Seasonal dynamics of the COS and CO₂ exchange of a managed temperate grassland

³ Felix Maximilian Spielmann¹, Albin Hammerle¹, Florian Kitz¹, Katharina Gerdel¹, Georg Wohlfahrt¹

⁴ ¹Department of Ecology, University of Innsbruck, Innsbruck, 6020, Austria

5 Correspondence to: Georg Wohlfahrt (Georg.Wohlfahrt@uibk.ac.at)

Abstract. Gross primary productivity (GPP), the CO_2 uptake by means of photosynthesis, cannot be measured directly on 6 7 ecosystem scale, but has to be inferred from proxies or models. One newly emerged proxy is the trace gas carbonyl sulfide 8 (COS). COS diffuses into plant leaves in a fashion very similar to CO₂, but is generally not emitted by plants. Laboratory 9 studies on leaf level gas exchange have shown promising correlations between the leaf relative uptake (LRU) of COS to CO₂ 10 under controlled conditions. However, in situ measurements including daily to seasonal environmental changes are required, 11 to test the applicability of COS as a tracer for GPP at larger temporal scales. To this end, we conducted concurrent 12 ecosystem scale CO2 and COS flux measurements above an agriculturally managed temperate mountain grassland. We also 13 determined the magnitude and variability of the soil COS exchange, which can affect the LRU on ecosystem level. The 14 cutting and removal of the grass at the site had a major influence on the soil flux as well as the total exchange of COS. The 15 grassland acted as a major sink for CO_2 and COS during periods of high leaf area. The sink strength decreased after the cuts 16 and the grassland turned into a net source for CO_2 and COS on ecosystem level. The soil acted as a small sink for COS when 17 the canopy was undisturbed, but also turned into a source after the cuts, which we linked to higher incident radiation hitting 18 the soil surface. However, the soil contribution was not large enough to explain the COS emission on ecosystem level, 19 hinting to an unknown COS source possibly related to dead plant matter degradation. Over the course of the season, we 20 observed a concurrent decrease of CO₂ and COS uptake on ecosystem level. With the exception of the short periods after the 21 cuts, the LRU under high light conditions was rather stable and indicates a high correlation between the COS flux and GPP 22 across the growing season.

23

24 1 Introduction

25 Carbonyl sulfide (COS) is the most abundant sulfur-containing gas in the atmosphere with tropospheric mole fractions of 26 ~500 ppt. Within the atmosphere, COS acts as a greenhouse gas with a 724 times higher direct radiative forcing efficiency as 27 CO₂ (Brühl et al., 2012). After reaching the stratosphere, it reacts to sulfur aerosols via oxidation and photolysis, hence 28 contributing to the backscattering of solar radiation and having a cooling effect on Earth's atmosphere (Krysztofiak et al., 2015; Whelan et al., 2018). The intra-seasonal atmospheric COS mole fraction follows the pattern of CO2 as terrestrial 29 30 vegetation acts as the largest known sink for both species (Montzka et al., 2007; Whelan et al., 2018; Le Quere et al., 2018). 31 However, the relative decrease in ambient mole fraction during summer of the northern hemisphere is 6 times stronger for COS than for CO2, (Montzka et al., 2007) as COS is generally not emitted by plants like CO2, which is released in respiration 32 33 processes.

34 The uptake of COS by plants is mostly mediated by the enzyme carbonic anhydrase (CA), but also photolytic enzymes like

35 Ribulose-1,5-bisphosphate-carboxylase/-oxygenase (Rubisco) (Lorimer and Pierce, 1989). This in turn means that COS and

- 36 CO₂ share a similar pathway into leaves through the boundary layer, the stomata and the cytosol, up to their reaction sites.
- 37 Compared to CO₂, COS is processed in a one-way reaction to H₂S and CO₂ (Protoschill-Krebs and Kesselmeier, 1992;Notni
- 38 et al., 2007) and therefore not released by plants, with the exception of severely stressed plants (Bloem et al., 2012;Gimeno

39 et al., 2017). That makes COS an interesting tracer for estimating the stomatal conductance and the gross uptake of CO₂,

40 referred to as gross primary production (GPP), on ecosystem level (Asaf et al., 2013;Kooijmans et al., 2017;Kooijmans et al.,

41 2019). However, to estimate GPP using COS, the relative uptake of COS to GPP deposition velocities (LRU) must be known

43
$$LRU = \frac{\frac{F_{COS}}{\chi_{COS}}}{\frac{F_{CO2}}{\chi_{CO2}}}$$
 (Eq.1)

beforehand (see Eq.1), so that GPP can be estimated on the basis of the COS flux.

42

 F_{COS} is the COS leaf flux (pmol m⁻² s⁻¹), F_{CO2} is the gross CO₂ uptake on leaf level (µmol m⁻² s⁻¹) and χ_{CO2} are the 44 45 ambient COS and CO₂ mole fractions in ppt and ppm, respectively. Leaf level studies for C_3 plants have estimated the LRU 46 to be around 1.7 with the 95% confidence interval between 0.7 and 6.2 (Whelan et al., 2018;Seibt et al., 2010;Sandoval-Soto 47 et al., 2005). The large spread of the LRU most likely originates from differences between plant species, for example, leaf 48 structure and plant metabolism (Wohlfahrt et al., 2012; Seibt et al., 2010), which questions the applicability of the concept of LRU in real-world ecosystems under naturally varying environmental conditions. It is also known that the LRU is just stable 49 under high light conditions, since the uptake of CO_2 by means of photosynthesis is a light driven process, while CA is able to 50 51 process COS independently of light conditions (Maseyk et al., 2014; Yang et al., 2018; Stimler et al., 2011). Any model of LRU should therefore reflect diurnal changes in light conditions. Kooijmans et al. (2019) recently discovered that the vapor 52 53 pressure deficit (VPD) appears to have a stronger control on F_{COS} than on F_{CO2}, in an evergreen needle forest. If generally 54 true, this would add further variability to the LRU and complicating the application of COS to estimate GPP. Besides inter-55 specific differences in LRU, the question remains unanswered if the LRU is also susceptible to seasonal changes of ecosystems for example, changes in species composition or phenology, which would further complicate the application of 56 57 COS in carbon cycle research. Maseyk et al. (2014) observed COS emissions on ecosystem scale over a winter wheat field 58 going into senescence, indicating that potentially strong sources of COS could distort LRU.

59 Since CA and other enzymes known to emit or take up COS are also present in microorganisms (Ogawa et al., 2013;Seefeldt 60 et al., 1995;Ensign, 1995;Smeulders et al., 2013;Whelan et al., 2018), recent studies have also quantified the contribution of 61 soils to the COS ecosystem flux (Kooijmans et al., 2017; Spielmann et al., 2019; Maseyk et al., 2014). COS soil fluxes could 62 modify the LRU on ecosystem level and hence inferred GPP, if they are substantial compared to COS canopy fluxes. Similar 63 to the ecosystem fluxes, the soil fluxes could not only be prone to diurnal, but also seasonal changes, depending on the 64 substrate availability, environmental conditions (e.g. soil temperature and moisture) (Liu et al., 2010), substrate quality and quantity, and changes in composition of the microbial communities (Kitz et al., 2019;Meredith et al., 2019). Recent studies 65 66 have also linked COS soil emissions to abiotic processes dependent on light and/or temperature (Whelan and Rhew, 67 2015;Kitz et al., 2019;Meredith et al., 2018).

The goal of our study was to provide new insights into the seasonal variability of COS fluxes on ecosystem, soil and canopy 68 69 level. To this end, we conducted a 6-month campaign on a managed temperate mountain grassland, measuring ecosystem as 70 well as soil COS fluxes. Since the grassland was cut four times during the campaign, we were able to observe multiple 71 growing cycles and investigate the diel and seasonal changes of the COS fluxes and the LRU in this highly dynamic 72 ecosystem. We hypothesize that (H1) the grassland, given its large CO_2 uptake capacity (Wohlfahrt et al. 2008), is a major 73 sink for COS and that the sink strength decreases over the course of the season, (H2) the drying of the cut grass leads to a release of COS, (H3) the LRU will change after the cuts, due to stressed plants and drying plant parts in the field, but is 74 75 otherwise stable, (H4) the cuts turn the soil into a COS source, due to the larger amount of light reaching the soil surface 76 (Kitz et al., 2017), but once a reasonably high leaf area index (LAI) has developed, COS is taken up by soil.

77 2 Methods

78 2.1 Study site and period

79 The study was conducted at an intensively managed mountain grassland in the municipal territory of Neustift (Austria) in 80 Stubai valley (FLUXNET ID: AT-Neu; doi: 10.18140/FLX/1440121). The grassland is situated at an elevation of 970 m a.s.l. in the middle of the flat valley bottom. The soil was classified as Fluvisol with an estimated depth of 1 m with the majority 81 82 of roots located within the first 10 cm. Measurements were conducted between June 01, 2015 and October 31, 2015 (183 83 days). The vegetation was described as Pastincao-Arrhenatheretum and consisted mainly of Dactylis glomerata, Festuca pratensis, Alopecurus pratensis, Trisetum flavescens, Ranunculus acris, Taraxacum offcinale, Trifolium repens, Trifolium 84 pratense, and Carum carvi (Kitz et al., 2017). During the campaign, the grassland was cut four times (June 02/ July 07/ 85 86 August 21/ October 01, 2015) and the biomass left to dry on the field for up to one day, before being removed as silage. 87 Each year, the field site was fertilized with solid manure and cattle slurry (Hörtnagl et al., 2018) at the end of the season 88 (October 07, 2015).

89 2.2 Leaf area index

90 The LAI was estimated from assessments of the average canopy height, which were related to destructive LAI 91 measurements, using the following sigmoid function:

92
$$LAI = \frac{1}{(1 + \exp(-(a_1DOY + a_2)))(b_1 - b_2)}$$
 (Eq.2)

93 where DOY is the day of the year and a1, a2, b1 and b2 are factors that were optimized for each growing period, for 94 example, before the first cut, between cuts and after fourth cut (Wohlfahrt et al., 2008). Additionally, biomass samples were 95 taken at 15 occasions, to assist with the LAI calculation.

96 2.3 Mole fraction measurements

97 The CO₂ (χ_{CO2}) and COS (χ_{COS}) mole fractions were measured using a Quantum Cascade Laser (QCL) Mini Monitor 98 (Aerodyne Research, Billerica, MA, USA) at a wavenumber of ca. 2056 cm⁻¹ and at a frequency of 10 Hz. To minimize the 99 effect of air temperature (T_{air}) changes on the instrument, we placed it in an insulated box which in turn was located in a 100 climate controlled instrument hut (30°C). The cooling of the laser was achieved by a chiller (ThermoCube 400, Solid State 101 Cooling Systems, Wappinger Falls, NY, USA).

102 We used ¼ inch Teflon[™] tubing, stainless steel fittings (SWAGELOK, Solon, OH, USA and FITOK, Offenbach, HE, Germany), Teflon Filters (Savilex, EdenPrarie, MN, USA) as well as COS-inert valves (Parker-Hannafin, Cleveland, OH, 103 104 USA) to ensure that only materials known not to interact with COS were used for the measurement and calibration airflow. 105 Since the data of the QCL and the sonic anemometer were saved on two separate PCs, a network time protocol software 106 (NTP, Meinberg, NI, Germany) was used to keep the time on both devices synchronized. We corrected known χ_{COS} drift issues of the QCL (Kooijmans et al., 2016) by doing half hourly calibrations for 1 min with a gas of known χ_{COS} . The gas 107 cylinders (working standards) used for the calibrations were either pressurized air (UN 1002) or nitrogen (UN 1066), which 108 109 were cross-compared (when working standard cylinders were full and close to empty) to an Aculife-treated aluminum 110 pressurized air cylinder obtained from the National Oceanic & Atmospheric Administration (NOAA). The latter was 111 analyzed by the central calibration laboratory of NOAA for its χ_{COS} using gas chromatography with mass spectrometric detection (GC-MS) on April 06, 2015. We then linearly interpolated between the offsets of the half hourly calibrations and 112 113 used the retrieved values to correct the high frequency COS data. Due to issues with the scale gas cylinder, no absolute concentrations were available before June 16, 2015. To increase the amount of available data for the first post cut period, we 114

- extrapolated the COS mole fractions to the day of the 1st cut. This was done on the basis of the measured CO₂ mole fractions 115
- and the mean half hourly ratio of the ambient CO₂ to COS mole fractions retrieved between June 16 and June 18, 2015. 116

117 2.3.1 Mole fraction measurements within the canopy

In order to investigate the χ_{COS} within the canopy, we used a multiplexer and 8 ¼ inch TeflonTM tubes to measure the χ_{COS} at 118 119 8 heights within and above the canopy i.e. at 2, 5, 10, 20, 30, 40, 50 & 250 cm height above ground with a tube length of 120 15 m for each height. The upper two intakes were located at the eddy covariance measurement and canopy height, respectively. Each height was measured for 1 min at 1 Hz and 2 l min⁻¹, while the other lines were each flushed at 2 l min⁻¹. 121 The χ_{COS} drift was also corrected by doing half hourly calibrations (see section 2.3). 122

123

2.4 COS soil fluxes 124

125 2.4.1 Soil chamber setup

126 To quantify soil COS fluxes, we installed four stainless steel (SAE grade: 316L) rings 5 cm into the soil. They remained on site for 112 days (June 10, 2015 - September 30, 2015). Two additional rings were installed on August 31, 2015 and the 127 128 October 02, 2015 to examine any long-term effects of the ring placement and to replace the original rings for the 129 measurements in September and October. The aboveground biomass within each ring was removed at the day of installation 130 and again at least one day prior to each measurement day. The roots within as well as the vegetation surrounding the rings were not removed and natural litter was left in place. At days without measurements the soil within the rings was covered by 131 132 fleece to prevent it from drying out.

To measure the soil fluxes, a transparent fused silica-glass chamber with a volume of approximately 4155 cm³ (Kitz et al., 133 134 2017) was placed into the water filled channel of the steel rings, while air was sucked through the chamber to the QCL at a 135 flow rate of 1.5 l min⁻¹. The chamber χ_{COS} was then compared with the ambient χ_{COS} above the chamber, using a second inlet to which we switched before the chamber measurement and after reaching stable readings inside the chamber. The intake 136 137 height of the ambient as well as the inlet of the chamber air were located at 0.12 m above the ground and thus within the 138 canopy height with the exception of measurements right after the cuts (see cutting dates in Section 2.1). Overall, 243 139 chamber measurements were conducted over the course of the campaign including day and nighttime measurements. Additional manual measurements included a hand-held sensor (WET-2, Delta-T Devices, Cambridge, England) to measure 140 soil water content (SWC) and soil temperature (T_{soil}) at a soil depth of 5 cm simultaneously with the soil chamber 141 measurements next to the rings. 142

143 2.4.1 COS soil flux calculation

144 The COS soil flux was calculated using the following equation:

145
$$F = \frac{q(\chi_{cos2} - \chi_{cos1})}{A}$$
 (Eq.3)

where F is the COS soil flux (pmol m⁻² s⁻¹), q denotes the flowrate in (mol s⁻¹), χ_{COS2} and χ_{COS1} are the chamber and ambient 146 χ_{cos} in ppt, respectively and A the soil surface area (0.032 m²) covered by the chamber. A more detailed description can be

148 found in Kitz et al. (2017).

147

2.4.2 COS soil exchange modelling 149

Due to the removal of the aboveground biomass and the consequent higher shortwave radiation reaching the soil surface in 150

151 the chambers, compared to the soil below the canopy, we simulated the soil COS exchange for natural conditions. The soil 152 flux was modelled using our measured soil fluxes and additionally retrieved soil and meteorological data - T_{soil}, soil water

- 153 content (SWC) at 5 cm depth next to the chambers and incident shortwave radiation reaching the soil surface (R_{SW-soil}) as
- 154 input for a random forest regression model (Liaw and Wiener, 2002). The soil fluxes were modelled on half hourly basis for
- 155 the whole duration of the measurement campaign to calculate the COS canopy fluxes from the difference of the COS
- 156 ecosystem and soil fluxes. To this end we used the scikit-learn (sklearn Ver. 0.19.1) package, the pandas library and the
- 157 Python Software Distribution Anaconda (Ver. 5.2.0) in the command shell Ipython (Ver. 6.4.0) based on the Programming
- 158 language Python (Ver. 3.3.5). We used the Beer-Lambert law to model R_{SW_soil} under undisturbed conditions as the
- 159 aboveground vegetation was removed to measure the COS exchange of bare soil:

160
$$R_{SW-soil} = R_{SW} \exp(-K \, LAI) \tag{Eq.4}$$

where $R_{SW-soil}$ (Wm⁻²) is the shortwave radiation (SW) reaching the soil surface, R_{SW} is the incoming SW radiation reaching the top of the canopy, LAI is the plant area index (Eq. 2) and K is the canopy extinction coefficient assuming a spherical leaf inclination distribution (Wohlfahrt et al., 2001), which was calculated using the following equation:

$$164 \quad K = \frac{1}{2\cos(\psi)} \tag{Eq.5}$$

165 where ψ is the zenith angle of the sun in radians.

166

A random forest with 1000 trees was grown which resulted in an out of bag (OOB) score of (0.82). The OOB score can be interpreted as a pseudo-R2 and is widely used in random forest analyses (regression and classification), especially in the absence of a proper test dataset. It uses the data not seen by the trees (random forest uses bootstrapping) as a test dataset. The optimal input parameters, including maximum tree depth, were determined with the function GridSearchCV from the sklearn package.

172 2.5 Ecosystem fluxes

173 **2.5.1 Setup for ecosystem fluxes**

The COS, CO_2 and H_2O ecosystem fluxes were obtained using the eddy covariance method (Aubinet et al., 1999;Baldocchi, 2014). We used a 3-axis sonic anemometer (Gill R3IA, Gill Instruments Limited, Lymington, UK) to obtain high-resolution data of the 3 wind components. The intake of the tube for the eddy covariance measurements was installed in close proximity to the sonic anemometer and insulated as well as heated above T_{air} to prevent condensation within the tube. The air was sucked to the QCL at a flowrate of 7 l min⁻¹ using a Vacuum Pump (Agilent Technologies, CA, USA).

179 2.5.2 Ecosystem flux calculation

In a first step we used a self-developed software to determine the time lag, introduced by the separation of tube intake and 180 181 the sonic anemometer and the tube length, between the QCL and sonic dataset (Hortnagl et al., 2010). The data were then processed using the software EdiRe (University of Edinburgh, UK) and Matlab2019a (MathWorks, MA, USA). We used the 182 laser drift corrected χ_{COS} data and linear detrending to process the data before following the procedure to correct for sensor 183 184 response, tube attenuation, path averaging and sensor separation following Gerdel et al. (2017). The random flux uncertainty was calculated following Langford et al. (2015).Nighttime NEE and COS fluxes were filtered for periods of low friction 185 velocity (u*). We determined the u* threshold (0.19 m s⁻¹) by running the change point detection algorithm of Barr et al. 186 (2013) on nighttime NEE (Fig. S9) and applied it on the nighttime NEE as well as the COS fluxes. 187

188 We estimated the COS canopy flux from the difference between the measured COS ecosystem and the modelled COS soil189 flux.

190 2.5.3 Flux partitioning and leaf relative uptake

191 The GPP on ecosystem level was determined using the FP+ model put forward by Spielmann et al. (2019). The model

estimates the GPP on the basis of nighttime net ecosystem exchange (NEE) measurements of CO_2 that are assumed to provide the temperature response of the ecosystem respiration (RECO) as well as a light dependency curve to estimate GPP based on the daytime NEE (Lasslop et al., 2010):

195
$$NEE = \frac{\alpha\beta R_{PAR}}{\alpha R_{PAR} + \beta} + rb \ e^{E_0(\frac{1}{T_{ref} - T_0} - \frac{1}{T_{air} - T_0})}$$
 (Eq.6)

where α denotes the canopy light utilization efficiency (µmol CO₂ µmol⁻¹ photons), β the maximum CO₂ uptake rate of the 196 canopy at light saturation (μ mol CO₂ m⁻² s⁻¹), R_{PAR} the incoming photosynthetic active radiation (μ mol m⁻² s⁻¹), rb the 197 ecosystem base respiration (µmol m⁻² s⁻¹) at the reference temperature T_{Ref} (°C), which is set to 15°C, T_{air} (°C) refers to the 198 air temperature and E₀ (°C) to the temperature sensitivity of RECO. T₀ was kept constant at -46.02°C. We did not use the 199 200 VPD modifier of beta put forward by Lasslop et al. (2010) as its value could not be estimated with confidence. We 201 determined the parameter E_0 by using nighttime data minimizing the root squared mean error. For the determination of the remaining five unknown model parameters of the two flux partitioning models we used DREAM, a multi-chain Markov 202 203 Chain Monte Carlo algorithm (for more detail see Spielmann et al. (2019)). We calculated the parameters for ~15 day windows but adjusted them to not overlap with a cut of the grassland. 204

The ecosystem relative uptake (ERU) was calculated using Eq. 1 substituting the GPP with the NEE and using the COS ecosystem flux for F_{COS} .

The FP+ model by Spielmann et al. (2019) extends the daytime FP (Eq.6) to also estimate the COS ecosystem fluxes by linking the GPP resulting from the first part on the right-hand side of Eq.6 with the COS exchange through:

$$209 \quad F_{COSmodel} = \frac{GPP \, LRU\chi_{COS}}{\chi_{CO2}} \tag{Eq.7}$$

210 developed by Sandoval-Soto et al. (2005), where $F_{COSmodel}$ is the modelled COS flux (pmol m⁻² s⁻¹), χ_{COS} (ppt) and χ_{CO2} (ppm)

211 are the measured ambient mole fractions of COS and CO₂ respectively and LRU (-) is the leaf relative uptake rate:

212
$$LRU = \iota e^{\left(\frac{\kappa}{R_{PAR}}\right)}$$
 (Eq.8)

where ι (-) corresponds to the LRU at high light intensity and the parameter κ (µmol m⁻² s⁻¹) governs the increase of LRU at low light conditions. While mathematically ι is only obtained at infinitely high PAR, in practice above about 700 µmol m⁻² s⁻¹ PAR (Kooijmans et al., 2019) only insignificant change is reported in other studies (Stimler et al., 2011). The light dependency of LRU originates from the fact that the COS uptake by the enzyme CA is light-independent, while the CO₂ uptake by Rubisco depends on solar radiation absorbed by leaf chlorophyll (Whelan et al., 2018;Kooijmans et al., 2019;Wohlfahrt et al., 2012).

The method stated above infers LRU solely on the basis of ecosystem scale fluxes, whereas other studies typically use branch/leaf chamber measurements (Yang et al., 2018) to determine the relationship between the COS and CO_2 uptake rates.

221 2.5.4 Linear perturbation analysis

- 222 The relative contribution of the parameters GPP, $F_{COSmodel}$, χ_{CO2} and χ_{COS} that drive ι (Eq. 7) were estimated through a linear 223 perturbation analysis (Stoy et al., 2006).
- 224 The changes in ι ($\delta\iota$) between the target and the reference window (before the 2nd cut, i.e. June 18, 2015-July 07, 2015) are

considered the total derivative of Eq. 7 and can be represented by a multivariate Taylors's expansion where the higher-order terms are neglected in this first-order analysis:

227
$$\delta\iota = \frac{\partial\iota}{\partial F_{\text{COSmod}}} dF_{\text{COSmod}} + \frac{\partial\iota}{\partial\chi_{COS}} d\chi_{COS} + \frac{\partial\iota}{\partial\text{GPP}} dGPP + \frac{\partial\iota}{\partial\chi_{CO2}} d\chi_{CO2}$$
(Eq.9)

228 The relative contributions of the parameters were determined by computing the partial derivatives of Eq. 7.

229	$\frac{\partial \iota}{\partial F_{cOSmod}} = \frac{\chi_{CO2}}{\chi_{COS}GPP}$	(Eq.10)
230	$\frac{\partial \iota}{\partial \chi_{COS}} = \frac{-\chi_{CO2} F_{cOSmod}}{\chi_{COS}^2 GPP}$	(Eq.11)
231	$\frac{\partial \iota}{\partial \text{GPP}} = \frac{\chi_{CO2}F_{COSmod}}{\chi_{COS}GPP^2}$	(Eq.12)
232	$\frac{\partial \iota}{\partial \chi_{CO2}} = \frac{F_{COSmod}}{\chi_{COS}GPP}$	(Eq.13)

234 2.6 Ancillary data

Supporting meteorological measurements included T_{air} (RFT-2, UMS, Munich, GER), T_{soil} (TCAV, Campbell Scientific, Logan, UT, USA), SWC (ML2x, Delta-T Devices, Cambridge, UK), incident solar radiation (CNR-1, Klipp and Zonen, Delft, NLD), incident photosynthetic active radiation (PAR) (BF2H, Delta-T Devices Ltd, Cambridge, UK) and the Normalized Difference Vegetation Index (NDVI) sensor (SRS-NDVI, Meter, Pullman, WA, USA). The data were recorded throughout the whole season as 1 min values and stored as half-hourly means and standard deviations.

240 3 Results

241 **3.1 Environmental conditions**

Air temperature ranged between -2 °C and 33 °C with a mean of 13 °C during the study period from 15th of May to first of 242 243 November (Fig. 1). While the majority of precipitation (total 360.5 mm) fell as rain, we observed an exceptionally late snow event on the 20th of May, which did not melt for almost two days (Fig. 1). Although the VPD reached values of above 2 kPa 244 during 25 days, and plant available water dropped below 50 % on 111 days during the campaign (Fig. 1), we did not observe 245 any relationship with COS (see Fig S1-S2). Due to the removal of the aboveground biomass, the cuts reduced LAI. They 246 247 also reduced the normalized difference vegetation index (NDVI) (Fig. 1), which is a measure of canopy greenness (Tucker, 248 1979). The NDVI further decreased in the subsequent days as a consequence of dying plant parts remaining at the field site 249 (Fig 2 panels a-c). This can also be observed in the webcam photos (Photo S1-S3).





Figure 1. Seasonal cycle of ancillary variables. Daily minimum, maximum and median (a) air and (b) soil temperatures (°C) indicated by the lower and upper end of the bars and the white circle, respectively. (c) Daily maximum incident shortwave radiation (W m^{-2}) reaching

253 the top of the canopy (black squares) and reaching the soil surface (white circles). (d) Daily minimum, maximum and median vapor

pressure deficit (kPA) indicated by the lower and upper end of the bars and the white circle, respectively. (e) Plant available water (%) depicted by black squares and cumulative precipitation (mm) depicted by open circles. (f) Modelled leaf area index (black lines), measured

256 LAI (grey squares) and normalized difference vegetation index (open circles).





Figure 2: The response of the daily midday medians of NDVI (yellow circles), COS (blue circles) and CO_2 (red circles) ecosystem fluxes around the 4 cutting events (a-d) of the grassland. The errorbars depict the respective median absolute deviations. The cuts are marked by a red dashed line.

261 **3.2 COS mole fractions above and within the canopy**

262 While the canopy depleted the ambient χ_{COS} during day as well as nighttime, we found that the χ_{COS} reached values as low as 263 134 ppt (depletion of 102 ppt with respect to the mole fraction at canopy height) during nighttime (see Fig. 3) at the bottom of the canopy in contrast to the midday χ_{COS} , which only went down to 389 ppt (depletion of 125 ppt with respect to the mole 264 fraction at canopy height). We observed a decrease in χ_{CO2} (up to 26 ppm) within the most upper layers of the canopy 265 compared to χ_{CO2} at canopy height during daytime, while χ_{CO2} increased within the lowest layers compared to χ_{CO2} at the 266 canopy height due to soil respiration. The above canopy χ_{COS} increased considerably starting at the onset of the day and 267 reached 587 ppt at 4 p.m. with a steep increase until 11 a.m. Over the course of the season the midday ambient χ_{COS} 268 269 decreased from 500 ±28 ppt from mid-June to mid-July to 405±29 ppt in October with the trend of increasing χ_{COS} starting at 270 the end of September (see Fig. S6).



271

Figure 3. Vertical gradient of the (a) COS and (b) CO_2 mole fraction (ppt and ppm, respectively) depicted by the background color between the soil and the eddy covariance tower at 250 cm for one day. The left y axis shows the log of the measurement divided by the canopy height (z/h). The white circles depict the incoming shortwave radiation (R_{SW}) in (W m⁻²). Plant area density (PAD) split into living (green) and dead (brown) plant material (c). Vertical gradient of the difference between the mole fraction at canopy height and each measurement height for (d) COS and (e) CO₂.

278 3.3 COS soil flux

The fluxes resulting from the soil chamber measurements ranged from -6.3 to 40.9 pmol $m^{-2}s^{-1}$, with positive fluxes denoting emission (see Fig. 4 panel d).

During nighttime (RSW = 0, n = 43), 74.4 % of the COS fluxes were negative, implying that the soil of the grassland acted 281 as a net sink for COS (range of -4.4 to 6.9 pmol m⁻²s⁻¹), whereas soils transitioned to a source in 88.5 % of all daytime 282 measurements ($R_{sw} > 0$, n = 200), reaching the highest fluxes of 40.9 pmol m⁻²s⁻¹ during midday (see Fig. 4 a-c and Fig. S3). 283 This diel pattern was maintained over the course of the season, however with decreasing maximum COS source strength of 284 285 the soil towards the end of the season (Fig. 4 a-c and Fig. S3). The random forest regression revealed that the most important 286 variable for predicting the soil fluxes was the incident shortwave radiation reaching the soil surface (R_{SW-soil}), accounting for more than 73.53 % of the total variance explained by the final model, while SWC and T_{soil} only accounted for 17.84 % and 287 288 8.62 %, respectively. The fast response of the COS soil fluxes to changes in R_{SW} can be seen in Fig. 4 a, where we observed a decrease of R_{SW-soil} as well as the COS soil flux during a cloudy period, even when the soil temperature still increased. Soil 289 fluxes estimated with the random forest regression ranged from -1.3 to 5.0 pmol m⁻²s⁻¹, reflecting the fact that under real-290 291 world conditions very little solar radiation reaches the soil surface. (Fig. 4 e). The resulting emissions peaked during daytime 292 shortly after the cuts when a high proportion of incident radiation was reaching the soil surface, while simulated nighttime fluxes were dominated by uptake (in 93 % of all cases) for the whole season. 293

294



295

Figure 4. COS soil fluxes (pmol m⁻²s⁻¹) originating from manual chamber measurements of three selected days (a), (b) and (c) depicted by black circles, incident shortwave radiation reaching the soil ($R_{SW-soil}$) depicted by the gray area and soil temperature (T_{soil}) depicted by empty black bordered squares. (d) Histogram of all conducted COS soil chamber observations with the dashed vertical lines depicting the 25, 50 and 75% quantile. (e) Season plot of the modelled COS soil fluxes ($F_{COSsoil}$) depicted by the black circles, incident shortwave radiation reaching the soil surface ($R_{SW-soil}$) depicted by grey circles and the black dashed lines depicting the cuttings of the grassland.

302 3.4 COS and CO₂ ecosystem-scale fluxes

The grassland acted as a net sink for COS during the majority of our study period with 80 % of the COS ecosystem fluxes 303 between -56.0 pmol m⁻²s⁻¹ and -4.5 pmol m⁻²s⁻¹ during daytime and -37.8 pmol m⁻²s⁻¹ and 9.2 pmol m⁻²s⁻¹ during nighttime. 304 We observed a net release of COS at the field site 11.2 % of the time. The net CO₂ fluxes ranged from -20.4 to 4.8 µmol m⁻ 305 2 s⁻¹ and -30.3 to 36.4 µmol m⁻²s⁻¹ for 80% of all observation during day and nighttime, with daytime net emissions occurring 306 after the cuttings of the grassland (Fig. 2 a-c and Fig. 5 a). While the COS nighttime fluxes remained unaffected by the cuts 307 308 (Fig. 5 c), the daytime fluxes showed a high variability (see Fig. 5 b). Especially after the cuts we observed a strong decline in COS uptake (Fig. 4 b) and the grassland even turned into a net source for COS in middays (Fig. 2 a-c) with a highest 309 emission flux of 26.8 pmol m⁻²s⁻¹ (midday median) in August after the cut. We observed COS emissions for up to 8 days 310 after the cut, when the dried litter had already been removed (Fig. 2 a-c). Compared to respiration processes outpacing GPP 311 312 almost instantaneously after the cuts, the grassland reached its peak COS emission on the day of the cut only in July, whereas the peak was reached five days after the cut in June and August (Fig. 2 a-c). The cut in October led to a reduction in 313 COS uptake, which declined across several days and did not recover, as the end of the season was reached (Fig. 2 d & Fig. 5 314 b). After the fertilization of the field in October the grassland also turned into a source for COS during midday hours for one 315 day (Fig. 5 b). Our flux measurements also included a time when the grassland was covered with snow (on May 20, 2015), 316 317 which reduced the COS (and CO₂) fluxes to values close to zero. Over the course of the season, we observed a decline in the magnitude of the daytime COS uptake from -50.6 ± 24.6 pmol m⁻²s⁻¹ during midday in the first week of May down to $-10.3 \pm$ 318 10.4 pmol m⁻²s⁻¹ in the last week of October, which was also correlated with the decline in the CO₂ sink strength from -19.9 319 \pm 8.0 µmol m⁻²s⁻¹ to -4.4 \pm 1.5 µmol m⁻²s⁻¹ (Fig. 5 a-b). We observed an increase in COS and CO₂ fluxes within the growing 320 321 phases after the cuts only up to an LAI of ~ 4 (-) (Fig. S4-S5), which then levelled out for COS and declined for CO₂ due to 322 ecosystem respiration compensating GPP.



Figure 5: Seasonal cycle of the half hourly CO₂ (a), COS daytime (b) and COS nighttime (c) ecosystem fluxes in μ mol m⁻²s⁻¹ and pmol m⁻²s⁻¹ depicted by black circles if they are above the limit of detection (LOD) and grey x's if they are below (Langford et al., 2015). The red circles depict the mean fluxes between 11 a.m. and 2 p.m. CET for (a & b) and between 11 p.m. and 2 a.m. for (c) that are above the LOD, while the red x's indicate means below the LOD. The red error bars depict the ±1standard deviation of the mean. The blue lines depict the running mean (5 days) for the mean fluxes. The black dashed lines depict the cuttings of the grassland.

330

The seasonal pattern of a decrease in COS sink strength was similar for nighttime fluxes (-18.0± 29.6 pmol m⁻²s⁻¹ to -10.6 ± 18.2 pmol m⁻²s⁻¹) (Fig. 5c). The mean nighttime respiration also decreased over the course of the season from 15.9 ± 28.2 µmol m⁻²s⁻¹ to 9.4 ± 17.5 µmol m⁻²s⁻¹ between May and October (Fig. 5a).

334 Periods between May and August of low (after cuts) and high (before cuts) LAI were compared as diel courses (Fig. 5). Over the course of the day, both periods were characterized by a mean uptake of COS (Fig 6 c & d). Even though the uptake was 335 similar during nighttime, the daytime pattern differed considerably. The modelled contribution of the soil to the ecosystem 336 337 scale COS flux under high LAI conditions (Fig. 6 d) was minor, contributing between 1.3 % and 5.5/5.7 % of the ecosystem flux during midday and morning/evening, respectively. In contrast, during low LAI conditions the soil contribution to the 338 339 ecosystem fluxes increased during daytime and contributed up to 82.4% of the mean hourly COS ecosystem flux (Fig 6. c). While the grassland acted as a stronger sink for COS during daytime at a high LAI, reaching peak mean uptake values of up 340 to -41.8 pmol m⁻² s⁻¹ \pm 16.8 pmol m⁻² s⁻¹ during midday, the mean daytime sink strength weakened and we observed close to 341 zero fluxes during midday in periods of low LAI. The magnitude of the soil flux $(2 \pm 1 \text{ pmol m}^{-2} \text{ s}^{-1})$ was not high enough to 342 explain the difference of up to 26.0 pmol $m^{-2}s^{-1}$ between the measured COS ecosystem flux and COS flux resulting from the 343 FP+ model (Fig 6 c), suggesting a missing COS source. For phases of high LAI we saw a good agreement between hourly 344 averaged modelled and measured COS ecosystem fluxes (Fig 6 d). While the grassland acted as a net sink for CO2 during 345 periods of high LAI (Fig. 6 b), a combination of a decline in GPP and an increase in daytime RECO, as more incoming 346 radiation was heating the soil surface, turned it into a net source during midday in periods of low LAI (Fig. 6 a). 347

348



Figure 6. Mean diel variation of the measured and modelled CO_2 (a & b) and COS (c & d) fluxes for phases of low (LAI <=3) (a & c) and high (LAI >= 4) (b & d) from May to August. The carats depict the modelled gross primary productivity (blue), the modelled ecosystem respiration (red) and the measured CO_2 ecosystem fluxes (black) in µmol m⁻²s⁻¹. The circles depict the modelled COS soil flux (yellow), the modelled COS ecosystem flux (turquoise) and the measured COS ecosystem fluxes (black) in pmol m⁻²s⁻¹. The red area depicts the difference between the measured ecosystem flux and the sum of the modelled fluxes. The grey areas depict the ±1 standard deviation of the mean for all the measured fluxes. The white bars depict the diel mean total incoming shortwave radiation (W m⁻²) while the grey bars indicate the diel mean shortwave radiation reaching the soil surface.

358

359 3.5 Leaf and ecosystem relative uptake

The LRU at high-light conditions, t, which we calculated using the FP+ algorithm increased from relatively stable precut 360 levels of 0.9-1. (-) before the 2^{nd} and the 1^{st} cut to up to 1.6 (-) after the 4^{th} cut (Fig. 7a). After the decrease in t between the 361 2^{nd} and the 3^{rd} cut, ι increased steadily until the 4^{th} cut, with the 3^{rd} cut seemingly not having an effect. The reason for the 362 increase in 1 after the 2nd and 4th cut was a stronger decrease in GPP than the COS uptake, while both decreased more evenly 363 after the 3rd cut (Fig. 7b). We observed 1 in the period before the 4th cut to be influenced not only by a decrease in COS 364 uptake, but also by a decrease in COS mole fraction (Fig 7b). The mean midday ERUs varied between 2.0 ± 0.1 (-) before 365 and 4.5 ± 0.4 (-) after the cuts. The larger difference between the ERU and t after the cuts reflect that we observed similar 366 respiration rates at low and high LAI (Fig 6a-b). 367

- 368 Under low light conditions, the LRU increased during pre- and post-cut phases in a similar manner with the last 15-day
- 369 period in October showing an earlier increase in the morning and evening (see Fig. S7).



Figure 7. (a) The seasonal cycle of t (black line) with the 95% confidence interval (gray area) resulting from the FP+ model and the midday mean (11 a.m. – 2 p.m. at PAR > 800 μ mol m₋₂ s₋₁) ecosystem relative uptake (ERU) (blue line) using the CO₂ ecosystem flux for the calculation windows (~15 days adjusted to cuts). The dashed black line depicts the progression of the leaf area index (LAI) of the grassland. (b) The contribution of the drivers (F_{COS}, χ_{COS} , GPP and χ_{CO2}) to the changes in t between all calculation windows and the reference period (DOY 169-188) resulting from the linear perturbation analysis compared to the observed change in t (δ t).

377 4 Discussion

378 4.1 COS mole fractions

The continuous seasonal decrease in above-canopy χ_{COS} was within the range of published records observing mole fractions to decrease from 465 (in summer) to 375 ppt (in winter) (Kuhn et al., 1999). This pattern is typical for the northern hemisphere and the COS drawdown by terrestrial ecosystems (Montzka et al., 2007). We found the lowest χ_{COS} at the end of September, which coincides with the lowest ambient mole fractions of COS, measured in Ireland, the closest COS observation site Mace Head (MHD) of NOAA, on the 6th of October (Fig. S6).

The extremely low COS canopy mole fractions we observed within the canopy, have also been reported by Rastogi et al. 384 385 (2018), who measured a mean χ_{COS} minimum of 152 ppt at 1 m above the soil within an old growth forest. Compared to the consistent decrease of COS below the canopy level during day and nighttime, the gradient for CO₂ reverses during nighttime 386 387 due to ongoing respiration processes while plants are not photosynthetically active. Even though the COS mole fraction at the layer closest to the soil were higher during day than during nighttime, the absolute decrease in COS was lower during 388 389 nighttime due to partial stomatal closure (Kooijmans et al., 2017;Campbell et al., 2017). The absolute difference in 390 concentrations during day and nighttime originate from changes in the height of the planetary boundary layer (PBL). While 391 the PBL is shallow during nighttime and the COS mole fraction decreases due to sink strength of the grassland, at the onset 392 of the day, the PBL layer height increases fast and COS rich air is transported down to the ecosystem (Fig. S12) (Campbell 393 et al., 2017). A similar steep increase until midday has also been observed by Rastogi et al. (2018). Even though CO_2 and 394 COS share a similar pathway into plants, reflected by their respective decrease in the mole fractions within the canopy, we 395 saw a difference at the lower levels of our gradient analysis during daytime. We only observed an increase in CO2 mole fractions, caused by the release of CO_2 through respiration processes in the soil, whereas COS mole fractions further 396 397 declined down to the soil surface. This supports our soil model, which predicted only minor COS fluxes under conditions of 398 high LAI, when only a small portion of incident radiation reaches the soil surface.

399 4.2 Soil fluxes

The nighttime soil chamber measurements compare well in terms of magnitude with the COS fluxes resulting from studies using dark chambers in agricultural and grassland sites (Whelan et al., 2018;Maseyk et al., 2014;Whelan and Rhew, 2016;Liu et al., 2010) and indicate the soil to be a small sink for COS. The current understanding of COS soil exchange links the COS consumption to soil biota e.g. bacteria and fungi, possessing the ubiquitous enzyme CA (Kesselmeier et al., 1999;Meredith et al., 2019). The origin of COS in soils on the other hand is still highly debated, but comparisons of untreated and sterilized soils suggest yet unknown abiotic processes (Meredith et al., 2019;Kitz et al., 2019).

The high COS emissions resulting from the soil chambers during daytime lie at the upper end of recently stated values of agricultural and grassland sites (Whelan et al., 2018;Kitz et al., 2017;Maseyk et al., 2014;Liu et al., 2010). Partly, this can be attributed to the type of chambers we used and their deployment. We allowed the full spectrum of incoming radiation to reach the soils surface, whereas most other studies used dark chambers. Therefore we were able to capture the influence of COS emission processes coupled to thermo- and photo production on our COS soil fluxes (Whelan and Rhew, 2015;Kitz et al., 2019;Meredith et al., 2018). This also led to lower peak soil emissions of COS at the end of the season, when the

412 incoming radiation declined.

The low COS mole fractions observed in the lowermost canopy layers just above the soil surface emphasize the importance of using air from within the canopy for soil chamber measurements and not COS richer air from above the canopy, which would increase the COS gradient and thus increase the uptake/decrease emission of COS to/from the soil.

416 Our modelled COS soil fluxes peak at about 12% of the maximum emissions retrieved from the soil chambers. This is owed 417 to the difference in incident radiation reaching the soil surface between the fluxes resulting from chamber measurements and 418 our model. For the chambers, the aboveground biomass was removed, whereas our modelled fluxes were adjusted for 419 undisturbed canopy conditions.

Another factor contributing to the high COS soil emissions might be the yearly fertilization using slurry, as high nitrogen content in soils has been linked to a higher source strength of COS (Kaisermann et al., 2018). This agrees well with the study of Kitz et al. (2019), who found a correlation between increased soil nitrogen content and soil COS emission in a laboratory experiment with samples taken from the grassland at two different dates (i.e. June and September).

425 4.3 Ecosystem fluxes

Our observations show that the agriculturally used grassland acted as a major sink for COS during the growing season. The fluxes fit well within or even exceeded the COS uptake rates of published grassland and agricultural sites during their growing phases (Billesbach et al., 2014;Whelan and Rhew, 2016;Geng and Mu, 2004). The late snow event that occurred in the peak growing season almost completely inhibited the exchange of CO_2 and COS, as the snow acted as a diffusion barrier for these compounds (Björkman et al., 2010).

431 The cuttings and the consecutive drying of the above ground plant material at the site had a major influence on the COS 432 exchange. COS emissions of a similar magnitude have also been reported at agricultural fields in phases of senescence 433 (Maseyk et al., 2014;Billesbach et al., 2014). Although the soil was a strong source for COS, caused by the high R_{soil} and 434 T_{soil} (Whelan and Rhew, 2015;Kitz et al., 2019;Meredith et al., 2018), and the sink strength of the grassland was low due to 435 the reduced aboveground biomass, soil fluxes did not explain the emission on ecosystem level (see Fig. 6a). As plants 436 contain precursors involved in COS emission processes, e.g. methionine and cysteine (Meredith et al., 2018), the plant litter 437 and dying plant parts remaining at the site after the cuts might be the missing source of COS. Laboratory tests of the soil of the grassland have shown that a mixing of dried litter and soil lead to a strong but short-lived emission peak of COS (Kitz et 438 439 al., 2019). We did not observe strong COS emissions after the last cut, as the incoming solar radiation, which we hypothesize 440 to amplify the degradation of sulfur containing compounds of plants, was reduced at the end of the season. Alternatively, the 441 cutting of the grassland might induce stress mediated COS production in the remaining living plant parts (Bloem et al.,

2012;Gimeno et al., 2017). The delay in the peak COS emissions at ecosystem scale after the cuts could indicate that some
yet unknown biotic or abiotic processes take several days to release COS.

The short-lived COS emission by yet unknown biotic or abiotic processes after the fertilization of the grassland towards the end of the growing season was likely triggered by the increase of available nitrogen (Kaisermann et al., 2018) and COS precursors introduced to the soil in the form of cattle slurry (Hörtnagl et al., 2018).

447 Due to the independence of CA to catalyze COS without R_{PAR} (Stimler et al., 2011), the grassland remained a sink for COS

448 during nighttime. Again, the soil sink was too small to explain the total COS exchange (Fig. 6), which indicates that the plant 449 stomata were not fully closed (Kooijmans et al., 2017) and were responsible for the majority of the COS uptake. The

450 minimum or residual stomatal conductances at the field site in Neustift have been reported to be between 10 and 65 mmol m

- 451 2 s⁻¹ depending on the species (Wohlfahrt, 2004).
- The large variability in COS nighttime fluxes (Fig. 5c) is due to the combination of low wind speeds and stable stratification, which results in highly intermittent CO_2 (Wohlfahrt et al., 2005) and COS fluxes compared to daytime. On half-hourly basis, even a nighttime net uptake of CO_2 has been reported at the field site, which is typically compensated for by large CO_2 emissions in a subsequent averaging period (Wohlfahrt et al., 2005). We also observed this pattern for COS.

Although we observed phases of high VPD and low SWC (Fig. 1), they did not lead to a decrease in CO_2 and COSecosystem fluxes (Fig. S1-S2), which has already been observed for the grasslands CO_2 and H_2O fluxes between 2001 and 2009. The species located at the site were insensitive to progressive drought conditions (Brilli et al., 2011).

459

460 **4.4 LRU**

The parameter ι of this study is placed at the lower end of a recent compilation of published leaf-level LRUs, that put 95% of all data between 0.7 (-) and 6.2 (-) with a median of 1.7 (-) (Whelan et al., 2018) and also lower than the LRU of 2.53 (-) estimated for grasslands by Seibt et al. (2010). Even the higher ι after the cuts was low compared to these studies. The seasonal trend of ι was strongly influenced by the cutting of the grass and can be attributed mainly to changes in the ratio of COS uptake to GPP. However, we also observed a strong decline in the ambient mole fraction of COS, which also had an equally strong influence on the change in ι as the COS flux for the 15 day window before the last cut (Fig 7 b).

467 Even though the changes in t can be explained, it is important to keep in mind that the grassland was a source for COS on

- 468 ecosystem level after the cuts. For the calculation of LRUs we had to remove the canopy flux data containing COS and/or
- 469 CO₂ emissions since they would yield negative values for ERU and LRU (see Eq.8). This indicates that the unknown source
- 470 strength after cuts likely decreases the post-cut ι 's.

471 5 Conclusion

472 Due to the management interventions at the grassland site, the leaf area development was decoupled from seasonal changes 473 in environmental forcing. This allowed us to measure concurrent CO_2 and COS fluxes at soil and ecosystem level for 474 multiple growing periods within one season. The LAI on seasonal scale as well as incoming solar radiation on hourly to 475 seasonal scales determined whether soils were a source or a sink for COS. The incoming shortwave radiation reaching the 476 soil surface had a decisive influence on the COS soil surface flux and thus supports our hypothesis H4. The covariance 477 between the daytime CO₂ and COS fluxes on daily to seasonal level was high and the fluxes only diverged after the cuts, leading to higher LRUs. Beside the perturbations of the ecosystem, the sink strength of the grassland was high for COS and 478 479 declined over the course of the season (H1). The COS emissions at ecosystem scale shortly after the cuts, which could not be 480 explained by the soil source, raise questions about other unknown mechanisms of COS production within ecosystems (H2). 481 With the exception of short periods after the cuts, the LRUs under high light conditions were relatively constant during the

482 season, indicating a good correlation between the COS flux and GPP under stable conditions (H3).

483 6. Data availability

484 Data and materials availability: https://doi.org/10.5281/zenodo.3886554

485 **7. Author contributions**

Felix M. Spielmann: Data curation, Formal analysis, Investigation, Methodology, Software, Visualization, Writing – original
 draft

- 488 Albin Hammerle: Data curation, Investigation, Software, Writing original draft
- 489 Florian Kitz: Data curation, Formal analysis, Investigation, Methodology, Software, Writing original draft

490 Katharina Gerdel: Investigation, Software, Writing - original draft

- 491 Georg Wohlfahrt: Conceptualization, Funding acquisition, Investigation, Methodology, Project administration, Software,
- 492 Supervision, Writing original draft

493 8. Competing interests

494 The authors declare no competing financial interests.

495 9. Acknowledgements

496 This study was financially supported by the Austrian National Science Fund (FWF; contracts P26931, P27176, and I03859),

497 the Tyrolean Science Fund (contract UNI-0404/1801), and the University of Innsbruck (Infrastructure funding by Research

498 Area Alpine Space-Man and Environment to G. W). Financial support to F. M. S. was provided through a PhD scholarship499 by the University of Innsbruck. We thank family Hofer (Neustift, Austria) for kindly granting us access to the study site.

500 COS flask data were provided by the Global Monitoring Division of the Nationals Oceanic and Atmospheric 501 Administration's Earth System Research Laboratory (NOAA ESRL/GMD). The authors declare no competing financial

502 interests.

503 10. References

- Asaf, D., Rotenberg, E., Tatarinov, F., Dicken, U., Montzka, S. A., and Yakir, D.: Ecosystem photosynthesis inferred from measurements of carbonyl sulphide flux, Nature Geoscience, 6, 186-190, 10.1038/ngeo1730, 2013.
- Aubinet, M., Grelle, A., Ibrom, A., Rannik, U., Moncrieff, J., Foken, T., Kowalski, A. S., Martin, P. H., Berbigier, P.,
 Bernhofer, C., Clement, R., Elbers, J., Granier, A., Grünwald, T., Morgenstern, K., Pilegaard, K., Rebmann, C.,
 Snijders, W., Valentini, R., and Vesala, T.: Estimates of the Annual Net Carbon and Water Exchange of Forests:
 The EUROFLUX Methodology, in: Advances in Ecological Research Volume 30, edited by: Fitter, A. H., and
 Raffaelli, D. G., Advances in Ecological Research, Academic Press, 113-175, 1999.
- Baldocchi, D.: Measuring fluxes of trace gases and energy between ecosystems and the atmosphere the state and future of
 the eddy covariance method, Glob Chang Biol, 20, 3600-3609, doi.org/10.1111/gcb.12649, 2014.
- Barr, A. G., Richardson, A. D., Hollinger, D. Y., Papale, D., Arain, M. A., Black, T. A., Bohrer, G., Dragoni, D., Fischer, M.
 L., Gu, L., Law, B. E., Margolis, H. A., McCaughey, J. H., Munger, J. W., Oechel, W., and Schaeffer, K.: Use of change-point detection for friction-velocity threshold evaluation in eddy-covariance studies, Agricultural and Forest Meteorology, 171, 31-45, 10.1016/j.agrformet.2012.11.023, 2013.
- 517 Billesbach, D. P., Berry, J. A., Seibt, U., Maseyk, K., Torn, M. S., Fischer, M. L., Abu-Naser, M., and Campbell, J. E.: 518 Growing season eddy covariance measurements of carbonyl sulfide and CO2 fluxes: COS and CO2 relationships in Agricultural 519 Southern Great Plains winter wheat, and Forest Meteorology, 184, 48-55, 520 10.1016/j.agrformet.2013.06.007, 2014.

- Björkman, M. P., Morgner, E., Cooper, E. J., Elberling, B., Klemedtsson, L., and Björk, R. G.: Winter carbon dioxide
 effluxes from Arctic ecosystems: An overview and comparison of methodologies, Global Biogeochemical Cycles,
 24, 10.1029/2009gb003667, 2010.
- Bloem, E., Haneklaus, S., Kesselmeier, J., and Schnug, E.: Sulfur Fertilization and Fungal Infections Affect the Exchange of
 H2S and COS from Agricultural Crops, Journal of Agricultural and Food Chemistry, 60, 7588-7596,
 10.1021/jf301912h, 2012.
- Brilli, F., Hörtnagl, L., Hammerle, A., Haslwanter, A., Hansel, A., Loreto, F., and Wohlfahrt, G.: Leaf and ecosystem
 response to soil water availability in mountain grasslands, Agricultural and Forest Meteorology, 151, 1731-1740,
 doi.org/10.1016/j.agrformet.2011.07.007, 2011.
- Brühl, C., Lelieveld, J., Crutzen, P. J., and Tost, H.: The role of carbonyl sulphide as a source of stratospheric sulphate
 aerosol and its impact on climate, Atmos. Chem. Phys., 12, 1239-1253, 10.5194/acp-12-1239-2012, 2012.
- Campbell, J. E., Whelan, M. E., Berry, J. A., Hilton, T. W., Zumkehr, A., Stinecipher, J., Lu, Y., Kornfeld, A., Seibt, U.,
 Dawson, T. E., Montzka, S. A., Baker, I. T., Kulkarni, S., Wang, Y., Herndon, S. C., Zahniser, M. S., Commane,
 R., and Loik, M. E.: Plant Uptake of Atmospheric Carbonyl Sulfide in Coast Redwood Forests, Journal of
 Geophysical Research-Biogeosciences, 122, 3391-3404, 10.1002/2016jg003703, 2017.
- Ensign, S. A.: Reactivity of Carbon Monoxide Dehydrogenase from Rhodospirillum rubrum with Carbon Dioxide, Carbonyl
 Sulfide, and Carbon Disulfide, Biochemistry, 34, 5372-5381, 10.1021/bi00016a008, 1995.
- Geng, C., and Mu, Y.: Carbonyl sulfide and dimethyl sulfide exchange between lawn and the atmosphere, Journal of
 Geophysical Research: Atmospheres, 109, 10.1029/2003jd004492, 2004.
- Gerdel, K., Spielmann, F. M., Hammerle, A., and Wohlfahrt, G.: Eddy covariance carbonyl sulfide flux measurements with a
 quantum cascade laser absorption spectrometer, Atmospheric Measurement Techniques, 10, 3525-3537,
 10.5194/amt-10-3525-2017, 2017.
- Gimeno, T. E., Ogee, J., Royles, J., Gibon, Y., West, J. B., Burlett, R., Jones, S. P., Sauze, J., Wohl, S., Benard, C., Genty,
 B., and Wingate, L.: Bryophyte gas-exchange dynamics along varying hydration status reveal a significant carbonyl
 sulphide (COS) sink in the dark and COS source in the light, New Phytologist, 215, 965-976, 10.1111/nph.14584,
 2017.
- Hortnagl, L., Clement, R., Graus, M., Hammerle, A., Hansel, A., and Wohlfahrt, G.: Dealing with disjunct concentration
 measurements in eddy covariance applications: A comparison of available approaches, Atmospheric Environment,
 44, 2024-2032, 10.1016/j.atmosenv.2010.02.042, 2010.
- Hörtnagl, L., Barthel, M., Buchmann, N., Eugster, W., Butterbach-Bahl, K., Díaz-Pinés, E., Zeeman, M., Klumpp, K., Kiese,
 R., Bahn, M., Hammerle, A., Lu, H., Ladreiter-Knauss, T., Burri, S., and Merbold, L.: Greenhouse gas fluxes over
 managed grasslands in Central Europe, Global Change Biology, 24, 1843-1872, 10.1111/gcb.14079, 2018.
- Kaisermann, A., Jones, S., Wohl, S., Ogée, J., and Wingate, L.: Nitrogen Fertilization Reduces the Capacity of Soils to Take
 up Atmospheric Carbonyl Sulphide, Soil Syst., 2, 10.3390/soilsystems2040062, 2018.
- Kesselmeier, J., Teusch, N., and Kuhn, U.: Controlling variables for the uptake of atmospheric carbonyl sulfide by soil, J.
 Geophys. Res.-Atmos., 104, 11577-11584, 10.1029/1999jd900090, 1999.
- Kitz, F., Gerdel, K., Hammerle, A., Laterza, T., Spielmann, F. M., and Wohlfahrt, G.: In situ soil COS exchange of a temperate mountain grassland under simulated drought, Oecologia, 1-10, 10.1007/s00442-016-3805-0, 2017.
- Kitz, F., Gómez-Brandón, M., Eder, B., Etemadi, M., Spielmann, F. M., Hammerle, A., Insam, H., and Wohlfahrt, G.: Soil
 carbonyl sulfide exchange in relation to microbial community composition: Insights from a managed grassland soil
 amendment experiment, Soil Biology and Biochemistry, 135, 28-37, 10.1016/j.soilbio.2019.04.005, 2019.
- Kooijmans, L. M. J., Uitslag, N. A. M., Zahniser, M. S., Nelson, D. D., Montzka, S. A., and Chen, H. L.: Continuous and
 high-precision atmospheric concentration measurements of COS, CO2, CO and H2O using a quantum cascade laser
 spectrometer (QCLS), Atmospheric Measurement Techniques, 9, 5293-5314, 10.5194/amt-9-5293-2016, 2016.
- Kooijmans, L. M. J., Maseyk, K., Seibt, U., Sun, W., Vesala, T., Mammarella, I., Kolari, P., Aalto, J., Franchin, A., Vecchi,
 R., Valli, G., and Chen, H.: Canopy uptake dominates nighttime carbonyl sulfide fluxes in a boreal forest, Atmos.
 Chem. Phys., 17, 11453-11465, 10.5194/acp-17-11453-2017, 2017.
- Kooijmans, L. M. J., Sun, W., Aalto, J., Erkkilä, K. M., Maseyk, K., Seibt, U., Vesala, T., Mammarella, I., and Chen, H.:
 Influences of light and humidity on carbonyl sulfide-based estimates of photosynthesis, Proc Natl Acad Sci U S A,
 10.1073/pnas.1807600116, 2019.
- Krysztofiak, G., Té, Y. V., Catoire, V., Berthet, G., Toon, G. C., Jégou, F., Jeseck, P., and Robert, C.: Carbonyl Sulphide
 (OCS) Variability with Latitude in the Atmosphere, Atmosphere-Ocean, 53, 89-101,
 10.1080/07055900.2013.876609, 2015.
- Kuhn, U., Ammann, C., Wolf, A., Meixner, F. X., Andreae, M. O., and Kesselmeier, J.: Carbonyl sulfide exchange on an
 ecosystem scale: soil represents a dominant sink for atmospheric COS, Atmospheric Environment, 33, 995-1008,
 Doi 10.1016/S1352-2310(98)00211-8, 1999.
- Langford, B., Acton, W., Ammann, C., Valach, A., and Nemitz, E.: Eddy-covariance data with low signal-to-noise ratio:
 time-lag determination, uncertainties and limit of detection, Atmospheric Measurement Techniques, 8, 4197-4213,
 10.5194/amt-8-4197-2015, 2015.
- Lasslop, G., Reichstein, M., Papale, D., Richardson, A. D., Arneth, A., Barr, A., Stoy, P., and Wohlfahrt, G.: Separation of net ecosystem exchange into assimilation and respiration using a light response curve approach: critical issues and global evaluation, Global Change Biology, 16, 187-208, 10.1111/j.1365-2486.2009.02041.x, 2010.
- Le Quere, C., Andrew, R. M., Friedlingstein, P., Sitch, S., Pongratz, J., Manning, A. C., Korsbakken, J. I., Peters, G. P.,
 Canadell, J. G., Jackson, R. B., Boden, T. A., Tans, P. P., Andrews, O. D., Arora, V. K., Bakker, D. C. E., Barbero,

- 585 L., Becker, M., Betts, R. A., Bopp, L., Chevallier, F., Chini, L. P., Ciais, P., Cosca, C. E., Cross, J., Currie, K., 586 Gasser, T., Harris, I., Hauck, J., Haverd, V., Houghton, R. A., Hunt, C. W., Hurtt, G., Ilyina, T., Jain, A. K., Kato, E., Kautz, M., Keeling, R. F., Goldewijk, K. K., Kortzinger, A., Landschutzer, P., Lefevre, N., Lenton, A., Lienert, 587 S., Lima, I., Lombardozzi, D., Metzl, N., Millero, F., Monteiro, P. M. S., Munro, D. R., Nabel, J., Nakaoka, S., 588 Nojiri, Y., Padin, X. A., Peregon, A., Pfeil, B., Pierrot, D., Poulter, B., Rehder, G., Reimer, J., Rodenbeck, C., 589 590 Schwinger, J., Seferian, R., Skjelvan, I., Stocker, B. D., Tian, H. Q., Tilbrook, B., Tubiello, F. N., van der Laan-591 Luijkx, I. T., van der Werf, G. R., van Heuven, S., Viovy, N., Vuichard, N., Walker, A. P., Watson, A. J., Wiltshire, 592 A. J., Zaehle, S., and Zhu, D.: Global Carbon Budget 2017, Earth System Science Data, 10, 405-448, 10.5194/essd-593 10-405-2018, 2018.
- Liaw, A., and Wiener, M.: Classification and Regression by RandomForest, R News, 2/3, 18-22, 2002.
- Liu, J., Geng, C., Mu, Y., Zhang, Y., Xu, Z., and Wu, H.: Exchange of carbonyl sulfide (COS) between the atmosphere and various soils in China, Biogeosciences, 7, 753-762, 10.5194/bg-7-753-2010, 2010.
- Lorimer, G. H., and Pierce, J.: Carbonyl sulfide: an alternate substrate for but not an activator of ribulose-1,5-bisphosphate
 carboxylase, The Journal of biological chemistry, 264, 2764-2772, 1989.
- Maseyk, K., Berry, J. A., Billesbach, D., Campbell, J. E., Torn, M. S., Zahniser, M., and Seibt, U.: Sources and sinks of
 carbonyl sulfide in an agricultural field in the Southern Great Plains, Proceedings of the National Academy of
 Sciences of the United States of America, 111, 9064-9069, 10.1073/pnas.1319132111, 2014.
- Meredith, L. K., Boye, K., Youngerman, C., Whelan, M., Ogee, J., Sauze, J., and Wingate, L.: Coupled Biological and
 Abiotic Mechanisms Driving Carbonyl Sulfide Production in Soils, Soil Syst., 2, 27, 10.3390/soilsystems2030037,
 2018.
- Meredith, L. K., Ogée, J., Boye, K., Singer, E., Wingate, L., von Sperber, C., Sengupta, A., Whelan, M., Pang, E., Keiluweit,
 M., Brüggemann, N., Berry, J. A., and Welander, P. V.: Soil exchange rates of COS and CO18O differ with the
 diversity of microbial communities and their carbonic anhydrase enzymes, The ISME Journal, 13, 290-300,
 10.1038/s41396-018-0270-2, 2019.
- Montzka, S. A., Calvert, P., Hall, B. D., Elkins, J. W., Conway, T. J., Tans, P. P., and Sweeney, C.: On the global distribution, seasonality, and budget of atmospheric carbonyl sulfide (COS) and some similarities to CO2, J. Geophys. Res.-Atmos., 112, 10.1029/2006jd007665, 2007.
- Notni, J., Schenk, S., Protoschill-Krebs, G., Kesselmeier, J., and Anders, E.: The missing link in COS metabolism: a model
 study on the reactivation of carbonic anhydrase from its hydrosulfide analogue, Chembiochem : a European journal
 of chemical biology, 8, 530-536, 10.1002/cbic.200600436, 2007.
- Ogawa, T., Noguchi, K., Saito, M., Nagahata, Y., Kato, H., Ohtaki, A., Nakayama, H., Dohmae, N., Matsushita, Y., Odaka,
 M., Yohda, M., Nyunoya, H., and Katayama, Y.: Carbonyl Sulfide Hydrolase from Thiobacillus thioparus Strain
 THI115 Is One of the β-Carbonic Anhydrase Family Enzymes, Journal of the American Chemical Society, 135,
 3818-3825, 10.1021/ja307735e, 2013.
- Protoschill-Krebs, G., and Kesselmeier, J.: ENZYMATIC PATHWAYS FOR THE CONSUMPTION OF CARBONYL
 SULFIDE (COS) BY HIGHER-PLANTS, Botanica Acta, 105, 206-212, 1992.
- Rastogi, B., Berkelhammer, M., Wharton, S., Whelan, M. E., Itter, M. S., Leen, J. B., Gupta, M. X., Noone, D., and Still, C.
 J.: Large Uptake of Atmospheric OCS Observed at a Moist Old Growth Forest: Controls and Implications for Carbon Cycle Applications, Journal of Geophysical Research-Biogeosciences, 123, 3424-3438, 10.1029/2018jg004430, 2018.
- Sandoval-Soto, L., Stanimirov, M., von Hobe, M., Schmitt, V., Valdes, J., Wild, A., and Kesselmeier, J.: Global uptake of
 carbonyl sulfide (COS) by terrestrial vegetation: Estimates corrected by deposition velocities normalized to the
 uptake of carbon dioxide (CO₂), Biogeosciences, 2, 125-132, 10.5194/bg-2-125-2005, 2005.
- Seefeldt, L. C., Rasche, M. E., and Ensign, S. A.: Carbonyl sulfide and carbon dioxide as new substrates, and carbon disulfide as a new inhibitor, of nitrogenase, Biochemistry, 34, 5382-5389, 10.1021/bi00016a009, 1995.
- Seibt, U., Kesselmeier, J., Sandoval-Soto, L., Kuhn, U., and Berry, J.: A kinetic analysis of leaf uptake of COS and its
 relation to transpiration, photosynthesis and carbon isotope fractionation, Biogeosciences, 7, 333-341, 2010.
- Smeulders, M. J., Pol, A., Venselaar, H., Barends, T. R. M., Hermans, J., Jetten, M. S. M., and Op den Camp, H. J. M.:
 Bacterial CS<sub>2</sub> Hydrolases from <span class="named-content genus-species"
 id="named-content-1">Acidithiobacillus thiooxidans Strains Are Homologous to the
 Archaeal Catenane CS<sub>2</sub> Hydrolase, Journal of Bacteriology, 195, 4046, 10.1128/JB.0062713, 2013.
- Spielmann, F. M., Wohlfahrt, G., Hammerle, A., Kitz, F., Migliavacca, M., Alberti, G., Ibrom, A., El-Madany, T. S., Gerdel,
 K., Moreno, G., Kolle, O., Karl, T., Peressotti, A., and Delle Vedove, G.: Gross Primary Productivity of Four
 European Ecosystems Constrained by Joint CO2 and COS Flux Measurements, Geophys. Res. Lett., 0,
 10.1029/2019gl082006, 2019.
- Stimler, K., Berry, J. A., Montzka, S. A., and Yakir, D.: Association between Carbonyl Sulfide Uptake and (18)Delta during
 Gas Exchange in C-3 and C-4 Leaves, Plant Physiology, 157, 509-517, 10.1104/pp.111.176578, 2011.
- Stoy, P. C., Katul, G. G., Siqueira, M. B. S., Juang, J. Y., Novick, K. A., McCarthy, H. R., Oishi, A. C., Uebelherr, J. M.,
 Kim, H. S., and Oren, R.: Separating the effects of climate and vegetation on evapotranspiration along a
 successional chronosequence in the southeastern US, Global Change Biology, 12, 2115-2135, 10.1111/j.13652486.2006.01244.x, 2006.
- Tucker, C. J.: Red and photographic infrared linear combinations for monitoring vegetation, Remote Sensing of
 Environment, 8, 127-150, <u>https://doi.org/10.1016/0034-4257(79)90013-0</u>, 1979.

- Whelan, M. E., and Rhew, R. C.: Carbonyl sulfide produced by abiotic thermal and photodegradation of soil organic matter
 from wheat field substrate, Journal of Geophysical Research-Biogeosciences, 120, 54-62, 10.1002/2014jg002661,
 2015.
- Whelan, M. E., and Rhew, R. C.: Reduced sulfur trace gas exchange between a seasonally dry grassland and the atmosphere,
 Biogeochemistry, 128, 267-280, 10.1007/s10533-016-0207-7, 2016.
- Whelan, M. E., Lennartz, S. T., Gimeno, T. E., Wehr, R., Wohlfahrt, G., Wang, Y., Kooijmans, L. M. J., Hilton, T. W., Belviso, S., Peylin, P., Commane, R., Sun, W., Chen, H., Kuai, L., Mammarella, I., Maseyk, K., Berkelhammer, M., Li, K. F., Yakir, D., Zumkehr, A., Katayama, Y., Ogée, J., Spielmann, F. M., Kitz, F., Rastogi, B., Kesselmeier, J., Marshall, J., Erkkilä, K. M., Wingate, L., Meredith, L. K., He, W., Bunk, R., Launois, T., Vesala, T., Schmidt, J. A., Fichot, C. G., Seibt, U., Saleska, S., Saltzman, E. S., Montzka, S. A., Berry, J. A., and Campbell, J. E.: Reviews and syntheses: Carbonyl sulfide as a multi-scale tracer for carbon and water cycles, Biogeosciences, 15, 3625-3657, 10.5194/bg-15-3625-2018, 2018.
- Wohlfahrt, G., Bahn, M., Tappeiner, U., and Cernusca, A.: A multi-component, multi-species model of vegetation atmosphere CO2 and energy exchange for mountain grasslands, Agricultural and Forest Meteorology, 106, 261 287, 10.1016/s0168-1923(00)00224-0, 2001.
- Wohlfahrt, G.: Modelling fluxes and concentrations of CO2, H2O and sensible heat within and above a mountain meadow
 canopy: A comparison of three Lagrangian models and three parameterisation options for the Lagrangian time
 scale, Boundary-Layer Meteorology, 113, 43-80, 10.1023/B:BOUN.0000037326.40490.1f, 2004.
- Wohlfahrt, G., Anfang, C., Bahn, M., Haslwanter, A., Newesely, C., Schmitt, M., Drosler, M., Pfadenhauer, J., and
 Cernusca, A.: Quantifying nighttime ecosystem respiration of a meadow using eddy covariance, chambers and
 modelling, Agricultural and Forest Meteorology, 128, 141-162, 10.1016/j.agrformet.2004.11.003, 2005.
- Wohlfahrt, G., Hammerle, A., Haslwanter, A., Bahn, M., Tappeiner, U., and Cernusca, A.: Seasonal and inter-annual
 variability of the net ecosystem CO2 exchange of a temperate mountain grassland: Effects of weather and
 management, Journal of Geophysical Research: Atmospheres, 113, 10.1029/2007jd009286, 2008.
- Wohlfahrt, G., Brilli, F., Hoertnagl, L., Xu, X., Bingemer, H., Hansel, A., and Loreto, F.: Carbonyl sulfide (COS) as a tracer
 for canopy photosynthesis, transpiration and stomatal conductance: potential and limitations, Plant Cell and
 Environment, 35, 657-667, 10.1111/j.1365-3040.2011.02451.x, 2012.
- Yang, F., Qubaja, R., Tatarinov, F., Rotenberg, E., and Yakir, D.: Assessing canopy performance using carbonyl sulfide
 measurements, Global Change Biology, 24, 3486-3498, doi:10.1111/gcb.14145, 2018.
- 678