

# High-resolution induced polarization imaging of biogeochemical carbon-turnover hot spots in a peatland

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## Abstract

Biogeochemical hot spots are defined as areas where biogeochemical processes occur with anomalously high reaction rates relative to their surroundings. Due to their importance in carbon and nutrient cycling, the characterization of hot spots is critical to ~~accurately predict~~predicting carbon budgets accurately in the context of climate change. However, biogeochemical hot spots are difficult to identify in the environment, as ~~sampling resolutions are often too coarse to find these areas in the subsurface. methods for in-situ measurements often directly affect the sensitive redox-chemical conditions.~~ Here, we present imaging results of a geophysical survey using the non-invasive induced polarization (IP) method to identify biogeochemical hot spots of carbon turnover in a minerotrophic wetland. To interpret the field-scale IP signatures, geochemical analyses were performed on freeze-core samples obtained in areas characterized by anomalously high and low IP responses. Our results reveal large variations in the electrical response, with the highest IP phase values ( $> 2018$  mrad) corresponding with high concentrations of phosphates ( $> 4000 \mu\text{M}$ ), an indicator of carbon turnover. Furthermore, we found a strong relationship between the electrical properties resolved in IP images and the dissolved organic carbon. Moreover, analysis of the freeze core ~~reveal~~reveals negligible concentrations of iron sulfides. The extensive geochemical and geophysical data presented in our study demonstrates that IP images can ~~asses~~track small scale changes in the biogeochemical activity in peat, and can be used to identify hot spots.

**Keywords:** biogeochemical carbon turnover; geophysical imaging methods; electrical conductivity; induced polarization; microbiologically active zones

## 1 Introduction

In terrestrial and aquatic ecosystems ~~'patches',~~ patches or areas that show disproportionately high biogeochemical reaction rates relative to the surrounding matrix are referred to as biogeochemical ~~'hot-spots'~~ 'hot spots' (McClain et al., 2003). Hot spots for turnover of redox-sensitive species (e.g., oxygen, nitrate or dissolved organic carbon) are often generated at interfaces between oxic and ~~annoxie~~ anoxic environments, where the local presence/absence of oxygen either ~~favours~~ favours or suppresses biogeochemical reactions such as aerobic respiration, denitrification, or oxidation/reduction of iron (McClain et al., 2003). Biogeochemical hot spots are important for nutrient and carbon cycling in various systems such as wetlands (Frei et al., 2010; 2012), lake sediments (Urban, 1994), the vadose zone (Hansen et al., 2014), hyporheic areas (Boano et al., 2014) or aquifers (Gu et al., 1998). Wetlands are distinct elements in the landscape, which are often located where ~~various~~ hydrological flow paths converge, such as at the bottoms of basin shaped catchments, local depressions or around rivers and streams (Cirmo and McDonnell 1997). Wetlands are attracting increasing interest because of their important contribution to water supply, water quality, nutrient cycling, and biodiversity (Costanza et al., 1997; 2017). Understanding microbial moderated cycling of nutrients and carbon in wetlands is critical, as these systems store a significant part of the global carbon through the accumulation of decomposed plant material (Kayranli et al., 2010). In wetlands, water table fluctuations as well as plant roots determine the vertical and horizontal distribution of oxic and anoxic areas (Frei et al., 2012; Gutknecht et al., 2006). Small scale subsurface flow processes in wetlands, moderated by micro-topographical structures (hollow and hummocks) (Diamond et al., 2020), can control the spatial presence of redox-sensitive solutes and formation of biogeochemical hot spots (Frei et al., 2010; 2012). Despite their relevance for the carbon and nutrient ~~cycles, the~~ field observations of the cycling, basic mechanisms controlling the formation and distribution of biogeochemical hot spots in space ~~is largely missing~~ are not well understood.

Biogeochemical active areas traditionally have been identified and localized through chemical analyses of point samples from the subsurface and ~~then~~ subsequent interpolation of the data in space (Morse et al., 2014; Capps and Flecker, 2013; Hartley and Schlesinger, 2000). However, such point-based sampling methods may either miss hot spots due to the low spatial resolution of sampling (McClain et al., 2003) or disturb the redox-sensitive conditions in the subsurface by bringing oxygen into anoxic areas during sampling. Non-invasive methods, such as geophysical techniques, have the potential to study subsurface biogeochemical ~~hot spots~~ activity

in-situ without interfering with the subsurface environment (e.g., Williams et al., 2005; 2009; Atekwana and Slater 2009; Flores Orozco et al., 2015; 2019; 2020). Geophysical methods permit to map large areas in 3D and still resolve subsurface physical properties with a high spatial resolution (Binley et al., 2015).

In particular, the induced polarization (IP) technique has recently emerged as a useful tool to delineate biogeochemical processes in the subsurface (e.g., Kemna et al., 2012; Kessouri et al., 2019; Flores Orozco et al., 2020). The IP method provides information about the electrical conductivity and the capacitive properties of the ground, which can be expressed, respectively, in terms of the real and imaginary components of the complex resistivity (Binley and Kemna, 2005). The method is commonly used to explore metallic ores because of the strong polarization response associated to metallic minerals (e.g., Marshall and Madden, 1959; Seigel et al., 2007). Pelton et al. (1978) and Wong (1979) proposed the first models linking the IP response to the size and content of metallic minerals. More recently, the role of chemical and textural properties in the polarization of metallic minerals has been investigated in detail (~~Revil et al., 2012, 2015a; 2015b, Bueker et al., 2018; 2019~~), based on further developments of Wong's model of a perfect conductor and reaction currents (Bueker et al., 2018; 2019); while Revil et al. (2012, 2015a, 2015b, 2017b, 2017c, 2018) have presented a new mechanistic model that takes into account the intragrain polarization and does not involve reactions currents. In porous media without a significant metallic content, the IP response can be related to the polarization of the electrical double layer formed at the grain-fluid interface (e.g., Waxman and Smits, 1968; Revil and Florsch, 2010; Revil, 2012). ~~In particular~~For instance, Revil et al. (~~2017~~2017a) carried out ~~complex conductivity~~IP measurements on a large set of soil samples, for which they report a linear ~~relation~~relationship between the magnitude of the polarization response and the cation exchange capacity (CEC), which ~~in turn~~ is related to surface area and surface charge density.

Since the early 2010s, various studies have explored the potential of IP measurement for the investigation of biogeochemical processes in the emerging field of biogeophysics (Slater and Atekwana, 2013). Laboratory studies on sediment samples examined the correlation between the spectral induced polarization (SIP) response and ~~X-iron sulfide~~sulfide precipitation caused by iron reducing bacteria (Williams et al., 2005; Ntarlagiannis et al., 2005; 2010; Slater et al., 2007; Personna et al., 2008; Zhang et al., 2010; Placencia Gomez et al., 2013; Abdel Aal et al., 2014). Further investigations in the laboratory have also revealed an increase in the polarization response accompanying the accumulation of microbial cells and biofilms (Davis et al., 2006; Abdel Aal et al., 2010a, 2010b; Albrecht et al., 2011; Revil et al., 2012; Zhang et al., 2013; Mellage et al., 2018; Rosier et al., 2019; Kessouri et al., 2019).

Motivated by these ~~successful laboratory measurements (i.e., Abdel Aal and Atekwana, 2014),~~observations, the IP method has also been used to characterize biogeochemical degradation of contaminants at the field scale (Williams et al., 2009; ~~Slater and Binley 2006;~~ Flores Orozco et al., 2011; 2012b; 2013; 2015; Maurya et al., 2017). ~~Natural accumulations~~Additionally, Wainwright et al. (2016) demonstrated the applicability of the IP imaging method to identify naturally reduced zones, i.e., hot spots, at the floodplain scale. These authors show that the accumulation of organic matter in areas with indigenous iron-reducing bacteria ~~might result~~results in naturally reduced zones and the accumulation of iron sulfide minerals, which are classical IP targets. In line with this argumentation, Abdel Aal and Atekwana (2014) argued that the biogeochemical precipitation of iron sulfide minerals. Wainwright et al. (2016) demonstrated the applicability of IP imaging to identify naturally reduced zones, i.e., hot spots~~sulfides control the high electrical conductivity and IP response observed in floodplains. Moreover~~hydrocarbon-impacted sites. Nonetheless, in a recent study, Flores Orozco et al. (2020) demonstrated the possibility to delineate biogeochemically active zones in a municipal solid waste landfill even in the absence of iron sulfides. Flores Orozco et al. (2020) argued that the high content of organic matter itself might explain both, the high polarization response and high rates of microbial activity; thus, opening the possibility to delineate biogeochemical hot spots that are not related to iron-reducing bacteria. This conclusion is consistent with previous studies performed in marsh and peat soils, areas with a high organic matter content and high microbial turnover rates (Mansoor and Slater, 2007; McAnallen et al., 2018). Peat soils are characterized by a high surface charge and have been suggested to enhance the IP response (Slater and Reeve, 2002). Mansoor and Slater (2007) concluded that the IP method is a useful tool to map iron cycling and microbial activity in marsh soils. Garcia-Artigas et al. (2020) demonstrated that bioclogging by bacteria increases the IP response accompanying wetlands treatment. Uhlemann et al. (2016) found differences in the electrical resistivity of peat according to saturation, microbial activity, and pore water conductivity; however, their study was limited to direct-current resistivity and did not investigate variations in the IP response. In contrast to these observations, laboratory studies have shown a low polarization response in samples with a high organic matter content, despite its high CEC (Schwartz and Furman, 2014). Based on field measurements, McAnallen et al. (2018) found that active peat is less polarizable due to variations in groundwater chemistry imposed by sphagnum mosses; while degrading peat resulted in low resistivity values and a high polarization response. Based on measurements with the Fourier Transform Infrared (FTIR) Spectroscopy in water samples, the authors concluded that the carbon-oxygen (C=O) double-

bound in degrading peat correlated with the polarization magnitude of the peat material. Based on laboratory investigations, Ponziani et al. (2011) also conclude that decomposition of peat occurs predominantly by aerobic respiration, i.e. using molecular oxygen as the terminal electron acceptor to oxidize organic matter. Thus decomposition rates are expected to be highest at the interface between the oxic and anoxic zones.

Based on these promising previous results, we ~~believe~~hypothesize that the IP method is a potentially useful tool for in-situ investigation of biogeochemical processes ~~and the mapping of biogeochemical hot spots~~. However, different responses observed in lab and field investigations do not offer a clear interpretation scheme of general validity. ~~This~~Additionally, it is not clear whether the IP method is only suited to characterize biogeochemical hot spots associated to iron-reducing bacteria, which favor the accumulation of iron sulfides. Hence, in this study ~~presents~~we present an extensive IP imaging dataset collected at a peatland site to investigate the controls on the IP response in biogeochemically active areas. IP monitoring results are compared to geochemical data obtained from the analysis of freeze cores and pore water ~~sample analysis~~samples. Our main objectives are (i) to assess the applicability of the IP method to spatially delimit highly active biogeochemical areas in the peat soil and (ii) to investigate whether the local IP response is related to the accumulation of iron sulfides or high organic matter turnover.

## 2 Material and methods

### 2.1 Study site

The study site is part of the Lehstenbach catchment located in the Fichtelgebirge mountains (Fig. 1a), a low mountain range in north-eastern Bavaria (Germany) close to the border to the Czech Republic. Various soil types including Dystric Cambisols, Haplic Podisols, and Histosols (i.e. peat soil) cover the catchment area of approximately 4.2 km<sup>2</sup>, situated on-top of variscan granite bedrock (Strohmeier et al., 2013). The catchment is bowl shaped (Fig. 1b), and minerotrophic riparian peatlands have developed around the major streams. The plot scale study site (Fig. 1c) is located in a riparian peatland draining into a nearby stream close to the ~~catchment's~~catchment's outlet (Fig. 1b).

The groundwater level in this area annually varies within the top 30 cm of the peat soil ~~and was 5-10 cm below the surface during the measurement campaign. The, and the local groundwater flow has a S-SW orientation (Durejka et al., 2019) towards a nearby drainage ditch. Permanently high water saturation of the peat soil favors the development of anoxic~~

biogeochemical processes close to the surface. Frei et al. (2012) demonstrated that hot spots, at the study area are related to the stimulation of iron reducing bacteria and accumulation of iron sulfides, which are generated by small scale subsurface flow processes and the spatial non uniform availability of electron acceptors and donors induced by the ~~peatland's~~ typical micro topography. ~~Transport limited and spatially non of the peatland. Non~~ uniform availability of electron acceptors and donors, ~~which to a large extent is controlled by subsurface flow patterns induced by the wetland's micro topography (hummocks and hollows),~~ in combination with labile carbon stocks are the primary drivers in generating biogeochemical hot spots in the peatland (Frei et al., 2012; Mishra and Riley, 2015).

## 2.2 Experimental plot and geochemical measurements

The experimental plot for the geophysical measurements covers approximately 160 m<sup>2</sup> (12.6 x 12.6 m, Fig. 1c) of the riparian peatland. Sphagnum Sp. (peat moss) and Molinia caerulea (purple moor-grass) dominate the vegetation, with the sphagnum and purple moor-grass abundance being higher in the Northern part of the plot (Fig. 2a and 2b). In the Southeastern region, where the sphagnum is less abundant, permanent surface runoff was observed (Fig. 2d). Peat thickness was measured with a 1 m resolution in E-W direction and 0.5 m resolution in N-S direction (along the IP profiles described below). To measure the thickness of the peat, a stainless-steel rod (0.5 cm in diameter) was pushed into the ground until it reached the granitic bedrock (similar to Parry et al., 2014). ~~Peat thickness was measured with a 1 m resolution in E-W direction and 0.5 m resolution in N-S direction. During the experiments, the~~ The local groundwater level was measured in two piezometers and was found at ~5-8 cm below the surface during the IP survey. Groundwater samples were collected at three different locations (S1, S2, and S3 indicated in Fig. 3) using a bailer. Pore water profiles were taken at S1, S2, and S3 at 5 cm intervals to a maximum depth of 50 cm below ground surface (bgs) using stainless steel mini-piezometers. All water samples were filtered through 0.45 µm filters and analyzed for fluoride, chloride, nitrite, bromide, nitrate, phosphate, and sulfate using an ion chromatograph (Compact IC plus 882, Metrohm GMBH). Dissolved organic carbon (DOC) was measured using a Shimadzu TOC analyzer via thermal combustion. Dissolved iron species (Fe<sup>2+</sup>) and total iron (Fe<sup>tot</sup>) concentrations were measured photometrically using the 1,10-phenanthroline method on pore water samples that had been stabilized with 1% vol/vol 1M HCl in the field (Tamura et al., 1974). Two freeze cores (see Fig. 2d) were extracted at locations S1 and S2 (Fig. 3) by pushing an 80-cm long stainless-steel tube into the peat. After the tube was installed, it was filled with a mixture of dry ice and ethanol. After around 20 minutes, the pipe with the frozen peat sample was extracted and stored on dry ice for transportation to the



laboratory at the University of Bayreuth. Both freeze cores were cut into 10 cm segments. Each segment was analyzed for reactive iron (1M HCl extraction and measured for  $\text{Fe}^{\text{tot}}$  as described above) (Canfield, 1989), reduced sulfur species using the total reduced inorganic sulfur (TRIS) method (Canfield et al., 1986) and carbon and nitrogen concentrations after combustion using a thermal conductivity detector. Peat samples were also analyzed by FTIR using a Vector 22 FTIR spectrometer (Bruker, Germany) in absorption mode; with subsequent baseline subtraction on KBr pellets (200 mg dried KBr and 2 mg sample). Thirty-two measurements were recorded per sample and averaged from 4.500 to 600  $\text{cm}^{-1}$  in a similar manner to Biester et al. (2014).

### 2.3 Non-invasive techniques: induced polarization measurements

The induced polarization (IP) imaging method, also known as complex conductivity imaging or electrical impedance tomography, is an extension of the electrical resistivity tomography (ERT) method (e.g., Kemna et al., 2012). As such, it is based on four-electrode measurements, where one pair of electrodes is used to inject a current (current dipole) and a second pair of electrodes is used to measure the resulting electrical potential (potential dipole). Modern devices can measure tens of potential dipoles simultaneously for a given current dipole, permitting the collection of dense data sets within a reasonable measuring time. This provides an imaging framework to gain information about lateral and vertical changes in the electrical properties of the subsurface. IP data can be collected in the frequency domain (FD), where an alternating current is injected into the ground where the polarization of the ground leads to a measurable phase shift between the injected periodic current and the measured voltage signals. ~~FD measurements can be conducted at different frequencies to gain information about the frequency dependence of the electrical properties of the subsurface. This approach is known as spectral IP (SIP).~~ From the ratio of the magnitudes of the measured voltage and the injected current, as well as the phase shift between the two signals, we can obtain the electrical transfer impedance. The inversion of imaging data sets, i.e. a large set of such four-point transfer-impedance measurements collected at different locations and with different spacing between electrodes along a profile, permits to solve for the spatial distribution of the electrical properties in the subsurface (see deGroot-Hedlin and Constable, 1990; Kemna et al., 2000; Binley and Kemna, 2005).

IP ~~imaging~~inversion results can be expressed in terms of the complex conductivity ( $\sigma^*$ ) or its inverse the complex resistivity ( $\sigma^*=1/\rho^*$ ). The complex conductivity can either be denoted in terms of its real ( $\sigma'$ ) and imaginary ( $\sigma''$ ) components, or in terms of its magnitude ( $|\sigma|$ ) and phase ( $\varphi$ ):  $\sigma^*=\sigma'+i\sigma''=|\sigma|e^{i\varphi}$ , (1)

where  $i = \sqrt{-1}$  is the imaginary unit,  $|\sigma| = \sqrt{\sigma'^2 + \sigma''^2}$  and  $\phi = \tan^{-1} \frac{\sigma''}{\sigma'}$ .

$\phi = \tan^{-1}(\sigma''/\sigma')$ . The real part of the complex conductivity is mainly related to the Ohmic conduction, while the imaginary part is mainly related to the polarization of the subsurface. For The conductivity ( $\sigma'$ ) is related to porosity, saturation, the conductivity of the fluid filling the pores and a more contribution of the surface conductivity (Lesmes and Frye, 2001). The polarization ( $\sigma''$ ) is only related to the surface conductivity taking place at the electrical double layer (EDL) at the grain-fluid interface. For a detailed description of the IP method, the reader is referred to the work of Ward (1988), Binley, and Kemna (2005), and Binley and Slater (2020).

The strongest polarization response is observed in the presence of electrically conducting minerals (e.g., iron) (e.g., Pelton et al., 1978) in the so-called electrode polarization (Wong et al., 1979). It arises from the different charge transport mechanisms in the electrical conductor (electronic or semiconductor conductivity) and the electrolytic conductivity of the surrounding pore fluid, which make the solid-liquid interface polarizable. Diffusion-controlled charging and relaxation processes inside the grain (e.g., Revil et al., 2018; 2019; Abdulsamad et al., 2020) or outside the grain in the electrolyte (e.g., Wong, 1979; Bückner et al., 2019) are considered as possible causes of the polarization response at low frequencies irrespective of the specific modeling approach. All mechanistic models predict an increase in the polarization response with increasing volume content of the conductive minerals (Wong, 1979; Revil et al. 2015a, 2015b, 2017a, 2017b, 2018; Qi et al., 2018; Bückner et al., 2018).

In the absence of electrical conductors, the polarization response is only related to the accumulation and polarization of ions in the EDL. Different models have been proposed to describe the polarization response as a function of grain size, surface area and surface charge (e.g., Schwarz, 1962; Schurr, 1964; Leroy et al., 2008). Alternatively, the membrane polarization related the IP response to variations in the geometry of the pores as well as the concentration and mobility of the ions (e.g., Marshall and Madden, 1959; Bückner and Hördt, 2013; Bückner et al., 2019). Regardless of the specific modeling approach, EDL polarization mechanisms strongly depend on the specific surface area of the material and the charge density at the surface (Revil, 2012; Waxman and Smits, 1968).

In this study, we conducted FDIP measurements at 1 Hz along 65 lines during a period of four days in July 2019. We used the DAS-1 instrument manufactured by Multi-Phase Technologies (now MTP-IRIS Inc.). We collected 64 N-S oriented lines (referred to as By 1 to By 64) with 20 cm spacing between each line. One additional line (By 68) was collected with a W-E



orientation, which intersects the N-S oriented lines at 3 m, as presented in Fig. 3. Each profile consisted of 64 stainless steel electrodes (3 mm diameter) with a separation of 20 cm between each electrode (Fig. 2c). Besides the short electrode spacing, the use of a dipole-dipole configuration with a unit dipole length of 20 cm warranted a high resolution within the upper 50 cm of the peat, where the biogeochemical hot spots were expected. We deployed a dipole-dipole skip-0 (i.e., the dipole length for each measurement is equal to the unit spacing of 20 cm) configuration; voltage measurements were collected across eight adjacent potential dipoles for each current dipole. The dipole-dipole configuration avoids the use of electrodes for potential measurements previously used for current injections to avoid contamination of the data caused by remnant polarization of electrodes. To evaluate data quality, reciprocal readings were collected along one profile every day (see, e.g., LaBrecque et al., 1996; Flores Orozco et al., 2012a; 2019). Reciprocal readings refer to data collected after interchanging current and potential dipoles. We used coaxial cables to connect the electrodes with the measuring device to minimize the distortion of the data due to electromagnetic coupling and cross-talking between the cables (e.g., Zimmermann et al., 2008, 2019; Flores Orozco et al., 2013), with the shields of all coaxial cable running together into one ground electrode (for further details see Flores Orozco et al., 2020, 2021).

## 3 Results

### 3.1 Data quality and processing

In Fig. 4, we present a modified pseudosection showing both normal (negative pseudodepth) and reciprocal (positive pseudodepth) readings in terms of apparent resistivity ( $\rho_a$ ) and apparent phase ( $\phi_a$ ) for the data collected along line By 25. Plots in Fig. 4 show consistency between the normal and reciprocal readings of apparent resistivity (4a) and phase (4b). The principle of reciprocity asserts that normal and reciprocal readings should be the same (e.g., Slater et al., 2000). Hence, we use here the misfit analysis of the discrepancy between normal and reciprocal readings to detect outliers, and to quantify data error (LaBrecque et al., 1996; Flores Orozco et al., 2012a; Slater and Binley, 2006; Slater et al., 2000). Figures 4c and 4d show the histograms of the normal-reciprocal misfits along line By 25 for both the resistance and phase ( $\Delta R$  and  $\Delta \phi$  respectively), which exhibit near-Gaussian distributions with low standard deviations (as expected for random noise) for both the normalized resistance ( $S_R=0.027$ ) and the apparent phase ( $S_\phi=1.1$  mrad). Readings exceeding these standard deviation values were considered as outliers (between 16 and 33% of the data at

~~the different lines) and were removed from the data set prior to the inversion. In this study, we~~  
~~quantified the error parameter for each line collected as normal and reciprocal pairs (using the~~  
~~approach outlined by Flores Orozco et al., 2012a) and computed the average value of the error~~  
~~parameters for the different lines to define the error model used for the inversion of all imaging~~  
~~data sets.~~

For the inversion of the IP imaging data set, we used CRTomo, a smoothness-constrained least-squares algorithm by Kemna (2000) that allows inversion of the data to a level of confidence specified by an error model. We used the resistance and phase error models described by Kemna (2000) and Flores Orozco et al. (2012a). The resistance ( $R$ ) error model is expressed as  $s(R)=a+bR$ , where  $a$  is the absolute error, which dominates at small resistances (i.e.,  $R < 0$ ), and  $b$  is the relative error, which dominates at high resistance values (LaBrecque et al., 1996; Slater et al., 2000). For the phase, the error model is also expressed as a function of the resistance  $s(\phi_a)=cR^d$ , where  $d<0$  in our study due to the ~~relativerelatively~~ low range in the measured resistances (see Flores Orozco et al., 2012a for further details). If  $d\rightarrow 0$ , the model reduces to the constant-phase-error model (Flores Orozco et al., 2012a) with  $s(\phi)=c$  described by Kemna (2000) and Slater and Binley (2006).

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outliers (between 16 and 33% of the data at the different lines) and were removed from the data set prior to the inversion of the entire data set.

Here, we present inversion results obtained using the error parameters,  $a = 0.001 \Omega$ ,  $b = 2.2\%$ ,  $c = 1 \text{ mrad}$ . For the imaging, we defined a cut-off value of the cumulated sensitivity of  $10^{2.75}$ , with pixels related to a lower cumulated sensitivity blanked in the images. The cumulated sensitivity values are a widely used parameter to assess the depth of investigation (Kemna et al., 2002; Flores Orozco et al., 2013).

### 3.2 Complex conductivity imaging results and their link to the peat thickness and land cover

The thickness of the peat in the plot was found to vary between 40 and 160 cm (Fig. 5). The thickness of the peat unit increased in the W-E direction, with much smaller variations in the N-S direction. Variations in the vegetation cover (as indicated by the three vegetation classes, abundant (*av*), moderate (*mv*), and sparse (*sv*)) do not seem to correspond with the variations in the peat thickness. Note that the N-S orientation of the majority of IP lines is approximately aligned with the direction of minimum changes in the peat thickness.

Figure 6 shows the imaging results of the N-S oriented profiles By 25, By 46, and the W-E oriented profile By 68 expressed in terms of the real ( $\sigma'$ ) and imaginary polarization ( $\sigma''$ ) part of the complex conductivity. These images reveal three main electrical units: (i) a shallow peat unit with high  $\sigma'$  ( $>5 \text{ mSm}^{-1}$ ) and high  $\sigma''$  ( $>80 \mu\text{Sm}^{-1}$ ) values in the top 10-20 cm bgs, (ii) an intermediate unit in the peat with moderate to low  $\sigma'$  ( $<5 \text{ mSm}^{-1}$ ) and moderate  $\sigma''$  ( $40-80 \mu\text{Sm}^{-1}$ ) values, and (iii) underneath it, a deep third unit characterized by moderate to low  $\sigma'$  ( $<5 \text{ mSm}^{-1}$ ) values and the lowest  $\sigma''$  ( $<40 \mu\text{Sm}^{-1}$ ) values. The low complex conductivity (both  $\sigma'$  and  $\sigma''$ ) in the third unit corresponds to corresponding to the granite bedrock. The compact structure of the granite, corresponding to low porosity, explains the observed low conductivity values ( $\sigma' < 5 \text{ mSm}^{-1}$ ) due to low surface charge and surface area. The shallow and intermediate electrical units are related to the relatively heterogeneous peat (Fig. 6), which is beyond the vertical change and lateral heterogeneities in the complex conductivity parameters. As shown in the plots of  $\sigma''$  in Fig. 6, the contact between the second and third unit boundary roughly corresponds to the contact between peat and granite measured using with the metal rod (represented by the dashed line in Fig. 5). This shows that indicates the electrical ability of IP imaging can accurately to resolve the contact with the bedrock even in areas where it is located geometry of the peat unit.

However, for the survey design used in this study,  $\sigma''$  images are not sensitive to materials deeper than  $\sim 1.25$  m bgs. Images of the electrical conductivity reveal much more considerable variability and a lack of clear contrasts between the peat and the granite materials, likely due to the weathering of the shallow granite unit (Lischeid et al., 2002, Partington et al., 2013).

~~The shallow and intermediate electrical units are related to the relatively heterogeneous peat (Fig. 6). In the first unit, the electrical images reveal lateral changes along the profiles oriented N-S (By 25 and By 46), while we generally observe higher polarization values ( $\sigma''$ ) in the northern part of the profiles than in the southern part. This lateral change corresponds to the W-E oriented profile By 68, where the polarization values are the highest around sampling point S1.~~

The phase of the complex conductivity ~~roughly~~ represents the ratio of the polarization ( $\sigma''$ ) relative to the Ohmic conduction ( $\phi = \sigma''/\sigma'$ ). ~~Therefore~~ Thus, it ~~has been argued that the phase represents the polarization effect better than the imaginary conductivity itself as it removes effects stemming from changes in fluid conductivity, saturation, and porosity~~ can also be used to represent the polarization response (Kemna et al., 2004; Ulrich and Slater 2004); Flores Orozco et al., 2020). Similar to the  $\sigma''$  images, the phase images presented in Fig. 7 resolve the three main units: (i) the shallow peat unit ~~in~~ within the top 10-50 cm is characterized by the highest values ( $\phi > 2218$  mrad), (ii) the intermediate unit still corresponding to peat, is characterized by moderate  $\phi$  values (between 13 and ~~2218~~ mrad), and (3) the ~~deep~~ third unit, associated to the granitic bedrock, related to the lowest  $\phi$  values ( $< 13$  mrad). The polarization images expressed in terms of  $\phi$  ~~solve the thickness of the peat with~~ show a higher resolution contrast between the peat and the granite units than the  ~~$\sigma'\sigma'$  (or  $\sigma''$ )~~ images. The histograms presented in Fig. 7 show the distribution of the phase values in the images, with a different color for model parameters extracted above and below the contact between peat and granite. The histograms highlight that the lowest phase values clearly correspond to the granite bedrock ( $< 13$  mrad), while higher phase values are characteristic of the peat unit.

Moreover, the shallow unit shows more pronounced lateral variations in the phase than in  $\sigma''$ , and patterns within the peat unit are more clearly defined: ~~Along~~ As observed in Fig. 6, along line By 25, the thickness of the first unit decreases from approx. 0.5 m at 2 m along the profile to 0 m around 10 m at the end of the profile. Along line By 46, the first unit is slightly thicker than 50 cm and shows the highest phase values ( $\sim 25$  mrad) between 0 and 6.5 m along the profile. Beyond 6.5 m, the polarizable unit becomes discontinuous with isolated polarizable ( $\sim 18$  mrad) zones, ~~which also extend~~ extending to a depth of 50 cm. The geometry of the

shallow, polarizable-~~shallow~~ unit is consistent with the corresponding results along line By 68, which crosses By 25 and By 46 at 3 m along these lines (S1 and S3 are located at these intersections). In particular, ~~it is clear that~~ the highest phase values ~~were~~are consistently found in the shallowest 50 cm in the peat unit, at the depth where ~~the presence of~~ biogeochemical hot spots ~~has~~have been reported in ~~previous studies (e.g., the study by~~ Frei et al., (2012).

~~The histograms presented in the right column of Fig. 7 show the distribution of the phase values in the images, with a different color for model parameters extracted above and below the contact between the peat and the granite. The histograms highlight that the lowest phase values clearly correspond to the granite bedrock ( $< 15$  mrad), while higher phase values are characteristic for the peat unit.~~

~~Fig.~~Figure 8 presents maps of the electrical parameters at different depths aiming ~~at~~to identify lateral changes in the ~~identification of a possible link to the vegetation cover: abundant (av), moderate (mv) and sparse vegetation (sv), observed at the field (Fig. 5).~~hot spots across the entire experimental plot. Such maps present the interpolation of values inverted in each profile. Along each profile, a value is obtained through the average of model parameters (conductivity magnitude and phase) within the surface and a depth of 20 cm (shallow maps) and between 100 and 120 cm (for deep maps). The western part of the experimental plot (between 0 and 4 m in X-direction and between 2 and 9 m in Y-direction) corresponds to a shallow depth to the bedrock (a peat thickness of  $\sim 50$ -70 cm) and is associated with ~~a~~high electrical response parameters in the shallow maps ( $\phi > 17$ 18 mrad,  $\sigma' > 7$  mSm $^{-1}$  and  $\sigma'' \geq 100$   $\mu$ Sm $^{-1}$ ). ~~In~~), which we can interpret here as the geometry of the biogeochemical hot spots. Another hot spot can be identified in the Northern part of the experimental plot, in the area with abundant vegetation; we observe a higher polarization response for the top 20 cm ( $\phi > 20$ 18 mrad and  $\sigma'' \geq 80$   $\mu$ Sm $^{-1}$ ) than, for instance, the one corresponding to the moderate vegetation located at the southern part. In contrast, the lowest polarization values ( $\phi < 15$  mrad, and  $\sigma'' < 80$   $\mu$ Sm $^{-1}$ ) values, which we interpret as biogeochemical inactive zones, are related to the area with sparse vegetation. ~~Thus, maps in Fig. 8 suggest that the biogeochemical process controlling the polarization response correlate to the vegetation. On the contrary, the electrical conductivity ( $\sigma'$ ) reveals only a weak dependence on the vegetation and permanent surface runoff.~~

~~To support our observation regarding the electrical response and~~Kleinebecker et al. (2009) suggest that besides climatic variables, biogeochemical characteristics of the peat influence the composition of vegetation in wetlands. Hence, we can use variations in the vegetation as a qualitative way to evaluate our interpretation of the IP imaging results. In Fig. 8j to 8l, we

present in Fig. 8j to 8l the histograms of the electrical parameters extracted at each one of the three vegetation features defined in the experimental plot (abundant, moderate and sparse vegetation). These histograms show, in general, that the location with sparse vegetation, where the water can be seen on the i.e., with permanent surface, presents runoff, is related to the lowest phase values (histogram peak at 13 mrad). Moderate vegetation corresponds with moderate phase and  $\sigma''$  values (histogram highest peak at 1718 mrad), while and  $70 \mu\text{Sm}^{-1}$ , respectively). In comparison, the abundant vegetation corresponds with the highest phase and  $\sigma''$  values (histogram peak at 2022 mrad and  $90 \mu\text{Sm}^{-1}$ , respectively) in the top 20 cm. The histogram of the three vegetation features in terms of  $\sigma'\sigma''$  values overlap overlaps with each other. However, we observe a strong correlation between  $\sigma''$  and the vegetation, with variations in  $\sigma''$  permitting to discriminate between areas with a surface cover with abundant and moderate sphagnum and purple moor grass.

### 3.3 Comparison of electrical and geochemical data parameters

The pore fluid conductivity measured in water samples retrieved from the piezometers show little variations with values ranging between  $6.7$  and  $10.4 \text{ mSm}^{-1}$ . The evaluation of the imaging results measured along profiles By 25, By 46, and By 68 were used to guide select the selection of locations for the freeze core and sampling of pore water and groundwater. Sampling points S1 and S3 were found defined in the highly polarizable parts of the uppermost peat unit (high  $\sigma'$  and  $\sigma''$  values). In contrast, sampling point S2 is located in an area characterized by particularly low polarization values. Figures 9a-e show the chemical parameters measured in the water samples, specifically chloride ( $\text{Cl}^-$ ), phosphate ( $\text{PO}_4^{3-}$ ), dissolved organic carbon (DOC), total iron ( $\text{Fe}_{\text{tot}} = \text{Fe}^{2+} + \text{Fe}^{3+}$ ), and pH; whereas Fig. 9f-j show the chemical parameters measured in the peat samples extracted from the freeze cores, namely, cation exchange capacity (CEC), concentrations of iron sulfide (FeS or  $\text{FeS}_2$ ), total reactive iron ( $\text{Fe}_{\text{tot}}$ ), potassium ( $\text{K}^+$ ), and sodium ( $\text{Na}^+$ ). The pore-fluid conductivity measured in water samples retrieved from the piezometers shows minor variation with values ranging between  $6.7$  and  $10.4 \text{ mSm}^{-1}$ . To facilitate the comparison of electrical parameters and geochemical data, Fig. 9k-m show the complex conductivity values parameters ( $\sigma'$ ,  $\sigma''$  and  $\phi$ ) at the sampling points S1, S2 and S3, which were extracted as vertical 1D profiles from the corresponding imaging results.

As observed in Fig. 6 and 7, the highest complex conductivity values ( $\sigma'\sigma''$ ) were highest resolved within the uppermost 10-20 cm and rapidly decreased with depth. Furthermore, the values of  $\phi$  and  $\sigma''$  in the top 20 cm at S1 and S3 are significantly higher than those at the location S2. High values of  $\phi$  and  $\sigma''$  at S1 and S3 correspond with high concentrations of



DOC, phosphate<sup>-</sup>, Fe<sub>tot</sub> in water samples, as well as with high K<sup>+</sup>, and Na<sup>+</sup> contents measured in soil materials extracted from the freeze cores. ~~Fig.~~Figure 9 reveals consistent patterns between geochemical and geophysical parameters: in the first 10 cm bgs close to the sampling points S1 and S3, we observe complex conductivity values ( $\sigma^2\sigma'$  and  $\sigma^{22}$ ) as well as chemical parameters, such as DOC, phosphate, (only at S1). Accordingly, as S1 Fe<sub>tot</sub> also reveals at least two times higher concentrations than those measured in S2.

Figure 10 shows the actual correlations between the complex conductivity and Cl<sup>-</sup>, DOC, and Fe<sub>tot</sub> concentrations measured in groundwater samples. In Fig. 10, we also provide a simple linear regression analysis to quantify ~~possible correlations~~the correlation between parameters. Figure 10 reveals that the phase has a weak to moderate correlation with DOC, Cl<sup>-</sup> and Fe<sub>tot</sub>. The conductivity ( $\sigma^2\sigma'$ ) shows a slightly stronger correlation with the DOC, the Cl<sup>-</sup> and total iron concentration than the polarization ( $\sigma^{22}$ ). The highest  $\sigma^{22}$  values (>100  $\mu\text{Sm}^{-1}$ ) are related to the highest DOC and total iron concentration. ~~Hence, it is possible to interpret the high phase values as those related to biogeochemical hot spots.~~

~~Figure 11 shows~~Further evidence on the presence of the biogeochemical hot spot interpreted at the position of S1 is available by the FTIR spectroscopy analysis of the freeze core samples presented in Fig. 11. The spectra show the absorbance intensity at different wavenumbers, C-O bond (~1050  $\text{cm}^{-1}$ ), C=O double-bond (~1640  $\text{cm}^{-1}$ ), carboxyl (~1720  $\text{cm}^{-1}$ ), and O-H bonds (~3400  $\text{cm}^{-1}$ ). The peaks are also indicated in Fig. 11 with the interpretation based on the typical values reported in peatlands, for instance, McAnallen et al. (2018) or Artz et al. (2008).

## 4 DISCUSSION

### 4.1 Biogeochemical interpretation

The geochemical and geoelectrical parameters presented in Fig. 6-7 and 9 ~~displayed~~reveal consistent patterns, with the highest values within the uppermost ~~20~~10 cm around S1 and S3. The high DOC, K<sup>+</sup> and phosphate concentrations in the uppermost peat layers and especially in the areas found to be biogeochemically active, strongly suggest that there is rapid decomposition of dead plant material in these areas (Bragazza et al., 2009). Ions such as K<sup>+</sup> and phosphate are essential plant nutrients, and phosphate species especially are often the primary limiting nutrient in peatlands (Hayati and Proctor, 1991). The presence of dissolved phosphate in pore waters suggests that (i) the plant uptake rate of this essential nutrient is exceeded by its production through the decomposition of plant material; and (ii) that organic matter turnover must be rapid indeed to deliver this amount of phosphate to the pore water. This is supported

by the DOC concentrations in pore waters exceeding 10 mM. DOC is produced as a decomposition product during microbial hydrolysis and oxidation of solid phase organic carbon via enzymes such as phenol oxidase (Kang et al., 2018). Enzymatic oxidation processes are enhanced by oxygen ingress via diffusion and, more importantly, by water table fluctuations that work as an ~~'oxygen pump'~~'oxygen pump' to the shallow subsurface (Estop-Aragonés et al., 2012). Thus, an increased DOC concentration in the pore water can be used as an indicator for microbial activity (Elifantz et al., 2011; Liu, 2013). The small amount of phosphate measured in the less active area ~~S3~~S2 can be explained by advective transport from the active area S1, which is ~~directly 'up-stream'~~direct 'up-stream' of S2. In this case, ~~advective~~advective water flow through the uppermost peat layers along the hydrological head gradient may have transported a small amount of reaction products from the biogeochemical source areas to the ~~'non-active'~~'non-active' area. The high DOC, Fe, K<sup>+</sup> and phosphate (only at S1) levels confirm our initial interpretation of the highly conductive and polarizable geophysical units within the first 20 - 50 cm bgs in the surroundings of S1 and S3 as biogeochemically active areas.

The high DOC concentrations are also likely to be directly or indirectly responsible for the Fe maximum in the upper layers. Dissolved Fe was predominantly found as Fe<sup>2+</sup> (reducing conditions) suggesting either that high labile DOC levels maintain a low redox potential, or that the dissolved Fe<sup>2+</sup> was complexed with the DOC limiting the oxidation kinetics enough so that Fe<sup>2+</sup> can accumulate in peat pore waters. The TRIS analysis clearly showed very low levels of sulfide minerals in both freeze cores, especially in the uppermost peat layers. This was unexpected considering the reducing conditions implied by the dominance of pore water Fe<sup>2+</sup>. We argue that the lack of sulfide minerals is due to insufficient H<sub>2</sub>S or HS<sup>-</sup> needed to form FeS or FeS<sub>2</sub>, or that the redox potential was not low enough to reduce sulfate to H<sub>2</sub>S or HS<sup>-</sup>. Both mechanisms are possible, as groundwater in the catchment ~~has~~has generally low sulfate concentrations, and yet sulfate was detected in peat pore water samples, which would not be expected if redox potentials were low enough to reduce sulfate to sulfide. The chemical analyses do not reveal any significant or systemic vertical gradient in mineral sulfide concentrations, as expected for the site (Frei et al., 2012). The maximum in extractable (reactive) solid phase Fe was also located in the ~~upper-most~~uppermost peat layer at the ~~'hot-spot'~~'hot spot' S1. This Fe was likely in the form of iron oxides or bound to/in the plant organic matter. Such iron rich layers typically form at the redox boundary between oxic and anoxic zones and can be quite highly dynamic depending on variations in the peatland water levels and oxygen ingress (Wang et al., 2017; Estop-Aragonés, 2013).

Similar to other peatlands, (Artz et al., 2008), the FTIR spectra show the presence of carbon-oxygen bonds such as C-O, C=O and COOH both at S1 and S2, furthermore, the peak intensities at S1 tend to decrease with the depth, while the peak intensities at S2 samples tend to increase ~~correlated to the IP response~~ in agreement with the increase in the polarization response (both phase and  $\sigma''$ ). This observation further supports our interpretation of the shallow 10 cm in IP images in the vicinity of S1 as a biogeochemical hot spot. However, such biogeochemical hot spot is not related to the accumulation of iron sulfides, which was suggested by Abdel Aal and Atekwana (2014) or Wainwright et al. (2016) as the main parameter controlling the high IP response. The phosphate and Fe could potentially form complexes with the O-H groups that show an absorbance peak at 1050 cm<sup>-1</sup> (Arai and Sparks, 2001; Parikh and Chorover, 2006). Furthermore, the iron can also form complexes with the carboxyl groups (absorbance at ~1720 cm<sup>-1</sup>).

#### 4.2 Correlation between the peat and the electrical signatures

The two electrical units observed within the peat indicate variations in the biogeochemical activity with depth. Thus, it is likely that the anomalies associated with the highest  $\sigma'$ ,  $\sigma''$ , and  $\phi$  values in the uppermost unit ~~characterized by the highest  $\sigma'$ ,  $\sigma''$ , and  $\phi$  values corresponds with an~~ correspond with the location of active biogeochemical ~~zone~~ zones, i.e., a hot spot. Consequently, the ~~second electrical unit associated with moderate  $\sigma'$ ,  $\sigma''$ , and  $\phi$  values is probably related to indicate~~ less biogeochemical active or even inactive zone in the peat. The third unit represents the granitic bedrock. The low metal content and the well-crystallized-crystallized form of the granite ~~leads lead~~ to low  $\sigma''$  values (here, < 40  $\mu\text{Sm}^{-1}$ ), as suggested by Marshall and Madden (1959).

The high polarization response of the biogeochemically active peat (here  $\sigma'' > 100 \mu\text{Sm}^{-1}$  and  $\phi > 18 \text{ mrad}$ ) is consistent with the measurements of McAnallen et al. (2018), who performed time-domain IP measurements in different peatlands. They suggest that the active peat is less polarizable due to the presence of the abundant sphagnum cover. They found that in the areas where the peat is actively accumulating, the ratio of the vascular plants and the non-vascular sphagnum is low, and therefore, the oxygen availability is low. However, the sphagnum is expected to exude a small amount of carbon into the peat, and Fenner et al. (2004) found that the sphagnum contributes to the DOC leachate to the pore water, which is contradictory to the model of McAnallen et al. (2018). ~~Our observations also show that the high DOC content correlates to an abundant sphagnum cover, in agreement with the observations by Fenner et al. (2004), but we cannot clearly tie this to exudates from roots or decomposition of decaying~~

organic matter. Moreover, at our study site the abundant sphagnum grows in conjunction with abundant purple moor grass. Figures 8-9 reveal an increase in the polarization response ( $\sigma''$ ,  $\phi$ ) with both the abundant vegetation as well as with DOC concentrations measured in pore water. Thus, supporting our interpretation that high polarization values ( $\sigma'' > 100 \mu\text{Sm}^{-1}$  and  $\phi > 20$  mrad) correspond to biogeochemical hot spots rather than vegetation type. Recent In agreement with Fenner et al. (2004), in our study, we also observe that high DOC content correlates with abundant sphagnum cover; which is also found in conjunction with abundant purple moor-grass. In this regard, recent studies have demonstrated an increase in the polarization response due to the accumulation of biomass and activity in the root system (e.g., Weigand and Kemna 2017; Tsukanov and Schwartz, 2020). However, the sphagnum does not have roots; ~~on~~thus, it cannot directly contribute to the ~~other hand,~~polarization response. McAnallen et al. (2018) suggest that the vascular purple moor-grass can contribute to the high IP, as the roots transport oxygen into the deeper area ~~which contributes to, increasing the increase in the polaritywettability~~ and normalized chargeability of the peat ~~(McAnallen et al., 2018).~~

Derived from the results and discussion above, we delineated the geometry of the hot spots. The map presented in Fig. 12 is based on the maps of phase and imaginary conductivity values at a depth of 10 and 20 cm. Hot spots interpreted at those areas exceeding both a phase value of 18 mrad and imaginary conductivity of  $100 \mu\text{Sm}^{-1}$  at the same time. Besides the geometry of the hot spots, Fig. 12 indicates that the hot spot activity attenuates with the depth.

#### 4.3 Possible polarization mechanisms

~~The low-frequency polarization response of subsurface materials is usually attributed to either electrode polarization of highly conductive metallic minerals (e.g., Pelton et al., 1978; Wong, 1979) or the polarization of the electrical double layer (EDL) covering charged surfaces of electrically non-conducting solid matter (e.g., Waxman and Smits, 1968; Revil and Florsch, 2010; Revil, 2012).~~

~~Electrode polarization arises from the conductivity contrast between the high electronic (or semi-conductor) conductivity of the metallic mineral and the much lower electrolytic conductivity of the surrounding pore fluid (Wong, 1979). This so-called charge polarization has its maximum at relatively high frequencies ( $\gg 1$  kHz) unless the conducting minerals are large (Bücker et al., 2018). If charge transfer reactions take place at the mineral-electrolyte interface, e.g., due to the presence of metallic ions in the electrolyte, the charge polarization is accompanied by a concentration polarization, which potentially shifts the maximum of the electrode polarization response into the low-frequency range (Bücker et al., 2018). The~~

~~magnitude of electrode polarization is proportional to the volume content of the conductive minerals (Wong, 1979).~~

In this study, we have found a strong correlation between the polarization response ( $\phi$  and  $\sigma''/\sigma'$ ) and  $\text{Fe}_{\text{tot}}$  in the solid phase and a less pronounced correlation between the polarization response and the concentration of dissolved iron in the liquid phase (see Fig. 10). In both groups of mechanistic polarization models, the phase value depends on the volumetric content of metallic particles (Wong, 1979; Revil 2015a, 2015b; Bückner et al., 2018, 2019) and therefore, the phase could reveal the possible metallic content in the peat. If the iron in the solid phase occurred in the form of highly conductive minerals, the two above correlations would clearly point to ~~an electrode~~ the polarization mechanism of perfect conductors described by Wong (1979) as a possible explanation for the observed response. Previous studies (e.g., Flores Orozco et al., 2011; 2013) ~~imply that~~ attributed the polarization of iron sulfides ( $\text{FeS}$  or  $\text{FeS}_2$ ) in sediments ~~polarize to such a polarization mechanism~~ as long as sufficient  $\text{Fe}^{2+}$  cations are available in the pore water. Such effect has been investigated in detail by Bückner et al. (2018; 2019), regarding the changes in the polarization response due to surface charge and reaction currents carried by redox reactions of metal ions at the mineral surface. However, in the case of the present study, the lack of sulfide, and the rather high  $\text{E}_{\text{h}}/\text{pH}$  (inferred from the presence of sulfate) in the pore water, do not favor the precipitation of conductive sulfides such as pyrite. Under these conditions, iron would rather precipitate as iron oxide or form iron-organic matter complexes. The electrical conductivity of most iron oxides is orders of magnitude smaller than the conductivity of sulfides (e.g., Cornell and Schwertmann, 1996), and is thus too low to explain an increased ~~electrode polarization~~ polarization based on a perfect-conductor polarization model (e.g., Wong, 1979; Bückner et al., 2018, 2019). The only highly conductive iron oxide is magnetite, with a conductivity similar to pyrite (Atekwana et al., 2016). Consequently, the presence of magnetite could explain ~~the electrodes~~ such a polarization. However, the low pH ( $\sim 5$ ) typical for peat systems does not favor the precipitation of magnetite, but rather ~~less conducting, and thus, less polarizable~~ iron (oxy)hydroxides such as ferrihydrite (Andrade et al., 2010; Linke and Gislason, 2018). Analysis of sediments of the freeze core did also not reveal magnetite. As indicated by the FTIR analysis, the iron might furthermore have built complexes with the carboxyl (absorbance at  $\sim 1720 \text{ cm}^{-1}$ ). ~~However, it is not clear if these iron complexes might have a sufficiently high electrical conductivity and—in the fashion of conductive iron minerals—permit electrode polarization mechanism.~~ Such moderately conductive iron minerals or iron-organic complexes might still cause a relatively strong polarization response as predicted by the polarization model developed by Revil et al. (2015)

and Misra et al. (2016a). In this model, which attributes the polarization response to a diffuse intra-grain relaxation mechanism, the polarization magnitude is mainly controlled by the volumetric content. In this model, the (moderate) particle conductivity only plays a secondary role (e.g., Misra et al. 2016b).

~~In addition to the polarization of conductive minerals, the polarization response can also be caused by the polarization of the electrical double layer (EDL). The EDL consists of an inner Stern layer of cations adsorbed to the charged solid surface and an outer diffuse layer. Together, these two layers screen the (usually) negative charge of the solid surface and represent a highly conductive and, thus, polarizable surface layer. Depending on the specific geometry of solid and liquid phases, EDL polarization can be understood in terms of the polarization of isolated EDL covered grains (e.g., Schwarz, Schurr, Leroy et al., 2008) or in terms of membrane polarization linked to EDL covered pore constrictions (e.g., Marshall and Madden, 1959; Bückner and Hördt, 2013; Bückner et al., 2019). Regardless of the specific mechanism, all EDL polarization mechanisms strongly depend on the specific surface area of the material and the charge density at the surface (Revil, 2012; Waxman and Smits, 1968).~~

The product of both surface charge density and specific surface area can be quantified by the Cation Exchange Capacity (CEC) of a material. As peat mainly consists of organic matter known to have a high CEC ~~–~~, even when compared to most clay minerals ~~–~~ (e.g., Schwartz and Furman, 2014; and references therein), ~~EDL~~the polarization of charged organic surfaces may explain the observed IP response. ~~However,–~~Additionally, Garcia-Artigas et al. (2020) concluded that bioclogging due to fine particles and biofilms increases the specific surface area and the CEC, resulting in this case, we would expect an increase ofin the polarization ~~magnitude with the CEC, which we did not observe in our data. The~~response. However, the CEC values ~~of~~measured in samples retrieved from the freeze core ~~samples all~~ vary ~~within~~in a narrow range ~~of~~between 5 ~~to~~and 25 meq/kg and we did not observe any correlation between CEC and changes in the polarization magnitude ( $\sigma''$ ,  $\sigma''$ ,  $\phi$ ). Such lack of correlation between the polarization effect and the CEC was also reported by Ponziani et al. (2011), who conducted ~~SIP~~spectral IP measurements on a set of peat samples, ~~have reported a similar observation previously. In~~ summary. Hence, the measured CEC is high enough to explain a rise in EDL-polarization; however, the (small) variation in CEC does not explain the observed variation in the polarization magnitude.

The pH of the pore fluid is also known to control the magnitude of EDL polarization; an increase of pH usually corresponds with an increase of the polarization magnitude (e.g., Skold et al.,



2011). At low pH values,  $H^+$  ions occupy (negative) surface sites and thus reduce the net surface charge of the EDL (e.g., Hördt et al., 2016; and references therein). Our data seems to show the opposite behavior: we found a lower pH in the highly polarizable anomalies at S1 and S3 compared to site S2 (the inactive and less polarizable location); while the pH increases at depth for decreasing values in the polarization (both  $\sigma''$  and  $\phi$ ). At the same time, variations in pH are within the range 4.45 and 5.77 and thus might not be sufficiently large to cause the observed variation in the polarization response.

Besides pH, pore fluid salinity plays a major role in the control of EDL polarization. Laboratory measurements on sand and sandstone samples indicated that an increase in salinity leads to an early increase of the imaginary conductivity, which is eventually followed by a peak and a decrease at very high salinities during later stages of the experiments (e.g., Revil and Skold, 2011; Weller et al., 2015). Hördt et al. (2016) provided a possible theoretical explanation of this behavior: In their membrane-polarization model, salinity controls the thickness of the (diffuse layer) of the EDL and depending on the specific geometry of the pores; there is an optimum thickness, which maximizes the magnitude of the polarization response. In the present study, we observed that an increase in salinity (as indicated by the high  $Cl^-$  concentrations within the uppermost 10 cm at all sampling locations) is associated with an increase in the polarization magnitude response (e.g., Revil and Skold, 2011; Weller et al., 2015; Hördt et al., 2016). However, the highest  $Cl^-$  concentrations were observed for the shallow layers at location S2, where we measured lower polarization magnitudes (in terms of  $\sigma''$ ,  $\phi$ ) compared to S1 and S3.

The strong correlation between the polarization response and the DOC suggests an, as yet not fully understood, causal relationship. A similar observation has recently been reported by Flores Orozco et al. (2020), who found a strong correlation between DOC the organic carbon content as a proxy of microbial activity and both  $\sigma'$  and  $\sigma''$  in a municipal waste landfill in Austria. Regarding the available carbon, McAnallen et al. (2018) reported a strong correlation between the occurrence of long-chained C=O double bonds and the total chargeability of peat material. The upper peat layers are exposed to oxygen leading to oxidation of the peat and formation of C=O double bonds at solid phase surfaces and in the pore water DOC. Such long-chained organic molecules have an increased polarity wettability and thus more readily attach (or even form at organic matter surfaces) to the surface of solid organic and mineral particles (Alonso et al., 2009). Based on a membrane-polarization model, Bucker et al. (2017) predict an increase of the polarization magnitude in the presence of wetting (i.e., long-chained) hydrocarbon in the free phase. The long-chained polar DOC attaches to the peat surface, similar

to polar hydrocarbon, and so it might provide extra surface charge, thus reducing the pore space and causing membrane polarization (Marshall and Madden, 1959).

As suggested by Vindedahl et al. (2016), organic matter can adsorb to the iron oxide surface via electrostatic attraction and provides a negatively charged macromolecular layer on the iron oxide. Such complexes could also explain the observed increase in the polarization response in the anomalies interpreted as biogeochemical hot spots. The point of zero charge of the peat is below pH 4 (Bakatula et al., 2018); while for iron (-oxide) is varying between ~5 and ~9 (Kosmulski et al., 2003). This means that the organic matter is probably negatively charged, and the iron oxide is most likely positively charged since the measured pH at the sample points is varying between 4.5 and 5.8 with lower values in the top 10 cm in the hot spot area. Hence, in the shallow 10 cm from S1 and S3, the pH favors the DOC to bond with the iron in the solid phase.

## 5 Conclusions

We investigated the applicability of induced polarization (IP) as a tool to identify and localize biogeochemically active areas or “hot spots” in peatlands. ~~IP imaging results revealed a high polarization response for areas that based on the solid and solution phase chemistry can be interpreted as biogeochemically active. The chemical analysis of the groundwater samples and the freeze core indicate that the top 10–20 cm correspond to the highest geochemical activity, whereas the deeper areas of the peat are less active. Accordingly, only the upper 10–20 cm can be defined as biogeochemical hot spots. Moreover, our geophysical survey shows that IP imaging can delineate the geometry of the hot spot with high spatial resolution. In this regard, indicators for increased biogeochemical activity (DOC, phosphate) correspond to a high IP response ( $\sigma''$  and phase values).~~

hot spots in peatlands. Although the exact polarization mechanism is not fully understood, our results reveal that the IP response of the peat changes with the level of biogeochemical activity. Thus, the IP method is capable of distinguishing between biogeochemical active and inactive zones within the peat. The phase and imaginary conductivity values show a contrast between these active and inactive zones and characterize the geometry of the hot spots even if iron sulfides are not present. ~~Previous studies have only investigated~~ The joint interpretation of chemical and geophysical data indicates that anomalous regions (characterized by phase values above 18 mrad and imaginary conductivity of 100  $\mu\text{Sm}^{-1}$ ) delineate the conductive properties geometry of the hot spots, which are limited to the top 10 cm bgs. Deeper areas (>10

cm) of the peat; ~~however, biogeochemical activity within the peat has not been investigated.~~  
are less active. In this regard, our study shows ~~a new alternative that the induced polarization~~  
method is able to characterize biogeochemical changes and their geometry within ~~the~~ peat with  
high resolution.

~~For further improvement and understanding of the effect of~~ Additionally, our study  
demonstrates the ability of the IP method to assess biogeochemically active zones even if they  
are not related to the microbiologically mediated accumulation of iron sulfides. We identify  
complexes of organic matter ~~such and iron as DOC or phosphate on~~ possible causes of the high  
polarization response, ~~further of the carbon turnover hot spots investigated in our study.~~ Further  
laboratory studies on peat samples with different ~~concentration~~ concentrations and  
~~mixture~~ mixtures of DOC, phosphate, and iron in the pore fluid are required to fully understand  
the effect in IP signatures due to iron-organic complexes and the control phosphate exerts over  
the related polarization process.

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## Data availability

All data are available from the corresponding author upon request.

## Competing interests.

The authors declare that they have no conflict of interest.

## Author contribution

AFO and TK designed the experimental set-up, TK conducted the field survey and analysis of  
the geophysical data. BG and SF conducted the geochemical measurements and their  
interpretation. AFO, MB and TK interpreted the geophysical signatures. TK lead the  
preparation of the draft, where SF, BG, MB and AFO contributed equally.

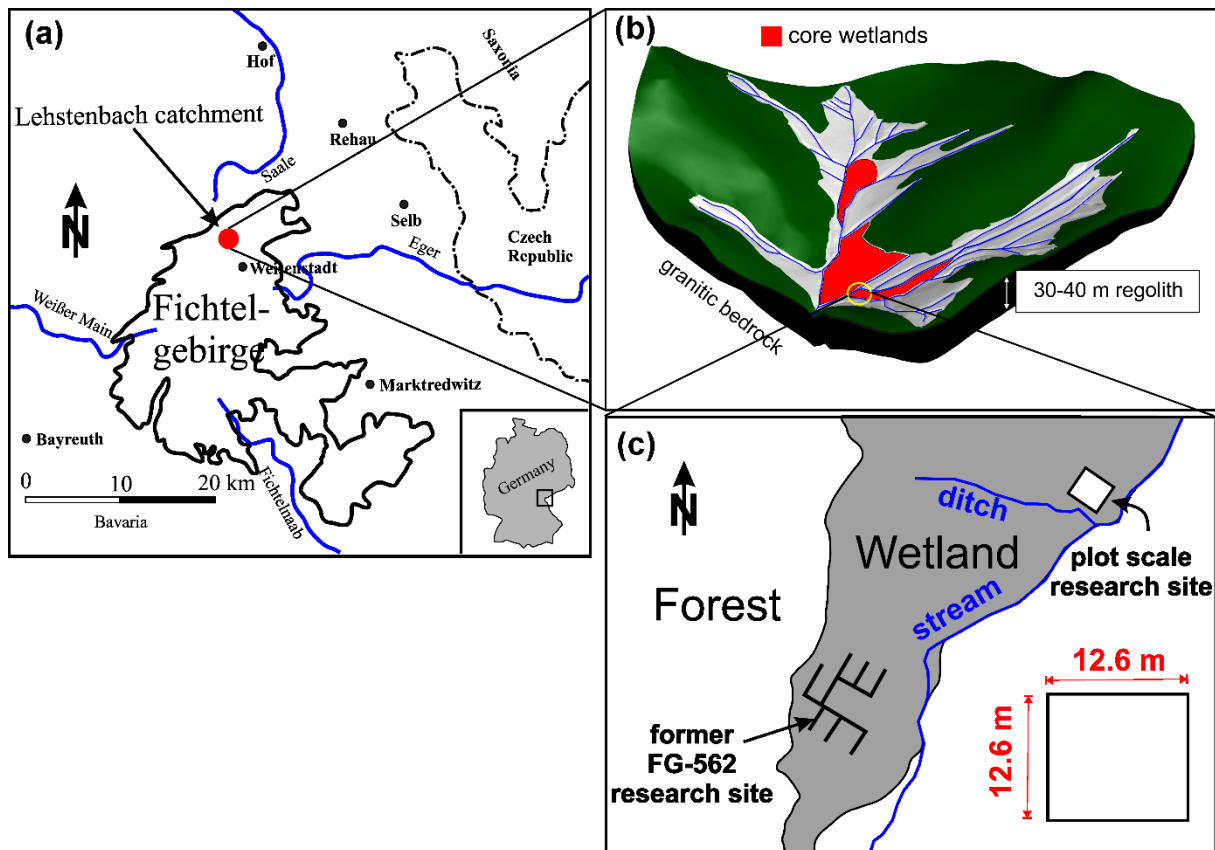


Figure 1: (a) General overview of the experimental plot located in the Fichtel Mountains and (b) structure of the bowl shaped Lehstenbach catchment, and (c) location of the experimental plot.

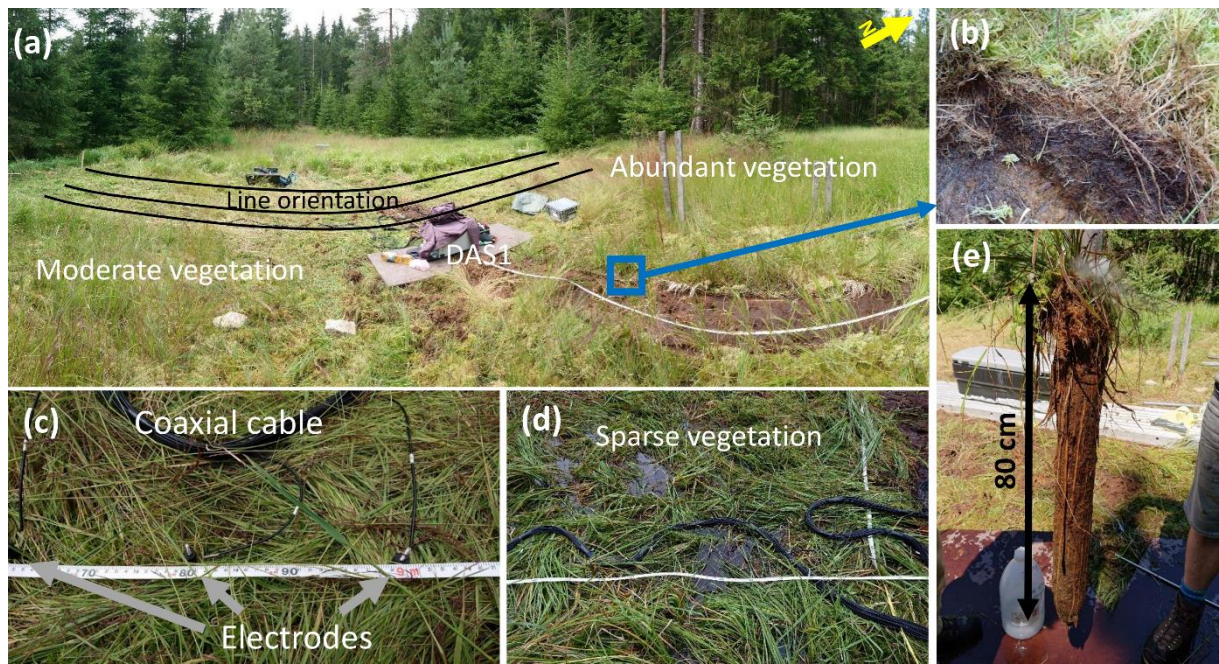
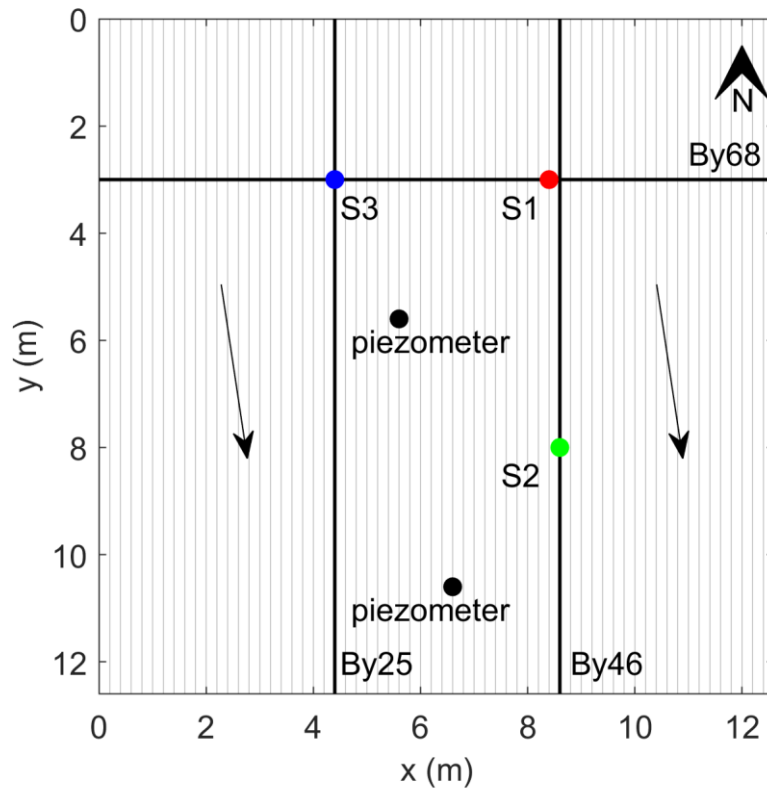


Figure 2: (a) Panoramic overview of the study site and the measurement setup. Pictures show the experimental setup and differences in the vegetation density between the northern and southern part of the experimental plot. The ~~spectral~~-induced polarization (~~SIP~~) lines appear distorted due to the panoramic view. (b) Sphagnum in the northern part of the experimental plot. (c) Coaxial cables and stainless steel electrodes used for ~~SIP~~ measurements. (d) Vegetation and the coaxial cable bundle used for IP measurements at the water covered area in the southeastern part of the experimental plot. (e) The freeze core shows the internal structure of the peat.



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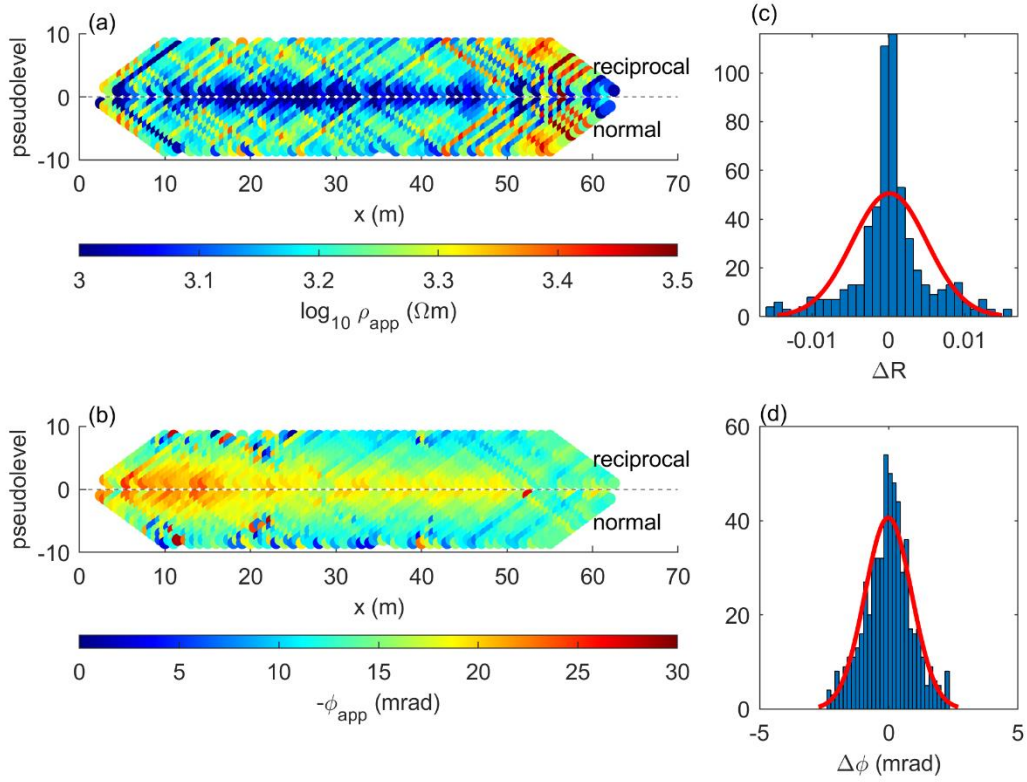


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*Figure 3: Schematic map of the experimental plot. The solid lines represent the measured profiles; the bold lines represent the position of the profiles discussed in this manuscript (By 25, By 46 and By 68). The arrows indicate the ground water flow direction. The points represent the locations of fluid (S1, S2 and S3) and freeze core (S1, S2) samples as well as the position of piezometric tubes, where the water level was measured.*

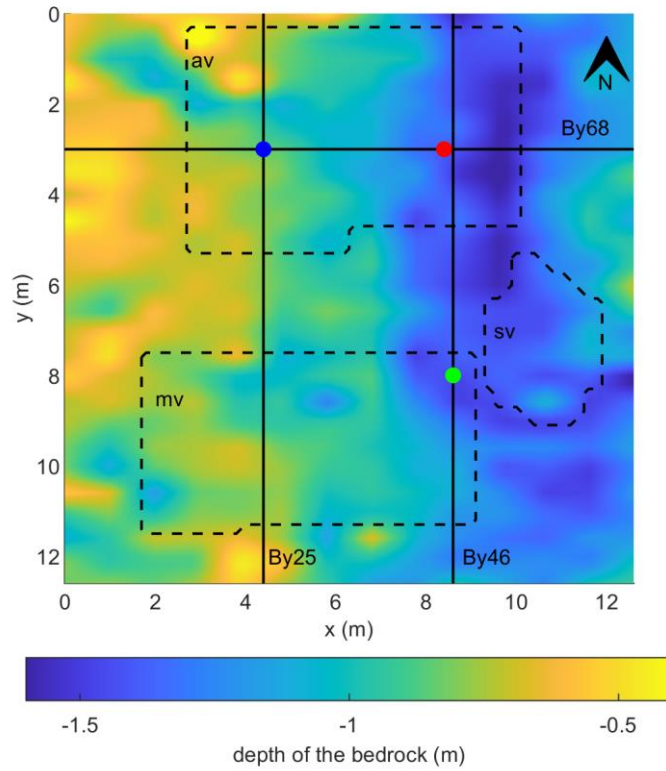
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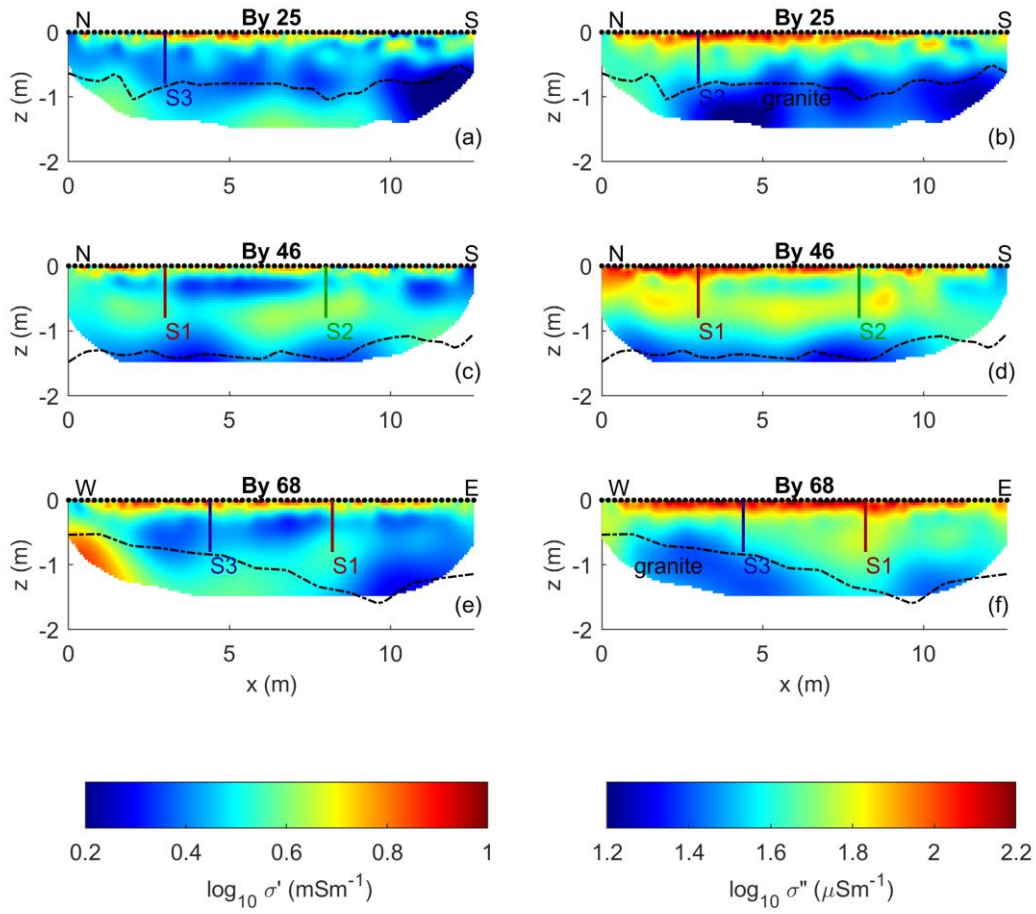
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*Figure 4: Raw data analysis. Raw-data pseudosections of (a) the apparent resistivity and (b) the apparent phase shift for measurements collected along profile By 25. Histograms of the normal-reciprocal misfits of (c) the measured resistance and (d) the apparent phase shift.*



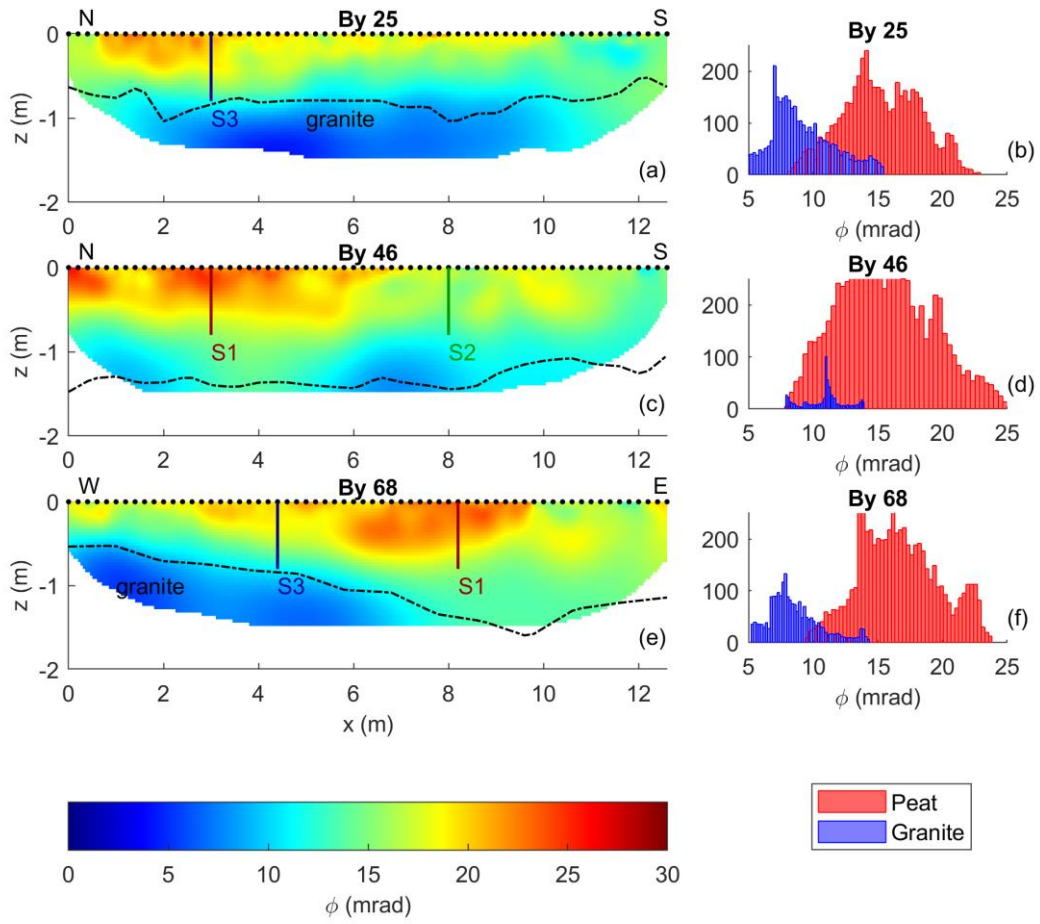
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*Figure 5: Variations in the thickness of the peat layer, i.e., depth to the granite bedrock. The positions of the three selected IP profiles By 25, By 46, and By 68 are indicated (solid lines) as well as the position of the sampling points and the geometry of the three classes of vegetation cover: abundant vegetation (av), moderate vegetation (mv), and sparse vegetation (sv).*



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Figure 6: Imaging results for data collected along profiles By 25 (a-b), By 46 (c-d), and By 68 (e-f) expressed as real  $\sigma'$  and imaginary  $\sigma''$  components of the complex conductivity. The dashed lines represent the contact between the peat and granite; the black dots show the electrode positions at the surface. The vertical lines represent the location of the fluid (S1, S2 and S3) and freeze core (S1, S2) samples.



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Figure 7: Imaging results for data collected along profiles By 25 (a), By 46 (c), and By 68 (e), expressed as phase values  $\phi$  of the complex conductivity. The dashed lines represent the contact between peat and granite; the black dots show the electrode positions at the surface. The vertical lines represent the location of the fluid (S1, S2, S3) and freeze core (S1, S2) samples. The histograms represent the phase values of the granite and peat extracted from the imaging results in (6b, 6d, 6f) according to the geometry of the dashed lines.

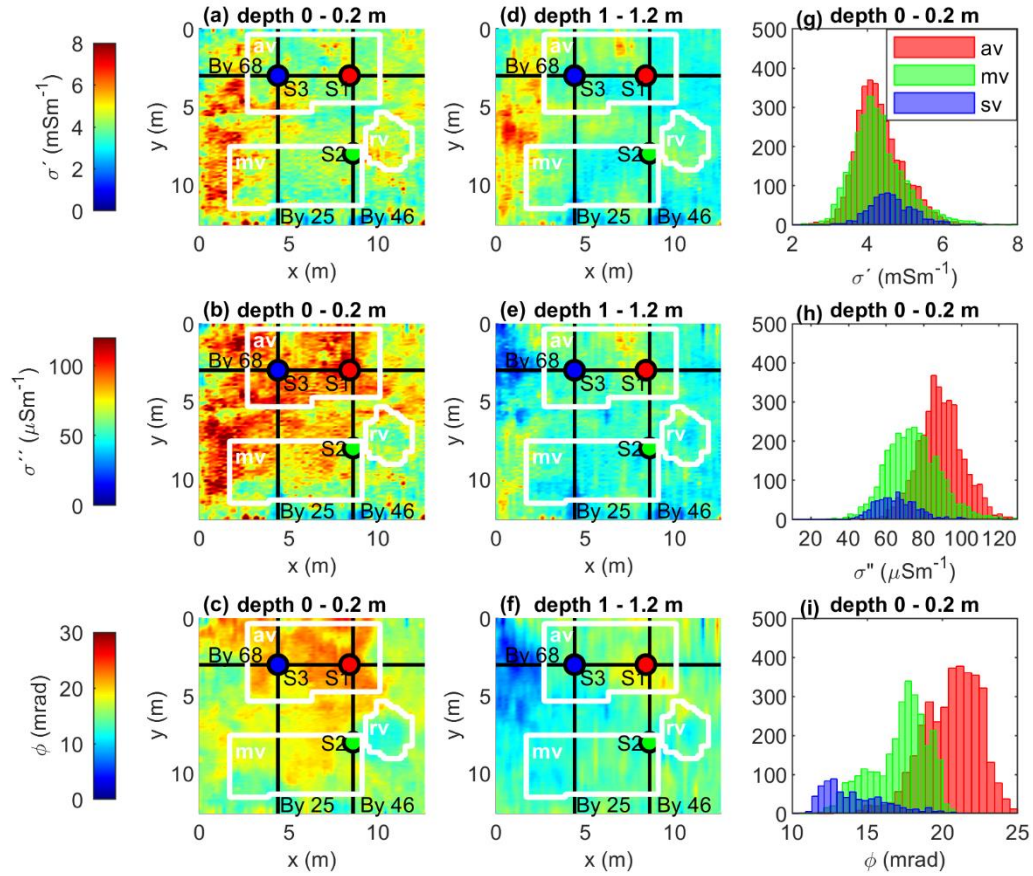


Figure 8: Maps of the complex conductivity at different depths. The black lines indicate the profiles By 25, By 46, and By 68. The dots represent the locations of the vertical sampling profiles S1, S2, and S3. The white lines outline areas classified as (av) abundant vegetation, (mv) moderate vegetation, (sv) sparse vegetation, and histograms of the complex-conductivity imaging results of the masked areas, the abundant vegetation (red bins), the moderate vegetation (green bins) and the sparse vegetation (blue bins).

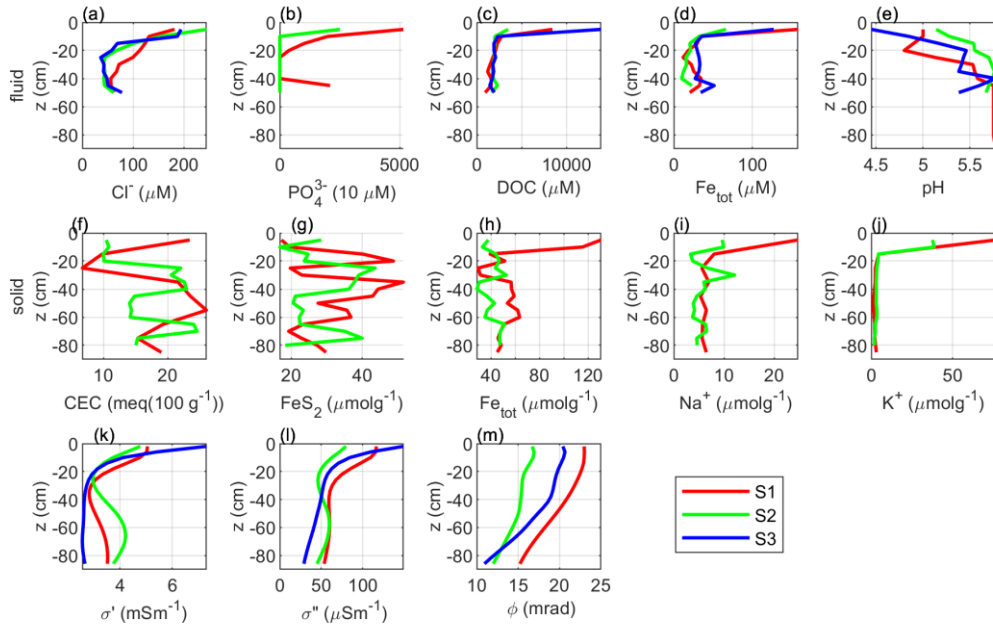


Figure 9: Results of geochemical analyses of water and soil samples. Fluid-sample analysis of the (a) chloride  $\text{Cl}^-$ , (b) phosphate  $\text{PO}_4^{3-}$ , (c) dissolved organic carbon, (d) total iron  $\text{Fe}_{\text{tot}}$ , and (e) pH. Freeze-core sample analysis of the (f) cation exchange capacity CEC, (g) iron sulfide  $\text{FeS}_2$ , (h) total iron  $\text{Fe}_{\text{tot}}$ , (i) sodium  $\text{Na}^+$ , and (j) potassium  $\text{K}^+$ . Imaging results at the three sampling locations in terms of (k) real component  $\sigma'$ , (l) imaginary component  $\sigma''$ , and (m) phase  $\phi$  of the complex conductivity.



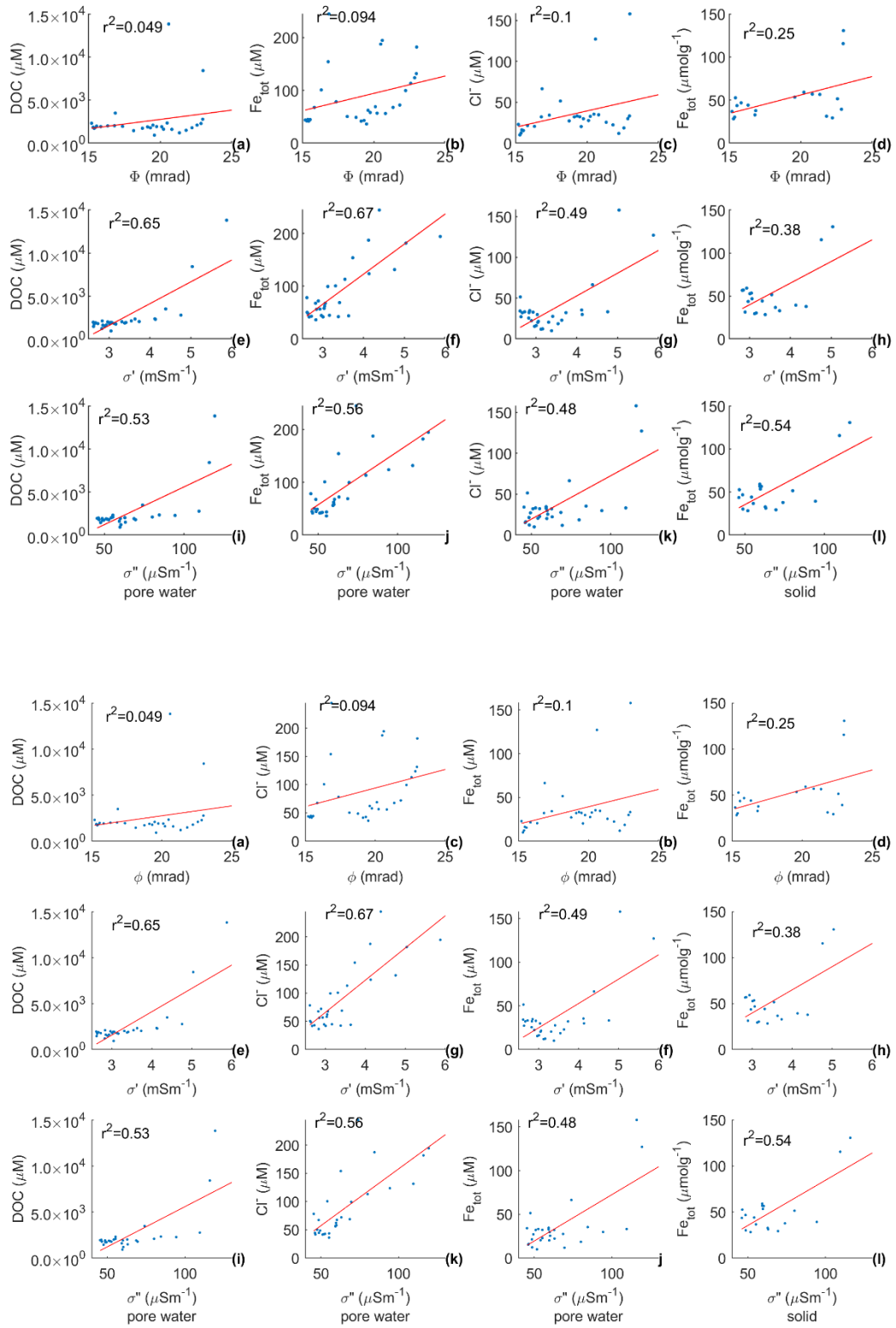
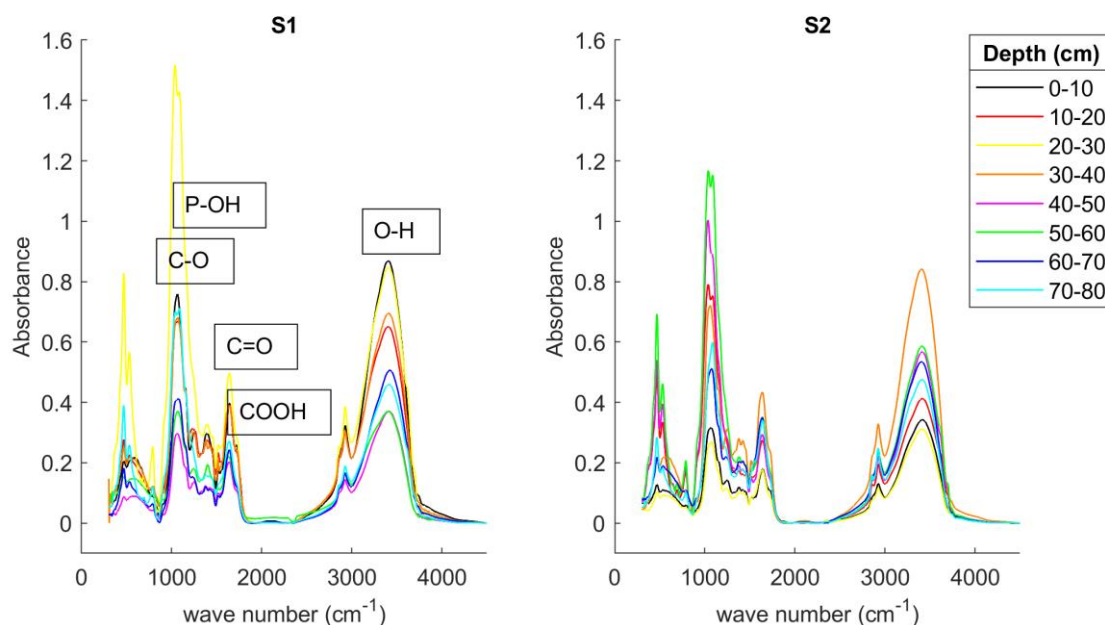


Figure 10: Correlations between the geophysical and geochemical parameters, phase ( $\phi$ ), the real ( $\sigma'$ ) and imaginary ( $\sigma''$ ) component of the complex conductivity (retrieved from the imaging results) and the biogeochemical analysis, expressed in terms of the dissolved organic carbon (DOC), and chloride ( $\text{Cl}^-$ ) content from the pore fluid samples and total iron ( $\text{Fe}_{\text{tot}}$ )

content from pore fluid in  $\mu\text{mol l}^{-1}$  and solid samples in  $\mu\text{mol g}^{-1}$ . The correlation coefficients of least square regressions analysis are shown in the top left corners of the subplots.

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Figure 11: Fourier transform infrared (FTIR) spectroscopy of the freeze core samples collected at S1 (left panel) and S2 (right panel). Each sample was extracted from the 10 cm segments. The lines represent the depth at every 10 cm between 0 and 80 cm below ground surface. The relevant peaks show the absorbance intensity, the interpretation is based on Artz et al. (2008), Arai and Sparks (2001), Parikh and Chorover (2006).

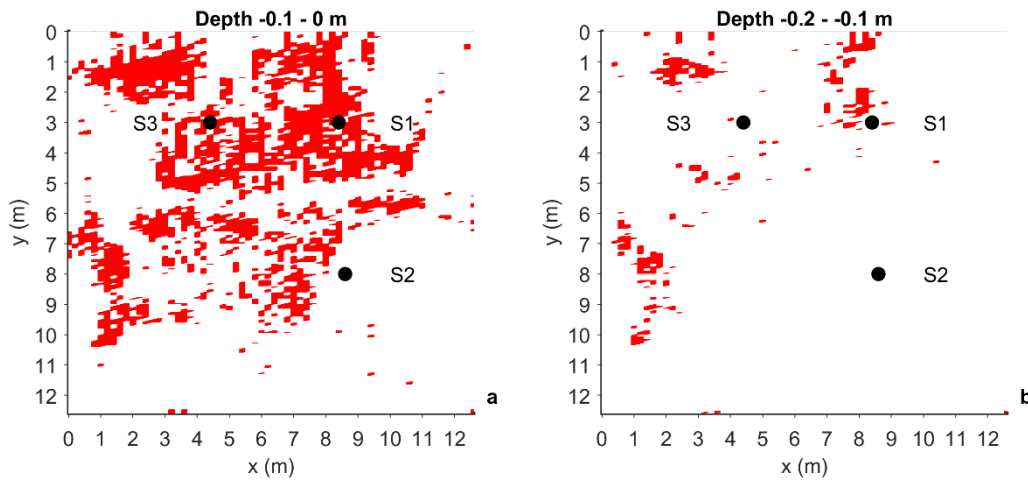


Figure 12: Imaging results in terms of imaginary component of the complex conductivity  $\sigma'' > 100 \mu\text{Sm}^{-1}$ , and phase  $\phi > 18 \text{ mrad}$ , indicating the hot spot geometry at depths of (a) 10 cm and at (b) 20 cm. The dots represent the locations of the vertical sampling profiles S1, S2, and S3.

## References

1. Abdel Aal, G. Z., & Atekwana, E. A. (2014). Spectral induced polarization (SIP) response of biodegraded oil in porous media. *Geophysical Journal International*, 196(2), 804-817.
2. Abdel Aal, G. Z., Atekwana, E. A., Rossbach, S., & Werkema, D. D. (2010a). Sensitivity of geoelectrical measurements to the presence of bacteria in porous media. *Journal of Geophysical Research: Biogeosciences*, 115(G3).
3. Abdel Aal, Gamal Z., Estella A. Atekwana, and Eliot A. Atekwana. "Effect of bioclogging in porous media on complex conductivity signatures." *Journal of Geophysical Research: Biogeosciences* 115, no. G3 (2010b).
4. Abdel Aal, G. Z., Atekwana, E. A., & Revil, A. (2014). Geophysical signatures of disseminated iron minerals: A proxy for understanding subsurface biophysicochemical processes. *Journal of Geophysical Research: Biogeosciences*, 119(9), 1831-1849.

5. Albrecht, R., Gourry, J. C., Simonnot, M. O., & Leyval, C. (2011). Complex conductivity response to microbial growth and biofilm formation on phenanthrene spiked medium. *Journal of Applied Geophysics*, 75(3), 558-564.
6. Alonso, D. M., Granados, M. L., Mariscal, R., & Douhal, A. (2009). Polarity of the acid chain of esters and transesterification activity of acid catalysts. *Journal of Catalysis*, 262(1), 18-26.
7. Andrade, Â. L., Souza, D. M., Pereira, M. C., Fabris, J. D., & Domingues, R. Z. (2010). pH effect on the synthesis of magnetite nanoparticles by the chemical reduction-precipitation method. *Quimica Nova*, 33(3), 524-527.
8. Arai, Y., & Sparks, D. L. (2001). ATR-FTIR spectroscopic investigation on phosphate adsorption mechanisms at the ferrihydrite-water interface. *Journal of Colloid and Interface Science*, 241(2), 317-326.
9. Artz, R. R., Chapman, S. J., Robertson, A. J., Potts, J. M., Laggoun-Défarge, F., Gogo, S., ... & Francez, A. J. (2008). FTIR spectroscopy can be used as a screening tool for organic matter quality in regenerating cutover peatlands. *Soil Biology and Biochemistry*, 40(2), 515-527.
10. Atekwana, E., Patrauchan, M., & Revil, A. (2016). *Induced Polarization Signature of Biofilms in Porous Media: From Laboratory Experiments to Theoretical Developments and Validation* (No. DOE-Okstate-SC0007118). Oklahoma State Univ., Stillwater, OK (United States).
11. Atekwana, E. A., & Slater, L. D. (2009). Biogeophysics: A new frontier in Earth science research. *Reviews of Geophysics*, 47(4).
12. Bakatula, E. N., Richard, D., Neculita, C. M., & Zagury, G. J. (2018). Determination of point of zero charge of natural organic materials. *Environmental Science and Pollution Research*, 25(8), 7823-7833.

~~12.13.~~ Biester, H., Knorr, K. H., Schellekens, J., Basler, A., & Hermanns, Y. M. (2014). Comparison of different methods to determine the degree of peat decomposition in peat bogs. *Biogeosciences*, 11(10), 2691-2707.

~~13.14.~~ Binley, A., & Kemna, A. (2005). DC resistivity and induced polarization methods. In *Hydrogeophysics* (pp. 129-156). Springer, Dordrecht.

~~14.15.~~ Binley, A., & Slater, L. (2020). *Resistivity and Induced Polarization: Theory and Applications to the Near-surface Earth*. Cambridge University Press.  
~~Hubbard, S. S., Huisman, J. A., Revil, A., Robinson, D. A., Singha, K., & Slater, L. D. (2015). The emergence of hydrogeophysics for improved understanding of subsurface processes over multiple scales. *Water resources research*, 51(6), 3837-3866.~~

16. Binley, A., Hubbard, S. S., Huisman, J. A., Revil, A., Robinson, D. A., Singha, K., & Slater, L. D. (2015). The emergence of hydrogeophysics for improved understanding of subsurface processes over multiple scales. *Water resources research*, 51(6), 3837-3866.

17. Boano, F., Harvey, J. W., Marion, A., Packman, A. I., Revelli, R., Ridolfi, L., & Wörmann, A. (2014). Hyporheic flow and transport processes: Mechanisms, models, and biogeochemical implications. *Reviews of Geophysics*, 52(4), 603-679.

~~16.18.~~ Bragazza, L., Buttler, A., Siegenthaler, A., & Mitchell, E. A. (2009). Plant litter decomposition and nutrient release in peatlands. *Geoph. Monog. Series*, 184, 99-110.

~~17.19.~~ Bucker, M., & Hördt, A. (2013). Analytical modelling of membrane polarization with explicit parametrization of pore radii and the electrical double layer. *Geophysical Journal International*, 194(2), 804-813.

~~18.20.~~ Bucker, M., Orozco, A. F., Hördt, A., & Kemna, A. (2017). An analytical membrane-polarization model to predict the complex conductivity signature of immiscible liquid hydrocarbon contaminants. *Near Surface Geophysics*, 15(6), 547-562.

- [19-21.](#) Bückner, M., Orozco, A. F., & Kemna, A. (2018). Electrochemical polarization around metallic particles—Part 1: The role of diffuse-layer and volume-diffusion relaxation. *Geophysics*, 83(4), E203-E217.
- [20-22.](#) Bückner, M., Undorf, S., Flores Orozco, A., & Kemna, A. (2019). Electrochemical polarization around metallic particles—Part 2: The role of diffuse surface charge. *Geophysics*, 84(2), E57-E73.
- [21-23.](#) Canfield, D. E. (1989). Reactive iron in marine sediments. *Geochimica et Cosmochimica Acta*, 53(3), 619-632.
- [22-24.](#) Canfield, D. E., Raiswell, R., Westrich, J. T., Reaves, C. M., & Berner, R. A. (1986). The use of chromium reduction in the analysis of reduced inorganic sulfur in sediments and shales. *Chemical geology*, 54(1-2), 149-155.
- [23-25.](#) Capps, K. A., & Flecker, A. S. (2013). Invasive fishes generate biogeochemical hotspots in a nutrient-limited system. *PLoS One*, 8(1), e54093.
- [24-26.](#) Cirno, C. P., & McDonnell, J. J. (1997). Linking the hydrologic and biogeochemical controls of nitrogen transport in near-stream zones of temperate-forested catchments: a review. *Journal of Hydrology*, 199(1-2), 88-120.
- [25-27.](#) Cornell, R. M., & Schwertmann, U. (1996). *The Iron Oxides*, VCH.
- [26-28.](#) Costanza, R., d'Arge, R., De Groot, R., Farber, S., Grasso, M., Hannon, B., ... & Raskin, R. G. (1997). The value of the world's ecosystem services and natural capital. *nature*, 387(6630), 253-260.
- [27-29.](#) Costanza, R., De Groot, R., Braat, L., Kubiszewski, I., Fioramonti, L., Sutton, P., ... & Grasso, M. (2017). Twenty years of ecosystem services: how far have we come and how far do we still need to go?. *Ecosystem services*, 28, 1-16.
- [28-30.](#) Davis, C. A., Atekwana, E., Atekwana, E., Slater, L. D., Rossbach, S., & Mormile, M. R. (2006). Microbial growth and biofilm formation in geologic media is



detected with complex conductivity measurements. *Geophysical Research Letters*,  
33(18).

[29-31.](#) Diamond, J. S., McLaughlin, D. L., Slesak, R. A., & Stovall, A. (2020).  
Microtopography is a fundamental organizing structure of vegetation and soil chemistry  
in black ash wetlands.

[30-32.](#) Durejka, S., Gilfedder, B. S., & Frei, S. (2019). A method for long-term high  
resolution 222Radon measurements using a new hydrophobic capillary membrane  
system. *Journal of environmental radioactivity*, 208, 105980.

[31-33.](#) Elifantz, H., Kautsky, L., Mor-Yosef, M., Tarchitzky, J., Bar-Tal, A., Chen, Y.,  
& Minz, D. (2011). Microbial activity and organic matter dynamics during 4 years of  
irrigation with treated wastewater. *Microbial ecology*, 62(4), 973-981.

[32-34.](#) Estop-Aragonés, C., Knorr, K. H., & Blodau, C. (2012). Controls on in situ  
oxygen and dissolved inorganic carbon dynamics in peats of a temperate fen. *Journal  
of Geophysical Research: Biogeosciences*, 117(G2).

[33-35.](#) Estop-Aragonés, C., Knorr, K. H., & Blodau, C. (2013). Belowground in situ  
redox dynamics and methanogenesis recovery in a degraded fen during dry-wet cycles  
and flooding. *Biogeosciences*, 10(1), 421-436.

[34-36.](#) Fenner, N., Ostle, N., Freeman, C., Sleep, D., & Reynolds, B. (2004). Peatland  
carbon efflux partitioning reveals that Sphagnum photosynthate contributes to the DOC  
pool. *Plant and Soil*, 259(1-2), 345-354.

[35-37.](#) Flores Orozco, A., Williams, K.H., Long, P.E., Hubbard, S.S. and Kemna, A.,  
2011. Using complex resistivity imaging to infer biogeochemical processes associated  
with bioremediation of an uranium-contaminated aquifer. *Journal of Geophysical  
Research: Biogeosciences*, 116(G3).

[36-38.](#) Flores Orozco, A., Kemna, A., & Zimmermann, E. (2012a). Data error  
quantification in spectral induced polarization imaging. *Geophysics*, 77(3), E227-E237.

- 37:39. Flores Orozco, A., Kemna, A., Oberdörster, C., Zschornack, L., Leven, C., Dietrich, P., & Weiss, H. (2012b). Delineation of subsurface hydrocarbon contamination at a former hydrogenation plant using spectral induced polarization imaging. *Journal of contaminant hydrology*, 136, 131-144.
- 38:40. Flores Orozco, A., Williams, K. H., & Kemna, A. (2013). Time-lapse spectral induced polarization imaging of stimulated uranium bioremediation. *Near Surface Geophysics*, 11(5), 531-544.
- 39:41. Flores Orozco, A., Velimirovic, M., Tosco, T., Kemna, A., Sapion, H., Klaas, N., ... & Bastiaens, L. (2015). Monitoring the injection of microscale zerovalent iron particles for groundwater remediation by means of complex electrical conductivity imaging. *Environmental Science & Technology*, 49(9), 5593-5600.
- 40:42. Flores Orozco, A., Kemna, A., Binley, A., & Cassiani, G. (2019). Analysis of time-lapse data error in complex conductivity imaging to alleviate anthropogenic noise for site characterization. *Geophysics*, 84(2), B181-B193.
- 41:43. Flores Orozco, A., Gallistl, J., Steiner, M., Brandstätter, C., & Fellner, J. (2020). Mapping biogeochemically active zones in landfills with induced polarization imaging: The Heferlbach landfill. *Waste Management*, 107, 121-132.
44. Flores Orozco, A., Aigner, L. and Gallistl, J., 2021. Investigation of cable effects in spectral induced polarization imaging at the field scale using multicore and coaxial cables. *Geophysics*, 86(1), pp.E59-E75.
- 42:45. Frei, S., Lischeid, G., & Fleckenstein, J. H. (2010). Effects of micro-topography on surface–subsurface exchange and runoff generation in a virtual riparian wetland—A modeling study. *Advances in Water Resources*, 33(11), 1388-1401.

- 43-46. Frei, S., Knorr, K. H., Peiffer, S., & Fleckenstein, J. H. (2012). Surface micro-topography causes hot spots of biogeochemical activity in wetland systems: A virtual modeling experiment. *Journal of Geophysical Research: Biogeosciences*, 117(G4).
47. Garcia-Artigas, R., Himi, M., Revil, A., Urruela, A., Lovera, R., Sendrós, A., ... & Rivero, L. (2020). Time-domain induced polarization as a tool to image clogging in treatment wetlands. *Science of The Total Environment*, 724, 138189.
- 44-48. deGroot-Hedlin, C., & Constable, S. (1990). ~~Occam's~~Occam's inversion to generate smooth, two-dimensional models from magnetotelluric data. *Geophysics*, 55(12), 1613-1624.
- 45-49. Gu, B., Liang, L., Dickey, M. J., Yin, X., & Dai, S. (1998). Reductive precipitation of uranium (VI) by zero-valent iron. *Environmental Science & Technology*, 32(21), 3366-3373.
- 46-50. Gutknecht, J. L., Goodman, R. M., & Balser, T. C. (2006). Linking soil process and microbial ecology in freshwater wetland ecosystems. *Plant and Soil*, 289(1-2), 17-34.
- 47-51. Hansen, D. J., McGuire, J. T., Mohanty, B. P., & Ziegler, B. A. (2014). Evidence of aqueous iron sulfid clusters in the vadose zone. *Vadose Zone Journal*, 13(3), 1-12.
- 48-52. Hartley, A. E., & Schlesinger, W. H. (2000). Environmental controls on nitric oxide emission from northern Chihuahuan desert soils. *Biogeochemistry*, 50(3), 279-300.
- 49-53. Hayati, A. A., & Proctor, M. C. F. (1991). Limiting nutrients in acid-mire vegetation: peat and plant analyses and experiments on plant responses to added nutrients. *The Journal of Ecology*, 75-95.
- 50-54. Hördt, A., Bairlein, K., Bielefeld, A., Bucker, M., Kuhn, E., Nordsiek, S. and Stebner, H., 2016. The dependence of induced polarization on fluid salinity and pH,

studied with an extended model of membrane polarization. *Journal of Applied Geophysics*, 135, pp.408-417.

[51-55.](#) Kang, H., Kwon, M. J., Kim, S., Lee, S., Jones, T. G., Johncock, A. C., ... & Freeman, C. (2018). Biologically driven DOC release from peatlands during recovery from acidification. *Nature communications*, 9(1), 1-7.

[52-56.](#) Kayranli, B., Scholz, M., Mustafa, A., & Hedmark, Å. (2010). Carbon storage and fluxes within freshwater wetlands: a critical review. *Wetlands*, 30(1), 111-124.

[53-57.](#) Kemna, A., 2000. Tomographic Inversion of Complex Resistivity: Theory and Application. Der Andere Verlag Osnabrück, Germany

[54-58.](#) Kemna, A., Binley, A., Ramirez, A., & Daily, W. (2000). Complex resistivity tomography for environmental applications. *Chemical Engineering Journal*, 77(1-2), 11-18.

[55-59.](#) Kemna, A., Vanderborght, J., Kulesa, B., & Vereecken, H. (2002). Imaging and characterisation of subsurface solute transport using electrical resistivity tomography (ERT) and equivalent transport models. *Journal of Hydrology*, 267(3-4), 125-146.

[56-60.](#) Kemna, A., Binley, A., & Slater, L. (2004). Crosshole IP imaging for engineering and environmental applications. *Geophysics*, 69(1), 97-107.

[57-61.](#) Kemna, A., Binley, A., Cassiani, G., Niederleithinger, E., Revil, A., Slater, L., ... & Kruschwitz, S. (2012). An overview of the spectral induced polarization method for near-surface applications. *Near Surface Geophysics*, 10(6), 453-468.

[58-62.](#) Kessouri, P., Furman, A., Huisman, J. A., Martin, T., Mellage, A., Ntarlagiannis, D., ... & Kemna, A. (2019). Induced polarization applied to biogeophysics: recent advances and future prospects. *Near Surface Geophysics*, 17(6-Recent Developments in Induced Polarization), 595-621.

63. [Kleinebecker, T., Hölzel, N., & Vogel, A. \(2008\). South Patagonian ombrotrophic bog vegetation reflects biogeochemical gradients at the landscape level. \*Journal of vegetation science\*, 19\(2\), 151-160.](#)
64. [Kosmulski, M., Maczka, E., Jartych, E., & Rosenholm, J. B. \(2003\). Synthesis and characterization of goethite and goethite–hematite composite: experimental study and literature survey. \*Advances in colloid and interface science\*, 103\(1\), 57-76.](#)
- ~~59~~65. [LaBrecque, D. J., Miletto, M., Daily, W., Ramirez, A., & Owen, E. \(1996\). The effects of noise on ~~Occam's~~ inversion of resistivity tomography data. \*Geophysics\*, 61\(2\), 538-548.](#)
- ~~60~~66. [Leroy, P., Revil, A., Kemna, A., Cosenza, P., & Ghorbani, A. \(2008\). Complex conductivity of water-saturated packs of glass beads. \*Journal of colloid and interface science\*, 321\(1\), 103-117.](#)
67. [Lesmes, D. P., & Frye, K. M. \(2001\). Influence of pore fluid chemistry on the complex conductivity and induced polarization responses of Berea sandstone. \*Journal of Geophysical Research: Solid Earth\*, 106\(B3\), 4079-4090.](#)
68. [Linke, T., & Gislason, S. R. \(2018\). Stability of iron minerals in Icelandic peat areas and transport of heavy metals and nutrients across oxidation and salinity gradients—a modelling approach. \*Energy Procedia\*, 146, 30-37.](#)
69. [Lischeid, G., Kolb, A., & Alewell, C. \(2002\). Apparent translatory flow in groundwater recharge and runoff generation. \*Journal of Hydrology\*, 265\(1-4\), 195-211.](#)
- ~~64~~70. [Liu, H. \(2013\). Thermal response of soil microbial respiration is positively associated with labile carbon content and soil microbial activity. \*Geoderma\*, 193, 275-281.](#)
- ~~62~~71. [Mansoor, N., & Slater, L. \(2007\). On the relationship between iron concentration and induced polarization in marsh soils. \*Geophysics\*, 72\(1\), A1-A5.](#)

- 63-72. Marshall, D. J., & Madden, T. R. (1959). Induced polarization, a study of its causes. *Geophysics*, 24(4), 790-816.
- 64-73. Maurya, P. K., Rønde, V. K., Fiandaca, G., Balbarini, N., Auken, E., Bjerg, P. L., & Christiansen, A. V. (2017). Detailed landfill leachate plume mapping using 2D and 3D electrical resistivity tomography-with correlation to ionic strength measured in screens. *Journal of Applied Geophysics*, 138, 1-8.
- 65-74. McAnallen, L., Doherty, R., Donohue, S., Kirmizakis, P., & Mendonça, C. (2018). Combined use of geophysical and geochemical methods to assess areas of active, degrading and restored blanket bog. *Science of the Total Environment*, 621, 762-771.
- 66-75. McClain, M. E., Boyer, E. W., Dent, C. L., Gergel, S. E., Grimm, N. B., Groffman, P. M., ... & McDowell, W. H. (2003). Biogeochemical hot spots and hot moments at the interface of terrestrial and aquatic ecosystems. *Ecosystems*, 301-312.
- 67-76. Mellage, A., Smeaton, C. M., Furman, A., Atekwana, E. A., Rezanezhad, F., & Van Cappellen, P. (2018). Linking spectral induced polarization (SIP) and subsurface microbial processes: Results from sand column incubation experiments. *Environmental science & technology*, 52(4), 2081-2090.
- 68-77. Mishra, U., & Riley, W. J. (2015). Scaling impacts on environmental controls and spatial heterogeneity of soil organic carbon stocks. *Biogeosciences Discussions (Online)*, 12(2).
78. Misra, S., Torres-Verdín, C., Revil, A., Rasmus, J., & Homan, D. (2016a). Interfacial polarization of disseminated conductive minerals in absence of redox-active species—Part 1: Mechanistic model and validation. *Geophysics*, 81(2), E139-E157.
79. Misra, S., Torres-Verdín, C., Revil, A., Rasmus, J., & Homan, D. (2016b). Interfacial polarization of disseminated conductive minerals in absence of redox-active species—Part 2: Effective electrical conductivity and dielectric permittivity Interfacial polarization due to inclusions. *Geophysics*, 81(2), E159-E176.



- 69.80. Morse, J. L., Werner, S. F., Gillin, C. P., Goodale, C. L., Bailey, S. W., McGuire, K. J., & Groffman, P. M. (2014). Searching for biogeochemical hot spots in three dimensions: Soil C and N cycling in hydopedologic settings in a northern hardwood forest. *Journal of Geophysical Research: Biogeosciences*, 119(8), 1596-1607.
- 70.81. Ntarlagiannis, D., Williams, K. H., Slater, L., & Hubbard, S. (2005). Low-frequency electrical response to microbial induced sulfid precipitation. *Journal of Geophysical Research: Biogeosciences*, 110(G2).
- 71.82. Ntarlagiannis, D., Doherty, R. and Williams, K.H., 2010. Spectral induced polarization signatures of abiotic FeS precipitation SIP signatures of FeS precipitation. *Geophysics*, 75(4), pp.F127-F133.
- 72.83. Parikh, S. J., & Chorover, J. (2006). ATR-FTIR spectroscopy reveals bond formation during bacterial adhesion to iron oxide. *Langmuir*, 22(20), 8492-8500.
- 73.84. Parry, L. E., West, L. J., Holden, J., & Chapman, P. J. (2014). Evaluating approaches for estimating peat depth. *Journal of Geophysical Research: Biogeosciences*, 119(4), 567-576.
85. Partington, D., Brunner, P., Frei, S., Simmons, C. T., Werner, A. D., Therrien, R., ... & Fleckenstein, J. H. (2013). Interpreting streamflow generation mechanisms from integrated surface-subsurface flow models of a riparian wetland and catchment. *Water Resources Research*, 49(9), 5501-5519.
- 74.86. Pelton, W. H., Ward, S. H., Hallof, P. G., Sill, W. R., & Nelson, P. H. (1978). Mineral discrimination and removal of inductive coupling with multifrequency IP. *Geophysics*, 43(3), 588-609.
- 75.87. Personna, Y. R., Ntarlagiannis, D., Slater, L., Yee, N., O'Brien, M., & Hubbard, S. (2008). Spectral induced polarization and electrodic potential monitoring of microbially mediated iron sulfid transformations. *Journal of Geophysical Research: Biogeosciences*, 113(G2).

- 76-88. Placencia-Gómez, E., Slater, L., Ntarlagiannis, D., & Binley, A. (2013). Laboratory SIP signatures associated with oxidation of disseminated metal sulfids. *Journal of Contaminant Hydrology*, 148, 25-38.
- 77-89. Ponziani, M., Slob, E. C., Ngan-Tillard, D. J. M., & Vanhala, H. (2011). Influence of water content on the electrical conductivity of peat. *International Water Technology Journal*, 1(1), 14-21.
90. Qi, Y., Soueid Ahmed, A., Revil, A., Ghorbani, A., Abdulsamad, F., Florsch, N., & Bonnenfant, J. (2018). Induced polarization response of porous media with metallic particles—Part 7: Detection and quantification of buried slag heaps. *Geophysics*, 83(5), E277-E291.
- 78-91. Revil, A. (2012). Spectral induced polarization of shaly sands: Influence of the electrical double layer. *Water Resources Research*, 48(2).
- 79-92. Revil, A., & Florsch, N. (2010). Determination of permeability from spectral induced polarization in granular media. *Geophysical Journal International*, 181(3), 1480-1498.
- 80-93. Revil, A., & Skold, M. (2011). Salinity dependence of spectral induced polarization in sands and sandstones. *Geophysical Journal International*, 187(2), 813-824.
- 81-94. Revil, A., Atekwana, E., Zhang, C., Jardani, A., & Smith, S. (2012). A new model for the spectral induced polarization signature of bacterial growth in porous media. *Water Resources Research*, 48(9).
- 82-95. Revil, A., Florsch, N., & Mao, D. (2015a). Induced polarization response of porous media with metallic particles—Part 1: A theory for disseminated semiconductors. *Geophysics*, 80(5), D525-D538.

- 83-96. Revil, A., Abdel Aal, G. Z., Atekwana, E. A., Mao, D., & Florsch, N. (2015b). Induced polarization response of porous media with metallic particles—Part 2: Comparison with a broad database of experimental data. *Geophysics*, 80(5), D539-D552.
- 84-97. Revil, A., Coperey, A., Shao, Z., Florsch, N., Fabricius, I. L., Deng, Y., ... & van Baaren, E. S. (2017a). Complex conductivity of soils. *Water Resources Research*, 53(8), 7121-7147.
98. Revil, A., Sleevi, M. F., & Mao, D. (2017b). Induced polarization response of porous media with metallic particles—Part 5: Influence of the background polarization. *Geophysics*, 82(2), E77-E96.
99. Revil, A., Mao, D., Shao, Z., Sleevi, M. F., & Wang, D. (2017c). Induced polarization response of porous media with metallic particles—Part 6: The case of metals and semimetals. *Geophysics*, 82(2), E97-E110.
100. Revil, A., Coperey, A., Mao, D., Abdulsamad, F., Ghorbani, A., Rossi, M., & Gasquet, D. (2018). Induced polarization response of porous media with metallic particles—Part 8: Influence of temperature and salinity. *Geophysics*, 83(6), E435-E456.
- 85-101. Rosier, C. L., Atekwana, E. A., Aal, G. A., & Patrauchan, M. A. (2019). Cell concentrations and metabolites enhance the SIP response to biofilm matrix components. *Journal of Applied Geophysics*, 160, 183-194.
- 86-102. Schurr, J. M. (1964). On the theory of the dielectric dispersion of spherical colloidal particles in electrolyte solution<sup>1</sup>. *The Journal of Physical Chemistry*, 68(9), 2407-2413.
- 87-103. Schwartz, N., & Furman, A. (2014). On the spectral induced polarization signature of soil organic matter. *Geophysical Journal International*, 200(1), 589-595.

~~88. Schwartz, N., Shalem, T., & Furman, A. (2014). The effect of organic acid on the spectral induced polarization response of soil. *Geophysical Journal International*, 197(1), 269-276.~~

~~89.104.~~ Schwarz, G. (1962). A theory of the low-frequency dielectric dispersion of colloidal particles in electrolyte solution1, 2. *The Journal of Physical Chemistry*, 66(12), 2636-2642.

~~90.105.~~ Seigel, H., Nabighian, M., Parasnis, D. S., & Vozoff, K. (2007). The early history of the induced polarization method. *The Leading Edge*, 26(3), 312-321.

~~106.~~ Skold, M., Revil, A., & Vaudelet, P. (2011). The pH dependence of spectral induced polarization of silica sands: Experiment and modeling. *Geophysical Research Letters*, 38(12).

~~91.107.~~ Slater, L., & Atekwana, E. (2013). Geophysical signatures of subsurface microbial processes. *Eos, Transactions American Geophysical Union*, 94(8), 77-78.

~~92.108.~~ Slater, L., & Binley, A. (2006). Synthetic and field-based electrical imaging of a zerovalent iron barrier: Implications for monitoring long-term barrier performance. *Geophysics*, 71(5), B129-B137.

~~93.109.~~ Slater LD, & Reeve A (2002) Investigating peatland stratigraphy and hydrogeology using integrated electrical geophysics. *Geophysics* 67(2):365–378

~~94.110.~~ Slater, L., Binley, A. M., Daily, W., & Johnson, R. (2000). Cross-hole electrical imaging of a controlled saline tracer injection. *Journal of applied geophysics*, 44(2-3), 85-102.

~~95.111.~~ Slater, L., Ntarlagiannis, D., Personna, Y. R., & Hubbard, S. (2007). Pore-scale spectral induced polarization signatures associated with FeS biomineral transformations. *Geophysical Research Letters*, 34(21).

~~96.~~112. Strohmeier, S., Knorr, K. H., Reichert, M., Frei, S., Fleckenstein, J. H., Peiffer, S., & Matzner, E. (2013). Concentrations and fluxes of dissolved organic carbon in runoff from a forested catchment: insights from high frequency measurements. *Biogeosciences*, 10(2), 905.

~~97.~~113. Tamura, H., Goto, K., Yotsuyanagi, T., & Nagayama, M. (1974). Spectrophotometric determination of iron (II) with 1, 10-phenanthroline in the presence of large amounts of iron (III). *Talanta*, 21(4), 314-318.

~~98. Trappe, J., & Kneisel, C. (2019). Geophysical and sedimentological investigations of Peatlands for the assessment of lithology and subsurface water pathways. *Geosciences*, 9(3), 118.~~

~~99.~~114. Tsukanov, K., & Schwartz, N. (2020). Relationship between wheat root properties and its electrical signature using the spectral induced polarization method. *Vadose Zone Journal*, 19(1), e20014.

~~100.~~115. Uhlemann, S.S., Sorensen, J.P.R., House, A.R., Wilkinson, P.B., Roberts, C., Gooddy, D.C., Binley, A.M. and Chambers, J.E., 2016. Integrated time-lapse geoelectrical imaging of wetland hydrological processes. *Water Resources Research*, 52(3), pp.1607-1625.

~~101.~~116. Ulrich, C., & Slater, L. (2004). Induced polarization measurements on unsaturated, unconsolidated sands. *Geophysics*, 69(3), 762-771.

~~102.~~117. Urban, N. R. Retention of sulfur in lake-sediments. in *Environmental Chemistry of Lakes and Reservoirs* (ed. Baker, L. A.) (American Chemical Society, 1994). 237, 323–369

118. Vindedahl, A. M., Strehlau, J. H., Arnold, W. A., & Penn, R. L. (2016). Organic matter and iron oxide nanoparticles: aggregation, interactions, and reactivity. *Environmental Science: Nano*, 3(3), 494-505.

- ~~403~~119. Wainwright, H. M., Flores Orozco, A., Bucker, M., Dafflon, B., Chen, J., Hubbard, S. S., & Williams, K. H. (2016). Hierarchical Bayesian method for mapping biogeochemical hot spots using induced polarization imaging. *Water Resources Research*, 52(1), 533-551.
- ~~404~~120. Wang, Y., Wang, H., He, J. S., & Feng, X. (2017). Iron-mediated soil carbon response to water-table decline in an alpine wetland. *Nature communications*, 8(1), 1-9.
- ~~405~~121. Ward, S. H. (1988, January). The resistivity and induced polarization methods. In *Symposium on the Application of Geophysics to Engineering and Environmental Problems 1988* (pp. 109-250). Society of Exploration Geophysicists.
- ~~406~~122. Waxman, M. H., & Smits, L. J. M. (1968). Electrical conductivities in oil-bearing shaly sands. *Society of Petroleum Engineers Journal*, 8(02), 107-122.
- ~~407~~123. Weigand, M., & Kemna, A. (2017). Multi-frequency electrical impedance tomography as a non-invasive tool to characterize and monitor crop root systems. *Biogeosciences*, 14(4).
- ~~408~~124. Weller, A., Zhang, Z. and Slater, L., 2015. High-salinity polarization of sandstones. *Geophysics*, 80(3), pp.D309-D318.
- ~~409~~125. Williams, K. H., Ntarlagiannis, D., Slater, L. D., Dohnalkova, A., Hubbard, S. S., & Banfield, J. F. (2005). Geophysical imaging of stimulated microbial biomineralization. *Environmental science & technology*, 39(19), 7592-7600.
- ~~410~~126. Williams, K. H., Kemna, A., Wilkins, M. J., Druhan, J., Arntzen, E., ~~N'Guessan~~N'Guessan, A. L., ... & Banfield, J. F. (2009). Geophysical monitoring of coupled microbial and geochemical processes during stimulated subsurface bioremediation. *Environmental science & technology*, 43(17), 6717-6723.



1282 [111:127.](#) Wong, J. (1979). An electrochemical model of the induced-polarization  
 1283 phenomenon in disseminated sulfid ores. *Geophysics*, 44(7), 1245-1265.  
 1284

1285 [112:128.](#) Zhang, C., Ntarlagiannis, D., Slater, L., & Doherty, R. (2010). Monitoring  
 1286 microbial sulfate reduction in porous media using multipurpose electrodes. *Journal of*  
 1287 *Geophysical Research: Biogeosciences*, 115(G3).  
 1288

1289 [113:129.](#) Zhang, C., Slater, L., & Prodan, C. (2013). Complex dielectric properties of  
 1290 sulfate-reducing bacteria suspensions. *Geomicrobiology Journal*, 30(6), 490-496.  
 1291

1292 [114:130.](#) Zimmermann, E., Kemna, A., Berwix, J., Glaas, W., & Vereecken, H. (2008).  
 1293 EIT measurement system with high phase accuracy for the imaging of spectral induced  
 1294 polarization properties of soils and sediments. *Measurement Science and Technology*,  
 1295 19(9), 094010.  
 1296

1297 [115:131.](#) Zimmermann, E., Huisman, J. A., Mester, A., & van Waasen, S. (2019).  
 1298 Correction of phase errors due to leakage currents in wideband EIT field measurements  
 1299 on soil and sediments. *Measurement Science and Technology*, 30(8), 084002.  
 1300