

List of all relevant changes made in the manuscript bg-2020-151-Munkes:

- (1) We went carefully through the manuscript and revised the language.
- (2) We performed the suggested corrections in the model formulations (Sect. 2) and revised in this context also Fig.1.
- (3) We rephrased potentially offensive expressions in the modelling part and extended the explanation of the dilemma of modellers (e.g., Sect.4.4, Conclusions).
- (4) We shortened the biological part (i.e. description of the tables) considerably and enhanced the readability by introducing bullet points, whenever the different species are considered (Sect.3).
- (5) We refer now systematically to the respective model assumptions when summarizing the current biological knowledge (Sect.3)
- (6) We extended the Section 4.2. (Abiotic Processes) according the reviewer's suggestions and discuss the interactions between cyanobacteria and the physical processes in more detail now.
- (7) We introduced a new subsection "Future perspectives" to the Discussion and elaborate on potential future changes of cyanobacteria blooms, including the potential impact of management actions.
- (8) The new subsection "Future perspectives" (Sect.4.4) additionally suggests now measures to enhance the reliability of projections.
- (9) We rewrote the Conclusions.
- (10) We added some extra explanation according to the reviewer's suggestions:
 - We explained the interactions between low-oxygen conditions and cyanobacteria processes (Sect.1)
 - We included a paragraph on sediment processes and phosphate feedbacks (Sect.1).
 - We inserted some extra explanations about photosynthetic processes, including alpha (Sect.3).
 - We extended the explanation of the connection between programmed cell death and biochemical cycling (Sect.3).
 - We explain now why assuming a fixed Redfield-ratio in models might be problematic (Discussion).
- (11) We inserted additional references to the revised manuscript.

Kiel, 6th of Nov., 2020
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Dear Perran Cook,

we herewith submit a substantially revised version of our manuscript titled 'Cyanobacteria Blooms in the Baltic Sea: A Review of Models and Facts'.

As you summarized, and of key importance towards an improved version of the manuscript was to put more effort into reconciling the reductionists approach of modellers and the more complex view of ecologists. Further, you raised the pressing question where increased complexity is needed to better reproduce key observations. Following these recommendations, we added more detail to an intertwined discussion of the modellers approach and the ecologists. However, rather than reconciling the modellers approach and the ecologists view it became even more apparent how big of a challenge is ahead of us on our way towards reliable model forecasts. We hope that, to this end, our review will be a first step in that it provides a comprehensive summary of the state-of-the-art.

Please note that we have made a serious effort to improve the accessibility of the information collated in this review (a total of more than 1000 changes following the advice of the very constructive reviews). Because we rewrote so much we invented the following system in our point by point responses: New, revised text is marked in font colour green. Blue denotes the original text into which we embedded our (green) changes.

Please note that the revised version of our manuscript with tracked-changes is best viewed with Adobe. We apologize for any inconvenience!

We hope that the improved manuscript now meets the standard for publication in Biogeosciences.

In any case we are grateful for your and the reviewer's work.

Yours Sincerely,
the authors

Point-to-point responses to the Reviewers:

Reviewer #1, Ute Daewel:

Dear Ute Daewel,

thank you very much for your time and effort. Based on your (and the other reviewer's help) we rewrote so much we invented the following system in our point-by-point responses: New, revised text is marked in font colour green. Blue denotes the original text into which we embedded our (green) changes.

Kind regards,
the authors

Ute Daewel: As the title already indicates, the study collects information about the processes related to cyanobacterial blooms in the Baltic Sea and how these are implemented in currently used marine ecosystem models. In that context the authors address processes related to growth, limitation processes as well as mortality. Overall the authors provide a good overview about the cyanobacteria implementation in marine ecosystem models as well as a thorough and very useful review on the available literature on cyanobacteria. However, by now I cannot unreservedly recommend the study for publication, as there are some, partly major, points that need to be addressed first.

First: The authors should have gotten in contact to the model developers prior to submission to verify the model descriptions and implementations. One correction has already been issued by Oleg Savchuk, and being one of the developers myself, I have to add some points as well that were not correctly described for the model I use (corrections listed below). This could have easily been avoided by one short communication beforehand. However, I encourage the authors to do so now and correct the section on models accordingly.

The authors: Thank you for your offer to contact you and your help! We have been in contact with the model developers and made corrections accordingly. For BALTSEM we corrected model parameters accordingly to comments of Oleg Savchuk (see response to the comment by Oleg). For ECOSMO we changed the model description accordingly to Ute Daewel (see below):

Ute Daewel: Temperature dependency for ECOSMO cyanobacteria: $T_{bg} = 1 / (1 + \exp(-T))$ with growth multiplied by T_{bg}

The authors: Changed in Fig. 1 and text. Page 6, line 3-6 now reads:

"The ERGOM-model stalls all cyanobacteria growth below 12 °C and sets maximum growth at temperatures exceeding 19 °C. In contrast, the assumed increase in growth with temperature is rather gradual in the CEMBS model. The ECOSMO-model includes a comparably weak temperature dependence for cyanobacteria growth (pers. comm. Ute Daewel)."

Ute Daewel: bg-Zooplankton in ECOSMO does not actively feed on itself, but Macrozooplankton feeds on Microzooplankton

The authors: Changed. Page 9, line 5ff now reads:

"..... the ECOSMO-model differentiates two zooplankton groups (micro- and macro zooplankton) and assumes that micro-zooplankton feeds on phytoplankton and detritus, while macro zooplankton feeds additionally on micro zooplankton."

Ute Daewel: Salinity constraints in ECOSMO:

The authors: After talking to Ute and in accordance with the respective paper we changed the sentence. Page 5, line 23f now reads:

"The SCOB1 and ECOSMO-models include an additional switch which shuts down cyanobacterial growth at salinities above 10 and 11.5 PSU, respectively."

Ute Daewel: positive buoyancy : sinking vel=-0.1 m/d
The authors: Changed in Tab.3, page 54.

Ute Daewel: Second: The authors give a very detailed review on the processes and impacts on cyanobacteria, in functional group type ecosystem models, however, the basic principle is simplification of the ecosystem. I would appreciate if the authors could more clearly connect the modelling section and the experimental section.

The authors: Throughout the document, we made changes in order to improve the link between observational biological evidence and the modelling sections. In each chapter about the biology of cyanobacteria there is, now, also a comparison of how this biological aspect is implemented in the studied models. More specifically respective additions have been added (in green):

Chapter 3.1.2 Temperature dependence. Page 10, line 21-25 now reads:

“In terms of temperature dependency, the model formulations differ widely and some models (e.g., SCOBI and BALTSEM) assume that cyanobacteria require higher temperatures than ordinary phytoplankton for optimal growth (Fig. 1). The respective model assumptions are roughly in agreement with experimental work: while cyanobacteria species typically have optimal growth at higher temperatures than dinoflagellates or diatoms (Paerl et al., 2011), there are only small differences between cyanobacteria and green algae (Lüring et al., 2013). However, there is considerable variation between species.”

Chapter 3.1.4 Light limitation. Page 13, line 12-26 now reads:

“Cyanobacterial and algal photosynthesis rates are significantly influenced i.a. by the combination of light intensity and temperature (Butterwick et al., 2005)..... This basic concept that algal growth is influenced by light and temperature is captured by all the models considered here - although it comes in different flavours: ECOSMO and CEMBS assume that the light requirement for cyanobacteria is higher than for other functional groups, while the other models do not distinguish between functional groups here (c.f., Sect..2.2).”

Chapter 3.1.5 Buoyancy. Page 14, line 17-21 now reads:

“Many cyanobacteria possess buoyancy regulation mechanisms, which enables them to actively control their position in the water column (Visser et al., 2016). In accordance, the ERGOM and ECOSMO model assume that cyanobacteria are buoyant, while the SCOBI model applies a lower sinking speed for cyanobacteria than for other phytoplankton. All respective velocities are in the order of centimetres per day and do not change with light intensity or nutrient availability (Tab. 3).”

Chapter 3.2 Loss Terms. Page 15, line 30-32 & page 16, line 1-7 now reads:

“There are a number of lethal threats for cyanobacteria cells: cell death can be caused by necrosis through adverse environmental conditions, such as insufficient light, nutrients or temperature, or by a programmed cell death (PCD) (Franklin, 2013). Cells can be infected by fungi, undergo viral lysis (Munn, 2011) or can be grazed by a diverse selection of zooplankton (Franklin, 2013). The following subsections list the various causes for cyanobacteria loss. Note that the biogeochemical models investigated in this study differentiate only grazing from other causes of cell losses. For all non-grazing related losses, the models generally assume a fixed loss rate which depends either linearly or quadratic on abundance. An exception is the BALTSEM model, where the mortality of phytoplankton depends on water temperature and is also inversely related to the Liebig minimum function (as a measure of unfavourable environmental conditions; c.f., Sect.2.3).”

Chapter 3.2.4 Grazing on cyanobacteria. Page 19, line 5-8 & line 26-31 now reads:

“Several studies suggested that there is hardly any grazing on cyanobacteria due to their toxicity, bioactive compounds that hamper digestion, bad taste, poor content of lipids, large filamentous size, and low food quality (Carey et al., 2012; Daewel and Schrum, 2013; Ger et al., 2016). All considered models agree in that cyanobacteria are grazed less than other phytoplankton, while the precise proportions vary between the models (Tab.4).

.....This rather low grazing pressure on cyanobacteria has a huge ecosystem impact, as, consequently, during cyanobacteria blooms a lower proportion of the primary production is consumed by larger grazers and therefore not transferred to higher trophic levels. These findings are represented in the models by assuming that grazing on diatoms or other phytoplankton is 2-4 times higher than grazing on cyanobacteria. There is, however, no

consensus yet on the exact formulation of zooplankton grazing in the current model generation and grazers are represented by a single (BALTSEM, ERGOM, SCOBI and CEMBS) or two (ECOSMO) functional groups.”

Chapter 3.3.1. Salinity constrains. Page 22, line 27f & 31f & page 23, line 1-6 now reads:

“The Baltic Sea features a wide range of salinities, ranging from 15-25 PSU in the north western part of the Baltic to 2-3 PSU in the Bothnian Bay..... By this means- salinity has the potential to control the occurrence of cyanobacteria species. Salinity thresholds are set in SCOBI and ECOSMO where growth is not permitted above 10 and 11.5 PSU, respectively. The other models do not include salinity constrains on simulated cyanobacteria. Field observations show that in most parts of the Baltic Sea large cyanobacteria blooms occur during summer, except in the relatively saline waters in the Kattegat and the Belt Sea. Thus, e.g., Rakko and Seppälä (2014) conclude, in line with the SCOBI and ECOSMO models, that high salinities seem to restrict the growth of Baltic Sea cyanobacteria and estimate a threshold around 10 PSU.”

Ute Daewel: The discussion section should be rewritten in a way were the model parameters and the observational finding are related to each other to actually address the “key differences between model approaches and observational evidence” (p. 23, l. 26f) and, if possible, provide recommendations for model improvements.

The authors: We addressed suggestions of the reviewer and discuss in more detail the biological findings in comparison with the model parameters in the Sect. 4.1 (Biogeochemical Processes). The newly introduced Subsection 4.4. (Future Perspectives) contains several suggestions for measures to improve the reliability of projections. Also, we rewrote the “Conclusions” to highlight potential steps forward. More specifically we added:

Chapter 4.1 Biogeochemical processes. Page 25, line 23-31 & page 26, line 1-4 now reads:

“To this end we regard particularly the various choices of growth permitting phosphate thresholds as critical, because we suspect that these parameter values might have a drastic effect on model based investigations of future projections and nutrient load scenarios (cf., Löptien and Dietze, 2020). Another potentially problematic assumption which might introduce substantial uncertainty in model-based projections of nutrient load scenarios is the fixed Redfield-ratio engrained in most models, and which does not account for the storage capacity of DIP by cyanobacteria. This might spuriously shrink their ecological niche. A related aspect, potentially introducing uncertainty, is the differentiation between dissolved inorganic (DIP) and dissolved organic phosphorus (DOP), which is currently not considered in the models. The ability of cyanobacteria to use DOP might increase their success under otherwise nutrient-replete conditions. We regard these effects as potentially important for nutrient reduction scenarios. For projections into our warming future we suspect that especially the very different temperature dependencies might introduce substantial uncertainty in the model responses. The largest uncertainties in model formulations are, however, related to processes that terminate blooms and require further research. To this end we want to highlight viruses that have been shown to be potentially very important but understudied agents in the system.”

Chapter 4.2 Abiotic oceanic processes. Page 26, line 6-33 & page 27, line 1-28 now reads:

“The occurrence of cyanobacteria is controlled by several abiotic factors, such as nutrient availability, light, temperature, salinity, mixed layer depth, currents, and upwelling. This control is both direct and indirect. E.g. temperature directly affects the speed of enzymatic reactions and as such photosynthesis rates and respiration. Temperature will also directly affect organism by changing the oxygen concentration in seawater and its viscosity. Indirect controls include feedback loops where temperature changes the interactions and competition in the feedback (e.g. by promoting competitors or predators). Similar effects exist for surface mixed layer dynamics which modulates the average light levels experienced by phytoplankton cells dispersed in the surface layer. A deepening may, e.g. promote buoyant species which manage to stay at the sun-lit surface while competitors and grazers are mixed downwards into dark ocean layers. Further complexity comes from abiotic nutrient transport which is determined by mixing processes and advection. Given the prominent controls of abiotic processes on pelagic ecosystem dynamics it is desirable to ensure utmost realism in reproducing abiotic drivers. CEMBS, ECOSMO, ERGOM and SCOBI attempt this by coupling to full general ocean circulation models (GCMs) which explicitly calculate three dimensional current fields in response to wind and buoyancy fluxes prescribed at the surface. The underlying GCMs (POP, HAMSOM, MOM and RCO, respectively) are conceptually very similar. In terms of spatial resolution, however, they span a wide range (1 to 5 m horizontally and 2 to 5 m vertically). This suggests that their capability of reproducing small-scale processes

such as local upwelling events will also span a wide range because, those models incapable of explicitly resolving the flow fields, need to parametrize their effect by mixing coefficients. The choice of choosing the appropriate coefficient is, however, very important and very difficult (e.g., Burchard et al., 2005).

Differences between the underlying ocean models can be expected to impact the simulation of cyanobacteria, besides the biogeochemical processes (cf. also, Reissmann et al., 2009): For example, the diazotrophic species *Aphanizomenon* sp. is able to utilise the upwelled DIP, despite relative cold temperatures in the upwelled waters (Nausch et al., 2009; Lips and Lips, 2008). Therefore, as upwelling events will affect the abundance of cyanobacteria, a correct representation in the underlying ocean model is of particular importance.....”

Chapter 4.4 Future Perspectives. Page 27, line 22-30 & Page 28, line 9-15 now reads:

“There is concern that cyanobacteria will benefit from climate change in decades to come (Karlberg and Wulff, 2013; Paerl and Otten, 2013; Visser et al., 2016). This concern is based, e.g., on the fact that the growth and the nitrogen fixation of cyanobacteria is generally favoured by high temperatures (cf., Sect 3.1.2). Further, the increased stratification and the lowered viscosity of seawater due to higher temperatures, might favour buoyant species, such as *Nodularia spumigena* and *Aphanizomenon flos-aquae* .

Accordingly, a number of numerical-models project a significant increase in cyanobacteria blooms in the future for the Baltic Sea (Hense et al., 2013; Neumann, 2010; Saraiva et al., 2019; Meier et al., 2012, 2019). In terms of actual numbers, however, the models diverge substantially.

..... how management actions to limit eutrophication in the Baltic Sea affect nitrogen fixation. The latter is especially of concern because the nitrogen input by cyanobacteria might be in the same order of magnitude as the scheduled nutrient reduction by the Baltic Sea Action Plan (BSAP): a reduction of nitrogen loads of 601.720 t (Helcom, 2007) is put up against an estimated annual nitrogen input of 434.000 t - 792.000 t by cyanobacteria (Wasmund et al., 2005).

We rate specifically the unclear relationship between excess P and cyanobacteria growth and the relation between bloom intensity and nitrogen fixation as key for studies to come.”

Chapter 5 Conclusion. Page 29, line 2-22 now reads:

“Our review summarises current knowledge on cyanobacteria in the Baltic Sea from the peer-reviewed literature. We take both, a ‘modelers’ and a ‘biologists’ perspective. We report that there is consensus between the five dissected biogeochemical models and biologists in that cyanobacteria have an advantage over other functional groups under nitrate deplete conditions. Further, there is consensus that the growth of cyanobacteria can be controlled by the availability of light and phosphate. Other than that, the specific formulations of the underlying assumptions diverge (cf., Table 3, 4 and Fig.1, 2).

The reasons for the large differences between specific model formulations is that there are still considerable knowledge gaps: processes such as the utilization (capabilities) of DOP and the stoichiometric elasticity (i.e. the potential to deviate 10 from the Redfield Ratio) along with the impact of viruses, programmed cell death and grazing are not comprehensively understood. Naturally, this translates into differing numerical representations of growth and decay of cyanobacteria assemblages. Further spread in the underlying assumptions is introduced by the, necessarily, reductionist approach of modelers, which often contrasts with the more complex findings of ecologists – a problem especially prominent because the most dominant cyanobacteria species of the Baltic Sea are diverse and feature very differing traits (cf., 4.4 Future Perspectives). To this end, the overarching question is how much complexity is needed for reliable projections.

In order to proceed, we advertise the use of variance-based sensitivity analyses (such as Sobol’s method (Sobol, 2001)) to determine the key parameters and processes from complex models - and to identify observations and suitable experimental designs for biological studies that may help to constrain these very parameters. By this and similar approaches we expect to make better use of the ever-increasing number of observations that is driven by a combination of persistence using established methods (such as satellite observations, (Bracher et al., 2009; Kahru et al., 2020) and innovation (such as proxy-based hindcasts, (Kaiser et al., 2020)). In summary, we are confident that the reliability of projections of cyanobacteria dynamics will improve along with the development of suitable model-data misfit metrics.”

Ute Daewel: Third: The manuscript gives the impression that parameterization and choice of functional groups happen at random or according to the modelers needs. While specific model parameters can indeed differ widely (“Somewhat disconcerting, the respective parameter choices differ substantially from one model to another” p. 24, l. 6f), so does observational evidence on which the parameter choices indeed are based. In most

cases the developers actually based their parameters on previous experimental publications or at least have a good reason for their assumptions. As this is usually explained in the corresponding publications, it would be more helpful to refer to these underlying reasons for choosing the parameters and revise the impression given in them.

The authors: *We apologize in case anyone who has written cyanobacteria code feels offended by our text. Our goal was to establish an understanding of the remaining problems, which are associated with simulating cyanobacteria. Please note that we do not rank one model approach over another and we do not deny that there are good reasons for any of the underlying model assumptions used to date. We reformulated the "Conclusions", which now explains the difficult situation of modelers and suggests measures to overcome these. Also, we deleted the phrase "somewhat disconcerting" as it seemed offending:*

Chapter 4.1 Biogeochemical processes. Page 25, line 22f now reads:

"The respective parameter choices differ substantially from one model to another, depending on the focus of the respective modeller (Table 3 and 4; Fig. 1 and 2)."

Ute Daewel: The general biological model structures are explained well, but I would consider a brief foray into the physics. This might be too extensive for a paper focusing solely on biological aspects, but some basic explanation of the most important physical variables could be useful for establishing context. Especially as the relationships between cyanobacteria and physics such as temperature are not only directly related as indicated by figure 1 & 2, but also indirectly through the ecosystem interactions as well as physical processes such as transport and upwelling.

The authors: *Agreed. We added an explanation of physical drivers to the revised version of the manuscript and refer now to the respective ocean models for all five biogeochemical models:*

Chapter 4.2 Abiotic oceanic processes. Page 26, line 6-23 now reads:

"The occurrence of cyanobacteria is controlled by several abiotic factors, such as nutrient availability, light, temperature, salinity, mixed layer depth, currents and upwelling. This control is both direct and indirect. E.g. temperature directly affects the speed of enzymatic reactions and as such photosynthesis rates and respiration. Temperature will also directly affect organism by changing the oxygen concentration in seawater and its viscosity. Indirect controls include feedback loops where temperature changes the interactions and competition in the feedback (e.g. by promoting competitors or predators). Similar effects exist for surface mixed layer dynamics which modulates the average light levels experienced by phytoplankton cells dispersed in the surface layer. A deepening may, e.g. promote buoyant species which manage to stay at the sun-lit surface while competitors and grazers are mixed downwards into dark ocean layers. Further complexity comes from abiotic nutrient transport which is determined by mixing processes and advection.

Given the prominent controls of abiotic processes on pelagic ecosystem dynamics it is desirable to ensure utmost realism in reproducing abiotic drivers. CEMBS, ECOSMO, ERGOM and SCOBI attempt this by coupling to full general ocean circulation models (GCMs) which explicitly calculate three dimensional current fields in response to wind and buoyancy fluxes prescribed at the surface. The underlying GCMs (POP, HAMSOM, MOM and RCO, respectively) are conceptually very similar. In terms of spatial resolution, however, they span a wide range (1 to 5nm horizontally and 2 to 5m vertically). This suggests that their capability of reproducing small-scale processes such as local upwelling events will also span a wide range because, those models incapable of explicitly resolving the flow fields, need to parametrize their effect by mixing coefficients. The choice of choosing the appropriate coefficient is, however, very important and very difficult (e.g., Burchard et al., 2005)."

Ute Daewel: Even though I found the ms well readable, the English needs substantial improvement. There are several occasions with wrong grammar, typos and somewhat odd expressions. Some are listed below, but, not being a native speaker myself, I would suggest the authors to edit the language thoroughly.

The authors: *We followed this suggestion. We have checked the manuscript again for spelling errors and odd expression and applied more than a hundred corrections.*

Ute Daewel: specific comments: p2, l 6: “[...] a reduction of loads will have no net effect on the nutrient budget because cyanobacteria will compensate...” Can you give a citation?

The authors: *This is a logical consequence in models which assume a fixed Redfield-ratio. We added the reference: Molot et al. (2014). Page 2, line 5f now reads:*

“A contrary view suggests that reduced loads will decrease primary productivity because nitrogen fixation is capped and cannot fully compensate reductions in nutrient loads (Molot et al., 2014).”

Ute Daewel: p2, l 12-16: “Some of the numerous studies on cyanobacteria [...] are motivated by concerns to run into low-oxygen conditions [...]” Please consider reformulation

The authors: *The sentence was reformulated for clarification. Page 2, line 10ff now reads:*

“Some of the numerous studies on cyanobacteria, which we will review in this study, are motivated by concerns to run into low-oxygen conditions triggered by global warming: warming decreases the solubility of oxygen in seawater which leads to lower oxygen concentrations.”

Ute Daewel: p2, l 22: comma

The authors: *Thanks.*

Ute Daewel: p2, l 30: reference from 2006: can you add a more up to date reference

The authors: *Agreed. We added Shimoda et al., 2016 and Taranu et al., 2012 (see below). Page 2, line 34ff now reads:*

“Despite the importance of cyanobacteria for the Baltic Sea ecosystem, the processes involved in the bloom formation of cyanobacteria are still not comprehensively understood (Hense and Beckmann, 2006; Shimoda and Arhonditsis, 2015; Taranu et al., 2012).”

Ute Daewel: p3, l 30 remove “so-called” as prognostic is a clearly defined concept A:

The authors: *Agreed. Page 4, line 1ff now reads:*

“CEMBS, ECOSMO, ERGOM, SCOB and BALTSEM are all mechanistic models as opposed to statistical models. They are, essentially, a set of partial differential equations that describe the temporal evolution of prognostic entities of relevance or interest.”

Ute Daewel: p4, l 7 I doubt that any modeler really does “ad-hoc” choices on parameters, please rephrase also see comment above.

The authors: *We rephrased the sentence. Page 4, line 12f now reads:*

“Typically, the respective parameters and formulations are based on abductive reasoning which introduces substantial uncertainty to the realism of the model dynamics.”

Ute Daewel: p4, l 23ff: “[...] cyanobacteria grow more slowly [...] and can in most models only thrive when nitrogen is no longer accessible to ordinary phytoplankton [...]”

The citations contain both modelling and observational studies. Does this assumption relate to models or observations?

The authors: *It is an assumption implicit to most models, supported by observational evidence. We changed the references to cite only modelling studies in the revised version of our manuscript to make the context clearer.*

Page 4, line 28-31 now reads:

“A basic concept of the current generation of biogeochemical models is generally the widespread paradigm that diazotrophic cyanobacteria grow more slowly than ordinary phytoplankton and can, therefore, in most models only thrive when nitrogen is no longer accessible to ordinary phytoplankton (LaRoche and Breitbarth, 2005; Hense and Beckmann, 2006; Deutsch et al., 2007).”

Ute Daewel: p4, l 29-31 Can you give a reference where this was done. The phytoplankton bloom dynamics is generally determined by nutrient availability, which is the obvious reason for the nitrogen depletion in surface waters.

The authors: *True – we added a sentence on bottom-up control. Page 4, line 32ff & page 5, line 1-3 now reads:*

“The phytoplankton bloom dynamics is generally determined by nutrient availability, which is the obvious reason for the nitrogen depletion in surface waters. Losses to phytoplankton abundances are set by sink terms which are designed to account for viral lysis, extracellular release, and zooplankton grazing. All models considered here resolve one functional group of zooplankton, with the exception of ECOSMO-model, which resolves two (both,

micro- and macro-zooplankton). As a general rule the model parameters associated to zooplankton growth (fuelled by grazing on phytoplankton) are tuned such that phytoplankton losses exceed the growth, which ultimately leads to a termination of blooms at the right time of the year.”

Ute Daewel: P5, l 6 remove “they”

The authors: Thanks.

Ute Daewel: P5, l15-16 please rephrase, its not clear which model does what.

The authors: We changed the sentence in the revised version of the manuscript to clarify the differences between the models. Page 5, line 21-25 now reads:

“The respective functional forms and thresholds, however, differ between models - with the ERGOM model requiring the highest temperatures to permit growth (Fig. 1a). The SCOB1 and ECOSMO-models include an additional switch which shuts down cyanobacterial growth at salinities above 10 and 11.5 PSU, respectively. Yet another level of complexity is added in SCOB1 where growth necessitates oxygen concentrations above 0.1 mlO₂=l, with growth gradually increasing above this oxygen threshold.”

Ute Daewel: P5, l20 comma

The authors: Thanks.

Ute Daewel: P9 Section 3: In this sections there are some paragraphs that basically just list the same numbers that are given in the tables. You might want to consider shortening these paragraphs to what is new and necessary and avoiding listing the same numbers in the text and in a table.

The authors: Agreed. We have shortened the sentences, removed superfluous information, and listed the content in the form of bullet points. Please find our changes at:

3.1.1. Maximum growth. Page 10, line 11-17 now reads:

Reformatted into bullet points to improve readability.

3.1.2. Temperature dependency. Page 11, line 1-9 now reads:

– *Aphanizomenon flos-aquae* has a somewhat wider optimal temperature range than *Nodularia spumigena*, spanning from 16-31 °C (Bugajev et al., 2015; Carey et al., 2012; Robarts and Zohary, 1987; Paerl and Otten, 2013; Degerholm et al., 2006). Temperatures which permit some growth are considerably lower than the optimal temperatures: Cirés and Ballot (2016) report that *Aphanizomenon flos-aquae* is able to grow at temperatures down to 10 °C. Üveges et al. (2012) measured intensive photosynthesis at even lower 2-5 °C.”

– For *Dolichospermum sp.* optimal temperatures for maximal growth lie between 18 and 25 °C. Growth starts at 10 °C, as reported by (Hellweger et al., 2016; Konopka and Brock, 1978; Robarts and Zohary, 1987; Paerl and Otten, 2013).

– *Nodularia spumigena* prefers 20-25 °C for optimal growth (Degerholm et al., 2006). Growth start at 5 °C (Nordin et al., 1980).”

3.1.4. Light limitation. Page 14, line 3-15 now reads:

– For *Aphanizomenon flos-aquae* optimal irradiance for photosynthesis are reported to be 6->33 Wm⁻² in laboratory and field experiments (Lehtimäki et al., 1997; Üveges et al., 2012). Photoinhibition for *Aphanizomenon flos-aquae* was reported at a light intensity of 99 Wm⁻² (Pechar et al., 1987).

– According to Eigemann et al. (2018) and Walsby and Booker (1980), *Dolichospermum sp.* prefers rather low irradiance values of 4 - 8 Wm⁻².

– Several scientists tested the optimal light intensity for *Nodularia spumigena*. They state that *Nodularia spumigena* grows best at the highest light level tested (23 - 66 Wm⁻²). Net growth was reported over a wide range of light intensities 5-114 Wm⁻² in laboratory experiments (Eigemann et al., 2018; Jodlowska and Latala, 2019; Lehtimäki et al., 1997; Nordin et al., 1980; Roleda et al., 2008). Nordin et al. (1980) observe that very high temperatures (35 °C) in combination with high light levels (114 Wm⁻²) can prohibit the growth of *Nodularia spumigena*. Consistently, Jodlowska and Latala (2019) report reduced filament concentration and reduced photosynthesis rates at a combination of high light intensities and high temperatures (30 °C). Noteworthy is, that Jodlowska and Latala (2019) did not find any photoinhibition of *Nodularia spumigena* until 153 Wm⁻².”

3.1.5. Buoyancy. Page 15, line 11-19 now reads:

“ – For *Aphanizomenon flos-aquae* very different floating velocities of 4-52 m d⁻¹ (Adam, 1999; Reynolds, 2006; Walsby et al., 1995) were measured. Walsby et al. (1995) also observed, that the floating velocities of *Aphanizomenon flos-aquae* vary depending on light conditions, with higher velocities under low light.
– In comparison *Dolichospermum* sp. is assumed to float much slower, as it does not form aggregates (Stal et al., 2003). Accordingly, Walsby et al. (1995) report respective floating velocities of 0.1-0.3 m d⁻¹ for *Dolichospermum* sp.. Note in this context, that *Dolichospermum* sp. occurs mainly at depth of 5-10 m.
– In the Baltic Sea, *Nodularia spumigena* tends to float to the sea surface and there is no evidence for large changes in floating velocities under varying light conditions (Adam, 1999). Recorded floating velocities range from 35-46 m d⁻¹ for *Nodularia spumigena* colonies (Adam, 1999; Walsby et al., 1995).”

3.3.1. Salinity constrains. Page 23, line 14-28 now reads:

“ – *Aphanizomenon flos-aquae* is known as a freshwater species (Laamanen et al., 2002). Accordingly, Rakko and Seppälä (2014) and Laamanen et al. (2002) measured rather low salinities of 0-5 PSU for optimal growth. Rakko and Seppälä (2014) describe this species as rather coastal, preferring less saline conditions. In line, Lehtimäki et al. (1997) state that *Aphanizomenon flos-aquae* is not able to tolerate salinities higher than 10 PSU. Consistently, its abundance decreases from the northern to the southern part of the Baltic proper.
– The taxa *Dolichospermum* originate from freshwater, with some strains adapted to brackish water (Brutemark et al., 2015). In agreement, in the Baltic Sea the specie *Dolichospermum flos-aquae* shows similar growth rates between 0-10 PSU and a strong decrease in growth rates at higher salinities (Moisander et al., 2002).
– For the species *Dolichospermum aphanizomenoides*, Moisander et al. (2002) report a wide range of salinities, from 0-10 PSU, which are related to similar growth rates. *Dolichospermum* taxa originate from freshwater, with some strains that are adapted to brackish water (Brutemark et al., 2015). In the Baltic Sea *Dolichospermum* spp. can grow in salinities between 0-10 PSU, but their growth rates decrease strongly above 10 PSU (Moisander et al., 2002).
– For different strains of *Nodularia spumigena* a wide range of tolerable salinities were reported: 0-20 PSU (Moisander et al., 2002; Lehtimäki et al., 1997), while Rakko and Seppälä (2014) and Nordin et al. (1980) narrow the optimal salinity range down to 5-10 PSU.”

3.3.2 Cyanobacteria akinetes. Page 24, line 17-32 now reads:

Reformatted into bullet points

Ute Daewel: P10 l23-24 Grammar, please rephrase the two sentences. (& comma)

The authors: We rephrased and shortened the sentences. Page 11, line 1-5- now reads:

“ – *Aphanizomenon flos-aquae* has a somewhat wider optimal temperature range than *Nodularia spumigena*, spanning from 16-31 °C (Bugajev et al., 2015; Carey et al., 2012; Robarts and Zohary, 1987; Paerl and Otten, 2013; Degerholm et al., 2006). Temperatures which permit some growth are considerably lower than the optimal temperatures: Cirés and Ballot (2016) report that *Aphanizomenon flos-aquae* is able to grow at temperatures down to 10 °C. Üveges et al. (2012) measured intensive photosynthesis at even lower 2-5 °C.”

Ute Daewel: P11,L8 “Similar to this,...”

The authors: Thanks

Ute Daewel: P15,L28-29 Please explain this sentence.

The authors: We rephrased the sentence to improve the explanation of the link between PCD and biogeochemical cycling. Page 16, line 31ff now reads:

“Also, cell death by PCD may facilitate biogeochemical cycling, through the regular death of cells by PCD and the resulting release of organic and inorganic matter, including the redistribution of fixed nitrogen (Berman-Frank et al., 2004).”

Ute Daewel: P18,L27 Reference?

The authors: see reference Eglite et al., 2019/ Wasmund et al., 2019 added to page 20, line 6ff now reads:
“Among the most important and abundant grazers in the Baltic Sea are copepods. Typical species of copepods in the Baltic Sea are e.g. *Acartia logiremis*, *Temora longicornis* or *Centropages hamatus* (Eglite et al., 2019; Wasmund et al., 2019)”.

Ute Daewel: P19,L5 “...,they are not able to...”

The authors: Thanks. Page 20, line 18f now reads:

“Overall, while copepods play an important grazing role in the ecosystem, they are not able to control cyanobacteria growth in the Baltic Sea (Sommer et al., 2006).”

Ute Daewel: P22, l14-15 revise sentence

The authors: Thanks. Please see: Page 23, line 26ff now reads:

“For different strains of *Nodularia spumigena* a wide range of tolerable salinities were reported: 0-20 PSU (Moisaner et al., 2002; Lehtimäki et al., 1997), while Rakko and Seppälä (2014) and Nordin et al. (1980) narrow the optimal salinity range down to 5-10 PSU.”

Ute Daewel: p23, l27: “Sect. 4.2 debates the impact of the oceanic processes to the Baltic Sea, respectively” In respect to what?

The authors: We deleted “,respectively” behind the word “Baltic Sea in the following sentence. Page 25, line 10f now reads:

“Sect. 4.2 debates the impact of the oceanic processes to the Baltic Sea. Some considerations about model assessment metrics are in sect. 4.3.”

Ute Daewel: p24, l9: “Another potentially problematic assumption [...] is the fixed Redfield-ratio[...]” Why and how is this problematic? Enhance

The authors: e.g., page 13, L 1-10 refers to the cyanobacteria’s storage capacity of DIP which is very difficult to account for when assuming a fixed Redfield-ratio. Not considering this in a model might shrink the ecological niche of cyanobacteria. Further, there is observational evidence that diazotrophs do not obey Redfield (e.g., Larsson et al, 2001). We rephrased the sentence and made the context clear in the revised version of the manuscript:

Chapter 4.1 Biogeochemical processes. Page 25, line 25-28 now reads:

“Another potentially problematic assumption which might introduce substantial uncertainty in model-based projections of nutrient load scenarios is the fixed Redfield-ratio engrained in most models, and which does not account for the storage capacity of DIP by cyanobacteria. This might spuriously shrink their ecological niche.”

Reviewer #2, Justus van Beusekom:

Dear Justus van Beusekom,
thank you very much for your time and effort. Based on your (and the other reviewer's help) we rewrote so much that we invented the following system in our point-by-point responses: New, revised text is marked in font colour green. Blue denotes the original text into which we embedded our (green) changes.

Kind regards,
the authors

Justus Van Beusekom: In this paper, the authors discuss Cyanobacteria blooms in the Baltic. These blooms may cause environmental problems including toxic events or large biomass productions leading to coastal anoxia. A correct modelling of such blooms is important to check measures to combat eutrophication. The authors aim to bring together both a modelers view and a biologists view. They compare 5 models that are used for political decision making regarding the approach to model cyano-blooms. Against this background, the factors that determine the cyanobacteria blooms in the real world are discussed. Based on the comparison of models and field/lab studies, the authors conclude that modelers tend to keep models simple (with good reasons), whereas in the real world cyano-blooms are complex with several species each with their specific requirements responding differently to changes in drivers (like nutrients, light, temperature, grazing). The paper is well written, but the discussion and outlook needs more substance. It does not come as a surprise that modelers tend to (have to) keep models simple and that biologists have an eye on complexity. Given this, I expect a more in-depth discussion on the next steps to be taken to overcome this schism. For instance, given the focus on those models that are used for policy purposes, the limits of using these models should be discussed and suggestions should be made how this dilemma can be solved. Could for instance the combination of "simple" biogeochemical models and conceptual models be a solution? And are examples available?

The authors: *We find your suggestions very constructive and changed the discussion and outlook accordingly. Specifically, we included an additional chapter: '4.4. Future Perspectives' to assess the possible impact of the formulation of cyanobacteria on future projections and make (now) suggestions on how to proceed to solve the dilemma in our revised manuscript. Further, we reformulated the "Conclusions", which now explains the difficult situation of modelers and suggests measures to overcome these.*

Specifically, we added:

4.4 Future Perspectives. Page 27, line 22-32 & page 28, line 1-34 now reads:

*"There is concern that cyanobacteria will benefit from climate change in decades to come (Karlberg and Wulff, 2013; Paerl and Otten, 2013; Visser et al., 2016). This concern is based, e.g., on the fact that the growth and the nitrogen fixation of cyanobacteria is generally favoured by high temperatures (cf, Sect 3.1.2). Further, the increased stratification and the lowered viscosity of seawater due to higher temperatures, might favour buoyant species, such as *Nodularia spumigena* and *Aphanizomenon flos-aquae*.*

Accordingly, a number of numerical-models project a significant increase in cyanobacteria blooms in the future for the Baltic Sea (Hense et al., 2013; Neumann, 2010; Saraiva et al., 2019; Meier et al., 2012, 2019). In terms of actual numbers, however, the models diverge substantially. E.g. the multi-model ensemble by Meier et al. (2012) projects a significant (but not specified) increase in cyanobacteria blooms, while the RCO-SCOBI-model projection of Meier et al. (2019) projects an increase of cyanobacteria blooms by 50 % or the complete loss of cyanobacteria blooms, depending on the respective nutrient load scenario. Sensitivity experiments of the CLC-model by Hense et al. (2013) estimate a future increase in cyanobacteria biomass of 2.3 fold, while the ESCOM model of Neumann (2010) projects an earlier start of cyanobacteria blooms, but lower concentrations. It is also not clear whether an increase in cyanobacteria biomass is also associated with an increase in nitrogen fixation. To this end numerical models give contradicting results: The multi-model ensemble by Saraiva et al. (2019) projects an increase in nitrogen fixation between 22-56 % or a decrease down to 0 %, depending on the nutrient load and climate scenario. Hense et al. (2013), on the other hand, estimate that nitrogen fixation will be twice as high in future, while Neumann (2010) project similar mean nitrogen fixation rates to today rates.

The large spread between models is a challenge for stake holders because it provides little guidance on the question how management actions to limit eutrophication in the Baltic Sea affect nitrogen fixation. The latter is especially of concern because the nitrogen input by cyanobacteria might be in the same order of magnitude as the scheduled nutrient reduction by the Baltic Sea Action Plan (BSAP): a reduction of nitrogen loads of 601.720 t (Helcom, 2007) is put up against an estimated annual nitrogen input of 434.000 t - 792.000 t by cyanobacteria (Wasmund et al., 2005). We rate specifically the unclear relationship between excess P and cyanobacteria growth and the relation between bloom intensity and nitrogen fixation as key for studies to come. The respective uncertainties, reported for the Baltic Sea, are consistent with findings by Löptien and Dietze (2020) who illustrate in a global model that future projections of diazotrophs may diverge considerably - depending on the specific model formulations that determine their ecological niche. We propose to extend related sensitivity studies to rate which model formulations are most impactful on future projections. It might well be the case that widely differing model formulations yield very similar (or very different) projections. The ultimate goal is to determine the minimum number of processes that need to be explicitly resolved in order to ensure predictive skill. For this purpose we rate variance based sensitivity analyses techniques (e.g., Sobol (2001)) as promising systematic approach. The major challenge in this respect is to keep the computational costs at bay. The use of surrogates (e.g., Preuss and von Toussaint (2017)) or spatially limited setups, can make such approaches computationally feasible. Another line of attack is the combination of laboratory, microcosm or field experiments with process-based mechanistic models. If, by clever design, the experimental work can quantify the effects of species traits and the interactions between species, then these information can be implemented in process based mechanistic models; which again can be tested against additional sets of control experiments. By this mean, such mechanistic models may provide robust nuclei on which more complex models may be built. In any case, and as being essential to all approaches, we advocate increased efforts to gather additional observational data. Respective observations are key to assess the predictive skill of traditional numerical models and/or to continue with statistical models based on e.g. machine learning techniques. Finally, we feel that in order to proceed in a cost-efficient manner, the modelling community must develop methods that allow for answering the question what kind of observations are actually needed to constrain their models - a task that is closely related to the development of model assessment metrics (cf., Sect. 4.3)."

5 Conclusion. Page 29, line 1-22 now reads:

Our review summarises current knowledge on cyanobacteria in the Baltic Sea from the peer-reviewed literature. We take both, a 'modelers' and a 'biologists' perspective. We report that there is consensus between the five dissected biogeochemical models and biologists in that cyanobacteria have an advantage over other functional groups under nitrate deplete conditions. Further, 5 there is consensus that the growth of cyanobacteria can be controlled by the availability of light and phosphate. Other than that, the specific formulations of the underlying assumptions diverge (cf., Table 3, 4 and Fig.1, 2).

The reasons for the large differences between specific model formulations is that there are still considerable knowledge gaps: processes such as the utilization (capabilities) of DOP and the stoichiometric elasticity (i.e. the potential to deviate 10 from the Redfield Ratio) along with the impact of viruses, programmed cell death and grazing are not comprehensively understood. Naturally, this translates into differing numerical representations of growth and decay of cyanobacteria assemblages. Further spread in the underlying assumptions is introduced by the, necessarily, reductionist approach of modelers, which often contrasts with the more complex findings of ecologist – a problem especially prominent because the most dominant cyanobacteria species of the Baltic Sea are diverse and feature very differing traits (cf., 4.4 Future 15 Perspectives). To this end, the overarching question is how much complexity is needed for reliable projections.

In order to proceed, we advertise the use of variance-based sensitivity analyses (such as Sobol's method (Sobol, 2001)) to determine the key parameters and processes from complex models - and to identify observations and suitable experimental designs for biological studies that may help to constrain these very parameters. By this and similar approaches we expect to make better use of the ever-increasing number of observations that is driven by a combination of persistence using established methods (such as satellite observations, (Bracher et al., 2009; Kahru et al., 2020) and innovation (such as proxy-based hindcasts, (Kaiser et al., 2020)). In summary, we are confident that the reliability of projections of cyanobacteria dynamics will improve along with the development of suitable model-data misfit metrics.

Minor comments:

Justus Van Beusekom: P1L12, delete commas around “: : :severe”

The authors: Thanks. Page 1, line 12 now reads:
“One particularly severe problem is eutrophication.”

Justus Van Beusekom: P2L4. This sentence is not very clear but very important. I would add a sentence, that explains the main question: How will future, management-induced changes in nutrient loads affect the blooms and how does this interact with expected warming?

The authors: We addressed this question in chapter ‘4.4. Future Perspectives’ (see answer above; Page 27, line 22-32 & page 28, line 1-34)

Justus Van Beusekom: P2L11 Phosphorus (not ..ous) P1L19: Sedimentary processes: this needs some explanation. Especially feedbacks between cyanos and P-release should at least be mentioned.

The authors: Thank you for your suggestion, we included a paragraph about sediment processes and P-release in our revised manuscript. Page 2, line 18-22 now reads:

“One problem that makes consequences of this chain of events so unpredictable and a precise quantitative process understanding so desirable is the existence of a positive feedback loop where low-oxygen conditions may drive P-release from the sediments. This excess P (which comes without the Redfield N equivalent to the system) may fuel additional cyanobacteria blooms (Conley et al., 2002; Savchuk, 2018; Stigebrandt et al., 2014; Vahtera et al., 2007b) thereby closing the positive feedback loop.”

Justus Van Beusekom: P3L4. Alternatives: so, in the end I expect suggestions on how superior alternatives can be developed.

The authors: Following your suggestions we added the chapter ‘4.4. Future Perspectives’ (see answer above. Page 27, line 22-32 & page 28, line 1-34)

Justus Van Beusekom: P4L6: prognostic variables: add s

The authors: Thanks.

Justus Van Beusekom: P5L31: Typo in .19_C? (dot 19)

The authors: Thanks.

Justus Van Beusekom: P6L32 PAR instead of RAR

The authors: Thanks.

Justus Van Beusekom: P11L30 POsubscript4, superscript 3 (you swapped both)

The authors: Thanks.

Justus Van Beusekom: P13, header: You discuss Pmax, but no mention is made of alpha. I suggest to add at least some words to this.

The authors: We rewrote the paragraph and have included some explanations about alpha in our revised manuscript. Page 13, line 12-21 & page 12, line 27-34 now reads:

“Cyanobacterial and algal photosynthesis rates are significantly influenced i.a. by the combination of light intensity and temperature (Butterwick et al., 2005). With increasing light intensity photosynthesis rate will increase until the saturation level is achieved (Ik) and the maximal photosynthesis rate (Pmax) is reached. The initial slope of this photosynthesis-irradiance curve (alpha) describes the performance of both light-harvesting and photosynthetic conversion efficiency. Alpha is species specific and rather temperature-independent over a wide range. Alpha is, however, a strong function of the highly variable carbon specific chlorophyll a content of cells owing to the central role of chlorophyll in photosynthesis.

Pmax, on the other hand, will be influenced i.a. by temperature. Below the temperature optima, P max increases non-linearly roughly doubling with each 10_C rise in temperature until a threshold temperature (Reynolds, 2006). Beyond the threshold excessive temperatures in combination with prolonged exposure to high light intensities may cause photo-inhibition and induce harmful effects in algae (Ibelings, 1996).”

.....“The species Nodularia spumigena and Aphanizomenon sp. seem to be well acclimated to relatively high PAR levels, especially at high temperatures (Sliwinska-Wilczewska et al., 2019). Their cell-specific Pmax values were the highest in Nodularia spumigena and Aphanizomenon sp. strains grown under the lowest light

intensity. Both species changed their Chl a -specific alpha depending on environmental conditions. Maximum alpha values for Nodularia spumigena and Aphanizomenon sp. were found at low light, low temperature and low salinity (10 $\mu\text{mol} \cdot \text{photons} \cdot \text{m}^{-2}$; 15 °C and 8 PSU) (Sliwinska-Wilczewska et al., 2019). Similar to this DeNobel et al. (1998) found that for Aphanizomenon flos-aquae and Anabaena sp. their alpha increased with decreasing irradiance but was always higher for Aphanizomenon flos-aquae than for Anabaena sp. . This was due to a higher chlorophyll a content in cells of Aphanizomenon flos-aquae than in Anabaena sp. .”

Justus Van Beusekom: P13L13 for Nodularia: space between the words

The authors: Thanks.

Justus Van Beusekom: P14L11: Should this be Table 10?

The authors: It's great that you noticed this. We corrected the reference to the table. Page 15, line 9f now reads: *“Listed below are the floating velocities for the considered cyanobacteria species of the Baltic Sea (Table 10 specifies the respective individual experiments).”*

Justus Van Beusekom: P15L18: Claessen et al describe (no s)

The authors: Thanks.

Justus Van Beusekom: P18L9 Inline -> In line ? (see also P21L33)

The authors: Thank you for this note. We corrected all wrong “in line” in the revised version.

Justus Van Beusekom: P18 L28: hamatusarexs ->hamatus?

The authors: Thank you for this note. You are correct with this species name. We changed the species name. Page 20, line 7ff now reads:

“Typical species of copepods in the Baltic Sea are e.g. Acartia logiremis , Temora longicornis or Centropages hamatus (Eglite et al., 2019; Wasmund et al., 2019).”

Justus Van Beusekom: P19L24: exceed -> excert?

The authors: Thanks.

Justus Van Beusekom: P21L31: specie ->species

The authors: Thanks.

Justus Van Beusekom: P21L32: add space after (2002)

The authors: Thanks.

Justus Van Beusekom: P23L27: processes on the Baltic. (delete respectively

The authors: Thanks, we deleted the respectively.

Justus Van Beusekom: P25L33: politic->policy?

The authors: That is right, thank you.

Justus Van Beusekom: P25L14: “A comparison of”. Do you refer to the present study?

The authors: Yes, we refer to models and field experiments studied in this manuscript. We changed the whole conclusion, such that it is easier to understand. Page 29, line 12-15 now reads:

Further spread in the underlying assumptions is introduced by the, necessarily, reductionist approach of modelers, which often contrasts with the more complex findings of ecologist – a problem especially prominent because the most dominant cyanobacteria species of the Baltic Sea are diverse and feature very differing traits (cf., 4.4 Future Perspectives). To this end, the overarching question is how much complexity is needed for reliable projections.

Justus Van Beusekom: P25L17ff: an unclear sentence. Maybe delete at the end “of which”?

The authors: That is right, thank you. We changed and shortened the whole chapter and deleted the sentence (See last answer for the conclusion. Page 29, line 1-22).

Comments #3, Oleg Savchuk:

Dear Oleg Savchuk,

thank you very much for your time and effort. Based on your (and the other reviewer's help) we rewrote so much that we invented the following system in our point-by-point responses: New, revised text is marked in font colour green. Blue denotes the original text into which we embedded our (green) changes.

Kind regards,
the authors

Oleg Savchuk: In my mind, this manuscript provides a very interesting and thorough coverage of an important topic.

The authors: *We thank Oleg Savchuk for this encouraging statement!*

Oleg Savchuk: Although I cannot quite agree with some of the author's statements related to modelling approach, the following notes of mine are by no means a formal review of the manuscript, but merely a correction of some confusion and/or misunderstanding. 1) In BALTSEM model, all the variables and biogeochemical fluxes are expressed in the weight, not molar units. As indicated in Table B2 (Savchuk, 2002), the phosphate half-saturation constants for the cyanobacteria growth and nitrogen fixation rates are 1.5 and 9.0 mg P m⁻³, that is about 0.05 and 0.3 mmol P m⁻³, respectively. Please, in your Table 03; at p. 11, lines 5-7; and elsewhere if necessary.

The authors: *Thank you very much for pointing this out! We adjusted Tab.3 and added a remark to the caption that the provided values for the BALTSEM-model were unit converted (compared to the original study). Additionally, the overall range of half-saturation constants used by the models changed slightly and we adjusted this in Sect. 3.1.3 Nutrient demands. Page 11, line 19 now reads:*

"The respective values for the half saturation constants envelope a large range from 0.05 - 0.5 mmolP/m³ (c.f., Table 3)."

Oleg Savchuk: 2) In BALTSEM model, both the mortality rates and sinking velocities of phytoplankton groups depend not only on the water temperature but are also inversely related to the Liebig minimum function as a measure of unfavorable environmental conditions; for cyanobacteria, accounting also for contribution of nitrogen fixation into their total primary production determined by both DIN uptake and nitrogen fixation. In result, neither mortality rate nor sinking velocity have a fixed value, contrary to what is now stated at p. 15, lines 4-6.

The authors: *We apologize and corrected this in the captions of Table 3 (page 54) and Table 4 (page 55) and in Sect.3.2., page 16, line 5ff now reads:*

Tab 3: *" Note that, as a peculiarity, the BALTSEM models uses different sinking rates for diatoms in spring and autumn and also the sinking speed depends on temperature, environmental conditions and morphology."*

Tab 4: *" The phytoplankton mortality rates in BALTSEM depend on temperature and the environmental conditions."*

Sect.3.2.

"For all non-grazing related losses, the models generally assume a fixed loss rate which depends either linearly or quadratic on abundance. An exception is the BALTSEM model, where the mortality of phytoplankton depends on water temperature and is also inversely related to the Liebig minimum function (as a measure of unfavourable environmental conditions (c.f., Sect.2.3)."