Dynamics of Rare Earth Elements and associated major and trace elements during Douglas-fir (*Pseudotsuga menziesii*) and European beech (*Fagus sylvatica* L.) litter degradation.

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Abstract. Given the diverse physico-chemical properties of elements, we hypothesize that their incoherent distribution across the leaf tissues, combined with the distinct resistance to degradation that each tissue exhibits, leads to distinct turnover rates among elements. Moreover, litter layers of different ages produce diverse chemical signatures in solution during the wet degradation. To verify our hypothesis, Na, K, Mg, Mn, Ca, Pb, Al and Fe were analysed together with the Rare Earth Elements (REE) in the solid fractions and in the respective leachates of fresh leaves and different litter layers of two forested soils developed under *Pseudotsuga menziesii* and *Fagus sylvatica* L. trees. The results from the leaching experiment were also compared to the *in situ* REE composition of the soil solutions to clarify the impact that the litter degradation processes may have on soil solution chemical compositions.

Both tree species showed similar biogeochemical processes dominating the element dynamics during the degradation of the litter. REE, Al, Fe and Pb were preferentially retained in the solid litter material, in comparison to the other cations, and that their concentrations increased over time during the degradation. Accordingly, different litter fractions produced different yields of elements and REE patterns in the leachates, indicating that the tree species and the age of the litter play a role in the chemical release during the degradation. In particular, the evolution of the REE patterns, relatively to the age of the litter layers, allowed us to deliver new findings on REE fractionation and mobilization during litter decay. Into specific, the degradation of the litter was characterised by a decrease in the Y/Ho ratio and an increase in the La_N/Yb_N ratio. The relationship between these ratios delivered information on the litter species-specific resistance to degradation, with Douglas-fir litter material showing a lower resistance.

During the degradation of the litter of the two tree species, two main differences were highlighted with the help of the REE: i) in *Pseudotsuga menziesii* the Eu behaviour appeared to be linked to the Ca during leaves senescence and ii) species-specific release of organic acids during the litter degradation leads to a more pronounced MREE enrichment in the *Fagus sylvatica* leachates.

Finally, we showed the primary control effect that white fungi may have in the Ce enrichment of soil solutions, which appears to be associated with the dissolution and/or direct transport of Ce-enriched MnO_2 particles accumulated on the surface of the old litter due to the metabolic functioning of these microorganisms. Similar MREE and HREE enrichments were also found in the leachates and the soil solutions, probably due to the higher affinity of these elements for the organic acids, which represent the primary products of the organic matter degradation.

1 Introduction

Nutrient cycling is key to forest ecosystem sustainability and productivity, especially in sites characterized by low fertility or degraded soils. There are three types of nutrient cycles, which relate to geochemical, biochemical or biogeochemical processes (Morris, 2004). The geochemical cycle encompasses all processes inherent to the introduction or removal of nutrients – excluding any kind of biological activity (e.g., input from aerosols, leaching of nutrients from rocks and their removal from the system through runoff). The biochemical cycle refers to processes involved in the transport and retention of nutrients inside the trees (such as the withdrawal of specific nutrients from leaves before senescence). The biogeochemical cycle encompasses the processes that occur outside of the trees and lead to the degradation of the organic waste material (such as exudates of leaves and stems, dead leaves and branches or even a whole tree) into its primary components and therefore to the release

of nutrients in a form that is reusable by trees (Morris, 2004). Organic matter degradation, which represents part of the biogeochemical cycle, is a major contributor to the nutrient stock available to trees in forest ecosystems (Staaf, 1980; Guo and Sims, 1999; Chadwick *et al.*, 1999; J.M. Pacyna, 2008; M. P. Krishna and M. Mohan, 2017) and in this context, litter degradation is known to play a key role in the replenishment of the nutritional pools of forests (Tagliavini *et al*, 2007). Nutrient release from litter is possibly regulated by various biotic and abiotic factors. Temperature, abundance of precipitation, species of decomposers (including the microfauna and microorganism communities), litter composition and chemical structure of its components are all factors that regulate the degradation rate and therefore the recirculation of the elements in a forest (Krishna and Mohan, 2017).

Trees absorb many nutrients to supply metabolic demands for growth, the immune system and reproduction but at the same time unnecessary elements, such as toxic metals, can also be absorbed and "trapped" in specific tree's compartments (Gomez *et al.*, 2018). Indeed, once absorbed, the elements are distributed within different tissues depending on their metabolic role or on their affinities with various compounds (Shan *et al.*, 2003; Ding *et al.*, 2005; Brioschi *et al*, 2013; Page and Feller, 2015; Ming Yuan *et al.*, 2017). This distribution could play a key role in processes involved in element turnovers in forest ecosystems especially during litter degradation, which potentially leads to a preferential release into the environment of some elements rather than others, depending on the substances to which they are bound. Litter degradation, indeed, would preferably promote the release of elements from more labile fractions making them available for tree uptake during the first stages of the degradation (Swift *et al*., 1979), while pools of elements that are trapped inside the most refractory tissues would remain unavailable for longer time spans.

The study of the biogeochemical processes involved in the distribution of the different classes of elements (toxic and nutrients) among the various leaf's tissues during the growth period, and their fractionation during the degradation of the litter is of crucial importance for a better understanding (and forecast) of the dynamics of the aforementioned classes of elements in forest ecosystems. To investigate such processes, Rare Earth Elements (REE) are interesting candidates due to their recognized use as tracers of geochemical processes, existing knowledge of their partitioning in plant tissues and recent studies related to their ecotoxicology as emerging pollutants (Liang *et al*. 2008; Xiaofei Li *et al*., 2013; Kyung Taek Rim *et al*., 2013). Indeed, knowledge of the processes involved in REE dynamics during litter degradation is of importance to environmental and social matters due to the increase in their environmental concentrations linked to their extraction in mining areas and exploitation in modern technologies. REE are a group of elements composed of lanthanides (from ^{57}La to ^{71}Lu) and ^{39}Y. Apart from Y, the other REE are usually divided into light (LREE: La to Nd), middle (MREE: Sm to Tb) and heavy (HREE: Dy to Lu) according to their atomic weight. To highlight the geochemical behaviour characteristics of the REE, it is convenient to consider the normalized concentrations rather than their absolute concentrations as is usually the case for other chemical elements. From the trends of the normalized concentrations (or patterns), it is then possible to determine the REE serial behaviour. These patterns can exhibit so-called "anomalies", which represent an enrichment (positive anomalies) or depletion (negative anomalies) in certain elements of the series, or also a fractionation occurring among the three groups of elements that are intimately linked to the environmental conditions. The characteristics of the patterns allow us to establish biogeochemical processes occurring in the system.

110   REE have already proven to be among the best-suited tracers for investigating Critical Zone processes such as: the origin of solid and dissolved load transported by stream (Aubert *et al.*, 2001; Hissler *et al.*, 2015a); metal adsorption in organic matter (Schijf and Zoll, 2011) and in bacterial cell walls (Takahashi *et al.*, 2005 and 2010); characterization of water-rock interaction and regolith weathering processes (Bau, 1996; Aubert *et al.*, 2001; Stille *et al.*, 2006, Ma *et al.*, 2011, Hissler *et al*, 2015b; Jin *et al.*, 2017; Laveuf and Cornu, 2009; Moragues-

115   Quiroga *et al.*, 2017, Vázquez-Ortega *et al.*, 2015 and 2016); as an indicator for atmospheric dust composition in leaves (Censi *et al.*, 2017); or wastewater spillage in freshwaters (Merschel *et al.* 2015; Hissler *et al.*, 2016); and metal mobilization and fractionation in the soil-plant continuum (Liang *et al.*, 2005 and 2008; Censi *et al.*, 2014a; Cheng *et al.*, 2014;; Srmhi *et al.*, 2009).

        Despite the large number of REE studies on plant tissues (Fu *et al.*, 1998; Wyttenbach *et al.*, 1998; Wei

120   *et al.*, 2001; Han *et al.*, 2005; Ding *et al.*, 2005; Brioschi *et al.*, 2013; Censi *et al.*, 2014; Zaharescu *et al.*, 2017), their dynamics during litter degradation is still scarcely known, since research has mainly focused on changes in the concentrations of elements according to the total mass loss of litter material during the duration of the experiment (Tyler, 2004; Brun *et al.*, 2008; Gautam *et al.*, 2020). To the best of our knowledge, no studies regarding the processes involved in the regulation of such dynamics have been carried out. Our aim is to elucidate

125   which processes may control the release and retention of REE in relation to major cations and other trace elements during the wet litter degradation – with a secondary focus on the qualitative impact of litter degradation on the REE patterns of soil solutions. We believe that REE environmental behaviour and its capacity to accurately trace biogeochemical processes add value to our understanding of nutrient cycles and in particular, could more precisely inform us about the processes that control the litter degradation stages and the release of nutrients in

130   forest ecosystems. Since the different leaf tissues (across which leaves distribute the uptaken elements during the living period) exhibit distinct resistance to degradation, we hypothesize that during litter decay the combination of such a distribution process with the different levels of degradation resistance of the tissues will lead to distinct turnover rates among the elements. Consequently, this leads to a specific chemical release into the environment depending on the degradation stage of litter and the different tissues among which the elements

135   are distributed. To test our hypothesis, we designed a field experiment in the forested Weierbach Experimental Catchment (Hissler *et al.*, 2021), relying on a series of biogeochemical tracers – including concentrations of Na, Mg, K, Ca, Mn, Fe, Pb, Al and REE measured in fresh leaves and different litter fractions of Douglas-fir (*Pseudotsuga menziesii)* and European beech (*Fagus Sylvatica* L.) grown on the same soil. Moreover, we carried out leaching experiments on these samples with ultrapure water (MilliQ) in order to observe how the different

140   fractions of litter can contribute to element release and sequestration and how this release may affect soil solution chemistry. Finally, we compared the leaching experiment results to the chemical composition of soil solutions collected from the two tree stands. If our hypothesis is confirmed, we expect the older litter fractions to show an enrichment in specific elements, which would be linked to their distribution in the most refractory tissues, limiting their release into the environment during the degradation. Subsequently, we expect that the solutions

145   obtained through the leaching experiment (hereafter called "leachates") will exhibit different chemical signatures according to the degree of degradation of the samples. On the contrary, if different litter fractions showed similar chemical compositions and similar chemical release during the leaching experiment, it would imply that the distribution of the elements among the tissues is coherent and, therefore, the degradation stage of the litter does not affect the chemical release during the litter decay.

2 Materials and methods

2.1 Study site

We selected two experimental plots in the Weierbach Experimental Catchment located in the Luxembourg Ardennes Massif, which have been monitored for ecohydrological purposes since 2012 (Hissler *et al.*, 2021). The "Be" profile (BeP) shows a deciduous cover of European beech (*Fagus sylvatica L.*), while the "Do" profile (DoP) is covered with Douglas-fir (*Pseudotsuga menziesii*). The altitude ranges from 450 to 500 m a.s.l. and the geological substratum consists of Devonian metamorphic slates covered by 70 to 100 cm of Pleistocene Periglacial Slope Deposit (PPSD) composed of past loamy aeolian deposition (Moragues-Quiroga *et al.*, 2017). The soil, which is developing below a Hemimoder type of humus (Jabiol *et al.*, 2013) in the first 50 cm of the PPSD, presents homogenous properties all over the catchment. It is at an early formation stage and classified as dystric cambisol according to the World Reference Base for soil resources (Juilleret *et al*, 2016).

2.2 Sampling and preparation

The different plant materials (i.e. fresh leaves and needles, as well as new and old litter) were collected on the same sampling day in May 2019 at both beech and Douglas-fir stands.

Fresh leaves (beech) and needles (Douglas-fir) (hereafter referred to as FL) were collected from 10 adult trees randomly selected per plot. The leaves were taken from different branches, accessible from the ground, at various heights and radial directions. All leaves were aggregated together in one sample per plot and stored in clean polypropylene bags. The litter material was collected from five different locations within an area of 500 m^2 of each experimental plot using a 25x25cm metallic frame and avoiding contamination by soil particles. During the collection, the different fractions of litter were sorted according to their degradation degree (Fig. 1) and stored in polypropylene bags.

In BeP, three litter fractions were identified: the new litter (OLn - unprocessed, unfragmented, light-brownish coloured), the old litter (OLv - slightly altered, bleached and softened, discoloured or dark-brownish coloured) and the fragmented litter (OF - partially decomposed and fragmented, grey-black coloured). For DoP only two fractions stood out: the new litter (OLn - unprocessed, unfragmented, light-brownish coloured) and the old litter (OLv - slightly altered, bleached and softened, grey-black coloured), whereas the fragmented litter layer was not sufficiently developed and was not considered in this study.

Douglas-fir (*Pseudotsuga menziesii*) samples

Douglas-fir fresh leaves (**Do FL**)   Douglas-fir new litter (**Do OLn**)   Douglas-fir old litter (**Do OLv**)

European beech (*Fagus sylvatica L.*) samples

Beech fresh leaves (**Be FL**)   Beech new litter (**Be OLn**)   Beech old litter (**Be OLv**)   Beech fragmented litter (**Be OF**)

Figure 1: Fresh leaves and litter of European beech and Douglas-fir collected in the Weierbach Experimental Catchment and sorted by degradation degree.

The atmospheric dust was collected in the Weierbach Experimental Catchment using a modified polypropylene version of the passive SIGMA-2 collectors produced by the German Meteorological Service in Freiburg, Germany (VDI 3787, 2010) installed at 1.5m above the ground. The SIGMA-2 passive sampler allows the sampling of atmospheric particles above 2.5µm size (Grobéty *et al*., 2010). The atmospheric dust represents an integrated sample exposed to the atmospheric deposition from September 2018 to May 2019. The particles were collected and stored in an acid-cleaned Teflon vessel during the whole exposure period and stored in a clean desiccator in the laboratory until its preparation before the analysis. After precise weighing of the atmospheric dust collected (2.3 mg), the sample was totally digested using concentrate ultrapure HNO_3, HF and $HClO_4$ acid mixture. The acids were then evaporated, and the residue was dissolved in a solution of HNO_3 (1% in volume) and stored at 4°C before the analysis.

Fresh leaves and litter materials were cleaned with a strong air flux and with a brush avoiding the use of water in order not to disturb the chemical signature and the biotic communities existing on the samples' surfaces. The samples were then dried in the oven at 40 °C and homogenized. Two aliquots were taken to perform the total chemical composition and the leaching experiment. The leaves and litter aliquots for the total chemical composition were preliminarily reduced to a fine powder and incinerated in closed ceramic cups at 550 °C before mineralization in order to destroy organic matter and to pre-concentrate the trace elements. We opted for a more convenient digestion protocol for the organic-derived samples by using Aqua Regia in a microwave-assisted oven (Anton Paar Multiwave PRO), which allows using higher amount of sample (250mg) and reaching high temperature (212°C) and pressure (25bars) conditions. Although Aqua Regia is not a total digestion method, among the different method tested, it delivered the best results without any precipitates nor suspended particles

in the digestates not only at the moment of the digestion but also long time after. The samples were then stored at 4°C before the analysis.

For the leaching experiment, 2L high-density polyethylene bottles were filled with the aliquots of leaves and litter fractions. 1L of ultrapure water was then added. There were two reasons for putting as much material as possible into the bottles: first, part of the samples had to be above the water level in order to enhance the aerobic degradation; and second, we wanted to be sure that the release of elements during the experiment period was abundant enough to be detectable with the instrumentations. The bottles were left with the cup partially open in order to allow gas exchanges to encourage bacterial and fungi activity. Samples were agitated for 1 hour/day for 7 days in an automatic vertical agitator (GFL type 3040) set at minimum speed to enhance the leaching process from all samples while avoiding further fragmentation. After one week, the leachates were separated from the solid material with a nylon sieve, filtered at 0.2 µm and acidified using HNO_3 (1% in volume). 50 ml aliquot from each leachate was evaporated in Savillex PFA vessels placed on a hot plate allowing the precipitation of the content. The residue was then sequentially mineralized with HF, HCl and HNO_3. After evaporation, the residue was then dissolved in HNO_3 (1% in volume) and samples were stored at 4°C before the analysis. All the chemicals used for the different digestion procedures were of ultrapure quality. Aliquots were also taken for DOC and pH measurement.

Thanks to the bi-weekly monitoring in place at the Weierbach catchment since 2009 (Hissler *et al.*, 2021), we can rely on the chemical composition of soil solutions collected between 2012 and 2014 at the two sampling locations at 20, 40 and 60 cm depths. The sampling was performed using Teflon/quartz suction cups (SDEC, Reignac-sur-Indre, France) connected to acid-clean 2L-Nalgene flasks under a vacuum of 0.8 bar.

2.3 Sample analysis

The concentrations of major cations and trace elements in all samples were analysed via Inductively Coupled Plasma - Mass Spectrometry (Agilent 7900). The measured isotopes were chosen with no isobaric interferences, and the polyatomic interferences were minimized by using the collision cell in Helium mode. Calibration standards and Quality Controls (QCs) were prepared with certified solutions (Chem-lab, Belgium and Merck, Belgium). QCs at low, medium and high levels of concentration of the calibration range were analysed each after ten samples to control the validity of the measurements. Internal standards were prepared with Chem-lab (Belgium) Rhenium and Rhodium certified solutions. To ensure the quality of the mineralization procedure, mineralization blanks were prepared following the same steps as for the samples and analysed. The limit of quantification (LoQ) for the different analyzed elements are reported in Table SI-1.

DOC was measured via a Teledyne Tekmar® Torch Combustion Analyser and pH with SenTix® 940 WTW.

2.4 REE normalization and anomaly calculations

All data presented in the following sections were normalized to the local atmospheric deposition (Table SI-2). This allowed us to compare our samples to a local reference for REE. As atmospheric dust is an important input of cations and nutrients (Reynolds *et al.*, 2006; Lequy *et al.*, 2012) we also expected it to be important in terms of REE input in forest ecosystems. Moreover, with this normalization, we could directly differentiate the vegetation contribution to the REE patterns observed for the litter samples.

Gadolinium (Gd), Europium (Eu) and Cerium (Ce) anomalies were calculated from equations 1 to 3, respectively:

$$Gd/Gd* = Gd_N/(0.33 \times Sm_N + 0.67 \times Tb_N) \tag{1}$$

<div style="margin-left:0">270</div>

$$Eu/Eu* = Eu_N/(0.5 \times Gd_N + 0.5 \times Sm_N) \tag{2}$$

$$Ce/Ce* = Ce_N/(0.5 \times La_N + 0.5 \times Pr_N) \tag{3}$$

with La_N, Ce_N, Pr_N, Sm_N, Eu_N, Gd_N and Tb_N indicating the REE normalized concentrations.

3 Results

3.1 Chemical composition of the fresh leaves and litter bulk samples

Concentrations of REE in fresh leaves and litter are reported in Table SI-3. When normalized to the local atmospheric deposition (hereafter referred to as "dust"), the REE patterns of bulk fresh leaves and litter material show some similarities between the two tree species. The REE concentrations increase with the age of the litter, with the lowest concentration in fresh leaves and the highest in the most degraded litter layers (Fig. 2a-b and Table SI-3). Additionally, both tree species present a depletion in HREE according to LREE and MREE (Fig. 3a-b). Y appears to behave coherently for both tree species during the litter degradation, as illustrated by the evolution of the Y/Ho ratios. Indeed, the samples show a Y enrichment according to the dust (Y/Ho = 25.93) that is higher in the fresh leaves with Y/Ho ratios equal to 34.34 and 34.3 for Do FL and Be FL respectively, and decreases progressively with the age of the litter showing the lowest values in the oldest litter fractions (23.58 and 25.43 in Do OLv and Be OF, respectively).
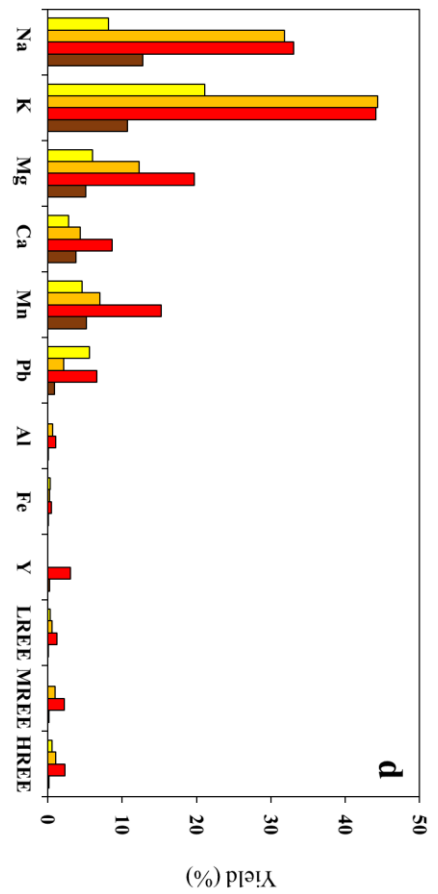
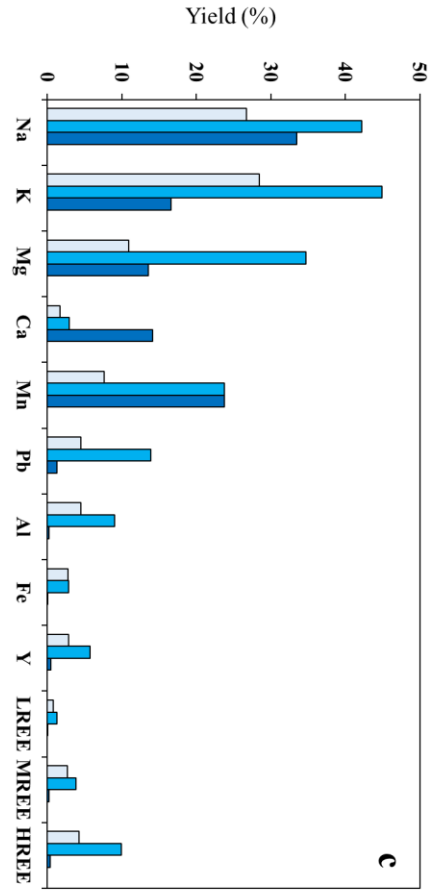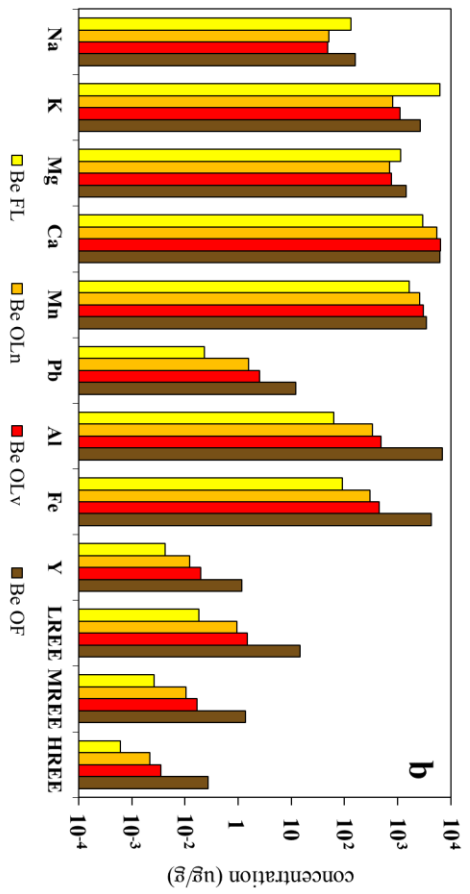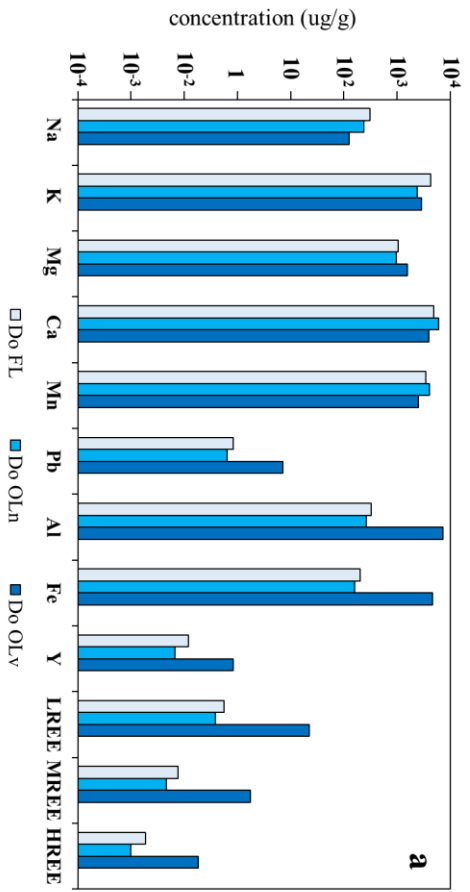**Figure 2:** Log$_{10}$ of the concentrations of the studied elements in fresh leaves and different litter fractions of (a) Douglas-fir and (b) European beech and element yields studied (% of the total mass) for the leaching experiment of (a) Douglas-fir and (b) European beech fresh leaves and litter samples.
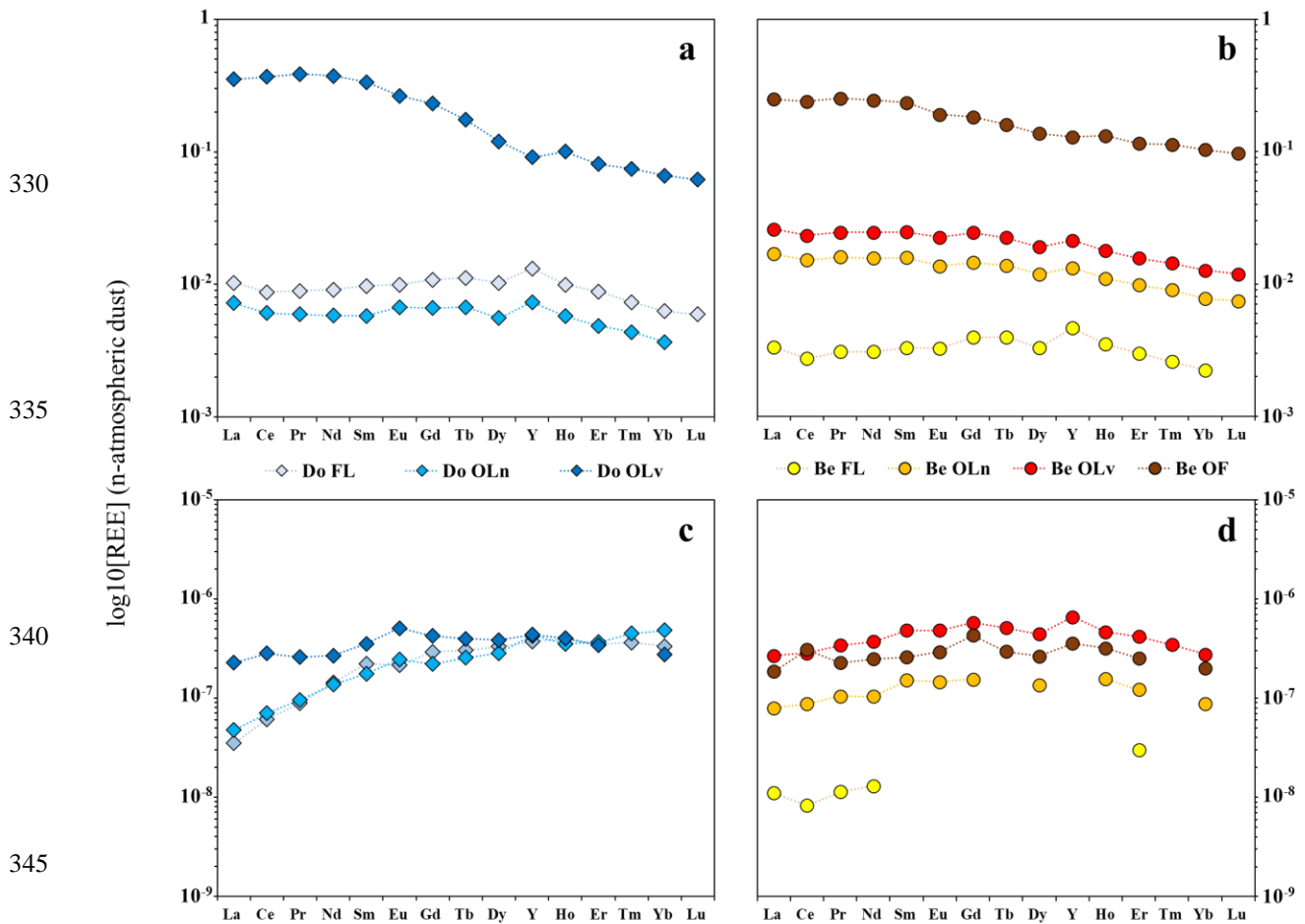
Figure 3: Patterns of Rare Earth Elements concentrations in samples of fresh leaves and litter of Douglas-fir (**a**) and beech (**b**); patterns of Rare Earth Elements concentrations in leachates of fresh leaves and litter of Douglas-fir (**c**) and beech (**d**). REE concentrations were normalized by the values in the local atmospheric dust.

In the bulk material, the REE concentration and the LREE enrichment also increase in line with the litter degradation stages but with differences between the Douglas-fir and the European beech. For the Douglas-fir samples, the REE total concentrations decrease from the fresh material to the Do OLn litter layer before drastically increasing at the Do OLv sample. The total REE concentrations are 0.78, 0.51 and 25.3 µg g^{-1} in Do FL, Do OLn and Do OLv, respectively. Here, the fractionations between the REE groups are not visible between the fresh leaves and the Do OLn litter layer. However, the fractionation between LREE, MREE and HREE in Do OLv are the highest for all the samples considered in this study, having La$_N$/Yb$_N$ and La$_N$/Gd$_N$ and Gd$_N$/Yb$_N$ ratios of 5.35 and 1.53 and 3.5, respectively. For the European beech, the REE total concentration in the fresh leaves is the lowest for all leaves and litter samples (Fig. 3b), and increases progressively at each degradation stage to reach its maximum value of 17.7 µg g^{-1} for the Be OF litter layer. The LREE enrichment increases during the degradation as illustrated by the La$_N$/Yb$_N$ ratios, which progressively evolve from 1.49 in the fresh leaves to 2.40 in the Be OF. The degradation of the beech litter also leads to an MREE enrichment as shown by the La$_N$/Gd$_N$ ratio evolving from 0.84 in Be FL to 1.16 in Be OLn and finally 1.36 in the Be OF.

The major elements and metals can be sorted into three different groups according to the evolution of their concentrations in the bulk samples of the different litter layers (Fig. 2a-b). This classification stays coherent for the two tree species. Na, K and Mg have their highest concentrations in the fresh leaves and the older litter

layers. Ca and Mn present a progressive increase of their concentration for the European beech, whereas they are less concentrated in the oldest litter layer of Douglas-fir in comparison to Do FL and Do OLn. The concentrations of the other trivalent metals (Fe, Al) evolve similarly to the REE with a progressive increase from the fresh leaves to the OL litters and an enrichment in the oldest litter layer for both species (Do OLV and Be OF). In contrast to the European beech, Douglas-fir fresh leaves present metals concentrations as high as in the Do OLn litter sample.

3.2 Chemical composition, pH and DOC content of leachates

The leaching experiment led to similar REE concentration ranges between the two tree species, except for the beech fresh leaves, which are one order of magnitude less concentrated (Table SI-4). The leachate patterns present strong differences to the REE characteristics of the bulk leaves and litter material and in between the two tree species.

The total REE concentrations of the Douglas-fir leachates are similar in Do FL and Do OLn samples and higher in the leachate of the Do OLv sample. As the HREE show very similar concentrations in the three leachates, the difference is mainly related to the concentration of LREE, as shown by the dust-normalized REE patterns (Fig. 3-c). Indeed, LREE are similarly depleted in the leachates of the two younger samples, showing La_N/Yb_N ratios of 0.10, while Do OLv presents a La_N/Yb_N ratio equal to 0.82. Noticeable are the Eu positive anomalies (Eu/Eu*) of 1.23 and 1.31 observed in the leachates of Do OLn and Do OLv, respectively. In Do OLv a slight positive Ce anomaly (Ce/Ce*) of 1.16 is also observed.

The REE concentrations in the leachates of beech samples increase from the fresh leaves to the highest stages of litter degradation but are higher for the Be OLv material. The dust-normalized REE patterns of beech leachates have an MREE enrichment in comparison to LREE and HREE where Gd shows the highest peak. The patterns of Be OLn and Be OLv leachates present very similar characteristics as indicated by their La_N/Gd_N ratios (0.46 and 0.43, respectively), Gd_N/Yb_N ratios (2.10 and 2.14, respectively) and the absence of any anomaly, whereas the Be OF leachate presents Ce and Gd positive anomalies (Ce/Ce*=1.49 and Gd/Gd*=1.52). In contrast, the leachate of the fresh beech leaves presents a Ce negative anomaly (Ce/Ce*=0.74).

Similar trends in the percentage of elements leached from the material of both tree species were observed. The percentage of leaching of the studied elements can be classified according to their valence Na, K > Mg, Mn > Ca, Pb > Al, Fe, REE, with the trivalent elements being less leached. However, some differences can be highlighted between Douglas-fir and beech.

The Douglas-fir material released a higher quantity of Na, Mg, Mn, Pb, Al, Fe, MREE and HREE compared to the beech. Major elements and Mn are preferentially released compared to trivalent metals and Pb, as shown in Figure 2c. The highest percentage of element release is observed from the Do OLn sample with Mn having similar values in Do OLn-L and Do OLv-L. An exception is made for Ca, which presents similar release percentages as those of the trivalent metals during the first stages of degradation and which shows the highest release from the Do OLv fraction.

In beech leachates, the elements show similar release trends as for Douglas-fir but the highest percentages of release for all elements are shown in Be OLv (Figure 2d) with Na and K having analogous values in Be OLn. In Be FL and Be OLn leachates, all the elements show lower release values than in Douglas-fir samples. Trivalent metals and Pb, similarly to those of Douglas-fir, show a low release from the solid material during the experiment

(in the case of Al, the release from fresh leaves is below the limit of quantification, as well as for many REE as shown in Table SI-4 and SI-5).

The Y/Ho ratios of the leachates of Douglas-fir litter can explain the appearance of a small Y depletion in the oldest litter sample. The leachate of Do OLn, which represents the stage of degradation that brings about the formation of the OLv fraction, indeed shows a Y/Ho ratio equal to 31.29 indicating a preferential release of Y (when compared to the neighbour) that leads to a lower-than-atmospheric dust value in the Do OLv sample (Y/Ho = 23.58 in Do OLv and Y/Ho = 25.93 in atmospheric dust).

In beech samples, we can observe a similar behaviour with values that are slightly higher. The Y/Ho ratio in the Be OLv leachate (Y/Ho = 36.59) justifies the absence of Y enrichment in the Be OF fraction, which instead shows a dust-like ratio (Y/Ho = 25.43).

The highest DOC concentrations (Table SI-4) were measured in the Douglas-fir leachates with values ranging from 10.39 mg L^{-1} in Do OLv to 29.37 mg L^{-1} in Do OLn, while beech leachates showed concentrations from 5.91 mg L^{-1} in Be OLn to 14.68 mg L^{-1} in Be OLv. Note that for both species, the highest DOC concentrations were measured in leachates of the second-to-last degradation stages with a decrease in the oldest fractions.

The pH appears to be inversely proportional to the DOC concentrations. Indeed, for the Douglas-fir leachates, the most acidic pH was found in Do OLn, which showed a pH = 4.26 (Table SI-4), while the pH of Do OLv was the highest with a value equal to 5.03. In beech leachates, the lowest pH was measured in Be OLv (pH = 4.07) and the highest in Be OLn (pH = 5.39).

3.3 Average REE in soil solutions

The average REE concentrations in soil solutions collected between 2012 and 2014 are reported in Table SI-6. The REE total concentrations differed by one order of magnitude and were lower under the Douglas-fir stand at 20 and 40 cm depth (ΣREE=0.88 μg L^{-1} and 0.92 μg L^{-1} in Do SS20 and Do SS40, respectively), whereas the highest concentration was observed in beech samples at 40 cm depth (ΣREE=6.70 μg L^{-1} in Be SS40).

The dust-normalized REE patterns show an MREE enrichment for all soil solutions and Ce positive anomalies at 20 and 40 cm depth. Among the MREE, Eu shows a peak in the soil solutions under the Douglas-fir, whereas Gd is more enriched in beech soil solutions. Do SS at 20 and 40 cm depth exhibit positive Ce anomalies (Ce/Ce* = 1.14 and 1.21, respectively) that disappear at 60 cm. Moreover, Douglas-fir samples also show an LREE depletion as indicated by the La_N/Yb_N ratios, which range from 0.50 to 0.66 in Do SS40 and Do SS60, respectively.

Under the beech, the Ce positive anomaly is higher at 20 cm and decreases with depth until it becomes negative in soil solutions at 60 cm depth (Ce/Ce* = 1.39, 1.20 and 0.82 in Be SS20, Be SS40 and Be SS60, respectively), whereas the LREE show a consistent depletion at 20 cm depth (La_N/Yb_N = 0.56) and a slight one at 60 cm depth (La_N/Yb_N = 0.85).

4 Discussion

4.1 REE fractionation during litter degradation: similarities and differences with the other elements

The leaching yields clearly illustrate that during the leaching experiment, REE are preferentially leached following the HREE>MREE>LREE order (Fig. 2c-d). This justifies the decreasing trends (from La to Lu)

observed in the REE patterns of litter fractions for both tree species and demonstrates the tendency of the litter to retain LREE rather than the other elements of the lanthanide series. This progressive LREE enrichment in the litter (Fig. 3a-b) proceeding towards degradation is more evident for the Douglas-fir samples as suggested by the La_N/Yb_N ratios in the Do OLv and Be OF samples (La_N/Yb_N=5.35 and La_N/Yb_N=2.40, respectively). The leaching experiment also showed that REE, together with the other trivalent metals (Fe and Al) and Pb, are preferentially held inside the solid material, while the other major elements and Mn are more easily released during litter degradation. This behaviour suggests a preferential fractionation of the studied nutrients (Na, K, Mg and Mn) into more labile tissues, probably due to their role in the metabolic functioning of the tissues of living leaves (Alejandro *et al.*, 2020; Sardans and Peñuelas, 2021; Shaul, 2002; Maathuis, 2013). This partitioning results in a concentration decrease of these elements during litter degradation, while Fe, Al, Pb and REE tend to remain embedded inside the most refractory tissues - increasing their concentration over time in the remnant litter material. In our experiment, all elements behave coherently according to their oxidation number (Fig. 2c-d). Monovalent elements (Na, K) are more likely to be released than the divalent elements (Mg, Mn), while trivalent metals (Al, Fe, REE) tend to stay more tightly bound to the solid residual fraction. Exceptions can be seen for Ca in the Douglas-fir samples and for Pb in the oldest litter fractions of both tree species.

Given that REE are preferentially retained in the recalcitrant tissues together with the other trivalent metals and Pb, the higher yields for these elements obtained from Douglas-fir samples during the leaching experiment might suggest a faster degradation process for the Douglas-fir litter when compared to the beech litter. The fact that LREE are preferentially retained in the litter of both tree species can be explained by referring to existing literature and by normalising the REE concentrations in the leachates to the ones of the respective litter material (Fig. 4).
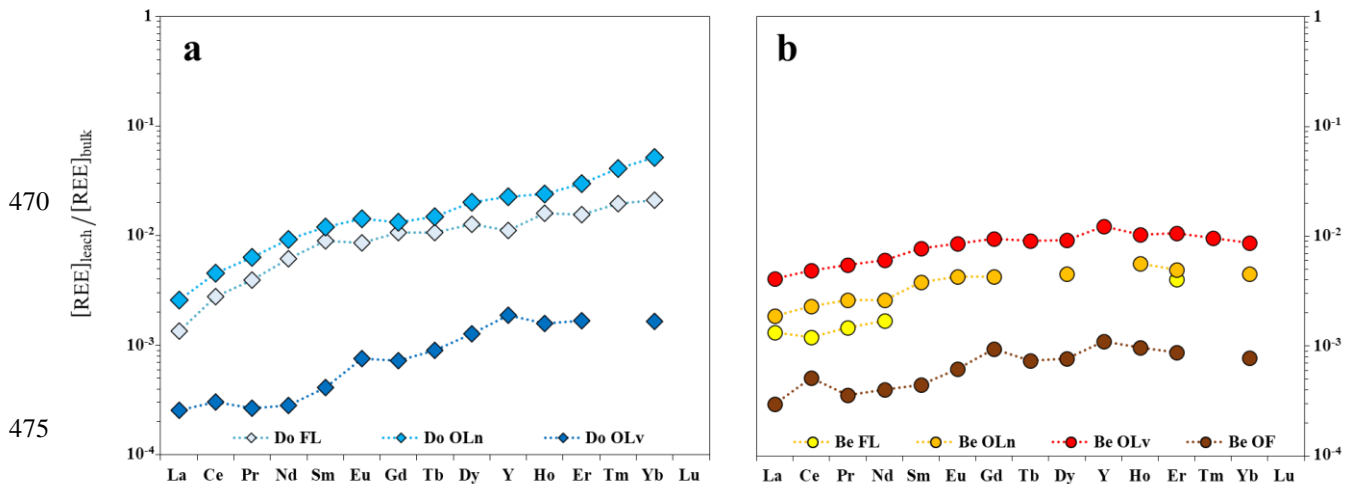
Figure 4. Patterns of the REE in leachates normalized to the REE concentrations in leaves and litter for Douglas-fir (**a**) and Beech (**b**) samples.

The patterns of REE in Douglas-fir samples show an HREE enrichment when compared to the other elements of the series ($0.05 \leq La_N/Yb_N \leq 0.15$ and $0.25 \leq Gd_N/Yb_N \leq 0.51$), indicating a preferential release of the heavy REE to the solution. In the European beech samples, the patterns are smoother between MREE and HREE

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(0.94≤Gd_N/Yb_N≤1.21), while they conserve the depletion in LREE when compared to the other groups
(0.32≤La_N/Gd_N≤0.44 and 0.38≤La_N/Yb_N≤0.48). In general, these patterns show a progressive increase from La to Lu that mirrors the trend of the stability constants of REE complexes with malate, EDTA, humic and fulvic acids (Suzuki *et al.*, 1980;; Pourret *et al.*, 2007; Marsac *et al.*, 2010; Sonke and Salters, 2005). Humic acids, fulvic acids and malic acid are indeed primary products of organic matter degradation (Adeleke *et al.*, 2017 and reference therein) and they have been reported to have metal chelating properties. The increasing release of REE from La to Lu in our leachates is likely due to an increase in the specific affinity - proceeding along the lanthanide series - of these elements towards the degradation products present in the aqueous solution (Schijf and Zoll, 2011). This suggests that the nephelauxetic effect (Juranic, 1988; Tchougréeff and Dronskowski, 2009) plays a key role in the mechanism of REE complexation with the dissolved organic ligands during the wet degradation of litter and is in line with the results shown by Sonke and Salters (2006), which experimentally demonstrated that the lanthanide contraction is responsible for a gradual increase in the complexation strength with humic substances when decreasing the ionic radius. The bond strength, indeed, increases with increasing ionic potential Z/r.

Differences in the atmospheric dust-normalized REE patterns of leachates between the two tree species can be explained by the nature of the ligands present in the solutions. Tang and Johannesson (2010) showed that the REE complexation with organic ligands in natural waters would produce patterns enriched in HREE when the majority of the ligands in solutions are represented by low molecular weight-dissolved organic compounds (such for instance citric, oxalic, malic, succinic, malonic and maleic acids) while a preponderant presence of heavy molecular weight-dissolved organic compounds (such as humic acid and fulvic acids) would produce an enrichment in MREE. Therefore, the results obtained indicate that the differences between the patterns of Douglas-fir and European beech leachates are likely due to the production of different classes of organic acids during the degradation of the samples.

Al, Fe, Pb and REE behaviours are coherent during litter degradation for both tree species, as their concentrations progressively increase towards the oldest litter fractions (Table SI-3 and Fig. 2a-b). We can explain such an accumulation in the oldest litter fractions by the binding with lignins. Lignins constitute the most degradation-resistant compounds in leaves. Their resilience lets these metals persist longer in the organic material than carbohydrates, lipids, proteins, hemicellulose and cellulose, which can degrade at a faster rate (Rahman *et al.*, 2013). Lignins are a class of organic polymers that have many functions in vascular plants. They provide structural support, improve cellular adhesion, enhance water transport and defence towards pathogens and are mainly situated in the cell walls of vascular and support tissues (Weng and Chapple, 2010; Leisola et al., 2012; Labeeuw *et al.,* 2015). Moreover, the chemical structure of these molecules exerts a strong control on litter decay rates (Talbot *et al.*, 2012). Increased lignin concentrations inhibit biological activity and linearly increase photo-degradation due to its wide spectrum of absorbance (Austin and Ballaré, 2010; Cogulet *et al.*, 2016).

While Fe, Al and Pb toxicity in plants is well-known (Imadi *et al.*, 2016; Bienfait, 1989; Woolhouse, 1983, Rout *et al.*, 2001; Sharma and Dubey, 2005; Pourrut *et al.*, 2011; Singh *et al.*, 2017; Miroslav Nikolic and Jelena Pavlovic, 2018), the toxicity of REE is not yet widely studied as their micropollutant nature has only recently emerged (Gwenzi *et al.*, 2018). However, the toxicity of REE in plants is far beyond the scope of this work, we limit ourselves to mentioning that researchers observed REE displaying redox-related toxicity mechanisms (Hassan Ragab El-Ramady, 2010; Pagano *et al.*, 2015; Tommasi *et al.*, 2021) and we assume that plants can trap REE in lignified tissues as a defence to avoid the toxicity-related events with the same mechanism

adopted for other potentially toxic metals, such as Pb and Al. Therefore, we propose that lignins constrain the REE

525   in the oldest litter fractions during the degradation of the leaves. Given the high affinity of such metals for oxygen, the absorption operated by lignins through the binding with the oxygen-bearing functional groups (such as phenolic, hydroxyl) may be the mechanism involved and would explain the accumulation of these metals in the oldest litter fractions. Therefore, during the living cycle of leaves, lignins are able to sequestrate the elements that show higher affinity for the exposed functional groups. As lignins are the most resistant tree components in forests,

530   they would prevent the release of the absorbed elements for longer during the litter degradation. The chemical elements that are more important for tree nutrition and metabolism would then be preferentially released to the soil solutions.

As shown by the evolution of the chemical composition of leaves and litter fractions along the different degradation stages, our hypothesis on the distribution of different elements among the different tissues is

535   confirmed. This finding is further corroborated by the result of the leaching experiment, which clearly confirms our hypothesis that during litter decay, the release of elements is linked to the degradation stage of the litter itself. As conjectured, elements partitioned in the most labile tissues are more easily released during the degradation process than those bound to more refractory tissues, which are instead accumulated over time.

According to our findings, two main REE fractionation processes are specific to a leaf's life-span:

540   (i) An inter-tissue fractionation occurring during the leave's "living period", through which recalcitrant tissues would preferentially absorb REE as a result of binding with lignins, developing a particular signature;

(ii) A degradation-driven fractionation, which has the different affinities of REE towards the products of the decay as the main factor for their partitioning between the remaining solid fraction of litter and the resulting solution.

4.2 Cerium anomalies in leachates

545   Another interesting aspect of our results is the presence of positive Ce anomalies in the leachates of the oldest litter fractions (Ce/Ce*=1.16 and Ce/Ce*=1.49 in Do OLv and Be OF leachates, respectively) and a slight W-type tetrad effect in the Be OF leachate. The tetrad effect can be defined as a graphical effect that divides the REE patterns into 4 segments so-called "tetrads" (T1 from La to Nd; T2 from Pm to Gd; T3 from Gd to Ho; T4 from Er to Lu), resulting from the increased stability at a quarter, half, three-quarter and complete filling of the 4f orbital

550   (McLennan, 1994). The tetrad effect is usually classified according to the shape of the patterns into the "W" type and "M" type.

Davranche et al. (2005) demonstrated that the REE complexation by organic acids inhibits the development of the tetrad effect and of Ce anomalies in the REE patterns of aqueous solutions. This is because the complexation operated by organic acids is not selective towards any specific lanthanide and therefore also Ce.

555   The REE complexation with the organic acids can therefore explain the absence of Ce anomalies and of the tetrad effect in the REE patterns of the leachates of the younger litter fractions (Do FL, Do OLn, Be FL, Be OLn, Be OLv), but it does not justify the positive Ce anomalies found in the patterns of the leachates of the oldest fractions (Do OLv and Be OF) and the W-type tetrad effect observed in the pattern of the Be OF leachate. The presence of both positive Ce anomalies and the tetrad effect can be explained by a biological-driven accumulation of

560   manganese oxides (MnO_2) particles on the surface of the components of the oldest litter layers and their subsequent transport into solution. Keiluweit et al. (2015) demonstrated how litter decomposition is controlled by the manganese redox cycle. The authors explained that, during the first three years of the litter degradation, specific

microorganisms (in particular fungi) are able to transform the Mn^{2+} supplied by the decomposing organic material into the more reactive Mn^{3+} form. This latter would be subsequently used by other microorganisms for the degradation of the aromatic compounds (such as lignin and tannins) through redox reactions with the litter components, which would give the Mn back under its reduced Mn^{2+} form. After the first few years, the excess of Mn^{3+} produced by the biological activity precipitates under the form of Mn^{3+} / Mn^{4+} oxides accumulating on the surface of the litter during more advanced stages of degradation (Keiluweit et al., 2015).

Unlike organic acids, manganese oxides are capable of a selective adsorption of Ce, along the other REE, with a mechanism of oxidative scavenging through which Ce is preferentially trapped onto the surface of the above-mentioned oxides (Bau, 1999; Bau and Koschinsky, 2009, Pourret and Davranche, 2013). The Ce enrichment linked to Mn oxides could be the reason for the formation of positive Ce anomalies in the waters that leached the litter material.

We conjecture that after a rainfall event, residual water that is deposited onto the surface of the oldest litter fraction has inherited a specific REE signature after passing through the younger litter layers above. We can assume that such a signature is similar to that of the leachates of the younger litter fractions recovered during our experiment (with the related MREE-HREE enrichment). Once the MnO_2 deposited onto the surface of the old litter interacts with this solution, it would preferentially adsorb Ce with the scavenging mechanism previously mentioned. A question mark here is related to the form (complexed or free ions) of the REE when they enter in contact with the MnO_2. We assume that their main form during such an interaction occurs mainly as free ions as their complexation with organic acids could inhibit the preferential adsorption of Ce onto MnO_2 as observed by Davranche *et al*. (2005 and 2008). Moreover, they also highlighted a process of REE-organic acids complexes dissociating with time and with decreasing HA/MnO_2 ratios. The reduced DOC concentrations (Table SI-4) and the presence of the MnO_2 in Do OLv and Be OF would then lead to the dissociation of the OA-REE complexes and to the re-adsorption of the REE onto the MnO_2 with a preferential intake of Ce. Note that when compared to the Do OLv leachate, the Be OF leachate shows lower DOC, a higher Ce anomaly and the presence of TE, which may be a direct effect of the decrease of the OA/MnO_2 ratios on the development of these specific REE features in the solutions during the litter degradation.

Interestingly, for both species the leachates of younger litter fractions (Do FL, Do OLn, Be FL, Be OLn and Be OLv) show higher DOC concentrations and lower pH than those of the oldest litter fractions (Do OLv and Be OF), as shown in Table SI-4. This strengthens the assumption that the REE patterns in leachates of fresh leaves and young litter fractions are shaped by the presence of organic acids, which confers the typical increasing trend from La to Lu and the absence of positive Ce/Ce* and of TE (Fig. 4). On the contrary, leachates of the oldest litter fractions show higher pH, lower DOC, positive Ce anomalies and TE (in Be OF leachate), indicating that the shapes of the REE patterns in the leachates of the oldest litter fractions are mainly resulting from the OA-REE dissociation accompanied by Ce-enriched MnO_2. This would explain both the increasing trend from La to Lu and the development of positive Ce anomalies (with a TE in the Be OF sample) in the leachates of the oldest litter fractions. We propose that the process leading to Ce enrichment in waters that are in contact with the oldest litter fractions occurs in three steps, as reported below (and more accurately in Figure 5):

i. Biologically-driven accumulation of MnO_2 particles onto the surface of the old litter components;
ii. Dissociation of OA-REE complexes and subsequent oxidative scavenging of Ce onto the MnO_2 particles' surface in the presence of stationary water in the litter surface during the degradation;

iii. Dissolution of Ce-enriched MnO_2 particles and/or their direct transport as MnO_2 nanoparticles into solution operated by incident waters characterized by higher water volume and higher turbulence, which may then "wash" the surface of the oldest litter layers. One or the combination of these two processes would lead to Ce enrichment in the solutions, thus developing a positive anomaly.

One may argue that the yields of Mn during the leaching experiment are higher in the Do OLn and Be OLv leachates where the Ce anomalies do not appear. Here it is important to consider not the overall concentration of Mn in the leachates, but rather the chemical form in which this element is present in the litter layers. We recall, in fact, that the formation of Mn oxides (which lead to the development of Ce enrichment) only occur during the last stages of the litter decay, in our case Do OLv and Be OF.

Stage I

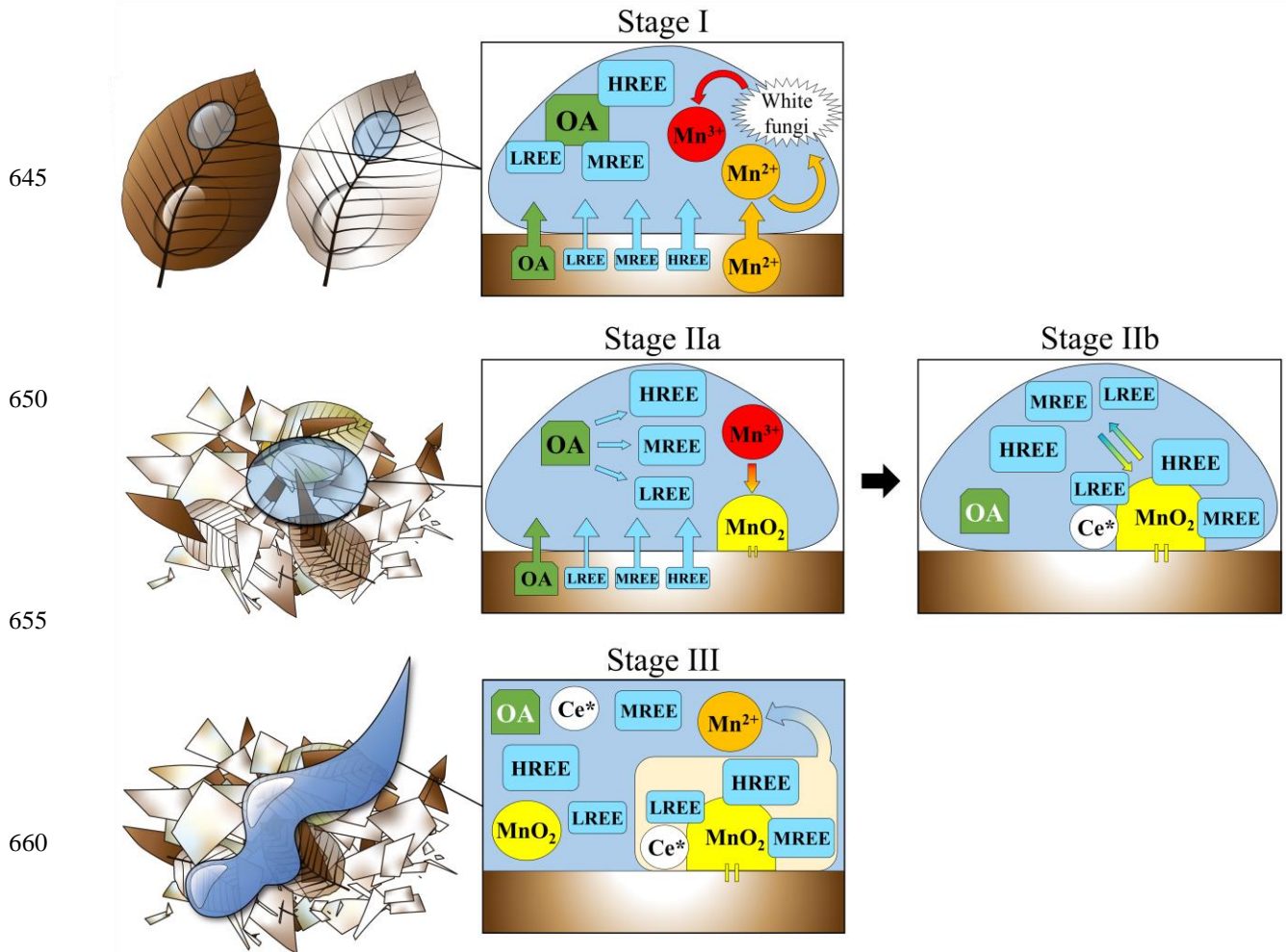## Stage IIa

Stage IIb

Stage III

Figure 5. Conceptual model representing the main processes treated in this study occurring during the litter degradation.

Stage I Do OLn, Be OLn and Be OLv degradation

- Preferential release of HREE and MREE linked to their affinity for organic acids
- REE complexation with organic acids and subsequent inhibition of Ce anomalies and Tetrad Effect
- Transformation of Mn^{2+} (coming from the litter) into Mn^{3+} operated by white fungi

Stage IIa Do OLv and Be OF degradation

- Preferential release of HREE and MREE linked to their affinity for organic acids
- REE complexation with organic acids and subsequent inhibition of Ce anomalies and Tetrad Effect
- Accumulation and precipitation of Mn^{3+} under the form of Mn oxides
- Decrease in the OA/MnO_2 ratios (due to Increased concentration of MnO_2 and decreased release of organic acids), lead to the REE-organic acids dissociation

Stage IIb Do OLv and Be OF degradation

- REE released from the organic acids in **Stage IIa** are re-adsorbed onto the MnO_2 particles positioned on the litter surface
- Scavenging of Ce and its subsequential enrichment on the Mn oxide surface

Stage III Do OLv and Be OF degradation

- Higher volume of water and higher turbulence lead to the dissolution and/or direct transportation of the Ce-enriched Mn oxides particles into solution, which inherits the enrichment in Ce and develops a tetrad effect (Be OF only)

For completeness of information, it is worth mentioning two other factors, which may play a role in the development of Ce positive anomalies in solution: the presence of siderophores and Fe oxides. Siderophores are a group of small molecules characterised by a strong affinity towards Fe^{3+} and that are among the strongest iron-chelating agents in nature. Plants (*Poaceae*), fungi and bacteria are able to synthetize and release these compounds to enhance the Fe assimilation in iron-deficiency conditions (Chennappa *et al.*, 2019). Accordingly, one would expect siderophores to be released by the above-mentioned organisms to enhance the Fe assimilation especially during the first stages of the degradation where the Fe concentrations are lower when compared to the oldest fractions. Therefore, it is precisely in those early stages of decay where one would expect a greater participation of siderophores in the geochemical behaviour of Ce. Kraemer *et al.* (2017) demonstrated that siderophores are able to scavenge Ce by oxidizing it to Ce (IV), forming stable complexes in solution and leading to the development of positive Ce anomalies. Anomalies that are not shown in the leachates of the younger litter fractions where one would expect them according to the iron-deficiency. However, the Ce enrichment occurs only in the leachates of the oldest litter fractions, suggesting that the process acting in these circumstances is timeframe-specific (during specific stages of the degradation) rather than being condition-specific (iron-deficiency). Other important aspects to consider are the much higher concentrations of manganese in the leachates when compared to iron and the different leaching yields that Fe shows between the two tree species (Table SI-4), while Ce anomaly instead shows the same dynamics.

Fe oxides have been shown to potentially oxidize Ce(III) to Ce(IV) and scavenge this element from the water column leading to the development of Ce positive anomalies in the oxide fractions as reported by Bau and Koschinsky (2009). Nevertheless, the Mn/Fe ratios in the leachates of our litter samples ($33.6 \leq Mn/Fe \leq 276.5$) are much higher than the ones of the samples treated by the above-mentioned authors ($1.57 \leq Mn/Fe \leq 1.65$), suggesting that Mn might be a better candidate to act in the oxidative scavenging of Ce during the degradation of the litter rather than Fe.

4.3 Behaviour of Ca and Eu during litter degradation

Calcium leaching yields are close to or even lower than those of trivalent metals during the first stages of Douglas-fir leaf degradation (Do FL and Do OLn), while its release increases from the oldest litter layer (Do OLv). In European beech samples, the leaching of Ca is not as low as for Douglas-fir samples but is lower than the other divalent elements (Mn and Mg). Calcium is involved in many plant's mechanisms and, among these, the stabilization of the cell wall structure is of vital importance. Indeed, it is an essential component of the calcium pectate, an insoluble molecule that forms polymers in between the cell walls linking them together (Bateman and Basham, 1976; Proseus and Boyer, 2012). The fact that calcium pectate is insoluble and that is positioned in between the cell walls makes this molecule less accessible to microorganisms during the initial stages of the degradation, leading to a reduced release of Ca. The fragmentation of the leaves and the decay of the weakest tissues during the first stages of degradation would therefore facilitate the accessibility of these insoluble components to the biological degradation (Norman, 1929), thus contributing to a higher Ca release from the oldest litter fractions. However, the stabilization of cell walls cannot explain the increase in Ca concentrations observed in the litter fractions. This latter, on the other hand, can be explained by the Ca-translocation mechanism occurring during the leaf's senescence described by Turpault *et al.* (2021). At high concentrations, Ca is a toxic element for trees and during the senescence is translocated from the other tree organs

to the leaves, where it crystalizes under the form of insoluble Ca-bearing biominerals and is released to the litter material during the leaf fall as a form of anti-toxicity mechanism.

Calcium accumulation during the leaf's senescence can also explain the development of the slight Eu enrichment found in the Do OLn fraction in comparison to the fresh leaves (Eu/Eu*=1.09 and 0.93 in Do OLn and Do FL respectively). The linkage between Ca and Eu in plants lies on the capability of Eu^{3+} to substitute Ca^{2+} in some physiological mechanisms due to their similar ionic radii. Stille *et al*. (2006) proposed that Eu positive anomalies in leaves can be related to its preferential uptake by plants. They argued that Eu^{3+} can follow the same Ca^{2+} fate in cells cytoplasm, in which Ca concentrations are controlled via oxalate crystals precipitation leading to the accumulation of Eu in leaves and the consequential development of a positive anomaly. In 2003, Gao *et al*. reported a preferential Eu accumulation in cell membranes and its capability to use calcium ion channels to enter inside cells and get absorbed by cytoplasmatic organelles. Ding *et al*. (2005) linked Eu enrichments in roots to the precipitation of Eu-enriched phosphate particles, while Brioschi *et al*. (2013) proposed that the origin of such anomalies in roots should be attributed to Ca-depleted soils where plants may suffer of Ca-deficiency and where Eu-substitution of Ca is responsible for the enrichments especially in soils characterised by high Eu/Ca ratios. In the case of the Weierbach soil, the Eu/Ca ratios in the first 60 cm of soil under the Douglas-fir stand ranged from 0.0005 to 0.0044 (Moragues-Quiroga *et al*., 2017) which, according to Brioschi *et al*. (2013), indicates a Ca depletion. Nonetheless, the slight enrichment in Eu is occurring in the Do OLn fraction and not in the fresh leaves where instead the Eu/Eu* is below 1. Then, it is our opinion that the Eu enrichment in the litter cannot be linked to the Ca-depleted soil as it does not occur during the leaf entire living period. In this study, the Eu enrichment in Do OLn indicates its involvement in Ca-dedicated biochemical pathways, which would lead to an anomalous accumulation of Eu in respect to the other REE in Douglas-fir litter. The fact the Eu/Eu* increases in Do OLn and not in Do FL, suggests that Eu is involved in the same process of Ca translocation that occurs during the leaves' senescence.

Moreover, the positive Eu anomalies observed in the Do OLn and Do OLv leachates (Figs.3c and 4a) suggest that at least part of the Eu is fractionated into slightly less degradation-resistant compounds - compared to the lignin where the other REE are supposed to stay bound, or into biogenic minerals, which are released from the cytoplasm after the cell wall breaks. The presence of these afore-mentioned compounds in the leachates and their subsequent digestion would deliver additional Eu content to the solution, leading to the development of positive anomalies in the leachates of Do OLv and Do OLn fractions. This would be in line with Gao *et al*. (2003), Turpault *et al* (2021) and partially with Stille *et al*. (2006). In fact, if a preferential absorption of Eu by trees had played a role in the development of a positive anomaly in the leaves, we would have observed an enrichment already in the Do FL, in which instead it does not occur.

Although, the reason for calcium behaving differently in the two tree species during the litter degradation cannot be explained with our experiment, it could be due to possible differences in the chemical and/or mechanical structures of the leaves or in the physiology of the tree species.

4.4 Rare Earth Elements as a proxy for litter degradation resistance?

Both tree species show a progressive decrease in the Y/Ho ratios, indicating that during the degradation of the litter material, Y decouples from Ho, as it is preferentially leached. This trend in the Y/Ho ratios is also accompanied by the enrichment in LREE proceeding towards decay in both species, as shown by the La_N/Yb_N

ratios. These ratios can be thought of as proxies for classifying resistance to the litter degradation of the two tree species in the Weierbach forest. As illustrated in Figure 6, the smaller the slope of the regression line, the lower the resistance. In accordance with this, Douglas-fir samples appear to be less resistant than the European beech ones. This is in line with the higher yields of trivalent metals we observed in the leachates of Douglas-fir samples as they are bound to the most resistant tissues. It is interesting to note that the fresh leaves from both species have a close position in the graph, indicating a similar stage of degradation (none) which changes over time.
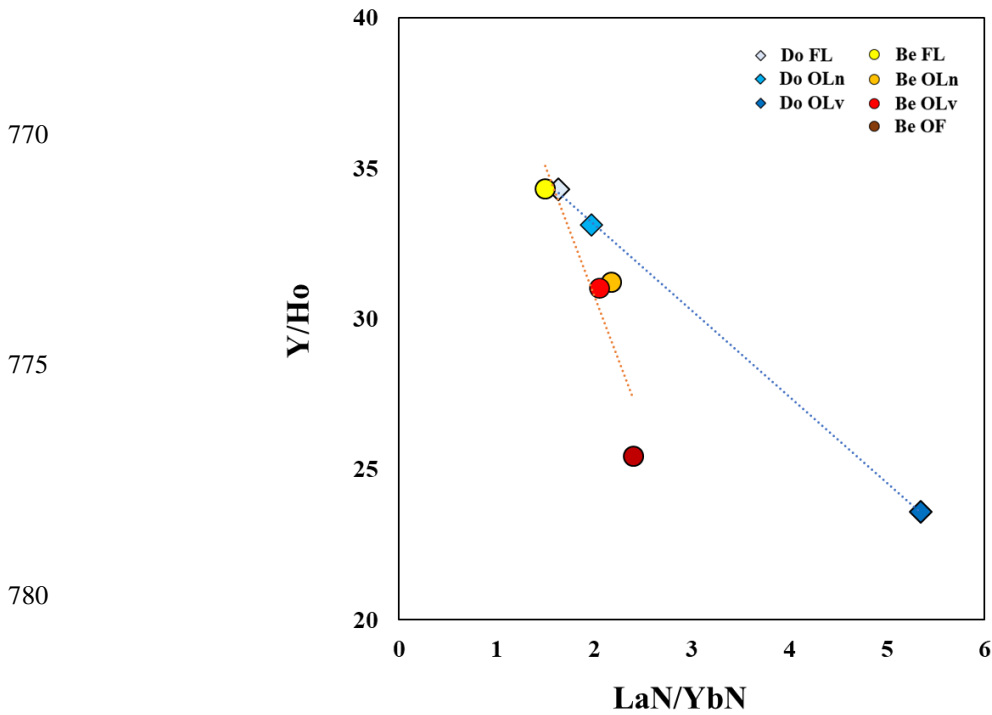
Figure 6. Y/Ho vs LaN/TbN ratios of fresh leaves and litter samples of Douglas-fir (blue scale) and European beech (red scale).

4.5 REE in soil solutions

The differences in the average soil solution REE patterns observed between the two experimental sites in the Weierbach catchment seem to be linked to the different REE release occurring during the degradation of the litter at each plot. Indeed, from a depth close to the litter layers (soil solution at 20 cm depth) to the deepest soil layer (soil solution at 60 cm depth), the evolution of the HREE enrichment, Ce anomaly and specific MREE (Gd and/or Eu) enrichments in soil solutions (Fig. 7) could be discussed according to similarities with the litter leachates (Fig. 3 c-d). It may be expected that if any degrading litter compound can contribute to the soil solution REE composition, it would be more easily observed close to the surface and would disappear progressively with depth, being diluted by the water-rock interaction processes and changing in redox conditions that control REE in soils (Braun et al., 1998; Laveuf and Cornu, 2009). Our results are in accordance with this expectation. For instance, particularities in the REE patterns of these litter leachates (especially for the last two stages of degradations for both species) seem to be mirrored by their respective soil solutions. Indeed, Eu and Gd enrichments, observed independently in both litter leachates (Do OLv and Be OF), were also found in the related

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soil solutions (1.08≤Gd/Gd*≤1.21 in Be SS and 1.06≤Eu/Eu*≤1.15 in Do SS). In both profiles, such anomalies are higher at 40 cm and decrease at 60 cm. This is in line with the leachate REE patterns normalized by the respective bulk litter concentrations reported in Figure 4. Indeed, in the patterns of Be OLv and Be OF, Gd is the most enriched, while in patterns of Do OLn and Do OLv Eu leads the MREE enrichment. This may indicate a preferential release of these two elements to the soil solutions during the natural leaching operated by rainfall and throughfall on Do OLn, Do OLv fractions in the Douglas-fir stand and of Be OLv, Be OF fractions in the beech stand.
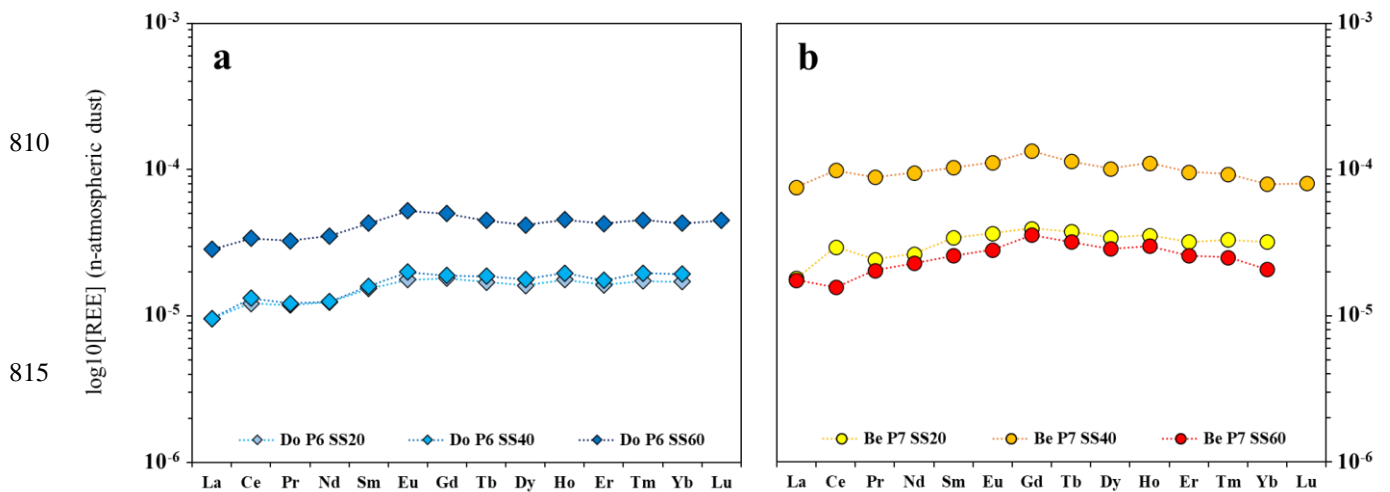
Figure 7. Patterns of the average Rare Earth Elements concentrations in soil solutions of DoP (**a**) and BeP (**b**). REE concentrations have been normalized by the values in the local atmospheric dust.

Moreover, REE patterns of soil solutions from both profiles show Ce positive anomalies (Ce/Ce*=1.14 in Do SS20 and Ce/Ce*= 1.39 in Be SS20) that are close to those in the leachates of the oldest litter fractions. The amplitude of these anomalies decreases with the increasing depth until they disappear at 60 cm. Here again, the natural leaching of the oldest litter material could lead to the desorption of REE from the MnO_2 or the direct transport into solution of Mn oxide nano-particles enriched in Ce, finally leading to a positive Ce anomaly in soil solutions. However, due to its redox sensitive nature, Ce dynamics are not easily understandable in soil solutions in which, due to the oxidative conditions we would expect there to be a depletion (negative Ce/Ce*) linked to the precipitation of Ce^{4+} as cerianite and adsorption onto Fe-Mn oxy-hydroxides.

Other important features of leachate patterns that are mirrored in the soil solutions are the HREE and MREE enrichments for the Douglas-fir and European beech stands, respectively, when compared to LREE. Indeed, average Douglas-fir soil solutions showed quite stable La_N/Yb_N ratios at all depths with values comprised between 0.50 (Do SS40) and 0.66 (Do SS60), which are in line with the La_N/Yb_N ratios of Douglas-fir litter leachates (0.10≤La_N/Yb_N≤0.82). Concerning the beech stand, La_N/Gd_N in average soil solutions shows values between 0.46 (Be SS20) and 0.57 (Be SS40), still in line with the values of beech litter leachates (0.43≤La_N/Gd_N≤0.51) obtained with our experiment. Similarities like these suggest a strong impact of the litter degradation on what is the REE signature of soil solutions, especially at shallower depths, and the fact that the anomalies tend to disappear in the deepest solutions strengthen this assumption.

It must be said that the same environmental conditions to which the litter is generally exposed are not found in the laboratory. Conditions under which a greater degradation efficiency would be expected and that were avoided due to limitations present in the laboratory (such as the limited exchange of gases with the atmosphere, limited light, greater volume of water per litter surface area, lower concentration of microorganisms). Additional *in-situ* studies regarding the REE dynamics in the Weierbach catchment's soils are necessary to better understand and quantify the real contribution of litter degradation to the REE composition of soil solutions in a forest ecosystem. Moreover, chromatographic analysis of the leachates and SEM analysis of litter surfaces could help, respectively, to elucidate what kind of REE ligands are present in the different leachates and to observe the existence (or not) of MnO_2 particles deposited on the surface of the oldest litter fractions.

5 Conclusions

We focused our attention on the role of forest vegetation on REE and major cations sequestration and release into and from leaf tissues during the litter degradation. As shown in our experiment and similarly for both tree species, major cations and nutrients like Na, Mg, K, and Mn are preferentially located in more labile tissues and are easily released during litter degradation, while Pb, Fe, Al and REE tend to be accumulated in the most recalcitrant tissues. We conjecture that such a sequestration in degradation-resistant tissues is imputable to the binding with lignins as the most resistant compounds in leaves.

Our results clearly show that litter degradation plays an important role in the REE dynamics in forest ecosystems. New findings related to REE dynamics during litter degradation and the potential of REE as complementary tracers for litter degradation processes were highlighted. The observation that the Ce anomaly and tetrad effect only occur in leachates of the oldest litter fractions can be linked to the accumulation of MnO_2 on the surface of the litter after the first years of degradation. In comparison to the major cations, REE presented marked differences during the degradation of the litters of European beech and Douglas-fir. In this latter, Eu seems to be involved in the same Ca translocation pathway that occurs during the leaf senescence. Moreover, the evolution of the La_N/Yb_N and Y/Ho ratios could be used as a proxy to analyse the resistance to the degradation of the leaves and litter between these two tree species.

Finally, the type of tree cover and the degradation stage of the litter are important parameters to consider when studying the chemistry of REE in forest soil waters. Similarities between the REE patterns of fresh leaves and litter leachates and REE patterns of soil solutions have been reported, possibly suggesting the importance of vegetation in determining the REE signatures in soil solutions. When compared to the other elements in the series, HREE are preferentially released from litter into solution due to the stronger affinity they have with the organic acids produced during the leaves' degradation stages. This would also explain the unexpected positive Ce anomaly that can be observed in the shallower soil solutions of the Weierbach experimental catchment in Luxembourg.

6 Data availability

The database used in this study is publicly available at zenodo.org (https://doi.org/10.5281/zenodo.5569559).

890 **10 References**

Adeleke, R., Nwangburuka, C. and Oboirien, B.: Origins, roles and fate of organic acids in soils: A review, South African J. Bot., 108, 393–406, doi:10.1016/j.sajb.2016.09.002, 2017.

Albers, D., Migge, S., Schaefer, M. and Scheu, S.: Decomposition of beech leaves (Fagus sylvatica) and spruce needles (Picea abies) in pure and mixed stands of beech and spruce, Soil Biol. Biochem., 36(1), 155–164,

895 doi:10.1016/j.soilbio.2003.09.002, 2004.

Alejandro, S., Höller, S., Meier, B. and Peiter, E.: Manganese in Plants: From Acquisition to Subcellular Allocation, Front. Plant Sci., 11(March), 1–23, doi:10.3389/fpls.2020.00300, 2020.

Amann, B. T., Mulqueen, P. and Horrocks, W. D. W.: A continuous spectrophotometric assay for the activation of plant NAD kinase by calmodulin, calcium(II), and europium(III) ions, J. Biochem. Biophys. Methods, 25(4),

900 207–217, doi:10.1016/0165-022X(92)90015-3, 1992.

Austin, A. T. and Ballaré, C. L.: Dual role of lignin in plant litter decomposition in terrestrial ecosystems, Proc. Natl. Acad. Sci. U. S. A., 107(10), 4618–4622, doi:10.1073/pnas.0909396107, 2010.

Bateman, D. F. and Basham, H. G.: Degradation of Plant Cell Walls and Membranes by Microbial Enzymes, Physiol. Plant Pathol., 316–355, doi:10.1007/978-3-642-66279-9_13, 1976.

905 Bau, M.: Rare-earth element mobility during hydrothermal and metamorphic fluid-rock interaction and the significance of the oxidation state of europium, Chem. Geol., 93(3–4), 219–230, doi:10.1016/0009-2541(91)90115-8, 1991.

Bau, M.: Controls on the fractionation of isovalent trace elements in magmatic and aqueous systems: Evidence from Y/Ho, Zr/Hf, and lanthanide tetrad effect, Contrib. to Mineral. Petrol., 123(3), 323–333, doi:10.1007/s004100050159, 1996.

Bau, M.: Scavenging of dissolved yttrium and rare earths by precipitating iron oxyhydroxide: Experimental evidence for Ce oxidation, Y-Ho fractionation, and lanthanide tetrad effect, Geochimica et Cosmochimica Acta, 63(1), 67-77, doi: 10.1016/S0016-7037(99)00014-9, 1999.

Bau, M. and Koschinsky, A.: Oxidative scavenging of cerium on hydrous Fe oxide: Evidence from the distribution of rare earth elements and yttrium between Fe oxides and Mn oxides in hydrogenetic ferromanganese crusts, Geochem. J., 43, 37–47, 2009.

Bienfait, H.: Prevention of stress in iron metabolism of plants, Acta Bot. Neerl., 38(2), 105–129, doi:/10.1111/j.1438-8677.1989.tb02035.x, 1989

Braun, J. J., Viers, J., Dupré, B., Polve, M., Ndam, J. and Muller, J. P.: Solid/liquid REE fractionation in the lateritic system of Goyoum, East Cameroon: The implication for the present dynamics of the soil covers of the humid tropical regions, Geochim. Cosmochim. Acta, 62(2), 273–299, doi:10.1016/S0016-7037(97)00344-X, 1998.

Brioschi, L., Steinmann, M., Lucot, E., Pierret, M. C., Stille, P., Prunier, J. and Badot, P. M.: Transfer of rare earth elements (REE) from natural soil to plant systems: Implications for the environmental availability of anthropogenic REE, Plant Soil, 366(1–2), 143–163, doi:10.1007/s11104-012-1407-0, 2013.

Brun, C. B., Åström, M. E., Peltola, P. and Johansson, M. B.: Trends in major and trace elements in decomposing needle litters during a long-term experiment in Swedish forests, Plant Soil, 306(1–2), 199–210, doi:10.1007/s11104-008-9572-x, 2008.

Censi, P., Saiano, F., Pisciotta, A. and Tuzzolino, N.: Geochemical behaviour of rare earths in Vitis vinifera grafted onto different rootstocks and growing on several soils, Sci. Total Environ., 473–474, 597–608, doi:10.1016/j.scitotenv.2013.12.073, 2014.

Chadwick, O. A., Derry, L. A., Vitousek, P. M., Huebert, B. J. and Hedin, L. O.: Changing sources of nutrients during four million years of ecosystem development, Nature, 397(6719), 491–497, doi:10.1038/17276, 1999.

Cidu, R., Buscaroli, A., Biddau, R., Da Pelo, S., Zannoni, D., Vianello, G., Dinelli, E., Vittori Antisari, L. and Carbone, S.: Dynamics of rare earth elements in water–soil systems: The case study of the Pineta San Vitale (Ravenna, Italy), Geoderma, 193–194, 52–67, doi:10.1016/j.geoderma.2012.10.009, 2013.

Cogulet, A., Blanchet, P. and Landry, V.: Wood degradation under UV irradiation: A lignin characterization, J. Photochem. Photobiol. B Biol., 158, 184–191, doi:10.1016/j.jphotobiol.2016.02.030, 2016.

Davranche, M., Pourret, O., Gruau, G., Dia, A., Jin, D. and Gaertner, D.: Competitive binding of REE to humic acid and manganese oxide: Impact of reaction kinetics on development of cerium anomaly and REE adsorption, Chem. Geol., 247(1–2), 154–170, doi:10.1016/j.chemgeo.2007.10.010, 2008.

Davranche, M., Pourret, O., Gruau, G., Dia, A. and Le Coz-Bouhnik, M.: Adsorption of REE(III)-humate complexes onto MnO_2: Experimental evidence for cerium anomaly and lanthanide tetrad effect suppression, Geochim. Cosmochim. Acta, 69(20), 4825–4835, doi:10.1016/j.gca.2005.06.005, 2005.

945　Dia, A., Gruau, G., Olivié-Lauquet, G., Riou, C., Molénat, J. and Curmi, P.: The distribution of rare earth elements in groundwaters: Assessing the role of source-rock composition, redox changes and colloidal particles, Geochim. Cosmochim. Acta, 64(24), 4131–4151, doi:10.1016/S0016-7037(00)00494-4, 2000.

Ding, S. M., Liang, T., Zhang, C. S., Yan, J. C. and Zhang, Z. L.: Accumulation and fractionation of rare earth elements (REEs) in wheat: Controlled by phosphate precipitation, cell wall absorption and solution
950　complexation, J. Exp. Bot., 56(420), 2765–2775, doi:10.1093/jxb/eri270, 2005.

El-ramady, H. R.: Ecotoxicology of rare earth elements : Ecotoxicology of rare earth elements within soil and plant environments, ." KG: VDM Verlag Dr. Muller Aktiengsellsellshaft & Co; 2010.

Gautam, M. K., Lee, K. S., Berg, B., Song, B. Y. and Yeon, J. Y.: Trends of major, minor and rare earth elements in decomposing litter in a cool temperate ecosystem, South Korea, Chemosphere, 222, 214–226,
955　doi:10.1016/j.chemosphere.2019.01.114, 2019.

Gómez, L., Contreras, A., Bolonio, D., Quintana, J., Oñate-Sánchez, L. and Merino, I.: Phytoremediation with trees, Adv. Bot. Res., 89, 281-321, ., 2019.

Grobéty, B., Gieré, R., Dietze, V. and Stille, P.: Airborne particles in the urban environment, Elements, 6(4), 229–234, doi:10.2113/gselements.6.4.229, 2010.

960　Guo, L. B. and Sims, R. E. H.: Litter decomposition and nutrient release via litter decomposition in New Zealand eucalypt short rotation forests, Agric. Ecosyst. Environ., 75(1–2), 133–140, doi:10.1016/S0167-8809(99)00069-9, 1999.

Gwenzi, W., Mangori, L., Danha, C., Chaukura, N., Dunjana, N. and Sanganyado, E.: Sources, behaviour, and environmental and human health risks of high-technology rare earth elements as emerging contaminants, Sci.
965　Total Environ., 636, 299–313, doi:10.1016/j.scitotenv.2018.04.235, 2018.

Hissler, C., Hostache, R., Iffly, J.F., Pfister, L., Stille, P.: Anthropogenic rare earth element fluxes into floodplains: coupling between geochemical monitoring and hydrodynamic-sediment transport modelling. C. R. Geoscience 347 294–303, doi: 10.1016/j.crte.2015.01.003, 2015a.

Hissler, C., Stille, P., Juilleret, J., Iffly, J. F., Perrone, T. and Morvan, G.: Elucidating the formation of terra
970　fuscas using Sr–Nd–Pb isotopes and rare earth elements, Appl. Geochemistry, 54, 85–99, doi:10.1016/j.apgeochem.2015.01.011, 2015b.

Hissler, C., Martínez-Carreras, N., Barnich, F., Gourdol, L., Iffly, J. F., Juilleret, J., Klaus, J. and Pfister, L.: The Weierbach experimental catchment in Luxembourg: A decade of critical zone monitoring in a temperate forest - from hydrological investigations to ecohydrological perspectives, Hydrol. Process., 35(5), 1–7,
975　doi:10.1002/hyp.14140, 2021.

Hissler, C., Stille, P. Iffly, J.F., Guignard, C., Chabaux, F., Pfister, L.: Origin and Dynamics of Rare Earth Elements during flood events in contaminated river basins: Sr-Nd-Pb evidence. Environ. Sci. Technol., 50(9), 4624-4631. doi:10.1021/acs.est.5b03660, 2016.

980 Imadi, S. R., Waseem, S., Kazi, A. G., Azooz, M. M. and Ahmad, P.: Aluminum Toxicity in Plants: An Overview, Plant Met. Interact. Emerg. Remediat. Tech., 1–20, doi:10.1016/B978-0-12-803158-2.00001-1, 2016.

Jabiol, B., Zanella, A., Ponge, J. F., Sartori, G., Englisch, M., van Delft, B., de Waal, R. and Le Bayon, R. C.: A proposal for including humus forms in the World Reference Base for Soil Resources (WRB-FAO), Geoderma, 192(1), 286–294, doi:10.1016/j.geoderma.2012.08.002, 2013.

Jin, L., Ma, L., Dere, A., White, T., Mathur, R. and Brantley, S. L.: REE mobility and fractionation during shale 985 weathering along a climate gradient, Chem. Geol., 466, 352–379, doi:10.1016/j.chemgeo.2017.06.024, 2017.

Juilleret, J., Dondeyne, S., Hissler, C., Vancampenhout, K. and Deckers, J.: Mind the gap: A classification system for integrating the subsolum into soil surveys, Geoderma, 264, 332–339, doi:10.1016/j.geoderma.2015.08.031, 2016.

Juranic, N.: Nephelauxetic Effect in Paramagnetic Shielding of Transition-Metal Nuclei in Octahedral d6 990 Complexes, J. Am. Chem. Soc., 111(21), 8326, doi:10.1021/ja00203a074, 1989.

Keiluweit, M., Nico, P., Harmon, M. E., Mao, J., Pett-Ridge, J. and Kleber, M.: Long-term litter decomposition controlled by manganese redox cycling, Proc. Natl. Acad. Sci. U. S. A., 112(38), E5253–E5260, doi:10.1073/pnas.1508945112, 2015.

Krishna, M. P. and Mohan, M.: Litter decomposition in forest ecosystems: a review, Energy, Ecol. Environ., 995 2(4), 236–249, doi:10.1007/s40974-017-0064-9, 2017.

Labeeuw, L., Martone, P. T., Boucher, Y. and Case, R. J.: Ancient origin of the biosynthesis of lignin precursors, Biol. Direct, 10(1), 1–21, doi:10.1186/s13062-015-0052-y, 2015.

Laveuf, C. and Cornu, S.: A review on the potentiality of Rare Earth Elements to trace pedogenetic processes, Geoderma, 154(1–2), 1–12, doi:10.1016/j.geoderma.2009.10.002, 2009.

1000 Leisola, M., Pastinen, O. and Axe, D. D.: Lignin--Designed Randomness, BIO-Complexity., 2012(3), 1–11, doi:10.5048/bio-c.2012.3, 2012.

Lequy, É., Conil, S. and Turpault, M. P.: Impacts of Aeolian dust deposition on European forest sustainability: A review, For. Ecol. Manage., 267, 240–252, doi:10.1016/j.foreco.2011.12.005, 2012.

Li, X., Chen, Z., Chen, Z. and Zhang, Y.: A human health risk assessment of rare earth elements in soil and 1005 vegetables from a mining area in Fujian Province, Southeast China, Chemosphere, 93(6), 1240–1246, doi:10.1016/j.chemosphere.2013.06.085, 2013.

Liang, T., Ding, S., Wenchong, S., Zhongyi, C., Zhang, C. and Li, H.: A review of fractionations of rare earth elements in plants, J. Rare Earths, 26(1), 7–15, doi:10.1016/S1002-0721(08)60027-7, 2008.

Liang, T., Zhang, S., Wang, L., Kung, H. Te, Wang, Y., Hu, A. and Ding, S.: Environmental biogeochemical behaviors of rare earth elements in soil-plant systems, Environ. Geochem. Health, 27(4), 301–311, doi:10.1007/s10653-004-5734-9, 2005.

McLennan, S.: Rare earth element geochemistry and the tetrad effect, Geochim. Cosmochim. Acta, 58(9), 2025–2033, doi:10.1016/0016-7037(94)90282-8, 1994.

Ma, L., Jin, L. and Brantley, S. L.: How mineralogy and slope aspect affect REE release and fractionation during shale weathering in the Susquehanna/Shale Hills Critical Zone Observatory, Chem. Geol., 290(1–2), 31–49, doi:10.1016/j.chemgeo.2011.08.013, 2011.

Maathuis, F. J. M.: Sodium in plants: Perception, signalling, and regulation of sodium fluxes, J. Exp. Bot., 65(3), 849–858, doi:10.1093/jxb/ert326, 2014.

Marsac, R., Davranche, M., Gruau, G. and Dia, A.: Metal loading effect on rare earth element binding to humic acid: Experimental and modelling evidence, Geochim. Cosmochim. Acta, 74(6), 1749–1761, doi:10.1016/j.gca.2009.12.006, 2010.

Moll, H., Sachs, S. & Geipel, G.: Plant cell (*Brassica napus*) response to europium(III) and uranium(VI) exposure. Environ Sci. Pollut. Res. 27, 32048–32061, dot:10.1007/s11356-020-09525-2, 2020.

Möller, P.: The distribution of rare earth elements and yttrium in water-rock interactions: field observations and experiments, Water-Rock Interaction. Water Science and Technology Library, vol 40, 97–123, doi:10.1007/978-94-010-0438-1_4, 2011.

Moragues-Quiroga, C., Juilleret, J., Gourdol, L., Pelt, E., Perrone, T., Aubert, A., Morvan, G., Chabaux, F., Legout, A., Stille, P. and Hissler, C.: Genesis and evolution of regoliths: Evidence from trace and major elements and Sr-Nd-Pb-U isotopes, Catena, 149, 185–198, doi:10.1016/j.catena.2016.09.015, 2017.

Morris, L.A. Nutrient cycling, Encyclopedia of Forest Sciences, 3, 1227–1235, ISBN 0-12-145160-7.

Nakada, R., Shibuya, T., Suzuki, K. and Takahashi, Y.: Europium anomaly variation under low-temperature water-rock interaction: A new thermometer, Geochemistry Int., 55(9), 822–832, doi:10.1134/S001670291709004X, 2017.

Nikolic, M. and Pavlovic, J.: Plant Responses to Iron Deficiency and Toxicity and Iron Use Efficiency in Plants, Plant Micronutrient Use Efficiency, 55-69, doi:10.1016/B978-0-12-812104-7.00004-6, 2018.

Norman, A. G.: The Biological Decomposition of Pectin, Ann. Bot., 43(170), 233–243, doi:10.1111/j.1469-8137.1941.tb07026.x, 1929.

Pacyna, J.M., Atmospheric Deposition, Encyclopedia of Ecology, 275-285, doi:10.1016/B978-008045405-4.00258-5, 2008.

Pagano, G., Aliberti, F., Guida, M., Oral, R., Siciliano, A., Trifuoggi, M. and Tommasi, F.: Rare earth elements in human and animal health: State of art and research priorities, Environ. Res., 142, 215–220, doi:10.1016/j.envres.2015.06.039, 2015.

Page, V. and Feller, U.: Heavy Metals in Crop Plants: Transport and Redistribution Processes on the Whole Plant Level, Agronomy, 5(3), 447–463, doi:10.3390/agronomy5030447, 2015.

1045   Pourret, O. and Davranche, M.: Rare earth element sorption onto hydrous manganese oxide: A modeling study, J. Colloid Interface Sci., 395(1), 18–23, doi:10.1016/j.jcis.2012.11.054, 2013.

Pourret, O., Davranche, M., Gruau, G. and Dia, A.: Rare earth elements complexation with humic acid, Chem. Geol., 243(1–2), 128–141, doi:10.1016/j.chemgeo.2007.05.018, 2007.

Pourrut, B., Shahid, M., Dumat, C., Winterton, P. and Pinelli, E.: Lead Uptake, Toxicity, and Detoxification in
1050   Plants, Rev. Environ. Contam. Toxicol., 213, 113–136, doi:10.1007/978-1-4419-9860-6_4, 2011.

Proseus, T. E. and Boyer, J. S.: Pectate chemistry links cell expansion to wall deposition in *Chara corallina*, Plant Signal. Behav., 7(11), 1490-1492, doi:10.4161/psb.21777, 2012.

Rahman, M. M., Tsukamoto, J., Rahman, M. M., Yoneyama, A. and Mostafa, K. M.: Lignin and its effects on litter decomposition in forest ecosystems, Chem. Ecol., 29(6), 540–553, doi:10.1080/02757540.2013.790380,
1055   2013.

Ramos, S. J., Dinali, G. S., Oliveira, C., Martins, G. C., Moreira, C. G., Siqueira, J. O. and Guilherme, L. R. G.: Rare Earth Elements in the Soil Environment, Curr. Pollut. Reports, 2(1), 28–50, doi:10.1007/s40726-016-0026-4, 2016.

Reynolds, R., Neff, J., Reheis, M. and Lamothe, P.: Atmospheric dust in modern soil on aeolian sandstone,
1060   Colorado Plateau (USA): Variation with landscape position and contribution to potential plant nutrients, Geoderma, 130(1–2), 108–123, doi:10.1016/j.geoderma.2005.01.012, 2006.

Rim, K. T., Koo, K. H. and Park, J. S.: Toxicological evaluations of rare earths and their health impacts to workers: A literature review, Saf. Health Work, 4(1), 12–26, doi:10.5491/SHAW.2013.4.1.12, 2013.

Rout, G., Samantaray, S., Das, P., Aluminium toxicity in plants : a review, Agronomie, 21(1), 3–21,
1065   doi:10.1051/agro:2001105, 2001.

Sardans, J. and Peñuelas, J.: Potassium Control of Plant Functions: Ecological and Agricultural Implications, Plants, 10(2), 419, doi:10.3390/plants10020419, 2021.

Schijf, J. and Zoll, A. M.: When dissolved is not truly dissolved-The importance of colloids in studies of metal sorption on organic matter, J. Colloid Interface Sci., 361(1), 137–147, doi:10.1016/j.jcis.2011.05.029, 2011.

1070   Shan, X., Wang, H., Zhang, S., Zhou, H., Zheng, Y., Yu, H. and Wen, B.: Accumulation and uptake of light rare earth elements in a hyperaccumulator *Dicropteris dichotoma*, Plant Sci., 165(6), 1343–1353, doi:10.1016/s0168-9452(03)00361-3, 2003.

Sharma, P. and Dubey, R. S.: Lead toxicity in plants, Braz. J. Plant Physiol.,, 17(1), doi:10.1590/S1677-04202005000100004, 2005.

1075   Shaul, O.: Magnesium transport and function in plants: The tip of the iceberg, BioMetals, 15(3), 309–323, doi:10.1023/A:1016091118585, 2002.

Shtangeeva, I. and Ayrault, S.: Effects of Eu and Ca on yield and mineral nutrition of wheat (*Triticum aestivum*) seedlings, Environ. Exp. Bot., 59(1), 49–58, doi:10.1016/j.envexpbot.2005.10.011, 2007.

Singh, S., Tripathi, D. K., Singh, S., Sharma, S., Dubey, N. K., Chauhan, D. K. and Vaculík, M.: Toxicity of
1080 aluminium on various levels of plant cells and organism: A review, Environ. Exp. Bot., 137, 177–193, doi:10.1016/j.envexpbot.2017.01.005, 2017.

Sonke, J. E. and Salters, V. J. M.: Lanthanide-humic substances complexation. I. Experimental evidence for a lanthanide contraction effect, Geochim. Cosmochim. Acta, 70(6), 1495–1506, doi:10.1016/j.gca.2005.11.017, 2006.

1085 Staaf, H.: Release of plant nutrients from decomposing leaf litter in a South Swedish beech forest, Ecography (Cop.), 3(2), 129–136, doi:10.1111/j.1600-0587.1980.tb00719.x, 1980.

Stille, P., Pierret, M. C., Steinmann, M., Chabaux, F., Boutin, R., Aubert, D., Pourcelot, L. and Morvan, G.: Impact of atmospheric deposition, biogeochemical cycling and water-mineral interaction on REE fractionation in acidic surface soils and soil water (the Strengbach case), Chem. Geol., 264(1–4), 173–186,
1090 doi:10.1016/j.chemgeo.2009.03.005, 2009.

Stille, P., Steinmann, M., Pierret, M. C., Gauthier-Lafaye, F., Chabaux, F., Viville, D., Pourcelot, L., Matera, V., Aouad, G. and Aubert, D.: The impact of vegetation on REE fractionation in stream waters of a small forested catchment (the Strengbach case), Geochim. Cosmochim. Acta, 70(13), 3217–3230, doi:10.1016/j.gca.2006.04.028, 2006.

1095 Suzuki, Y., Yokoi, S., Katoh, M., Minato, M. and Takizawa, N.: STABILITY CONSTANTS OF RARE-EARTH COMPLEXES WITH SOME ORGANIC LIGANDS, Rare Earths Mod. Sci. Technol., (1), 121–126, doi:10.1007/978-1-4613-3054-7, 1980.

Tagliavini, M., Tonon, G., Scandellari, F., Quiñones, A., Palmieri, S., Menarbin, G., Gioacchini, P. and Masia, A.: Nutrient recycling during the decomposition of apple leaves (Malus domestica) and mowed grasses in an
1100 orchard, Agric. Ecosyst. Environ., 118(1–4), 191–200, doi:10.1016/j.agee.2006.05.018, 2007.

Takahashi, Y., Châtellier, X., Hattori, K. H., Kato, K. and Fortin, D.: Adsorption of rare earth elements onto bacterial cell walls and its implication for REE sorption onto natural microbial mats, Chem. Geol., 219(1–4), 53–67, doi:10.1016/j.chemgeo.2005.02.009, 2005.

Takahashi, Y., Yamamoto, M., Yamamoto, Y. and Tanaka, K.: EXAFS study on the cause of enrichment of
1105 heavy REEs on bacterial cell surfaces, Geochim. Cosmochim. Acta, 74(19), 5443–5462, doi:10.1016/j.gca.2010.07.001, 2010.

Talbot, J. M., Yelle, D. J., Nowick, J. and Treseder, K. K.: Litter decay rates are determined by lignin chemistry, Biogeochemistry, 108(1–3), 279–295, doi:10.1007/s10533-011-9599-6, 2012.

Tang, J. and Johannesson, K. H.: Ligand extraction of rare earth elements from aquifer sediments: Implications
1110 for rare earth element complexation with organic matter in natural waters, Geochim. Cosmochim. Acta, 74(23), 6690–6705, doi:10.1016/j.gca.2010.08.028, 2010.

Tchougréèff, A. and Drownkowski, R.: Nephelauxetic Effect Revisited, Int. J. Quantum Chem., 109(11), 2606–2621, doi:10.1002/qua.21989, 2009.

Tommasi, F., Thomas, P. J., Pagano, G., Perono, G. A., Oral, R., Lyons, D. M., Toscanesi, M. and Trifuoggi, M.: Review of Rare Earth Elements as Fertilizers and Feed Additives: A Knowledge Gap Analysis, Arch. Environ. Contam. Toxicol., 81, 531-540, doi:10.1007/s00244-020-00773-4, 2021.

Turpault, M. P., Kirchen, G., Calvaruso, C., Redon, P. O. and Dincher, M.: Exchanges of major elements in a deciduous forest canopy, Biogeochemistry, 152(1), 51–71, doi:10.1007/s10533-020-00732-0, 2021.

Tyler, G.: Rare earth elements in soil and plant systems - A review, Plant Soil, 267, 191–206, doi:10.1007/s11104-005-4888-2, 2004.

Vázquez-Ortega, A., Zapata-Ríos, X., Rasmussen, C., McIntosh, J., Brooks, P. D., Perdrial, J., Amistadi, M. K., Chorover, J., Schaap, M., Harpold, A. and Pelletier, J. D.: Rare earth elements as reactive tracers of biogeochemical weathering in forested rhyolitic terrain, Chem. Geol., 391, 19–32, doi:10.1016/j.chemgeo.2014.10.016, 2015.

Vázquez-Ortega, A., Huckle, D., Perdrial, J., Amistadi, M. K., Durcik, M., Rasmussen, C., McIntosh, J. and Chorover, J.: Solid-phase redistribution of rare earth elements in hillslope pedons subjected to different hydrologic fluxes, Chem. Geol., 426, 1–18, doi:10.1016/j.chemgeo.2016.01.001, 2016.

Weng, J. K. and Chapple, C.: The origin and evolution of lignin biosynthesis, New Phytol., 187(2), 273–285, doi:10.1111/j.1469-8137.2010.03327.x, 2010.

Wenming, D., Xiangke, W., Xiaoyan, B., Aixia, W., Jingzhou, D. and Tao, Z.: Comparative study on sorption/desorption of radioeuropium on alumina, bentonite and red earth: Effects of pH, ionic strength, fulvic acid, and iron oxides in red earth, Appl. Radiat. Isot., 54(4), 603–610, doi:10.1016/S0969-8043(00)00311-0, 2001.

Woolhouse, H. W.: Toxicity and Tolerance in the Responses of Plants to Metals, Physiol. Plant Ecol. III, 245–300, doi:10.1007/978-3-642-68153-0_8, 1983.

Yuan, M., Liu, C., Liu, W. S., Guo, M. N., Morel, J. L., Huot, H., Yu, H. J., Tang, Y. T. and Qiu, R. L.: Accumulation and fractionation of rare earth elements (REEs) in the naturally grown Phytolacca americana L. in southern China, Int. J. Phytoremediation, 20(5), 415–423, doi:10.1080/15226514.2017.1365336, 2018.

Zaharescu, D. G., Burghelea, C. I., Dontsova, K., Presler, J. K., Maier, R. M., Huxman, T., Domanik, K. J., Hunt, E. A., Amistadi, M. K., Gaddis, E. E., Palacios-Menendez, M. A., Vaquera-Ibarra, M. O. and Chorover, J.: Ecosystem Composition Controls the Fate of Rare Earth Elements during Incipient Soil Genesis, Sci. Rep., 7(February), 1–15, doi:10.1038/srep43208, 2017.

Zeng, F., Tian, H. E., Wang, Z., An, Y., Gao, F., Zhang, L., Li, F. and Shan, L.: Effect of rare earth element europium on amaranthin synthesis in *Amarathus caudatus* seedlings, Biol. Trace Elem. Res., 93(1–3), 271–282, doi:10.1385/BTER:93:1-3:271, 2003.

Zhu, Z., Liu, C. Q., Wang, Z. L., Liu, X. and Li, J.: Rare earth elements concentrations and speciation in rainwater from Guiyang, an acid rain impacted zone of Southwest China, Chem. Geol., 442, 23–34, doi:10.1016/j.chemgeo.2016.08.038, 2016.