

1 **Factors controlling *Carex brevicuspis* leaf litter**  
2 **decomposition and its contribution to surface soil organic**  
3 **carbon pool at different water levels**

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18

19 **Abstract.** Litter decomposition plays a vital role in wetland carbon cycling. However, the contribution  
20 of aboveground litter decomposition to the wetland soil organic carbon (SOC) pool has not yet been  
21 quantified. Here, we conducted a *Carex brevicuspis* leaf litter input experiment to clarify the intrinsic  
22 factors controlling litter decomposition and quantify its contribution to the SOC pool at different water  
23 levels. The *Carex* genus is ubiquitous to global freshwater wetlands. We sampled this plant leaf litter at  
24 -25, 0, and +25 cm relative to the soil surface over 280 days and analysed leaf litter decomposition and  
25 its contribution to the SOC pool. The percentage litter dry weight loss and the instantaneous litter dry  
26 weight decomposition rate were the highest at +25 cm water level (61.8%, 0.01307d<sup>-1</sup>), followed by the  
27 0 cm water level (49.8%, 0.00908 d<sup>-1</sup>), and the lowest at -25 cm water level (32.4%, 0.00527 d<sup>-1</sup>).  
28 Significant amounts of litter carbon, nitrogen, and phosphorus were released at all three water levels.  
29 Litter input significantly increased the soil microbial biomass and fungal density but had nonsignificant  
30 impacts on soil bacteria, actinomycetes, and the fungal/bacterial concentrations at all three water levels.  
31 Compared with litter removal, litter **addition** increased the SOC by 16.93%, 9.44%, and 2.51% at the  
32 +25 cm, 0 cm, and -25 cm water levels, respectively. Hence, higher water levels facilitate the release of  
33 organic carbon from leaf litter into the soil via water leaching. In this way, they increase the soil carbon  
34 pool. At lower water levels, soil carbon is lost due to the slower litter decomposition rate and active  
35 microbial (actinomycete) respiration. Our results revealed that the water level in natural wetlands  
36 influenced litter decomposition mainly by leaching and microbial activity, by extension, and affected  
37 the wetland surface carbon pool.

38 **Key words:** *Carex brevicuspis*; decomposition; leaf litter; soil surface organic carbon pool; water level

## 39 **1 Introduction**

40 Wetlands are important terrestrial carbon pools. Depending on the definition of “wetland”, they contain  
41 between 82 and 158 Pg SOC, (Kayranli et al., 2010; Kochy et al., 2015). The surface soil organic carbon  
42 (SOC) pool (S-SOCP) and its turnover are sensitive to climate, topography, and hydrological conditions  
43 (Wang et al., 2016; Zhang et al., 2017; Pinto et al., 2018).

44 Leaf litter decomposition is a major biotic carbon input route from vegetation to S-SOCP in wetland  
45 ecosystems (Whiting and Chanton, 2001; Moriyama et al., 2013). However, the reported impacts of litter  
46 decomposition on the soil carbon pool are highly variable (Bowden et al., 2014; Cao et al., 2020). Litter

47 input destabilised carbon storage by stimulating soil mineralisation and increasing labile soil carbon  
48 fractions (microbial biomass carbon [MBC], soil dissolved organic carbon [DOC]), and enzyme activity  
49 in the freshwater marshland of Northeast China (Song et al., 2014). It also promoted soil carbon loss via  
50 CO<sub>2</sub> emissions and microbial activity in alpine and coastal wetlands (Gao et al., 2016; Liu et al., 2017).  
51 In contrast, a study has recently found that litter decomposition stabilised the soil carbon pool after  
52 processing by soil microbes in the Jiaozhou Bay wetland (Sun et al., 2019).

53 Litter decomposition is a physicochemical process that reduces litter to its elemental chemical  
54 constituents (Berg and Mcclaugherty, 2003). Litter decomposition rates are determined mainly by  
55 environmental factors (climatic and soil conditions), litter quality (litter composition such as C, N, and  
56 lignin content) and decomposer organisms (microorganisms and invertebrates) (Yan et al., 2018; Yu et  
57 al., 2020). A previous study showed that regional and global environmental conditions explain > 51% of  
58 the variation in litter decomposition rate (Zhang et al., 2019). In wetland ecosystems, the water level  
59 ecosystem processes determine soil aerobic and anaerobic conditions which, in turn, affect the microbial  
60 decomposition of litter and SOC decomposition (Liu et al., 2017; Yan et al., 2018). An earlier study  
61 reported that high soil moisture content and long flooding periods facilitate litter decomposition by  
62 promoting leaching, fragmentation, and microbial activity (Van de Moortel et al., 2012). The water level  
63 may contribute to soil physicochemical conditions which, in turn, regulate litter decomposition (Xie et  
64 al., 2016b). Leaf litter contributes more to soil organic carbon than fine roots (Cao et al., 2020), litter also  
65 strongly influences root decomposition rates, particularly near the surface (Hoyos-Santillan et al., 2015).  
66 However, the contribution of litter decomposition to the **S-SOCP** pool has seldom been quantified.

67 Peng et al. reported that the organic carbon density in Dongting Lake wetland soil at 1 m depth was 127.3  
68 ± 36.1 t hm<sup>-2</sup> and the carbon density in the 0–30 cm topsoil was 46.5 ± 19.7 t hm<sup>-2</sup> (Peng et al., 2005).  
69 *Carex brevicuspis* is a dominant species in the Dongting Lake wetland and has large carbon reserves  
70 (~6.5 × 10<sup>6</sup> t y<sup>-1</sup>) (Kang et al., 2009). However, due to the dam construction upstream of Dongting Lake,  
71 the water regime varies considerably (early water withdrawal and decline of groundwater in non-flood  
72 season) in recent years, leading to a significant carbon loss in this floodplain wetland (Hu et al., 2018;  
73 Deng et al., 2018).

74 Here, we investigated *C. brevicuspis* **leaf litter** decomposition and its contribution to the SOC pool at  
75 three water levels (-25 cm, 0 cm, and +25 cm relative to the soil surface) to find the factors controlling  
76 *C. brevicuspis* leaf litter decomposition and quantify the contribution of litter decomposition to the SOC

77 pool. We tested the following hypotheses. Firstly, the water level has a significant effect on litter  
78 decomposition. Secondly, the intrinsic factors that control litter decomposition rate at three water levels  
79 are different. Thirdly, the contribution of leaf decomposition to S-SOCP is relatively higher at the +25  
80 cm water level.

## 81 **2 Materials and methods**

### 82 **2.1 Soil core collection and leaf litter preparation**

83 Dongting Lake (28°30'–30°20' N, 111°40'–113°10' E) is the second-largest freshwater lake in China. It  
84 is connected to the Yangtze River via tributaries. Dongting Lake wetlands are characterised by large  
85 seasonal fluctuations in water level ( $\leq 15$  m) and are completely flooded during June–October and  
86 exposed during November–May (Chen et al., 2016). Soil cores (40 cm diameter  $\times$  50 cm length) were  
87 taken from the wetland. Leaf litter was collected in May 2017 from an undisturbed *Carex brevicuspis*  
88 community at the sampling site (29°27'2.02" N, 112°47'32.28" E) of the Dongting Lake Station for  
89 Wetland Ecosystem Research, which is part of the China Ecosystem Research Network. The litter was  
90 cleaned with distilled water, oven-dried at 60 °C to a constant weight, and cut into pieces 5–10 cm long.  
91 Pre-weighed litter samples (5 g;  $10.73 \pm 0.28$  g kg<sup>-1</sup> N,  $0.89 \pm 0.04$  g kg<sup>-1</sup> P,  $40.23 \pm 2.6\%$  organic C, and  
92  $17.83 \pm 0.25\%$  lignin) were placed into 10 cm  $\times$  15 cm 1 mm mesh nylon bags. This mesh size excluded  
93 macroinvertebrates but permitted microbial colonisation and litter fragment leaching (Xie et al., 2016a).

### 94 **2.2 Experimental design**

95 There were three water level treatments (-25 cm, 0 cm, and +25 cm relative to the soil surface) nested by  
96 two litter treatments (input vs removal) and three replicates. The experiment was conducted in nine  
97 cement ponds (2 m  $\times$  2 m  $\times$  1 m) at the Dongting Lake Station for Wetland Ecosystem Research. For the  
98 -25 cm treatment, the water level was 25 cm below the soil surface. For the 0 cm treatment, the soil was  
99 fully wetted with belowground water (the belowground water was extracted from the well in the  
100 experiment site by a water pump) but without surface pooling. For the +25 cm treatment, the water level  
101 was 25 cm above the soil surface. Water levels were adjusted weekly using belowground water (TOC:  
102 3.44 mg L<sup>-1</sup>; TN: 0.001 mg L<sup>-1</sup>; TP: 0.018 mg L<sup>-1</sup>). Three soil core sets were placed in each pond. One  
103 was designated the litter removal control (S), the second was distributed on the soil surface with 15 litter

104 bags to observe the effects of leaf litter input on soil carbon pool (L), and the third was distributed on the  
105 soil surface with 15 litter bags to monitor the litter decomposition rate and process (D) (Fig. 1). Litter  
106 bags were laid flat on the surface of the soil. Each litter bag was not filled, and there are a little overlap  
107 between the litter bags where there is no litter. All the litter bags were fixed to the soil surface with  
108 bamboo sticks. The experiment started on 20 August 2017 and lasted 280 d. By that time, no further  
109 significant change in litter dry weight was observed. Before incubation, three litter and three soil samples  
110 (SOC: 63.32 g kg<sup>-1</sup>) were collected to determine their initial quality. Litter bags were randomly collected  
111 from treatment D after 20 d, 40 d, 60 d, 80 d, 100 d, 130 d, 160 d, 190 d, 220 d, 250 d, and 280 d. After  
112 collection, the litter samples were separated, cleaned with distilled water, and oven-dried at 60 °C to a  
113 constant weight ( $\pm 0.01$  g). All samples were pulverised and passed through a 0.5-mm mesh screen for  
114 litter quality analysis. At the end of incubation, the surface soil (0–5 cm, ~600 g FW) was collected to  
115 eliminate the influences of root decomposition on the soil organic pool. The soil samples were placed in  
116 aseptic sealed plastic bags and transported to the laboratory. The samples were sieved (< 2 mm),  
117 thoroughly mixed, and divided into three subsamples. The first subsample (~150 g) was stored at -20 °C  
118 and freeze-dried for phospholipid fatty acid (PLFA) analysis. The second one (~150 g) was stored at 4 °C  
119 for MBC and DOC measurements. The third subsample (~300 g) was air-dried for physicochemical  
120 analysis.

### 121 **2.3 Litter quality analyses**

122 Litter organic carbon content was analysed by the H<sub>2</sub>SO<sub>4</sub>-K<sub>2</sub>Cr<sub>2</sub>O<sub>7</sub> heat method. Litter nitrogen was  
123 extracted by Kjeldahl digestion and quantified with a flow injection analyser (AA3; Seal Analysisten  
124 GmbH, Langenselbold, Germany) (Xie et al., 2017). Litter phosphorus content was quantified by the  
125 molybdenum-antimony anti-spectrophotometric method. The lignin content was measured by hydrolysis  
126 (72% H<sub>2</sub>SO<sub>4</sub>) (Graça et al., 2005; Xie et al., 2017).

### 127 **2.4 Soil quality analyses**

#### 128 **2.4.1 Soil chemical analyses**

129 SOC was determined by wet oxidation with KCr<sub>2</sub>O<sub>7</sub> + H<sub>2</sub>SO<sub>4</sub> and titration with FeSO<sub>4</sub> (Xie et al., 2017).

130 Soil DOC was extracted with K<sub>2</sub>SO<sub>4</sub> and measured with a TOC analyser (TOC-VWP; Shimadzu Corp.,  
131 Kyoto, Japan). MBC was analysed by chloroform fumigation, K<sub>2</sub>SO<sub>4</sub> extraction, and measured with a  
132 TOC analyser (TOC-VWP, Shimadzu Corp., Kyoto, Japan) (Tong et al., 2017).

## 133 2.4.2 Soil microbial composition

134 The total and specific microbial group biomass values and the microbial community structure were  
135 estimated by phospholipid fatty acid (PLFA) analysis. The PLFAs were extracted from 8 g of freeze-  
136 dried soil and analysed as previously described (Zhao et al., 2015). The concentrations of each PLFA was  
137 calculated relative to that of the methyl nonadecanoate (19:0) internal standard. The PLFAs for the  
138 following groups were determined: bacterial biomass, sum of i15:0, a15:0, 15:0, i16:0, 16:1u7, i17:0,  
139 a17:0, 17:0, cy17:0, and cy19:0; actinomycete biomass, sum of 10 Me 16:0, 10 Me17:0, and 10 Me 18:0;  
140 and fungal biomass, 18:2 ω6 and 18:1 ω9. The total microbial biomass was represented by the sum of the  
141 bacterial, fungal, and actinomycete biomass values. The ratios of fungal to bacterial lipids (F/B) were  
142 used to evaluate the microbial community structure (Bossio and Scow, 1998; Wilkinson et al., 2002;  
143 Zhao et al., 2015). We calculated PLFA mass content first, PLFA (ng g<sup>-1</sup> dry soil) = (Response of PLFA  
144 / Response of 19:0 internal standard) × concentration of 19:0 internal standard × (volume of sample /  
145 mass of soil). Concentration of 19:0 internal standard: 5 μg ml<sup>-1</sup>, volume of sample: 200 μl, mass of soil:  
146 8g dry soil. And then we calculated PLFA molar mass concentration, PLFA (n mol g<sup>-1</sup> dry soil) = PLFA  
147 (ng g<sup>-1</sup> dry soil) / relative molecular mass.

## 148 2.5 Data processing

### 149 2.5.1 Litter decomposition rate

150 The percentage of litter dry weight loss was calculated as follows (Zhang et al., 2019):

$$151 L_t = \frac{M_0 - M_t}{M_0} \times 100\% (1)$$

152 where  $L_t$  is the percentage litter dry weight loss at time  $t$  (%),  $M_t$  is the litter dry matter weight at the time  
153  $t$  (g), and  $M_0$  is the initial dry matter weight (g).

154 The instantaneous litter dry mass decay rate ( $k$ ) was calculated based on the Olson negative exponential  
155 attenuation model and double exponential decay model (Olson, 1963; Berg, 2014):

156  $M_{t_n} = M_{t_{n-1}} e^{-k_n(t_n - t_{n-1})}$  (2)

157 where  $M_{t_n}$  is the litter dry matter weight at nth sampling (g),  $M_{t_{n-1}}$  is the litter dry matter weight at  
158 (n-1)th sampling (g),  $t_n - t_{n-1}$  is the time between the nth and (n-1)th sampling,  $k_n$  is the instantaneous  
159 decomposition rate at the nth sampling.

### 160 **2.5.2 Relative release index**

161 The relative release indices (RRIs) of C, N, and P from the plant litter were calculated as follows (Zhang  
162 et al., 2019):

163 
$$RRI_t = \frac{M_0 \times C_0 - M_t \times C_t}{M_0 \times C_0} \times 100\% \quad (3)$$

164 where  $C_t$  is the concentration of an element in the litter at time  $t$ ,  $C_0$  is the initial concentration of an  
165 element in the litter, and  $M_t$  is the litter dry matter weight at time  $t$  (g). CRRI, NRRI, PRRI, and LRRI  
166 represent the carbon, nitrogen, phosphorus, and lignin RRIs, respectively. A positive RRI indicates a net  
167 release of the element during litter decomposition whilst a negative RRI indicates a net accumulation of  
168 the element during litter decomposition.

### 169 **2.5.3 Contribution of litter-C input to the SOC pool**

170 The contribution of litter-C input to the SOC pool was calculated as follows (Lv and Wang, 2017):

171 
$$LC = \frac{SOC_L - SOC_S}{SOC_i} \times 100\% \quad (4)$$

172 where  $LC$  is the contribution of the litter-C input to SOC pool,  $SOC_L$  is the SOC concentration for the  
173 litter input treatment,  $SOC_S$  is the SOC concentration for the treatment without litter input, and  $SOC_i$  is  
174 the initial SOC content before the experimental treatments.

### 175 **2.6 Statistical analyses**

176 The percentage of litter dry weight losses and the instantaneous decomposition rates were compared  
177 among the three water levels by repeated ANOVA analyses. The water level was the main factor, and  
178 time was the repeated factor. The intrinsic litter decomposition rate-limiting factor was analysed by the  
179 stepwise regression method in a multiple regression model. The surface soil chemical components and  
180 the microbial community structure were compared by two-way ANOVA. Treatment (with or without  
181 litter input) and water level were the main factors. The percentage differences in litter dry weight loss,

182 the instantaneous decomposition rates, the soil chemical components, and the microbial community  
183 structure were evaluated by LSD at the 0.05 significance level. The data were expressed as means  $\pm$   
184 standard error. All statistical analyses were performed in SPSS 21 (IBM Corp., Armonk, NY, USA).

### 185 **3 Results**

#### 186 **3.1 litter decomposition process**

187 The percentage of litter dry weight loss was the highest for the +25 cm water level treatment through the  
188 entire litter decomposition period followed by the 0 cm water level treatment. The percentage of litter  
189 dry weight loss was the lowest for the -25 cm water level treatment ( $P < 0.01$ ; Fig. 2a). After 280 d  
190 decomposition, the percentage litter dry weight loss values under the +25 cm, 0 cm, and -25 cm water  
191 level treatments were 61.8%, 49.8% and 32.4%, respectively.

192 The instantaneous decomposition rate at each measurement time point was calculated based on the Olson  
193 negative exponential attenuation model and double exponential decay model. The instantaneous  
194 decomposition rate was highest at initial and slowly decreased and stabilised for all three water levels.  
195 The maximum decomposition rates for the -25 cm, 0 cm, and +25 cm water levels were  $0.00527 \text{ d}^{-1}$ ,  
196  $0.00908 \text{ d}^{-1}$ , and  $0.01307 \text{ d}^{-1}$ , respectively (Fig. 2b).

#### 197 **3.2 Intrinsic litter decomposition rate-limiting factor**

198 During the entire decomposition process, CRRI, NRRI, PRRI, and LRRI significantly increased with the  
199 water level. Litter carbon and lignin were always released at all three water levels whilst at -25 cm,  
200 nitrogen and phosphorus enrichment appeared in the middle stage (Fig. 3a–3d). At the start of the  
201 experiment, neither the C/N nor the lignin/N ratio significantly differed at the three water levels. At the  
202 middle stage, however, both the C/N and lignin/N ratios were significantly lower at the -25 cm water  
203 level than they were at the 0 cm and -25 cm water levels (Fig. 3e–3f).

204 The multiple regression model of the instantaneous litter decomposition rate and the litter properties  
205 showed that at the -25 cm water levels, the main decomposition rate-limiting factor was the lignin  
206 concentration whilst at the 0 cm and +25 cm water level, the main litter decomposition rate-limiting  
207 factor was the lignin/N ratio (Table 1).



### 208 3.3 Soil surface microbial community structure

209 Under both litter input and litter removal conditions, the bacterial, fungal, and microbial biomass levels  
210 were the highest under the 0 cm water level treatment; however, these parameters showed nonsignificant  
211 differences between +25 cm above and below water level treatments ( $P > 0.05$ ; Fig. 4a, 4b, and 4f). The  
212 actinomycete biomass was the highest under the -25 cm water level treatment, followed by that under the  
213 0 cm water level treatment. It was the lowest under the +25 cm water level treatment (Fig. 4c). Litter  
214 input significantly stimulated fungal and microbial biomass at all three water levels but only significantly  
215 stimulated bacterial and actinomycete biomass at the -25 cm water level ( $P < 0.05$ ; Fig. 4a–4c and 4e).  
216 Under litter input conditions, the fungal/bacteria ratio was the highest at the 0 cm water level, followed  
217 by the +25 cm water level. It was the lowest under the -25 cm water level treatment. Under litter removal  
218 conditions, however, the fungal/bacteria ratio was significantly higher under the -25 cm water level  
219 treatment than it was under the 0 cm and +25 cm water level treatments ( $P < 0.05$ ; Fig. 4d).

### 220 3.4 Contribution of leaf decomposition to the soil surface carbon pool

221 The SOC, MBC, and DOC concentrations were significantly affected by the water level. The SOC and  
222 MBC were the highest at the 0 cm water level and the lowest at the -25 cm water level ( $P < 0.01$ ; Fig. 5a  
223 and 5b). The DOC was the highest at the -25 cm water level and the lowest at the +25 cm water level ( $P$   
224  $< 0.01$ ; Fig. 5c).

225 Compared with the litter removal group, the SOC concentrations were significantly higher for the litter  
226 input group at the +25 cm and 0 cm water levels. Relative to the litter removal group, the DOC  
227 concentrations were significantly higher for the litter input group at the 0 cm- and -25 cm water levels  
228 ( $P < 0.001$ ; Fig. 5a and 5c). The contribution of the litter-C input to the S-SOCP was the highest for the  
229 +25 cm water level treatment (16.93%), intermediate for the 0 cm water level treatment (9.44%), and the  
230 lowest for the -25 cm water level treatment (2.51%) ( $P < 0.001$ ; Fig. 5d).

## 231 4 Discussion

### 232 4.1 Environmental control of litter decomposition

233 The water level significantly influenced *C. brevicuspis* leaf litter decomposition ( $P < 0.001$ ). The

234 instantaneous decomposition rates ( $k$ ) were the highest for the +25 cm water level treatment, intermediate  
235 for the 0 cm water level treatment, and the lowest for the -25 cm water level treatment (Fig. 2b). Hence,  
236 the percentage litter dry weight loss and the decomposition rate increased with the water level, which  
237 supported our first hypothesis. The wetland water level strongly affects litter leaching and microbial  
238 decomposition (Peltoniemi et al., 2012). Related research showed that the wetland water level strongly  
239 affects litter leaching and microbial decomposition (Peltoniemi et al., 2012). Molles et al. (1995) also  
240 found that compared with the terrestrial environment, in wetland, water promotes litter leaching and  
241 microbial metabolism, thereby accelerating litter decomposition. Moreover, water infiltration into litter  
242 also increases relative leaching loss (Molles et al., 1995). Here, the high litter decomposition rate  
243 measured for the +25 cm water level treatment may be explained primarily by litter leaching. This finding  
244 was consistent with results reported for *Carex cinerascens* litter decomposition in Poyang Lake (Zhang  
245 et al., 2019) and *Calamagrostis angustifolia* litter decomposition on the Sanjiang Plain (Sun et al., 2012).  
246 The high soil total microbial, bacterial and fungal biomass levels at the 0 cm water level could account  
247 for the rapid litter decomposition observed there. Certain microorganisms are vital to the decomposition  
248 process (Yarwood, 2018). Fungi are primary litter decomposers as they fragment dead plant tissues by  
249 breaking down lignin and cellulose. Bacteria are secondary decomposers that utilise the simpler  
250 compounds generated by fungal activity (de Boer et al., 2005; Bani et al., 2019). Microbial decomposers  
251 generally flourish in humid environments. At the 0 cm water level, microbial activity explains most of  
252 the litter decomposition. While at the -25 cm water level, there are comparatively few microbial  
253 decomposers, and decomposition is very slow.

#### 254 **4.2 Intrinsic factors controlling litter decomposition**

255 The instantaneous decomposition rate was highest at initial and slowly decreased and stabilised for all  
256 three water levels. (Fig. 2b). Water-soluble components and non-lignin carbohydrates are preferentially  
257 and quickly decomposed at the **initial** of decomposition (Davis et al., 2003). Here, a multiple regression  
258 model of the instantaneous litter decomposition rate and litter properties showed that the internal limiting  
259 factors affecting the rate of *C. brevicuspis* leaf litter decomposition varied with the water level. The lignin  
260 concentration determined the litter decomposition rate for the -25 cm water level treatment whilst the  
261 lignin/N ratio regulated the litter decomposition rate for the 0 cm and +25 cm water level treatment. This

262 discovery upheld our second hypothesis and was consistent with the findings of Zhang et al. who reported  
263 that wetland ecosystems decomposed *Carex cinerascens* lignin much earlier and faster than terrestrial  
264 ecosystems (Zhang et al., 2019). Here, we found that the lignin content was the major internal limiting  
265 factor of the *C. brevicuspis* leaf litter decomposition rate at -25 cm water level. At the 0 cm and +25 cm  
266 water level, N is rapidly lost, and the L/N ratio significantly increases. Thus, L/N is the main internal  
267 limiting factor at the 0 cm and +25 cm water levels. A few studies have shown that the lignin content is  
268 a key factor limiting terrestrial plant and hygrophyte litter decomposition (Yue et al., 2016; Zhang et al.,  
269 2018). Therefore, the amount of carbon that the litter can return to the ecosystem is closely associated  
270 with the plant lignin content. The lignin content of *C. brevicuspis* leaf litters is ~10% less than that of  
271 other wetland plants such as *Miscanthus sacchariflorus* (~30%) (Xie et al., 2016), *Spartina alterniflora*  
272 (~40%) (Yan et al., 2019), and terrestrial plants such as willow (~25%), larch (~38%), and cypress (~28%)  
273 (Yue et al., 2016), so the *C. brevicuspis* leaf litter is more easily leached and then contributes more to the  
274 SOC pool. Furthermore, in Dongting lake wetland, the *Carex* genus covers a large area (~23,950 hm<sup>2</sup>)  
275 and generates abundant litter (~36,547 t) (Kang et al., 2009). Thus, *C. brevicuspis* litter may potentially  
276 return large amounts of carbon to the soil.

### 277 **4.3 Contribution of leaf decomposition to the soil surface carbon pool**

278 Litter decomposition is the main pathway by which nutrients are transferred from the plants to the soil.  
279 Litter affects the SOC, the stabilisation of which affects other soil properties such as sorption, nutrient  
280 availability, pH, and water holding capacity (Brady and Weil, 2008). The results of this study showed  
281 that litter addition increases SOC in a manner that varies with the water level. The contribution of litter-  
282 C input to the S-SOCP was the highest under the +25 cm water level treatment (16.93%), intermediate  
283 under the 0 cm water level treatment (9.44%), and the lowest under the -25 cm water level treatment  
284 (2.51%). For this reason, flooding conditions are conducive to litter carbon input into the soil. These  
285 findings corroborated our third hypothesis. In addition, litter input had a similar effect on soil DOC at  
286 the 0 cm and -25 cm water levels. Therefore, litter decomposition contributes mainly soluble carbon to  
287 the soil (Zhou et al., 2015). However, this DOC is also readily lost and decomposed (Sokol and Bradford,  
288 2019; Gomez-Casanovas et al., 2020). This fact accounts for the significantly lower relative DOC under  
289 the +25 cm water level treatment here. Wetlands have comparatively larger but also more unstable S-

290 SOCPs than terrestrial environments. In wetlands, water level fluctuations could readily cause carbon  
291 loss (Gao et al., 2016; Chen et al., 2018). The SOC differences among three water levels were caused by  
292 different soil mineralization in different environments. Soil mineralization in aerobic environment (-25  
293 cm) was significantly higher than that in the flooded environment (0 cm, +25 cm) (Qiu et al., 2018), so  
294 the SOC at -25 cm water level was lower than the other two water levels. Nevertheless, we considered  
295 mainly aboveground litter in this experiment. Hence, the influence of underground litter (root)  
296 decomposition on the SOC pool should be investigated in future research (Sokol and Bradford, 2019;  
297 Lyu et al., 2019).

## 298 **5 Conclusion**

299 In this study, we quantified the contribution of leaf litter decomposition on soil surface organic carbon  
300 pools (S-SOCPs) under different water level conditions. Appropriate flooding (+25 cm water level  
301 treatment in our study) can significantly promote the decomposition of litter and contribute about 16.93%  
302 organic carbon to S-SOCPs. Under waterlogging condition (0 cm water level), litter decomposition,  
303 which mainly controlled by microbial activity, contributed 9.44% organic carbon to S-SOCP. However,  
304 under relative drought conditions (-25 cm water level treatment in our study), litter decomposition only  
305 contributes about 2.51% organic carbon to S-SOCP, which is largely ascribed to the slower  
306 decomposition rate and soil carbon lost by microbe metabolism (i.e., actinomycetes). We also found that  
307 lignin or lignin/N content were intrinsic factors controlling the litter decomposition rate in *Carex*  
308 *brevicuspis*. In Dongting Lake floodplain, the groundwater decline due to climate change and human  
309 disturbance would slow down the return rate of organic carbon from leaf litter to the soil, and facilitate  
310 the S-SOCP loss.

## 311 **Data availability**

312 The data used in this paper are stored in the open-access online database Figshare and can be accessed  
313 using the following link: <https://doi.org/10.6084/m9.figshare.12758387.v1> (Zhu et al. 2020).

## 314 **Conflict of interest**

315 The authors declare that they have no conflict of interest.

## 316 **Author contributions**

317 Lianlian Zhu designed experiments, collected samples, acquired, analysed, interpreted data, and wrote  
318 the manuscript. Zhengmiao Deng designed experiments, interpreted data, and revised the manuscript.  
319 Yonghong Xie designed experiments and revised the manuscript. Xu Li, Feng Li, Xincheng Chen and

320 Yeai Zou collected samples and revised the manuscript. Chengyi Zhang and Wei Wang interpreted data  
321 and revised the manuscript.

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### 332 **References**

333 Berg, B., and Mcclaugherty, C.: Plant litter: decomposition, humus formation, carbon sequestration,  
334 2003.

335 Berg, B.: Decomposition patterns for foliar litter - A theory for influencing factors, *Soil Biology &*  
336 *Biochemistry*, 78, 222-232, 10.1016/j.soilbio.2014.08.005, 2014.

337 Bossio, D. A., and Scow, K. M.: Impacts of carbon and flooding on soil microbial communities:  
338 Phospholipid fatty acid profiles and substrate utilization patterns, *Microb. Ecol.*, 35, 265-278,  
339 10.1007/s002489900082, 1998.

340 Bowden, R. D., Deem, L., Plante, A. F., Peltre, C., Nadelhoffer, K., and Lajtha, K.: Litter Input  
341 Controls on Soil Carbon in a Temperate Deciduous Forest, *Soil Science Society of America Journal*,  
342 78, S66-S75, 10.2136/sssaj2013.09.0413nafsc, 2014.

343 Cao, J. B., He, X. X., Chen, Y. Q., Chen, Y. P., Zhang, Y. J., Yu, S. Q., Zhou, L. X., Liu, Z. F., Zhang, C.  
344 L., and Fu, S. L.: Leaf litter contributes more to soil organic carbon than fine roots in two 10-year-old  
345 subtropical plantations, *Science of the Total Environment*, 704, 8, 10.1016/j.scitotenv.2019.135341,  
346 2020.

347 Chen, H. Y., Zou, J. Y., Cui, J., Nie, M., and Fang, C. M.: Wetland drying increases the temperature  
348 sensitivity of soil respiration, *Soil Biology & Biochemistry*, 120, 24-27, 10.1016/j.soilbio.2018.01.035,  
349 2018.

350 Chen, X. S., Deng, Z. M., Xie, Y. H., Li, F., Hou, Z. Y., and Wu, C.: Consequences of Repeated  
351 Defoliation on Belowground Bud Banks of *Carex brevicuspis* (Cyperaceae) in the Dongting Lake  
352 Wetlands, China, *Frontiers in Plant Science*, 7, 10.3389/fpls.2016.01119, 2016.

353 Deng, Z. M., Li, Y. Z., Xie, Y. H., Peng, C. H., Chen, X. S., Li, F., Ren, Y. J., Pan, B. H., and Zhang, C.  
354 Y.: Hydrologic and Edaphic Controls on Soil Carbon Emission in Dongting Lake Floodplain, China, *J.*  
355 *Geophys. Res.-Biogeosci.*, 123, 3088-3097, 10.1029/2018jg004515, 2018.

356 Gao, J. Q., Feng, J., Zhang, X. W., Yu, F. H., Xu, X. L., and Kuzyakov, Y.: Drying-rewetting cycles  
357 alter carbon and nitrogen mineralization in litter-amended alpine wetland soil, *Catena*, 145, 285-290,

358 10.1016/j.catena.2016.06.026, 2016.

359 Graça, M. A. S., Bärlocher, F., and Gessner, M. O.: Methods to Study Litter Decomposition, Springer  
360 Netherlands, 2005.

361 Hoyos-Santillan, J., Lomax, B. H., Large, D., Turner, B. L., Boom, A., Lopez, O. R., and Sjogersten, S.:  
362 Getting to the root of the problem: litter decomposition and peat formation in lowland Neotropical  
363 peatlands, *Biogeochemistry*, 126, 115-129, 10.1007/s10533-015-0147-7, 2015.

364 Hu, J. Y., Xie, Y. H., Tang, Y., Li, F., and Zou, Y. A.: Changes of Vegetation Distribution in the East  
365 Dongting Lake After the Operation of the Three Gorges Dam, China, *Frontiers in Plant Science*, 9, 9,  
366 10.3389/fpls.2018.00582, 2018.

367 Kang, W. X., Tian, H., Jie-Nan, H. E., Hong-Zheng, X. I., Cui, S. S., and Yan-Ping, H. U.: Carbon  
368 Storage of the Wetland Vegetation Ecosystem and Its Distribution in Dongting Lake, *Journal of Soil &  
369 Water Conservation*, 2009.

370 Kayranli, B., Scholz, M., Mustafa, A., and Hedmark, A.: Carbon Storage and Fluxes within Freshwater  
371 Wetlands: a Critical Review, *Wetlands*, 30, 111-124, 10.1007/s13157-009-0003-4, 2010.

372 Kochy, M., Hiederer, R., and Freibauer, A.: Global distribution of soil organic carbon - Part 1: Masses  
373 and frequency distributions of SOC stocks for the tropics, permafrost regions, wetlands, and the world,  
374 *Soil*, 1, 351-365, 10.5194/soil-1-351-2015, 2015.

375 Liu, S. L., Jiang, Z. J., Deng, Y. Q., Wu, Y. C., Zhao, C. Y., Zhang, J. P., Shen, Y., and Huang, X. P.:  
376 Effects of seagrass leaf litter decomposition on sediment organic carbon composition and the key  
377 transformation processes, *Sci. China-Earth Sci.*, 60, 2108-2117, 10.1007/s11430-017-9147-4, 2017.

378 Lv, F. C., and Wang, X. D.: Contribution of Litters to Soil Respiration : A Review, *Soils*, 49, 225-231,  
379 2017.

380 Moriyama, A., Yonemura, S., Kawashima, S., Du, M. Y., and Tang, Y. H.: Environmental indicators for  
381 estimating the potential soil respiration rate in alpine zone, *Ecol. Indic.*, 32, 245-252,  
382 10.1016/j.ecolind.2013.03.032, 2013.

383 **Olson, J. S.: Energy- storage and balabce of producers and decomposers in ecological- systems,**  
384 ***Ecology*, 44, 322-&, 10.2307/1932179, 1963.**

385 **Peng, P. Q., Zhang, W. J., Tong, C. L., Qiu, S. J., and Zhang, W. C.: Soil C, N and P contents and their**  
386 **relationships with soil physical properties in wetlands of Dongting Lake floodplain, *The journal of***  
387 ***applied ecology*, 16, 1872-1878, 2005.**

388 Pinto, O. B., Vourlitis, G. L., Carneiro, E. M. D., Dias, M. D., Hentz, C., and Nogueira, J. D.:  
389 Interactions between Vegetation, Hydrology, and Litter Inputs on Decomposition and Soil CO<sub>2</sub> Efflux  
390 of Tropical Forests in the Brazilian Pantanal, *Forests*, 9, 17, 10.3390/f9050281, 2018.

391 Qiu, H. S., Ge, T. D., Liu, J. Y., Chen, X. B., Hu, Y. J., Wu, J. S., Su, Y. R., and Kuzyakov, Y.: Effects of  
392 biotic and abiotic factors on soil organic matter mineralization: Experiments and struc tural modeling  
393 analysis, *Eur. J. Soil Biol.*, 84, 27 34, 10.1016/j.ejsobi.2017.12.003, 2018.

394 Song, Y. Y., Song, C. C., Tao, B. X., Wang, J. Y., Zhu, X. Y., and Wang, X. W.: Short-term responses of  
395 soil enzyme activities and carbon mineralization to added nitrogen and litter in a freshwater marsh of  
396 Northeast China, *Eur. J. Soil Biol.*, 61, 72-79, 10.1016/j.ejsobi.2014.02.001, 2014.

397 Sun, X. L., Kong, F. L., Li, Y., Di, L. Y., and Xi, M.: Effects of litter decomposition on contents and  
398 three-dimensional fluorecence spectroscopy characteristics of soil labile organic carbon in coastal  
399 wetlands of Jiaozhou Bay, China, *The journal of applied ecology*, 30, 563-572, 10.13287/j.1001-  
400 9332.201902.036, 2019.

401 Tong, C., Cadillo-Quiroz, H., Zeng, Z. H., She, C. X., Yang, P., and Huang, J. F.: Changes of

402 community structure and abundance of methanogens in soils along a freshwater-brackish water  
403 gradient in subtropical estuarine marshes, *Geoderma*, 299, 101-110, 10.1016/j.geoderma.2017.03.026,  
404 2017.

405 Van de Moortel, A. M. K., Du Laing, G., De Pauw, N., and Tack, F. M. G.: The role of the litter  
406 compartment in a constructed floating wetland, *Ecol. Eng.*, 39, 71-80, 10.1016/j.ecoleng.2011.11.003,  
407 2012.

408 Wang, X. L., Xu, L. G., and Wan, R. R.: Comparison on soil organic carbon within two typical wetland  
409 areas along the vegetation gradient of Poyang Lake, China, *Hydrol. Res.*, 47, 261-277,  
410 10.2166/nh.2016.218, 2016.

411 Whiting, G. J., and Chanton, J. P.: Greenhouse carbon balance of wetlands: methane emission versus  
412 carbon sequestration, *Tellus Ser. B-Chem. Phys. Meteorol.*, 53, 521-528, 10.1034/j.1600-  
413 0889.2001.530501.x, 2001.

414 Wilkinson, S. C., Anderson, J. M., Scardelis, S. P., Tisiafouli, M., Taylor, A., and Wolters, V.: PLFA  
415 profiles of microbial communities in decomposing conifer litters subject to moisture stress, *Soil*  
416 *Biology & Biochemistry*, 34, 189-200, 2002.

417 Xie, Y., Xie, Y., Chen, X., Li, F., Hou, Z., and Li, X.: Non-additive effects of water availability and  
418 litter quality on decomposition of litter mixtures, *Journal of Freshwater Ecology*, 31, 153-168,  
419 10.1080/02705060.2015.1079559, 2016a.

420 Xie, Y. J., Xie, Y. H., Hu, C., Chen, X. S., and Li, F.: Interaction between litter quality and simulated  
421 water depths on decomposition of two emergent macrophytes, *J. Limnol.*, 75, 36-43,  
422 10.4081/jlimnol.2015.1119, 2016b.

423 Xie, Y. J., Xie, Y. H., Xiao, H. Y., Chen, X. S., and Li, F.: Controls on Litter Decomposition of  
424 Emergent Macrophyte in Dongting Lake Wetlands, *Ecosystems*, 20, 1383-1389, 10.1007/s10021-017-  
425 0119-y, 2017.

426 Yan, J. F., Wang, L., Hu, Y., Tsang, Y. F., Zhang, Y. N., Wu, J. H., Fu, X. H., and Sun, Y.: Plant litter  
427 composition selects different soil microbial structures and in turn drives different litter decomposition  
428 pattern and soil carbon sequestration capability, *Geoderma*, 319, 194-203,  
429 10.1016/j.geoderma.2018.01.009, 2018.

430 Yu, X. F., Ding, S. S., Lin, Q. X., Wang, G. P., Wang, C. L., Zheng, S. J., and Zou, Y. C.: Wetland plant  
431 litter decomposition occurring during the freeze season under disparate flooded conditions, *Science of*  
432 *the Total Environment*, 706, 9, 10.1016/j.scitotenv.2019.136091, 2020.

433 Zhang, L., Zhou, G. S., Ji, Y. H., and Bai, Y. F.: Grassland Carbon Budget and Its Driving Factors of the  
434 Subtropical and Tropical Monsoon Region in China During 1961 to 2013, *Scientific Reports*, 7, 11,  
435 10.1038/s41598-017-15296-7, 2017.

436 Zhang, Q. J., Zhang, G. S., Yu, X. B., Liu, Y., Xia, S. X., Ya, L., Hu, B. H., and Wan, S. X.: Effect of  
437 ground water level on the release of carbon, nitrogen and phosphorus during decomposition of *Carex*  
438 *cinerascens* Kukenth in the typical seasonal floodplain in dry season, *Journal of Freshwater Ecology*,  
439 34, 305-322, 10.1080/02705060.2019.1584128, 2019.

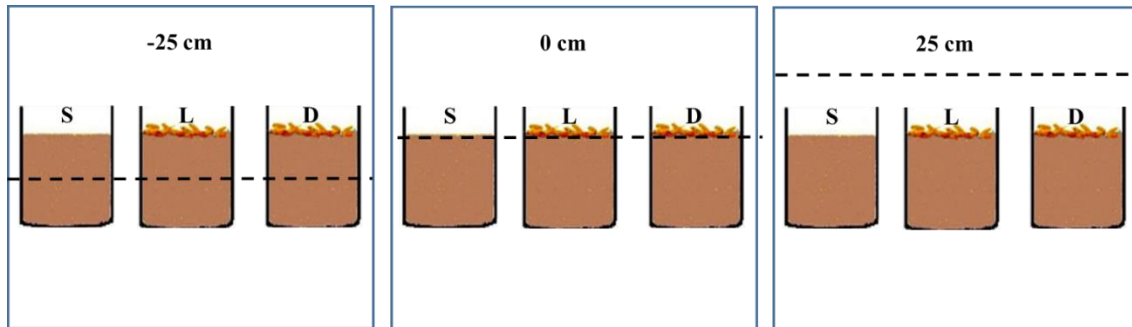
440 Zhao, J., Zeng, Z. X., He, X. Y., Chen, H. S., and Wang, K. L.: Effects of monoculture and mixed  
441 culture of grass and legume forage species on soil microbial community structure under different levels  
442 of nitrogen fertilization, *Eur. J. Soil Biol.*, 68, 61-68, 10.1016/j.ejsobi.2015.03.008, 2015.

443

444 **Table 1: Multiple regression model of instantaneous litter decomposition rate and litter**  
 445 **properties**

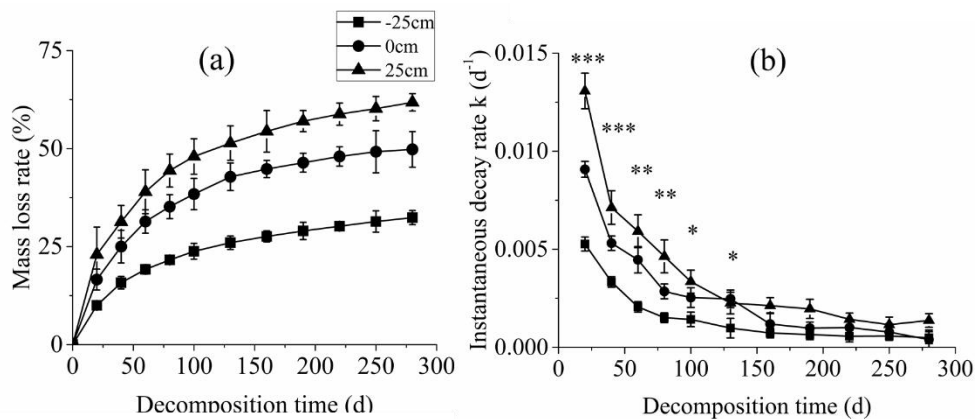
Water level (cm)	Multiple regression model	<i>F</i>	<i>R</i> <sup>2</sup>	<i>P</i>
-25	$R = -0.715L - 0.443C + 0.033$	5.738	0.727	0.006
0	$R = -0.928LN - 0.233CN + 0.023$	5.928	0.927	< 0.001
+25	$R = -0.717LN + 0.016$	9.543	0.793	0.002

446 where *R* is the litter instantaneous decomposition rate, *L* is the lignin concentration, *CN* is the carbon-to-  
 447 nitrogen ratio (*C/N*, g g<sup>-1</sup>), and *LN* is the lignin-to-nitrogen ratio (lignin/*N*, g g<sup>-1</sup>). All indicators used to analyse  
 448 the model was referred to the content at each time point.



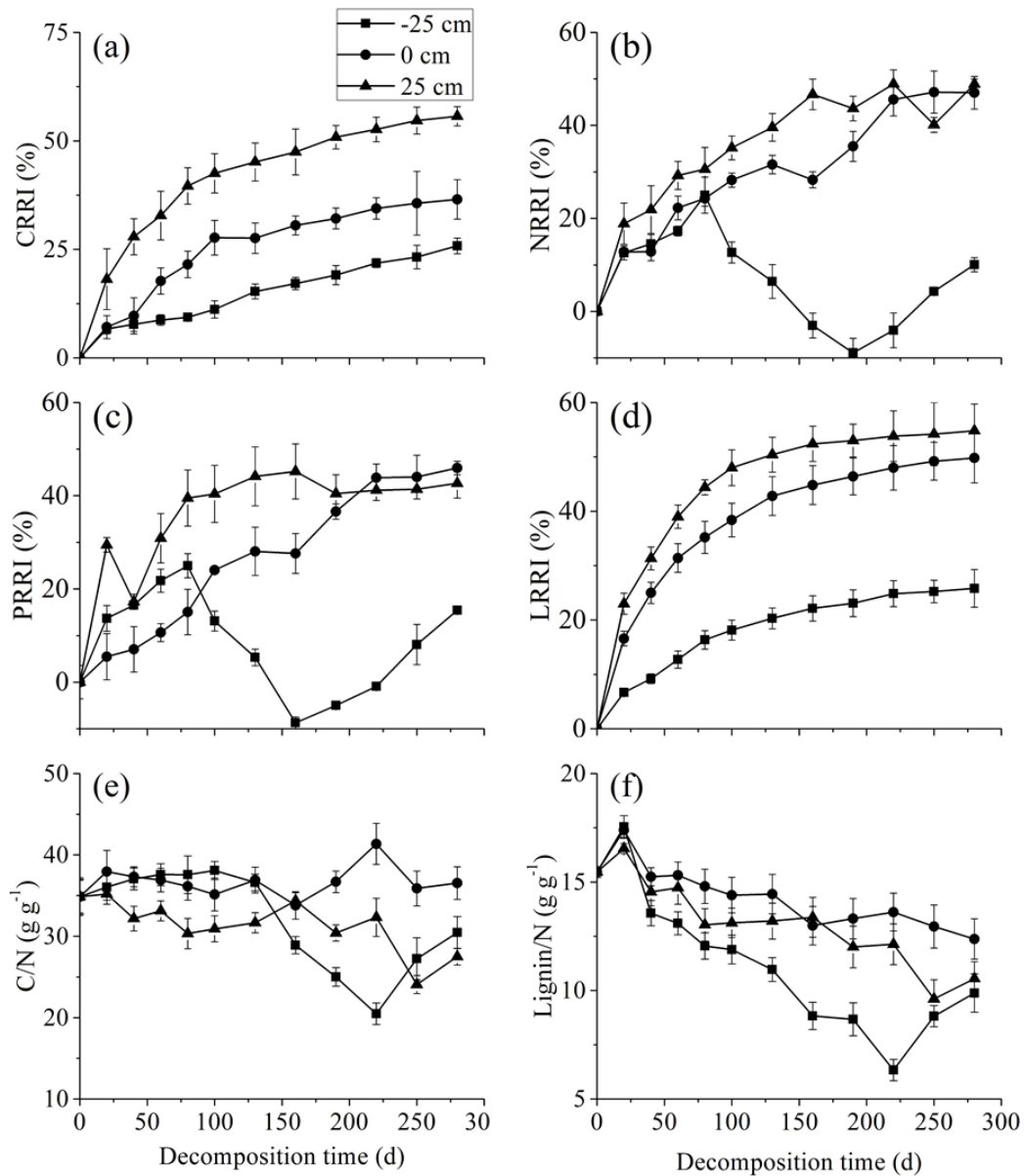
449  
 450 **Figure 1: Schematic diagram of the experimental setup. The dotted line represents the water level.**  
 451 **L** represents litter which was distributed on the soil surface in 15 litter bags to observe the effects of leaf litter  
 452 input on soil carbon pool; **S** represents soil which was designated the litter removal control; **D** represents  
 453 decomposition which was distributed on the soil surface in 15 litter bags to monitor the litter decomposition  
 454 rate and process.

455



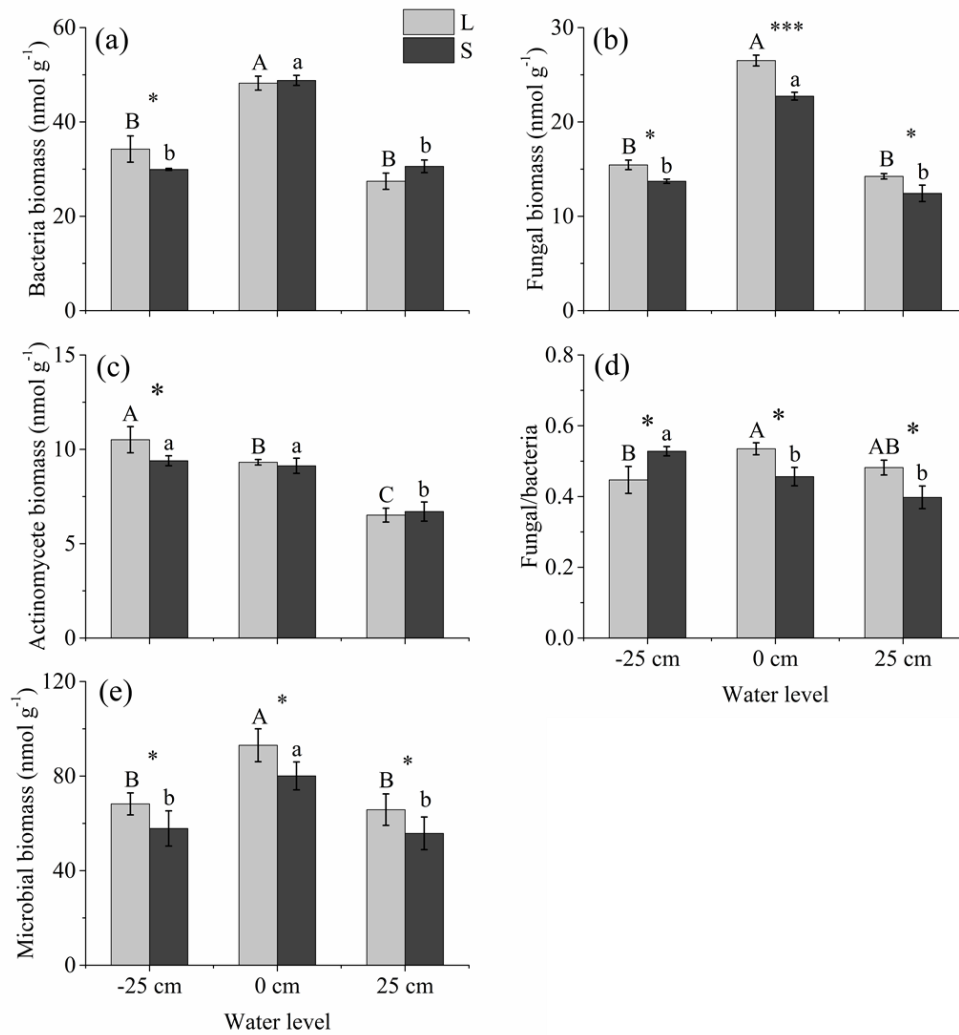
456  
 457 **Figure 2: Percentage litter dry weight loss and decomposition rate during *C. brevicuspis* decomposition at**  
 458 **three water levels (-25 cm, 0 cm, and +25 cm). \*, \*\*, and \*\*\* represent significant differences of the litter**  
 459 **instantaneous decay rate among the three water levels at the 0.05, 0.01, and 0.001 significance levels,**  
 460 **respectively.**





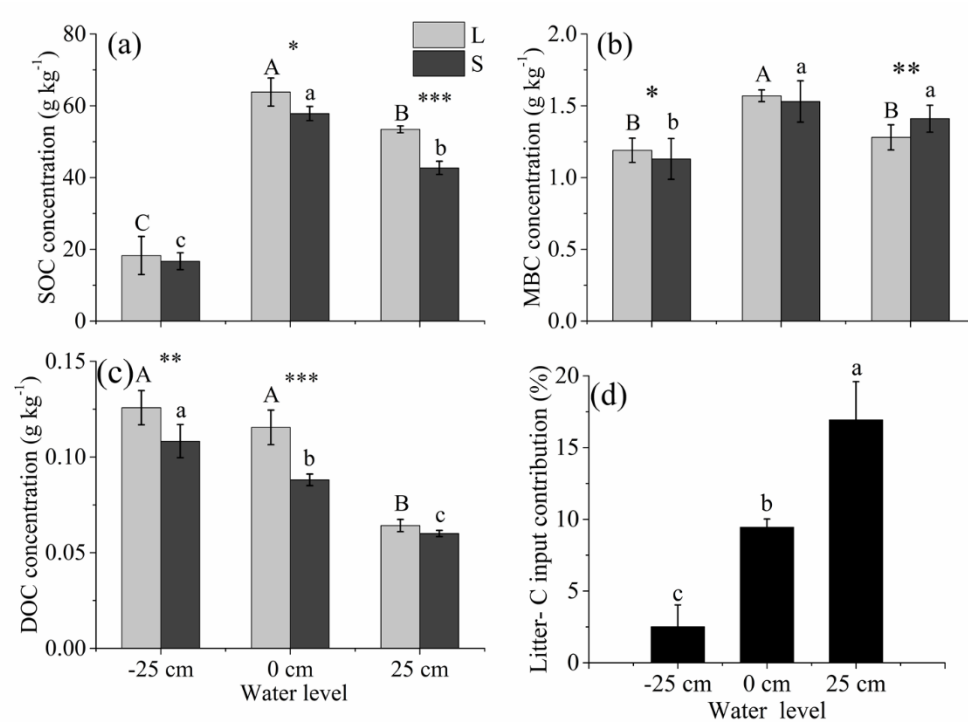
461

462 **Figure 3: Percentage (mean ± SE) of carbon relative release index (CRRRI), nitrogen relative release index**  
 463 **(NRRRI), phosphorus relative release index (PRRI), lignin relative release index (LRRRI), C/N ratio, and**  
 464 **lignin/N ratio at three water levels (-25 cm, 0 cm, and +25 cm).**



465

466 **Figure 4: Microbial community structure under litter input and litter removal at three water levels. Different**  
 467 **uppercase letters among vertical bars indicate significant differences among the three water levels in the litter**  
 468 **input (L) group. Different lowercase letters indicate significant differences among the three water levels in**  
 469 **the litter removal (S) group. The significance level is  $\alpha = 0.05$ . \*, \*\*, and \*\*\* represent significant differences**  
 470 **between the litter input (L) and litter removal (S) groups at the 0.05, 0.01, and 0.001**  
 471 **significance levels, respectively.**



472

473 **Figure 5: Concentrations of SOC (a), MBC (b), DOC (c) between the litter input (L) and litter removal (S)**  
 474 **groups and the litter-C input contribution (d) under three water levels at the end of the experiment. Different**  
 475 **uppercase letters among vertical bars indicate significant differences among the three water levels in the**  
 476 **litter input (L) group. Different lowercase letters indicate significant differences among the three water levels in**  
 477 **the litter removal (S) group. The significance level is  $\alpha = 0.05$ . \*, \*\*, and \*\*\* represent significant differences**  
 478 **between the litter input (L) and litter removal (S) groups at the 0.05, 0.01, and 0.001**  
 479 **significance levels, respectively.**