



An investigation of grazing behaviors that result in winter phytoplankton biomass accumulation

Mara Freilich¹, Alexandre Mignot², Glenn Flierl³, and Raffaele Ferrari³

¹MIT-WHOI Joint Program in Oceanography & Applied Ocean Science and Engineering, Cambridge, MA, USA

²Mercator Ocean International, Ramonville-Saint-Agne, France

³Department of Earth Atmospheric and Planetary Science, Massachusetts Institute of Technology, Cambridge, MA

Correspondence: Mara Freilich (maraf@mit.edu)

Abstract. Recent observations have shown that phytoplankton biomass increases in the North Atlantic during winter, even when the mixed layer is deepening and light is limited. Current theories suggest that this is due to a release from grazing pressure. Here we demonstrate that the often-used grazing models that are linear at low phytoplankton concentration do not allow for a wintertime increase in phytoplankton biomass. However, certain mathematical formulations of grazing that are quadratic (or more generally non-linear) in phytoplankton concentration at low concentrations can reproduce the fall to spring transition in phytoplankton, including wintertime biomass accumulation. We illustrate this point with a minimal model for the annual cycle of North Atlantic phytoplankton designed to simulate phytoplankton concentration as observed by BioGeoChemical-Argo (BGC-Argo) floats in the North Atlantic. This analysis provides a mathematical framework for assessing hypotheses of phytoplankton bloom formation.

10 1 Introduction

One of the most prominent biological events in the surface ocean is the North Atlantic spring bloom (Boss *et al.*, 2008; Siegel *et al.*, 2014; Cole *et al.*, 2015). Each spring, in an event that is distinctive in satellite ocean color observations (Siegel *et al.*, 2014), there is a rapid accumulation of phytoplankton in the ocean surface layer across the North Atlantic. A bloom occurs when the phytoplankton growth rates are sufficiently faster than the loss rates over a sustained time period (Sverdrup, 1953). The large annual cycle in the phytoplankton population in the North Atlantic occurs in the context of large seasonal cycles in atmospheric conditions that drive changes in mixed layer depth, surface irradiance, and upper layer temperature. How these environmental factors interact with ecological processes to produce a bloom is still being debated (Fischer *et al.*, 2014).

The traditional theory of phytoplankton population dynamics in the North Atlantic attributes the spring bloom to the release of phytoplankton from light limitation, which causes phytoplankton growth rates to increase. This has become known as the “critical depth hypothesis” (Sverdrup, 1953) because the theory states that phytoplankton can begin to grow when the mixed layer has shoaled sufficiently so that the light-dependent phytoplankton growth terms are larger than the phytoplankton loss terms, which are assumed to be constant throughout the winter and into the spring. Other hypotheses have demonstrated that the release of phytoplankton from light limitation can occur even while the mixed layer is being cooled due to a reduction in turbulence at fronts (Taylor and Ferrari, 2011; Mahadevan *et al.*, 2012).



25 An alternative hypothesis proposed by *Behrenfeld* (2010) focuses on changes in loss rates rather than growth rates. The
“disturbance-recovery hypothesis” states that even though phytoplankton growth rates are very low in the wintertime, due
primarily to light limitation, loss rates decrease even faster as the mixed layer deepens due to decreasing phytoplankton-
zooplankton encounter rates. This hypothesis was formulated as an explanation of recent observations of increasing phyto-
plankton stocks in the wintertime (*Behrenfeld*, 2010; *Boss and Behrenfeld*, 2010). Wintertime biomass accumulation is in-
30 consistent with the critical depth hypothesis, which assumes that the winter growth rates are smaller than the constant loss
rates.

The critical depth hypothesis and the disturbance-recovery hypothesis differ in their predictions of the evolution of winter
loss rates. Process-level understanding and quantification of phytoplankton population loss rates is challenging, because it
is very difficult to directly measure the factors that contribute to loss at large scales. Phytoplankton loss is due primarily
35 to grazing by zooplankton or mortality due to viral lysis and secondarily to respiration and natural mortality (*Landry and
Calbet*, 2004). Loss due to grazing depends on both the concentration of phytoplankton and zooplankton populations and to
the interactions between them. Autonomous measurements from satellites and floats have made quantification of phytoplankton
biomass possible over large spatial and temporal scales (*Siegel et al.*, 2002; *Boss et al.*, 2008; *Mignot et al.*, 2018). No such
equivalent measurements exist for zooplankton populations.

40 The interactions between phytoplankton and zooplankton are quantified through mathematical relationships which express
the rate of phytoplankton consumption by zooplankton as a function of phytoplankton concentration. There are many functional
responses that are supported by experiments and theory and that have been used to represent grazing in numerical simulations
and to interpret observations (*Gentleman et al.*, 2003; *Laufkötter et al.*, 2015). The most commonly used functional responses
increase linearly or quadratically and saturate to a constant rate at high concentrations (*Gentleman et al.*, 2003).

45 During the spring bloom, phytoplankton accumulation is exponential due to the rapid increase in growth rates that makes
loss processes relatively much smaller. In the wintertime, the observed phytoplankton accumulation is slower and leading
hypotheses of phytoplankton bloom formation differ in their predictions both of phytoplankton population dynamics and of
phytoplankton loss rates. Comparing phytoplankton-zooplankton models with different representations of grazing against the
observations of biomass accumulation during sub-optimal growth conditions, such as during the wintertime, may constrain
50 the range of appropriate grazing functions for winter conditions or even the winter-spring transition. Here, we demonstrate
that the disturbance-recovery hypothesis requires a grazing function that decreases more rapidly than linearly at low prey
concentrations. We show that a model with a quadratic grazing function at low winter phytoplankton concentrations captures
the full annual cycle of phytoplankton biomass in the North Atlantic, i.e. both weak wintertime biomass accumulation and an
explosive springtime bloom. Our aim is to provide empirically motivated guidance for the formulation and testing of grazing
55 models.



2 Predator-prey decoupling

In this section we formulate a simple ecosystem model and examine different grazing functions to clarify the relationship between grazing rates and mixed layer depth during winter conditions (Figure 1).

Marine planktonic ecosystem dynamics can be coarsely represented as an interaction between three compartments: nutrients, phytoplankton, and zooplankton. These broad compartments integrate across all the chemical and biological diversity observed in the ocean and are defined by their interactions with each other. In the simple formulation adopted here, the nutrients are consumed by phytoplankton, the zooplankton consume phytoplankton, and the plankton are converted back to nutrients when they die. The set of equations that describe these interactions for the concentrations of nutrients (n), phytoplankton (p), and zooplankton (z) as a function of the ocean depth ζ take the form:

$$\begin{aligned}
 \frac{Dn}{Dt} &= -\mu(n,t)e^{K_d\zeta}p + d_p p + (1-a)g(p)z + d_z z^2 + \frac{\partial}{\partial \zeta} \kappa \frac{\partial n}{\partial \zeta}, \\
 \frac{Dp}{Dt} &= \mu(n,t)e^{K_d\zeta}p - g(p)z - d_p p + \frac{\partial}{\partial \zeta} \kappa \frac{\partial p}{\partial \zeta}, \\
 \frac{Dz}{Dt} &= a g(p)z - d_z z^2 + \frac{\partial}{\partial \zeta} \kappa \frac{\partial z}{\partial \zeta}.
 \end{aligned} \tag{1}$$

The three compartments are modelled in terms of their carbon content and thus have the same units. The vertical coordinate, ζ , is zero at ocean surface and negative below. All compartments are mixed in the vertical by ocean turbulence at a rate set by the diffusivity κ . The phytoplankton specific growth rate depends on nutrients, according to the function $\mu(n)$, and decays exponentially with depth due to the absorption of light with depth with an attenuation coefficient K_d . Phytoplankton mortality, $-d_p p$, is linear in p . Zooplankton mortality, $-d_z z^2$, is quadratic in z to account for the drop in grazing by higher trophic levels when food is scarce; this choice has the additional property of preventing extinction of zooplankton in winter. The grazing of phytoplankton by zooplankton is linear in z and proportional to p according to the grazing function $g(p)$. Zooplankton are messy eaters and ingest only a fraction $a < 1$ of $g(p)z$.

We will focus on analyzing the model in equation 1 during the wintertime period through the bloom onset. During this period, we can make a few simplifying assumptions. First, we will assume that turbulence is strong enough to keep all compartments well mixed in the vertical over a mixed layer of depth H . This assumption holds if the turbulence mixes all compartments throughout H on a timescale faster than any biological timescale (*Taylor and Ferrari, 2011*). Second, we will assume that winter growth is not nutrient limited ($n \gg n_0$) and thus $\mu(n)$ saturates to a constant mixed layer-averaged value μ_0 (we will not make this assumption about a time and nutrient independent growth rate in section 3). For simplicity, we also assume that nutrients are exactly conserved in the mixed layer and model the nutrient concentration implicitly as $n = N_{max} - p - z$, where N_{max} is the total carbon content which gets redistributed across the three compartments. Finally, we assume that the mixed layer is deep relative to the depth of light penetration ($H K_d \gg 1$). All these assumptions are appropriate in winter, the focus of our study, but they are less defensible in other seasons when turbulence is weak (*Taylor and Ferrari, 2011*).

We formulate a bulk mixed layer model by employing these simplifying assumptions and taking the vertical average of the equations in (1) over the mixed layer depth $H(t)$,

$$\begin{aligned}
 \frac{\partial p}{\partial t} &= \left(\frac{1}{K_d H(t)} \mu_0 - d_p \right) p - g(p)z - s^+ p \\
 \frac{\partial z}{\partial t} &= g(p)z - d_z z^2 - s^+ z,
 \end{aligned} \tag{2}$$

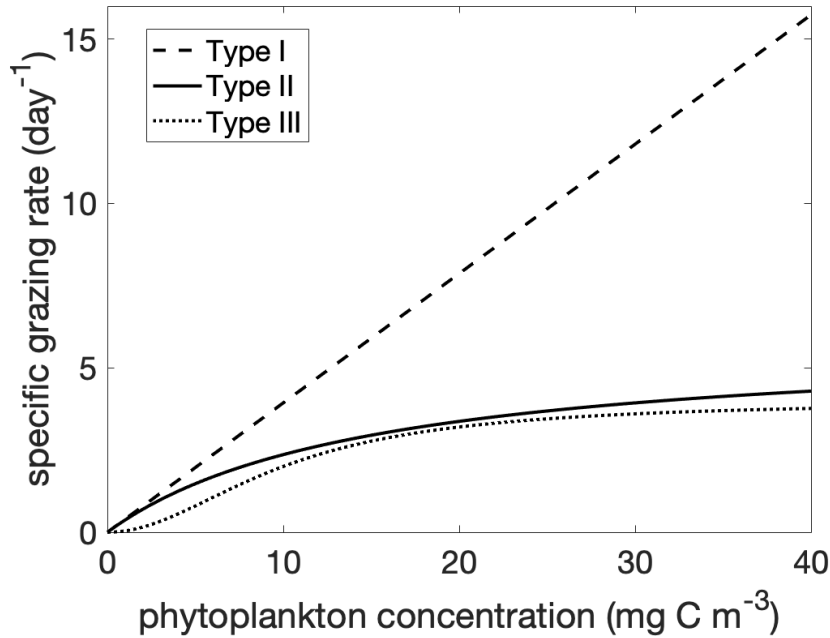


Figure 1. Grazing rate $g(p)$ as a function of phytoplankton concentration for Holling Type I, II, and III functional responses. The parameters g_0 and p_0 are given in Table 1 for the Holling type II and III functional responses. The forms of the Holling type II and III is as in equation 6. The form of the Holling type I is the linearized type II, $g_{H_1}(p) = g_0/p_0 p$.

where p and z are the constant mixed layer concentrations of phytoplankton and zooplankton, respectively. The term $\frac{1}{K_d H(t)}$ is the average light over the mixed layer, which is computed as the integral of the light level over the mixed layer depth divided by the mixed layer depth.

90 The term s^+ appears when taking the vertical average of the mixing term in equation (1). It represents the dilution of phytoplankton and zooplankton that results from the turbulent entrainment of water without biomass across the mixed layer base and is given by,

$$s^+ = \begin{cases} \frac{1}{H} \frac{dH}{dt} & \frac{dH}{dt} > 0 \\ 0 & \frac{dH}{dt} \leq 0 \end{cases} \quad (3)$$

We can derive an equation for the standing stock of biomass in the mixed layer by taking a vertical integral of the equations
 95 in (1). Introducing $P = Hp$ and $Z = Hz$ to represent the total biomass of phytoplankton and zooplankton respectively we have

$$\begin{aligned} \frac{\partial P}{\partial t} &= \left(\frac{1}{K_d H} \mu_0 - d_p \right) P - g(P/H)Z - s^- P \\ \frac{\partial Z}{\partial t} &= g(P/H)Z - \frac{1}{H} d_z Z^2 - s^- Z \end{aligned} \quad (4)$$



In contrast to the average concentration, the total biomass does not change due to the physical effects of dilution. However, when the mixed layer shoals, biomass is lost below the mixed layer and the total biomass decreases at a rate given by s^-

$$s^- = \begin{cases} 0 & \frac{dH}{dt} > 0 \\ \frac{dH}{dt} & \frac{dH}{dt} \leq 0 \end{cases} \quad (5)$$

In the following subsections we will analyze the phenology of phytoplankton for different choices of grazing functions (Figure 1). The linear (Holling type I) grazing function assumes that the plankton-specific grazing rate (units of per day) increases linearly with phytoplankton concentration, $g_{HI}(p) = g_0 p$. The saturating functional responses are linear at low prey concentration and saturate at high prey concentrations. An example saturating response is the Holling type II functional response, $g_{HII}(p) = g_0 \frac{p}{p_0 + p}$. This functional response assumes that processing of food and searching for food are mutually exclusive behaviors (Visser, 2007; Kiørboe et al., 2018). The parameter g_0 is a function of processing time and the parameter p_0 is a function of both search and processing time. This parsimonious theoretical basis and ability to fit the parameters from experimental data has made this functional response one of the most commonly used (Verity, 1991; Kiørboe et al., 2018). The Holling type III functional response has a reduction in grazing at low prey concentration. One formulation is a sigmoidal function, $g_{HIII}(p) = g_0 \frac{p^2}{p_0^2 + p^2}$ which is quadratic in p for low p . This type III functional response can be derived as a generalization of the type II response where the search time is a linear function of prey concentration. Effectively, there is a prey refuge at low concentration because it is more difficult for predators to find each prey item. There are other possible mechanisms for a type III functional response, including a threshold response by predators (Mullin et al., 1975; Ohman, 1984) and prey switching (Vallina et al., 2014). To compare the functional responses, we formulate the zooplankton specific grazing rate as a function of phytoplankton concentration

$$g(p) = g_0 \frac{(p/p_0)^{k-1}}{1 + (p/p_0)^{k-1}}. \quad (6)$$

The exponent k determines the degree of non-linearity of the functional response. The Holling type II functional response is $k = 2$ and the Holling type III response is $k = 3$. The parameter p_0 is a half saturation constant. When $p = p_0$, the grazing is at half of the maximum rate $g(p_0) = \frac{g_0}{2}$.

2.1 Grazing linear in phytoplankton concentration for constant zooplankton concentration: Sverdrup's model

Phytoplankton are known to respond faster than zooplankton to environmental changes (Fileman and Leakey, 2005). Sverdrup (1953) assumed that such an assumption applied to the rapid onset of the spring bloom and proposed to model the phytoplankton growth rate according to equation (2), but setting $g(p)z = \frac{g_0}{p_0} z_0 p$ with z_0 the constant zooplankton concentration before the bloom onset,

$$\frac{\partial p}{\partial t} = \left(\frac{1}{K_d H(t)} \mu_0 - d_p - g_0 z_0 / p_0 \right) p. \quad (7)$$

Sverdrup focused on the time at the end of winter when the mixed layer starts shoaling in response to spring atmospheric conditions and thus could ignore the entrainment, i.e. $s^+ = 0$. Under these assumptions the mixed layer depth $H(t)$ is the only



time dependent parameter which can determine whether the phytoplankton concentration is exponentially decaying (winter conditions) or exponentially increasing (spring bloom onset). This gave rise to the widely applied ‘critical depth hypothesis’ which states that phytoplankton accumulation starts when the mixed layer shoals beyond a critical depth,

$$H_c = \frac{\mu_0}{(d_p + g_0 z_0 / p_0) K_d}. \quad (8)$$

While the critical depth hypothesis has become the most widely accepted framework to interpret the onset of spring blooms—but there are growing objections *Behrenfeld* (2010)—it is not very useful to make quantitative predictions. The criterion requires knowledge of the grazing rate at the end of winter before bloom onset, which is very difficult to measure. Sometimes this obstacle is overcome by assuming that $g_0 z_0 \ll d_p$, in which case the critical depth dependence on grazing can be ignored. However the assumption is likely inappropriate for most blooms. For example, assuming a typical attenuation coefficient of $K_d = 0.05 \text{ m}^{-1}$ in the winter North Atlantic (*Organelli et al.*, 2017), where bloom onset is often observed at a critical depth of around 200 m (as reported in *Siegel et al.* (2002)), the ratio of growth to mortality rate, $\frac{d_p + g_0 z_0 / p_0}{\mu_0}$, is predicted to be close to be 0.1. Mortality timescales of phytoplankton are believed to be much longer than ten times their division rates implying that grazing, not mortality, dominates phytoplankton losses at bloom onset. A theory of blooms must therefore include a predictions of the zooplankton concentrations and their grazing rates at the end of winter, if it is to make falsifiable predictions. Additionally, on seasonal timescales there is substantial variation in zooplankton concentrations so a theory that includes variable phytoplankton and zooplankton concentrations is necessary. The goal of the next two sections is to present two models of grazing with a focus on wintertime conditions.

2.2 Grazing linear in phytoplankton concentration: $g(p) \sim p$

Consider first the saturating (type II) grazing function. In winter, prey concentrations are very low and this function is approximately linear $g_{HII}(p) \approx \frac{g_0}{p_0} p$ (Figure 2b). During the wintertime, as the mixed layer deepens, water from below the mixed layer is entrained, decreasing the concentration of the phytoplankton and zooplankton ($s^+ > 0$) but not their standing stock ($s^- = 0$).

$$\begin{aligned} \frac{\partial P}{\partial t} &= \frac{1}{H} \left(\frac{\mu_0}{K_d} - \frac{g_0}{p_0} Z \right) P - d_p P \\ \frac{\partial Z}{\partial t} &= \frac{1}{H} \left(a \frac{g_0}{p_0} P - d_z Z \right) Z \end{aligned} \quad (9)$$

Assuming the natural mortality of phytoplankton is negligibly small, the growth and grazing terms in the P and Z equations have the same dependence on mixed layer depth H and thus any increase in H does not reduce grazing any more than it reduces the growth of phytoplankton. Consider for example a population in equilibrium, i.e. $\frac{dP}{dt} = \frac{dZ}{dt} = 0$. The equilibrium populations are $Z^* \approx \frac{\mu_0 p_0}{K_d g_0}$ and $P^* = \frac{d_z p_0}{a g_0} Z^* = \frac{d_z \mu_0 p_0^2}{K_d a g_0^2}$ are independent of H , and thus an equilibrium population will remain in equilibrium even as the mixed layer deepens (Figure 2a). If phytoplankton biomass decreases at some point in winter then subsequent changes in mixed layer depth cannot trigger any biomass accumulation as long as the biological parameters μ_0 , a , g_0 , d_p , d_z , and K_d remain constant.

It could be rebutted that winter accumulation is possible if zooplankton mortality is represented as a linear, rather than quadratic loss term. In that case, as the mixed layer deepens, zooplankton biomass loss rates would not decrease as quickly as

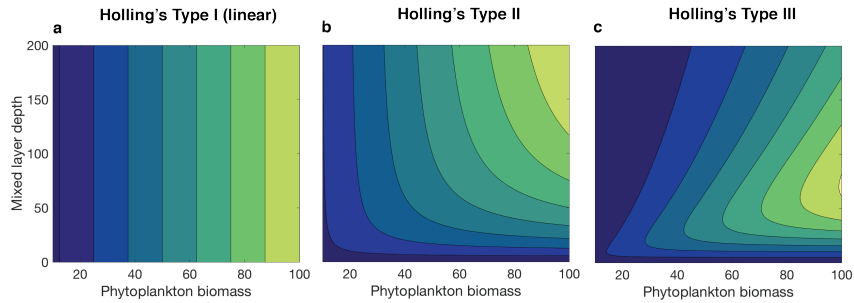


Figure 2. Grazing rate as a function of phytoplankton biomass and mixed layer depth for mixed layer integrated models (e.g. equations 4). Blues are low rates of change of biomass, yellows are high rates of change of biomass. (a) Holling type I (linear) (b) Holling type II (saturating) (c) Holling type III (inflection at low concentrations). Only in the case of Holling type III functional response is there a decrease in the grazing rate as the mixed layer increases. This occurs at low values of phytoplankton biomass and deep mixed layers, as required for the disturbance-recovery hypothesis. The grazing rates with Holling type I functional response is independent of mixed layer depth. At high biomass there is also a decrease in grazing rate at both deeper and shallower mixed layers with a type III functional response and at shallow mixed layers with a type II functional response.

160 the rate of zooplankton grazing on phytoplankton, eventually reaching a crossing over point at which there would be a net loss of zooplankton biomass and consequently an increase in phytoplankton biomass. This is the case of Lotka-Volterra predatory-prey dynamics in a variable environment (*Yorke and Anderson Jr, 1973; Dubois, 1975*). However, this model is problematic because a linear zooplankton mortality at low concentrations is only defensible in the absence of grazing by higher trophic levels. Such grazing is what is implicitly modelled with a quadratic mortality term such as the one used in equation (9).

165 **2.3 Grazing quadratic in phytoplankton concentration: $g(p) \sim p^2$**

The situation is different if we prescribe a phytoplankton grazing function with a dependence on p that is stronger than linear. The Holling type III functional response is a popular choice and can be written as $g_{HIII}(p) = g_0 \frac{p^2}{p^2 + p_0^2}$, which can be approximated as $g_{HIII}(p) \approx \frac{g_0}{p_0^2} p^2$ at low prey concentration. With this functional response, the rate of change of biomass is given by,

170
$$\begin{aligned} \frac{\partial P}{\partial t} &= \frac{1}{H} \left(\frac{\mu_0}{K_d} P - \frac{g_0}{H p_0^2} Z P^2 \right) - d_p P \\ \frac{\partial Z}{\partial t} &= \frac{1}{H} \left(a \frac{g_0}{H p_0^2} Z P^2 - d_z Z^2 \right). \end{aligned} \quad (10)$$

In this case, the grazing rate decreases faster than the phytoplankton growth rate as the mixed layer deepens due to the additional $\frac{1}{H}$ factor in the grazing term (Figure 2c). This opens the possibility of a net increase in phytoplankton biomass due to deepening of the mixed layer, consistent with the scenario invoked in the disturbance-recovery hypothesis (*Behrenfeld, 2010; Behrenfeld and Boss, 2014*). This is the key result of this paper. In what follows, we will use observations to explore the implications of
 175 this insight beyond the low phytoplankton limit.



3 Modeling the annual cycle

We aim to demonstrate that when implemented in a full NPZ model, a grazing function with a quadratic (or higher) dependence on phytoplankton concentration at low p is sufficient to reproduce both wintertime biomass accumulation and a spring bloom. In order to model the full annual cycle, we utilize a more realistic phytoplankton growth rate that depends on nutrient concentration and has a yearly cycle. We replace the growth term $\frac{\mu_0}{K_d H(t)}$ in (2) with

$$\mu(t, n) = \mu_0 \frac{n}{n_0 + n} \left[\frac{3}{10} \sin \left(\frac{t + 10}{365} \pi \right) + 1 \right] \frac{1}{K_d H(t)} \left(1 - e^{-K_d H(t)} \right). \quad (11)$$

This growth rate has temporal dependence through the mixed layer depth and through the surface irradiance. It also depends on nutrient concentration through the function $n/(n + n_0)$ which varies throughout the year and is close to one in winter when p and z are small. Using this formulation we now test the impact of the grazing function on the yearly evolution of biomass and compare with in-situ observations.

Float measurements of the phytoplankton annual cycle in the North Atlantic

We calibrate the NPZ model using the averaged annual cycle of phytoplankton biomass as observed by BGC-Argo floats in the high-latitude North Atlantic (Mignot *et al.*, 2018). Our model ignores the effect of lateral heterogeneity or restratification on phytoplankton dynamics. In order to relate the model results to empirical data, we followed Mignot *et al.* (2018) and selected observations where vertical mixing dominates over lateral transport, i.e. regions where lateral density gradients that drive horizontal flows are weak. This was done by restricting the analysis to floats that did not cross into different water masses (defined as a change in water mass properties in T-S space). Twelve annual cycles that met this criterion were observed during the period 2013-2016 between the latitudes of 50°N and 65°N.

We estimated phytoplankton concentration $p(t)$ from backscatter measurements. The mixed layer depth $H(t)$ is defined as the depth at which the potential density increases by 0.03 kg m⁻³ from the potential density at 10m. As in Mignot *et al.* (2018) the net phytoplankton population growth rate was then calculated using the observed phytoplankton concentration and mixed layer depth as

$$r_p = \frac{1}{P} \int_{-H}^0 \frac{\partial p}{\partial t} d\zeta = \frac{1}{P} \left(\frac{\partial P}{\partial t} - p(-H) \frac{\partial H}{\partial t} \right). \quad (12)$$

In contrast to Mignot *et al.* (2018), the accumulation rate was computed over the mixed layer rather than the productive layer. In order to account for interannual and regional variability in bloom timing, we rescaled the time axis of each individual float time series to account for variability in the start and end dates of winter and spring each year. The rescaled time is defined as $\tau = \frac{t-t_1}{t_2-t_1}$ where t_1 is the onset of weak winter accumulation (the first time in the year when the growth rate is positive for at least 24 consecutive days) and t_2 is the onset of spring (the first time in the year when the mixed layer shoals for at least 24 consecutive days) (Mignot *et al.*, 2018). The average population growth rates was then estimated by averaging over all float time series as a function of the time τ . The result is then plotted in Figure 3 as a function of calendar days setting $\tau = 0$ as the median of all t_1 and $\tau = 1$ as the median of all t_2 .



Model parameters

The NPZ model equations (2) are solved replacing μ_0 with $\mu(n, t)$ as given in equation (11) and using the yearly time-series of $H(t)$ estimated from the average from all float measurements. The total nitrogen pool is assumed to be constant at $N_{max} = 30 \text{ mg C m}^{-3}$ which implies that phytoplankton and zooplankton are immediately remineralized once they die so that $n + p + z = N_{max}$ at all times and one needs equations only for p and z , while the nutrient concentration can be inferred from $n = N_{max} - p - z$. Some parameters are prescribed based on reasonably well established values found in the literature: $\mu_0 = 0.8 \text{ day}^{-1}$ (Eppley, 1972; Geider *et al.*, 1998; Bissinger *et al.*, 2008), $a = 0.5$ (Landry *et al.*, 1984; Moore *et al.*, 2001), $n_0 = 4 \text{ mg C m}^{-3}$ (Moore *et al.*, 2001), and $K_d = 0.05 \text{ m}^{-1}$. However other parameters relating to grazing and zooplankton and phytoplankton mortality are much more uncertain (see Table 1). These parameters and the initial p and z concentrations are therefore calibrated by fitting the model accumulation rate and concentrations to observations over the full annual cycle using a trust-region-reflective least-squares algorithm (Coleman and Li, 1996). The accumulation rates are smoothed before fitting with a five-point Savitzky-Golay filter. The best fit parameters values are given in Table 1.

Table 1. Parameters used in figures 3 and 4. Parameters above the line were prescribed based on literature values. Parameters below the line were fit by linear least squares parameter fitting of the phytoplankton growth rates.

Parameter	Significance (units)	Type II	Type III
u_0	maximum growth rate (day^{-1})	0.8	0.8
a	zooplankton assimilation efficiency	0.5	0.5
n_0	nutrient half saturation constant (mg C m^{-3})	4	4
N_{max}	deep nutrient concentration (mg C m^{-3})	30	30
K_d	attenuation coefficient (m^{-1})	0.05	0.05
g_0	maximum grazing rate (day^{-1})	5.9	4.0
p_0	grazing saturation factor (mg C m^{-3})	15	10
d_z	zooplankton mortality rate ($\text{day}^{-1} \text{mg C}^{-1} \text{m}^3$)	3.5	2.4
d_p	phytoplankton linear mortality rate (day^{-1})	0.004	0.01

The temporal rescaling used to average the timeseries creates a spurious peak in net population growth rate at the beginning of winter. Throughout the winter there is variability in accumulation rates among individual timeseries, including some negative values, even when the average is positive. Our choice to define the start of winter as the period when all timeseries have positive accumulation rates creates the spurious maximum in the observations at that time. We remove this artifact before parameter fitting by interpolating linearly from day 315 to day 4.

Prior values for the biological parameters were chosen based on estimates from the literature (Moore *et al.*, 2001; Behrenfeld and Boss, 2014). We tested the sensitivity of our estimates to the priors by systematically varying the initial parameter choice within the range of values reported in empirical studies. While the fitting algorithm found multiple local minima, all the biologically sensible ones cluster around the values given in Table 1.

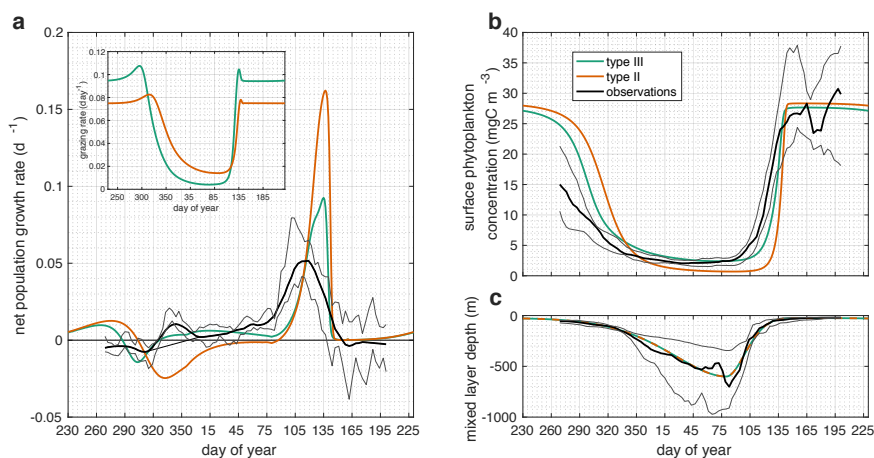


Figure 3. (a) Net mixed layer population growth rate in observations and model. Inset is grazing rate $g(p)$. The thin black line from day 315 to day 4 shows the interpolated growth rate used in parameter fitting. (b) Annual cycle of phytoplankton surface concentration in observations and models (c) Mixed layer depth. The observations, black line, are the median quantity measured by Argo floats with the interquartile range shown in grey lines. The green line is the model in equations 1 with a Holling type III functional response. The orange line is the model with a Holling type II functional response.

Comparison of model and observations

Using either Holling type II or III grazing functions, the model with the best fit parameters generates a spring bloom with a rapid increase in phytoplankton concentration and biomass that coincides with the spring shoaling of the mixed layer (Figure 3b). However, the Holling type III model results in net positive phytoplankton population growth through the winter, while the Holling type II model does not (Figure 3a). The commonly used grazing functions of Holling type I and II do not satisfy the requirement of non-linear dependence of grazing on phytoplankton at low phytoplankton concentrations, and thus cannot capture the observed wintertime biomass accumulation, while the Holling type III functional response has the appropriate nonlinear dependence (Holling, 1959).

During the winter, the phytoplankton concentration is larger when using the type III grazing function than when using the type II grazing function (Figure 3b). Despite this, the winter grazing rate is lower with the type III grazing function (Figure 3a). In order to compensate for the larger winter grazing with the type II function, the parameter fitting procedure infers a much lower linear phytoplankton mortality d_p for that case (Table 1).

During the summertime, the mixed layer depth is fairly constant and phytoplankton and zooplankton populations are close to equilibrium. This model does not include export from the mixed layer through sinking or migrating particles. Instead, any carbon export from the mixed layer only occurs when the mixed layer is shoaling due to biomass being left in the stratified layer below the new mixed layer.



The modeled relationship between phytoplankton and zooplankton shows notable differences between the two grazing func-
245 tions. This is best illustrated by plotting the temporal evolution of the two communities in a $z - p$ plane as shown in Figure 4.
At the end of winter, zooplankton are at slightly higher concentration with type III than type II grazing, because they have fared
better throughout the winter by feeding on a larger phytoplankton population. Zooplankton respond slowly to the explosive
spring phytoplankton bloom with the type II grazing, resulting in higher phytoplankton growth rates and a lower zooplankton
concentration. By contrast, with the type III grazing, zooplankton are strongly coupled to phytoplankton and start grazing as
250 soon as the bloom gets going reducing its amplitude. Importantly the rate of increase of phytoplankton concentration during
the spring bloom is slower than exponential (Mignot *et al.*, 2018), consistent with the prediction of the disturbance-recovery
hypothesis (Behrenfeld and Boss, 2014). With both grazing functions, the spring bloom populations are out of equilibrium with
phytoplankton concentrations being higher and zooplankton concentrations being lower than at equilibrium.

The simple $n - p - z$ model used here is an imperfect representation of the observations. For example, the model only
255 includes one phytoplankton type and one zooplankton type, which precludes both the succession of different phytoplankton
types during the spring and summer and the presence of a microbial loop that could reduce the flow of carbon up the food
chain (Azam *et al.*, 1983). The bulk zero-dimensional model assumes that phytoplankton and zooplankton concentrations are
uniform in the mixed layer and zero below, a defensible approximation for winter conditions—the focus of this study—when
the mixed layer is deep and turbulent mixing is strong, but not in other seasons when turbulence is weak (Taylor and Ferrari,
260 2011). The deficiencies of the bulk model are evident at the spring bloom onset, which is slightly delayed in the model relative
to the observations, occurring once the mixed layer has shoaled rather than during mixed layer shoaling. In observations, blooms
start as soon as turbulent mixing subsides because phytoplankton is no longer mixed away from the surface, while there is a lag
of days to weeks before the mixed layer restratifies and shoals (Taylor and Ferrari, 2011). The bulk model is also problematic
in summer when the mixed layer is shallower than the euphotic layer and some of the productivity takes place below the
265 mixed layer base where the model assumes $p = z = 0$. Despite these deficiencies, bulk mixed layer models have been shown
to qualitatively reproduce the full annual cycle of plankton dynamics in other regions (c.f. Evans and Parslow (1985)) and are
especially appropriate for our work which focuses on phytoplankton growth in winter.

4 Discussion

Our work suggests that the winter accumulation of biomass recently documented from float observations in the North At-
270 lantic (Behrenfeld, 2010; Mignot *et al.*, 2018), while much weaker than that the spring and summer accumulation (Lutz *et al.*,
2007; Uitz *et al.*, 2010), reveals otherwise hard to document top-down controls on phytoplankton populations. By studying
winter time phytoplankton population dynamics, when growth conditions are less than optimal, we have been able to make
inferences about the rate of zooplankton grazing. We demonstrated that there must be non-linearity in the interaction between
the zooplankton and the phytoplankton at low phytoplankton concentrations in order to release the phytoplankton from grazing
275 pressure. A quadratic grazer response function at low phytoplankton biomass is sufficient for phytoplankton biomass accumu-
lation, although higher order nonlinearities would also reproduce the observed dynamics.

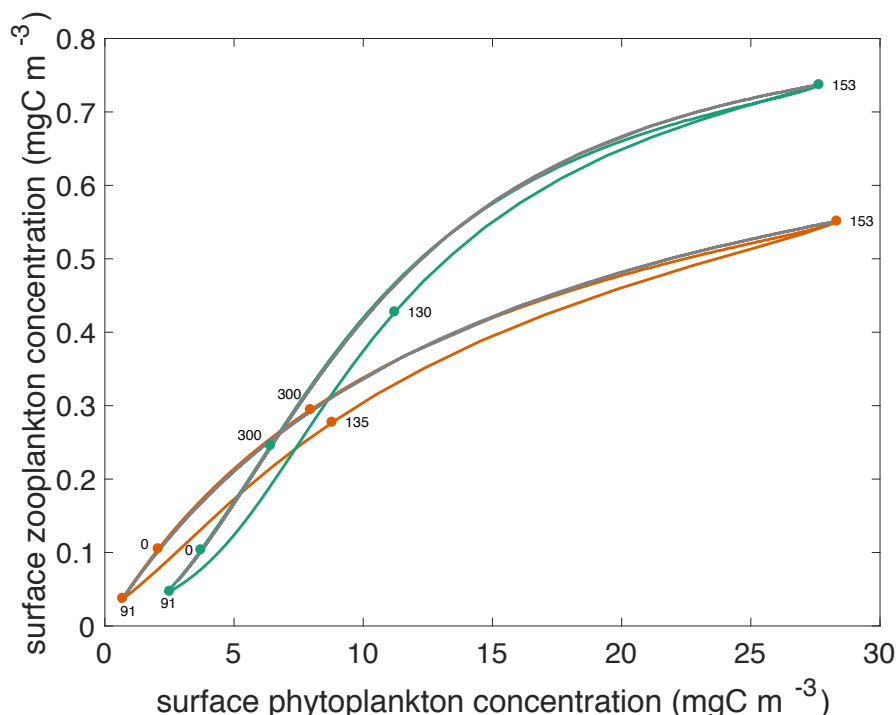


Figure 4. Phytoplankton and zooplankton concentrations through the annual cycle for both models. Green curve shows concentrations that result from the type III model. Orange curve shows concentrations that result from the type II model. The labeled dots indicate day of year for particular locations in the phase portrait. Over the annual cycle, the phytoplankton and zooplankton populations transit these curves counter-clockwise. The grey curves show the steady state concentrations throughout the annual cycle.

280 Relatively little is known about phytoplankton loss through grazing (*Dolan and McKeon, 2005*) in comparison to the other factors that control the dynamics of phytoplankton populations like macro- and micronutrients, light availability, and temperature (*Eppley, 1972*). Our work suggests that winter conditions may offer a unique opportunity to study phytoplankton grazing in the field. In the wintertime, cell division rates decrease because of light limitation due to both deepening of the mixed layer and decrease in sea surface light. In order for phytoplankton accumulation rates to be positive while cell division rates are declining, the phytoplankton loss rates must decrease faster than division rates. While grazing is not the only concentration-dependent process, it is an interesting and compelling example of one process that could lead to biomass accumulation during the wintertime and it is the fundamental tenet of the “disturbance-recovery hypothesis” (*Behrenfeld, 2010*).

285 The wintertime growth is not only important to sustain phytoplankton populations in winter, but it is believed to play a crucial role for the development of the subsequent spring bloom. We showed that a non-linear coupling between zooplankton and phytoplankton results in larger wintertime populations of both zooplankton and phytoplankton at the end of winter than would occur with a linear coupling. Furthermore a non-linear coupling results in a faster acceleration in zooplankton grazing



once phytoplankton concentrations increase during a bloom. The combination of more abundant wintertime populations and
290 stronger/more rapid coupling between phytoplankton and zooplankton populations curb explosive phytoplankton growth.

There are other possible explanations for wintertime biomass accumulation beyond the dilution of phytoplankton. The
biological functions encapsulated in the NPZ model parameters may vary over time. For example, the zooplankton assimilation
rate, a , could depend on the nutrient content of the prey introducing an alternative nonlinear effect (*Landry et al.*, 1984; *Irigoién*
et al., 2005). In our model, the only time-dependent terms are light availability, which has little influence on wintertime
295 biomass accumulation, and mixed layer depth, which drives the wintertime biomass accumulation. Finally, there is evidence
that wintertime growth can be triggered by mixed layer instabilities that occasionally restratify the mixed layer during the
winter and thus increase the light available for phytoplankton (*Karimpour et al.*, 2018). However this cannot be the unique
explanation, because float observations presented in (*Mignot et al.*, 2018) and reviewed here show many examples of wintertime
accumulation where these mixed layer dynamics did not seem to apply.

300 The sensitive dependence of phytoplankton phenology on the rate of grazing by higher trophic levels at low concentrations
provides a powerful quantitative framework in which to evaluate theories of plankton phenology. Observations of wintertime
phytoplankton biomass accumulation have been interpreted as evidence of a release from grazing pressure in deep mixed
layers, but little attention has been given to the key role played by the choice of grazing functions in these theories. Some
studies have used a Holling type III grazing function (*Behrenfeld and Boss*, 2014; *Yang et al.*, 2020) while others relied on a
305 prey switching formulation, where the zooplankton preferentially consumes the most common type of phytoplankton (*Llort*
et al., 2015). The observational evidence for a lower bound on phytoplankton concentrations (*Lessard and Murrell*, 1998) ought
to be studied within the framework presented here. The interpretation of the response of phytoplankton to sudden environmental
perturbations on subseasonal timescales, such as storms (*Behrenfeld and Boss*, 2018), will also require a careful assessment
of the grazing functions which control how fast zooplankton grazing responds to increases in phytoplankton concentrations.
310 Last, but not least, this framework ought to be applied to test the predictions of different theories of bloom onset (*Verity et al.*,
1993; *Morison et al.*, 2020; *Mojica et al.*, 2020). While our work pointed out the key role played by the choice of grazing
functions in such theories and models, it is important to point out that state of the art Earth System Models often use multi-
species ecosystem models (*Laufkötter et al.*, 2015). Multispecies models do not necessarily result in the same dynamics as the
single-species functional responses used here (*Gentleman et al.*, 2003), but our result that phytoplankton phenology is very
315 sensitive to the degree of non-linearity in growth and mortality functions is very likely to hold for more complex ecosystem
models as well.

It is worth commenting on the ecological underpinnings for the different models of grazing. The grazing functions used in our
model represent the coupling between all species of each trophic level of phytoplankton and zooplankton; the phytoplankton
class includes all autotrophs, while the zooplankton class includes all grazers that consume phytoplankton. A nonlinear decrease
320 in grazing rates at low prey concentration has been observed in the lab studies of aquatic vertebrates and invertebrates and in
theoretical studies (*Real*, 1977; *Barrios-O'Neill et al.*, 2016), but it is not clear how these results translate to full populations.
Similarly, copepods go into diapause in the wintertime (*Baumgartner and Tarrant*, 2017), and this effectively reduces grazing
pressure during winter; however, microzooplankton account for the majority of phytoplankton mortality in the ocean (*Landry*



and Calbet, 2004). Other mechanisms described by the Holling type III functional response include prey switching (Vallina
325 *et al.*, 2014), predator learning (Holling, 1966), and prey refuges (Taylor, 2013). While there are few structural prey refuges in
an oceanic mixed layer, a patchy environment can also provide a type of prey refuge. A Holling type III functional response
can arise due to non-random grazing behavior when population dynamics are integrated over a patchy environment (Nachman,
2006; Morozov, 2010).

5 Conclusions

330 A reduction in the grazing rate at low phytoplankton concentration has been proposed as the mechanism to explain the emerging
observation that biomass often increases, albeit weakly, during the wintertime when mixed layers deepen (Behrenfeld, 2010).
It has also been pointed out that the critical depth hypothesis fails to capture wintertime growth because it implicitly assumes
that loss rates are constant either because they are dominated by constant respiration or by grazing by a constant zooplankton
population. Previous modeling results have not acknowledged that a reduction in grazing pressure through dilution of plankton
335 populations in deep mixed layers requires a grazing function that decreases faster than linearly in phytoplankton concentration
at low concentrations.

While our analysis focused on wintertime conditions, we believe that more attention on the functional form of grazing
functions may shed light on other phases of phytoplankton phenology as well. Observations show a tight coupling between
the evolution of phytoplankton and zooplankton populations in all seasons Stelfox-Widdicombe *et al.* (2000); Karayanni *et al.*
340 (2005). This coupling has been interpreted as evidence that zooplankton grazing pressure can respond very rapidly to any
changes in phytoplankton concentrations (Behrenfeld and Boss, 2018). To the extent that these interpretations are correct, our
study suggests that observations can therefore be used to infer the functional form of grazing functions, an aspect of plankton
ecology that is otherwise very to quantify.

Observational validation of the functional forms of grazing functions is key to build confidence in predictions based on
345 biogeochemical models. Different models can be tuned to provide reasonable estimates of the annual cycle of phytoplankton
biomass, like our NPZ model with a more linear grazing function. However, in order to make predictions that are robust to
changing conditions, it is important that models have the correct functional dependencies. Tuning of model parameters is no
guarantee of model performance in an evolving environment that has not been observed yet. Climate change may reshape
North Atlantic phytoplankton populations and primary production (Balaguru *et al.*, 2018) due to increasing surface tempera-
350 ture, shoaling mixed layer depths, and increasing upper ocean stratification (Edwards and Richardson, 2004). Predicting and
quantifying the impact of these changes requires robust model formulations, not models tuned to present climate conditions.

Code and data availability. All code and data are available at https://github.com/mara-freilich/grazing_functions_bg
(DOI: 10.5281/zenodo.4282657)



355 *Author contributions.* All authors conceptualized the research. AM curated the BGC Argo data. MAF wrote the model code and performed the simulations and model analysis. MAF, GF, and RF performed the mathematical analysis. MAF wrote the manuscript with input from all co-authors.

Competing interests. The authors declare that they have no conflict of interest.

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360 References

- Azam, F., T. Fenchel, J. G. Field, J. Gray, L. Meyer-Reil, and F. Thingstad, The ecological role of water-column microbes in the sea, *Marine Ecology Progress Series*, pp. 257–263, 1983.
- Balaguru, K., S. C. Doney, L. Bianucci, P. J. Rasch, L. R. Leung, J.-H. Yoon, and I. D. Lima, Linking deep convection and phytoplankton blooms in the northern Labrador Sea in a changing climate, *PLOS ONE*, 13(1), 1–17, <https://doi.org/10.1371/journal.pone.0191509>, 2018.
- 365 Barrios-O'Neill, D., R. Kelly, J. T. Dick, A. Ricciardi, H. J. MacIsaac, and M. C. Emmerson, On the context-dependent scaling of consumer feeding rates, *Ecology Letters*, 19(6), 668–678, 2016.
- Baumgartner, M. F., and A. M. Tarrant, The physiology and ecology of diapause in marine copepods, *Annual Review of Marine Science*, 9, 387–411, 2017.
- Behrenfeld, M. J., Abandoning Sverdrup's critical depth hypothesis on phytoplankton blooms, *Ecology*, 91(4), 977–989, 2010.
- 370 Behrenfeld, M. J., and E. S. Boss, Resurrecting the ecological underpinnings of ocean plankton blooms, *Annual Reviews in Marine Science*, 2014.
- Behrenfeld, M. J., and E. S. Boss, Student's tutorial on bloom hypotheses in the context of phytoplankton annual cycles, *Global Change Biology*, 24(1), 55–77, <https://doi.org/10.1111/gcb.13858>, 2018.
- Bissinger, J. E., D. J. Montagnes, J. harples, and D. Atkinson, Predicting marine phytoplankton maximum growth rates from temperature: Improving on the eppley curve using quantile regression, *Limnology and Oceanography*, 53(2), 487–493, 2008.
- 375 Boss, E., and M. Behrenfeld, In situ evaluation of the initiation of the North Atlantic phytoplankton bloom, *Geophysical Research Letters*, 37(18), 2010.
- Boss, E., D. Swift, L. Taylor, P. Brickley, R. Zaneveld, S. Riser, M. Perry, and P. Strutton, Observations of pigment and particle distributions in the western North Atlantic from an autonomous float and ocean color satellite, *Limnology and Oceanography*, 53(5part2), 2112–2122, 2008.
- 380 Cole, H. S., S. Henson, A. P. Martin, and A. Yool, Basin-wide mechanisms for spring bloom initiation: how typical is the North Atlantic?, *ICES Journal of Marine Science*, 72(6), 2029–2040, 2015.
- Coleman, T. F., and Y. Li, An interior trust region approach for nonlinear minimization subject to bounds, *SIAM Journal on Optimization*, 6(2), 418–445, 1996.
- 385 Dolan, J., and K. McKeon, The reliability of grazing rate estimates from dilution experiments: Have we over-estimated rates of organic carbon consumption by microzooplankton?, *Ocean Science*, 1(1), 1–7, 2005.
- Dubois, D. M., A model of patchiness for prey—predator plankton populations, *Ecological Modelling*, 1(1), 67–80, 1975.
- Edwards, M., and A. J. Richardson, Impact of climate change on marine pelagic phenology and trophic mismatch, *Nature*, 430(7002), 881, 2004.
- 390 Eppley, R. W., Temperature and phytoplankton growth in the sea, *Fish. Bull.*, 70(4), 1063–1085, 1972.
- Evans, G. T., and J. S. Parslow, A model of annual plankton cycles, *Biological Oceanography*, 3(3), 327–347, 1985.
- Fileman, E., and R. Leakey, Microzooplankton dynamics during the development of the spring bloom in the north-east Atlantic, *JMBA-Journal of the Marine Biological Association of the United Kingdom*, 85(4), 741–754, 2005.
- Fischer, A. D., E. A. Moberg, H. Alexander, E. F. Brownlee, K. R. Hunter-Cevera, K. J. Pitz, S. Z. Rosengard, and H. M. Sosik, Sixty years 395 of Sverdrup: A retrospective of progress in the study of phytoplankton blooms, *Oceanography*, 27(1), 222–235, 2014.



- Geider, R. J., H. L. MacIntyre, and T. M. Kana, A dynamic regulatory model of phytoplankton acclimation to light, nutrients, and temperature, *Limnology and Oceanography*, 43(4), 679–694, 1998.
- Gentleman, W., A. Leising, B. Frost, S. Strom, and J. Murray, Functional responses for zooplankton feeding on multiple resources: a review of assumptions and biological dynamics, *Deep Sea Research Part II: Topical Studies in Oceanography*, 50(22-26), 2847–2875, 2003.
- 400 Holling, C. S., Some characteristics of simple types of predation and parasitism, *The Canadian Entomologist*, 91(7), 385–398, 1959.
- Holling, C. S., The functional response of invertebrate predators to prey density, *The Memoirs of the Entomological Society of Canada*, 98(S48), 5–86, 1966.
- Irigoin, X., K. Flynn, and R. Harris, Phytoplankton blooms: a ‘loophole’ in microzooplankton grazing impact?, *Journal of Plankton Research*, 27(4), 313–321, 2005.
- 405 Karayanni, H., U. Christaki, F. Van Wambeke, M. Denis, and T. Moutin, Influence of ciliated protozoa and heterotrophic nanoflagellates on the fate of primary production in the northeast Atlantic Ocean, *Journal of Geophysical Research: Oceans*, 110(C7), 2005.
- Karimpour, F., A. Tandon, and A. Mahadevan, Sustenance of phytoplankton in the subpolar North Atlantic during winter, *Journal of Geophysical Research: Oceans*, 2018.
- Kjørboe, T., E. Saiz, P. Tiselius, and K. H. Andersen, Adaptive feeding behavior and functional responses in zooplankton, *Limnology and*
- 410 *Oceanography*, 63(1), 308–321, 2018.
- Landry, M., R. Hassett, V. Fagerness, J. Downs, and C. Lorenzen, Effect of food acclimation on assimilation efficiency of *Calanus pacificus*, *Limnology and Oceanography*, 29(2), 361–364, 1984.
- Landry, M. R., and A. Calbet, Microzooplankton production in the oceans, *ICES Journal of Marine Science*, 61(4), 501–507, 2004.
- Laufkötter, C., et al., Drivers and uncertainties of future global marine primary production in marine ecosystem models, *Biogeosciences*, 12,
- 415 6955–6984, 2015.
- Lessard, E. J., and M. C. Murrell, Microzooplankton herbivory and phytoplankton growth in the northwestern Sargasso Sea, *Aquatic Microbial Ecology*, 16(2), 173–188, 1998.
- Llort, J., M. Lévy, J.-B. Sallée, and A. Tagliabue, Onset, intensification, and decline of phytoplankton blooms in the Southern Ocean, *ICES Journal of Marine Science*, 72(6), 1971–1984, 2015.
- 420 Lutz, M. J., K. Caldeira, R. B. Dunbar, and M. J. Behrenfeld, Seasonal rhythms of net primary production and particulate organic carbon flux to depth describe the efficiency of biological pump in the global ocean, *Journal of Geophysical Research: Oceans*, 112(C10), 2007.
- Mahadevan, A., E. D’Asaro, C. Lee, and M. J. Perry, Eddy-driven stratification initiates North Atlantic spring phytoplankton blooms, *Science*, 337(6090), 54–58, 2012.
- Mignot, A., R. Ferrari, and H. Claustre, Floats with bio-optical sensors reveal what processes trigger the North Atlantic bloom, *Nature*
- 425 *Communications*, 9(1), 190, 2018.
- Mojica, K. D., C. A. Carlson, and M. J. Behrenfeld, Regulation of low and high nucleic acid fluorescent heterotrophic prokaryote subpopulations and links to viral-induced mortality within natural prokaryote-virus communities, *Microbial Ecology*, 79(1), 213–230, 2020.
- Moore, J. K., S. C. Doney, J. A. Kleypas, D. M. Glover, and I. Y. Fung, An intermediate complexity marine ecosystem model for the global domain, *Deep Sea Research Part II: Topical Studies in Oceanography*, 49(1-3), 403–462, 2001.
- 430 Morison, F., G. Franzè, E. Harvey, and S. Menden-Deuer, Light fluctuations are key in modulating plankton trophic dynamics and their impact on primary production, *Limnology and Oceanography Letters*, 2020.
- Morozov, A. Y., Emergence of Holling type III zooplankton functional response: bringing together field evidence and mathematical modelling, *Journal of Theoretical Biology*, 265(1), 45–54, 2010.



- Mullin, M. M., E. F. Stewart, and F. J. Fuglister, Ingestion by planktonic grazers as a function of concentration of food, *Limnology and Oceanography*, 20(2), 259–262, 1975.
- Nachman, G., A functional response model of a predator population foraging in a patchy habitat, *Journal of Animal Ecology*, 75(4), 948–958, 2006.
- Ohman, M. D., Omnivory by *Euphausia pacifica*: The role of copepod prey., *Marine Ecology Progress Series*, 19(1), 125–131, 1984.
- Organelli, E., H. Claustre, A. Bricaud, M. Barbieux, J. Uitz, F. D’Ortenzio, and G. Dall’Omo, Bio-optical anomalies in the world’s oceans: An investigation on the diffuse attenuation coefficients for downward irradiance derived from Biogeochemical Argo float measurements, *Journal of Geophysical Research: Oceans*, 122(5), 3543–3564, 2017.
- Real, L. A., The kinetics of functional response, *The American Naturalist*, 111(978), 289–300, 1977.
- Siegel, D., S. Doney, and J. Yoder, The North Atlantic spring phytoplankton bloom and Sverdrup’s critical depth hypothesis, *Science*, 296(5568), 730–733, 2002.
- Siegel, D., K. Buesseler, S. Doney, S. Sailley, M. J. Behrenfeld, and P. Boyd, Global assessment of ocean carbon export by combining satellite observations and food-web models, *Global Biogeochemical Cycles*, 28(3), 181–196, 2014.
- Stelfox-Widdicombe, C. E., E. S. Edwards, P. H. Burkill, and M. A. Sleigh, Microzooplankton grazing activity in the temperate and subtropical NE Atlantic: summer 1996, *Marine Ecology Progress Series*, 208, 1–12, 2000.
- Sverdrup, H., On conditions for the vernal blooming of phytoplankton, *J. Cons. Int. Explor. Mer*, 18(3), 287–295, 1953.
- Taylor, J. R., and R. Ferrari, Shutdown of turbulent convection as a new criterion for the onset of spring phytoplankton blooms, *Limnology and Oceanography*, 56(6), 2293–2307, <https://doi.org/10.4319/lo.2011.56.6.2293>, 2011.
- Taylor, R. J., *Predation*, Springer Science & Business Media, 2013.
- Uitz, J., H. Claustre, B. Gentili, and D. Stramski, Phytoplankton class-specific primary production in the world’s oceans: Seasonal and interannual variability from satellite observations, *Global Biogeochemical Cycles*, 24(3), 2010.
- Vallina, S. M., B. Ward, S. Dutkiewicz, and M. Follows, Maximal feeding with active prey-switching: A kill-the-winner functional response and its effect on global diversity and biogeography, *Progress in Oceanography*, 120, 93–109, 2014.
- Verity, P. G., Measurement and simulation of prey uptake by marine planktonic ciliates fed plastidic and aplastidic nanoplankton, *Limnology and Oceanography*, 36(4), 729–750, 1991.
- Verity, P. G., D. K. Stoecker, M. E. Sieracki, and J. R. Nelson, Grazing, growth and mortality of microzooplankton during the 1989 North Atlantic spring bloom at 47 N, 18 W, *Deep Sea Research Part I: Oceanographic Research Papers*, 40(9), 1793–1814, 1993.
- Visser, A. W., Motility of zooplankton: fitness, foraging and predation, *Journal of Plankton Research*, 29(5), 447–461, 2007.
- Yang, B., E. S. Boss, N. Haëntjens, M. C. Long, M. J. Behrenfeld, R. Eveleth, and S. C. Doney, Phytoplankton phenology in the North Atlantic: Insights from profiling float measurements, *Frontiers in Marine Science*, 7, 139, 2020.
- Yorke, J. A., and W. N. Anderson Jr, Predator-prey patterns, *Proceedings of the National Academy of Sciences*, 70(7), 2069, 1973.