stock bars) and small (length< 1 mm)/large (length> 1 mm) copepod ratio (dashed line). (*)Stations sampled during the night.

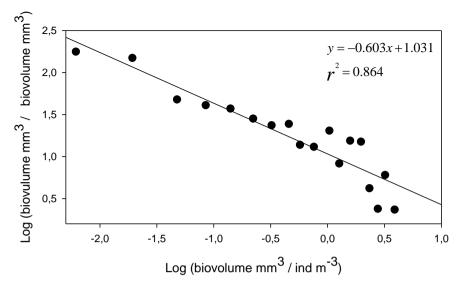


Figure 6: Normalized biomass size spectrum (NBSS) of mesozooplankton at Station ION1. Normalized biomasses in the successive size classes (black dots) and lineal regression (straight line) giving the slope value.

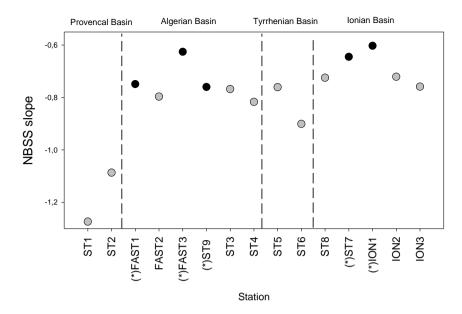


Figure 7: NBSS slope values of mesozooplankton obtained for all stations during the PEACETIME survey Black dots (night samples) and grey dots (day samples)

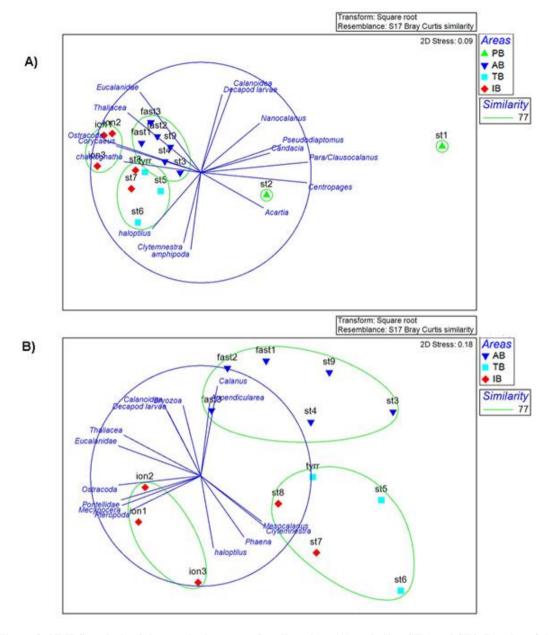


Figure 8: NMDS analysis of the zooplankton taxa for all stations (A) excluding ST 1 and ST2 (B): plot of the stations and the taxa correlated at >0.65 with the axes. Colour of the stations represents the areas identified by the PCA in the environmental analysis (see Fig. 2). This analysis was performed on the zooplankton collected with the data from N_{200} . PB: Provencal Basin, AB= Algerian Basin, TB = Tyrrhenian Basin, IB = Ionian Basin.

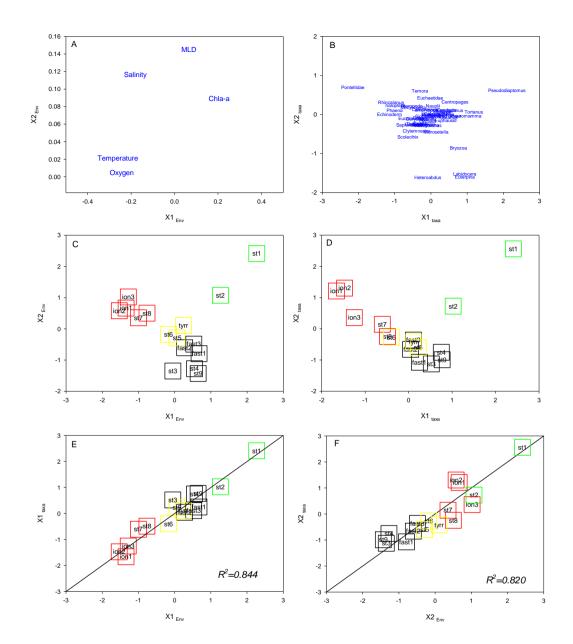


Figure 9: Co-inertia analysis. Ordination on the plans (1, 2) of the environmental variables (A) and the abundance of the zooplankton taxa (B) and of the stations in the 'Environment system' (C) and in the 'Zooplankton system' and plots of the stations on the first (C) and second (D) axes of the two systems. The line represents the equality between the coordinates on the two systems. Coloured squares identify the different regions: green = PB, black = AB, yellow = TB and red = IB.

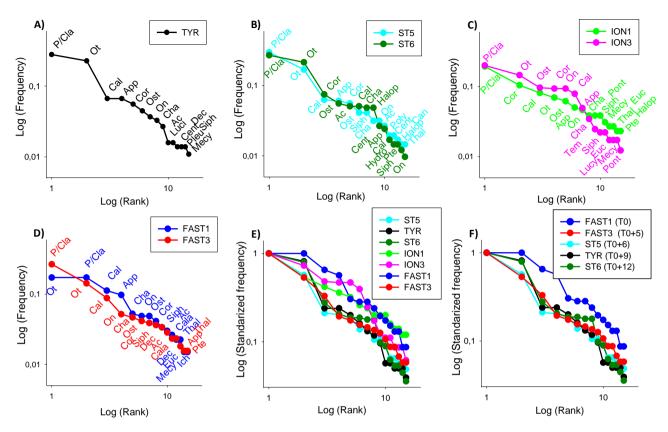


Figure 10: Rank frequency diagram at stations TYR (A), ST5 and 6 (B)ION (C) FAST (D) and Log standardized frequency for all stations (E) and stations influenced by dust deposition (F). Ac: Acartia spp.; Cal: Calanoid copepods; Cala: Calanus spp.; Cent: Centropages spp.; Cor: Corycaeus spp.; Euc: Eucalanus spp.; Halop: Haloptilusspp; Luci: Lucicutia spp.; Mecy: Mecynocera spp.; On: Oncaea spp.; Ot: Oithona spp.; P/Cla: Para/Clausocalanus spp.; Pleu: Pleuromamma spp.; Pont:Pontellidae; Tem: Temora spp.; App: Appendicularia; Cha: Chaetognatha; Dec: Decapods; Hydro: Hydrozoans; Ich: Ichtyoplankton; Ost: Ostracods; Poly: Polychaeta; Pte: Pteropods; Siph: Siphonophores; Thal: Thaliaceans.

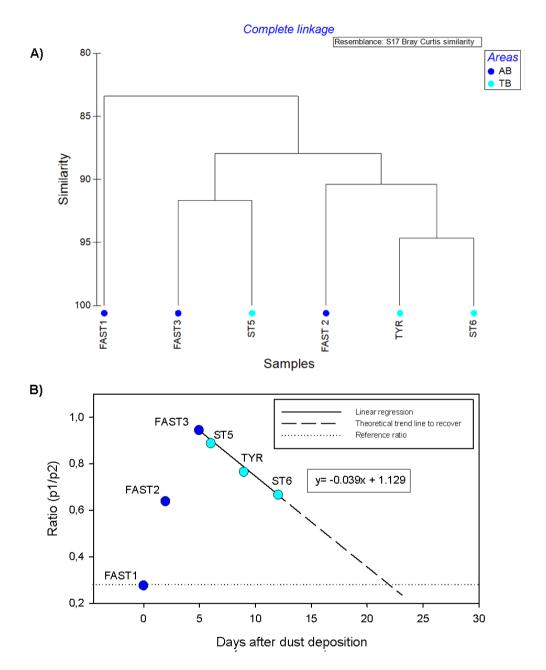


Figure 11. Cluster analysis on rank frequency diagrams (A) and changing trends in the p1/p2 ratio (B) on the stations impacted by wet dust deposition.

Table 1. Sampled stations during the PEACETIME survey: geographical information, date and time of zooplankton net sampling.

48. Algerian Basin; PB: Provencal Basin; TB: Tyrrhenian Basin; IB: Ionian Basin.

				Date	Time
Station ID	Area	lat (N)	long (E)	(DD/MM/YYYY)	(HH:MM)
St1	PB	41°53,51	6°20,00	12/05/2017	11:30
St2	PB	40°30,37	6°43,79	13/05/2017	9:30
St3	AB	39°8,00	7°41,01	14/05/2017	9:15
St4	AB	37°58,99	7°58,61	15/05/2017	9:15
St5	TB	38°57,19	11°1,40	16/05/2017	7:05
TYR	TB	39°20,39	12°35,57	19/05/2017	23:00
St6	TB	38°48,46	14°29,98	22/05/2017	10:15
St7	IB	36°39,49	18°9,29	24/05/2017	2:00
ION1	IB	35°29,38	19°46,51	26/05/2017	21:59
ION2	IB	35°29,38	19°46,51	27/05/2017	8:50
ION3	IB	35°29,38	19°46,51	28/05/2017	8:45
St8	IB	36°12,62	16°37,86	30/05/2017	9:05
St9	AB	38°8,08	5°50,45	01/06/2007	23:00
FAST1	AB	37°56,81	2°54,99	04/06/2017	22:15
FAST2	AB	37°56,81	2°54,99	06/06/2017	9:50
FAST3	AB	37°56,81	2°54,99	08/06/2017	23:45

Table 2. Overview of the main characteristics of the wet dust events occurring during PEACETIME. Zooplankton sampling was carried out very close to a CTD cast except at FAST2 where the sampling was done between two casts respectively 9 hours after the first cast ^(a) and 16 hours before the second



Stations impacted by dust and cruise visit duration	Crusie strategy with regard to dust events	Dates, geographical characteristics and intensity of the dust events predicted by the model and by observations	Zooplankton sampling Date	Iron in aerosol ng m ^{.3}	Nutrients below the nutricline NO3 (PO ₄ (Surface Primary production mg C m ⁻³ d ⁻¹	Water column (0-250) average Chi-a concentration mg m ⁻³	Depth range of the DCM strata (m)	Mean concentration of ChI-a on DCM strata mg m ⁻³	Ratio fluorecence phytoplanktor F _{micro} : F _{nano} : F _{picc} within the DCM strata
		From 3 to 5 June; Impacted area: Betwen Baleares and Algerian coast; Predicted flux from models: 0.5 g m² (Guieu et al., accepted, Supp Info figure SI5);	FAST1: 04-06-2017	245.3	224 246	2.44	0,12	60-90	0,42	27:45:28
Wet dust event FAST 02 to 07 June	position detemined on board acording to metereologial event	FAST schedule and On-board atmospheric dust deposition observations confirmed a on determined on weak wet dust deposition g m² (Guieu et al., accepted). In sediment tra	FAST2: 06-06-2017	266.0	808 239	2.85	0,14 ^(a)	60-100 ^(a) 70-90 ^(b)	0,38 ^(a)	25:43:32 ^(a) 50:30:20 ^(b)
		Water column observations (nutrients, trace Wan Wambeke et al., in prep, Tovar-Sánchez et al. 2020, Bressac et al., in prep) show a clear imprint of the atmospheric deposition.	FAST3: 08-06-2017	44.9	135 113	2.04	0,10	70-90	0,42	20:49:31
Wet dust	TB stations schedule before	From 10 to 12 may; Impacted area: whole southern Tyrrhenian sea;	ST5: 16-05-2017	57.3	841 148	1.68	0,12	70-80	0,55	21:48:30
event Tyrrhenian 16	the cruise. Model predicted a dust event 6 days before	Predicted flux from models: >1 g m ⁻² (Desboeufs et al. in prep) Dust event was confirmed by alumimium, iron and Lithogenic Si measured (sedin ent tramps at TYR with a lithogenic	TYR: 19-05-2017	162.3	n.d 127	1.77	0,11	70-80	0,61	33:40:27
to 22May	to 22May the arrival	flux betweend 1000m was 150-200 mg m ⁻² (Bressac et al., in prep.)	ST6: 22-05-2017	189.8	488 136	1.66	0,07	70-80	0,36	7:44:49
References of the data	Dulac (pers.com) Desboeufs et al. (in prep) Guieu et al. (accepted) .	Desboeufs et al. (in prep) Guieu et al. (accepted) Bressac et al. (in prep) Tovar-Sánchez et al. (2020) van Wambeke et al. (in prep)		Tovar-Sánchez et al. (2020)	van Wambeke et al. (in prep)	E. Maranon and M. Perez- Lorenzo	J.Uitz, C. Dimier	J.Uitz, C. Dimier	J.Uitz, C. Dimier	J.Uitz, C. Dimierl

Table 3. Summary table of Spearman rank correlations. T° = temperature; Sal= salinity; Chl-a= Chlorophyll; MLD= Mix layer depth; pp= primary production. Bold characters indicate significant Rs value (p<0.05)

			Correlation o	oeff.		
<mark>Abundance</mark>	<mark>7°</mark>	<mark>Sal</mark>	<mark>Chl-a</mark>	<mark>DO</mark>	<mark>MLD</mark>	<mark>PP</mark>
C<200	<mark>-0,49</mark>	<mark>-0,43</mark>	<mark>0,32</mark>	<mark>-0,61</mark>	<mark>-0,16</mark>	<mark>-0,37</mark>
C ₂₀₀₋₃₀₀	<mark>-0,58</mark>	<mark>-0,37</mark>	<mark>0,48</mark>	<mark>-0,58</mark>	<mark>0,08</mark>	-0,24
C ₃₀₀₋₅₀₀	<mark>-0,51</mark>	<mark>-0,19</mark>	<mark>0,52</mark>	<mark>-0,45</mark>	<mark>0,21</mark>	<mark>-2,28</mark>
C ₅₀₀₋₁₀₀₀	<mark>-0,56</mark>	<mark>-0,50</mark>	<mark>0,23</mark>	<mark>-0,49</mark>	<mark>-0,06</mark>	<mark>0,05</mark>
C ₁₀₀₀₋₂₀₀₀	<mark>0,29</mark>	<mark>0,01</mark>	<mark>-0,28</mark>	<mark>0,33</mark>	<mark>-0,34</mark>	<mark>0,35</mark>
C>2000	<mark>-0,12</mark>	<mark>-0,53</mark>	<mark>-0,15</mark>	<mark>0,08</mark>	<mark>-0,50</mark>	<mark>-0,16</mark>
Total abundance	<mark>-0,67</mark>	<mark>-0,44</mark>	<mark>0,56</mark>	<mark>-0,68</mark>	<mark>0,08</mark>	-0,28
<u>Biomass</u>						
C _{<200}	<mark>-0,61</mark>	<mark>-0,48</mark>	<mark>0,42</mark>	<mark>-0,71</mark>	<mark>-0,08</mark>	-0,36
C ₂₀₀₋₃₀₀	<mark>-0,52</mark>	<mark>-0,29</mark>	<mark>0,52</mark>	<mark>-0,51</mark>	0,12	<mark>-0,14</mark>
C ₃₀₀₋₅₀₀	<mark>-0,49</mark>	<mark>-0,18</mark>	<mark>-0,53</mark>	<mark>-0,46</mark>	<mark>0,19</mark>	-0,27
C ₅₀₀₋₁₀₀₀	<mark>-0,45</mark>	<mark>-0,43</mark>	<mark>0,17</mark>	<mark>-0,41</mark>	<mark>-0,11</mark>	<mark>0,14</mark>
C ₁₀₀₀₋₂₀₀₀	<mark>0,24</mark>	<mark>-0,05</mark>	<mark>-0,37</mark>	<mark>0,32</mark>	<mark>-0,39</mark>	<mark>0,30</mark>
C _{>2000}	<mark>-0,18</mark>	<mark>-0,61</mark>	-0,10	<mark>-0,02</mark>	<mark>-0,53</mark>	-0,10
total biomass	<mark>-0,58</mark>	<mark>-0,62</mark>	<mark>0,24</mark>	<mark>-0,43</mark>	<mark>-0,27</mark>	<mark>0,08</mark>

Table 4: Results of the one-way ANOVAs performed to test differences between areas (PB, AB, IB and TB) in abundance and biomass data for the different zooplankton size classes, for total zooplankton (cumulative of all size classes) and for mesozooplankton (ESD between 200 and 2000 μm) between the areas. Significant p-value <0.05 are marked in bold. *ns*= non-significant difference. Values of F and p in italic mark where Dunnett's test was used. In the post-hoc analysis homogeneous group with the lowest and highest values are noted with "a" and "b" respectively. PB= Provencal basin, AB= Algerian basin, TB= Tyrrhenian basin, IB= Ionian basin.

9	3	5

	Abundance					Biomass						
			S	heffé p	ost-hoc	2			Sl	neffé po	st-hoc	<u> </u>
Size class	f	P	<mark>PB</mark>	<mark>AB</mark>	TB	<mark>IB</mark>	F	p	<mark>PB</mark>	<mark>AB</mark>	TB	IB
C200	3.19	0.067	<mark>ns</mark>	<mark>ns</mark>	<mark>ns</mark>	<mark>ns</mark>	<mark>3.64</mark>	0.048	<mark>a</mark>	a	a	<mark>a</mark>
C200-300	3.46	0.055	<mark>ns</mark>	<mark>ns</mark>	<mark>ns</mark>	<mark>ns</mark>	2.55	0.109	<mark>ns</mark>	<mark>ns</mark>	<mark>ns</mark>	<mark>ns</mark>
C300-500	<mark>4.4</mark>	0.029	<mark>b</mark>	<mark>ab</mark>	<mark>ab</mark>	a	5.03	0.020	<mark>b</mark>	<mark>a</mark>	<mark>ab</mark>	<mark>a</mark>
C500-1000	<i>3.01</i>	<u>0.076</u>	<mark>ns</mark>	<mark>ns</mark>	<mark>ns</mark>	<mark>ns</mark>	1.75	0.214	<mark>ns</mark>	<mark>ns</mark>	<mark>ns</mark>	<mark>ns</mark>
C1000-2000	14.77	0.000	<mark>a</mark>	<mark>b</mark>	<mark>ab</mark>	b	17.87	0.000	<mark>a</mark>	<mark>b</mark>	<mark>ab</mark>	b
C>2000	9.25	0.002	<mark>a</mark>	<mark>b</mark>	<mark>ab</mark>	<mark>ab</mark>	11.63	0.001	<mark>a</mark>	<mark>b</mark>	<mark>a</mark>	<mark>a</mark>
Total	5.51	0.015	<mark>b</mark>	<mark>ab</mark>	<mark>ab</mark>	<mark>a</mark>	3.2	<u>0.066</u>	<mark>ns</mark>	<mark>ns</mark>	<mark>ns</mark>	<mark>ns</mark>
Total mesozooplankton (200- 2000 μm)	5.03	0.020	<mark>b</mark>	<mark>ab</mark>	<mark>ab</mark>	a	<u>1.06</u>	<u>0.405</u>	<mark>ns</mark>	<mark>ns</mark>	<mark>ns</mark>	ns_

Groups	<mark>E</mark> r	nvironmenta	<mark>l variables</mark>	Zooplankton taxa abundances				
	t	P(perm)	<mark>Unique Perms</mark>	t	P(perm)	Unique Perms		
PB, AB	3,78	0,044	<mark>28</mark>	2,08	0,049	28		
PB, TB	<mark>3,24</mark>	<mark>0,101</mark>	<mark>10</mark>	<mark>2,01</mark>	<mark>0,094</mark>	<mark>10</mark>		
PB, IB	5,65	0,043	<mark>21</mark>	<mark>2,47</mark>	<mark>0,056</mark>	<mark>21</mark>		
AB, TB	1,79	0,014	<mark>84</mark>	1,65	0,008	84		
AB, IB	5,91	0,001	<mark>400</mark>	1,67	0,004	404		
TB, IB	4,59	0,016	<mark>56</mark>	1,57	0,045	<mark>56</mark>		
TB+st7 and 8, ION ST	1.65	0.159	<mark>56</mark>	1,90	0,019	<mark>56</mark>		

Table 6. Estimated grazing, respiration and excretion rates of zooplankton based on allometric models (see methods) and their impact on the phytoplankton stock and production along the PEACETIME survey transect.

	Provenca	al basin			Algeria	n basin			Tyrrheni	an Basin		Id	onian Ba	asin	
	st1	st2	fast1	fast2	fast3	st9	st3	st4	st5	st6	st8	st7	ion1	ion2	ion3
Grazing impact Phytoplankton stock (mg C m ⁻²)	1749	1632	1554	1691	1412	1805	1161	1458	1526	933	1582	1212	1376	1587	1587
Primary Production (mgC m ⁻² d ⁻¹)	295	155	229	184	297	303	165	225	197	190	289	187	266	279	304
$ZCD\ (mgC\ m^{\text{-}2}\ d^{\text{-}1})$	280	155	274	263	249	228	224	278	202	145	195	205	204	244	177
Grazing impact on Phyto. stock (%)	16,0	9,5	17,7	15,6	17,7	12,7	19,3	19,1	13,3	15,6	12,4	17,0	14,8	15,4	11,2
Grazing impact on PP (%)	94,8	99,9	119,7	143,3	83,9	75,4	135,6	123,7	102,5	76,7	67,6	109,7	76,5	87,6	58,3
Respiration Respiration (mg C m ⁻² d ⁻¹) % of Primary productio	n	2 64,3	95,3	90,1	86,2	81,3	83,8	100,2	78,7	62,9	75,6	77,0	72,4	94,7	71,6
respired by zooplanktor NH4 zooplankton	38,0	41,4	41,5	49,0	29,0	26,8	50,6	44,5	39,8	33,1	26,1	41,0	27,1	33,9	23,5
contribution Excretion (mg N-NH4 m ⁻² d ⁻¹)	17,7	9,2	13,6	12,9	12,3	16,2	12,0	14,3	11,3	9,1	10,9	11,0	10,4	13,6	10,3
Phytoplankton needs (mgN m ⁻² d ⁻¹)	50,2	26,4	39,0	31,3	50,6	51,6	28,2	38,3	33,6	32,4	49,2	31,9	45,3	47,4	51,8
N demand (%)	35,2	34,9	34,9	41,1	24,3	31,5	42,6	37,4	33,6	28,0	22,1	34,6	22,9	28,8	19,9
PO4 zooplankton contribution Excretion															
$(mg P-PO4 m^{-2} d^{-1})$	2,3	1,3	2,0	1,9	1,8	1,7	1,8	2,1	1,6	1,3	1,6	1,6	1,5	2,0	1,5
Phytoplankton needs (mg P m ⁻² d ⁻¹)	8,6	4,5	6,7	5,3	8,6	8,8	4,8	6,5	5,7	5,5	8,4	5,4	7,7	8,1	8,8
P demand (%)	27,3	29,7	30,4	35,9	21,3	19,5	36,8	32,5	28,6	23,5	18,6	29,6	19,7	24,1	16,6

Table 7: Comparison of zooplankton biomass and abundance in different areas of the Mediterranean Sea. ** wet weight

Area	Sampling period	Net mesh size (µm)	Layer (m)	Biomass (mg m ⁻³)	Abundance (ind m ⁻³)	Reference
NWMS - Provencal and	Sampling period	(μιιι)	(111)	Bioiliass (ilig ili)	Abundance (ind in)	Reference
Ligurian Seas NWMS - Provencal and	Feb 2013	120	0-250	12.3 (1.9-42.3)	608 (21-2548)	Donoso et al. (2017)
Ligurian Seas	Apr 2013	120	0-250	64.5 (13.9-197.8)	3668 (850-7205)	Donoso et al. (2017)
NWMS - Gulf of Lions shelf NWMS - Gulf of Lions	Mar/Apr 1998	80-200	0-200	9.56 ± 4.73		Gaudy et al. (2003)
shelf	Jan 1999	80-200	0-200	4.73 ± 2.53		Gaudy et al. (2003)
NWMS - Provencal sea	Mar 1969	200	0-200	0.4 - 53		Nival et al. (1975)
NWMS - Provencal sea	Apr 1969	200	0-200	10 - 210		Nival et al. (1975)
NWMS - Provencal sea	Spring 2008	200	0-200	13.15 ± 2.5	1731	Mazzocchi et al. (2014)
NWMS - Provencal sea	Jul 1999	200	0-300		383	Siokou et al. (2019)
NWMS - Provencal sea	May/Jun 2017	100-200	0-300	5.5 ± 2.1	1638 ± 433	this study
SWMS - Algerian sea	Jul-Aug 1997	200	0-200	8.2 (2.1-34.5)	370 (36-844)	Riandey et al. (2005)
SWMS - Algerian sea SWMS- Algero Provencal	Jul 1999	<mark>200</mark>	<mark>0-300</mark>		<mark>197</mark>	Siokou et al. (2019
sea sea	Jun/Jul 2008	200	0-200	5.4	1561 ± 205	Nowaczyk et al. 2011
SWMS- Algerian sea	May/Jun 2017	100-200	0-300	6.6 ± 0.6	1254 ± 191	This study
						Fonda Umani and de
Tyrrhenian Sea	Autumn 1986	200	0-50	3.6 - 32		Olazábal (1988)
Coastal Tyrrhenian sea	1984-2006	200	0-50		1708	Mazzocchi et al. (2011)
Tyrrhenian sea	Sep/Oct 1963	60-300	0-700	0.15-0.3		Cited from Champalbert, (1996)
Tyrrhenian sea	Jun/Jul 2008	200	0-200	3.2	1250	Nowaczyk et al. 2011
Tyrrhenian sea	Jun 1968	Not specify	0-200	5.8**		Cited from Kovalev et al. (2003)
Tyrrhenian sea	May/Jun 2017	100-200	0-300	4.8 ± 1.1	1398 ± 108	This study
Ionian sea	Apr/May 1999	200	0-100	6.0 ± 0.8 (eastern)		Mazzochi et al. (2003)
Ionian sea	Apr/May 1999	200	0-100	8.2 to 13.4 (western)		Mazzochi et al. (2003)
Ionian sea	Spring 1992	200	0-300		219	Mazzochi et al. (2003)
Ionian sea	Spring 1999	200	0-300		193	Mazzochi et al. (2003)
Ionian sea	Spring 2008	200	0-200	2.73	213	Mazzocchi et al. (2014)
Ionian sea	Autumn 2008	200	0-200	3.25	338	Mazzocchi et al. (2014)
Ionian sea	Jun/Jul 2008	200	0-200	8	1181 ± 630	Nowaczyk et al. 2011
Ionian sea	Jul 1999	200	0-300		146	Siokou et al. (2019
Ionian sea	May/Jun 2017	100-200	0-300	5.1 ± 0.5	1003±76	This study