Ideas and perspectives: A strategic assessment of methane and nitrous oxide measurements in the marine environment

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- 50 Abstract. In the current era of rapid climate change, accurate characterization of climate-
- relevant gas dynamics namely production, consumption and net emissions is required for all
- 52 biomes, especially those ecosystems most susceptible to the impact of change. Marine
- environments include regions that act as net sources or sinks for numerous climate-active trace
- 54 gases including methane (CH_4) and nitrous oxide (N_2O). The temporal and spatial distributions
- of CH_4 and N_2O are controlled by the interaction of complex biogeochemical and physical
- processes. To evaluate and quantify how these mechanisms affect marine CH_4 and N_2O cycling
- 57 requires a combination of traditional scientific disciplines including oceanography,
- 58 microbiology, and numerical modeling. Fundamental to these efforts is ensuring that the
- 59 datasets produced by independent scientists are comparable and interoperable. Equally critical is
- 60 transparent communication within the research community about the technical improvements
- for required to increase our collective understanding of marine CH_4 and N_2O . An Ocean Carbon &
- 62 Biogeochemistry (OCB) sponsored workshop was organized to enhance dialogue and
- 63 collaborations pertaining to marine CH₄ and N₂O. Here, we summarize the outcomes from the
- 64 workshop to describe the challenges and opportunities for near-future CH₄ and N₂O research in
- 65 the marine environment.

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68 1. Background

The most abundant greenhouse gases in the troposphere, excluding water vapor, are carbon 69 dioxide (CO_2) , methane (CH_4) , and nitrous oxide (N_2O) . Together they account for more than 70 80% of the total radiative forcing (IPCC, 2013) and their current tropospheric mole fractions and 71 72 rates of increase are unprecedented in recent Earth history (Ciais et al., 2013; Burke et al., 2020; Fig. 1a and 1b). While CO_2 is the most abundant of the three greenhouse gases, CH_4 and N_2O 73 both have a higher warming potential than CO₂ (Montzka et al., 2011). To accurately constrain 74 the contribution of CH_4 and N_2O to Earth's radiation budget and their representation in 75 predictive models requires their sources and sinks to be quantified with high resolution at the 76 global scale. 77

78 The oceans are a fundamental component of the global climate system and are a net source of 79 tropospheric CH₄ and N₂O at the global scale, although local to regional budgets may include both source and sink components. There are far fewer marine measurements of dissolved CH_4 80 81 and N₂O than of dissolved CO₂ and while there is substantial international coordination with regard to CO₂ analysis, calibration and data reporting, no such coordination yet exists for CH₄ 82 83 and N_2O (Wilson et al. 2018). Given the increasing prominence of climate change on scientific 84 and societal agendas, greater coordination among the marine CH_4 and N_2O scientific community 85 to provide more targeted measurements and increase the quality and interoperability of CH₄ and N₂O observations is particularly timely. 86

87 Despite the lack of an international coordinating framework, there have been important advances in our understanding of marine CH₄ and N₂O in numerous research disciplines, ranging 88 89 from cellular metabolism and model microbial systems to large-scale modeling. For example, 90 recent work identified novel microorganisms and metabolic pathways in the production of N_2O (Trimmer et al., 2016; Caranto and Lancaster, 2017) and CH₄ (Repeta et al. 2016; Bižić et al., 91 2020). Earth system models now incorporate improved N_2O parameterizations to better resolve 92 the ocean's role in the global N₂O cycle (Battaglia and Joos, 2018). New techniques enable the 93 94 discrimination of ancient and modern dissolved CH₄ (Sparrow et al., 2018) and the transfer of CH₄-derived carbon to other carbon pools (Pohlman et al., 2011; Garcia-Tigreros and Kessler, 95 96 2018). Other technological and analytical advances include improved near-continuous

spectroscopic analysis that yield greater sampling resolution in surface waters (e.g. Gülzow et
al., 2011; Arévalo-Martínez et al., 2013; Erler et al., 2015) and the deployment of analytical
devices on robotic vehicles (Nicholson et al., 2018).

These scientific advances and an improvement in the quantity and quality of CH₄ and N₂O 100 observations are timely given that large areas of both the open and coastal ocean remain under-101 sampled (Fig. 1c and 1d). Limited observations contribute to uncertainty in marine CH₄ and 102 N₂O inventories, their rates of production and consumption, and their emissions. The uncertainty 103 associated with CH_4 and N_2O inventories is particularly problematic given that the marine 104 environment is susceptible to an accelerating rate of anthropogenic change that will continue to 105 modify the global cycles of carbon and nitrogen into the future. Environmental impacts on 106 marine CH₄ and N₂O distributions include increasing seawater temperatures, decreasing 107 concentrations of dissolved oxygen (O_2) , acidification, retreat of ice and mobilization of carbon 108 substrates from former permafrost, altering coastal run-off, and eutrophication (IPCC, 2019). 109 These impacts will undoubtedly alter future CH₄ and N₂O exchange with the atmosphere, but the 110 directions and magnitudes of these modified fluxes remains insufficiently understood. 111 112 The need to resolve the marine CH₄ and N₂O inventories prompted an evaluation of the collective ability of the international scientific community to accurately determine the 113 114 distribution and emissions of CH₄ and N₂O, and the determining physical-biogeochemical factors. This became the focus of a marine CH₄ and N₂O workshop hosted by the Ocean Carbon 115 116 and Biogeochemistry (OCB) program at Lake Arrowhead, California in October 2018. The workshop considered CH₄ and N₂O equally on the same agenda, even though nearly all field, 117 118 laboratory, and modeling studies examine these trace gases separately. The rationale for this dual approach is that CH₄ and N₂O share common considerations of the physical, chemical, and 119 120 microbial processes that dictate their water-column distributions (Bakker et al., 2014; Bodelier and Steenbergh, 2014). In addition, many of the analytical procedures for quantifying CH₄ and 121 N_2O and the subsequent data quality assurances share many common requirements. The 122 opportunity to bring a large research community together to increase dialogue and encourage the 123 cross-fertilization of ideas was thus considered very valuable. This article articulates the 124 125 workshop outcomes framed in the context of current marine CH₄ and N₂O research and explores

126 future research opportunities and challenges.

128 2. Coordination of oceanic CH₄ and N₂O measurements

Our understanding of the temporal and spatial distributions of oceanic CH₄ and N₂O derives 129 130 from over five decades of open ocean and coastal observations, including targeted expeditions, repeat hydrographic surveys, and time-series monitoring, each of which has been crucial to the 131 development of our current knowledge (Fig. 2). Targeted programs have enabled invaluable 132 insights into the role of oxygen deficient zones in N₂O cycling (Babbin et al., 2015; Bourbonnais 133 et al., 2017; Frey et al., 2020) and the exploration of CH₄-rich seeps and vents (Foucher et al., 134 2009; Suess, 2010; Boetius and Wenzhöfer, 2013). Basin-scale repeat hydrographic surveys 135 (e.g. the international GO-SHIP program) have facilitated extensive water-column mapping to 136 identify relevant water masses and evaluate ventilation rates (Fig. 2d) (de la Paz et al., 2017). 137 Other oceanic surveys have focused exclusively on surface sampling, using continuous 138 139 equilibrator systems connected to various gas analyzers to yield high-resolution surface concentration fields of CH₄ and N₂O (Gülzow et al., 2013; Erler et al., 2015; Kodovska et al., 140 2016; Thornton et al., 2016a; Pohlman et al., 2017). In contrast, sustained long-term time-series 141 measurements of CH_4 and N_2O at fixed monitoring stations are relatively few, but they span a 142 143 range of latitudes and biogeochemical provinces (Fig. 2a and 2b). The time-series observations provide the contextual background for seasonal and interannual variation that allow long-term 144 145 temporal trends and episodic events to be identified and evaluated (Farías et al., 2015; Wilson et al., 2017; Ma et al., 2019). Overall, the majority of measurements enable the variability in 146 147 marine CH₄ and N₂O to be quantified at the mesoscale or greater (i.e. from hundreds of kilometers to ocean basins), with monthly to annual resolution, but there are substantially fewer 148 149 datasets at the sub-mesoscale level (i.e. <10 km and hours to days) (Fig. 3). A major reason for 150 the limited sampling at the sub-mesoscale level is that it necessitates high-resolution 151 measurements to resolve the heterogeneous variability that exists at these time-space scales. 152 Such analyses have only recently become technically feasible (discussed in more detail in 153 Section 6).

Until recently there has been no formal coordination of observations across the CH_4 and N_2O scientific community. In response to this, a Scientific Committee on Oceanic Research (SCOR) Working Group was initiated in 2014 entitled: '*Dissolved N₂O and CH₄*: Working towards a global network of ocean time series measurements'. A major goal of the SCOR Working Group was to unite the international community in joint activities conceived to improve and inform

seagoing CH₄ and N₂O analyses. An important activity was the preparation and distribution of 159 160 common, combined gaseous CH_4 and N_2O standards to twelve international laboratories, with 161 the aim of improving and standardizing calibration (Bullister et al., 2017). A subsequent intercomparison of discrete seawater samples included the use of these standards and revealed the 162 variability between laboratories. While there were some encouraging results from the 163 intercomparison, such as the agreement between individual laboratories using contrasting 164 165 techniques, overall a large range was observed in CH₄ and N₂O concentration data generated by the participating laboratories (Wilson et al., 2018). Such analytical discrepancies weaken our 166 collective ability as a community to evaluate temporal-spatial variability in marine CH₄ and N₂O. 167 The discrepancies also highlighted the need for Standard Operating Protocols (SOPs) for CH₄ 168 and N₂O analyses to facilitate standardization of sampling, measurement, and calibration, as well 169 as the reporting of data and accompanying metadata in common repositories. The SOPs are 170 currently in preparation with intended publication on the Ocean Best Practices network. 171 172 A data repository for oceanic CH_4 and N_2O data known as the MarinE MEthane and NiTrous Oxide database (MEMENTO) was established in 2009 (Bange et al., 2009; Kock and Bange, 173 174 2015). MEMENTO is now sufficiently mature to support descriptions of the broad-scale surface distributions of CH₄ and N₂O (e.g. Suntharalingam et al., 2012; Zamora and Oschlies, 2014; 175 176 Buitenhuis et al., 2018; Battaglia and Joos, 2018). Machine-learning mapping also recently identified the various contributions of physical and biogeochemical predictor variables for CH₄ 177 178 (e.g. depth, primary production; Weber et al., 2019; Fig 4b) and N₂O distributions (e.g. chlorophyll, sea surface temperature, apparent oxygen utilization, and mixed-layer depth; Yang 179 180 et al., 2020; Fig. 4a). The application of gas transfer algorithms to the extrapolated oceanic CH_4 and N₂O distributions helped decrease the uncertainty in estimates of global air-sea exchange 181 182 fluxes (Fig. 4c), thereby fulfilling one of the key goals of MEMENTO (Bange et al., 2009). Net global open ocean emissions of N₂O are now similarly estimated at 3–5 Tg N yr⁻¹ by both Yang 183 184 et al. (2020) and the Global Nitrous Oxide Project (Tian et al., 2020). In comparison, net global ocean CH₄ emissions from machine-learning mapping were estimated at 6-12 Tg CH₄ yr⁻¹ 185 (Weber et al., 2019), compared to 9-22 Tg CH₄ yr⁻¹ in the most up-to-date CH₄ synthesis 186 187 (Saunois et al., 2020). However, the narrower range for machine-learning derived CH_4 emissions retains high uncertainty in regions such as the Arctic, where emissions are highly 188 189 heterogeneous and compounded by seasonal ice cover. Identifying the causes for uncertainty in

190 high emission regions will greatly aid future sampling campaigns, as is discussed in the

191 following sections.

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3. Methane in marine environments

In the surface waters of tropical and temperate oceans, a number of factors contribute to the 194 low supersaturation of CH₄ including direct aerobic production arising from the degradation of 195 methylated sulfur compounds by phytoplankton (Klintzsch et al., 2019) and methyl phosphonate 196 in phosphorus-depleted waters (Karl et al. 2008, Sosa et al., 2020), indirect production via 197 grazing (Schmale et al., 2018) and abiotic photoproduction (Li et al., 2020). A recent study 198 199 demonstrated that CH₄ production by cyanobacteria is linked to general cell metabolism and does not rely on the presence of methylated precursor compounds (Bižić et al., 2020). Deep 200 201 within the ocean's pelagic interior, CH₄ is weakly undersaturated reflecting depletion via microbial oxidation (Reeburgh 2007; Weber et al., 2019). Towards the coastline, CH_4 202 supersaturation increases by orders of magnitude (Figure 5b), reflecting terrestrial inputs (e.g. 203 river and groundwater), increased organic matter loading (Borges et al., 2018), and CH_4 204 205 diffusion and ebullition from shallow anoxic methane rich sediments (Zhang et al., 2008; Borges et al., 2016; Upstill-Goddard and Barnes, 2016). Supersaturation of CH₄ occurs frequently in the 206 207 Arctic Ocean and its relatively shallow marginal seas with the most extreme values observed in the Eurasian Arctic (e.g. Shakhova et al., 2010; Damm et al., 2015; Kosmach et al., 2015; 208 209 Thornton et al, 2016a; Lorensen et al., 2016; Fenwick et al., 2017; Lapham et al., 2017). Terrestrial and subsea permafrost are potential CH₄ sources to shelf waters in addition to CH₄ 210 211 hydrates that are found in marginal shelves globally (Ruppel and Kessler, 2017). Large point source CH₄ emissions, such as seafloor gas seeps can be large sources to the atmosphere in small 212 213 localized areas (e.g. Thornton et al., 2020), but these sites remain particularly difficult to 214 parameterize in models. This reflects limited observations and a poor understanding of their spatial distributions, the driving mechanisms, and the wider context within the carbon cycle. For 215 216 example, the upwelling of cold, nutrient-rich water that accompanies CH₄ ascending the water column stimulates CO₂ consumption by photosynthesizing phytoplankton, rendering such CH₄ 217 218 seeps an overall net sink for climate-forcing gases (Pohlman et al., 2017). Recent work using thermal infrared satellite retrievals indicates increased high-latitude oceanic CH₄ release in late 219 220 autumn, coincident with pycnocline breakdown and a deepening of the ocean mixed layer depth

221 thereby bringing deep CH₄ to the surface (Yurganov et al., 2019). This is especially notable in 222 the Kara and Barents Seas, but the remote observations have not yet been confirmed by surface 223 ocean measurements which are difficult and therefore rare, except during the Arctic summer. 224 Seabed CH_4 emissions are hypothesized to increase in a warming ocean through the decomposition of gas hydrates, the degradation of subsea permafrost under some high-latitude 225 seas, and the increased biodegradation of sediment carbon (Romanovskii et al., 2005; Biastoch et 226 227 al., 2011; Lapham et al., 2013; Ruppel and Kessler, 2017; Borges et al., 2019). Effort is thus focused on quantifying the fraction of CH_4 generated in or released from marine sediments that 228 ultimately enters the atmosphere, particularly on shallow continental shelves and in coastal 229 230 ecosystems. Natural stable isotopes have been used to inform spatial and temporal changes in dissolved CH₄ concentrations (e.g. Pack et al., 2011; Mau et al., 2012; Weinstein et al., 2016; 231 232 Leonte et al., 2017; Chan et al., 2019) and incubation experiments with added stable isotopes and radiotracers have helped elucidate how oxidation (anaerobically in sediments and aerobically in 233 234 the water column), ebullition (where CH₄ pore water partial pressure exceeds sediment hydrostatic pressure), and subsequent bubble dissolution in the water column interact to mitigate 235 236 CH₄ emissions to air (Steinle et al., 2015; Jordan et al., 2020). The information deriving from these various approaches is inherently different but complementary. Isotope tracer incubations 237 238 provide snapshots of rates specific to the methanotrophic community and CH_4 concentration at the time of sampling, whereas concentrations and isotopic gradients are used to infer *in situ* rates 239 240 integrated over space and time. A recent study deployed a remotely operated vehicle to examine the isotopic fractionation of CH_4 during bubble ascent and used this to constrain the extent of 241 242 bubble dissolution (Leonte et al., 2018). This work demonstrated an experimental approach established for broadly constraining water column CH₄ cycling directly from a surface research 243 244 vessel.

Despite the range of analytical and experimental approaches available, determining whether the origin of the emitted CH₄ is seafloor release or aerobic production in the upper water column remains problematic. To date there is no straightforward way to routinely distinguish between seafloor derived and water column generated CH₄ for all locations. Even so, stable carbon and hydrogen isotope measurements (i.e. δ^{13} C-CH₄ and δ^{2} H-CH₄) combined with ancillary data may provide valuable source information. For example, combining these measurements with the ratio of CH₄ to higher order hydrocarbons (e.g. ethene (C₂H₄) and ethane (C₂H₆)) can be used to infer

252 for example, whether the origin of the CH_4 is thermogenic, sub-seafloor, or biogenic within the 253 water column (Whiticar, 1999; Pohlman et al., 2009; Lan et al., 2019). Continuous shipboard 254 measurement of CH₄ isotopes in surface water (e.g. Pohlman et al., 2017) and in the atmospheric boundary layer (Pankratova et al., 2019; Berchet et al., 2020) are now possible and they have 255 been used in combination with atmospheric inversion models to characterize and discriminate 256 marine-emitted CH₄ from other sources (Berchet et al., 2020). Application of this method to 257 258 land-based monitoring stations appears promising for apportioning CH₄ emissions from various 259 marine regions and sources (Thonat et al., 2019). Additionally, in regions where aerobic CH_4 oxidation is substantial, the resulting isotopic fractionation generates measurable vertical and/or 260 261 horizontal seawater gradients that can also be used to identify contrasting biogenic CH₄ sources (Leonte et al., 2020). However, the general overlap in isotope compositions of sediment CH₄ 262 (e.g. Thornton et al., 2016b; Sapart et al., 2017) can complicate purely isotope-based 263

264 determinations of sources.

Measurements of the natural radiocarbon content of dissolved oceanic CH_4 , while being 265 highly specialized and requiring substantial amounts of ship time and processing (Kessler and 266 267 Reeburgh, 2005; Sparrow and Kessler, 2017), provide valuable source information because the ¹⁴C-CH₄ measurements are normalized to the same δ^{13} C value and are unaffected by the extent 268 of oxidation. The bubbles sampled from hydrate and active seafloor seeps are largely devoid of 269 270 radiocarbon (Pohlman et al., 2009; Kessler et al., 2008; Douglas et al., 2016). However, CH_4 in sediments can also be derived from more modern or recently deposited organic material and an 271 272 exact determination of individual contributions is hard to achieve (Kessler et al., 2008; Sparrow et al., 2018). The powerful insights made by radiocarbon-CH₄ investigations would be further 273 274 strengthened by concurrent sampling of other analytes that offer CH₄ source information, such as clumped isotopes. Isotope clumping, the co-occurrence of two or more of the less abundant 275 isotopes in a molecule (e.g. ¹³C and ²H or ¹H and ²H), provides unique information on marine 276 CH₄ sources (Stolper et al., 2014; Wang et al, 2015; Douglas et al., 2017; Young et al., 2017; 277 278 Labidi et al., 2020). In this approach, the isotopic deviations in samples from their random 279 probability distributions can give insight into formation temperature and the extent of 280 biochemical disequilibrium. However, the sample size required for a clumped isotope analysis in the oceanic environment away from areas of seafloor emission is large and exceeds the 281 already demanding volume requirements for ¹⁴C analyses by 1–2 orders of magnitude (Douglas 282

et al., 2017). While the requirement of large sample size and lengthy measurement time
currently preclude their more widespread application, clumped isotope measurements offer
future promise in refining our understanding of the processes of marine CH₄ production and
consumption.

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4. Nitrous oxide in marine environments

289 The large-scale spatial distribution of N₂O in the global ocean is reasonably well-established. The highest open ocean N₂O values are in upwelling environments, where concentrations extend 290 up to micromolar levels (Arévalo-Martínez et al., 2015) and production rates can be as high as 291 120 nM d^{-1} (Frey et al., 2020). The highly elevated N₂O concentrations can be proximal to 292 regions with some of the lowest recorded N₂O concentrations, in the cores of O₂ deficient zones. 293 This coexistence of the highest and lowest observed N₂O concentrations over vertical distances 294 of tens of meters make upwelling regions a focal point for N₂O research, particularly since O₂ 295 deficient ocean zones are increasing in size (Stramma et al., 2011). In contrast, in the surface 296 297 waters of the expansive oligotrophic ocean gyres, N_2O is weakly supersaturated (103-105%) 298 with respect to atmospheric equilibrium (Weiss et al., 1992; Wilson et al., 2017, Charpentier et al., 2010). Nitrous oxide becomes more highly saturated in the surface waters of equatorial 299 300 upwelling regions due to the upward advection of N₂O-rich waters (Arévalo-Martínez et al., 2017). For the Arctic Ocean, the data indicate low net N₂O emissions, with some areas acting as 301 302 net N₂O sources and others as N₂O sinks (Fenwick et al., 2017, Zhang et al., 2015). Several parameters control net N₂O emissions from the ocean, including temperature, 303 salinity, dissolved O₂, apparent oxygen utilization (AOU), nutrients, and microbial community 304 305 abundance and composition. A recent modeling study trained with just three of these variables 306 (chlorophyll, O₂, and AOU) accounted for 60% of the observed variability in oceanic N₂O 307 concentrations (Yang et al., 2020; Fig. 5a), highlighting the importance of N_2O in productive upwelling systems. Correlations between N₂O and environmental variables provide some insight 308 into the factors controlling its distribution, but they provide no information about the 309 310 microorganisms or metabolic pathways involved. Microbial production of N₂O occurs during 311 the metabolic processes of nitrification and denitrification (Stein and Yung, 2003). To determine which process dominates N₂O production at any given location requires the application of 312 multiple methodological approaches, ideally in parallel. 313

314 One of the most commonly used approaches is the incubation of discrete water samples under *in situ* conditions with stable isotope (15 N) addition such as 15 N enriched NH₄⁺, NO₂⁻ or 315 NO_3^- to measure N₂O production rates from nitrification and denitrification (e.g. Ji et al., 2017). 316 These approaches also provide insight into the microorganisms involved. For example, N₂O 317 resulting from archaeal NH_4^+ oxidation is mostly formed from a combination of NH_4^+ and 318 another N compound (e.g. NO_2^{-}) whereas bacteria produce N₂O from NH_4^{+} alone (Santoro et al., 319 320 2011, Stieglmeier et al., 2014; Carini et al. 2018; Lancaster et al., 2018; Frey et al. 2020). Unfortunately, as with all incubation-based approaches ¹⁵N techniques are subject to bottle 321 artifacts, and the strong dependence of N₂O production and consumption on ambient O₂ 322 323 increases the potential for contamination during the collection and manipulation of anoxic deep seawaters. Incubation based rate measurements are also compromised by abiotic N₂O 324 production via chemodenitrification, specifically the reduction of NO₂⁻ coupled to Fe²⁺ 325 oxidation, as observed in high Fe environments (Ostrom et al., 2016; Buchwald et al., 2016; 326 Wankel et al., 2017). These issues highlight the need for incubation techniques that mitigate the 327 effect of experimental artifacts (Stewart et al., 2012). 328

329 In addition to isotope addition and incubation, natural abundance water-column measurements of N₂O concentrations, isotopes, and isotopomers yield valuable rate and process 330 331 information. These measurements are free from experimental artifacts and can be used to integrate over appropriate temporal and spatial scales. For example, nitrification in sunlit waters 332 333 has been inferred from N₂O distributions (Dore and Karl, 1996), and N₂O production close to the ocean surface is a large contributor to the uncertainty in oceanic N₂O emissions (Ward et al., 334 335 1982; Zamora and Oeschlies, 2014). Isotopomers are isomers having the same number of each isotope of each element but differing in their structural positions. Nitrous oxide isotopomers are 336 337 increasingly used, sometimes in combination with box models, to estimate the rates of different N₂O production pathways, in the upwelling systems off southern Africa (Frame et al., 2014) and 338 Peru (Bourbonnais et al., 2017). There is however some disagreement about whether isotopomer 339 signatures are robust indicators of the formation pathway (Yoshida and Toyoda, 2000; Sutka et 340 al., 2006) or whether there is fractionation during production (Schmidt et al., 2004; Casciotti et 341 342 al., 2018). Greater clarity is therefore required in the use of N_2O isotopes and isotopomers to infer metabolic pathways of N₂O formation. Notwithstanding this issue, field measurements of 343 N₂O isotopes and/or isotopomers have the potential to greatly increase current experimental 344

capabilities and robustness (Yu et al., 2020). However, the development of spectroscopic gas analysis systems that have been so advantageous to CH_4 research has been slower for N₂O. This is due to the higher costs and the increased complexity of the laser systems, although progress is being made to improve instrumental precision, and to decrease matrix effects and spectral interferences (e.g. Harris et al., 2019).

A better understanding of the microorganisms responsible for N₂O production and 350 351 consumption is fundamental to deriving more accurate estimates of process rates. For example, the metabolic activity of ammonia oxidizing archaea can exceed that of ammonia oxidizing 352 bacteria in the ocean (Santoro et al., 2010; Löscher et al., 2012; Fuchsman et al., 2017). The 353 354 differing sensitivities of these archaea and bacteria to dissolved O_2 (Stahl and de la Torre, 2012; Hink et al., 2017) are a critical factor in evaluating the microbial response to changing 355 356 environmental conditions, as shown for the terrestrial environment (Prosser at al., 2020). Therefore, to understand the impact of deoxygenation on oceanic N₂O emission requires a better 357 358 understanding of both archaeal and bacterial metabolisms and their environmental niches. Fieldbased sequencing not only characterizes the community but can highlight potential metabolic 359 360 pathways when they might not otherwise be inferred. For example, transcripts encoding for N₂O consumption (nosZ) have repeatedly been identified in the oxic water column, despite 361 362 denitrification being an anaerobic metabolic process (Wyman et al., 2013; Sun et al., 2017). The transcription of nosZ has been also located in highly dynamic O₂ permeable coastal sediments 363 364 (Marchant et al., 2017). Denitrification under aerobic conditions is attributed to fluctuations in O_2 , NO_3^- , organic matter and other parameters that affect the availability of electron donors and 365 366 acceptors which ultimately influences whether a coastal environment is a net source or sink of N₂O, as discussed in the next section. 367

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5. CH₄ and N₂O in shallow marine environments

Coastal and other shallow (<50 m) marine systems are globally relevant CH₄ and N₂O source regions. However, their emission rates to the atmosphere are weakly constrained in comparison with the open ocean. Several factors contribute to the uncertainty, including the high diversity of coastal and shallow marine ecosystems and lack of consistency in adequately defining them, locally heterogeneous conditions causing strong spatial and temporal concentration gradients, highly uncertain spatial distribution of CH₄ seeps, a bias towards studies in the northern

hemisphere, and incomplete or sometimes inappropriate sampling strategies (Al-Haj and 376 377 Fulweiler, 2020). Until these issues are resolved it will remain difficult to adequately define the 378 contribution from shallow marine systems to global CH₄ and N₂O budgets. An important illustration of this is reflected in the prevailing view that large geological sources (e.g. seeps, 379 380 mud volcanoes, and hydrates) are the main contributors to marine CH_4 emissions (Ciais et al., 2013). The most recent modeled estimate of global marine CH_4 emissions (6–12 Tg CH_4 vr⁻¹) 381 382 reported that near-shore environments (depths of 0-50 m) contribute a large and highly uncertain diffusive flux (Weber et al., 2019). A study of coastal ecosystems, in this case defined as shelf, 383 estuarine, and tidally influenced rivers, estimated them to contribute 7 Tg CH_4 yr⁻¹ (Anderson et 384 al., 2010) while another estimated 1–7 Tg CH₄ yr⁻¹ for estuaries alone (Borges and Abril, 2011). 385 Similar uncertainties exist for N₂O. Estimates of coastal N₂O emissions (which include coastal, 386 estuarine, and riverine sources) range from 0.1–2.9 Tg N yr⁻¹ (Ciais et al., 2013), although a 387 recent review of N₂O production across a range of estuarine habitats placed N₂O fluxes at the 388 lower end of these estimates (0.17–0.95 Tg N yr⁻¹) (Murray et al., 2015). Based on these data, 389 coastal systems account for around one third of total marine N₂O emissions (Yang et al., 2020). 390 391 The direct quantification of CH₄ and N₂O emissions from shallow coastal ecosystems has historically involved using gas concentrations measured in discrete water and air samples 392 393 combined with a gas transfer velocity (k_w) . For the coastal and open ocean, the dominant driver of gas exchange is wind speed (e.g. Nightingale et al., 2000; Wanninkhof, 2014) whereas in 394 395 nearshore, shallow water environments the interaction of water, depth, and tidal current speeds may be a major contributor to near surface turbulence. Several k_w parameterizations are now in 396 397 use for coastal waters (e.g. Raymond and Cole 2001; Kremer et al., 2003; Zappa et al., 2003; Borges and Abril, 2011; Ho et al. 2011; Rosentreter et al., 2017; Jeffrey et al., 2018) which 398 399 increases the uncertainties associated with CH₄ and N₂O emissions. For example, a fivefold variation in CH₄ emissions from a single system occurred when applying different 400 401 parameterizations to the measured gradients in CH₄ (Ferrón et al., 2007). To constrain emissions over small areas, continuous air-sea fluxes can be measured using 402 403 free-floating chambers (e.g. Bahlmann et al., 2015; Rosentreter et al., 2018; Yang et al., 2018; 404 Murray et al., 2020), but issues related to turbulence modification may still generate flux artifacts (Upstill-Goddard, 2006). To overcome these problems in the future, a greater reliance on direct 405

406 and robust continuous techniques for air-sea flux measurement, such as eddy covariance (e.g.

407 Podgrajsek et al., 2016), that avoid any need for k_w , will be necessary. Eddy-covariance measurements also capture both diffusive and ebullitive flux components (Thornton et al., 2020). 408 409 Combining this approach with new analytical techniques such as cavity enhanced absorption spectroscopy (CEAS) and non-dispersive infrared (NDIR) should continue to improve the 410 quality of CH₄ and N₂O flux estimates (McDermitt et al., 2011; Nemitz et al., 2018; Maher et al., 411 2019). Indeed, eddy flux towers aboard ships (Thornton et al., 2020) and in coastal locations 412 (Yang et al., 2016; Gutiérrez-Loza et al., 2019) are now being equipped with CH₄ 413 instrumentation that enables the integration of CH₄ fluxes over large areas. There are fewer N₂O 414 flux estimates made with CEAS and NDIR and the implementation of N₂O sensors on eddy flux 415 towers remains limited. Recently, N₂O emissions from Eastern Boundary Upwelling Systems 416 were quantified using inversion modeling based on atmospheric measurements from coastal 417 418 monitoring stations highlighting the potential of this approach to constrain N₂O emissions from remote oceanographic regions that have significant spatial and temporal heterogeneity (Ganesan 419 et al., 2020; Babbin et al., 2020). Inverse modeling of atmospheric measurements was also 420 421 recently used to constrain CH₄ emissions from the East Siberian Arctic Shelf (Tohjima et al., 2020) 422

Coastal measurements of CH₄ and N₂O also require the collection of ancillary data such as 423 424 water-column depth, tidal motions (Rosentreter et al., 2018; Huang et al., 2019; Pfeiffer-Hebert et al., 2019), and other information relating to diel processes (Maher et al., 2016). Such data are 425 426 important because for example, the magnitude of CH₄ and N₂O fluxes vary over a diel period depending on the redox environment as a result of tidal effects and changes in inorganic N and 427 428 O₂ availability (Seitzinger and Kroeze, 1998; Call et al., 2015; Vieillard and Fulweiler, 2014; Maher et al., 2015; Murray et al., 2015; Foster and Fulweiler, 2019). The magnitude of CH₄ and 429 430 N₂O fluxes also varies over longer temporal scales (seasonally to yearly) due to additional factors such as groundwater inputs, adjacent land-use, dissolved O₂, organic matter content and 431 quality, and macrofaunal distributions (Barnes and Upstill-Goddard, 2011; Upstill-Goddard and 432 Barnes, 2016; Gelesh et al., 2016; Bonaglia et al., 2017; Borges et al., 2018; Wells et al., 2018; 433 Ray et al., 2019; Al-Haj and Fulweiler, 2020; Reading et al., 2020). To determine the 434 435 contributing factors and resolve the spatial distributions, mobile sampling platforms such as small vessels (Müller et al., 2016; Brase et al., 2017; Tait et al., 2017), and autonomous vehicles 436 437 (Manning et al., 2019) are essential. Recent improvements in gas sensors and in technology such

- 438 as sonar and ebullition sensors will further increase our ability to measure dynamic fluxes
- (Maher et al., 2019; Lohrberg et al., 2020). Improvements to the quality and quantity of CH₄ and
- N_2O measurements in coastal systems will enable the development of iterative forecast models,
- 441 further improving estimates of global coastal CH₄ and N₂O fluxes.
- 442

443 **6.** Leveraging culture studies to further our ecosystem understanding

A more complete understanding of marine CH₄ and N₂O necessitates closer integration between 444 biogeochemistry, model requirements, and targeted microbiological studies involving both single 445 microorganism isolates and enrichment cultures. Marine CH₄ and N₂O budgets deriving from 446 both 'bottom-up' (e.g. emissions inventories, ocean and terrestrial process models) and 'top-447 down' (e.g. inverse analyses of atmospheric trace-gas measurements) approaches would greatly 448 449 benefit from more highly constrained metabolic processes. Specifically, this includes rates of CH₄ or N₂O production and consumption for key model microorganisms, and the kinetic 450 parameters associated with these metabolic rates. Reliable inventories of key microbially 451 mediated process rates will improve the robustness of Earth System models used for predicting 452 453 climate-mediated changes to marine CH₄ and N₂O emissions.

For N₂O, laboratory studies quantifying microbial process rates, such as for nitrification and 454 455 denitrification, are relatively few (e.g. Frame and Casciotti 2010; Santoro et al. 2011; Löscher et al. 2012; Ji et al. 2015; Qin et al., 2017). Consequently, models largely continue to use process 456 457 rates optimized using water column concentrations of N₂O, O₂, and related nitrogen cycle quantities (e.g. Battaglia and Joos, 2018; Buitenhuis et al., 2018; Landolfi et al., 2017). Future 458 459 model parameterizations for N₂O will require information on the variability of microbial process yields derived from culture studies with controlled varying conditions of O₂ (Goreau et al. 1980, 460 461 Frame and Casciotti 2010, Löscher et al. 2012; Ji et al., 2018), pH (Breider et al., 2019; Hopkins et al. 2020), temperature, and nutrients. Automated incubation systems have measured N₂O 462 production kinetics and yield as functions of the concentrations of O_2 and total ammonia nitrogen 463 (Molstad et al., 2007; Hink et al., 2017). Quantifying the physiology of relevant microorganisms 464 and connecting them to environmental characteristics will provide insights into why, for 465 466 example, some shallow marine habitats act as N_2O sinks while others are N_2O sources, or how N₂O is produced in well oxygenated open-ocean waters, as compared to oxygen deficient zones. 467

468 For CH_4 , a key requirement to relate *in situ* CH_4 production with transport to atmospheric emissions is our ability to accurately determine rates of CH_4 oxidation. Fundamental issues 469 470 include the challenges of cultivating methanotrophs and of replicating environmental conditions 471 such as pressure and the chemistry of CH_4 gas bubbles. The increased emphasis on CH_4 dynamics in shallow water environments highlighted in Section 5, must be supported by culture-472 based measurements of CH₄ oxidation that control for temperature, O₂ and other important 473 474 variables. In comparison to CH₄ oxidation, culture-based studies are used increasingly to identify organisms capable of aerobic CH_4 production and their underlying metabolic pathways 475 (Carini et al., 2014; Klintzsch et al; 2019; Bižić et al., 2020). 476

477 Specific cellular yields and consumption rates of CH_4 and N_2O are not the sole objective of culturing experiments. Cultivation of microorganisms involved in CH₄ and N₂O production and 478 consumption provides vital information into the physiology, metabolism, and interactions of 479 environmentally relevant clades. When combined with genomic approaches, insights can 480 481 therefore be gained into the diversity and global distribution of organisms involved in CH₄ and N_2O cycling. For CH₄ some unexpected physiologies have been revealed (Ettwig et al., 2010; 482 483 Haroon et al., 2013; Ettwig et al., 2016), which has directed research into sources and sinks of CH₄ in the natural environment. Similarly, our understanding of how and when ammonia 484 485 oxidizers produce N₂O has been facilitated by studies of cultured nitrifiers and detailed analysis of their biochemistry (Stahl and de la Torre, 2012; Caranto and Lancaster, 2017). Recent 486 487 combinations of cultivation studies with environmental genomics, albeit largely for terrestrial systems, have revealed a variety of denitrifiers, many of which are only involved in specific 488 489 denitrification steps (Ganesh et al., 2014; Lycus et al, 2017; Hallin et al, 2018; Marchant et al., 490 2018; Conthe et al, 2019).

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492 **7.** Outlook and priorities for marine CH₄ and N₂O measurements

This perspectives article has assessed the collective ability of the scientific community to determine the spatial variability of marine CH_4 and N_2O distributions, the underlying mechanisms that determine this variability, and the resulting sea-to-air emissions. Shallow marine environments and oxygen deficient zones are widely recognized as deserving of greater attention because they have high CH_4 and N_2O concentrations with inherently high uncertainties that complicate any assessment of their emissions to air (Bange et al., 1994; Bange et al., 1996;

499 Bakker et al., 2014; James et al., 2016; Borges et al., 2016; Tian et al., 2020). Fortunately, recent 500 technological advances that have increased our ability to conduct high-resolution measurements 501 allow an optimistic outlook for making substantial progress in quantifying the CH₄ and N₂O budgets of these ecosystems. Even so, the inherent complexity of shallow marine environments 502 clearly warrants a strategically coordinated approach to optimize the value of future studies. 503 Issues to consider include identifying the locations of complementary sampling sites, 504 505 standardizing sampling strategies and techniques, and agreeing the use of common ancillary measurements that set the broad biogeochemical context (Bange et al., 2019). In contrast to the 506 open ocean, measurement campaigns in shallow water environments are amenable to the use of 507 eddy covariance flux towers, and they have the potential to lever resources from existing 508 observation networks, which in North America include the Long-Term Ecological Research 509 510 network (LTER) and the National Estuarine Research Reserve (NERR) System (Novick et al., 2018). Indeed, such activities are already underway; an increasing number of flux towers are 511 being equipped for CH₄ measurements (Torn et al., 2019) and future efforts should focus on the 512 inclusion of N_2O (see Section 5). 513

514 We are encouraged that the Global Carbon Project with its objective of developing a complete picture of the global carbon cycle including interactions and feedbacks has expanded to 515 516 include CH_4 (Saunois et al., 2020) and is now incorporating N₂O (Tian et al., 2020). These Projects compile the most recent data from peer-reviewed analyses of the sources and sinks of 517 518 atmospheric CH₄ and N₂O from both natural and human activities. For example, the aquatic components of the recent Global Carbon Project N₂O budget reported emissions from the open 519 520 ocean, inland waters, estuaries and coastal zones. Low-oxygen oceanic regions associated with eastern-boundary upwelling zones, and the coastal ocean were identified as key regions with 521 522 significant N₂O variability requiring more detailed assessment via measurement campaigns and 523 model analyses (Tian et al., 2020). Contribution to the Global Carbon Project and similar 524 initiatives will identify areas of synergistic CH₄ and N₂O research for oceanographers and other Earth observation scientists (Ganesan et al., 2019). Furthermore, as highlighted in Section 6, 525 field observations alone are insufficient to improve the robustness of Earth System models and 526 527 leveraging laboratory-based microbial process studies is highly recommended.

The success of any coordinated CH_4 and N_2O research program relies heavily on having uniformly high confidence in the various resulting datasets and their interoperability, and we identify three key initiatives that are paramount to ensuring this:

531 (i) The first is to develop and adopt Standard Operating Protocols (SOPs) to help obtain intercomparable CH₄ and N₂O datasets of the highest possible accuracy and precision. 532 Currently, there is no community-defined level of analytical uncertainty to characterize high 533 534 quality CH₄ and N₂O measurements. However, attaining an analytical agreement between multiple laboratories of $\leq 1\%$ is considered achievable for the repeat oceanographic surveys and 535 time-series observations (Fig. 3). For context, an analytical agreement of $\leq 1\%$ would permit the 536 537 ocean's response to the increasing tropospheric CH_4 and N_2O mole fractions to be resolved on timescales of 10 and 5 years, respectively. These values are based on the changes in surface 538 539 ocean CH₄ and N₂O concentrations that are predicted to occur due to the ongoing increase in tropospheric CH₄ and N₂O mole fractions at a seawater temperature of 20°C and a salinity of 35 540 g kg⁻¹, and assuming all sources and sinks remaining constant. In our recent marine CH_4 and 541 N₂O inter comparison exercise we concluded that the diversity of analytical procedures 542 543 employed by the participants was a major cause of high variability between the reported concentrations, highlighting an urgent requirement for CH₄ and N₂O SOPs (Wilson et al., 2018). 544 545 Consequently, these SOPs are now being compiled, and they will be freely available via the Ocean Best Practices System. 546

547 (ii) The second is increased regularity of intercomparison exercises through the periodic 548 distribution of consensus material, i.e. water samples in which CH_4 and N_2O concentrations are 549 known with high confidence, obtained by pooling analyses from several laboratories with 550 demonstrated analytical capability. These will help the scientific community to monitor data 551 comparability and accuracy, particularly in the case of highly elevated concentrations of CH_4 and 552 N_2O , i.e. those exceeding atmospheric equilibrium concentrations by at least an order of 553 magnitude.

(iii) The third activity builds on the previous initiative and calls for the production of Global Data Products for dissolved CH_4 and N_2O measurements. To date, individual CH_4 and N_2O measurements are represented at the global scale by the MEMENTO database which has been very successful at compiling CH_4 and N_2O datasets and making them readily accessible to the modeling community. However, the MEMENTO database does not currently include a Global

559 Data Product that includes publicly accessible quality controlled dissolved CH₄ and N₂O 560 datasets. The international marine carbon science community has widely embraced such an 561 approach for fCO₂, by submitting data to the Surface Ocean CO₂ Atlas (SOCAT), which was initiated in response to the need for a quality controlled, publicly available, global surface CO₂ 562 dataset (e.g. Bakker et al., 2016). Due to the fewer measurements, a similar data product for 563 marine CH₄ and N₂O would be needed every ~5 years. We consider the production of Global 564 565 Data Products for dissolved CH₄ and N₂O to be essential for supporting future global modeling efforts and to enhance field observations. 566

The benefits of pursuing the three activities described above have already been clearly demonstrated for carbon system measurements in the ocean. The intercomparability and high accuracy and precision of carbon system measurements was achieved by streamlining methodological approaches, universally adopting agreed SOPs, production of reference material, and following community-driven quality control procedures (Dickson et al., 2007, Dickson et al, 2010). It is encouraging to see the marine CH₄ and N₂O community beginning to move in a similar direction.

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589 **References**

- Al-Haj, A. N. and Fulweiler, R. W.: A synthesis of methane emissions from shallow vegetated
 coastal ecosystems, Glob. Change Biol., https://doi.org/10.1111/gcb.15046, 2020.
- Anderson, B., Bartlett, K., Frolking, S., Hayhoe, K., Jenkins, J. and Salas, W.: Methane and
 nitrous oxide emissions from natural sources, Office of Atmospheric Programs, US EPA,
 EPA 430-R-10-001, Washington DC, 2010.
- 595 Arévalo–Martínez, D. L., Beyer, M., Krumbholz, M., Piller, I., Kock, A., Steinhoff, T.,
- 596 Körtzinger, A., and Bange, H. W.: A new method for continuous measurements of oceanic
- and atmospheric N_2O , CO and CO_2 : Performance of off-axis integrated cavity output
- spectroscopy (OA-ICOS) coupled to non-dispersive infrared detection (NDIR), Ocean Sci.,
 9, 1071–1087, 2013.
- Arevalo–Martínez, D. L., Kock, A., Löscher, C. R., Schmitz, R. A. and Bange, H. W.: Massive
 nitrous oxide emissions from the tropical South Pacific Ocean, Nat. Geoscience, 8, 530,
 2015.
- Arévalo-Martínez, D. L., Kock, A., Steinhoff, T., Brandt, P., Dengler, M., Fischer, T.,
- Körtzinger, A. and Bange, H. W.: Nitrous oxide during the onset of the Atlantic cold tongue,
 J. Geophys. Res.: Oceans, 122,171–184, 2017.
- Babbin, A. R., Bianchi, D., Jayakumar, A., and Ward, B. B.: Rapid nitrous oxide cycling in the
 suboxic ocean, Science, 348, 1127–1129, 2015.
- Babbin, A. R., Boles, E. L., Mühle, J. and Weiss, R. F.: On the natural spatio-temporal
 heterogeneity of South Pacific nitrous oxide, Nat. Comm., 11, 1–9, 2020.
- Bahlmann, E., Weinberg, I., Lavrič, J. V., Eckhardt, T., Michaelis, W., Santos, R. and Seifert, R.:
 Tidal controls on trace gas dynamics in a seagrass meadow of the Ria Formosa lagoon
 (southern Portugal), Biogeosciences 12:1683–1696, 2015.
- Bakker, D. C. E., Bange, H. W., Gruber, N., Johannessen, T., Upstill-Goddard, R. C., Borges, A.
 V., Delille, B., Loscher, C. R., Naqvi, S. W. A., Omar, A. M. and Santana-Casiano, M.: Airsea interactions of natural long-lived greenhouse gases (CO₂, N₂O, CH₄) in a changing
- climate. In: Liss, P., Johnson, M, ed. Ocean-Atmosphere Interactions of Gases and Particles.
 Berlin: Springer-Verlag, 113–169, 2014.
- Bakker, D. C., Pfeil, B., Landa, C. S., Metzl, N., O'brien, K. M., Olsen, A., Smith, K., Cosca, C.,
 Harasawa, S., Jones, S.D. and Nakaoka, S. I.: A multi-decade record of high-quality fCO(2)
 data in version 3 of the Surface Ocean CO2 Atlas (SOCAT), Earth Sys.Science Data, 8, 383–
 413, 2016.
- Bange, H. W., Bartell, U. H., Rapsomanikis, S., and Andreae, M. O.: Methane in the Baltic and
 North Seas and a reassessment of the marine emissions of methane, Global Biogeochem. Cy.,
 8, 465–480, 1994.
- Bange, H. W., Rapsomanikis, S., and Andreae, M. O.: Nitrous oxide in coastal waters, Global
 Biogeochem. Cy., 10, 197–207, 1996.
- Bange, H. W., Bell, T. G., Cornejo, M., Freing, A., Uher, G., Upstill-Goddard, R. C. and Zhang,
 G.: MEMENTO: a proposal to develop a database of marine nitrous oxide and methane
 measurements, Environ. Chem., 6, 195–197, 2009.
- Bange, H. W., Arévalo-Martínez, D. L., de la Paz, M., Farías, L., Kaiser, J., Kock, A., Law, C.
- 631 S., Rees, A. P., Rehder, G., Tortell, P. D., Upstill-Goddard, R. C., and Wilson, S. T.: A
- harmonized nitrous oxide (N_2O) ocean observation network for the 21st Century, Front. Mar.
- 633 Sci., 6:157. doi: 10.3389/fmars.2019.00157, 2019.

- Barnes, J. and Upstill-Goddard, R.C.: N₂O seasonal distributions and air-sea exchange in UK
 estuaries: Implications for the tropospheric N₂O source from European coastal waters, J.
 Geophys. Res.:-Biogeosci., 116(G1), 2011.
- Battaglia, G. and Joos, F.: Marine N₂O emissions from nitrification and denitrification
 constrained by modern observations and projected in multimillennial global warming
 simulations, Global Biogeochem. Cy., 32, 92–121, https://doi.org/10.1002/2017GB005671,
- **640** 2018.
- Beman, J. M., Chow, C. E., King, A. L., Feng, Y., Fuhrman, J. A., Andersson, A., Bates, N. R.,
 Popp, B. N. and Hutchins, D. A.: Global declines in oceanic nitrification rates as a
 consequence of ocean acidification, P. Natl. Acad. Sci. USA, 108: 208–213, 2011.
- Berchet, A., Pison, I., Crill, P. M., Thornton, B., Bousquet, P., Thonat, T., Hocking, T.,
 Thanwerdas, J., Paris, J. D., and Saunois, M.: Using ship-borne observations of methane
 isotopic ratio in the Arctic Ocean to understand methane sources in the Arctic, Atmos. Chem.
 Physics, 20(6), 2020.
- Biastoch, A., Treude, T., Rüpke, L.H., Riebesell, U., Roth, C., Burwicz, E. B., Park, W., Latif,
 M., Böning, C.W., Madec, G., and Wallmann, K.: Rising Arctic Ocean temperatures cause
 gas hydrate destabilization and ocean acidification, Geophys. Res. Lett., 38, L08602,
 doi:10.1029/2011GL047222, 2011.
- Bižić, M., Klintzsch, T., Ionescu, D., Hindiyeh, M.Y., Gunthel, M., Muro-Pastor, A. M., Eckert,
 W., Urich, T., Keppler, F., and Grossart, H.-P.: Aquatic and terrestrial cyanobacteria produce
 methane, Science Advances, 6, eaax5343, 2020.
- Bodelier, P. L. and Steenbergh, A. K.: Interactions between methane and the nitrogen cycle in
 light of climate change, Curr. Opin. Environ. Sustain., 9, 26–36, 2014.
- Bonaglia, S., Brüchert, V., Callac, N., Vicenzi, A., Fru, E. C., and Nascimento, F. J. A.: Methane
 fluxes from coastal sediments are enhanced by macrofauna, Sci. Rep. 7:1–10, 2017.
- Boetius, A. and Wenzhöfer, F.: Seafloor oxygen consumption fuelled by methane from cold
 seeps, Nat. Geoscience, 6, 725–734, 2013.
- Borges A. V. and Abril, G.: Carbon dioxide and methane dynamics in estuaries, In: Wolanski, E.
 and McLusky, D. (eds) Treatise on Estuarine and Coastal Science, Vol. 5: Biogeochemistry,
 Academic Press, Waltham, pp 119-161, 2011.
- Borges, A. V., Champenois, W., Gypens, N., Delille, B., and Harlay, J.: Massive marine methane
 emissions from near-shore shallow coastal areas, Sci. Rep., 6, 27908, https://doi.org/ARTN
 2790810.1038/srep27908, 2016.
- Borges A. V., Speeckaert, G., Champenois, W., Scranton, M. I. and Gypens N.: Productivity and
 temperature as drivers of seasonal and spatial variations of dissolved methane in the Southern
 Bight of the North Sea, Ecosystems, 21, 583–599, doi: 10.1007/s10021-017-0171-7, 2018.
- Borges, A. V., Royer, C., Lapeya Martin, J., Champenois, W. and Gypens, N.: Response to the
 European 2018 heatwave of marine methane emission in the Southern North Sea, Cont. Shelf
- 672 Res., 190, 104004, doi: 10.1016/j.csr.2019.104004, 2019.
- Bourbonnais, A., Letscher, R. T., Bange, H. W., Echevin, V., Larkum, J., Mohn, J., Yoshida, N.,
 and Altabet, M. A.: N₂O production and consumption from stable isotopic and concentration
 data in the Peruvian coastal upwelling system, Global Biogeochem. Cy., 31, 678–698,
 https://doi.org/10.1002/2016GB005567, 2017.
- Brase, L., Bange, H. W., Lendt, R., Sanders, T., and Dähnke, K.: High resolution measurements
- of nitrous oxide (N_2O) in the Elbe Estuary, Front. Mar. Sci., 4, 162, doi:
- 679 10.3389/fmars.2017.00162, 2017.

- Breider, F., Yoshikawa, C., Makabe, A., Toyoda, S., Wakita, M., Matsui, Y., Kawagucci, S.,
 Fujiki, T., Harada, N., and Yoshida, N.: Response of N₂O production rate to ocean
- acidification in the western North Pacific, Nature Climate Change, 9, 954–958, 2019.
- Buchwald, C., Grabb, K., Hansel, C. M., and Wankel, S. D.: Constraining the role of iron in
 environmental nitrogen transformations: Dual stable isotope systematics of abiotic NO₂⁻
 reduction by Fe (II) and its production of N₂O, Geochim. Cosmochim. Acta, 186, 1–12,
- **686** 2016.
- Buitenhuis, E. T., Suntharalingam, P., and Le Quéré, C.: Constraints on global oceanic emissions
 of N₂O from observations and models, Biogeosciences, 15, 2161–2175, doi: 10.5194/bg-152161-2018, 2018.
- Bullister, J. L., Wisegarver, D. P., and Wilson, S. T.: The production of Methane and Nitrous
 Oxide Gas Standards for Scientific Committee on Ocean Research (SCOR) Working Group
 #143. pp 1–9, 2016.
- Burke, K. D., Williams, J. W., Chandler, M. A., Haywood, A. M., Lunt, D. J., and Otto-Bliesner,
 B. L.: Pliocene and Eocene provide best analogs for near-future climates, P. Natl. Acad. Sci.
- 695 USA, 115, 13288–13293, 2018.
- Call, M., Maher, D. T., Santos, I. R., Ruiz-Halpern, S., Mangion, P., Sanders, C. J., Erler, D. V.,
 Oakes, J. M., Rosentreter, J., Murray, R. and Eyre, B. D.: Spatial and temporal variability of
 carbon dioxide and methane fluxes over semi-diurnal and spring-neap-spring timescales in a
 mangrove creek, Geochim. Cosmochim. Acta, 150, 211–225, 2015.
- Caranto, J. D. and Lancaster, K. M.: Nitric oxide is an obligate bacterial nitrification
 intermediate produced by hydroxylamine oxidoreductase, P. Natl. Acad. Sci. USA, 114,
 8217–8222, 2017.
- Carini, P., White, A.E., Campbell, E.O., and Giovannoni, S. J.: Methane production by
 phosphate-starved SAR11 chemoheterotrophic marine bacteria, Nat. Comm., 5, 1–7, 2014.
- Carini, P., Dupont, C. L., and Santoro, A. E.: Patterns of thaumarchaeal gene expression in culture and diverse marine environments, Environ. Microbiol., 20, 2112–2124, https://doi.org/10.1111/1462-2920.14107, 2018.
- Casciotti, K. L., Forbes, M., Vedamati, J., Peters, B., Martin, T., and Mordy, C. W.: Nitrous
 oxide cycling in the Eastern Tropical South Pacific as inferred from isotopic and
 isotopomeric data, Deep Sea Res. Part II, 156, 155–167,
- 711 https://doi.org/10.1016/J.DSR2.2018.07.014, 2018.
- Chan, E. W., Shiller, A. M., Joung D. J., Arrington, E. C., Valentine, D. L., Redmond, M. C.,
 Breier, A., Socolofsky, A., and Kessler J. D.: Investigations of aerobic methane oxidation in
 two marine seep environments: Part 1 Chemical Kinetics, J. Geophys. Res.-Oceans,
 doi:10.1029/2019jc015594, 2019.
- Charpentier, J., Farías, L., and Pizarro, O.: Nitrous oxide fluxes in the central and eastern South
 Pacific, Global Biogeochem. Cy., 24, GB3011, doi:10.1029/2008GB003388, 2010.
- 718 Ciais, P., Sabine, C., Bala, G., Bopp, L., Brovkin, V., Canadell, J., Chhabra, A., DeFries, R.,
- Galloway, J., Heimann, M., Jones, C., Quéré, C. Le, Myneni, R. B., Piao, S. and Thornton, P.
 Carbon and Other Biogeochemical Cycles. In: Climate Change 2013: The Physical Science
 Basis. Contribution of Working Group 1 to the Fifth Assessment Report of the
- 722 Intergovernmental Panel on Climate Change. Stocker T, Qin D, Plattner G-K, Tignor M,
- Allen S, Boschung J, Nauels A, Xia Y, Bex V, Midgley P (eds) Cambridge University Press,
- 724 Cambridge, UK; New York, NY, 2013.

- 725 Conthe, M., Lycus, P., Arntzen, M. Ø., da Silva, A. R., Frostegård, Å., Bakken, L. R., van
- Loosdrecht, M. C.: Denitrification as an N₂O sink, Water Res., 151, 381–387, 2019.
- Damm, E., Thoms, S., Beszczynska-Möller, A., Nöthig, E. M., and Kattner, G.: Methane excess
 production in oxygen-rich polar water and a model of cellular conditions for this paradox,
 Polar Science, 9, 327–334, 2015.
- de la Paz, M., García-Ibáñez, M. I., Steinfeldt, R., Ríos, A. F., and Pérez, F. F.: Ventilation
 versus biology: What is the controlling mechanism of nitrous oxide distribution in the North
 Atlantic?, Global Biogeochem. Cyc., 31, 745–760, doi: 10.1002/2016GB005507, 2017.
- Dickey, T. D.: Emerging ocean observations for interdisciplinary data assimilation systems, J.
 Mar. Sys., 40, 5–48, 2003.
- Dickson, A. G., Sabine, C. L. and Christian, J. R. (Eds.): Guide to Best Practices for Ocean CO₂
 Measurements. PICES Special Publication, 2007.
- 737 Dickson, A.G.: Standards for ocean measurements, Oceanogr., 23, 34–47, 2010.
- Dore, J. E. and Karl, D. M.: Nitrification in the euphotic zone as a source for nitrite, nitrate, and
 nitrous oxide at Station ALOHA, Limnol. Oceanogr., 41, 1619–1628, 1996.
- Douglas, P. M. J., Stolper, D. A., Smith, D. A., Anthony, K. W., Paull, C. K., Dallimore, S.,
 Wik, M., Crill, P. M., Winterdahl, M., Eiler, J. M., and Sessions, A. L.: Diverse origins of
 Arctic and Subarctic methane point source emissions identified with multiply-substituted
 isotopologues, Geochim. Cosmochim. Acta, 188, 163–188, 2016.
- Douglas, P. M., Stolper, D. A., Eiler, J. M., Sessions, A. L., Lawson, M., Shuai, Y., Bishop, A.,
 Podlaha, O. G., Ferreira, A. A., Neto, E. V. S., and Niemann, M.: Methane clumped isotopes:
 Progress and potential for a new isotopic tracer, Organic Geochem., 113, 262–282, 2017.
- Frler, D. V., Duncan, T. M., Murray, R., Maher, D. T., Santos, I. R., Gatland, J. R., Mangion, P.,
 and Eyre, B. D.: Applying cavity ring-down spectroscopy for the measurement of dissolved
 nitrous oxide concentrations and bulk nitrogen isotopic composition in aquatic systems:
 Correcting for interferences and field application, Limnol. Oceanogr.: Methods, 13, 391–401,
- 751 2015.
- Ettwig, K. F., Butler, M. K., Le Paslier, D., Pelletier, E., Mangenot, S., Kuypers, M. M.,
 Schreiber, F., Dutilh, B. E., Zedelius, J., de Beer, D., and Gloerich, J.: Nitrite-driven
 anaerobic methane oxidation by oxygenic bacteria, Nature, 464, 543–548, 2010.
- Ettwig, K. F., Zhu, B., Speth, D., Keltjens, J. T., Jetten, M. S., and Kartal, B.: Archaea catalyze
 iron-dependent anaerobic oxidation of methane, P. Natl. Acad. Sci. USA, 113, 12792–12796,
 2016.
- Farías, L., Besoain, V., and García-Loyola, S.: Presence of nitrous oxide hotspots in the coastal
 upwelling area off central Chile: an analysis of temporal variability based on ten years of a
 biogeochemical time series, Environ. Res. Lett., 10, p.044017, 2015.
- Fenwick, L., Capelle, D., Damm, E., Zimmermann, S., Williams, W. J., Vagle, S., and Tortell, P.
 D.: Methane and nitrous oxide distributions across the North American Arctic Ocean during
- summer, 2015, J. Geophys. Res.-Oceans, 122, 390–412, doi:10.1002/2016JC012493, 2017.
- Ferrón, S., Ortega, T., Gómez-Parra, A. and Forja, J. M.: Seasonal study of dissolved CH₄, CO₂
 and N₂O in a shallow tidal system of the bay of Cádiz (SW Spain), J. Mar. Sys., 66, 244–257, 2007.
- 767 Foster, S. Q. and Fulweiler, R. W.: Estuarine sediments exhibit dynamic and variable
- biogeochemical responses to hypoxia, J Geophys. Res.-Biogeosciences, 124, 737–758, 2019.

- Foucher, J. P., Westbrook, G. K., Boetius, A. N., Ceramicola, S. I., Dupré, S., Mascle, J.,
 Mienert, J., Pfannkuche, O., Pierre, C. and Praeg D.: Structure and drivers of cold seep
- ecosystems, Oceanography, 22, 92–109, 2009.
- Frame C. and Casciotti K. L.: Biogeochemical controls and isotopic signatures of nitrous oxide
 production by a marine ammonia-oxidizing bacterium, Biogeosciences 7, 2695–2709, 2010.
- Frame, C. H., Deal, E., Nevison, C. D. and Casciotti, K. L.: N₂O production in the eastern South
 Atlantic: Analysis of N₂O stable isotopic and concentration data, Global Biogeochem. Cyc.,
- 776 28, 1262–1278, 2014.
- Frey, C., Bange, H. W., Achterberg, E. P., Jayakumar, A., Löscher, C. R., Arévalo-Martínez, D.
 L., León-Palmero, E., Sun, M., Xie, R. C., Oleynik, S., and Ward, B. B.: Regulation of
 nitrous oxide production in low oxygen waters off the coast of Peru, Biogeosci., 17, 2263–
 2287, 2020.
- Fuchsman, C. A., Devol, A. H., Saunders, J. K., McKay, C., and Rocap, G.: Niche partitioning of
 the N cycling microbial community of an offshore oxygen deficient zone, Front. Microbiol.,
 8, 2384, https://doi.org/10.3389/fmicb.2017.02384, 2017.
- Ganesan, A. L., Schwietzke, S., Poulter, B., Arnold, T., Lan, X., Rigby, M., et al.: Advancing
 scientific understanding of the global methane budget in support of the Paris Agreement,
 Global Biogeochem. Cyc., 33. https://doi.org/10.1029/ 2018GB006065, 2019.
- Ganesan, A.L., Manizza, M., Morgan, E.J., Harth, C.M., Kozlova, E., Lueker, T., Manning, A.J.,
 Lunt, M.F., Mühle, J., Lavric, J.V. and Heimann, M.: Marine nitrous oxide emissions from
 three Eastern Boundary Upwelling Systems inferred from atmospheric observations,
 Geophys. Res. Lett.,doi: 10.1029/2020GL087822, 2020.
- Ganesh, S., Parris, D. J., DeLong, E. F., and Stewart, F.J.: Metagenomic analysis of sizefractionated picoplankton in a marine oxygen minimum zone, The ISME J., 8, 187–211,
 2014.
- Garcia-Tigreros, F. and Kessler, J. D.: Limited acute influence of aerobic methane oxidation on
 ocean carbon dioxide and pH in Hudson Canyon, northern US Atlantic margin, J. Geophys.
 Res.: Biogeosci., 123, 2135–2144, 2018.
- Gelesh, L., Marshall, K., Boicourt, W., and Lapham, L.: Methane concentrations increase in
 bottom waters during summertime anoxia in the highly eutrophic estuary, Chesapeake Bay,
 USA, Limnol. Oceanogr., 61, S253–S266, 2016.
- Goreau, T. J., Kaplan, W. A., Wofsy, S. C., McElroy, M. B., Valois, F. W., and Watson, S.W.:
 Production of NO₂⁻ and N₂O by nitrifying bacteria at reduced concentrations of oxygen.
 Appl. Environ. Microbiol., 40, 526–532, 1980.
- Gülzow, W., Rehder, G., Schneider, B., Schneider, J., Deimling, V., and Sadkowiak, B.: A new
 method for continuous measurement of methane and carbon dioxide in surface waters using
 off-axis integrated cavity output spectroscopy (ICOS): An example from the Baltic Sea,
 Limnol. Oceanogr. Methods, 9, 176–184, 2011.
- Gutiérrez-Loza, L., Wallin, M. B., Sahlée, E., Nilsson, E., Bange, H. W., Kock, A. and
 Rutgersson, A.: Measurement of air-sea methane fluxes in the Baltic Sea using the eddy
 covariance method, Front. Earth Sci., 7, 93, doi: 10.3389/feart.2019.00093, 2019.
- Hallin, S., Philippot, L., Löffler, F. E., Sanford, R. A., and Jones, C. M.: Genomics and ecology
 of novel N₂O-reducing microorganisms, Trends Microbiol., 26, 43–55, 2018.
- Haroon, M.F., Hu, S., Shi, Y., Imelfort, M., Keller, J., Hugenholtz, P., Yuan, Z., and Tyson,
- G.W.: Anaerobic oxidation of methane coupled to nitrate reduction in a novel archaeal
- lineage, Nature, 500, 567–570, https://doi.org/10.1038/nature12375, 2013.

- 815 Harris, S. J., Liisberg, J., Longlong Xia, Wei, J., Zeyer, K., Yu, L., Barthel, M., Wolf, B., Kelly,
- B. F. J., Cendón, D. I., Blunier, T., Six, J., and Mohn, J.: N₂O isotopocule measurements
 using laser spectroscopy: Analyzer characterization and intercomparison, Atmos. Meas.
 Tech. https://doi.org/10.5194/amt-2019-451, 2019.
- Hink, L., Lycus, P., Gubry-Rangin, C., Frostegård, Å., Nicol, G. W., Prosser, J. I. and Bakken, L.
 R.: Kinetics of NH₃-oxidation, NO-turnover, N₂O-production and electron flow during
 oxygen depletion in model bacterial and archaeal ammonia oxidizers, Environ. Microbiol.,
- 822 19, 4882–4896, 2017.
- Ho, D. T., Schlosser, P., Orton, P. M.: On factors controlling air–water gas exchange in a large
 tidal river, Estuar. Coasts, 34, 1103–1116, 2011.
- Hopkins, F. E., Suntharalingam, P., Gehlen, M., Andrews, O., Archer, S. D., Bopp, L.,
 Buitenhuis, E., Dadou, I., Duce, R., Goris, N. and Jickells, T.: The impacts of ocean
 acidification on marine trace gases and the implications for atmospheric chemistry and
 climate, Proc. R. Soc. A, 476: 20190769, doi:10.1098/rspa.2019.0769, 2020.
- Huang, J., Luo, M., Liu, Y., Zhang, Y., and Tan, J.: Effects of tidal scenarios on the methane
 emission dynamics in the subtropical tidal marshes of the Min River Estuary in Southeast
 China, Internat. J. Environ. Res. Public Health, 16, 2790, 2019.
- IPCC: In Climate Change: The Physical Science Basis. Contribution of Working Group I to the
 Fifth Assessment Report of the Intergovernmental Panel on Climate Change 2013. Stocker,
- T. F., Qin, D., Plattner, G.-K., Tignor, M., Allen, S. K., Boschung, J., Nauels, A., Xia, Y.,
- Bex ,V., Midgley, P. M. (Eds.) Cambridge University Press; 20131535
- IPCC, 2019: Summary for Policymakers. In: IPCC Special Report on the Ocean and Cryosphere
 in a Changing Climate [H.-O. Pörtner, D.C. Roberts, V. Masson-Delmotte, P. Zhai, M.
 Tignor, E. Poloczanska, K. Mintenbeck, M. Nicolai, A. Okem, J. Petzold, B. Rama, N.
 Weyer (Eds.).
- James, R. H., Bousquet, P., Bussmann, I., Haeckel, M., Kipfer, R., Leifer, I., Niemann, H.,
- Ostrovsky, I., Piskozub, J., Rehder, G. and Treude, T.: Effects of climate change on methane
 emissions from seafloor sediments in the Arctic Ocean: A review, Limnol. Oceanogr., 61,
 S283–S299, 2016.
- Jeffrey, L. C., Maher, D. T., Santos, I. R., Call, M., Reading, M. J., Holloway, C. and Tait, D. R.:
 The spatial and temporal drivers of pCO₂, pCH₄ and gas transfer velocity within a subtropical
 estuary, Estuar. Coast Shelf Sci., 208, 83–95, 2018.
- Ji, Q., Babbin, A. R., Jayakumar, A., Oleynik, S. and Ward, B. B.: Nitrous oxide production by
 nitrification and denitrification in the Eastern Tropical South Pacific oxygen minimum zone,
 Geophys. Res. Lett., 42, 10–755, 2015.
- Ji, Q. and Ward, B. B.: Nitrous oxide production in surface waters of the mid-latitude North
 Atlantic Ocean, J. Geophys. Res.: Oceans, 122, 2612–2621,
 https://doi.org/10.1002/2016IC012467_2017
- 852 https://doi.org/10.1002/2016JC012467, 2017.
- Ji, Q., Buitenhuis, E., Suntharalingam, P., Sarmiento, J. L., and Ward, B. B.: Global nitrous
 oxide production determined by oxygen sensitivity of nitrification and denitrification, Global
 Biogeochem. Cy., 32, 1790–1802, https://doi.org/10.1029/2018GB005887, 2018.
- Jordan, S. F., Treude, T., Leifer, I., Janßen, R., Werner, J., Schulz-Vogt, H. and Schmale, O.:
- 857 Bubble-mediated transport of benthic microorganisms into the water column: Identification
- of methanotrophs and implication of seepage intensity on transport efficiency, Scientific Reports, 10, 1–15, 2020.

- Karl, D. M., Beversdorf, L., Björkman, K. M., Church, M. J., Martinez, A., and Delong, E. F.:
 Aerobic production of methane in the sea, Nat. Geosci., 1, 473–478, 2008.
- Kessler, J. D. and Reeburgh, W. S.: Preparation of natural methane samples for stable isotope
 and radiocarbon analysis, Limnol. Oceanogr.: Methods, 3, 408–418,
 doi:10.4319/lom.2005.3.408, 2005.
- Kessler, J. D., Reeburgh, W. S., Valentine, D. L., Kinnaman, F. S., Peltzer, E. T., Brewer, P. G.,
 Southon, J., and Tyler, S. C.: A survey of methane isotope abundance (¹⁴C, ¹³C, ²H) from five
 nearshore marine basins that reveals unusual radiocarbon levels in subsurface waters, J.
 Geophys. Res.-Oceans, 113(C12), doi:10.1029/2008jc004822, 2008.
- Klintzsch, T., Langer, G., Nehrke, G., Wieland, A., Lenhart, K., and Keppler, F.: Methane
 production by three widespread marine phytoplankton species: release rates, precursor
 compounds, and potential relevance for the environment, Biogeosciences, 16, 4129–4144,
 2019.
- Kock, A. and Bange, H. W.: Counting the ocean's greenhouse gas emissions, Eos Earth & Space
 Science News, 96, 10–13, 2015.
- Kodovska, F. G. T., Sparrow, K. J., Yvon-Lewis, S. A., Paytan, A., Dimova, N. T., Lecher, A.
 and Kessler, J. D.: Dissolved methane and carbon dioxide fluxes in Subarctic and Arctic
 regions: Assessing measurement techniques and spatial gradients, Earth Planet. Sci. Lett.
 436, 43–55, 2016.
- Kosmach, D. A., Sergienko, V. I., Dudarev, O. V., Kurilenko, A. V., Gustafsson, O., Semiletov,
 I. P., and Shakhova, N. E.: Methane in the surface waters of Northern Eurasian marginal
 seas. In Doklady Chemistry Vol. 465, No. 2, 281–285. Pleiades Publishing.
- Kremer, J. N., Reischauer, A., D'Avanzo, C.: Estuary-specific variation in the air-water gas
 exchange coefficient for oxygen, Estuaries, 26, 829–836, 2003.
- Labidi, J., Young, E. D., Giunta, T., Kohl, I. E., Seewald, J., Tang, H., Lilley, M. D., and Fruhgreen, G. L.: Methane thermometry in deep-sea hydrothermal systems: evidence for reordering of doubly-substituted isotopologues during fluid cooling, Geochim. Cosmochim.
 Acta, 88, 248–261, 2020.
- Lan, X., Tans, P., Sweeney, C., Andrews, A., Dlugokencky, E., Schwietzke, S., Kofler, J.,
 McKain, K., Thoning, K., Crotwell, M., and Montzka, S.: Long-term measurements show
 little evidence for large increases in total U.S. methane emissions over the past decade,
 Geophys. Res. Lett., 46, 4991–4999. https://doi.org/10.1029/2018GL081731, 2019.
- Lancaster, K. M., Caranto, J. D., Majer, S. H., and Smith, M. A.: Alternative bioenergy: Updates
 to and challenges in nitrification metalloenzymology, Joule, 2, 421–441.
- https://doi.org/10.1016/j.joule.2018.01.018, 2018.
- Landolfi, A., Somes, C. J., Koeve, W., Zamora, L. M., and Oschlies, A.: Oceanic nitrogen
 cycling and N₂O flux perturbations in the Anthropocene, Global Biogeochem. Cy., 31, 1236–
 1255, doi:10.1002/2017GB005633, 2017.
- Lapham, L., Wilson, R., Riedel, M., Paull, C. K. and Holmes, M. E.: Temporal variability of in situ methane concentrations in gas hydrate-bearing sediments near Bullseye Vent, Northern Cascadia Margin, Geochem. Geophys. Geosys., 14, 2445–2459, 2013.
- 901 Lapham, L., Marshall, K., Magen, C., Lyubchich, V., Cooper, L.W., and Grebmeier, J. M.:
- Dissolved methane concentrations in the water column and surface sediments of Hanna Shoal
- and Barrow Canyon, Northern Chukchi Sea, Deep Sea Res., 144, 92–103, 2017.
- Leonte, M., Kessler, J. D., Kellermann, M. Y., Arrington, E. C., Valentine, D. L., and Sylva, S.
- P.: Rapid rates of aerobic methane oxidation at the feather edge of gas hydrate stability in the

- waters of Hudson Canyon, US Atlantic Margin, Geochim. Cosmochim. Acta, 204, 375–387,
 doi:10.1016/j.gca.2017.01.009, 2017.
- Leonte, M., Wang, B., Socolofsky, S. A., Mau, S., Breier, J. A., and Kessler, J. D.: Using carbon
 isotope fractionation to constrain the extent of methane dissolution into the water column
 surrounding a natural hydrocarbon gas seep in the Northern Gulf of Mexico, Geochem.
 Geophys. Geosys. 19, 4459–4475, doi:10.1029/2018gc007705, 2018.
- Leonte, M., Ruppel, C. D., Ruiz-Angulo, A., and Kessler, J. D.: Surface methane concentrations
- along the Mid-Atlantic Bight driven by aerobic subsurface production rather than seafloor
 gas seeps, J. Geophys. Res.: Oceans, 125, e2019JC015989, 2020.
- Lohrberg, A., Schmale, O., Ostrovsky, I., Niemann, H., Held, P. and Schneider von Deimling, J.:
 Discovery and quantification of a widespread methane ebullition event in a coastal inlet
 (Baltic Sea) using a novel sonar strategy, Sci. Rep. 10, 4393, 2020.
- Lorenson, T. D., Greinert, J. and Coffin, R. B.: Dissolved methane in the Beaufort Sea and the
 Arctic Ocean, 1992–2009; sources and atmospheric flux, Limnol. Oceanogr., 61, S300–S323,
 2016.
- Löscher, C., Kock, A., Koenneke, M., LaRoche, J., Bange, H. W. and Schmitz, R. A.: Production
 of oceanic nitrous oxide by ammonia-oxidizing archaea, Biogeosci., 9, 2419–2429, 2012.
- Lycus, P., Bøthun, K. L., Bergaust, L., Shapleigh, J. P., Bakken, L. R., and Frostegård, Å.:
 Phenotypic and genotypic richness of denitrifiers revealed by a novel isolation strategy, The
 ISME J., 11, 2219–2232, 2017.
- Ma, X., Lennartz, S. T. and Bange, H. W.: A multi-year observation of nitrous oxide at the
 Boknis Eck Time Series Station in the Eckernförde Bay (southwestern Baltic Sea),
 Biogeosci., 16, 4097–4111, https://doi.org/10.5194/bg-16-4097-2019, 2019.
- Maher, D. T., Cowley, K., Santos, I. R., Macklin, P., and Eyre, B. D.: Methane and carbon
 dioxide dynamics in a subtropical estuary over a diel cycle: Insights from automated in situ
 radioactive and stable isotope measurements, Mar. Chem., 168, 69–79, 2015.
- Maher, D. T., Sippo, J. Z., Tait, D. R., Holloway, C., and Santos, I. R., Pristine mangrove creek
 waters are a sink of nitrous oxide, Sci. Rep., 6, 25701, 2016.
- Maher, D. T., Drexl, M., Tait, D. R., Johnston, S. G. and Jeffrey, L. C.: iAMES: An inexpensive
 automated methane ebullition sensor, Environ. Sci. Technol., 53, 6420–6426, 2019.
- 936 Manning, C. C., Preston, V. L., Jones, S. F., Michel, A. P. M., Nicholson, D. P., Duke, P. J.,
- Ahmed, M. M. M., Manganini, K., Else, B. G. T., and Tortell, P.D.: River inflow dominates
 methane emissions in an Arctic coastal system, Geophys. Res. Lett., 47, e2020GL087669
 https://doi.org/10.1029/2020GL087669, 2020
- Marchant, H. K., Ahmerkamp, S., Lavik, G., Tegetmeyer, H. E., Graf, J., Klatt, J. M.,
 Holtappels, M., Walpersdorf, E. and Kuypers, M. M.: Denitrifying community in coastal
 sediments performs aerobic and anaerobic respiration simultaneously, The ISME J., 11,
 1799–1812, 2017.
- Marchant, H. K., Tegetmeyer, H. E., Ahmerkamp, S., Holtappels, M., Lavik G., Graf, J.,
 Schreiber, F., Mussmann, M., Strous M., and Kuypers M. M. M.: Metabolic specialization of
 denitrifiers in permeable sediments controls N₂O emissions, Environ. Microbiol., 20, 4486–
 4502, doi: 10.1111/1462-2920.14385, 2018.
- Mau, S., Heintz, M. B., and Valentine, D. L.: Quantification of CH₄ loss and transport in
- dissolved plumes of the Santa Barbara Channel, California, Cont. Shelf Res., 32, 110–120,
 doi: 10.1016/j.csr.2011.10.016, 2012.

- McDermitt, D., Burba, G., Xu, L., Anderson, T., Komissarov, A., Riensche, B., Schedlbauer, J.,
 Starr, G., Zona, D., Oechel, W., Oberbauer, S. and Hastings, S.: A new low-power, open-path
 instrument for measuring methane flux by eddy covariance, Applied Physics B, 102, 391–
 405, 2011
- Molstad, L., Dörsch, P. and Bakken, L.R.: Robotized incubation system for monitoring gases
 (O₂, NO, N₂O N₂) in denitrifying cultures, J. Microbiol. Methods, 71, 202–211, 2007.
- Montzka, S. A., Dlugokencky, E. J. and Butler, J. H.: Non-CO₂ greenhouse gases and climate
 change, Nature, 476, 43–50, 2011.
- Müller, D., Bange, H. W., Warneke, T., Rixen, T., Müller, M., Mujahid, A., and Notholt, J.:
 Nitrous oxide and methane in two tropical estuaries in a peat–dominated region of
 northwestern Borneo, Biogeosci., 13, 2415–2428, https://doi.org/10.5194/bg-13–2415–2016,
 2016.
- Murray, R. H., Erler, D. V., and Eyre, B. D.: Nitrous oxide fluxes in estuarine environments:
 response to global change, Glob. Change Biol., 21, 3219–3245, 2015.
- Murray, R., Erler, D. V., Rosentreter, J., Wells, N. S., and Eyre, B. D.: Seasonal and spatial
 controls on N₂O concentrations and emissions in low-nitrogen estuaries: Evidence from three
 tropical systems, Mar. Chem., 103779, 2020.
- Nemitz, E., Mammarella, I., Ibrom, A., Aurela, M., Burba, G. G., Dengel, S., Gielen, B., Grelle,
 A., Heinesch, B., Herbst, M., Hörtnagl, L., Klemedtsson, L., Lindroth, A., Lohila, A.,
- McDermitt, D. K., Meier, P., Merbold, L., Nelson, D., Nicolini, G., Nilsson, M. B., Peltola.
 O., Rinne, J., and Zahniser, M.: Standardisation of eddy-covariance flux measurements of
 methane and nitrous oxide, Int. Agrophysics, 32, 517–549, 2018.
- Nicholson, D. P., Michel, A. P., Wankel, S. D., Manganini, K., Sugrue, R. A., Sandwith, Z. O.,
 and Monk, S.A.: Rapid mapping of dissolved methane and carbon dioxide in coastal
 ecosystems using the ChemYak autonomous surface vehicle, Environ. Sci. Tech., 52, 13314–
 13324, 2018.
- Nightingale, P. D., Malin, G., Law, C. S., Watson, A. J., Liss, P. S., Liddicoat, M. I., Boutin, J.,
 and Upstill-Goddard, R. C.: In situ evaluation of air-sea gas exchange parameterizations
 using novel conservative and volatile tracers, Global Biogeochem. Cy., 14, 373–387, 2000.
- Novick, K. A., Biederman, J. A., Desai, A. R., Litvak, M. E., Moore, D. J., Scott, R. L. and Torn,
 M. S.: The AmeriFlux network: A coalition of the willing, Agricultural and Forest
 Meteorology, 249, 444–456, 2018.
- Ostrom, N. E., Gandhi, H., Trubl, G., and Murray, A. E.: Chemodenitrification in the
 cryoecosystem of Lake Vida, Victoria Valley, Antarctica. Geobiol., 14, 575–587, 2016.
- Pack, M. A., Heintz, M. B., Reeburgh, W. S., Trumbore, S. E., Valentine, D. L., Xu, X., and
 Druffel, E. R.: A method for measuring methane oxidation rates using low levels of ¹⁴Clabeled methane and accelerator mass spectrometry, Limnol. Oceanogr.: Methods, 9, 245–
 260. doi:10.4319/lom 2011.9.245.2011
- 988 260, doi:10.4319/lom.2011.9.245, 2011.
- 989 Pankratova, N., Belikov, I., Skorokhod, A., Belousov, V., Arta- monov, A., Repina, I., and 990 Shishov, E.: Measurements and data processing of atmospheric CO₂, CH₄, H₂O and δ^{13} C-991 CH₄ mixing ratio during the ship campaign in the East Arctic and the Far East seas in autumn
- 2016, IOP Conf. Ser.: Earth Environ. Sci., 231, 012041, https://doi.org/10.1088/1755 1315/231/1/012041, 2019.
- Pfeiffer-Herbert, A. S., Prahl, F. G., Peterson, T. D., and Wolhowe, M.: Methane dynamics
 associated with tidal processes in the Lower Columbia River, Estuaries and Coasts, 42,
 1249–1264, 2019.

- 997 Podgrajsek, E., Sahlee, E., Bastviken, D., Natchimuthu, S., Kljun, N., Chmiel, H. E.,
- Klemedtsson, L., Rutgersson, A.: Methane fluxes from a small boreal lake measured with the
 eddy covariance method, Limnol. Oceanogr., 61, S41–S50, 2016.
- Pohlman, J. W., Kaneko, M., Heuer, V. B., Coffin, R. B., and Whiticar, M.: Methane sources and
 production in the northern Cascadia margin gas hydrate system, Earth Planetary Sci. Lett.,
 287, 504–512, 2009.
- Pohlman, J. W., Bauer, J. E., Waite, W. F., Osburn, C. L., and Chapman, N. R.: Methane
 hydrate-bearing seeps as a source of aged dissolved organic carbon to the oceans, Nat.
 Geosci., 4, 37–41, 2011.
- Pohlman, J. W., Greinert, J., Ruppel, C., Silyakova, A., Vielstädte, L., Casso, M., Mienert, J.,
 and Bünz, S.: Enhanced CO₂ uptake at a shallow Arctic Ocean seep field overwhelms the
 positive warming potential of emitted methane, P. Natl. Acad. Sci. USA, 114, 5355–5360,
 2017.
- Prosser, J. I., Hink, L., Gubry-Rangin, C., and Nicol, G. W.: Nitrous oxide production by
 ammonia oxidizers: Physiological diversity, niche differentiation and potential mitigation
 strategies, Glob. Change Biol., 26, 103–118, 2020.
- Qin, W., Meinhardt, K. A., Moffett, J. W., Devol, A. H., Armbrust, E.V., Ingalls, A.E. and Stahl,
 D. A:. Influence of oxygen availability on the activities of ammonia-oxidizing archaea,
 Environ. Microbiol. Reports, 9, 250–256, 2017.
- Ray, N. E., Maguire, T. J., Al-Haj, A. N., Henning, M. C., and Fulweiler, R. W.: Low
 greenhouse gas emissions from oyster aquaculture, Environ. Sci. Technol., 53, 9118–9127,
 2019.
- Raymond, P. A. and Cole, J. J.: Gas exchange in rivers and estuaries: Choosing a gas transfer velocity, Estuaries, 24, 312–317, 2001.
- Reading, M. J., Tait, D. R., Maher, D. T., Jeffrey, L. C., Looman, A., Holloway, C., Shishaye, H.
 A., Barron, S., and Santos, I. R.: Land use drives nitrous oxide dynamics in estuaries on
 regional and global scales, Limnol. Oceanogr., https://doi.org/10.1002/lno.11426, 2020.
- 1024 Reeburgh, W. S.: Oceanic methane biogeochemistry, Chem. Rev., 107: 486–513, 2007.
- Repeta, D. J., Ferrón, S., Sosa, O. A., Johnson, C. G., Repeta, L. D., Acker, M., DeLong, E. F.,
 and Karl, D. M.: Marine methane paradox explained by bacterial degradation of dissolved
 organic matter, Nat. Geosci., 9, 884–887, 2016.
- Romanovskii, N. N., Hubberten, H. W., Gavrilov, A. V., Eliseeva, A. A., and Tipenko, G. S.:
 Offshore permafrost and gas hydrate stability zone on the shelf of East Siberian Seas, GeoMar. Lett., 25, 167–182, 2005.
- Rosentreter, J. A., Maher, D. T., Ho, D. T., Call, M., Barr, J. G., and Eyre, B. D.: Spatial and
 temporal variability of CO₂ and CH₄ gas transfer velocities and quantification of the CH₄
 microbubble flux in mangrove dominated estuaries, Limnol. Oceanogr., 62, 561–578, 2017.
- Rosentreter, J. A., Maher, D. T., Erler, D. V., Murray, R. H., and Eyre, B. D.: Methane emissions
 partially offset "blue carbon" burial in mangroves, Science advances 4, eaao4985, 2018.
- Ruppel, C. D. and Kessler, J. D.: The interaction of climate change and methane hydrates, Rev.
 Geophys., 55, 126–168, doi:10.1002/2016RG000534, 2017.
- Santoro, A. E., Casciotti, K. L., and Francis, C. A.: Activity, abundance and diversity of
 nitrifying archaea and bacteria in the central California Current, Environ. Microbiol., 12,
 1989–2006, 2010.
- Santoro, A. E., Buchwald, C., McIlvin, M. R., and Casciotti, K. L.: Isotopic signature of N(2)O
 produced by marine ammonia-oxidizing archaea, Science, 333, 1282–1285, 2011.

- 1043 Sapart, C. J., Shakhova, N., Semiletov, I., Jansen, J., Szidat, S., Kosmach, D., Dudarev, O., van
- der Veen, C., Egger, M., Sergienko, V., Salyuk, A., Tumskoy, V., Tison, J.-L., and
- 1045 Röckmann, T.: The origin of methane in the East Siberian Arctic Shelf unraveled with triple
 1046 isotope analysis, Biogeosciences, 14, 2283–2292, 2017.
- Saunois, M., Bousquet, P., Poulter, B., Peregon, A., Ciais, P., Canadell, J.G., Dlugokencky, E. J.,
 Etiope, G., Bastviken, D., Houweling, S., and Janssens-Maenhout, G.: The global methane
 budget 2000-2012, Earth System Science Data, 8, 697–751, 2016.
- Saunois et al.: The Global Methane Budget 2000–2017, Earth Syst. Sci. Data Discuss.,
 https://doi.org/10.5194/essd-2019-128, 2020.
- Schmale, O., Wage, J., Mohrholz, V., Wasmund, N., Grawe, U., Rehder, G., Labrenz, M., LoickWilde, N.: The contribution of zooplankton to methane supersaturation in the oxygenated
 upper waters of the central Baltic Sea, Limnol. Oceanogr.: 63:412–430, 2018.
- Schmidt, H. L., Werner, R. A., Yoshida, N., and Well, R.: Is the isotopic composition of nitrous
 oxide an indicator for its origin from nitrification or denitrification? A theoretical approach
 from referred data and microbiological and enzyme kinetic aspects, Rapid Comm. Mass
 Spec., 18, 2036–2040, 2004.
- Schutte, C. A., Wilson, A. M., Evans, T., Moore, W. S., and Joye, S. B.: Methanotrophy controls
 groundwater methane export from a barrier island, Geochim. Cosmochim. Acta, 179, 242–
 256, 2016.
- Seitzinger, S. P. and Kroeze, C.: Global distribution of nitrous oxide production and N inputs in
 freshwater and coastal marine ecosystems, Global Biogeochem. Cy., 12, 93–113, 1998.
- Shakhova, N., Semiletov, I., Leifer, I., Salyuk, A., Rekant, P., and Kosmach, D.: Geochemical
 and geophysical evidence of methane release over the East Siberian Arctic Shelf, J. Geophys.
 Res: Oceans, 115, doi:10.1029/2009JC005602, 2010.
- Sosa, O. A., Burrell, T. J., Wilson, S. T., Foreman, R. K., Karl, D. M., and Repeta, D. J.:
 Phosphonate cycling supports methane and ethylene production and supersaturation in the
 phosphate-depleted western North Atlantic Ocean, Limnol. Oceanogr., doi:
 10.1002/lno.11463, 2020.
- Sparrow, K. J. and Kessler, J. D.: Efficient collection and preparation of methane from low
 concentration waters for natural abundance radiocarbon analysis, Limnol.
 Oceanogr.:Methods, 15, 601–617, doi:10.1002/lom3.10184, 2017.
- Sparrow, K. J., Kessler, J. D., Southon, J. R., Garcia-Tigreros, F., Schreiner, K.M., Ruppel, C.
 D., Miller, J. B., Lehman, S. J., and Xu, X.: Limited contribution of ancient methane to
 surface waters of the U.S. Beaufort Sea shelf, Sci. Adv., 4, doi:10.1126/sciadv.aao4842,
 2018.
- Stahl, D. A. and de la Torre, J.R.: Physiology and diversity of ammonia-oxidizing archaea.
 Annu. Review Microbiol., 66, 83–101, 2012.
- Stein, L. Y. and Yung, Y. L.: Production, isotopic composition, and atmospheric fate of
 biologically produced nitrous oxide, Annu. Rev. Earth Planetary Sci., 31, 329–356, 2003.
- Steinle, L., Graves, C. A., Treude, T., Ferré, B., Biastoch, A., Bussmann, I., Berndt, C., Krastel,
 S., James, R. H., Behrens, E. and Böning, C.W.: Water column methanotrophy controlled by
 a rapid oceanographic switch, Nat. Geoscience, 8, 378–382, 2015.
- 1085 Stewart, F. J., Dalsgaard, T., Young, C. R., Thamdrup, B., Revsbech, N. P., Ulloa, O., Canfield,
- 1086 D. E., and DeLong, E. F.: Experimental incubations elicit profound changes in community 1087 transcription in OMZ bacterioplankton, PloS One, 7, e37118, 2012.

- Stieglmeier, M., Mooshammer, M., Kitzler, B., Wanek, W., Zechmeister-Boltenstern, S.,
 Richter, A., and Schleper, C.: Aerobic nitrous oxide production through N-nitrosating hybrid formation in ammonia-oxidizing archaea, The ISME J., 8, 1135–1146, 2014.
- Stolper, D. A., Sessions, A. L., Ferreira, A. A., Neto, E. S., Schimmelmann, A., Shusta, S. S.,
 Valentine, D. L. and Eiler, J. M.: Combined ¹³C–D and D–D clumping in methane: methods
 and preliminary results, Geochim. Cosmochim. Acta, 126, 169–191, 2014.
- 1094 Stramma, L., Johnson, G. C., Sprintall, J. and Mohrholz, V.: Expanding oxygen-minimum zones 1095 in the tropical oceans, Science, 320, 655–658, 2008.
- Suess, E.: Marine cold seeps, In Handbook of Hydrocarbon and Lipid Microbiology, (Eds
 Timmis, K. N.) pp. 187–203, Springer, Berlin, 2010.
- Sun, X., Jayakumar, A., and Ward, B. B.: Community composition of nitrous oxide consuming
 bacteria in the oxygen minimum zone of the Eastern Tropical South Pacific, Front.
 Microbiol., 8, 1183. https://doi.org/10.3389/fmicb.2017.01183, 2017.
- Suntharalingam, P., Buitenhuis, E., Le Quere, C., Dentener, F., Nevison, C., Butler, J. H., Bange,
 H. W., and Forster, G.: Quantifying the impact of anthropogenic nitrogen deposition on
- 1103 oceanic nitrous oxide, Geophys. Res. Lett., 39, L07605, doi:10.1029/2011gl050778, 2012.
- Sutka, R. L., Ostrom, N. E., Ostrom, P. H., Breznak, J. A., Gandhi, H., Pitt, A. J., and Li, F.:
 Distinguishing nitrous oxide production from nitrification and denitrification on the basis of
 isotopomer abundances, Appl. Environ. Microbiol., 72, 638–644, 2006.
- Tait, D. R., Maher, D. T., Wong, W., Santos, I. R., Sadat–Noori, M., Holloway, C., and Cook,
 P.L.: Greenhouse gas dynamics in a salt–wedge estuary revealed by high resolution cavity
 ring–down spectroscopy observations, Environ. Sci. Tech., 51, 13771–13778, 2017.
- 1110 Thonat, T., Saunois, M., Pison, I., Berchet, A., Hocking, T., Thornton, B., Crill, P., and 1111 Bousquet, P.: Assessment of the theoretical limit in instrumental detectability of Arctic 1112 methane sources using $\delta^{13}C_{CH4}$ atmospheric signals, Atmos. Chem. Phys., 19, 12141–12161, 1113 2019.
- Thornton, B. F., Geibel, M. C., Crill, P. M., Humborg, C., and Mörth, C,-M.: Methane fluxes
 from the sea to the atmosphere across the Siberian shelf seas, Geophys. Res. Letts., 43,
 doi:10.1002/2016GL068977, 2016a.
- Thornton, B. F., Wik, M., and Crill, P. M.: Double-counting challenges the accuracy of highlatitude methane inventories, Geophys. Res. Lett., 43,12,569–12,577,
 doi:10.1002/2016GL071772, 2016b.
- Thornton, B. F., Prytherch, J., Andersson, K., Brooks, I. M., Salisbury, D., Tjernström, M., and
 Crill, P. M.: Shipborne eddy covariance observations of methane fluxes constrain Arctic sea
 emissions, Science Advances, 6, doi:10.1126/sciadv.aay7934, 2020.
- Tian, H., Xu, R., Canadell, J. G. et al. A comprehensive quantification of global nitrous oxide
 sources and sinks, Nature, 586, 248–256, 2020.
- Tohjima, Y., Zeng, J., Shirai, T., Niwa, Y., Ishidoya, S., Taketani, F., Sasano, D., Kosugi, N.,
 Kameyama, S., Takashima, H. and Nara, H.: Estimation of CH₄ emissions from the East
 Siberian Arctic Shelf based on atmospheric observations aboard the R/V Mirai during fall
- 1128 cruises from 2012 to 2017, Polar Sci., doi:10.1016/j.polar.2020.100571, 2020.
- 1129 Torn, M. S., Biraud, S., Agarwal, D., Keenan, T. F., Chan, S., Christianson, D.S., Chu, H.,
- 1130 McNicol, G., Papale, D., Pastorello, G. and Stover, D. B.: AmeriFlux: Flux Synthesis and the
- 1131 Year of Methane. Abstract, American Geophysical Union Fall Meeting, TH13H, 2019.

- Trimmer, M., Chronopoulou, P. M., Maanoja, S. T., Upstill-Goddard, R. C., Kitidis, V., and
 Purdy, K. J.: Nitrous oxide as a function of oxygen and archaeal gene abundance in the North
 Pacific, Nat. Comm., 7, 1–10, 2016.
- Upstill-Goddard, R. C.: Air-sea gas exchange in the coastal zone, Est., Coastal Shelf Sci., 70,
 388–404, 2006.
- Upstill-Goddard, R. C. and Barnes, J.: Methane emissions from UK estuaries: Re-evaluating the
 estuarine source of tropospheric methane from Europe, Mar. Chem., 180, 14–23, 2016.
- Valentine, D. L.: Emerging topics in marine methane biogeochemistry, Annu. Rev. Mar. Sci., 3,
 147–171, 2011.
- Vieillard, A. M. and Fulweiler, R. W.: Tidal pulsing alters nitrous oxide fluxes in a temperate
 intertidal mudflat, Ecology, 95, 1960–1971, 2014.
- Wanninkhof, R.: Relationship between wind speed and gas exchange over the ocean revisited,
 Limnol. Oceanogr. Methods, 12, 351–362, 2014.
- Wang, D. T., Gruen, D. S., Lollar, B. S., Hinrichs, K. U., Stewart, L. C., Holden, J. F., Hristov,
 A. N., Pohlman, J. W., Morrill, P. L., Könneke, M., and Delwiche, K. B.: Nonequilibrium
 clumped isotope signals in microbial methane, Science, 348, 428–431, 2015.
- Wankel, S. D., Ziebis, W., Buchwald, C., Charoenpong, C., de Beer, D., Dentinger, J., Xu, Z.,
 and Zengler, K.: Evidence for fungal and chemodenitrification based N₂O flux from nitrogen
 impacted coastal sediments, Nat. Comm., 8, 1–11, 2017.
- Ward, B. B., Olson, R. J., and Perry, M. J.: Microbial nitrification rates in the primary nitrite
 maximum off southern-California, Deep-Sea Research, 29, 247–255, 1982.
- Weber, T., Wiseman, N. A., and Kock, A.: Global ocean methane emissions dominated by
 shallow coastal waters, Nat. Communications, 10, 1–10, 2019.
- 1155 Weinstein, A., Navarrete, L., Ruppel, C., Weber, T. C., Leonte, M., Kellermann, M. Y.,
- Arrington, E. C., Valentine, D. L., Scranton, M. I., and Kessler, J. D.: Determining the flux
 of methane into Hudson Canyon at the edge of methane clathrate hydrate stability, Geochem.
 Geophys. Geosys., 17, 3882–3892, doi:10.1002/2016gc006421, 2016.
- Weiss, R. F., F. A. Von Woy, and P. K. Salameh (1992), Surface water and atmospheric carbon dioxide and nitrous oxide Observations by shipboard automated gas chromatography: Results from expeditions between 1977 and 1990, Rep. S/0 92–11, Carbon Dioxide Inf. Anal. Cent., Oak Ridge Natl. Lab., Oak Ridge, Tenn.
- Wells, N. S., Maher, D. T., Erler, D. V., Hipsey, M., Rosentreter, J. A. and Eyre, B. D.: Estuaries
 as sources and sinks of N₂O across a land use gradient in subtropical Australia, Global
 Biogeochem. Cy., 32, 877–894, 2018.
- Whiticar, M. J.: Carbon and hydrogen isotope systematics of bacterial formation and oxidation of
 methane, Chem. Geology, 161, 291–314, 1999.
- Wilson, S. T., Ferrón, S., and Karl, D. M.: Interannual variability of methane and nitrous oxide in
 the North Pacific Subtropical Gyre, Geophys. Res. Lett., 44, 9885–9892,
- 1170 https://doi.org/10.1002/2017GL074458, 2017.
- Wilson, S.T., Bange, H.W., Arévalo-Martínez, D. L., Barnes, J., Borges, A., Brown, I., Bullister,
 J. L. et al.: An intercomparison of oceanic methane and nitrous oxide measurements,
 Biogeosci. 15, 5801, 5007, doi: 10,5104/bg.15,5801,2018,2018
- 1173 Biogeosci., 15, 5891–5907, doi: 10.5194/bg-15-5891-2018, 2018.
- 1174 Wyman, M., Hodgson, S., and Bird, C.: Denitrifying alphaproteobacteria from the Arabian Sea
- that express nosZ, the gene encoding nitrous oxide reductase, in oxic and suboxic waters,
 Appl. Environ. Microbiol., 79, 2670–2681, 2013.

- Yang, M., Bell, T. G., Hopkins, F. E., Kitidis, V., Cazenave, P. W., Nightingale, P. D., Yelland,
 M. J., Pascal, R. W., Prytherch, J., Brooks, I. M., and Smyth, T. J.: Air–sea fluxes of CO₂ and
 CH₄ from the Penlee Point Atmospheric Observatory on the south-west coast of the UK,
 Atmos. Chem. Physics, 16, 5745–5761, 2016.
- Yang, W.-B., Yuan, C.-S., Huang, B.-Q., Tong, C. and Yang, L.: Emission characteristics of
 greenhouse gases and their correlation with water quality at an estuarine mangrove
 ecosystem-the application of an in situ on-site NDIR monitoring technique, Wetlands, 38,
- 1184 723–738, 2018.
- Yang, S., Chang, B. X., Warner, M., Weber, T. S., Bourbonnais, A., Santoro, A. E., Kock, A.,
 Sonnerup, R., Bullister, J., Wilson, S. T., and Bianchi D.: New global reconstruction reduces
 the uncertainty of oceanic nitrous oxide emissions and reveals a vigorous seasonal cycle, P.
 Natl. Acad. Sci. USA, doi:10.1073/pnas.1921914117, 2020.
- Yoshida, N. and Toyoda, S.: Constraining the atmospheric N₂O budget from intramolecular site
 preference in N₂O isotopomers, Nature, 405, 330–334, 2000.
- Young, E. D., Kohl, I. E., Lollar, B. S., Etiope, G., Rumble III, D., Li, S., Haghnegahdar, M. A.,
 Schauble, E. A., McCain, K. A., Foustoukos, D. I. and Sutclife, C.: The relative abundances
 of resolved ¹²CH₂D₂ and ¹³CH₃D and mechanisms controlling isotopic bond ordering in
 abiotic and biotic methane gases, Geochim. Cosmochim. Acta, 203, 235–264, 2017.
- Yu, L., Harris, E., Lewicka-Szczebak, D., Barthel, M., Blomberg, M.R., Harris, S.J., Johnson,
 M.S., Lehmann, M.F., Liisberg, J., Müller, C., Ostrom, N.E., Six, J., Toyoda, S., Yoshida, N.,
 and Mohn, J.: What can we learn from N₂O isotope data?- Analytics, processes and
 modelling. Rapid Comms. Mass Spec. doi: 10.1002/rcm.8858, 2020.
- Yurganov L., Muller-Karger F., and Leifer I.: Methane increase over the Barents and Kara Seas
 after the autumn pycnocline breakdown: satellite observations, Adv. Polar Sci., 30, 382–390.
 doi: 10.13679/j.advps.2019.0024, 2019.
- Zamora, L. M. and Oschlies, A.: Surface nitrification: A major uncertainty in marine N₂O
 emissions, Geophys. Res. Lett., 2014GL060556, doi: 10.1002/2014gl060556, 2014.
- Zappa, C. J., Raymond, P. A., Terray, E. A., and McGillis, W. T.: Variation in surface turbulence
 and the gas transfer velocity over a tidal cycle in a macro-tidal estuary, Estuaries, 26, 1401–
 1415, 2003.
- Zhang, G., Zhang, J., Liu, S., Ren, J., Xu, J., and Zhang, F.: Methane in the Changjiang (Yangtze River) Estuary and its adjacent marine area: riverine input, sediment release and atmospheric fluxes, Biogeochemistry, 91, 71–84, 2008.
- Zhang, J., Zhan, L., Chen, L., Li, Y. and Chen, J.: Coexistence of nitrous oxide undersaturation
 and oversaturation in the surface and subsurface of the western Arctic Ocean, J. Geophys.
- 1212 Res., 120, 8392–8401, doi:10.1002/2015JC011245, 2015.
- 1213





1215 Figure 1. Atmospheric values of (a) CH₄ and (b) N₂O with the black lines reconstructed from

- 1216 ice-core measurements (Etheridge et al., 1998; Machida et al., 1995) and the colored lines from
- 1217 Mauna Loa Observatory (https://www.esrl.noaa.gov/gmd/dv/data/). Global maps of marine (c)
- 1218 CH₄ and (d) N₂O measurements available from the MEMENTO database
- 1219 (https://memento.geomar.de/). The 2018 workshop focused on the marine contribution to
- 1220 atmospheric CH_4 and N_2O and the underlying microbial and biogeochemical control
- 1221 mechanisms.





Figure 2. Repeat oceanic observations include both (a, b) fixed location time-series monitoring observations and (c,d) hydrographic surveys. Together, such field observation programs helps resolve temporal variability ranging from months to years and spatial variability at the ocean basin scale (see Fig. 3). The Station ALOHA data derive from Wilson et al. (2018), the Station 18 data derive from Farías et al. (2015), and the P16 transect was conducted in 2015 by the NOAA PMEL Tracer Group as part of the GO-SHIP program. The data shown in the plots are N_2O concentrations, either as ΔN_2O (i.e. deviation from equilibrium value) or absolute values.



1236

1237 Figure 3. Time-space scale diagram illustrating various physical, biological, and climatological 1238 processes relevant to marine CH₄ and N₂O (adapted from Dickey, 2003). To date, the majority of marine CH₄ and N₂O measurements resolve variability at the mesoscale level or higher. 1239 Recent technological developments and the need to resolve concentrations and fluxes in shallow 1240 1241 water environments will increase the number of measurements conducted at the submesoscale level (see Fig. 5). The low resolution oceanographic surveys are more likely to achieve a high 1242 level of analytical accuracy compared to high resolution coastal measurements, however this is 1243 compensated for by high temporal resolution achieved by underway sampling. 1244





Figure 4. Distributions and emissions of marine CH₄ and N₂O, (a) Air-sea N₂O disequilibrium 1247 mapped using a Regression Forest model (adapted from Yang et al., 2020), (b) Air-sea CH₄ 1248 disequilibrium mapped using an Artificial Neural Network model (adapted from Weber et al., 1249 2019). For consistency with the original publications, the air-sea disequilibrium is shown in 1250 1251 different units for N₂O (partial pressure) and CH₄ (concentration). (c) A summary of global ocean CH₄ and N₂O emissions estimated by Yang et al. (2020) and Weber et al. (2019), 1252 1253 compared to the estimates of the IPCC 5th Annual Report (IPCC AR5) and the Global Methane Budget (Saunois et al., 2016). 1254



Figure 5. Key environmental predictors of surface ocean CH_4 and N_2O gradients. (a) Excess airsea N_2O is best predicted by O_2 concentrations in the subsurface water-column (base of the mixed layer to a depth of 100 m) (adapted from Yang et al., 2020). (b) Excess CH_4 is best predicted by seafloor depth, reflecting the supply from anoxic sediments (adapted from Weber et al., 2019). The grey dots represent individual data points and the red dots with error bars represent mean ±1s.d. of binned data, using O_2 bins of 10 µM width and seafloor depth bins of 10 m width.