1	Biogeochemical and plant trait mechanisms drive enhanced methane emissions
2	in response to whole-ecosystem warming
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27 Abstract

28 Climate warming perturbs ecosystem carbon (C) cycling, causing both positive and negative feedbacks on greenhouse gas emissions. In 2016, we began a tidal marsh field experiment in two 29 30 vegetation communities to investigate the mechanisms by which whole-ecosystem warming alters C gain, 31 via plant-driven sequestration in soils, and C loss, primarily via methane (CH₄) emissions. Here, we 32 report the results from the first four years. As expected, warming of 5.1 °C more than doubled CH₄ 33 emissions in both plant communities. We propose this was caused by a combination of four mechanisms: 34 (i) a decrease in the proportion of CH_4 consumed by CH_4 oxidation, (ii) more C substrates available for 35 methanogenesis, (iii) reduced competition between methanogens and sulfate reducing bacteria, and (iv) 36 indirect effects of plant traits. Plots dominated by Spartina patens consistently emitted more CH₄ than 37 plots dominated by Schoenoplectus americanus, indicating key differences in the roles these common 38 wetland plants play in affecting anerobic soil biogeochemistry and suggesting that plant composition can 39 modulate coastal wetland responses to climate change.

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41 **1. Introduction**

42 Methane (CH₄) is a potent greenhouse gas that contributes to 15-19 % of total greenhouse gas 43 radiative forcing (IPCC, 2013) and has a sustained-flux global warming potential that is 45 times that of CO2 on a 100-year timescale (Neubauer & Megonigal, 2015). Wetlands are the largest natural source of 44 45 CH₄ to the atmosphere and were recently identified as the largest source of uncertainty in the global CH₄ 46 budget (Saunois et al., 2016). Recent estimates calculate that CH₄ emissions from vegetated coastal 47 wetlands offset 3.6 of the 12.2 million metric tons (MMT) of CO₂ equivalents accumulated by these 48 ecosystems each year (EPA, 2017). Despite this, there is still a substantial knowledge gap regarding how 49 global change factors, such as climate warming, will alter coastal wetland CH_4 emissions (Mcleod et al., 50 2011) even though these feedbacks have the potential to shift coastal wetlands from being a net sink of C 51 to a net source (Al-Haj and Fulweiler, 2020; Bridgham et al., 2006).

52	The net flux of CH ₄ to the atmosphere from any ecosystem represents the balance between the
53	amount of CH ₄ produced (methanogenesis), the amount of CH ₄ oxidized (methanotrophy), and the rate of
54	CH ₄ transport from the soil. In coastal wetlands, methanogenesis occurs through three pathways: (i)
55	hydrogenotrophic methanogenesis (i.e. CO_2 reduction) in which H_2 is the electron donor and CO_2 is the
56	electron acceptor, (ii) acetoclastic methanogenesis, in which acetate (CH ₄ COOH) is split into CH ₄ and
57	CO_2 and (iii) methylotrophic methanogenesis in which methylated compounds are converted to CH_4 and
58	CO ₂ (Conrad, 2020; Oremland et al., 1982; Schlesinger and Bernhardt, 2020). Rates of methanogenesis
59	are driven by low redox conditions and substrate availability, while aerobic CH ₄ oxidation requires both
60	O ₂ and CH ₄ as substrates. Roots and rhizomes in wetland ecosystems influence methane-related substrates
61	through at least two mechanisms: (i) deposition of organic compounds that support multiple pathways of
62	heterotrophic microbial respiration, including methanogenesis, and (ii) release of O ₂ that simultaneously
63	promotes CH ₄ oxidation and regeneration of competing electron acceptors such as Fe(III) and SO ₄
64	(Philippot et al., 2009; Stanley and Ward, 2010). Root exudates, which typically include low-molecular-
65	weight compounds, may either be more readily used by microbes than existing soil C (Kayranli et al.,
66	2010; Megonigal et al., 1999) or may prime microbial use of soil C (Basiliko et al., 2012; Philippot et al.,
67	2009; Robroek et al., 2016; Waldo et al., 2019). Root exudates can also decrease CH ₄ oxidation by
68	stimulating use of O ₂ by other aerobic microbes (Lenzewski et al., 2018; Mueller et al., 2016).
69	Consequently, wetland CH ₄ emissions are strongly linked to a wide variety of plant traits that govern the
70	supply of reductive (organic carbon) and oxidative (O ₂) substrates to soils (Moor et al., 2017; Mueller et
71	al., 2020).
72	Although it is understood that wetland plants are a primary control on CH ₄ emissions and that much
73	of their influence is mediated through conditions in the rhizosphere (Waldo et al., 2019), there are
74	surprisingly few data, especially from coastal wetlands, that couple plant responses to the dynamics of
75	electron donors (organic C), electron acceptors (O ₂ , SO ₄), and the rates of competing (sulfate reduction vs
76	methanogenesis) or opposing (CH ₄ production vs CH ₄ oxidation) microbial processes. The general lack of
77	process data on wetland CH_4 cycling makes it difficult to forecast ecosystem responses to climate change. 3

78 For example, the well-documented observation that warming increases wetland methane emissions can be 79 either amplified or dampened depending on changes in plant activity (e.g. primary production) or plant 80 traits (e.g. community composition) (Mueller et al., 2020). Vegetation composition has been shown to be 81 a stronger control on CH₄ emissions than ~ 1 °C of warming in northern peatlands (Ward et al., 2013) and 82 Chen et al. (2017) proposed that warming effects on plant functional types can drive C flux responses that 83 cannot otherwise be explained by abiotic conditions. In freshwater marshes, plant species and growth 84 trends have also been linked to seasonal shifts in pools of dissolved CH₄ and DIC (Ding et al., 2005; 85 Stanley and Ward, 2010) and methanogenesis dynamics (Sorrell et al., 1997). 86 Tidal wetlands are particularly good model systems for determining the mechanisms by which warming alters CH₄ emissions. Not only will the CH₄ cycle respond to the direct effects of warming, but 87 88 the temperature effects on the outcome of competition for electron acceptors is relatively easily observed 89 because of the abundance of SO₄. Thermodynamic theory in which terminal electron acceptors (TEAs) 90 are used in order of decreasing thermodynamic yield is commonly interpreted to mean that a system will 91 support only one form of anaerobic respiration at time, with acetoclastic and hydrogenotrophic 92 methanogenesis occurring only when pools of more energetically favorable TEAs have been depleted 93 (Conrad, 2020; Schlesinger and Bernhardt, 2020). However, in real systems with spatial and temporal 94 variability in the supply of electron donor substrates and TEAs, all forms of anaerobic metabolism occur 95 simultaneously (Megonigal et al. 2004, Bridgham et al., 2013). Much of this spatial and temporal 96 variation arises from the distribution and activity of roots and rhizomes as mediated by the rhizosphere 97 (Neubauer et al., 2008). Global change factors such as warming will further affect the spatial distribution 98 of key metabolic substrates. In addition, the relatively-limited species diversity in saline tidal wetlands 99 allows species-level effects on CH_4 cycling to be delineated more easily than in diverse freshwater 100 wetlands.

Methane flux measurements are a metric of broader shifts in redox potential and biogeochemical
 cycling, as they are sensitive to virtually all processes that regulate availability of electron donors and
 electron acceptors. Emissions are commonly predicted to increase with future climate warming, including 4

from coastal wetlands (Al-Haj and Fulweiler, 2020), but there is minimal prior understanding of the underlying mechanisms, which was the focus of this study. Our objectives were to explore the mechanisms that drive enhanced CH₄ emissions under warming. To accomplish this, we measured monthly CH₄ emissions from 2016 through 2019 and coupled these flux measurements with analysis of porewater biogeochemistry and vegetation biomass and composition.

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110 **2. Materials and Methods**

111 <u>2.1 Site Description and Experimental Design</u>

112 The Salt Marsh Accretion Response to Temperature eXperiment (SMARTX) was established in the 113 Smithsonian's Global Change Research Wetland (GCReW) in 2016. GCReW is part of Kirkpatrick 114 Marsh, a microtidal, brackish high marsh on the western shore of the Chesapeake Bay, USA (38°53' N, 115 76°33' W). Soils are organic (>80 % organic matter) to a depth of 5 m, which is typical of high marshes 116 in the Chesapeake Bay and elsewhere. The very low mineral content (<20%) affects methane dynamics 117 because negligible competition between methanogens and iron-reducing bacteria for electron donors is 118 expected in the absence of a significant pool of poorly crystalline iron oxides (Roden and Wetzel, 1996), 119 as has been documented previously at this site (Weiss et al., 2004). Soil bulk density in the upper 60 cm averages 0.124 g cm³ and ranges from 0.079 to 0.180 g cm³. The relatively uniform bulk density of the 120 121 soil profile reflects the uniform soil organic matter content and the fact that bulk density becomes largely 122 independent of organic matter and mineral content once organic matter content exceeds 50% (Holmquist 123 et al., 2018). The marsh is typically saturated to within 5-15 cm of the soil surface, but inundation 124 frequency varies across the site, from 10-20 % of high tides in high elevation areas to 30-60 % of high 125 tides in low elevation areas.

SMARTX consists of six replicate transects, three located in each of the two dominant annual plant communities (Fig. S1). In the C₃-dominated community (herein the 'C₃ community') the C₃ sedge *Schoenoplectus americanus* (herein *Schoenoplectus*) composes more than 90 % of the aboveground biomass (Table 1). In the C₄-dominated community (herein the 'C₄ community'), 75 % of the

- 130 aboveground biomass was initially composed of two C₄ grasses (Spartina patens and Distichlis spicata,
- 131 herein Spartina and Distichlis, respectively). However, by 2019, Spartina and Distichlis declined to 56 %

132 of the aboveground biomass (Table 1).

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Table 1. Relative contribution to total aboveground biomass from C_3 sedges (*Schoenoplectus americanus*) and C_4 grasses (*Spartina patens* and *Distichlis spicata*) in each plant community. Values are means and SE (n = 12).

Each transect is an active warming gradient consisting of unheated ambient plots and plots that

	C_3 community		C_3 community C_4 community	
Year	% C ₃	% C ₄	% C ₃	% C ₄
2016	93 (3)	8 (3)	8 (2)	76 (6)
2017	91 (3)	9 (3)	10 (3)	64 (4)
2018	95 (1)	5 (1)	15 (4)	65 (7)
2019	93 (2)	4(1)	23 (5)	56 (6)

134 are heated to 1.7 °C, 3.4 °C, and 5.1 °C above ambient. All plots are 2 x 2 meters with a 20 cm-wide buffer around the perimeter. Aboveground plant-surface temperature is elevated via infrared heaters and 135 136 soil temperature is elevated via vertical resistance cables (Rich et al., 2015). Soils are heated to a depth of 137 1.5 m, which is the depth most vulnerable to climate or human disturbance (Pendleton et al., 2012). 138 Aboveground and belowground temperature variation are assessed via thermocouples embedded in 139 acrylic plates at plant canopy level and inserted into the soil, respectively, and the temperature gradient is 140 maintained by integrated microprocessor-based feedback control (Rich et al., 2015). Noyce et al. (2019) 141 provides additional details of the heating system. Warming began on 1 Jun 2016 and has continued year-142 round. 143 2.2 Methane flux measurements 144 Methane emissions were measured monthly year-round from May 2016 to Dec 2019 using a static chamber system. One permanent 160 cm² aluminum base was inserted 10 cm into the soil in each plot in 145 146 Apr 2016. On each measurement date, clear chambers (40 x 40 x 40 cm) were gently placed on top of 147 each base and secured with compression clips. Chambers consisted of an aluminum frame with 148 transparent sides made of polychlorotrifluoroethylene film (Honeywell International) and closed-cell 149 foam on the base. Depending on the height of the vegetation at the time of measurement, chambers were 150 stacked up to four high (total height of 40 to 160 cm) (Fig. S2). The advantage of this stacking method is

151 that it uses the minimal chamber volume necessary, while also allowing for plant growth. After 152 placement, the chambers were left open for at least 10 minutes, to minimize disturbance effects and allow 153 air inside the chamber to return to ambient conditions. During data collection, chambers were covered 154 with a transparent polycarbonate top equipped with sampling tubes, a fan to circulate air inside the 155 chambers, a PAR sensor, and thermocouples. The sealed chamber was covered with a foil shroud to block 156 out all light and to minimize changes in temperature and relative humidity during the measurement 157 period. An UltraPortable Greenhouse Gas Analyzer (Los Gatos Research, CA) was used to measure 158 headspace CH₄ concentrations for 5 min. Plots were accessed from permanent boardwalks elevated 15 cm 159 above the soil surface to avoid compressing the surrounding peat and altering diffusive CH_4 emissions. 160 Fluxes were calculated as the slope of the linear regression of CH₄ concentration over time. The 40 fluxes 161 where p > 0.05 were assigned a value of $\frac{1}{2}$ the limit of detection of the system (Wassmann et al., 2018; 162 Table S1). This was 1 % of fluxes during the growing season and 4.5 % of fluxes over the remaining 163 months. For 2017-2019, monthly measurements were scaled to annual estimates by regressing CH₄ 164 emissions against daily mean soil temperature and day of year (as a proxy for phenological status). 165 Annual estimates were not calculated for 2016 because flux measurements did not start until May. 166 2.3 Porewater sampling and analysis 167 Porewater samples were collected in May, Jul, and Sep of each year using stainless steel 'sippers'

168 permanently installed in each plot. Each sipper consisted of a length of stainless-steel tubing, crimped and 169 sealed at the end, with several slits (approximate width 0.8 mm) cut in the bottom 2 cm. The aboveground 170 portion of each sipper was connected to Tygon Masterflex® tubing capped with a 2-way stopcock. In 171 May 2016, duplicate clusters of sippers were installed in each of the 30 plots at 20, 40, 80, and 120 cm 172 below the soil surface. An additional set of 10 cm-deep sippers was installed in 2017. In this study we 173 defined samples from 10-20 cm as "rooting zone" samples and samples from 40-120 cm as "deep peat" 174 samples. On sampling dates, porewater sitting in the sippers was drawn up and discarded, after which 60 175 mL of porewater from each depth (30 mL from each sipper) was withdrawn and stored in syringes 176 equipped with 3-way stopcocks. A 10 mL-aliquot of each sample was filtered through a pre-leached 0.45 7

µm syringe-mounted filter, preserved with 5 % zinc acetate and sodium hydroxide, and frozen for future
SO₄ and Cl analysis. Dissolved CH₄ was extracted from 15 mL of porewater in the syringe by drawing 15
mL of ambient air and shaking vigorously for 2 min to allow the dissolved CH₄ to equilibrate with the
headspace. Headspace subsamples were then immediately analyzed on a Shimadzu GC-14A gas
chromatograph equipped with a flame ionization detector. 3 mL of porewater was used to measure pH
using a Fisher Scientific accumet electrode (13-620-290). The remaining porewater was used to measure
H₂S, and NH₄; those data are not reported here.

184 SO₄ and Cl were measured on a Dionex ICS-2000 ion chromatography system (2016-2018) or a

185 Dionex Integrion (2019). On the Dionex ICS-2000 samples were separated using an A11 column with 30

186 mM of KOH as eluent; on the Dionex Integrion samples were separated using an A11-4 μ m-fast column

187 with 35 mM KOH. Sulfate depletion (Sulfate_{Dep}) was calculated based on measured porewater

188 concentrations of SO_4 (SO_{4pw}) and Cl (Cl_{pw}) and the constant molar ratio of Cl to SO_4 in surface seawater

189 ($R_{sw} = 19.33$; Bianchi, 2006) using the following equation: Sulfate_{Dep} = Cl_{pw}/R_{sw} - SO_{4pw}. If driven only

190 by seawater inputs, the ratio of Cl to SO₄ would remain constant, but under anaerobic conditions SO₄ can

191 be reduced by sulfate reducing bacteria, altering this ratio. As a result, SO₄ depletion can be used as a

192 proxy for SO₄ reduction rates.

193 <u>2.4 Plant biomass measurements</u>

194 Measurements of *Schoenoplectus, Spartina*, and *Distichlis* biomass were conducted during peak

195 biomass of each year (29 Jul – 2 Aug) as described by Noyce et al. (2019). *Schoenoplectus* biomass was

196 estimated using non-destructive allometric techniques (Lu et al., 2016) in 900-cm² quadrats, and *Spartina*

197 and *Distichlis* biomass were estimated through destructive harvest of 25-cm² subplots.

198 <u>2.5 Data analysis</u>

199 Statistics were conducted in R (version 3.6.3). Methane flux (Fig. S3) and porewater data were log

200 transformed to become normally distributed prior to statistical analyses. The 'growing season' was

- 201 defined as May through Sep based on *Schoenoplectus* growth trends (Fig. S4). Pearson's correlations
- were used to test the relationships between CH₄ flux and soil temperature, and CH₄ flux and plant 8

203	biomass. Responses of CH ₄ emissions to vegetation type and warming treatment were analyzed using
204	linear mixed models with vegetation community and warming treatment as categorical variables, and
205	plot and year as random effects. P-values were calculated using Satterthwaite's method and Tukey's
206	post-hoc tests were used to compare individual means. Porewater data was averaged per year and then
207	analyzed using one-way ANOVAs to determine the effects of warming treatment or plant community,
208	applying Tukey's HSD test for post-hoc analyses.
209	
210	3. Results
211	3.1 Environmental conditions, site characteristics, and experiment performance
212	The growing season of 2016 was the hottest of the four years, with growing season temperatures
213	averaging >1 $^{\circ}$ C above the other three years (Table 2). While 2017 through 2019 had similar summer
214	temperatures, they had very different precipitation regimes; 2018 was much wetter on average and 2019
215	was slightly drier (Table 2). During all years, temperatures in the experimental plots were successfully
216	shifted by the target differentials of +1.7, +3.4, and +5.1 °C above the ambient plots (Fig. 1 (top); Noyce
217	et al., 2019). Porewater pH ranged from 6.4 to 6.8 across the measurement period, with no effect of
218	temperature treatment ($p > 0.1$; data not shown). There was no difference in soil bulk density between the
219	ambient and +5.1 °C plots after 4.5 years of warming ($p = 0.54$; data not shown).
220	

Table 2. Growing season	Year	Mean aboveground	Mean soil	Total
(May-Sep) temperature and		temperature (°C)	temperature (°C)	precipitation (cm)
precipitation. Temperature	2016	24.7 (0.3)	22.4 (0.2)	51.0
data are means (SE) of daily	2017	22.0 (0.3)	20.7 (0.2)	51.1
averages from ambient plots	2018	23.7 (0.3)	20.8 (0.2)	86.4
and precipitation is total from	2019	23.6 (0.3)	20.9 (0.2)	43.7
May through September.		· ·	· · ·	

221 <u>3.2 Methane fluxes</u>

222 Methane emissions increased with soil temperature ($R^2 = 0.41$, *p* <0.001) (Fig. 1). Emissions from 223 all treatments had strong seasonal trends; fluxes were highest in the C₃ community in Jun through Aug

- and peak fluxes in the C₄ community were shifted about a month later to Jul through Sep (Fig. S3).
- 225 Whole-ecosystem warming increased CH₄ emissions throughout the growing season ($F_{3,400} = 5.1$, p =
- 226 0.002; Fig. 2). Across all four years, 5.1 °C of warming more than doubled growing season emissions,
- 227 from 624 to 1413 μ mol CH₄ m⁻² d ($p_{adj} = 0.02$; Fig. 2).



Figure 1. Bottom: CH_4 emissions from each plot versus the soil temperature at the time of measurement. Top: Density plot depicting the range of soil temperatures in each treatment, delineated by color: ambient (blue), +1.7°C (green), +3.4°C (yellow), +5.1°C (red).





Figure 2. Comparison of CH₄ emissions from each warming treatment during the growing season (May-Sep). Means include both the C₃ and C₄ community and all years of measurement. Error bars indicate SE. Horizonal bars indicate means that are significantly different and the corresponding p_{adj} .

230 Mean CH₄ emissions were higher from the C₄ community than from the C₃ community both during the

231 growing season ($F_{1,22} = 13.6$, p = 0.001; Fig. 3a) and on an annual basis ($F_{1,22} = 8.5$, p = 0.008; Fig. 3b).

- 232 Mean annual CH₄ emissions ranged from 58 mmol CH₄ m^{-2} yr⁻¹ (ambient) to 343 mmol CH₄ m^{-2} yr⁻¹ (+5.1
- $^{\circ}$ C) in the C₃ community and from 55 mmol CH₄ m⁻² yr⁻¹ (ambient) to 879 mmol CH₄ m⁻² yr⁻¹ (+5.1 $^{\circ}$ C) in
- the C₄ community (Table S2). Under ambient conditions, growing season CH₄ fluxes were almost twice
- as large from C₄ plots, whereas under low warming (1.7 to 3.4 °C) this difference increased to more than
- three times as large (Fig. 3a). From 2017-2019, CH₄ emissions were positively related to Spartina and 10

- 237 *Distichlis* aboveground biomass across all warming treatments and negatively related to *Schoenoplectus*
- biomass (Fig. 4a,b). In 2016, however, the direction of those relationships in both plant communities were
- the exact opposite, with *Spartina* and *Distichlis* biomass negatively related, and *Schoenoplectus* biomass
- 240 positively related, to CH₄ emissions (Fig. 4a,b).



Figure 3. Comparison of CH₄ emissions from the C₃ community dominated by *Schoenoplectus* (open bars) and the C₄ community dominated by *Spartina* and *Distichlis* (grey bars). (a) During the growing season (May-Sep) and (b) scaled to a year. Means are averaged across all sampling dates for 2017 – 2019. Error bars indicate SE. Asterisks indicate significant differences between C₃ and C₄ means at a given temperature (* $p_{adj} < 0.05$, ** $p_{adj} < 0.01$).



Figure 4. Mean growing season (May-Sep) CH₄ emissions from each plot versus the biomass of (a) C_3 (*Schoenoplectus*) and (b) C_4 (*Spartina* and *Distichlis*) plants. All regressions are significant at p = 0.05.

241 <u>3.3 Porewater chemistry</u>

Under ambient conditions, porewater collected from the C4 community had more dissolved CH4 (F1,22 242 243 = 18.4, p < 0.001; Fig. 5a,b), less SO₄ (F_{1,22} = 29.1, p < 0.001; Fig 6a), and similar salinity (p = 0.068) 244 compared to the C_3 community. In the C_3 community, warming increased dissolved CH₄ in both the 245 rooting zone porewater (10-20 cm) ($F_{3,44} = 2.85$, p = 0.048; Fig. 5a) and in the deep peat (40-120 cm) $(F_{3,44} = 6.23, p = 0.001; Fig. 5b)$. Dissolved CH₄ concentrations were relatively similar in the ambient, 246 +1.7 °C, and +3.4 °C treatments, but more than doubled with +5.1 °C of warming in both the rooting zone 247 (59 to 125 μ mol CH₄ L⁻¹, $p_{adj} < 0.001$) and the deeper porewater (43 to 1254 μ mol CH₄ L⁻¹, $p_{adj} < 0.001$). 248 249 In the C_4 community there was minimal effect of warming treatment on porewater in the rooting zone 250 $(F_{3,44} = 0.442, p = 0.72; Fig. 5a)$, but all levels of warming decreased dissolved CH₄ below 40 cm $(F_{3,44} = 0.442, p = 0.72; Fig. 5a)$ 251 =129.3, p < 0.001), with concentrations in the +3.4 and +5.1 plots less than a third of the concentrations 252 in the ambient plots (155 vs 56 and 40 μ mol CH₄ L⁻¹, $p_{adj} < 0.001$; Fig. 5b).



Figure 5. Comparison of dissolved CH₄ from the C₃ community dominated by *Schoenoplectus* (open bars) and the C₄ community dominated by *Spartina* and *Distichlis* (grey bars). (a) In the dominant rooting zone (10-20 cm) and (b) below the rooting zone (40-120 cm). Means are averaged across all sampling dates for 2016 - 2019. Error bars indicate SE. Letters indicate temperature treatments that are significantly different from each other ($p_{adj} < 0.05$) within the same plant community.



Figure 6. Comparison of sulfate concentrations and estimated sulfate depletion from the C₃ community dominated by *Schoenoplectus* (open bars) and the C₄ community dominated by *Spartina* and *Distichlis* (grey bars). (a) Sulfate availability throughout the entire soil profile and (b) sulfate depletion in the rooting zone. Means are averaged across all sampling dates for 2016 – 2019. Error bars indicate SE. Letters indicate temperature treatments that are significant different from each other ($p_{adj} < 0.05$) within the same plant community.

253 In the C₃ community, SO₄ concentrations decreased with warming ($F_{3.44} = 3.76$, p = 0.017), but 254 warming effects on SO₄ cycling in the C₄ community were more mixed with +3.4 °C increasing SO₄ (p_{adi}) 255 = 0.048) but no other treatments having large effects (Fig. 6a). In all plots, the measured concentrations of 256 rooting-zone SO₄ were lower than expected based on salinity (Fig. 6b), indicating that SO₄ reduction 257 occurred. In both plant communities, the +5.1 °C treatments increased this SO₄-depletion effect compared 258 to ambient, though the effect was stronger in the C₃ community (p < 0.001) than the C₄ community (p =259 (0.04). (Fig. 6b). Dissolved CH₄ was highest in both plant communities when SO₄ concentrations were < 5 260 mmol $SO_4 L^{-1}$ (Fig. S5).

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262 **4. Discussion**

263 Soil temperature (both seasonal and experimental) and plant traits were both strong drivers of CH₄ 264 emissions from this site. This follows prior field, mesocosm, and incubation studies across a variety of 265 wetlands, in which temperature has been shown to be a strong predictor of CH_4 emissions (e.g. Al-Haj and Fulweiler, 2020; van Bodegom and Stams, 1999; Christensen et al., 2003; Dise et al., 1993; Fey and 266 267 Conrad, 2000; Liu et al., 2019; Ward et al., 2013; Yang et al., 2019; Yvon-Durocher et al., 2014) and in 268 which plant functional type has an interacting effect (e.g. Chen et al., 2017; Duval & Radu, 2018; Liu et 269 al., 2019; Mueller et al., 2020; Ward et al., 2013). Methane emissions are a function of the balance 270 between methanogenesis, CH₄ oxidation, and CH₄ transport, so explaining these results requires some 271 combination of stimulation of methanogenesis, reduction of CH₄ oxidation, or increase in CH₄ transport. 272 Prior data from brackish wetlands are limited, but incubation studies of freshwater wetland soils 273 typically show large increases in CH₄ fluxes with warming (van Bodegom and Stams, 1999; Duval and 274 Radu, 2018; Hopple et al., 2020; Inglett et al., 2012; Sihi et al., 2017; Wilson et al., 2016), indicating that 275 warming alters belowground processes. Though there is some evidence that rhizosphere temperature 276 alters CH₄ transport through rice aerenchyma (Hosono and Nouchi, 1997), any transport-driven effects in 277 this ecosystem would be transient unless there was a simultaneous increase in net CH_4 production (i.e. an 278 increase in methanogenesis that was not completely offset by methanotrophy). Instead, we observed a 14

sustained increase in CH₄ emissions, suggesting large shifts in anaerobic metabolism, especially with +5.1°C of warming. We propose four potential and non-exclusive mechanisms to explain the temperaturedriven increase in CH₄ emissions: (1) shifted ratios of CH₄ production to oxidation, (2) increased substrate availability, (3) reduced competition with sulfate reducers for H₂ and organic C, and (4) indirect plant trait effects (Fig. 7).



Figure 7. Schematic of mechanisms driving enhanced CH₄ emissions in response to warming. SCAM = *Schoenoplectus americanus*. SPPA = *Spartina patens*. Left: Processes under ambient conditions. Plants add organic compounds to the soil, which are transformed into other low-molecular-weight organic compounds. This pool, and processed soil organic matter, support terminal respiration processes dominated by SO₄ reduction over CH₄ production in organic-rich brackish marsh soils. Plants also transport O₂ which supports oxidation of a fraction of the CH₄ before it can be transported out of the soil. Right: Processes under warmed conditions. 1) Rates of CH₄ production increase more than rates of CH₄ oxidation. 2) Substrate availability increases as plants add more rhizodeposits and organic matter is more rapidly fermented to low-molecular-weight organic compounds and H₂. 3) The pool of electron donors available to methanogens increases as SO₄ reducers become SO₄ limited. 4) The dominant plant species have different effects on these processes with *S. americanus* driving a net increase in O₂ transport and *S. patens* driving a net increase in rhizodeposits.

284 <u>4.1 Whole-ecosystem warming promotes methanogenesis over CH₄ oxidation</u>

- Holding the supply of substrates and transport properties of the system constant, warming is expected
- to increase rates of CH₄ production relative to CH₄ oxidation due solely to differences in the temperature
- 287 dependence of each process (Megonigal et al., 2016). In wetland soils, the average Q₁₀ of methanogenesis

288	is 4.1 compared to 1.9 for aerobic CH4 oxidation (Segers, 1998), which means that a system starting with
289	a given initial ratio between the two processes will become increasingly dominated by methanogenesis as
290	soils warm. A corollary to this expected pattern is that the ratio of the two processes should be constant if
291	the Q ₁₀ responses are similar, an outcome that was supported with <i>in situ</i> measurements of the two
292	processes in a tidal freshwater forested wetland (Megonigal & Schlesinger, 2002). We did not quantify
293	the temperature dependence of CH ₄ production and oxidation in the present study, but based on the
294	literature (Segers, 1998) it is likely that methanogenic activity increased more than aerobic
295	methanotrophic activity in direct response to warming (Fig. 7, mechanism 1). Evidence for this is that
296	rhizosphere pools of porewater CH ₄ were highest in the warmest treatment (Fig. 5a); because this
297	occurred despite either no change or an increase in aboveground biomass (Noyce et al., 2019), which
298	would by itself have lowered porewater CH4 due to venting (plant transport), it indicates that CH4
299	production increased relative to the sum of aerobic and anaerobic methane oxidation.
300	4.2 Whole-ecosystem warming increases substrate availability for methanogens
301	Methanogenesis can be the terminal step of anaerobic decomposition, but a consortium of microbes is
302	required to break down soil organic matter to electron donor substrates that methanogens can metabolize.
303	The final step in any decomposition pathway involves the flow of electrons from organic matter (electron
304	donors) to a TEA. Under anaerobic conditions, this is accomplished by microbes that tend to specialize in
305	one TEA and compete for organic C as an electron donor (Megonigal et al., 2004). Consequently, the
306	supply of both electron donors and TEAs regulates the multi-step process of anaerobic decomposition and
307	thus ultimately controls CH4 emissions. For all pathways, methanogenic activity is typically limited by
308	the supply of electron donors, including low-molecular-weight organic compounds (e.g. acetate,
309	Neubauer & Craft, 2009) and H ₂ , a product of organic matter fermentation. We propose that whole-
310	ecosystem warming increases the availability of previously limited C substrates in two aspects (Fig. 7,
311	mechanism 2).

312 First, warming may directly influence C availability through biochemical kinetics. Even if organic inputs remained constant, warming likely accelerates fermentation of soil organic matter, increasing 313 314 substrate availability for methanogens. Second, the warmed plots had longer growing seasons than the 315 unheated controls (Noyce et al., 2019). This presumably increased inputs of root exudates and fresh 316 detritus, accelerating all forms of heterotrophic microbial respiration by providing organic material that is 317 decomposed into low-molecular-weight organic C compounds and H₂ (Philippot et al., 2009), stimulating 318 growing season CH₄ emissions from warmed plots. In 2017, we observed that gross primary production 319 was positively correlated with CH_4 emissions and that this effect increased with warming (Fig. S6). Prior 320 studies have also linked CH₄ production or emissions to rates of photosynthesis (Vann and Megonigal, 321 2003), periods of active growth (Chen et al., 2017; Ward et al., 2013), and plant senescence which 322 coincides with a pulse input of labile C from plants to soils (Bardgett et al., 2005).

323 Temperature-accelerated biochemical kinetics and increased electron-donor supply are mechanisms 324 that can increase methanogenesis without necessarily shifting methanogenic pathways. However, shifts in 325 the balance between hydrogenotrophic, acetoclastic, and methylotrophic methanogenesis pathways can be 326 expected with warming. For example, in Arctic permafrost, methylotrophic methanogenesis was found to 327 be more sensitive to warming than the other pathways (de Jong et al., 2018). Such shifts can be quantified 328 with future analyses of H_2 and low-molecular-weight organic compounds (e.g. Bridgham et al., 2013; 329 Yang et al., 2016), isotopic tracing of specific methanogenic pathways (e.g. Blaser & Conrad, 2016; 330 Conrad, 2005; Neumann et al., 2016; Whiticar, 1999), and molecular community analyses (e.g. Bridgham 331 et al., 2013; He et al., 2015; Wilson et al., 2016).

332 4.4 Whole-ecosystem warming reduces competition with sulfate reducers

333 While low-molecular-weight organic compounds are electron donors for acetoclastic methanogenic

respiration, they are also substrates for other microbial groups such as SO₄ reducers (Megonigal et al.,

335 2004; Ye et al., 2014). As a result, consumption of the limited organic carbon supply by SO₄ reducers

336 should (and often does) limit methanogenic activity, such that terminal microbial respiration is typically

337	dominated by SO ₄ reduction in brackish marshes (Sutton-Grier et al., 2011). Similarly, SO ₄ reducers are
338	more efficient than methanogens at competing for the H ₂ required for CO ₂ reduction (Kristjansson et al.,
339	1982). We did not measure rates of SO ₄ reduction in this study but can use SO ₄ depletion as a proxy;
340	more SO ₄ depletion indicates that more SO ₄ reduction has occurred. Warming generally increased SO ₄
341	depletion, especially in the plots dominated by Schoenoplectus (Fig. 6b). Differences in SO ₄ depletion
342	between plots is not driven by SO ₄ inputs because the only supply of SO ₄ is the tidal flow, which is the
343	same for all plots in each community of the experiment. Instead, higher rates of SO ₄ reduction are most
344	likely driven by some combination of electron donor supply and kinetics. While SO ₄ reducers likely
345	benefited from the increased availability of electron donors, as described above, the kinetics of SO_4
346	reduction also respond strongly to temperature (Weston and Joye, 2005).
347	When SO ₄ concentrations drop below a threshold concentration, SO ₄ reduction becomes SO ₄ -limited,
348	rather than electron donor-limited (Megonigal et al., 2004). A review of the coastal wetland CH ₄ literature
349	estimated this threshold at 4 mmol SO ₄ (Poffenbarger et al., 2011), a value that is consistent with patterns
350	of porewater CH_4 and SO_4 at the GCReW site (Keller et al. 2009). As SO_4 and O_2 are the dominant
351	electron-accepting compounds that suppress methanogenesis in this organic soil, this drawdown then
352	releases the methanogens from substrate competition (Fig. 7, mechanism 3). Here, we show that SO ₄ is
353	typically below 4 mmol in the $+5.1$ plots in the C ₃ community and in all plots in the C ₄ community (Fig.
354	6, Fig. S5). The drawdown of SO ₄ may also reduce rates of anaerobic CH ₄ oxidation (Hinrichs and
355	Boetius, 2003). Van Hulzen et al. (1999) proposed a multi-phase system in a warming incubation
356	experiment, observing that first methanogens are out-competed for substrates by other microbes, next
357	CH4 production increases as the supply of inhibiting TEA decreases, and finally TEA availability is
358	reduced to the point that methanogenesis is controlled only by the supply of electron donors. Warming in
359	this study decreased the time required for the system to pass through the first two phases (van Hulzen et
360	al., 1999). In our experiment, this final phase of increased methanogenic activity occurs when SO ₄
361	concentrations dip below 4 mmol SO ₄ L^{-1} , which occurs most often in the +5.1 °C plots, especially in the
362	C ₃ community. This interpretation is also supported by the long-term record of porewater chemistry from 18

an allied experiment at the site, demonstrating that porewater CH₄ concentrations increase as SO₄
concentrations decrease (Keller et al., 2009).

Methanogens may also have a competitive advantage over SO₄ reducers for electron donor consumption at warmer temperatures (van Hulzen et al., 1999). Sulfate reducers and methanogens have very similar K_M values for acetate, but the K_M for acetoclastic methanogenesis may decrease with temperature whereas K_M values for SO₄ reducers increase with temperature (van Bodegom and Stams, 1999). If this is the case in our system then warming would allow methanogens to use a greater proportion of the available substrates.

371 <u>4.5 Plant traits modify warming effects on CH₄ cycling</u>

372 The three biogeochemical mechanisms we propose to explain a warming-induced increase in CH₄

area emissions should interact strongly with plant responses to warming. Relationships between plant

374 functional groups and CH₄ emissions have been demonstrated through field studies in other wetland

375 ecosystems such as peatlands (Bubier et al., 1995; Ward et al., 2013) and in tidal wetland mesocosms (Liu

et al., 2019; Martin & Moseman-Valtierra, 2017; Mueller et al., 2020). We provide field evidence that

377 two species with distinct plant traits -- Schoenoplectus and Spartina -- have strikingly different effects on

378 CH₄ emissions from brackish wetlands. Spartina-dominated communities had consistently higher CH₄

379 emissions under both ambient and warmed conditions (Fig. 3). In most years, Schoenoplectus biomass

380 was negatively correlated with CH₄ emissions, while *Spartina/Distichlis* biomass was positively

381 correlated. Vegetation effects are typically strongest during the growing season, when the plants are

actively altering rhizosphere biogeochemistry (van der Nat & Middelburg, 1998; Ward et al., 2013),

383 which is consistent with our observations in this study.

As with warming effects, plant-driven shifts in CH₄ emissions are the result of differing rates of CH₄ production, oxidation, transport, or a combination of these processes, but sustained differences in emissions cannot be attributed only to transport, as discussed previously. Instead, the stimulation of CH₄ emissions is likely due to changes in the plant-mediated supply of electron acceptors and electron donors. In a field environment, differentiating between species-specific effects and underlying environmental conditions can be difficult, but mesocosm studies that control all environmental factors have also found species-specific effects on CH₄ cycling (e.g. Liu et al., 2014). Plants can alter CH₄ cycling by adding O₂ (electron acceptor) or C substrates (electron donors) to the rhizosphere, altering the redox state. We propose that *Schoenoplectus* is a net oxidizer of the rhizosphere and that *Spartina* is a net reducer, and thus their presence and productivity have opposing effects on CH₄ emissions (Fig. 7, mechanism 4).

394 4.5.1 Schoenoplectus oxidizes the rhizosphere, increasing CH₄ oxidation

395 Species vary in their capacity to support aerobic CH₄ oxidation (van der Nat & Middelburg, 1998) 396 and Schoenoplectus appears to support higher rates of aerobic CH₄ oxidation than Spartina (Mueller et 397 al., 2020). Scirpus lacustris is morphologically similar to Schoenoplectus americanus studied here and 398 has been demonstrated to have substantial rhizosphere oxidation capacity, especially during the growing 399 season (van der Nat & Middelburg, 1998). Consequently, these plants likely exert stronger control on 400 rates of CH₄ oxidation than rates of methanogenesis (van der Nat & Middelburg, 1998). We hypothesize 401 that the relatively high capacity of *Schoenoplectus* to transport O_2 held the CH₄ emissions stimulation 402 caused by modest levels of warming $(+1.7 \text{ to } +3.4 \text{ }^\circ\text{C})$ to rates similar to under ambient conditions (Fig. 403 3). At high warming (+5.1 °C), however, *Schoenoplectus* community CH₄ emissions drastically increase 404 (Fig. 3). We suggest that this is due to the combined effects of the three mechanisms discussed 405 previously, namely the differences in the Q10-values of CH4 production and CH4 oxidation, the increased 406 supply of organic substrate through plant productivity, and the decrease in competition for electron 407 donors due to SO₄ depletion. Collectively, when the ecosystem is warmed above current ambient 408 conditions by 5 °C or more, enhanced stimulation of CH₄ production starts to offset some of the 409 Schoenoplectus oxidation effect. This also offers an explanation for the positive correlation between 410 Schoenoplectus biomass and CH₄ emissions observed in 2016 as that was the hottest of the four years in 411 this study.

412 4.5.2 Spartina reduces the rhizosphere, increasing CH₄ production

413 The variability in quality and quantity of root exudates between plant functional types is well known 414 to affect microbial community composition and activity (Devn et al., 2008). Methanogenesis responses to 415 warming in incubation studies are related to the lignin and cellulose content of the peat, which in turn 416 depends on the plant functional type from which the peat developed (Duval and Radu, 2018). Although 417 warming is likely increasing substrate availability across the whole experiment, the production of labile, 418 low-molecular-weight C substrates through fermentation is less sensitive to temperature above 25 °C than 419 below this threshold (Neubauer & Craft, 2009; Weston & Joye, 2005). Microorganisms may also 420 preferentially use freshly produced (i.e. labile) organic carbon compounds as electron donors (DeLaune et al., 2014) and consequently warming effects on CH₄ production should be strongest in a system where 421 422 rates of root exudation and turnover are most rapid. We propose that root exudation and turnover explain 423 the positive correlation between plant biomass and CH_4 emissions that we observed in the C_4 community. 424 Multiple years of porewater chemistry at this site show that *Spartina*-dominated communities have higher 425 DOC and dissolved CH₄ than adjacent Schoenoplectus communities (Keller et al., 2009; Marsh et al., 426 2005). Though we did not directly measure root exudation, porewater DOC is partially derived from root 427 exudates and has been used as a proxy to understand the responses of root exudates to global change 428 factors (Dieleman et al., 2016; Fenner et al., 2007; Jones et al., 2009). 429 We observed a simultaneous increase in dissolved CH₄ at the soil surface and a decrease in dissolved 430 CH_4 at depth in the warmed C_4 plots. As with the observed trends in CH_4 emissions, there are multiple 431 mechanisms that could cause a shift in porewater CH₄ concentrations. Of the four mechanisms outlined 432 above, perhaps the simplest explanation is an increase in labile C at shallow depths and a decrease in 433 deeper soil. This is consistent with DOC depth profiles from this the C4 community in which porewater 434 DOC increases with warming in shallow samples but decreases with warming in deep samples (Fig. S7). 435 This shallowing of peak DOC concentrations could be due to a warming-induced increase in

evapotranspiration, leading to slower downward hydrologic transport of DOC-rich surface porewater to

21

437 lower depths, or a warming-induced shallowing of the root system, leading to a shift in the location of438 root exudates.

439 In most years, *Spartina* biomass was positively correlated with CH₄ emissions, supporting our 440 hypothesis that Spartina favors net CH₄ production. However, in 2016 Spartina biomass and CH₄ 441 emissions were negatively correlated. Prior work at this site has indicated that Spartina/Distichlis biomass 442 is more negatively affected by hot and dry growing conditions than *Schoenoplectus* (Novce et al., 2019) 443 due in part because the Spartina/Distichlis (C4) communities are less frequently inundated. The 2016 444 growing season was substantially warmer than average (Table 2) and the heating treatments were 445 initialized on 1 Jun of that year, after the annual plants had already established and may have developed adaptations to ambient, rather than elevated, temperature conditions. The combination of these two effects 446 447 likely led to heat stress, reducing the root exudates supplied to the rhizosphere microbial community 448 (Heckathorn et al., 2013) and thus minimizing the Spartina stimulation effect.

449 <u>4.6 Comparisons with prior data</u>

450 Methane emissions have been measured at the GCReW site previously, but this study represents the most 451 comprehensive dataset collected to date, and is thus particularly useful for advancing the process-based 452 understanding needed to improve prognostic models. Overall, our flux estimates are lower than those 453 reported previously. The earliest CH₄ fluxes were measured in a single month (July) in Schoenoplectusdominated plots and reported to be 331 to 6883 μ mol m⁻² d⁻¹ (Dacey et al., 1994), much higher than our 454 range of 359 to 1651 µmol m⁻² d⁻¹ for ambient temperature *Schoenoplectus* plots in July. Similarly, Marsh 455 et al. (2005) reported mean growing season (May-Oct) CH₄ emissions from this site of $846 \pm 111 \,\mu$ mol 456 CH₄ m⁻² d⁻¹, whereas we measured $656 \pm 79 \mu$ mol CH₄ m⁻² d⁻¹ over the same months. Finally, Pastore et 457 458 al. (2017) estimated average annual fluxes in their Schoenoplectus-dominated ambient CO₂ plots as $3.1 \pm$ 1.7 g CH₄ m⁻² yr⁻¹, compared to our estimates of 1.6 ± 0.3 g CH₄ m⁻² yr⁻¹ for *Schoenoplectus* plots. The 459 460 different estimates by these studies may be partly due to interannual variability as demonstrated in our 461 data where 2018 had substantially higher fluxes than any of the surrounding years (Table S2).

462 The annual estimates reported here for ambient temperature plots trended lower than published 463 mean CH₄ emissions for mesohaline tidal marshes. Our plots ranged from 0.7 to 9.3 g CH₄ m⁻² yr⁻¹ (mean = 9.3), compared to the range of 3.3 to 16.4 g CH₄ m⁻² yr⁻¹ (mean = 16.4) reported by Poffenbarger et al. 464 465 (2011). This difference may be explained by the fact that there was significant within-class variation in 466 the oligohaline and mesohaline salinity classes that was unexplained and their assessment was based on 467 too few data points to fully capture the variation that is expected to exist in the mesohaline class. Indeed, 468 subsequent studies have documented fluxes well below 3 g CH₄ m⁻² yr⁻¹ (Krauss and Whitbeck, 2012), 469 and even negative fluxes (Al-Haj and Fulweiler, 2020). We hypothesize that the low fluxes measured at 470 our site reflect Schoenoplectus americanus traits that favor CH₄ oxidation more than CH₄ production, and 471 that the high end of our range was limited by the high soil elevation (i.e. deep water table) of areas 472 dominated by Spartina patens, off-setting the influence of S. patens traits that favor CH₄ production. 473 4.7 Implications for tidal wetland carbon cycling 474 Warming accelerates rates of CH₄ emissions from brackish marshes, especially during the growing 475 season. This is driven by both direct and indirect warming effects and mediated by soil biogeochemistry, 476 but the magnitude of the warming effect is also dependent on traits of the plant species that dominate the 477 plant community. Communities dominated by Spartina patens increase net CH₄ emissions in response to 478 smaller increments of warming than communities dominated by Schoenoplectus americanus. Spartina-479 dominated sites may thus have a higher likelihood of shifting from a net C sink to a net C source under 480 future warming conditions, due to this increased loss of C as CH₄. However, this effect could be mitigated 481 if these high-elevation Spartina marshes become dominated by Schoenoplectus in response to predicted 482 accelerated sea-level rise (Kirwan and Guntenspergen, 2012). In addition, Spartina traits are plastic and 483 influenced by factors such as soil redox conditions (Kludze and DeLaune, 1994), salinity (Crozier and

- 484 DeLaune, 1996), and water level (Liu et al., 2019), all of which can be expected to change plant-mediated
- 485 effects on CH₄ biogeochemistry. Further studies are needed to thoroughly assess the range of

486	environmental conditions under which Spartina is a net reducer and Schoenoplectus is a net oxidizer as
487	proposed by the present study.
488	
489	Data availability
490	All data is available from the corresponding author upon request.
491	
492	Author contribution
493	GLN and JPM designed the study, GLN collected and analyzed the data, and GLN and JPM wrote the
494	paper.
495	
496	Competing interests
497	The authors declare that they have no conflict of interest.
498	
499	Acknowledgements
500	This manuscript is based upon work supported by the U.S. Department of Energy, Office of Science,
501	Office of Biological and Environmental Research Program (DE-SC0014413 and DE-SC0019110), the
502	National Science Foundation Long-Term Research in Environmental Biology Program (DEB-0950080,
503	DEB-1457100, and DEB-1557009), and the Smithsonian Institution. Roy Rich designed the warming
504	infrastructure and maintains it with the assistance of Gary Peresta. We also thank technicians in the SERC
505	Biogeochemistry Lab for assistance with porewater collection and analysis.
506	
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