



1 **Ideas and perspectives: A strategic assessment of methane and nitrous oxide measurements**
2 **in the marine environment**

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48 **Abstract.** In the current era of rapid climate change, accurate characterization of climate-
49 relevant gas dynamics - namely production, consumption and net emissions - is required for all
50 biomes, especially those ecosystems most susceptible to the impact of change. Marine
51 environments include regions that act as net sources or sinks for a number of climate-active trace
52 gases including methane (CH₄) and nitrous oxide (N₂O). The temporal and spatial distributions
53 of CH₄ and N₂O are controlled by the interaction of complex biogeochemical and physical
54 processes. To evaluate and quantify the importance of these mechanisms relevant to marine CH₄
55 and N₂O cycling requires a combination of traditional scientific disciplines including
56 oceanography, microbiology, and numerical modeling. Fundamental to all of these efforts is
57 ensuring that the datasets produced by independent scientists around the world are comparable
58 and interoperable. Equally critical is transparent communication within the research community
59 about the technical improvements required to increase our collective understanding of marine
60 CH₄ and N₂O. An Ocean Carbon & Biogeochemistry (OCB) sponsored workshop was organized
61 to enhance dialogue and collaborations pertaining to marine CH₄ and N₂O. Here, we summarize
62 the outcomes from the workshop to describe the challenges and opportunities for near-future
63 CH₄ and N₂O research in the marine environment.

64



65

66 **1. Background**

67 The most abundant greenhouse gases in the troposphere, excluding water vapor, are carbon
68 dioxide (CO₂), methane (CH₄), and nitrous oxide (N₂O). Together they account for more than
69 80% of the total radiative forcing (IPCC, 2013) and their current tropospheric mole fractions and
70 rates of increase are unprecedented in recent Earth history (Ciais et al., 2013; Burke et al., 2020;
71 Fig. 1a). While CO₂ is the most abundant of the three greenhouse gases, CH₄ and N₂O both have
72 a higher warming potential than CO₂ (Montzka et al., 2011). To accurately constrain the
73 contribution of CH₄ and N₂O to Earth's radiation budget requires their sources and sinks to be
74 quantified with high resolution at the global scale.

75 The oceans are a fundamental component of the global climate system and are a net source of
76 tropospheric CH₄ and N₂O at the global scale, although local to regional budgets may include
77 both source and sink aspects. There are far fewer marine measurements of dissolved CH₄ and
78 N₂O than of dissolved CO₂ and while there is substantial international coordination with regard
79 to CO₂ analysis, calibration and data reporting, no such coordination yet exists for CH₄ and N₂O
80 (Wilson et al. 2018). Given the increasing prominence of climate change on scientific and
81 societal agendas, greater coordination among the marine CH₄ and N₂O scientific community to
82 provide more targeted measurements and increase the quality and interoperability of CH₄ and
83 N₂O observations is particularly timely.

84 Despite the lack of an international coordinating framework, there have been important
85 advances in our understanding of marine CH₄ and N₂O in numerous research disciplines, ranging
86 from cellular metabolism and model microbial systems to large-scale modeling. For example,
87 recent work identified novel microorganisms and metabolic pathways in the production of N₂O
88 (Trimmer et al., 2016; Caranto and Lancaster, 2017) and CH₄ (Repeta et al. 2016; Bižić et al.,
89 2020). Earth system models now incorporate improved N₂O parameterizations to better resolve
90 the ocean's role in the global N₂O cycle (Battaglia and Joos, 2018). New techniques enable the
91 discrimination of ancient and modern dissolved CH₄ (Sparrow et al., 2018) and the transfer of
92 CH₄-derived carbon to other carbon pools (Pohlman et al., 2011; Garcia-Tigreros and Kessler,
93 2018). Other technological and analytical advances include improved near-continuous
94 spectroscopic analysis that yield greater sampling resolution in surface waters (e.g. Gülzow et



95 al., 2011; Arévalo-Martínez et al., 2013; Erler et al., 2015) and the deployment of analytical
96 devices on robotic vehicles (Nicholson et al., 2018).

97 These scientific advances and an improvement in the quantity and quality of CH₄ and N₂O
98 observations are timely given that large areas of both the open and coastal ocean remain under-
99 sampled (Fig. 1b). This leads to uncertainty in oceanic CH₄ and N₂O inventories, their rates of
100 production and consumption, and their emissions. This is problematic given that the marine
101 environment is susceptible to an accelerating rate of anthropogenic change that will continue to
102 modify the global cycles of carbon and nitrogen into the future. Environmental impacts on
103 marine CH₄ and N₂O distributions include increasing seawater temperatures, decreasing
104 concentrations of dissolved oxygen (O₂), acidification, retreat of ice and mobilization of carbon
105 substrates from former permafrost, altering coastal run-off, and eutrophication (IPCC, 2019).
106 These impacts will undoubtedly alter future CH₄ and N₂O exchange with the atmosphere, but the
107 directions and magnitudes of these modified fluxes remains insufficiently understood.

108 A need to resolve the uncertainties prompted an evaluation of the collective ability of the
109 international scientific community to accurately determine the distribution and emissions of CH₄
110 and N₂O, and the physical-biogeochemical factors that determine them. This became the focus
111 of a marine CH₄ and N₂O workshop hosted by the Ocean Carbon and Biogeochemistry (OCB)
112 program at Lake Arrowhead, California in October 2018. The workshop considered CH₄ and
113 N₂O equally on the same agenda, even though nearly all field, laboratory, and modeling studies
114 examine these trace gases separately. The rationale for this dual approach is that CH₄ and N₂O
115 share common considerations of the physical, chemical, and microbial processes that dictate their
116 water-column distributions (Bakker et al., 2014; Bodelier and Steenbergh, 2014). In addition,
117 many of the analytical procedures for quantifying CH₄ and N₂O and the subsequent data quality
118 assurances share many common requirements. The opportunity to bring a large research
119 community together to increase dialogue and encourage the cross-fertilization of ideas was thus
120 considered very valuable. This article articulates the workshop outcomes framed in the context
121 of current marine CH₄ and N₂O research and explores future research opportunities and
122 challenges.

123

124 **2. Coordination of oceanic CH₄ and N₂O measurements**



125 Our understanding of the temporal and spatial distributions of oceanic CH₄ and N₂O derives
126 from over five decades of open ocean and coastal observations, including targeted expeditions,
127 repeat hydrographic surveys, and time-series monitoring, each of which has been crucial to the
128 development of our current knowledge (Fig. 2). Targeted programs have enabled invaluable
129 insights into the role of oxygen deficient zones in N₂O cycling (Babbin et al., 2015; Bourbonnais
130 et al., 2017; Frey et al., 2020) and the exploration of CH₄-rich seeps and vents (Foucher et al.,
131 2009; Suess, 2010; Boetius and Wenzhöfer, 2013). Basin-scale repeat hydrographic surveys
132 (e.g. the international GO-SHIP program) have facilitated extensive water-column mapping to
133 identify relevant water masses and evaluate ventilation rates (e.g. de la Paz et al., 2017). Other
134 oceanic surveys have focused exclusively on surface sampling, using continuous equilibrator
135 systems connected to various gas analyzers to yield high-resolution surface concentration fields
136 of CH₄ and N₂O (Gülzow et al., 2013; Erlen et al., 2015; Kodovska et al., 2016; Thornton et al.,
137 2016a; Pohlman et al., 2017). In contrast, sustained long-term time-series measurements of CH₄
138 and N₂O at fixed monitoring stations are relatively few, but they span a range of latitudes and
139 biogeochemical provinces (Fig. 2). The time-series observations provide the contextual
140 background for seasonal and interannual variation that allow long-term temporal trends and
141 episodic events to be identified and evaluated (Farías et al., 2015; Wilson et al., 2017; Ma et al.,
142 2019). Overall, the majority of measurements enable the variability in marine CH₄ and N₂O to
143 be quantified at the mesoscale or greater (i.e. from hundreds of kilometers to ocean basins), with
144 monthly to annual resolution but there are substantially fewer datasets at the sub-mesoscale level
145 (i.e. <10 km and hours to days) (Fig. 3). A major reason for the limited sampling at the sub-
146 mesoscale level is that it necessitates high-resolution measurements to resolve the heterogeneous
147 variability that exists at these time-space scales. Such analyses have only recently become
148 technically feasible (discussed in more detail in Section 6).

149 Until recently there has been no formal coordination of observations across the CH₄ and N₂O
150 scientific community. In response to this, a Scientific Committee on Oceanic Research (SCOR)
151 Working Group was initiated in 2014 entitled: '*Dissolved N₂O and CH₄: Working towards a
152 global network of ocean time series measurements*'. A major goal of the SCOR Working Group
153 was to unite the international community in joint activities conceived to improve and inform
154 seagoing CH₄ and N₂O analyses. An important activity was the preparation and distribution of
155 common, combined gaseous CH₄ and N₂O standards to twelve international laboratories, with



156 the aim of improving and standardizing calibration (Bullister et al., 2017). A subsequent inter-
157 comparison of discrete seawater samples included the use of these standards and revealed the
158 variability between laboratories. While there were some encouraging results from the
159 intercomparison, such as the agreement between individual laboratories using contrasting
160 techniques, overall a large range was observed in CH₄ and N₂O concentration data generated by
161 the participating laboratories (Wilson et al., 2018). Such analytical discrepancies weaken our
162 collective ability as a community to evaluate temporal-spatial variability in marine CH₄ and N₂O.
163 The discrepancies also highlighted the need for Standard Operating Protocols (SOPs) for CH₄
164 and N₂O analyses to facilitate standardization of sampling, measurement, and calibration, as well
165 as the reporting of data and accompanying metadata in common repositories. The SOPs are
166 currently in preparation with intended publication on the Ocean Best Practices network.

167 A data repository for oceanic CH₄ and N₂O data known as the MarinE Methane and Nitrous
168 Oxide database (MEMENTO) was established in 2009 (Bange et al., 2009; Kock and Bange,
169 2015). MEMENTO is now sufficiently mature to support descriptions of the broad-scale surface
170 distributions of CH₄ and N₂O (e.g. Suntharalingam et al., 2012; Zamora and Oschlies, 2014;
171 Buitenhuis et al., 2018; Battaglia and Joos, 2018). Machine-learning mapping recently identified
172 CH₄ and N₂O distributions and various physical and biogeochemical predictor variables (e.g.
173 depth, temperature, salinity, O₂, nutrients, primary production) (Weber et al., 2019; Yang et al.,
174 2020, Fig. 4). The application of gas transfer algorithms to the extrapolated oceanic CH₄ and
175 N₂O distributions helped decrease the uncertainty in estimates of global air-sea exchange fluxes
176 (Fig. 4c), thereby fulfilling one of the key goals of MEMENTO (Bange et al., 2009). Net global
177 open ocean emissions of N₂O are now similarly estimated at 3–5 Tg N yr⁻¹ by both Yang et al.
178 (2020) and the Global Nitrous Oxide Project (Tian et al., 2020). In comparison, net global ocean
179 CH₄ emissions from machine-learning mapping were estimated at 6–12 Tg CH₄ yr⁻¹ (Weber et
180 al., 2019), compared to 9–22 Tg CH₄ yr⁻¹ in the most up-to-date CH₄ synthesis (Saunio et al.,
181 2020). However, the narrower range for machine-learning derived CH₄ emissions retains high
182 uncertainty in regions such as the Arctic, where emissions are highly heterogeneous and
183 compounded by seasonal ice cover. Identifying the causes for uncertainty in high emission
184 regions will greatly aid future sampling campaigns, as is discussed in the following sections.

185

186 3. Methane in marine environments



187 In the surface waters of tropical and temperate oceans, the low supersaturation of CH₄ is
188 driven by aerobic production arising from the decomposition of methyl phosphonate in
189 phosphorus-depleted waters (Karl et al. 2008, Sosa et al., 2020), the degradation of methylated
190 sulfur compounds by phytoplankton (Klitzsch et al., 2019), and other processes (Schmale et al.,
191 2018). Deep within the ocean's pelagic interior, CH₄ is weakly undersaturated reflecting
192 depletion via microbial oxidation (Reeburgh 2007; Weber et al., 2019). Towards the coastline,
193 CH₄ supersaturation increases by orders of magnitude (Figure 5b), reflecting terrestrial inputs
194 (e.g. river and groundwater) and CH₄ diffusion and ebullition from shallow anoxic methane rich
195 sediments (Zhang et al., 2008; Borges et al., 2016; Upstill-Goddard and Barnes, 2016).
196 Supersaturation of CH₄ occurs frequently in the Arctic Ocean and its relatively shallow marginal
197 seas with the most extreme values observed in the Eurasian Arctic (e.g. Shakhova et al., 2010;
198 Damm et al., 2015; Kosmach et al., 2015; Thornton et al., 2016a; Fenwick et al., 2017).
199 Terrestrial and subsea permafrost are potential CH₄ sources to shelf waters in addition to CH₄
200 hydrates that are found in marginal shelves globally (Ruppel and Kessler, 2017). Large point
201 source CH₄ emissions, such as seafloor gas seeps can be large sources to the atmosphere in small
202 localized areas (e.g. Thornton et al., 2020), but these sites remain particularly difficult to
203 parameterize in models. This reflects limited observations and a poor understanding of their
204 spatial distributions, the driving mechanisms, and the wider context within the carbon cycle. For
205 example, the upwelling of cold, nutrient-rich water that accompanies CH₄ ascending the water
206 column stimulates CO₂ consumption by photosynthesizing phytoplankton, rendering such CH₄
207 seeps an overall net sink for climate-forcing gases (Pohlman et al., 2017). Recent work using
208 thermal infrared satellite retrievals indicates increased high-latitude oceanic CH₄ release in late
209 autumn, coincident with pycnocline breakdown and a deepening of the ocean mixed layer depth
210 thereby bringing deep CH₄ to the surface (Yurganov et al., 2019). This is especially notable in
211 the Kara and Barents Seas, but the remote observations have not yet been confirmed by surface
212 ocean measurements which are difficult and therefore rare, except during the Arctic summer.

213 Seabed CH₄ emissions are hypothesized to increase in a warming ocean through the
214 decomposition of gas hydrates, the degradation of subsea permafrost under some high-latitude
215 seas, and the increased biodegradation of sediment carbon (Romanovskii et al., 2005; Biastoch et
216 al., 2011; Ruppel and Kessler, 2017; Borges et al., 2019). Effort is thus focused on quantifying
217 the fraction of CH₄ generated in or released from marine sediments that ultimately enters the



218 atmosphere, particularly on shallow continental shelves and in coastal ecosystems. Natural
219 stable isotopes have been used to inform spatial and temporal changes in dissolved CH₄
220 concentrations (e.g. Pack et al., 2011; Mau et al., 2012; Weinstein et al., 2016; Leonte et al.,
221 2017; Chan et al., 2019) and incubation experiments with added stable isotopes and radiotracers
222 have helped elucidate how oxidation (anaerobically in sediments and aerobically in the water
223 column), ebullition (where CH₄ pore water partial pressure exceeds sediment hydrostatic
224 pressure), and subsequent bubble dissolution in the water column interact to mitigate CH₄
225 emissions to air (Steinle et al., 2015; Jordan et al., 2020). The information deriving from these
226 various approaches is inherently different but complementary. Isotope tracer incubations provide
227 snapshots of rates specific to the methanotrophic community and CH₄ concentration at the time
228 of sampling, whereas concentrations and isotopic gradients are used to infer *in situ* rates
229 integrated over space and time. A recent study deployed a remotely operated vehicle to examine
230 the isotopic fractionation of CH₄ during bubble ascent and used this to constrain the extent of
231 bubble dissolution (Leonte et al., 2018). This work demonstrated an experimental approach
232 established for broadly constraining water column CH₄ cycling directly from a surface research
233 vessel.

234 Despite the range of analytical and experimental approaches available, determining whether
235 the origin of the emitted CH₄ is seafloor release or aerobic production in the upper water column
236 remains problematic. To date there is no straightforward way to routinely distinguish between
237 seafloor derived and water column generated CH₄ for all locations. Even so, stable carbon and
238 hydrogen isotope measurements (i.e. $\delta^{13}\text{C-CH}_4$ and $\delta^2\text{H-CH}_4$) combined with ancillary data may
239 provide valuable source information. For example, combining these measurements with the ratio
240 of CH₄ to higher order hydrocarbons (e.g. ethene (C₂H₄) and ethane (C₂H₆)) can be used to infer
241 for example, whether the origin of the CH₄ is thermogenic, sub-seafloor, or biogenic within the
242 water column (Whiticar, 1999; Pohlman et al., 2009; Lan et al., 2019). Continuous shipboard
243 measurement of CH₄ isotopes in surface water (e.g. Pohlman et al., 2017) and in the atmospheric
244 boundary layer (Pankratova et al., 2019; Berchet et al., 2020) are now possible and they have
245 been used in combination with atmospheric inversion models to characterize and discriminate
246 marine-emitted CH₄ from other sources (Berchet et al., 2020). Application of this method to
247 land-based monitoring stations appears promising for apportioning CH₄ emissions from various
248 marine regions and sources (Thonat et al., 2019). Additionally, in regions where aerobic CH₄



249 oxidation is substantial, the resulting isotopic fractionation generates measurable vertical and/or
250 horizontal seawater gradients that can also be used to identify contrasting biogenic CH₄ sources
251 (Leonte et al., 2020). However, the general overlap in isotope compositions of sediment CH₄
252 (e.g. Thornton et al., 2016b; Sapart et al., 2017) can complicate purely isotope-based
253 determinations of sources.

254 Measurements of the natural radiocarbon content of dissolved oceanic CH₄, while being
255 highly specialized and requiring substantial amounts of ship time and processing (Kessler and
256 Reeburgh, 2005; Sparrow and Kessler, 2017), provide valuable source information because the
257 ¹⁴C-CH₄ measurements are normalized to the same δ¹³C value and are unaffected by the extent
258 of oxidation. The bubbles sampled from hydrate and active seafloor seeps are largely devoid of
259 radiocarbon (Pohlman et al., 2009; Kessler et al., 2008; Douglas et al., 2016). However, CH₄ in
260 sediments can also be derived from more modern or recently deposited organic material and an
261 exact determination of individual contributions is hard to achieve (Kessler et al., 2008; Sparrow
262 et al., 2018). The powerful insights made by radiocarbon-CH₄ investigations would be further
263 strengthened by concurrent sampling of other analytes that offer CH₄ source information, such as
264 clumped isotopes. Isotope clumping, the co-occurrence of two or more of the less abundant
265 isotopes in a molecule (e.g. ¹³C and ²H or ¹H and ²H), provides unique information on marine
266 CH₄ sources (Stolper et al., 2014; Wang et al., 2015; Douglas et al., 2017; Young et al., 2017;
267 Giunta et al. 2019). In this approach, the isotopic deviations in samples from their random
268 probability distributions can give insight into formation temperature and the extent of
269 biochemical disequilibrium. However, the sample size required for a clumped isotope analysis
270 in the oceanic environment away from areas of seafloor emission is large and exceeds the
271 already demanding volume requirements for ¹⁴C analyses by 1–2 orders of magnitude (Douglas
272 et al., 2017). While the requirement of large sample size and lengthy measurement time
273 currently preclude their more widespread application, clumped isotope measurements offer
274 future promise in refining our understanding of the processes of marine CH₄ production and
275 consumption.

276

277 **4. Nitrous oxide in marine environments**

278 The large-scale spatial distribution of N₂O in the global ocean is reasonably well-established.
279 The highest open ocean N₂O values are in upwelling environments, where concentrations extend



280 up to micromolar levels (Arévalo-Martínez et al., 2015) and production rates can be as high as
281 120 nM d^{-1} (Frey et al., 2020). The highly elevated N_2O concentrations can be proximal to
282 regions with some of the lowest recorded N_2O concentrations, in the cores of O_2 deficient zones.
283 This coexistence of the highest and lowest observed N_2O concentrations over vertical distances
284 of tens of meters make upwelling regions a focal point for N_2O research, particularly since O_2
285 deficient ocean zones are increasing in size (Stramma et al., 2011). In contrast, in the surface
286 waters of the expansive oligotrophic ocean gyres, N_2O is weakly supersaturated (103-105%)
287 with respect to atmospheric equilibrium (Weiss et al., 1992; Wilson et al., 2017, Charpentier et
288 al., 2010). Nitrous oxide becomes more highly saturated in the surface waters of equatorial
289 upwelling regions due to the upward advection of N_2O -rich waters (Arévalo-Martínez et al.,
290 2017). For the Arctic Ocean, the data indicate low net N_2O emissions, with some areas acting as
291 net N_2O sources and others as N_2O sinks (Fenwick et al., 2017, Zhang et al., 2015).

292 Several parameters control net N_2O emissions from the ocean, including temperature,
293 salinity, dissolved O_2 , apparent oxygen utilization (AOU), nutrients, and microbial community
294 abundance and composition. A recent modeling study trained with just three of these variables
295 (chlorophyll, O_2 , and AOU) accounted for 60% of the observed variability in oceanic N_2O
296 concentrations (Yang et al., 2020), highlighting the importance of N_2O in productive upwelling
297 systems. Correlations between N_2O and environmental variables provide some insight into the
298 factors controlling its distribution, but they provide no information about the microorganisms or
299 metabolic pathways involved. Microbial production of N_2O occurs during the metabolic
300 processes of nitrification and denitrification (Stein and Yung, 2003). To determine which
301 process dominates N_2O production at any given location requires the application of multiple
302 methodological approaches, ideally in parallel.

303 One of the most commonly used approaches is the incubation of discrete water samples
304 under *in situ* conditions with stable isotope (^{15}N) addition such as ^{15}N enriched NH_4^+ , NO_2^- or
305 $\text{NO}_2^-/\text{NO}_3^-$ to measure N_2O production rates from nitrification and denitrification, respectively
306 (e.g. Ji et al., 2017). These approaches also provide insight into the microorganisms involved.
307 For example, N_2O resulting from archaeal NH_4^+ oxidation is mostly formed from a combination
308 of NH_4^+ and another N compound (e.g. NO_2^-) whereas bacteria produce N_2O from NH_4^+ alone
309 (Santoro et al., 2011, Stieglmeier et al., 2014; Carini et al. 2018; Lancaster et al., 2018; Frey et
310 al. 2020). Unfortunately, as with all incubation-based approaches ^{15}N techniques are subject to



311 bottle artifacts, and the strong dependence of N₂O production and consumption on ambient O₂
312 increases the potential for contamination during the collection and manipulation of anoxic deep
313 seawaters. Incubation based rate measurements are also compromised by abiotic N₂O
314 production via chemodenitrification, specifically the reduction of NO₂⁻ coupled to Fe²⁺
315 oxidation, as observed in high Fe environments (Ostrom et al., 2016; Buchwald et al., 2016;
316 Wankel et al., 2017). These issues highlight the need for incubation techniques that mitigate the
317 effect of experimental artifacts (Stewart et al., 2012).

318 In addition to isotope addition and incubation, natural abundance water-column
319 measurements of N₂O concentrations, isotopes, and isotopomers yield valuable rate and process
320 information. These measurements are free from experimental artifacts and can be used to
321 integrate over appropriate temporal and spatial scales. For example, nitrification in sunlit waters
322 has been inferred from N₂O distributions (Dore and Karl, 1996), and N₂O production close to the
323 ocean surface is a large contributor to the uncertainty in oceanic N₂O emissions (Ward et al.,
324 1982; Zamora and Oeschlies, 2014). Isotopomers are isomers having the same number of each
325 isotope of each element but differing in their structural positions. Nitrous oxide isotopomers are
326 increasingly used, sometimes in combination with box models, to estimate the rates of different
327 N₂O production pathways, in the upwelling systems off southern Africa (Frame et al., 2014) and
328 Peru (Bourbonnais et al., 2017). There is however some disagreement about whether isotopomer
329 signatures are robust indicators of the formation pathway (Yoshida and Toyoda, 2000; Sutka et
330 al., 2006) or whether there is fractionation during production (Schmidt et al., 2004; Casciotti et
331 al., 2018). Greater clarity is therefore required in the use of N₂O isotopes and isotopomers to
332 infer metabolic pathways of N₂O formation. Notwithstanding this issue, field measurements of
333 N₂O isotopes and/or isotopomers have the potential to greatly increase current experimental
334 capabilities and robustness (Yu et al., 2020). However, the development of spectroscopic gas
335 analysis systems that have been so advantageous to CH₄ research has been slower for N₂O. This
336 is due to the higher costs and the increased complexity of the laser systems, although progress is
337 being made to improve instrumental precision, and to decrease matrix effects and spectral
338 interferences (e.g. Harris et al., 2019).

339 A better understanding of the microorganisms responsible for N₂O production and
340 consumption is fundamental to deriving more accurate estimates of process rates. For example,
341 the metabolic activity of ammonia oxidizing archaea can exceed that of ammonia oxidizing



342 bacteria in the ocean (Santoro et al., 2010; Löscher et al., 2012; Fuchsman et al., 2017). The
343 differing sensitivities of these archaea and bacteria to dissolved O₂ (Stahl and de la Torre, 2012;
344 Hink et al., 2017) are a critical factor in evaluating the microbial response to changing
345 environmental conditions, as shown for the terrestrial environment (Prosser et al., 2020).
346 Therefore, to understand the impact of deoxygenation on oceanic N₂O emission requires a better
347 understanding of both archaeal and bacterial metabolisms and their environmental niches. Field-
348 based sequencing not only characterizes the community but can highlight potential metabolic
349 pathways when they might not otherwise be inferred. For example, transcripts encoding for N₂O
350 consumption (*nosZ*) have repeatedly been identified in the oxic water column, despite
351 denitrification being an anaerobic metabolic process (Wyman et al., 2013; Sun et al., 2017). The
352 transcription of *nosZ* has been also located in highly dynamic O₂ permeable coastal sediments
353 (Marchant et al., 2017). Denitrification under aerobic conditions is attributed to fluctuations in
354 O₂, NO₃⁻, organic matter and other parameters that affect the availability of electron donors and
355 acceptors which ultimately influences whether a coastal environment is a net source or sink of
356 N₂O, as discussed in the next section.

357

358 **5. CH₄ and N₂O in shallow marine environments**

359 Coastal and other shallow (<50 m) marine systems are globally relevant CH₄ and N₂O source
360 regions. However, their emission rates to the atmosphere are weakly constrained in comparison
361 with the open ocean. Several factors contribute to the uncertainty, including the high diversity of
362 coastal and shallow marine ecosystems and lack of consistency in adequately defining them,
363 locally heterogeneous conditions causing strong spatial and temporal concentration gradients,
364 highly uncertain spatial distribution of CH₄ seeps, a bias towards studies in the northern
365 hemisphere, and incomplete or sometimes inappropriate sampling strategies (Al-Haj and
366 Fulweiler, 2020). Until these issues are resolved it will remain difficult to adequately define the
367 contribution from shallow marine systems to global CH₄ and N₂O budgets. An important
368 illustration of this is reflected in the prevailing view that large geological sources (e.g. seeps,
369 mud volcanoes, and hydrates) are the main contributors to marine CH₄ emissions (Ciais et al.,
370 2013). The most recent modeled estimate of global marine CH₄ emissions (6–12 Tg CH₄ yr⁻¹)
371 reported that near-shore environments (depths of 0–50 m) contribute a large and highly uncertain
372 diffusive flux (Weber et al., 2019). A study of coastal ecosystems, in this case defined as shelf,



373 estuarine, and tidally influenced rivers, estimated them to contribute 7 Tg CH₄ yr⁻¹ (Anderson et
374 al., 2010) while another estimated 1–7 Tg CH₄ yr⁻¹ for estuaries alone (Borges and Abril, 2011).
375 Similar uncertainties exist for N₂O. Estimates of coastal N₂O emissions (which include coastal,
376 estuarine, and riverine sources) range from 0.1–2.9 Tg N yr⁻¹ (Ciais et al., 2013), although a
377 recent review of N₂O production across a range of estuarine habitats placed N₂O fluxes at the
378 lower end of these estimates (0.17–0.95 Tg N yr⁻¹) (Murray et al., 2015). Based on these data,
379 coastal systems account for around one third of total marine N₂O emissions (Yang et al., 2020).

380 The direct quantification of CH₄ and N₂O emissions from shallow coastal ecosystems has
381 historically involved using gas concentrations measured in discrete water and air samples
382 combined with a gas transfer velocity (k_w). For the coastal and open ocean, the dominant driver
383 of gas exchange is wind speed (e.g. Nightingale et al., 2000; Wanninkhof, 2014) whereas in
384 nearshore, shallow water environments the interaction of water, depth, and tidal current speeds
385 may be a major contributor to near surface turbulence. Several k_w parameterizations are now in
386 use for coastal waters (e.g. Raymond and Cole 2001; Kremer et al., 2003; Zappa et al., 2003;
387 Borges and Abril, 2011; Ho et al. 2011; Rosentreter et al., 2017; Jeffrey et al., 2018) which
388 increases the uncertainties associated with CH₄ and N₂O emissions. For example, a fivefold
389 variation in CH₄ emissions from a single system occurred when applying different
390 parameterizations to the measured gradients in CH₄ (Ferrón et al., 2007).

391 In order to constrain emissions over small areas continuous air-sea fluxes can be measured
392 using free-floating chambers (e.g. Bahlmann et al., 2015; Rosentreter et al., 2018; Yang et al.,
393 2018; Murray et al., 2020), but issues related to turbulence modification may still generate flux
394 artifacts (Upstill-Goddard, 2006). To overcome these problems in the future, a greater reliance
395 on direct and robust continuous techniques for air-sea flux measurement, such as eddy
396 covariance (e.g. Podgrajsek et al., 2016) that avoids any need for k_w , will be necessary.
397 Combining this with new analytical techniques such as cavity enhanced absorption spectroscopy
398 (CEAS) and non-dispersive infrared (NDIR) should continue to improve the quality of such
399 estimates (McDermitt et al., 2011; Nemitz et al., 2018; Maher et al., 2019). Indeed, eddy flux
400 towers aboard ships (Thornton et al., 2020) and in coastal locations (Yang et al., 2016; Gutiérrez-
401 Loza et al., 2019) are now being equipped with CH₄ instrumentation that enables the integration
402 of CH₄ fluxes over large areas. There are fewer N₂O flux estimates made with CEAS and NDIR
403 and the implementation of N₂O sensors on eddy flux towers remains limited. Recently, N₂O



404 emissions from three major Eastern Boundary Upwelling Systems were quantified using
405 atmospheric measurements from coastal monitoring stations highlighting their ability to attain
406 multi-year time-series measurements (Ganesan et al., 2020).

407 Flux towers at fixed locations provide a stable instrument platform and facilitate the
408 collection of ancillary data such as water-column depth, tidal motions (Rosentreter et al., 2018;
409 Huang et al., 2019; Pfeiffer-Hebert et al., 2019), and other information relating to diel processes
410 (Maher et al., 2016). Such data are important because for example, the magnitude of CH₄ and
411 N₂O fluxes vary over a diel period depending on the redox environment as a result of tidal
412 effects and changes in inorganic N and O₂ availability (Seitzinger and Kroeze, 1998; Call et al.,
413 2015; Vieillard and Fulweiler, 2014; Maher et al., 2015; Murray et al., 2015; Foster and
414 Fulweiler, 2019). The magnitude of CH₄ and N₂O fluxes also varies over longer temporal scales
415 (seasonally to yearly) due to additional factors such as groundwater inputs, adjacent land-use,
416 dissolved O₂, organic matter content and quality, and macrofaunal distributions (Barnes and
417 Upstill-Goddard, 2011; Upstill-Goddard and Barnes, 2016; Gelesh et al., 2016; Bonaglia et al.,
418 2017; Borges et al., 2018; Wells et al., 2018; Ray et al., 2019; Al-Haj and Fulweiler, 2020;
419 Reading et al., 2020). To determine the contributing factors and resolve the spatial distributions,
420 mobile sampling platforms such as small vessels (Müller et al., 2016; Brase et al., 2017; Tait et
421 al., 2017), and autonomous vehicles (Manning et al., 2019) are essential. Recent improvements
422 in gas sensors and in technology such as sonar and ebullition sensors will further increase our
423 ability to measure dynamic fluxes (Maher et al., 2019; Lohrberg et al., 2020). Improvements to
424 the quality and quantity of CH₄ and N₂O measurements in coastal systems will enable the
425 development of iterative forecast models, further improving estimates of global coastal CH₄ and
426 N₂O fluxes.

427

428 **6. Leveraging culture studies to further our ecosystem understanding**

429 A more complete understanding of marine CH₄ and N₂O necessitates closer integration between
430 biogeochemistry, model requirements, and targeted microbiological studies involving both single
431 microorganism isolates and enrichment cultures. Marine CH₄ and N₂O budgets deriving from
432 both ‘bottom-up’ (e.g. emissions inventories, ocean and terrestrial process models) and ‘top-
433 down’ (e.g. inverse analyses of atmospheric trace-gas measurements) approaches would greatly
434 benefit from more highly constrained metabolic processes. Specifically, this includes rates of



435 CH₄ or N₂O production and consumption for key model microorganisms, and the kinetic
436 parameters associated with these metabolic rates. Reliable inventories of key microbially
437 mediated process rates will improve the robustness of Earth System models used for predicting
438 climate-mediated changes to marine CH₄ and N₂O emissions.

439 For N₂O, laboratory studies quantifying microbial process rates, such as for nitrification and
440 denitrification, are relatively few (e.g. Frame and Casciotti 2010; Santoro et al. 2011; Löscher et
441 al. 2012; Ji et al. 2015; Qin et al., 2017). Consequently, models largely continue to use process
442 rates optimized using water column concentrations of N₂O, O₂, and related nitrogen cycle
443 quantities (e.g. Battaglia and Joos, 2018; Buitenhuis et al., 2018; Landolfi et al., 2017). Future
444 model parameterizations for N₂O will require information on the variability of microbial process
445 yields derived from culture studies with controlled varying conditions of O₂ (Goreau et al. 1980,
446 Frame and Casciotti 2010, Löscher et al. 2012; Ji et al., 2018), pH (Breider et al., 2019; Hopkins
447 et al. 2020), temperature, and nutrients. Automated incubation systems have measured N₂O
448 production kinetics and yield as functions of the concentrations of O₂ and total ammonia nitrogen
449 (Molstad et al., 2007; Hink et al., 2017). Quantifying the physiology of relevant microorganisms
450 and connecting them to environmental characteristics will provide insights into why, for
451 example, some shallow marine habitats act as N₂O sinks while others are N₂O sources, or how
452 N₂O is produced in well oxygenated open-ocean waters, as compared to oxygen deficient zones.

453 For CH₄, a key requirement to relate *in situ* CH₄ production with transport to atmospheric
454 emissions is our ability to accurately determine rates of CH₄ oxidation. Fundamental issues
455 include the challenges of cultivating methanotrophs and of replicating environmental conditions
456 such as pressure and the chemistry of CH₄ gas bubbles. The increased emphasis on CH₄
457 dynamics in shallow water environments highlighted in Section 5, must be supported by culture-
458 based measurements of CH₄ oxidation that control for temperature, O₂ and other important
459 variables. In comparison to CH₄ oxidation, culture-based studies are used increasingly to
460 identify organisms capable of aerobic CH₄ production and their underlying metabolic pathways
461 (Carini et al., 2014; Klintzsch et al; 2019; Bižić et al., 2020).

462 Specific cellular yields and consumption rates of CH₄ and N₂O are not the sole objective of
463 culturing experiments. Cultivation of microorganisms involved in CH₄ and N₂O production and
464 consumption provides vital information into the physiology, metabolism, and interactions of
465 environmentally relevant clades. When combined with genomic approaches, insights can



466 therefore be gained into the diversity and global distribution of organisms involved in CH₄ and
467 N₂O cycling. For CH₄ some unexpected physiologies have been revealed (Ettwig et al., 2010;
468 Haroon et al., 2013; Ettwig et al., 2016), which has directed research into sources and sinks of
469 CH₄ in the natural environment. Similarly, our understanding of how and when ammonia
470 oxidizers produce N₂O has been facilitated by studies of cultured nitrifiers and detailed analysis
471 of their biochemistry (Stahl and de la Torre, 2012; Caranto and Lancaster, 2017). Recent
472 combinations of cultivation studies with environmental genomics, albeit largely for terrestrial
473 systems, have revealed a variety of denitrifiers, many of which are only involved in specific
474 denitrification steps (Ganesh et al., 2014; Lycus et al., 2017; Hallin et al., 2018; Marchant et al.,
475 2018; Conthe et al., 2019).

476

477 **7. Outlook and priorities for marine CH₄ and N₂O measurements**

478 This perspectives article has assessed the collective ability of the scientific community to
479 determine the spatial variability of marine CH₄ and N₂O distributions, the underlying
480 mechanisms that determine this variability, and the resulting sea-to-air emissions. Shallow
481 marine environments and oxygen deficient zones are widely recognized as deserving of greater
482 attention because they have high CH₄ and N₂O concentrations with inherently high uncertainties
483 that complicate any assessment of their emissions to air (Bange et al., 1994; Bange et al., 1996;
484 Bakker et al., 2014; James et al., 2016; Borges et al., 2016; Tian et al., 2020). Fortunately, recent
485 technological advances that have increased our ability to conduct high-resolution measurements
486 allow an optimistic outlook for making substantial progress in quantifying the CH₄ and N₂O
487 budgets of these ecosystems. Even so, the inherent complexity of shallow marine environments
488 clearly warrants a strategically coordinated approach to optimize the value of future studies.
489 Issues to consider include identifying the locations of complementary sampling sites,
490 standardizing sampling strategies and techniques, and agreeing the use of common ancillary
491 measurements that set the broad biogeochemical context (Bange et al., 2019). In contrast to the
492 open ocean, measurement campaigns in shallow water environments are amenable to the use of
493 eddy covariance flux towers, and they have the potential to lever resources from existing
494 observation networks, which in North America include the Long-Term Ecological Research
495 network (LTER) and the National Estuarine Research Reserve (NERR) System (Novick et al.,
496 2018). Indeed, such activities are already underway; an increasing number of flux towers are



497 being equipped for CH₄ measurements (Torn et al., 2019) and future efforts should focus on the
498 inclusion of N₂O (see Section 5).

499 We are encouraged that the Global Carbon Project with its objective of developing a
500 complete picture of the global carbon cycle including interactions and feedbacks has expanded to
501 include CH₄ (Saunio et al., 2020) and is now incorporating N₂O (Tian et al., 2020). These
502 Projects compile the most recent data from peer-reviewed analyses of the sources and sinks of
503 atmospheric CH₄ and N₂O from both natural and human activities. For example, the aquatic
504 components of the recent Global Carbon Project N₂O budget reported emissions from the open
505 ocean, inland waters, estuaries and coastal zones. Low-oxygen oceanic regions associated with
506 eastern-boundary upwelling zones, and the coastal ocean were identified as key regions with
507 significant N₂O variability requiring more detailed assessment via measurement campaigns and
508 model analyses (Tian et al., 2020). Coordinating with global initiatives such as the Global
509 Carbon Project and identifying other areas of synergistic CH₄ and N₂O research of mutual
510 benefit to oceanographers and scientists studying other biomes serve to strengthen the scientific
511 achievements of all involved (Ganesan et al., 2019). Furthermore, as highlighted in Section 6,
512 field observations alone are insufficient to improve the robustness of Earth System models and
513 leveraging laboratory-based microbial process studies is highly recommended.

514 The success of any coordinated CH₄ and N₂O research program relies heavily on having
515 uniformly high confidence in the various resulting datasets and their interoperability, and we
516 identify three key initiatives that are paramount to ensuring this:

517 (i) The first is to develop and adopt Standard Operating Protocols (SOPs) to help obtain
518 intercomparable CH₄ and N₂O datasets of the highest possible accuracy and precision. In our
519 recent marine CH₄ and N₂O inter comparison exercise we concluded that the diversity of
520 analytical procedures employed by the participants was a major cause of high variability between
521 the reported concentrations, highlighting an urgent requirement for CH₄ and N₂O SOPs (Wilson
522 et al., 2018). Consequently, these SOPs are now being compiled, and they will be freely
523 available via the Ocean Best Practices System.

524 (ii) The second is the regular, routine inter comparison of measurements, by periodically
525 distributing to the community “consensus material”, i.e. water samples in which CH₄ and N₂O
526 concentrations are known with high confidence, obtained by pooling analyses from several
527 laboratories with demonstrated analytical capability. These will allow us to routinely monitor



528 data inter comparability and accuracy, particularly in the case of highly elevated concentrations
529 of CH₄ and N₂O, i.e. those exceeding atmospheric equilibrium concentrations by at least an order
530 of magnitude.

531 (iii) The third activity is increased use and support for MEMENTO. Until now the main
532 function of MEMENTO has been as a data repository. In this regard, it has been very valuable in
533 supporting the modeling components of CH₄ and N₂O research (see Section 3). We encourage a
534 much more widespread, routine, use of this data facility, with submitted data produced according
535 to the SOPs and inter comparison procedures. To maintain its relevance, MEMENTO must
536 continue to build its activities and develop into an ‘ocean CH₄ and N₂O Atlas’. The international
537 marine carbon science community has widely embraced such an approach for CO₂, by
538 submitting data to the Surface Ocean CO₂ Atlas (SOCAT), which was initiated in response to the
539 need for a quality controlled, publicly available, global surface CO₂ dataset (e.g. Bakker et al.,
540 2016). We believe establishing a similar data product for marine CH₄ and N₂O to be essential
541 for supporting future global modeling efforts and to enhance and reward community
542 engagement.

543 The benefits of pursuing the three activities described above have already been clearly
544 demonstrated for carbon system measurements in the ocean. The intercomparability and high
545 accuracy and precision of carbon system measurements was achieved by streamlining
546 methodological approaches, universally adopting agreed SOPs, production of reference material,
547 and following community-driven quality control procedures (Dickson et al., 2007, Dickson et al,
548 2010). It is encouraging to see the marine CH₄ and N₂O community beginning to move in a
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550

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562

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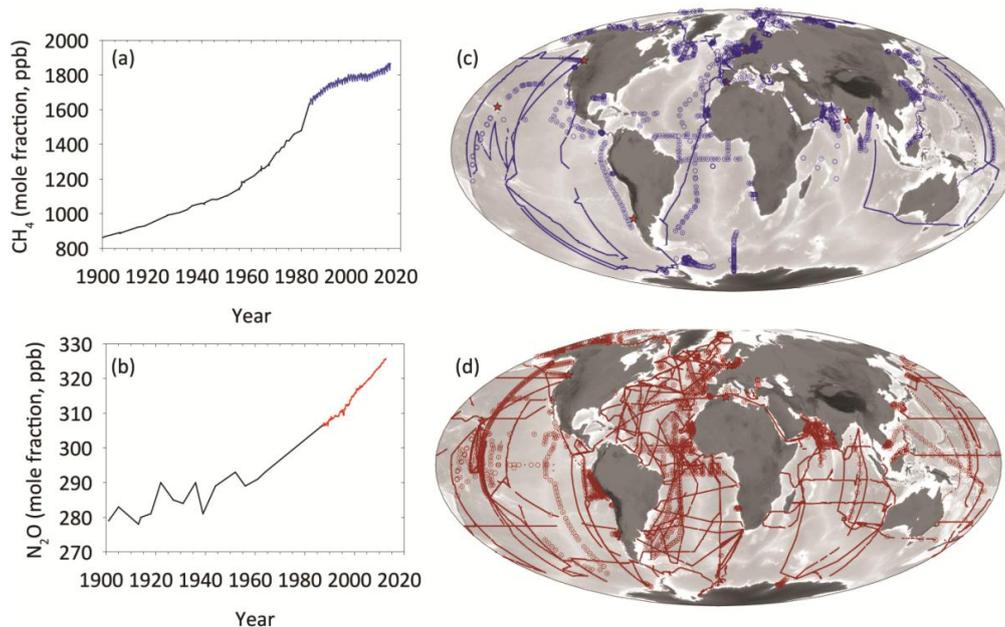
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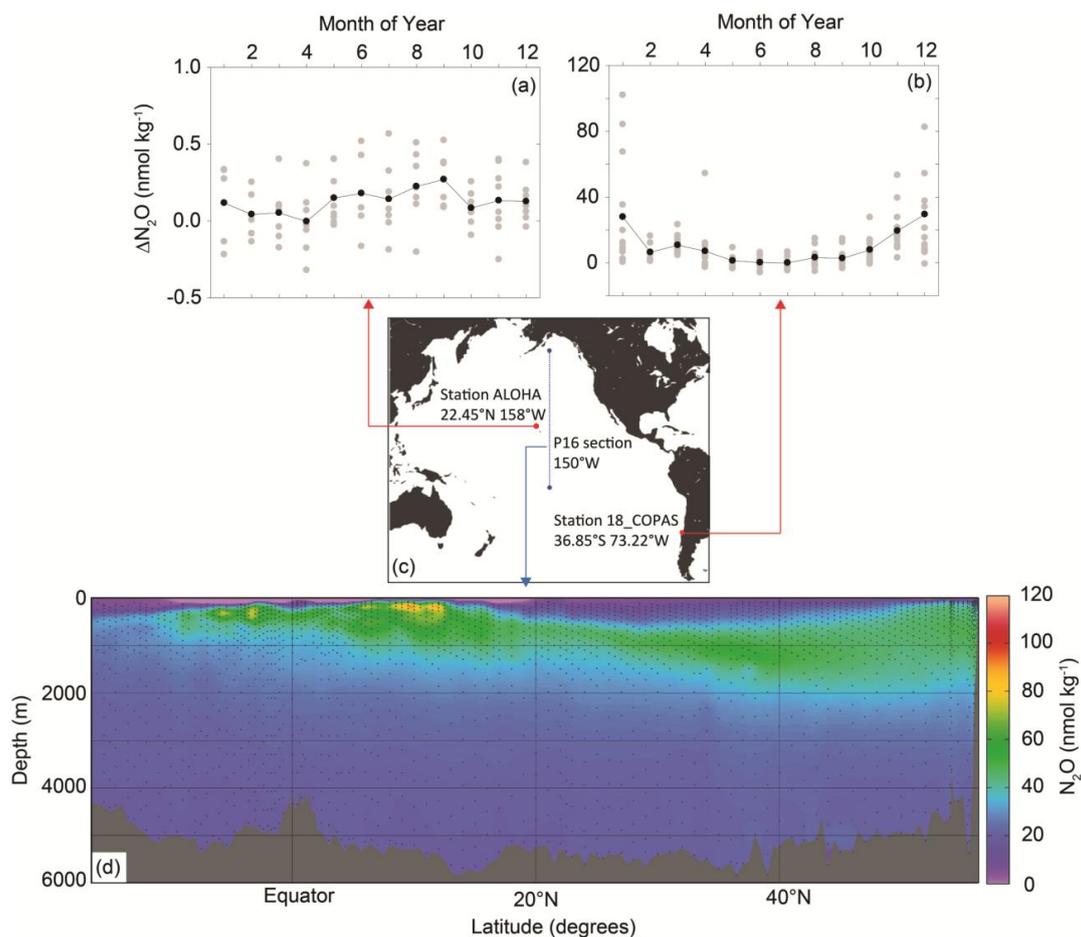
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1175 Figure 1. Atmospheric values of (a) CH₄ and (b) N₂O with the black lines reconstructed from
1176 ice-core measurements (Etheridge et al., 1998; Machida et al., 1995) and the colored lines from
1177 Mauna Loa Observatory (<https://www.esrl.noaa.gov/gmd/dv/data/>). Global maps of marine (c)
1178 CH₄ and (d) N₂O measurements available from the MEMENTO database
1179 (<https://memento.geomar.de/>). The 2018 workshop focused on the marine contribution to
1180 atmospheric CH₄ and N₂O and the underlying microbial and biogeochemical control
1181 mechanisms.

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1184 Figure 2. Repeat oceanic observations include both (a, b) fixed location time-series monitoring
1185 observations and (c,d) hydrographic surveys. Together, such field observation programs helps
1186 resolve temporal variability ranging from months to years and spatial variability at the ocean
1187 basin scale (see Fig. 3). The Station ALOHA data derive from Wilson et al. (2018), the Station
1188 18 data derive from Fariás et al. (2015), and the P16 transect was conducted in 2015 by the
1189 NOAA PMEL Tracer Group as part of the GO-SHIP program. The data shown in the plots are
1190 N_2O concentrations, either as ΔN_2O (i.e. deviation from equilibrium value) or absolute values.

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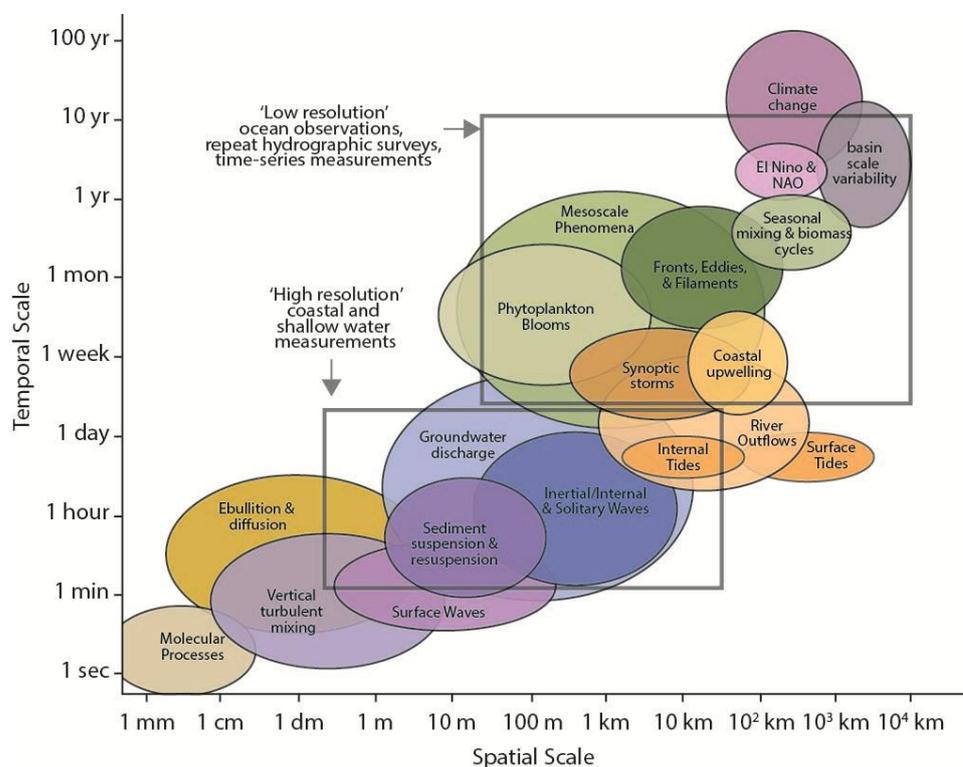
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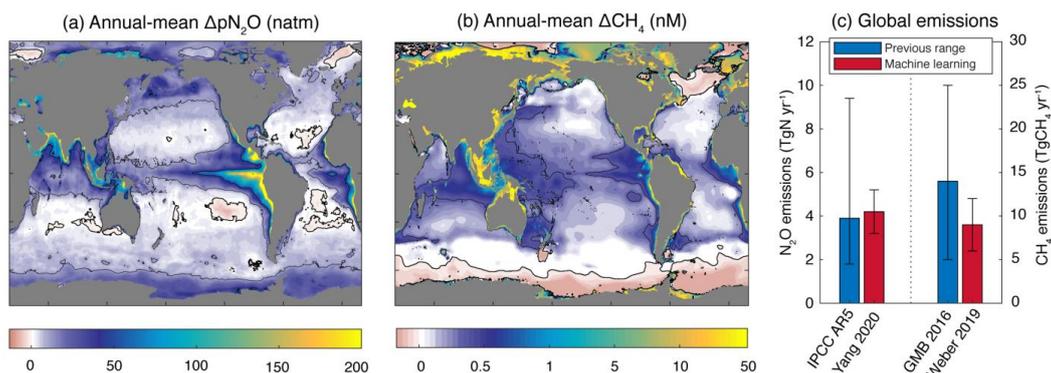


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Figure 3. Time-space scale diagram illustrating various physical, biological, and climatological processes relevant to marine CH₄ and N₂O (adapted from Dickey, 2003). To date, the majority of marine CH₄ and N₂O measurements resolve variability at the mesoscale level or higher. Recent technological developments and the need to resolve concentrations and fluxes in shallow water environments will increase the number of measurements conducted at the sub mesoscale.



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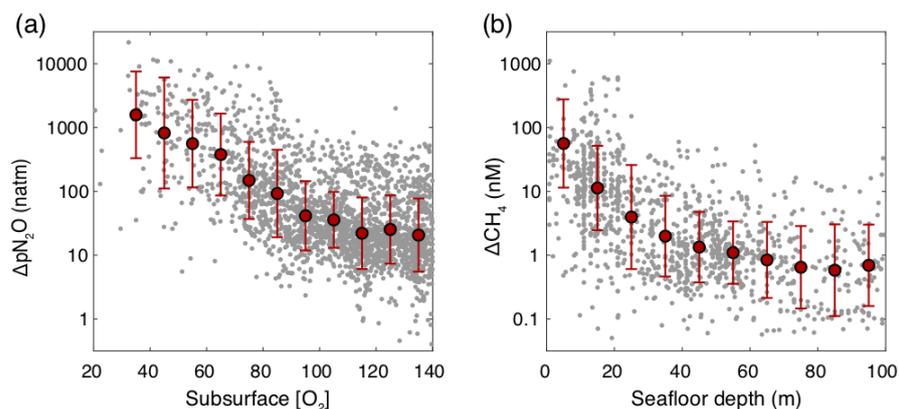
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1209 Figure 4. Distributions and emissions of marine CH_4 and N_2O , (a) Air-sea N_2O disequilibrium
1210 mapped using a Regression Forest model (adapted from Yang et al., 2020), (b) Air-sea CH_4
1211 disequilibrium mapped using an Artificial Neural Network model (adapted from Weber et al.,
1212 2019). For consistency with the original publications, the air-sea disequilibrium is shown in
1213 different units for N_2O (partial pressure) and CH_4 (concentration). (c) A summary of global
1214 ocean CH_4 and N_2O emissions estimated by Yang et al. (2020) and Weber et al. (2019),
1215 compared to the estimates of the IPCC 5th Annual Report (IPCC AR5) and the Global Methane
1216 Budget (Saunois et al., 2016).

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1220 Figure 5. Key environmental predictors of surface ocean CH₄ and N₂O gradients. (a) Excess air-
1221 sea N₂O is best predicted by O₂ concentrations in the subsurface water-column (base of the
1222 mixed layer to a depth of 100 m) (adapted from Yang et al., 2020). (b) Excess CH₄ is best
1223 predicted by seafloor depth, reflecting the supply from anoxic sediments (adapted from Weber et
1224 al., 2019). The grey dots represent individual data points and the red dots with error bars
1225 represent mean ± 1 s.d. of binned data, using O₂ bins of 10 μM width and seafloor depth bins of
1226 10 m width.

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