

~~Greenhouse gas emissions from tropical coastal wetlands and their
alternative agricultural lands: Where significant mitigation gains lie~~
Soil greenhouse gas fluxes from tropical coastal wetlands and
alternative agricultural land uses

Naima Iram¹, Emad Kavehei¹, Damien. T. Maher², Stuart. E. Bunn¹, Mehran Rezaei Rashti¹, Bahareh Shahrabi Farahani¹, Maria Fernanda Adame¹

¹Australian Rivers Institute, Griffith University, Brisbane, 4111, Australia

²School of Environment, Science and Engineering, Southern Cross University, Lismore, 2480, Australia

Correspondence to: Naima Iram (naima.iram@griffithuni.edu.au)

Abstract. Tidal coastal wetlands are crucial in regulating the global carbon budgets through carbon sequestration and greenhouse gas (GHG; CO₂, CH₄ and N₂O) emissions. The conversion of tidal coastal wetlands to agriculture land alters soil processes changing GHG emissions. However, our understanding about GHG emissions associated with land-use change of coastal wetland is limited. We measured soil GHG fluxes from mangroves, saltmarsh and freshwater tidal forest and their alternative agricultural lands including sugarcane and ponded pastures. We investigated seasonal variations in soil GHG fluxes between June 2018 and February 2020 in tropical Australia. The wet ponded pasture had by far the highest CH₄ emissions with 1,231±386 mg m⁻² d⁻¹, which were 200-fold higher than any other land use. Agricultural lands were the most significant sources of N₂O emissions with 55±9 mg m⁻² d⁻¹ from dry ponded pasture (wet-hot period) and 11±3 mg m⁻² d⁻¹ from sugar cane (hot-dry period), coinciding with fertilisation. The N₂O fluxes from the tidal coastal wetlands ranged between -0.55±0.23 and 2.76±0.45 mg m⁻² d⁻¹ throughout the study period. The highest CO₂ fluxes of 20±1 g m⁻² d⁻¹ were from the dry ponded pasture during the wet-hot period, while the saltmarsh had the lowest CO₂ fluxes having an uptake of -1.19±0.08 g m⁻² d⁻¹ in the dry-hot period. Overall, agricultural lands had significantly higher total cumulative GHG emissions (CH₄ + N₂O) of 7142 to 56,124 CO₂-eq kg ha⁻¹ y⁻¹ compared to those of any type of tidal coastal wetlands, which ranged between 144 and 884 CO₂-eq kg ha⁻¹ y⁻¹. Converting agricultural land, particularly wet ponded pasture, to tidal coastal wetlands could provide large GHG mitigation gains and potential financial incentives. Converting unproductive sugarcane land or pastures (especially ponded ones) to coastal wetlands could provide significant GHG mitigation.

1 Introduction

Coastal wetlands are at the interface between terrestrial and marine ecosystems and account for 10% of the global wetland area (Lehner and Döll 2004; Yang et al. 2017). Tropical coastal wetlands are highly productive ecosystems with significant potential for providing various services such as water quality improvement, biodiversity and carbon sequestration and storage (Lal, 2008; Duarte et al. 2013; Mitsch et al. 2013). Coastal wetlands, such as mangroves and marshes can accumulate a considerable amount of organic carbon in their sediments (Kauffman et al. 2020). Therefore, coastal wetlands have five times higher carbon storage potential than terrestrial forests (McLeod et al. 2011; Sjögersten et al. 2014). However, the anoxic soil conditions that promote carbon sequestration in coastal wetlands can also favour emissions of potent greenhouse gas (GHGs), e.g. CH_4 and N_2O which contribute to global warming (Whalen, 2005; Conrad, 2009).

Coastal wetlands exchange three main GHGs with the atmosphere: carbon dioxide (CO_2), methane (CH_4) and nitrous oxide (N_2O). Sediments of coastal wetlands have spatially and temporally diverse conditions which affect the microbial processes that regulate their soil atmospheric GHGs exchange (Bauza et al. 2002; Whalen, 2005). Emission of CO_2 from wetlands can be considered recycling of the carbon fixed through primary production, which is partly returned to the atmosphere through respiration (Oertel et al. 2016). Emission of CH_4 is a product of anaerobic and aerobic respiration in wetland soil (Angle et al., 2017; Saunio et al. 2020). Nitrous oxide fluxes are mainly driven by soil moisture and mediated by microbial activity under anoxic condition through denitrification and during aerobic condition through nitrification (Ussiri and Lal 2013). The total GHG budget of coastal wetlands is the result of the balance between aerobic and anaerobic sediment conditions that influence GHG fluxes from the main sources including soil and vegetation. Therefore, there are high uncertainties around global wetland GHG emissions (Kirschke et al. 2013; Oertel et al. 2016).

Despite potential GHG emissions from coastal wetlands, these are still likely to be lower than those from their alternative agricultural land uses. Agricultural practices are responsible for large amounts of GHG from their construction and throughout their productive life. Firstly, when wetlands are converted to agricultural land, the oxidation of sequestered carbon in the organic rich soils release significant amounts of CO_2 (Kauffman et al. 2015, 2018), contributing substantially to emissions caused by land use change (Ciais et al. 2013). Secondly, removing tidal flow and reverting coastal wetlands to freshwater ecosystems, such as during the creation of ponded pastures or dams, could result in high CH_4 and N_2O emissions (Martin et al. 2015; Maucieri et al. 2017; Capocci et al. 2019). Thirdly, the impoundment of ponds and dams can dramatically increase both CH_4 and N_2O emissions which are largely unaccounted for in GHG budgets from agriculture (Ollivier et al. 2019). Finally, the use of N fertilisers in intensive agricultural systems result in significant N_2O emissions (Rashti et al. 2015). Fertilisers may also increase CO_2 emissions by changing the balance between carbon and nitrogen within the aerobic condition of agricultural lands. Comprehensive studies accounting for simultaneous measurement of three main GHGs from wetlands and their alternate agricultural lands are very limited.

Land-use change affects different soil properties such as microbial communities (Van Leeuwen et al, 2017), enzyme activity (Niemi et al, 2005), soil water content (Grover et al, 2012), nutrient and terminal electron acceptor availability, and inundation (Xu, Wong and Reef, 2020), all of which affect GHG emissions (Pouyat et al, 2007). These emissions are likely to be highest in tropical conditions as temperature is one of the main drivers (Oertel et al, 2016). Of the three main GHGs, CO₂ has been well studied and incorporated into global climate change models, while models that include CH₄ and N₂O still have many uncertainties (Arneeth et al, 2010; Zachle and Dalmonech, 2011; Flato et al, 2014). Emissions of CH₄ and N₂O need to be accounted for when determining the net radiative balance of an ecosystem due to their high radiative forcing potential of these gases compared to CO₂. Some of the uncertainties in these models arise from the role of natural wetlands as "sinks" or "sources", which can only be established when all GHG emissions have been accounted for (Cobb et al., 2012), with only a few complete GHG budgets compiled for tropical wetlands (e.g. Mitsch et al. 2013; Wang et al, 2014; Gütlein et al., 2018; Harris et al, 2018).

Reducing GHG emissions in tropical coastal regions can be achieved through different management and restoration mechanisms, such as the reinstallation of tidal inundation on unused agricultural land (Kroeger et al, 2017). Reinstating tidal exchange to previously drained, or ponded freshwater agricultural land can mitigate GHG emissions through several mechanisms. Tidal coastal wetlands contain sulphate, which can inhibit or reduce the production of CH₄ through the competitive metabolic edge of sulphate reducers over methanogens (Poffenbarger et al, 2011). Tidal coastal wetlands also have high primary productivity rates leading to above ground and below ground carbon production and storage (Burden et al, 2013). Finally, tidal exchange also facilitates sediment carbon accumulation through increased sediment accretion due to sediment supply through tidal inundation (Burden et al, 2013). However, the land use change related GHG emissions could be useful for initiating restoration strategies that rely upon financial incentives such as carbon credits, as these could be a valuable mitigation strategy.

The emission factors or changes in GHG emission, from an agricultural land-use to a coastal wetland are required to evaluate the mitigation capacity of a restoration project. However, this information is very scarce in tropical regions (IPCC 2013). In mid 2000s, Australian greenhouse office initiated a national carbon accounting scheme to track sinks and sources of GHG fluxes. The purpose of the scheme was to help Australia stay on track to reduce GHG emission targets set by Kyoto Protocol (Beringer et al., 2013). However, large knowledge gaps were found in GHG flux estimates mainly due to spatial variation in ecosystems and soil types across Australia (Baldoek et al., 2012). Therefore, GHG flux assessment from different land covers is important to develop emissions reduction policies. This study will fill in this knowledge gap to improve wetland accounting in GHG mitigation strategies (IPCC 2013). Additionally, the information on GHG emissions from land use management particularly in tropical climate would improve our understanding of reducing GHG emissions by different management implication paradigms (IPCC 2013).

In this study, we measured the annual GHG fluxes from different land-use types including natural tidal coastal wetlands (freshwater tidal forest, saltmarsh, and mangroves) and agricultural lands (sugarcane plantation and ponded pastures) in tropical Australia. We aimed to assess the potential benefit of restoring wetlands in agricultural landscapes for decreasing GHG emissions. Based on these case studies, our objective was to provide some management implication to reduce GHG emissions from agricultural lands in tropical regions. We hypothesised that GHG emissions from agricultural lands would be larger than those of natural wetlands and that emissions would be higher in the hot wet season and during high tides.

Coastal wetlands are found at the interface of terrestrial and marine ecosystems and account for 10% of the global wetland area (Lehner and Döll 2004). They are highly productive and provide various ecosystem services such as water quality improvement, biodiversity, and carbon sequestration (Duarte et al, 2013). For instance, mangroves can accumulate five times more soil carbon than terrestrial forests (Kauffman et al, 2020). However, the high productivity and anoxic soil conditions that promote carbon sequestration can also favour potent greenhouse gas emissions (GHGs), including CO₂, CH₄ and N₂O (Whalen, 2005; Conrad, 2009).

The GHG emissions in coastal wetlands primarily result from microbial processes in the soil-water-atmosphere interface (Bauza et al, 2002; Whalen, 2005). The emission of CO₂ is a result of respiration, where fixed carbon by photosynthesis is partially released back into the atmosphere (Oertel et al., 2016). Emissions of CH₄ result from anaerobic and aerobic respiration by methanogenic bacteria, mostly in waterlogged conditions (Angle et al., 2017; Sauniois et al, 2020). Finally, N₂O emissions are caused by denitrification in anoxic conditions and nitrification in aerobic soils, both driven by nitrogen content and soil moisture (Ussiri and Lal 2013). Thus, the total GHG emissions from a wetland are driven by environmental conditions that favour these microbial processes, all of which result in highly variable emissions from wetlands worldwide (Kirschke et al., 2013; Oertel et al. 2016).

Despite potential high GHG emissions from coastal wetlands, these are likely to be lower than those from alternative agricultural land uses (Knox et al., 2015), which emit GHGs from their construction throughout their productive lives. Firstly, when wetlands are converted to agricultural land, the oxidation of sequestered carbon in the organic-rich soils release significant amounts of CO₂ (Drexler, de Fontaine, & Deverel, 2009; Hooijer et al, 2012). Secondly, removing tidal flow and converting coastal wetlands to freshwater systems, such as during the creation of ponded pastures, dams or agricultural ditches, can result in very high CH₄ emissions (Deemer et al., 2016; Grinham et al, 2018; Ollivier et al, 2019). For instance, agricultural ditches contribute up to 3% of the total anthropogenic CH₄ emissions globally (Peacock et al., 2021). Finally, the use of fertilisers significantly increases N₂O emissions (Rashti et al, 2015). Thus, emissions of GHG from land-use change can be mitigated through the reversal of these activities, for instance, reduction of fertiliser use and the reinstallation of tidal flow on unused agricultural land (Rashti et al, 2016; Kroeger et al. 2017).

134 This study measured the annual GHG fluxes (CO₂, CH₄ and N₂O) from three natural coastal wetlands (mangroves,
135 saltmarsh and freshwater tidal forests) and two agricultural land use sites (sugarcane plantation and pasture) in tropical
136 Australia. The objectives were to assess the difference in GHG fluxes throughout different seasons that characterise tropical
137 climates (dry-cool, dry-hot and wet-hot) and to identify environmental factors associated with these GHG fluxes. These data
138 will inform emission factors for converting wetlands to agricultural land uses and vice versa, filling in a knowledge gap
139 identified in Australia (Baldock et al., 2012) and tropical regions worldwide (IPCC, 2013).

141 2 Materials and Methods

142 2.1 Study sites

143 2.1 Study sites

144 The study area is located within the Herbert River catchment in Queensland, northeast Australia (Fig 1a). The region has a
145 tropical climate with mean monthly minimum temperature ranging from 14 to 23°C and mean monthly maximum
146 temperature ranging from 25 to 33°C (Australian Bureau of Meteorology, ABM, 2020; 1968-2020). The average rainfall is
147 2,158 mm y⁻¹ with the highest rainfall of 476 mm in February (ABM 2020; 1968-2020).

148 Wetlands in Queensland had undergone deforestation because of rapid agricultural development in the last century (Griggs-
149 2017). These wetlands were converted to agricultural land, mainly for sugarcane farming and grazing (WetlandInfo, 2016).

150 The study area is located within the Herbert River catchment in Queensland, northeast Australia (Fig 1a). The Herbert basin
151 covers 9,842 km², from which 56% is used for grazing mainly ponded pastures, 31% is conserved natural land use including
152 wetlands and forestry, 8% is sugarcane and 4% is other land uses (WetlandInfo, 2020), uses (Department of Science and
153 Environment, QLD, DES, WetlandInfo, 2020). Wetlands in this region were heavily deforested in the past century (1943-
154 1996) due to rapid agricultural development, primarily for sugarcane farming (Griggs, 2018). Before clearing, the land was
155 mostly covered by rainforest and coastal wetlands, mainly *Melaleuca* forest, grass and sedge swamps (Johnson, Ebert, &
156 Murray, 1999).

157 The region has a tropical climate with mean monthly minimum temperature ranging from 14 to 23°C and mean monthly
158 maximum temperature ranging from 25 to 33°C (Australian Bureau of Meteorology, ABM, 2020; 1968-2020). The average
159 rainfall is 2,158 mm y⁻¹ with the highest rainfall of 476 mm in February (ABM 2020; 1968-2020).

161 We selected five sites including three natural wetlands (Fig. 1b); a freshwater tidal forest (18°53'45" S,
162 146°15'52"E), a saltmarsh (18°53'43" S, 146°15'52"E) and a mangrove forest (18°53'42" S, 146°15'51"E), and two common
163 agricultural land use types in the area, sugarcane plantation (18°53'44. '6" S, 146°15'53.2"E) and a ponded pasture for fodder
164 grazing. The pasture contained different levels of ponding including shallow ponds (50-100 cm depth) and wet grassy areas

165 hereafter called "wet ponded pasture" (' 18°43'8"S, 146°15'50"E) and a "dry ponded pasture" (18°43'7"S, 146°15'50"E). All
166 the sites were located within the same property at Insulator Creek (18°53'44.6"S 146°15'53.2"E, Fig. 1A) except for the
167 ponded pasture, which was located about 20 km north at Mungalla Station (18°42'26"S, 146°15'18.37"E, Fig. 1A). The
168 selected freshwater tidal freshwater wetland was dominated by *Melaleuca quinquenervia* trees, a forest type commonly
169 known as "tea tree swamp". Seawards we sampled a saltmarsh dominated by *Sueda salsa* and *Sporobolus spp* followed by
170 mangrove forests dominated by *Avicennia marina* with few plants of *Rhizophora stylosa* (Fig. 1B). Landwards, the wetlands
171 were adjacent to a sugarcane farming area of ~110 ha. The sugarcane was fertilised once a year with urea at a rate of 135 kg
172 N ha⁻¹ and harvested during May-June, while foliage was left on the soil surface (trash blanket) after harvest. The ponded
173 pastures in Mungalla Station extended over 2,500 ha and supported ~900 cattle throughout the year by providing fodder to
174 cattle during dry periods. The selected ponded pasture was covered by *Eichhornia crassipes* (water hyacinth) and
175 *Hymenachnae amplexicaulis* (Fig. 1C).

176
177 We conducted four sampling campaigns on the selected sites for soil and gas sampling for three periods of the year
178 which characterise the main climatic variability of the region: wet and hot (17-24 February 2020), dry and cool (17 June
179 2018; 31 May–7 June 2019), and a dry and hot (21-29 Oct 2018; Table 1). We conducted measurements on three days for
180 each land use and ecosystem type (Livesley et al. 2009), except for the first sampling during the dry-cool period of 2018,
181 when only mangroves, saltmarsh and sugarcane were surveyed for one day. The effect of tidal inundation on GHG emissions
182 of mangroves, saltmarsh and freshwater tidal forest was also tested during the cool-dry period of 2019 by measuring GHG
183 emissions during a low (0.7m), and a high tide (2.8m) (tidal range from Lucinda town, Lat: 18° 31' S; 146°23'E). Each of the
184 five sites was sampled during three periods dry-cool (May-September), dry-hot (October-December) and wet-hot (January-
185 April; Table 1). During each time, soil physicochemical properties and GHG fluxes were measured as detailed below.

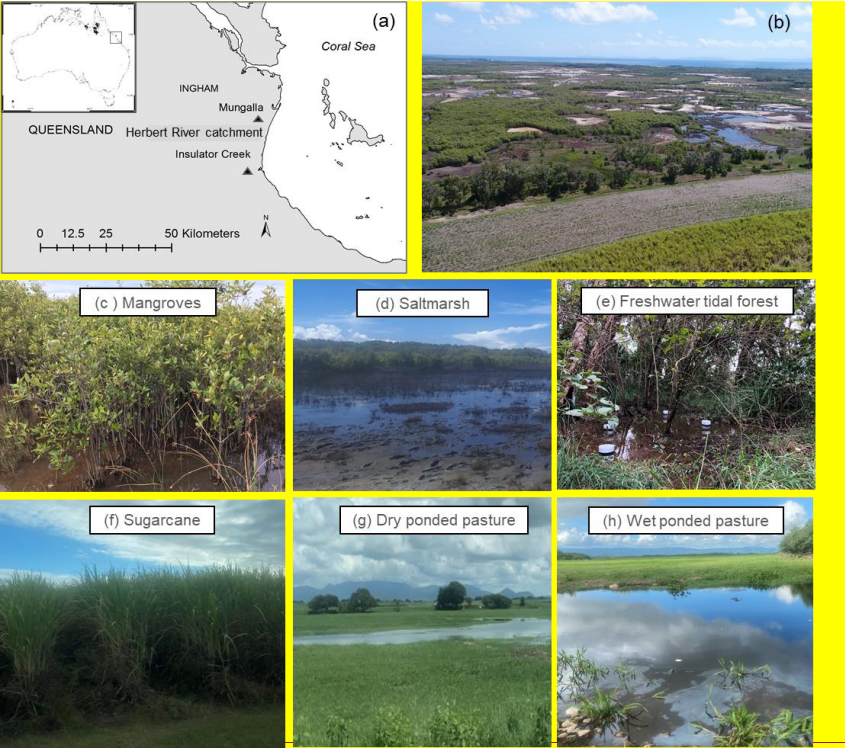
186

187 Table 1. Mean daily air temperature and rainfall at the Ingham weather station during sampling.

<u>Season</u>	<u>Study period</u>	<u>Daily min</u> <u>temperature</u> <u>(°C)</u>	<u>Daily max</u> <u>temperature</u> <u>(°C)</u>	<u>Rainfall</u> <u>(mm d⁻¹)</u>
<u>Dry-cool</u>	<u>17/06/2018</u>	<u>13.4 - 14.6</u>	<u>27.7 - 28.2</u>	<u>0</u>
<u>Dry-hot</u>	<u>23-29/10/2018</u>	<u>15.7 - 21.1</u>	<u>32.2 - 36.2</u>	<u>0</u>
<u>Dry-cool</u>	<u>31/05 to 6/06/2019</u>	<u>10.9 - 17.5</u>	<u>21.6 - 28.2</u>	<u>0-25</u>
<u>Wet-hot</u>	<u>17-22/02/2020</u>	<u>23.9 - 25.3</u>	<u>33.6 - 34.5</u>	<u>0-86</u>

188
189

<u>Season</u>	<u>Study period</u>	<u>Daily min Temp</u> <u>(°C)</u>	<u>Daily max</u> <u>temperature °C</u>	<u>Rainfall</u> <u>(mm d⁻¹)</u>
<u>Wet and hot</u>	<u>17-22 Feb, 2020</u>	<u>23.9 - 25.3</u>	<u>33.6 - 34.5</u>	<u>0-86</u>
<u>Dry and cool</u>	<u>17 June 2018</u>	<u>13.4 - 14.6</u>	<u>27.7 - 28.2</u>	<u>0</u>
<u>Dry and cool</u>	<u>31 May to 6 June 2019</u>	<u>10.9 - 17.5</u>	<u>21.6 - 28.2</u>	<u>0-25</u>
<u>Dry and hot</u>	<u>23-29 Oct 2018</u>	<u>15.7 - 21.1</u>	<u>32.2 - 36.2</u>	<u>0</u>



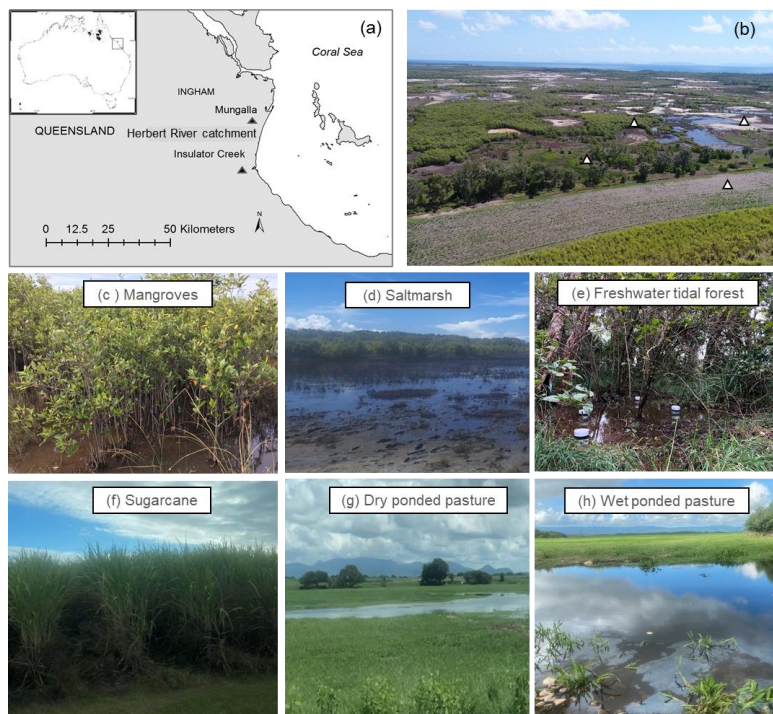


Figure 1: a) Location of sampling sites (Insulator Creek and Mungalla) in the Herbert River catchment, northeast Australia, (b) natural wetlands adjacent to sugarcane farm in Insulator Ck, and (c) mangroves, (d) saltmarsh, (e) freshwater tidal forest, (f) sugarcane, (g) dry ponded pasture and (h) wet ponded pasture. Pictures by N. Iram and MF Adame.

2.2 Soil sampling and analysis of physicochemical properties

Soil physicochemical characteristics were measured by taking composite soil samples ($n = 5$; 0-30 cm) next to each gas chamber location from all study sites during the dry-hot season. The samples were obtained by inserting an open steel corer to a depth of 30 cm; the core was divided into three depths: 0-10cm, 10-20cm and 20-30 cm. Soil samples were dried in the oven at 105 °C for 48 h to determine volumetric water content through gravimetric analysis. The volumetric water content was divided by total soil porosity to determine water-filled pore space (WFPS). Total soil porosity was calculated through the equation: $\text{total soil porosity} = 1 - (\text{soil bulk density} / 2.65)$ where soil particle density was assumed at 2.65 g cm^{-3} (Rashti et al, 2015). Soil texture analysis (% sand, % silt, % clay) was carried for each site following a simplified method for particle

size determination (Kettler et al, 2001). Soil electrical conductivity (EC) and pH were measured using a conductivity meter (WP-84 TPS, Australia) in soil/water slurry at 1:5. Soil subsamples were air-dried, sieved (2mm), ground (Retch™ mill) and analysed for %N and %C using elemental analyser connected to a gas isotope ratio mass spectrometer (EA-Delta V Advantage IRMS, Griffith University). Additionally, soil samples from the top 10 cm were collected in each sampling event to measure gravimetric soil moisture content and bulk density.

2.3 Greenhouse gas fluxes

The static manual gas chambers were used to measure GHG (CO₂, CH₄ and N₂O) fluxes from each site (Hutchinson and Mosier, 1981; Kavehei et al, 2021). The gas chambers were made of high density, round polyvinyl chloride pipe and consisted of two units: a base (r = 12 cm, h = 18 cm) and a detachable collar (r = 12 cm, h = 12 cm). On each sampling date, five chambers were installed at random locations ~ 5cm deep in the soil a day before taking samples to minimise the soil disturbance because of installation (Rashti et al, 2015). The selected locations were carefully observed to avoid including crab burrows. The depth of bases was recorded from five points within each chamber to calculate the headspace volume. At the start of the experiment, gas chambers were closed, and a sample was taken, at time zero with a 20 ml tight syringe and transferred to a 12mL vacuumed exetainers (Exetainer, Labco Ltd., High Wycombe, UK). Samples were collected in the same manner over 1 hour following the sealing of the chamber. The gas samples were collected between 9:00 to 11:00 am, which is the optimum time for minimising the diurnal variation effect on daily GHG emissions (Reeves et al, 2016). Additionally, GHG sampling on this timing minimised the variability in cumulative seasonal fluxes for intermittent manual flux measurements (Wang et al., 2011; Deng et al, 2012; Reeves et al, 2016; Rashti et al, 2015).

A propagation soil thermometer (Gardman, 64704) was inserted at ~5cm depth outside of each chamber 15 minutes before the reading to record the soil temperature for each gas sampling (Kavehei et al, 2020). The GHG concentrations of all samples were analysed within two weeks of sampling with a gas chromatograph (Shimadzu GC 2010 Plus). For N₂O analysis, an electron capture detector was used with helium as carrier gas while CH₄ concentration was analysed on flame ionisation detector with nitrogen as a carrier gas. For CO₂ determination, the gas chromatograph was equipped with a thermal conductivity detector. Peak areas of the samples were compared against standard curves for the determination of GHG concentrations (Chen et al, 2012). The obtained volumetric unit concentrations were converted to mass-based units using the Ideal Gas Law (Hutchinson and Mosier, 1981; Kavehei et al, 2021). Daily cumulative GHG fluxes were calculated by modifying the equation described by Shaaban et al. (2015) as following (Eq. 1);

$$\text{Seasonal cumulative GHG fluxes} = \sum_{i=1}^n (R_i \times 24 \times D_i \times 17.381)$$

where; R_i = Gas emission rate ($\text{mg m}^{-2} \text{hr}^{-1}$ for CO_2 and $\mu\text{g m}^{-2} \text{hr}^{-1}$ for CH_4 and N_2O), D_i = number of the sampling days in a season and 17.38 = number of weeks in each season assuming three seasons prevailed over an annual cycle. Annual cumulative GHG fluxes were calculated by integrating seasonal cumulative GHG fluxes. Total cumulative GHG emissions reported in our research represent $\text{CH}_4 + \text{N}_2\text{O}$ fluxes. It is customary to exclude CO_2 in total GHG fluxes for C projects. For the first sampling period during the hot and dry season (21–29 October 2018), gas samples were collected at 0, 20, 40 and 60 minutes from all chambers to perform linearity test for measuring increase or decrease in the concentration of the gas with time. For subsequent experiments, linearity test was performed on subset chambers for each site (Rashti et al, 2016) and R^2 value of > 0.7 was recorded for all tested samples with a linear trend for CO_2 , CH_4 and N_2O over the experimental period. For comparing GHG effects of CH_4 and N_2O fluxes, CO_2 -equivalent ($\text{CO}_2\text{-eq}$), the measurements in our study only represented soil fluxes, mainly respiration, because the chambers were incubated in the dark, thus do not represent a full budget that would include primary production.

We measured GHG fluxes (CO_2 , CH_4 and N_2O) at each site for three consecutive days during each sampling period except for the dry-cool period of 2018, when mangroves, saltmarsh and sugarcane were surveyed for one day. The sampling was done between 9:00 to 11:00 am, representing the mean daily temperatures, thus, minimising variability of cumulative seasonal fluxes based on intermittent manual flux measurements (Reeves et al, 2016). Additionally, we assessed the variability of our measurements with tidal inundation in mangroves and saltmarsh, which were regularly inundated (~10-30 cm). For this, we measured GHG emissions during a low (0.7m) and a high tide (2.8m; Lucinda, 18° 31' S; 146° 23 'E) in the dry-cool period of 2019. We found that CH_4 fluxes did not significantly vary between the low and high tide within all coastal wetlands. Contrarily, for saltmarsh, CO_2 was taken during the high tide ($1.12 \pm 0.24 \text{ g m}^{-2} \text{ d}^{-1}$) but emitted ($0.69 \pm 0.4 \text{ g m}^{-2} \text{ d}^{-1}$) during the low tide ($F_{1,28} = 20.06$, $p < 0.001$). Finally, for N_2O , fluxes differed in all coastal wetlands, with higher uptakes in the high tide for mangroves ($F_{1,28} = 38.28$, $p < 0.001$; $F_{1,28} = 13.53$, $p = 0.001$) and higher release for saltmarsh ($F_{1,28} = 38.31$, $p < 0.001$) during low tide (Table S4). These results suggested that for CO_2 and N_2O fluxes, there was a probability of variation depending on the time of sampling. Thus, further sampling was conducted only during low tides.

We used static, manual gas chambers made of high-density, round polyvinyl chloride pipe, which consisted of two units: a base ($r = 12 \text{ cm}$, $h = 18 \text{ cm}$) and a detachable collar ($h = 12 \text{ cm}$; Hutchinson and Mosier, 1981; Kavehei et al, 2022). The chambers had lateral holes that could be left covered with rubber bungs at low water levels and left open at high water levels to allow for water movement between sampling events. When the wetlands were inundated for the experiments, we used PVC extensions ($h = 18 \text{ cm}$). Five chambers were set ~ 5cm deep in the soil at random locations one day before sampling to minimise the disturbance of installation during the experiment (Rashti et al, 2015). The chambers were selectively located on soil with minimal vegetation, roots, and crab burrows. We were careful not to tramp around the chambers during installation and sampling. The fact that emissions were not significantly different among days ($p > 0.05$) provided us with confidence that disturbance due to installation was not problematic.

At the start of the experiment, gas chambers were closed. A sample was taken at time zero and then after one hour with a 20 ml syringe and transferred to a 12 mL-vacuumed exetainer (Exetainer, Labco Ltd., High Wycombe, UK). During the dry-hot season, linearity tests of GHG fluxes with time were conducted by sampling at 0, 20, 40 and 60 min (Rashti et al., 2016). For the rest of the experiments, linearity tests were performed in one of the five chambers at each site; R^2 values were consistently above 0.70. During each experiment, soil temperature was measured next to each chamber. At the end of the experiment, the depth of the base was recorded from five points within each chamber to calculate the headspace volume. The obtained volumetric unit concentrations were converted to mass-based units using the Ideal Gas Law (Hutchinson and Mosier, 1981).

The GHG concentrations of all samples were analysed within two weeks of sampling with a gas chromatograph (Shimadzu GC-2010 Plus). For N_2O analysis, an electron capture detector was used with helium as the carrier gas, while CH_4 was analysed on a flame ionisation detector with nitrogen as the carrier gas. For CO_2 determination, the gas chromatograph was equipped with a thermal conductivity detector. Peak areas of the samples were compared against standard curves to determine concentrations (Chen et al., 2012). Seasonal cumulative GHG fluxes were calculated by modifying the equation described by Shaaban et al. (2015; Eq. 2):

$$\text{Seasonal cumulative GHG fluxes} = \sum_{i=1}^n (R_i \times 24 \times D_i \times 17.381)$$

Equation 2

Where;
 R_i = Gas emission rate ($mg\ m^{-2}\ hr^{-1}$ for CO_2 and $\mu g\ m^{-2}\ hr^{-1}$ for CH_4 and N_2O),
 D_i = number of the sampling days in a season,
 17.38 = number of weeks in each period, assuming these conditions were representative of the annual cycle (see Table 1).
 Annual cumulative soil GHG fluxes ($CH_4 + N_2O$) were calculated by integrating cumulative seasonal fluxes. These estimations did not account for soil CO_2 values as our methodology with dark chambers only accounted for emissions from respiration and excluded uptake from primary productivity. The $CO_{2-equivalent}$ (CO_{2-eq}) values were estimated by multiplying CH_4 and N_2O emissions by 25 and 298, respectively (Solomon, 2007), which represent the radiative balance of these gases (Neubauer, 2021).

2.4 Statistical analyses

GHG flux data were analysed for normality through Kolmogorov-Smirnov and Shapiro-Wilk tests. When data were not normal, they were transformed (\log , $1/x$) to comply with the assumptions of normality and homogeneity of variances. Despite transformations, some variables were not normally distributed; thus, the differences between sites and seasons were

300 analysed with the non-parametric Kruskal-Wallis test and Mann-Whitney U Test. The data which met the normality
301 assumptions were analysed for spatial and temporal differences with one-way Analyses of Variance (ANOVA), where site
302 and season were the predictive factors and replicate (gas chamber) was the random factor of the model. Additionally, a
303 Pearson correlation test was run to evaluate the correlation of GHG with measured environmental factors. The data were
304 analysed using a statistical program, SPSS (v25, IBM, New York, USA) and values were presented as mean \pm standard error.

305 **3 Results**

306 **3.1 Soil physicochemical properties**

307 Soil physical and chemical parameters varied significantly among the sites. Gravimetric moisture content was highest in the
308 ponded pasture wet (55%) and lowest in the sugarcane (12%). Freshwater tidal wetlands and saltmarsh had similar moisture
309 content (Table 2). All tested soils were acidic with mangroves having the highest pH value with 6 ± 0.1 , followed by
310 saltmarsh with 5.8 ± 0.1 . The lowest electrical conductivity (EC) was recorded for the ponded pasture with 247 ± 38 and
311 $190\pm39 \mu\text{S cm}^{-1}$ for the dry and wet pasture, respectively. For the top 20 cm soil, the natural wetlands had significantly
312 higher EC (1418 ± 104 , 8049 ± 276 and $8930\pm790 \mu\text{S cm}^{-1}$ for tidal freshwater wetland, saltmarsh and mangroves,
313 respectively) compared to the agricultural land ($190\pm39 \mu\text{S cm}^{-1}$, 247 ± 38 and $382\pm11 \mu\text{S cm}^{-1}$ for wet and dry ponded
314 pasture and sugarcane, respectively).

315 ~~The mean bulk density of the top 30 cm soil of the saltmarsh ($1.4\pm0.1 \text{ g cm}^{-3}$), sugarcane ($1.5\pm0.1 \text{ g cm}^{-3}$) and~~
316 ~~mangroves ($1.9\pm0.1 \text{ g cm}^{-3}$) was similar, while the freshwater tidal wetland had the lowest value ($0.6\pm0.1 \text{ g cm}^{-3}$).~~ Soil bulk
317 density was highest in sugarcane ($1.5 \pm 0.1 \text{ g cm}^{-3}$) and lowest in the freshwater tidal wetland ($0.6 \pm 0.1 \text{ g cm}^{-3}$). For all
318 measured ecosystems, %C was highest in the top 10 cm of the soil and decreased below 10 cm (Table 2). The highest %C
319 was recorded in the freshwater tidal wetland ($5.1\pm0.6\%$) and lowest in the saltmarsh ($1.2\pm0.1\%$). Soil %N ranged from
320 0.1 ± 0.0 to $0.4\pm0.1\%$ at all sites, except in the freshwater tidal wetland, where it reached values of $0.6\pm0.0\%$ in the top 10 cm
321 (Table 2).

322
323
324

325 Table 2. Physicochemical characteristics for the soil of natural wetlands and agricultural land use types for the top 30 cm of
 326 soil in tropical Australia. Fw = Freshwater tidal forest, C= carbon, N = Nitrogen, EC = Electrical Conductivity. Values are
 327 mean ± standard error (5 replicates from each site)

Ecosystem	Depth (cm)	Gravimetric moisture content (%)		pH		EC (µs cm ⁻¹)		Bulk density (g cm ⁻³)		C (%)		N (%)	
		Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE
Mangroves	0-10	41.7	1.1	5.9	0.1	12550	524	1.1	0.1	2.3	0.1	0.2	0.0
	10-20	34.6	0.7	5.9	0.3	12164	905	1.3	0.0	1.7	0.2	0.1	0.0
	20-30	31.3	0.6	6.2	0.1	5560	365	1.9	0.1	0.9	0.1	0.1	0.0
	Mean	35.9	1.2	6.0	0.1	8930	790	1.9	0.1	1.6	0.2	0.1	0.0
Saltmarsh	0-10	25.6	1.2	5.8	0.2	8442	435	1.1	0.0	1.4	0.1	0.1	0.0
	10-20	26.6	0.3	5.8	0.1	8666	437	1.5	0.1	1.3	0.1	0.1	0.0
	20-30	26.4	0.2	5.9	0.3	7040	316	1.6	0.0	1.0	0.3	0.1	0.0
	Mean	26.2	0.4	5.8	0.1	8049	276	1.4	0.1	1.2	0.1	0.1	0.0
Fw tidal forest	0-10	33.4	0.5	4.4	0.2	1099	17	0.5	0.1	7.8	0.1	0.6	0.0
	10-20	24.9	0.6	4.2	0.0	1272	164	0.7	0.0	5.4	0.0	0.5	0.0
	20-30	22.4	0.7	4.2	0.1	1882	47	0.8	0.0	2.2	0.1	0.1	0.0
	Mean	26.9	1.3	4.3	0.1	1418	104	0.6	0.1	5.1	0.6	0.4	0.1
Sugarcane	0-10	9.1	0.4	5.7	0.1	429	12	1.3	0.1	1.5	0.1	0.1	0.0
	10-20	12.1	0.6	5.3	0.3	365	11	1.5	0.1	1.5	0.1	0.1	0.0
	20-30	13.7	0.2	4.7	0.2	351	2	1.6	0.1	1.3	0.1	0.1	0.0
	Mean	11.7	0.6	5.2	0.2	382	11	1.5	0.1	1.4	0.1	0.1	0.0
Dry ponded pasture	0-10	12.4	0.3	4.1	0.0	378	21	0.8	0.1	3.1	0.3	0.3	0.0
	10-20	13.6	0.1	4.4	0.1	279	60	1.2	0.1	1.6	0.4	0.1	0.0
	20-30	14.5	0.7	4.4	0.3	84	4	1.3	0.2	1.6	0.2	0.1	0.0
	Mean	13.5	0.3	4.3	0.1	247	38	1.1	0.1	2.1	0.3	0.2	0.0
Wet ponded pasture	0-10	52.1	0.4	4.8	0.0	358	71	0.6	0.1	3.6	0.3	0.3	0.0
	10-20	47.7	0.4	4.9	0.1	117	11	1.3	0.0	1.7	0.1	0.1	0.0
	20-30	46.4	0.2	5.1	0.1	95	6	1.3	0.0	1.5	0.1	0.1	0.0
	Mean	48.7	0.7	4.9	0.0	190	39	1.1	0.1	2.3	0.3	0.2	0.0

3.2 Greenhouse gas fluxes

Soil emissions for CO₂ were significantly different among sites and times of the year ($t = 155.09$, $n = 237$, $p < 0.001$; Fig. 2a). Highest CO₂ emissions were measured during the wet-hot period in the dry ponded pasture, where values reached $20.31 \pm 1.95 \text{ g m}^{-2} \text{ d}^{-1}$ while the lowest values were measured in the saltmarsh, the only site where the soils were a sink of CO₂ with an uptake rate of $-0.59 \pm 0.15 \text{ g m}^{-2} \text{ d}^{-1}$ (Fig. 2a). In the ponded pastures, CO₂ emissions were higher when dry, with cumulative CO₂ emissions of $5,748 \pm 303 \text{ g m}^{-2} \text{ y}^{-1}$ compared to wet ponded pastures with $2,163 \pm 465 \text{ g m}^{-2} \text{ y}^{-1}$. For wetlands, cumulative annual CO₂ emissions were highest in freshwater tidal forests with $2,213 \pm 284 \text{ g m}^{-2} \text{ y}^{-1}$, followed by mangroves with $1,493 \pm 111 \text{ g m}^{-2} \text{ y}^{-1}$ and lowest at the saltmarsh with uptake rates of $-264 \pm 29 \text{ g m}^{-2} \text{ y}^{-1}$. During high and low tide, emissions of CO₂ in the hot-dry season were similar for mangroves ($F_{1,28} = 2.911$, $p = 0.099$) and freshwater tidal forest ($F_{1,28} = 0.563$, $p = 0.459$; Table 3). However, saltmarsh had significantly different lower CO₂ fluxes during the high tide ($F_{1,28} = 20.060$, $p < 0.001$) with $-1.12 \pm 0.24 \text{ g m}^{-2} \text{ d}^{-1}$ compared to $-0.69 \pm 0.4 \text{ g m}^{-2} \text{ d}^{-1}$ during the low tide (Table 3).

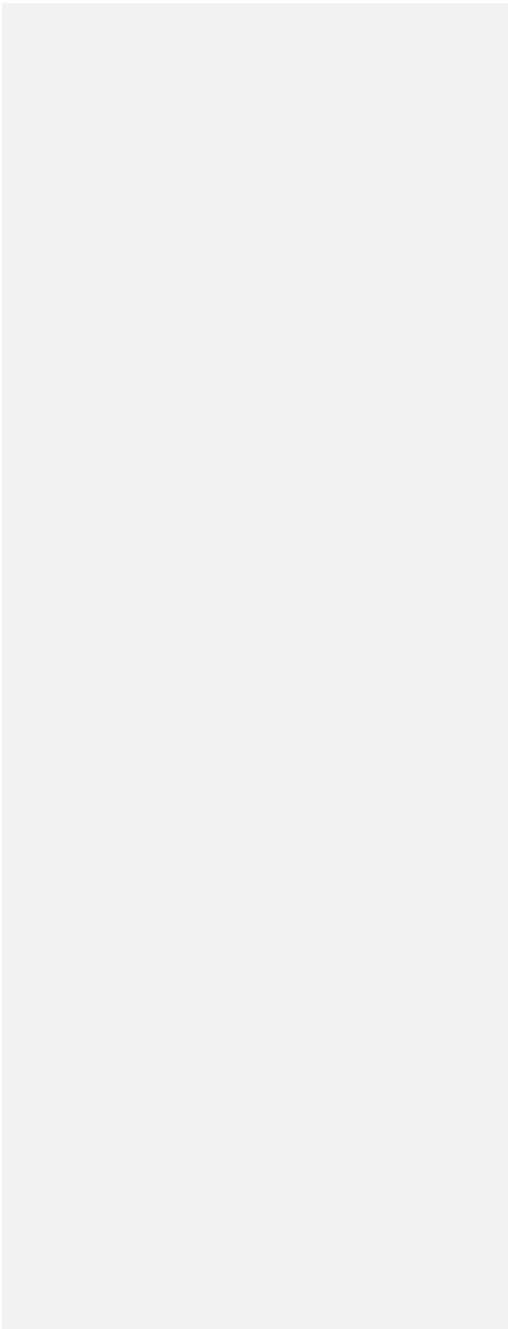
For CH₄ fluxes, significant differences were observed among sites and seasons ($t = 182.33$, $n = 237$, $p < 0.001$). The differences between different sites were substantial, with wet ponded pasture having significantly higher ($p < 0.001$) CH₄ emissions than any site, with rates ~200 times higher throughout the measured period (Fig. 2b). For tidal coastal wetlands, emissions of CH₄ were highest during the wet-hot season in all the sites except for the mangroves, which had similar emissions throughout the year (Fig. 2b). Overall, cumulative annual CH₄ emissions were $209 \pm 36 \text{ g m}^{-2} \text{ y}^{-1}$ for the wet ponded pasture followed by mangroves ($0.73 \pm 0.13 \text{ g m}^{-2} \text{ y}^{-1}$), dry ponded pasture ($0.15 \pm 0.03 \text{ g m}^{-2} \text{ y}^{-1}$), freshwater tidal forest ($0.14 \pm 0.03 \text{ g m}^{-2} \text{ y}^{-1}$), saltmarsh ($0.04 \pm 0.01 \text{ g m}^{-2} \text{ y}^{-1}$) and sugarcane ($-0.04 \pm 0.02 \text{ g m}^{-2} \text{ y}^{-1}$). For tidal coastal wetlands, CH₄ emissions did not differ significantly among mangroves ($F_{1,28} = 1.539$, $p = 0.225$), saltmarsh ($F_{1,28} = 0.007$, $p = 0.934$), and freshwater tidal forest ($F_{1,28} = 2.052$, $p = 0.163$) between low and high tide (Table 3).

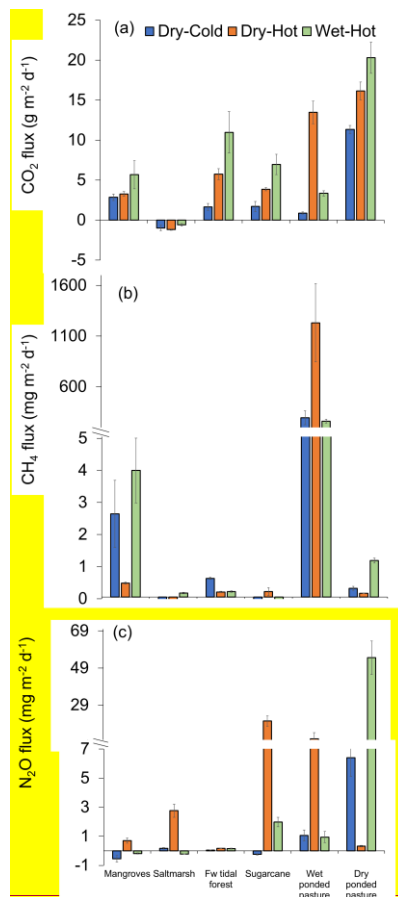
For N₂O fluxes, highest emissions ($55 \pm 9 \text{ mg m}^{-2} \text{ d}^{-1}$) were measured in the dry ponded pasture in the hot-wet season followed by sugarcane ($20 \pm 3 \text{ mg m}^{-2} \text{ d}^{-1}$) during the hot-dry period which coincides with the post-fertilisation period (Fig. 2c). Overall, dry ponded pastures had the highest cumulative annual N₂O emissions ($7.99 \pm 2.26 \text{ mg m}^{-2} \text{ d}^{-1}$), followed by sugarcane ($2.37 \pm 0.68 \text{ mg m}^{-2} \text{ d}^{-1}$), wet ponded pasture ($1.32 \pm 0.33 \text{ mg m}^{-2} \text{ d}^{-1}$), saltmarsh ($0.33 \pm 0.11 \text{ mg m}^{-2} \text{ d}^{-1}$), freshwater tidal forests ($0.04 \pm 0.0 \text{ mg m}^{-2} \text{ d}^{-1}$) and finally, mangroves ($0.02 \pm 0.04 \text{ mg m}^{-2} \text{ d}^{-1}$). However, these differences were only statistically significant when considering the interaction between time of the year and site ($t = 100.21$, $n = 237$, $p < 0.001$). For tidal coastal wetlands, N₂O fluxes for the low versus high tide were significantly different for mangroves ($F_{1,28} = 38.283$, $p < 0.001$) with $-0.74 \pm 0.17 \text{ mg m}^{-2} \text{ d}^{-1}$ of N₂O uptake during high tide compared to $0.15 \pm 0.06 \text{ mg m}^{-2} \text{ d}^{-1}$ release during low tide (Table 3). Saltmarsh showed the opposite trend with significantly higher N₂O uptake ($F_{1,28} = 38.313$, $p < 0.001$) with $-0.15 \pm 0.04 \text{ mg m}^{-2} \text{ d}^{-1}$ during high tide compared to $0.04 \pm 0.01 \text{ mg m}^{-2} \text{ d}^{-1}$ release during low tide (Table 3).

0.001) during low tide (Table 3). Like saltmarsh, the freshwater tidal forest had significantly higher N₂O uptake during low compared to high tide ($F_{1,28} = 13.529, p = 0.001$; Table 3).

The wet pasture had the highest total cumulative soil GHG emissions (CH₄ + N₂O) with 56,124 CO₂eq kg ha⁻¹ y⁻¹ followed by dry pasture 23,890 CO₂eq kg ha⁻¹ y⁻¹ and sugarcane 7,142 CO₂eq kg ha⁻¹ y⁻¹. While coastal wetlands had comparatively very lower total cumulative soil GHG emissions with 884, 235 and 144 CO₂eq kg ha⁻¹ y⁻¹ for saltmarsh, mangroves and freshwater tidal forests, respectively. Overall, the three coastal wetlands measured in this study had lower total cumulative GHG emissions at 1,263 CO₂eq kg ha⁻¹ yr⁻¹ compared to the alternate agricultural land uses, which emitted 87,156 CO₂eq kg ha⁻¹ yr⁻¹.

Formatted: Justified, Indent: First line: 1.27 cm, Line spacing: 1.5 lines





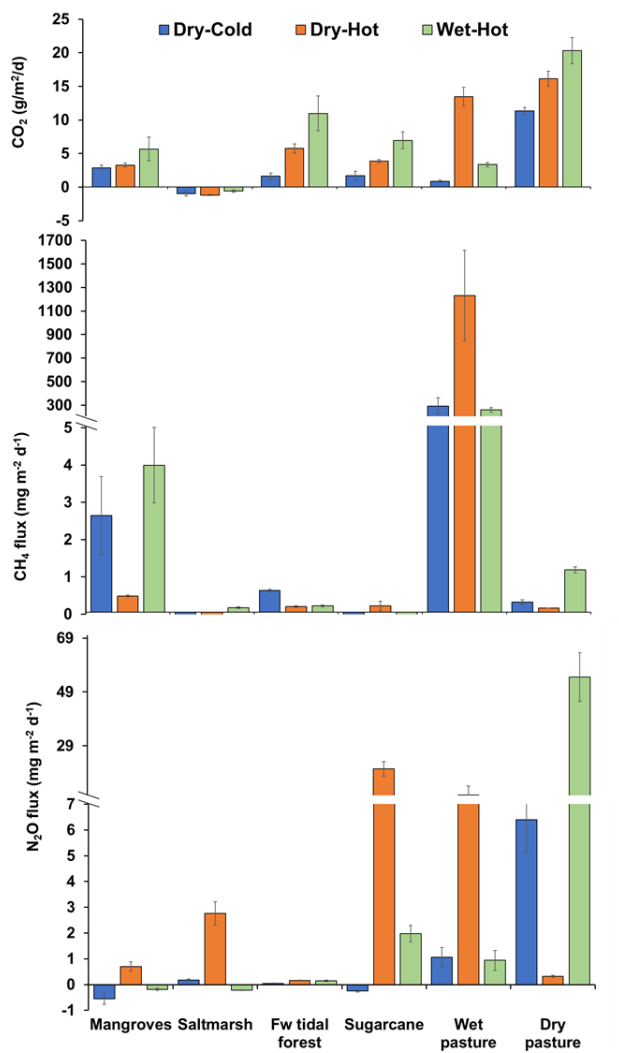


Figure 2: Greenhouse gas fluxes of (a) CO₂ (g m⁻² d⁻¹), (b) CH₄ (mg m⁻² d⁻¹) and (c) N₂O (mg m⁻² d⁻¹) from soils of tropical coastal wetlands (mangroves, saltmarsh, freshwater (Fw) tidal forest and their alternative land uses (sugarcane and ponded pastures) during three periods of the year: dry-cold, dry-hot and wet-hot

380

Table 3. Greenhouse gas (GHG) fluxes from soils of tropical coastal wetlands: mangroves, saltmarsh, and freshwater (Fw) tidal forest during high and low tide during a dry-hot season

GHG	Land-use type	High tide		Low tide	
		Mean	SE	Mean	SE
CO ₂ (g m ⁻² d ⁻¹)	Mangroves	2.55	0.37	3.25	0.57
	Saltmarsh	1.12	0.24	0.69	0.40
	Fw tidal forest	2.97	1.35	5.35	2.68
CH ₄ (mg m ⁻² d ⁻¹)	Mangroves	3.38	0.98	236	73
	Saltmarsh	-0.13	0.06	-25	6
	Fw tidal forest	1.10	0.52	457	108
N ₂ O (mg m ⁻² d ⁻¹)	Mangroves	-0.74	0.17	0.15	0.06
	Saltmarsh	0.19	0.06	-0.14	0.04
	Fw tidal forest	0.06	0.01	-0.25	0.16

3.3 Greenhouse gas emissions and environmental factors

385

Overall, we found not one single parameter could explain GHG emissions from all sites except land-use. The CO₂ emissions were not significantly correlated to bulk density ($R^2=0.026$ $p=0.918$ $n=18$), % WFPS ($R^2=-0.003$ $p=0.99$ $n=18$), or soil temperature ($R^2=0.296$ $p=0.233$, $n=18$). Similarly, soil CH₄ emissions were not correlated with bulk density ($R^2=-0.096$ $p=0.706$ $n=18$), % WFPS ($R^2=0.224$ $p=0.372$, $n=18$) or soil temperature ($R^2=0.286$ $p=0.25$ $n=18$). Finally, no correlation was found between N₂O emissions and bulk density ($R^2=-0.349$ $p=0.156$ $n=18$), % WFPS ($R^2=-0.34$ $p=0.168$ $n=18$), or soil temperature ($R^2=-0.241$ $p=0.335$ $n=18$). [See full raw dataset at Table 1S and S4.](#)

Formatted: Not Highlight

390 4 Discussion

395

Across the agricultural land-use types in our study area, very high GHG emissions were measured. The highest emitter was by far the wet-ponded pastures, with CH₄ emissions that were 200 times higher than any other measured land uses. The dry pasture was also a high emitter of CO₂ at 20.31 ± 1.95 g m⁻² d⁻¹ and of N₂O at 55 ± 9 mg m⁻² d⁻¹, especially during hot periods. Natural wetlands had significantly lower CH₄ emissions, with saltmarshes having the lowest. Overall, natural wetlands had very low total cumulative GHG emissions at 1.263 CO_{2-eq} kg ha⁻¹ yr⁻¹ compared to their alternate agricultural land uses, which emitted 87.156 CO_{2-eq} kg ha⁻¹ yr⁻¹. These results confirm our hypothesis that coastal tidal wetlands, even freshwater ones, can be a viable land use to reduce GHG emissions from current agricultural land.

In this study, we found that the three coastal tropical wetlands measured in this study (mangroves, saltmarshes and freshwater tidal forests) had significantly lower GHG emissions compared to two alternative land uses common in tropical Australia (sugar cane and grazing pastures). Notably, we found that coastal wetlands had 200 times lower CH₄ emissions and seven times lower N₂O compared to wet pastures and sugarcane soils, respectively. While future studies should measure GHG from other wetlands, land uses, and within other tropical regions, these results support the idea that the management or conversion of unused agricultural land could be converted to coastal wetlands could result in significant GHG mitigation.

The GHG emissions varied with season, with an overall increase in emissions during the hottest and wettest time of the year. The emissions of CO₂ and N₂O were highest when temperatures were > 38°C. Similar results have been shown in terrestrial forests, where N₂O emissions increased with temperature, explaining 86% of the flux variations (Schindlbacher et al., 2004). Emissions of CO₂ were associated with temperature, although the correlation was not significant, meaning that differences in temperature within season would affect emissions. Highest CO₂ emissions from all land-use types during the early wet season could be attributed to "Birch effect" which refers to short term but a substantial increase of respiration from soils under the effect of precipitation during early wet season (Fernandez-Bou et al., 2020). These findings are in accordance with recent studies which reported that CO₂ fluxes from subtropical mangroves were largest when the temperature was highest, and in periods of the year with reduced salinity (Liu and Lai 2019). The soil from mangroves within our study sites were always sources of CO₂, partly because the incubation was done in dark chambers.

The variability of GHG fluxes was best explained by land use and wetland type; however, some trends with seasons were evident. For instance, CO₂ and N₂O emissions were lowest during the dry-cool periods. Reduced emissions at low temperatures are expected as the temperature is a main driver of any metabolic process, including respiration and nitrification-denitrification. Mangroves tend to have higher CO₂ emissions as temperature increases (Liu and Lai 2019), and terrestrial forests have significantly higher N₂O emissions during warm seasons (Schindlbacher et al., 2004). Emissions of CH₄ also tend to increase with temperature as the activity of soil methane-producing microbes (Ding et al., 2004) and the availability of carbon is higher in warmer conditions (Yvon-Durocher et al., 2011). However, as most of the studies on GHG fluxes, were conducted in temperate and subtropical locations where differences in temperature throughout the year are much larger than those in tropical regions. For tropical regions, increased GHG emissions are likely to be strongly affected by the "Birch effect", which refers to short-term but a substantial increase of respiration from soils under the effect of precipitation during the early wet season (Fernandez-Bou et al., 2020).

Emissions of CH₄ were not significantly associated with the environmental factors measured in this study. However, CH₄ emissions were highest during the hot-dry season. Increased CH₄ emissions with temperature could be attributed to the increase of the activity of methane-producing microbes in the soil (Ding et al., 2004). Additionally, high temperatures increase plant growth, providing the substrate for CH₄ production by root exudates (Yvon-Durocher et al., 2011). The

relatively low CH₄ emissions from all the natural wetlands could be attributed to the presence of terminal electron acceptors like iron, sulphate, manganese and nitrate which result in low rates of methanogenesis (Fumoto et al, 2008; Kögel-Knabner et al, 2010; Sahrawat, 2004). For example, sulphate-reducing bacteria outcompete methane-producing bacteria (methanogens) in the presence of high sulphate concentrations in tidal wetlands, resulting in low CH₄ production. Additionally, competition between methanogens and methanotrophs (CH₄-consuming bacteria), could result in a net balance of low CH₄ production despite freshwater conditions (Maietta et al. 2020).

The variability of GHG fluxes was best explained by land use and wetland type; however, some trends with seasons were evident. For instance, CO₂ and N₂O emissions were lowest during the dry-cool periods. Reduced emissions at low temperatures are expected as the temperature is a main driver of any metabolic process, including respiration and nitrification-denitrification. Mangroves tend to have higher CO₂ emissions as temperature increases (Liu and Lai 2019), and terrestrial forests have significantly higher N₂O emissions during warm seasons (Schindlbacher et al, 2004). Emissions of CH₄ also tend to increase with temperature as the activity of soil methane-producing microbes (Ding et al, 2004) and the availability of carbon is higher in warmer conditions (Yvon-Durocher et al, 2011). However, as most of the studies on GHG fluxes, were conducted in temperate and subtropical locations where differences in temperature throughout the year are much larger than those in tropical regions. For tropical regions, increased GHG emissions are likely to be strongly affected by the “Birch effect”, which refers to short-term but a substantial increase of respiration from soils under the effect of precipitation during the early wet season (Fernandez-Bou et al, 2020).

The main factor associated with GHG fluxes was land use and type of wetland. Notably, coastal wetlands, even the freshwater tidal forests, had much lower emissions compared to the wet pastures. This large difference could be attributed to the presence of terminal electron acceptors in the soils (e.g. iron, sulphate, manganese) of the coastal wetlands, which could inhibit methanogenesis (Kögel-Knabner et al, 2010; Sahrawat, 2004). Sulphate-reducing bacteria are also likely to outcompete methane-producing bacteria (methanogens) in the presence of high sulphate concentrations in tidal wetlands, resulting in low CH₄ production. Competition between methanogens and methanotrophs may result in a net balance of low CH₄ production despite freshwater conditions (Maietta et al. 2020). Additionally, microorganism living within the bark of *Melaleuca* trees can consume CH₄ (Jeffrey et al, 2021), so it is possible that similar bacteria within the soil could reduce CH₄ emissions. Interestingly, variability within CH₄ fluxes among sites was very high, despite them being very close to each other (Fig. 1b). These differences highlight the importance of land use in GHG fluxes, which are likely to significantly alter the microbial community composition and abundance, which can change rapidly over small spatial scales (Martiny et al, 2006; Drenovsky et al, 2009).

Our results are consistent with other studies, which have shown the importance of land use in GHG emissions. For instance, in a Mediterranean climate, drained agricultural land use types, pasture and corn, were larger CO₂ emitters compared to restored wetlands (Knox et al. 2015). Clearing of wetlands for agricultural development, such as the drainage of peatlands, results in very high CO₂ emissions (Nieveen et al. 2005; Veenendaal et al. 2007; Hirano et al. 2012), and restoration of these wetlands could decrease these emissions (Cameron et al. 2020). Additionally, some of the wetland types, such as marshes, were occasional sinks of CO₂ and CH₄, consistent with previous studies where intertidal wetlands sink of GHG at least under some conditions or during some times of the year (Knox et al. 2015; Maher et al. 2016).

We found high variability in CH₄ emissions within land use types that were very close to each other (Fig. 1b). These differences might be attributed to the microbial community composition and abundance, which can change rapidly over small spatial scales (Martiny et al. 2006; Drenovsky et al. 2010). In general, emissions of CH₄ were best explained by land use type. Our study is consistent with the recent findings, which reported very high variations in GHG fluxes from mature, converted, and restored land use types for mangroves (Cameron et al. 2020). Our findings also corroborate studies on GHG emissions in a Mediterranean climate, where drained agricultural land use types, pasture and corn were the largest CO₂ emitters compared to restored wetlands (Knox et al. 2015). Our findings are also in agreement with other studies which reported drained, and degraded peatlands were large net sources of CO₂ (Nieveen et al. 2005; Veenendaal et al. 2007; Hirano et al. 2012). Overall, our study provides evidence that despite GHG emissions from wetlands, these are lower than for some agricultural land uses. Additionally, some of the wetland types, such as marshes were consistently found to be sinks of CO₂ and CH₄. This has been previously reported, with studies finding that some intertidal wetlands can be sinks of GHG at least under some conditions or during some times of the year (Knox et al. 2015; Maher et al. 2016).

The GHG emissions from wetlands have an extensive range. For CO₂ fluxes, they can range between -139 and 22,000 mg m⁻² d⁻¹ (Stadmark and Leonardson 2005; Morse et al. 2012), for CH₄, from -1 to 418 mg m⁻² d⁻¹ (Allen et al. 2007; Mitsch et al. 2013; Cabezas et al. 2018), and for N₂O, from -0.3 to 3.9 mg m⁻² d⁻¹ (Hernandez and Mitsch 2006; Morse et al. 2012). The GHG fluxes measured in this study are within the lower end, with ranges from -1191 to 10,970 mg m⁻² d⁻¹ for CO₂, from -0.2 to 3.9 mg m⁻² d⁻¹ for CH₄, and -0.2 to 2.8 mg m⁻² d⁻¹ for N₂O.

The fluxes measured in the coastal wetlands of this study (-1,191 to 10,970 mg m⁻² d⁻¹ for CO₂, -0.2 to 3.9 mg m⁻² d⁻¹ for CH₄, and -0.2 to 2.8 mg m⁻² d⁻¹ for N₂O) are within the range of those measured in other wetlands, worldwide. For CO₂, fluxes can range between -139 and 22,000 mg m⁻² d⁻¹ (Stadmark and Leonardson 2005; Morse et al. 2012), for CH₄, from -1 to 418 mg m⁻² d⁻¹ (Allen et al. 2007; Mitsch et al. 2013; Cabezas et al. 2018), and for N₂O, from -0.3 to 3.9 mg m⁻² d⁻¹ (Hernandez and Mitsch 2006; Morse et al. 2012). Despite being in tropical regions, the fluxes from this study were not notably higher compared to wetlands in other climates. The general lower nitrogen pollution in Australia's soils and waterways compared to other countries may partially explain the lower emissions. However, the GHG flux measurements from this study did not account for the effects of vegetation, which can alter fluxes. For instance, some plant species of rice

500 paddies (Timilsina et al., 2020) and *Miscanthus sinensis* (Lenhart et al., 2019) can increase N₂O emissions, and some tree species can facilitate CH₄ efflux from the soil (Pangala et al. 2013). Finally, changes in emissions between low and high tides were detected for CO₂ and N₂O. Thus, future studies that include vegetation and changes within tidal cycles will improve GHG flux estimates for coastal wetlands.

4.1 Management implications

505 Under the Paris Agreement, Australia has committed to reducing GHG emissions 26 - 28% below its 2005 levels by 2030. Any GHG mitigation strategy should be based on robust GHG flux quantification from different land-use scenarios. With annual emissions of 153.0 million tonnes of carbon dioxide equivalent (Mt CO_{2-eq} y⁻¹), Queensland is a major GHG emitter in Australia (~ 28.7% of the total in 2016; www.stateoftheenvironment.des.qld.gov.au). Of these emissions, about 18.3 Mt CO_{2-eq} y⁻¹ (14%) are attributed to agriculture, while land-use change, and forestry emitting another 12.1 Mt CO_{2-eq} y⁻¹ (www.stateoftheenvironment.des.qld.gov.au). (DES, 2016). Production of CH₄ from ruminant animals, mostly cattle, contributed 82% of agriculture-related emissions (www.daf.qld.gov.au). (DES, 2016).

510 This study provides evidence for three management actions to reduce GHG emissions. The conversion of agricultural lands both ponded pastures and sugarcane to intertidal wetlands including mangroves, marshes and freshwater tidal wetlands seems to be a promising land use change management option for GHG mitigation. This study supports the application of three management actions that could reduce GHG emissions. First, the conversion of ponded pastures to coastal wetlands is likely to reduce soil GHG emissions. Our results showed that wet pastures emit 56 ton CO_{2-eq} ha⁻¹ y⁻¹ of total GHG (CH₄ + N₂O) compared with 0.2 ton CO_{2-eq} ha⁻¹ y⁻¹, 0.1 ton CO_{2-eq} ha⁻¹ y⁻¹ and 0.9 ton CO_{2-eq} ha⁻¹ y⁻¹ from mangroves, freshwater tidal forest, and saltmarshes, respectively. Our results showed that wet and dry ponded pastures emitted 56 CO_{2-eq} ton ha⁻¹ y⁻¹ and 24 CO_{2-eq} ton ha⁻¹ y⁻¹ of total GHG (CH₄ + N₂O) compared with 0.2 CO_{2-eq} ton ha⁻¹ y⁻¹, 0.1 CO_{2-eq} ton ha⁻¹ y⁻¹ and 0.9 CO_{2-eq} ton ha⁻¹ y⁻¹ from mangroves, freshwater tidal forest and saltmarshes respectively. This implies that up to 56 CO_{2-eq} ton ha⁻¹ y⁻¹ emissions could be potentially avoided by restoring wet ponded pastures to coastal wetlands. Furthermore, financial incentives through the inclusion of restored agricultural lands in C markets could be an alternative income source for farmers. For instance, our results suggested that wet pasture landowners could get ~AUD \$ 860 ha⁻¹yr⁻¹ assuming carbon value of AUD \$15.37 per ton of CO_{2-eq} (Australian Emission Reduction Fund, December 2018). The carbon mitigation for GHG emissions from soil solely could provide ~ AUD 860 ha⁻¹ yr⁻¹, assuming a carbon value of AUD 15.37 per ton of CO_{2-eq} (Australian Government Clean Energy Regulator, 2018). This mitigation could be added up to the carbon sequestration through sediment accumulation and tree growth that results from wetland restoration. Recent studies suggested that legal enablers are in place for the conversion of ponded pastures to tidal wetlands (Bell-James and Lovelock 2019) supporting our recommendation for inclusion of these ecosystems in carbon mitigation programs. However, actual GHG mitigation gains could be even greater, if accounted for the carbon sequestration in the vegetation and soil after wetland restoration.

525

530

Formatted: Not Highlight

Formatted: Not Highlight

Another management option would be to reducing the area of wet pastures to minimise CH₄ emissions. For example, our study showed that dry pastures produced significantly less CH₄ ~0.005 kg ha⁻¹ d⁻¹ than wet pastures 6 kg ha⁻¹ d⁻¹. Assuming the average cow produces 141 g CH₄ per day (McGinn et al, 2004) and our study area supported around 900/2500 ha cattle throughout the year, cattle would be responsible for only 0.05 kg ha⁻¹ d⁻¹ of CH₄ emissions. This means that nearly 99% of the CH₄ emissions came from the wet pastures while dry pasture and grazing cattle had a low share in total CH₄ emissions. Therefore, land use management of wet pastures which are used to feed grazing cattle in Queensland may be a significant opportunity to reduce agriculture-related CH₄ emissions.

A second management option would be to reduce the time pastures are kept under water. Dry pastures produced significantly less CH₄ with ~0.005 kg ha⁻¹ d⁻¹ than wet pastures with 6 kg ha⁻¹ d⁻¹. For comparison, an average cow produces 141 g CH₄ d⁻¹ (McGinn et al, 2004), and our study area supported around 900 cattle over 2,500 ha throughout the year, equivalent to 19 kg ha⁻¹ y⁻¹ compared to 2 kg ha⁻¹ y⁻¹ and 2090 kg ha⁻¹ y⁻¹ CH₄ from dry and wet pasture respectively. This implies that nearly 99% of the CH₄ emissions came from wet pastures, while dry pasture and grazing cattle had a low share in total CH₄ emissions. Therefore, land use management of wet pastures which are used to feed grazing cattle in Queensland may be a significant opportunity to reduce agriculture-related CH₄ emissions. Future studies should increase the number of sites of ponded pastures to account for variability in hydrology, fertilisation, and cattle use. However, the very high difference (2-3 orders of magnitude) between dry and ponded pastures provides confidence that pasture management could provide significant GHG mitigation throughout the year.

Finally, fertiliser management in sugarcane could reduce N₂O emissions. Higher N₂O emissions of 17.63 mg m⁻² d⁻¹ were measured in sugarcane following fertilisation, during the dry-hot season. Comparatively, natural wetlands had low N₂O emissions (0.16 to 2.79 mg m⁻² d⁻¹), even with the saltmarsh being an occasional sink. Thus, improved management of fertiliser applications could result in GHG emission mitigation through reduced N₂O emissions. Some activities include split application of nitrogen fertiliser in combination with low irrigation, reduction in fertiliser application rates, substitution of nitrate-based fertiliser for urea (Rashti et al, 2015), removing mulch layer before fertiliser application (Pinheiro et al, 2019; Xu et al, 2019) or conversion of unproductive sugarcane to coastal wetlands.

5 Conclusion

To our best knowledge, this is the first study to report GHG emissions mitigation gains from a wide range of tropical natural wetlands including mangroves, saltmarshes and freshwater tidal wetlands and their alternative agricultural land use types. The significantly higher emissions from agricultural land use types as compared to tropical coastal wetlands were most likely due to land use as any other physical or chemical factors could not explain the differences observed. Restoration of wet ponded pastures and sugarcane to coastal tidal wetlands, even freshwater tidal forests, has the potential to mitigate total GHG emissions (CH₄ + N₂O) derived from agricultural activities. The GHG emissions from ponded pastures were particularly

565 alarming, with values 200 fold than any other land use. However, these massive emissions provide an opportunity to target
ponded pastures for consideration in GHG mitigation programs, which could deliver financial incentives for farmers, and
many co-benefits derived from coastal wetland restoration. Overall, the data from this study will contribute significantly to
our understanding of land use effects on GHG budgets in Australia, and in similar tropical regions around the world.
The GHG emissions from three coastal wetlands in tropical Australia (mangroves, saltmarsh and freshwater tidal forests)
were consistently lower than those from two common agricultural land use of the region (sugarcane and pastures) throughout
three climatic conditions (dry-cool, dry-hot and wet-hot). Pondered pastures, which emitted 200 times more CH₄, and
570 sugarcane emitted seven times more than any natural coastal wetland. If these high emissions are persistent in other locations
and within other tropical regions, conversion of pastures and sugarcane to similar coastal wetlands could provide significant
GHG mitigation. As nations try to reach their emission reduction targets, projects aimed at converting or restoring coastal
wetland can financially benefit farmers and provide additional co-benefits derived from coastal wetland restoration.

575 **Author contribution**

Iram, N. and M.F. Adame designed the project, Iram, N, B. Shahrabi Farahani and E. Kavehei carried out experiments, Iram,
N., E. Kavehei and M.F. Adame analysed the data. Iram, N prepared the manuscript with contributions from D.T. Maher,
S.E., Bunn, M. Rezaei Rashti and M.F. Adame.

580 **Competing interests**

The authors declare that they have no conflict of interest.

Acknowledgements

We acknowledge the Traditional Owners of the land in which the field study was conducted, especially the Nywaigi people
585 from Mungalla Station, where this study was conducted. We are also thankful to Sam and Santo Lamari for allowing us to
work in their property and for sharing their local knowledge. We are thankful to Charles Cadier and Julieta Gamboa for their
contribution in the field. This project was financially supported by an Advance Queensland Industry Research Fellowship to
MF Adame.

590 **References**

Allen, D.E., Dalal, R.C., Rennenberg, H., Meyer, R.L., Reeves, S., and Schmidt, S.: Spatial and temporal variation of nitrous oxide and methane flux between subtropical mangrove sediments and the atmosphere, *Soil Biol. Biochem.*, 39, 622–631, <https://doi.org/10.1016/j.soilbio.2006.09.013>, 2007.

Angle, J. C., Morin, T. H., Solden, L. M., Narrowe, A. B., Smith, G. J., Borton, M. A., Rey-Sanchez, C., Daly, R. A.,
595 Mirfenderesgi, G., Hoyt, D. W., Riley, W. J., Miller, C. S., Bohrer, G., and Wrighton, K. C.: Methanogenesis in oxygenated soils is a substantial fraction of wetland methane emissions, *Nat. Commun.*, 8,1-9, <https://doi.org/10.1038/s41467-017-01753-4>, 2017.

Armeth, A., Sitch, S., Bondeau, A., Butterbach-Bahl, K., Foster, P., Gedney, N., de Noblet-Ducoudré, N., Prentice, I. C., Sanderson, M., Thonicke, K., Wania, R., and Zaehle, S.: From biota to chemistry and climate: towards a comprehensive
600 description of trace gas exchange between the biosphere and atmosphere, *Biogeosciences*, 7, 121–149, <https://doi:10.5194/bg-7-121-2010>, 2010.

Australian Government Bureau of Meteorology: <http://www.bom.gov.au/qld/flood/brochures/herbert/herbert.shtml>, last access: 25 March 2020.

Australian Government Bureau of Meteorology: <http://www.bom.gov.au/>, last access: 24 March 2020.

605 Australian Government Clean Energy Regulator: <http://www.cleanenergyregulator.gov.au/Infohub/Markets/buying-accus/australian-carbon-credit-unit-market-updates/december-2018#f4>, last access: 6 February 2021, 2020.

Baldock, J. A., Wheeler, I., McKenzie, N., and McBratney, A.: Soils and climate change: potential impacts on carbon stocks and greenhouse gas emissions, and future research for Australian agriculture, *Crop Pasture Sci.*, 63, 269-283, <http://dx.doi.org/10.1071/CP11170>, 2012.

610 Bauza, J. F., Morell, J. M., and Corredor, J. E.: Biogeochemistry of nitrous oxide production in the red mangrove (*Rhizophora mangle*) forest sediments, *Estuar. Coast. Shelf S.*, 55, 697-704, <https://doi.org/10.1006/ecss.2001.0913>, 2002.

Bell-James, J. and Lovelock, C. E.: Legal barriers and enablers for reintroducing tides: an Australian case study in reconverting ponded pasture for climate change mitigation, *Land Use Policy*, 88, 1-9, <https://doi.org/10.1016/j.landusepol.2019.104192>, 2019.

615 Beringer, J., Livesley, S. J., Randle, J., Hutley, L. B.: Carbon dioxide fluxes dominate the greenhouse gas exchanges of a seasonal wetland in the wet–dry tropics of northern Australia, *Agr. Forest Meteorol.*, 182-183, 239–247, <https://doi.org/10.1016/j.agrformet.2013.06.008>, 2013.

Beringer, J., Livesley, S.J., Randle, J., and Hutley, L. B.: Carbon dioxide fluxes dominate the greenhouse gas exchanges of a seasonal wetland in the wet–dry tropics of northern Australia, *Agr. Forest Meteorol.*, 182-183, 239-247,
620 <https://doi.org/10.1016/j.agrformet.2013.06.008>, 2013.

Burden, A., Garbutt, R. A., Evans, C. D., Jones, D. L., and Cooper, D. M.: Carbon sequestration and biogeochemical cycling in a saltmarsh subject to coastal managed realignment, *Estuar. Coast. Shelf S.*, 120, 12-20, <https://doi.org/10.1016/j.ecss.2013.01.014>, 2013.

Cabezas, A., Mitsch, W. J., MacDonnell, C., Zhang, L., Bydalek, F., and Lasso, A.: Methane emissions from mangrove soils in hydrologically disturbed and reference mangrove tidal creeks in southwest Florida, *Ecol. Eng.*, 114, 57-65, <https://doi.org/10.1016/j.ecoleng.2017.08.041>, 2018.

Cameron, C., Hutley, L. B., Munksgaard, N. C., Phan, S., Aung, T., Thinn, T., Aye, W. M., and Lovelock, C. E.: Impact of an extreme monsoon on CO₂ and CH₄ fluxes from mangrove soils of the Ayeyarwady Delta, Myanmar, *Sci. Total Environ.*, 760, 1-11, <https://doi.org/10.1016/j.scitotenv.2020.143422>, 2021.

Capocci, M., Barba, J., Seyfferth, A. L., and Vargas, R.: Experimental influence of storm-surge salinity on soil greenhouse gas emissions from a tidal salt marsh, *Sci. Total Environ.*, 686, 1164-1172, <https://doi.org/10.1016/j.scitotenv.2019.06.032>, 2019.

Chen, G.C, Tam, N. F.Y., and Yong, Y.: Spatial and seasonal variations of atmospheric N₂O and CO₂ fluxes from a subtropical mangrove swamp and their relationships with soil characteristics, *Soil Biol. Biochem.*, 48, 175-181, <https://doi.org/10.1016/j.soilbio.2012.01.029>, 2012.

Ciais, P., Sabine, C., Bala, G., Bopp, L., Brovkin, V., Canadell, J., Chhabra, A., DeFries, R., Galloway, J., Heimann, M., Jones, C., Le Quéré, C., Myneni, R. B., Piao, S., and Thornton, P.: Carbon and other biogeochemical Cycles, In: *Climate change 2013: The physical science basis, Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*, Cambridge University Press, Cambridge, United Kingdom and New York, United States, 105 pp., 2013.

Cobb, A., Agus, F., Warren, M., Applegate, G., Ryan, Z., Engel, V., Handayani, E. P., Hooijer, A., Husen, E., Jauhiainen, J., Kawaroe, M., Kusmana, C., Naito, R., Osaki, M.: Greenhouse gas fluxes and flux changes from land-use dynamics in tropical wetlands, in: *Tropical Wetlands for Climate Change Adaptation and Mitigation: Science and Policy Imperatives with Special Reference to Indonesia*, edited by: Murdiyarso, D., Kauffman, J. B., Warren, M., Pramova, E., and Hergoualc'h, K., Centre for International Forestry Research (CIFOR), Bogor, Indonesia, 5-9, Working Paper No. 91, https://www.cifor.org/publications/pdf_files/WPapers/WP91Murdiyarso.pdf, 2012.

Conrad, R.: The global methane cycle: recent advances in understanding the microbial processes involved, *Env. Microbiol. Rep.*, 1, 285-292, <https://doi.org/10.1111/j.1758-2229.2009.00038.x>, 2009.

Deng, J., Zhou, Z., Zheng, X., Liu, C., Yao, Z., Xie, B., Cui, F., Han, S., and Zhu, J.: Annual emissions of nitrous oxide and nitric oxide from rice-wheat rotation and vegetable fields: a case study in the Tai-Lake region, China, *Plant Soil*, 360, 37-53, <https://doi.org/10.1007/s11104-012-1223-6>, 2012.

Ding, W., Cai, Z., and Tsuruta, H.: Cultivation, nitrogen fertilization, and set-aside effects on methane uptake in a drained marsh soil in Northeast China, *Glob. Change Biol.*, 10, 1801-1809, <https://doi.org/10.1111/j.1365-2486.2004.00843.x>, 2004.

Drenovsky, R. E., Steenwerth, K. L., Jackson, L. E., and Scow, K. M.: Land use and climatic factors structure regional patterns in soil microbial communities, *Global Ecol. Biogeogr.*, 19, 27-39, <https://doi.org/10.1111/j.1466-8238.2009.00486.x>, 2009.

Duarte, C. M., Losada, I. J., Hendriks, I. E., Mazarrasa, I., and Marba, N.: The role of coastal plant communities for climate change mitigation and adaptation, *Nat. Clim. Change*, 3, 961–968, <https://doi.org/10.1038/nclimate1970>, 2013.

Fernandez-Bou, A.S., Dierick, D., Allen, M. F., Harmon, T. C.: Precipitation-drainage cycles lead to hot moments in soil carbon dioxide dynamics in a Neotropical wet forest, *Glob. Change Biol.*, 26, 5303–5319, <https://doi.org/10.1111/gcb.15194>, 2020.

Flato, G., Marotzke, J., Abiodun, B., Braconnot, P., Chou, S. C., Collins, W., Cox, P., Driouech, F., Emori, S., Eyring, V., Forest, C., Gleckler, P., Guilyardi, E., Jakob, C., Kattsov, V., Reason, C., Rummukainen, M.: Evaluation of climate models, In: *Climate change 2013: The physical science basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*, Cambridge University Press, Cambridge, United Kingdom and New York, United States, 125 pp, 2013.

Fumoto, T., Kobayashi, K., Li, C., Yagi, K., and Hasegawa, T.: Revising a process-based biogeochemistry model (DNDC) to simulate methane emission from rice paddy fields under various residue management and fertilizer regimes, *Glob. Change Biol.*, 14, 382–402, <https://doi.org/10.1111/j.1365-2486.2007.01475.x>, 2007.

Griggs, P. D. (2018). Too much water: drainage schemes and landscape change in the sugar-producing areas of Queensland, 1920–90. *Australian Geographer*, 49(1), 81-105, <https://doi.org/10.1080/00049182.2017.1336965>. 2018.

Grover, S. P. P., Livesley, S. J., Hutley, L. B., Jamali, H., Fest, B., Beringer, J., Butterbach-Bahl, K., and Arndt, S. K.: Land use change and the impact on greenhouse gas exchange in north Australian savanna soils, *Biogeosciences*, 9, 423-437, <https://doi.org/10.5194/bg-9-423-2012>, 2012.

Güttelein, A., Gerschlauser, F., Kikoti, I., and Kiese, R.: Impacts of climate and land use on N₂O and CH₄ fluxes from tropical ecosystems in the Mt. Kilimanjaro region, Tanzania, *Glob. Change Biol.*, 24, 1239–1255, <https://doi.org/10.1111/gcb.13944>, 2017.

Harris, E., Ladreiter-Knauss, T., Butterbach-Bahl, K., Wolf, B., and Bahn, M.: Land-use and abandonment alters methane and nitrous oxide fluxes in mountain grasslands, *Sci. Total Environ.*, 628–629, 997–1008, <https://doi.org/10.1016/j.scitotenv.2018.02.119>, 2018.

Hirano, T., Segah, H., Kusin, K., Limin, S., Takahashi, H., and Osaki, M.: Effects of disturbances on the carbon balance of tropical peat swamp forests, *Glob. Change Biol.*, 18, 3410–3422, <https://doi.org/10.1111/j.1365-2486.2012.02793.x>, 2012.

Hutchinson, G.L. and Mosier, A.R.: Improved soil cover method for field measurements of nitrous oxide fluxes, *Soil Sci. Soc. Am. J.*, 45, 311–316, <https://doi.org/10.2136/sssaj1981.03615995004500020017x>, 1981.

Jackson, W. J., Argent, R. M., Bax, N. J., Clark, G. F., Coleman, S., Cresswell, I. D., Emmerson, K. M., Evans, K., Hibberd, M. F., Johnston, E. L., Keywood, M. D., Klekociuk, A., Mackay, R., Metcalfe, D., Murphy, H., Rankin, A., Smith, D. C.,

and Wienecke, B.: Australia State of the Environment Report, Australian Government Department of the Environment and Energy, Canberra, <https://soe.environment.gov.au>, 2017.

690 Kauffman, J. B., Adame, M. F., Arifanti, V. B., Schile-Beers, L. M., Bernardino, A. F., Bhomia, R. K., Donato, D. C., Feller, I. C., Ferreira, T. O., Garcia, M. del C. J., MacKenzie, R. A., Megonigal, J. P., Murdiyarso, D., Simpson, L., and Trejo, H. H.: Total ecosystem carbon stocks of mangroves across broad global environmental and physical gradients, *Ecol. Monogr.*, 90, 1-18, <https://doi.org/10.1002/ecm.1405>, 2020.

Kauffman, J. B., Bernardino, A. F., Ferreira, T. O., Bolton, N. W., Gomes, L. E. de O., and Nobrega, G. N.: Shrimp ponds lead to massive loss of soil carbon and greenhouse gas emissions in northeastern Brazilian mangroves, *Ecol. Evol.*, 8, 5530–5540, <https://doi.org/10.1002/ece3.4079>, 2018.

695 Kauffman, J. B., Trejo, H. H., Garcia, M. del C. J., Heider, C., and Contreras, W. M.: Carbon stocks of mangroves and losses arising from their conversion to cattle pastures in the Pantanos de Centla, Mexico, *Wetl. Ecol. Manag.*, 24, 203–216, <https://doi.org/10.1007/s11273-015-9453-z>, 2015.

Kavehei, E., Iram, N., Rashti, M. R., Jenkins, G. A., Lemckert, C., and Adame, M. F.: Greenhouse gas emissions from stormwater bioretention basins, *Ecol. Eng.*, 159, 1-9, <https://doi.org/10.1016/j.ecoleng.2020.106120>, 2021.

700 Kettler, T. A., Doran, J. W., and Gilbert, T. L.: Simplified method for soil particle-size determination to accompany soil-quality analyses, *Soil Sci. Soc. Am. J.*, 65, 849-852, <https://doi.org/10.2136/sssaj2001.653849x>, 2001.

Kirschke, S., Bousquet, P., Ciais, P., Saunois, M., Canadell, J. G., Dlugokencky, E. J., Bergamaschi, P., Bergmann, D., Blake, D. R., Bruhwiler, L., Cameron-Smith, P., Castaldi, S., Chevallier, F., Feng, L., Fraser, A., Heimann, M., Hodson, E.

705 L., Houweling, S., Josse, B., Fraser, P. J., Krummel, P. B., Lamarque, J., Langenfelds, R. L., le Quere, C., Naik, V., O'Doherty, S., Plamer, P. I., Pison, I., Plummer, D., Poulter, B., Prinn, R. G., Rigby, M., Ringeval, B., Santini, M., Schmidt, M., Shindell, D. T., Simpson, I. J., Spahni, R., Steele, L. P., Strobe, S. A., Sudo, K., Szopa, S., van der Werf, G. R., Voulgarakis, A., van Weele, M., Weiss, R. F., Williams, J. E., and Zeng, G.: Three decades of global methane sources and sinks. *Nat. Geosci.*, 6, 813–823, <https://doi.org/10.1038/ngeo1955>, 2013.

710 Kögel-Knabner, I., Amelung, W., Cao, Z., Fiedler, S., Frenzel, P., Jahn, R., Kalbitz, K., Kölbl, A., and Schlöter, M.: Biogeochemistry of paddy soils, *Geoderma*, 157, 1–14, <https://doi.org/10.1016/j.geoderma.2010.03.009>, 2010.

Kroeger, K. D., Crooks, S., Moseman-Valtierra, S., and Tang, J.: Restoring tides to reduce methane emissions in impounded wetlands: A new and potent blue carbon climate change intervention, *Scientific reports*, 7, 1-12, <https://doi.org/10.1038/s41598-017-12138-4>, 2017.

715 Lal R.: Carbon sequestration. *Philosophical Transactions Royal Society B*, 363, 815–830, <https://doi.org/10.1098/rstb.2007.2185>, 2008.

Lehner, B., and Döll, P.: Development and validation of a global database of lakes, reservoirs and wetlands, *J. Hydrol.*, 296, 1-22, <https://doi.org/10.1016/j.jhydrol.2004.03.028>, 2004.

Liu, J., and Lai, D. Y.: Subtropical mangrove wetland is a stronger carbon dioxide sink in the dry than wet seasons, *Agric. For. Meteorol.*, 278, 107644, <https://doi.org/10.1016/j.agrformet.2019.107644>, 2019.

720

- Livesley, S. J., Kiese, R., Miehe, P., Weston, C. J., Butterbach-Bahl, K., and Arndt, S. K. Soil–atmosphere exchange of greenhouse gases in a Eucalyptus marginata woodland, a clover-grass pasture, and Pinus radiata and Eucalyptus globulus plantations. *Glob. Chang. Biol.* 15(2), 425–440, <https://doi.org/10.1111/j.1365-2486.2008.01759.x>, 2009.
- 725 Maher DT, Sippo JZ, Tait DR, Holloway C, Santos IR. Pristine mangrove creek waters are a sink of nitrous oxide. *Sci. Rep.* 6, 25701. doi:10.1038/srep25701. 2016.
- Maietta, C. E., Hondula, K. L., Jones, C. N., and Palmer, M. A.: Palmer. "Hydrological conditions influence soil and methane-cycling microbial populations in seasonally saturated wetlands." *Front. Environ. Sci.* 8:593942, doi: 10.3389/fenvs.2020.593942, 2020.
- Martin, R.M. and Moseman-Valtierra, S.: Greenhouse Gas Fluxes Vary Between Phragmites Australis and Native
 730 Vegetation Zones in Coastal Wetlands Along a Salinity Gradient, *Wetlands*, 35, 1021–1031, <https://doi.org/10.1007/s13157-015-0690-y>, 2015.
- Martiny, J. B. H., Bohannan, B. J. M., Brown, J. H., Colwell, R. K., Fuhrman, J. A., Green, J. L., Horner-Devine, M. C., Kane, M., Krumins, J. A., Kuske, C. R., Morin, P. J., Naeem, S., Øvreås, L., Reysenbach, A. L., Smith, V. H. and Staley, J. T.: Microbial biogeography: putting microorganisms on the map, *Nat. Rev. Microbiol.*, 4, 102–112,
 735 <https://doi.org/10.1038/nrmicro1341>, 2006.
- Maucieri, C., Zhang, Y., McDaniel, M. D., Borin, M., and Adams, M. A.: Short-term effects of biochar and salinity on soil greenhouse gas emissions from a semi-arid Australian soil after re-wetting, *Geoderma*, 307, 267–276, <https://doi.org/10.1016/j.geoderma.2017.07.028>, 2017.
- McLeod, E., Chmura, G. L., Bouillon, S., Salm, R., Björk, M., Duarte, C. M., Lovelock, C. E., Schlesinger, W. H. and
 740 Silliman, B. R.: A blueprint for blue carbon: Toward an improved understanding of the role of vegetated coastal habitats in sequestering CO₂, *Front. Ecol. Environ.* 9, 552–560, <https://doi.org/10.1890/110004>, 2011.
- Mitsch, W. J., Bernal, B., Nahlik, A. M., Mander, Ü., Zhang, L., Anderson, C. J., Jørgensen, S. E., and Brix, H.: Wetlands, carbon, and climate change. *Landscape Ecol.*, 28, 583–597. <https://doi.org/10.1007/s10980-012-9758-8>, 2013.
- Morse, J. L., Ardón, M., and Bernhardt, E. S.: Greenhouse gas fluxes in southeastern US coastal plain wetlands under
 745 contrasting land uses, *Ecol. Appl.*, 22, 264–280, <https://doi.org/10.1890/11-0527.1>, 2012.
- Niemi, R.M., Vepsäläinen, M., Wallenius, K., Simpanen, S., Alakukku, L., and Pietola, L.: Temporal and soil depth-related variation in soil enzyme activities and in root growth of red clover (*Trifolium pratense*) and timothy (*Phleum pratense*) in the field, *Appl. Soil Ecol.*, 30, 113–125, <https://doi.org/10.1016/j.apsoil.2005.02.003>, 2005.
- Nieveen, J.P., Campbell, D.I., Schipper, L.A., Blair, I.J.: Carbon exchange of grazed pasture on a drained peat soil, *Glob. Chang. Biol.*, 11, 607–618, <https://doi.org/10.1111/j.1365-2486.2005.00929.x>, 2005.
- 750 Oertel, C., Matschullat, J., Zurba, K., Zimmermann, F., and Erasmi, S.: Greenhouse gas emissions from soils—A review, *Geochemistry*, 76, 327–352, <https://doi.org/10.1016/j.chemer.2016.04.002>, 2016.
- Ollivier, Q. R., Maher, D. T., Pitfield, C. and Macreadie, P. I.: Punching above their weight: Large release of greenhouse gases from small agricultural dams, *Glob. Chang. Biol.*, 25, 721–732, doi:10.1111/gcb.14477, 2019.

- Pinheiro, P. L., Recous, S., Dietrich, G., Weiler, D. A., Schu, A. L., Bazzo, H. L. S., and Giacomini, S. J.: N₂O emission increases with mulch mass in a fertilised sugarcane cropping system, *Biol. Fertil. Soils*, 55, 511-523, <https://doi.org/10.1007/s00374-019-01366-7>, 2019.
- Poffenbarger, H. J., Needelman, B. A. and Megonigal, J. P.: Salinity Influence on methane emissions from tidal marshes. *Wetlands*. 31, 831–842, <https://doi.org/10.1007/s13157-011-0197-0>, 2011.
- Pouyat, R.V., Yesilonis, I.D., Russell-Anelli, J., and Neerchal, N.K.: Soil chemical and physical properties that differentiate urban land-use and cover types, *Soil Sci. Soc. Am. J.* 71, 1010–1019, <https://doi.org/10.2136/sssaj2006.0164>, 2007.
- Rashti, M. R., Wang, W. J., Harper, S. M., Moody, P. W., Chen, C. R., Ghadiri, H., and Reeves, S. H.: Strategies to mitigate greenhouse gas emissions in intensively managed vegetable cropping systems in subtropical Australia, *Soil Res.*, 53, 475-484, <https://doi.org/10.1071/SR14355>, 2015.
- Rashti, M. R., Wang, W. J., Reeves, S. H., Harper, S. M., Moody, P. W., and Chen, C. R.: Linking chemical and biochemical composition of plant materials to their effects on N₂O emissions from a vegetable soil, *Soil Biol. Biochem.*, 103, 502-511, <https://doi.org/10.1016/j.soilbio.2016.09.019>, 2016.
- Reeves, S., Weijin W, Barry S, and Neil H.: Quantifying nitrous oxide emissions from sugarcane cropping systems: Optimum sampling time and fr-equency, *Atmospheric environment*, 136, 123-133, <https://doi.org/10.1016/j.atmosenv.2016.04.008>, 2016.
- Sahrawat, K.L.: Terminal electron acceptors for controlling methane emissions from submerged rice soils, *Commun. Soil Sci. Plant Anal.* 35, 1401–1413, <https://doi.org/10.1081/CSS-120037554>, 2004.
- Saunois, M., R. Stavert, A., Poulter, B., Bousquet, P., G. Canadell, J., B. Jackson, R., A. Raymond, P., J. Dlugokencky, E., Houweling, S., K. Patra, P., Ciais, P., K. Arora, V., Bastviken, D., Bergamaschi, P., R. Blake, D., Brailsford, G., Bruhwiler, L., M. Carlson, K., Carrol, M., Castaldi, S., Chandra, N., Crevoisier, C., M. Crill, P., Covey, K., L. Curry, C., Etiope, G., Frankenberg, C., Gedney, N., I. Hegglin, M., Höglund-Isaksson, L., Hugelius, G., Ishizawa, M., Ito, A., Janssens-Maenhout, G., M. Jensen, K., Joos, F., Kleinen, T., B. Krummel, P., L. Langenfelds, R., G. Laruelle, G., Liu, L., MacHida, T., Maksyutov, S., C. McDonald, K., McNorton, J., A. Miller, P., R. Melton, J., Morino, I., Müller, J., Murguia-Flores, F., Naik, V., Niwa, Y., Noce, S., O'Doherty, S., J. Parker, R., Peng, C., Peng, S., P. Peters, G., Prigent, C., Prinn, R., Ramonet, M., Regnier, P., J. Riley, W., A. Rosentreter, J., Segers, A., J. Simpson, I., Shi, H., J. Smith, S., Paul Steele, L., F. Thornton, B., Tian, H., Tohjima, Y., N. Tubiello, F., Tsuruta, A., Viogy, N., Voulgarakis, A., S. Weber, T., Van Weele, M., R. Van Der Werf, G., F. Weiss, R., Worthy, D., Wunch, D., Yin, Y., Yoshida, Y., Zhang, W., Zhang, Z., Zhao, Y., Zheng, B., Zhu, Q., Zhu, Q. and Zhuang, Q.: The global methane budget 2000-2017, *Earth Syst. Sci. Data*, 12, 1561–1623, doi:10.5194/essd-12-1561-2020, 2020.
- Shaaban, M., Peng, Q. A., Bashir, S., Wu, Y., Younas, A., Xu, X., Rashti, M.R., Abid, M., Zafar-ul-Hye, M., Núñez-Delgado, A. and Hu, R. Restoring effect of soil acidity and Cu on N₂O emissions from an acidic soil. *J. Envir. Manage.*, 15250, 109535 (2019).<https://doi.org/10.1016/j.jenvman.2019.109535>

- Sjögersten, S., Black, C.R., Evers, S., Hoyos-Santillan, J., Wright, E.L. and Turner, B.L.: Tropical wetlands: a missing link in the global carbon cycle?, *Glob. Biogeochem. Cycles*, 28, 1371–1386, <https://doi.org/10.1002/2014GB004844>, 2014.
- 790 Smith, P., Martino, D., Cai, Z., Gwary, D., Janzen, H., Kumar, P., McCarl, B., Ogle, S., O'Mara, F., Rice, C., Scholes, B., Sirotenko, O.: Agriculture, In: *Climate change 2007: Mitigation, Contribution of Working Group III to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*, Cambridge University Press, Cambridge, United Kingdom and New York, United States, 43 pp., 2007.
- 795 Stadmark, J., and Leonardson, L.: Emissions of greenhouse gases from ponds constructed for nitrogen removal. *Ecological Engineering*, 25, 542-551. <https://doi.org/10.1016/j.ecoleng.2005.07.004>, 2005.
- Ussiri, D., and Lal, R.: *Soil emission of nitrous oxide and its mitigation*. Springer Science & Business Media, Germany, 378, ISBN 978-94-007-5364-8, 2012.
- Van Leeuwen, J. P., Djukic, I., Bloem, J., Lehtinen, T., Hemerik, L., de Ruiter, P. and Lair, G.: Effects of land use on soil microbial biomass, activity and community structure at different soil depths in the Danube floodplain, *Eur. J. Soil Biol.*, 79, 14–20, <https://doi.org/10.1016/j.ejsobi.2017.02.001>, 2017.
- 800 Veenendaal E. M., Kolle, O., Leffelaar, P.A., Schrier-Uijl, Huissteden, J., V., Moller, F., and Berendse, F.: CO₂ exchange and carbon balance in two grassland sites on eutrophic drained peat soils, *Biogeosciences*, 4, 1027–1040, <https://doi.org/10.5194/bg-4-1027-2007>, 2007.
- Wang, J., Song, C., Zhang, J., Wang, L., Zhu, X., Shi, F.: Temperature sensitivity of soil carbon mineralisation and nitrous oxide emission in different ecosystems along a mountain wetland-forest ecotone in the continuous permafrost of Northeast China, *Catena*, 121, 110–118, <https://doi.org/10.1016/j.catena.2014.05.007>, 2014.
- 805 Wang, W., Dalal, R. C., Reeves, S. H., Butterbach-Bahl, and Kiese, R.: Greenhouse gas fluxes from an Australian subtropical cropland under long-term contrasting management regimes, *Glob. Chang. Biol.*, 17, 3089-3101, <https://doi.org/10.1111/j.1365-2486.2011.02458.x>, 2011.
- 810 WetlandInfo, 2020. Herbert River drainage sub-basin — facts and maps, <https://wetlandinfo.des.qld.gov.au/wetlands/facts-maps/sub-basin-herbert-river/> viewed 20 March 2020,
- Whalen, S.: Biogeochemistry of methane exchange between natural wetlands and the atmosphere, *Environ. Eng. Sci.*, 22, 73–94, <https://doi.org/10.1089/ees.2005.22.73>, 2005.
- Xu, C., Han, X., Ru, S., Cardenas, L., Rees, R. M., Wu, D., and Meng, F.: Crop straw incorporation interacts with N fertiliser on N₂O emissions in an intensively cropped farmland, *Geoderma*, 341, 129-137, <https://doi.org/10.1016/j.geoderma.2019.01.014>, 2019.
- 815 Xu, C., Wong, V. N. L., and Reef, R. E.: Effect of inundation on greenhouse gas emissions from temperate coastal wetland soils with different vegetation types in southern Australia, *Sci. Total Environ.*, 763, 142949, <https://doi.org/10.1016/j.scitotenv.2020.142949>, 2021.
- 820 Yang, W. B., Yuan, C. S., Tong, C., Yang, P., Yang, L., and Huang, B. Q.: Diurnal variation of CO₂, CH₄, and N₂O emission fluxes continuously monitored in-situ in three environmental habitats in a subtropical estuarine wetland, *Marine Pollution Bulletin*, 119, 289-298, <https://doi.org/10.1016/j.marpolbul.2017.04.005>, 2017.

Yvon-Durocher, G., Montoya, J.M., Woodward, G., Jones, J.I., and Trimmer, M.: Warming increases the proportion of primary production emitted as methane from freshwater mesocosms, *Glob. Change Biol.*, 17, 1225-1234, <https://doi.org/10.1111/j.1365-2486.2010.02289.x>, 2011.

Zaehle, S., and Dalmonech, D.: Carbon-nitrogen interactions on land at global scales: Current understanding in modelling climate biosphere feedbacks, *Curr. Opin. Environ. Sustain*, 3, 311–320, <https://doi.org/10.1016/j.cosust.2011.08.008>, 2011.

IPCC: Agriculture in Climate change (2007). Mitigation Contribution of Working Group III to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. New York: Cambridge University Press; 2007.

Jeffrey, L. C., Maher, D. T., Chiri, E., Leung, P. M., Nauer, P. A., Arndt, S. K., ... & Johnston, S. G. (2021). Bark-dwelling methanotrophic bacteria decrease methane emissions from trees. *Nature Communications*, 12(1), 1-8.

Lenhart, K.; Behrendt, T.; Greiner, S.; Steinkamp, J.; Well, R.; Gieseemann, A.; Keppler, F. Nitrous oxide effluxes from plants as a potentially important source to the atmosphere. *New Phytol.* **2019**, 221, 1398–140.

Timilsina, A., Bizimana, F., Pandey, B., Yadav, R. K. P., Dong, W., & Hu, C. (2020). Nitrous oxide emissions from paddies: understanding the role of rice plants. *Plants*, 9(2), 180.