

1      **Reviews and syntheses: Heterotrophic fixation of inorganic carbon –**  
2      **significant but invisible flux in environmental carbon cycling**

3

4                   **Supplementary Information**

5

6      Alexander Braun<sup>1</sup>, Marina Spona-Friedl<sup>1</sup>, Maria Avramov<sup>1</sup>, Martin Elsner<sup>1,2</sup>, Federico Baltar<sup>3</sup>,  
7      Thomas Reinthaler<sup>3</sup>, Gerhard J. Herndl<sup>3,4</sup> & Christian Griebler<sup>1,3\*</sup>

8

9      <sup>1</sup> Helmholtz Zentrum München, Institute of Groundwater Ecology, Ingolstaedter Landstrasse 1, D-85764  
10     Neuherberg, Germany

11     <sup>2</sup> Technical University of Munich, Department of Analytical Chemistry and Water Chemistry, Munich, Germany

12     <sup>3</sup> University of Vienna, Department of Functional and Evolutionary Ecology, Althanstrasse 14, 1090 Vienna,  
13     Austria

14     <sup>4</sup> Department of Marine Microbiology and Biogeochemistry, Royal Netherlands Institute for Sea Research,  
15     Utrecht University, PO Box 59, 1790 AB Den Burg, The Netherlands

16     \* Author for correspondence: christian.griebler@univie.ac.at

17

18

19

20

21

22

23

24

25

26

27

28

29 **Table SI-1:** Global standing stock of organic carbon in living biomass and contribution from anaplerotic CO<sub>2</sub>  
 30 fixation (only anaplerosis is considered here; other mechanisms of heterotrophic CO<sub>2</sub> fixation were neglected).  
 31 In heterotrophs, a conservative estimate of 1-5% of the cell carbon is assumed to originate from inorganic  
 32 carbon fixation (see references in text).

Continental habitats	Carbon biomass [Pg C]	Carbon in biomass derived from anaplerotic CO <sub>2</sub> fixation [Pg C]	References for carbon biomass
Terrestrial animals	0.6	0.006 – 0.03	(Bar-On et al. 2018)
Soil fungi	12	0.12 – 0.6	(Bar-On et al. 2018)
Terrestrial protists	1.6	0.016 – 0.8	(Bar-On et al. 2018)
Soil prokaryotes (upper 100 cm of soil)	23.2	0.23 – 1.16	(Xu, Thornton and Post 2013)
Continental subsurface prokaryotes	2.4 – 12.6*	0.024 – 0.63	(Magnabosco et al. 2018)
Heterotrophic prokaryotes in freshwater and saline inland surface waters	0.013**	0.00013 – 0.00065	(Whitman et al. 1998)
<b>Marine and oceanic habitats</b>			
Marine Animals	2	0.02 – 0.1	(Bar-On et al. 2018)
Marine protists	2	0.02 – 0.1	(Bar-On et al. 2018)
Marine fungi	0.3	0.003 – 0.015	(Bar-On et al. 2018)
Marine planktonic heterotrophic prokaryotes	1.4 – 3.5***	0.014 – 0.175	(Whitman et al. 1998)
Subseafloor sedimentary prokaryotes	1.5 – 22	0.015 – 1.1	(Kallmeyer et al. 2012, Schippers et al. 2005)
Prokaryotes of the oceanic crust	0.5 – 5	0.005 – 0.25	(Bar-On et al. 2018)
<b>Total heterotrophic carbon biomass</b>	<b>47 – 85</b>	<b>0.47 – 4.96</b>	

33  
 34 \* Cell abundances ( $2 – 6 \times 10^{29}$  cells) from Magnabosco et al. (2018) were converted into cell carbon using the  
 35 carbon conversion factors 12 fg C cell<sup>-1</sup> and 21 fg C cell<sup>-1</sup> (Wilhartitz et al. 2009, Griebler et al. 2002) for the  
 36 minimum and maximum values of the range, respectively. In favor of a conservative estimate, quite low carbon  
 37 conversion factors were used (at the lower end of the carbon content values for freshwater prokaryotic cells  
 38 reported in literature).

39  
 40 \*\* Cell abundance ( $2.3 \times 10^{26}$  cells) from Whitman et al. (1998) were converted into cell carbon using a carbon  
 41 conversion factor of 57 fg C cell<sup>-1</sup>, which is the arithmetic mean of the minimum and maximum of a range of  
 42 values (6 to 107 fg C cell<sup>-1</sup>) reported for freshwater lakes and rivers of different trophic states in literature  
 43 (Pedrós-Alió and Brock 1982, Bjørnsen 1986, Simon 1987, Lever et al. 2015).

44  
 45 \*\*\* Cell abundances were converted into cell carbon using the carbon conversion factors 12 fg C cell<sup>-1</sup> and  
 46 30 fg C cell<sup>-1</sup> (Fukuda et al. 1998) for the minimum and maximum values, respectively.

49 **Table SI-2:** Annual global heterotrophic carbon biomass production and contribution from heterotrophic CO<sub>2</sub>  
 50 fixation (via anaplerosis).

	<b>Annual heterotrophic C-biomass production [Pg C yr<sup>-1</sup>]</b>	<b>Anaplerotically fixed carbon [Pg C yr<sup>-1</sup>] <sup>§</sup></b>	
<b><u>Marine and oceanic habitats</u></b>			
Marine and freshwater	2.4 – 76 *	0.024 – 3.8	(Cole, Findlay and Pace 1988, del Giorgio and Duarte 2002)
Oceanic subseafloor	0.1 – 9.8 **	0.001 – 0.49	(Schippers et al. 2005)
<b><u>Continental habitats</u></b>			
Aquifers and unsaturated subsurface	0.12 – 26.3 †	0.0012 – 1.315	(Magnabosco et al. 2018, Griebler et al. 2014)
Soils	31.3 – 133.2 ‡	0.313 – 6.66	(Prentice et al. 2001, Manzoni et al. 2012, Hashimoto et al. 2015, Potter and Klooster 1998)
<b>Total heterotrophic C-biomass production</b>	<b>34 – 245</b>	<b>0.34 – 12.3</b>	

\* Bacterial carbon production (BCP) rates from 54 marine and freshwater studies (Cole et al. 1988) were converted from [mg C m<sup>-2</sup>d<sup>-1</sup>] into [Pg C yr<sup>-1</sup>] and extrapolated to global scale using a world water surface area of 361,419,000 km<sup>2</sup> (<http://www.worldatlas.com/aatlas/infopage/oceans.htm>).

\*\* The total number of living cells [ $1.3 \times 10^{29}$ ] was divided by the turnover time of subseafloor bacteria [0.25–22 yrs], multiplied by the mean carbon content per cell [19 fg C], and converted from [fg C] to [Pg C]. All data as given in Schippers et al. (2005).

† The range of bacterial carbon production rates [fg C L<sup>-1</sup> yr<sup>-1</sup>] from 14 groundwater wells (sampled in spring and autumn) located in an oligotrophic porous aquifer in the Bavarian Alps (close to Mittenwald in Southern Germany) was divided by the corresponding bacterial abundance [cells L<sup>-1</sup>] to obtain BCP rates per cell (data from Griebler et al. 2014). The minimum and the maximum values of these cell-specific BCP rates were then multiplied by the minimum and the maximum estimated total number of prokaryotes in the continental subsurface [ $2\text{--}6 \times 10^{29}$  cells] from Magnabosco et al. (2018), respectively, and carbon mass units were converted from [fg] to [Pg]. Note: since comprehensive, global data on microbial carbon production in aquifers are currently still missing, the level of uncertainty of this estimate is high. Therefore, in order to avoid overestimation, and in favor of obtaining a most conservative estimate, we selected out of the available data only those production rates, which were determined in pristine, highly oligotrophic environments. If all other data from the dataset in Griebler et al. (2014), in total 88 wells throughout Germany, sampled twice, as well as the data from four other available studies with sites in the USA, Austria and Denmark (Thorn and Ventullo 1988, Kazumi and Capone 1994, Albrechtsen and Winding 1992, Wilhartz et al. 2009) were to be included, a much higher estimate of the global annual heterotrophic carbon biomass production in aquifers would be obtained, ranging from 0.06 to 4,829 Pg C yr<sup>-1</sup>, and corresponding to 0.001 – 386 Pg C yr<sup>-1</sup> of anaplerotically fixed carbon each year.

‡ Global terrestrial heterotrophic respiration in soils [55 Pg C yr<sup>-1</sup>] from Prentice et al. (2001) was extrapolated to carbon biomass production assuming that respiration accounts for 30–62% of the total carbon consumed

79 (corresponding to a carbon use efficiency (CUE) of 38-70%) in the course of organic matter decomposition in  
80 different types of soils (Manzoni et al. 2012).

81  
82 <sup>§</sup> It was assumed that 1-5% of the annually produced carbon biomass of heterotrophs originate from anaplerotic  
83 CO<sub>2</sub> fixation (see ref. in the text of the main MS). A fraction of 1% was applied to the minimum, and 5% to the  
84 maximum value of the C-biomass production ranges in this table, respectively.

85

86

87 **References**

88 Albrechtsen, H.-J., and Winding A.: Microbial biomass and activity in subsurface sediments from  
89 Vejen, Denmark. *Microb. Ecol.*, 23, 303-317, 1992.

90 Bar-On, Y. M., Phillips, R., and Milo, R.: The biomass distribution on Earth. *PNAS*, 115, 6506-6511,  
91 2018.

92 Bjørnsen, P. K.: Automatic determination of bacterioplankton biomass by image analysis. *Applied  
93 and Environ. Microbiol.*, 51, 1199-1204, 1986.

94 Cole, J. J., Findlay, S. E. G., and Pace, M. L. Bacterial production in fresh and saltwater ecosystems : a  
95 cross-system overview. *Mar. Ecol. Prog. Ser.*, 43, 1-10, 1988.

96 del Giorgio, P. A., and Duarte, C. M.: Respiration in the open ocean. *Nature*, 420, 379-384, 2002.

97 Falkowski, P. G., Barber, R. T., and Smetacek, V.: Biogeochemical controls and feedbacks on ocean  
98 primary production. *Science*, 281, 200-206, 1998.

99 Fukuda, R., Ogawa, H., Nagata, T., and Koike, I.: Direct determination of carbon and nitrogen  
100 contents of natural bacterial assemblages in marine environments. *Appl. Environ. Microbiol.*, 64,  
101 3352-3358, 1998.

102 Griebler, C., Mindl, B., Slezak, D., and Geiger-Kaiser, M.: Distribution patterns of attached and  
103 suspended bacteria in pristine and contaminated shallow aquifers studied with an in situ sediment  
104 exposure microcosm. *Aquat. Microb. Ecol.*, 28, 117-129, 2002.

105 Griebler, C., Hahn, H. J., Stein, H., Kellermann, C., Fuchs, A., Steube, C., Berkhoff, S., and Briemann,  
106 H.: Development of a biological assessment scheme and criteria for groundwater ecosystems  
107 (Entwicklung biologischer Bewertungsmethoden und -kriterien für Grundwasserökosysteme). Report  
108 to the German Federal Environmental Agency (UBA); UFOPLAN grant no. 3708 23 200, ISSN: 1862-  
109 4804, 153 pp., 2014.

110 Hashimoto, S., Carvalhais, N., Ito, A., Migliavacca, M., Nishina, K., and Reichstein, M.: Global  
111 spatiotemporal distribution of soil respiration modeled using a global database. *Biogeosci.*, 12, 4121-  
112 4132, 2015.

113 Kallmeyer, J., Pockalny, R., Adhikari, R. R., Smith, D. C., and D'Hondt, S.: Global distribution of  
114 microbial abundance and biomass in subseafloor sediment. *PNAS*, 109, 16213-16216, 2012.

115 Kazumi, J., and Capone D. G.: Heterotrophic microbial activity in shallow aquifer sediments of Long  
116 Island, New York. *Microb. Ecol.*, 28, 19-37, 1994.

117 Kieft, T. L., and Simmons K. A.: Allometry of animal-microbe interactions and global census of animal-  
118 associated microbes. *Proceedings of the Royal Society of London B: Biol. Sci.*, 282, 1-8, 2015.

- 119 Lever, M. A., Rogers, K. L., Lloyd, K. G., Overmann, J., Schink, B., Thauer, R. K., Hoehler, T. M. &  
120 Jørgensen, B. B.: Life under extreme energy limitation: a synthesis of laboratory- and field-based  
121 investigations. *FEMS Microbiol. Rev.*, 39, 688-728, 2015.
- 122 Magnabosco, C., Lin, L. H., Dong, H., Bomberg, M., Ghiorse, W., Stan-Lotter, H., Pedersen, K., Kieft, T.  
123 L., van Heerden, E., and Onstott, T. C. The biomass and biodiversity of the continental subsurface.  
124 *Nature Geoscience*, 11, 707-717, 2018.
- 125 Manzoni, S., Taylor, P., Richter, A., Porporato, A., and Ågren, G. I.: Environmental and stoichiometric  
126 controls on microbial carbon-use efficiency in soils. *New Phytologist*, 196, 79-91, 2012.
- 127 McMahon, S., and Parnell J. Weighing the deep continental biosphere. *FEMS Microbiol. Ecol.*, 87,  
128 113-120, 2014.
- 129 Pedrós-Alió, C., and Brock T. D.: Assessing biomass and production of bacteria in eutrophic lake  
130 Mendota, Wisconsin. *Appl. Environ. Microbiol.*, 44, 203-218, 1982.
- 131 Potter, C. S., and Klooster S. A.: Interannual variability in soil trace gas(CO<sub>2</sub>, N<sub>2</sub>O, NO) fluxes and  
132 analysis of controllers on regional to global scales. *Global Biogeochem. Cycl.*, 12, 621-635, 1998.
- 133 Prentice, I. C., G. D. Farquhar, M. J. R. Fasham, M. L. Goulden, M. Heimann, V. J. Jaramillo, H. S.  
134 Kheshgi, C. Le Quéré, R. J. Scholes, and D. W. R. Wallace.: The carbon cycle and atmospheric carbon  
135 dioxide. In *Climate Change 2001: The Scientific Basis. Contribution of Working Group I to the Third*  
136 *Assessment Report of the Intergovernmental Panel on Climate Change* eds. J. T. Houghton, Y. Ding,  
137 D. J. Griggs, M. Noguer, P. J. van der Linden, X. Dai, K. Maskell & C. A. Johnson, 183-237. Cambridge,  
138 United Kingdom and New York, NY, USA: Cambridge University Press, 2001.
- 139 Schippers, A., Neretin, L. N., Kallmeyer, J., Ferdelman, T. G., Cragg, B. A., Parkes, R. J. & Jørgensen, B.  
140 B.: Prokaryotic cells of the deep sub-seafloor biosphere identified as living bacteria. *Nature*, 433,  
141 861-864, 2005.
- 142 Simon, M.: Biomass and production of small and large free-living and attached bacteria in Lake  
143 Constance. *Limnol. Oceanogr.*, 32, 591-607, 1987.
- 144 Thorn, P. M., and Ventullo, R. M. Measurement of bacterial growth rates in subsurface sediments  
145 using the incorporation of tritiated thymidine into DNA. *Microb. Ecol.*, 16, 3-16, 1988.
- 146 Whitman, W. B., Coleman, D. C. & Wiebe, W. J.: Prokaryotes: The unseen majority. *PNAS*, 95, 6578-  
147 6583, 1998.
- 148 Wilhartitz, I. C., Kirschner, A. K. T., Stadler, H., Herndl, G. J., Dietzel, M., Latal, C., Mach, R. L., and  
149 Farnleitner, A. H.: Heterotrophic prokaryotic production in ultra-oligotrophic alpine karst aquifers  
150 and ecological implications. *FEMS Microbiol. Ecol.*, 68, 287-299, 2009.
- 151 Xu, X., Thornton, P. E., and Post, W. M.: A global analysis of soil microbial biomass carbon, nitrogen  
152 and phosphorus in terrestrial ecosystems. *Global Ecol. Biogeograph.*, 22, 737-749, 2013.