Dear Editor,

We thank the referees for their supportive and constructive comments to our manuscript. We have responded to the comments point by point as follows. The revised sentences or contents are underlined.

Responses to comment 1 are following:

General comments:

The manuscript is clearly written and structured. The study is well designed and explained considering how complex experiments tend to get when increasing the number of environmental drivers. Also, the rationale behind using a “environmental clustering” approach is clearly stated and the co-variation of the environmental drivers is well deduced from previous studies and literature. It is interesting to see in this study how much variation in response to nutrient availability and light there is, even though there are clear differences in PIC, POC quota and growth between ambient and future scenarios. These differences are put into context and discussed well in the manuscript. There is however, one critical point that I think is not well discussed and needs to be considered in the discussion. In the beginning of the discussion, the Authors make the comparison between ocean observations of Coccolithophores and try to highlight discrepancies to the lab experiment. While these comparisons are nearly impossible, because laboratory conditions are so different from natural conditions, I feel that this point merits further discussion. Environmental variation due to different geographical regions affects the environmental history an organism has experienced and thus how an organism can and will react to changes in the environment. In addition within species and within functional group variation in plastic responses of growth and other traits is well established (other publications by Zhang et al have already shown this as well) and can affect how "a species” responds to environmental change. Since the Authors want to make a conclusion what their study tells us about how a cosmospolitan and biogeochemically relevant group of phytoplankton react to future ocean scenarios and not only conclude something about the combination and co-variation of environmental drivers, I feel that ecological variability and environmental variability should be taken into account in the discussion.

One way to approach this "gap" of knowledge could be a discussion about what experimental conditions (based on the given study) could now be focused on to further characterize the responses of other coccolithophore strains that i) come from environmentally different regions, ii) are more recently isolated and thus not acclimatized to long times spent in the laboratory.

Response: We agree with the suggestions from the referee, and have revised the discussion and added related analysis on this aspect with further references to extra literatures at lines 545–560 in the marked-up manuscript version (below): 'Different *E. huxleyi* strains displayed optimal responses to a broad range of temperature or $CO_2$ level, and *E. huxleyi* strains isolated from different regions showed local adaptation to temperature or $CO_2$ level (Zhang et al., 2014; 2018). Strain-specific responses of growth, POC and PIC production rates in *E. huxleyi* isolated from different regions to changing seawater carbonate chemistry have also been documented (Langer et al., 2009). It has been suggested that inter-strain genetic variability has greater potential to induce larger phenotypic differences than the phenotypic plasticity of a single strain cultured under a broad range of variable environmental conditions (Blanco-Ameijeiras et al., 2016). On the other hand, the genetic adaptation to culture experimental conditions over time may no longer accurately represent the cells in the sea, as reflected in a diatom (Guan and Gao, 2008). Phytoplankton species that had been maintained under laboratory conditions might have lost original traits and display different responses to environmental changes (Lakeman et al., 2009). The strain used in this study has been kept in the laboratory for about 30 years, and the data obtained in this work can hardly reflect relation to its biogeographic origin.'

Specific comments:

Line 120: here the Authors imply, that their study will help understand how biogeochemically relevant phytoplankton change in future climate change scenarios but this is not adequately discussed later on (see general comments).

Reponse: We have revised this part as indicated at lines 537–540 in the marked-up manuscript version (below): 'We have to admit that results from laboratory experiments can hardly extrapolate to natural conditions. Nevertheless, our data provide mechanistic understanding of the combined effects of ocean climate change drivers, which can be useful in analyzing field observations.'

Line 161: "adding low light" is misleading. Would it be possible to say that light was reduced?

Response: Agree. We have reworded the words, and changed ‘added’ to ‘supplied’ at lines 164 –167 in the marked-up manuscript version (below).

Line 151 and 161: it would be good to have an idea about nutrient and light concentrations here already. The information following in line 190 comes a bit late and could even be combined as later on the pCO2 manipulation is in focus.

Response: We added one sentence: ‘Initial DIN and DIP concentration were 24 μmol $L^{-1}$ and 1.5 μmol $L^{-1}$, respectively, and initial light intensity was 60 μmol photons $m^{-2}$ $s^{-1}$.’ in lines 154–155 in the marked-up manuscript version (below).

Line 175: I do not fully follow the rationale behind adding the nutrient limitation stepwise

Response: we added “Such stepwise reduction of nutrients levels would be useful for us to analyze effects of nitrate and phosphate separately, and be expected to have implications for the cells episodically exposed to different levels of nutrients in the sea.” in lines 180–183 in the marked-up manuscript version (below).

PIC quota Line 434 ff: I stumbled over the way that the effect of future ocean scenarios are increasing PIC quotas followed by the explanation of how PIC is reduced with increasing pCO2. It would be helpful if there was one more sentence that relates the different results. In addition it could be helpful to highlight in Fig. 2-6, what parts of all of the results are used for the ambient-future comparison. Then the in-between data that are very interesting could become more clear.

Response: We added one sentence: ‘However, the opposite results were found under the elevated $CO_2$ treatment alone.’ in lines 443–444 in the marked-up manuscript version (below). We added these contents ‘The results shown in the black column were used for the ambient-future comparison in figure 2’ in figure legends of figures 3–6 (lines 1056–1057, 1065–1067, 1076–1077, 1086–1087 in the marked-up manuscript version (below)).

Line 531: Please see the general comment: here the discussion should go further because not only oceanic conditions may be different but ecologically within species and functional groups there

are many differences that can affect the results.

Response: We agree with the suggestions of this referee. Please see the response to general comment 1, we have revised the discussion and added related analysis on this aspect with further references to extra literatures (lines 545–560).

Line 612 ff.: I see how considering TEP as part of POC quota is important. But then the Authors also say that it is negligible. As it is written currently, the two sentences contradict each other a bit. Consider rephrasing.

Response: We have deleted these contents 'However, released organic compounds should be negligible, since they are usually photorespiration-dependent (Beardall, 1989; Obata et al., 2013)' after lines 666–667 in the marked-up manuscript version (below)).

Line 620ff: consider moving this part of the discussion about RNA and protein metabolism to where cell cycle is already discussed in line 580. Could fit better together.

Response: Agree. We have moved the contents of the discussion about RNA and protein metabolism to where cell cycle is discussed in lines 616–629 in the marked-up manuscript version (below).

Line 643: The conclusion about competitive ability comes a bit "sudden". Consider mentioning the implications of nutrient uptake on competitive ability earlier in the discussion where phosphate and nutrient uptake related changes are discussed.

Response: Agree. We have move the contents 'While substantial evolutionary responses to multiple drivers may help further, our results imply that decreased phosphate availability along with progressive ocean acidification and warming in surface ocean may reduce the competitive capability of *E. huxleyi* in oligotrophic waters.' to where phosphate uptake is discussed in lines 643–646 in the marked-up manuscript version (below).

Technical/language comments:

Line 234: "taken" should be "took"

Response: Thanks. 'taken' is changed to 'took' in line 239.

Line 571: should say nutrient-replete

Response: Thanks. ‘nutrient-replicate’ is changed to ‘nutrient-replete’ in line 600.

Line 598: On the other hand to what? Please rephrase

Response: ‘On the other hand’ is changed to ‘Meanwhile’ in line 646.

Line 620: type: “a” is missing

Response: Thanks. ‘a’ is added in line 616.

Fig. 1: Please indicate in the legend that experimental steps were done in a consecutive manner. Also this might be helpful to mention again in Fig S1. Visually this implies that the steps are done in parallel, but in the methodological description they are explained as being done one after another.

Response: Thanks. We added these contents ‘Experimental steps were done in a consecutive manner.’ in line 1028, and in lines 1090–1091.

Responses to comment 2 are following:

1. The authors refer the manipulated conditions as "future conditions" in the discussion. Therefore, it would be better to justify why these environmental conditions represent the future global change scenario. For example, the irradiance levels and the nutrient concentrations set up for the experiment are not within the ranges listed in Table S1. The physiological response of *E. huxleyi* would be different under different levels of environmental conditions (i.e. irradiance and nutrient). How will the results of this study be extrapolated to the future global change scenario?

Response: Under the LNLP condition, initial DIN concentration was 8 μmol $L^{-1}$ and initial DIP concentration was about 0.5 μmol $L^{-1}$. During the incubation, DIN and DIP concentrations reduced to about 2.7 μmol $L^{-1}$ and 0.1 μmol $L^{-1}$, respectively, at the end of the incubation (Table 2). DIN and DIP were 0–4.9 μmol $L^{-1}$ and 0.1–0.3 μmol $L^{-1}$, respectively, under the future conditions (Table S1). So, nutrient concentrations were within the ranges listed in Table S1 at the end of the incubation where cell concentration, cellular POC and PIC quotas were measured. In addition, high light intensity was 240 μmol photons $m^{-2}$ $s^{-1}$ during the cultures, and was also within the ranges of irradiance under the future conditions where irradiance was 156–455 μmol photons $m^{-2}$ $s^{-1}$ (Table S1).

We agree that the physiological response of *E. huxleyi* would be different under different levels of environmental conditions. 'We have to admit that results from laboratory experiments can hardly extrapolate to natural conditions. Nevertheless, our data provide mechanistic understanding of the combined effects of ocean climate change drivers, which can be useful in analyzing field observations.' These contents were added in lines 537–540 in the marked-up manuscript version (below).

2. The coccolithophore *Emiliania huxleyi* is a cosmopolitan species. Previous studies have shown strain-specific responses of *E. huxleyi* to environmental changes (especially ocean acidification). I would suggest the authors to expand the discussion on Table 5 a little further.

Response: As mentioned in response to general comment 1, strain-specific responses in growth rate, POC and PIC production rates of *E. huxleyi* to a range of $CO_2$ or temperature have been reported by Langer et al. (2009) and Zhang et al. (2014; 2018). In addition, Blanco-Ameijeiras et al. (2016) examined variability in cellular contents of POC and PIC, magnesium (Mg) and strontium (Sr) of 13 *E. huxleyi* strains under identical culture conditions. We added related analysis on this aspect with further references to extra literatures in lines 545–560 in the marked-up manuscript version (below).

Some other specific comments:

Lines 163: "low nitrogen was added: : :" I don't think this is a correct expression of introducing low nitrate concentration. Could the authors also specify how the nitrate concentration was reduced? The same for line 164, "low phosphate was added..".

Response: Agree. We changed 'added' to 'supplied' in lines 163–167.

Line 269: The cell diameter was measured for the whole coccosphere, with coccoliths attached. However, both PIC quota and PIC/POC ratio was changed by different experimental manipulations, especially by alteration of pCO2. This would have also resulted in changes in coccolith thickness. I was wondering if the authors have considered this when calculating the cell-volume normalized particulate organic elemental quotas.

Response: We agree with the suggestions of this referee. We have calculated the cell-volume normalized POC and PIC quotas in figures S6 and S7. POC (or PIC) quota and the cell-volume normalized POC (or PIC) quota showed similar trends in response to different environmental conditions (Figures 4; 5; S6; S7). We added 'and the cell-volume normalized POC quotas' in line 653 in the marked-up manuscript version (below).

Lines 527-531: This sentence is too long, please split to two.

Response: This sentence was reduced to 'Our results from laboratory experiments with multiple drivers experiment instead predicted a different trend with progressive ocean climate changes.' in lines 533–535 in the marked-up manuscript version (below).

Line 556: "low-pH inhibited growth.." Here the authors indicate it was mainly the effects of pH instead of changing pCO2, please add some explanations on this.

Response: We added these contents 'In ocean acidification condition, the negative effect of low pH on growth rate of the same *E. huxeyi* strain PML B92/11 was larger than the positive effect of high $CO_2$ concentration (Bach et al., 2011). Our data further showed that low-pH inhibited growth to lesser extent under the high light than under low light (Fig. 3e; Table 2).' in lines 581–585 in the marked-up manuscript version (below).

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Bach, L. T., Riebesell, U., and Schulz, K. G.: Distinguishing between the effects of ocean acidification and ocean carbonation in the coccolithophore *Emiliania huxleyi*, Limnol. Oceanogr., 56, 2040–2050, doi: 10.4319/lo.2011.56.6.2040, 2011.

Line 559: Please add a reference after the sentence "photosynthesis under the high light regime could generate more energy-conserving compounds".

Response: Fernández et al. (1996) reported that high light intensity facilitates carbohydrate accumulation and low light intensity reduces cellular carbohydrate content. So we cited this reference in line 587.

Fernández, E., Fritz, J. J., and Balch, W. M.: Chemical composition of the coccolithophorid *Emiliania huxleyi* under light-limited steady state growth. J. Exp. Mar. Biol. Ecol., 207, 149–160.

doi: 10.1016/S0022-0981(96)02657-3, 1996.

Line 561: Please specify the strain of the *E. huxleyi* examined in Jin et al., as well as in line 569.
Response: Jin et al. (2017) examine responses of growth rate, POC and PIC quotas of *E. huxleyi* strains PML B92/11 and CCMP 2090 under different levels of incident solar radiation. '*E. huxleyi*' was replaced by '*E. huxleyi* strains PML B92/11 and CCMP 2090' in line 589, and by '*E. huxleyi* strain PML B92/11' in line 598.

Line 575: Why was PIC quota increased under high light? Please add some explanations.
Response: One explanation could be that high light intensity makes cells to remove $H^+$ faster and then reduce the negative effects of low pH on calcification of *E. huxleyi* (Jin et al., 2017). These contents 'increased light levels can partially counteract the negative effects of OA on calcification' were changed to 'high light intensity could make cells to remove $H^+$ faster and then reduce the negative effects of low pH on calcification of *E. huxleyi* (Jin et al., 2017)' in lines 606–607 in the marked-up manuscript version (below).

Line 617: The sentence "released organic compounds should be negligible: : :" contradicts to the previous expression of "over-synthesis of cellular organic carbon might be released as dissolved organic carbon.." in lines 612-613.

Response: Thanks. We have deleted these contents 'However, released organic compounds should be negligible, since they are usually photorespiration-dependent (Beardall, 1989; Obata et al., 2013)' in lines 666–667.

Fig. 1 Please label the symbols in the graph for a better understanding of the treatments.
Response: Thanks. We have done in Figure 1

Fig. S1 I think it would be better to move this figure to the main manuscript, instead of being in the supplementary materials, in order to make a better understanding of the step-wise experimental design.
Response: We try to move the figure S1 to the main manuscript, whereas we find that figure S1 and figure 1 seem to repeat in terms of the experiment setup. So we would like to keep the figure S1 in the supplemental information. If the referee persists in, we will do it.

Fig. S11 How was the RNA concentration measured? This is not presented in the methods section.
Response: The sentence: 'In this study, RNA content per cell was verified by a SYBR Green method (Berdalet et al., 2005).' is added in line 617–618 in the marked-up manuscript version (below).

Berdalet, E., Roldán, C., Olivar, M. P., and Lysnes, K.: Quantifying RNA and DNA in planktonic organisms with SYBR Green II and nucleases. Part A. Optimisation of the assay, Sci. Mar., 69, 1–16, doi: 10.3989/scimar.2005.69n11, 2005.

## A list of all relevant changes

### Materials and Methods:

**Lines 154–155:** Add 'Initial DIN and DIP concentrations were 24 μmol $L^{-1}$ and 1.5 μmol $L^{-1}$, respectively, and initial light intensity was 60 μmol photons $m^{-2}$ $s^{-1}$.'

**Lines 163–164:** Add ', 60 μmol photons $m^{-2}$ $s^{-1}$' and ', 240 μmol photons $m^{-2}$ $s^{-1}$'.

**Lines 164–167:** 'added' is changed to 'supplied'.

**Lines 180–183:** Add 'Such stepwise reduction of nutrients levels would be useful for us to analyze effects of nitrate and phosphate separately, and be expected to have implications for the cells episodically exposed to different levels of nutrients in the sea.'

**Line 239:** 'taken' is changed to 'took'.

### Results

**Lines 443–444:** Add a sentence: 'However, the opposite results were found under the elevated $CO_2$ treatment alone.'

### Discussion

**Line 535:** 'change' is changed to 'changes'.

**Lines 535–537:** Delete ', suggesting that some key elements of understanding phytoplankton responses to changing conditions that would enable researchers to connect laboratory studies and field observations are missing.'

**Lines 537–540:** Add these contents: 'We have to admit that results from laboratory experiments can hardly extrapolate to natural conditions. Nevertheless, our data provide mechanistic understanding of the combined effects of ocean climate change drivers, which can be useful in analyzing field observations.'

**Lines 545–560:** Add these contents: 'Different *E. huxleyi* strains displayed optimal responses to a broad range of temperature or $CO_2$ level, and *E. huxleyi* strains isolated from different regions showed local adaptation to temperature or $CO_2$ level (Zhang et al., 2014; 2018). Strain-specific responses of growth, POC and PIC production rates in *E. huxleyi* isolated from different regions to changing seawater carbonate chemistry have also been documented (Langer et al., 2009). It has been suggested that inter-strain genetic variability has greater potential to induce larger phenotypic

differences than the phenotypic plasticity of a single strain cultured under a broad range of variable environmental conditions (Blanco-Ameijeiras et al., 2016). On the other hand, the genetic adaptation to culture experimental conditions over time may no longer accurately represent the cells in the sea, as reflected in a diatom (Guan and Gao, 2008). Phytoplankton species that had been maintained under laboratory conditions might have lost original traits and display different responses to environmental changes (Lakeman et al., 2009). The strain used in this study has been kept in the laboratory for about 30 years, and the data obtained in this work can hardly reflect relation to its biogeographic origin.'

**Lines 581–584:** Add these contents: ' In ocean acidification condition, the negative effect of low pH on growth rate of the same *E. huxeyi* strain PML B92/11 was larger than the positive effect of high $CO_2$ concentration (Bach et al., 2011). Our data further showed that'.

**Line 584:** Delete 'Interestingly,'

**Line 587:** Add '(Fernández et al., 1996).', and change 'which' to 'This'.

**Line 589:** Delete 'was' and add 'strains PML B92/11 and CCMP 2090'.

**Line 590:** Add 'were'.

**Line 598:** Add 'strain PML B92/11'.

**Line 600:** Change 'replicate' to 'replete'.

**Lines 605–606:** Delete: 'increased light levels can partially counteract the negative effects of OA on calcification'.

**Lines 606–607:** Add 'high light intensity could make cells to remove $H^+$ faster and then reduce the negative effect of low pH on calcification of *E. huxleyi* (Jin et al., 2017)'

**Lines 616–629:** Add these contents: 'Synthesis of RNA is a large biochemical sink for phosphate in *E. huxleyi* and other primary producers (Dyhrman, 2016). In this study, RNA content per cell was verified by a SYBR Green method (Berdalet et al., 2005). Compared to HNHP conditions, HNLP-grown cells had only 7.8% of total RNA (Fig. S11). This indicates that decreased availability of phosphate strongly decreased RNA synthesis, which would consequently extend the interphase of the cell cycle where calcification occurs (Müller et al., 2008). This could explain why PIC quotas were enhanced by decreased phosphate availability (Fig. 5). Similarly, decreased

availability of nitrate decreased protein (or PON) synthesis (Fig. S10), which can also block cells in the interphase of the cell cycle, and increase the time available for calcification in *E. huxleyi* (Vaulot et al., 1987). Consistently with this, lower rates of assimilation or organic matter production in *E. huxleyi* in LNHP than in HNHP treatments are consistent with more energy being reallocated to use for calcification (Nimer and Merrett, 1993; Xu and Gao, 2012).'

**Lines 643–646:** Add these contents: 'While substantial evolutionary responses to multiple drivers may help further, our results imply that decreased phosphate availability along with progressive ocean acidification and warming in surface ocean may reduce the competitive capability of *E. huxleyi* in oligotrophic waters.'

**Line 646:** 'On the other hand' is changed to 'Meanwhile'.

**Line 653:** Add 'and the cell-volume normalized POC quotas'.

**Line 654:** Add 's' and 'S6;'.

**Lines 666–667:** Delete these contents: 'However, released organic compounds should be negligible, since they are usually photorespiration-dependent (Beardall, 1989; Obata et al., 2013).'

**Lines 668–679:** Delete these contents: 'Synthesis of RNA is a large biochemical sink for phosphate in *E. huxleyi* and other primary producers (Dyhrman, 2016). Compared to HNHP conditions, HNLP-grown cells had only 7.8% of total RNA (Fig. S11). This indicates that decreased availability of phosphate strongly decreased RNA synthesis, which would consequently extend the interphase of the cell cycle where calcification occurs (Müller et al., 2008). This could explain why PIC quotas were enhanced by decreased phosphate availability (Fig. 5). Similarly, decreased availability of nitrate decreased protein (or PON) synthesis (Fig. S10), which can also block cells in the interphase of the cell cycle, and increase the time available for calcification in *E. huxleyi* (Vaulot et al., 1987). Consistently with this, lower rates of assimilation or organic matter production in *E. huxleyi* in LNHP than in HNHP treatments are consistent with more energy being reallocated to use for calcification (Nimer and Merrett, 1993; Xu and Gao, 2012).'

**Lines 688–692:** Delete these contents: 'While substantial evolutionary responses to multiple drivers may help further, our results imply that decreased phosphate availability along with progressive ocean acidification and warming in surface ocean may reduce the competitive capability of *E. huxleyi* in oligotrophic waters.'

## References

**Lines 724–727:** Add ‘Bach, L. T., Riebesell, U., and Schulz, K. G.: Distinguishing between the effects of ocean acidification and ocean carbonation in the coccolithophore *Emiliania huxleyi*, Limnol. Oceanogr., 56, 2040–2050, doi: 10.4319/lo.2011.56.6.2040, 2011.’

**Lines 731–734:** Add ‘Berdalet, E., Roldán, C., Olivar, M. P., and Lysnes, K.: Quantifying RNA and DNA in planktonic organisms with SYBR Green II and nucleases. Part A. Optimisation of the assay, Sci. Mar., 69, 1–16, doi: 10.3989/scimar.2005.69n11, 2005.’

**Lines 735–736:** Delete ‘Beardall, J.: Photosynthesis and photorespiration in marine phytoplankton, Aquat. Bot., 34, 105–130, doi: 10.1016/0304-3770(89)90052-1, 1989.’

**Lines 801–804:** Add ‘Fernández, E., Fritz, J. J., and Balch, W. M.: Chemical composition of the coccolithophorid *Emiliania huxleyi* under light-limited steady state growth. J. Exp. Mar. Biol. Ecol., 207, 149–160. doi: 10.1016/S0022-0981(96)02657-3, 1996.’

**Lines 822–825:** Add ‘Guan, W., and Gao, K.: Light histories influence the impacts of solar ultraviolet radiation on photosynthesis and growth in a marine diatom, *Skeletonema costatum*, J. Photoch. Photobio. B., 91, 151–156, doi: 10.1016/j.jphotobiol.2008.03.004, 2008.’

**Lines 852–854:** Add ‘Lakeman, M. B., von Dassow, P., and Cattolico, R. A.: The strain concept in phytoplankton ecology, Harmful Algae, 8, 746–758, doi: 10.1016/j.hal.2008.11.011, 2009.’

**Lines 855–857:** Add ‘Langer, G., Nehrke, G., Probert, I., Ly, J., and Ziveri, P.: Strain-specific responses of *Emiliania huxleyi* to changing seawater carbonate chemistry, Biogeosciences, 6, 2637–2646, doi: 10.5194/bg-6-2637-2009.’

**Lines 905–908:** Delete ‘Obata, T., Schoenefeld, S., Krahnert, I., Bergmann, S., Scheffel, A., and Fernie, A. R.: Gas-chromatography mass-spectrometry (GC-MS) based metabolite profiling reveals mannitol as s major storage carbohydrate in the coccolithophorid alga *Emiliania huxleyi*, Metabolites, 3, 168–184, doi: 10.3390/metabo3010168, 2013.’

**Lines 996–999:** Add ‘Zhang, Y., Bach, L. T., Lohbeck, K. T., Schulz K. G., Listmann, L., Klapper R., and Riebesell, U.: Population-specific responses in physiological rates of *Emilinia huxleyi* to a broad $CO_2$ range, Biogeosciences, 15, 3691–3701, doi: 10.5194/bg-15-3691-2018, 2018.’

**Lines 1003–1006:** Add ‘Zhang, Y., Klapper, R., Lohbeck, K. T., Bach, L. T., Schulz, K. G., Reusch, T. B. H., and Riebesell, U.: Between- and within-population variations in thermal reaction

norms of the coccolithophore *Emiliania huxleyi*, Limnol. Oceanogr., 59, 1570–1580, doi: 10.4319/lo.2014.59.5.1570, 2014.'

## Figure Legends

**Lines 1023–1026:** 'added' is changed to 'supplied'.

**Lines 1028:** Add 'Experimental steps were done in a consecutive manner.'

**Lines 1056–1057, 1065–1067, 1076–1077, 1086–1087:** Add 'The results shown in the black column were used for the ambient-future comparison in figure 2.'

**Lines 1090–1091:** Add 'Experimental steps were done in a consecutive manner.'

**Lines 1251–1268:** Change 'Figure 1'.

**Lines 1276–1287:** Change 'Figure 2'.

**Lines 1290–1310:** Change 'Figure 3'.

**Lines 1319–1337:** Change 'Figure 4'.

**Lines 1347–1367:** Change 'Figure 5'.

**Lines 1377–1395:** Change 'Figure 6'.

# Reduced growth with increased quotas of particulate organic and inorganic carbon in the coccolithophore *Emiliania huxleyi* under future ocean climate change conditions

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Running head: Response of *E. huxleyi* to multiple drivers

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Keywords: $CO_2$; coccolithophore; functional trait plasticity; light; multiple drivers; nutrients; ocean acidification; warming.

**Abstract**

Effects of ocean acidification and warming on marine primary producers can be modulated by other environmental factors, such as levels of nutrients and light. Here, we investigated the interactive effects of five oceanic environmental drivers ($CO_2$, temperature, light, dissolved inorganic nitrogen and phosphate) on growth rate, particulate organic (POC) and inorganic (PIC) carbon quotas of the cosmopolitan coccolithophore *Emiliania huxleyi*. Population growth rate increased with increasing temperature (16 to 20 °C) and light intensities (60 to 240 µmol photons $m^{-2}$ $s^{-1}$), but decreased with elevated $pCO_2$ concentrations (370 to 960 µatm) and reduced availability of nitrate (24.3 to 7.8 µmol $L^{-1}$) and phosphate (1.5 to 0.5 µmol $L^{-1}$). POC quotas were predominantly enhanced by combined effects of increased $pCO_2$ and decreased availability of phosphate. PIC quotas increased with decreased availability of nitrate and phosphate. Our results show that concurrent changes in nutrient concentrations and $pCO_2$ levels predominantly affected growth, photosynthetic carbon fixation and calcification of *E. huxleyi*, and imply that plastic responses to progressive ocean acidification, warming and decreasing availability of nitrate and phosphate reduce population growth rate while increasing cellular quotas of particulate organic and inorganic carbon of *E. huxleyi*, ultimately affecting coccolithophore-related ecological and biogeochemical processes.

## 1 Introduction

Ocean acidification (OA), due to continuous oceanic absorption of anthropogenic $CO_2$, is occurring alongside ocean warming. This in turn, leads to shoaling in the upper mixed layer (UML) and a consequent reduction in the upward transport of nutrients into the UML. These ocean changes expose phytoplankton cells within the UML to multiple simultaneous stressors or drivers, and organismal responses to these drivers can affect both trophic and biogeochemical roles of phytoplankton (see reviews by Boyd et al., 2015; Gao et al., 2019 and literatures therein). While most studies on the effects of ocean global climate changes on marine primary producers have focused on organismal responses to one, two or three environmental drivers, there is an increasing awareness of the need to measure the combined effects of multiple drivers (see reviews by Riebesell and Gattuso, 2015; Boyd et al., 2018; Gao et al., 2019; Kwiatkowski et al., 2019). For this purpose, several manipulative experimental approaches have been recommended (Boyd et al., 2018). One approach using many unique combinations of different numbers of drivers showed that both short and long-term growth responses were, on average, explained by the dominant single driver in a multi-driver environment, but this result relies on having many (>5) drivers with known or measured large-effect single drivers (Brennan and Collins, 2015; Brennan et al., 2017). For experiments with multiple drivers where interactions are likely to preclude making predictions from single drivers, where average responses are not the most informative ones, or where logistics preclude using a very large number of multi-driver environments, Boyd et al. (2010) suggested an 'environmental cluster' method where key drivers (such as temperature, light intensity, nutrient concentration, $CO_2$ and Fe) are covaried within experiments, allowing the investigation of physiological responses of phytoplankton to concurrent changes of the clustered

drivers. This approach examines responses to projected overall environmental shifts rather than pulling apart the biological or statistical interactions between responses to individual drivers. To our knowledge, studies to date have employed such a driver clustering approach to investigate responses of diatoms *Fragilariopsis cylindrus*, *Thalassiosira pseudonana*, *Skeletonema costatum*, and the prymnesiophyte *Phaeocystis antarctica* to combinations of drivers projected for 2100 (Xu et al., 2014a; Xu et al., 2014b; Boyd et al., 2016).

An environmental cluster approach is especially useful when drivers are known to interact in terms of the organismal responses they elicit, as is the case for OA, light levels, and key nutrients acting on population growth rate and carbon fixation (Boyd et al., 2016). For example, in the cosmopolitan coccolithophore *Emiliania huxleyi*, interactive effects of OA and light showed that OA increased population growth rate and photosynthetic carbon fixation under low light, whereas it slightly lowered population growth rate and photosynthetic carbon fixation under high light (Zondervan et al., 2002; Kottmeier et al., 2016). In addition, photosynthetic carbon fixation was further enhanced by longer light exposure at high $pCO_2$ levels (Zondervan et al., 2002). On the other hand, OA can exacerbate the negative impact of solar UV radiation on photosynthetic carbon fixation and calcification in *E. huxleyi* under nutrient-replete conditions (Gao et al., 2009), but can increase calcification (coccolith volume) and particulate organic carbon (POC) quota under phosphate-limited conditions (Leonardos and Geider, 2005; Müller et al., 2017), demonstrating that the effects of OA on calcification is likely nutrient-dependent. On the other hand, ocean warming, which occurs alongside OA, is known to increase coccolith length, POC, particulate organic nitrogen (PON) and inorganic carbon (PIC) production rates of several *E. huxleyi* strains (Rosas-Navarro et al., 2016; Feng et al., 2017). Warming

has also been shown to increase the optimal $pCO_2$ levels for growth, POC and PIC production rates (Sett et al., 2014). In one case warming was found to compensate for the negative impact of OA on growth rate under low light intensity (Feng et al., 2008). Nevertheless, decreased photosynthetic carbon fixation and calcification at reduced carbonate saturation state (lowered $Ca^{2+}$ concentrations) were exacerbated by warming treatment (Xu et al., 2011). Overall, there is strong evidence that understanding the plastic responses of this key calcifier to ocean changes requires investigating responses to the overall expected shift in the environment, in addition to the detailed studies to date on individual drivers, due to the sheer number of interactions between individual drivers on traits that affect the trophic and biogeochemical roles of *E.huxleyi.*

Despite known interactions among two- and three-way combinations of OA, temperature, light, phosphate levels and nitrogen levels, there have been few empirical studies investigating effects of the larger cluster projected for future surface ocean changes. The data to date show that interactions among drivers can affect both the direction and magnitude of trait changes in biogeochemically important taxa. In addition, based on single or two-driver studies, changes in temperature, $pCO_2$, light, dissolved inorganic nitrogen (DIN) and phosphate (DIP) in combination are predicted to affect primary productions (Barton et al., 2016; Monteiro et al., 2016; Boyd et al., 2018; Gao et al., 2019; Kwiatkowski et al., 2019). Understanding the trait-based responses of cocolithophores to future ocean changes is important for projections of changes in the biogeochemical roles of phytoplankton, such as biological carbon pump efficiency (Rost and Riebesell, 2004).

In order to understand the combined effects of $pCO_2$, temperature, light, dissolved inorganic nitrogen (DIN) and phosphate (DIP) on functional traits, we incubated

*Emiliania huxleyi* (Lohmann) under different combinations of environmental conditions that represented subsets of, and eventually the complete set of environments for, this environmental driver cluster. We recently examined the interactive effects of light intensity and $CO_2$ level on growth rate, POC and PIC quotas of *E. huxleyi* under nutrients replete, low DIN, or low DIP concentrations (Zhang et al., 2019). Light, $CO_2$, DIN and DIP levels usually change simultaneously with temperature, and temperature modulated responses of *E. huxleyi* to other environmental drivers (Gafar and Schulz, 2018; Tong et al., 2019). In addition, warming or cooling can direcly influence the activity of enzymes, thus directly modulating metabolic rates (Sett et al., 2014). Because of the overwhelming evidence that temperature can act as a general modulator of organismal responses, we use the present study to examine how the addition of temperature as a key driver in the environmental change cluster can modulate the combined effects of $CO_2$, light and nutrients. We found that future ocean scenario treatments with OA, warming, increased light and reduced availability of nutrients led to lower growth rate and larger POC and PIC quotas of *E. huxleyi*.

## 2 Materials and Methods

### 2.1 Experimental setup

*Emiliania huxleyi* strain PML B92/11 was originally isolated from coastal waters off Bergen, Norway, and obtained from the Plymouth algal culture collection, UK. The average levels of $pCO_2$, temperature, light, dissolved inorganic nitrate (DIN) and phosphate (DIP) were set up according to recorded data in Norwegian coastal waters during 2000 to 2007 and projected for 2100 in high-latitudes (Larsen et al., 2004; Locarnini et al., 2006; Omar et al., 2010; Boyd et al., 2015) (Table S1). *E. huxleyi* was

cultured with a 12 h/12 h light/dark cycle in thermo-controlled incubators in Aquil medium, which was prepared according to Sunda et al. (2005) with the addition of 2200 µmol $L^{-1}$ bicarbonate to achieve the total alkalinity (TA) of 2200 µmol $L^{-1}$. Initial DIN and DIP concentrations were 24 µmol $L^{-1}$ and 1.5 µmol $L^{-1}$, respectively, and initial light intensity was 60 µmol photons $m^{-2}$ $s^{-1}$. The experiment was conducted in five steps (Fig. 1). Considering ocean acidification and warming as the key drivers for ocean climate changes, we first established 4 "baseline" treatments where the $pCO_2$ and temperature drivers were combined in a fully factorial way: low $pCO_2$ + low temperature (LCLT), high $pCO_2$ + low temperature (HCLT), low $pCO_2$ + high temperature (LCHT), and high $pCO_2$ + high temperature (HCHT). Since reduced availability of nutrients and increased light exposures are triggered by warming-enhanced stratification, we then added additional single or pairs of drivers to each of these "baseline" treatments (Fig. S1). In step 1, low light (LL, 60 µmol photons $m^{-2}$ $s^{-1}$) was ~~added~~ supplied; in step 2, high light (HL, 240 µmol photons $m^{-2}$ $s^{-1}$) was ~~added~~ exposed. HL was then maintained for the rest of the experiment. In step 3, low nitrogen was ~~added~~ supplied and high phosphate levels were maintained (LNHP). In step 4, low phosphate was ~~added~~ supplied and high nitrogen levels were restored (HNLP). In step 5, both nitrogen and phosphate were low (LNLP), respectively (Figs. 1 and S1). In all cases, the cells were acclimated to each unique stressor cluster for at least 14–16 generations before physiological and biochemical parameters were measured. Although this stepwise design introduces a historical effect, physiological traits are generally reported after 10 to 20 generations acclimation to OA treatment (Perrin et al., 2016; Tong et al., 2016; Li et al., 2017), so the historical effects here are similar to those that would be introduced with standard methods in other physiology studies (Tong et al., 2016; Zhang et al., 2019). Since individually reduced availability

of nitrate or phosphate decreased growth, did not change POC quota, and enhanced PIC quota under optimal light intensity (HL in this study) in the same *E. huxleyi* strain (Zhang et al., 2019), we hypothesized that combination of DIN and DIP limitation would result in similar trend under the $pCO_2$ and/or temperature combined treatments. Therefore, we added stepwise nitrate and/or phosphate drivers (Fig. 1). Such stepwise reduction of nutrients levels would be useful for us to analyze effects of nitrate and phosphate separately, and be expected to have implications for the cells episodically exposed to different levels of nutrients in the sea.

For step 1, $NO_3^-$ and $PO_4^{3-}$ were modified to 24 μmol $L^{-1}$ and 1.5 μmol $L^{-1}$, respectively, which is the HNHP treatment in the synthetic seawater (Sunda et al., 2005) (Fig. S1). The seawater was dispensed into 4 glass bottles, and 2 bottles of seawater were placed at 16 °C (LT) in an incubator (HP400G-XZ, Ruihua, Wuhan), and aerated for 24 h with filtered (PVDF 0.22 μm pore size, Haining) air containing 400 μatm (LC) or 1000 μatm $pCO_2$ (HC). Another 2 bottles of seawater were maintained at 20 °C (HT) in the other chamber and also aerated with LC or HC air as described above. The dry air/$CO_2$ mixture was humidified with deionized water prior to the aeration to minimize evaporation. The LCLT, HCLT, LCHT and HCHT seawaters (Figs. 1a and S1) were then filtered (0.22 μm pore size, Polycap 75 AS, Whatman) and carefully pumped into autoclaved 250 mL polycarbonate bottles (Nalgene, 4 replicate flasks for each of LCLT, HCLT, LCHT and HCHT, a total of 16 flasks at the beginning of the experiment) with no headspace to minimize gas exchange. The flasks were inoculated at a cell density of about 150 cells $mL^{-1}$. The volume of the inoculum was calculated (see below) and the same volume of seawater was taken out from the bottles before inoculation. The samples were initially cultured at 60 μmol photons $m^{-2}$ $s^{-1}$ (LL) of photosynthetically active radiation (PAR)

(measured using a PAR Detector, PMA 2132 from Solar Light Company) under LCLT, HCLT, LCHT and HCHT conditions for 8 generations (6 days) (d), and then the samples were diluted to their initial concentrations and grown for another 8 generations (6 d) (Fig. 1a). Samples in culture bottles were mixed twice a day at 9:00 a.m. and 5:00 p.m. At the end of the incubation, sub-samples were taken for measurements of cell concentration, POC and TPC quotas, TA, pH and nutrient concentrations.

In step 2, samples grown under the previous conditions were transferred at the end of the cultures from 60 (LL) to 240 μmol photons $m^{-2}$ $s^{-1}$ (HL) of PAR with initial cell concentrations of 150 cells $mL^{-1}$, and acclimated to the HL for 8 generations (5 d in 16 °C environment, 4 d in 20 °C environment) (Fig. 1b). The cultures were then diluted to achieve initial cell concentration and incubated at the HL for another 8 generations (the fifth day in 16 °C environment and the fourth day in 20 °C environment) before sub-samples were taken for measurements.

In step 3, step 4 and step 5, $NO_3^-$ and $PO_4^{3-}$ concentrations were set to be 8 μmol $L^{-1}$ and 1.5 μmol $L^{-1}$ for the LNHP treatment, and 24 μmol $L^{-1}$ and 0.5 μmol $L^{-1}$ for the HNLP treatment, and 8 μmol $L^{-1}$ and 0.5 μmol $L^{-1}$ for the LNLP treatment, respectively (Fig. 1c,d,e). The LCLT, HCLT, LCHT and HCHT were step 1 conditions, now we are into step 3, 4 and 5. Under 240 μmol photons $m^{-2}$ $s^{-1}$ (HL) of PAR, cell samples with an initial concentration of 150 cells $mL^{-1}$ were transferred from HNHP condition (step 2) to LNHP conditions (step 3) and acclimated to LNHP conditions for 8 generations (5 d in 16 °C environment, 4 d in 20 °C environment) (Fig. 1c). The cultures were then diluted back to initial cell concentrations and incubated in the LNHP conditions (step 3) for a further 8 generations. On the last day

of the incubation (the fifth day in 16 ºC environment and the fourth day in 20 ºC environment), sub-samples were taken for measurements of the parameters.

After that, cell samples were transferred stepwise from HNHP conditions (step 2, Fig. 1b) to HNLP conditions (step 4, Fig. 1d), then from HNLP conditions to LNLP conditions (step 5, Fig. 1e). Cell samples were acclimated for 8 generations at HNLP and LNLP conditions, respectively, and followed by another 8 generation incubations for 4 d at HT and 5 d at LT. On the fourth day (for populations in high temperature environments) or the fifth day (for populations in low temperature environments), sub-samples were taken for measurements (Fig. 1d,e). At low nutrient concentrations, maximal cell concentrations were limited by nutrients (Rouco et al., 2013; Rokitta et al., 2016). To check whether cells sampled were in exponential growth at each nutrient level, we examined cell concentrations every day at LCHT, or LCLT and high light conditions (Fig. S2). We found that cell concentrations were in the exponential growth phase during the 1$^{st}$ and 5$^{th}$ days at HT, and during the 1$^{st}$ and 7$^{th}$ days at LT. In this study, we ~~taken~~ took samples in the 4$^{th}$ day at HT and in the 5$^{th}$ day at LT, and thus cells sampled were in the exponential growth phase of *E. huxleyi*.

In the previous work (Zhang et al., 2019), we transferred *E. huxleyi* cells stepwise from 80 µmol photons $m^{-2}$ $s^{-1}$ to 120 µmol photons $m^{-2}$ $s^{-1}$, then to 200 µmol photons $m^{-2}$ $s^{-1}$, to 320 µmol photons $m^{-2}$ $s^{-1}$ and to 480 µmol photons $m^{-2}$ $s^{-1}$ at both LC and HC levels under HNHP, LNHP or HNLP conditions, respectively. In this study, we transferred the same strain from LL to HL under HNHP condition, and then from HNHP to LNHP or HNLP, and from HNLP to LNLP under HL conditions under 4 "baseline" $CO_2$ and temperature treatments, in an effort to elucidate interactive and combined effects of temperature, $CO_2$, DIN and DIP (Table S2), in contrast the previous work carried out under constant temperature (Zhang et al., 2019).

## 2.2 Nutrient concentrations and carbonate chemistry measurements

In the first and last days of the incubations, 20 mL samples for determination of inorganic nitrogen and phosphate concentrations were taken at the same time using a filtered syringe (0.22 μm pore size, Haining) and measured by using a scanning spectrophotometer (Du 800, Beckman Coulter) according to Hansen and Koroleff (1999). The nitrate was reduced to nitrite by zinc cadmium reduction and then total nitrite concentration was measured. In parallel, 25 mL samples were taken for determination of total alkalinity (TA) after being filtered (0.22 μm pore size, Syringe Filter) under moderate pressure using a pump (GM-0.5A, JINTENG) and stored in the dark at 4 °C for less than 7 d. TA was measured at 20 °C by potentiometric titration (AS-ALK1+, Apollo SciTech) according to Dickson et al. (2003). Samples for $pH_T$ (total scale) determinations were syringe-filtered (0.22 μm pore size), and the bottles were filled from bottom to top with overflow and closed immediately without headspace. The $pH_T$ was immediately measured at 20 °C by using a pH meter (Benchtop pH, Orion 8102BN) which was calibrated with buffers (Tris•HCl, Hanna) at pH 4.01, 7.00 and 10.00. Carbonate chemistry parameters were calculated from TA, $pH_T$, phosphate (at 1.5 μmol $L^{-1}$ or 0.5 μmol $L^{-1}$), temperature (at 16 °C or 20 °C), and salinity using the $CO_2$ system calculation in MS Excel software (Pierrot et al., 2006). $K_1$ and $K_2$, the first and second carbonic acid constants, were taken from Roy et al. (1993).

## 2.3 Cell concentration measurements

In the last day of the incubation, ~25 mL samples (8 samples) were taken at the same time (about 1:00 p.m.). Cell concentration and cell diameter (D) were measured using

a Z2 Coulter Particle Count and Size Analyzer (Beckman Coulter). The diameter of detected particles was set to be 3 to 7 μm in the instrument, which excludes detached coccoliths (Müller et al., 2012). Cell concentration was also measured by microscopy (ZEISS), and variation in measured cell concentration between two methods was ± 7.9% (Zhang et al., 2019). Average growth rate ($\mu$) was calculated for each replicate according to the equation: $\mu = (\ln N_1 - \ln N_0) / d$, where $N_0$ was 150 cells $mL^{-1}$ and $N_1$ was the cell concentration in the last day of the incubation, $d$ was the growth period in days. *E. huxleyi* cells were spherical and its cell volume with coccoliths was calculated according to the equation: $V = 3.14 \times (4/3) \times (D/2)^3$.

**2.4 Total particulate (TPC) and particulate organic (POC) carbon measurements**

100 mL samples for determination of TPC and POC quotas were filtered onto GF/F filters (pre-combusted at 450 °C for 6 h) at the same time in each treatment. TPC and POC samples were stored in the dark at –20 °C. For POC measurements, samples were fumed with HCl for 12 h to remove inorganic carbon, and samples for TPC measurements were not treated with HCl. All samples were dried at 60 °C for 12 h, and analyzed using a Thermo Scientific FLASH 2000 CHNS/O elemental analyzer (Thermo Fisher, Waltham, MA). Particulate inorganic carbon (PIC) quota was calculated as the difference between TPC quota and POC quota. POC and PIC production rates were calculated by multiplying cellular contents with $\mu$ ($d^{-1}$), respectively. Variations in measured carbon content between the four replicates were calculated to be 1–24% in this study.

**2.5 Data analysis**

299 Firstly, we examined the interactions of temperature, $pCO_2$ and light under nutrient-
300 replete (HNHP) conditions. Here, the effects of temperature, $pCO_2$, light intensity and
301 their interaction on growth rate, POC and PIC quotas were tested using a three-way
302 analysis of variance (ANOVA). Secondly, we examined the effects of nutrient
303 limitation in the different $pCO_2$ and temperature environments under the high light
304 intensity (HL). Here, the effects of temperature, $pCO_2$, dissolved inorganic nitrogen
305 (DIN), dissolved inorganic phosphate (DIP) and their interaction on growth rate, POC
306 and PIC quotas were tested using a four-way ANOVA. Finally, a one-way ANOVA
307 was used to test the differences in growth rate, POC and PIC quotas between present
308 (defined as low levels of $pCO_2$, temperature and light along with high levels of DIN
309 and DIP (LC LT LL HN HP)) and future ocean (defined as higher levels of $pCO_2$,
310 temperature, and light along with low levels of DIN and DIP (HC HT HL LN LP))
311 scenarios. A Tukey post hoc test was performed to identify the differences between
312 two temperatures, two $pCO_2$ levels, two DIN or two DIP treatments. Normality of
313 residuals was conducted with a Shapiro-Wilk's test, and a Levene test was conducted
314 graphically to test for homogeneity of variances. A generalized least squares (GLS)
315 model was used to stabilize heterogeneity if variances were non-homogeneous. All
316 statistical calculations were performed using *R* (R version 3.5.0).

317 In order to quantify the individual effect of nitrate concentration or phosphate
318 concentration on the physiological and biochemical parameters, we calculated the
319 change ratio ($R$) of physiological rates according to the equation: $R = | M_{\text{LNHP or HNLP}}$
320 $- M_{\text{HNHP}} | / M_{\text{HNHP}}$, where $M_{\text{LNHP or HNLP or HNHP}}$ respresents measured trait values in
321 LNHP or HNLP or HNHP conditions, and the '|' denotes the absolute value
322 (Schaum et al., 2013). We then calculated the expected growth rate, POC quota and
323 PIC quota in LNLP conditions based on the measured trait values in HNHP

conditions and the change ratios in LNHP and HNLP conditions according to a linear model: $E_{LNLP} = (1 - R_{LNHP} - R_{HNLP}) \times M_{HNHP}$ for growth rate and POC quota; $E_{LNLP} = (1+R_{LNHP}+R_{HNLP}) \times M_{HNHP}$ for PIC quota (Brennan and Collins, 2015). We tested the significant differences between the expected trait values ($E_{LNLP}$) and the measured trait values ($M_{LNLP}$) in LNLP conditions by a one-way ANOVA (Fig. S3). We also calculated the extent of synergy between LNHP and HNLP on growth rate, POC quota and PIC quota according to equation: $S = | E_{LNLP} - M_{HNHP} | / M_{HNHP}$. Please see the discussion section for more information.

## 3 Results

### 3.1 Carbonate chemistry parameters and nutrient concentrations

During the incubations, $pH_T$ values increased due to organismal activity by, on average, 0.03 ± 0.01 in LCLT, by 0.01 ± 0.01 in HCLT, by 0.02 ± 0.01 in LCHT and by 0.02 ± 0.01 in HCHT conditions (Fig. 1f–j; Table 1). Correspondingly, seawater $pCO_2$ concentrations decreased by 8.8% ± 1.1% in LCLT, by 6.1% ± 4.4% in HCLT, by 6.6% ± 1.7% in LCHT, and by 5.4% ± 3.6% in HCHT conditions, respectively (Fig. 1k–o; Table 1).

During the incubations, dissolved inorganic nitrogen (DIN) concentrations decreased by 28.7% ± 6.7% in HNHP and LL (Fig. 1p), by 26.8% ± 5.9% in HNHP and HL (Fig. 1q), by 71.1% ± 3.3% in LNHP (Fig. 1r), by 32.9% ± 5.6% in HNLP (Fig. 1s), and by 69.8% ± 3.2% in LNLP conditions (Fig. 1t; Table 2). Dissolved inorganic phosphate (DIP) concentrations decreased by 62.2% ± 16.5% in HNHP and LL (Fig. 1u), by 71.3% ± 6.7% in HNHP and HL (Fig. 1v), by 61.0% ± 5.2% in LNHP (Fig. 1w), by 83.8% ± 5.4% in HNLP (Fig. 1x), and by 86.3% ± 1.4% in LNLP conditions (Fig. 1y; Table 2).

Overall, while organismal activity affected nutrient levels during growth cycles as expected, the high and low nutrient treatments remained different at all times (Table 2). Organismal activity had minimal effects on carbonate chemistry (see Fig. 1).

### 3.2 Population growth rate

Growth rate was significantly lower under the future scenario (HCHT HL LNLP: high levels of $pCO_2$, temperature and light as well as low levels of nutrients) than under the present scenario (LCLT LL HNHP: low levels of $pCO_2$, temperature and light alongside high levels of nutrients) (one-way ANOVA, $F = 52.6$, $p < 0.01$) (Figs. 2a and 3a,d; Table 2). The effect of increasing $pCO_2$ on growth rate is negative at low light or low nutrients levels, which can be seen by comparing population growth in all of the HC regimes with their paired LC regimes (Figs. 3a,b,e and S4). The extent of reduction in population growth rate depends on which other stressors are present. Compared to present atmospheric $pCO_2$ levels (LC, Fig. 3a), growth rates under ocean acidification (HC, Fig. 3b) decreased by an average of 17.4% ± 1.3% in HNHP and LL, and by an average of 4.4% ± 1.1% in HNHP and HL conditions (three-way ANOVA, both $p < 0.01$; Tukey post hoc test, both $p < 0.01$) (Fig. 3e; Tables 2 and 3), by 7.6% ± 2.6% in LNHP, by 21.4% ± 0.2% in HNLP, and by 32.1% ± 0.5% in LNLP conditions under the HL, respectively (four-way ANOVA, all $p < 0.01$; Tukey post hoc test, all $p < 0.01$) (Fig. 3a,b,e; Tables 2 and 4).

Across all HT/LT (high/low temperature) regime pairs, population growth rate is faster in the HT regimes, indicating that increasing temperature from 16 to 20 °C increases population growth rate in *E. huxleyi* (Figs. 3a,c,f and S4). Compared to the low temperature (LT, Fig. 3a), growth rates at the high temperature (HT, Fig. 3c) increased by 7.7% ± 0.7% in HNHP and LL, and by 34.0% ± 0.4% in HNHP and HL

conditions (three-way ANOVA, both $p < 0.01$; Tukey post hoc test, both $p < 0.01$) (Fig. 3a,c,f; Tables 2 and 3), by 42.4% ± 0.4% in LNHP, by 33.5% ± 0.5% in HNLP, and by 40.4% ± 3.1% in LNLP conditions under HL (four-way ANOVA, all $p < 0.01$; Tukey post hoc test, all $p < 0.01$) (Fig. 3a,c,f; Tables 2 and 4). Compared to low $pCO_2$ and low temperature (LCLT, Fig. 3a), growth rates in high $pCO_2$ and high temperature environments (HCHT, Fig. 3d) increased by 3.9% ± 0.9% in HNHP and LL, and by 31.1% ± 0.1% in HNHP and HL conditions (three-way ANOVA, both $p < 0.01$; Tukey post hoc test, both $p < 0.01$) (Fig. 3a,d,g; Tables 2 and 3), by 38.6% ± 0.1% in LNHP and by 17.1% ± 1.7% in HNLP, whereas growth rate decreased by 12.1% ± 2.2% in LNLP conditions under HL, respectively (four-way ANOVA, all $p < 0.01$; Tukey post hoc test, all $p < 0.01$) (Fig. 3a,d,g; Tables 2 and 4). These results show that high $pCO_2$, low nitrate and low phosphate concentrations collectively reduced the population growth rate in *E. huxleyi*, though elevated temperature could counteract this response.

The effects of reduced availability of nutrients on growth are nutrient-specific (Fig. 3). Compared to HNHP and HL, growth rates in LNHP decreased by 3.0–12.1% (all $p < 0.05$ at LCLT, HCLT, LCHT and HCHT conditions) (Fig. 3h; Tables 2 and 4). In contrast, HNLP did not significantly affect growth in LC conditions ($p > 0.1$ in LCLT and LCHT conditions) (Fig. 3a,c,i), but did lower population growth rate by 11.3–19.2% in HC conditions (both $p < 0.01$ at HCLT and HCHT conditions) (Fig. 3b,d,i). Unsurprisingly, when both nitrate and phosphate levels were reduced, growth rates always decreased by larger extent compared to environments where they were reduced individually (Fig. 3h,i,j). Compared to growth rates in HNHP and HL, growth rates in LNLP were 4.8–10.2% lower in LC environments, and 34.7–40.3% lower in HC environments (Tukey post hoc test, all $p < 0.01$ at LCLT, HCLT, LCHT and

HCHT conditions) (Fig. 3a–d,j; Tables 2 and 4). In summary, nitrate and phosphate limitation exacerbated the impacts of OA and warming on population growth rate.

### 3.3 POC quota

Cellular POC quotas were two-fold larger under the future scenario (HCHT HL LNLP) than under the current scenario (LCLT LL HNHP) (one-way ANOVA, $F = 96.1$, $p < 0.01$, Figs. 2b and 4a,d). The effect of increasing $pCO_2$ on POC quota is positive, regardless of other drivers present, which can be seen by comparing POC quotas in all of the HC regimes with their paired LC regimes (Figs. 4a,b,e and S4), though the extent of increase in POC quota depends on which other stressors are present. Compared to current atmospheric $pCO_2$ level (LC, Fig. 4a), POC quotas under ocean acidification (Fig. 4b) increased by 40.3% ± 10.1% in HNHP and LL (Tukey post hoc test, $p < 0.01$), by 13.8% ± 10.1% in HNHP and HL ($p = 0.47$), by 33.2% ± 11.1% at LNHP, by 109.4% ± 14.0% in HNLP and by 87.3% ± 10.8% in LNLP conditions under HL, respectively (four-way ANOVA, all $p < 0.01$; Tukey post hoc test, all $p < 0.01$) (Fig. 4a,b,e; Tables 2 and 4).

The effect of elevated temperature on POC quota can be seen by comparing POC quota in all of the HT regimes with their paired LT regimes (Figs. 4a,c,f and S4). Across all HT/LT regime pairs, POC quotas did not show significant differences between the HT and LT regimes under HNHP and LL, HNHP and HL, LNHP, HNLP and LNLP conditions under HL, respectively (Tukey post hoc test, all $p > 0.1$) (Fig. 4a,c,f). This demonstrated that increasing temperature within the test range had no significant effect on POC quota. The combined effects of increasing $pCO_2$ and temperature on POC quotas were nutrient dependent. Compared to low $pCO_2$ and low temperature (LCLT, Fig. 4a), POC quotas at high $pCO_2$ and high temperature (HCHT,

Fig. 4d) did not show significant differences in HNHP and LL ($p$ = 0.79), in HNHP and HL ($p$ = 0.99), and in LNHP and HL ($p$ = 0.99), but increased by 52.2% ± 20.6% in HNLP and by 45.6% ± 14.8% in LNLP conditions under HL (Tukey post hoc test, both $p < 0.01$) (Fig. 4a,d,g; Tables 2 and 4). These data showed that high $pCO_2$ and low phosphate concentrations enhanced POC quotas of *E. huxleyi*, and that their combined effects were partly reduced by rising temperature.

The effects of nutrient reduction on POC quota are nutrient specific (Fig. 4). Compared to HNHP and HL, POC quotas in LNHP did not show a significant difference (all $p > 0.1$ at LCLT, HCLT, LCHT and HCHT) (Fig. 4a–d,h; Tables 2 and 4). At LC, POC quotas did not significantly differ between HNHP, HNLP and LNLP conditions (Tukey post hoc test, all $p > 0.1$) (Fig. 4a,c,i,j). In contrast, in HC, they were 43.3–78.2% larger in HNLP or LNLP than in HNHP (all $p < 0.01$) (Fig. 4b,d,i,j; Table 2).

### 3.4 PIC quota

Cellular PIC quotas were significantly larger in the future scenario with high levels of $pCO_2$, temperature and light along with low nutrients concentrations, than PIC quotas in the present scenario with low levels of $pCO_2$, temperature and light along with relatively high nutrients concentrations (one-way ANOVA, F = 63.6, $p < 0.01$) (Figs. 2c and 5a,d). However, the opposite results were found under the elevated $CO_2$ treatment alone. The effect of increasing $pCO_2$ on PIC quota is negative, regardless of presence of other drivers. By comparing PIC quota in all of the HC regimes with their paired LC regimes (Figs. 5a,b,e and S4), the effects of elevated $pCO_2$ level are clear, though the extent of reduction in PIC quota depends on which other stressors are present. Compared to present atmospheric $pCO_2$ levels (LC, Fig. 5a), PIC quotas

under ocean acidification (Fig. 5b) are reduced by 31.8% ± 17.1% in HNHP and LL, by 34.3% ± 10.0% in HNHP and HL, by 25.0% ± 3.8% in LNHP, by 22.8% ± 6.3% in HNLP and by 44.6% ± 0.9% in LNLP conditions under HL, respectively (Tukey post hoc test, all $p < 0.05$) (Fig. 5a,b,e; Tables 2–4). The extent of reduction in PIC quota is larger under LNLP conditions.

The effects of rising temperature on PIC quota were nutrient dependent, and can be seen by comparing PIC quotas in the HT regimes with those in their paired LT regimes (Figs. 5a,c,f and S4). Compared to low temperature (LT, Fig. 5a), PIC quotas at high temperature (HT, Fig. 5c) did not show significant differences in HNHP and LL, in HNHP and HL, in LNHP, and in HNLP conditions (Tukey post hoc test, all $p > 0.05$), whereas they decreased by 27.9% ± 8.4% in LNLP conditions under HL (Tukey post hoc test, $p < 0.01$) (Fig. 5a,c,f; Tables 2–4). The combined effects of rising $pCO_2$ and temperature on PIC quota are negative, regardless of which other drivers are present (Fig. 5a,d,g). Compared to low $pCO_2$ and low temperature (LCLT, Fig. 5a), PIC quotas in high $pCO_2$ and high temperature (HCHT, Fig. 5d) declined by 11.1% ± 10.9% in HNHP and LL ($p = 0.96$), by 32.5% ± 2.4% in HNHP and HL ($p < 0.01$), by 42.2% ± 3.2% in LNHP ($p < 0.01$), by 10.2% ± 7.7% in HNLP ($p = 0.92$), and by 45.3% ± 5.9% in LNLP conditions under HL, respectively ($p < 0.01$) (Fig. 5a,d,g; Table 2).

Effects of both nitrate and phosphate reduction on PIC quota are positive, regardless of levels of $pCO_2$ and temperature for the range used here (Fig. 5h,i,j). Compared to HNHP and HL, PIC quotas were larger in LNHP (Tukey post hoc test, $p < 0.01$ in LCLT, HCLT and LCHT conditions; $p = 0.73$ at HCHT condition) (Fig. 5h), in HNLP, and in LNLP conditions, respectively (all $p < 0.01$ at LCLT, HCLT, LCHT and HCHT conditions) (Fig. 5a–d,i,j; Table 2). In addition, PIC quotas were larger in

LNLP than in HNLP conditions (Tukey post hoc test, $p < 0.01$ in LCLT and HCLT conditions; $p = 0.06$ in LCHT; $p = 0.21$ in HCHT conditions) (Fig. 5a–d,i,j). These data showed that low nitrate and phosphate concentrations act synergistically to increase PIC quotas, which was moderated under the high $pCO_2$.

### 3.5 PIC / POC value

The ratio of PIC to POC (PIC/POC value) was not significantly different between the future scenario (HCHT HL LNLP) and the current scenario (LCLT LL HNHP) (one-way ANOVA, $F = 0.3$, $p = 0.60$) (Figs. 2d and 6a,d). The PIC / POC value followed the same trend as for PIC quotas described above. The effect of increasing $pCO_2$ on PIC / POC value was negative, regardless of which other drivers were present (Figs. 6a,b,e and S4), but the extent of reduction in PIC / POC value depended on presence of other drivers. Compared to current atmospheric $pCO_2$ levels (LC, Fig. 6a), PIC / POC values under ocean acidification (HC, Fig. 6b) decreased by 50.7% ± 18.2% in HNHP and LL, by 41.8% ± 15.4% in HNHP and HL, by 43.9% ± 5.8% in LNHP, by 63.0% ± 4.2% in HNLP, and by 70.7% ± 2.0% in LNLP conditions under HL, respectively (Tukey post hoc test, all $p < 0.05$) (Fig. 6a,b; Table 2).

The effect of rising temperature on PIC / POC value was nutrient dependant (Figs. 6a,c,f and S4). Compared to low temperature (LT, Fig. 6a), PIC / POC values at high temperature (HT, Fig. 6c) did not show significant differences in HNHP and LL, in HNHP and HL, in LNHP, and in LNLP conditions (Tukey post hoc test, all $p > 0.1$), whereas they increased by 39.0% ± 8.9% in HNLP conditions (Tukey post hoc test, $p = 0.006$) (Fig. 6a,c,f; Table 2). The combined effects of elevated $pCO_2$ and temperature on PIC / POC values were negative (Fig. 6a,d,g). Relative to low $pCO_2$ and low temperature (LCLT, Fig. 6a), PIC / POC values at high $pCO_2$ and high

499 temperature (HCHT, Fig. 6d) did not show significant differences in HNHP and LL,
500 and in HNHP and HL conditions (Tukey post hoc test, both $p > 0.1$), but they
501 decreased by 39.9% ± 3.0% in LNHP, by 40.6% ± 5.8% in HNLP, and by 67.8% ±
502 3.1% in LNLP conditions under HL, respectively (Tukey post hoc test, all $p < 0.01$)
503 (Fig. 6a,d,g; Table 2).

504 Across all LNHP/HNHP (low/high nitrate) regime pairs, PIC / POC values were
505 higher in the LNHP regime (Fig. 6h), though the extent of increase in PIC / POC
506 values depended on $pCO_2$ or temperature levels. Compared to HNHP and HL, PIC /
507 POC values in LNHP were about 106.0% ± 13.0% larger (Tukey post hoc test, $p <$
508 0.05 in LCLT and LCHT conditions; $p > 0.05$ in HCLT and HCHT conditions) (Fig.
509 6a–d, h; Table 2). The effect of phosphate on PIC / POC value also depended on
510 $pCO_2$ levels (Fig. 6i). In LC, PIC / POC values were larger in HNLP than in HNHP ($p$
511 = 0.22 at LCLT; $p < 0.05$ at LCHT conditions), and in LNLP than in LP ($p < 0.01$ at
512 LCLT; $p = 0.09$ in LCHT conditions) (Fig. 6a,c). In HC conditions, PIC / POC values
513 did not show significant differences among HNHP, HNLP and LNLP conditions
514 (Tukey post hoc test, all $p > 0.05$ in HCLT and HCHT conditions) (Fig. 6b,d; Table 2).
515

516 **4 Discussion**

517 Understanding effects of multiple drivers is helpful for improving how
518 coccolithorphores are represented in models (Krumhardt et al., 2017). Responses of
519 growth, POC and PIC quotas to ocean acidification have been shown to be modulated
520 by temperature (Gafar and Schulz, 2018; Tong et al., 2019), light intensity or light
521 period (light : dark cycle) (Jin et al., 2017; Bretherton et al., 2019), DIN or DIP
522 concentrations (Müller et al., 2017), combinations of light intensity and nutrients
523 availability (Zhang et al., 2019) (Table 5). Following up our previous study (Zhang et

al., 2019), we added temperature as a key driver of 5 drivers (Table S2), and explored how temperature changes would modulate the combined effects of $CO_2$, light, DIN and DIP that we previously reported. Our data showed that a future ocean climate change cluster (increasing $CO_2$, temperature, and light levels along with decreasing DIN and DIP levels) can lower growth rate with increased POC and PIC quota per cell (Fig. 2) as a result of plastic responses to the drivers. In contrast, observations of coccolithophore Chl *a* increased from 1990 to 2014 in the North Atlantic, and rising $CO_2$ and temperature has been aassociated with accelerated growth of coccolithophores since 1965 in the North Atlantic (Rivero-Calle et al., 2015; Krumhardt et al., 2016). Our results from laboratory experiments with multiple drivers experiment instead predicted a different trend with progressive ocean climate changes~~,. suggesting that some key elements of understanding phytoplankton responses to changing conditions that would enable researchers to connect laboratory studies and field observations are missing.~~ We have to admit that results from laboratory experiments can hardly extrapolate to natural conditions. Nevertheless, our data provide mechanistic understanding of the combined effects of ocean climate change drivers, which can be useful in analyzing field observations.

It should also be noted that regional responses to ocean global changes could differ due to chemical and physical environmental differences and species and strain variability among different oceans or regions (Blanco-Ameijeiras et al., 2016; Gao et al., 2019), and that this could also explain discrepancies between experiments and observations. Different *E. huxleyi* strains displayed optimal responses to a broad range of temperature or $CO_2$ level, and *E. huxleyi* strains isolated from different regions showed local adaptation to temperature or $CO_2$ level (Zhang et al., 2014; 2018). Strain-specific responses of growth, POC and PIC production rates in *E. huxleyi*

isolated from different regions to changing seawater carbonate chemistry have also been documented (Langer et al., 2009). It has been suggested that inter-strain genetic variability has greater potential to induce larger phenotypic differences than the phenotypic plasticity of a single strain cultured under a broad range of variable environmental conditions (Blanco-Ameijeiras et al., 2016). On the other hand, the genetic adaptation to culture experimental conditions over time may no longer accurately represent the cells in the sea, as reflected in a diatom (Guan and Gao, 2008). Phytoplankton species that had been maintained under laboratory conditions might have lost original traits and display different responses to environmental changes (Lakeman et al., 2009). The strain used in this study has been kept in the laboratory for about 30 years, and the data obtained in this work can hardly reflect relation to its biogeographic origin.

The decreased availability of nitrate or phosphate individually reduced growth rate and increased PIC quota, respectively, in this experiment. Furthermore, under LNLP and high $pCO_2$ levels, measured growth rates were significantly lower than the expected values estimated on the basis of the values in LNHP and HNLP conditions (Fig. S3a). This indicates synergistic negative effects of LN and LP on growth rate, an evidence that colimitation of N and P is more severe than that by N or P alone. Here, the extent of synergy between LN and LP on growth rate was calculated to be 8.6%±2.8% at low temperature and to be 40.6%±3.8% at high temperature (Fig. S3a), suggesting modulating effect of temperature on response of growth rate to nutrient limitations (Thomas et al., 2017). Similarly, at LNLP and low $pCO_2$ level, the measured PIC quota was significantly larger than the expected value (Fig. S3c), indicating synergistic positive effects of LN and LP on PIC quota, with the extent of

synergy being 31.4%±3.9% at low temperature. LN and LP did not synergistically act to reduce POC quota.

While there were always interactions among stressors, increased temperature itself sped up population growth to a relatively consistent value at high light, regardless of nutrient limitation, with statistically significant but small differences over the different nutrient regimes (Fig. 3f). Rising $pCO_2$ level not only decreased the absolute values of growth rate, but also reduced the positive effect of high temperature on growth. In addition, elevated $pCO_2$ also altered patterns of growth responses to changes in light and nutrient levels (Fig. 3e–g). In ocean acidification condition, the negative effect of low pH on growth rate of the same *E. huxeyi* strain PML B92/11 was larger than the positive effect of high $CO_2$ concentration (Bach et al., 2011). Our data further showed that ~~Interestingly,~~ low-pH inhibited growth to lesser extent under the high light than under low light (Fig. 3e; Table 2). One possible explanation for this could be that photosynthesis under the high light regime could generate more energy-conserving compounds (Fernández et al., 1996)~~,~~. ~~which~~ This results in faster $pCO_2$ removal and counteracts the negative effects of low pH. This interaction between low pH and high light was also observed when *E. huxleyi* ~~was~~ strains PML B92/11 and CCMP 2090 were grown under incident sunlight (Jin et al., 2017).

Increases in temperature reduced PIC quotas under some conditions (high light (HL), HL-LNHP and HL-LNLP) (Fig. 5f), suggesting that the ratio of N:P is important in modulating calcification under warming. One striking result is the consistent negative effect of high $pCO_2$ on growth and PIC quota, regardless of other stressors. While $pCO_2$ levels affected the absolute PIC values, the combination of high $pCO_2$ and warming did not affect the responses to light and nutrients once the direct reduction in PIC quota due to increased $pCO_2$ was taken into account (Fig. 5g).

It has been documented that PIC quotas of *E. huxleyi* strain PML B92/11 reduced at high $pCO_2$ due to suppressed calcification (Riebesell and Tortell, 2011). This knowledge has been based on experiments under nutrient-~~replicate~~replete or constant conditions without consideration of multiple drivers. In this work, PIC quota of *E. huxleyi* under OA were raised with increased light intensity and decreased availability of nutrients (Figs. 2 and 5). These results are consistent with other studies (Perrin et al., 2016; Jin et al., 2017), which reported that nutrient limitations enhanced calcification, and ~~increased light levels can partially counteract the negative effects of OA on calcification~~ high light intensity could make cells to remove $H^+$ faster and then reduce the negative effect of low pH on calcification of *E. huxleyi* (Jin et al., 2017). Our data also indicate that effects of ocean climate change on calcification of *E. huxleyi* are more complex than previously thought (Meyer and Riebesell, 2015). It is worth noting that the observed higher POC and PIC quotas under future ocean climate change scenario could be attributed to cell cycle arrest of a portion of the community (Vaulot et al., 1987). Decreased availabilities of nitrate and phosphate can extend the G1 phase where photosynthetic carbon fixation and calcification occurred, and lead to lower dark respiration which reduces carbon consumption (Vaulot et al., 1987; Müller et al., 2008; Gao et al., 2018).

Synthesis of RNA is a large biochemical sink for phosphate in *E. huxleyi* and other primary producers (Dyhrman, 2016). In this study, RNA content per cell was verified by a SYBR Green method (Berdalet et al., 2005). Compared to HNHP conditions, HNLP-grown cells had only 7.8% of total RNA (Fig. S11). This indicates that decreased availability of phosphate strongly decreased RNA synthesis, which would consequently extend the interphase of the cell cycle where calcification occurs (Müller et al., 2008). This could explain why PIC quotas were enhanced by decreased

phosphate availability (Fig. 5). Similarly, decreased availability of nitrate decreased protein (or PON) synthesis (Fig. S10), which can also block cells in the interphase of the cell cycle, and increase the time available for calcification in *E. huxleyi* (Vaulot et al., 1987). Consistently with this, lower rates of assimilation or organic matter production in *E. huxleyi* in LNHP than in HNHP treatments are consistent with more energy being reallocated to use for calcification (Nimer and Merrett, 1993; Xu and Gao, 2012).

Low phosphate concentrations can induce high affinity phosphate uptake in *E. huxleyi* (Riegman et al., 2000; Dyhrman and Palenik, 2003; McKew et al., 2015). This mechanism enables *E. huxleyi* to take up phosphate efficiently at low $pCO_2$ concentrations, so that no significant difference in growth rate was observed between HNLP and HNHP treatments (Fig. 3a,c). However, at high $pCO_2$, low phosphate concentration (HNLP) lowered growth of *E. huxleyi* relative to HNHP (Fig. 3a–d; Table 2). While the affinity of *E. huxleyi* for phosphate under different $pCO_2$ levels has not been studied, the extra energetic cost of coping with stress from high $pCO_2$ could limit the energy available for the active uptake of phosphate. In addition, the activity of alkaline phosphatase, which might work to reuse released organic P, decreases at low pH (Rouco et al., 2013). Finally, the enlarged cell volume in HC and HNLP (or LNLP) conditions may further reduce nutrient uptake by cells due to reduced surface to volume ratios, and lower cell division rates (Fig. S5) (Finkel, 2001). While substantial evolutionary responses to multiple drivers may help further, our results imply that decreased phosphate availability along with progressive ocean acidification and warming in surface ocean may reduce the competitive capability of *E. huxleyi* in oligotrophic waters. Meanwhile, ~~On the other hand,~~ HNLP also affected expressions of genes related to nitrogen metabolism due to the tight stoichiometric

coupling of nitrogen and phosphate metabolism (Rokitta et al., 2016). Decreased availability of nitrate further limited nitrogen metabolism of *E. huxleyi* (Rokitta et al., 2014), which lowered the overall biosynthetic activity and reduced cellular PON quotas (Fig. S10). These explain the synergistic inhibitions of low-pH, low-phosphate and low-nitrate on growth of *E. huxleyi* (Fig. 3).

POC quotas and the cell-volume normalized POC quotas were larger at high $pCO_2$ than at low $pCO_2$ under all treatments (Figs. 4; S6; Table 2), which could be a combined outcome of increased photosynthetic carbon fixation (Zondervan et al., 2002; Hoppe et al., 2011; Tong et al., 2019) and reduced cell division (present work), leading to pronounced increase of POC quotas in the cells grown under low phosphate (HNLP) and high $pCO_2$ (Fig. 4). At HNLP and high $pCO_2$ levels, photosynthetic carbon fixation proceeds whereas cell division rate decreases (Figs. 3 and 4), so reallocation of newly produced particulate organic carbon (POC) could be slowed down (Vaulot et al., 1987). In this case, over-synthesis of cellular organic carbon might be released as dissolved organic carbon (DOC), which can coagulate to transparent exopolymer particles (TEP) and attach to cells (Biermann and Engel, 2010; Engel et al., 2015). When cells were filtered on GF/F filters, any TEP would not have be separated from the cells and would have contributed to the measured POC quota in this study. ~~However, released organic compounds should be negligible, since they are usually photorespiration-dependent (Beardall, 1989; Obata et al., 2013).~~

~~Synthesis of RNA is large biochemical sinks for phosphate in *E. huxleyi* and other primary producers (Dyhrman, 2016). Compared to HNHP conditions, HNLP-grown cells had only 7.8% of total RNA (Fig. S11). This indicates that decreased availability of phosphate strongly decreased RNA synthesis, which would consequently extend the interphase of the cell cycle where calcification occurs (Müller et al., 2008). This~~

~~could explain why PIC quotas were enhanced by decreased phosphate availability (Fig. 5). Similarly, decreased availability of nitrate decreased protein (or PON) synthesis (Fig. S10), which can also block cells in the interphase of the cell cycle, and increase the time available for calcification in *E. huxleyi* (Vaulot et al., 1987). Consistently with this, lower rates of assimilation or organic matter production in *E. huxleyi* in LNHP than in HNHP treatments are consistent with more energy being reallocated to use for calcification (Nimer and Merrett, 1993; Xu and Gao, 2012).~~

Large PIC quotas of coccolithophores may facilitate accumulation of calcium carbonate in the deep ocean and increase the contribution of $CaCO_3$ produced by coccolithophores to calcareous ooze in the pelagic ocean (Hay, 2004). Due to $CaCO_3$ being more dense than organic carbon, larger PIC quotas may facilitate effective transport of POC to deep oceans, leading to vertical DIC or $CO_2$ gradients of seawater (Milliman, 1993; Ziveri et al., 2007). While the effects of global ocean climate changes on physiological processes of phytoplankton can be complex, our results promote our understanding on how a cosmopolitan coccolithophore responds to future ocean environmental changes through plastic trait change. ~~While substantial evolutionary responses to multiple drivers may help further, our results imply that decreased phosphate availability along with progressive ocean acidification and warming in surface ocean may reduce the competitive capability of *E. huxleyi* in oligotrophic waters.~~

*Data availability.* The data are available upon request to the corresponding author (Kunshan Gao).

*Author contributions*. YZ, KG designed the experiment. YZ performed this experiment. All authors analysed the data, wrote and improved the manuscript.

*Competing interests*. The authors declare that they have no conflict of interest.

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## Figure Legends

**Figure 1.** Four "baseline" environments were used where $pCO_2$ and temperature (temp) were combined in all pairwise combinations: low $pCO_2$ + low temp (LCLT, △), high $pCO_2$ + low temp (HCLT, ✳), low $pCO_2$ + high temp (LCHT, □) and high $pCO_2$ + high temp (HCHT, ○). Additional stressors were then added to each of the four "baseline" environments. In step 1, low light (LL) was ~~added~~ supplied. In step 2, high light (HL) was ~~added~~ supplied. HL was then maintained for the rest of the experiment. In step 3, low nitrogen was ~~added~~ supplied and high phosphate levels were restored (LNHP). In step 4, low phosphate was ~~added~~ supplied and high nitrogen levels were restored (HNLP). In step 5, both nitrogen and phosphate were low (LNLP). Experimental steps were done in a consecutive manner. At each step, we measured cell concentration (**a**–**e**), medium $pH_T$ value (**f**–**j**), medium $pCO_2$ level (**k**–**o**), dissolved inorganic nitrogen (DIN) (**p**–**t**) and phosphate (DIP) (**u**–**y**) concentrations in the media in the beginning and at the end of the incubations. Respectively, LC and HC represent $pCO_2$ levels of about 370 and 960 µatm; LT and HT 16 and 20 °C; LL and HL 60 and 240 µmol photons $m^{-2}$ $s^{-1}$ of photosynthetically active radiation (PAR); HN and LN 24.3 and 7.8 µmol $L^{-1}$ $NO_3^-$ at the beginning of the incubation; HP and LP 1.5 and 0.5 µmol $L^{-1}$ $PO_4^{3-}$ at the beginning of the incubations. The samples were taken in the last day of the cultures in each treatment. The values were indicated as the means ± sd of 4 replicate populations for each treatment.

**Figure 2.** Growth rate (**a**), particulate organic (POC, **b**) and inorganic (PIC, **c**) carbon quotas, PIC / POC value (**d**) and cell volume (**e**) of *Emiliania huxleyi* grown under the present (defined as low levels of $pCO_2$, temperature and light along with high levels of nutrients) and the future (defined as higher levels of $pCO_2$, temperature, and light

1043 along with low levels of nutrients due to ocean acidification, warming and shoaling of
1044 upper mixing layer) scenarios. Data were obtained after cells were acclimated to
1045 experimental conditions for 14–16 generations and means ± sd of 4 replicate
1046 populations. Different letters (a, b) in each panel represent significant differences
1047 between future and present ocean conditions (Tukey Post hoc, $p < 0.05$).
1048
1049 **Figure 3.** Growth rates of *E. huxleyi* grown in LCLT (**a**), HCLT (**b**), LCHT (**c**) and
1050 HCHT (**d**) conditions, and the ratio of growth rate at HC to LC (**e**), HT to LT (**f**),
1051 HCHT to LCLT (**g**), LNHP to HNHP (**h**), HNLP to HNHP (**i**) and LNLP to HNHP (**j**).
1052 Data were obtained after cells were acclimated to experimental conditions for 14–16
1053 generations and means ± sd of 4 replicate populations. Horizontal lines in panels (**e**)–
1054 (**j**) showed the value of 1. Different letters (a, b, c, d) in panels (**a**)–(**d**) represent
1055 significant differences between different nutrient treatments (Tukey Post hoc, $p <$
1056 0.05). The results shown in the black column were used for the ambient-future
1057 comparison in figure 2. Detailed experimental conditions were shown in Figure 1.
1058
1059 **Figure 4.** POC quota of *E. huxleyi* grown in LCLT (**a**), HCLT (**b**), LCHT (**c**) and
1060 HCHT (**d**) conditions, and the ratio of POC quota at HC to LC (**e**), HT to LT (**f**),
1061 HCHT to LCLT (**g**), LNHP to HNHP (**h**), HNLP to HNHP (**i**) and LNLP to HNHP (**j**).
1062 Data were obtained after cells were acclimated to experimental conditions for 14–16
1063 generations and means ± sd of 4 replicate populations. Horizontal lines in panels (**e**)–
1064 (**j**) showed the value of 1. Different letters (a, b) in panels (**a**)–(**d**) represent significant
1065 differences between different nutrient treatments (Tukey Post hoc, $p < 0.05$). The
1066 results shown in the black column were used for the ambient-future comparison in
1067 figure 2. Detailed experimental conditions were shown in Figure 1.

**Figure 5.** PIC quota of *E. huxleyi* grown in LCLT (**a**), HCLT (**b**), LCHT (**c**) and HCHT (**d**) conditions, and the ratio of PIC quota at HC to LC (**e**), HT to LT (**f**), HCHT to LCLT (**g**), LNHP to HNHP (**h**), HNLP to HNHP (**i**) and LNLP to HNHP (**j**). Data were obtained after cells were acclimated to experimental conditions for 14–16 generations and means ± sd of 4 replicate populations. Horizontal lines in panels (**e**)–(**j**) showed the value of 1. Different letters (a, b, c) in panels (**a**)–(**d**) represent significant differences between different nutrient treatments (Tukey Post hoc, $p < 0.05$). The results shown in the black column were used for the ambient-future comparison in figure 2. Detailed experimental conditions were shown in Figure 1.

**Figure 6.** PIC / POC value of *E. huxleyi* grown in LCLT (**a**), HCLT (**b**), LCHT (**c**) and HCHT (**d**) conditions, and the ratio of (PIC / POC value) at HC to LC (**e**), HT to LT (**f**), HCHT to LCLT (**g**), LNHP to HNHP (**h**), HNLP to HNHP (**i**) and LNLP to HNHP (**j**). Data were obtained after cells were acclimated to experimental conditions for 14–16 generations and means ± sd of 4 replicate populations. Horizontal lines in panels (**e**)–(**j**) showed the value of 1. Different letters (a, b, c) in panels (**a**)–(**d**) represent significant differences between different nutrient treatments (Tukey Post hoc, $p < 0.05$). The results shown in the black column were used for the ambient-future comparison in figure 2. Detailed experimental conditions were shown in Figure 1.

**Figure S1.** Flow chart of the experimental processes. Experimental steps were done in a consecutive manner. Detailed experimental conditions were shown in Figure 1.

**Figure S2.** Representative curves for the time course for cell concentrations of *E. huxleyi* under low $pCO_2$ (LC), high (HT) or low (LT) temperatures, and high light (HL) conditions with varying levels of nutrients: HNHP (**a**), LNHP (**b**), HNLP (**c**) and LNLP (**d**), respectively. Arrow indicates the day when samples were taken in each treatment. Data were means ± sd of 4 replicate populations. Detailed experimental conditions were shown in Figure 1.

**Figure S3.** Comparison of growth rate (**a**), POC quota (**b**) and PIC quota (**c**) between the expected (calculated) values and the measured values under the LNLP treatments. Different letters (a, b) in each "baseline" environment (LCLT, HCLT, LCHT or HCHT) represent significant differences (Tukey Post hoc, $p < 0.05$). Detailed experimental conditions were shown in Figure 1.

**Figure S4.** Heatmap of the changes in growth rate, POC quota, PIC quota and PIC:POC in each treatment. Values in the present scenario (LC LT LL HNHP) were considered as the control. A minus sign indicates the reduction in these parameters.

**Figure S5.** Cell volume of *E. huxleyi* grown in LCLT (**a**), HCLT (**b**), LCHT (**c**) and HCHT (**d**) conditions, and its correlation with POC quota (**e**) and PIC quota (**f**). Data were obtained after cells were acclimated to experimental conditions for 14–16 generations and means ± sd of 4 replicate populations in panels (**a**)–(**d**). Each point in panels (**e**) and (**f**) indicates an individual replicate from all experiment. Different letters (a, b, c) in panels (**a**)–(**d**) represent significant differences between different nutrient treatments (Tukey Post hoc, $p < 0.05$).

**Figure S6.** Normalized POC quota of *E. huxleyi* to cell volume in LCLT (**a**), HCLT (**b**), LCHT (**c**) and HCHT (**d**) conditions. Data were obtained after cells were acclimated to experimental conditions for 14–16 generations and means ± sd of 4 replicate populations. Different letters (a, b) in each panel represent significant differences between different nutrient treatments (Tukey Post hoc, $p < 0.05$).

**Figure S7.** Normalized PIC quota of *E. huxleyi* to cell volume in LCLT (**a**), HCLT (**b**), LCHT (**c**) and HCHT (**d**) conditions. Data were obtained after cells were acclimated to experimental conditions for 14–16 generations and means ± sd of 4 replicate populations. Different letters (a, b, c) in each panel represent significant differences between different nutrient treatments (Tukey Post hoc, $p < 0.05$).

**Figure S8.** POC production rate of *E. huxleyi* in LCLT (**a**), HCLT (**b**), LCHT (**c**) and HCHT (**d**) conditions, and the ratio of POC production rate at HC to LC (**e**), HT to LT (**f**), HCHT to LCLT (**g**), LNHP to HNHP (**h**), HNLP to HNHP (**i**) and LNLP to HNHP (**j**). Data were obtained after cells were acclimated to experimental conditions for 14–16 generations and means ± sd of 4 replicate populations. Horizontal lines in panels (**e**)–(**j**) showed the value of 1. Different letters (a, b, c) in panels (**a**)–(**d**) represent significant differences between different nutrient treatments (Tukey Post hoc, $p < 0.05$).

**Figure S9.** PIC production rate of *E. huxleyi* in LCLT (**a**), HCLT (**b**), LCHT (**c**) and HCHT (**d**) conditions, and the ratio of PIC production rate at HC to LC (**e**), HT to LT (**f**), HCHT to LCLT (**g**), LNHP to HNHP (**h**), HNLP to HNHP (**i**) and LNLP to HNHP (**j**). Data were obtained after cells were acclimated to experimental conditions

for 14–16 generations and means ± sd of 4 replicate populations. Horizontal lines in panels (**e**)–(**j**) showed the value of 1. Different letters (a, b, c) in panels (**a**)–(**d**) represent significant differences between different nutrient treatments (Tukey Post hoc, $p < 0.05$).

**Figure S10.** PON quota of *E. huxleyi* in LCLT (**a**), HCLT (**b**), LCHT (**c**) and HCHT (**d**) conditions, and the ratio of PON quota at HC to LC (**e**), HT to LT (**f**), HCHT to LCLT (**g**), LNHP to HNHP (**h**), HNLP to HNHP (**i**) and LNLP to HNHP (**j**). Data were obtained after cells were acclimated to experimental conditions for 14–16 generations and means ± sd of 4 replicate populations. Horizontal lines in panels (**e**)–(**j**) showed the value of 1. Different letters (a, b) in panels (**a**)–(**d**) represent significant differences between different nutrient treatments (Tukey Post hoc, $p < 0.05$).

**Figure S11.** Normalized RNA quota of *E. huxleyi* to POC quota in HNHP and HNLP conditions. Data were obtained after cells were acclimated to experimental conditions for 14–16 generations and means ± sd of 4 replicate populations. Different letters (a, b) represent significant differences between different nutrient treatments (Tukey Post hoc, $p < 0.05$).

**Table 1.** Carbonate chemistry parameters at the end of the incubation. The values are means ± sd of 4 replicate populations. LL and HL represent 60 and 240 µmol photons $m^{-2}$ $s^{-1}$ of photosynthetically active radiation (PAR), respectively; HN and LN represent 24.3 and 7.8 µmol $L^{-1}$ DIN in the beginning of the incubation; HP and LP represent 1.5 and 0.5 µmol $L^{-1}$ DIP in the beginning of the incubation, respectively.

| | | | $pCO_2$ (µatm) | pH (total scale) | TA (µmol $L^{-1}$) | DIC (µmol $L^{-1}$) | $HCO_3^-$ (µmol $L^{-1}$) | $CO_3^{2-}$ (µmol $L^{-1}$) | $CO_2$ (µmol $L^{-1}$) |
|---|---|---|---|---|---|---|---|---|---|
| 16 | LL-HNHP | LC | 371±17 | 8.07±0.02 | 2266±19 | 2017±9 | 1823±6 | 180±8 | 13.4±0.6 |
| | | HC | 918±21 | 7.73±0.02 | 2248±45 | 2149±39 | 2027±35 | 90±5 | 33.3±0.7 |
| | HL-HNHP | LC | 387±22 | 8.06±0.02 | 2297±12 | 2050±17 | 1857±20 | 179±6 | 14.0±0.8 |
| | | HC | 972±11 | 7.71±0.01 | 2283±34 | 2189±31 | 2066±29 | 88±3 | 35.2±0.4 |
| | HL-LNHP | LC | 393±20 | 8.05±0.02 | 2273±9 | 2033±3 | 1845±9 | 174±7 | 14.3±0.7 |
| | | HC | 1012±13 | 7.69±0.01 | 2263±28 | 2177±25 | 2057±24 | 84±2 | 36.7±0.5 |
| | HL-HNLP | LC | 395±19 | 8.06±0.02 | 2318±5 | 2073±12 | 1879±16 | 179±6 | 14.3±0.7 |
| | | HC | 958±63 | 7.70±0.01 | 2205±69 | 2117±71 | 1999±69 | 84±1 | 34.7±2.3 |
| | HL-LNLP | LC | 375±24 | 8.06±0.01 | 2181±78 | 1947±77 | 1767±73 | 167±3 | 13.6±0.9 |
| | | HC | 1014±46 | 7.68±0.01 | 2198±73 | 2118±73 | 2002±69 | 79±2 | 36.7±1.7 |
| 20 | LL-HNHP | LC | 349±16 | 8.09±0.02 | 2257±14 | 1963±4 | 1741±6 | 210±8 | 11.3±0.5 |
| | | HC | 899±40 | 7.74±0.02 | 2257±53 | 2130±45 | 1994±40 | 107±7 | 29.0±1.3 |
| | HL-HNHP | LC | 363±11 | 8.08±0.01 | 2281±16 | 1990±18 | 1770±19 | 208±2 | 11.7±0.3 |
| | | HC | 947±24 | 7.72±0.01 | 2248±21 | 2130±19 | 1998±18 | 102±3 | 30.6±0.8 |
| | HL-LNHP | LC | 362±18 | 8.08±0.02 | 2262±12 | 1973±13 | 1756±16 | 206±7 | 11.7±0.6 |
| | | HC | 970±10 | 7.71±0.01 | 2271±31 | 2155±28 | 2021±25 | 102±3 | 31.4±0.3 |
| | HL-HNLP | LC | 370±14 | 8.08±0.01 | 2314±3 | 2023±10 | 1800±14 | 211±4 | 12.0±0.4 |
| | | HC | 946±47 | 7.71±0.01 | 2200±72 | 2088±72 | 1960±68 | 98±2 | 30.6±1.5 |
| | HL-LNLP | LC | 350±18 | 8.08±0.01 | 2193±71 | 1912±68 | 1701±63 | 200±5 | 11.3±0.6 |
| | | HC | 977±59 | 7.70±0.01 | 2192±78 | 2086±79 | 1959±76 | 95±2 | 31.6±1.9 |

**Table 2.** Final nitrate and phosphate concentrations (N : P, μmol $L^{-1}$), growth rate ($d^{-1}$), POC and PIC quotas (pg C $cell^{-1}$), and PIC / POC value. Values in the brackets represent final DIN and DIP concentrations, and standard deviation of 4 replicate populations for growth rate, POC and PIC quotas, and PIC / POC value. Detailed information was shown in Table 1.

| $pCO_2$ | T | Light | Final N : P | Growth rate | POC quota | PIC quota | PIC/POC |
|---|---|---|---|---|---|---|---|
| LC | LT | LL | HNHP (17.1 : 0.7) | 0.96 (0.012) | 1.80 (0.14) | 0.38 (0.09) | 0.21 (0.07) |
| | | HL | HNHP (17.3 : 0.5) | 1.09 (0.006) | 2.50 (0.28) | 0.62 (0.05) | 0.25 (0.05) |
| | | HL | LNHP (2.5 : 0.6) | 1.00 (0.013) | 2.07 (0.25) | 0.90 (0.02) | 0.44 (0.05) |
| | | HL | HNLP (15.4 : 0.1) | 1.08 (0.006) | 2.42 (0.08) | 0.83 (0.04) | 0.34 (0.01) |
| | | HL | LNLP (2.4 : 0.1) | 0.99 (0.003) | 2.62 (0.25) | 1.62 (0.14) | 0.63 (0.11) |
| HC | LT | LL | HNHP (18.6 : 0.9) | 0.79 (0.012) | 2.52 (0.33) | 0.26 (0.06) | 0.10 (0.04) |
| | | HL | HNHP (18.2 : 0.5) | 1.04 (0.012) | 2.85 (0.36) | 0.41 (0.06) | 0.15 (0.04) |
| | | HL | LNHP (2.0 : 0.6) | 0.92 (0.026) | 2.75 (0.23) | 0.68 (0.03) | 0.25 (0.03) |
| | | HL | HNLP (15.5 : 0.1) | 0.85 (0.002) | 5.06 (0.34) | 0.64 (0.05) | 0.13 (0.01) |
| | | HL | LNLP (2.7 : 0.1) | 0.67 (0.005) | 4.91 (0.28) | 0.90 (0.01) | 0.18 (0.01) |
| LC | HT | LL | HNHP (16.6 : 0.3) | 1.03 (0.006) | 1.58 (0.11) | 0.43 (0.02) | 0.27 (0.01) |
| | | HL | HNHP (17.3 : 0.3) | 1.46 (0.004) | 2.15 (0.28) | 0.52 (0.07) | 0.25 (0.06) |
| | | HL | LNHP (2.1 : 0.5) | 1.42 (0.004) | 1.68 (0.05) | 0.79 (0.04) | 0.47 (0.03) |
| | | HL | HNLP (17.0 : 0.1) | 1.44 (0.004) | 2.09 (0.03) | 1.00 (0.05) | 0.48 (0.03) |
| | | HL | LNLP (2.1 : 0.1) | 1.39 (0.038) | 2.02 (0.05) | 1.17 (0.13) | 0.58 (0.07) |
| HC | HT | LL | HNHP (16.7 : 0.4) | 0.99 (0.008) | 1.54 (0.12) | 0.34 (0.05) | 0.22 (0.04) |
| | | HL | HNHP (17.9 : 0.5) | 1.43 (0.001) | 2.57 (0.06) | 0.42 (0.02) | 0.16 (0.01) |
| | | HL | LNHP (2.4 : 0.6) | 1.38 (0.009) | 1.97 (0.03) | 0.52 (0.03) | 0.27 (0.01) |
| | | HL | HNLP (17.1 : 0.1) | 1.27 (0.018) | 3.68 (0.50) | 0.74 (0.06) | 0.20 (0.02) |
| | | HL | LNLP (2.2 : 0.1) | 0.87 (0.022) | 3.81 (0.39) | 0.89 (0.10) | 0.20 (0.04) |

**Table 3.** Results of three-way ANOVAs of the effects of temperature (T), $pCO_2$ (C) and light intensity (L) and their interaction on growth rate, POC and PIC quotas, and PIC / POC value. Significant values were marked in bold.

| | | T | C | L | T×C | T×L | C×L | T×C×L |
|---|---|---|---|---|---|---|---|---|
| Growth rate | F | 20037.5 | 477.4 | 23625.8 | 120.0 | 1550.9 | 34.0 | 86.4 |
| | *p* | **<0.01** | **<0.01** | **<0.01** | **<0.01** | **<0.01** | **<0.01** | **<0.01** |
| POC quota | F | 27.1 | 54.4 | 62.0 | 7.4 | 1.9 | < 0.1 | 6.1 |
| | *p* | **<0.01** | **<0.01** | **<0.01** | **0.01** | 0.18 | 0.83 | **0.02** |
| PIC quota | F | 0.4 | 38.6 | 47.6 | 2.3 | 6.6 | 1.6 | 1.1 |
| | *p* | 0.56 | **<0.01** | **<0.01** | 0.14 | **0.02** | 0.22 | 0.31 |
| PIC / POC value | F | 9.9 | 443.6 | 2.0 | 0.8 | 10.0 | 0.6 | 0.3 |
| | *p* | **<0.01** | **<0.01** | 0.17 | 0.38 | **<0.01** | 0.46 | 0.60 |

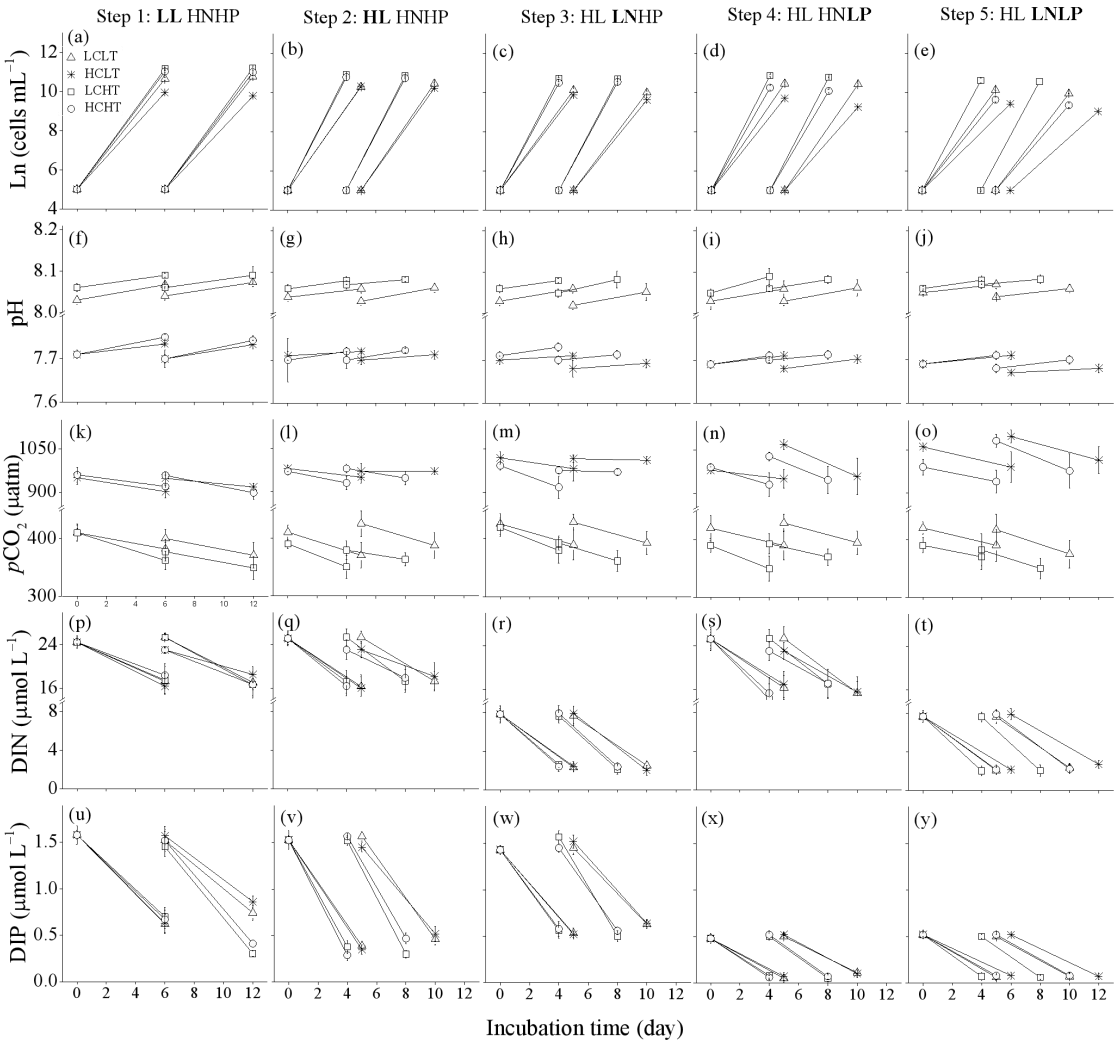
**Table 4.** Results of four-way ANOVAs of the effects of temperature (T), $pCO_2$ (C), dissolved inorganic nitrate (N) and phosphate (P) concentrations and their interaction on growth rate, POC and PIC quotas, and PIC / POC value. Significant values were marked in bold.

| | Growth rate | | POC quota | | PIC quota | | PIC / POC value | |
|---|---|---|---|---|---|---|---|---|
| | F | *p* | F | *p* | F | *p* | F | *p* |
| T | 500026.0 | **<0.01** | 297.4 | **<0.01** | 30.2 | **<0.01** | 82.8 | **<0.01** |
| C | 5798.0 | **<0.01** | 162.8 | **<0.01** | 376.2 | **<0.01** | 787.3 | **<0.01** |
| N | 4542.0 | **<0.01** | 157.0 | **<0.01** | 84.4 | **<0.01** | 127.6 | **<0.01** |
| P | 5347.0 | **<0.01** | 206.5 | **<0.01** | 474.6 | **<0.01** | 0.1 | 0.74 |
| T×C | 6899.0 | **<0.01** | 52.2 | **<0.01** | 0.2 | 0.68 | 7.2 | **<0.01** |
| T×N | 510.0 | **<0.01** | 5.6 | **0.02** | 60.0 | **<0.01** | 7.9 | **<0.01** |
| T×P | 39.0 | **<0.01** | 5.2 | **0.03** | 9.4 | **<0.01** | 16.2 | **<0.01** |
| C×N | 1265.0 | **<0.01** | 107.2 | **<0.01** | 9.5 | **<0.01** | 3.1 | 0.09 |
| C×P | 1718.0 | **<0.01** | 174.1 | **<0.01** | 14.7 | **<0.01** | 88.0 | **<0.01** |
| N×P | 179.0 | **<0.01** | 19.7 | **<0.01** | 10.7 | **<0.01** | 14.3 | **<0.01** |
| T×C×N | 35.0 | **<0.01** | <0.1 | 0.81 | 0.2 | 0.67 | 1.9 | 0.17 |
| T×C×P | 27.0 | **<0.01** | 5.5 | **0.02** | 0.1 | 0.71 | 1.0 | 0.31 |
| T×N×P | 96.0 | **<0.01** | <0.1 | 0.80 | 15.7 | **<0.01** | 3.3 | 0.08 |
| C×N×P | 241.0 | **<0.01** | 0.4 | 0.56 | 8.2 | **<0.01** | 1.2 | 0.28 |
| T×C×N×P | 105.0 | **<0.01** | 3.9 | 0.05 | 22.4 | **<0.01** | 4.5 | **0.04** |

**Table 5.** List of the physiological responses of *E. huxleyi* to the concurrent changes in multiple drivers investigated by the laboratory incubations in the published studies. '↑' represents increase, '↓' represents decrease, and 'n' represents no significant change to simultaneous changes in multiple drivers. C, T, L, N and P represent $CO_2$ (μatm), temperature (°C), light intensity (μmol photons $m^{-2}$ $s^{-1}$), dissolved inorganic nitrogen and phosphate (μmol $L^{-1}$), respectively. Simultaneous changes in multiple drivers were marked in bold. [1] represents De Bodt et al., (2010), [2] Borchard et al., (2011), [3] Sett et al., (2014), [4] Gafar and Schulz, (2018), [5] Tong et al., (2019), [6] Jin et al., (2017), [7] Bretherton et al., (2019), [8] Rost et al., (2002), [9] Feng et al., (2008), [10] Müller et al. (2012), [11] Perrin et al., (2016), [12] Leonardos and Geider, (2005), [13] Matthiessen et al., (2012), [14] Zhang et al., (2019), [15] this study.

| Strain | C | T | L | N | P | μ | POC | PIC | PIC: POC | Cite |
|---|---|---|---|---|---|---|---|---|---|---|
| AC481 | **380 to 750** | **13 to 18** | 150 | 32 | 1 | n | ↑ | ↓ | ↓ | [1] |
| PML B92/11 | **300 to 900** | **14 to 18** | 300 | 29 | 1 | ↑ | n | ↓ | ↓ | [2] |
| PML B92/11 | **400 to 1000** | **10 to 20** | 150 | 64 | 4 | ↑ | ↑ | ↓ | ↓ | [3] |
| PML B92/11 | **400 to 1000** | **10 to 20** | 150 | 64 | 4 | ↑ | ↓ | ↓ | | [4] |
| PML B92/11 | **400 to 1000** | **15 to 24** | 190 | 100 | 10 | ↑ | ↑ | ↓ | ↓ | [5] |
| CCMP2090 | **395 to 1000** | 20 | **57 to 567** | 110 | 10 | ↑ | ↑ | | | [6] |
| NZEH | **390 to 1000** | 20 | **175 to 300** | 100 | 10 | ↓ | ↑ | ↑ | ↑ | [7] |
| PCC124-3 | **390 to 1000** | 20 | **175 to 300** | 100 | 10 | ↑ | n | ↑ | ↑ | [7] |
| PCC70-3 | **390 to 1000** | 20 | **175 to 300** | 100 | 10 | ↑ | n | ↑ | ↑ | [7] |
| PML B92/11 | **140 to 880** | 15 | **80 to 150** | 100 | 6 | ↑ | ↑ | ↓ | ↓ | [8] |
| PML B92/11 | **395 to 1000** | 20 | **54 to 457** | 110 | 10 | ↑ | ↑ | ↓ | ↓ | [6] |
| PML B92/11 | **400 to 1000** | 20 | **50 to 1200** | 64 | 4 | ↑ | ↑ | ↑ | | [4] |
| RCC962 | **390 to 1000** | 20 | **175 to 300** | 100 | 10 | ↓ | ↑ | n | ↓ | [7] |
| CCMP371 | **375 to 750** | **20 to 24** | **50 to 400** | 100 | 10 | ↑ | n | ↓ | ↓ | [9] |
| B62 | **280 to 1000** | 20 | 300 | **88 to 9** | 4 | | ↑ | ↓ | ↓ | [10] |
| RCC911 | 400 | 20 | **30 to 140** | **100 to 5** | 6 | ↑ | ↑ | ↑ | ↑ | [11] |
| RCC911 | 400 | 20 | **30 to 140** | 100 | **6 to 0.6** | ↑ | ↑ | ↑ | ↑ | [11] |
| PML92A | **360 to 2000** | 18 | **80 to 500** | 200 | **6.7 to 40** | n | ↑ | | | [12] |
| A | **460 to** | 16 | 130 | **17 to 9** | **0.2 to 0.5** | | ↓ | ↓ | | [13] |

| | | | | | | | | | | |
|---|---|---|---|---|---|---|---|---|---|---|
| PML B92/11 | **1280 410 to 920** | 20 | **80 to 480** | **100 to 8** | 10 | ↓ | ↓ | ↑ | ↑ | [14] |
| PML B92/11 | **410 to 920** | 20 | **80 to 480** | 100 | **10 to 0.4** | ↓ | ↑ | n | ↓ | [14] |
| PML B92/11 | **370 to 960** | **16 to 20** | **60 to 240** | **24 to 8** | **1.5 to 0.5** | ↓ | ↑ | ↑ | n | [15] |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Step 2: HL HNHP
Step 3: HL LNHP
Step 4: HL HNLP
Step 5: HL LNLP
(a)
LCLT
HCLT
LCHT
HCHT
Ln (cells mL$^{-1}$)
pH
$p$CO$_2$ (µatm)
DIN (µmol L$^{-1}$)
DIP (µmol L$^{-1}$)
Incubation time (day)

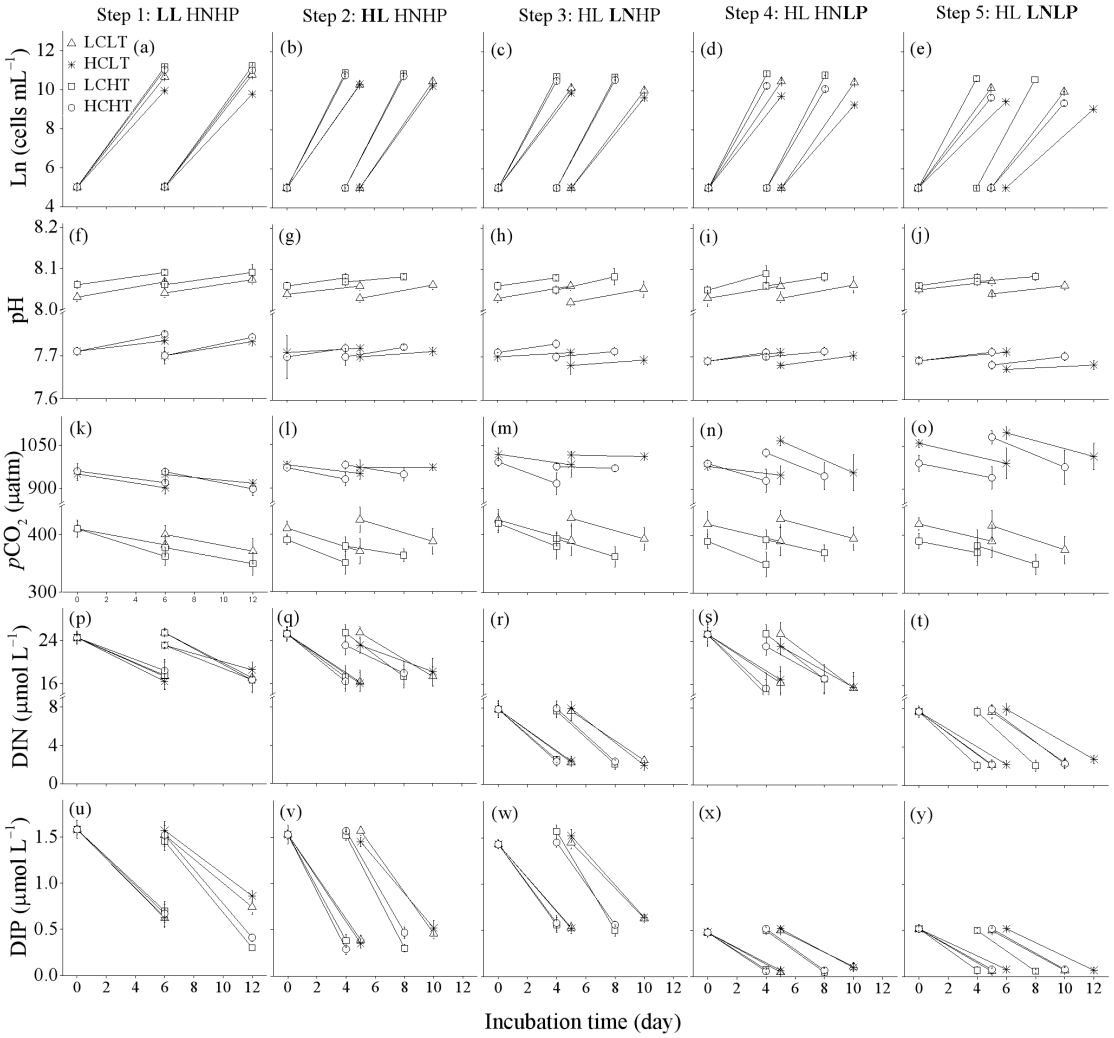
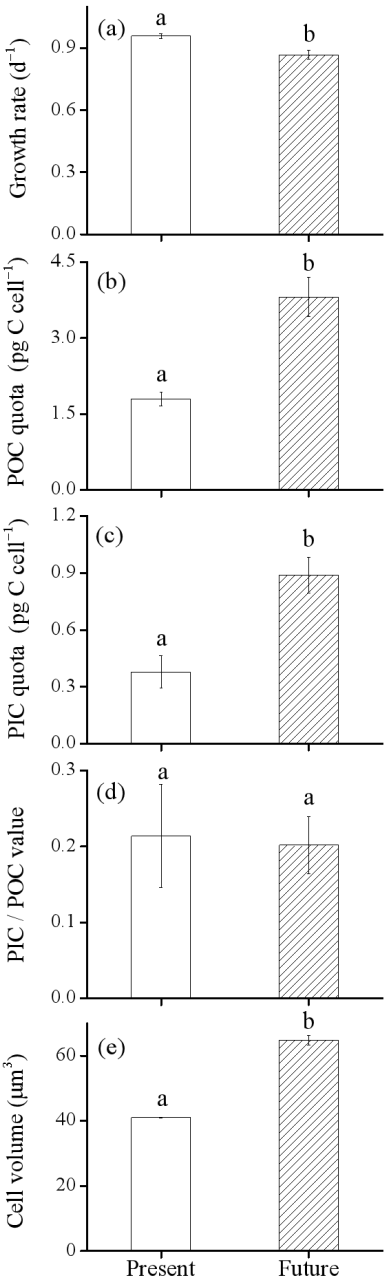

Step 1: LL HNHP
Step 2: HL HNHP
Step 3: HL LNHP
Step 4: HL HNLP
Step 5: HL LNLP
LCLT
HCLT
LCHT
HCHT
Ln (cells mL$^{-1}$)
pH
$pCO_2$ (µatm)
DIN (µmol L$^{-1}$)
DIP (µmol L$^{-1}$)
Incubation time (day)

Figure 1

1275

1276

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1280

1281

1282

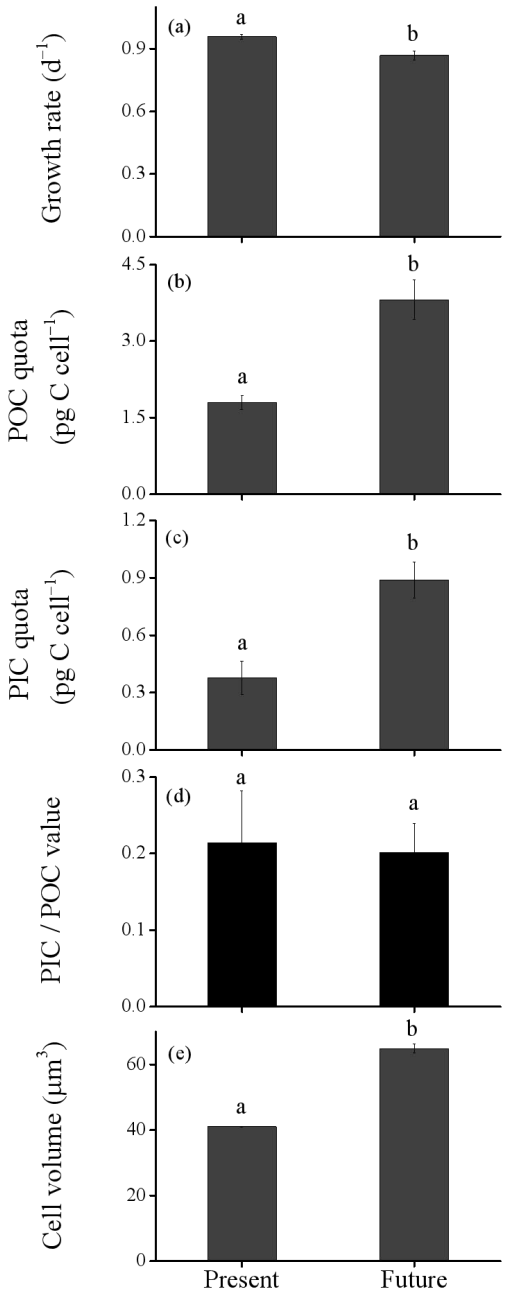
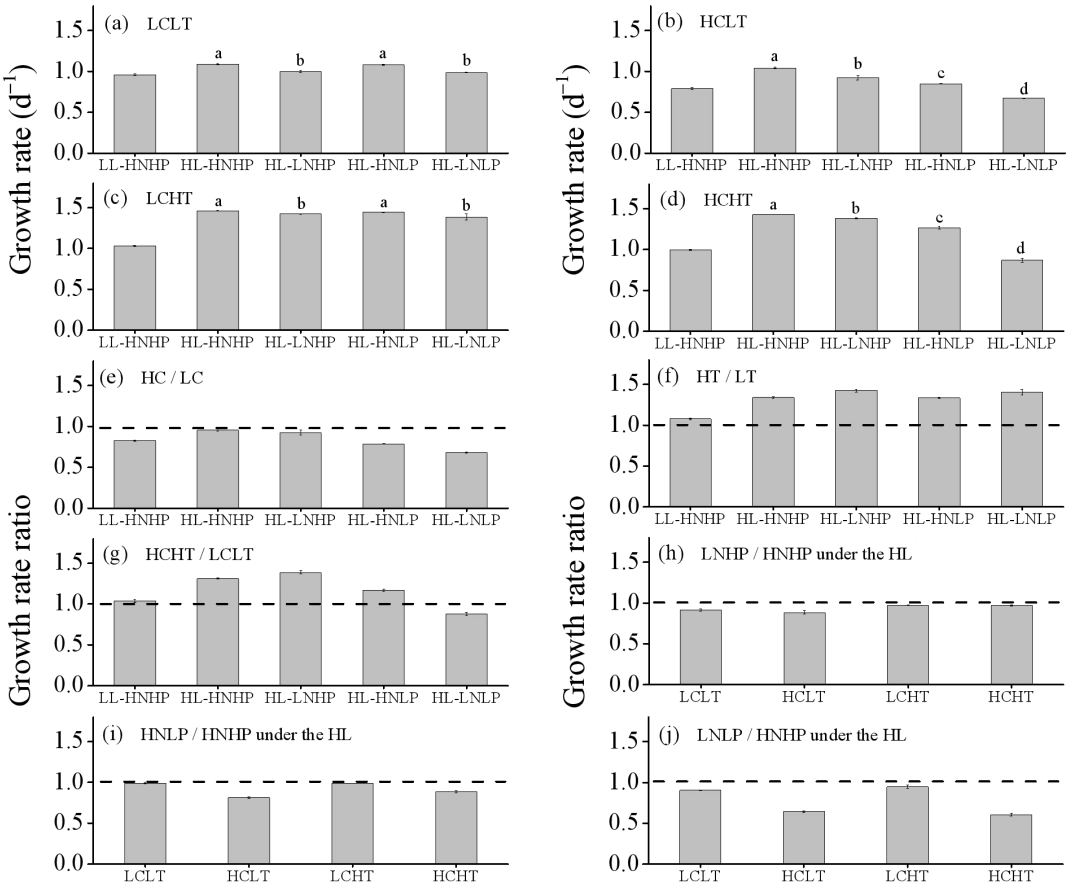

(a)
Growth rate (d$^{-1}$)
0.0
0.3
0.6
0.9
a
b
(b)
POC quota (pg C cell$^{-1}$)
0.0
1.5
3.0
4.5
a
b
(c)
PIC quota (pg C cell$^{-1}$)
0.0
0.3
0.6
0.9
1.2
a
b
(d)
PIC / POC value
0.0
0.1
0.2
0.3
a
a
(e)
Cell volume (μm$^3$)
0
20
40
60
a
b
Present
Future

Figure 2


(a) LCLT
(b) HCLT
(c) LCHT
(d) HCHT
(e) HC / LC
(f) HT / LT
(g) HCHT / LCLT
(h) LNHP / HNHP under the HL
(i) HNLP / HNHP under the HL
(j) LNLP / HNHP under the HL
Growth rate (d$^{-1}$)
Growth rate ratio
LL-HNHP
HL-HNHP
HL-LNHP
HL-HNLP
HL-LNLP
LCLT
HCLT
LCHT
HCHT

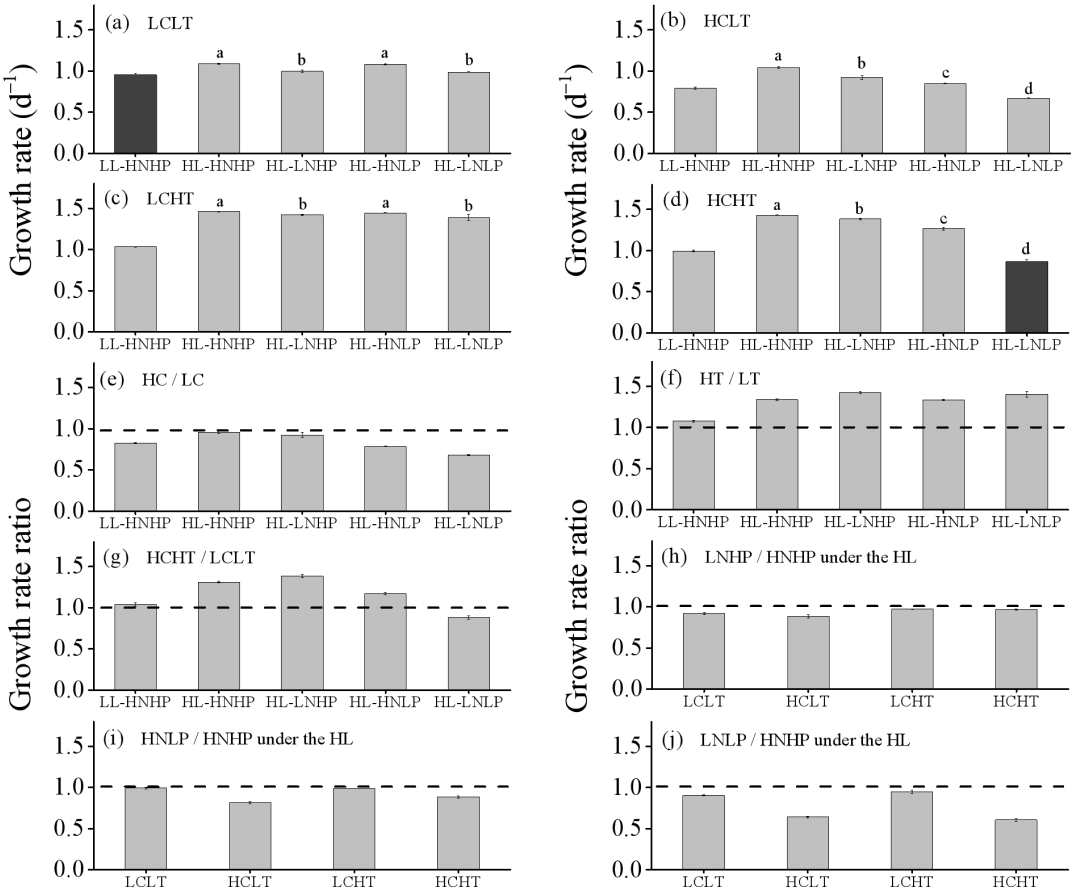
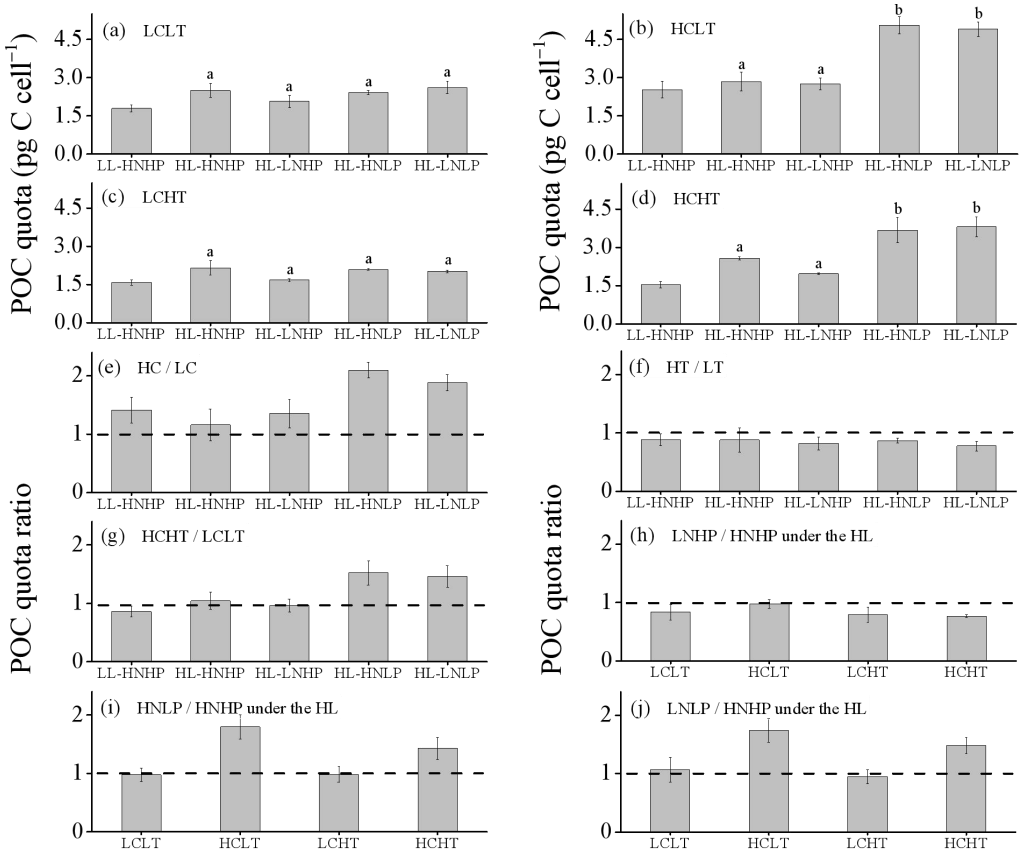

(a) LCLT
(b) HCLT
(c) LCHT
(d) HCHT
(e) HC / LC
(f) HT / LT
(g) HCHT / LCLT
(h) LNHP / HNHP under the HL
(i) HNLP / HNHP under the HL
(j) LNLP / HNHP under the HL
Growth rate (d$^{-1}$)
Growth rate ratio
LL-HNHP
HL-HNHP
HL-LNHP
HL-HNLP
HL-LNLP
LCLT
HCLT
LCHT
HCHT

Figure 3


(a) LCLT
(b) HCLT
(c) LCHT
(d) HCHT
(e) HC / LC
(f) HT / LT
(g) HCHT / LCLT
(h) LNHP / HNHP under the HL
(i) HNLP / HNHP under the HL
(j) LNLP / HNHP under the HL
POC quota (pg C cell$^{-1}$)
POC quota ratio
LL-HNHP
HL-HNHP
HL-LNHP
HL-HNLP
HL-LNLP
LCLT
HCLT
LCHT
HCHT

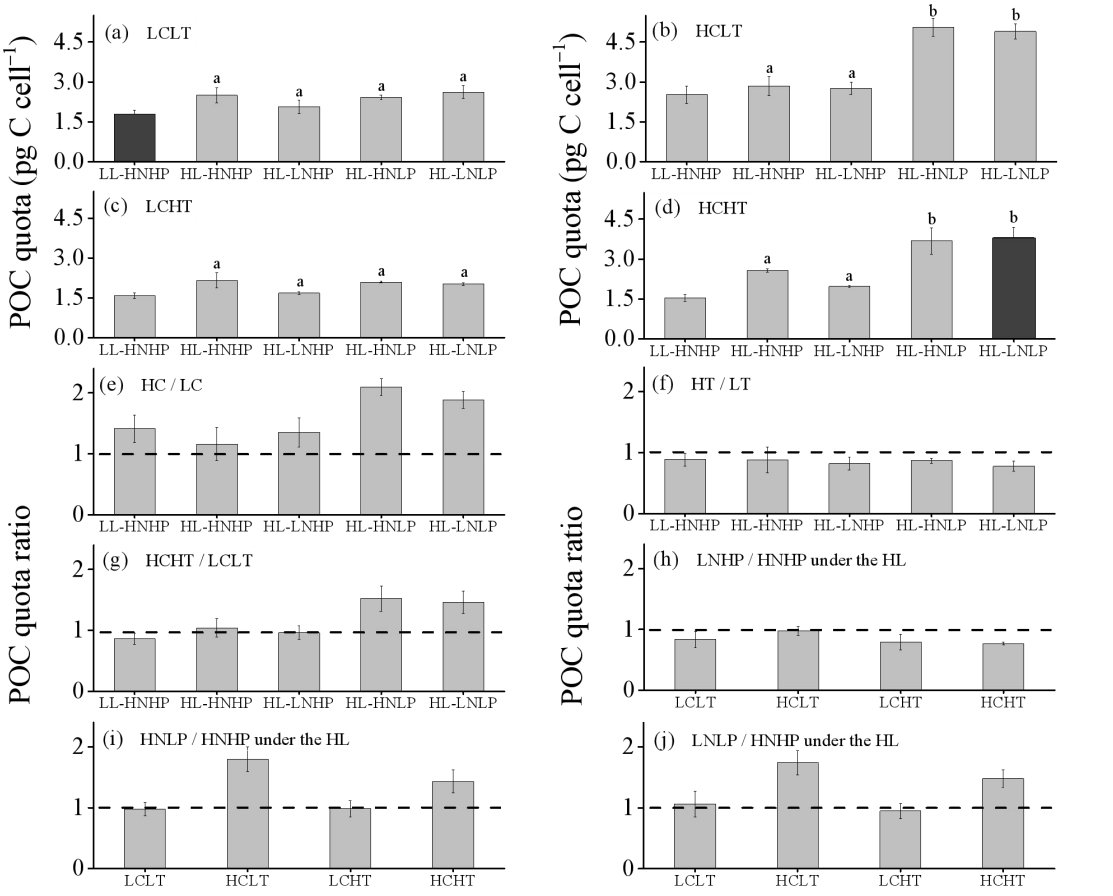
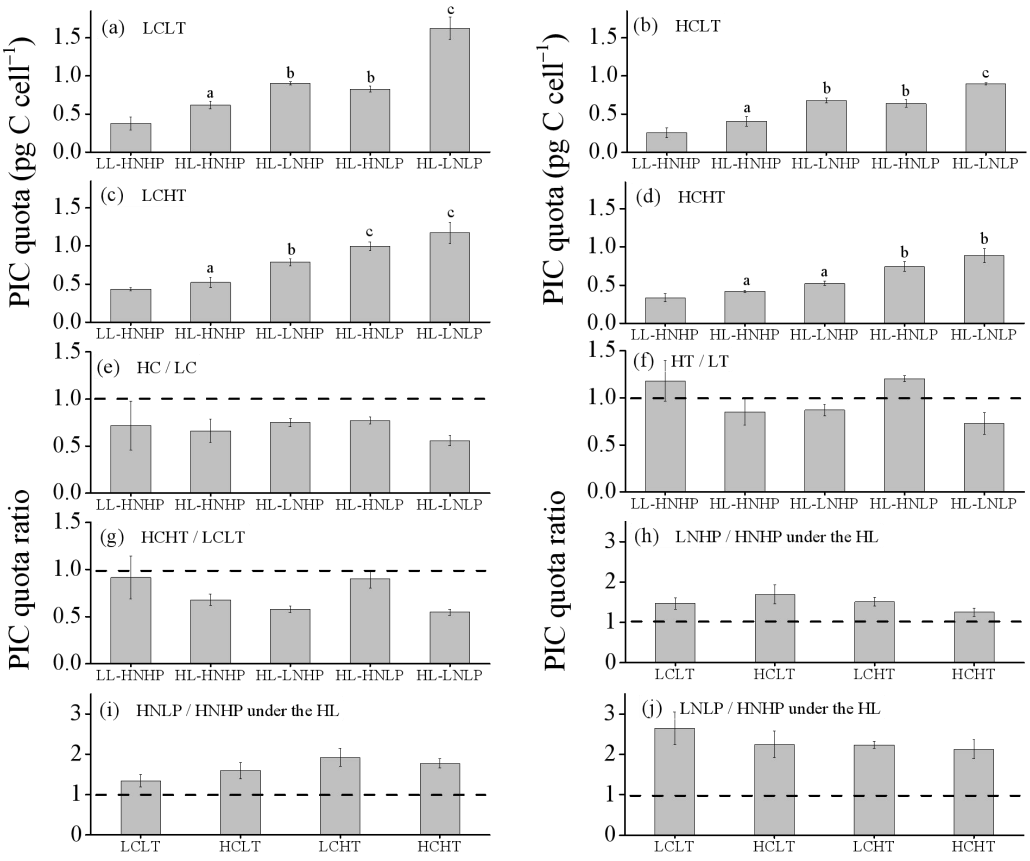

(a) LCLT
(b) HCLT
(c) LCHT
(d) HCHT
(e) HC / LC
(f) HT / LT
(g) HCHT / LCLT
(h) LNHP / HNHP under the HL
(i) HNLP / HNHP under the HL
(j) LNLP / HNHP under the HL
POC quota (pg C cell$^{-1}$)
POC quota ratio
LL-HNHP
HL-HNHP
HL-LNHP
HL-HNLP
HL-LNLP
LCLT
HCLT
LCHT
HCHT

Figure 4


(a) LCLT
(b) HCLT
(c) LCHT
(d) HCHT
(e) HC / LC
(f) HT / LT
(g) HCHT / LCLT
(h) LNHP / HNHP under the HL
(i) HNLP / HNHP under the HL
(j) LNLP / HNHP under the HL
PIC quota (pg C cell$^{-1}$)
PIC quota ratio
LL-HNHP
HL-HNHP
HL-LNHP
HL-HNLP
HL-LNLP
LCLT
HCLT
LCHT
HCHT

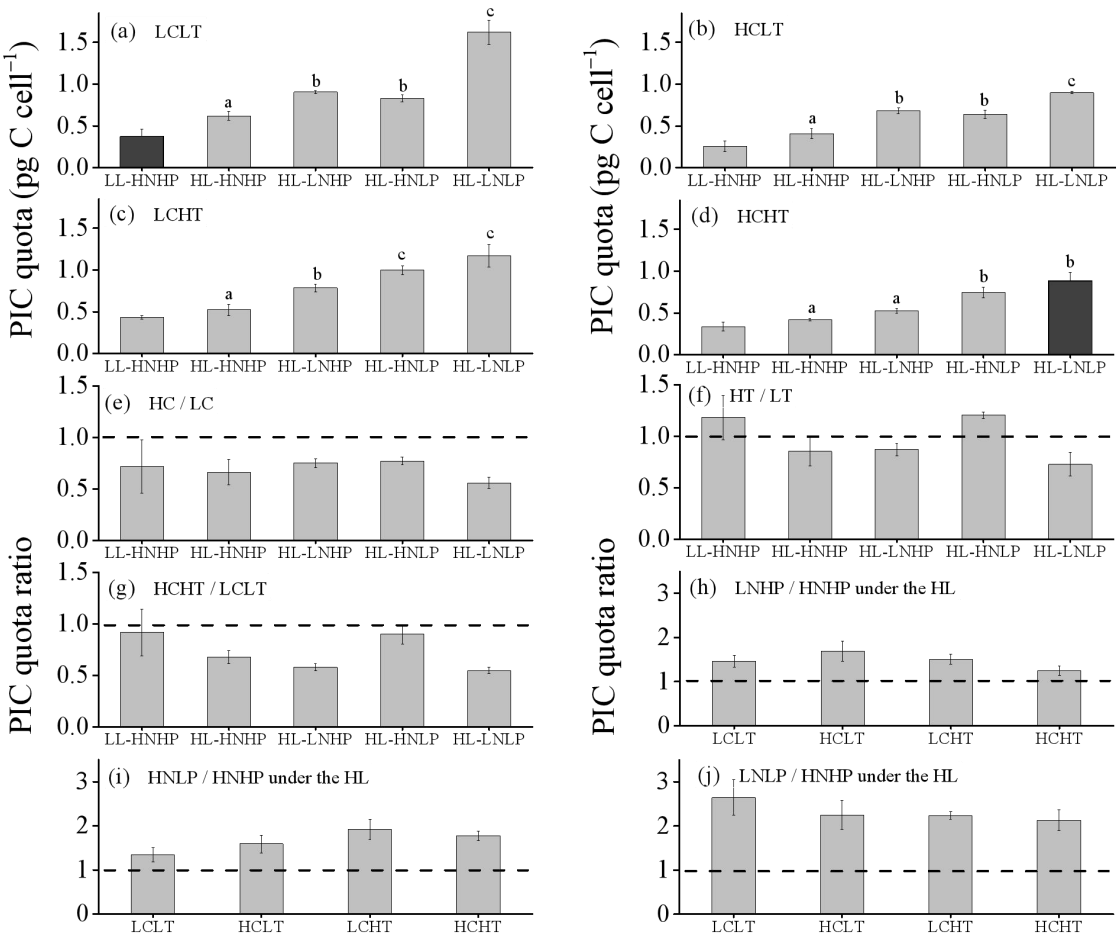
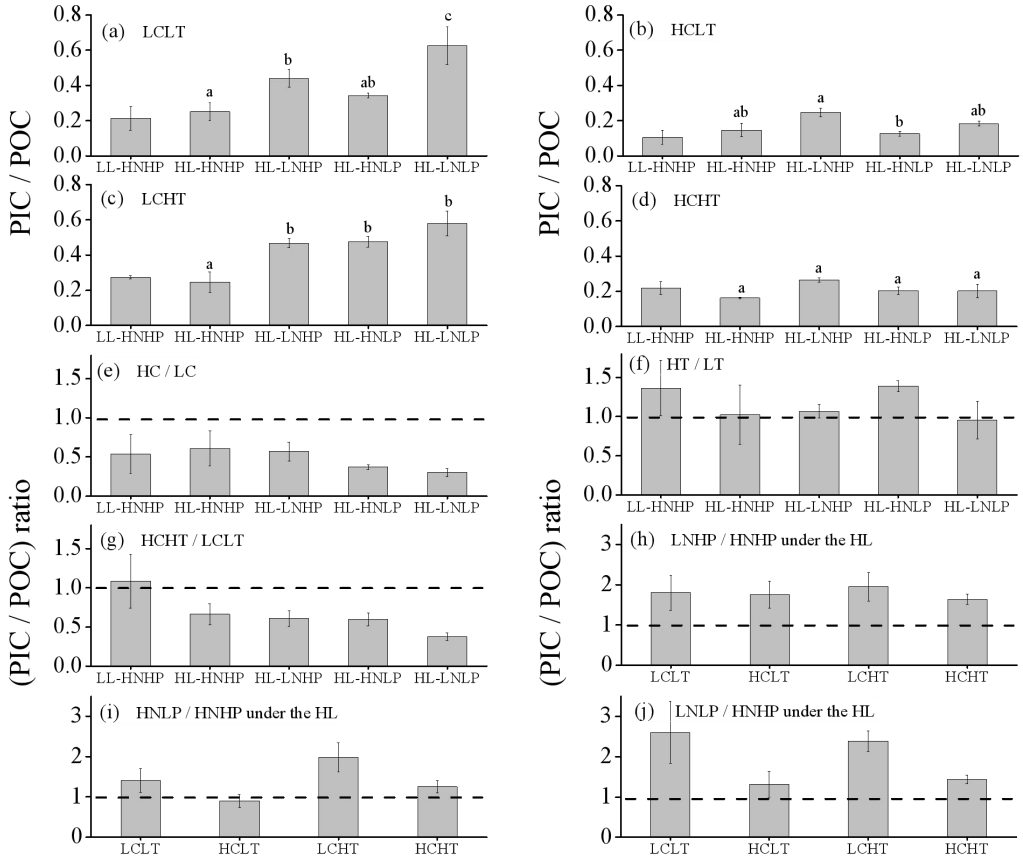

PIC quota (pg C cell$^{-1}$)
PIC quota ratio
(a) LCLT
(b) HCLT
(c) LCHT
(d) HCHT
(e) HC / LC
(f) HT / LT
(g) HCHT / LCLT
(h) LNHP / HNHP under the HL
(i) HNLP / HNHP under the HL
(j) LNLP / HNHP under the HL
LL-HNHP
HL-HNHP
HL-LNHP
HL-HNLP
HL-LNLP
LCLT
HCLT
LCHT
HCHT

Figure 5


(a) LCLT
(b) HCLT
(c) LCHT
(d) HCHT
(e) HC / LC
(f) HT / LT
(g) HCHT / LCLT
(h) LNHP / HNHP under the HL
(i) HNLP / HNHP under the HL
(j) LNLP / HNHP under the HL
PIC / POC
(PIC / POC) ratio
LL-HNHP
HL-HNHP
HL-LNHP
HL-HNLP
HL-LNLP
LCLT
HCLT
LCHT
HCHT

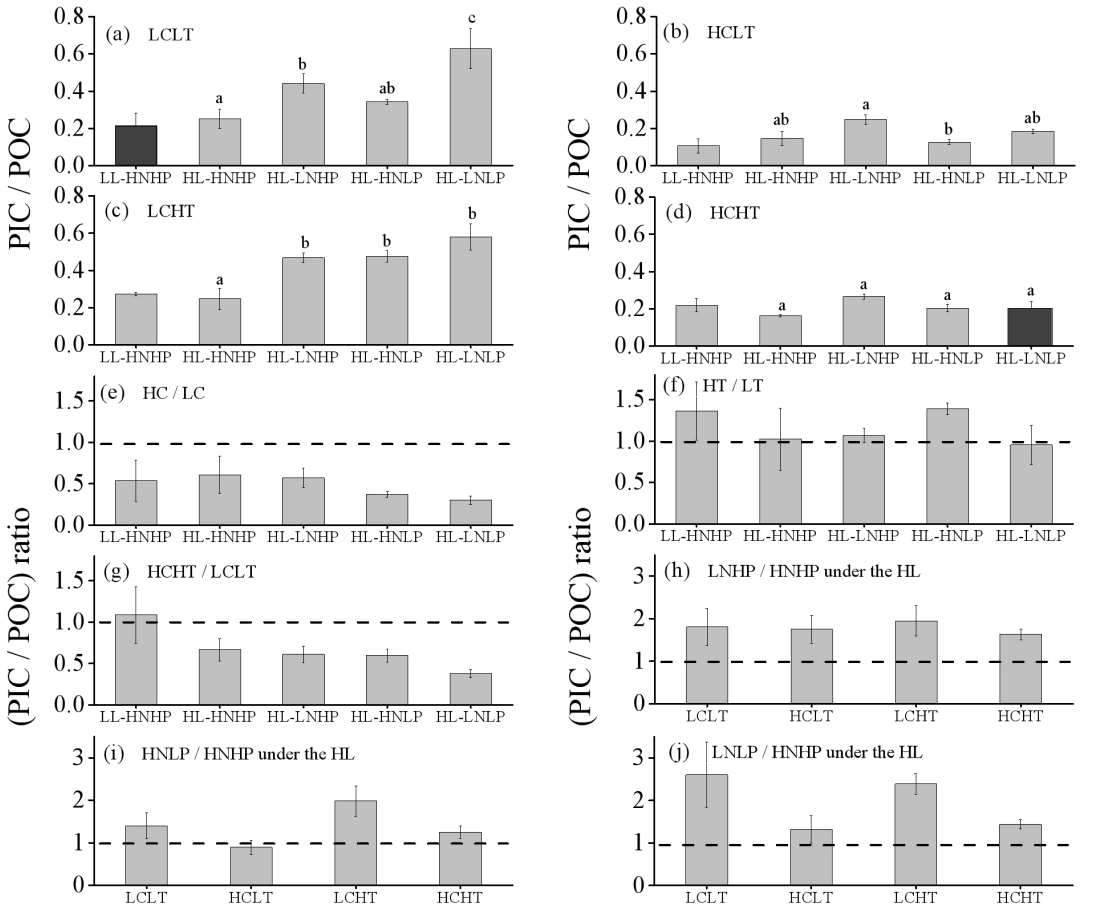

PIC / POC
(PIC / POC) ratio
(a) LCLT
(b) HCLT
(c) LCHT
(d) HCHT
(e) HC / LC
(f) HT / LT
(g) HCHT / LCLT
(h) LNHP / HNHP under the HL
(i) HNLP / HNHP under the HL
(j) LNLP / HNHP under the HL
LL-HNHP
HL-HNHP
HL-LNHP
HL-HNLP
HL-LNLP
LCLT
HCLT
LCHT
HCHT

Figure 6