



# The Cretaceous physiological adaptation of angiosperms to a declining pCO<sub>2</sub> : a trait-oriented modelling approach

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**Abstract.** The Cretaceous evolution of angiosperm leaves towards higher vein densities enables unprecedented leaf stomatal conductance. Still, simulating and quantifying the impact of such change on plant productivity and transpiration in the peculiar environmental conditions of the Cretaceous remains challenging. Here, we address this issue by combining a paleo proxybased model with a fully atmosphere-vegetation model that couples stomatal conductance to carbon assimilation. Based on

- 5 the fossil record, we build and evaluate three consistent pre-angiosperm vegetation parameterizations under two end-members scenarios of  $pCO_2$  (280 ppm and 1120 ppm) for the mid-Cretaceous : a reduction of hydraulic or photosynthetic capacity and a combination of both, supported by a likely coevolution of stomatal conductance and photosynthetic biochemistry. Our results suggest that decreasing hydraulic or/and photosynthetic capacities always generates a reduction of transpiration that is predominantly the result of plant productivity variations, modulated by light, water availability in the soil and atmospheric
- 10 evaporative demand. The high  $pCO_2$  acts as a fertilizer on plant productivity that bolsters plant transpiration and water-use efficiency. However, we show that pre-angiosperm physiology does not allow vegetation to grow under low  $pCO_2$  because of a positive feedback between leaf stomatal conductance and leaf area index. Our modelling approach stresses the need to better represent paleovegetation physiological traits. It also confirms the hypothesis of a likely evolution of angiosperms from a stage of low hydraulic and photosynthetic capacities at high  $pCO_2$  to a stage of high hydraulic and photosynthetic capacities linked
- 15 to leaves more and more densely irrigated together with a more efficient biochemistry at low  $pCO_2$ .

# 1 Introduction

Vegetation plays a pivotal role in the climate system as it controls water and energy fluxes at the interface between land surfaces and the atmosphere through albedo (Charney et al., 1975; Port et al., 2016; Brovkin et al., 2009), roughness length, and evapo-transpiration capabilities (e.g., Bathiany et al., 2010; Betts et al., 1997; Kleidon et al., 2000; Gibbard et al., 2005). Specifically,

20 evapotranspiration (ET), i.e. the sum of soil evaporation, vegetation evaporation and vegetation transpiration fluxes, is a key term in the continental hydrological cycle, and has been shown to control moisture convergence and convection, thereby precipitation patterns in the tropics either in present-day-like configurations (Sun and Barros, 2015; Fraedrich et al., 1999) or for past climates (e.g., Braconnot et al., 1999; Brovkin et al., 2006). It is therefore mandatory that earth system models designed to simulate climate evolution, either in the past or in the future, account for plant traits that alter these fluxes, despite the numerous





- 25 challenges involved (Fisher and Koven, 2020). Including vegetation traits in land surface models is a growing field of research (Kattge et al., 2020), but mostly dedicated to better represent present-day vegetation and its response to human-induced climate change (Scheiter et al., 2013; Dury et al., 2018; Peng et al., 2020; Davin and de Noblet-Ducoudré, 2010). The problem has an additional degree of complexity for million-years-old ("deeptime") climates, for which the vegetation physiological traits have been very different from the present. As shown recently for the extinct vegetation of the late Paleozoic, fossil plants provide
- 30 invaluable information regarding "paleo-traits" that can be in turn included in land surface models (White et al., 2020; Richey et al., 2021).

Another case-study for testing the impact of pivotal changes in physiological traits on the vegetation characteristics is the Cretaceous angiosperm radiation (Boyce and Lee, 2010; Boyce et al., 2010; De Boer et al., 2012). With more than 295,000 species described, angiosperms represent more than 95 % of modern terrestrial vascular plant diversity (Christenhusz and

- Byng, 2016). One of the most emblematic changes in vegetation through Earth history is their diversification at the expanse of gymnosperms during the Cretaceous (Condamine et al., 2020). The macroflora fossil record reveals significant changes in leaf anatomy between the early Cretaceous and the late Cretaceous associated with the rise of angiosperms. Specifically, measurements on fossil leaves show that vein density (D<sub>v</sub>) increased from 3.1 mm mm<sup>-2</sup> for the oldest angiosperms leaves from the Aptian-Albian (115 Ma) to 9.8 mm mm<sup>-2</sup> by the Maastrichtian-Paleocene (66 Ma), while gymnosperms D<sub>v</sub> remained
- 40 centered around 2.4 mm mm<sup>-2</sup> over the same period (Feild et al., 2011a). Contemporaneous changes in stomatal size (S) and density ( $D_s$ ) are observed in the fossil record (De Boer et al., 2012) showing a strong correlation with vein density. While veins allow the plants to efficiently transport water from the soil close to the site of transpiration i.e stomata, the latter allow the leaves to efficiently exchange carbon and water with the atmosphere. Studies have already shown that  $D_v$  is a reliable marker of hydraulic capacity (Brodribb et al., 2007; Brodribb and Feild, 2010; Feild et al., 2011a, b; Boyce et al., 2010; Lee
- and Boyce, 2010; Boyce et al., 2009; Boyce and Lee, 2010, 2017) and pioneer modelling studies have explored the impact of such a variation of vein density on the transpiration rate in present-day conditions (Boyce et al., 2010; Lee and Boyce, 2010; Boyce and Lee, 2010, 2017). Based on the closed link between vein density, hydraulic and photosynthetic capacities (Brodribb et al., 2007), linear relationships have been inferred between D<sub>v</sub> and (1) the transpiration rate on one hand (Boyce et al., 2009; Boyce and Lee, 2017; Boyce et al., 2010), and (2) the maximum rate of carboxylation (V<sub>cmax</sub>), involved in photosynthesis, on
- 50 the other hand (Boyce and Lee, 2010; Lee and Boyce, 2010). Sensitivity tests carried out with a land surface model coupled to an atmospheric circulation model showed that angiosperms with modern traits bolster the current hydrological cycle, ensuring seasonally high levels of transpiration and precipitation rates in the tropics (Lee and Boyce, 2010; Boyce and Lee, 2010, 2017; Boyce et al., 2010).

Even if a lot of functional traits evolved during the angiosperm radiation, in this paper we will focus on changes of leaf

stomatal conductance and photosynthetic capacity. In particular, since they are highly related to environmental changes, our approach relies on the integration of fossil traits in a land surface model in order to quantify how transpiration and photosynthesis responded to the angiosperm leaf trait evolution in the peculiar environmental conditions of the Cretaceous (different paleogeography and high atmospheric  $CO_2$  concentrations). Land surface models used in climate modelling do not explicitly represent traits such as vein densities but rather directly simulate the operational stomatal conductance at the leaf scale ( $g_s$ )





- 60 (Farquhar et al., 1980; Ball et al., 1987; Krinner et al., 2005; Yin and Struik, 2009). However, based on relationships obtained from extant and fossil leaves that links maximal anatomic stomatal conductance to  $H_2O(g_{anat}^{max})$  to  $D_v$  (Brodribb et al., 2007; Brodribb and Feild, 2010), they can be parameterized to represent the changes in stomatal conductance that occured with the angiosperm radiation. Depending on the choices made, these parameterizations can reflect changes on plant traits controlling hydraulic capacity, photosynthetic capacity or both. The latter is supported by our knowledge of modern plant processes and
- in particular of a coevolution between stomatal conductance and  $V_{cmax}$  (Franks and Beerling, 2009a; De Boer et al., 2012). Although there is no fossil evidence of changes in photosynthetic capacity, we assume that this covariation is a time-invariant property and that such relationship was the same in the past. Moreover, atmospheric pCO<sub>2</sub> is known to control the degree of stomatal opening and closing at very short-term (Jarvis et al., 1999). Several studies have reported that the  $g_s/g_{anat}^{max}$  ratio is relatively stable around 20-40 %, an optimal range within which changes in guard cell turgor pressure are most efficient
- <sup>70</sup> in controlling quick stomatal conductance changes (Dow et al., 2014; Dow and Bergmann, 2014; McElwain et al., 2016). Therefore, it is likely that pCO<sub>2</sub> also drives changes in  $g_{anat}^{max}$  on long time scales. Paleo-CO<sub>2</sub> reconstructions based on proxies (plant fossils and isotopes) and geochemical models show that, despite a large spread, pCO<sub>2</sub> was high throughout the Cretaceous, ranging from 500 ppm to 2000 ppm. Values peaked during the mid-Cretaceous (Cenomanian-Turonian, ca. 95 Ma), then steadily declined towards the Cretaceous-Paleogene boundary (Fletcher et al., 2008; Wang et al., 2014). It is thus important
- to consider this varying pCO<sub>2</sub> when exploring the mechanisms of the angiosperm physiological changes. Finally, the goal of our study is to evaluate the combined environmental and physiological-induced response of plants evolving higher  $D_v$  with a coupled atmosphere-vegetation model. This experimental design considers both reducing plant hydraulic or/and photosynthetic capacities and applying different forcing in pCO<sub>2</sub>, allowing the modeled  $g_s$  to account for long-term plant adaptation to their changing environment.

#### 80 2 Methods

### 2.1 IPSL atmosphere-land surface model

In this study, we use LMDz and ORCHIDEE versions embedded in the IPSL-CM5A2 model (Sepulchre et al., 2020). LMDz is the atmosphere general circulation model of the Institut Pierre-Simon Laplace earth system model. LMDz couples a dynamical core solving the primitive equations of conservation with a set of physical parameterizations of the radiative and convective

- 85 schemes. The spatial resolution of LMDz is the regular 3.75 x 1.875° longitude-latitude grid. LMDz has 39 vertical levels to describe vertical processes (Hourdin et al., 2013). ORCHIDEE is the land surface component of the IPSL earth system model (Krinner et al., 2005). When coupled with LMDz, it has the same horizontal resolution, and a 4-meter soil water depth represented with a two-layer bucket model. ORCHIDEE simulates water and energy exchanges (Rosnay and Polcher, 1998; Ducoudré et al., 1993), and the key processes of the terrestrial carbon cycle such as photosynthesis, carbon allocation, or soil
- 90 organic matter decomposition for thirteen Plant Functional Types (PFTs). Each PFT gathers a set of plant species with similar functional characteristics that will be translated into a common set of parameter values. Photosynthesis, water and energy



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exchanges are computed every 30 minutes while carbon-related slow processes are computed on a daily basis (Krinner et al., 2005; Sitch et al., 2003).

#### 2.1.1 Stomatal conductance in ORCHIDEE

- 95 Stomatal conductance is computed per canopy layer, with layer depth increasing exponentially from top to bottom of the canopy. This feature enables to account for the response to light radiation extinction through the canopy. Leaf stomatal conductance modelling is coupled to photosynthesis based on original formulations from Ball et al. (1987) and Leuning et al. (1995). Within the model used in ORCHIDEE (Yin and Struik, 2009), leaf operational stomatal conductance to H<sub>2</sub>O, g<sub>s</sub> (mol  $m^{-2}$ [leaf] s<sup>-1</sup>), depends on the net carbon assimilation A (molCO<sub>2</sub> m<sup>-2</sup>[leaf] s<sup>-1</sup>), the day respiration R<sub>d</sub> (molCO<sub>2</sub> m<sup>-2</sup>[leaf]
- 100 s<sup>-1</sup>), the intercellular CO<sub>2</sub> partial pressure C<sub>i</sub> (bar), the C<sub>i</sub>-based CO<sub>2</sub> compensation point in the absence of R<sub>d</sub>, C<sup>\*</sup><sub>i</sub> (bar), a factor fcpl, describing the strength of the coupling between A and g<sub>s</sub>, which is function of the leaf-to-air vapor pressure deficit and that we will further name "hydraulic capacity", and the residual stomatal conductance to H<sub>2</sub>O, g<sub>0</sub> (mol m<sup>-2</sup>[leaf] s<sup>-1</sup>) to account for a non-zero conductance when the carbon assimilation is zero (Farquhar et al., 1980; Ball et al., 1987; Yin and Struik, 2009):

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$$g_s = g_0 + \frac{A + R_d}{C_i - C_i^{\star}} * fcpl$$
 (1)

The semi-empirical formalism of  $g_s$  allows to account for both the structural conductance, linked to leaves morphologic and physiologic traits (e.g.  $D_v$ ,  $D_s$  and S) developed on the long-term plant evolution history, and the dynamical conductance, related to the short-term stomatal opening and closing depending on the environment. A is limited by either the Rubisco enzyme (RuBP) activity ( $V_c$ ) or the electron transport ( $V_j$ ):

$$110 \quad A = \min(V_c, V_j) \tag{2}$$

 $V_c$  represents the ability of the RuBP to fix CO<sub>2</sub> and initiate the Calvin cycle. It relies on  $V_{cmax}$ ,  $K_c$  and  $K_o$  the Michaelis-Menten constants of Rubisco for CO<sub>2</sub> and O<sub>2</sub> respectively (bar) and O the oxygen partial pressure (bar):

$$V_c = \left[\frac{(C_i - C_i^{\star})V_{cmax}}{C_i + K_c \left(1 + \frac{O}{K_O}\right)} - R_d\right] * \left[1 - w_{soil}^{lim}\right]$$
(3)

 $V_{cmax}$  is parameterized for each PFT as a function of leaf age (Krinner et al., 2005).  $V_j$  represents the regeneration rate of the 115 RuBP. The latter depends on the radiation and the electron transport :

$$V_j = \left[\frac{(C_i - C_i^{\star})J}{4C_i + 8C_i^{\star}} - R_d\right] * \left[1 - w_{soil}^{lim}\right]$$

$$\tag{4}$$

Where J is the actual rate of electron transport (Yin and Struik, 2009), dependent on irradiance and  $V_{jmax}$ , the maximum rate of electron transport at saturating light (mol e- m<sup>-2</sup> s<sup>-1</sup>).  $V_{jmax}$  is directly proportional to  $V_{cmax}$  with the ratio  $V_{cmax}/V_{jmax}$  varying with surface monthly temperature (Kattge and Knorr, 2007). Then, when discussing the changes in  $V_{cmax}$ , we consider the proportional changes in  $V_{jmax}$ , in the following. A increases with  $V_{cmax}$  and pCO<sub>2</sub>, until to reach a plateau where light



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and temperature become the limiting factors. Both  $V_c$  and  $V_j$  are a function of the soil moisture stress for transpiration  $w_{soil}^{lim}$ , that ultimately controls photosynthesis. This factor depends on both the soil moisture and the root profile given per PFT, trees having deeper roots than grasses.  $w_{soil}^{lim}$  is unitless and ranges between 0 for fully moist soil and 1 for maximal water stress. The transpiration rate Tr (mm d<sup>-1</sup>) is a function of the potential evapotranspiration, that is, the atmospheric evaporative demand, modulated by a serie of resistances :

$$Tr = \frac{1}{r_c + r_a}\rho(q_s^{\star} - q) \tag{5}$$

where  $r_a$  and  $r_c$  are the aerodynamic and canopy resistances respectively (s m<sup>-1</sup>),  $\rho$  is the air density (kg m<sup>-3</sup>),  $q_s^*$  is the saturation specific humidity of the evaporating surface (kg kg<sup>-1</sup>), function of the temperature, and q is the specific humidity of the surrounding air (kg kg<sup>-1</sup>). Calculation of the transpiration requires the integration of  $g_s$  at the canopy level which is  $g_c$  (molH<sub>2</sub>O m<sup>-2</sup>[ground] s<sup>-1</sup>), that is over the leaf area index (LAI) and expressed as :

$$g_c = \int_{l=0}^{l=LAI} g_s(l) dl$$

$$g_c = \frac{1}{r_c}$$
(6)
(7)

#### 2.1.2 Summary of dynamical coupling between stomatal conductance and photosynthesis in a given environment

- At the leaf level, the structural resistance (Fig. 1, in orange) represents traits (veins and stomata) that evolve at long time scales.
  135 The structural resistance drives the maximum theoretical conductance of water (g<sup>max</sup><sub>anat</sub>). The dynamical resistance (Fig. 1, in dark blue) controls the stomatal closure on a very short time scale. Plants can also slightly adapt their V<sub>cmax</sub> and V<sub>jmax</sub>, in absence of nitrogen stress, on seasonal timescale and in a greater range on evolutionary long time scales. External forcing are pCO<sub>2</sub>, light, rainfall, relative humidity, wind stress and soil moisture stress for transpiration (Fig. 1, in light blue). The coupling factor (fcpl) controls the sensitivity of stomatal conductance to the surrounding environment : a low fcpl implies that g<sub>s</sub> is less
  140 sensitive to C<sub>i</sub>, A and the vapor pressure deficit than a high fcpl. The end limit of the assimilation process is the available energy from light that controls the quantity of RuBP that can be regenerated (V<sub>j</sub> in Eq. (4)). At high pCO<sub>2</sub>, C<sub>i</sub> is maintained at
- a sufficient level to feed the carboxylation rate ( $V_c$ ) and then, for a sufficient  $V_{cmax}$ , light becomes the main limiting factor even for a small conductance (Eq. (2) to (4)). So, the structural resistance is not a limiting factor and plants will close the stomata to limit the water loss. Likewise,  $V_c$  rate increases, with both  $C_i$  and  $V_{cmax}$ , and the level of  $V_{cmax}$  for which  $V_j$  becomes
- the main limiting factor decreases (Eq. (2)). Because nitrogen acquisition and RuBP protein maintenance are highly energetic for the plants, they tend to optimize the  $V_{cmax}$  and  $V_{jmax}$  to reach the point where  $V_c$  and  $V_j$  are colimiting (Maire et al., 2012). On the contrary, at low pCO<sub>2</sub> structural conductance (that drives the maximum conductance) can become a limiting factor since, even with full stomata opening, it can limit  $C_i$  under the level to feed the carboxylation rate.  $V_{cmax}$  and  $V_{jmax}$ should also be increased to maintain a sufficient level of carboxylation ( $V_c$ ) (lower efficiency of the oxidation/carboxylation

150 ratio).





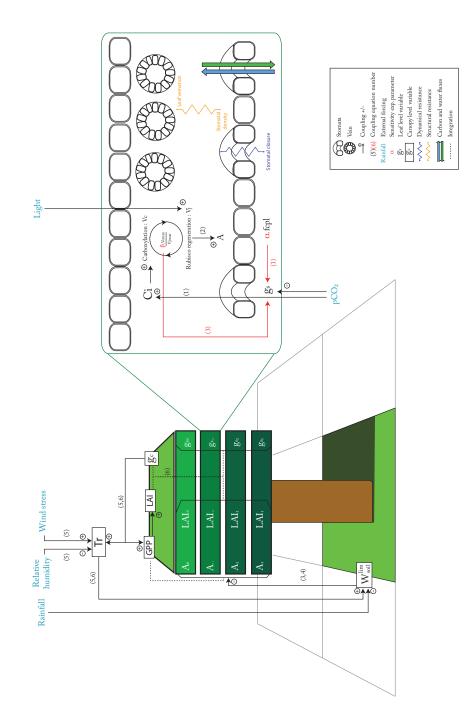


Figure 1. Schematized coupling between stomatal conductance and photosynthesis from the leaf to the canopy scale in the ORCHIDEE model.

At the canopy level (Fig. 1), the canopy conductance  $g_c$  depends on both leaf level conductance  $g_s$  and LAI. As LAI is a function of plant productivity, there is an additional positive feedback: a change in  $g_s$  will impact GPP and then LAI in the

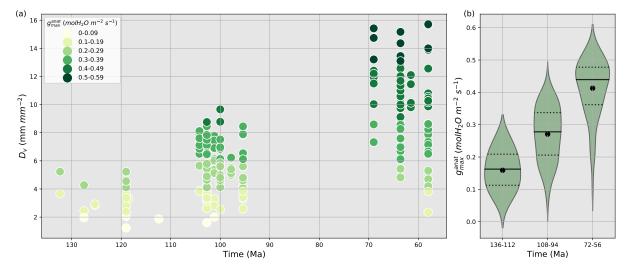




same direction, which will amplify the initial effect of g<sub>s</sub> on g<sub>c</sub>. Another external forcing is the soil water availability (w<sup>lim</sup><sub>soil</sub>). A low soil water content will induce a water stress limiting the V<sub>cmax</sub> and V<sub>jmax</sub> that will indirectly also reduce g<sub>s</sub>. Hence,
arid ecosystems will be less sensitive to structural change in g<sub>s</sub> than ecosystems without a large hydric stress. On the contrary, they will be affected in the same way for direct changes on V<sub>cmax</sub> and V<sub>jmax</sub>.

# 2.2 Fossil evidence of increasing angiosperm hydraulic and photosynthetic capacities

Vein density as well as stomatal size and density are both used to reconstruct past variations of g<sup>max</sup><sub>anat</sub> (Franks and Beerling, 2009a, b; Brodribb et al., 2007; Brodribb and Feild, 2010; De Boer et al., 2012; Franks and Farquhar, 2001). Here, we have
160 chosen to account for D<sub>v</sub> changes rather than D<sub>s</sub> and S. Indeed, using D<sub>v</sub> is a good proxy to constrain g<sup>max</sup><sub>anat</sub> since D<sub>v</sub> and D<sub>s</sub> are correlated and that observed relationship between D<sub>v</sub> and g<sup>max</sup><sub>anat</sub> gives the highest correlation coefficient (Feild et al., 2011b). Vein densities from angiosperm fossils published in Feild et al. (2011b) record a 2 to 5-fold increase in angiosperm D<sub>v</sub> during the Cretaceous (Fig. 2a), compared to early angiosperms and non-angiosperm (Table S1).



**Figure 2.** (a) Time evolution of fossil vein densities (mm mm<sup>-2</sup>, y-axis) adapted from Feild et al. (2011a) and the corresponding maximal stomatal conductance to H<sub>2</sub>O (mol m<sup>-2</sup> s<sup>-1</sup>, filled circles) calculated by the anatomic relationship developed by Brodribb et al. (2007) and Brodribb and Feild (2010). (b) Subdivision of maximal stomatal conductance to H<sub>2</sub>O (mol m<sup>-2</sup> s<sup>-1</sup>) by time periods. The solid line is the median, the dotted lines are the first and third quartiles, and the point is the mean value for each time period.

 $D_v$  allows to reconstruct the maximal water that can flow through the stomata  $g_{anat}^{max}$  (mol m<sup>-2</sup> s<sup>-1</sup>) using the relationship developed by Brodribb et al. (2007) and Brodribb and Feild (2010) :

$$g_{max}^{anat} = \frac{12760}{\nu} \Delta \Psi_{leaf} \left( \frac{\pi}{2} \sqrt{\frac{650^2}{D_v^2} + y^2} \right)^{-1.27} \tag{8}$$

Where  $\nu$  is the leaf-to-air vapor pressure deficit (MPa),  $\Delta \Psi_{leaf}$  is the leaf water potential gradient (MPa),  $D_v$  is the vein density (mm mm<sup>-2</sup>) and y is the distance from vein terminals to epidermis ( $\mu$ m).  $\nu$  and  $\Delta \Psi_{leaf}$  are set to 0.002 MPa and





0.4 MPa respectively, values that are typical for temperate-tropical environments (Brodribb and Feild, 2010). An estimation of
y is 70-130 μm, producing a span of predicted values encompassing the likely morphological variability of leaves (Brodribb and Feild, 2010). The lower y is, the closer the vein is to the stomata and the higher g<sup>max</sup><sub>anat</sub> is. The highest increase in g<sup>max</sup><sub>anat</sub> is obtained for the largest variation of D<sub>v</sub> over time combined with the smallest y. Conversely, the lowest increase in g<sup>max</sup><sub>anat</sub> is obtained for the smallest variation of the highest D<sub>v</sub> values over time, combined with the highest y (Table S1). From the early to the late Cretaceous, the 2 to 5-fold increase in angiosperm D<sub>v</sub> corresponds to a 3 to 5-fold increase in g<sup>max</sup><sub>anat</sub> (Fig. 2b and Table S1). Thus, fossil D<sub>v</sub> provides an estimation of the increase in g<sup>max</sup><sub>anat</sub> over time. Our land surface model does not explicitly represent vegetation traits nor g<sup>max</sup> but only g<sub>w</sub>, the operational stomatal conductance. However, based on the strong

explicitly represent vegetation traits nor  $g_{anat}^{max}$  but only  $g_s$ , the operational stomatal conductance. However, based on the strong relationship between  $g_{anat}^{max}$  and  $g_s$  (Dow et al., 2014; Dow and Bergmann, 2014), one assumes that variations of  $g_s$  due to the long-term evolution of hydraulic and photosynthetic capacities together with that of environmental factors such as pCO<sub>2</sub> would reflect into proportional changes on  $g_{anat}^{max}$ .

# 180 2.3 Experimental setup

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To assess the impact of angiosperm leaf evolution on transpiration and photosynthesis, we performed climate model simulations while varying plant hydraulic or photosynthetic capacity, or both for two contrasted  $pCO_2$  levels (Table 1). The model continental boundary conditions include mid-Cretaceous (115 Ma) paleogeography reconstructions from Sewall et al. (2007). Global vegetation distribution (Fig. 3) is set by establishing a first-order estimate of correspondence between biomes, inferred from paleobotanical data (Sewall et al., 2007), and PFTs (Table S2).

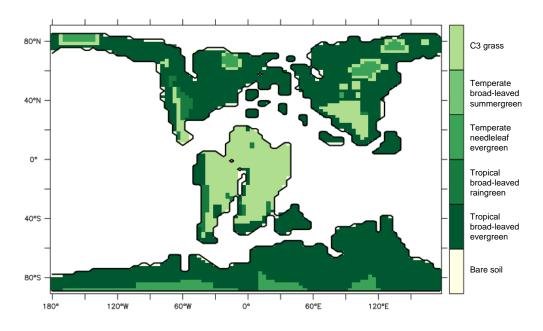


Figure 3. Vegetation distribution in the Aptian simulation configuration : map showing the dominant PFT for each grid cell. For all experiments, this distribution is fixed along the time.



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This configuration allows the carbon cycle to be activated and the LAI to be prognostically calculated as a function of dynamic carbon allocation (Krinner et al., 2005). We set the solar radiation at 99 % of the current value and the orbitals parameters as today. The ocean is prescribed through sea-surface temperatures obtained from an unpublished Aptian fully-coupled simulation obtained with the ESM IPSL-CM5A2 (Sepulchre et al., 2020). To assess the impact of angiosperm trait evolution on transpiration and photosynthesis, we consider (i) two different parameters applied to angiosperm-PFTs only, and (ii) one external forcing ( $pCO_2$ ), that alter the stomata conductance in our land-surface model (Table 1).

Table 1. Definition of the experiments

Name	Parameters representing the fraction of current plant capaci- ties for angiosperms PFTs		pCO <sub>2</sub> (ppm)	Description
	Hydraulic capacity $(\alpha)$	Photosynthetic capacity $(\beta)$		
ANGIO(1120)	1	1	1120	Cretaceous world with the high hydraulic and pho- tosynthetic capacities of modern angiosperms
NOANGIOh(1120)	$1/_{5}$	1	1120	Cretaceous world without the high hydraulic capac- ity of modern angiosperms
NOANGIOp(1120)	1	1/5	1120	Cretaceous world without the high photosynthetic capacity of modern angiosperms
NOANGIOhp(1120	) $\sqrt{1/5}$	$\sqrt{1/5}$	1120	Cretaceous world without the high hydraulic and photosynthetic capacities of modern angiosperms
ANGIO(280)	1	1	280	Cretaceous world with the high hydraulic and pho- tosynthetic capacities of modern angiosperms, un- der preindustrial pCO <sub>2</sub>
NOANGIOh(280)	$1/_{5}$	1	280	Cretaceous world without the high hydraulic ca- pacity of modern angiosperms, under preindustrial pCO <sub>2</sub>
NOANGIOp(280)	1	$1/_{5}$	280	Cretaceous world without the high photosynthetic capacity of modern angiosperms, under preindus-trial pCO <sub>2</sub>
NOANGIOhp(280)	$\sqrt{1/5}$	$\sqrt{1/5}$	280	Cretaceous world without the high hydraulic and photosynthetic capacities of modern angiosperms, under preindustrial pCO <sub>2</sub>





Changes in hydraulic capacity is considered via a change in the coupling factor (fcpl) that describes the slope of the relationship between  $g_s$  and the dynamical driving parameters (namely A, pCO<sub>2</sub> and vapor pressure deficit). Because fcpl is related to the structural conductance, the change in  $g_{anat}^{max}$  can then be empirically represented by a parameter  $\alpha$ , modulating fcpl in the 195  $g_s$  formulation (Eq. (1)). Changes in photosynthetic capacity are taken into account via changes in  $V_{cmax}/V_{jmax}$  ratio (Eq. (2) to (4)). To this end, we use a parameter  $\beta$  that acts directly on V<sub>cmax</sub> (Eq. (3)). We carry out three different factorial experiments using the upper value of changes in  $g_{anat}^{max}$  induced from  $D_v$  proxy as depicted earlier. (1)  $\alpha$  is set to 1/5 while  $\beta$ is 1, accounting for a direct decrease of flowering plant hydraulic capacity by a factor of 5 (NOANGIOh, Table 1). (2)  $\beta$  is set to 1/5 while  $\alpha$  is 1, corresponding to a direct decrease of angiosperm photosynthetic capacity by a factor of 5 (NOANGIOP, Table 1). This sensitivity experiment is supported by the coupling between stomatal conductance and assimilation in the model 200 and by several studies suggested to mimic pre-angiosperm capacities by decreasing modern  $V_{cmax}$  by a factor of 5 (Boyce and Lee, 2010; Lee and Boyce, 2010). Since plant hydraulic and photosynthetic capacities are likely to have co-evolved over the Cretaceous (Franks and Beerling, 2009a; De Boer et al., 2012), (3) we set  $\alpha$  and  $\beta$  to  $\sqrt{1/5}$ , i.e with half the forcing simultaneously applied to the hydraulic and the photosynthetic capacities (NOANGIOhp, Table 1). These three sensitivity tests model three likely Cretaceous worlds without modern-like angiosperms. We perform an additional experiment where the 205 parameters  $\alpha$  and  $\beta$  are set to 1 in order to keep the modern hydraulic and photosynthetic capacity of extant angiosperms (ANGIO, Table 1). Since pCO<sub>2</sub> simultaneously impacts hydraulic and photosynthetic traits evolution on the long-term (Franks and Beerling, 2009a), we repeat this set of experiments for two different extreme  $pCO_2$  forcings. At 1120 ppm, we refer to the mid-Cretaceous (115 Ma) estimates, contemporary with the beginning of the angiosperm radiation. At 280 ppm, we make a sensitivity test to assess the response of pre-angiosperm vegetation to a preindustrial-like pCO<sub>2</sub>. Assuming the  $g_s/g_{anat}^{max}$  ratio 210 is a constant at long-time scales because of plants structural adaptation to their environment, we consider that all the changes in the model operational  $g_s$  arising from the above-mentioned pre-angiosperm parameterizations reflect that of the maximal

anatomic stomatal conductance. The experiments were run for 60 years, a sufficient time to balance gross primary productivity and evapotranspiration. We analyse and show the averaged last 10 years of simulations.

# 215 3 Results

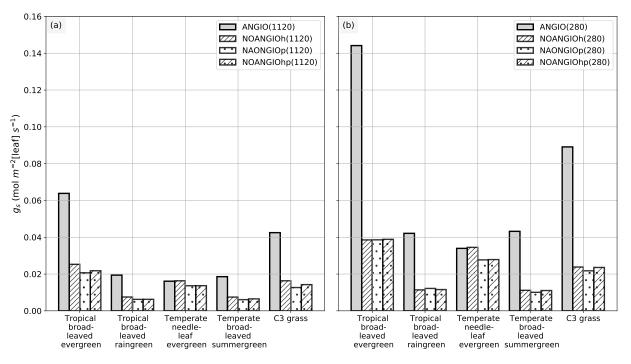
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The leaf stomatal conductance to H<sub>2</sub>O ( $g_s$ , mol m<sup>-2</sup>[leaf] s<sup>-1</sup>) is given by a unit of foliar surface where exposure to sunlight is maximal. Figure 4 shows that leaf stomatal conductance decreases as we sequentially reduce hydraulic (NOANGIOh) or photosynthetic (NOANGIOp) capacities, or both (NOANGIOhp), to mimic the pre-angiosperm world. Reducing the high hydraulic and/or photosynthetic capacities from modern angiosperm leads to a 3-time drop of leaf stomatal conductance on average, a factor in the lowest bracket of the range expected from  $g_{anat}^{max}$  inferred from the fossil record (Fig. 2b). Within every PFT group,  $g_s$  decrease is similar amongst sensitivity experiments and validates our chosen combinations of values for  $\alpha$  and  $\beta$ . The maximal decrease in  $g_s$  is simulated for the tropical broad-leaved evergreen and the C3 grass PFTs which also depict the largest stomatal conductance in the reference ANGIO simulations. Hydraulic and photosynthetic parameters have not been altered for the temperate needleleaf evergreen PFT, as they correspond to gymnosperms. Thus, gymnosperm  $g_s$  was expected





to be constant for each of the p $CO_2$  group of experiments. However,  $g_s$  slightly decreases in the sensitivity experiments for this PFT, suggesting a feedback from the atmosphere-vegetation coupling that ultimately altered soil moisture stress and stomatal conductance.



**Figure 4.** Leaf stomatal conductance to  $H_2O g_s$  (mol m<sup>-2</sup>[leaf] s<sup>-1</sup>) per PFT : (a) ANGIO(1120) (gray bar), NOANGIOh(1120) (diagonal hatched bar), NOANGIOp(1120) (dotted bar), and NOANGIOhp(1120) (diagonal hatched and dotted bar); (b) ANGIO(280) (gray bar), NOANGIOh(280) (diagonal hatched bar), NOANGIOp(280) (dotted bar), and NOANGIOhp(280) (diagonal hatched and dotted bar). The leaf stomatal conductance to  $H_2O$  is the daylight average over the year and the surface grid where each PFT is found.

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Simulations forced with "Cretaceous-like" pCO<sub>2</sub> (1120 ppm) show systematically lower  $g_s$  values than simulations with pCO<sub>2</sub> set at preindustrial (280 ppm). This illustrates the model reproducing the expected closure of stomata as a response to high pCO<sub>2</sub>, in direct link with Eq. (3) and (4). This decrease in  $g_s$  with increasing pCO<sub>2</sub> is the highest (52 % on average) for experiments with the modern-like photosynthetic and hydraulic capacities (ANGIO(1120) versus ANGIO(280)). For these experiments, tropical broad-leaved evergreen PFT reacts to the high pCO<sub>2</sub> with  $g_s$  values dropping from 0.14 mol m<sup>-2</sup>[leaf] s<sup>-1</sup> at 280 ppm to 0.06 mol m<sup>-2</sup>[leaf] s<sup>-1</sup> at 1120 ppm, while that of C3 grasses drops from 0.09 to 0.04 mol m<sup>-2</sup>[leaf] s<sup>-1</sup>. At 1120 ppm, as the leaf stomatal conductance of ANGIO runs is already strongly reduced compared to that at 280 ppm, the

relative decrease in  $g_s$  when modern hydraulic and photosynthetic capacities are reduced is also less important : - 66 % for tropical broad-leaved evergreen at 1120 ppm to be compared to -73 % for the same PFT at 280 ppm (Fig. 4).

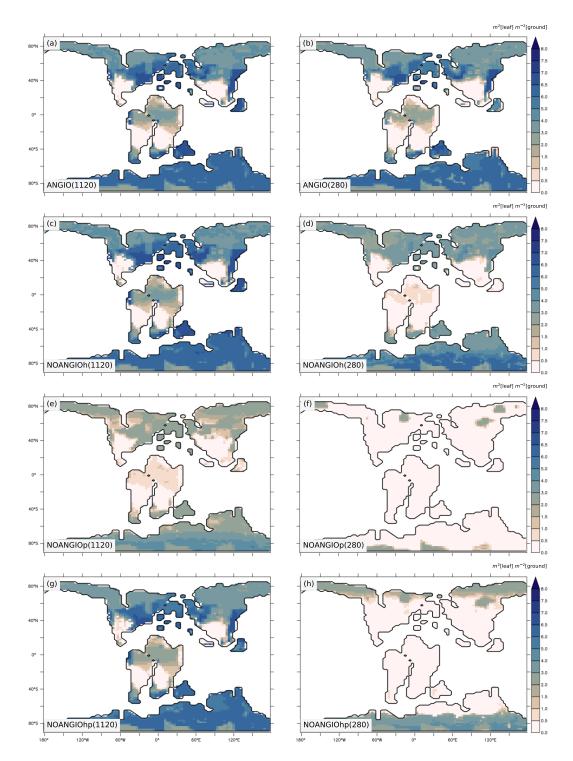




Annual mean leaf area index (LAI) provides an indication of the vegetation ability to grow given the prescribed boundary conditions, i.e. Cretaceous paleogeography, varying  $pCO_2$  and interactions with the atmosphere. For the modern-like vegetation 240 (ANGIO), figures 5a and b show that the highest LAI values are simulated for regions dominated by tropical broad-leaved evergreen, that are set in mid-to-high latitudes in the peculiar Cretaceous configuration (Fig. 3). LAI is also slightly higher at 1120 ppm than at 280 ppm, as a response to  $CO_2$  fertilization effect on photosynthesis that increases biomass production (Fig. 1). The vegetation response to the prescribed perturbations of fcpl and  $V_{cmax}$  strongly differs depending on the pCO<sub>2</sub>. At high pCO<sub>2</sub>, LAI is unchanged with the sole perturbation of the hydraulic capacity (fcpl\*1/5, NOANGIOh, Fig. 5c) meaning that even with a reduced  $g_s$  at the leaf level, the high pCO<sub>2</sub> allows for the fertilization effect to drive biomass growing (C<sub>i</sub> 245 in Eq. (1)). Conversely, V<sub>cmax</sub> perturbation (NOANGIOp, Fig. 5e) induces a strong decrease in the vegetation cover of all angiosperm PFTs, with LAI dropping to values ranging from 1 to 4 m<sup>2</sup>[leaf] m<sup>-2</sup>[ground] compared to LAI ranging from 1 to 8 m<sup>2</sup>[leaf] m<sup>-2</sup>[ground] for ANGIO(1120). Lastly, when both fcpl and V<sub>cmax</sub> perturbations are combined (fcpl\* $\sqrt{1/5}$ and  $V_{cmax} * \sqrt{\frac{1}{5}}$ , NOANGIOhp, Fig. 5g), LAI is mostly unchanged, suggesting that in a high pCO<sub>2</sub> context, slightly reduced 250 hydraulic and photosynthetic capacities do not affect actual assimilation. At high pCO<sub>2</sub>, the impact of reducing  $V_{cmax}$  on photosynthesis is limited because the limiting factor is likely  $V_i$  (and then light) rather than  $V_c$  (and  $C_i$ ) (Eq. (2)). At low pCO<sub>2</sub>, LAI depicts a much stronger response to perturbations. Reducing the hydraulic capacity leads to a global drop of the vegetation cover, with grass-dominated regions averaging 1 m<sup>2</sup>[leaf] m<sup>-2</sup>[ground] and tree-dominated regions barely reaching  $5 \text{ m}^2$ [leaf] m<sup>-2</sup>[ground] except in the very southern latitudes, where no significant change is simulated (Fig. 5d). The strongest signal occurs with the reduction of photosynthetic capacity through  $V_{cmax}$  (NOANGIOp, Fig. 5f). Apart from the unchanged 255 needleleaf PFT, the modified vegetation cannot grow in NOANGIOp experiment at 280 ppm, as shown by LAI values close to zero for every angiosperm PFTs. Simulated LAI in NOANGIOhp shows that at a lower  $V_{cmax}$  associated with a lower fcpl, tropical PFTs also globally collapse, although vegetation is maintained at high latitude (Fig. 5h). At 280 ppm, intercellular  $CO_2$  partial pressure (Eq. (1) to (4)) becomes more limiting than the solar energy whatever the vegetation prescribed. The latitudinal gradient of LAI reduction testifies to another limiting factor to be integrated, which plays at low latitudes but not at 260 high latitudes. Figure S1b shows that the soil moisture stress for transpiration (Fig. 1, Eq. (3) and (4)) is larger in the tropics to mid-latitudes and near zero at high latitudes. Water in the soil available for transpiration is thus a limiting factor of plant growth in the tropics and mid-latitudes at low pCO<sub>2</sub>.







**Figure 5.** Annual mean LAI (m<sup>2</sup>[leaf] m<sup>-2</sup>[ground]) for (a) ANGIO(1120), (b) ANGIO(280), (c) NOANGIOh(1120), (d) NOANGIOh(280), (e) NOANGIOp(1120), (f) NOANGIOp(280), (g) NOANGIOhp(1120) and (h) NOANGIOhp(280).





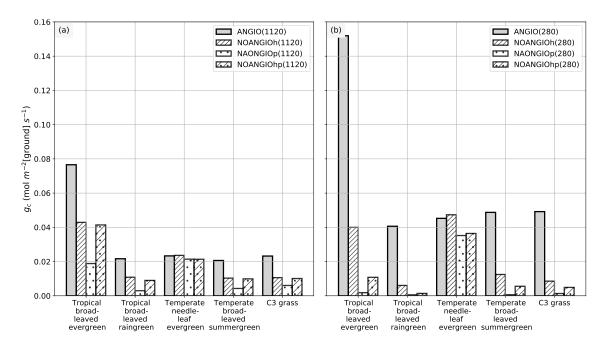


Figure 6. The canopy stomatal conductance to  $H_2O$ ,  $g_c$  (mol m<sup>-2</sup>[ground] s<sup>-1</sup>) per PFT : (a) ANGIO(1120) (gray bar), NOANGIOh(1120) (diagonal hatched bar), NOANGIOp(1120) (dotted bar), and NOANGIOhp(1120) (diagonal hatched and dotted bar); (b) ANGIO(280) (gray bar), NOANGIOh(280) (diagonal hatched bar), NOANGIOp(280) (dotted bar), and NOANGIOhp(280) (diagonal hatched and dotted bar). The canopy stomatal conductance to  $H_2O$  is the daylight average over the year and the surface grid where each PFT is found.

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Albeit the fossil record and empirical models can help to infer the foliar maximal stomatal conductance to H<sub>2</sub>O, the land surface model is required to explore the stomatal conductance to  $H_2O$  at the canopy level that is given by Eq. (6) and (7):  $g_c$  is inversely proportional to the canopy resistance  $r_c$ . The canopy stomatal conductance is given by surface unit of ground and integrates the stomatal conductance for shadow leaves at the bottom and sunny leaves at the top of the canopy. Figures 4 and 6 show significant differences between  $g_s$  and  $g_c$  in respect to the pCO<sub>2</sub> prescribed: while  $g_s$  decreases by a factor of 3 for each pre-angiosperm sensitivity experiment compared to the reference (Fig. 4), the decrease in  $g_c$  is not equal through 270 experiments (Fig. 6). The dramatic drop in  $g_c$  when reducing the high photosynthetic capacity (NOANGIOp) is well explained by the significant LAI decrease (Fig. 5e and f). Besides, the effect of pCO<sub>2</sub> is more complex at the canopy scale. Indeed,  $g_c$ is still reduced in the perturbed experiments, but is not systematically lower at 1120 ppm than at 280 ppm (Fig. 6) as opposed to  $g_s$  (Fig. 4). NOANGIOh simulations do not show a change in  $g_c$  between 280 ppm and 1120 ppm. A decrease in fcpl by a factor of 5 implies a reduction of the LAI at 280 ppm (Fig. 5d) because plants cannot assimilate enough carbon to sustain their 275 biomass ( $C_i$  in Eq. (1) to (4)) whereas it is not the case at 1120 ppm (Fig. 5c). Indeed, at the canopy scale, the closing of the stomata at high  $pCO_2$  is compensated by the decrease in LAI at low  $pCO_2$ . In the NOANGIOp and NOANGIOp experiments,  $g_c$  is higher at 1120 ppm than at 280 ppm. By increasing the carbon assimilation during photosynthesis (Eq. (1)), high pCO<sub>2</sub> allows for higher vegetation cover (Fig. 5e and g) and higher canopy stomatal conductance whereas low  $pCO_2$  together with low photosynthetic capacity imply a collapse of the vegetation (Fig. 5f and h).





- 280 LAI changes described earlier act as a feedback between g<sub>s</sub> and g<sub>c</sub> (Fig. 1). At low pCO<sub>2</sub>, the LAI decrease between the ANGIO and any of the three perturbed experiments (Fig. 5d, f and h) strengthens the initial decrease in g<sub>s</sub> (positive feedback, Fig. 4b and 6b). Mean g<sub>s</sub> decreases by 69 %, 73 % and 69 % while mean g<sub>c</sub> decreases by 72 %, 97 % and 91 % respectively for NOANGIOh, NOANGIOp and NOANGIOhp compared to ANGIO (Fig. 4b and 6b). Indeed, decreasing V<sub>cmax</sub> under low pCO<sub>2</sub> implies a reduction of the plant capacity to assimilate carbon (Eq. (3)) and directly impacts the GPP (Fig. in S2f and h) and then the LAI at the canopy scale (Fig. 5f and h). However, decreasing fcpl and thus g<sub>s</sub> reduce the CO<sub>2</sub> concentration at the
- chloroplast level ( $C_i$  in Eq. (1)) and have only an indirect effect on GPP (Fig. S2d) and thus on LAI (Fig. 5d). At high pCO<sub>2</sub>, the LAI is almost sustained for NOANGIOh and NOANGIOhp compared to ANGIO (Fig. 5a, c and g) because the assimilation remains high when  $C_i$  is not the limiting factor (Fig. S2a, c and g). The latter lessens the initial decrease in  $g_s$  (Fig. 6a) on  $g_c$ (Fig. 4a). Nevertheless, NOANGIOp(1120) experiment shows a much lower  $g_c$  than the two previous experiments because of

290 the direct impact of decreasing  $V_{cmax}$  on the LAI (Fig. 5e). Therefore, comparing  $g_c$  of perturbed simulations to that of the reference allows us to account for the structural conductance linked to plant trait evolution at the canopy level.

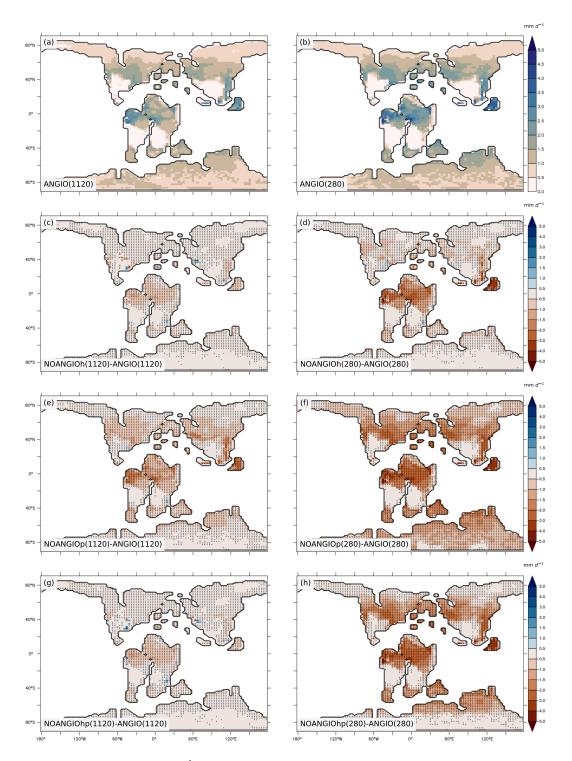
As shown in Eq. (5), transpiration rate is controlled by (i) the atmospheric evaporative demand, that depends on air humidity, the surface temperature as well as the aerodynamic resistance, and (ii) the capacity of plants to transpire, driven by the canopy conductance. For both pCO<sub>2</sub> cases, experiments with modern-like vegetation depict the highest transpiration rates in the tropics and the mid-latitudes, where they reach up to 2.5 and 3 mm day<sup>-1</sup> at 1120 and 280 ppm, respectively (Fig. 7a and b). These regions have optimal conditions in water availability in the soil (Fig. S1b) and light, resulting in higher canopy stomatal conductance (Fig. S3a and b) together with a high atmospheric evaporative demand (Fig. S4a and b) as a response to high

temperatures. Transpiration is also slightly strengthened (+ 0.2 mm day<sup>-1</sup>) at low pCO<sub>2</sub> compared to high pCO<sub>2</sub> (Fig. 7a and b) as a consequence of higher  $g_c$  (Fig. 4 and 6).

- Parameterizing the vegetation without the modern angiosperms hydraulic and photosynthetic capacities systematically leads to lower transpiration rates (Fig. 7). Overall, in line with changes depicted for LAI and  $g_c$ , transpiration rates react stronger in a 280 ppm world than in a 1120 ppm world to decreasing hydraulic and photosynthetic capacities. At 280 ppm, NOANGIOh shows a decrease of 0.6 mm d<sup>-1</sup> (- 44 %) in transpiration compared to ANGIO, especially over equatorial Gondwana and
- 305 paleo Southeast Asia, whereas the decrease is limited to 0.3 mm d<sup>-1</sup> (- 24 %) at 1120 ppm (Fig. 7c and d) as a response to  $g_c$  changes described earlier (Fig. 6). Transpiration also significantly drops when photosynthetic capacity alone is reduced (NOANGIOp, Fig. 7e and f). At 1120 ppm, transpiration drops by 0.5 mm d<sup>-1</sup> (-53 %) (Fig. 7e). The signal is stronger at 280 ppm, where a complete collapse of transpiration is simulated (Fig. 7f). This latter result is a direct consequence of the LAI collapse described earlier (Fig. 5e and f). Combining reduction in photosynthetic and hydraulic capacities (NOANGIOp)
- leads to little decreases in transpiration rate at 1120 ppm (Fig. 7g), comparable to NOANGIOh (Fig. 7c), because the high pCO<sub>2</sub> prevents the decrease in carbon assimilation (Fig. 5g) and canopy stomatal conductance (Fig. 6). Conversely, at low pCO<sub>2</sub>, the limitation of C<sub>i</sub> (Eq. (1), (3) and (4)) implies a collapse of LAI (Fig. 5h) and g<sub>c</sub> (Fig. 6) over the tropics and midlatitudes, and transpiration is near zero for NOANGIOhp, with a decrease of 1 mm d<sup>-1</sup> (-81 %) compared to ANGIO (Fig. 7h).







**Figure 7.** Annual mean transpiration rate (mm d<sup>-1</sup>) for (a) ANGIO(1120) and (b) ANGIO(280), anomalies of annual mean transpiration rate (mm d<sup>-1</sup>) for (c) NOANGIOh(1120) vs ANGIO(1120), (d) NOANGIOh(280) vs ANGIO(280), (e) NOANGIOp(1120) vs ANGIO(1120), (f) NOANGIOp(280) vs ANGIO(280), (g) NOANGIOhp(1120) vs ANGIO(1120) and (h) NOANGIOhp(280) vs ANGIO(280). The t-test 95 % confidence level anomalies are given by dots.





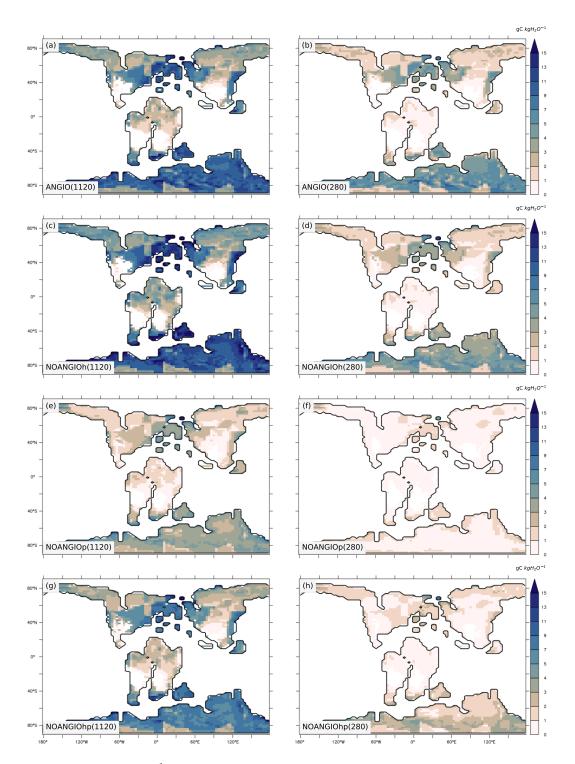
- 315 Transpiration anomalies come from variations in the atmospheric evaporative demand (Fig. S4) combined with that of the canopy stomatal conductance (Fig. S3), arising from g<sub>s</sub> and LAI changes which are modulated by the soil moisture stress for transpiration and light. Transpiration anomalies are located over the paleo-tropics (Fig. 7c and d) for NOANGIOh compared to ANGIO experiments because decreasing hydraulic capacity by a factor of 5 acts mainly on wet regions where plants efficiently transpire in the ANGIO world (Fig. 1 and Eq. (1) to (4)), while arid belt regions are less sensitive to any change in g<sub>c</sub>. However, mid-latitude do not show transpiration anomalies because the decrease in g<sub>c</sub> (Fig. 6, S3c and d) is compensated by the increase
- in atmospheric evaporative demand (Fig. S4c and d) that bolsters transpiration rate. In contrast, decreasing photosynthetic capacity (NAONGIOp) acts more globally on terrestrial plants and explains the widespread transpiration anomalies (Fig. 7e and f), always modulated by the atmospheric evaporative demand (Fig. S4e and f). At 1120 ppm, NOANGIOhp shows the same pattern as NOANGIOh (Fig. 7c and g) because plants are more sensitive to reduction in fcpl than to reduction in  $V_{cmax}$  (C<sub>i</sub>
- 325 is not limiting) while the decrease in transpiration is more extended at 280 ppm (Fig. 7h) because it arises mainly from the decrease of  $V_{cmax}$  at low pCO<sub>2</sub> that drives carbon assimilation to decline (Eq. (2) and (3)).

The annual mean water use efficiency (WUE) is the ratio of annual mean GPP to annual mean transpiration over each grid point (gC kgH<sub>2</sub>O<sup>-1</sup>). It pictures the capacity of vegetation to maximise carbon uptake while minimizing water loss and thus is a good indicator of plants adaptation to their environment. For the modern-like vegetation (ANGIO), maximal values of WUE are found in the mid to high-latitudes where optimal conditions of water and light (Fig. 8a and b, Fig. S1b) are available for the vegetation. WUE increases with pCO<sub>2</sub> : while it barely exceeds 10 gC kgH<sub>2</sub>O<sup>-1</sup> at 280 ppm, it reaches more than 15 gC kgH<sub>2</sub>O<sup>-1</sup> at 1120 ppm. This is explained by the CO<sub>2</sub> fertilization effect generated by the high pCO<sub>2</sub> on C<sub>i</sub> (Eq. (2) to (4)) that strengthens plant productivity. At 1120 ppm, the highest value of WUE is found for NAONGIOh, with a 1.7 gC kgH<sub>2</sub>O<sup>-1</sup> (+

- 335 30 %) increase compared to ANGIO (Fig. 8a and c). Indeed, reducing the hydraulic capacity does not imply a change in GPP which remains similar to that of ANGIO(1120) (Fig. S2a and c) but rather generates a slight decrease in transpiration rate (Fig. 7c), explaining the increase of WUE. Therefore, plants with lower hydraulic capacities than today are better adapted to the high pCO<sub>2</sub> environment. For NOANGIOhp(1120), the WUE is partly reduced compared to ANGIO(1120) but stays at a high level (Fig. 8a and g). It results from the larger decrease in GPP than in transpiration rate (Fig. S2g and 7g), as GPP is more sensitive to
- 340 the  $V_{cmax}$  reduction (Eq. (2)) than transpiration is to the fcpl reduction (Eq. (1) and (5)). In contrast, WUE is largely degraded in NOANGIOp(1120) compared to ANGIO(1120) (Fig, 8a and e). Both GPP (Fig. S2a and e) and transpiration rate (Fig. 7a and e) significantly drop when reducing photosynthetic capacity (NOANGIOp) at high pCO<sub>2</sub>. However,  $V_{cmax}$  acts directly on the carbon assimilation (Eq. (2) and (3)) while it is more indirect on the stomatal conductance, thus implying that the reduction of  $V_{cmax}$  has a larger effect on GPP than on transpiration rate. In contrast to experiments at high pCO<sub>2</sub>, ANGIO(280) gives
- the highest WUE at low pCO<sub>2</sub>. NOANGIOh(280) depicts a lower WUE than ANGIO(280) (Fig. 8b and d) which demonstrates that plants with lower hydraulic capacity than today are less adapted to the low pCO<sub>2</sub> environment. At low pCO<sub>2</sub>, the low C<sub>i</sub> decreases GPP (Eq. (3), Fig. S2b and d) while it increases  $g_s$  (Eq. (1), Fig. 4), modulating the transpiration decrease (Fig. 7d).







**Figure 8.** Annual mean WUE ( $gC kgH_2O^{-1}$ ) for a) ANGIO(1120), b) ANGIO(280), c) NOANGIOh(1120), d) NOANGIOh(280), e) NOANGIOp(1120), f) NOANGIOp(280), g) NOANGIOhp(1120) and h) NOANGIOhp(280).





However, at low pCO<sub>2</sub>, WUE collapses to very low values for NOANGIOp and NOANGIOhp (Fig. 8f and h), that is driven by the large decrease in GPP when combining the low C<sub>i</sub> to the reduction of V<sub>cmax</sub> (Fig. S2f and h). Once the photosynthetic
capacity is decreased, changes in GPP are the main contributor to the changes in WUE whatever the pCO<sub>2</sub> level prescribed.

# 4 Discussion

### 4.1 How to better account for the pre-angiosperm conductance traits in land surface models?

Fossil maximal anatomic stomatal conductance has been widely used to estimate the maximum of water flow through the stomata before and after the angiosperm radiation. Still, determining how a 5-time increase in maximal anatomic stomatal conductance translates into actual flux at the top of the canopy is challenging. We show that the complete response of the vegetation to evolving physiological and morphological traits is modulated by environmental factors such as pCO<sub>2</sub>, light and water availability in the soil (Fig. 1).

The simplest representation of pre-angiosperm vegetation is to account for the decrease of maximal anatomic stomatal conductance by a factor of 5, consistent with the fossil records, directly by applying this factor to fcpl in the calculation of the leaf stomatal conductance (Eq. (1)). Our results show that decreasing fcpl of angiosperms at high pCO<sub>2</sub> does not change plant

- 360 leaf stomatal conductance (Eq. (1)). Our results show that decreasing fcpl of angiosperms at high pCO<sub>2</sub> does not change plant photosynthesis (Fig. 5c and S2c) but slightly decreases transpiration (Fig. 7c), driving WUE to increase (Fig. 8c) compared to the modern angiosperm prescription. Hence, a lower maximal stomatal conductance at high pCO<sub>2</sub> appears as an advantage compared to modern angiosperm because of a better optimization of carbon uptake over water loss. At low pCO<sub>2</sub>, both transpiration (Fig. 7d) and photosynthesis are decreased because of the positive feedbacks of the LAI on the canopy stomatal
- 365 conductance, which strengthens the initial reduction of leaf stomatal conductance compared to the modern vegetation. Despite the absence of a direct proxy for fossil plant maximum  $V_{cmax}$ , several studies have suggested to mimic the pre-angiosperm capacities by decreasing modern  $V_{cmax}$  (Boyce and Lee, 2010; Lee and Boyce, 2010), rather than fcpl, by a factor of 5. This approach is supported by our knowledge of modern plant processes that stomatal conductance interacts with assimilation in order to optimize carbon gain against water loss (Bonan, 2015) and is made possible by the coupling between stomatal con-
- 370 ductance and photosynthetic capacity in our land-surface model (Farquhar et al., 1980; Ball et al., 1987; Krinner et al., 2005; Yin and Struik, 2009). However, applying this method leads vegetation cover to decrease at high pCO<sub>2</sub> (Fig. 5e and S2e) or even collapse at low pCO<sub>2</sub> (Fig. 5f and S2f), which is not recorded in the fossil record. As a consequence, transpiration rates (Fig. 7e and f) and WUE (Fig. 8e and f) significantly decrease at high pCO<sub>2</sub> and are zero at low pCO<sub>2</sub> compared to the modern vegetation. Hence, taking into account stomatal conductance reduction only through V<sub>cmax</sub> reduction does not appear to be
- adequate. However, several studies suggested a decrease of  $V_{cmax}$  with increasing pCO<sub>2</sub> (Ainsworth and Rogers, 2007) driven by (i) a coevolution between stomatal conductance and  $V_{cmax}$  through time (Franks and Beerling, 2009a; De Boer et al., 2012) and (ii) the photosynthesis coordination theory that states that plants optimize  $V_{cmax}$  to be near the co-limitation between carboxylation rate and RuBp regeneration (Maire et al., 2012). This theory has been recently improved considering also the cost related to stomatal conductance (Stocker et al., 2020). This  $V_{cmax}$  limitation is related to the high energetic (and then
- 380 respiration) cost needed to maintain a high level of Rubisco (acquisition of nitrogen). Rather than the two extreme cases that





decrease hydraulic or photosynthetic capacity of angiosperms by a factor of 5, we consider a covariation of fcpl and V<sub>cmax</sub>, by applying half the forcing given by the fossil records (i.e. √1/5) to fcpl directly and the other half to V<sub>cmax</sub>. Experimental studies on extant plant types (Lin et al., 2015) have shown differences in water-use strategy between modern angiosperm trees and gymnosperm trees. They argue that modern angiosperm trees have 2 times higher stomatal conductance sensitivity response to driving factors than gymnosperm trees, showing that our choice for fcpl\*√1/5 together with V<sub>cmax</sub>\*√1/5 seems the most realistic. When applying this factor jointly to fcpl and V<sub>cmax</sub> at a high pCO<sub>2</sub>, our results suggest that vegetation is barely impacted, with a slight reduction of GPP (Fig. S2g), that remains enough to sustain the LAI (Fig. 5g) and a relatively

- high WUE (Fig. 8g) compared to the modern vegetation (Fig. 5a and 8a). Conversely at low pCO<sub>2</sub>, LAI collapses (Fig. 5h) and GPP, transpiration and WUE reach zero (Fig. S2h, 7h and 8h) as a response to carbon assimilation drop.
- 390 At 280 ppm, sensitivity tests with a low photosynthetic capacity or together with a low hydraulic capacity can be considered as extrema where plants cannot grow (Fig. 5f and h, S2f and h) whereas experiment with modern trait vegetation corresponds to a maximum in plant productivity and transpiration (Fig. 5b, 7b and 8b). At 1120 ppm, two sets of parameters simulate a sustainable paleovegetation productivity : lowered hydraulic capacity only or lowered hydraulic and photosynthetic capacities together (Fig. 5c and g, S2c and g). However, lowering only the hydraulic capacity while keeping the high V<sub>cmax</sub> as in
- 395 the modern vegetation induces a nitrogen cost. Although our model does not represent the nitrogen cycle, we infer that this supplementary cost would lead to a decrease in productivity when simulating the pre-angiosperm vegetation with a modern photosynthetic capacity together with a low hydraulic capacity. Taking into account the differentiated response of vegetation to a declining pCO<sub>2</sub>, we suggest that simultaneously decreasing fcpl and V<sub>cmax</sub> by a factor of  $\sqrt{1/5}$  is the most realistic representation of pre-angiosperm vegetation. The latter is consistent with previous study of Franks and Beerling (2009a) suggesting
- 400 that past fluctuations in pCO<sub>2</sub> acted as a forcing on both V<sub>cmax</sub> and stomatal conductance. Finally, our study confirms the hypothesis that paleovegetation has evolved from a likely stage with a relatively low stomatal conductance (Fig. 4a) and V<sub>cmax</sub> under high pCO<sub>2</sub> while sustaining high productivity (Fig. 5g, 7g and 8g) to a stage of high stomatal conductance (Fig. 4b) and V<sub>cmax</sub> as in the modern vegetation, in order to preserve high productivity under low pCO<sub>2</sub> (Fig. 5b, 7b and 8b). The study also shows that a reasonable way to account for pre-angiosperm conductance traits is to use
- 405 a model that fully describes the coupling between stomatal conductance and plant productivity from leaf to the canopy scale. Furthermore, we show that decreasing hydraulic and/or photosynthetic capacities does not coincide with a decrease of the leaf operational stomatal conductance to the same extent. Indeed, accounting for a decrease by a factor of 5, given by the maximal bound of the range expected from the maximal anatomic stomatal conductance, leaf stomatal conductance is only 3-time lower than the reference. We have a complex response because of the coupling between stomatal conductance and assimilation.
- 410 While some previous studies directly decrease the transpiration rate by the factor of vein density changes from the fossil record (Boyce et al., 2009; Boyce and Lee, 2017; Boyce et al., 2010), we suggest to explicitly represent changes in hydraulic and photosynthetic capacities. Nevertheless, when embedding parallel changes on pCO<sub>2</sub>, leaf stomatal conductance of plants with reduced hydraulic and photosynthesis capacities at high pCO<sub>2</sub> is nearly 6-time lower than the reference one at low pCO<sub>2</sub>. It confirms that accounting for pCO<sub>2</sub> changes is mandatory to model the evolution of angiosperm leaf traits.





# 415 **4.2** Do the high hydraulic and photosynthetic capacities of angiosperms provide a selective advantage compared to the other plants under decreasing pCO<sub>2</sub>?

Our work relies upon the assumption that stomata aperture maximizes carbon gain while minimizing water loss (Bonan, 2015). Both photosynthesis and stomatal conductance to H<sub>2</sub>O are sensitive to environmental variables such as light, pCO<sub>2</sub> and water availability in the soil (Fig. 1). As pCO<sub>2</sub> has varied a lot through the Cretaceous (Fletcher et al., 2008; Wang et al., 2014),
plants had to adjust their stomatal conductance. Our study suggests that having a high stomatal conductance, which means a large vein density and a high V<sub>cmax</sub>, provides little advantage compared to a low stomatal conductance under high pCO<sub>2</sub>. In contrast, having a high stomatal conductance under low pCO<sub>2</sub> may confer a competitive advantage over plants with limited stomatal conductance to assimilate carbon. But this higher stomatal conductance should be linked to an increase in V<sub>cmax</sub> to sustain growth. In that sense, fossil records provide evidence of plant traits evolution during the angiosperm radiation. Fossils
of basal angiosperms show that vein density was as low as the other plant types because having a high vein density under high pCO<sub>2</sub> was not necessary for the plants to grow. Then, the pCO<sub>2</sub> likely declined (Fletcher et al., 2008; Wang et al., 2014). At that time, we confirm the hypothesis that angiosperms evolved towards leaves more and more densely irrigated together with a more efficient biochemistry that allowed them to have an increasingly stomatal conductance and photosynthetic capacity to

430 given a competitive advantage to angiosperms compared to gymnosperms dominating the vegetation of the period to colonize almost all the terrestrial ecosystems. Our results are consistent with that of Franks and Beerling (2009a), which have shown that WUE co-evolves positively with variations in  $pCO_2$  over the Phanerozoic : periods with high  $pCO_2$  strengthen GPP, meanwhile a potential decrease of transpiration rate by the closing of the stomata. They also show that even after the evolution of angiosperm leaf morphology and biochemistry, WUE is estimated to have been at its lowest level since the Carboniferous.

counteract the effect of  $pCO_2$  decrease on carbon assimilation. Among others, this evolution of physiological leaves traits has

- Our model consistently represents the range of WUE deducted by Franks and Beerling (2009a) under different  $pCO_2$ : between 5 and 9 gC kgH<sub>2</sub>O<sup>-1</sup> (Fig. 8a and b). Moreover, there is a likely co-adaptation of stomatal traits and leaf venation which implies a better optimisation of carbon gain against water loss (De Boer et al., 2012). Progressively, under decreasing pCO<sub>2</sub>, angiosperms with high stomatal density and low stomatal size (Franks and Beerling, 2009a, b) likely invested increasingly energy in building more and more veins to sustain the higher stomatal conductance and then carbon assimilation, while other
- 440 plants did not. The innovation of angiosperms in densely water transport networks could have become a necessity to support higher stomatal conductance and prevent plant desiccation (De Boer et al., 2012).

#### 4.3 What are the limitations of our modelling choices?

As a first step toward understanding the impact of trait evolution on the hydraulic and photosynthetic capacities of angiosperms, we have chosen to simulate the Aptian (115 Ma) because this time period corresponds to the first step of increasing vein density found in the fossil record (Feild et al., 2011a). To get an exhaustive view of the angiosperm evolution, future studies will benefit from considering similar experiments with boundary conditions set several million years before and after the Aptian. Specifically, exploring cold and warm extremes of the Cretaceous, such as the Cenomanian-Turonian (95 Ma) and the Maastrichtian





(70 Ma) would be valuable, as climate and  $pCO_2$  have been shown to vary a lot during these periods (Ladant and Donnadieu, 2016).

- 450 The vegetation map we used (Fig. 3) results from two efforts of (i) compilation and spatialization of the Aptian paleobotanical records (Sewall et al., 2007) and (ii) conversion of the fossil data into plant functional types combination (Table S1). Each of these two steps include uncertainties that can propagate into our results, but can be hardly quantified. We acknowledge that the prescribed vegetation cover, especially in the tropics, can potentially alter the radiative balance and the hydrological cycle (e.g., Port et al., 2016). It is however unlikely that the Aptian vegetation cover would be very different from the one provided
- 455 by Sewall et al. (2007), given the compilation effort made for this reconstruction. Further studies could still circumvent this potential issue by running a fully dynamical vegetation model, i.e. by allowing PFTs to spatially settle in regions where the simulated climate is the most appropriate.

Recent studies about Paleozoic vegetation transitions indicate that differences in transpiration rates also arise from the ratio of carbon over nitrogen (White et al., 2020; Richey et al., 2021). As mentioned earlier, the ORCHIDEE version we used does not

460 explicitly represent the nitrogen cycle, preventing us from considering the additional cost to maintain high photosynthetic capacity with lower hydraulic capacity. The recent developments of a new version of ORCHIDEE that does include the nitrogen cycle (Vuichard et al., 2019) will help to account for this process, provided that good constraints can be obtained regarding the C:N ratio of Cretaceous vegetation and soils.

Through the use of the coupled LMDZ and ORCHIDEE models, our approach includes the pivotal coupling between atmo-

- 465 sphere and vegetation. However by using fixed sea-surface temperatures, we neglect the feedbacks from the ocean-atmosphere coupling that could occur as a response to simulated changes in vegetation cover. Although sensitivity experiments with strong changes in vegetation suggested ocean feedbacks could play a significant role on the continental hydrological cycle (Davin and de Noblet-Ducoudré, 2010), our choice was motivated by (i) the will to focus on first-order continental processes, (ii) the computing cost required to equilibrate fully coupled simulations that typically require more than 3000 simulated years (Sepul-
- 470 chre et al., 2020), the fact that comparable studies either used fixed-SSTs (Boyce and Lee, 2010; Lee and Boyce, 2010) or slab oceans (White et al., 2020).

Finally our parameterization of stomatal conductance in ORCHIDEE from Yin and Struik (2009) (Eq. (1)) is semi-empirical. A refinement of our modelling approach would be to use a stomatal conductance model based on optimisation theory, that explicitly describes the stomata functioning so as to optimise carbon gain against water loss (Medlyn et al., 2011; Buckley and

475 Mott, 2013; Buckley, 2017). In particular, the model we use here simply links external forcing to leaf stomatal conductance by an empirical term of coupling fcpl that describe all the processes related to stomatal conductance. The  $\alpha$  factor applied in Eq. (1) to the coupling factor (fcpl) does not fully represent the change in the maximal anatomic stomatal conductance as it should be considering changes in vein density (stomatal density and size). It emphasizes the need, in the future, to improve the parameterization of stomatal conductance in global mode by explicitly modelling both structural and dynamic conductance.





#### 480 5 Conclusions

In line with recent studies focusing on Paleozoic vegetation transitions (White et al., 2020; Richey et al., 2021), the purpose of our study is to better represent past vegetation in earth system models by implementing "paleo-traits" in the vegetation parameterizations. Our approach involves an atmosphere-vegetation model, which couples stomatal conductance and carbon assimilation, with an ecophysiological model based on angiosperm fossil records. Here, it allows us to evaluate three different paleovegetation prescriptions under two end-member scenarios of pCO<sub>2</sub> for the Cretaceous. We show that the simulated vegetation cover, transpiration rate and water use efficiency are sensitive to the paleovegetation trait prescribed. Only accounting for hydraulic capacity reduction provides no significant change in LAI, GPP and transpiration, while slightly increasing WUE at high pCO<sub>2</sub>. In contrast, global transpiration decreases at low pCO<sub>2</sub> because of the positive feedback between LAI and stomatal conductance. On the other hand, only accounting for photosynthetic capacity reduction gives a substantial decrease or even a

- 490 collapse of vegetation at high or low pCO<sub>2</sub> respectively, which is not recorded in the fossil. Combining a reduction of hydraulic capacity with that of photosynthetic capacity does not affect the plant productivity and LAI at high pCO<sub>2</sub> while vegetation collapses at low pCO<sub>2</sub>. All the results taken together demonstrate that under high pCO<sub>2</sub> the reduced stomatal conductance of the pre-angiosperm vegetation is not a limiting factor on productivity. It also shows that high values of V<sub>cmax</sub> as observed in modern angiosperms do not enhance plant productivity, whereas maintaining the high V<sub>cmax</sub> likely requires higher leaf nitrogen
- 495 concentration and higher energy demand. Therefore, the combining decrease of hydraulic and photosynthetic capacities seems the most realistic physiological parameterization for pre-angiosperms in the specific high pCO<sub>2</sub> context of the Cretaceous. This is supported by evidence of coevolution inferred from previous studies (Franks and Beerling, 2009a, b) and the ratio of stomatal conductance between modern angiosperms and gymnosperms from in-situ experiments (Lin et al., 2015). This result is also consistent with recent studies on coordination theory (Maire et al., 2012; Stocker et al., 2020).
- 500 Our study also suggests that pre-angiosperm vegetation with low hydraulic and photosynthetic capacities was adapted to high  $pCO_2$  by sustaining productivity and a high WUE. Conversely, it was not adapted to lower  $pCO_2$  as GPP collapses. Modelling the full coupling between GPP and stomatal conductance allows to understand why increasing both structural conductance and maximum photosynthetic capacity, even at an expense of a possible increasing water loss, was a selective advantage with decreasing  $pCO_2$  and is the likely explanation for observed increasing structural conductance of angiosperms since the Cre-
- taceous, consistent with previous studies (Franks and Beerling, 2009a, b; Boyce and Zwieniecki, 2012; Brodribb and Feild, 2010). From a low stomatal conductance (low vein density and  $V_{cmax}$ ) similar to that of gymnosperms under high pCO<sub>2</sub>, angiosperms evolve towards a high stomatal conductance (high vein density and  $V_{cmax}$ ) to counteract the effect of the pCO<sub>2</sub> decrease on carbon assimilation.

While this study provides clues on how to account for angiosperm evolutionary traits in paleoclimate simulations, further work

510 is needed to assess the potential climate effects of the Cretaceous angiosperm leaf evolution, especially on the hydrological cycle and the energy balance at the land surface. Furthermore, allowing dynamic vegetation would be an important future refinement of this research to model feedbacks between vegetation and climate. With such a model, we may question the veracity of the reciprocity with the human-induced increase in pCO<sub>2</sub>. Building leaves with high vein density and  $V_{cmax}$  induces





an investment in nutrients and energy that could be an extra cost for plants without benefit under a higher pCO<sub>2</sub>. That would suggest that plants should evolve back to a reduced stomatal conductance and  $V_{cmax}$ . However, the pace of the current increase in pCO<sub>2</sub> is dramatically higher than the million-year time scale of Cretaceous changes, and likely incompatible with a genetic plant adaptation. Whether extant plants will be able to adjust their physiological and morphological traits or not to the human perturbation is a challenging question for future studies.

Data availability. We provide the simulation outputs in the online repository : http://doi.org/10.5281/zenodo.4773541

520 *Author contributions.* JB, PS, NVIO and NVUI designed the work. Under supervision of PS, NVIO and NVUI, JB performed the simulations, processed the data, wrote the manuscript and drew the figures, except Fig. 1 drawn by PS. All authors reviewed and approved the manuscript.

Competing interests. The authors declare that they have no conflict of interest.

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#### References

- Ainsworth, E. A. and Rogers, A.: The response of photosynthesis and stomatal conductance to rising [CO2]: mechanisms and environmental interactions, Plant Cell Environ., 30, 258–270, https://doi.org/10.1111/j.1365-3040.2007.01641.x, 2007.
- 530 Ball, J. T., Woodrow, I. E., and Berry, J. A.: A model predicting stomatal conductance and its contribution to the control of photosynthesis under different environmental conditions, in: Progress in photosynthesis research, pp. 221–224, Springer, https://doi.org/https://doi.org/10.1007/978-94-017-0519-6\_48, 1987.
  - Bathiany, S., Claussen, M., Brovkin, V., Raddatz, T., and Gayler, V.: Combined biogeophysical and biogeochemical effects of large-scale forest cover changes in the MPI earth system model, Biogeosciences, 7, 1383–1399, https://doi.org/https://doi.org/10.5194/bg-7-1383-2010, 2010.
- 535 2

540

Betts, R. A., Cox, P. M., Lee, S. E., and Woodward, F. I.: Contrasting physiological and structural vegetation feedbacks in climate change simulations, Nature, 387, 796–799, https://doi.org/https://doi.org.insu.bib.cnrs.fr/10.1038/42924, 1997.

Bonan, G.: Ecological climatology: concepts and applications, Cambridge University Press, 2015.

Boyce, C. K. and Lee, J.-E.: An exceptional role for flowering plant physiology in the expansion of tropical rainforests and biodiversity, P. Roy. Soc. B-Biol. Sci., 277, 3437–3443, https://doi.org/10.1098/rspb.2010.0485, 2010.

- Boyce, C. K. and Lee, J.-E.: Plant evolution and climate over geological timescales, Annu. Rev. Earth Pl. Sc., 45, 61–87, https://doi.org/https://doi.org/10.1146/annurev-earth-063016-015629, 2017.
  - Boyce, C. K. and Zwieniecki, M. A.: Leaf fossil record suggests limited influence of atmospheric CO2 on terrestrial productivity prior to angiosperm evolution, P. Natl. Acad. Sci. USA, 109, 10403–10408, https://doi.org/https://doi.org/10.1073/pnas.1203769109, 2012.
- 545 Boyce, C. K., Brodribb, T. J., Feild, T. S., and Zwieniecki, M. A.: Angiosperm leaf vein evolution was physiologically and environmentally transformative, P. Roy. Soc. B-Biol. Sci., 276, 1771–1776, https://doi.org/https://doi.org/10.1098/rspb.2008.1919, 2009.
  - Boyce, C. K., Lee, J.-E., Feild, T. S., Brodribb, T. J., and Zwieniecki, M. A.: Angiosperms Helped Put the Rain in the Rainforests: The Impact of Plant Physiological Evolution on Tropical Biodiversity1, Ann. Mo. Bot. Gard., 97, 527–540, https://doi.org/https://doi.org/10.3417/2009143, 2010.
- 550 Braconnot, P., Joussaume, S., Marti, O., and De Noblet, N.: Synergistic feedbacks from ocean and vegetation on the African monsoon response to mid-Holocene insolation, Geophys. Res. Lett., 26, 2481–2484, https://doi.org/https://doi.org/10.1029/1999GL006047, 1999.
  - Brodribb, T. J. and Feild, T. S.: Leaf hydraulic evolution led a surge in leaf photosynthetic capacity during early angiosperm diversification, Ecol. Lett., 13, 175–183, https://doi.org/https://doi.org/10.1111/j.1461-0248.2009.01410.x, 2010.

Brodribb, T. J., Feild, T. S., and Jordan, G. J.: Leaf maximum photosynthetic rate and venation are linked by hydraulics, Plant Physiol., 144,

- 555 1890–1898, https://doi.org/https://doi.org/10.1104/pp.107.101352, 2007.
  - Brovkin, V., Claussen, M., Driesschaert, E., Fichefet, T., Kicklighter, D., Loutre, M.-F., Matthews, H. D., Ramankutty, N., Schaeffer, M., and Sokolov, A.: Biogeophysical effects of historical land cover changes simulated by six Earth system models of intermediate complexity, Clim. Dynam., 26, 587–600, https://doi.org/10.1007/s00382-005-0092-6, 2006.
- Brovkin, V., Raddatz, T., Reick, C. H., Claussen, M., and Gayler, V.: Global biogeophysical interactions between forest and climate, Geophys.
  Res. Lett., 36, https://doi.org/10.1029/2009GL037543, 2009.
  - Buckley, T. N.: Modeling stomatal conductance, Plant physiology, 174, 572–582, https://doi.org/https://doi.org/10.1104/pp.16.01772, 2017.
     Buckley, T. N. and Mott, K. A.: Modelling stomatal conductance in response to environmental factors, Plant, cell & environment, 36, 1691–1699, https://doi.org/https://doi.org/10.1111/pce.12140, 2013.



580



- Charney, J., Stone, P. H., and Quirk, W. J.: Drought in the Sahara: a biogeophysical feedback mechanism, Science, 187, 434–435, https://doi.org/https://doi.org/10.1126/science.187.4175.434, 1975.
  - Christenhusz, M. J. and Byng, J. W.: The number of known plants species in the world and its annual increase, Phytotaxa, 261, 201–217, https://doi.org/10.11646/phytotaxa.261.3.1, 2016.
  - Condamine, F. L., Silvestro, D., Koppelhus, E. B., and Antonelli, A.: The rise of angiosperms pushed conifers to decline during global cooling, P. Natl. Acad. Sci. USA, 117, 28 867–28 875, https://doi.org/https://doi.org/10.1073/pnas.2005571117, 2020.
- 570 Davin, E. L. and de Noblet-Ducoudré, N.: Climatic impact of global-scale deforestation: Radiative versus nonradiative processes, J. Climate, 23, 97–112, https://doi.org/10.1175/2009JCLI3102.1, 2010.
  - De Boer, H. J., Eppinga, M. B., Wassen, M. J., and Dekker, S. C.: A critical transition in leaf evolution facilitated the Cretaceous angiosperm revolution, Nat. Commun., 3, 1–11, https://doi.org/https://doi.org.insu.bib.cnrs.fr/10.1038, 2012.
- Dow, G. J. and Bergmann, D. C.: Patterning and processes: how stomatal development defines physiological potential, Curr. Opin. Plant
   Biol., 21, 67–74, https://doi.org/10.1016/j.pbi.2014.06.007, 2014.
  - Dow, G. J., Bergmann, D. C., and Berry, J. A.: An integrated model of stomatal development and leaf physiology, New Phytol., 201, 1218–1226, https://doi.org/https://doi.org/10.1111/nph.12608, 2014.
  - Ducoudré, N. I., Laval, K., and Perrier, A.: SECHIBA, a new set of parameterizations of the hydrologic exchanges at the land-atmosphere interface within the LMD atmospheric general circulation model, J. Climate, 6, 248–273, https://doi.org/https://doi.org/10.1175/1520-0442(1993)006<0248:SANSOP>2.0.CO;2, 1993.
- Dury, M., Mertens, L., Fayolle, A., Verbeeck, H., Hambuckers, A., and François, L.: Refining species traits in a dynamic vegetation model to project the impacts of climate change on tropical trees in Central Africa, Forests, 9, 722, https://doi.org/https://doi.org/10.3390/f9110722, 2018.
- Farquhar, G. D., von Caemmerer, S. v., and Berry, J. A.: A biochemical model of photosynthetic CO 2 assimilation in leaves of C 3 species,
  Planta, 149, 78–90, https://doi.org/https://doi.org/10.1007/BF00386231, 1980.
  - Feild, T. S., Brodribb, T. J., Iglesias, A., Chatelet, D. S., Baresch, A., Upchurch, G. R., Gomez, B., Mohr, B. A., Coiffard, C., Kvacek, J., et al.: Fossil evidence for Cretaceous escalation in angiosperm leaf vein evolution, P. Natl. Acad. Sci. USA, 108, 8363–8366, https://doi.org/https://doi.org/10.1073/pnas.1014456108, 2011a.
- Feild, T. S., Upchurch, G. R., Chatelet, D. S., Brodribb, T. J., Grubbs, K. C., Samain, M.-S., and Wanke, S.: Fossil evidence for low gas exchange capacities for Early Cretaceous angiosperm leavesEarly Angiosperm Leaf Gas Exchange, Paleobiology, 37, 195–213, https://doi.org/https://doi.org/10.1666/10015.1, 2011b.
  - Fisher, R. A. and Koven, C. D.: Perspectives on the future of land surface models and the challenges of representing complex terrestrial systems, J. Adv. Model. Earth Sy., 12, e2018MS001453, https://doi.org/https://doi.org/10.1029/2018MS001453, 2020.
- Fletcher, B. J., Brentnall, S. J., Anderson, C. W., Berner, R. A., and Beerling, D. J.: Atmospheric carbon dioxide linked with Mesozoic and
  early Cenozoic climate change, Nat. Geosci., 1, 43, https://doi.org/https://doi.org.insu.bib.cnrs.fr/10.1038/ngeo.2007.29, 2008.
- Fraedrich, K., Kleidon, A., and Lunkeit, F.: A green planet versus a desert world: Estimating the effect of vegetation extremes on the atmosphere, J. Climate, 12, 3156–3163, https://doi.org/https://doi.org/10.1175/1520-0442(1999)012<3156:AGPVAD>2.0.CO;2, 1999.
  - Franks, P. and Beerling, D.: CO2-forced evolution of plant gas exchange capacity and water-use efficiency over the Phanerozoic, Geobiology, 7, 227–236, https://doi.org/https://doi.org/10.1111/j.1472-4669.2009.00193.x, 2009a.
- 600 Franks, P. J. and Beerling, D. J.: Maximum leaf conductance driven by CO2 effects on stomatal size and density over geologic time, P. Natl. Acad. Sci. USA, 106, 10343–10347, https://doi.org/10.1073/pnas.0904209106, 2009b.



605

615



- Franks, P. J. and Farquhar, G. D.: The effect of exogenous abscisic acid on stomatal development, stomatal mechanics, and leaf gas exchange in Tradescantia virginiana, Plant Physiol., 125, 935–942, https://doi.org/https://doi.org/10.1104/pp.125.2.935, 2001.
- Gibbard, S., Caldeira, K., Bala, G., Phillips, T. J., and Wickett, M.: Climate effects of global land cover change, Geophys. Res. Lett., 32, https://doi.org/https://doi.org/10.1029/2005GL024550, 2005.
- Hourdin, F., Foujols, M.-A., Codron, F., Guemas, V., Dufresne, J.-L., Bony, S., Denvil, S., Guez, L., Lott, F., Ghattas, J., et al.: Impact of the LMDZ atmospheric grid configuration on the climate and sensitivity of the IPSL-CM5A coupled model, Clim. Dynam., 40, 2167–2192, https://doi.org/https://doi.org/10.1007/s00382-012-1411-3, 2013.
- Jarvis, A., Mansfield, T., and Davies, W. J.: Stomatal behaviour, photosynthesis and transpiration under rising CO2, Plant Cell Environ., 22, 639–648, https://doi.org/https://doi.org/10.1046/j.1365-3040.1999.00407.x, 1999.
  - Kattge, J. and Knorr, W.: Temperature acclimation in a biochemical model of photosynthesis: a reanalysis of data from 36 species, Plant Cell Environ., 30, 1176–1190, https://doi.org/10.1111/j.1365-3040.2007.01690.x, 2007.

Kattge, J., Bönisch, G., Díaz, S., Lavorel, S., Prentice, I. C., Leadley, P., Tautenhahn, S., Werner, G. D., Aakala, T., Abedi, M., et al.: TRY plant trait database–enhanced coverage and open access, Glob. Change Biol, 26, 119–188, https://doi.org/https://doi.org/10.1111/gcb.14904, 2020.

- Kleidon, A., Fraedrich, K., and Heimann, M.: A green planet versus a desert world: Estimating the maximum effect of vegetation on the land surface climate, Climatic Change, 44, 471–493, https://doi.org/https://doi.org/10.1023/A:1005559518889, 2000.
- Krinner, G., Viovy, N., de Noblet-Ducoudré, N., Ogée, J., Polcher, J., Friedlingstein, P., Ciais, P., Sitch, S., and Prentice, I. C.: A dynamic global vegetation model for studies of the coupled atmosphere-biosphere system, Global Biogeochem. Cy., 19, https://doi.org/https://doi.org/10.1029/2003GB002199, 2005.
  - Ladant, J.-B. and Donnadieu, Y.: Palaeogeographic regulation of glacial events during the Cretaceous supergreenhouse, Nat. Commun., 7, 1–9, https://doi.org/https://doi.org/10.1038/ncomms12771, 2016.

- 625 Leuning, R., Kelliher, F. M., De Pury, D., and Schulze, E.-D.: Leaf nitrogen, photosynthesis, conductance and transpiration: scaling from leaves to canopies, Plant Cell Environ., 18, 1183–1200, https://doi.org/https://doi.org/10.1111/j.1365-3040.1995.tb00628.x, 1995.
  - Lin, Y.-S., Medlyn, B. E., Duursma, R. A., Prentice, I. C., Wang, H., Baig, S., Eamus, D., De Dios, V. R., Mitchell, P., Ellsworth, D. S., et al.: Optimal stomatal behaviour around the world, Nat. Clim. Change, 5, 459–464, https://doi.org/https://doi org.insu.bib.cnrs.fr/10.1038/nclimate2550, 2015.
- 630 Maire, V., Martre, P., Kattge, J., Gastal, F., Esser, G., Fontaine, S., and Soussana, J.-F.: The coordination of leaf photosynthesis links C and N fluxes in C 3 plant species, PloS one, 7, e38 345, https://doi.org/https://doi.org/10.1371/journal.pone.0038345, 2012.
  - McElwain, J. C., Yiotis, C., and Lawson, T.: Using modern plant trait relationships between observed and theoretical maximum stomatal conductance and vein density to examine patterns of plant macroevolution, New Phytol., 209, 94–103, https://doi.org/https://doi.org/10.1111/nph.13579, 2016.
- 635 Medlyn, B. E., Duursma, R. A., Eamus, D., Ellsworth, D. S., Prentice, I. C., Barton, C. V., Crous, K. Y., De Angelis, P., Freeman, M., and Wingate, L.: Reconciling the optimal and empirical approaches to modelling stomatal conductance, Global Change Biology, 17, 2134–2144, https://doi.org/10.1111/j.1365-2486.2010.02375.x, 2011.
  - Peng, S.-S., YUE, C., and CHANG, J.-F.: Developments and applications of terrestrial biosphere model, Chinese Journal of Plant Ecology, 44, 436, https://doi.org/https://doi.org/10.17521/cjpe.2019.0315, 2020.

Lee, J.-E. and Boyce, K.: Impact of the hydraulic capacity of plants on water and carbon fluxes in tropical South America, J. Geophys. Res.-Atmos., 115, https://doi.org/https://doi.org/10.1029/2010JD014568, 2010.





- Port, U., Claussen, M., and Brovkin, V.: Radiative forcing and feedback by forests in warm climates–a sensitivity study, Earth Syst. Dynam.,
   7, 535–547, https://doi.org/10.5194/esd-7-535-2016, 2016.
  - Richey, J. D., Montañez, I. P., White, J. D., DiMichele, W. A., Matthaeus, W. J., Poulsen, C. J., Macarewich, S. I., and Looy, C. V.: Modeled physiological mechanisms for observed changes in the late Paleozoic plant fossil record, Palaeogeogr. Palaeocl., 562, 110056, https://doi.org/10.1016/j.palaeo.2020.110056, 2021.
- 645 Rosnay, P. d. and Polcher, J.: Modelling root water uptake in a complex land surface scheme coupled to a GCM, Hydrol. Earth Syst. Sc., 2, 239–255, https://doi.org/https://doi.org/10.5194/hess-2-239-1998, 1998.
  - Scheiter, S., Langan, L., and Higgins, S. I.: Next-generation dynamic global vegetation models: learning from community ecology, New Phytol., 198, 957–969, https://doi.org/10.1111/nph.12210, 2013.
- Sepulchre, P., Caubel, A., Ladant, J.-B., Bopp, L., Boucher, O., Braconnot, P., Brockmann, P., Cozic, A., Donnadieu, Y., Dufresne, J.-L.,
- et al.: IPSL-CM5A2-an Earth system model designed for multi-millennial climate simulations, Geosci. Model Dev., 13, 3011-3053, https://doi.org/https://doi.org/10.5194/gmd-13-3011-2020, 2020.
  - Sewall, J. v., Van De Wal, R., Zwan, K. v., Oosterhout, C. v., Dijkstra, H., and Scotese, C.: Climate model boundary conditions for four Cretaceous time slices, Clim. Past, 3, 647–657, https://doi.org/https://doi.org/10.5194/cp-3-647-2007, 2007.
  - Sitch, S., Smith, B., Prentice, I. C., Arneth, A., Bondeau, A., Cramer, W., Kaplan, J. O., Levis, S., Lucht, W., Sykes, M. T., et al.: Evaluation
- of ecosystem dynamics, plant geography and terrestrial carbon cycling in the LPJ dynamic global vegetation model, Glob. Change Biol.,
   9, 161–185, https://doi.org/10.1046/j.1365-2486.2003.00569.x, 2003.
  - Stocker, B. D., Wang, H., Smith, N. G., Harrison, S. P., Keenan, T. F., Sandoval, D., Davis, T., and Prentice, I. C.: P-model v1. 0: an optimality-based light use efficiency model for simulating ecosystem gross primary production, Geosci. Model Dev., 13, 1545–1581, https://doi.org/https://doi.org/10.5194/gmd-13-1545-2020, 2020.
- 660 Sun, X. and Barros, A. P.: Impact of Amazonian evapotranspiration on moisture transport and convection along the eastern flanks of the tropical Andes, Q. J. Roy. Meteor. Soc., 141, 3325–3343, https://doi.org/https://doi.org/10.1002/qj.2615, 2015.
  - Vuichard, N., Messina, P., Luyssaert, S., Guenet, B., Zaehle, S., Ghattas, J., Bastrikov, V., and Peylin, P.: Accounting for carbon and nitrogen interactions in the global terrestrial ecosystem model ORCHIDEE (trunk version, rev 4999): multi-scale evaluation of gross primary production, Geosci. Model Dev., 12, 4751–4779, https://doi.org/https://doi.org/10.5194/gmd-12-4751-2019, 2019.
- 665 Wang, Y., Huang, C., Sun, B., Quan, C., Wu, J., and Lin, Z.: Paleo-CO2 variation trends and the Cretaceous greenhouse climate, Earth-Sci. Rev., 129, 136–147, https://doi.org/10.1016/j.earscirev.2013.11.001, 2014.
  - White, J. D., Montañez, I. P., Wilson, J. P., Poulsen, C. J., McElwain, J. C., DiMichele, W. A., Hren, M. T., Macarewich, S., Richey, J. D., and Matthaeus, W. J.: A process-based ecosystem model (Paleo-BGC) to simulate the dynamic response of Late Carboniferous plants to elevated O2 and aridification, Am. J. Sci., 320, 547–598, https://doi.org/https://doi.org/10.2475/09.2020.01, 2020.
- 670 Yin, X. and Struik, P.: C3 and C4 photosynthesis models: An overview from the perspective of crop modelling, NJAS-Wagen. J. Life Sc., 57, 27–38, https://doi.org/https://doi.org/10.1016/j.njas.2009.07.001, 2009.