

Dear editor and reviewers:

We would like to thank you for your comments on our manuscript which has remarkably improved our revised manuscript. We have thoroughly revised the manuscript with responses to the comments point by point as shown in the response letter.

The main changes made in the revised version are as follows:

- (1) We have added plots of the net oxygen evolution and dark respiration rates under culture conditions and plotted $rETR_{max}$, net oxygen evolution and dark respiration rates versus growth rate and made some corresponding changes in the methods, results and discussion sections as suggested.
- (2) Adding more details about culture pH and the measurement of growth rate as suggested.
- (3) Correcting false statements of RbcL and the plot of relative protein concentrations and adding some discussion on the increased RbcL concentration at elevated pCO_2 level in winter as suggested.

All changes are marked in red.

Correspondence about this paper should be directed to Futian Li and Juntian Xu at the following address, phone and fax number and email address.

Sincerely yours,

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Response to the reviews

Reviewer 1

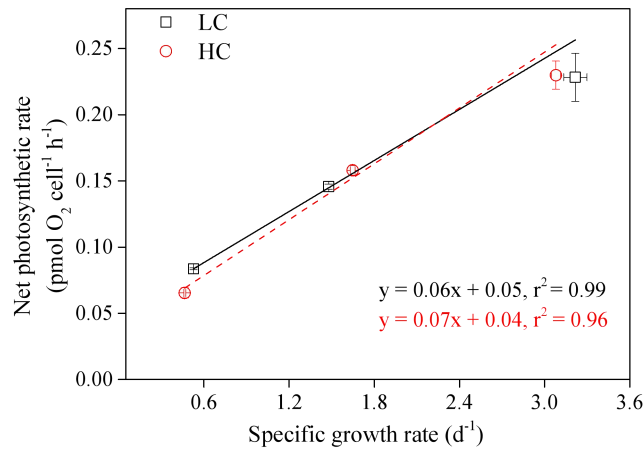
Comments:

I am very happy to see a study of the interaction of increased pCO₂ with temperature and photoperiod
5 on a model diatom. These multifactorial experiments are challenging but necessary. To limit the
(infinite) range of possible combinations the authors matched photoperiod to expected seasonal
temperature.

I think the data, as presented, is valuable, but under-analyzed. I offer some suggestions for some cross
plots of physiological performance at growth condition with achieved growth rate. What is the growth
10 return on O₂ evolution across the conditions?

Response: We thank the reviewer for the useful comments and suggestions that have greatly improved
this manuscript.

The following figure is the relationship between net photosynthetic rates (pmol O₂ cell⁻¹ h⁻¹) and
specific growth rates (d⁻¹) and we will add the figure and the relationships between other physiological
15 parameters and growth in the revised MS.



Abstract: Good

Materials and Methods:

2.3 The authors need to describe whether the growth rates taken every 2 days were averaged within
 20 each growth condition, or whether the maximum measured growth rate for a condition is taken. The
 presented growth rates are fast for a diatom.

Response: We are sorry for the ambiguous description. More details were added in the manuscript.
 Cultures were diluted every 3 days and cell concentration was counted two and three days after the
 dilution, and growth rates were calculated based on the concentrations. The data showed in Fig. 1 were
 25 averaged growth rates calculated for three times at different dilution days.

Diatoms have diverse cell sizes and thus growth rates of different species vary a lot. For *Skeletonema
 costatum*, the average growth rate is around 1.0-2.0 d⁻¹ under normal (20°C, 100 μmol photons m⁻² s⁻¹,
 12L: 12D) conditions (Balzano et al., 2011; Sakshaug and Andresen, 1986), which accords with the
 growth rates (1.6 d⁻¹) under autumn condition (15°C, 150μmol photons m⁻² s⁻¹, 12L: 12D) in this study.

30 In addition, the average growth rate could increase to 2.4 d⁻¹ under 12L: 12D light and dark cycle, when

temperature rise to 25 °C (Zhang et al., 2020). In the present study, for the summer condition, daylength is 16 h, and temperature is 25 °C, which could lead to higher growth rate.

' 2.7 Measurements of PSII Proteins' or '2.7 PSII Protein Measurements'

(current header is technically correct but is archaic usage).

35 **Response:** Corrected.

Figures/Results; Figure 1: Amazing growth rate under summer conditions; the fastest I have ever seen I think for a diatom.

Response: Please see our former responses (line 27-35).

Figure 2: Lovely data, congratulations. Very surprising switch of the OA effect in winter. Suggested
40 additional plot: V at growth $p\text{CO}_2$? After all, most of the curve as plotted is above even the OA range of $p\text{CO}_2$. So a hypothetical V_{max} may not be as important as the achieved V at growth conditions. Then, I would suggested plotting growth rate vs. V at growth condition ($p\text{CO}_2$ and light).

Response: We appreciate the reviewer for the useful suggestion. DIC concentrations in the culture media are about 2 and 2.1 mmol/L in ambient and OA environment respectively, and in the P-C curve,
45 DIC concentration range from 0 to 4 mmol/L. Photosynthetic rate increased significantly with increasing DIC concentration when DIC is lower than 2 mmol/L, and the rate is relative constant when DIC is higher than 2 mmol/L. In the present study V_{max} obtained from the P-C curve showed similar pattern and value with V at growth condition.

Figure 4: Without standard curves, be very cautious in interpretation of the immunoblotting data. The
50 example blot result shows near-saturation (non-linear response of signal to target abundance) for many
of the bands. So the Y axis dynamic range of the greyscale plots may be considerably compressed
relative to the actual change in protein target abundance. Once a band is black, it cannot get any blacker.
It is also very surprising that in winter OA increased RbcL signal.

Response: In this experiment we just calculated the relative value of each protein. The data provide us
55 with a general trend, not the accurate concentration as the reviewer mentioned, among different
treatments. And we will add a caveat in the revised MS. Actin (internal control) could correct the
experimental error in the process of quantitative sample loading of protein, to ensure the accuracy of the
experimental results. The greyscale plots were measured according to density and the area of bands
(TanonImage software). Although they are all black, the densities and areas are different in most
60 conditions.

For the increased RbcL signal under OA condition in winter, although some researchers found RubisCO
contents decreased at OA (Losh et al., 2013; Endo et al., 2015), RbcL expression of different diatoms
(*Thalassiosira pseudonana*, *Phaeodactylum tricornutum*, and *T. weissflogii*) were found slightly
increased under OA in nitrogen-replete condition (Hong et al., 2017). And phytoplankton communities
65 showed enhanced Rubisco expression in 800 ppm treatment compared with 350 ppm (Tortell et al.,
2000). A coastal isolated *T. pseudonana* and *Emiliana huxleyi* also showed higher RbcL contents in
higher CO₂ level, although the offshore isolated *T. pseudonana* showed no significant difference
between ambient and high CO₂ conditions (McCarthy et al., 2012). And low temperature could increase
the relative amount of RbcL (Devos et al., 1998). We will add further discussion in the revised MS.

70 **References:**

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- Devos N, Ingouff M, Loppes R, et al. Rubisco adaptation to low temperatures: a comparative study in psychrophilic and mesophilic unicellular algae. *Journal of Phycology*, 1998, 34: 655-660.
- 75 Endo H, Sugie K, Yoshimura T, et al. Effects of CO₂ and iron availability on *rbcL* gene expression in Bering Sea diatoms. *Biogeosciences*, 2015, 12: 2247-2259.
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- Sakshaug E, Andresen K. Effect of light regime upon growth rate and chemical composition of a clone
85 of *Skeletonema costatum* from the Trondheimsfjord, Norway. *Journal of Plankton Research*, 1986, 8: 619-637.
- Tortell P D, Rau G H, Morel F M M. Inorganic carbon acquisition in coastal Pacific phytoplankton communities. *Limnology and Oceanography*, 2000, 45: 1485-1500.

Zhang L, Li H, Wu M, Li F, and Xu J.: Effects of seawater acidification on photosynthetic
90 physiological characteristics of *Skeletonema costatum* at different temperatures, Journal of Jiangsu
Ocean University, 2020, 29: 1-7.

Reviewer 2

Comments:

The manuscript titled “Physiological responses of *Skeletonema costatum* to the interactions of seawater
95 acidification and combination of photoperiod and temperature” described a research attempting to
explore the impact of high pCO₂ (or ocean acidification, OA) under different seasons (combination of
photoperiod and temperature) on the diatom *S. costatum*. The experiments are well conceived, and
methods are clearly presented. The most interesting observation is that high pCO₂ (OA) does not
uniformly impact *S. costatum* under different seasons: with somewhat negative impact on winter
100 conditions. The authors showed the interesting observation, but the experimental design and data quality
can be improved.

Here are some questions and suggestions for the authors.

Response: We thank the reviewer for the recognition of the value of our work and the valuable
comments.

105 On the experimental setup, the authors stated that the cell culture pH did not change over 0.05 units in
the 3d of one generation (section 2.3), so one very basic question is: what are the pH values and ranges
for the six different conditions (i.e. three seasons and two pCO₂)? Since the manuscript is about how

OA impacts *S. costatum* differently during different seasons, the acidification information, which can be presented easily as pH, is very critical to this whole article, however, this information is missing.

110 **Response:** The pH of culture media under different treatments are presented below. And the information will be added in the revised MS.

Treatments	W-LC	W-HC	A-LC	A-HC	S-LC	S-HC
pH	8.10 ± 0.01	7.85 ± 0.01	8.14 ± 0.01	7.89 ± 0.01	8.19 ± 0.02	7.89 ± 0.02

Following the cell culture pH question, the authors measured the photosynthesis (P) vs DIC curve at pH 8.12 (section 2.5) and very likely they did the same with P-I curve. It would be better if the authors measure the responses under lower pH for high pCO₂ treated cells, according to the high pCO₂ (OA) conditions. It should be expected that the pH is lower under OA conditions, and *S. costatum* acclimated to OA conditions may not photosynthesize better under the experimental condition with higher pH (8.12). As a result, the presented P-I and P-DIC curves for *S. costatum* cultured in HC (OA conditions) may not reflect their real physiological status in terms of photosynthesis under OA conditions. Also please note that Tris buffer is known to change pH significantly with different temperature, so it is important to measure or calculate the pH at certain temperature.

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Response: Researchers determine the P-C curves of OA treated cells under either ambient pH or culture pH (HC conditions). As the reviewer mentioned, P-C curves determined under culture pH could reflect the real physiological status under OA conditions. However, the percentages of DIC species (CO₂, HCO₃⁻, CO₃²⁻) differ in media with different pH, which is inconvenient to compare K_m of LC and HC cells. Thus, some studies prefer setting same pH for LC and HC conditions to compare V_{max} and K_m at same pH (Nakajima et al., 2013; Shi et al., 2017). For P-I curves, the measurement time was much

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shorter than that used for P-C curves, so the pH wouldn't change markedly and Tris buffer was not used for P-I curves in the present study. Cells were resuspended in pre-aerated fresh medium under cultured condition (i.e. pH 8.14 for LC cells, 7.9 for HC cells) to measure P-I curves. Details for P-I measurements were added in the revised MS. We are sorry we missed the effects of Tris buffer on pH at different temperature. For the P-C curves measurements in the present study, Tris was added in the culture medium, and the pH of the medium used for resuspending cells was adjusted at room temperature.

In the results session, the authors only mentioned and cited Fig. 1, while Figure 2, 3, 4 are listed at the end of the manuscript, none of those was referred in the text. In the tables presented, Table 2 and Table 3 do not show any units.

Response: We are sorry for the mistakes, Figure 2, 3, 4 are referred in the text now and units are added in Table 2 and 3 now.

In section 3.5 where the authors present the "PSII protein concentrations", RbcL was included as key PSII proteins. Such claim should be red-flagged, likewise the statement of "RbcL is related to the function of QA". Inclusion of RbcL in PSII proteins is also found in the Abstract and Discussion. The authors should make sure what RbcL really does with creditable citations before writing assumptions or conclusions about RbcL.

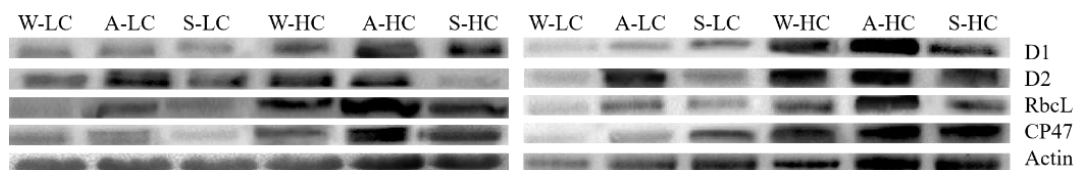
Response: We apologize for the vague statement regarding the RbcL. It's revised in section 3.5, Abstract and Discussion.

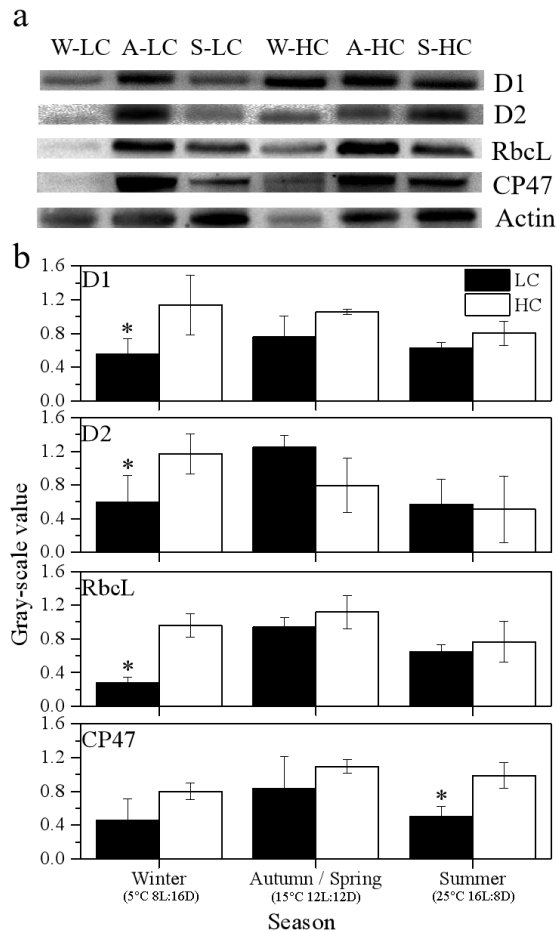
The description of methodology “Values of Actin were divided by other densitometric scanning values of protein to calculate Gray-scale values” should be modified to indicate the supposing meaning of normalizing density to Actin.

Response: Corrected. It has been revised as “Actin was used as internal control in order to correct the experimental error in the process of quantitative sample loading of protein, to ensure the accuracy of the experimental results.”

With the data presented in Figure 4, panel (a) and panel (b) do not seem to agree with each other. The western blot data does not look like a representative of the statistical data. For example, in W-HC (winter high pCO₂) condition, the D1 density is much higher than Actin (Fig 4a), so such value is greater than 2 if analyzed using ImageJ, however, the data presented in statistics showed a value very tightly close to 0.9 (Fig 4b). It would be nice if all raw data (immunoblots) are presented to support the statistics in Fig 4b.

Response: We thank the reviewer for pointing this out. We checked the data and revised the mistake. In Fig. 4 panel (a), the D1 value is 1.55 according to TanonImage software. The immunoblots data and revised Fig. 4 are presented as following:





Other minor concerns:

Section 2.7, “1000 and 34.5 are constants”, what are the units? Or at least provide the unit of “where C
165 represents total chlorophyll concentration”.

Response: The unit of chlorophyll concentration is $\mu\text{g} / \text{ml}$. It is revised in the manuscript.

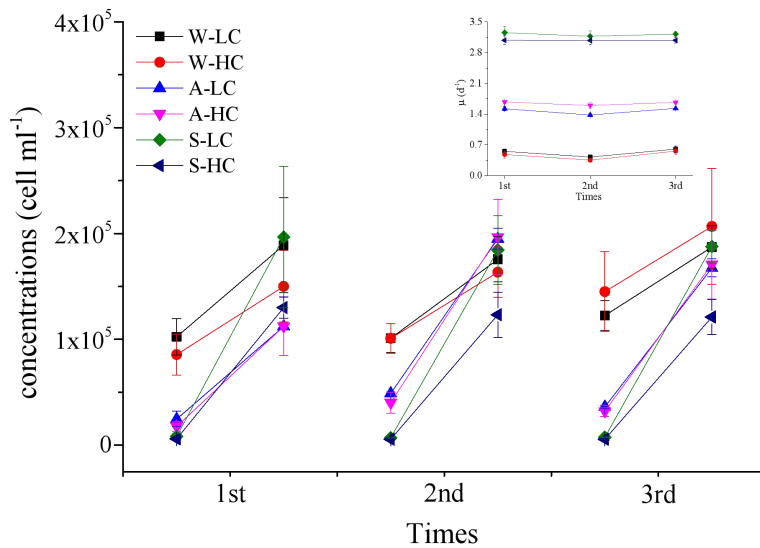
The use of letter “C” is ambiguous in the text. C was also used as carbon in line 219: “C fixation”.

Response: “C fixation” is revised as “carbon fixation”.

For the measurement of specific growth rate, more details on how data were collected would be helpful.

170 It would be better to have the raw data, cell concentration vs time (days), presented.

Response: More details will be added in the revised MS. Cells concentration was diluted every 3 days and cell concentration was counted two and three days after the dilution, such growth rate data in Fig. 1 were calculated for three times at different dilution days. The following figure presents cell concentrations and specific growth rate at three different times.



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Line 229, “initial slop” should be “initial slope”.

Response: Corrected.

References

Shi Q, Xiahou W, Wu H. Photosynthetic responses of the marine diatom *Thalassiosira pseudonana* to

180 CO₂-induced seawater acidification. *Hydrobiologia*, 2017, 788: 361-369.

Nakajima K, Tanaka A, Matsuda Y. SLC4 family transporters in a marine diatom directly pump bicarbonate from seawater. *Proceedings of the National Academy of Sciences*, 2013, 110: 1767-1772.

Physiological responses of *Skeletonema costatum* to the interactions of seawater acidification and combination of photoperiod and temperature

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Abstract. Ocean acidification (OA), which is a major environmental change caused by increasing atmospheric CO₂, has considerable influences on marine phytoplankton. But few studies have investigated interactions of OA and seasonal changes in temperature and photoperiod on marine diatoms. In the present study, a marine diatom *Skeletonema costatum* was cultured under two different CO₂ levels (LC, 400 µatm; HC, 1000 µatm) and three different combinations of temperature and photoperiod length (8:16 L:D with 5 °C, 12:12 L:D with 15 °C, 16:8 L:D with 25 °C), simulating different seasons in typical temperate oceans, to investigate the combined effects of these factors. The results showed that specific growth rate of *S. costatum* increased with increasing temperature and daylength. However, OA showed contrasting effects on growth and photosynthesis under different combinations of temperature and daylength: while positive effects of OA were observed under spring and autumn conditions, it significantly decreased growth (11 %) and photosynthesis (21 %) in winter. In addition, OA alleviated the negative effect of low temperature and short daylength on RbcL and key PSII proteins (D1 and D2). These data indicated that future ocean acidification may show differential effects on diatoms in different cluster of other factors.

Key words. diatom, growth, photosynthesis, CO₂, temperature and photoperiod

1 Introduction

25 Ocean acidification (OA) is one of major environmental changes caused by increasing atmospheric CO₂, which has directly raised from 280 ppm in preindustrial era to higher than 400 ppm at present (Friedlingstein et al., 2019). It is predicted that surface seawater pH would drop 0.3–0.5 and 0.5–0.7 units by the year 2100 and 2300 respectively (Caldeira and Wickett, 2003). It has been suggested that calcifying organisms, such as coral reefs and coccolithophores, are vulnerable to OA due to

the decreased calcification at elevated CO₂ (Albright et al., 2016). The responses of non-calcifying organisms such as diatoms to OA vary widely among taxonomic groups which may be detrimental, negligible or even beneficial (Gao and Campbell, 2014). Consequently, the abundance of marine phytoplankton and community structure might be altered by OA (Gattuso et al., 2015).

Diatoms are ubiquitous photosynthetic phytoplankton which account for about 20 % of global primary productivity, and thus play a crucial role in the global cycling of carbon and silicon (Falkowski et al., 2004). To overcome the limited aqueous CO₂ concentration in seawater, they have developed CO₂-concentrating mechanisms (CCMs) (Spalding, 2007). Decreased photosynthetic affinity for dissolved inorganic carbon (DIC) and activity of CCM related enzymes are generally found under increased CO₂ condition (Raven and Beardall, 2014). For phytoplankton assemblages, elevated CO₂ could lead to increases in chlorophyll *a* concentrations and the abundance of diatoms (Johnson et al., 2013). Species and strain specificity are observed in studies on physiological responses of diatoms to OA, which might be caused by the balance between positive effects of elevated CO₂ and negative effects of decreased pH (Langer et al., 2009; Li et al., 2016). In addition, acclimation and adaptation processes, i.e. the timescale of diatoms exposed to OA, could also influence the physiological effects of OA (Wu et al., 2014; Li et al., 2017). Moreover, other environmental factors, such as temperature (Seebah et al., 2014), light (Gao et al., 2012), nutrients (Li et al., 2015) and clusters of multiple factors (Xu et al., 2014a; Xu et al., 2014b), are shown to have interaction with OA on diatoms.

Diatoms are widespread across oceans, thus they would experience different photoperiods. Photoperiod controls the total light dose received by phytoplankton and thus could remarkably influence the physiological performance such as growth and lipid content of microalgae (Wahidin et al., 2013). For Antarctic sea ice microalgae *Chlamydomonas* sp., continuous illumination stimulates higher growth and nutrient absorption rates than successive darkness condition (Xu et al., 2014c). Growth rate of *Chlamydomonas reinhardtii* is gradually enhanced following the increasing photoperiod (Hsieh et al., 2018). In contrast, *Alexandrium minutum* grows faster under short daylength relative to longer and even continuous daylength (Wang et al., 2019). Moreover, different photoperiods could influence intracellular carbon demand of microalgae, which has a stronger regulation effect on CCMs compared with effects of changes in CO₂ supply (Rost et al., 2006).

Under the combined influence of photoperiod and OA, physiological performance of phytoplankton might be different from that under single factor. For example, continuous light moderates the negative effect of OA on coccolithophore growth, although species isolated from different regions show diverse responses (Bretherton et al., 2019). The changes of photoperiod are often accompanied by increase or decrease temperature, and impacts of OA on diatoms can also be changed by temperature. For example, under OA condition, decreased metabolic activity is observed in *Phaeodactylum tricorutum* when temperature elevates (Bautista et al., 2018), while elevated temperature enhances the growth rate of *Nitzschia lecointei* (Torstensson et al., 2013).

But limited studies have investigated interactions between OA and combination of temperature and photoperiod (i.e. conditions in different seasons) on diatoms. *Skeletonema costatum* is a widespread, eurythermal and euryhaline diatom species, which frequently causes red tide. We hypothesized the effect of OA on *S. costatum* may be modulated by

photoperiod and temperature. In the present study, we investigate the physiological performance of marine diatom *Skeletonema costatum* under two different CO₂ levels and three combinations of temperature and photoperiod, which simulated different seasons in typical temperate oceans (winter, 5 °C with 8:16 L:D; spring or autumn, 15 °C with 12:12 L:D; summer, 25 °C with 16:8 L:D).

2 Materials and methods

2.1 Culture conditions

The diatom *Skeletonema costatum* in this study was isolated from Gaogong Island, Lianyungang, Jiangsu province (34°70' 74.95"N, 119°49'26.47"E). Before being used in experiments, the cells were cultured in autoclaved natural seawater enriched with f/2 medium (Guillard and Ryther, 1962). Semi-continuous cultures were maintained in 500 ml Erlenmeyer flasks with a filter unit (Millex GP, Merck, USA) in order to aerate sterile air. Triplicate independent cultures were set for each treatment at the light intensity of 150 μmol photons m⁻² s⁻¹.

2.2 Experimental setup

In order to evaluate effects of pCO₂ levels and different combination of temperature and photoperiod on *S. costatum*, cells were cultured under winter (5 °C with light: dark cycle of 8:16 h), spring / autumn (15 °C with 12:12 h), summer (25 °C with 16:8 h) conditions independently with two pCO₂ levels (400 ppm, LC; 1000 ppm, HC), simulating temperature and daylength conditions of different seasons in typical temperate oceans. Temperatures and light intensity (150 μmol photons m⁻² s⁻¹) were controlled by illumination incubators (GXZ-500B, Ningbo, China). Cells were inoculated in cultures with fresh medium which was aerated with ambient air (400 ppm) or CO₂-enriched air (1000 ppm). The high pCO₂ level was manipulated by a CO₂ plant incubator (HP 1000 G-D, Ruihua Instruments, Wuhan, China). Cultures were kept at exponential phase by diluting every 3 d, and cells concentrations were controlled below 2×10⁵ cell ml⁻¹ in order to minimize the effect of cell metabolism on carbonate chemistry in medium. The changes in culture pH was less than 0.05 during the 3 d (8.10 ± 0.01 for LC and 7.85 ± 0.01 for HC in winter; 8.14 ± 0.01 for LC and 7.85 ± 0.01 for HC in spring / autumn; 8.19 ± 0.02 for LC and 7.89 ± 0.02 for HC in summer). After acclimating to different treatments for at least 40 generations, following parameters were measured.

2.3 Growth measurement

To estimate the growth of *S. costatum*, triplicate samples (1 ml each) were collected from each treatment at 48 and 72 h after dilution and fixed with 10 μl Lugol's solution, then a plankton counting chamber (DSJ-01, Xundeng Instruments, Xiamen, China) was used to count cells directly under an optical microscope (DM500, Leica, Germany). The specific growth rate was

calculated as: $\mu = (\ln N_t - \ln N_0) / (t - t_0)$, where N_t represents the cell concentration (cells ml⁻¹) at time t ; N_0 represents the cell concentration at time t_0 , $t - t_0 = 1$ d. The growth rates were averaged from 3 dilution processes within each growth condition.

2.4 Chlorophyll *a* and BSi measurements

95 Samples were filtered onto GF / F filters (25 mm, Whatman, UK), and chlorophyll *a* were extracted with 4 ml of methanol at 4 °C for 24 h in darkness. An ultraviolet spectrophotometer (Ultrospect 3300 pro, Amersham Bioscience, Sweden) was used to detect the absorption values of supernatant under 632 nm, 665 nm and 750 nm after centrifuging (Biofuge primo R, Thermo, Germany). The chlorophyll *a* concentration (pg cell⁻¹) of *S. costatum* was calculated by the equation of Ritchie (2006).

100 Samples (200 ml) for biogenic silica (BSi) measurement (pmol cell⁻¹) were filtered onto polycarbonate filters (0.8 μm, Merck Millipore, Germany) by polysulfone filter funnel (25 mm, Pall Corporation, UK), and filters were then dried at 80°C for 24 h. BSi on the filter was digested by 4 ml of 0.2 M NaOH in boiling bath for 40 min, and were neutralized with 1 ml of 1M HCl when cooled. The supernatant (1 ml) was diluted with 4 ml of milli-Q water, and then 2 ml of molybdate soln and 3 ml of reducing agent were added into tubes. The absorption was measured at 810 nm by an ultraviolet spectrophotometer (Ultrospect 3300 pro, Amersham Bioscience, Sweden) after the color developing for 2-3 h (Brzezinski and Nelson, 1995).

105 2.5 Photosynthesis and respiration measurements

The net photosynthetic rate under culture condition, photosynthetic oxygen evolution rate versus light intensity (P-I) curve and photosynthesis versus DIC concentration (P-C) curve of *S. costatum* were measured through a Clark-type oxygen electrode (Oxygraph+, Hansatech, UK), in which temperature was controlled by a thermostatic water bath (DHX-2005, China).

110 For measurement of net photosynthesis under culture condition, light intensity was set as 150 μmol photons m⁻² s⁻¹ (light intensity during culture), provided by a halogen lamp (QVF135, Philips, Netherlands). Sample of 50 ml was filtered (< 0.02 MPa) onto a cellulose acetate membrane (Xinya Instruments, Shanghai, China). Due to the shorter measurement time than that of P-C curve, cells were filtered and resuspended in 5 ml pre-aerated fresh medium (without buffer) under cultured condition, which was then used to determine oxygen evolution rate and cell concentration. Oxygen consumption was measured under darkness which was realized by covering an opaque box on the reaction chamber.

115 2.6 P-C curve measurement

For P-C curve, photosynthetic oxygen evolution was measured under same light intensity as culture. Sample of 50 ml was filtered and resuspended in 5 ml fresh C_i-free f / 2 tris buffered medium (pH 8.12), which was then used to determine oxygen

120 evolution rate and cell concentration. Same pH level (pH 8.12) was set for both LC and HC treatments in order to compare K_m and V_{max} at same percentages of DIC species. The remaining intracellular C_i was depleted by exposing cells to culture light intensity for 20-30 min. Net photosynthetic rate was determined after adding calculated amounts of $NaHCO_3$ stock solution (0.025 M and 0.2 M). The final concentrations of DIC were 0, 0.025, 0.05, 0.1, 0.2, 0.4, 1, 2 and 4 mM. Net photosynthetic rates under different DIC concentration were fitted by Michaelis-Menten equation.

2.7 P-I curve measurement

125 For P-I curve, oxygen consumption rates in darkness and net photosynthetic oxygen evolution at 7 different light intensities (0, 10, 20, 50, 100, 200, 500, 1000 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) were identified. Light intensity was achieved by adjusting the distance between the halogen lamp and oxygen electrode chamber. Cells were also filtered and resuspended in pre-aerated fresh medium in a similar way to photosynthesis measurement under culture condition. Photosynthetic rate and light intensity data were fitted according to Henley (1993): $P = P_m \times \tanh(\alpha \times \text{PAR} / P_m) + R_d$, where PAR is irradiance, P is photosynthetic rate, P_m is light-saturated photosynthetic rate, α is initial slope of P-I curve, R_d is dark respiration rate. I_k (saturating irradiance for photosynthesis) and I_c (light compensation point) were also calculated by: $I_k = P_m / \alpha$, $I_c = R_d / \alpha$. Cells were measured under 8 different PAR levels (0, 10, 20, 50, 100, 200, 500, 1000 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) lasting for 10 s each using a hand-held fluorometer (AquaPen-C AP-P 100, Chech) for rapid light curves (RLCs) measurement. Relative electron transport rates (rETR) of *S. costatum* were measured, and were estimated as Wu, et al. (2010):
130 $rETR = \text{PAR} \times Y(\text{II}) \times 0.5$, where PAR represents the photon flux density of actinic light; $Y(\text{II})$ represents the effective quantum yield of PSII, 0.5 is based on the assumption that PSII receives half of all absorbed quanta. RLCs were fitted as: $P = \text{PAR} / (a \times \text{PAR}^2 + b \times \text{PAR} + c)$, where P represents rETR; a, b and c are model parameters. The relative photoinhibition ratio of rETR was calculated as: $\text{Inh} (\%) = (rETR_{\text{max}} - rETR_x) / rETR_{\text{max}} \times 100 (\%)$, where $rETR_x$ is the value of rETR at 1000 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$.

140 2.8 Protein measurements

Ribulose-1,5-bisphosphate carboxylase/oxygenase (RubisCO) large subunit binding protein (RbcL) and PSII proteins (PsbA (D1), PsbD (D2), and PsbB (CP47)) were measured under different $p\text{CO}_2$ levels and combinations of temperature and daylength. D1 and D2 proteins are located in reaction center, while CP47 is a junction of antenna, and RbcL is a component of RubisCO which is a key enzyme in the CO_2 fixation process. For the relative value of protein measurements, cells were filtered and resuspended in 2 ml of extracting medium (50 mM Tris-HCl, pH 7.6, 5.0 mM MgCl_2 , 10 mM NaCl, 0.4 M sucrose, and 0.1 % BSA) according to Ma et al. (2019). After cell disruption and centrifugation, supernatant liquid was used to measured chlorophyll concentration ($\mu\text{g ml}^{-1}$) according to Arnon (1949): $C = D_{652} \times 1000 \div 34.5 \times T$, where C represents total chlorophyll concentration, D_{652} represents absorption value in 652 nm, T represents dilution ratio, 1000 and 34.5 are

constants. Same concentration of chlorophyll (2.4 micrograms) per lane was used for 12 % sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE, Mini PROTEAN, Bio-rad, America) at 150 V for 1 h, and the proteins were transferred into polyvinylidene difluoride (PVDF) membranes which were then immersed in blocking solution with antibodies (D1, D2, CP47, RbcL and Actin; Agrisera) for 1 h, and successively, goat anti-rabbit secondary antibodies were used. Blots were developed by using enhance chemluminescence luminescence (ECL) reagent and were quantified with a chemiluminescence detection system (Tanon 5500, Shanghai, China). Actin was used as internal control in order to correct the experimental error in the process of quantitative sample loading of protein, to ensure the accuracy of the experimental results. And the data provide us with a general trend, not accurate concentrations, among different treatments.

2.9 Data analyses

Data were analyzed with IBM SPSS Statistics 24 and are presented as mean \pm SD (standard deviation). One-way ANOVA was used to compare differences among combination of temperature and photoperiod treatments. The independent-samples *t*-test was applied to compare differences between two pCO₂ levels. General linear model was conducted to assess the interactive effects of CO₂ level and combination of temperature and photoperiod on growth rate, rETR, photosynthesis and respiration, contents of chlorophyll *a* and BSi and proteins. When *P* values were under 0.05, tukey test was used for *post hoc* analysis.

3 Results

3.1 Specific growth rate

Growth rate of *S. costatum* ranged from 0.47 ± 0.01 to 3.22 ± 0.08 d⁻¹ under different treatments and increased significantly with increasing temperature and daylength (*P* < 0.05) regardless the pCO₂ level (Fig. 1). In summer season, elevated pCO₂ showed no significant effects, however, it remarkably influenced the growth rate in other seasons. Elevated pCO₂ enhanced the growth rate by 11 % in spring and autumn (*P* < 0.001), while the reverse pattern was found in winter (*P* < 0.001). General linear model indicated that the season and CO₂ level had a notable interaction on specific growth rate (*P* < 0.001, Table 1).

3.2 Chlorophyll *a* and BSi contents

Under ambient CO₂ condition, chlorophyll *a* content was enhanced by increased temperature and daylength (Table 2), and the content was 22 % higher in summer compared with winter (*P* = 0.008). When CO₂ was elevated, chlorophyll *a* content in winter was 42 % and 32 % lower than that in spring and summer respectively (*P* = 0.001, *P* = 0.004). Elevated pCO₂ decreased chlorophyll *a* content in winter (*P* = 0.022) while enhanced it in spring (*P* = 0.002) and had no significant impact in summer. A significant interaction between season and CO₂ can be found (*P* < 0.001) (Table 1).

A different trend was detected for BSi content (Table 2). Under ambient pCO₂, BSi content decreased with higher temperature

180 along with longer daylength and the value in winter was significantly higher than that in spring and summer ($P=0.005$, 0.002 respectively). Higher pCO₂ decreased BSi significantly in winter ($P = 0.016$) and spring ($P = 0.007$) while had no significant influence on the content in summer ($P = 0.3$). There is a significant interaction between season and CO₂ on BSi content ($P < 0.05$) (Table 1).

3.3 Photosynthesis and respiration

185 Net photosynthetic oxygen evolution and dark respiration rates showed similar patterns under same CO₂ condition (Fig. 2). The lowest photosynthesis and respiration rates were observed under winter condition, and maximal rates were observed in summer at each pCO₂ level. Both photosynthesis and respiration rates increased with increasing temperature and daylength ($P < 0.05$). Elevated pCO₂ inhibited net photosynthetic rate under winter condition ($P = 0.0013$) while photosynthesis was enhanced by elevated pCO₂ in spring and autumn ($P = 0.006$). In addition, higher pCO₂ stimulated dark respiration rate in spring and autumn ($P < 0.001$). Both photosynthesis and respiration were not impacted by higher pCO₂ in summer. Interaction between season and CO₂ on net photosynthetic rate was detected ($P = 0.035$). Positive relationships of dark respiration or net photosynthetic rates and growth rate were observed (Fig. 5a, b).

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3.4 P-C curve

Combination of temperature and photoperiod in different seasons had a significant effect on V_{max} ($P < 0.001$, Table 1, Fig. 3b), cells under spring / autumn season condition had the highest V_{max} at both pCO₂ levels, but the lowest V_{max} was found in winter at elevated pCO₂ and in summer at ambient pCO₂. Elevated pCO₂ significantly enhanced the values in higher temperature and longer daylength treatments (129 % for spring / autumn, 130 % for summer when compared with ambient CO₂ level). With respect to K_m, different season had no significant effect on it at ambient pCO₂, while K_m in summer was lowest among the seasons at elevated pCO₂ (70 % and 69 % lower than winter and spring / autumn respectively). Elevated pCO₂ had significant influences on K_m under same season besides summer and the values increased by 143 % and 113 % in winter and spring / autumn respectively when compared with low pCO₂ ($P = 0.021$, $P = 0.003$ in winter and spring / autumn). Interaction between CO₂ and combination of temperature and photoperiod on parameters of P-C curves were detected (Table 1).

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3.5 P-I curve

205 Net photosynthetic oxygen evolution rates increased with increasing light intensity initially and then reached the plateaus in all seasons and the curves under winter condition reached the plateaus much earlier than the other two seasons (Fig. 4).

Higher temperature and prolonged daylength had a main effect on P_{\max} , R_d , I_k and I_c . However, elevated $p\text{CO}_2$ instead of season had main effect on α (Table 1). P_{\max} was enhanced when temperature increasing with prolonged daylength ($P < 0.05$) except when the summer season was compared with spring and autumn condition at elevated $p\text{CO}_2$ (Table 3). Effects of elevated $p\text{CO}_2$ was only observed in spring and autumn ($P = 0.03$). I_k increased remarkably in summer under both $p\text{CO}_2$ treatments which was similar as R_d at elevated $p\text{CO}_2$ ($P < 0.05$). There was a significant interaction between CO_2 and combination of temperature and photoperiod on R_d (Table 1).

At higher $p\text{CO}_2$ level, $r\text{ETR}_{\max}$ values were significantly different among different seasons ($P < 0.05$), and the highest value was found in spring and autumn (52 % and 14% higher than winter and summer, Table 3). Elevated $p\text{CO}_2$ decreased $r\text{ETR}_{\max}$ in winter and summer ($P < 0.001$ in winter, $P = 0.01$ in summer). $r\text{ETR}_{\max}$ had positive correlation with growth rate when temperature and daylength increased from winter to spring / autumn condition. However, as temperature and daylength continuously increasing from spring / autumn to summer, the correlation became negative (Fig. 5c). Interactions between the two factors were detected on $r\text{ETR}_{\max}$ ($P < 0.001$, Table 1). Photoinhibitions were found in RLCs of cells under all treatments. As the season processed from winter to summer, photoinhibitions were alleviated significantly in both $p\text{CO}_2$ levels ($P < 0.05$). In spring and autumn, the inhibition at higher $p\text{CO}_2$ was significantly decreased compared with ambient $p\text{CO}_2$ condition ($P = 0.039$, Table 3).

3.5 PSII protein concentrations

The relative value of RbcL and key PSII proteins (PsbA (D1), PsbD (D2), and PsbB (CP47)) were quantified in different seasons under ambient and elevated CO_2 conditions (Fig. 6). At ambient $p\text{CO}_2$ level, the highest contents of all four proteins were detected in spring and autumn. Elevated $p\text{CO}_2$ significantly enhanced RbcL, D1 and D2 protein contents in winter ($P < 0.05$). In addition, higher $p\text{CO}_2$ led to increase of CP47 in summer ($P = 0.006$).

4. Discussion

Phytoplankton like diatoms have already evolved several strategies to cope with different temperature and daylength in temperate oceans, where variations of season are evident. However, the ongoing elevated $p\text{CO}_2$ combining with changes in temperature and daylength is a new stress on diatoms and we know little about their interactions. Therefore, we examined the combined effects of $p\text{CO}_2$ and seasonal changes in temperature and photoperiod on the physiological performance of a typical marine diatom *S. costatum*.

4.1 Physiological responses of *S. costatum* to different combinations of temperature and photoperiod

In the present study, the growth rate of *S. costatum* increased with increasing temperature and daylength regardless of the $p\text{CO}_2$ level (Fig. 1). Previous studies showed that most phytoplankton, such as *Chlamydomonas reinhardtii*, *Trichodesmium* or *Alexandrium catenella*, grew faster under prolonged photoperiods (Cai and Gao, 2015) although *Alexandrium minutum*

grew faster under shorter photoperiods (Wang et al., 2019). For *S. costatum*, a remarkably higher contribution of HCO_3^- to the overall carbon uptake was observed under light dark cycles compared with continuous light, and a shorter photoperiod led to lower photosynthetic affinity for inorganic carbon (Rost et al., 2006). Basically, the Chl *a* quota in microalgae increases with decreasing daylength, however, our results exhibited inverse pattern (Table 2). The inconsistency might be caused by the different temperatures set in studies, which is another main environmental factor affecting the growth of diatoms.

Increasing temperature may lead to various changes in growth rate depending on whether the temperature is optimal for the species. For *S. costatum*, its growth rate has been shown to increase with temperature up to 30 °C (Ebrahimi and Salarzadeh, 2016). Zhang et al. (2020) also found that the growth rate of *S. costatum* could increase with temperature from 5 °C to 30 °C and then drop sharply. The underlying mechanism is that elevated temperature promotes *S. costatum* metabolic rates when nutrients are abundant. This could be shown by the relationship between respiration and growth. Higher mitochondrial respiration can result in higher growth rate theoretically (Geider and Osborne, 1989), since this process provides ATP and carbon skeletons (Raven et al., 2017). It seems that temperature shows the dominant effect compared with daylength.

Rubisco is an important enzyme for carbon fixation, psychrophilic diatoms utilize increasing abundance of RbcL protein to maintain high cellular enzymatic rates and growth rate at low temperature (Young et al., 2015). However, for diatoms in temperate area, the amount of RbcL protein decreased in low temperature along with short daylength compared with higher temperature and longer daylength in our results. Low abundance of RbcL was in line with slower growth rate in winter condition.

4.2 Effect of ocean acidification under different seasons

Our results showed that the impacts of elevated pCO_2 on *S. costatum* depended on seasonal changes in temperature and photoperiod. High pCO_2 had enhanced the growth of *S. costatum* in spring / autumn and reduced it in winter, while no significant effects were detected in summer (Fig. 1). CO_2 concentration mechanisms (CCMs) is energy-dependent and high pCO_2 down-regulate CCMs of most phytoplankton including *S. costatum*, so the saved energy could be used for growth (Raven et al., 2017). Higher initial slope of the P-I curve at elevated pCO_2 might be partly responsible for the higher growth rate compared with that at ambient pCO_2 (Table 3). But in winter, growth decreased under OA condition, although photosynthesis and respiration had no significant changes. This is because the combination of biochemical and biophysical CCMs may cause the lack of a positive response to elevated pCO_2 under near-optimal growth conditions (Passow, 2015). In addition, when other environmental factors are stressful, the sensitivity of diatoms to CO_2 and temperature is prominent (Taucher et al., 2015). The shorter daylength and low temperature simulating winter condition in present study can be seen as stressors, under which *S. costatum* was more sensitive to elevated pCO_2 . When temperature and photoperiod are optimal, positive or neutral effects of higher pCO_2 were observed. However, different patterns were reported for other species, such as *E. huxleyi* and the macroalgae *Ulva linza* (Bretherton et al., 2019; Yue et al., 2019). For these two species, reduced growth

rate at elevated pCO₂ were found when the daylength was longest. High temperature might accelerate nutrient uptake and metabolic rates, which may alleviate the negative effects of longer daylength under higher pCO₂ environment (Bretherton et al., 2019; Yue et al., 2019). Maximum photosynthetic rate increased significantly under higher pCO₂ in spring and autumn condition. This was consistent with the higher photosynthetic efficiency and growth rate (Table 3, Fig. 1). Meanwhile, K_m increased under high CO₂ concentration which means the affinity for CO₂ decrease and thus CCMs are down-regulated (Fig. 3).

Silicification directly relates to cell division and growth, and is independent of photoperiod (Brzezinski, 1992). BSi contents generally increased with decreasing growth rate when any limiting factors such as temperature, light or ammonium exist (Martin et al., 2000). In the present study, elevated pCO₂ mitigated the negative effects of temperature and photoperiod limitation on BSi content (Table 2). Higher BSi contents in winter under ambient CO₂ condition can intensify the ballasting effects and thus impact the sinking rate of organic matters produced by diatoms.

Lavaud et al. (2016) indicated that PSII activity and phosphorylation of thylakoid protein may play a crucial role in controlling the change of the photosynthetic activity. Higher pCO₂ induced higher photosynthetic efficiency in PSII, with increased contents of D1 and D2 proteins in winter (Fig. 6). Proteins will be degraded and synthesized rapidly after damage. Therefore, the decline in growth under winter condition might result from the increased metabolic costs of photoprotection and elevated D1 turnover under the combination of short daylength limitation and low temperature (Hoppe et al., 2015).

Although RbcL decreased with elevated CO₂ in some studies that might be caused by the decrease in RuBP concentration for diatoms (Endo et al., 2015), McCarthy et al. (2012) observed an increase in Rubisco concentration with higher CO₂, which is in line with the present study under winter condition. Light and temperature could affect RbcL amount in phytoplankton. RbcL increased slightly with higher CO₂ at low light intensity condition, while it decreased slightly at higher light intensity in the research of Levitan et al. (2010). In the present study, the combination of low temperature and short daylength might lead to a complex trend of RbcL, elevated CO₂ might stimulate the sensitivity of *S. costatum* to low temperature which lead to a steep rise of RbcL in winter.

For diatoms, a mounting body of studies has pay attention to the effect of interactions of ocean acidification and other environmental factors such as light intensity, UV, temperature, nutrient limitation, salinity or photoperiod (Gao et al., 2012; Yue et al., 2019), and the results showed positive, negative or neutral effects (Xu et al., 2014c; Li et al., 2017). However, few studies combined elevated pCO₂ with seasonal changes in seawater physical and chemical characters on marine diatoms. In this study, temperature and photoperiod were chosen as seasonal factors to investigate the combined effects of pCO₂ and these two factors on physiology of *S. costatum*. Our results suggested temperature and photoperiod could mediate effects of elevated pCO₂ on the typical diatom *S. costatum*. Positive effects of OA on growth and photosynthesis were observed in spring and autumn, while negative effects were found in winter condition. To better understand how global climate changes would affect marine diatoms in the future, it is necessary to explore the interactive effects of ocean acidification with seasonal changes in seawater characters.

Author Contributions

JX and FL conceived and designed the experiments; HL and TX carried out the experiments; HL, FL and JX analyzed data; HL wrote the draft of the paper; FL, JM and JX revised the manuscript and approved this version for submission.

305 Competing interests

The authors declare that they have no conflict of interest.

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Table 1: Significance test results of growth, BSi, photosynthetic and respiration rates, parameters of P-I (P_{\max} is the maximum net photosynthetic rate, α is the photosynthetic efficiency, R_d is the dark respiration rate), P-C (K_m is half-saturation constant and V_{\max} is the inorganic carbon-saturated maximal rate of photosynthesis) and RLCs curves ($rETR_{\max}$, which is the maximum relative electron rate) for combination of temperature and photoperiod (season), CO_2 , and their interactions.

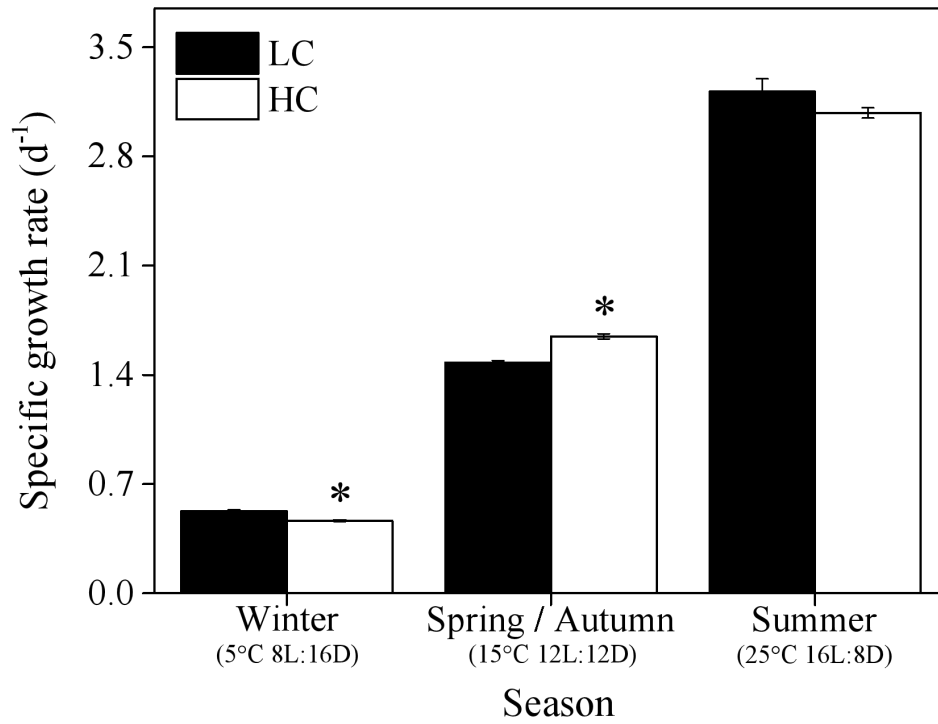
Parameter	Season		CO_2		Season * CO_2	
	F	<i>p</i>	F	<i>p</i>	F	<i>p</i>
Specific growth rate	7662.1	<0.001	0.3	0.569	27.3	<0.001
BSi	22.6	<0.001	22.5	<0.001	10.9	<0.001
Photosynthesis	452.1	<0.001	0.1	0.735	4.5	0.0345
Dark respiration	51.9	<0.001	7.6	0.018	1.9	0.182
P_{\max}	85.5	<0.001	6.3	0.028	3.4	0.069
α	1.8	0.211	6.2	0.028	1.5	0.262
R_d	7.3	0.010	3.3	0.097	4.2	0.044
V_{\max}	88.4	<0.001	136.8	<0.001	49.9	<0.001
K_m	20.5	<0.001	35.7	<0.001	6.7	0.011
$rETR_{\max}$	85.3	<0.001	98.5	<0.001	26.0	<0.001

Table 2: Chl *a* ($pg\ cell^{-1}$) and BSi ($pmol\ cell^{-1}$) contents of *S. costatum* acclimated to ambient and elevated pCO_2 in different seasons. The data are mean \pm SD values of triplicate cultures ($n = 3$). Different lowercases represent significant differences ($P < 0.05$) between two CO_2 levels under same season (*t*-test).

Treatments	Chl <i>a</i>		BSi	
	LC	HC	LC	HC
Winter	0.18 \pm 0.008 ^a	0.15 \pm 0.013 ^b	0.035 \pm 0.003 ^a	0.025 \pm 0.003 ^b
Spring / Autumn	0.19 \pm 0.007 ^a	0.26 \pm 0.014 ^b	0.025 \pm 0.002 ^a	0.019 \pm 0.001 ^b
Summer	0.22 \pm 0.015 ^a	0.23 \pm 0.024 ^a	0.023 \pm 0.001 ^a	0.024 \pm 0.002 ^a

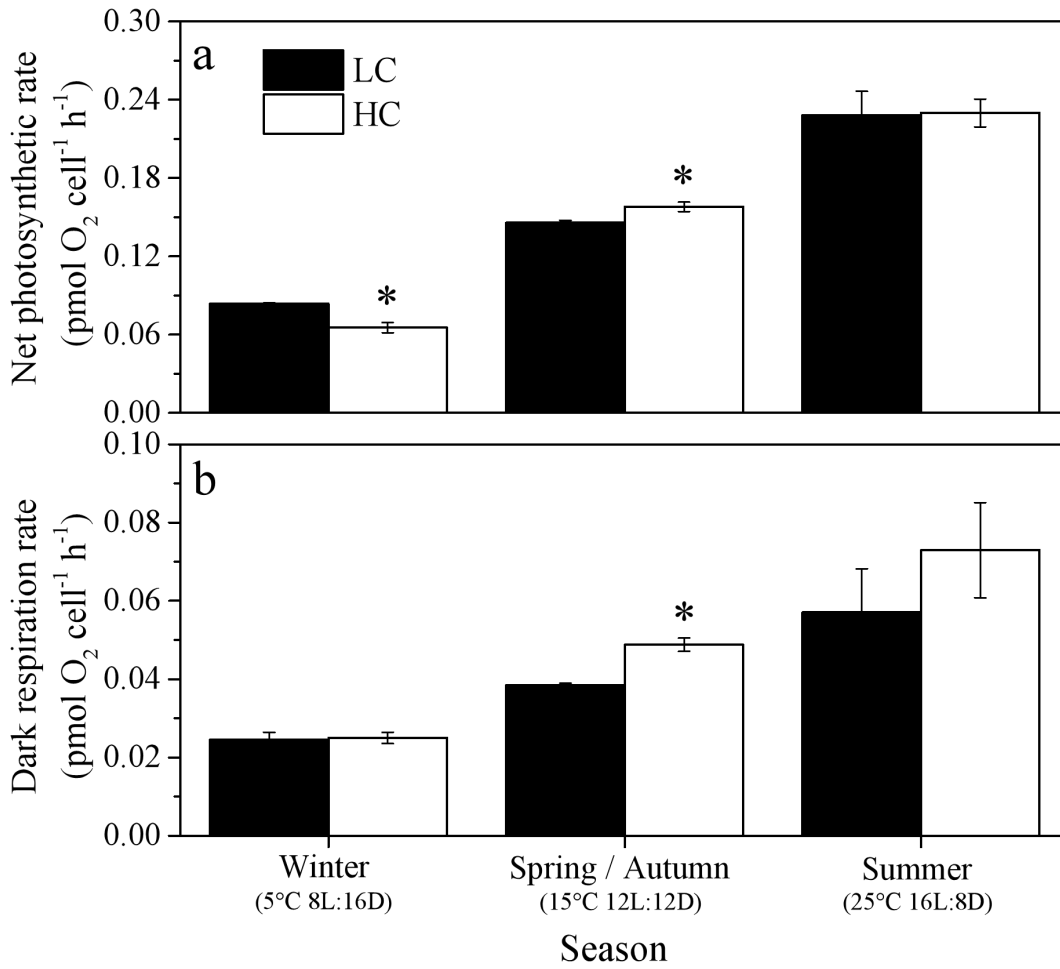
Table 3: Photosynthetic parameters fitted from $P_{(O_2)}$ -I and rapid light curves for *S. costatum* acclimated to ambient and elevated pCO_2 in different seasons. P_{\max} ($pmol\ O_2\ cell^{-1}\ h^{-1}$) is the maximum net photosynthetic rate, α is the photosynthetic efficiency, R_d ($pmol\ O_2\ cell^{-1}\ h^{-1}$) is the dark respiration rate, I_k ($\mu mol\ photons\ m^{-2}\ s^{-1}$) is the photosynthetic saturated light intensity, I_c ($\mu mol\ photons\ m^{-2}\ s^{-1}$) is light compensation point, $rETR_{\max}$ is the maximum relative electron rate and Inh is the relative photoinhibition ratio of rETR. Different lowercases represent significant differences ($P < 0.05$) between two CO_2 levels under same season (*t*-test).

	P_{\max}	$\alpha(O_2)$	R_d	$I_k(O_2)$	I_c	$I_k(ETR)$	$\alpha(ETR)$	$rETR_{\max}$	Inh (%)
Winter-LC	0.13 \pm 0.01 ^a	0.0012 \pm 0.00007 ^a	0.011 \pm 0.006 ^a	112.4 \pm 17.3 ^a	9.8 \pm 5.1 ^a	302.7 \pm 8.3 ^a	0.21 \pm 0.008 ^a	59.0 \pm 1.76 ^a	147.3 \pm 6.7 ^a
Winter-HC	0.11 \pm 0.02 ^a	0.0012 \pm 0.00045 ^a	0.003 \pm 0.002 ^a	97.5 \pm 21.7 ^a	7.2 \pm 3.3 ^a	193.6 \pm 10.7 ^b	0.22 \pm 0.005 ^a	40.3 \pm 1.43 ^b	135.0 \pm 16.8 ^a
Spring-LC	0.23 \pm 0.04 ^a	0.0011 \pm 0.00012 ^a	0.007 \pm 0.003 ^a	204.1 \pm 28.0 ^a	5.7 \pm 1.9 ^a	373.0 \pm 8.5 ^a	0.20 \pm 0.0005 ^a	67.4 \pm 1.59 ^a	68.7 \pm 6.9 ^a
Spring-HC	0.31 \pm 0.03 ^b	0.0016 \pm 0.00019 ^b	0.019 \pm 0.007 ^a	192.3 \pm 19.9 ^a	11.1 \pm 3.5 ^a	386.6 \pm 10.2 ^a	0.21 \pm 0.005 ^b	61.6 \pm 3.77 ^a	54.5 \pm 4.3 ^b
Summer-LC	0.35 \pm 0.05 ^a	0.0011 \pm 0.00009 ^a	0.016 \pm 0.008 ^a	336.7 \pm 66.3 ^a	14.5 \pm 7.1 ^a	465.1 \pm 18.6 ^a	0.12 \pm 0.018 ^a	57.1 \pm 0.04 ^a	28.9 \pm 4.3 ^a
Summer-HC	0.41 \pm 0.05 ^a	0.0013 \pm 0.00018 ^a	0.032 \pm 0.011 ^a	322.8 \pm 36.1 ^a	25.4 \pm 10.5 ^a	462.3 \pm 16.8 ^a	0.16 \pm 0.009 ^b	53.8 \pm 1.29 ^b	27.1 \pm 2.7 ^a



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Figure 1: Specific growth rate of *S. costatum* acclimated to ambient (LC, black bars) and elevated pCO₂ (HC, white bars) under different combination of temperature and photoperiod conditions. The data are mean ± SD values of triplicate cultures (n = 3). Asterisks represent significant differences ($P < 0.05$) between two CO₂ levels under same season condition (*t*-test).



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Figure 2: Net photosynthetic (a) and dark respiration rates (b) of *S. costatum* acclimated to ambient (LC, black bars) and elevated pCO₂ (HC, white bars) under different combination of temperature and photoperiod conditions. The data are mean ± SD values of triplicate cultures (n = 3). Asterisks represent significant differences (P < 0.05) between two CO₂ levels under same season condition (t-test).

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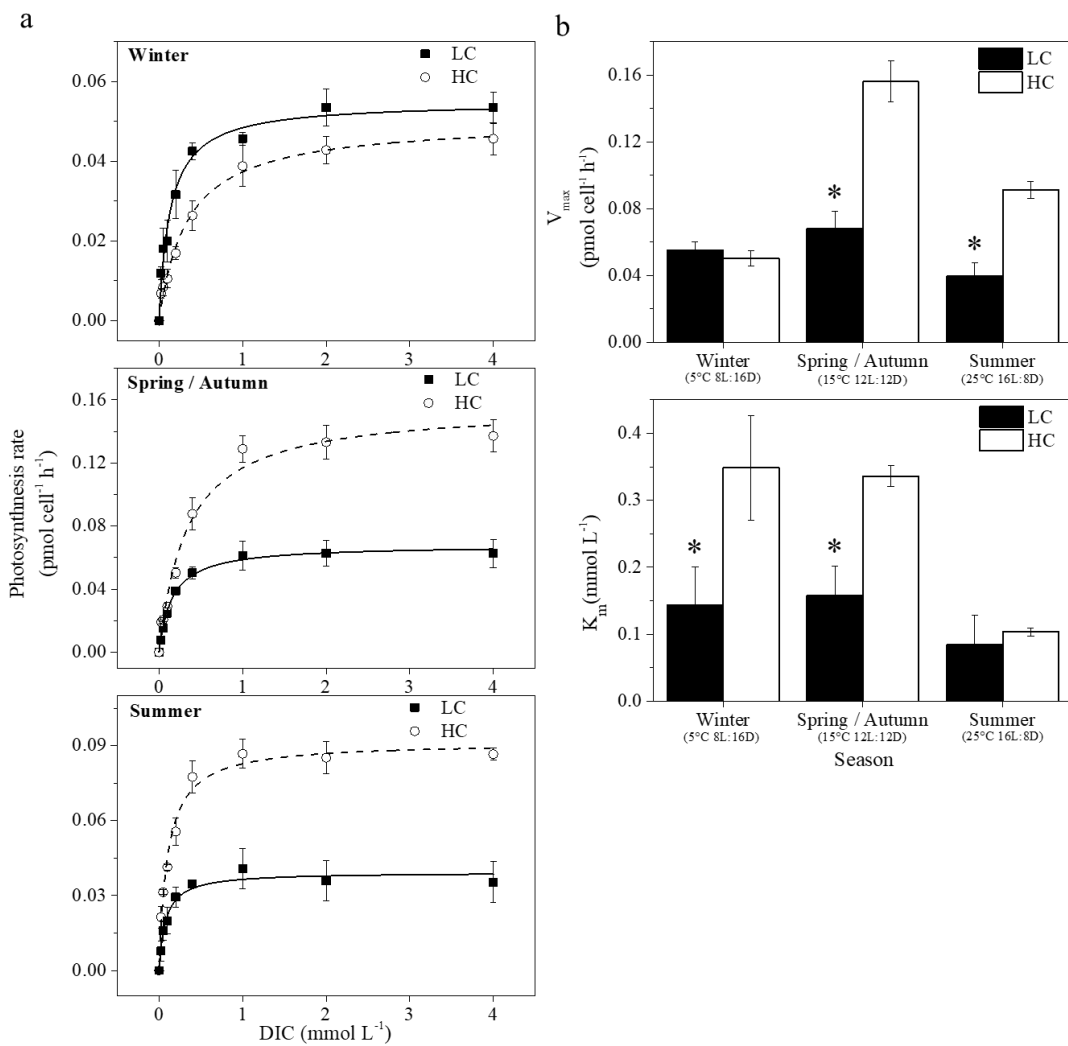
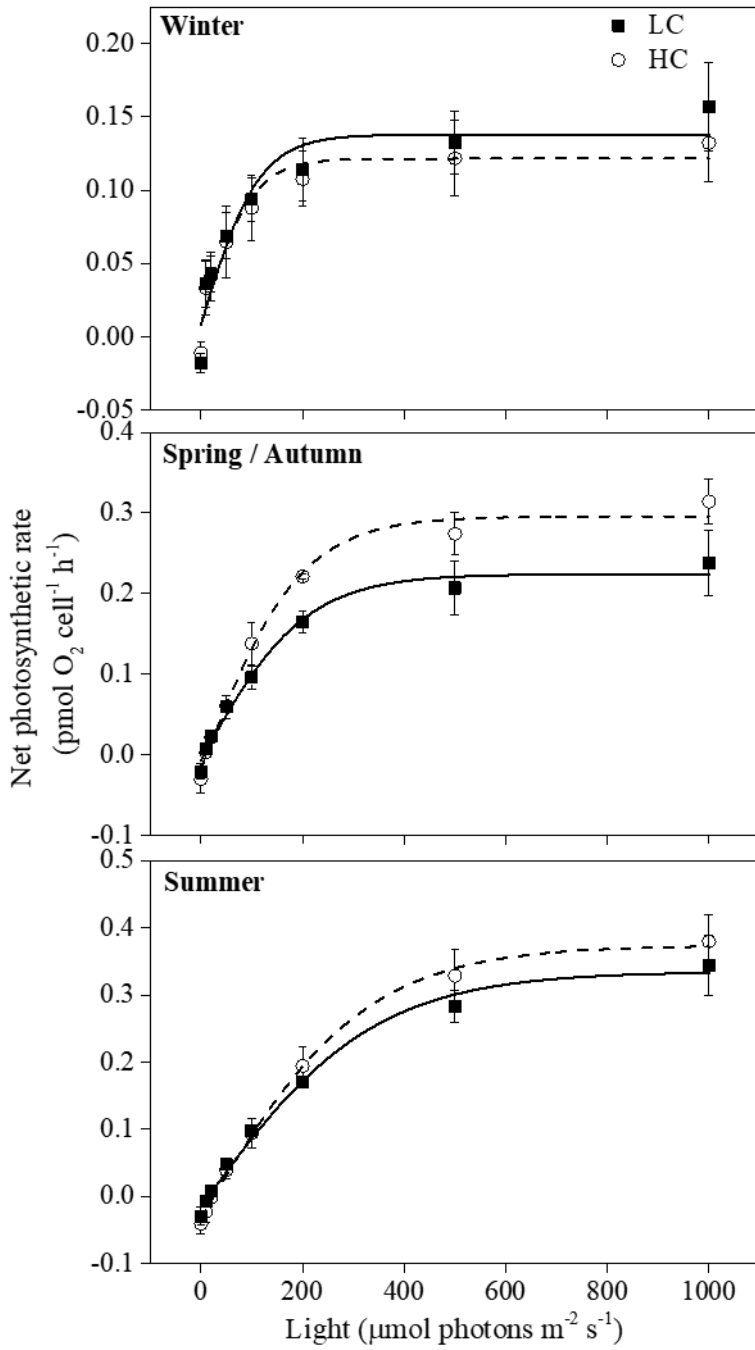


Figure 3: (a) Photosynthesis-inorganic carbon (P-C) curves of cells acclimated to ambient and elevated pCO₂ in different seasons. (b) Photosynthetic parameters of P - C curves at ambient (LC, black bars) and elevated pCO₂ (HC, white bars) in different seasons. K_m is half-saturation constant and V_{max} is the inorganic carbon-saturated maximal rate of photosynthesis. The data are mean ± SD values of triplicate cultures (n = 3). Asterisks represent significant differences (P < 0.05) between two CO₂ levels under same season (t-test).

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495 **Figure 4:** Photosynthesis-light curves (P-I curves) of cells acclimated to ambient and elevated pCO₂ in different seasons. The data are mean ± SD values of triplicate cultures (n = 3).

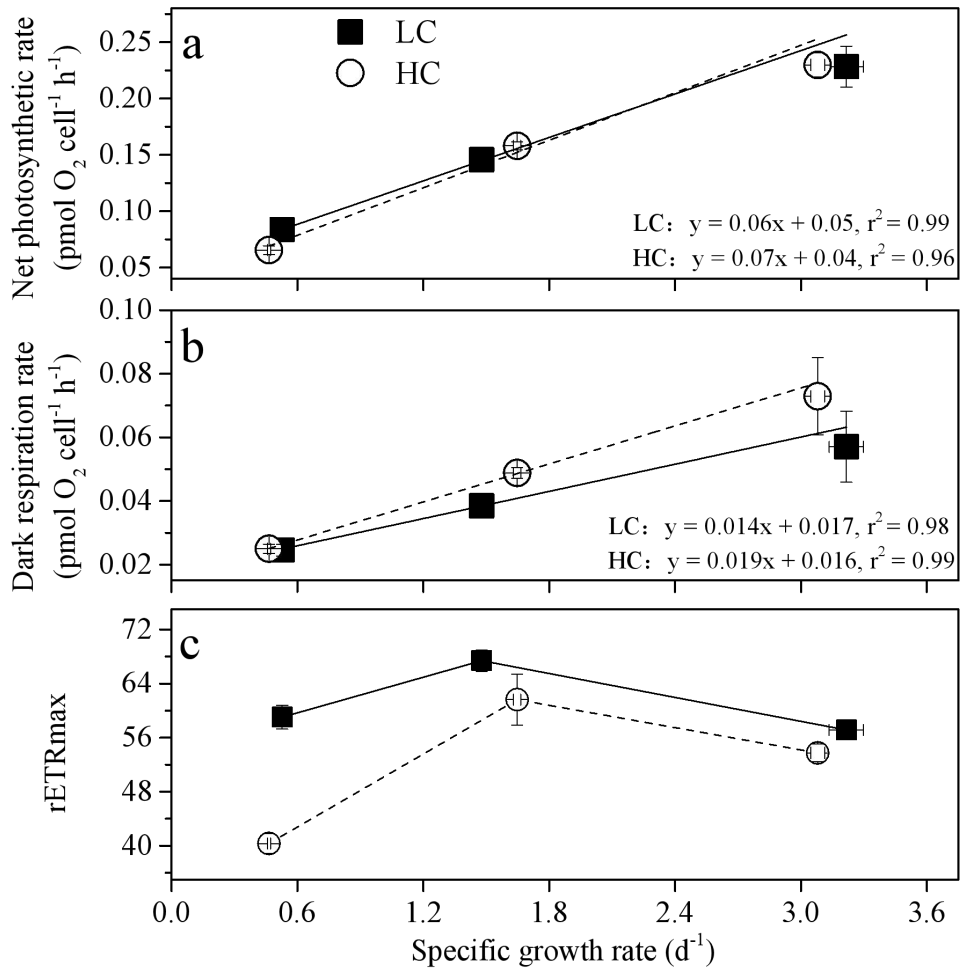


Figure 5: The relationship between net photosynthetic rate (a), dark respiration (b), rETR_{max} (c) and specific growth rate of *S. costatum* acclimated to ambient (LC, black square) and elevated pCO₂ (HC, white circle) under different combination of temperature and photoperiod conditions. The data are mean ± SD values of triplicate cultures (n = 3).

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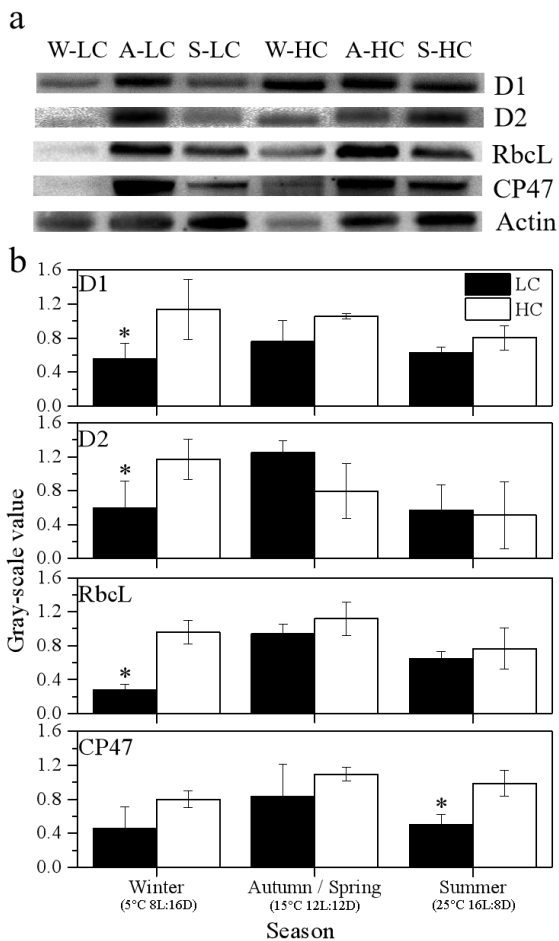


Figure 6: (a) Immunoblot using antibodies against **RbcL** and key **PSII** proteins (PsbA (D1), PsbD (D2), and PsbB (CP47)) isolated from *S. costatum* acclimated to ambient and elevated pCO₂ in different season levels (W for winter, A for autumn, S for summer). Each line was loaded with similar amounts of proteins. (b) Quantitative analysis of proteins. The relative abundance of each band was estimated by densitometric scanning of the exposed films. Asterisks represent significant differences ($P < 0.05$) between two CO₂ levels under same season (t -test).

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