Global nutrient cycling by commercially-targeted marine fish

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Abstract. Throughout the course of their lives fish ingest food containing essential elements, including nitrogen (N), phosphorus (P) and iron (Fe). Some of these elements are retained in the fish body to build new biomass, which acts as a stored reservoir of nutrients, while the rest is excreted or egested, providing a recycling flux to water. Fishing activity has modified the fish biomass distribution worldwide and consequently may have altered fish-mediated nutrient cycling, but this possibility remains largely unassessed, mainly due to the difficulty of estimating global fish biomass and metabolic rates. Here we quantify the role of commercially-targeted marine fish between 10g and 100kg (CTF_{10g}^{100kg}) in the cycling of N, P and Fe in the global ocean, and its change due to fishing activity, by using a global size-spectrum model of marine fish populations calibrated to observations of fish catches. Our results show that the amount of nutrients potentially stored in the global pristine CTF_{10g}^{100kg} biomass ~~was~~ is generally small compared to the ambient surface nutrient concentrations but might be significant in the nutrient-poor regions of the world: the North Atlantic for P, the oligotrophic gyres for N and the High Nutrient Low Chlorophyll (HNLC) regions for Fe. Similarly, the rate of nutrient ~~removed~~ removal from the ocean through fishing is globally small compared to the inputs, but can be important locally especially for Fe in the equatorial Pacific and along the western margin of South America and Africa. ~~This model allowed us to compute the spatial distribution of the cycling of elements by the~~ We also estimate that the cycling rate of elements through CTF_{10g}^{100kg} biomass ~~at pristine and global peak catch state, which is relatively small compared to the estimated primary production demand for nutrients and estimated export production of nutrients. Pristine cycling (excretion + egestion) accounted for less than 2.7~~ was on the order of 3% of the primary productivity demand for N, P and Fe globally. ~~Relative to the~~, prior to industrial fishing. The corresponding export of nutrients ~~, modeled global pristine CTF_{10g}^{100kg} egestion represents on average~~ by egestion of fecal matter by CTF_{10g}^{100kg} was 2.3% (N), 3.0% ~~and 1.1-22% for N, P and Fe (low-high estimates), respectively, with a higher fraction~~ (P) and 1-22% (Fe) of the total particulate export flux, and was generally more significant in the low-export oligotrophic tropical gyres. Our study ~~highlights the~~ supports a significant, direct role of the CTF_{10g}^{100kg} fraction of the ~~icthyosphere (i. e. does not include non-commercial~~ ichthyosphere in global nutrient cycling, most notably for Fe, which has been substantially modified by industrial fishing. Although we were not able to estimate the roles of smaller species such as mesopelagic fish ~~) on nutrient storage and cycling, and the potential role of fishing activities on this cycling , which~~ because of the sparsity of observational data, fishing is also likely to have altered their biomass significantly through trophic cascades, with impacts on biogeochemical cycling that could be of ~~importance in regions~~

~~of low nutrient concentration, high fish biomass and/or high productivity demand, and especially at the more local scale for Fe~~comparable magnitude to the changes we assess here.

1 Introduction

Nutrient elements are used by organisms to construct the molecules they need to grow and metabolise, but are often scarce, limiting growth rates (Moore et al., 2013; Sterner and Elser, 2002). Plankton and bacteria dominate the cycling of nutrients in the ocean (Sarmiento and Gruber, 2006) but an increasing number of studies recognize the contribution of animals to biogeochemical cycles ~~(e.g., Saba et al., 2021)~~(e.g., Wilson et al., 2009; ?; Saba et al., 2021). Locally, it has been shown that marine animals can significantly impact the supply and storage of nutrients with consequences for primary production (Cavan et al., 2019; Roman et al., 2014), and interact with the cycling of elements through direct and indirect pathways (Vanni, 2002; Atkinson et al., 2017; Allgeier et al., 2017). For instance, Leroux and Schmitz (2015) described a theoretical framework in which animals control the flux of nutrients up the trophic chain through predation and release of waste products ~~and also affect~~ while also affecting the cycling of nutrients through non-consumptive effects (e.g. prey selection and stress induced in preys). In addition, since animals can swim and move in the water column, they are also able to transport nutrients from one place to another, over distances that increase with animal size (Hall et al., 2007; Vanni, 2002; Roman et al., 2014). But thus far there has been little effort to estimate how the global fish population, which we term "~~ichtyosphere~~ichthyosphere", influences large-scale nutrient cycling in the ocean.

During their lifecycle, fish assimilate, store and recycle essential elements that they need to build their body tissues. This storage of nutrients within fish biomass is important for human nutrition as wild-caught fish globally provide essential proteins and other micronutrients (Hicks et al., 2019). Apart from a ~~direct~~ dietary interest for humans, the ~~accumulation of nutrients in fish tissues could be significant for primary productivity compared to nutrients otherwise available dissolved in the water since nutrients stored~~ reservoir of nutrients comprised by fish biomass can play a role in ecosystem function as a nutrient stockpile, the importance of which depends on how much nutrients are available otherwise (Allgeier et al., 2016). For example, the nutrients embedded in fish biomass are not directly available for primary producers~~. As an example of this~~, which can appear as a type of competition for resources ~~, Hjerne and Hansson (2002) showed that fish may compete with primary producers for P in the Baltic Sea, when their biomass increases. In contrast to the accumulation of nutrients in biomass~~(Hjerne and Hansson, 2002). But at the same time, the cycling of elements by fish ~~may act~~ acts as a source of nutrients to primary producers. ~~On one hand,~~, as fish recycle elements through the excretion of dissolved bioavailable components~~that may support part of the primary production. Cycling~~.

The cycling of N and P by fish has often been studied in freshwater systems but little is known for the global ocean (Schindler and Eby, 1997; Vanni, 2002; Vanni et al., 2006; Griffiths, 2006). For instance, McIntyre et al. (2008) showed that fish are able to create hotspots of recycled nutrients in streams that could meet more than 75% of the ~~algae and microbes~~ algal and microbial requirement for N~~. On the other hand~~, and Allgeier et al. (2014) showed that fish are a fundamental nutrient source to coral reefs. Additionally, fish egest particulate products that can be mineralized and enhance productivity or that can sink to depth

and export elements (Davison et al., 2013; Saba and Steinberg, 2012), so that they are no longer available for primary producers.

As such, fish biomass can act as a bank account for nutrients, depositing nutrients when fish are feeding rapidly and withdrawing nutrients to the water column when metabolism exceeds predation. Finally, the egestion of particulate products by fish has been shown to modify the stoichiometry of biogenic particles, including dramatic changes of Fe:C, implying that egested material may also modify the relative availability of nutrients through the water column (Le Mézo and Galbraith, 2020).

The amount of nutrient stored and cycled by fish can vary with different environmental and physiological factors, in space and time (e.g., Halvorson and Small, 2016; Prabhu et al., 2016; Francis and Côté, 2018; Czamanski et al., 2011; Allgeier et al., 2014). In addition to natural variations, anthropogenic activities, mostly fishing, modify the storage and cycling of nutrients by the ichthyosphere. For instance, Layman et al. (2011) and Allgeier et al. (2016) analyzed the cycling of N and P by fish in fished and un-fished coastal sites of the Bahamas and the Caribbean, respectively. Layman et al. (2011) showed lower recycling rates of nutrients by fish in fished sites, due to biomass reduction and habitat fragmentation. Beyond fish biomass reduction, Allgeier et al. (2016) stressed the role of community size structure that, influenced by fishing, also led to reduction in nutrient storage and cycling.

Although numerous works have identified significant local effects, little is known about the contribution of the ichthyosphere to nutrient budgets at the global scale. Maranger et al. (2008) used global fish catch data to estimate the total removal of N by commercial fisheries. They integrated a spatial component in their analysis by computing N removal in 58 Large Marine Ecosystems (LMEs) and compared it to fertilizer runoff in each off these LMEs. Moreno and Haffa (2014) took a similar approach to estimate the amount of Fe removed each year by fishing from 1950 to 2010. They also used biomass estimates from literature to quantify the amount of Fe in the global fish biomass, and the amount of Fe cycled by this biomass each year. However, the total inventories and cycling rates have remained unquantified due to the lack of reliable global fish biomass and metabolism estimates.

Recently Bianchi et al. (2021) used a dynamical global model of marine commercially-targeted fish (CTF), constrained by observed fish catches, to estimate the total CTF biomass and cycling rates, and their distribution in the world's oceans. Our study builds on Bianchi et al. (2021) by using an updated version of the model to estimate the role of CTF in the global cycling of the three most important growth-limiting nutrients: N, P and Fe (Fig. 1). Section 2 lays out the method we use to investigate the nutrient dynamics for the total CTF biomass for body sizes between 10 g and 100 kg, hereafter CTF_{10g}^{100kg}, using model simulations for both a reconstructed state prior to industrial fishing ("pristine") and a simulated global peak catch. We then present and discuss the results as a series of relatively self-contained sections by topic. Section 3 details the total nutrient content in CTF_{10g}^{100kg}, Section 4 focuses on nutrient cycling rates by CTF_{10g}^{100kg}, Section 5 discusses an extension of the CTF_{10g}^{100kg} estimates to all fish from 1g to 1000kg, and Section 6 presents the impact of fishing on CTF_{10g}^{100kg} (Fig. 1). Section 7 concludes the paper.
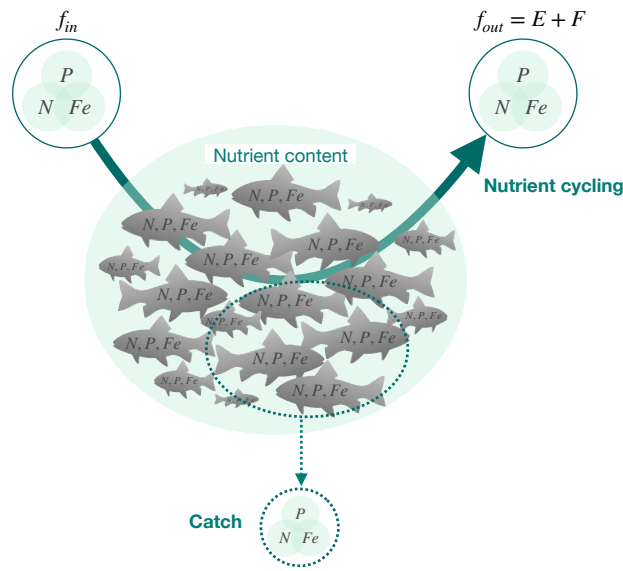
Figure 1. ~~Icthyosphere interactions with nutrients~~The ichthyosphere role in global nutrient cycling. ~~Schematic of~~ The diagram schematically indicates the cycling of nutrients (N, P and Fe) ~~by~~ through the global fish biomass (solid arrow), the ~~nutrient content of this~~ nutrients contained within the biomass, and the removal of nutrients from the ocean through fishing (dashed arrow). f_{in} and f_{out} are the fluxes in and out of the global fish biomass, respectively. E ~~is for~~ indicates excretion (release of dissolved compounds) and F ~~is for~~ indicates egestion (release of solid feces).

2   Methods

2.1   Model description and simulations

95   We used an ensemble of simulations ~~following the same method as in Bianchi et al. (2021). We used global fish biomass estimates, under pristine and fished ocean conditions, simulated by the~~ from the BiOeconomic mArine Trophic Size-spectrum (BOATS) model (Carozza et al., 2017). ~~BOATS represents~~, also used by Bianchi et al. (2021).

2.1.1   The model

BOATS provides a global, size-based numerical simulation of commercially-targeted marine organisms (~~here simply called~~
100   including molluscs and crustaceans, here collectively termed "fish") ~~, larger than 10g and under 100kg, hereby called CTF_{10g}^{100kg}~~, by coupling an ecological ~~and~~ module with a fishery economics ~~model~~ module (Carozza et al., 2016, 2017). The ecological model is based on processes derived from macro-ecological theory ~~(Carozza 2016). It is~~ (Carozza et al., 2016), parameterized through a Bayesian Monte Carlo approach ~~, that compares~~ that optimizes model performance by minimizing the discrepancy between observed and simulated catch ~~in LMEs (Carozza et al. 2017).~~ and biomass for globally-distributed LMEs
105   (Carozza et al., 2017). Modeled fish are divided into 3 "super-species" groups defined by the asymptotic mass of fish: small (0.3
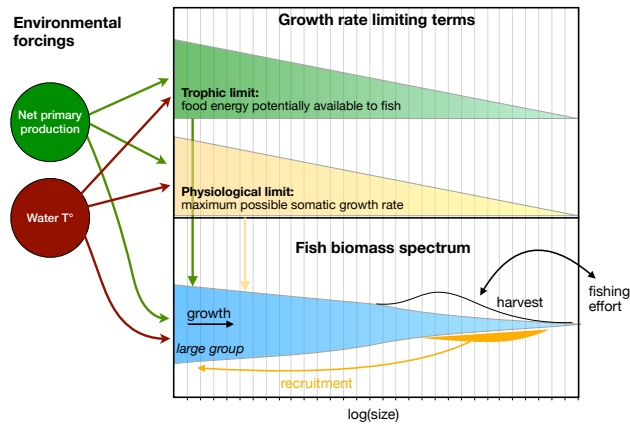
Figure 2. Schematic overview of the BOATS model. The red, green, and black arrows indicate dependencies of model components on external forcings. The top panel indicates the energetic limits of growth as a function of fish size, while the bottom panel illustrates the three size spectra of fish groups (for simplicity only the large group is represented), their internal dynamics, and link to economics via harvest and the interactive effort.

kg), medium (8.5 kg) and large (100 kg). Individual sizes are binned logarithmically into 30 mass classes ranging from 10 g to 100 kg, with all three size groups starting at 10 g (Carozza et al., 2016). The three groups are not intended to represent the entire marine ecosystem, but rather the sum of all species that have been commercially harvested (and are therefore accounted for in harvest records and stock assessments, which are used to constrain the model). The underlying philosophy of the model is that, although these very diverse species differ widely in their biological strategies, all are competing for food energy ultimately provided by the fixation of organic carbon through photosynthesis (which has been shown to limit fish harvests (**??**)), while inhabiting the same environment, which therefore makes them subject to the same basic ecological constraints. The constraints we apply in the model are the impacts of water temperature on growth, mortality, and phytoplankton size, and net primary production. Although this biologically 'coarse-grained' approach precludes resolution of species-level dynamics, it is solidly-founded in bioenergetic principles, and is well-suited to providing a global view of the entire ecosystem on long timescales, given that it is likely to be relatively robust under any changes in the distribution, abundance or evolution of commercial stocks under historical fishing pressure (Guiet et al., 2020). Figure 2 provides a schematic overview of the model structure.

The BOATS model was deliberately developed to represent marine organisms over the size range most heavily targeted by fisheries, since this is the range for which fisheries data can offer useful constraints on the ecosystem function (Carozza et al., 2016, 2017). The starting point of 10 g coincides roughly with the weight of mature anchovy (approximately 11 cm in length according to Pauly and Tsukayama (1984)), while above 100 kg the growing significance of mammals in the ocean makes a strictly ectothermic model less capable of capturing the full marine animal community (Hatton et al., 2021).

Fishing effort and catch are computed assuming open-access dynamics and based on the Gordon-Schaefer fishery economics
125 model (Gordon, 1954; Schaefer, 1954) with increasing catchability over time ~~(Galbraith et al., 2017).~~ due to technological progress (Galbraith et al., 2017).

The model represents fish on a two-dimensional grid, i.e. longitude and latitude, which is divided in regular 1° x 1° grid cells. Thus, the model does not resolve the vertical dimension, but sums all ecosystem productivity and biomass within the water column at each horizontal point. Given that the model does not resolve interactions in space between individuals, this reduction
130 in dimensionality does not - on its own - introduce any bias. The model is forced by monthly climatologies of observed net primary production and surface ocean temperature (Dunne et al., 2007).

Galbraith et al. (2019) hypothesizes that fish growth ~~may be affected by Fe~~ is reduced under iron scarcity in the wild, and demonstrates that the implementation of a simple form of ~~Fe~~ iron limitation of fish in BOATS ~~better represents~~ improves the simulated fish catch in ~~HNLC regions~~ oceanic regions known to have low iron concentrations. We thus used a different
135 version of the model ~~with the~~ from Bianchi et al. (2021) that includes an Fe limitation of fish growth~~describe~~, using surface nitrate concentrations as a proxy for iron limitation as described in Galbraith et al. (2019). ~~The model runs on a 1-degree two-dimensional grid, forced by monthly climatologies of observed net primary production and surface ocean temperature (Carozza et al., 2016). Fish are divided into 3 size groups defined by the asymptotic mass of fish; small (0.3 kg), medium (8.5 kg) and large (100 kg) and, each size-group is divided into 30 mass classes ranging from 10 g to 100 kg (Carozza et al., 2016).~~
140 ~~From~~

2.1.2 Simulations

We use the parameter sets selected in Bianchi et al. (2021) to run our updated Fe-limited model version. Briefly, from the Monte Carlo ensemble of ~~simulations~~10,~~we select 31 (similar to Bianchi et al. (2021)~~000 simulations, Bianchi et al. (2021) selected the parameter combinations (31 in total) that best match the historical ~~harvest in LMEs as observed and~~ catch maxima
145 across LMEs as reconstructed by the Sea Around Us Project ~~(Pauly and Zeller, 2016).~~ (SAUP) (Pauly and Zeller, 2016), while simultaneously falling within the acceptable bounds for the catch:biomass ratio as constrained by stock estimates (Ricard et al., 2012). We then perform simulations with our Fe-limited verision of the model using these 31 model parameter combinations. Each simulation includes 200 years without catch to estimate pristine biomass at equilibrium, followed by 220 years with fishing driven by the only increase of the catchability of biomass at 7%.y^{-1} to reproduce the historical progression of the global
150 fishery ~~(Bianchi et al., 2021).~~ (Galbraith et al., 2017). The simulations slighlty underestimate the observed LME catch peak of $\sim 110 Mt.y^{-1}$ and the variation between LME peaks corresponds to observation with a squared Pearson product-moment correlation coefficient $r^2 = 0.42, p - value < 10^{-9}$, when averaged across all ensemble members (Suppp. Figs. A1 and A2). More details on the observational constraints and on the Monte Carlo approach used here can be found in Bianchi et al. (2021). From the 31 simulations ~~, we here~~ of the ensemble, we analyze the global biomass and cycling rates at pristine state and at the
155 time of the global peak catch.

2.1.3 ~~Nutrient content of fish~~

The mean nutrient content of the global

2.2 Nutrient content of fish

To estimate the quantity of a nutrient X stored in fish biomass, we convert our modeled biomass estimates from wet weight to carbon (C) weight and multiply by an average mass ratio of nutrient to carbon, (X:C). Prior work has suggested that body nutrient concentration of fish may be affected by several factors such as body size, ontogeny, species, sex, diet, temperature or water nutrient concentration (e.g., Halvorson and Small, 2016; Prabhu et al., 2016; Allgeier et al., 2017), with species appearing to be the most important factor (Allgeier et al., 2020). Among these factors, our model could potentially account for change during ontogeny as organisms grow in size, but analysis of the available data (see supplement) shows little to no systematic variation of specific nutrient content with size. We thus assumed constant nutrient proportions throughout food webs.

Although the model implicitly includes molluscs and crustaceans, they represent only a small proportion of the commercial catch between 10g and 100kg (from SAU data invertebrates comprised about <14% of total catch). Additionally, the measured nutrient content of molluscs and crustaceans falls within the uncertainty range around the mean value for fish (Supp. Tables A2-A3). As a result, we did not attempt to account for invertebrates separately, but applied the fish nutrient ratios to all CTF_{10g}^{100kg} biomass is calculated using mean values of 0.6 ± 0.2% P and 2.8 ± 0.4% N in fish wet weight (Table1). We estimate the average body Fe content of fish to be about 21 μmolFe/molC based on whole body Fe measurements (Galbraith et al., 2019; Prabhu et al but as the biomass. The Fe content of fish is poorly constrained (few whole body measurements), so we rather use the 95% confidence-interval from available data, which ranges between 10 and 200 μmolFe/molC (Galbraith et al., 2019). This way we can estimate the effects of the uncertainty on fish body Fe:C.

Body nutrient concentration of fish may be affected by several factors such as body size, ontogeny, speciation, sex, diet, temperature or water nutrient concentration (e.g., Halvorson and Small, 2016; Prabhu et al., 2016; Allgeier et al., 2017). Among these our model could best account for change during ontogeny as organisms grow in size. Yet, analysis of data (see supplement) shows little to no systematic variation of specific nutrient content with size. While other factors can't be represented in our model, some of their effect are included in the uncertainty around the values in table 1. We assumed constant nutrient proportions throughout food webs and focused on the removal by fishing (Fig. 1).

2.3 Nutrient cycling by fish

Although the model non-explicit represents all organisms between 10g and 100kg, including molluscs and crustaceans, hereby called "fish", we used the nutrient content values of fish as they represent the largest proportion of the commercial catch. This may result in an slight overestimation of N and P content, and an underestimation of the Fe content of the modeled fish biomass, part of which is included in the uncertainties we computed (Table ?? and Supp. Table ??).

Table 1. Mean nutrient content in fish, zooplankton and phytoplankton, and mean absorption efficiencies (A) of N, P and Fe for fish used in this study.

	% N in ww	% P in ww	C/N (molC/molN)	C/P (molC/molP)	Fe:C (μmolFe/molC)
Fish					
Global ± std (range)	[a]2.8± 0.4	[a]0.6± 0.2	[a]4.6 (3.4-6.4)	[a]49 (29-82)	[c,e]21 (10-200)
Zooplankton					
Global± std (range or Fe-poor/Fe-rich, low/high)	[b]1.4± 0.3	[b]0.14± 0.04	[a]4.7 (2.9-7.7)	[a]140 (84-231)	[c]30.6 (8.2/85, 4.1/248)
Phytoplankton					
Global (Fe-poor/Fe-rich, low/high)	From relationships in Galbraith and Martiny (2015) (Eqs. 4)				[f]60.5 ([c]5/92, [f]2.13/258)

	C	N	P	Fe
Fish mean absorption efficiency (A)	[a]0.88	[a,b]0.86	[a,b]0.71	[d]0.24

Data from [a]Czamanski et al. (2011) (% converted back to wet weight using a 25% dry weight in wet weight (Galbraith et al., 2019), geometric mean, P absorption efficiency is computed from the linear regression between predator and prey C/P ratio), [b]Schindler and Eby (1997), [c]Galbraith et al. (2019), [d]Thodesen et al. (2001), [e]Prabhu et al. (2016), [f]Moore et al. (2013). "ww" is for wet weight, Fe-poor and Fe-rich refer to the conditions in which the organisms lived, low/high are the low and high estimates from gathered data in Galbraith et al. (2019). Standard deviations are the arithmetic standard deviations, associated with the arithmetic mean. Ratio mean values are geometric means and the ranges are the 95% confidence interval, except for fish Fe:C for which the range is estimated from Galbraith et al. (2019).

2.3.1 ~~Nutrient cycling by fish~~

First, we define the ~~excretion and egestion terms considering a single fish. Figure 3a represents~~ terms we use by considering the fate of a nutrient element when ingested by ~~a fish .~~ an individual fish (Fig. 3a). A fish i ~~assimilates part of the nutrients it ingests , AI_i, with~~ ingests a mass flux of a nutrient, I_i ~~the amount ingested, while the rest~~ (e.g. $gN.d^{-1}$), of which a fraction α (between 0 and 1) is absorbed across the gut to produce an absorption flux, A_i, while the remainder is egested in the form of ~~fecal pellets, $F_i = (1 - A)I_i$~~ feces, $F_i = (1 - \alpha)I_i$ (Fig.3). ~~The absorption efficiency , A,~~ Note that absorption efficiency here is defined as the ~~proportion of an element that goes across the gut of fish, i.e. ingestion net of~~ difference between ingestion and egestion. Published estimates of absorption efficiencies of fish are listed in Table 1. ~~Part of the elements~~ A fraction of the element absorbed across the fish gut ~~are~~ is then used for growth and reproduction, ~~$Pr_i = \alpha AI_i$, where α~~ $G_i = \gamma A_i = \gamma \alpha I_i$, where γ is the somatic assimilation efficiency. The trophic efficiency, ~~te~~ τ, is commonly defined as the ~~amount of an element that is integrated into new biomass following ingestion~~ ratio of the production of new biomass to the ingestion rate, and is equal to ~~αA~~ $\gamma \alpha$. The remainder is used for maintenance and excreted back to the water; ~~$E_i = (1 - \alpha)AI_i$~~ $E_i = (1 - \gamma)\alpha I_i$ (Fig. 3a,b). ~~In our model ensemble, te has an average value of 17% for wet biomass, which is directly proportionnal to carbon content (Bianchi et al., 2021)~~ We define the cycling rate of a nutrient through the fish as $I_i - G_i = E_i + F_i$, which can be thought of as the biologically-processed outputs of the fish.

~~Second, at the community level we~~ Based on these terms, we follow Bianchi et al. (2021) to estimate the total flux of ~~elements~~
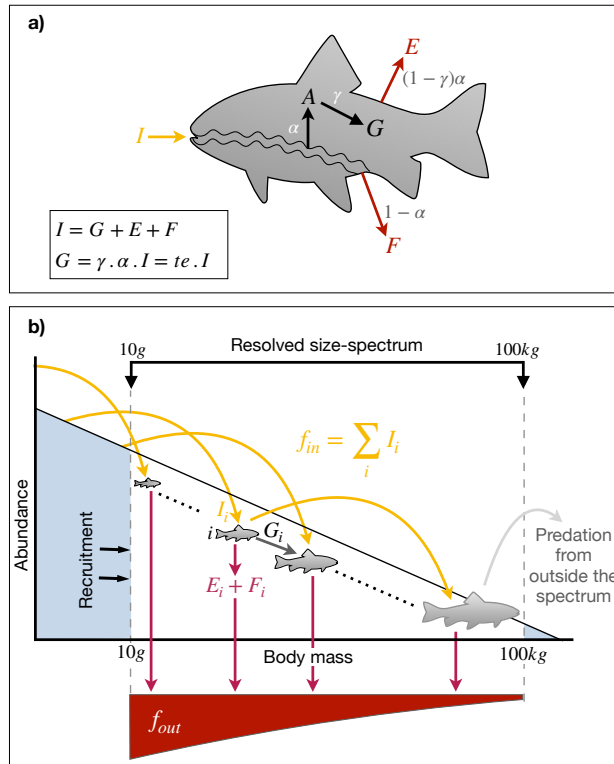
Figure 3. Schematic of the flow of elements a) within an individual and b) through the size-spectrum. I = ingestion, ~~Pr~~G = ~~production~~growth and reproduction, ~~I_A~~A = ~~part of ingestion that is assimilated~~absorption, E = excretion, F = egestion (feces), ~~A~~α = ~~assimilation~~absorption efficiency~~net of egestion~~, i.e. ~~absorption~~assimilation efficiency net of egestion, ~~α~~γ = somatic assimilation efficiency, i.e ~~part assimilated~~assimilation allocated to growth, f_{in} = flux entering the spectrum, f_{out} = flux going out of the spectrum

~~by computing the input flux through all feeding on unresolved prey, i. e. "outside" the spectrum~~ carbon through the CTF_{10g}^{100kg} based on the community average τ and growth rates $G_i(C)$ of all simulated fish within the size spectrum. We calculate the ingestion for each fish within the size bins by dividing $G_i(C)$ by $\tau(C)$ (which is 0.17 when averaged across the ensemble). We then subtract the growth $G_i(C)$, to arrive at the carbon cycling rate, and sum over all fish (Fig. 3b). ~~By doing so, we avoid accounting for internal cycling of nutrients within the spectrum. We use~~ We note that, at steady state for the community level there is no net growth since new production is balanced by predation, but by subtracting $G_i(C)$ we ensure that there is no double-counting of internal cycling within the simulated size spectrum through predation on simulated fish. Because the model mean predator-prey mass ratio ~~(PPMR), about 6.0 10³ in our ensemble , to select all the fish in the spectra whose size allows them to feed only on organisms smaller than the smallest fish of the spectrum (10g). We also assume that the flux entering the spectrum from predation on larger unresolved organisms is compensated by a related external mortality, i.e. by~~ for the ensemble is $0.6 \; 10^4$, there is very little predation within the resolved size spectrum (there are only four orders of magnitude across the smallest to the largest size bin), so that the subtraction of $G_i(C)$ could lead to a small systematic underestimate in our cycling

estimates. However, we also consider that predation by large predators not resolved by the model, including very large fish, marine mammals or birds, will cause a portion of the resolved fish growth to be cycled by other organisms outside of the CTF_{10g}^{100kg}. Any consequent underestimate is therefore likely to be significantly less than the value of $\tau(C)$ (i.e. significantly less than 17%). Thus, our carbon cycling rate equation for CTF_{10g}^{100kg} is given by the output flux equation:

$$f_{out}(C) = \sum_i (I_i - Pr(C) - G_i(C)) = \sum_i (E_i(C) + F_i(C)) = f_{in} - Pr \tag{1}$$

(C)-$\sum_i(G_i(C))$ We can then use the input rate of carbon to the fish population, $f_{in}(C)$, to calculate the cycling terms for any nutrient element based on the stoichiometry of the ingested organic matter and element-specific absorption. We do so by taking the nutrient X to carbon mass ratio within the prey, $(X:C)_{prey}$, and the absorption efficiency of nutrient X, α_X, to compute:

$$f_{in}(X) = f_{in}(C)(X:C)_{prey} \quad ; \quad E(X) = \sum_i (\alpha_X I_i(X) - G_i(X)) \quad ; \quad F(X) = f_{in}(X)(1 - \alpha_X) \tag{2}$$

where $f_{in}(X)$ is the ingestion flux of nutrient X into all fish, and E(X) and F(X) are the excreted and egested fluxes of nutrient X, respectively, for all fish.

Note that we separate the ingested fraction, I, and egested fraction, F, using a constant mean absorption coefficient for each nutrient, α_X (Table 1).

As indicated by eq.2, the cycling rate we calculate for an element X will be proportional to the average value of $(X:C)_{prey}$. To maintain tractability, we assumed that zooplankton provide a representative indication of the stoichiometry of the fish food source and we used mean zooplankton N and P nutrient content to compute the cycling of these elements through the fish biomass (Table 1). For zooplankton Fe content, spatial variability appears to be more important than for N and P, given significant differences between Fe-rich and Fe-poor regions (Table 1, Galbraith et al. (2019)). In order to be thorough, we computed Fe cycling in three different ways based on the various computation of the Fe:C distribution of zooplankton: 1) we used a mean Fe:C in zooplankton of 30.6 μmolFe/molC, 2) we used a spatial variation between Fe-rich and Fe-poor regions and used the Fe:C mean values in Fe-poor and Fe-rich regions, respectively, from Table 1, 3) we used the same spatial variation, but with the low and high Fe:C estimates of zooplankton from Table 1. For the spatial variations of Fe:C, we assumed that Fe-poor conditions are encountered in HNLC regions, which are determined by a concentration of surface nitrate $[NO_3^-]$ larger than $5 mmolN.m^{-3}$. In order to take into account the gradient between these regions, we locally weighted zooplankton

Fe content using a Michaelis-Menten function:

$$(Fe:C)_{zoo} = (Fe:C)_{zoo}^{Fe-rich\,or\,max} + ((Fe:C)_{zoo}^{Fe-poor\,or\,low} - (Fe:C)_{zoo}^{Fe-rich\,or\,high})\frac{[NO_3^-]_{surf}}{5 + [NO_3^-]_{surf}} \tag{3}$$

2.3.1 ~~Primary producers demand, nutrient concentrations and export and atmospheric deposition~~

250 ~~In our analysis, we~~

2.4 Primary producers demand, nutrient concentrations, export and atmospheric deposition

We compare the nutrient cycling by fish to the nutrient demand by primary producers, in order to provide a rough characterization of its magnitude. We use an averaged satellite-based primary productivity (PP) (Dunne et al., 2007) to compute the PP demand for N, P and Fe. We predict the C:P and C:N ratios in phytoplankton using empirical relationships with PO_4^{3-} and NO_3^-

255 surface concentrations as described in Galbraith and Martiny (2015):

$$(N:C)_{phyto} = 12.5\% + 3\%\frac{[NO_3^-]_{surf}}{0.32 + [NO_3^-]_{surf}} \qquad ; \qquad (P:C)_{phyto} = 0.6\% + 0.69\%[PO_4^{3-}]_{surf} \tag{4}$$

~~with nutrient concentrations~~ where nutrient concentrations are in μmol/L.

Similarly to zooplankton, the Fe:C of phytoplankton is computed by allowing variation in stoichiometric ratios between the mean value found in Fe-poor conditions and the mean value found in Fe-rich conditions, or between the high and low phyto-

260 plankton Fe:C estimates (Table 1), using a Michaelis-Menten equation ~~:~~

$$(Fe:C)_{phy} = (Fe:C)_{phy}^{Fe-rich\,or\,high} + ((Fe:C)_{phy}^{Fe-poor\,or\,low} - (Fe:C)_{phy}^{Fe-rich\,or\,high})\frac{[NO_3^-]_{surf}}{5 + [NO_3^-]_{surf}}$$

~~We did not use a global mean value for phytoplankton, as previous studies and data has clearly showed that phytoplankton Fe:C values varies significantly between Fe-poor and Fe-rich areas (Table 1). We~~ analogous to equation 4.

We then use the phytoplankton nutrient ratios to compute the export of nutrients from a satellite-based estimate of total C

265 export (Dunne et al., 2007).

We use the World Ocean Atlas observed PO_4^{3-} and NO_3^- water concentrations (Locarnini et al., 2010), and dissolved Fe concentrations simulated by the biogeochemical model TOPAZ2 (Dunne et al., 2013) to compare the fish biomass nutrient content to the surface ocean ambient concentrations of nutrients, and to compute the stoichiometric ratios ~~in equation 4and ??~~(e.g. in eq.4). The TOPAZ2 model represents the cycles of different elements from carbon to calcite and Fe with 30 different

270 tracers and the dynamics of three groups of phytoplankton. Surface concentrations are computed using the 2002-2019 annual mean euphotic depth from MODIS-Aqua (2018). ~~Compared with the single estimation from surface by MODIS, $[PO_4^{3-}]$ and $[NO_3^-]$ observations are discretized along the water column, with 32 levels with a 10m interval or more. Computation of coarse $[PO_4^{3-}]$ and $[NO_3^-]$ over the euphotic layer leads to rough edges of the nutrient concentration maps (Fig. 5b).~~

~~Finally, we use~~ Finally, we compare the rate of nutrients removal by fishing at the time of global peak catch to current atmo-

275 spheric deposition fields of soluble N (Brahney et al., 2015) and Fe (Mahowald et al., 2009) (Supp. Fig. B1~~, ref [1]~~)~~to compare to~~

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[1]~~http://www.geo.cornell.edu/eas/PeoplePlaces/Faculty/mahowald/~~

3 Fish biomass: a living pool of nutrients

280 Our results show that the ~~quantity of~~ nutrients contained within ~~the~~ CTF_{10g}^{100kg} biomass ~~represents~~ represent a non-negligible proportion of the ambient dissolved concentrations ~~of the respective nutrients~~ in areas where these concentrations in seawater are low and/or where ~~the~~ CTF_{10g}^{100kg} biomass is high.
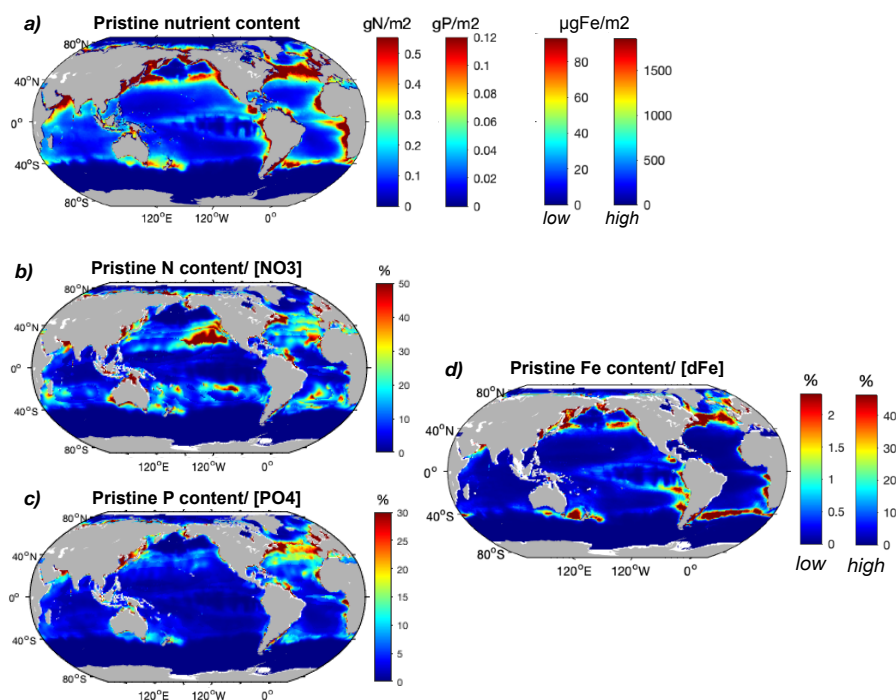
Figure 4. Modeled commercial pristine fish biomass mean nutrient content and relative to surface nutrient concentrations. a) P content (mmolP/m2), N content (mmolN/m2) and Fe content (low and high estimates, μmolFe/m2) of the global pristine CTF_{10g}^{100kg} biomass and, b) N content relative to surface NO_3^- concentrations (%), c) P content relative to surface PO_4^{3-} concentrations (%), and d) Fe content (low and high estimates) relative to modeled surface dissolved Fe (%) from the TOPAZ model (Dunne et al., 2013). All surface concentrations are integrated over the 2002-2019 annual mean euphotic depth (MODIS-Aqua, 2018).

The highest amounts of N, P and Fe in the pristine fish biomass are located in the most productive regions along the coasts, where most simulated fish biomass occurs (Fig. 5a). Globally, the estimated pristine biomass of CTF_{10g}^{100kg}, which represents 285 2.5 ± 0.8 Gtons of wet biomass, contains 69 ± 31 Tg of N, 15 ± 14 Tg of P and $0.012 - 0.23$ Tg of Fe~~(Table S??)~~, of which

about half is found in the Large Marine Ecosystems (LMEs) (Table ~~??).~~ 2). Note that, in our computations, fish biomass is the only term that varies spatially since the nutrient contents of fish, (X:C), are held globally constant (see Methods section).
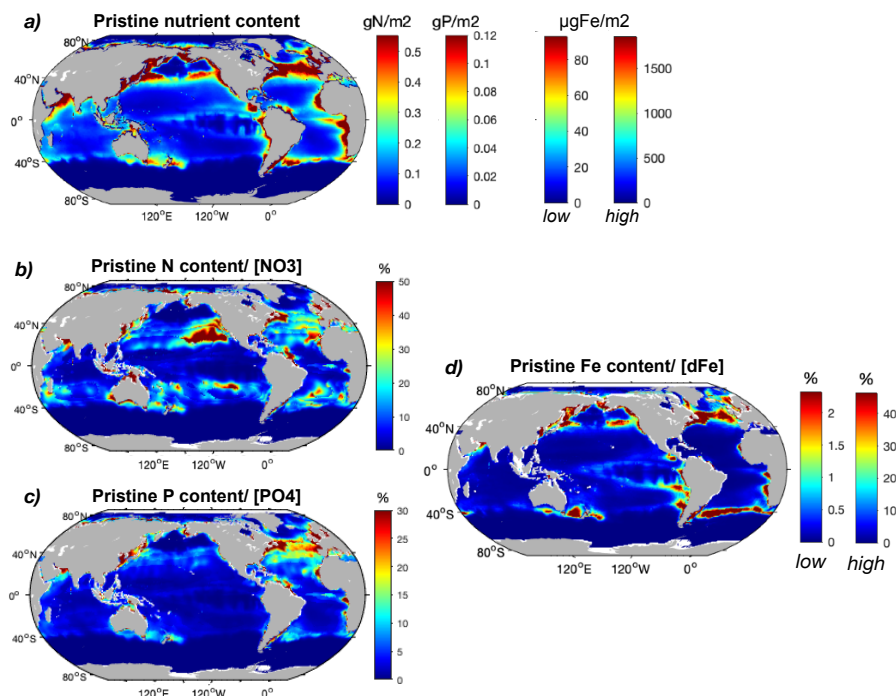
Figure 5. Modeled commercial pristine fish biomass mean nutrient content and relative to surface nutrient concentrations. a) P content (mmolP/m2), N content (mmolN/m2) and Fe content (low and high estimates, μmolFe/m2) of the global pristine CTF_{10g}^{100kg} biomass and, b) N content relative to surface NO_3^- concentrations (%), c) P content relative to surface PO_4^{3-} concentrations (%), and d) Fe content (low and high estimates) relative to modeled surface dissolved Fe (%) from the TOPAZ model(Dunne et al., 2013). All surface concentrations are integrated over the 2002-2019 annual mean euphotic depth(MODIS-Aqua, 2018). The N, P and Fe content of fish are represented on a single map given that we used a globally constant nutrient ratio for each element, so that all spatial variation is caused by fish biomass.

The N content of ~~the~~ CTF_{10g}^{100kg} biomass ~~appears more significant compared to~~ is locally on the same order of magnitude as ambient surface NO_3^- concentrations, ~~up to more than~~ exceeding 50% ~~,~~ of $[NO_3^-]$ in the oligotrophic gyres where nutrient concentrations are low, and in coastal ~~areas~~ shelves where large fish biomass accumulates ~~in the pristine ocean~~ prior to industrial fishing (Fig. 5b~~), thus explaining the high percentage on average over the LMEs (Table ??~~, Table 2). The amount of P in ~~the~~ CTF_{10g}^{100kg} biomass represents a high proportion of available PO_4^{3-} in the North Atlantic ocean, which is relatively P-poor, ~~up to more than~~ exceeding 30% in some areas (Fig. 5b). The ratio also ~~reaches values larger than~~ exceeds 20% in the western North Pacific and in a few locations such as in the Arabian sea. CTF_{10g}^{100kg} biomass stores much higher Fe compared to dissolved surface concentrations in the ~~subarctic~~ highly productive subtropical front waters of the North Pacific, ~~subarctic~~

North Atlantic, ~~along 40°S and in coastal areas~~ and Southern Oceans, with relative values ~~as high as 20~~exceeding 50% for the ~~low~~ high-end estimate (Fig. 5d). ~~The ratio for Fe is particularly low~~ Despite low surface Fe concentrations in the Southern Ocean, the proportion in CTF_{10g}^{100kg} is particularly low due to the low modeled fish biomass~~(Galbraith et al., 2019), and~~, while in the tropical Atlantic ~~where~~ the input of Fe from dust ~~is the largest~~ greatly overwhelms the iron in fish (Mahowald et al., 2009; Myriokefalitakis et al., 2016).

~~Both low ambient dissolved nutrient concentrations and high fish biomass, as well as a combination of both, result in a significant~~ In summary, the storage of nutrients by ~~the~~ CTF_{10g}^{100kg} biomass ~~compared to nutrient availability. Thus, in the areas where these conditions occurs~~can be quite significant, compared to the dissolved nutrient inventory of the euphotic zone, but only where ambient dissolved nutrient concentrations are low and/or fish biomass is high. In these areas, CTF_{10g}^{100kg} biomass ~~has the potential to act as a significant~~ could be expected to have a greater potential as a source (if stored nutrients are made available) or ~~a~~ sink (if the nutrients cannot be used by primary producers) of nutrients.

Table 2. Table of values in LMEs from the model ensemble simulations in the pristine state and at the global peak catch. This table contains integrated values of: ~~1.~~ nutrient content in CTF_{10g}^{100kg} biomass (Tg) ~~, 2.~~ and the ratio of nutrient content in CTF_{10g}^{100kg} biomass with surface nutrient concentrations (%), ~~3. the amount of nutrient cycled by the CTF_{10g}^{100kg} biomass (Tg/yr), 4. the ratio of this cycling with the global primary producers demand for these nutrients (%), 5. the amount of nutrient egested by the CTF_{10g}^{100kg} biomass (Tg/yr) and 6. its ratio with the exported nutrient quantities (%), 7. the amount of nutrients removed by fishing in LMEs (Tg/yr) and 8. the ranges of values of the global nutrients inputs to the ocean from the literature (Tg/yr).~~

Content (Tg)

Pristine 37.4±17.2 8.1±7.5 (6.3±2.1)10^{-3} 0.13±0.04 *At global peak catch* 9.8±5.4 2.2±2.2 (1.6±0.7)10^{-3} (3.3±1.4)10^{-2}

Pristine

At global peak catch

At global peak catch 56±35 4.2±2.5 0.03±0.07 0.21±0.07

Pristine

At global peak catch

Pristine 2.1±1.1 2.6±1.3 1.1±2.3 20.1±5.0 *At global peak catch* 1.6±1.0 2.1±1.2 0.8±1.7 15.7±5.0 *LMEs* 2.8±0.4 0.6±0.2 (4.7)10^{-4} (

3.1 Comparison to previous estimates

~~The differences between our estimates and previous studies~~ Our estimates for the N and Fe contents of the ichthyosphere differ from previous studies to some degree, which can be explained by ~~large~~ differences in the ~~biomass of fish, which is difficult~~

~~to accurately get at the global scale, and, especially for Fe, by the uncertainty on~~ estimates of global fish biomass and/or uncertainty regarding fish nutrient contents ~~on which we lack measurements~~(we were not able to find prior estimates for P).

The amount of N stored within the global fish biomass has been previously estimated to be about 23 Tg (Allgeier et al., 2017)[1], which is about 66% less than our global pristine estimate of 68.7 ± 30.5 TgN (Table ~~S??~~2). Their computation is based on an estimation of fish biomass of 0.9 Gtons (Jennings et al., 2008) while our ensemble pristine CTF_{10g}^{100kg} biomass is 2.5 ± 0.8 Gtons ~~, which is~~ (a smaller biomass estimate than that of Bianchi et al. (2021) due to Fe-limitation of fish), a difference of biomass of 64%~~discussed by Bianchi et al. (2021)~~. Additionally, we used a mean N content of 2.8%, slightly higher than their value of 2.6%, because we used measurements made only on wild fish ~~only~~ and did not try to account for all the catch diversity in organisms. ~~We only added the species~~To reflect this uncertainty, we indicate the species-related uncertainty around the mean value we use, $\pm0.4\%$, which covers the value used in Maranger et al. (2008), ~~to~~for our calculations in Tables ~~?? and ??~~2 and 4.

For Fe, Moreno and Haffa (2014) estimated that the global fish biomass stored between 0.07 and 0.7 Tg~~of Fe~~, which is a ~~slightly higher range than ours~~ roughly threefold higher than our range of 0.012-0.23 Tg of Fe (Table 1). To compute these values, Moreno and Haffa (2014) used an estimated fish biomass of 0.9-2 Gtons, which is lower than our modeled estimate of 2.5 Gtons for commercially-targeted fish only due to a conservative maximum estimation (Wilson et al., 2009). However, they used a range of Fe:C values, 0.073-0.324 gFe per kg of wet weight (ww) for ray-finned fish, that is about 3-12 times larger ~~to~~than our 10-200 μmolFe/molC range, equivalent to 0.006-0.12 gFe/kgww (assuming 12.5% C in ww). We are more confident in our compilation of Fe:C values, which is updated, more complete, and only based on peer-reviewed studies (Galbraith et al., 2019)~~, however~~. Nonetheless, the differences highlighted here, and the large range of estimates (low-high values) ~~stress out the high~~highlight the large uncertainty on the Fe content of whole fish and the ~~lack of~~great need for more measurements in this domain.

Note that our modeled estimates are likely to be more reliable in LMEs since fish biomass in these regions is better constrained by fish catch data. ~~An additional table with the global estimates is provided in the supplementary material (Supp. Table ??).The new biomass estimate this model provides explains the large difference with previous calculations of the N content of fish, biomass estimates that are discussed in details in Bianchi et al. (2021). We did not find previous work on the P content of the global fish biomass and its distribution in the World's oceans. As for Fe, in addition to biomass, the high uncertainty on the Fe:C content of CTF_{10g}^{100kg} strongly impact the estimations of the total Fe content.~~

3.2 Nutrient content variations in fish~~and limitations of our study~~

Many factors ~~are responsible for~~contribute to variations of fish body nutrient concentrations~~(Table 1)~~, which we ~~can't all model but include to the best of our abilities in the uncertainties around the mean values we use.~~cannot model explicitly but instead capture within our uncertainty estimates. Among the different influencing factors, fish species ~~can be of importance, with~~is important, for instance bony fish ~~having~~contain larger quantities of P ~~in their body~~compared to other fishes (El-Sabaawi et al.,

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[1]It seems that there is a typo in Allgeier et al. (2017). Their estimate is based on Jennings et al. (2008)'s wet biomass estimate of $9 \cdot 10^8$ tons of fish and they used a 2.6% of N in fish, thus the total N harvest should be 23.4 Tg and not 233.4 Tg

2016). Species can also vary in terms of the size of storage components (e.g., Czamanski et al., 2011) and the number and size of bones if vertebrates, which generally increases with adult size (Sterner and Elser, 2002). Variations in body nutrient contents

345 can also be explained by sex and life stage, e.g. decreased Fe body ~~burden~~ content occurs in female rainbow trout during sexual maturation (Shearer, 1984), ~~and variations in the storage components between organisms (e.g., Czamanski et al., 2011) such as the number and size of bones of vertebrates, which generally increases with size (Sterner and Elser, 2002). Studies also showed~~ though these would tend to average out at the population level. Studies have also shown that ontogeny may affect nutrient content~~with~~, for example juveniles ~~having~~ have less P than adults (Pilati and Vanni, 2007) ~~and the Fe content varying~~

350 ~~through~~ while Fe content varies throughout the life cycle of salmon (Shearer et al., 1994).
~~Our model only integrates size as a differentiating factor~~ Because our model uses size to differentiate between fish, ~~so~~ we analyzed aquatic ~~animals~~ animal body nutrient and body size data from Vanni et al. (2017) for any existing relationship between size and nutrient content. We found no relationship between body N content and size, and only a weak but significant relationship between body P and size (Supp. Fig. C1). In this data set, the changes of P content with size seem to be more

355 related to the difference between vertebrates and invertebrates, and to be significant for benthic organisms more than pelagic organisms. A recent study as indeed showed that the taxonomic identity is prominent in driving nutrient content variations compared to size (Allgeier et al., 2020). In addition, Hjerne and Hansson (2002) found no significant changes in the N and P content of fish with species (sprat and herring), fish size, seasons or different areas of the Baltic Sea, as did Griffiths (2006) for the P content of different fish species in lakes. ~~Available~~ The scant available data for Fe, does not allow us to draw conclusions

360 on the variations in Fe content with size. ~~Our model does not include distinctions between fish speciesor diet so we did not test the effect of theses factors on nutrient storage but included the uncertainty on the mean nutrient content of fish in our calculations of the total uncertainty (Tables 1, ?? and Supp. Table ??). Consequently, we did not include variations of body nutrient content with size, the only factor included in our model, in our calculations but simply used the mean values in Table 1 and the uncertainties around these values.~~

365 Although nutrient ratios show many fascinating variations between species, these are small relative to variations of fish biomass density in the ocean. Thus, most of the spatial variations of the nutrient content of fish are likely to be due to variations in fish abundance rather species assemblages, and the uncertainty related to nutrient ratios is likely to be small compared to the uncertainty on fish biomass.

4   Nutrient cycling by the commercial fish biomass

370 ~~Nutrients excreted by fish can directly be used by phytoplankton and bacteria as they often are in a bioavailable form (e.g., Vanni et al., 2006 egested material tends to sink rapidly (Wotton and Malmqvist, 2001).~~ Because they are capable of large-scale movement and alter the stoichiometry of particles, fish can have ~~different impacts than grazing. We gauge these potential~~ impacts on nutrient cycling that differ from single-celled heterotrophic plankton. In this section, we gauge the potential scale of these impacts by estimating the rates at which fish cycle nutrients, what fraction of primary productivity (PP) this cycling represents, and how

375 much it can contribute to the export of nutrients from the euphotic zone as sinking egested materials. Note that this is ~~intended only~~ to illustrate the potential magnitude, which could be built ~~only~~ upon with coupled fish biogeochemistry modeling.

The global cycling, i.e. excretion plus egestion, of nutrients by the pristine CTF_{10g}^{100kg} biomass represents about 210 ± 113 Tg of N per year, 15.6 ± 8.0 Tg of P per year and 0.12-0.77 Tg of Fe per year, of which about half is in the LMEs (Table ~~??~~ ~~and Supp. Table ??~~3). Like nutrient storage, modeled cycling by fish is larger where the biomass is higher (Fig. 6 and Supp.

380 Fig. D1). The three different ways of computing Fe cycling by the commercial fish biomass show similar spatial patterns, but ~~larger Fe cycling~~ Fe cycling is larger when using the weighted spatial variation between the low and high Fe:C estimates in zooplankton (Fig. 6~~, Table 1~~). The spatially weighted computations ~~strongly reflect~~ suggest the possibility that Fe cycling by CTF_{10g}^{100kg} might be reduced in HNLC regions (principally in the Southern Ocean and the subarctic Pacific Ocean).
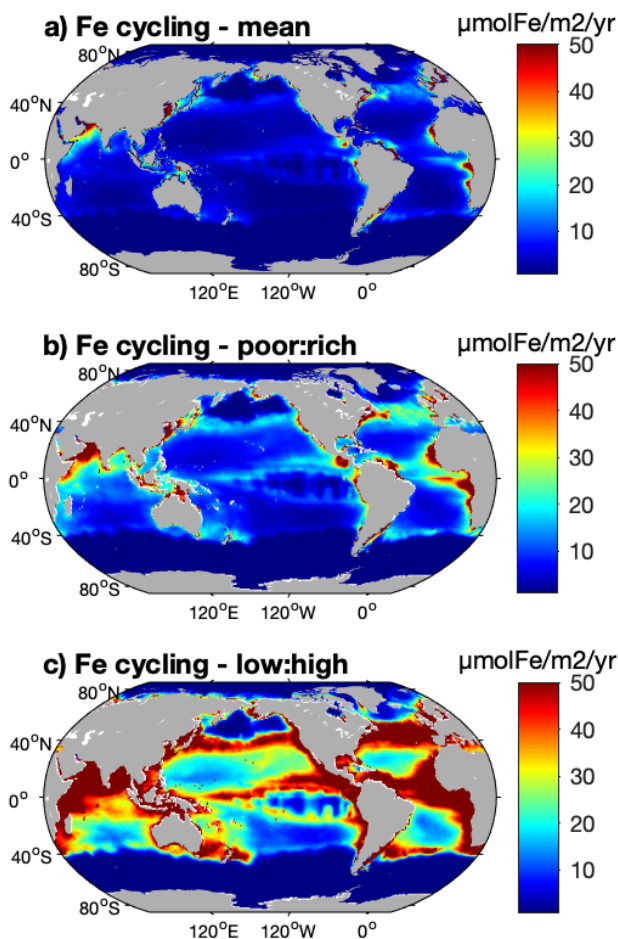
Figure 6. Iron cycling by ~~the~~ CTF_{10g}^{100kg} biomass computed using a) a mean Fe:C in zooplankton, b) a spatial variation between zooplankton Fe:C mean values in Fe-poor and Fe-rich areas and based on [NO3] concentrations, and c) the same spatial variation but using the low and high estimates of zooplankton Fe:C from data. (For more details see Methods)

4.1 Nutrient cycling by commercial fish ~~and~~ relative to primary ~~producers demand~~production

385 ~~Our analysis shows that if the nutrients are readily available, fish only represent a small source of recycled elements, larger in the regions where primary producers (PP) demand and fish biomass are high and in areas where the concentration of nutrients is low. Indeed, he~~ The modeled N cycling by pristine CTF_{10g}^{100kg} biomass contributes on average to $2.2 \pm 1.2\%$ of the N demand of ~~PP~~ primary producers (PP) in LMEs (Table ~~??~~3), and ~~globally~~ generally accounts for less than 5% of the demand, except in some coastal areas where it can be as high as 14% (Fig. 7a). ~~Overall~~Similarly, the modeled pristine P cycling ~~accounts for~~ by

390 CTF_{10g}^{100kg} represents 1.2% of the total P demand in the LMEs (Table 3), less than 4% of the global P demand ~~,~~ with a larger contribution in the North and equatorial Atlantic coastal regions, and larger than 6% contributions in some coastal areas (Fig. 7b). ~~This P cycling by CTF_{10g}^{100kg} represents 1.2% of the total P demand in the LMEs (Table ??).~~ The high-end estimate of the Fe cycling by CTF_{10g}^{100kg} relative to PP demand for Fe is slightly more significant than the ratios for N and P, as it represents up to 13% of the PP demand for Fe in some coastal areas, and less than 10% everywhere else (Fig. 7e).

395 ~~This model framework allowed us to analyze the fish nutrient cycling at the global scale, and our global~~ Our global estimates are broadly consistent with the order of magnitude influence of fish estimated by prior local studies. For all the coral reefs in the ocean, Allgeier et al. (2014) estimated that the total fish community supply about 1.2 Tg of N per year, which is about 0.6% of our global pristine estimate (0.8% compared to global cycling at peak catch), or 1.2% of the LMEs pristine estimate (2.1% of our N cycling by ~~the~~ CTF_{10g}^{100kg} biomass at global peak catch) (Table ~~??~~3)). This is consistent with the

400 fact that coral reefs cover about 255000-600000 km^2 (Spalding and Grenfell, 1997), which is about 0.4-0.8% of the LMEs area. Hernández-León et al. (2008) estimated that zooplankton supply about 1780 TgN/yr worldwide, representing 12-23% of the requirements of phytoplankton and bacteria. This estimated zooplankton cycling is about 11 times our modeled recycling rate by the pristine CTF_{10g}^{100kg} biomass and 6-12 times what CTF_{10g}^{100kg} cycling could provide for primary producers. ~~The Sheldon spectrum has a slope of 0 for the biomass~~ Given that the biomass distribution of all marine organisms ~~(?),~~assuming

405 ~~has a slope of 0 (Hatton et al., 2021),~~(Supp. Fig. E1), and assuming that zooplankton mass ranges ~~from 10^{-12}g to 10g it already represents~~ over 12 orders of magnitude, zooplankton biomass would be expected to be roughly 3 times ~~more orders of magnitude compared to CTF_{10g}^{100kg}, and likely as much more cycling~~the biomass of the total fish biomass in the 10 g to 100 kg size range. In addition, due to their higher metabolic and ingestion rates (Maldonado et al., 2016; Griffiths, 2006), zooplankton cycling rates are likely to be much higher than fish cycling rates, also exemplified by our modeled cycling size spectrum which

410 has a negative slope (Supp. Fig. E2). ~~Consequently, due to their high biomass, high metabolic rates and trophic proximity to primary producers (Maldonado et al., 2016; Griffiths, 2006), small plankton mediates most of the nutrient recycling.~~ McIntyre et al. (2008) showed that fish excretion can be important in supplying N and P to primary producers when conditions of high fish biomass and high PP demand or low ambient nutrient concentrations are combined, and when nutrient inputs from anthropogenic sources are low. Our results indeed suggest an increased contribution of fish cycling when these conditions are

415 combined. However, the strength of this contribution also depends on the timing between the ~~the~~release of nutrients by fish and the ~~the productivity needs~~ primary producers demand for these nutrients~~as we will discuss a little more in the case of nutrient~~
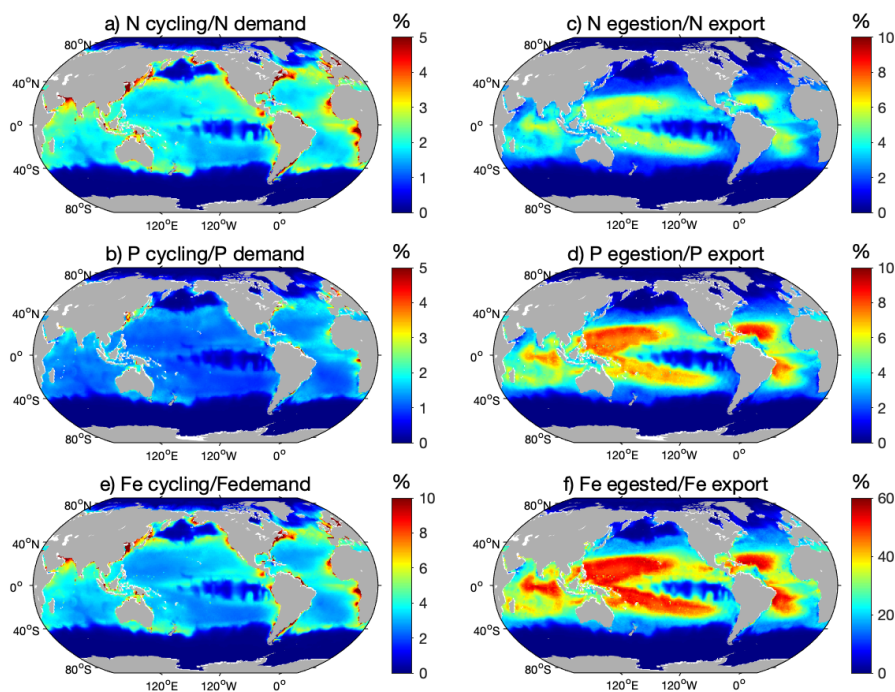
Figure 7. Total nutrient cycling and egestion relative to export. Ratio (%) between the amount of nutrients cycled by the modeled pristine global CTF_{10g}^{100kg} biomass and estimated primary producers demand for a) N and b) P, ratios of the modeled amount of nutrient egested by the pristine CTF_{10g}^{100kg} biomass and export at the base of the euphotic zone of c) N and d) P, and high-end estimate of the ratio (%) between the modeled amount of e) Fe cycled by the modeled pristine global CTF_{10g}^{100kg} biomass and estimated primary producers Fe demand, and f) Fe egested by the pristine CTF_{10g}^{100kg} biomass and Fe export at the base of the euphotic zone. The high-end estimates are obtained using Fe cycling computed from the weighted spatial variation between the low and high Fe:C values of zooplankton, and the weighted spatial variation between the averaged Fe:C ratios of phytoplankton in Fe-poor and Fe-rich conditions.

~~As for Fe, the~~ For Fe, CTF_{10g}^{100kg} cycling represents a more important fraction of the PP demand compared to N and P,

420   but large uncertainties remain in its computation. At the global scale, Moreno and Haffa (2014) estimated that the amount of Fe excreted by the commercial marine fish biomass ranged between 0.4-1.5 TgFe per year. Our modeled estimate range of 0.12-0.77 TgFe/yr, based on the high and low estimates (Fig. 7e and Supp. Fig.D1), is lower but overlapping. The difference can in part be attributed to the lower Fe:C values we used for zooplankton (Table 1), and highlights the uncertainty on the Fe cycling computation (Fig. 6). In the Southern Ocean, whales have been shown to contribute to a maximum of 0.2-0.3% of the

425   phytoplankton demand for Fe in a pre-whaling ecosystem and no more than 0.03-0.04% in a post-whaling ecosystem, making their contribution negligible compared to that of zooplankton (>70% for microzooplankton only) (Maldonado et al., 2016). With a modeled contribution of about 0.05-0.5% of the phytoplankton demand for Fe in the Southern Ocean, modeled pristine

CTF_{10g}^{100kg} cycling coherently might be able to sustain a larger part of primary productivity than the current whale population, but still far less than zooplankton as discussed before for N cycling.

430

~~Similarly to nutrient contents, some but not all types of variability in cycling rates are accounted for in this study. Excretion varies with body size (Vanni and McIntyre, 2016; Allgeier et al., 2015; Hall et al., 2007; Schindler and Eby, 1997), which is taken into account here through the variations of metabolic and production rates within the size-spectrum in the model (Carozza et al., 2017). This allows nutrient cycling to vary with size in our calculations. The effect of temperature on the energy flow and thus nutrient cycling is also integrated in the model (Carozza et al., 2017). Even though taxonomy, diet, ontogeny, and body nutrient content also influence fish recycling (e.g., Vanni and McIntyre, 2016; Allgeier et al., 2015; El-Sabaawi et al., 2016; Moody et were not included in our study apart from the uncertainty on zooplankton nutrient content (Table 1).~~ Finally, fish movements also allow the transport of nutrients ~~and constitute~~ laterally in the ocean, constituting a sink of nutrients where the fish forage and a source of nutrients where the fish excrete, egest or die (Vanni et al., 2013; Francis and Côté, 2018), an effect we do not explicitly include here.

4.2 Nutrient export by feces

Our results show that ~~the export of faecal~~ fish fecal material has the potential to affect the distribution of nutrients within the water column, especially in regions of low export intensity. Egested nutrients are integrated into ~~faecal~~ fecal pellets that sink out of the surface layer and are recycled at ~~larger~~ greater depths than if bound to smaller particles (Wotton and Malmqvist, 2001; Turner, 2015), especially fish fecal pellets that can sink faster and deeper than marine snow and phytodetritus (Saba and Steinberg, 2012). Figure 7c-d,f quantifies how much CTF_{10g}^{100kg} egestion may contribute to the export of N, P and Fe to depth using the mean absorption efficiencies in Table 1 and assuming exported nutrient ratios, without fish, are on average equal to those of phytoplankton. For all the nutrients, CTF_{10g}^{100kg}-mediated export accounts for a larger part of the export in the warm, low export regions of the world oceans, i.e. the tropical gyres, where it can contribute up to 50% of the exported Fe for the high-end estimate (Fig. 7f), 6% of the exported N and 10% of the exported P (Fig. 7c,d). Globally, modeled pristine CTF_{10g}^{100kg} biomass egests 29.4 ± 15.9 Tg of N, 4.5 ± 2.3 Tg of P and 0.009-0.59 Tg of Fe each year, which on average roughly accounts for $2.3 \pm 1.2\%, 3.0 \pm 1.5\%$ and 1.1-22% of the export of N, P and Fe, respectively, out of the euphotic zone (~~Supp. Table ??~~Table 3).

These results are in agreement with Davison et al. (2013) who showed that the contribution of mesopelagic fish to the carbon export (via respiration, excretion, egestion and death) is higher in regions where the total export is small. However, Davison et al. (2013) also showed that locally, in the California Current, the active transport of C by mesopelagic fish alone, which we do not model, represents about 15-17% of the total carbon export at depth~~, with spatial and temporal variations~~. In their modeling study, Aumont et al. (2018) estimated that, globally, diurnal vertical migration of epipelagic organisms (all migrating fish and zooplankton) contributes to the flux of carbon to depth of about 18% of the passive flux. ~~Egesting~~ Fish egesting and respiring at depth ~~transports~~ transport significant amounts of carbon, and thus also transfers nutrients from the surface to deeper

Table 3. Table of values from the model ensemble simulations in the pristine state and at the global peak catch at both the global scale and in LMEs. This table contains integrated values of: the amount of nutrient cycled by the CTF_{10g}^{100kg} biomass (Tg/yr), the ratio of this cycling with the global primary producers demand for these nutrients (%), the amount of nutrient egested by the CTF_{10g}^{100kg} biomass (Tg/yr) and its ratio with the exported nutrient quantities (%).

		N	P	Fe (low estimate)	Fe (high estimate)
Total Cycling (Tg/yr)					
Pristine	*Global*	210 ± 113	15.6 ± 8.0	0.12 ± 0.25	0.77 ± 0.19
	LME	101 ± 55	7.5 ± 3.9	0.06 ± 0.12	0.36 ± 0.09
At global peak catch	*Global*	145 ± 88	10.8 ± 6.3	0.08 ± 0.18	0.56 ± 0.17
	LME	56 ± 35	4.2 ± 2.5	0.03 ± 0.07	0.21 ± 0.07
Cycling/PP demand (%)					
Pristine	*Global*	1.5 ± 0.83	0.91 ± 0.47	0.16 ± 0.33	2.7 ± 0.66
	LME	2.2 ± 1.2	1.2 ± 0.6	0.26 ± 0.53	4.0 ± 1.0
At global peak catch	*Global*	1.2 ± 0.75	0.76 ± 0.44	0.12 ± 0.26	2.2 ± 0.68
	LME	1.5 ± 0.9	0.85 ± 0.5	0.15 ± 0.33	2.6 ± 0.8
Egestion (Tg/yr)					
Pristine	*Global*	29.4 ± 15.9	4.5 ± 2.3	$(9.1 \pm 18.9)10^{-2}$	0.59 ± 0.14
	LME	14.1 ± 12.3	2.2 ± 1.1	$(4.4 \pm 9.1)10^{-2}$	$(2.7 \pm 0.7)10^{-1}$
At global peak catch	*Global*	20.4 ± 12.3	3.1 ± 1.8	$(6.3 \pm 13.5)10^{-2}$	0.42 ± 0.13
	LME	7.8 ± 4.9	1.2 ± 0.7	$(2.4 \pm 5.2)10^{-2}$	$(1.6 \pm 0.5)10^{-1}$
Egestion/Export (%)					
Pristine	*Global*	2.3 ± 1.2	3.0 ± 1.5	1.1 ± 2.3	21.7 ± 5.3
	LME	2.1 ± 1.1	2.6 ± 1.3	1.1 ± 2.3	20.1 ± 5.0
At global peak catch	*Global*	2.1 ± 1.2	2.7 ± 1.6	1.0 ± 2.1	19.5 ± 6.0
	LME	1.6 ± 1.0	2.1 ± 1.2	0.8 ± 1.7	15.7 ± 5.0

layers, ~~which would probably~~ a process that would have increased the contribution of fish to the export of nutrient ~~in this study~~ if represented in the model.

More than ~~their~~ the fish contribution to total export, ~~fish~~ their effect on particles may be most relevant for stoichiometric changes, especially for Fe. Indeed, since the absorption efficiency of Fe is smaller than that of C, the Fe:C in feces will be greater than in the ingested particles ~~(Le Mézo and Galbraith, 2020). Thus,~~, and exported fecal material will have a greater Fe:C than biogenic sinking particles made of phytoplankton aggregates or dead organisms (Le Mézo and Galbraith, 2020).

5 Extended size-spectrum and total fish biomass

470 ~~In this study, we only considered the~~ Until now we have considered CTF_{10g}^{100kg} ~~, which is the target range of the~~ as represented explicitly by the BOATS model, ~~but it does not include all fish in terms of size and non-commercial biomass. First, a more inclusive size range would be from small larvae , about 1g, to sharks, about~~ which ignores non-commercially targeted fish as well as the small and large ends of the fish size range. We provide a rough estimate of how the total fish biomass within a more inclusive marine size-spectrum spanning 1g larvae to 10^6g ~~, i. e. $CTF_{1g}^{10^6 g}$, which encompasses~~ sharks (F_{1g}^{1000kg}) would

475 compare to our estimates in two steps. First, we take the Bianchi et al. (2021) estimate that the CTF_{10g}^{100kg} is supported by roughly half of the total primary production that could be available to this size range (best estimate 58%), from which we calculate that if the non-commercial fraction were included the total biomass estimate would be about 5.2 Gt. We then consider that the expanded size range includes 6 orders of ~~magnitudes~~ magnitude compared to the 4 orders of magnitude of the standard BOATS size range. ~~We do not include planktonic larvae as small as $10^{-5}g - 10^{-1}g$ because these size classes tend to be

480 dominated by zooplankton (Hatton et al., 2021). We first estimate the amount of nutrients stored within this extended marine spectrum, $CTF_{1g}^{10^6 g}$, by analyzing~~ In BOATS, the size spectrum of ~~fish abundance in BOATS. The size spectrum of abundance~~ abundance has a slope of about -1, ~~and~~ giving the biomass size spectrum a slope of 0 (Fig. SE1) ~~as predicted by the~~ consistent with the observed Sheldon spectrum (Hatton et al., 2021). ~~So by extension to smaller and larger sizes~~ Thus, by extension, we estimate that the ~~$CTF_{1g}^{10^6 g}$~~ biomass contains about 1.5 times (6 orders of magnitude versus 4 orders of magnitude) more

485 biomass ~~than the CTF_{10g}^{100kg} biomass, which represents 6.9 Gtons of~~ , giving a total of 7.8 Gt of wet biomass. Overall, we estimate that F_{1g}^{1000kg} exceeds CTF_{10g}^{100kg} by a factor of 3.1.

~~These estimates do not take into account non-commercial fish species, especially mesopelagic fish on which there are poor constraints. Bianchi et al. (2021) estimated that the commercially-targeted fish represent only half of the total fish biomass. Consequently, we could estimate that $CTF_{1g}^{10^6 g}$ biomass in the pristine state represents about 20.7 Gtons of wet biomass in this~~

490 ~~model. The extended size-spectrum estimate of all commercial and non-commercial fish biomass is likely as low as 2-times and high as 8-times the CTF biomass within BOATS' range.~~

~~By extending the size spectrum , in addition to increasing fish biomass the icthyosphere would~~ The extended size spectrum contains more biomass, but it also contains more small fish than ~~in~~ the standard size range of BOATS, a combination that would change the cycling rates of the total fish biomass. Since smaller fish are shown to have higher cycling rates than large ones

495 (?), we expect ~~a higher cycling estimates by the global fish biomass . Indeed, in~~ higher cycling of nutrients by the extended size spectrum than would be implied by the larger biomass alone. In the pristine ocean modeled here by BOATS, the C cycling ~~size-spectrum has~~ rates decrease with size following a slope of $-0.37 molC.m^{-2}.g^{-1}$ ~~, and of $-0.57 molC.m^{-2}.g^{-1}$ at the global peak catch~~ (Supp. Fig. E2). ~~Based on these slopes,~~ Based on this slope, and the biomass slope of 0, we would expect that the ~~global fish biomass, including non-commercial fish species, between 1g and 10^6g, would cycled about 4~~ CTF_{1g}^{1000kg}

would have cycled about 2.4 times more C in the pristine ocean ~~and about 3 times more C at global peak catch~~ compared to the CTF_{10g}^{100kg} ~~biomass represented in BOATS in the same conditions.~~ , with the 1g-10g size range cycling 40% more C than the rest of the spectrum combined. Taken all together, the total cycling by F_{1g}^{1000kg} would have been a factor of 4.8 greater than the CTF_{10g}^{100kg} rates discussed throughout the paper above. Assuming the additional organisms eat prey with similar body nutrient contents as the organisms already modeled, all of the nutrient cycling rates presented above would be increased proportionally.

6   Fish catch: anthropogenic extraction of nutrients from the ocean

As fishing activity represents a direct removal of nutrients from the ocean, we estimated how much nutrients were extracted at the global peak catch and how these extraction rates would compare to nutrient inputs to the ocean ~~since our model spatially represents fish at the global scale. The majority of the catch takes place where fish biomass is modeled to be high: the North Atlantic and Pacific, the east equatorial Pacific, around 40S and along the coasts. Globally,~~ . Globally, we estimate that modeled fishing activity removes 5.4 ± 0.7 ~~TgN/yr~~ $TgN.yr^{-1}$, 1.2 ± 0.3 ~~TgP/yr~~ $TgP.yr^{-1}$ and $0.09 - 1.8$ ~~10^{10} gFe/yr~~ $10^{10} gFe.yr^{-1}$ from the ocean at the time of global peak catch ~~(Supp. Table ??)~~, of which a little less than 50% in LMEs (Table ~~??)~~. 4). Although our model is calibrated to agree reasonably well with observed catch maxima in LMEs over the years 1950 to 2010 (Pauly and Zeller, 2016; Bianchi et al., 2021), the total catch varies between ensemble members and tends to be overestimated in the open ocean, even with the Fe-limitation we added in this study, for reasons that remain unclear. In addition, the catch estimate we present is at the time of the global peak catch in idealized simulations, rather than being historically accurate. Consequently, the simulated average catch at global peak (196.2 ± 57 Tg wet weight) is higher than the peak catch estimated from fishery observations (130 ± 65 Tg), so our estimates may exceed actual wild capture extractions by 50%, with the most significant overestimates in the open ocean (see Bianchi et al. (2021) for more details).
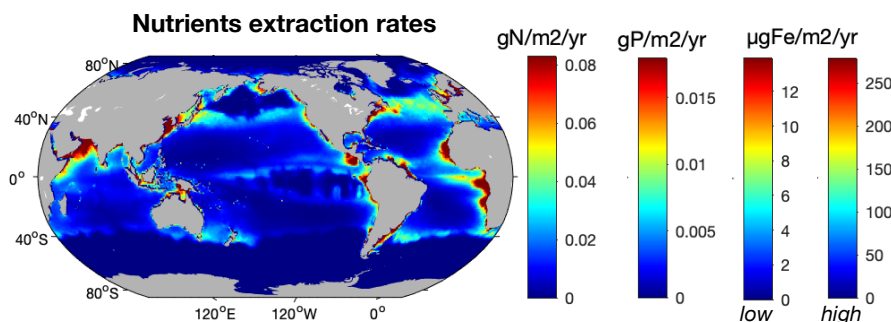
Figure 8. Distribution of the modeled amount of N (gN/m2/yr), P (gP/m2/yr) and Fe (μgFe/m2/yr) extracted from the ocean at the time of the global peak catch. The two colorbars for Fe represent the low and high estimates based on the 95% confidence interval for Fe:C values in fish (10-200 μmolFe/molC).

6.1 Nitrogen

Our estimate of N extraction from the CTF_{10g}^{100kg} catch is ~~in coherent with previous estimates~~ consistent with previous computations, and the differences mostly reflects the ~~choice of~~ fish biomass estimates~~in previous studies compared to the one used to calibrate this model~~. For example, using catch data, Maranger et al. (2008) estimated the amount of N returned to land via fishing in LMEs to be about 0.9 TgN/yr in the 60's and 2.3 TgN/yr in 2000. Our estimate of the N content of fish catch at global peak catch in LMEs is 2.8 ± 0.4 Tg of N per year, which is slightly larger than Maranger et al. (2008) 2000's estimate. ~~The difference might be explained by the fact that our model is calibrated to fit observed catches over the years 1950 to 2010 (Pauly and Zeller, 2016; Bianchi et al., 2021) and the catch estimate we use is at the time of the global peak catch. Moreover, our mean N content of fish (2.8% of wet weight) (Table 1) induces a difference of about 0.2 Tg N per year.~~ Allgeier et al. (2017) also estimated the amount of N globally harvested to be about 2.072 Tg of N per year using Maranger et al. (2008)'s 2.6% N content and the FAO catch data. The FAO data only contains reported catches and consequently is lower than the SAUP data used in Maranger et al. (2008) and used to calibrate BOATS, which explains part of the difference~~with our estimate of 5.4 ± 0.7 TgN/yr at the global scale (see Bianchi et al. (2021) for more details).~~.

Our study framework allows to spatially compare extracted nutrients to nutrient inputs, which was not the case in previous work. For N, even though N extraction by fishing can be significant locally compared to N deposition at the surface, N extraction is negligible compared to the other sources of N to the surface layers. Figure 9a compares the amount of N extracted by fishing to the modeled soluble atmospheric N deposition from Brahney et al. (2015). Globally, fishing removal of N is smaller than current modeled atmospheric deposition of soluble N, with the higher values, up to more than 60% of the N deposition, in the southern equatorial Pacific, along the western margins of Africa and South America and in the Arabian Sea, where catch is high and deposition is low (Supp. Fig. B1). However, most of N supply to the surface ocean occurs through vertical diffusion and mixing of the upper layers (Sarmiento and Gruber, 2006), which likely accentuates the fact that N extraction by fishing is insignificant at the global scale.

6.2 Phosphorus

Similarly to N, our estimate of P extraction by fishing is coherent with previous work and shows that it is very small compared to inputs of P to the ocean ~~andd~~ and resupply from vertical mixing and diffusion in the water column. We estimate that the amount of P removed by fishing at the global scale amounts to 1.2 ± 0.3 TgP/yr, of which 0.6 ± 0.2 TgP/yr occur in the LMEs (Table ~~?? and Supp. Table ??~~4). Huang et al. (2020) estimated that wild and aquaculture fisheries, including finfish, crustaceans and molluscs, represented 1.1 Tg of P in 2016, which ~~agrees very well with~~ is superficially similar to our estimate. However, ~~we should note that their calculation is based on a catch of 169 Tg of biomass containing finfish, crustaceans and molluscs, while our estimated catch of 196.2 ± 57 Tg of wet weight, is larger than~~ the global amount of catch ~~we modeled is higher, 196.2 ± 57 Tg of wet weight~~they used of 169 Tg, even though ~~Huang et al. (2020) also considers~~ they considered aquaculture in addition to wild captures. Our catch estimate at global peak catch ~~likely overestimate catch (Bianchi et al., 2021)~~overestimates the high seas catch as discussed at the beginning of this section and in~~Bianchi et al. (2021)~~. Additionally, our estimate is solely based

on fish P content (Table 1), which may slightly overestimate the amount of P extracted by fishing activity since crustaceans and molluscs have lower P content than finfish (Huang et al., 2020).

555 Contrary to N, the modeled removal of P from harvest would largely exceeds the atmospheric deposition of soluble P as P inputs to the ocean mostly occur through riverine inputs (Table 4). Consequently, catch transfers P from the ocean to land where as P supply to the ocean is mostly occurring in the coastal areas, with possible impacts on the P budget of the open ocean (Huang et al., 2020). But similarly to N, vertical diffusion and mixing of the upper layers supplies P to the surface ocean in quantities that most likely render P extraction by

560 fishing relatively insignificant.

6.3 Iron

Fe extraction by fishing activity is within the range of previous estimates but large uncertainties remain due to uncertainty regarding the Fe:C of fish. Moreno and Haffa (2014) investigated the extent to which commercial catch has globally translocated Fe from the ocean to land. They estimated the global rate of translocation of Fe to

565 be between 0.007 and 0.03 Tg in 2010. Our modeled global range of Fe removal is about $0.0009 - 0.018$ TgFe/yr. Our lower estimated values can once again be explained by the difference in the Fe:C ratios used for fish. Even though our estimate is lower, it shows that locally Fe extraction can be significant compared to Fe inputs from dust deposition. Although the high-end estimate of Fe extracted is globally small relative to modeled soluble Fe deposition from Mahowald et al. (2009), it reaches values larger than 100% in the coastal eastern equatorial Pacific and in some other

570 coastal areas such as Western South Africa, Northern Europe and Canada (Fig. 9b), where modeled Fe deposition is small and harvest is high (Fig. SB1). Contrary to N and P, Fe has a much shorter residence time and thus is subjects to local perturbations, among which Fe extraction by fishing could be important.

6.4 Local and time-dependent nutrient budgets

Nutrient budgets are subject to perturbations in space and in time that can modify the relative strength of the nutrient extraction

575 by fishing activity. Some local nutrient budgets have been investigated to compare the amount of nutrient extracted by fishing to the nutrient loads (e.g. Hjerne and Hansson (2002)). If we were to do similar budgets, at the global scale, assuming all P inputs come from rivers and atmospheric deposition, which represents 48.5 TgP/yr (Table 4 and B1), then the extracted P flux represents 2.5% of the global input flux (1.2% over the LMEs). For N, global catch represents about 2% of the combined N inputs from atmospheric deposition (49.6 TgN/yr),

580 rivers (80 TgN/yr) and N_2-fixation (140 TgN/yr) (Tables 4 and B1).

Note that fish extracted from a given area may have foraged elsewhere, especially large fish able to undertake long-distance migrations like tuna, salmon or sharks (e.g., Afonso et al., 2017; Gresh et al., 2000). Consequently, the ratios between extracted nutrients and nutrient deposition may be over- or under-estimated, thus over- or under-estimating the role of fishing as a local sink of nutrients (Vanni et al., 2013). In addition, the relative timing of fishing effort along with phytoplankton growth, nutrient

Table 4. Table of values in LMEs from the model ensemble simulations in the pristine state and at the global peak catch. This table contains integrated values of: the amount of nutrients removed by fishing in LMEs (Tg/yr) and the ranges of values of the global nutrients inputs to the ocean from the literature (Tg/yr).

	N	P	Fe (low estimate)	Fe (high estimate)
Catch (Tg/yr)				
Global	5.4 ± 0.7	1.2 ± 0.3	$9.1 10^{-4}$	$1.8 10^{-2}$
LMEs	2.8 ± 0.4	0.6 ± 0.2	$(4.7)10^{-4}$	$(9.3)10^{-3}$

Global inputs to the ocean (Tg/yr)	N	P	Fe
Soluble depostion	$16 - 63^{a,c,e,h,j}$	$0.1 - 0.5^{a,b,c,d,i,j}$	$0.6 - 13.4^{b,c,f,g,k}$
Rivers	80^{h}	$0.93 - 48^{i}$	$0.08 - 0.09^{k}$
N_2 fixation	140^{h}	-	-
Iceberg melting	-	-	$0.09 - 0.1^{k}$

[a]Brahney et al. (2015), [b]Mahowald et al. (2009), [c]Okin et al. (2011), [d]Myriokefalitakis et al. (2016), [e]Fowler et al. (2013), [f]Ito (2015) , [g]Wang et al. (2015), [h]Gruber and Galloway (2008), [i]Benitez-Nelson (2000) , [j]Kanakidou et al. (2012), [k]Moreno and Haffa (2014)
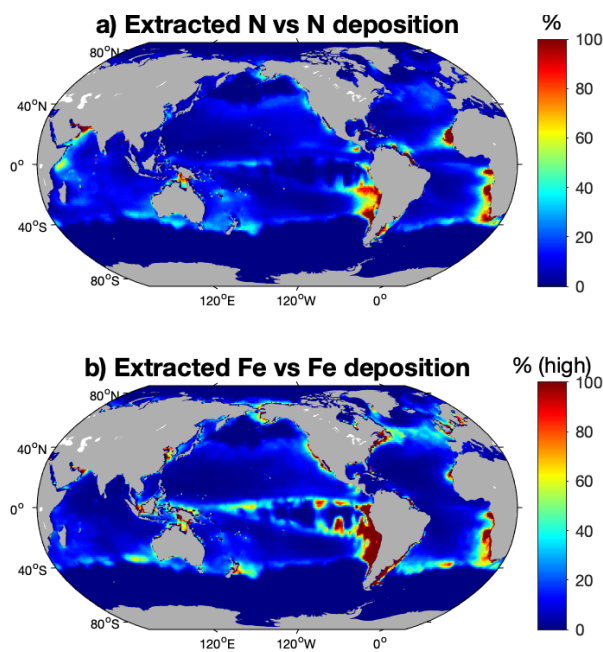
Figure 9. Ratio (%) between simulated extracted nutrients and current aeolian soluble nutrients inputs at the surface of the ocean for a) N and b) Fe. "low" and "high" refer to the use of 10 μmolFe/molC or 200 μmolFe/molC in fish, respectively.

585   ~~inputs~~ input seasonality and residence times may also modify the importance of fishing activity as a sink of nutrients (e.g., Francis and Côté, 2018; Vanni et al., 2006).

6.5   ~~Fishing and~~ Reductions in nutrient cycling caused by fishing

Fishing has had a dramatic influence on nutrient cycling by CTF_{10g}^{100kg}, as it ~~removes a lot~~ has permanently removed a large amount of biomass, especially in the large size classes~~, which our analysis clearly shows~~. In our ensemble of simulations,
590   the cycling rates decrease by about 30% for the three elements considered at the time of the global peak catch (Table ~~?? and~~ ~~Supp. Table ??~~3), due to the global reduction in fish biomass of about 60%. ~~The reduction in total nutrient cycling is not as~~ ~~pronounced as the biomass reduction between the pristine state and the global peak catch state because the remaining fish~~ ~~at the global peak catch are smaller and have higher mass-specific metabolic rates (Bianchi et al., 2021). By targeting mostly~~ ~~large fish and specific species, fishing modifies the community size structure and trophic interactions thereby changing the~~
595   ~~animal-mediated nutrient cycling (e.g., Schmitz et al., 2010; Vanni, 2002). In our model ensemble, only changes in the size~~ ~~spectrum structure due to fishing are accounted for (Bianchi et al., 2021). Large fish are targeted first, and~~ Since large fish are heavily targeted by fisheries, the size-spectrum of CTF ~~is thus~~ appears truncated at larger size class at the time of the global peak catch compared to a pristine state (Supp. Fig. E1). This reduction of the mean community size enhances the cycling of elements ~~, because~~ as smaller animals tend to have higher metabolic rates ~~. However, the reduction from pristine biomass to~~
600   ~~biomass at the global peak catch predominates, and globally fish cycling is reduced~~ (e.g. Schmitz et al., 2010; Vanni, 2002), so that the reduction in overall cycling rate is roughly half the reduction of biomass.

    Finally, as the size classes below 10g are not resolved (Fig. 2) we are not able to account for biomass changes that fishing might induces through trophic cascades, or how it reverberates up to fish through food supply (Dupont et al., submitted), which has the potential to further modify fish-mediated nutrient cycling.

605 ## 7   Conclusions

In this study, we estimate the amount of ~~nutrients,~~ N, P and Fe ~~,~~ contained in and cycled by the global CTF_{10g}^{100kg} biomass, both in its pristine state and at the time of the global peak catch. The overall contribution of this commercial ~~icthyosphere~~ ~~is small but~~ ichthyosphere to oceanic nutrient cycling is relatively small, but is more significant in regions of low ambient nutrient concentrations, high fish biomass and low export production. ~~Catch~~ The industrial fish catch generally represents a
610   small extraction of nutrients globally compared to external inputs, ~~but~~ though it removes significant amount of P from the open ocean compared to external inputs ~~that mainly occurs from rivers. However,~~ (mainly riverine). In general, local cycling of N and P ~~cycling~~ by fish is less significant than Fe cycling by fish because N and P are resupplied globally through ~~resuspension~~ ~~by mixing~~ large-scale circulation processes, while Fe cycling is ~~much~~ more local and susceptible to perturbations through rapid scavenging for example. In addition, poor absorption of Fe by fish leads to an enrichment of the Fe content of fecal matter.
615   Globally, nutrient cycling by the modeled CTF_{10g}^{100kg} biomass is small compared to primary producers demand for these nutrients. The highest contributions are found close to the coasts where fish biomass and productivity demand are high. Fish

egestion of nutrients via faecal pellets is ~~most important~~ likely to comprise the largest fraction of the sinking flux in regions of low export production, i.e. the ~~tropical gyres, especially for Fe. However, fecal~~ subtropical gyres. Fecal pellets may also significantly impact the stoichiometry of sinking particles, especially for Fe, with consequences for mesopelagic organisms.

620 ~~Finally, even though these contributions of fish do not appear to be very significant at the global scale they are~~ Our study provides a first global glimpse of nutrient cycling by the ichthyosphere. Although the contribution of CTF_{10g}^{100kg} simulated by our model tends to be on the order of only a few percent of total surface nutrient budgets and fluxes, we estimate that these would be a factor of 3 or more larger for the 1 g to 1000 kg range, including all fish. Furthermore, the role of fish in shaping the ecosystem processes through top-down pressure on their prey may be of similar or greater magnitude than

625 the quantities estimates here (Frank et al., 2005; Baum and Worm, 2009; **?**; Kavanagh and Galbraith, 2018). Fish can also be highly relevant at the local scale, ~~especially for Fe as mentioned before~~for example by changing the vertical distribution of Fe in the water column as mentioned above, and as highlighted by many studies on fish in coral reefs~~for example. Fish contributions to nutrient cycles may also be enhanced by top-down effects and trophic cascades that operate on longer time-scales (Kavanagh and Galbraith, 2018). This study does not account for several factors~~. Unresolved factors, such as fish

630 migrations~~that would alter the results , especially as in response~~, would alter our results and the sensitivity of fish-mediated nutrient cycling to warming and deoxygenation due to climate change (e.g., Lefort et al., 2015; Lotze et al., 2019). There remains much to be learned about the role of fish in global nutrient cycling.

Appendix A: ~~Global values and nutrient inputs to the ocean~~Model-data comparison

~~Table~~The Bayesian Monte Carlo approach was not performed for the updated model with Fe limitation that we use in this study. Instead, we took the 31 combinations of selected parameters from the Monte Carlo approach on the standard version of the model performed in Bianchi et al. (2021) and run 31 simulations with the updated Fe-limited version of the model. The Fe limitation decrease global fish biomass slightly (Galbraith et al., 2019) so that we underestimate the global LME peak catch of $\sim 110 Mt.yr^{-1}$ (Fig. A1). The comparison of modeled versus observed peak catch across LMEs gives an r^2 of ~~global values from the model ensemble simulations in the pristine state and at the global peak catch. This table contains globally integrated values of: **1.** nutrient content in fish biomass~~ 0.42 ~~(Tg), **2.** the ratio~~ $p-value < 10^-9$) for the ensemble average, while the ensemble meber with the best fit to observations has an r^2 of ~~nutrient content in fish with surface nutrient conentrations~~ 0.57 ~~(%), **3.** the amount of nutrient cycled by the~~ $p-value < 10^-9$) (Fig.A2)(Bianchi et al., 2021).
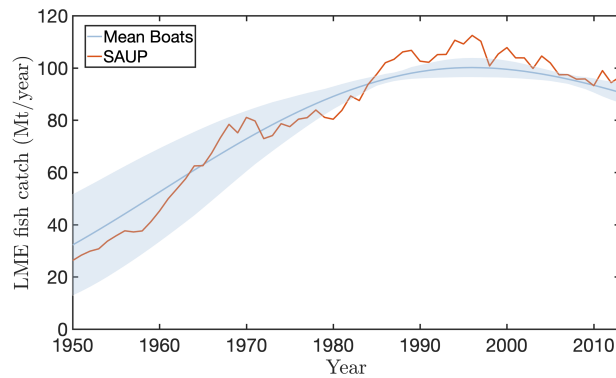
Figure A1. ~~Modeled~~ fish ~~biomass (Tg/yr), **4.**~~ catch from the ~~ratio~~ 31 simulations ~~of this cycling with~~ the ~~global primary producers demand for these nutrients (%), **5.** the amount of nutrient egested by the fish biomass (Tg/yr) and **6.** its ratio with the exported nutrient quantities (%), and **7.** the amount of nutrients removed via harvest at the global scale (Tg/yr)~~ ensemble from 1950 to 2010s compared to SAUP reconstructed catch data.

~~**N P Fe (low estimate)Fe (high estimate)**~~

~~Pristine~~ 68.7±30.5 14.9±13.6 (1.2±0.4)10^{-2} 0.23±0.07At global peak catch 26.2±14.7 5.7±5.8 (4.4±1.9)10^{-3} (8.8±3.8)10^{-2}

~~Pristine~~ 21.7±9.6 7.1±6.5 0.50±0.16 9.9±3.1At global peak catch 7.9±4.4 2.3±2.4 0.17±0.07 3.4±1.5Pristine 210±113 15.6±8.0 0.12±0.25 0.77±0.19At global peak catch 145±88 10.8±6.3 0.08±0.18 0.56±0.17
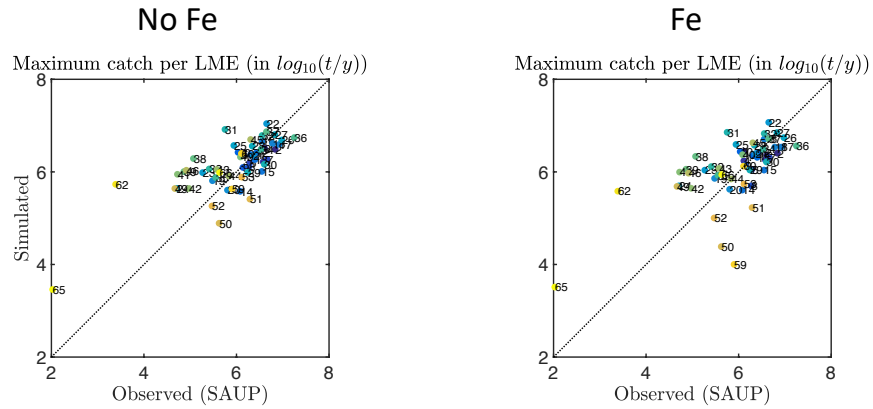
Figure A2. Comparison between LMEs maximum catch from data and from the model ensemble in the ensemble of simulations with no Fe limitation and in the ensemble of simulations with Fe limitation. Every number represents one LME.

Pristine 1.5±0.83 0.91±0.47 0.16±0.33 2.7±0.66 *At global peak catch* 1.2±0.75 0.76±0.44 0.12±0.26 2.2±0.68

Appendix B: Global values and nutrient inputs to the ocean

Pristine 29.4±15.9 4.5±2.3 (9.1±18.9)10^{-2} 0.59±0.14 *At global peak catch* 20.4±12.3 3.1±1.8 (6.3±13.5)10^{-2} 0.42±0.13

Pristine 2.3±1.2 3.0±1.5 1.1±2.3 21.7±5.3 *At global peak catch* 2.1±1.2 2.7±1.6 1.0±2.1 19.5±6.0

Global 5.4±0.7 1.2±0.3 9.1$10^{-4}$ 1.8$10^{-2}$
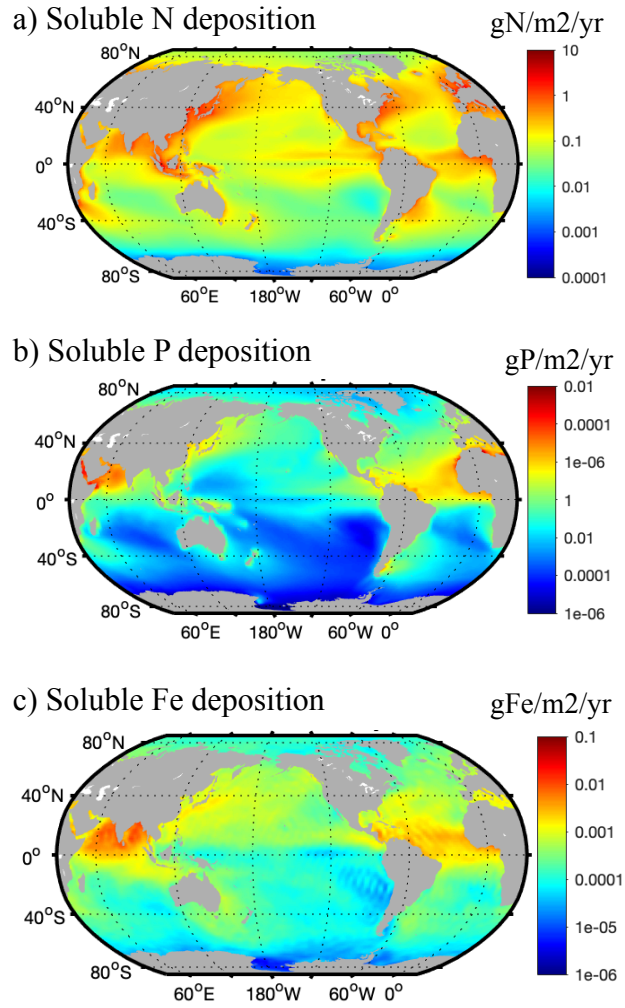
Figure B1. Modeled deposition fields of soluble a) N (gN/m2/yr), b) P (gP/m2/yr) and c) Fe (gFe/m2/yr) used to make Figure 9. N and P fields are from Brahney et al. (2015), Fe field is from Mahowald et al. (2009).

Table B1. Inputs of nutrients to the ocean.

Sources (Tg/yr)	N	P	Fe	Reference	Comment
Atmospheric deposition	63	0.32	0.36	Okin et al. (2011)	
	-	0.17	-	Myriokefalitakis et al. (2016)	bioavailable P
	29.4	-	-	Fowler et al. (2013)	
	-	-	13.4	Ito (2015)	
	-	-	8.4	Wang et al. (2015)	With Anthropogenic input
	50	-	-	Gruber and Galloway (2008)	NO_3^- and NH_4^+
	-	0.31	-	Benitez-Nelson (2000)	pre-anthropogenic, soluble reactive P
	16 (6.4)	0.35 (0.02)	-	Kanakidou et al. (2012)	organic soluble (anthropogenic contribution)
	36.6	-	-	Kanakidou et al. (2012)	Total inorganic N
	-	0.24 (0.034)	-	Mahowald et al. (2009)	Inorganic P (anthropogenic contribution)
	-	-	0.6-2	Moreno and Haffa (2014)	
Rivers	-	0.93-4.7	-	Benitez-Nelson (2000)	pre-anthropogenic
	-	23-48	-	Benitez-Nelson (2000)	Total with anthropogenic
	80	-	-	Gruber and Galloway (2008)	
	-	-	0.08-0.09	Moreno and Haffa (2014)	
N₂ fixation	140	-	-	Gruber and Galloway (2008)	
Iceberg melting	-	-	0.09-0.1	Moreno and Haffa (2014)	

Appendix C: Body nutrient content versus body size

	Nitrogen				Phosphorus			
	Intercept	Slope	R	p-value	Intercept	Slope	R	p-value
All	9.9	0.08	0.07	4.3e-5**	2.4	0.42	0.58	3.4e-299**
Invertebrate								
All	9.3	-0.07	-0.05	0.0138*	1.3	0.12	0.34	4.2e-63**
Pelagic	8.0	-0.37	-0.25	6.1e-24**	0.9	0.01	0.04	0.113
Benthic	10.2	0.07	0.05	0.207	1.7	0.43	0.66	3.3e-87**
Vertebrate								
All	10.4	-0.41	-0.29	2e-21**	3.2	0.37	0.29	8.4e-22**
Pelagic	11.4	0.27	0.26	0.058	2.7	0.04	0.05	0.74
Benthic	10.3	-0.58	-0.39	3.6e-37**	3.3	0.45	0.34	4.0e-28**

Figure C1. Body N (% of dry weight) and body P (% of dry weight), as a function of body mass (log(g)) for pelagic (purple) and benthic (blue) invertebrates and for pelagic (orange) and benthic (red) vertebrates. Regression lines for each type of organisms are shown in the same color. The dashed black line is the global regression line. Regression coefficients are given in the underneath table. Data from Vanni et al. (2017).

Appendix D: Cycling ~~computation and~~ of N and P, Fe cycling computations versus PP demand for Fe
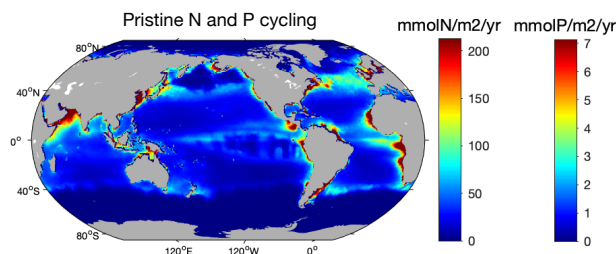
Figure D1. N and P ~~pristine~~ cycling prior to industrial fishing.

Appendix E: ~~Fe cycling versus PP demand for Fe: different computations~~
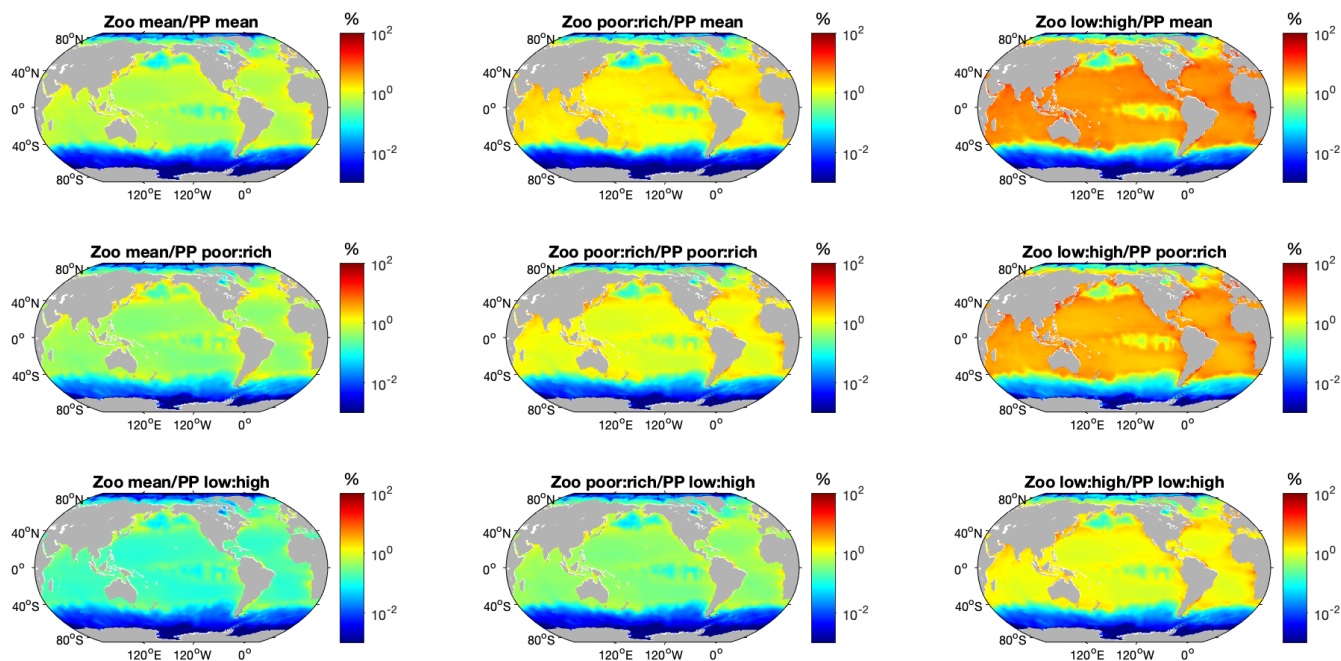
Figure D1. Fe cycling divided by phytoplankton demand for Fe. Columns distinguish the Fe cycling computation: left = using a mean Fe:C in zooplankton, middle = using a linear interpolation between the zooplankton mean Fe:C in Fe-rich and in Fe-poor conditions, right = using a linear interpolation between the zooplankton low Fe:C and high Fe:C estimates, interpolations are based on nitrate concentrations as a proxy for HNLC/non-HNLC areas. Lines distinguish the Fe demand of phytoplankton computation: top = linear interpolation between mean Fe:C values in Fe-rich and Fe-poor conditions, bottom = linear interpolation between low and high Fe:C estimates (Table 1).

 Appendix E: Size-spectrum of abundance, biomass and C cycling
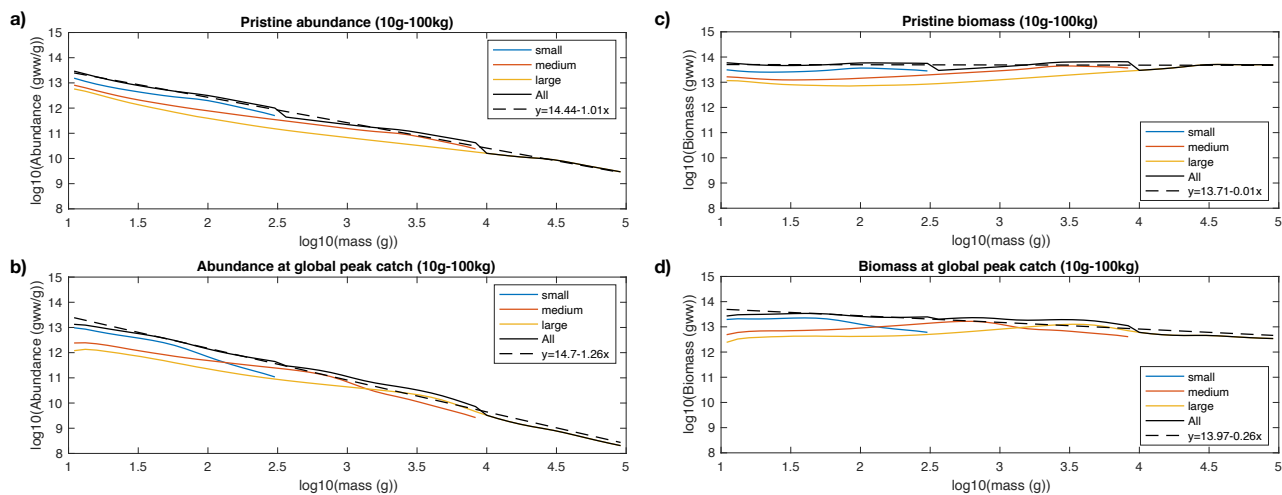
Figure E1. Size-spectrum of modeled CTF_{10g}^{100kg} abundance a) in the pristine state and b) at global peak catch, and size-spectrum of modeled CTF_{10g}^{100kg} biomass c) in the pristine state and d) at global peak catch
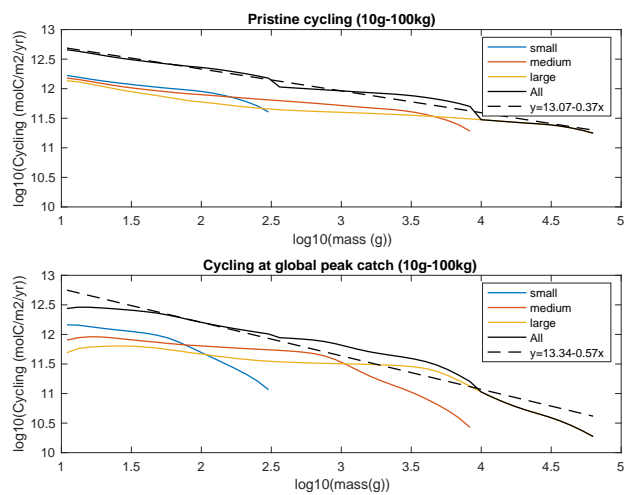
Figure E2. Size spectrum of modeled CTF_{10g}^{100kg} C cycling a) in the pristine state and b) at global peak catch.

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References

Afonso, A. S., Garla, R., and Hazin, F. H. V.: Tiger sharks can connect equatorial habitats and fisheries across the Atlantic Ocean basin, PLOS ONE, 12, e0184 763, https://doi.org/10.1371/journal.pone.0184763, https://dx.plos.org/10.1371/journal.pone.0184763, 2017.

Allgeier, J. E., Layman, C. A., Mumby, P. J., and Rosemond, A. D.: Consistent nutrient storage and supply mediated by diverse fish communities in coral reef ecosystems, Global Change Biology, 20, 2459–2472, https://doi.org/10.1111/gcb.12566, 2014.

Allgeier, J. E., Wenger, S. J., Rosemond, A. D., Schindler, D. E., and Layman, C. A.: Metabolic theory and taxonomic identity predict nutrient recycling in a diverse food web, Proceedings of the National Academy of Sciences, 112, E2640–E2647, https://doi.org/10.1073/pnas.1420819112, http://www.pnas.org/lookup/doi/10.1073/pnas.1420819112, 2015.

Allgeier, J. E., Valdivia, A., Cox, C., and Layman, C. A.: Fishing down nutrients on coral reefs, Nature Communications, 7, 1–5, https://doi.org/10.1038/ncomms12461, http://dx.doi.org/10.1038/ncomms12461, 2016.

Allgeier, J. E., Burkepile, D. E., and Layman, C. A.: Animal pee in the sea: consumer-mediated nutrient dynamics in the world's changing oceans, Global Change Biology, 23, 2166–2178, https://doi.org/10.1111/gcb.13625, 2017.

Allgeier, J. E., Wenger, S., and Layman, C. A.: Taxonomic identity best explains variation in body nutrient stoichiometry in a diverse marine animal community, Scientific Reports, 10, 1–10, https://doi.org/10.1038/s41598-020-67881-y, https://doi.org/10.1038/s41598-020-67881-y, 2020.

Atkinson, C. L., Capps, K. A., Rugenski, A. T., and Vanni, M. J.: Consumer-driven nutrient dynamics in freshwater ecosystems: from individuals to ecosystems, Biological Reviews, 92, 2003–2023, https://doi.org/10.1111/brv.12318, 2017.

Aumont, O., Maury, O., Lefort, S., and Bopp, L.: Evaluating the Potential Impacts of the Diurnal Vertical Migration by Marine Organisms on Marine Biogeochemistry, Global Biogeochemical Cycles, 32, 1622–1643, https://doi.org/10.1029/2018GB005886, 2018.

Baum, J. K. and Worm, B.: Cascading top-down effects of changing oceanic predator abundances, Journal of Animal Ecology, 78, 699–714, https://doi.org/10.1111/j.1365-2656.2009.01531.x, http://doi.wiley.com/10.1111/j.1365-2656.2009.01531.x, 2009.

Benitez-Nelson, C. R.: The biogeochemical cycling of phosphorus in marine systems, Earth Science Reviews, 51, 109–135, https://doi.org/10.1016/S0012-8252(00)00018-0, 2000.

Bianchi, D., Carozza, D. A., Galbraith, E. D., Guiet, J., and DeVries, T.: Estimating global biomass and biogeochemical cycling of marine fish with and without fishing, Science Advances, 7, https://doi.org/10.1126/sciadv.abd7554, 2021.

Brahney, J., Mahowald, N., Ward, D. S., Ballantyne, A. P., and Neff, J. C.: Is atmospheric phosphorus pollution altering global alpine Lake stoichiometry?, Global Biogeochemical Cycles, 29, 1369–1383, https://doi.org/10.1002/2015GB005137, https://onlinelibrary.wiley.com/doi/abs/10.1002/2015GB005137, 2015.

Carozza, D. A., Bianchi, D., and Galbraith, E. D.: The ecological module of BOATS-1.0: A bioenergetically constrained model of marine upper trophic levels suitable for studies of fisheries and ocean biogeochemistry, Geoscientific Model Development, 9, 1545–1565, https://doi.org/10.5194/gmd-9-1545-2016, 2016.

Carozza, D. A., Bianchi, D., and Galbraith, E. D.: Formulation, General Features and Global Calibration of a Bioenergetically-Constrained Fishery Model, PLOS ONE, 12, e0169 763, https://doi.org/10.1371/journal.pone.0169763, http://dx.plos.org/10.1371/journal.pone.0169763, 2017.

Cavan, E. L., Belcher, A., Atkinson, A., Hill, S. L., Kawaguchi, S., McCormack, S., Meyer, B., Nicol, S., Ratnarajah, L., Schmidt, K., Steinberg, D. K., Tarling, G. A., and Boyd, P. W.: The importance of Antarctic krill in biogeochemical cycles, Nature Communications, 10, 1–13, https://doi.org/10.1038/s41467-019-12668-7, http://dx.doi.org/10.1038/s41467-019-12668-7, 2019.

Czamanski, M., Nugraha, A., Pondaven, P., Lasbleiz, M., Masson, A., Caroff, N., Bellail, R., and Tréguer, P.: Carbon, nitrogen and phosphorus elemental stoichiometry in aquacultured and wild-caught fish and consequences for pelagic nutrient dynamics, Marine Biology, 158, 2847–2862, https://doi.org/10.1007/s00227-011-1783-7, 2011.

710   Davison, P. C., Checkley, D. M., Koslow, J. A., and Barlow, J.: Carbon export mediated by mesopelagic fishes in the northeast Pacific Ocean, Progress in Oceanography, 116, 14–30, https://doi.org/10.1016/j.pocean.2013.05.013, 2013.

Dunne, J. P., Sarmiento, J. L., and Gnanadesikan, A.: A synthesis of global particle export from the surface ocean and cycling through the ocean interior and on the seafloor, Global Biogeochemical Cycles, 21, n/a–n/a, https://doi.org/10.1029/2006GB002907, http://doi.wiley.com/10.1029/2006GB002907, 2007.

715   Dunne, J. P., John, J. G., Shevliakova, S., Stouffer, R. J., Krasting, J. P., Malyshev, S. L., Milly, P. C., Sentman, L. T., Adcroft, A. J., Cooke, W., Dunne, K. A., Griffies, S. M., Hallberg, R. W., Harrison, M. J., Levy, H., Wittenberg, A. T., Phillips, P. J., and Zadeh, N.: GFDL's ESM2 global coupled climate-carbon earth system models. Part II: Carbon system formulation and baseline simulation characteristics, Journal of Climate, 26, 2247–2267, https://doi.org/10.1175/JCLI-D-12-00150.1, 2013.

Dupont, L., Le Mézo, P., Clerc, C., Maury, O., Aumont, O., Ethé, C., and Bopp, L.: High trophic level feedbacks on ocean biogeochemistry

720   under climate change, submitted.

El-Sabaawi, R. W., Warbanski, M. L., Rudman, S. M., Hovel, R., and Matthews, B.: Investment in boney defensive traits alters organismal stoichiometry and excretion in fish, Oecologia, 181, 1209–1220, https://doi.org/10.1007/s00442-016-3599-0, 2016.

Fowler, D., Coyle, M., Skiba, U., Sutton, M., Cape, J. N., Reis, S., Sheppard, L., Jenkins, A., Grizetti, B., Galloway, J. N., Vitousek, P., Leach, A., Bouwman, L., Butterbach-Bahl, K., Dentener, F., Stevenson, D., Amann, M., and Voss, M.: The global nitrogen cycle in the

725   21th century, Philisophical Transactions of the Royal Society of London, B Biological Sciences, 368, 20130 165, 2013.

Francis, F. T. and Côté, I. M.: Fish movement drives spatial and temporal patterns of nutrient provisioning on coral reef patches, Ecosphere, 9, https://doi.org/10.1002/ecs2.2225, 2018.

Frank, K. T., Petrie, B., Choi, J. S., and Leggett, W. C.: Trophic cascades in a formerly cod-dominated ecosystem., Science (New York, N.Y.), 308, 1621–3, https://doi.org/10.1126/science.1113075, http://www.ncbi.nlm.nih.gov/pubmed/15947186, 2005.

730   Galbraith, E. D. and Martiny, A. C.: A simple nutrient-dependence mechanism for predicting the stoichiometry of marine ecosystems, Proceedings of the National Academy of Sciences, 112, https://doi.org/10.1073/pnas.1423917112, 2015.

Galbraith, E. D., Carozza, D. A., and Bianchi, D.: A coupled human-Earth model perspective on long-term trends in the global marine fishery, Nature Communications, 8, 1–7, https://doi.org/10.1038/ncomms14884, http://dx.doi.org/10.1038/ncomms14884, 2017.

Galbraith, E. D., Le Mézo, P. K., Solanes Hernandez, G., Bianchi, D., and Kroodsma, D.: Growth limitation of marine fish by low iron

735   availability in the open ocean, Frontiers in Marine Science, 6, 1–13, https://doi.org/10.3389/fmars.2019.00509, https://www.frontiersin.org/article/10.3389/fmars.2019.00509/full, 2019.

Gordon, H. S.: The Economic Theory of a Common-Property Resource: The Fishery, Education + Training, 62, 124–142, 1954.

Gresh, T., Lichatowich, J., and Schoonmaker, P.: An Estimation of Historic and Current Levels of Salmon Production in the Northeast Pacific Ecosystem: Evidence of a Nutrient Deficit in the Freshwater Systems of the Pacific Northwest, Fisheries, 25,

740   15–21, https://doi.org/10.1577/1548-8446(2000)025<0015:AEOHAC>2.0.CO;2, http://doi.wiley.com/10.1577/1548-8446(2000)025%3C0015:AEOHAC%3E2.0.CO;2, 2000.

Griffiths, D.: The direct contribution of fish to lake phosphorus cycles, Ecology of Freshwater Fish, 15, 86–95, https://doi.org/10.1111/j.1600-0633.2006.00125.x, 2006.

Gruber, N. and Galloway, J. N.: An Earth-system perspective of the global nitrogen cycle, Nature, 451, 293–296, https://doi.org/10.1038/nature06592, 2008.

Guiet, J., Galbraith, E., Bianchi, D., and Cheung, W.: Bioenergetic influence on the historical development and decline of industrial fisheries, ICES Journal of Marine Science, 77, 1854–1863, 2020.

Hall, R. O. J., Koch, B. J., Marshall, M. C., Taylor, B. W., How, L. M. T., Koch, B. J., Marshall, M. C., and Taylor, B. W.: How body size mediates the role of animals in nutrient cycling in aquatic ecosystems, The structure and function of aquatic ecosystems, pp. 286–305, https://doi.org/http://dx.doi.org/10.1017/CBO9780511611223, 2007.

Halvorson, H. M. and Small, G. E.: Observational field studies are not appropriate tests of consumer stoichiometric homeostasis, Freshwater Science, 35, 1103–1116, https://doi.org/10.1086/689212, https://www.journals.uchicago.edu/doi/10.1086/689212, 2016.

Hatton, I. A., Heneghan, R. F., Bar-On, Y. M., and Galbraith, E. D.: The global ocean size spectrum from bacteria to whales, Science Advances, 7, 1–9, https://doi.org/10.1126/sciadv.abh3732, 2021.

Hernández-León, S., Fraga, C., and Ikeda, T.: A global estimation of mesozooplankton ammonium excretion in the open ocean, Journal of Plankton Research, 30, 577–585, https://doi.org/10.1093/plankt/fbn021, 2008.

Hicks, C. C., Cohen, P. J., Graham, N. A. J., Nash, K. L., Allison, E. H., D'Lima, C., Mills, D. J., Roscher, M., Thilsted, S. H., Thorne-Lyman, A. L., and MacNeil, M. A.: Harnessing global fisheries to tackle micronutrient deficiencies, Nature, 574, 95–98, https://doi.org/10.1038/s41586-019-1592-6, 2019.

Hjerne, O. and Hansson, S.: The role fish and fisheries in the Baltic sea nutrient dyanmics, Limnology and Oceanography, 47, 1023–1032, 2002.

Huang, Y., Ciais, P., Goll, D. S., Sardans, J., Peñuelas, J., Cresto-Aleina, F., and Zhang, H.: The shift of phosphorus transfers in global fisheries and aquaculture, Nature Communications, 11, 1–10, https://doi.org/10.1038/s41467-019-14242-7, http://dx.doi.org/10.1038/s41467-019-14242-7, 2020.

Ito, A.: Atmospheric Processing of Combustion Aerosols as a Source of Bioavailable Iron, Environmental Science & Technology Letters, 2, 70–75, https://doi.org/10.1021/acs.estlett.5b00007, https://pubs.acs.org/doi/10.1021/acs.estlett.5b00007, 2015.

Jennings, S., Mélin, F., Blanchard, J. L., Forster, R. M., Dulvy, N. K., Wilson, R. W., Jennings, S., Dulvy, N. K., Wilson, R. W., Melin, F., Blanchard, J. L., Forster, R. M., Dulvy, N. K., Wilson, R. W., Mélin, F., Blanchard, J. L., Forster, R. M., Dulvy, N. K., and Wilson, R. W.: Global-scale predictions of community and ecosystem properties from simple ecological theory, Proceedings of the Royal Society B: Biological Sciences, 275, 1375–1383, https://doi.org/10.1098/rspb.2008.0192, 2008.

Kanakidou, M., Duce, R. A., Prospero, J. M., Baker, A. R., Benitez-Nelson, C., Dentener, F. J., Hunter, K. A., Liss, P. S., Mahowald, N., Okin, G. S., Sarin, M., Tsigaridis, K., Uematsu, M., Zamora, L. M., and Zhu, T.: Atmospheric fluxes of organic N and P to the global ocean, Global Biogeochemical Cycles, 26, 1–12, https://doi.org/10.1029/2011GB004277, 2012.

Kavanagh, L. and Galbraith, E.: Links between fish abundance and ocean biogeochemistry as recorded in marine sediments, PLoS ONE, 13, 1–22, https://doi.org/10.1371/journal.pone.0199420, 2018.

Layman, C. A., Allgeier, J. E., Rosemond, A. D., Dahlgren, C. P., and Yeager, L. A.: Marine fisheries declines viewed upside down: Human impacts on consumer-driven nutrient recycling, Ecological Applications, 21, 343–349, https://doi.org/10.1890/10-1339.1, 2011.

Le Mézo, P. K. and Galbraith, E. D.: The fecal iron pump : impact of animals on the Fe stoichiometry of marine sinking particles, Limnology and Oceanography, pp. 1–39, 2020.

Lefort, S., Aumont, O., Bopp, L., Arsouze, T., Gehlen, M., and Maury, O.: Spatial and body-size dependent response of marine pelagic communities to projected global climate change, Global Change Biology, 21, 154–164, https://doi.org/10.1111/gcb.12679, 2015.

Leroux, S. J. and Schmitz, O. J.: Predator-driven elemental cycling: the impact of predation and risk effects on ecosystem stoichiometry, Ecology and Evolution, 5, 4976–4988, https://doi.org/10.1002/ece3.1760, http://doi.wiley.com/10.1002/ece3.1760, 2015.

Locarnini, R. A., Mishonov, A. V., Antonov, J. I., Boyer, T. P., Garcia, H. E., Baranova, O. K., Zweng, M. M., and Johnson, D. R.: World Ocean Atlas 2009, vol. 1, Temperature, NOAA Atlas NESDIS, 68, 2010.

Lotze, H. K., Tittensor, D. P., Bryndum-Buchholz, A., Eddy, T. D., Cheung, W. W., Galbraith, E. D., Barange, M., Barrier, N., Bianchi, D., Blanchard, J. L., Bopp, L., Büchner, M., Bulman, C. M., Carozza, D. A., Christensen, V., Coll, M., Dunne, J. P., Fulton, E. A., Jennings, S., Jones, M. C., Mackinson, S., Maury, O., Niiranen, S., Oliveros-Ramos, R., Roy, T., Fernandes, J. A., Schewe, J., Shin, Y. J., Silva, T. A., Steenbeek, J., Stock, C. A., Verley, P., Volkholz, J., Walker, N. D., and Worm, B.: Global ensemble projections reveal trophic amplification of ocean biomass declines with climate change, Proceedings of the National Academy of Sciences of the United States of America, 116, 12 907–12 912, https://doi.org/10.1073/pnas.1900194116, 2019.

Mahowald, N. M., Engelstaedter, S., Luo, C., Sealy, A., Artaxo, P., Benitez-Nelson, C., Bonnet, S., Chen, Y., Chuang, P. Y., Cohen, D. D., Dulac, F., Herut, B., Johansen, A. M., Kubilay, N., Losno, R., Maenhaut, W., Paytan, A., Prospero, J. M., Shank, L. M., and Siefert, R. L.: Atmospheric Iron Deposition: Global Distribution, Variability, and Human Perturbations, Annual Review of Marine Science, 1, 245–278, https://doi.org/10.1146/annurev.marine.010908.163727, 2009.

Maldonado, M. T., Surma, S., and Pakhomov, E. A.: Southern Ocean biological iron cycling in the pre-whaling and present ecosystems, Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences, 374, https://doi.org/10.1098/rsta.2015.0292, 2016.

Maranger, R., Caraco, N., Duhamel, J., and Amyot, M.: Nitrogen transfer from sea to land via commercial fisheries, Nature Geoscience, 1, 111–113, https://doi.org/10.1038/ngeo108, http://www.nature.com/articles/ngeo108, 2008.

McIntyre, P. B., Flecker, A. S., Vanni, M. J., Hood, J. M., Taylor, B. W., and Thomas, S. A.: Fish distributions and nutrient recycling in streams: can fish create biogeochemical hotspots?, Ecology, 89, 2335–2346, 2008.

MODIS-Aqua, M.: NASA goddard space flight center, ocean ecology laboratory, ocean biology processing group, Moderate-resolution Imaging Spectroradiometer (MODIS) Aqua Euphotic Depth Data; 2018 Reprocessing. NASA OB. DAAC, Greenbelt, MD, USA, https://doi.org/data/10.5067/AQUA/MODIS/L3M/ZLEE/2018, 2018.

Moody, E. K., Corman, J. R., Elser, J. J., and Sabo, J. L.: Diet composition affects the rate and N: P ratio of fish excretion, Freshwater Biology, 60, 456–465, https://doi.org/10.1111/fwb.12500, 2015.

Moore, C. M., Mills, M. M., Arrigo, K. R., Berman-Frank, I., Bopp, L., Boyd, P. W., Galbraith, E. D., Geider, R. J., Guieu, C., Jaccard, S. L., Jickells, T. D., La Roche, J., Lenton, T. M., Mahowald, N. M., Marañón, E., Marinov, I., Moore, J. K., Nakatsuka, T., Oschlies, A., Saito, M. A., Thingstad, T. F., Tsuda, A., and Ulloa, O.: Processes and patterns of oceanic nutrient limitation, Nature Geoscience, 6, 701–710, https://doi.org/10.1038/ngeo1765, http://www.nature.com/doifinder/10.1038/ngeo1765, 2013.

Moreno, A. R. and Haffa, A. L. M.: The Impact of Fish and the Commercial Marine Harvest on the Ocean Iron Cycle, PLoS ONE, 9, e107 690, https://doi.org/10.1371/journal.pone.0107690, https://dx.plos.org/10.1371/journal.pone.0107690, 2014.

Myriokefalitakis, S., Nenes, A., Baker, A. R., Mihalopoulos, N., and Kanakidou, M.: Bioavailable atmospheric phosphorous supply to the global ocean: A 3-D global modeling study, Biogeosciences, 13, 6519–6543, https://doi.org/10.5194/bg-13-6519-2016, 2016.

Nugraha, A., Pondaven, P., and Tréguer, P.: Influence of consumer-driven nutrient recycling on primary production and the distribution of N and P in the ocean, Biogeosciences, 7, 1285–1305, https://doi.org/10.5194/bg-7-1285-2010, 2010.

Okin, G. S., Baker, A. R., Tegen, I., Mahowald, N. M., Dentener, F. J., Duce, R. A., Galloway, J. N., Hunter, K., Kanakidou, M., Kubilay, N., Prospero, J. M., Sarin, M., Surapipith, V., Uematsu, M., and Zhu, T.: Impacts of atmospheric nutrient deposition on marine productivity:
820   Roles of nitrogen, phosphorus, and iron, Global Biogeochemical Cycles, 25, 1–10, https://doi.org/10.1029/2010GB003858, 2011.

Pauly, D. and Tsukayama, I.: On the seasonal growth, monthly recruitment and monthly biomass of Peruvian anchoveta (Engraulis ringens) from 1961 to 1979., 1984.

Pauly, D. and Zeller, D.: Catch reconstructions reveal that global marine fisheries catches are higher than reported and declining, Nature Communications, 7, 10 244, https://doi.org/10.1038/ncomms10244, http://www.nature.com/articles/ncomms10244, 2016.

825 Pilati, A. and Vanni, M. J.: Ontogeny, diet shifts, and nutrient stoichiometry in fish, Oikos, 116, 1663–1674, https://doi.org/10.1111/j.0030-1299.2007.15970.x, http://doi.wiley.com/10.1111/j.0030-1299.2007.15970.x, 2007.

Prabhu, A. J. P., Schrama, J. W., and Kaushik, S. J.: Mineral requirements of fish: A systematic review, Reviews in Aquaculture, 8, 172–219, https://doi.org/10.1111/raq.12090, 2016.

Ricard, D., Minto, C., Jensen, O. P., and Baum, J. K.: Examining the knowledge base and status of commercially exploited marine species
830   with the RAM Legacy Stock Assessment Database, Fish and fisheries, 13, 380–398, 2012.

Roman, J., Estes, J. A., Morissette, L., Smith, C., Costa, D., McCarthy, J., Nation, J. B., Nicol, S., Pershing, A., and Smetacek, V.: Whales as marine ecosystem engineers, Frontiers in Ecology and the Environment, 12, 377–385, https://doi.org/10.1890/130220, 2014.

Saba, G. K. and Steinberg, D. K.: Abundance, composition, and sinking rates of fish fecal pellets in the santa barbara channel, Scientific Reports, 2, 1–6, https://doi.org/10.1038/srep00716, 2012.

835 Saba, G. K., Burd, A. B., Dunne, J. P., Hernández-León, S., Martin, A. H., Rose, K. A., Salisbury, J., Steinberg, D. K., Trueman, C. N., Wilson, R. W., and Wilson, S. E.: Toward a better understanding of fish-based contribution to ocean carbon flux, Limnol. Oceanogr, 9999, 1–26, https://doi.org/10.1002/lno.11709, https://aslopubs.onlinelibrary.wiley.com/doi/full/10.1002/lno.11709%0Ahttps://aslopubs.onlinelibrary.wiley.com/doi/abs/10.1002/lno.11709%0Ahttps://aslopubs.onlinelibrary.wiley.com/doi/10.1002/lno.11709, 2021.

Sarmiento, J. L. and Gruber, N.: Ocean biogeochemical dynamics, Princeton University Press, 2006.

840 Schaefer, M. B.: Some aspects of the dynamics of populations important to the management of the commercial marine fisheries, https://doi.org/10.1007/BF02464432, 1954.

Schindler, D. E. and Eby, L. A.: Stoichiometry of Fishes and Their Prey : Implications for Nutrient Recycling, Ecology, 78, 1816–1831, 1997.

Schmitz, O. J., Hawlena, D., and Trussell, G. C.: Predator control of ecosystem nutrient dynamics, Ecology Letters, 13, 1199–1209,
845   https://doi.org/10.1111/j.1461-0248.2010.01511.x, http://doi.wiley.com/10.1111/j.1461-0248.2010.01511.x, 2010.

Shearer, K. D.: Changes in Elemental Composition of Hatchery-Reared Rainbow Trout, Salmo gairdneri , Associated with Growth and Reproduction, Canadian Journal of Fisheries and Aquatic Sciences, 41, 1592–1600, https://doi.org/10.1139/f84-197, 1984.

Shearer, K. D., ÅSgård, T., Andorsdöttir, G., and Aas, G. H.: Whole body elemental and proximate composition of Atlantic salmon (Salmo salar) during the life cycle, Journal of Fish Biology, 44, 785–797, https://doi.org/10.1111/j.1095-8649.1994.tb01255.x, 1994.

850 Spalding, M. D. and Grenfell, A. M.: New estimates of global and regional coral reef areas, Coral Reefs, 16, 225–230, https://doi.org/10.1007/s003380050078, http://link.springer.com/10.1007/s003380050078, 1997.

Sterner, R. W. and Elser, J. J.: Ecological stoichiometry: the biology of elements from molecules to the biosphere, Princeton university press, 2002.

Thodesen, J., Storebakken, T., Shearer, K. D., Rye, M., Bjerkeng, B., and Gjerde, B.: Genetic variation in mineral absorption of large Atlantic
855   salmon (Salmo salar) reared in seawater, Aquaculture, 194, 263–271, https://doi.org/10.1016/S0044-8486(00)00525-1, 2001.

Turner, J. T.: Zooplankton fecal pellets, marine snow, phytodetritus and the ocean's biological pump, Progress in Oceanography, 130, 205–248, https://doi.org/10.1016/j.pocean.2014.08.005, http://dx.doi.org/10.1016/j.pocean.2014.08.005http://linkinghub.elsevier.com/retrieve/pii/S0079661114001281, 2015.

Vanni, M. J.: Nutrient Cycling by Animals in Freshwater Ecosystems, Annual Review of Ecology and Systematics, 33, 341–370, https://doi.org/10.1146/annurev.ecolsys.33.010802.150519, http://www.annualreviews.org/doi/10.1146/annurev.ecolsys.33.010802.150519, 2002.

Vanni, M. J. and McIntyre, P. B.: Predicting nutrient excretion of aquatic animals with metabolic ecology and ecological stoichiometry: A global synthesis, Ecology, 97, 3460–3471, https://doi.org/10.1002/ecy.1582, 2016.

Vanni, M. J., Bowling, A. M., Dickman, E. M., Hale, R. S., Higgins, K. A., Horgan, M. J., Knoll, L. B., Renwick, W. H., and Stein, R. A.: Nutrient cycling by fish supports relatively more primary production as lake productivity increases, Ecology, 87, 1696–1709, https://doi.org/10.1890/0012-9658(2006)87[1696:NCBFSR]2.0.CO;2, 2006.

Vanni, M. J., Boros, G., and McIntyre, P. B.: When are fish sources versus sinks of nutrients in lake ecosystems?, Ecology, 94, 2195–2206, 2013.

Vanni, M. J., McIntyre, P. B., Allen, D., Arnott, D. L., Benstead, J. P., Berg, D. J., Brabrand, Å., Brosse, S., Bukaveckas, P. A., Caliman, A., Capps, K. A., Carneiro, L. S., Chadwick, N. E., Christian, A. D., Clarke, A., Conroy, J. D., Cross, W. F., Culver, D. A., Dalton, C. M., Devine, J. A., Domine, L. M., Evans-White, M. A., Faafeng, B. A., Flecker, A. S., Gido, K. B., Godinot, C., Guariento, R. D., Haertel-Borer, S., Hall, R. O., Henry, R., Herwig, B. R., Hicks, B. J., Higgins, K. A., Hood, J. M., Hopton, M. E., Ikeda, T., James, W. F., Jansen, H. M., Johnson, C. R., Koch, B. J., Lamberti, G. A., Lessard-Pilon, S., Maerz, J. C., Mather, M. E., McManamay, R. A., Milanovich, J. R., Morgan, D. K., Moslemi, J. M., Naddafi, R., Nilssen, J. P., Pagano, M., Pilati, A., Post, D. M., Roopin, M., Rugenski, A. T., Schaus, M. H., Shostell, J., Small, G. E., Solomon, C. T., Sterrett, S. C., Strand, Ø., Tarvainen, M., Taylor, J. M., Torres-Gerald, L. E., Turner, C. B., Urabe, J., Uye, S. I., Ventelä, A. M., Villeger, S., Whiles, M. R., Wilhelm, F. M., Wilson, H. F., Xenopoulos, M. A., and Zimmer, K. D.: A global database of nitrogen and phosphorus excretion rates of aquatic animals, https://doi.org/10.1002/ecy.1792, 2017.

Wang, R., Balkanski, Y., Boucher, O., Bopp, L., Chappell, A., Ciais, P., Hauglustaine, D., Peñuelas, J., and Tao, S.: Sources, transport and deposition of iron in the global atmosphere, Atmospheric Chemistry and Physics, 15, 6247–6270, https://doi.org/10.5194/acp-15-6247-2015, 2015.

Wilson, R. W., Millero, F. J., Taylor, J. R., Walsh, P. J., Christensen, V., Jennings, S., and Grosell, M.: Contribution of fish to the marine inorganic carbon cycle, Science, 323, 359–362, 2009.

Wotton, R. S. and Malmqvist, B.: Feces in Aquatic Ecosystems, BioScience, 51, 537–544, https://doi.org/10.1641/0006-3568(2001)051[0537:FIAE]2.0.CO;2, 2001.