

# **Soil greenhouse gas fluxes from tropical coastal wetlands and alternative agricultural land uses**

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**Abstract.** Coastal wetlands are essential for regulating the global carbon budget through soil carbon sequestration and greenhouse gas fluxes (GHG: CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O). The conversion of coastal wetlands to agricultural land alters these fluxes' magnitude and direction (uptake/release). However, the extent and drivers of change of GHG fluxes is still unknown for many 15 tropical regions. We measured soil GHG fluxes from three natural coastal wetlands: mangroves, saltmarsh, and freshwater tidal forests, and two alternative agricultural land use, sugarcane farming and pastures for cattle grazing (ponded and dry conditions). We assessed variations throughout different climatic conditions (dry-cool, dry-hot and wet-hot) within two years 20 of measurements (2018-2020) in tropical Australia. The wet pasture had by far the highest CH<sub>4</sub> emissions with  $1,231 \pm 386$  mg m<sup>-2</sup> d<sup>-1</sup>, which were 200-fold higher than any other site. Dry pastures and sugarcane were the highest emitters of N<sub>2</sub>O with 25  $55 \pm 9$  mg m<sup>-2</sup> d<sup>-1</sup> (wet-hot period) and  $11 \pm 3$  mg m<sup>-2</sup> d<sup>-1</sup> (hot-dry period, coinciding with fertilisation), respectively. Dry pastures were also the highest emitters of CO<sub>2</sub> with  $20 \pm 1$  g m<sup>-2</sup> d<sup>-1</sup> (wet-hot period). The three coastal wetlands measured had lower emission, with saltmarsh uptake of  $-0.55 \pm 0.23$  of N<sub>2</sub>O and  $-1.19 \pm 0.08$  g m<sup>-2</sup> d<sup>-1</sup> of CO<sub>2</sub> during the dry-hot period. During the sampled period, sugarcane and pastures had higher total cumulative soil GHG emissions (CH<sub>4</sub> + N<sub>2</sub>O) of 7,142 and 56,124 CO<sub>2</sub>eq kg ha<sup>-1</sup> y<sup>-1</sup> compared to coastal wetlands with 144 to 884 CO<sub>2</sub>eq kg ha<sup>-1</sup> y<sup>-1</sup>. Restoring unproductive sugarcane land or pastures (especially ponded ones) to coastal wetlands could provide significant GHG mitigation.

## 1 Introduction

Coastal wetlands are found at the interface of terrestrial and marine ecosystems and account for 10% of the global wetland area (Lehner and Döll, 2004). They are highly productive and provide various ecosystem services such as water quality improvement, biodiversity, and carbon sequestration (Duarte et al., 2013). For instance, mangroves can accumulate five times 30 more soil carbon than terrestrial forests (Kauffman et al., 2020). However, the high productivity and anoxic soil conditions that promote carbon sequestration can also favour potent greenhouse gas emissions (GHGs), including CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O (Whalen, 2005; Conrad, 2009).

The GHG emissions in coastal wetlands primarily result from microbial processes in the soil-water-atmosphere 35 interface ((Bauza et al., 2002) Whalen, 2005). The emission of CO<sub>2</sub> is a result of respiration, where fixed carbon by photosynthesis is partially released back into the atmosphere (Oertel et al., 2016). Emissions of CH<sub>4</sub> result from anaerobic and aerobic respiration by methanogenic bacteria, mostly in waterlogged conditions (Angle et al., 2017; Saunois et al., 2020). Finally, N<sub>2</sub>O emissions are caused by denitrification in anoxic conditions and nitrification in aerobic soils, both driven by 40 nitrogen content and soil moisture (Ussiri and Lal 2013). Thus, the total GHG emissions from a wetland are driven by environmental conditions that favour these microbial processes, all of which result in highly variable emissions from wetlands worldwide (Kirschke et al., 2013; Oertel et al., 2016).

Despite potential high GHG emissions from coastal wetlands, these are likely to be lower than those from alternative agricultural land use (Knox et al., 2015), which emit GHGs from their construction throughout their productive lives. Firstly, 45 when wetlands are converted to agricultural land, the oxidation of sequestered carbon in the organic-rich soils release significant amounts of CO<sub>2</sub> (Drexler et al., 2009; Hooijer et al., 2012). Secondly, removing tidal flow and converting coastal wetlands to freshwater systems, such as during the creation of ponded pastures, dams or agricultural ditches, can result in very high CH<sub>4</sub> emissions (Deemer et al., 2016; Grinham et al., 2018; Ollivier et al., 2019). For instance, agricultural ditches contribute up to 3% of the total anthropogenic CH<sub>4</sub> emissions globally (Peacock et al., 2021). Finally, the use of fertilisers 50 significantly increases N<sub>2</sub>O emissions (Rezaei Rashti et al., 2015). Thus, emissions of GHG from land-use change can be mitigated through the reversal of these activities, for instance, reduction of fertiliser use and the reinstallation of tidal flow on unused agricultural land (Kroeger et al., 2017; Rezaei Rashti et al., 2015).

This study measured the annual GHG fluxes (CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O) from three natural coastal wetlands (mangroves, 55 saltmarsh and freshwater tidal forests) and two agricultural land use sites (sugarcane plantation and pasture) in tropical Australia. The objectives were to assess the difference in GHG fluxes throughout different seasons that characterise tropical climates (dry-cool, dry-hot and wet-hot) and identify environmental factors associated with these GHG fluxes. These data will

inform emission factors for converting wetlands to agricultural land uses and vice versa, filling in a knowledge gap identified in Australia (Baldock et al., 2012) and tropical regions worldwide (IPCC, 2013).

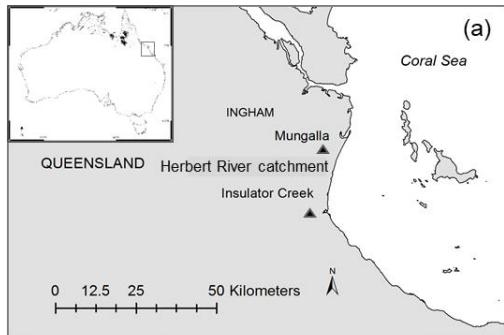
## 60 2 Materials and Methods

### 2.1 Study sites

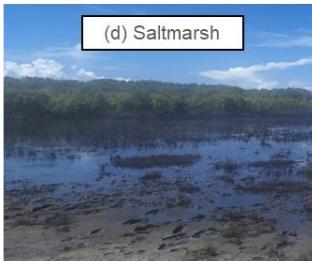
The study area is located within the Herbert River catchment in Queensland, northeast Australia (Fig 1a). The region has a tropical climate with a mean monthly minimum temperature from 14 to 23°C and mean monthly maximum temperature from 25 to 33°C (Australian Bureau of Meteorology, ABM, 2020; 1968-2020; Table S3). The average rainfall is 2,158 mm  $y^{-1}$ , with the highest values of 476 mm during February (ABM 2020; 1968-2020; Table S3). The Herbert basin covers 9,842 km<sup>2</sup>, from which 56% is grazing, 31% is conserved wetlands and forestry, 8% is sugarcane, and 4% is other land uses (Department of Science and Environment, DES, WetlandInfo, 2020). Wetlands in this region were heavily deforested in the past century (1943-1996) due to rapid agricultural development, primarily for sugarcane farming (Griggs, 2018). Before clearing, the land was mostly covered by rainforest and coastal wetlands, mainly *Melaleuca* forest, grass and sedge swamps (Johnson et al., 1999).

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We selected five sites, including three natural coastal wetlands (Fig. 1): a mangrove forest (18° 53' 42" S, 146° 15' 51" E), a saltmarsh (18° 53' 43" S, 146° 15' 52" E) and a freshwater tidal forest (18° 53' 45" S, 146° 15' 52" E), and two common agricultural land use types of the region, a sugarcane crop (18° 53' 44.6" S, 146° 15' 53.2" E) and a pasture for fodder grazing. The pasture had different levels of inundation, some areas were covered with shallow ponds (50-100 cm depth), some were wet (hereafter "wet pasture"; 18° 43' 8" S, 146° 15' 50" E) and others were dry (hereafter, "dry pasture"; 18° 43' 7" S, 146° 15' 50" E). The natural coastal wetlands and the sugarcane site were located within the same property < 200 m apart at Insulator Creek (Fig. 1B), while the ponded pasture was 20 km north at Mungalla Station (Fig. 1A). The mangroves were dominated by *Avicennia marina* with few plants of *Rhizophora stylosa*, and the saltmarsh was dominated by *Sueda salsa* and *Sporobolus spp.*. Landwards, the freshwater tidal freshwater forest, a wetland commonly known as "tea tree swamp", was dominated by *Melaleuca quinquenervia* trees. While the mangroves and saltmarsh are directly submerged by tides (5- 30 cm), the tidal freshwater forest is indirectly affected by tidal fluctuations, such as during large spring tides, when tidal water can push groundwater above the forest thus forming "supra-tidal" wetlands. The coastal wetlands were adjacent to a sugarcane farm of ~110 ha (Fig. 1b). The sugarcane is fertilised once a year with urea at a rate of 135 kg N ha<sup>-1</sup> and harvested during May-June, while the foliage is left on the soil surface (trash blanket) after harvest. The ponded pastures in Mungalla Station is a 250 ha farm and support ~900 cattle by providing fodder during dry periods. The selected ponded pastures were covered by *Eichhornia crassipes* (water hyacinth) and *Hymenachne amplexicaulis* (Fig. 1g-h). Each of the five sites was sampled during three periods: dry-cool (May-September), dry-hot (October-December) and wet-hot (January-April; Table 1). During each time, soil physicochemical properties and GHG fluxes were measured as detailed below.



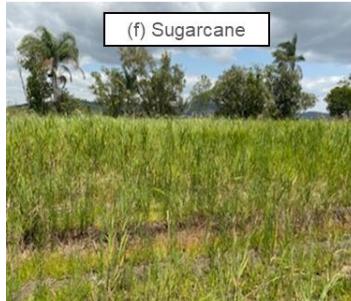
(c) Mangroves



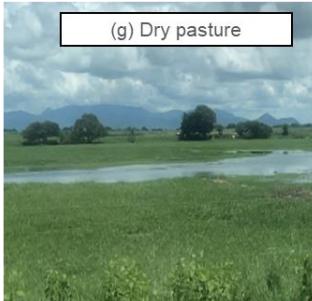
(d) Saltmarsh



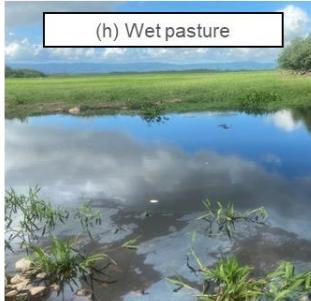
(e) Freshwater tidal forest



(f) Sugarcane



(g) Dry pasture



(h) Wet pasture

Figure 1: a) Location of sampling sites (Insulator Creek and Mungalla) in the Herbert River catchment, northeast Australia, (b) natural wetlands adjacent to sugarcane in Insulator Ck and sampling locations, and (c) mangroves, (d) saltmarsh, (e) freshwater tidal forest, (f) sugarcane, (g) dry pasture and (h) wet pasture. Pictures by N. Iram and MF Adame.

Table 1. Mean daily air temperature and rainfall range (Ingham, weather station 32078) during sampling.

Season	Study period	Daily min temperature (°C)	Daily max temperature (°C)	Rainfall (mm d <sup>-1</sup> )
Dry-cool	17/06/2018	13.4 - 14.6	27.7 - 28.2	0
Dry-hot	23-29/10/2018	15.7 - 21.1	32.2 - 36.2	0
Dry-cool	31/05 to 6/06/2019	10.9 - 17.5	21.6 - 28.2	0-25
Wet-hot	17-22/02/2020	23.9 - 25.3	33.6 - 34.5	0-86

## 2.2 Soil sampling and analysis of physicochemical properties

100 Soil physicochemical characteristics were measured in composite soil samples next to each gas chamber location (n = 5; 0-30 cm) for all study sites during the dry-hot season. The samples were obtained by inserting an open steel corer to a depth of 30 cm; the core was divided into three depths: 0-10, 10-20 and 20-30 cm. Soil samples were oven-dried at 105 °C for 48 h to determine volumetric water content through gravimetric analysis. The volumetric water content was divided by total soil porosity to determine water-filled pore space (WFPS). Total soil porosity was calculated using an equation from Rezaei Rashti  
105 et al. (2015, Eq.1):

$$\text{Total soil porosity} = 1 - [(\text{soil bulk density (mg cm}^{-3}) / 2.65)]$$

Equation 1.

110 Soil texture analysis (% sand, silt, clay) was done with a simplified method for particle size determination (Kettler et al., 2001). Soil electrical conductivity (EC) and pH were measured using a conductivity meter (WP-84 TPS, Australia) in soil/water slurry at 1:5. Soil subsamples were air-dried, sieved (2 mm), ground (Retch<sup>TM</sup> mill) and analysed for %N and %C using an elemental analyser connected to a gas-isotope ratio mass spectrometer (EA-Delta V Advantage IRMS, Griffith University). Additionally, soil samples from the top 10 cm were collected during each sampling to measure gravimetric soil  
115 moisture content and bulk density.

## 2.3 Greenhouse gas fluxes

We measured GHG fluxes (CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O) at each site for three consecutive days during each sampling period except for the dry-cool period of 2018, when mangroves, saltmarsh and sugarcane were surveyed for one day. The sampling was done  
120 between 9:00 to 11:00 am, representing the mean daily temperatures, thus, minimising variability of cumulative seasonal fluxes

based on intermittent manual flux measurements (Reeves et al., 2016). Additionally, we assessed the variability of our measurements with tidal inundation in mangroves and saltmarsh, which were regularly inundated (~10-30 cm) during the hot-dry season. However, due to logistic constraints, further sampling was conducted only during low tides.

125 We used static, manual gas chambers made of high-density, round polyvinyl chloride pipe, which consisted of two units: a base ( $r = 12$  cm,  $h = 18$  cm) and a detachable collar ( $h = 12$  cm; (Hutchinson and Mosier, 1981; Kavehei et al., 2021). The chambers had lateral holes that could be left covered with rubber bungs at low water levels and left open at high water levels to allow water movement between sampling events. When the wetlands were inundated for the experiments, we used PVC extensions ( $h = 18$  cm). Five chambers were set ~ 5cm deep in the soil, separated one to two meters from each other, 130 selectively located on soil with minimal vegetation, roots, and crab burrows. The chambers were set one day before sampling to minimise installation disturbance during the experiment (Rezaei Rashti et al., 2015). We were careful not to tramp around the chambers during installation and sampling. The fact that emissions were not significantly different among days ( $p > 0.05$ ) provided us with confidence that disturbance due to installation was not problematic.

135 At the start of the experiment, gas chambers were closed. A sample was taken at time zero and then after one hour with a 20 ml syringe and transferred to a 12 mL-vacuumed container (Exetainer, Labco Ltd., High Wycombe, UK). During the dry-hot season, linearity tests of GHG fluxes with time were conducted by sampling at 0, 20, 40 and 60 min at all chambers (Rezaei Rashti et al., 2016). For the rest of the experiments, linearity tests were performed in one chamber per site;  $R^2$  values were consistently above 0.70. During each experiment, soil temperature was measured next to each chamber. For each 140 experiment, the base depth was recorded from five points within each chamber to calculate headspace volume. The obtained volumetric unit concentrations were converted to mass-based units using the Ideal Gas Law (Hutchinson and Mosier, 1981).

145 The GHG concentrations of all samples were analysed within two weeks of sampling with a gas chromatograph (Shimadzu GC-2010 Plus). For  $N_2O$  analysis, an electron capture detector was used with helium as the carrier gas, while  $CH_4$  was analysed on a flame ionisation detector with nitrogen as the carrier gas. For  $CO_2$  determination, the gas chromatograph was equipped with a thermal conductivity detector. Peak areas of the samples were compared against standard curves to determine concentrations (Chen et al., 2012). Seasonal cumulative GHG fluxes were calculated by modifying the equation described by Shaaban et al. (2015; Eq. 2):

$$\text{Seasonal cumulative GHG fluxes (mg or } \mu\text{g m}^{-2} \text{ yr}^{-1}) = \sum_{i=1}^n (R_i \times 24 \times D_i \times 17.38) \quad \text{Equation 2}$$

150

Where;

$R_i$  = Gas emission rate ( $mg\ m^{-2}\ hr^{-1}$  for  $CO_2$  and  $\mu g\ m^{-2}\ hr^{-1}$  for  $CH_4$  and  $N_2O$ ),

$D_i$  = Mean daily GHG emission rate in a season ( $mg\ m^{-2}\ d^{-1}$  for  $CO_2$  and  $\mu g\ m^{-2}\ d^{-1}$  for  $CH_4$  and  $N_2O$ ) during low tide

17.38 = number of weeks in each season, assuming these conditions were representative of the annual cycle (see Table 1).

155 Annual cumulative soil GHG fluxes ( $\text{CH}_4 + \text{N}_2\text{O}$ ) were calculated by integrating cumulative seasonal fluxes. These estimations did not account for soil  $\text{CO}_2$  values as our methodology with dark chambers only accounted for emissions from respiration and excluded uptake by primary productivity. The  $\text{CO}_2$ -equivalent ( $\text{CO}_2\text{-eq}$ ) values were estimated by multiplying  $\text{CH}_4$  and  $\text{N}_2\text{O}$  emissions by 25 and 298, respectively (Solomon, 2007), which represented the radiative balance of these gases (Neubauer, 2021). For annual cumulative soil GHG flux calculations from coastal wetlands, we used GHG fluxes measured 160 during low tide; therefore, our values did not incorporate the effect of tidal fluctuations. The spatial and temporal replication of this study targeted spatial variation within soil type (< 50 cm, five chambers), days (three days per sampling) and seasons (three seasons per year). However, our replication within land use and wetland type was limited; thus, generalisations for all wetlands and land uses should be done acknowledging this limitation.

## 165 2.4 Statistical analyses

GHG flux data were tested for normality through Kolmogorov-Smirnov and Shapiro-Wilk tests. The data was then analysed 170 for spatial and temporal differences with one-way Analyses of Variance (ANOVA), where site and season were the predictive factors and the replicate (chamber) was the random factor of the model. When data were not normal, they were transformed ( $\log_{10}$  or  $1/x$ ) to comply with the assumptions of normality and homogeneity of variances. Some variables were not normally distributed despite transformations and were analysed with the non-parametric Kruskal-Wallis test and Mann-Whitney U Test. A Pearson correlation test was run to evaluate the correlation of GHG with measured environmental factors. Analyses were done with SPSS (v25, IBM, New York, USA), and values are presented as mean  $\pm$  standard error (SE).

## 3 Results

### 3.1 Soil physicochemical properties

175 Soil physical and chemical parameters (mean values 0-30 cm) varied among sites (Table 2, see full results of statistical analyses in Supplementary Material). As expected, gravimetric moisture content was highest in the coastal wetlands and wet pasture (> 26%) and lowest in the sugarcane and the dry pasture (< 14%). All soils were acidic, especially the freshwater tidal forest and the wet pastures with values < 5 throughout the sediment column; mangroves had the highest pH with  $6.0 \pm 0.1$ . The lowest EC was recorded in the pastures ( $247 \pm 38$  and  $190 \pm 39 \mu\text{S cm}^{-1}$  for the dry and wet pasture, respectively), and highest in the 180 three natural coastal wetlands with  $1,418 \pm 104$ ,  $8,049 \pm 276$  and  $8,930 \pm 790 \mu\text{S cm}^{-1}$  for the tidal freshwater forest, saltmarsh and mangroves, respectively.

Soil bulk density was highest in sugarcane ( $1.5 \pm 0.1 \text{ g cm}^{-3}$ ) and lowest in the freshwater tidal forest ( $0.6 \pm 0.1 \text{ g cm}^{-3}$ ). For all sites, %C was highest in the top 10 cm of the soil and decreased with depth, with the highest values in the freshwater

tidal forest ( $5.1 \pm 0.6\%$ ) and lowest in the saltmarsh ( $1.2 \pm 0.1\%$ ). Soil %N ranged from  $0.1 \pm 0.0$  to  $0.4 \pm 0.1\%$  in all sites,  
185 except in the freshwater tidal wetland, where it reached values of  $0.6 \pm 0.0\%$  in the top 10 cm (Table 2).

### 3.2 Greenhouse gas fluxes

Soil emissions for CO<sub>2</sub> were significantly different among sites and times of the year ( $t = 155.09$ ,  $n = 237$ ,  $p < 0.001$ ; Fig. 2a). The highest CO<sub>2</sub> emissions were measured during the wet-hot period in the dry pasture, where values reached  $20308 \pm 1951$  mg m<sup>-2</sup> d<sup>-1</sup> while the lowest values were measured in the saltmarsh, the only site that acted as a sink of CO<sub>2</sub> with an uptake rate  
190 of  $-594 \pm 152$  mg m<sup>-2</sup> d<sup>-1</sup>. In the pastures, CO<sub>2</sub> emissions were twice as high when dry with cumulative annual emissions of  $5,748 \pm 303$  g m<sup>-2</sup> y<sup>-1</sup> compared to when wet, with  $2,163 \pm 465$  g m<sup>-2</sup> y<sup>-1</sup>. For the coastal wetlands, cumulative annual CO<sub>2</sub> emissions were highest in freshwater tidal forests with  $2,213 \pm 284$  g m<sup>-2</sup> y<sup>-1</sup>, followed by mangroves with  $1,493 \pm 111$  g m<sup>-2</sup> y<sup>-1</sup> and lowest at the saltmarsh with uptake rates of  $-264 \pm 29$  g m<sup>-2</sup> y<sup>-1</sup>.

195 For CH<sub>4</sub> fluxes, significant differences were observed among sites and seasons ( $t = 182.33$ ,  $n = 237$ ,  $p < 0.001$ ). The differences between different sites were substantial, with wet pasture having significantly higher CH<sub>4</sub> emissions than any other site, with rates ~200 times higher (Fig. 2b). For tidal coastal wetlands, emissions of CH<sub>4</sub> were highest during the wet-hot season in all the sites except for the mangroves, which had similar emissions throughout the year (Fig. 2b). Overall, cumulative annual CH<sub>4</sub> emissions were  $209 \pm 36$  g m<sup>-2</sup> y<sup>-1</sup> for the wet pasture followed by mangroves ( $0.73 \pm 0.13$  g m<sup>-2</sup> y<sup>-1</sup>), dry pasture  
200 ( $0.15 \pm 0.03$  g m<sup>-2</sup> y<sup>-1</sup>), freshwater tidal forest ( $0.14 \pm 0.03$  g m<sup>-2</sup> y<sup>-1</sup>), saltmarsh ( $0.04 \pm 0.01$  g m<sup>-2</sup> y<sup>-1</sup>), and sugarcane ( $-0.04 \pm 0.02$  g m<sup>-2</sup> y<sup>-1</sup>).

205 For N<sub>2</sub>O fluxes, the highest emissions ( $54.6 \pm 9.0$  mg m<sup>-2</sup> d<sup>-1</sup>) were from the dry pasture in the wet-hot season, followed by sugarcane ( $20.5 \pm 2.7$  mg m<sup>-2</sup> d<sup>-1</sup>) during the hot-dry period, which coincides with the post-fertilisation months (Fig. 2c). Overall, dry pastures had the highest cumulative annual N<sub>2</sub>O emissions ( $7.99 \pm 2.26$  g m<sup>-2</sup> y<sup>-1</sup>), followed by sugarcane ( $2.37 \pm 0.68$  g m<sup>-2</sup> y<sup>-1</sup>), wet pasture ( $1.32 \pm 0.33$  mg m<sup>-2</sup> d<sup>-1</sup>), saltmarsh ( $0.33 \pm 0.11$  mg m<sup>-2</sup> d<sup>-1</sup>), freshwater tidal forests ( $0.04 \pm 0.0$  g m<sup>-2</sup> y<sup>-1</sup>) and finally, mangroves ( $0.02 \pm 0.04$  g m<sup>-2</sup> y<sup>-1</sup>). However, these differences were only significant when considering the interaction between time of the year and site ( $t = 100.21$ ,  $n = 237$ ,  $p < 0.001$ ).

210 The CH<sub>4</sub> fluxes did not vary significantly between the low and high tide within all coastal wetlands. Contrarily, for saltmarsh, CO<sub>2</sub> was taken during the high tide ( $1.12 \pm 0.24$  g m<sup>-2</sup> d<sup>-1</sup>) but emitted ( $0.69 \pm 0.4$  g m<sup>-2</sup> d<sup>-1</sup>) during the low tide ( $F_{1,28} = 20.06$ ,  $p < 0.001$ ). Finally, for N<sub>2</sub>O, fluxes differed in all coastal wetlands, with higher uptakes in the high tide for mangroves ( $F_{1,28} = 38.28$ ,  $p < 0.001$ ;  $F_{1,28} = 13.53$ ,  $p = 0.001$ ) and higher emissions in saltmarsh ( $F_{1,28} = 38.31$ ,  $p < 0.001$ ) during low tide (Table S4). These results suggested that there was a likely variability for CO<sub>2</sub> and N<sub>2</sub>O fluxes, depending on the time of  
215 sampling.

The wet pasture had the highest total cumulative soil GHG emissions ( $\text{CH}_4 + \text{N}_2\text{O}$ ) with 56,124  $\text{CO}_2\text{eq kg ha}^{-1} \text{yr}^{-1}$  followed by dry pasture 23,890  $\text{CO}_2\text{eq kg ha}^{-1} \text{yr}^{-1}$  and sugarcane 7,142  $\text{CO}_2\text{eq kg ha}^{-1} \text{yr}^{-1}$ . While coastal wetlands had comparatively lower cumulative soil GHG emissions with 884, 235 and 144  $\text{CO}_2\text{eq kg ha}^{-1} \text{yr}^{-1}$  for saltmarsh, mangroves and freshwater tidal forests, respectively. Overall, the three coastal wetlands measured in this study had lower total cumulative GHG emissions at 1,263  $\text{CO}_2\text{eq kg ha}^{-1} \text{yr}^{-1}$  compared to the alternate agricultural land uses, which emitted 87,156  $\text{CO}_2\text{eq kg ha}^{-1} \text{yr}^{-1}$ .

### 3.3 Greenhouse gas emissions and environmental factors

Overall, we found that not one single parameter measured in this could explain GHG fluxes for all sites except land-use. The  $\text{CO}_2$  emissions were not significantly correlated with bulk density ( $R^2 = 0.026, p = 0.918, n = 18$ ), % WFPS ( $R^2 = -0.003, p = 0.99, n = 18$ ), or soil temperature ( $R^2 = 0.296, p = 0.233, n = 18$ ). Soil  $\text{CH}_4$  emissions were neither correlated with bulk density ( $R^2 = -0.096, p = 0.706, n = 18$ ), % WFPS ( $R^2 = 0.224, p = 0.372, n = 18$ ) or soil temperature ( $R^2 = 0.286, p = 0.25, n = 18$ ). Finally, no correlation was found between  $\text{N}_2\text{O}$  emissions and bulk density ( $R^2 = -0.349, p = 0.156, n = 18$ ), % WFPS ( $R^2 = -0.34, p = 0.168, n = 18$ ), or soil temperature ( $R^2 = -0.241, p = 0.335, n = 18$ ). Full raw dataset of GHG fluxes were provided in Table S1.

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Table 2. Physicochemical characteristics for the soil of natural coastal wetlands, sugarcane and pastures (dry and ponded) for the top 30 cm of soil in tropical Australia. C= carbon, N = Nitrogen, EC = Electrical Conductivity. Values are mean  $\pm$  standard error ( $n = 5$ ).

Site	Depth (cm)	Gravimetric moisture content (%)	pH	EC ( $\mu\text{s cm}^{-1}$ )	Bulk density ( $\text{g cm}^{-3}$ )	%C	%N
Mangroves	0-10	41.7 $\pm$ 1.1	5.9 $\pm$ 0.1	12,550 $\pm$ 524	1.14 $\pm$ 0.05	2.3 $\pm$ 0.1	0.18 $\pm$ 0.01
	10-20	34.6 $\pm$ 0.7	5.9 $\pm$ 0.3	12,164 $\pm$ 5,560	1.34 $\pm$ 0.03	1.7 $\pm$ 0.2	0.12 $\pm$ 0.01
	20-30	31.3 $\pm$ 0.6	6.2 $\pm$ 0.1	5,560 $\pm$ 365	1.95 $\pm$ 0.12	0.9 $\pm$ 0.1	0.07 $\pm$ 0.01
	Mean	35.9 $\pm$ 1.2	6.0 $\pm$ 0.1	8,930 $\pm$ 790	1.48 $\pm$ 0.10	1.6 $\pm$ 0.2	0.12 $\pm$ 0.01
Saltmarsh	0-10	25.6 $\pm$ 1.2	5.8 $\pm$ 0.2	8,442 $\pm$ 435	1.12 $\pm$ 0.04	1.4 $\pm$ 0.1	0.11 $\pm$ 0.01
	10-20	26.6 $\pm$ 0.3	5.8 $\pm$ 0.1	8,666 $\pm$ 437	1.47 $\pm$ 0.05	1.3 $\pm$ 0.1	0.12 $\pm$ 0.01
	20-30	26.4 $\pm$ 0.2	5.9 $\pm$ 0.3	7,040 $\pm$ 316	1.56 $\pm$ 0.03	1.0 $\pm$ 0.3	0.10 $\pm$ 0.02
	Mean	26.2 $\pm$ 0.4	5.8 $\pm$ 0.1	8,049 $\pm$ 276	1.38 $\pm$ 0.06	1.2 $\pm$ 0.1	0.11 $\pm$ 0.01
Freshwater tidal forest	0-10	33.4 $\pm$ 0.5	4.4 $\pm$ 0.2	1,099 $\pm$ 17	0.46 $\pm$ 0.05	7.8 $\pm$ 0.1	0.62 $\pm$ 0.03
	10-20	24.9 $\pm$ 0.6	4.2 $\pm$ 0.0	1,272 $\pm$ 164	0.71 $\pm$ 0.02	5.4 $\pm$ 0.0	0.46 $\pm$ 0.04
	20-30	22.4 $\pm$ 0.7	4.2 $\pm$ 0.1	1,882 $\pm$ 47	0.83 $\pm$ 0.03	2.2 $\pm$ 0.1	0.10 $\pm$ 0.00
	Mean	26.9 $\pm$ 1.3	4.3 $\pm$ 0.1	1,418 $\pm$ 104	0.59 $\pm$ 0.05	5.1 $\pm$ 0.6	0.39 $\pm$ 0.06
Sugarcane	0-10	9.1 $\pm$ 0.4	5.7 $\pm$ 0.1	429 $\pm$ 12	1.35 $\pm$ 0.08	1.5 $\pm$ 0.1	0.10 $\pm$ 0.00
	10-20	12.1 $\pm$ 0.6	5.3 $\pm$ 0.3	365 $\pm$ 11	1.46 $\pm$ 0.06	1.5 $\pm$ 0.1	0.12 $\pm$ 0.01
	20-30	13.7 $\pm$ 0.2	4.7 $\pm$ 0.2	351 $\pm$ 2	1.64 $\pm$ 0.10	1.3 $\pm$ 0.1	0.10 $\pm$ 0.00
	Mean	11.7 $\pm$ 0.6	5.2 $\pm$ 0.2	382 $\pm$ 11	1.48 $\pm$ 0.05	1.4 $\pm$ 0.1	0.11 $\pm$ 0.00
Pasture	Dry	0-10	12.4 $\pm$ 0.3	4.1 $\pm$ 0.0	378 $\pm$ 21	0.78 $\pm$ 0.06	3.1 $\pm$ 0.3
		10-20	13.6 $\pm$ 0.1	4.4 $\pm$ 0.1	279 $\pm$ 60	1.21 $\pm$ 0.14	1.6 $\pm$ 0.4
		20-30	14.5 $\pm$ 0.7	4.4 $\pm$ 0.3	84 $\pm$ 4	1.32 $\pm$ 0.19	1.6 $\pm$ 0.2
		Mean	13.5 $\pm$ 0.3	4.3 $\pm$ 0.1	247 $\pm$ 38	1.10 $\pm$ 0.10	2.1 $\pm$ 0.3
	Wet	0-10	52.1 $\pm$ 0.4	4.8 $\pm$ 0.0	358 $\pm$ 71	0.62 $\pm$ 0.06	3.6 $\pm$ 0.3
		10-20	47.7 $\pm$ 0.4	4.9 $\pm$ 0.1	117 $\pm$ 11	1.30 $\pm$ 0.02	1.7 $\pm$ 0.1
		20-30	46.4 $\pm$ 0.2	5.1 $\pm$ 0.1	95 $\pm$ 6	1.31 $\pm$ 0.02	1.5 $\pm$ 0.1
		Mean	48.7 $\pm$ 0.7	4.9 $\pm$ 0.0	190 $\pm$ 39	1.07 $\pm$ 0.09	2.3 $\pm$ 0.3

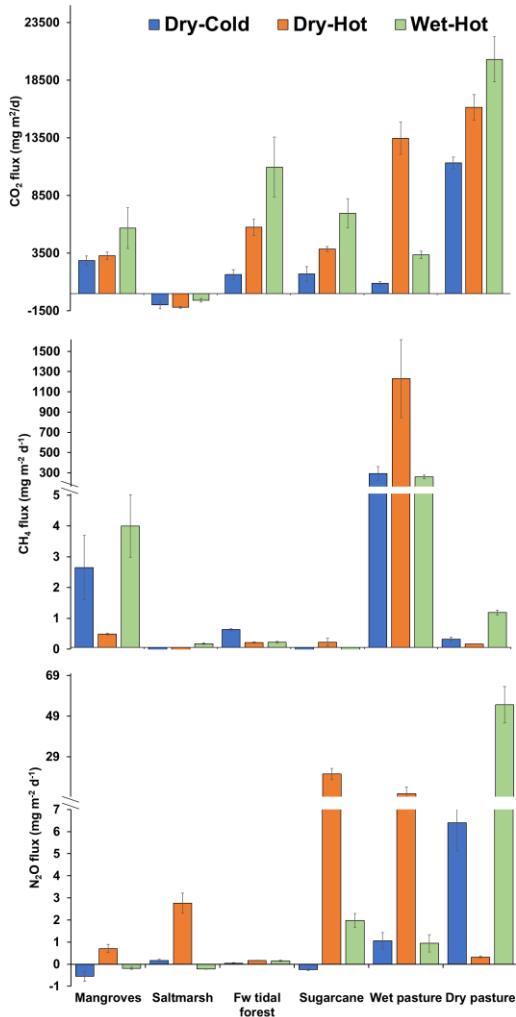


Figure 2: Greenhouse gas fluxes of ( $\text{mg m}^{-2} \text{ d}^{-1}$ ) (a)  $\text{CO}_2$  (b)  $\text{CH}_4$  and (c)  $\text{N}_2\text{O}$  from the soils of tropical coastal wetlands: mangroves, saltmarsh, freshwater (Fw) tidal forest, and two alternative land uses: sugarcane and pastures (wet and dry) during three periods of the year: dry-cold, dry-hot and wet-hot

#### 4 Discussion

The soils of the three coastal wetlands measured in this study (mangroves, saltmarshes and freshwater tidal forests) had significantly lower GHG emissions than those from two alternative land uses common in tropical Australia (sugar cane and grazing pastures). Notably, we found that coastal wetlands had 200 times lower CH<sub>4</sub> emissions and seven times lower N<sub>2</sub>O compared to wet pastures and sugarcane soils, respectively. These results support our hypothesis that the management or conversion of unused sugarcane land and ponded pastures in tropical Australia could be restored to coastal wetlands and result in significant GHG mitigation.

The variability of GHG fluxes was best explained by land use and wetland type; however, some trends with seasons were evident. For instance, CO<sub>2</sub> and N<sub>2</sub>O emissions were lowest during the dry-cool periods. Reduced emissions at low temperatures are expected as the temperature is a primary driver of any metabolic process, including respiration and nitrification-denitrification. Mangroves tend to have higher CO<sub>2</sub> emissions as temperature increases (Liu and Lai, 2019), and forests have significantly higher N<sub>2</sub>O emissions during warm seasons (Schindlbacher et al., 2004). Emissions of CH<sub>4</sub> also tend to increase with temperature as the activity of soil methane-producing microbes (Ding et al., 2004) and the availability of carbon is higher in warmer conditions (Yvon-Durocher et al., 2011). However, most studies to date on GHG fluxes have been conducted in temperate locations, where temperature differences throughout the year are smaller than in tropical regions. For tropical regions, increased GHG emissions are likely to be strongly affected by the "Birch effect", which refers to short-term but a substantial increase of respiration from soils under the effect of precipitation during the early wet season (Fernandez-Bou et al., 2020).

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The main factor associated with GHG fluxes was land use and type of wetland. Notably, coastal wetlands, even the freshwater tidal forests, had much lower emissions than wet pastures. This significant difference could be attributed to terminal electron acceptors in the soils (e.g. iron, sulphate, manganese) of the coastal wetlands, which could inhibit methanogenesis (Kögel-Knabner et al., 2010; Sahrawat, 2004). Sulphate reducing bacteria are also likely to outcompete methane-producing bacteria (methanogens) in the presence of high sulphate concentrations in tidal wetlands, resulting in low CH<sub>4</sub> production. Competition between methanogens and methanotrophs may result in a net balance of low CH<sub>4</sub> production despite freshwater conditions (Maietta et al., 2020). Additionally, microorganisms living within the bark of *Melaleuca* trees can consume CH<sub>4</sub> (Jeffrey et al., 2021), so it is possible that similar bacteria could reduce CH<sub>4</sub> emissions in the soil. Interestingly, variability within CH<sub>4</sub> fluxes among sites was very high, despite being very close to each other (Fig. 1b). These differences highlight the importance of land use in GHG fluxes, which are likely to significantly alter the microbial community composition and abundance, changing rapidly over small spatial scales (Drenovsky et al., 2010; Martiny et al., 2006).

Our results are consistent with other studies, showing the importance of land use in GHG emissions. For instance, in a Mediterranean climate, the drained agricultural land use types, pasture and corn, were larger CO<sub>2</sub> emitters than restored wetlands(Knox et al., 2015). Clearing of wetlands for agricultural development, such as the drainage of peatlands, results in very high CO<sub>2</sub> emissions (Hirano et al., 2012; Nieven et al., 2005; Veenendaal et al., 2007), and restoration of these wetlands could decrease these emissions(Cameron et al., 2021). Additionally, some of the wetland types, such as marshes, were occasional sinks of CO<sub>2</sub> and CH<sub>4</sub>, consistent with previous studies where intertidal wetlands sink of GHG at least under some conditions or during some times of the year (Knox et al., 2015; Maher et al., 2016).

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The fluxes measured in the coastal wetlands of this study, -1191 to 10,970 mg m<sup>-2</sup> d<sup>-1</sup> for CO<sub>2</sub>, -0.3 to 3.9 mg m<sup>-2</sup> d<sup>-1</sup> for CH<sub>4</sub>, and -0.2 to 2.8 mg m<sup>-2</sup> d<sup>-1</sup> for N<sub>2</sub>O, were within the range of those measured in other subtropical/tropical wetlands, worldwide (except for the negative CO<sub>2</sub> fluxes in saltmarsh soils, Table 3). For CO<sub>2</sub>, fluxes can range between 44 and 11,328 mg m<sup>-2</sup> d<sup>-1</sup>, for CH<sub>4</sub>, from 0.03 to 1255 mg m<sup>-2</sup> d<sup>-1</sup> and for N<sub>2</sub>O, from 0.1 to 279 mg m<sup>-2</sup> d<sup>-1</sup> (Table 3). Despite being in tropical regions, GHG fluxes from this study were lower compared to other climates (Table 3). Contrary to previous studies, CO<sub>2</sub> uptake by saltmarsh soil was ere-likely to be linked with dark CO<sub>2</sub> fixation in wetland soils (Akinyede et al., 2020; Mar Lynn et al., 2017). Wetland soils exhibit autotrophic bacteria which contribute to dark CO<sub>2</sub> fixation at ~311 mg m<sup>-2</sup> d<sup>-1</sup> however these rates could vary depending upon abundance and diversity of microbial communities (Akinyede et al., 2020). Further studies exploring presence and abundance of CO<sub>2</sub> fixing bacteria in saltmarsh soils is recommended. The general lower nitrogen pollution in Australia's soils and waterways compared to other countries may partially explain the lower emissions. However, the GHG flux measurements from this study did not account for the effects of vegetation, which can alter fluxes. For instance, some plant species of rice paddies (Timilsina et al., 2020) and *Misanthus sinensis* (Lenhart et al., 2019) can increase N<sub>2</sub>O emissions, and some tree species can facilitate CH<sub>4</sub> efflux from the soil (Pangala et al., 2013). Finally, changes in emissions between low and high tides were detected for CO<sub>2</sub> and N<sub>2</sub>O. Thus, future studies that include vegetation and changes within tidal cycles will improve GHG flux estimates for coastal wetlands.

Table: 3. Comparison of GHG fluxes ( $\text{mg m}^{-2} \text{ d}^{-1}$ ) with other studies

Reference	Climate	Country	Ecosystem	CO <sub>2</sub> fluxes	CH <sub>4</sub> fluxes	N <sub>2</sub> O fluxes
Allen et al., 2011	Subtropical	Australia	Mangroves estuary	-	1.5-51	-
(Cabezas et al., 2018)	Subtropical	USA	Mangroves estuary	-	0.3-2.2	-
Li and Mitsch, 2016	Subtropical	USA	Flooded brackish marsh	-	$212 \pm 51$	-
Morse, Ardón and Bernhardt, 2012	Subtropical	USA	Forested wetlands	7224-11328	118-1255	46-279
Musenze et al., 2014	Subtropical	Australia	Mangroves estuary	-	5-448	0.1-3.4
Whiting and Chanton, 2001	Subtropical	USA	Typha marsh	409-477	189-264	-
Mitsch et al., 2001	Tropical	South Africa	Seasonally flooded wetland	-	$264 \pm 29$	-
Krithika et al., 2008	Tropical	India	Mangroves	-	25-50	-
Kristensen et al., 2008	Tropical	Tanzania	Mangroves	44-3521	1.9-6.5	-
Biswas et al., 2007	Tropical	India	Mangroves estuary	-	0.03-2.16	-
Purvaja et al., 2004	Tropical	India	Mangroves estuary	-	10-85	-
Kreuzwieser et al., 2003	Tropical	Australia	Mangroves	-	0.6-11	-

Kiese and Butterbach-Bahl, 2002	Tropical Australia	Tropical forest	rain	2208-3288	1.9-3.2	
Purvaja and Ramesh, 2000	Tropical India	Mangroves	-	63-434	-	
Sotomayor et al. 1994	Tropical Puerto Rico	Mangroves	-	5-110	-	
Barnes et al., 2006	Tropical India	Mangroves	-	9-15	-	
Melling et al. 2012	Tropical Malaysia	Peat swamp forest	3384	21-29		
This study	Tropical Australia	Freshwater forest	tidal	1640-10970	0.16-0.59	-0.19-0.7
This study	Tropical Australia	Saltmarsh	-594-(-1191)	-0.25-0.12	-0.22-2.76	
This study	Tropical Australia	Mangroves	2852-5669	0.44-3.95	0.04-0.16	
Oertel et al., 2016	(Sub) Tropical Global	Wetlands	-	-1.08-1169	-	
Oertel et al., 2016	Temperate Global	Wetlands	-	-1.49-1510	-	
Oertel et al., 2016	Mediterranea n	Wetlands	-	-	-2.6-9.4	
Al-Haj and Fulweiler, 2020	- Global	Mangroves	-	-1.1-1169	-0.2-6.3	
Al-Haj and Fulweiler, 2020	- Global	Saltmarshes	6844-34983	0.38-3002	-7.39-28.52	
Rosentreter et al., 2021	- Global	Mangroves	4563-30800	-0.69-10.78	-1.69-4.65	
Rosentreter et al., 2021	- Global	Saltmarshes	3802-20914	107-168	4.96	
IPCC 2013	Tropical Global	Swamp forests		30.76-2149		

300 Note: Hyphen means no data was available; GHG fluxes as CO<sub>2</sub>-C, CH<sub>4</sub>-C and N<sub>2</sub>O-N were multiplied by 3.66, 1.34 and 1.57 respectively to calculate CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O fluxes (National Greenhouse Accounts Factors, Australian Government Department of Industry, Science, Energy and Resources. 2020).

#### 4.1 Management implications

305 Under the Paris Agreement, Australia has committed to reducing GHG emissions 26 - 28% below its 2005 levels by 2030. With annual emissions of 153 million tonnes of carbon dioxide equivalent (Mt CO<sub>2</sub>-eq y<sup>-1</sup>), Queensland is a major GHG emitter in Australia (~ 28.7% of the total in 2016; DES, 2016). Of these emissions, about 18.3 Mt CO<sub>2</sub>-eq y<sup>-1</sup> (14%) are attributed to agriculture, while land-use change and forestry emit 12.1 Mt CO<sub>2</sub>-eq y<sup>-1</sup> (DES, 2016). Production of CH<sub>4</sub> from ruminant animals, primarily cattle, contribute 82% of agriculture-related emissions (DES, 2016). Therefore, any GHG mitigation strategy from 310 the land-use change could be important for Australia to achieve its national goals.

This study supports the application of three management actions that could reduce GHG emissions. First, the conversion of ponded pastures to coastal wetlands is likely to reduce soil GHG emissions. Our results showed that wet pastures emit 56 ton CO<sub>2</sub>-eq ha<sup>-1</sup> y<sup>-1</sup> of the total GHG (CH<sub>4</sub> + N<sub>2</sub>O) compared with 0.2 ton CO<sub>2</sub>-eq ha<sup>-1</sup> y<sup>-1</sup>, 0.1 ton CO<sub>2</sub>-eq ha<sup>-1</sup> y<sup>-1</sup> and 0.9 ton CO<sub>2</sub>-eq ha<sup>-1</sup> y<sup>-1</sup> from mangroves, freshwater tidal forest, and saltmarshes, respectively. This implies that about 55 ton CO<sub>2</sub>-eq ha<sup>-1</sup> y<sup>-1</sup> emissions from the soils could be potentially avoided by converting wet pastures to coastal wetlands. The carbon mitigation for GHG emissions only from soil could provide ~ AUD 894 ha<sup>-1</sup> yr<sup>-1</sup>, assuming a carbon value of AUD 15.99 per ton of CO<sub>2</sub>-eq (Australian Government Clean Energy Regulator, 2021). This mitigation could be added up to the carbon sequestration through sediment accumulation and tree growth that results from wetland restoration. Legal enablers in 320 Queensland are in place to manage unproductive agricultural land this way (Bell-James and Lovelock, 2019) and could provide an alternative income source for farmers.

A second management option would be to reduce the time pastures are kept underwater. Dry pastures produced 325 significantly less CH<sub>4</sub> with ~0.005 kg ha<sup>-1</sup> d<sup>-1</sup> than wet pastures with 6 kg ha<sup>-1</sup> d<sup>-1</sup>. For comparison, an average cow produces 141 g CH<sub>4</sub> d<sup>-1</sup> (McGinn et al., 2011), and our study farm supported around 900 cattle over 250 ha throughout the year, equivalent to 185 kg ha<sup>-1</sup> y<sup>-1</sup> compared to 2 kg ha<sup>-1</sup> y<sup>-1</sup> and 2,090 kg ha<sup>-1</sup> y<sup>-1</sup> CH<sub>4</sub> from dry and wet pasture respectively. This implies that nearly 92% of the CH<sub>4</sub> emissions came from wet pastures, while dry pasture and grazing cattle had a low share in total CH<sub>4</sub> emissions in this case scenario. Therefore, land use management of wet pastures may be an opportunity to reduce 330 agriculture-related CH<sub>4</sub> emissions. Future studies should increase the number of sites of ponded pastures to account for variability in hydrology, fertilisation, and on-farm cattle density. However, the exceptionally large difference (2-3 orders of

magnitude) between dry and ponded pastures and other coastal wetlands provides confidence that pasture management could provide significant GHG mitigation.

335 Finally, fertiliser management in sugarcane could reduce  $\text{N}_2\text{O}$  emissions. Higher  $\text{N}_2\text{O}$  emissions of  $17.6 \text{ mg m}^{-2} \text{ d}^{-1}$  were measured in the sugarcane crop following fertilisation during the dry-hot season. Comparatively, natural wetlands had low  $\text{N}_2\text{O}$  emissions ( $0.16$  to  $2.79 \text{ mg m}^{-2} \text{ d}^{-1}$ ), and even the saltmarsh was an occasional sink. Thus, improved management of fertiliser applications could result in GHG emission mitigation. Some activities include split application of nitrogen fertiliser in combination with low irrigation, reduction in fertiliser application rates, the substitution of nitrate-based fertiliser for urea  
340 (Rezaei Rashti et al., 2015), removing mulch layer before fertiliser application (Pinheiro et al., 2019; Xu et al., 2019; Zaehle and Dalmonech, 2011) or conversion of unproductive sugarcane to coastal wetlands.

## 5 Conclusion

The GHG emissions from three coastal wetlands in tropical Australia (mangroves, saltmarsh and freshwater tidal forests) were consistently lower than those from two common agricultural land use of the region (sugarcane and pastures) throughout three  
345 climatic conditions (dry-cool, dry-hot and wet-hot). Ponded pastures emitted 200 times more  $\text{CH}_4$ , and sugarcane emitted seven times more than any natural coastal wetland measured. If these high emissions are persistent in other locations and within other tropical regions, conversion of pastures and sugarcane to similar coastal wetlands could provide significant GHG mitigation. As nations try to reach their emission reduction targets, projects aimed at converting or restoring coastal wetlands can financially benefit farmers while providing additional co-benefits  
350

### Author contribution

Iram, N. and M.F. Adame designed the project, Iram, N, B. Shahrabi Farahani and E. Kavehei carried out experiments, Iram, N., E. Kavehei and M.F. Adame analysed the data. Iram, N and M.F. Adame prepared the manuscript with contributions from  
355 D.T. Maher, S.E., Bunn, and M. Rezaei Rashti.

### Competing interests

The authors declare that they have no conflict of interest.

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## References

Akinyede, R., Taubert, M., Schrumpf, M., Trumbore, S. and Küsel, K.: Rates of dark CO<sub>2</sub> fixation are driven by microbial  
370 biomass in a temperate forest soil, *Soil Biol. Biochem.*, 150, 107950, doi:10.1016/J.SOILBIO.2020.107950, 2020.

Allen, D., Dalal, R. C., Rennenberg, H. and Schmidt, S.: Seasonal variation in nitrous oxide and methane emissions from  
375 subtropical estuary and coastal mangrove sediments, Australia, *Plant Biol.*, 13(1), 126–133, doi:10.1111/j.1438-8677.2010.00331.x, 2011.

Angle, J. C., Morin, T. H., Soden, L. M., Narrowe, A. B., Smith, G. J., Borton, M. A., Rey-Sanchez, C., Daly, R. A.,  
380 Mirfenderesgi, G., Hoyt, D. W., Riley, W. J., Miller, C. S., Bohrer, G. and Wrighton, K. C.: Methanogenesis in oxygenated  
soils is a substantial fraction of wetland methane emissions, doi:10.1038/s41467-017-01753-4, n.d.

Barnes, J., Ramesh, R., Purvaja, R., Rajkumar, A. N., Kumar, B. S., Krithika, K., Ravichandran, K., Uher, G. and Upstill-  
385 Goddard, R.: Tidal dynamics and rainfall control N<sub>2</sub>O and CH<sub>4</sub> emissions from a pristine mangrove creek, *Geophys. Res. Lett.*, 33(15), doi:10.1029/2006GL026829, 2006.

Bauza, J. F., Morell, J. M. and Corredor, J. E.: Biogeochemistry of nitrous oxide production in the red mangrove (Rhizophora  
mangle) forest sediments, *Estuar. Coast. Shelf Sci.*, 55(5), 697–704, doi:10.1006/ecss.2001.0913, 2002.

Bell-James, J. and Lovelock, C. E.: Legal barriers and enablers for reintroducing tides: An Australian case study in reconvertig  
390 ponded pasture for climate change mitigation, *Land use policy*, 88, 104192, doi:10.1016/J.LANDUSEPOL.2019.104192,  
2019.

Biswas, H., Mukhopadhyay, S. K., Sen, S. and Jana, T. K.: Spatial and temporal patterns of methane dynamics in the tropical  
395 mangrove dominated estuary, NE coast of Bay of Bengal, India, *J. Mar. Syst.*, 68(1–2), 55–64,  
doi:10.1016/j.jmarsys.2006.11.001, 2007.

Cabezas, A., Mitsch, W. J., MacDonnell, C., Zhang, L., Bydałek, F. and Lasso, A.: Methane emissions from mangrove soils  
400 in hydrologically disturbed and reference mangrove tidal creeks in southwest Florida, *Ecol. Eng.*, 114, 57–65,  
doi:10.1016/j.ecoleng.2017.08.041, 2018.

Cameron, C., Hutley, L. B., Munksgaard, N. C., Phan, S., Aung, T., Thinn, T., Aye, W. M. and Lovelock, C. E.: Impact of an  
extreme monsoon on CO<sub>2</sub> and CH<sub>4</sub> fluxes from mangrove soils of the Ayeyarwady Delta, Myanmar, *Sci. Total Environ.*, 760,  
143422, doi:10.1016/J.SCITOTENV.2020.143422, 2021.

Chen, G. C., Tam, N. F. Y. and Ye, Y.: Spatial and seasonal variations of atmospheric N<sub>2</sub>O and CO<sub>2</sub> fluxes from a subtropical  
405 mangrove swamp and their relationships with soil characteristics, *Soil Biol. Biochem.*, 48, 175–181,  
doi:10.1016/j.soilbio.2012.01.029, 2012.

Deemer, B. R., Harrison, J. A., Li, S., Beaulieu, J. J., Delsontro, T., Barros, N., Bezerra-Neto, J. F., Powers, S. M., Santos, M.  
A. Dos and Vonk, J. A.: Greenhouse Gas Emissions from Reservoir Water Surfaces: A New Global Synthesis, *Bioscience*,  
66(11), doi:10.1093/biosci/biw117, 2016.

Ding, W. X., Cai, Z. C. and Tsuruta, H.: Cultivation, nitrogen fertilization, and set-aside effects on methane uptake in a drained

marsh soil in Northeast China, *Glob. Chang. Biol.*, 10(10), 1801–1809, doi:10.1111/j.1365-2486.2004.00843.x, 2004.

Drenovsky, R. E., Steenwerth, K. L., Jackson, L. E. and Scow, K. M.: Land use and climatic factors structure regional patterns in soil microbial communities, *Glob. Ecol. Biogeogr.*, 19(1), 27–39, doi:10.1111/j.1466-8238.2009.00486.x, 2010.

Drexler, J. Z., De Fontaine, C. S. and Deverel, S. J.: THE LEGACY OF WETLAND DRAINAGE ON THE REMAINING  
405 PEAT IN THE SACRAMENTO-SAN JOAQUIN DELTA, CALIFORNIA, USA., n.d.

Duarte, C. M., Losada, I. J., Hendriks, I. E., Mazarrasa, I. and Marbà, N.: The role of coastal plant communities for climate change mitigation and adaptation, *Nat. Clim. Chang.*, 3(11), 961–968, doi:10.1038/nclimate1970, 2013.

Fernandez-Bou, A. S., Dierick, D., Allen, M. F. and Harmon, T. C.: Precipitation-drainage cycles lead to hot moments in soil carbon dioxide dynamics in a Neotropical wet forest, *Glob. Chang. Biol.*, 26(9), 5303–5319, doi:10.1111/gcb.15194, 2020.

410 Griggs, P. D.: Too much water: drainage schemes and landscape change in the sugar-producing areas of Queensland, 1920–90, *Aust. Geogr.*, 49(1), 81–105, doi:10.1080/00049182.2017.1336965, 2018.

Grinham, A., Albert, S., Deering, N., Dunbabin, M., Bastviken, D., Sherman, B., Lovelock, C. E. and Evans, C. D.: The importance of small artificial water bodies as sources of methane emissions in Queensland, Australia, *Hydrol. Earth Syst. Sci.*, 22(10), 5281–5298, doi:10.5194/hess-22-5281-2018, 2018.

415 Hirano, T., Segah, H., Kusin, K., Limin, S., Takahashi, H. and Osaki, M.: Effects of disturbances on the carbon balance of tropical peat swamp forests, *Glob. Chang. Biol.*, 18(11), 3410–3422, doi:10.1111/j.1365-2486.2012.02793.x, 2012.

Hooijer, A., Page, S., Jauhainen, J., Lee, W. A., Lu, X. X., Idris, A. and Anshari, G.: Subsidence and carbon loss in drained tropical peatlands, *Biogeosciences*, 9(3), 1053–1071, doi:10.5194/bg-9-1053-2012, 2012.

Hutchinson, G. L. and Mosier, A. R.: Improved Soil Cover Method for Field Measurement of Nitrous Oxide Fluxes, *Soil Sci. Soc. Am. J.*, 45(2), 311–316, doi:10.2136/sssaj1981.03615995004500020017x, 1981.

420 Jeffrey, L. C., Maher, D. T., Chiri, E., Leung, P. M., Nauer, P. A., Arndt, S. K., Tait, D. R., Greening, C. and Johnston, S. G.: Bark-dwelling methanotrophic bacteria decrease methane emissions from trees, , doi:10.1038/s41467-021-22333-7, n.d.

Johnson, A. K. L., Ebert, S. P. and Murray, A. E.: Distribution of coastal freshwater wetlands and riparian forests in the Herbert River catchment and implications for management of catchments adjacent the Great Barrier Reef Marine Park, *Environ. Conserv.*, 26(3), 229–235, doi:10.1017/S0376892999000314, 1999.

425 Kauffman, J. B., Adame, M. F., Arifanti, V. B., Schile-Beers, L. M., Bernardino, A. F., Bhomia, R. K., Donato, D. C., Feller, I. C., Ferreira, T. O., Jesus Garcia, M. del C., MacKenzie, R. A., Megonigal, J. P., Murdiyarso, D., Simpson, L. and Hernández Trejo, H.: Total ecosystem carbon stocks of mangroves across broad global environmental and physical gradients, *Ecol. Monogr.*, 90(2), doi:10.1002/ecm.1405, 2020.

430 Kavehei, E., Iram, N., Rezaei Rashti, M., Jenkins, G. A., Lemckert, C. and Adame, M. F.: Greenhouse gas emissions from stormwater bioretention basins, *Ecol. Eng.*, 159, doi:10.1016/j.ecoleng.2020.106120, 2021.

Kettler, T. A., Doran, J. W. and Gilbert, T. L.: Simplified Method for Soil Particle-Size Determination to Accompany Soil-Quality Analyses, *Soil Sci. Soc. Am. J.*, 65(3), 849–852, doi:10.2136/sssaj2001.653849x, 2001.

Kiese, R. and Butterbach-Bahl, K.: N 2 O and CO 2 emissions from three different tropical forest sites in the wet tropics of

435 Queensland, Australia. [online] Available from: [www.elsevier.com/locate/soilbio](http://www.elsevier.com/locate/soilbio), n.d.

Kirschke, S., Bousquet, P., Ciais, P., Saunois, M., Canadell, J. G., Dlugokencky, E. J., Bergamaschi, P., Bergmann, D., Blake, D. R., Bruhwiler, L., Cameron-Smith, P., Castaldi, S., Chevallier, F., Feng, L., Fraser, A., Heimann, M., Hodson, E. L., Houweling, S., Josse, B., Fraser, P. J., Krummel, P. B., Lamarque, J. F., Langenfelds, R. L., Le Quéré, C., Naik, V., O'doherty, S., Palmer, P. I., Pison, I., Plummer, D., Poulter, B., Prinn, R. G., Rigby, M., Ringeval, B., Santini, M., Schmidt, M., Shindell, D. T., Simpson, I. J., Spahni, R., Steele, L. P., Strode, S. A., Sudo, K., Szopa, S., Van Der Werf, G. R., Voulgarakis, A., Van Weele, M., Weiss, R. F., Williams, J. E. and Zeng, G.: Three decades of global methane sources and sinks, *Nat. Geosci.*, 6(10), 813–823, doi:10.1038/ngeo1955, 2013.

Knox, S. H., Sturtevant, C., Matthes, J. H., Koteen, L., Verfaillie, J. and Baldocchi, D.: Agricultural peatland restoration: Effects of land-use change on greenhouse gas (CO<sub>2</sub> and CH<sub>4</sub>) fluxes in the Sacramento-San Joaquin Delta, *Glob. Chang. Biol.*, 21(2), 750–765, doi:10.1111/gcb.12745, 2015.

Kögel-Knabner, I., Amelung, W., Cao, Z., Fiedler, S., Frenzel, P., Jahn, R., Kalbitz, K., Kölbl, A. and Schloter, M.: Biogeochemistry of paddy soils, *Geoderma*, 157(1–2), 1–14, doi:10.1016/J.GEODERMA.2010.03.009, 2010.

Kreuzwieser, J., Buchholz, J. and Rennenberg, H.: Emission of Methane and Nitrous Oxide by Australian Mangrove Ecosystems, *Plant Biol.*, 5(4), 423–431, doi:10.1055/s-2003-42712, 2003.

Kristensen, E., Flindt, M. R., Uломи, S., Borges, A. V., Abril, G. and Bouillon, S.: Emission of CO<sub>2</sub> and CH<sub>4</sub> to the atmosphere by sediments and open waters in two Tanzanian mangrove forests, *Mar. Ecol. Prog. Ser.*, 370, 53–67, doi:10.3354/meps07642, 2008.

Krithika, K., Purvaja, R. and Ramesh, R.: Current Science Association Fluxes of methane and nitrous oxide from an Indian mangrove., 2008.

Kroeger, K. D., Crooks, S., Moseman-Valtierra, S. and Tang, J.: Restoring tides to reduce methane emissions in impounded wetlands: A new and potent Blue Carbon climate change intervention, *Sci. Rep.*, 7(1), doi:10.1038/s41598-017-12138-4, 2017.

Lehner, B. and Döll, P.: Development and validation of a global database of lakes, reservoirs and wetlands, *J. Hydrol.*, 296(1–4), 1–22, doi:10.1016/j.jhydrol.2004.03.028, 2004.

Lenhart, K., Behrendt, T., Greiner, S., Steinkamp, J., Well, R., Giesemann, A. and Keppler, F.: Nitrous oxide effluxes from plants as a potentially important source to the atmosphere, *New Phytol.*, 221(3), 1398–1408, doi:10.1111/nph.15455, 2019.

Li, X. and Mitsch, W. J.: Methane emissions from created and restored freshwater and brackish marshes in southwest Florida, USA, *Ecol. Eng.*, 91, 529–536, doi:10.1016/j.ecoleng.2016.01.001, 2016.

Liu, J. and Lai, D. Y. F.: Subtropical mangrove wetland is a stronger carbon dioxide sink in the dry than wet seasons, *Agric. For. Meteorol.*, 278, 107644, doi:10.1016/J.AGRFORMAT.2019.107644, 2019.

Maher, D. T., Sippo, J. Z., Tait, D. R., Holloway, C. and Santos, I. R.: Pristine mangrove creek waters are a sink of nitrous oxide OPEN, *Nat. Publ. Gr.*, doi:10.1038/srep25701, 2016.

Maietta, C. E., Hondula, K. L., Jones, C. N. and Palmer, M. A.: Hydrological Conditions Influence Soil and Methane-Cycling Microbial Populations in Seasonally Saturated Wetlands, *Front. Environ. Sci.*, 8, doi:10.3389/fenvs.2020.593942, 2020.

Mar Lynn, T., Ge, T., Yuan, H., Wei, X., Wu, X., Xiao, K., Kumaresan, D., San Yu, S., Wu, J. and Whiteley, A. S.: Soil Carbon-Fixation Rates and Associated Bacterial Diversity and Abundance in Three Natural Ecosystems, , doi:10.1007/s00248-016-0890-x, 2017.

Martiny, J. B. H., Bohannan, B. J. M., Brown, J. H., Colwell, R. K., Fuhrman, J. A., Green, J. L., Horner-Devine, M. C., Kane, M., Krumins, J. A., Kuske, C. R., Morin, P. J., Naeem, S., Øvreås, L., Reysenbach, A. L., Smith, V. H. and Staley, J. T.: Microbial biogeography: Putting microorganisms on the map, *Nat. Rev. Microbiol.*, 4(2), 102–112, doi:10.1038/nrmicro1341, 2006.

McGinn, S. M., Turner, D., Tomkins, N., Charmley, E., Bishop-Hurley, G. and Chen, D.: Methane Emissions from Grazing Cattle Using Point-Source Dispersion, *J. Environ. Qual.*, 40(1), 22–27, doi:10.2134/jeq2010.0239, 2011.

Mitsch, W. J., Bernal, B., Nahlik, A. M., Mander, Ü., Zhang, L., Anderson, C. J., Jørgensen, S. E. and Brix, H.: Wetlands, carbon, and climate change, *Landsc. Ecol.*, 28(4), 583–597, doi:10.1007/s10980-012-9758-8, 2013.

Musenze, R. S., Werner, U., Grinham, A., Udy, J. and Yuan, Z.: Methane and nitrous oxide emissions from a subtropical estuary (the Brisbane River estuary, Australia), *Sci. Total Environ.*, 472, 719–729, doi:10.1016/j.scitotenv.2013.11.085, 2014.

Neubauer, S. C.: Global Warming Potential Is Not an Ecosystem Property, *Ecosystems*, doi:10.1007/s10021-021-0063, n.d.

Nieveen, J. P., Campell, D. I., Schipper, L. A. and Blair, I. J.: Carbon exchange of grazed pasture on drained peat soil, *Glob. Chang. Biol.*, 11(4), 607–618, doi:10.1111/j.1365-2486.2005.00929.x, 2005.

Oertel, C., Matschullat, J., Zurba, K., Zimmermann, F. and Erasmi, S.: Greenhouse gas emissions from soils—A review, *Chemie der Erde*, 76(3), 327–352, doi:10.1016/j.chemer.2016.04.002, 2016.

Ollivier, Q. R., Maher, D. T., Pitfield, C. and Macreadie, P. I.: Punching above their weight: Large release of greenhouse gases from small agricultural dams, *Glob. Chang. Biol.*, 25(2), 721–732, doi:10.1111/gcb.14477, 2019.

Pangala, S. R., Moore, S., Hornibrook, E. R. C. and Gauci, V.: Trees are major conduits for methane egress from tropical forested wetlands, *New Phytol.*, 197(2), 524–531, doi:10.1111/nph.12031, 2013.

Pinheiro, P. L., Recous, S., Dietrich, G., Weiler, D. A., Schu, A. L., Bazzo, H. L. S. and Giacomini, S. J.: N<sub>2</sub>O emission increases with mulch mass in a fertilized sugarcane cropping system, *Biol. Fertil. Soils*, 55(5), 511–523, doi:10.1007/s00374-019-01366-7, 2019.

Purvaja, R. and Ramesh, R.: Human impacts on methane emission from mangrove ecosystems in India., n.d.

Purvaja, R., Ramesh, R. and Frenzel, P.: Plant-mediated methane emission from an Indian mangrove, *Glob. Chang. Biol.*, 10(11), 1825–1834, doi:10.1111/j.1365-2486.2004.00834.x, 2004.

Reeves, S., Wang, W., Salter, B. and Halpin, N.: Quantifying nitrous oxide emissions from sugarcane cropping systems: Optimum sampling time and frequency, *Atmos. Environ.*, 136, 123–133, doi:10.1016/j.atmosenv.2016.04.008, 2016.

Rezaei Rashti, M., Wang, W. J., Harper, S. M., Moody, P. W., Chen, C. R., Ghadiri, H. and Reeves, S. H.: Strategies to mitigate greenhouse gas emissions in intensively managed vegetable cropping systems in subtropical Australia, *Soil Res.*, 53(5), 475–484, doi:10.1071/SR14355, 2015.

Rezaei Rashti, M., Wang, W. J., Reeves, S. H., Harper, S. M., Moody, P. W. and Chen, C. R.: Linking chemical and

biochemical composition of plant materials to their effects on N<sub>2</sub>O emissions from a vegetable soil, *Soil Biol. Biochem.*, 103, 502–511, doi:10.1016/j.soilbio.2016.09.019, 2016.

505 Rosentreter, J. A., Al-Haj, A. N., Fulweiler, R. W. and Williamson, P.: Methane and Nitrous Oxide Emissions Complicate Coastal Blue Carbon Assessments, *Global Biogeochem. Cycles*, 35(2), doi:10.1029/2020GB006858, 2021.

Sahrawat, K. L.: Terminal electron acceptors for controlling methane emissions from submerged rice soils, *Commun. Soil Sci. Plant Anal.*, 35(9–10), 1401–1413, doi:10.1081/CSS-120037554, 2004.

Saunois, M., Stavert, A. R., Poulter, B., Bousquet, P., Canadell, J. G., Jackson, R. B., Raymond, P. A., Dlugokencky, E. J.,

510 Houweling, S., Patra, P. K., Ciais, P., Arora, V. K., Bastviken, D., Bergamaschi, P., Blake, D. R., Brailsford, G., Bruhwiler, L., Carlson, K. M., Carroll, M., Castaldi, S., Chandra, N., Crevoisier, C., Crill, P. M., Covey, K., Curry, C. L., Etiope, G., Frankenberg, C., Gedney, N., Hegglin, M. I., Höglund-Isaksson, L., Hugelius, G., Ishizawa, M., Ito, A., Janssens-Maenhout, G., Jensen, K. M., Joos, F., Kleinen, T., Krummel, P. B., Langenfelds, R. L., Laruelle, G. G., Liu, L., Machida, T., Maksyutov, S., McDonald, K. C., McNorton, J., Miller, P. A., Melton, J. R., Morino, I., Müller, J., Murguia-Flores, F., Naik, V., Niwa, Y.,

515 Noce, S., O'Doherty, S., Parker, R. J., Peng, C., Peng, S., Peters, G. P., Prigent, C., Prinn, R., Ramonet, M., Regnier, P., Riley, W. J., Rosentreter, J. A., Segers, A., Simpson, I. J., Shi, H., Smith, S. J., Steele, L. P., Thornton, B. F., Tian, H., Tohjima, Y., Tubiello, F. N., Tsuruta, A., Viovy, N., Voulgarakis, A., Weber, T. S., van Weele, M., van der Werf, G. R., Weiss, R. F., Worthy, D., Wunch, D., Yin, Y., Yoshida, Y., Zhang, W., Zhang, Z., Zhao, Y., Zheng, B., Zhu, Q., Zhu, Q. and Zhuang, Q.: The Global Methane Budget 2000–2017, *Earth Syst. Sci. Data*, 12(3), 1561–1623, doi:10.5194/essd-12-1561-2020, 2020.

520 Schindlbacher, A., Zechmeister-Boltenstern, S. and Butterbach-Bahl, K.: Effects of soil moisture and temperature on NO, NO<sub>x</sub> and N<sub>2</sub>O emissions from European forest soils, *J. Geophys. Res.*, 109, 17302, doi:10.1029/2004JD004590, 2004.

Shaaban, M., Peng, Q. an, Hu, R., Wu, Y., Lin, S. and Zhao, J.: Dolomite application to acidic soils: a promising option for mitigating N<sub>2</sub>O emissions, *Environ. Sci. Pollut. Res.*, 22(24), 19961–19970, doi:10.1007/s11356-015-5238-4, 2015.

525 Solomon and S.: Climate Change : The Physical science Basis, *Clim. Chang. Phys. Sci. Basis* [online] Available from: <http://ci.nii.ac.jp/naid/20001628955/en/> (Accessed 13 August 2021), 2007.

Timilsina, A., Bizimana, F., Pandey, B., Kailash, R., Yadav, P., Dong, W. and Hu, C.: plants Nitrous Oxide Emissions from Paddies: Understanding the Role of Rice Plants, , doi:10.3390/plants9020180, n.d.

Veenendaal, E. M., Kolle, O., Leffelaar, P. A., Schrier-Uijl, A. P., Van Huissteden, J., Walssem, J. Van, Möller, F. and 530 Berendse, F.: CO<sub>2</sub> exchange and carbon balance in two grassland sites on eutrophic drained peat soils. [online] Available from: [www.biogeosciences.net/4/1027/2007/](http://www.biogeosciences.net/4/1027/2007/), 2007.

Whiting, G. J. and Chanton, J. P.: Greenhouse carbon balance of wetlands: Methane emission versus carbon sequestration, *Tellus, Ser. B Chem. Phys. Meteorol.*, 53(5), 521–528, doi:10.3402/tellusb.v53i5.16628, 2001.

Xu, C., Han, X., Ru, S., Cardenas, L., Rees, R. M., Wu, D., Wu, W. and Meng, F.: Crop straw incorporation interacts with N 535 fertilizer on N<sub>2</sub>O emissions in an intensively cropped farmland, *Geoderma*, 341, 129–137, doi:10.1016/J.GEODERMA.2019.01.014, 2019.

Formatted: Normal

Yvon-Durocher, G., Montoya, J. M., Woodward, G., Jones, J. I. and Trimmer, M.: Warming increases the proportion of primary production emitted as methane from freshwater mesocosms, *Glob. Chang. Biol.*, 17(2), 1225–1234, doi:10.1111/j.1365-2486.2010.02289.x, 2011.

540 Zaehle, S. and Dalmonech, D.: Carbon-nitrogen interactions on land at global scales: Current understanding in modelling climate biosphere feedbacks, *Curr. Opin. Environ. Sustain.*, 3(5), 311–320, doi:10.1016/j.cosust.2011.08.008, 2011.

Al-Haj, A. N., and Fulweiler, R. W.: A synthesis of methane emissions from shallow vegetated coastal ecosystems. *Global Change Biology*, 26(5), 2988–3005. <https://doi.org/10.1111/gcb.15046>, 2020.

545 Akinyede, R., Taubert, M., Schrumpf, M., Trumbore, S. and Küsel, K.: Rates of dark CO<sub>2</sub> fixation are driven by microbial biomass in a temperate forest soil, *Soil Biol. Biochem.*, 150, 107950, doi:10.1016/J.SOILBIO.2020.107950, 2020.

Allen, D., Dalal, R. C., Rennenberg, H. and Schmidt, S.: Seasonal variation in nitrous oxide and methane emissions from subtropical estuary and coastal mangrove sediments, Australia, *Plant Biol.*, 13(1), 126–133, doi:10.1111/j.1438-8677.2010.00331.x, 2011.

550 Angle, J. C., Morin, T. H., Soden, L. M., Narrope, A. B., Smith, G. J., Borton, M. A., Rey-Sanchez, C., Daly, R. A., Mirfenderesgi, G., Hoyt, D. W., Riley, W. J., Miller, C. S., Bohrer, G. and Wrighton, K. C.: Methanogenesis in oxygenated soils is a substantial fraction of wetland methane emissions, doi:10.1038/s41467-017-01753-4, n.d.

Barnes, J., Ramesh, R., Purvaja, R., Rajkumar, A. N., Kumar, B. S., Krithika, K., Ravichandran, K., Uher, G. and Upstill-Goddard, R.: Tidal dynamics and rainfall control N<sub>2</sub>O and CH<sub>4</sub> emissions from pristine mangrove creek, *Geophys. Res. Lett.*, 33(15), doi:10.1029/2006GL026829, 2006.

Bauza, J. F., Morell, J. M. and Corredor, J. E.: Biogeochemistry of nitrous oxide production in the red mangrove (Rhizophora mangle) forest sediments, *Estuar. Coast. Shelf Sci.*, 55(5), 697–704, doi:10.1006/ecss.2001.0913, 2002.

Bell-James, J. and Lovelock, C. E.: Legal barriers and enablers for reintroducing tides: An Australian case study in reconvert ing ponded pasture for climate change mitigation, *Land use policy*, 88, 104192, doi:10.1016/J.LANDUSEPOL.2019.104192, 560 2019.

Biswas, H., Mukhopadhyay, S. K., Sen, S. and Jana, T. K.: Spatial and temporal patterns of methane dynamics in the tropical mangrove dominated estuary, NE coast of Bay of Bengal, India, *J. Mar. Syst.*, 68(1–2), 55–64, doi:10.1016/j.jmarsys.2006.11.001, 2007.

565 Cabezas, A., Mitsch, W. J., MacDonnell, C., Zhang, L., Bydałek, F. and Lasso, A.: Methane emissions from mangrove soils in hydrologically disturbed and reference mangrove tidal creeks in southwest Florida, *Ecol. Eng.*, 114, 57–65, doi:10.1016/j.ecoleng.2017.08.041, 2018.

Cameron, C., Hutley, L. B., Munksgaard, N. C., Phan, S., Aung, T., Thinn, T., Aye, W. M. and Lovelock, C. E.: Impact of an extreme monsoon on CO<sub>2</sub> and CH<sub>4</sub> fluxes from mangrove soils of the Ayeyarwady Delta, Myanmar, *Sci. Total Environ.*, 760, 143422, doi:10.1016/J.SCITOTENV.2020.143422, 2021.

570 Chen, G. C., Tam, N. F. Y. and Ye, Y.: Spatial and seasonal variations of atmospheric N<sub>2</sub>O and CO<sub>2</sub> fluxes from a subtropical

mangrove swamp and their relationships with soil characteristics, *Soil Biol. Biochem.*, 48, 175–181, doi:10.1016/j.soilbio.2012.01.029, 2012.

Deemer, B. R., Harrison, J. A., Li, S., Beaulieu, J. J., Delsontro, T., Barros, N., Bezerra-Neto, J. F., Powers, S. M., Santos, M. A. Dos and Vonk, J. A.: Greenhouse Gas Emissions from Reservoir Water Surfaces: A New Global Synthesis, *Bioscience*, 575 66(11), doi:10.1093/biosci/biw117, 2016.

Ding, W. X., Cai, Z. C. and Tsuruta, H.: Cultivation, nitrogen fertilization, and set-aside effects on methane uptake in a drained marsh soil in Northeast China, *Glob. Chang. Biol.*, 10(10), 1801–1809, doi:10.1111/j.1365-2486.2004.00843.x, 2004.

Drenovsky, R. E., Steenwerth, K. L., Jackson, L. E. and Scow, K. M.: Land use and climatic factors structure regional patterns in soil microbial communities, *Glob. Ecol. Biogeogr.*, 19(1), 27–39, doi:10.1111/j.1466-8238.2009.00486.x, 2010.

580 Drexler, J. Z., De Fontaine, C. S. and Deverel, S. J.: THE LEGACY OF WETLAND DRAINAGE ON THE REMAINING PEAT IN THE SACRAMENTO-SAN JOAQUIN DELTA, CALIFORNIA, USA., n.d.

Duarte, C. M., Losada, I. J., Hendriks, I. E., Mazarrasa, I. and Marbà, N.: The role of coastal plant communities for climate change mitigation and adaptation, *Nat. Clim. Chang.*, 3(11), 961–968, doi:10.1038/nclimate1970, 2013.

585 Fernandez-Bou, A. S., Dierick, D., Allen, M. F. and Harmon, T. C.: Precipitation-drainage cycles lead to hot moments in soil carbon dioxide dynamics in a Neotropical wet forest, *Glob. Chang. Biol.*, 26(9), 5303–5319, doi:10.1111/gcb.15194, 2020.

Griggs, P. D.: Too much water: drainage schemes and landscape change in the sugar-producing areas of Queensland, 1920–90, *Aust. Geogr.*, 49(1), 81–105, doi:10.1080/00049182.2017.1336965, 2018.

590 Grinham, A., Albert, S., Deering, N., Dunbabin, M., Bastviken, D., Sherman, B., Lovelock, C. E. and Evans, C. D.: The importance of small artificial water bodies as sources of methane emissions in Queensland, Australia, *Hydrol. Earth Syst. Sci.*, 22(10), 5281–5298, doi:10.5194/hess-22-5281-2018, 2018.

Hirano, T., Segah, H., Kusin, K., Limin, S., Takahashi, H. and Osaki, M.: Effects of disturbances on the carbon balance of tropical peat swamp forests, *Glob. Chang. Biol.*, 18(11), 3410–3422, doi:10.1111/j.1365-2486.2012.02793.x, 2012.

Hooijer, A., Page, S., Jauhainen, J., Lee, W. A., Lu, X. X., Idris, A. and Anshari, G.: Subsidence and carbon loss in drained tropical peatlands, *Biogeosciences*, 9(3), 1053–1071, doi:10.5194/bg-9-1053-2012, 2012.

595 Hutchinson, G. L. and Mosier, A. R.: Improved Soil Cover Method for Field Measurement of Nitrous Oxide Fluxes, *Soil Sci. Soc. Am. J.*, 45(2), 311–316, doi:10.2136/sssaj1981.03615995004500020017x, 1981.

Jeffrey, L. C., Maher, D. T., Chiri, E., Leung, P. M., Nauer, P. A., Arndt, S. K., Tait, D. R., Greening, C. and Johnston, S. G.: Bark-dwelling methanotrophic bacteria decrease methane emissions from trees, , doi:10.1038/s41467-021-22333-7, n.d.

Johnson, A. K. L., Ebert, S. P. and Murray, A. E.: Distribution of coastal freshwater wetlands and riparian forests in the Herbert 600 River catchment and implications for management of catchments adjacent the Great Barrier Reef Marine Park, *Environ. Conserv.*, 26(3), 229–235, doi:10.1017/S037689299000314, 1999.

Kauffman, J. B., Adame, M. F., Arifanti, V. B., Schile-Beers, L. M., Bernardino, A. F., Bhomia, R. K., Donato, D. C., Feller, I. C., Ferreira, T. O., Jesus Garcia, M. del C., MacKenzie, R. A., Megonigal, J. P., Murdiyarso, D., Simpson, L. and Hernández Trejo, H.: Total ecosystem carbon stocks of mangroves across broad global environmental and physical gradients, *Ecol.*

605 Monogr., 90(2), doi:10.1002/ecm.1405, 2020.

Kavehei, E., Iram, N., Rezaei Rashti, M., Jenkins, G. A., Lemckert, C. and Adame, M. F.: Greenhouse gas emissions from stormwater bioretention basins, *Ecol. Eng.*, 159, doi:10.1016/j.ecoleng.2020.106120, 2021.

Kettler, T. A., Doran, J. W. and Gilbert, T. L.: Simplified Method for Soil Particle-Size Determination to Accompany Soil-Quality Analyses, *Soil Sci. Soc. Am. J.*, 65(3), 849–852, doi:10.2136/sssaj2001.653849x, 2001.

610 Kiese, R. and Butterbach-Bahl, K.: N 2 O and CO 2 emissions from three different tropical forest sites in the wet tropics of Queensland, Australia. [online] Available from: [www.elsevier.com/locate/soilbio](http://www.elsevier.com/locate/soilbio), n.d.

Kirschke, S., Bousquet, P., Ciais, P., Saunois, M., Canadell, J. G., Dlugokencky, E. J., Bergamaschi, P., Bergmann, D., Blake, D. R., Bruhwiler, L., Cameron-Smith, P., Castaldi, S., Chevallier, F., Feng, L., Fraser, A., Heimann, M., Hodson, E. L., Houweling, S., Josse, B., Fraser, P. J., Krummel, P. B., Lamarque, J. F., Langenfelds, R. L., Le Quéré, C., Naik, V., O'doherty, S., Palmer, P. I., Pison, I., Plummer, D., Poulter, B., Prinn, R. G., Rigby, M., Ringeval, B., Santini, M., Schmidt, M., Shindell, D. T., Simpson, I. J., Spahni, R., Steele, L. P., Strode, S. A., Sudo, K., Szopa, S., Van Der Werf, G. R., Voulgarakis, A., Van Weele, M., Weiss, R. F., Williams, J. E. and Zeng, G.: Three decades of global methane sources and sinks, *Nat. Geosci.*, 6(10), 813–823, doi:10.1038/ngeo1955, 2013.

Knox, S. H., Sturtevant, C., Matthes, J. H., Koteen, L., Verfaillie, J. and Baldocchi, D.: Agricultural peatland restoration: 620 Effects of land-use change on greenhouse gas (CO2 and CH4) fluxes in the Sacramento-San Joaquin Delta, *Glob. Chang. Biol.*, 21(2), 750–765, doi:10.1111/gcb.12745, 2015.

Kögel-Knabner, I., Amelung, W., Cao, Z., Fiedler, S., Frenzel, P., Jahn, R., Kalbitz, K., Kölbl, A. and Schloter, M.: Biogeochemistry of paddy soils, *Geoderma*, 157(1–2), 1–14, doi:10.1016/J.GEODERMA.2010.03.009, 2010.

Kreuzwieser, J., Buchholz, J. and Rennenberg, H.: Emission of Methane and Nitrous Oxide by Australian Mangrove 625 Ecosystems, *Plant Biol.*, 5(4), 423–431, doi:10.1055/s-2003-42712, 2003.

Kristensen, E., Flindt, M. R., Uломи, S., Borges, A. V., Abril, G. and Bouillon, S.: Emission of CO2 and CH4 to the atmosphere by sediments and open waters in two Tanzanian mangrove forests, *Mar. Ecol. Prog. Ser.*, 370, 53–67, doi:10.3354/meps07642, 2008.

Kritihika, K., Purvaja, R. and Ramesh, R.: Current Science Association Fluxes of methane and nitrous oxide from an Indian 630 mangrove., 2008.

Kroeger, K. D., Crooks, S., Moseman-Valtierra, S. and Tang, J.: Restoring tides to reduce methane emissions in impounded wetlands: A new and potent Blue Carbon climate change intervention, *Sci. Rep.*, 7(1), doi:10.1038/s41598-017-12138-4, 2017.

Lehner, B. and Döll, P.: Development and validation of a global database of lakes, reservoirs and wetlands, *J. Hydrol.*, 296(1–4), 1–22, doi:10.1016/j.jhydrol.2004.03.028, 2004.

635 Lenhart, K., Behrendt, T., Greiner, S., Steinkamp, J., Well, R., Giesemann, A. and Keppler, F.: Nitrous oxide effluxes from plants as a potentially important source to the atmosphere, *New Phytol.*, 221(3), 1398–1408, doi:10.1111/nph.15455, 2019.

Li, X. and Mitsch, W. J.: Methane emissions from created and restored freshwater and brackish marshes in southwest Florida, USA, *Ecol. Eng.*, 91, 529–536, doi:10.1016/j.ecoleng.2016.01.001, 2016.

Liu, J. and Lai, D. Y. F.: Subtropical mangrove wetland is a stronger carbon dioxide sink in the dry than wet seasons, *Agric. For. Meteorol.*, 278, 107644, doi:10.1016/J.AGRFORMAT.2019.107644, 2019.

Maher, D. T., Sippo, J. Z., Tait, D. R., Holloway, C. and Santos, I. R.: Pristine mangrove creek waters are a sink of nitrous oxide *OPEN, Nat. Publ. Gr.*, doi:10.1038/srep25701, 2016.

Maietta, C. E., Hondula, K. L., Jones, C. N. and Palmer, M. A.: Hydrological Conditions Influence Soil and Methane-Cycling Microbial Populations in Seasonally Saturated Wetlands, *Front. Environ. Sci.*, 8, doi:10.3389/fenvs.2020.593942, 2020.

Mar Lynn, T., Ge, T., Yuan, H., Wei, X., Wu, X., Xiao, K., Kumaresan, D., San Yu, S., Wu, J. and Whiteley, A. S.: Soil Carbon-Fixation Rates and Associated Bacterial Diversity and Abundance in Three Natural Ecosystems, , doi:10.1007/s00248-016-0890-x, n.d.

Martiny, J. B. H., Bohannan, B. J. M., Brown, J. H., Colwell, R. K., Fuhrman, J. A., Green, J. L., Horner-Devine, M. C., Kane, M., Krumins, J. A., Kuske, C. R., Morin, P. J., Naeem, S., Øvreås, L., Reysenbach, A. L., Smith, V. H. and Staley, J. T.: Microbial biogeography: Putting microorganisms on the map, *Nat. Rev. Microbiol.*, 4(2), 102–112, doi:10.1038/nrmicro1341, 2006.

McGinn, S. M., Turner, D., Tomkins, N., Charmley, E., Bishop-Hurley, G. and Chen, D.: Methane Emissions from Grazing Cattle Using Point-Source Dispersion, *J. Environ. Qual.*, 40(1), 22–27, doi:10.2134/jeq2010.0239, 2011.

Mitsch, W. J., Bernal, B., Nahlik, A. M., Mander, Ü., Zhang, L., Anderson, C. J., Jørgensen, S. E. and Brix, H.: Wetlands, carbon, and climate change, *Landsc. Ecol.*, 28(4), 583–597, doi:10.1007/s10980-012-9758-8, 2013.

Musenze, R. S., Werner, U., Grinham, A., Udy, J. and Yuan, Z.: Methane and nitrous oxide emissions from a subtropical estuary (the Brisbane River estuary, Australia), *Sci. Total Environ.*, 472, 719–729, doi:10.1016/j.scitotenv.2013.11.085, 2014.

Neubauer, S. C.: Global Warming Potential Is Not an Ecosystem Property, *Ecosystems*, doi:10.1007/s10021-021-0063, n.d.

Nieveen, J. P., Campell, D. I., Schipper, L. A. and Blair, I. J.: Carbon exchange of grazed pasture on drained peat soil, *Glob. Chang. Biol.*, 11(4), 607–618, doi:10.1111/j.1365-2486.2005.00929.x, 2005.

Oertel, C., Matschullat, J., Zurba, K., Zimmermann, F. and Erasmi, S.: Greenhouse gas emissions from soils—A review, *Chemie der Erde*, 76(3), 327–352, doi:10.1016/j.chemer.2016.04.002, 2016.

Ollivier, Q. R., Maher, D. T., Pitfield, C. and Macreadie, P. I.: Punching above their weight: Large release of greenhouse gases from small agricultural dams, *Glob. Chang. Biol.*, 25(2), 721–732, doi:10.1111/gcb.14477, 2019.

Pangala, S. R., Moore, S., Hornibrook, E. R. C. and Gauci, V.: Trees are major conduits for methane egress from tropical forested wetlands, *New Phytol.*, 197(2), 524–531, doi:10.1111/nph.12031, 2013.

Pinheiro, P. L., Recous, S., Dietrich, G., Weiler, D. A., Schu, A. L., Bazzo, H. L. S. and Giacomini, S. J.: N<sub>2</sub>O emission increases with mulch mass in a fertilized sugarcane cropping system, *Biol. Fertil. Soils*, 55(5), 511–523, doi:10.1007/s00374-019-01366-7, 2019.

Purvaja, R. and Ramesh, R.: Human impacts on methane emission from mangrove ecosystems in India., n.d.

Purvaja, R., Ramesh, R. and Frenzel, P.: Plant-mediated methane emission from an Indian mangrove, *Glob. Chang. Biol.*, 10(11), 1825–1834, doi:10.1111/j.1365-2486.2004.00834.x, 2004.

Reeves, S., Wang, W., Salter, B. and Halpin, N.: Quantifying nitrous oxide emissions from sugarcane cropping systems: Optimum sampling time and frequency, *Atmos. Environ.*, 136, 123–133, doi:10.1016/j.atmosenv.2016.04.008, 2016.

675 Rezaei Rashti, M., Wang, W. J., Harper, S. M., Moody, P. W., Chen, C. R., Ghadiri, H. and Reeves, S. H.: Strategies to mitigate greenhouse gas emissions in intensively managed vegetable cropping systems in subtropical Australia, *Soil Res.*, 53(5), 475–484, doi:10.1071/SR14355, 2015.

Rezaei Rashti, M., Wang, W. J., Reeves, S. H., Harper, S. M., Moody, P. W. and Chen, C. R.: Linking chemical and biochemical composition of plant materials to their effects on N<sub>2</sub>O emissions from a vegetable soil, *Soil Biol. Biochem.*, 103, 680 502–511, doi:10.1016/j.soilbio.2016.09.019, 2016.

Rosentreter, J. A., Al-Haj, A. N., Fulweiler, R. W. and Williamson, P.: Methane and Nitrous Oxide Emissions Complicate Coastal Blue Carbon Assessments, *Global Biogeochem. Cycles*, 35(2), doi:10.1029/2020GB006858, 2021.

Sahrawat, K. L.: Terminal electron acceptors for controlling methane emissions from submerged rice soils, *Commun. Soil Sci. Plant Anal.*, 35(9–10), 1401–1413, doi:10.1081/CSS-120037554, 2004.

685 Saunois, M., Stavert, A. R., Poulter, B., Bousquet, P., Canadell, J. G., Jackson, R. B., Raymond, P. A., Dlugokencky, E. J., Houweling, S., Patra, P. K., Ciais, P., Arora, V. K., Bastviken, D., Bergamaschi, P., Blake, D. R., Brailsford, G., Bruhwiler, L., Carlson, K. M., Carroll, M., Castaldi, S., Chandra, N., Crevoisier, C., Crill, P. M., Covey, K., Curry, C. L., Etiope, G., Frankenberg, C., Gedney, N., Hegglin, M. I., Höglund-Isaksson, L., Hugelius, G., Ishizawa, M., Ito, A., Janssens-Maenhout, G., Jensen, K. M., Joos, F., Kleinen, T., Krummel, P. B., Langenfelds, R. L., Laruelle, G. G., Liu, L., Machida, T., Maksyutov, S., McDonald, K. C., McNorton, J., Miller, P. A., Melton, J. R., Morino, I., Müller, J., Murguia-Flores, F., Naik, V., Niwa, Y., Noce, S., O'Doherty, S., Parker, R. J., Peng, C., Peng, S., Peters, G. P., Prigent, C., Prinn, R., Ramonet, M., Regnier, P., Riley, W. J., Rosentreter, J. A., Segers, A., Simpson, I. J., Shi, H., Smith, S. J., Steele, L. P., Thornton, B. F., Tian, H., Tohjima, Y., Tubiello, F. N., Tsuruta, A., Viovy, N., Voulgarakis, A., Weber, T. S., van Weele, M., van der Werf, G. R., Weiss, R. F., Worthy, D., Wunch, D., Yin, Y., Yoshida, Y., Zhang, W., Zhang, Z., Zhao, Y., Zheng, B., Zhu, Q., Zhu, Q. and Zhuang, Q.: 690 The Global Methane Budget 2000–2017, *Earth Syst. Sci. Data*, 12(3), 1561–1623, doi:10.5194/essd-12-1561-2020, 2020.

Schindlbacher, A., Zechmeister-Boltenstern, S. and Butterbach-Bahl, K.: Effects of soil moisture and temperature on NO, NO<sub>x</sub>, and N<sub>2</sub>O emissions from European forest soils, *J. Geophys. Res.*, 109, 17302, doi:10.1029/2004JD004590, 2004.

700 Shaaban, M., Peng, Q. an, Hu, R., Wu, Y., Lin, S. and Zhao, J.: Dolomite application to acidic soils: a promising option for mitigating N<sub>2</sub>O emissions, *Environ. Sci. Pollut. Res.*, 22(24), 19961–19970, doi:10.1007/s11356-015-5238-4, 2015.

Solomon and S.: Climate Change : The Physical science Basis, Clim. Chang. Phys. Sci. Basis [online] Available from: <http://ci.nii.ac.jp/naid/20001628955/en/> (Accessed 13 August 2021), 2007.

Timilsina, A., Bizimana, F., Pandey, B., Kailash, R., Yadav, P., Dong, W. and Hu, C.: plants Nitrous Oxide Emissions from Paddies: Understanding the Role of Rice Plants, , doi:10.3390/plants9020180, n.d.

705 Veenendaal, E. M., Kolle, O., Leffelaar, P. A., Schrier-Uijl, A. P., Van Huissteden, J., Walssem, J. Van, Möller, F. and Berendse, F.: CO<sub>2</sub> exchange and carbon balance in two grassland sites on eutrophic drained peat soils. [online] Available

from: [www.biogeosciences.net/4/1027/2007/](http://www.biogeosciences.net/4/1027/2007/), 2007.

Whiting, G. J. and Chanton, J. P.: Greenhouse carbon balance of wetlands: Methane emission versus carbon sequestration, *Tellus, Ser. B Chem. Phys. Meteorol.*, 53(5), 521–528, doi:10.3402/tellusb.v53i5.16628, 2001.

710 Xu, C., Han, X., Ru, S., Cardenas, L., Rees, R. M., Wu, D., Wu, W. and Meng, F.: Crop straw incorporation interacts with N fertilizer on N<sub>2</sub>O emissions in an intensively cropped farmland, *Geoderma*, 341, 129–137, doi:10.1016/J.GEODERMA.2019.01.014, 2019.

Yvon-Durocher, G., Montoya, J. M., Woodward, G., Jones, J. I. and Trimmer, M.: Warming increases the proportion of primary production emitted as methane from freshwater mesocosms, *Glob. Chang. Biol.*, 17(2), 1225–1234,

715 doi:10.1111/j.1365-2486.2010.02289.x, 2011.

Zaehle, S. and Dalmonech, D.: Carbon-nitrogen interactions on land at global scales: Current understanding in modelling climate biosphere feedbacks, *Curr. Opin. Environ. Sustain.*, 3(5), 311–320, doi:10.1016/j.cosust.2011.08.008, 2011.