



1 **Title:** The relative importance of photodegradation and biodegradation of terrestrially derived
2 dissolved organic carbon across four lakes of differing trophic status
3

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48 **Abstract**

49 Outgassing of carbon dioxide (CO₂) from freshwater ecosystems comprises 12-25% of the total
50 carbon flux from soils and bedrock. This CO₂ is largely derived from both biodegradation and
51 photodegradation of terrestrial dissolved organic carbon (DOC) entering lakes from wetlands and
52 soils in the watersheds of lakes. In spite of the significance of these two processes in regulating
53 rates of CO₂ outgassing, their relative importance remains poorly understood in lake ecosystems.
54 In this study, we used groundwater from the watersheds of one subtropical and three temperate
55 lakes of differing trophic status to simulate the effects of increases in terrestrial DOC from storm
56 events. We assessed the relative importance of biodegradation and photodegradation in oxidizing
57 DOC to CO₂. We measured changes in DOC concentration, the optical characteristics of the
58 DOC (SUVA₃₂₀ and S_r), dissolved oxygen, and dissolved inorganic carbon (DIC) in short-term
59 experiments from May-August, 2016. In all lakes, photodegradation led to larger changes in
60 DOC and DIC concentrations and optical characteristics than biodegradation. A descriptive
61 discriminant analysis showed that in brown-water lakes, photodegradation led to the largest
62 declines in DOC concentration. In these brown-water systems, ~30% of the DOC was processed
63 by sunlight and ~2% was photo mineralized. In addition to documenting the importance of
64 photodegradation in lakes, these results also highlight how lakes in the future may respond to
65 changes in DOC inputs.

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67



68 **Introduction**

69 Lakes are closely linked to their surrounding terrestrial ecosystems. As the lowest point
70 in the landscape, they receive a significant influx of terrestrially-derived dissolved organic
71 carbon (DOC) and nutrients (Williamson et al., 2009; Wilkinson et al., 2013). Climate and land
72 use changes are altering the link between lakes and their surrounding landscapes by
73 strengthening the flow of material during extreme rain events and large wildfires, or weakening
74 it during extended periods of drought (Strock et al., 2016; Williamson et al., 2016). Long-term
75 changes in DOC concentrations are variable and appear to be regionally controlled. In
76 northeastern North American and western European lakes, there has been as much as a doubling
77 of DOC concentrations due to recovery from anthropogenic acidification and climate change
78 (Monteith et al., 2007; Williamson et al., 2015; de Wit et al., 2016). However, DOC
79 concentrations in Greenland lakes (Saros et al., 2015) and the Mississippi River (Duan et al.,
80 2017) have been decreasing. A long-term study of the Florida Everglades showed that some
81 study sites were decreasing in DOC concentration, but the majority of sites were not changing
82 (Julian et al., 2017). As DOC inputs into aquatic ecosystems have increased, stabilized, or
83 decreased, long-term studies have focused on understanding the mechanisms behind the change,
84 but less research has addressed the fate of DOC once it enters a lake.

85 By attenuating light in the water column and also providing a source of energy, DOC
86 serves an important role in lakes by regulating the balance between photosynthesis and
87 respiration (Williamson et al., 1999), and thus the flux of CO₂ to the atmosphere (Cole et al.,
88 1994). Previous studies indicated that most lakes are net heterotrophic, where the breakdown of
89 organic carbon exceeds production (Kling et al., 1991; Cole et al., 1994). Estimates suggest that
90 lakes respire about half of the annual 2 gigaton flux of carbon to the oceans each year as CO₂



91 (Cole et al., 1994; Tranvik et al., 2009; Tranvik, 2014). The traditional paradigm has been that
92 the dominant mechanism causing the release of excess CO₂ from lakes is bacterial respiration of
93 DOC (biodegradation), with photomineralization accounting for only 10% of bacterial rates
94 (Granéli et al., 1996; del Giorgio et al., 1997; Jonsson et al., 2001). However, research on over
95 200 Arctic lakes, rivers, and streams revealed that sunlight dominated the processing of DOC,
96 and photomineralization rates were on average 5x greater than dark bacterial respiration rates
97 (Cory et al., 2014). In addition, the source of inland water CO₂ remains uncertain (Raymond et
98 al., 2013; Lapierre et al., 2013; Weyhenmeyer et al., 2015) and predicting DOC reactivity has
99 been challenging (Evans et al., 2017). Quantifying the dominant degradation pathways for
100 terrestrial DOC from a range of lakes will improve estimates of carbon fluxes, particularly
101 mineralization rates that currently have a high degree of uncertainty (Hanson et al., 2014).

102 Many past studies have focused on testing the effects of photodegradation and
103 biodegradation on DOC quantity individually, but they have not simultaneously evaluated how
104 these two processes alter the absorbance characteristics of DOC, hereafter referred to as colored
105 dissolved organic matter (CDOM) (Granéli et al., 1996; Koehler et al., 2014; Vachon et al.,
106 2016a). The effects of sunlight on DOC are not isolated to only increasing mineralization rates;
107 photodegradation can also decrease the color and molecular weight of DOC, which can increase
108 light availability and the subsequent bacterial respiration of DOC (Bertilsson and Tranvik, 2000;
109 Amado et al., 2003; Chen and Jaffé, 2016). Cory et al. (2014) found the dominant degradation
110 process for Arctic lakes to be partial photodegradation, suggesting that in lakes, sunlight-driven
111 changes in CDOM without undergoing complete mineralization may dominate DOC processing.

112 Since light attenuation varies so strongly among lakes of differing trophic status, testing
113 the relative importance of DOC processing via photodegradation or biodegradation with



114 mechanistic experiments is needed. Previous research on DOC degradation has primarily
115 occurred in high DOC lakes, but in clear-water lakes, 1% of surface UV-A and
116 photosynthetically active radiation (PAR), which are the primary wavelengths active in
117 photodegradation (Osburn et al., 2001), may reach up to 5-7 m for UV-A and 12-14 m for PAR
118 in oligotrophic lakes and depths of >45-50 m in some of the clearest lakes in the world, such as
119 Lake Tahoe (Rose et al., 2009a; Rose et al., 2009b). Geographic location and time of year
120 influence the amount of solar radiation lakes receive. In the subtropics, PAR and UV light have
121 high intensity across the spectrum year-around, whereas in temperate regions those wavelengths
122 are strongest during the summer months.

123 Watershed land use and lake trophic status have also been shown to influence DOC
124 composition and reactivity (Lu et al., 2013; Hosen et al., 2014; Larson et al., 2014; Evans et al.,
125 2017). DOC from forested systems was more reactive when compared to disturbed environments
126 and had different optical properties (Lu et al., 2012; Williams et al., 2015; Evans et al., 2017).
127 Studies examining how terrestrial DOC inputs are processed in lakes are needed, especially with
128 the increasing frequency of extreme rain events (Rahmstorf and Coumou, 2011; Westra et al.,
129 2014; Fischer and Knutti, 2015). Future climate change projections suggest that for northern
130 ecosystems a 10% increase in precipitation could lead to a 30% increase in the mobilization of
131 soil organic matter (de Wit et al., 2016). Extreme rain events deliver fresh DOC not exposed to
132 prior sunlight into lakes, which can lead to significant reductions in light availability, as well as
133 increases in thermal stability and lake heterotrophy (Jennings et al., 2012; Klug et al., 2012; de
134 Eyto et al., 2016; Zwart et al., 2016). As DOC concentrations change globally, understanding the
135 processes that determine the fate of DOC will help predict the systems most likely to release
136 more CO₂.



137 Here our aim was to 1) determine the relative importance of photodegradation and
138 biodegradation for altering terrestrial DOC quantity and CDOM from lakes of varying trophic
139 status, 2) quantify the percentage of the initial DOC pool that was photomineralized, partially-
140 photodegraded, biodegraded or remained unprocessed, and 3) compare the effects of
141 photodegradation on DOC quantity and CDOM across four lakes to understand differences in
142 how terrestrial DOC from the watersheds of different lake types responds to photodegradation.
143 Since lakes are closely linked to their surrounding landscape (i.e. soils and vegetation), we
144 collected terrestrial DOC from the watershed of three temperate lakes and one subtropical lake,
145 all varying in trophic status. This soil organic matter represents the current and future inputs of
146 organic material. We studied changes in the concentration of DOC, dissolved inorganic carbon
147 (DIC), and dissolved oxygen (DO) and measured changes in CDOM. We hypothesized that
148 photodegradation would be more important than biodegradation in all lakes, but the strongest
149 responses to sunlight would be observed in the brown-water lakes.

150

151 **1. Methods**

152 **1.1 Study Sites and Samplers**

153 Groundwater samples were collected from the watersheds immediately adjacent to four
154 lakes used in this study (Table 1). All of the lakes are small, with a surface area ≤ 0.48 km² and a
155 maximum depth ranging from 12.5 m in Lake Waynewood to 24 m in Lake Giles. The three
156 temperate lakes (Giles – oligotrophic; Lacawac – brown-water; Waynewood – eutrophic) are in
157 close proximity, located on the Pocono Plateau in northeastern Pennsylvania. Lake Annie
158 (brown-water) is a subtropical, sinkhole lake located on the Lake Wales Ridge in south-central
159 Florida. These lakes were selected because of their variability in the dominant vegetation types



160 in their watersheds that lead to differences in DOC concentration and quality (Table 1). Annie,
161 Giles, and Lacawac are all seepage lakes within protected watersheds, and there have been no
162 significant changes in land use or land cover over the past thirty years. The watersheds of Giles
163 and Lacawac have > 90% cover of mixed and northern hardwood-conifer forests, with oak trees
164 dominating the watershed at Giles, while hemlocks represent the highest proportion of
165 Lacawac's watershed (Moeller et al., 1995). Annie is surrounded by well-drained sandy soils and
166 the major vegetation types include a mixed-scrub community, pinelands, and oak forests (Gaiser,
167 2009). Both Annie and Lacawac are brown-water lakes with moderate DOC concentrations and
168 lower transparency (Table 1). A higher percentage of wetlands (7% for Annie and 25% for
169 Lacawac) in their watersheds likely contribute to their darker color compared to the other lakes
170 (Moeller et al., 1995; H. Swain *unpublished data*). Waynewood is the most eutrophic lake and
171 has the largest watershed with runoff from dairy farms upstream that feeds into the lake through
172 an inlet stream. The forest surrounding Waynewood is similarly dominated by oak and hemlock
173 trees, but there is overall less total forest cover in the watershed than Lacawac and Giles, and
174 there are more homes adjacent to the lake (Moeller et al., 1995).
175



176 **Table 1.** Summary characteristics of the four study lakes in May-August 2013–2016 (mean ±
 177 SD). Abbreviations: Chl-*a* (chlorophyll-a), DOC (dissolved organic carbon), GW DOC (initial
 178 groundwater DOC), PAR (photosynthetically active radiation, 400-700 nm), UV-A (ultraviolet
 179 A radiation, 380 nm), UV-B (ultraviolet B radiation, 320 nm), RT (residence time).

Lake	Lat. (°)	Long. (°)	Lake area (km ²)	Max. depth (m)	Chl- <i>a</i> (µg L ⁻¹) ± (SD)	Lake DOC (mg L ⁻¹) ± (SD)	GW DOC (mg L ⁻¹) ± (SD)	pH ± (SD)	1% UV-B depth (m) ± (SD)	1% UV-A depth (m) ± (SD)	1% PAR depth (m) ± (SD)	RT (yr)
Lacawac	41° 22' N	75° 17' W	0.21	13	1.9 (1.4)	5.2 (0.8)	59.4 (6.1)	6.6 ⁺	0.4 (0.1)	0.9 (0.2)	5.7 (0.6)	3.3
Annie	27° 12' N	81° 20' W	0.36	20.7	4.0 (1.5)	9.4 (2.5)	20.7 (0.5)	5.5 (0.3)	0.5*	1.3*	4.5 (1.6)	2
Wayne-wood	41° 23' N	75° 21' W	0.28	12.5	5.3 (3.7)	6.4 (1.0)	7.6 (0.3)	7.5 ⁺	0.3 (0.1)	0.7 (0.2)	4.3 (0.9)	0.42
Giles	41° 22' N	75° 5' W	0.48	24	1.1 (0.7)	2.3 (0.3)	6.0 (0.6)	6.2 ⁺ (0.3)	2.0 (0.5)	4.7 (1.2)	14.4 (2.1)	5.6

180 *Indicates estimates from a single profile in March 2012. ⁺pH data in Lacawac and Waynewood
 181 are from 2015 only and from 2015-2016 in Giles.

182
 183 Samplers were used to collect groundwater as a proxy for terrestrial DOC runoff entering
 184 the lakes. The samplers were installed in close proximity to the Pocono lakes near small inlet
 185 streams in sandy or bog areas on 6 July 2015 (~1 year prior to experiments). The groundwater
 186 sampler consisted of 1m sections of 7.6cm diameter PVC pipe installed to a depth of 60-81cm
 187 below ground. 0.5cm holes were drilled in the sides with a fine mesh covering the holes to let
 188 shallow groundwater in but exclude large particulates. At Lake Annie, a groundwater sampler
 189 was installed on 17 March 2016 on the south side of the lake near a small, intermittent inlet
 190 stream. The groundwater sampler near Lake Annie was a 3m section of PVC pipe installed
 191 slightly deeper to 2m below ground to allow continuous access to groundwater during the dry
 192 season.



193 On 7 May 2016, 10 L of water was collected using a peristaltic pump from the
194 groundwater samplers at all of the Pocono lakes in acid-washed 18 L bottles. Groundwater
195 samples from Annie were collected from the sampler monthly (25 April, 31 May, 27 June, and 1
196 Aug 2016) prior to starting the experiments and shipped overnight on ice to Pennsylvania. All
197 groundwater samples were kept cold (4 °C) and dark until filtered to avoid sunlight exposure
198 prior to the start of the experiments. Samples for the May experiments were filtered on May 8,
199 2016 through a 0.7 µm Whatman GF/F filter. The remaining 8 L of groundwater for the June,
200 July, and August experiments for each Pocono lake were filtered in a similar manner over the
201 next 14 days. Samples were kept cold and dark until the experiments started. Samples for June,
202 July, and August were re-filtered with a 0.7 µm Whatman GF/F filter prior to the start of those
203 experiments. The initial DOC concentration of the groundwater for each lake varied at the start
204 of each experiment, but it was always higher than the in-lake DOC concentration (Table 1).

205

206 **1.2 Sampling Design and Variables Analyzed**

207 To determine the relative importance of photodegradation and biodegradation for
208 processing DOC, we designed three treatments in a manner similar to Cory et al., (2014): 1)
209 photodegradation only, 2) biodegradation only, and 3) control. From each treatment, five
210 different variables were measured including DOC concentration, DIC concentration, DO
211 concentration, SUVA₃₂₀, and S_r. The different variables measured in each treatment required the
212 use of different containers for the sample water. Samples for DOC analysis (concentration and
213 CDOM) were deployed in acid-washed, muffled 35 mL quartz tubes sealed with silicone
214 stoppers. The quartz tubes had an average transmittance of 96% of solar UV-A and 87% of solar
215 UV-B, which allowed for an accurate representation of *in-situ* solar radiation levels (SFig. 1,



216 Morris and Hargreaves, 1997). However, the quartz tubes were not gas tight, so samples for
217 dissolved inorganic carbon (DIC) and dissolved oxygen (DO) analysis were deployed in gas tight
218 borosilicate vials (Labco, Ceredigion, UK). The borosilicate vials had a volume of 12 mL but
219 were filled to 10 mL due to safety concerns with mercury chloride (see below). A clean 10 mL
220 pipette was used to carefully transfer water into the borosilicate vials. Borosilicate glass has a
221 sharp cut-off at 320 nm and transmits <5% UV-B, but it transmits an average of 63% of UV-A
222 radiation and 90% of PAR (SFig. 1, Reche et al., 1999).

223 Water samples for all of the treatments were initially filtered through ashed 0.7 μm
224 Whatman GF/F filters one day prior to the start of each monthly experiment. For the
225 photodegradation and control treatments detailed below, samples for DO and DIC analysis were
226 treated with 0.35 mL of 1% mercury chloride (HgCl_2) to kill the microbial community. Samples
227 for DOC concentration and CDOM analysis (SUVA_{320} and S_r) for the same treatments were
228 sterile filtered with a 0.2 μm membrane filter (Sterivex MilliporeSigma, Burlington, MA USA)
229 pre-rinsed with 100 mL of DI water and 50 mL of sample water instead of using HgCl_2 because
230 adding HgCl_2 altered the optical scans. Sterile filtering has previously been shown to remove the
231 majority of microbes present, and water samples remained sterile for one week following this
232 procedure (Moran et al., 2000; Fasching and Battin, 2011). For the biodegradation treatment,
233 water samples were inoculated with 100 μL of unfiltered groundwater that was collected 1 day
234 prior to the start of each monthly experiment. By adding a fresh inoculum of groundwater each
235 month, we aimed to re-stimulate the microbial community and assess the short-term response of
236 biodegradation. Treatments were deployed in triplicate for each lake (i.e. 3 DOC quartz tubes, 3
237 DO borosilicate vials, and 3 DIC borosilicate vials for each treatment). Here, we included a
238 summary of the three experimental treatments that were designed as follows:



239 a) *Photodegradation Only*: Water for DOC concentration and CDOM analysis (SUVA₃₂₀
240 and S_r) was sterile filtered and stored in quartz tubes (n = 3 replicates). Water for DIC
241 and DO analysis was treated with 1% HgCl₂ and stored in borosilicate vials (n = 6
242 replicates; 3 replicates for DIC and 3 replicates for DO analysis).

243 b) *Biodegradation Only*: Water for all analyses was inoculated with 100 µL of unfiltered
244 groundwater. Water samples for DOC concentration and CDOM analysis were stored in
245 quartz tubes (n = 3 replicates). Water samples for DIC and DO analysis were stored in
246 borosilicate vials (n = 6 replicates; 3 replicates for DIC and 3 replicates for DO analysis).
247 Both the quartz tubes and borosilicate vials were wrapped with multiple layers of
248 aluminum foil to eliminate light exposure.

249 c) *Control*: Water for DOC concentration and CDOM analysis was sterile filtered and
250 stored in quartz tubes (n = 3 replicates). Water for DIC and DO analysis was treated with
251 1% HgCl₂ and stored in borosilicate vials (n = 6 replicates; 3 replicates for DIC and 3
252 replicates for DO analysis). All samples were wrapped in aluminum foil (dark).

253

254 The experimental treatments for each lake were deployed for seven days at the surface of
255 Lake Lacawac in May, June, July, and August of 2016 (for exact sampling dates see SI, Table 1).
256 Samples were kept at the lake surface using floating racks, and samples from each lake were
257 randomly distributed across the racks. The deployment design ensured that samples stayed at the
258 surface and dipped no deeper than 2 cm in the water column. After the one-week exposure, racks
259 were collected from the surface of Lake Lacawac and samples were immediately transferred into
260 coolers and returned to the lab. We assessed the response of terrestrially derived DOC to
261 photodegradation and biodegradation by measuring changes in the concentrations of DOC, DIC,



262 and DO, and the absorbance properties ($SUVA_{320}$ and S_r) of the CDOM. All samples were
263 analyzed within 72 hours of collection.

264 Dissolved organic carbon concentrations were analyzed using a Shimadzu TOC-V_{CPH}
265 Total Organic Analyzer with an ASI-V auto sampler. External acidification was used for each
266 sample and triplicate measurements were performed following the methods of Sharp (1993).
267 Dissolved inorganic carbon concentrations (as CO₂) were measured with a Shimadzu GC-8A
268 Gas Chromatograph using helium as the carrier gas. Samples were acidified using 0.1 N H₂SO₄
269 and then stripped with nitrogen gas prior to injection. Dissolved oxygen was measured using a
270 modified Winkler titration (Parson et al., 1984). Samples for gas measurements (DO and DIC)
271 were kept in a 21°C water bath for 30 minutes prior to analysis. These samples were well mixed
272 just prior to analysis. The absorbance properties of CDOM were analyzed using a Shimadzu UV
273 1800 scanning spectrophotometer at 25°C. Raw absorbance scans were generated from 800 to
274 200 nm using a 1 cm cuvette and were blank corrected with ultra-pure DI water. From the
275 absorbance scans, the spectral slope ratio (S_r) was calculated following Helms et al., (2008). The
276 DOC specific ultraviolet absorbance at 320 nm ($SUVA_{320}$) was calculated following methods in
277 Williamson et al., (2014). S_r can be used as a proxy for the molecular weight of the DOC, while
278 $SUVA_{320}$ can be used as a proxy for DOC color and aromatic carbon content (Helms et al., 2008,
279 Williamson et al., 2014).

280 Due to differences between the borosilicate vials and quartz tubes, the DIC and DO
281 samples were spectrally corrected for the amount of light they received (SI, SFig. 1). Total
282 cumulative energy exposure over the monthly incubations was calculated from a BSI Model
283 GUV-521 (Biospherical Instruments, San Diego, CA) radiometer with cosine irradiance sensors
284 that have a nominal bandwidth of 8 nm for 305 nm, 320 nm, 340 nm, 380 nm, and 400-700 nm



285 (PAR). Daily irradiance for UV-B, UV-A, and PAR were calculated using 15-minute averages of
286 1-second readings from a GUV radiometer located near Lake Lacawac over the 7-day
287 experiments. The area under the curve was calculated by multiplying the measurement frequency
288 (900 sec) by the average of two adjacent time step readings. These values were then summed
289 over the exposure period to calculate the total cumulative energy exposure for each sample.
290 Readings from a profiling BIC sensor (Biospherical Instruments, San Diego, CA) were then used
291 to calculate the percent of the deck cell at the surface rack incubation depth (0.02 m) in Lake
292 Lacawac.

293

294 ***1.3 Explanation of Calculations and Statistical Analysis***

295 To determine the fate of terrestrial DOC in the four lakes, we used the measured changes
296 (i.e. final – control) in DOC and DIC concentrations to identify four pools of DOC:
297 photomineralized, partially photodegraded, biodegraded, and unprocessed. Each pool was
298 converted to a carbon basis, and we assumed a conversion of 0.5 moles CO₂ for each mole of
299 DOC consumed (Cory et al., 2014). The amount of carbon photomineralized (converted to CO₂)
300 was calculated as the concentration of DIC produced [DIC*2] by sunlight (i.e. carbon that was
301 completely oxidized by sunlight). The amount of carbon partially photodegraded represents the
302 remainder of the carbon pool that was processed by sunlight (but not completely oxidized to
303 CO₂) and was calculated as the total DOC processed by sunlight minus the amount
304 photomineralized [Total Photodegraded – Photomineralized]. The amount of carbon biodegraded
305 was calculated as the concentration of DOC lost in the biodegradation treatments. The
306 unprocessed carbon was calculated as the fraction of the carbon pool that was not processed by
307 either sunlight or microbes [Control DOC – Photomineralized – Partially Photodegraded –



308 Biodegraded]. Each process was determined for each lake and each month. Here we report the
309 average response across all four months for each DOC pool.

310 While we carried out monthly experiments (May-August), here we report the average
311 response across the open-water season (i.e. all four months) to provide a more complete picture
312 of DOC processing. The downside of this approach is that it potentially increases variation in
313 variables associated with DOC processing, since such processing may vary across the season.
314 However, there was not a strong seasonal response to photodegradation or biodegradation in all
315 of our study variables (SI Fig. 3). Furthermore, the majority of the terrestrial DOC was collected
316 on a single date and time (except for Lake Annie).

317 Final treatments were compared relative to the dark and killed (1% HgCl₂) control
318 treatments, as those samples were deployed at the surface of the lake with the photodegradation
319 and biodegradation treatments. We used a t-test to determine whether the photodegradation
320 samples for all of the variables were significantly different from the biodegradation samples (n =
321 12 for each treatment) in each lake. Photodegradation and biodegradation samples were analyzed
322 separately using a one-way ANOVA to assess differences between lakes. A post-hoc Tukey's
323 multiple comparison test (Sigma Plot 14.0) was used to determine if there were significant
324 differences in the response variables between the lakes to the photodegradation and
325 biodegradation treatments. A descriptive discriminant analysis (DDA) was used to classify the
326 four lakes based on changes in DOC, DIC, DO, SUVA₃₂₀, and S_r measurements due to
327 photodegradation. Since these five measures are likely to be highly correlated with one another,
328 DDA is a good choice since it considers these relationships simultaneously in the analysis
329 (Sherry 2006). In this case, DDA, works by producing linear combinations of the five measured
330 variables (DOC, DIC, DO, SUVA₃₂₀, and S_r). The first linear combination provides the best



331 separation of the four lakes, followed by subsequent linear combinations for axes that are
332 orthogonal (Sherry, 2006). Linear combinations are weighted more heavily by variables that are
333 better able to discriminate between the lakes. In the figures and tables below, we report these
334 data as either average measured changes (i.e. concentrations) or average percent changes and
335 have indicated where appropriate. Data for this experiment were analyzed in either Sigma Plot
336 14.0 (Fig. 1, Table 2) or Systat version 10.2 (Fig. 4).

337

338 **2. Results**

339 Throughout the results and discussion, the use of the lake names is to present the data in a
340 meaningful manner, but it is important to recognize that the actual water samples originated from
341 groundwater samples adjacent to each lake.

342

343 ***2.1 Photodegradation and biodegradation responses in each lake***

344 Photodegradation altered DOC quantity and CDOM significantly more than
345 biodegradation for terrestrial DOC from the watersheds of all four lakes (Table 2, Fig. 1). For the
346 photodegradation only treatments, exposure to sunlight resulted in significant production of DIC
347 and increases in S_r , as well as significant decreases in DO, DOC, and $SUVA_{320}$ relative to the
348 biodegradation treatments. The only significant effect of biodegradation on terrestrial DOC was
349 a reduction in DO concentrations compared to the dark control (Fig. 1c). In all other cases, the
350 biodegradation treatments were not significantly different than the control, and the average
351 percent change was close to 0.

352 The terrestrial DOC from the brown-water lakes (Lacawac and Annie) typically followed
353 similar patterns to each other, while the terrestrial DOC from the oligotrophic and eutrophic



354 lakes (Giles and Waynewood) responded more similarly to each other. In the brown-water lakes,
355 we observed a stronger response in DOC quantity (i.e. DOC, DIC, and DO), while the changes in
356 DOC quantity were much more muted in the oligotrophic and eutrophic lakes. The responses of
357 S_r changes in each lake due to sunlight did not differ significantly. All four lakes showed a strong
358 response to changes in terrestrial CDOM (i.e. $SUVA_{320}$ and S_r).
359



360 **Table 2.** A summary of the mean (\pm SD) final concentration of DOC, DIC, DO, SUVA₃₂₀ and S_r
 361 in photodegradation (Photo), biodegradation (Bio), and control experimental treatments in
 362 groundwater samples from the watersheds of lakes Lacawac, Annie, Giles, and Waynewood. The
 363 mean (\pm SD) initial concentration for each variable is also depicted. The P/B column list the
 364 results of a t-test to determine whether photodegradation samples were significantly different
 365 from the biodegradation samples ($n = 12$ for each treatment for the four months). Bolded values
 366 indicate the Photo treatments that were statistically different from the Bio treatments ($p < 0.05$).

Analysis	Treatment	Lacawac (Mean \pm SD)	P/B p-value	Annie (Mean \pm SD)	P/B p-value	Giles (Mean \pm SD)	P/B p-value	Waynewood (Mean \pm SD)	P/B p-value
DOC ($\mu\text{moles L}^{-1}$)	Photo	3600 \pm 330	p < 0.001	1270 \pm 211	p < 0.001	692 \pm 123	p < 0.001	883 \pm 73.3	p < 0.001
	Bio	4910 \pm 674		1810 \pm 45.7		608 \pm 99.0		765 \pm 93.8	
	Control	5110 \pm 628		1820 \pm 76.9		630 \pm 102		783 \pm 73.8	
DIC ($\mu\text{moles L}^{-1}$)	Photo	54 \pm 8.2	p < 0.001	41.9 \pm 11.4	p < 0.001	20.4 \pm 1.9	p < 0.001	32.2 \pm 7.3	p = 0.02
	Bio	16.1 \pm 5.0		25.3 \pm 7.2		17.7 \pm 3.0		27.1 \pm 8.0	
	Control	13.8 \pm 4.6		30.4 \pm 18.2		15.3 \pm 2.1		27.8 \pm 3.5	
DO ($\mu\text{moles L}^{-1}$)	Photo	278 \pm 62.4	p < 0.001	419 \pm 25.9	p < 0.001	536 \pm 35.6	p = 0.09	522 \pm 49.0	p < 0.001
	Bio	556 \pm 46.4		533 \pm 42.2		556 \pm 34.3		577 \pm 76.9	
	Control	660 \pm 29.4		656 \pm 32.1		688 \pm 60.9		702 \pm 57.3	
SUVA ₃₂₀ ($\text{m}^{-1}/\text{mg L}^{-1}$)	Photo	4.3 \pm 0.4	p < 0.001	2.4 \pm 0.4	p < 0.001	2.4 \pm 0.2	p < 0.001	1.8 \pm 0.2	p < 0.001
	Bio	5.3 \pm 0.2		3.8 \pm 0.1		4.8 \pm 0.3		3.2 \pm 0.2	
	Control	5.1 \pm 0.2		3.8 \pm 0.1		4.7 \pm 0.2		3.2 \pm 0.1	
S _r	Photo	1.1 \pm 0.0	p < 0.001	1.3 \pm 0.1	p < 0.001	1.4 \pm 0.1	p < 0.001	1.2 \pm 0.1	p < 0.001
	Bio	0.7 \pm 0.1		0.8 \pm 0.0		0.9 \pm 0.1		0.8 \pm 0.1	
	Control	0.7 \pm 0.1		0.8 \pm 0.0		0.9 \pm 0.1		0.9 \pm 0.1	

367

368 Sunlight caused average (\pm SD) DOC losses relative to the control treatments of 30.5 \pm
 369 11.5% and 28.9 \pm 8.3% in Lacawac and Annie, respectively (Fig. 1a). In Giles and Waynewood,
 370 we observed an average of 9.6 \pm 6.5% and 13.4 \pm 6.2% increase in DOC concentration,
 371 respectively following exposure to sunlight. When we compared lakes within each treatment,
 372 there were no significant differences in DOC concentration due to sunlight in Giles vs.
 373 Waynewood, whereas Annie and Lacawac were significantly different from the prior two lakes
 374 and from each other (ANOVA: $F_{1,3} = 70.9$, $p < 0.001$).

375 Decreases in DOC concentration due to photodegradation could lead to mineralization
 376 (i.e. DIC production; Fig. 1b) and therefore oxidation (i.e. DO consumption; Fig. 1c). We



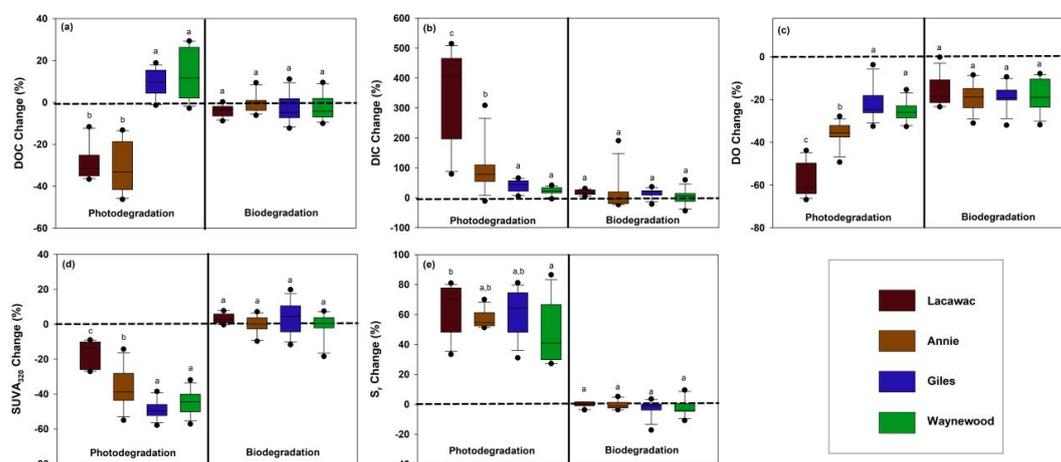
377 observed the production of DIC due to sunlight in all of our lakes (Fig. 1b). In Lacawac and
378 Annie, the average (\pm SD) percent increases in DIC relative to the control treatments were $350 \pm$
379 160% and $96.0 \pm 79.0\%$, respectively. The average percent increases relative to controls in Giles
380 and Waynewood were $40.7 \pm 19.4\%$ and $23.2 \pm 12.7\%$ respectively. The DIC percent change
381 was similar between Giles and Waynewood, and both were statistically different from Annie and
382 Lacawac. The percent DIC change in Lacawac was significantly higher than Annie (ANOVA:
383 $F_{1,3} = 36.4$, $p < 0.001$).

384 In all lakes, both photodegradation and biodegradation led to decreases in DO
385 concentrations (Fig. 1c). Average DO losses due to biodegradation for all four lakes ranged from
386 15 to 18%. DO losses due to photodegradation were more variable. The average DO loss from
387 sunlight in Lacawac and Annie was $58.2 \pm 7.8\%$ and $35.9 \pm 5.4\%$, respectively. In Giles and
388 Waynewood, we observed average DO losses of $21.6 \pm 7.9\%$ and $25.6 \pm 4.7\%$ respectively.
389 While the largest losses of DO due to sunlight were observed in Annie and Lacawac, there was
390 no significant difference between Annie and Waynewood. Giles and Lacawac were significantly
391 different from the other two lakes and from each other (ANOVA: $F_{1,3} = 73.9$, $p < 0.001$).

392 Changes in CDOM due to biodegradation were minimal in all of the lakes (Fig. 1d & 1e).
393 In contrast, photodegradation caused significant changes in all of the lakes, but the magnitude of
394 the change varied by lake. $SUVA_{320}$ decreased in all lakes due to sunlight, but the largest changes
395 were observed in the oligotrophic and eutrophic lakes (Fig. 1d). Average $SUVA_{320}$ values
396 decreased between 16.8% in Lacawac and 48.9% in Giles. The response in Annie and
397 Waynewood were similar, whereas Lacawac and Giles were significantly different from the prior
398 two lakes and each other (ANOVA: $F_{1,3} = 39.7$, $p < 0.001$). In all lakes, S_r increased due to
399 sunlight (Fig. 1e). Average percent increases for the lakes ranged from 46.4% in Waynewood to



400 65.1% in Lacawac. For S_r , the response between Lacawac and Waynewood were significantly
401 different, but those lakes were no different compared to the remaining lakes (ANOVA: $F_{1,3} = 3.1$,
402 $p = 0.04$).
403



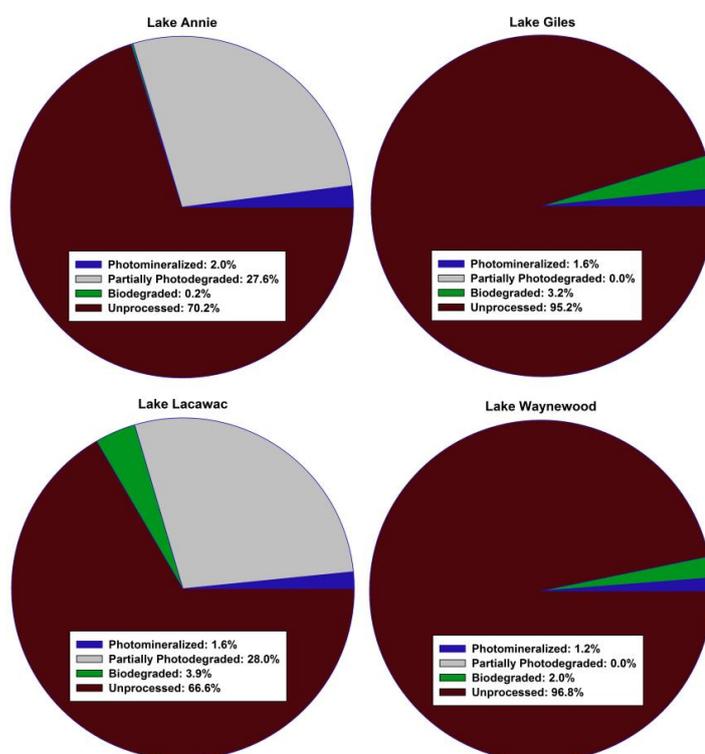
404
405 **Figure 1.** The monthly average percent change from the dark and killed control treatments
406 (dashed line) in each lake for photodegradation (left) and biodegradation (right) for (a) DOC, (b)
407 DIC, (c) DO, (d) $SUVA_{320}$, and (e) S_r . Statistical differences ($p < 0.05$) between lakes are
408 indicated by different letters above each boxplot. For each boxplot $n = 12$ replicates.
409

410 2.2 Fate of DOC

411 Of the four pools of carbon we identified in the groundwater samples entering our study
412 lakes, we found the average amount of carbon processed by sunlight ranged from 1.2% to ~30%
413 (Fig. 2). Carbon in Giles and Waynewood ($< 2\%$) showed little response to sunlight, whereas the
414 response in Annie and Lacawac (~30%) was much higher over the 7-day experiments. The
415 dominant pathway through which sunlight interacted with DOC was through partial
416 photodegradation in these latter two lakes. About 2% of the carbon pool was photomineralized in
417 the brown water lakes. The amount of carbon processed via biodegradation was minimal in all



418 lakes (ranging from 0.2–4%). The fraction of the unprocessed carbon pool ranged from a low of
419 66% for Lacawac to a high of 97% for Waynewood. An average of 3.4 to 34% of the carbon
420 pool was processed in one week.



421

422 **Figure 2.** A summary of the average fate of carbon in the groundwater samples from our study
423 lakes (see methods section for explanation of calculations). All terms were converted to a carbon
424 basis. Photomineralized describes the amount of carbon completely mineralized to CO₂ by
425 sunlight. Partially photodegraded describes the amount of carbon processed by sunlight minus
426 the amount photomineralized. Biodegraded describes the amount of carbon lost through
427 biodegradation. Unprocessed carbon describes the remaining carbon that was not processed by
428 photodegradation or biodegradation.

429

430 **2.3 DOC response by lake trophic status**

431 For the descriptive discriminant analysis (DDA) to classify the lakes, we found that the
432 five metrics were strongly correlated with one another (Table 3). In general, the changes in DOC,



433 DIC, and DO were more strongly correlated with one another than with $SUVA_{320}$ and S_r and vice
434 versa (Table 3). We will refer to the changes in DOC, DIC, and DO as “DOC quantity” and the
435 changes in $SUVA_{320}$ and S_r as “CDOM” for brevity.

436

437 **Table 3.** Pearson correlations between the measured changes in the five metrics: DOC, DIC, DO,
438 $SUVA_{320}$, and S_r .

	DOC	DIC	DO	$SUVA_{320}$
DIC	-0.934			
DO	0.869	-0.837		
$SUVA_{320}$	-0.705	-0.671	-0.666	
S_r	-0.027	0.021	0.163	-0.319

439

440 DDA produced three functions (axes) with canonical correlations of 0.961, 0.753, and
441 0.181 (Fig. 3). Collectively, the entire model was significant (Wilks' $\lambda = 0.032$; $F_{15, 108} = 17.79$; p
442 < 0.001). Effect size was calculated following Sherry and Henson (2010) as $1 - \text{Wilks' } \lambda$, and
443 therefore the overall model explains 96.8% of the variation among lakes. Functions 1 through 3
444 and 2 through 3 were significant ($p < 0.001$ for both). Function 3 was not significant ($p = 0.710$)
445 and therefore is not discussed further. Functions 1 through 3 collectively explain 92.4% of the
446 shared variance while functions 2 through 3 collectively explain 56.7% of the shared variance.

447 Function 1 represents a new variate that is a linear combination of the changes in the five
448 variables that best discriminates the lakes from one another. This new variate is composed
449 mainly of DOC, with a function coefficient of 0.465 and a structure coefficient of 0.821 (Table
450 4). Of note are also DIC, DO, and $SUVA_{320}$ that had smaller function coefficients (< 0.45), but
451 had large structure coefficients (> 0.45). This result suggests that Function 1 is mainly related to
452 DOC quantity. Function 2, also a new variate that is a linear combination of the five measured
453 changes, is composed mainly of $SUVA_{320}$ (function coefficient = 0.985 and structure coefficient

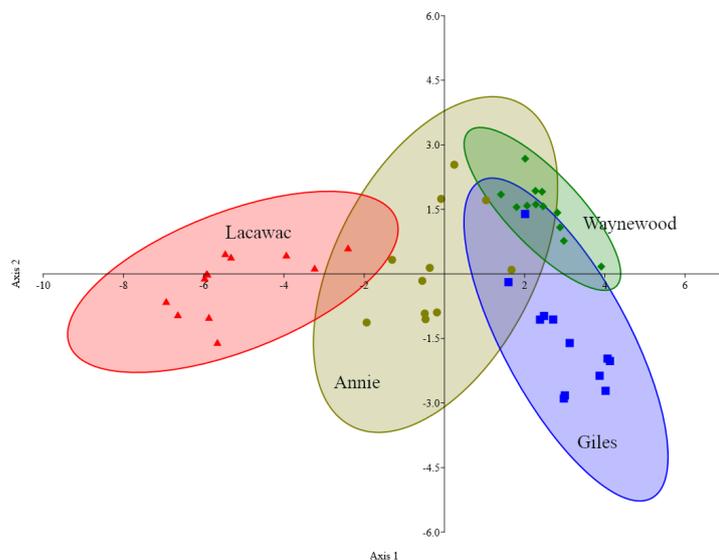


454 = 0.719; Table 4). Function 2 is orthogonal to Function 1 and together they discriminate the four
 455 lakes (Fig. 3).

456
 457 **Table 4.** The solution for changes in measured independent variables that predict the dependent
 458 variable, lake. Structure coefficients (r_s) and communality coefficients greater than $|0.45|$ are in
 459 bold. Coeff = standardized canonical function coefficient; r_s = structure coefficient; r_s^2 = squared
 460 structure coefficient.
 461

Variable	Function 1			Function 2		
	Coeff.	r_s	r_s^2 (%)	Coeff.	r_s	r_s^2 (%)
DOC	0.465	0.821	67.40	0.639	0.278	40.83
DIC	-0.337	-0.703	11.36	-0.059	-0.216	0.35
DO	0.440	0.679	19.36	-0.124	0.009	1.54
SUVA ₃₂₀	-0.139	-0.473	1.93	0.985	0.719	97.02
S _r	0.244	0.068	5.95	-0.238	-0.434	5.66

462



463

464 **Figure 3.** Canonical plot scores and 95% confidence ellipses from descriptive discriminant
 465 analysis of the measured changes (i.e. treatment minus control) in the five variables (DOC, DIC,
 466 DO, SUVA₃₂₀, and S_r) and four lakes: Annie (olive circles), Giles (blue squares), Lacawac (red
 467 triangles), and Waynewood (green diamonds). Only photodegradation samples were included in
 468 this analysis.



469 DDA correctly classified 89.4% of the samples to their collection site (Fig. 3). One
470 sample from Annie was incorrectly assigned to Waynewood, two samples from Giles were
471 incorrectly assigned to Waynewood, and two samples from Lacawac were incorrectly assigned
472 to Annie. All of the Waynewood samples were correctly classified.

473

474 **3. Discussion**

475 **3.1 Comparing the relative importance of photodegradation and biodegradation**

476 Despite a large number of studies examining the effects of either photodegradation or
477 biodegradation on DOC processing, very few have conducted simultaneous *in-situ* experiments
478 of the relative importance of both processes for transforming DOC from the watersheds of a
479 range of different lakes. Our results indicate that sunlight was the primary process in the surface
480 waters responsible for degrading terrestrial DOC from the watershed of all four lakes.
481 Biodegradation played a minimal role in changing the DOC quantity and CDOM. We observed
482 decreases in DOC, DO, and SUVA₃₂₀ due to sunlight and saw increases in DIC and S_r. The loss
483 of DOC, as well as a shift to more photobleached, and lower molecular weight organic material
484 is consistent with prior studies on these lakes that evaluated just the effects of sunlight (Morris
485 and Hargreaves, 1997). Exceptions to DOC loss due to photodegradation occurred in Giles and
486 Waynewood. In these lakes, we observed an increase in average DOC concentrations. In Giles,
487 there was significant production of DOC in June and July. In Waynewood, significant production
488 occurred in May and July. We speculate that this production may be due to the lysing of any
489 microbes remaining in solution. Increases may also be attributed to interactions with iron. We
490 have no measurable evidence, but a number of samples from Giles and Waynewood contained a



491 red precipitate at the conclusion of the one-week experiments. Previously iron-bound DOC could
492 have been released back into the water.

493 Dissolved oxygen was the lone variable where biodegradation led to decreases in DO
494 relative to the controls, but the differences between lakes were not significant. We attributed the
495 changes in DO to the “sloppy feeding” of bacteria, where they produce DOC through exudates
496 and then assimilate it (Evans et al., 2017). The above results are similar to observations in Arctic
497 and tropical waters in that photodegradation was more important than biodegradation on short
498 time scales (Cory et al., 2014; Chen and Jaffé, 2014; Amado et al., 2003). Interestingly, we
499 found that terrestrial DOC from the watersheds of lakes of different trophic status was processed
500 differently, resulting in DIC production and DOC degradation for the brown-water lakes
501 (Lacawac and Annie), but greater changes in $SUVA_{320}$ for the oligotrophic and eutrophic lakes
502 (Giles and Waynewood). This highlights the need to account for lake trophic status in predicting
503 DOC processing and CO_2 emissions from lakes.

504

505 ***3.2 Dominant degradation process***

506 Based on our study design we were able to identify four pools of carbon:
507 photomineralized, partially photodegraded, biodegraded, and unprocessed. The dominant
508 degradation pathway across all lakes was partial photodegradation (i.e. loss of DOC, but no
509 mineralization), although the size of each carbon pool varied by lake. In the brown-water lakes,
510 ~28% of the total carbon pool was partially photodegraded and ~2% was photomineralized. In
511 the oligotrophic and eutrophic lakes ~1.4% of the carbon was photodegraded and none of the
512 carbon was photomineralized.



513 Observations in Toolik Lake showed 70% of the total carbon pool being processed by
514 sunlight during the open water period (~3 months) (Cory et al., 2014). Other estimates have
515 found that photomineralization of DOC accounts for only 8-14% of total water column CO₂
516 production (Granéli et al., 1996; Jonsson et al., 2001; Koehler et al., 2014; Vachon et al., 2016b).
517 We observed 30% of the carbon pool being processed by sunlight within one week in our lakes
518 and this was restricted to the brown-water lakes. Similar to Toolik Lake, the dominant
519 degradation process was partial photodegradation. Partial photodegradation can alter CDOM and
520 stimulate subsequent bacterial respiration. Degradation of CDOM can have important effects for
521 downstream ecosystems if it can be further processed and released as CO₂ or instead is buried or
522 exported downstream (Weyhenmeyer et al., 2012; Catalan et al., 2016; Chen and Jaffe, 2014;
523 Biddanda and Cotner, 2003). It is thus important to include all sunlight-driven degradation
524 processes to fully account for its relative importance.

525 Differences between the responses observed in the Arctic and our temperate/subtropical
526 lakes are most likely explained by the initial concentration and quality of terrestrially derived
527 DOC. In the Arctic, glacial meltwater can be highly photolabile and dominated by seasonal
528 inputs of DOC from shallow or deep soils (Cory et al., 2014; Spencer et al., 2014; and Kaiser et
529 al., 2017). In temperate regions, DOC tends to contain more humic and fulvic acids derived from
530 soils, which may be less photolabile than Arctic DOC. Additionally, we did not integrate our
531 results over the entire water column because the samples were analyzed on the surface of a single
532 lake. Over the entire water column, photodegradation could have processed additional carbon. In
533 clear-water lakes, DOC may be photodegraded down to the 1% UV-A attenuation depth (Osburn
534 et al., 2001), which ranged from 0.7-4.7 m in our study lakes (Table 1).

535



536 **3.3 Response of lakes to photodegradation**

537 With an increase in extreme precipitation events, terrestrial DOC inputs are likely to
538 increase in many aquatic ecosystems (Rahmstorf and Coumou, 2011; Westra et al., 2014). By
539 using groundwater as a proxy of terrestrial inputs from the watersheds of different types of lakes,
540 we simulated the effects of storm events and compared the sensitivity of different terrestrial
541 DOC sources to photodegradation. Interestingly, we found DOC from the watersheds of
542 oligotrophic and eutrophic lakes showed stronger changes in CDOM, compared to DOC from the
543 watersheds of the brown-water lakes that showed significantly larger changes in DOC quantity.
544 This difference may be due to the more allochthonous nature of the brown-water DOC, which is
545 highly photolabile, resulting in greater changes in DOC quantity due to its ability to absorb UV
546 radiation (Bertilsson and Tranvik, 2000). The less allochthonous and more microbially derived
547 DOC from the watersheds of the eutrophic and oligotrophic lakes may be less photolabile with
548 fewer UV-absorbing chromophores. Results of the DDA may be helpful in predicting changes in
549 other lakes based on their trophic status. $SUVA_{320}$ is the variable most likely to change due to
550 photodegradation in eutrophic and oligotrophic lakes. In contrast, DOC concentration is the
551 variable most likely to change in brown-water lakes due to photodegradation. Both results (DOC
552 and $SUVA_{320}$) highlight how lakes of varying trophic status respond to photodegradation. These
553 results can be used to predict how lakes not included in this study will respond to increased DOC
554 concentrations (i.e. browning).

555 Across our study lakes, changes in DIC production scaled linearly with initial
556 groundwater DOC concentration. Lacawac had the highest initial DOC concentration ($59.4 \pm$
557 6.1) and the highest average DIC production, while Giles had the lowest initial DOC
558 concentration (6.0 ± 0.6) and the lowest average DIC production. This suggests that DOC



559 concentration plays a critical role in determining the fate of DOC (Leech et al., 2014; Lapierre et
560 al., 2013). Recent research has also reported that residence time controls organic carbon
561 decomposition across a wide range of freshwater ecosystems (Catalan et al., 2016, Evans et al.,
562 2017). However, extreme precipitation events may shorten the residence time of lakes,
563 effectively flushing out fresh DOC and preventing significant in-lake degradation from occurring
564 (de Wit et al., 2018). For the terrestrial DOC from the oligotrophic and eutrophic lakes, a
565 significant fraction was not degraded, which may mean that terrestrial inputs from these
566 watersheds undergoes less immediate in-lake processing and instead is exported downstream.
567 Our results indicate that differences in the fate and processing of DOC from the watersheds of a
568 range of lake types have important implications for determining which lakes may release more
569 CO₂ versus export DOC downstream (Weyhenmeyer et al., 2012; Zwart et al., 2015;
570 Weyhenmeyer and Conley, 2017).

571 Even though we observed similar responses to photodegradation in the brown-water lakes
572 (Fig. 1), the magnitude of the response varied and may have been related to the initial DOC
573 concentration. Initial concentrations (mg L⁻¹) of terrestrial DOC from Lacawac (59.4 ± 6.1) were
574 almost 3x higher than Annie (20.7 ± 0.5). Average DOC losses for both lakes due to
575 photodegradation were ~35%. The main difference between Lacawac and Annie was the DIC
576 percent change due to photodegradation (Fig. 1). Average percent increases in DIC for Lacawac
577 were close to 400%, whereas in Annie it was ~85%. Despite the fact that both Annie and
578 Lacawac are brown-water lakes, their different DIC production rates indicate that certain types of
579 terrestrial DOC may be more photolabile than others and capable of outgassing large amounts of
580 CO₂. The DDA analysis did also pick out the separation between Lacawac and Annie primarily
581 on axis 1 (DOC). The responses in Annie shared similarities with the other 3 lakes while



582 Lacawac only overlapped with Annie. When put in the context of the entire DOC pool for each
583 lake, photomineralization accounted for 2% of the carbon loss. We anticipated that terrestrial
584 DOC from subtropical lakes would undergo additional microbial processing due to the higher
585 temperatures year-round. In a comparison between boreal Swedish and tropical Brazilian lakes,
586 Graneli et al., (1998) also found strong similarities in changes of DOC concentrations and DIC
587 production between lakes from the different latitudes. A weak significant correlation between
588 DOC concentration and DIC production has also been observed in Amazon clear water systems
589 (Amado et al., 2003)

590

591 **Conclusions**

592 Here we showed that photodegradation can be more important than biodegradation in
593 processing watershed inputs of terrestrial DOC on short time scales in the surface waters of a
594 lake. The responses that we observed varied with lake trophic status. Quantitative changes in
595 DOC, DIC, and DO were strongest in the terrestrial DOC from the watersheds of the brown-
596 water lakes, whereas the largest changes in $SUVA_{320}$ were observed in the terrestrial DOC from
597 the watersheds of the eutrophic and oligotrophic lakes. Consistent with prior studies, we found
598 that sunlight can impact not only changes in the concentration, but also the absorbance properties
599 of the DOC pool. We observed a range of 1.2 to 34% of the carbon pool processed in one week.
600 As DOC concentrations increase in some aquatic ecosystems, the potential for increased CO_2
601 outgassing due to photo-mineralization also increases. On short time scales, sunlight had
602 important impacts on our study lakes. Future studies should focus on additional lakes, longer
603 timescales, and integrating DIC production throughout the water column.



604 Over the next century, DOC concentrations in northern boreal lakes are projected to
605 increase by 65% (Larsen et al., 2011). Thus, understanding the fate of terrestrial sourced organic
606 material will be essential for predicting the ecological consequences for lakes and downstream
607 ecosystems (Solomon et al., 2015; Williamson et al., 2015; Finstad et al., 2016). Improving
608 estimates of organic carbon processing in lakes will be an important component of creating more
609 complete carbon budgets (Hanson et al., 2004; 2014) and global estimates of CO₂ emissions can
610 be more accurately scaled to reflect the ability of lakes to act as CO₂ sinks or sources as
611 browning continues (Lapierre et al., 2013, Evans et al., 2017).

612

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622

623 **Data Availability:**

624 Data and metadata will be made available in the Environmental Data Initiative repository. Data
625 archiving will be led by C. Dempsey and J. Brentrup.

626



627 **Author Contribution Statement**

628 CMD, JAB, and CEW designed the study with help from LBK, EEG, and HMS. CMD, JAB,
629 SM, and HMS collected the water samples and ran the experiments. DPM provided the
630 analytical equipment for measuring DIC and DOC. CMD and JAB analyzed the data, and CMD
631 and MTG conducted the statistical and DDA analyses. CMD and JAB wrote the manuscript with
632 contributions from all of the authors.

633



References

- 634
635
636 Amado, A.M., Farjalla, V.F., Esteves, F.A., and Bozelli, R.L. DOC photo-oxidation in clear
637 water Amazonian aquatic ecosystems. *Amazonia* 17 (3/4): 513-523, 2003.
- 638 Bertilsson, S. and L. J. Tranvik. Photochemical transformation of dissolved organic matter in
639 lakes. *Limnology and Oceanography* 45: 753–762. doi:10.4319/lo.2000.45.4.0753, 2000.
- 640 Biddanda, B.A. and Cotner, J.B. Enhancement of Dissolved Organic Matter Bioavailability by
641 Sunlight and Its Role in the Carbon Cycle of Lakes Superior and Michigan. *Journal of Great
642 Lakes Research* 29 (2): 228-241, 2003.
- 643 Catalan, N., R. Marcé, D. N. Kothawala, and L. J. Tranvik. Organic carbon decomposition rates
644 controlled by water retention time across inland waters. *Nature Geoscience* 9: 501–504.
645 doi:10.1038/ngeo2720, 2006.
- 646 Cole, J. J., N. F. Caraco, G. W. Kling, and T. K. Kratz. Carbon dioxide supersaturation in the
647 surface waters of lakes. *Science* 265: 1568–1570, 1994.
- 648 Cory, R. M., C. P. Ward, B. C. Crump, and G. W. Kling. Sunlight controls water column
649 processing of carbon in arctic fresh waters. *Science* 345: 925–928.
650 doi:10.1126/science.1253119, 2014.
- 651 Chen, M. and R. Jaffe. Photo- and bio-reactivity patterns of dissolved organic matter from
652 biomass and soil leachates and surface waters in a subtropical wetland. *Water Research* 61:
653 181-190. DOI: 10.1016/j.watres.2014.03.075, 2014.
- 654 Chen, M., Jaffé, R. Quantitative assessment of photo- and bio-reactivity of chromophoric and
655 fluorescent dissolved organic matter from biomass and soil leachates and from surface
656 waters in a subtropical wetland. *Biogeochemistry* 129: 273–289.
657 <https://doi.org/10.1007/s10533-016-0231-7>, 2016.
- 658 de Eyto, E., E. Jennings, E. Ryder, K. Sparber, M. Dillane, C. Dalton, and R. Poole. Response of
659 a humic lake ecosystem to an extreme precipitation event: physical, chemical, and biological
660 implications. *Inland Waters* 6: 483–498. doi:10.5268/IW-6.4.875, 2016.
- 661 de Wit, H. A., S. Valinia, G. A. Weyhenmeyer, and others. Current Browning of Surface Waters
662 Will Be Further Promoted by Wetter Climate. *Environ. Sci. Technol. Lett.* 3: 430–435.
663 doi:10.1021/acs.estlett.6b00396, 2016.
- 664 de Wit, H.A., Couture, R.M., Jackson-Blake, L., Futter, M.N., Valinia, S., Austnes, K., Guerrero,
665 J., and Lin, Y. Pipes or chimneys? For carbon cycling in small boreal lakes, precipitation
666 matters most. *Limnology and Oceanography Letters* 3:275-284, 2018.
- 667 del Giorgio, P. A., J. J. Cole, and A. Cimleris. Respiration rates in bacteria exceed
668 phytoplankton production in unproductive aquatic systems. *Nature* 385: 148–151, 1997.
- 669 Duan, S., He, Y., Kaushai, S.S., Bianchi, T.S., Ward, N.D., and Guo, L. Impact of wetland
670 decline on decreasing dissolved organic carbon concentrations along the Mississippi River
671 continuum. *Frontiers in Marine Science* 3:280. doi: 10.3389/fmars.2016.00280, 2017.
- 672 Evans, C.D., Futter, M.N., Moldan, F., Valinia, S., Frogbrook, Z., and Kothawala, D.N.
673 Variability in organic carbon reactivity across lake residence time and trophic gradients.
674 *Nature Geosciences* 3: 832-837. doi: 10.1038/NGEO3051, 2017.
- 675 Fasching, C., and T. J. Battin. Exposure of dissolved organic matter to UV-radiation increases
676 bacterial growth efficiency in a clear-water Alpine stream and its adjacent groundwater.
677 *Aquatic Sciences* 74: 143–153. doi:10.1007/s00027-011-0205-8, 2011.
- 678 Finstad, A. G., T. Andersen, S. Larsen, K. Tominaga, S. Blumentrath, H. A. de Wit, H.
679 Tømmervik, and D. O. Hessen. From greening to browning: Catchment vegetation



- 680 development and reduced S-deposition promote organic carbon load on decadal time scales
681 in Nordic lakes. *Sci. Rep.* 6: 31944–9. doi:10.1038/srep31944, 2016.
- 682 Fischer, E. M., and R. Knutti. Anthropogenic contribution to global occurrence of heavy
683 precipitation and high-temperature extremes. *Nature Climate Change* 5: 560–564.
684 doi:10.1038/nclimate2617, 2015.
- 685 Gaiser, E.E., Deyrup, N.D., Bachmann, R.W., Battoe, L.E. and Swain, H.M. Effects of climate
686 variability on transparency and thermal structure in subtropical, monomictic Lake Annie,
687 Florida. *Fundamental and Applied Limnology*. Vol 175 (3): 217-230, 2009.
- 688 Granéli, W., M. Lindell, and L. Tranvik. Photo-oxidative production of dissolved inorganic
689 carbon in lakes of different humic content. *Limnology and Oceanography* 41: 698–706.
690 doi:10.4319/lo.1996.41.4.0698, 1996.
- 691 Granéli, W., Lindell, M., de Faria, B.M., and de Assis Esteves, F. Photoproduction of dissolved
692 inorganic carbon in temperate and tropical lakes – dependence on wavelength band and
693 dissolved organic carbon concentration. *Biogeochemistry* 43: 175-195, 1998.
- 694 Hanson, P. C., A. I. Pollard, D. L. Bade, K. Predick, S. R. Carpenter, and J. A. Foley. A model of
695 carbon evasion and sedimentation in temperate lakes. *Global Change Biology* 10: 1285–
696 1298. doi:10.1111/j.1529-8817.2003.00805.x, 2004.
- 697 Hanson, P. C., M. L. Pace, S. R. Carpenter, J. J. Cole, and E. H. Stanley. Integrating Landscape
698 Carbon Cycling: Research Needs for Resolving Organic Carbon Budgets of Lakes.
699 *Ecosystems* 18: 363–375. doi:10.1007/s10021-014-9826-9, 2014.
- 700 Helms, J. R., Stubbins, A., Ritchie, J. D., Minor, E. C., Kieber, D. J., and Mopper, K. Absorption
701 spectral slopes and slope ratios as indicators of molecular weight, source, and
702 photobleaching of chromophoric dissolved organic matter, *Limnol. Oceanogr.*, 53, 955–969,
703 2008.
- 704 Hosen, J.D., O.T. McDonough, C.M. Febria, and M.A. Palmer. Altered stream dissolved organic
705 matter composition and bioavailability with urbanization. *Environmental Science &*
706 *Technology*. 48:7817-7824, 2014.
- 707 Jennings, E., S. Jones, L. Arvola, and others. Effects of weather-related episodic events in lakes:
708 an analysis based on high-frequency data. *Freshwater Biology* 57: 589–601.
709 doi:10.1111/j.1365-2427.2011.02729.x, 2012.
- 710 Jonsson, A., M. Meili, A.-K. Bergström, and M. Jansson. Whole-lake mineralization of
711 allochthonous and autochthonous organic carbon in a large humic lake (Oerträsket, N.
712 Sweden). *Limnology and Oceanography* 46: 1691–1700, 2001.
- 713 Julian II, P., Gerber, S., Wright, A.L., Gu, B., and Osborne, T.Z. Carbon pool trends and
714 dynamics within a subtropical peatland during long-term restoration. *Ecological Processes*
715 6:43. DOI 10.1186/s13717-017-0110-8, 2017.
- 716 Kaiser, K., Candedo-Oropeza, M., McMahon, R., and Amon, R.M. Origins and transformations
717 of dissolved organic matter in large Arctic rivers. *Scientific Reports* 7, 13064, 2017.
- 718 Kling, G. W., G. W. Kipphut, and M. C. Miller. Arctic lakes and rivers as gas conduits to the
719 atmosphere: implications for tundra carbon budgets. *Science* 251:298-301, 1991.
- 720 Klug, J. L., D. C. Richardson, H. A. Ewing, and others. Ecosystem Effects of a Tropical Cyclone
721 on a Network of Lakes in Northeastern North America. *Environmental Science &*
722 *Technology* 46: 11693–11701. doi:10.1021/es302063v, 2012.
- 723 Koehler, B., T. Landelius, G. A. Weyhenmeyer, N. Machida, and L. J. Tranvik. Sunlight-induced
724 carbon dioxide emissions from inland waters. *Global Biogeochemical Cycles* 28: 696–711.
725 doi:10.1002/2014GB004850, 2014.



- 726 Lapiere, J.-F., F. Guillemette, M. Berggren, and P. A. del Giorgio. Increases in terrestrially
727 derived carbon stimulate organic carbon processing and CO₂ emissions in boreal aquatic
728 ecosystems. *Nat Comms* 4. doi:10.1038/ncomms3972, 2013.
- 729 Larsen, S., T. Andersen, and D. O. Hessen. Climate change predicted to cause severe increase of
730 organic carbon in lakes. *Global Change Biology* 17: 1186–1192. doi:10.1111/j.1365-
731 2486.2010.02257.x, 2011.
- 732 Larson, J., Frost, P., Xenopoulos, M., Williams, C., Morales-Williams, A., Vallazza, J., Nelson,
733 J., and Richardson, W. Relationships Between Land Cover and Dissolved Organic Matter
734 Change Along the River to Lake Transition. *Ecosystems*. 17. 10.1007/s10021-014-9804-2,
735 2014.
- 736 Leech, D., M. Snyder, and R. Wetzel. Alterations in the photomineralization of allochthonous
737 DOM related to elevated atmospheric CO₂. *Inland Waters* 4: 147–156. doi:10.5268/IW-
738 4.2.626, 2014.
- 739 Lu, Y. H., J. E. Bauer, E. A. Canuel, Y. Yamashita, R. M. Chambers, and R. Jaffe´.
740 Photochemical and microbial alteration of dissolved organic matter in temperate headwater
741 streams associated with different land use, *J. Geophys. Res. Biogeosci.*, 118, 566–580,
742 doi:10.1002/jgrg.20048, 2013.
- 743 Madronich, S. UV radiation in the natural and perturbed atmosphere. Pages 17–69 in M. Tevini,
744 editor. *Environmental effects of UV (ultraviolet) radiation*. Lewis Publisher, Boca Raton,
745 Florida, 1993.
- 746 Moeller, R.E., Williamson, C.E., Hargreaves, B.R., and Morris, D.P. *Limnology of lakes*
747 *Lacawac, Giles, and Waynewood 1989-1993: An introduction to the core lakes of the*
748 *Pocono comparative lakes program*. Lehigh University, 1995.
- 749 Monteith, D. T., J. L. Stoddard, C. D. Evans, and others. Dissolved organic carbon trends
750 resulting from changes in atmospheric deposition chemistry. *Nature* 450: 537–540.
751 doi:10.1038/nature06316, 2007.
- 752 Moran, M. A., W. M. Sheldon Jr, and R. G. Zepp. Carbon loss and optical property changes
753 during long-term photochemical and biological degradation of estuarine dissolved organic
754 matter. *Limnology and Oceanography* 45: 1254–1264. doi:10.4319/lo.2000.45.6.1254, 2000.
- 755 Morris, D.P. and Hargreaves, B.R. The role of photochemical degradation of dissolved organic
756 carbon in regulating the UV transparency of three lakes on the Pocono Plateau. *Limnology*
757 *and Oceanography* 42(2): 239-249, 1997.
- 758 Osburn, C. L., H. E. Zagarese, D. P. Morris, B. R. Hargreaves, and W. E. Cravero. Calculation of
759 spectral weighting functions for the solar photobleaching of chromophoric dissolved organic
760 matter in temperate lakes. *Limnology and Oceanography* 46: 1455–1467.
761 doi:10.4319/lo.2001.46.6.1455, 2001.
- 762 Parsons, T. R., Y. Maita, and C. M. Lalli. *A manual for chemical and biological methods for*
763 *seawater analysis*, Pergamon Press, 1984.
- 764 Rahmstorf, S., and D. Coumou. Increase of extreme events in a warming world. *Proceedings of*
765 *the National Academy of Sciences of the United States of America* 108: 17905–17909.
766 doi:10.1073/pnas.1101766108, 2011.
- 767 Raymond, P. A., D. L. Hartmann, R. Lauerwald, and others. Global carbon dioxide emissions
768 from inland waters. *Nature* 503: 355–359. doi:10.1038/nature12760, 2013.
- 769 Reche, I., Pace, M.L, and Cole, J.J. Relationship of trophic and chemical conditions to
770 photobleaching of dissolved organic matter in lake ecosystems. *Biogeochemistry* 44: 259-
771 280, 1999.



- 772 Rose, K.C., Williamson, C.E., Saros, J.E., Sommaruga, R., and Fisher, J.M. Differences in UV
773 transparency and thermal structure between alpine and subalpine lakes: implications for
774 organisms. *Photochemical and Photobiological Sciences*. Volume 8(9): 1244-56.
775 doi:10.1039/b9056, 2009a.
- 776 Rose, K.C., Williamson, C.E., Schladow, S. G., Winder, M., and Oris, J.T. Patterns of spatial and
777 temporal variability of UV transparency in Lake Tahoe, California-Nevada. *Journal of*
778 *Geophysical Research Biogeosciences*. Volume 114(G2). doi:10.1029/2008JG000816, 2009,
779 2009b.
- 780 Saros, J. E., C. L. Osburn, R.M.Northington, S.D. Birkel, J.D. Auger, C. A. Stedmon, and N. J.
781 Anderson. Recent decrease in DOC concentrations in Arctic lakes of southwest Greenland,
782 *Geophys. Res. Lett.*, 42, 6703–6709, doi:10.1002/2015GL065075, 2015.
- 783 Sharp, J.H., and others. Procedures subgroup report. *Marine Chemistry*. Volume 41. 1993. pp 37-
784 49, 1993.
- 785 Sherry, A. Discriminant analysis in counseling psychology research. *The Counseling*
786 *Psychologist* 34: 661-683, 2006.
- 787 Sherry, A., and R. K. Henson. Conducting and interpreting canonical correlation analysis in
788 personality research: A user-friendly primer. *Journal of Personality Assessment* 84: 37-48,
789 2010.
- 790 Solomon, C. T., S. E. Jones, B. C. Weidel, and others. Ecosystem Consequences of Changing
791 Inputs of Terrestrial Dissolved Organic Matter to Lakes: Current Knowledge and Future
792 Challenges. *Ecosystems* 18: 376–389. doi:10.1007/s10021-015-9848-y, 2015.
- 793 Spencer, R.G.M., Guo, W. Raymond, P.A., Dittmar, T., Hood, E., Fellman, J., and Stubbins, A.
794 Source and biolability of ancient dissolved organic matter in glacier and lake ecosystems on
795 the Tibetan Plateau. *Geochimica et Cosmochimica Acta*. Vol 142:1-64-74, 2014.
- 796 Strock, K. E., J. E. Saros, S. J. Nelson, S. D. Birkel, J. S. Kahl, and W. H. McDowell. Extreme
797 weather years drive episodic changes in lake chemistry: implications for recovery from
798 sulfate deposition and long-term trends in dissolved organic carbon. *Biogeochemistry* 127:
799 353–365. doi:10.1007/s10533-016-0185-9, 2016.
- 800 Tranvik, L. J., J. A. Downing, J. B. Cotner, and others. Lakes and reservoirs as regulators of
801 carbon cycling and climate. *Limnology and Oceanography* 54: 2298–2314, 2009.
- 802 Tranvik, L. Carbon cycling in the Arctic. *Science* 345: 870–870. doi:10.1126/science.1258235,
803 2014.
- 804 Vachon, D., C. T. Solomon, and P. A. del Giorgio. Reconstructing the seasonal dynamics and
805 relative contribution of the major processes sustaining CO₂ emissions in northern lakes.
806 *Limnology and Oceanography* 62: 706–722. doi:10.1002/lno.10454, 2016a.
- 807 Vachon, D., J.-F. Lapierre, and P. A. del Giorgio. Seasonality of photochemical dissolved
808 organic carbon mineralization and its relative contribution to pelagic CO₂ production in
809 northern lakes. *J. Geophys. Res. Biogeosci.* 121: 864–878. doi:10.1002/2015JG003244,
810 2016b.
- 811 Westra, S., H. J. Fowler, J. P. Evans, and others. Future changes to the intensity and frequency of
812 short-duration extreme rainfall. *Reviews of Geophysics* 52: 522–555.
813 doi:10.1002/2014RG000464, 2014.
- 814 Weyhenmeyer, G. A., M. Fröberg, E. Karlun, M. Khalil, D. Kothawala, J. Temnerud, and L. J.
815 Tranvik. Selective decay of terrestrial organic carbon during transport from land to sea.
816 *Global Change Biology* 18: 349–355, 2012.
- 817 Weyhenmeyer, G. A., S. Kosten, M. B. Wallin, L. J. Tranvik, E. Jeppesen, and F. Roland.



- 818 Significant fraction of CO₂ emissions from boreal lakes derived from hydrologic inorganic
819 carbon inputs. *Nature Geosci.* doi:10.1038/ngeo2582, 2015.
- 820 Weyhenmeyer, G. A., and D. J. Conley. Large differences between carbon and nutrient loss rates
821 along the land to ocean aquatic continuum-implications for energy:nutrient ratios at
822 downstream sites. *Limnology and Oceanography* 10: 141–11. doi:10.1002/lno.10589, 2017.
- 823 Wilkinson, G. M., M. L. Pace, and J. J. Cole. Terrestrial dominance of organic matter in north
824 temperate lakes. *Global Biogeochemical Cycles* 27: 43–51. doi:10.1029/2012GB004453,
825 2013.
- 826 Williams, C.J., Frost, P.C., Morales-Williams, A.M., Larson, J.H., Richardson, W.B., Chiandet,
827 A.S. and Xenopoulos, M.A. Human activities cause distinct dissolved organic
828 matter composition across freshwater ecosystems. *Glob Change Biol*, 22: 613–626.
829 doi:10.1111/gcb.13094, 2016.
- 830 Williamson, C. E., D. P. Morris, M. L. Pace, and O. G. Olson. Dissolved organic carbon and
831 nutrients as regulators of lake ecosystems: Resurrection of a more integrated paradigm.
832 *Limnology and Oceanography* 44: 795–803. doi:10.4319/lno.1999.44.3_part_2.0795, 1999.
- 833 Williamson, C. E., J. E. Saros, W. F. Vincent, and J. P. Smol. Lakes and reservoirs as sentinels,
834 integrators, and regulators of climate change. *Limnology and Oceanography* 54: 2273–2282,
835 2009.
- 836 Williamson, C.E., J.A. Brentrup, J. Zhang, W.H. Renwick, B.R. Hargreaves, L.B. Knoll, E.P.
837 Overholt, and K.C. Rose. Lakes as sensors in the landscape: Optical metrics as scalable
838 sentinel responses to climate change. *Limnology and Oceanography* 59: 840–850, 2014.
- 839 Williamson, C. E., E. P. Overholt, R. M. Pilla, T. H. Leach, J. A. Brentrup, L. B. Knoll, E. M.
840 Mette, and R. E. Moeller. Ecological consequences of long- term browning in lakes. *Sci.*
841 *Rep.* 5. doi:10.1038/srep18666, 2015.
- 842 Williamson, C. E., E. P. Overholt, J. A. Brentrup, and others. Sentinel responses to droughts,
843 wildfires, and floods: effects of UV radiation on lakes and their ecosystem services.
844 *Frontiers in Ecology and the Environment* 14: 102–109. doi:10.1002/fee.1228, 2016.
- 845 Zwart, J. A., N. Craig, P. T. Kelly, S. D. Sebestyen, C. T. Solomon, B. C. Weidel, and S. E.
846 Jones. Metabolic and physiochemical responses to a whole-lake experimental increase in
847 dissolved organic carbon in a north-temperate lake. *Limnology and Oceanography* 61: 723–
848 734. doi:10.1002/lno.10248, 2015.