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Interactive comment on "Factors controlling Carex brevicuspis leaf litter decomposition and its contribution to surface soil organic carbon pool at different water levels" by Lianlian Zhu et al.

Lianlian Zhu et al.

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Comment 2.1 Line 20 and 46: I recommend including (Cao et al. 2020), and references therein, that address aboveground litter decomposition on SOC pools. Response 2.1 Thank you for your recommendation, we had cited (Bowden et al., 2014; Cao et al., 2020) in the revised manuscript in line 46. Comment 2.2 Line 30: The SOC increase due to litter application (Figure 5d) appears to be calculated from Figure 5a, but I could not reconcile the value for -25 cm. While Figure 5 does seem to support that litter increases SOC, I have two concerns about this presentation. First, the differences in Figure 5a for the 0 cm water levels is labelled as significant, but the error bars clearly

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gestion. We have add the value into the manuscript, and cited the references (Kayranli

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et al., 2010; Kochy et al., 2015). The sentences have been rephrased as follows: Wetlands are important terrestrial carbon pools. They contain between 82 and 158 Pg SOC, which depending on the definition of "wetland" (Kayranli et al., 2010; Kochy et al., 2015). Please see Line40-41. Comment 2.4 Line 52: The Sun et al., 2019 study does not support the statement that litter decomposition stabilized the soil organic carbon pool. Litter decomposition made DOC more mobile and labile, which the authors suggested could lead to SOC stability after processing by soil microbes. Response 2.4 We are sorry for the mistake. The sentences have been rephrased as: In contrast, a recent study found that litter decomposition stabilized the soil carbon pool after processing by soil microbes in the Jiaozhou Bay wetland (Sun et al., 2019). Please see Line51-52. Comment 2.5 Lines 54-56: Aerts (1997) addresses litter decomposition in non-wetland sites where shredder invertebrates (detritovores) are important, but their role in wetland settings is more uncertain (Inkley, Wissinger, and Baros 2008). Shredding would be an important physio-chemical control on DOC leaching. Response 2.5 We thank the reviewer very much for the commendation and suggestion. The references have been changed, and the sentences have been rephrased as follows: Litter decomposition is a physicochemical processes that reduces litter to its elemental chemical constituents (Berg and McClaugherty, 2003). Litter decomposition rates are determined mainly by environmental factors (climatic and soil conditions), litter quality (litter composition such as C, N, and lignin content) and decomposer organisms (microorganisms and invertebrates) (Yu et al., 2020; Yan et al., 2018). Please see Line 53-57. Comment 2.6 Line 62: Zhang 2019 supports the statement that water levels affected microbial activity, but leaching and fragmentation were only discussed, not measured. Response 2.6 We are sorry for the ambiguity. The references have been changed to (Van de Moortel et al., 2012), which designed a leaching experiment to clarify the leaching process of litter decomposition. Please see Line 62.

Comment 2.7 Line 64: This is a mischaracterization of the Upton, 2018 study. Perhaps a better reference is (Hoyos-Santillan et al. 2015). However, clarification is needed because Hoyos-Santillan states that roots (not litter) are the main source of SOC in

peatlands, but litter strongly influences root decomposition rates, particularly near the surface. Response 2.7 We are sorry for our carelessness. The sentence has been rephrased as follows: Leaf litter contributes more to soil organic carbon than fine roots (Cao et al., 2020), litter also strongly influences root decomposition rates, particularly near the soil surface (Hoyos-Santillan et al., 2015). Please see Line 64-66.

Comment 2.8 Line 151. The Olson (1963) simple decay model assumes constant k, which you demonstrated is not a constant. Although use of this decay model is common in the literature, it is an oversimplification. This does not adversely affect your comparative analysis, but the paper would be strengthened with a more sophisticated analysis, such as a double exponential decay model (Berg 2014 or Wider and Lang 1982). Response 2.8 Thank you for the constructive suggestions. We have modified the model based on your suggestion to highlight the instantaneous rate variation of litter decomposition. The model is: M_(t_n)=M_(t_(n-1)) e^(ãĂŰ-kãĂŮ_n (t_n-t_(n-1)) 1))) Where M (t n) is the litter dry matter weight at nth sampling (g), M (t (n-1)) is the litter dry matter weight at (n-1)th sampling (g), t n-t (n-1)is the time between the nth and (n-1)th sampling, k nis the instantaneous decomposition rate at the nth sampling. (Please see line 150-155) This model would be more accurate. The result was as following: The instantaneous litter decomposition rate was highest at initial and slowly decreased and stabilized at all three water levels. The maximum decomposition rates at the -25 cm, 0 cm, and +25 cm water levels were 0.00527 d-1, 0.00908 d-1, and 0.01307 d-1, respectively (Fig. 2b). (Please see line 189-192)

Based on the change of the instantaneous decomposition rate, we recalculated the multiple regression model which was used to analyze the intrinsic litter decomposition rate-limiting factor. The models are as follows (Table 1):

Water level (cm) Multiple regression model F R2 P -25 R=-0.715L-0.443C+0.033 5.738 0.727 0.006 0 R=-928LN-0.233CN+0.023 5.928 0.927 < 0.001 +25 R=-0.717LN+0.016 9.543 0.793 0.002

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about 2.51% organic carbon to S-SOCP, which is largely ascribe to the slower decomposition rate and soil carbon lost by metabolism of the microbes (i.e. actinomycete).

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Second, the intrinsic limiting factors may be different among three water levels. Third,

the contribution of leaf decomposition to S-SOCP was relatively higher at the +25 cm water level. Comment 2.16 Line 102: Did the litter bags float? Did you need to pin them in contact with the soil surface? Response 2.16 We are sorry for the negligence. All litter bags were fixed to the soil surface with bamboo sticks. And the sentence has been added in section 2.2 in line 107-108. Comment 2.17 Line 105 - 107: Clarify how many soil cores were used in each pond for each purpose and how they were prepared (e.g. were soils blended prior to starting the experiment). The text is confusing. Response 2.17 We have clarified that all the soil cores were undisturbed soil. The experiment was conducted in nine cement ponds (2 m \times 2 m \times 1 m) (see Line 107-108 in revised manuscript). Three soil core sets were placed in each pond. One was designated the litter removal control (S), the second was distributed on the soil surface in 15 litter bags to observe the effects of leaf litter input on soil carbon pool (L), and the third was distributed on the soil surface in 15 litter bags to monitor the litter decomposition rate and process (D) in Line 113-116 in revised manuscript.

Comment 2.18 Line 183: You use capital letters (Fig 2A) in text references, but lower-case letters in the Figures. Response 2.18 We are sorry for the mistake. The capital letters in the text references have been changed into lower-case letters. Comment 2.19 Line 187/188 and Line 247: I would not interpret your data that decomposition rates "rapidly increased" – the decomposition rate at time t=0 is undefined. Response 2.19 We are sorry for our obscure writing. The sentences have been rephrased as: The instantaneous litter dry weight decomposition rate was highest at initial and slowly decreased and stabilized at all three water levels. Please see line 189-190. Comment 2.20 Figure 4: Are these figures reporting mass? The units are nmol g-1. Response 2.20 We are sorry for that we didn't clearly define the calculation method about microbial community structure. These figures were used to report the PLFA molar mass concentration. This is a common way to show the microbial content (Zhao et al., 2015). We calculated PLFA mass content first, PLFA (ng/g dry soil) = (Response of unkown PLFA/ Response of 19:0 internal standard)×concentration of 19:0 internal standard: 5

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2020. Zhao, J., Zeng, Z. X., He, X. Y., Chen, H. S., and Wang, K. L.: Effects of

monoculture and mixed culture of grass and legume forage species on soil microbial community structure under different levels of nitrogen fertilization, Eur. J. Soil Biol., 68,

61-68, 10.1016/j.ejsobi.2015.03.008, 2015.

Please also note the supplement to this comment: https://bg.copernicus.org/preprints/bg-2020-266/bg-2020-266-AC2-supplement.pdf

Interactive comment on Biogeosciences Discuss., https://doi.org/10.5194/bg-2020-266, 2020.

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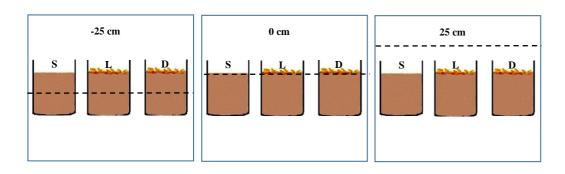


Fig. 1. Figure 1: Schematic diagram of the experimental setup. The dotted line represents the water level. L represents litter which was distributed on the soil surface in 15 litter bags to observe the effect

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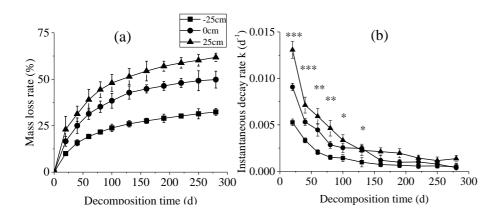


Fig. 2. Figure 2: Percentage litter dry weight loss and decomposition rate during C. brevicuspis decomposition at three water levels (-25 cm, 0 cm, and +25 cm). *, **, and *** represent significant difference

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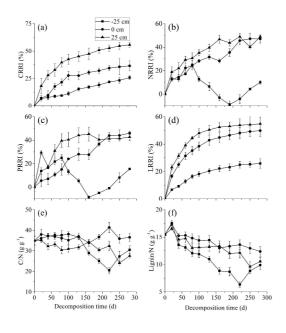


Fig. 3. Figure 3: Percentage (mean \pm SE) of carbon relative release index (CRRI), nitrogen relative release index (NRRI), phosphorus relative release index (PRRI), lignin relative release index (LRRI), C/N ra

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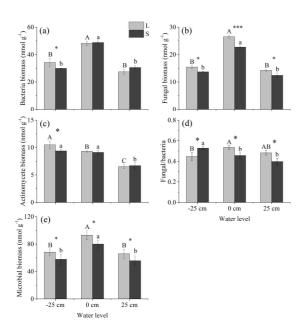


Fig. 4. Figure 4: Microbial community structure under litter input and litter removal at three water levels. Different uppercase letters among vertical bars indicate significant differences among the three wa

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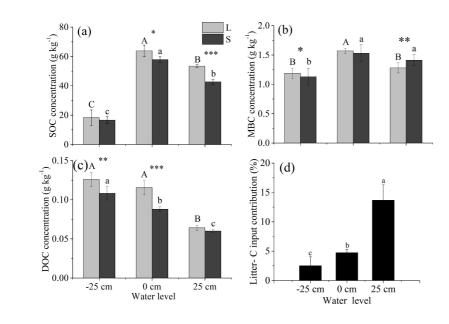


Fig. 5. Figure 5: Concentrations of SOC (A), MBC (B), DOC (C) between the litter input (L) and litter removal (S) groups and the litter-C input contribution (D) under three water levels at the end of the expe

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