Greenhouse gas fluxes in mangrove forest soil in an Amazon estuary

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Abstract: Tropical mangrove forests are important carbon sinks, the soil being the main carbon reservoir. Understanding the variability and the key factors that control fluxes is critical to accounting for greenhouse gas (GHG) emissions, particularly in the current scenario of global climate change. This study is the first to quantify methane (CH_4) and carbon dioxide (CO_2) emissions using a dynamic chamber in a natural mangrove soil of the Amazon. The plots for the trace gases study were allocated at contrasting topographic heights. The results showed that the mangrove soil of the Amazon estuary is a source of CO_2 (6.66 g CO_2 m^{-2} d^{-1}) and CH_4 (0.13 g CH_4 m^{-2} d^{-1}) to the atmosphere. The CO_2 flux was higher in the high topography (7.858 g CO_2 m^{-2} d^{-1}) than in the low topography (4.734 g CO_2 m^{-2} d^{-1}) in the rainy season, and CH_4 was higher in the low topography (0.128 g CH_4 m^{-2} d^{-1}) than in the high topography (0.014 g CH_4 m^{-2} d^{-1}) in the dry season. However, in the dry period, the low topography soil produced more CH_4. Soil organic matter, carbon and nitrogen ratio (C/N), and redox potential influenced the annual and seasonal variation of CO_2 emissions; however, they did not affect CH_4 flux. The mangrove soil of the Amazon estuary produced 35.4 Mg $CO_{2\text{-eq}}$ ha^{-1} y^{-1}. A total of 2.16 kg CO_2 m^{-2} y^{-1} needs to be sequestered by the mangrove ecosystem to counterbalance CH_4 emissions.

1 Introduction

The mangrove areas are estimated to be the main contributors to greenhouse gas emissions in marine ecosystems (Allen et al., 2011; Chen et al., 2012). However, mangrove forests are highly productive due to a high nutrient turnover rate (Robertson et al., 1992) and have mechanisms that maximize carbon gain and minimize water loss through plant transpiration (Alongi and Mukhopadhyay, 2015). A study conducted in 25 mangrove forests (between 30° latitude and 73° longitude) revealed that these forests are the richest in carbon storage in the tropics, containing on average 1,023 Mg C ha^{-1} of which 49 to 98% is present in the soil (Donato et al., 2011).

The estimated soil CO_2 outgassing, in tropical estuarine areas is 16.2 Tg C y^{-1} (Alongi, 2009). However, soil efflux measurements from tropical mangroves revealed emissions ranging from 2.9 to 11.0 g CO_2 m^{-2} d^{-1} (Castillo et al., 2017; Chen et al., 2014; Shiau and Chiu, 2020). In situ CO_2 production is related to the water input of terrestrial, riparian, and groundwater brought by rainfall (Rosentreter et al., 2018b). Due to the periodic tidal movement, the mangrove ecosystem is daily flooded, leaving the soil anoxic and consequently reduced, favoring methanogenesis (Dutta et al., 2013). Thus, estuaries are considered hotspots for CH_4 production and emission (Bastviken et al., 2011; Borges et al., 2015). The organic material decomposition by methanogenic bacteria in anoxic environments, such as sediments, inner suspended particles, zooplankton gut (Reeburgh, 2007; Valentine, 2011), and the impact of freshwater should change the electron flow from sulfate-reducing bacteria to methanogenesis (Purvaja et al., 2004), which also results in CH_4 formation. On the other hand, an ecosystem with salinity levels above 18 ppt may show an absence of CH_4 emissions (Poffenbarger et al., 2011), since methane dissolved in pores is typically oxidized anaerobically by sulfate (Chuang et al., 2016). Currently the uncertainty in emitted CH_4 values in vegetated coastal wetlands is approximately 30% (EPA, 2017). Soil flux measurements from tropical mangroves revealed emissions ranging from 0.3 to 4.4 mg CH_4 m^{-2} d^{-1} (Castillo et al., 2017; Chen et al., 2014; Kreuzwieser et al., 2003).

The production of greenhouse gases from soils is mainly driven by biogeochemical processes. Microbial activities and gas production are related to soil properties, including total carbon and nitrogen concentrations, moisture, porosity, salinity, and redox potential (Bouillon et al., 2008; Chen et al., 2012). Due to the dynamics of tidal movements, mangrove soils may become saturated and present a reduced oxygen availability, or suffer total aeration caused by the ebb tide. Studies attribute soil carbon flux responses to moisture perturbations because of seasonality and flooding events (Banerjee et al., 2016), with fluxes being dependent on tidal extremes (high tide and low tide), and flood duration (Chowdhury et al., 2018). In addition, phenolic compounds inhibit microbial activity and help keep organic carbon intact, thus leading to the accumulation of organic matter in mangrove forest soils (Friesen et al., 2018).

The Amazonian coastal areas in the State of Pará (Brazil) cover 2,176.8 km^2 where mangroves develop under the macro-tide regime (Souza Filho, 2005), representing approximately 85% of the entire area of Brazilian mangroves (Herz, 1991). The objective of this study is to investigate the monthly flux of CO_2 and CH_4 from the soil, at two topographic heights, in a pristine mangrove area in the Mojuim River Estuary, belonging to the Amazon biome. The gas fluxes were studied together with the analysis of the vegetation structure and soil physical-chemical parameters.

2 Material and Methods

2.1 Study site

This study was conducted in the Amazonian coastal zone, Macaca Island (-0.746491 latitude and -47.997219 longitude), located in the Mojuim River estuary, at the Mocapajuba Marine Extractive Reserve, municipality of São Caetano de Odivelas (Figure 1), state of Pará (Brazil). The Macaca island has an area of 1,322 ha of pristine mangroves, which belongs to a mangrove area of 2,177 km^2 in the state of Pará (Souza Filho, 2005). The climate is type Am (tropical monsoon) according to the Köppen classification (Peel et al., 2007). The climatological data were obtained from the Meteorological Database for Teaching and Research of the National Institute of Meteorology (INMET). The area has a rainy season from January to June (2,296 mm of precipitation) and a dry season from July to December (687 mm). March and April were the rainiest months with 505 and 453 mm of precipitation, while October and November were the driest (53 and 61 mm, respectively). The minimum temperatures occur in the rainy period (26 °C) and the maximum in the dry period (29 °C). The Mojuim estuary has a macrotidal regime, with an average amplitude of 4.9 m during spring tide and 3.2 m during low tide (Rollnic et al., 2018). During the wet season the Mojuim River has a flow velocity of 1.8 m s^{-1} at the ebb tide and 1.3 m s^{-1} at the flood tide. During the dry season, the maximum currents reach 1.9 m s^{-1} at the flood and 1.67 m s^{-1} at the ebb tide (Rocha, 2015). The annual mean salinity is 26.95 PSU (Valentim et al., 2018).

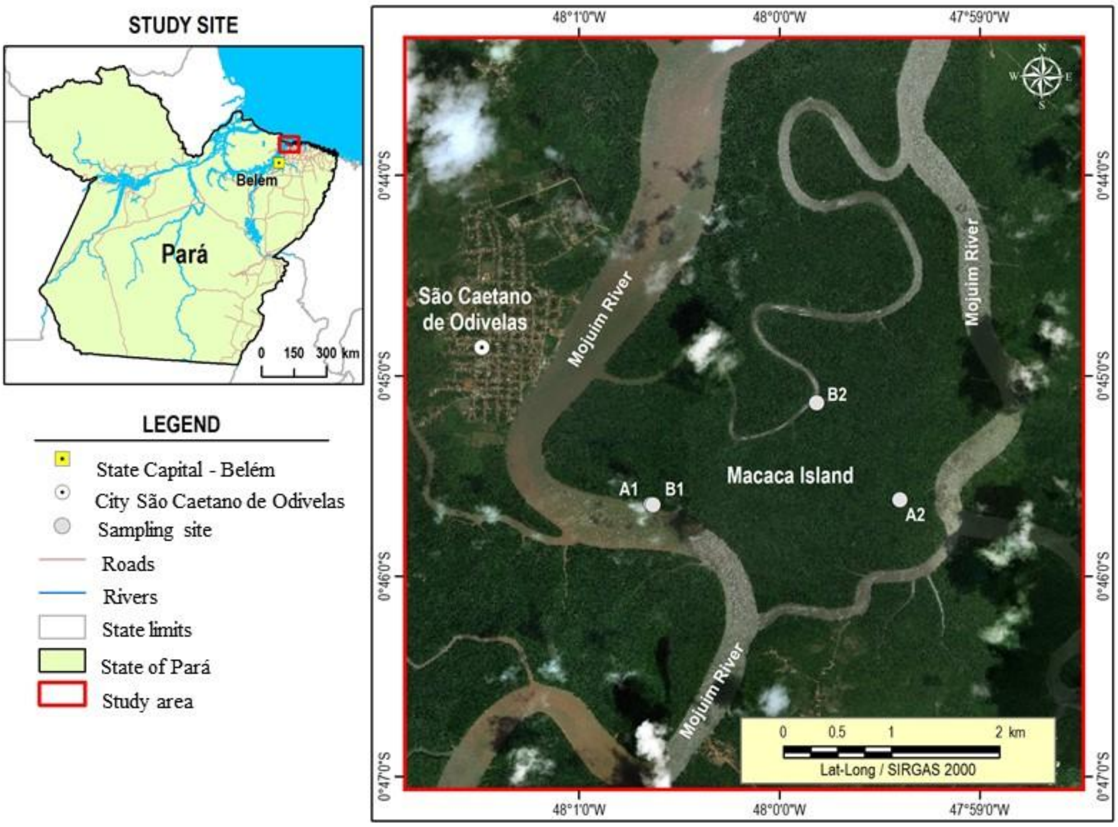

Figure 1. The Macaca Island located in the mangrove coast of Northern Brazil, Municipality of São Caetano de Odivelas (state of Pará), with sampling points at low (plot B1 and plot B2) and high topographies (plot A1 and plot A2). Image Source: © Google Earth

The Mojuim River region is geomorphologically formed by partially submerged river basins consequent of the increase in the relative sea level during the Holocene (Prost et al., 2001) associated with the formation of mangroves, dunes, and beaches (El-Robrini et al., 2006). This river forms the entire watershed of the municipality of São Caetano de Odivelas and borders the municipality of São João da Ponta (Figure 1). Before reaching the estuary, the Mojuim River crosses an area of a dryland forest highly fragmented by family farming, forming remnants of secondary forest (< 5.0 ha) of various ages (Fernandes and Pimentel, 2019). The population economically exploited the estuary, primarily by artisanal fishing, crab (*Ucides cordatus* L.) extraction, and oyster farms.

The flora of the mangrove area of Macaca Island is little anthropized and comprises the plant genera *Rhizophora*, *Avicenia*, *Laguncularia*, and *Acrostichum* (Ferreira, 2017; França et al., 2016). The estuarine plains are influenced by macrotide dynamics and can be physiographically divided into four sectors (França et al., 2016). The Macaca Island is ranked as being from the fourth sector, which implies having woods of adult trees of the genus *Ryzophora* with an average height of 10 to 25 m, being located at an elevation of 0 to 5 m, and having silt-clay soil (França et al., 2016).

Four sampling plots were selected in the Macaca Island (Figure 1) on 19/05/2017, when the moon was in the waning quarter phase: two plots where flooding occurs every day (plots B1 and B2; Figure 1), called low topography (Top_Low), and two plots where flooding occurs only at high tides during the solstice and on the high tides of the rainy season of the new and full moons (plots A1 and A2; Figure 1), called high topography (Top_High).

2.2 Greenhouse gas flux measurements

In each plot, eight Polyvinyl Chloride rings with 0.20 m diameter and 0.12 m height were randomly installed within a circumference with a diameter of 20 m. The rings had an area of 0.028 m^{-2} (volume of 3.47 L), were fixed 0.05 m into the ground, and remained in place until the study was completed. Once a month, the Greenhouse gas flux was measured during periods of waning or crescent moon, as these are the times when the soil in the low topography is more exposed. To avoid the influence of mangrove roots on the gas fluxes, the rings were placed in locations without any seedlings or aboveground mangrove roots. CO_2 and CH_4 concentrations (ppm) were measured using the dynamic chamber methodology (Norman et al., 1997; Verchot et al., 2000), sequentially connected to a Los Gatos Research portable gas analyzer (Mahesh et al., 2015). The device was calibrated monthly with a high quality standard gas (500 ppm CO_2; 5 ppm CH_4). The rings were sequentially closed for three minutes with a PVC cap, being connected to the analyzer through two 12.0 m polyethylene hoses. The gas concentration was measured every two seconds and automatically stored by the analyzer. CO_2 and CH_4 fluxes were calculated from the linear regression of increasing/decreasing CO_2 and CH_4 concentrations within the chamber, usually between one and three minutes after the ring cover was placed (Frankignoulle, 1988; McEwing et al., 2015). The flux is considered zero when the linear regression reaches an $R^2 < 0.30$ (Sundqvist et al., 2014). However, in our analyses, most regressions reached $R^2 > 0.70$, and the regressions were weak and considered zero in only 6% of the samples. At the end of each flux measurement, the height of the ring above ground was measured at four equidistant points with a ruler. The seasonal data were analyzed by comparing the average monthly fluxes in the wet season and dry season separately.

2.3 Vegetation structure and biomass

The floristic survey was conducted in October 2017 using circular 1,256.6 m^2 plots (Kauffman et al., 2013) divided into four 314.15 m^2 subplots, which is the equivalent to 0.38 ha (Figure 1), at the same topographies as the gas flux analysis. We recorded the diameter above the aerial roots, the diameter of the stem, and total height of all trees with DBH (diameter at breast height; m) greater than 0.05m. The allometric equations (Howard et al., 2014) to calculate tree biomass (aboveground biomass; AGB) were: $AGB = 0.1282 * DBH^{2.6}$ ($R^2 = 0.92$) for *R. mangle*; $AGB = 0.140 * DBH^{2.4}$ ($R^2 = 0.97$) for *A. germinans*; and Total $AGB = 0.168 * \rho * DBH^{2.47}$ ($R^2 = 0.99$), where $\rho_{R.\ mangle} = 0.87$; $\rho_{A.\ germinans} = 0.72$ (ρ = wood density).

2.4 Soil sampling and environmental characterization

Four soil samples were collected with an auger at a depth of 0.10 m in all the studied plots for gas flux measurements in July 2017 (beginning of the dry season) and January 2018 (beginning of the rainy season; Figure 1). Before the soil samples were removed, pH and redox potential (Eh; mV) were measured with a Metrohm 744 equipment by inserting the platinum probe directly into the intact soil at a depth of 0.10 m (Bauza et al., 2002). The soil samples collected in the field were transported to the laboratory (Chemical Analysis Laboratory of the *Museu Paraense Emílio Goeldi*) in thermal boxes containing ice. The soil samples were analyzed on the day after collection at the laboratory, and the samples were kept in a freezer. Salinity (Sal; ppt) was measured with PCE-0100, and soil moisture (Sm; %) by the residual gravimetric method (EMBRAPA, 1997).

Organic Matter (OM; g kg^{-1}), Total Carbon (T_C; g kg^{-1}) and Total Nitrogen (T_N; g kg^{-1}) were calculated by volumetry (oxidoreduction) using the Walkley-Black method (Kalembasa and Jenkinson, 1973). Microbial carbon (C_{mic}; mg kg^{-1}) and microbial nitrogen (N_{mic}; mg kg^{-1}) were determined through the 2.0 min of Irradiation-extraction method of soil by microwave technique (Islam and Weil, 1998). Microwave heated soil extraction proved to be a simple, fast, accurate, reliable, and safe method to measure soil microbial biomass (Araujo, 2010; Ferreira et al., 1999; Monz et al., 1991). The C_{mic} was determined by dichromate oxidation (Kalembasa and Jenkinson, 1973; Vance et al., 1987). The N_{mic} was analyzed following the method described by Brookes et al. (1985), changing fumigation to irradiation, which uses the difference between the amount of T_N in irradiated and non-irradiated soil. We used the flux conversion factor of 0.33 (Sparling and West, 1988) and 0.54 (Almeida et al., 2019; Brookes et al., 1985), for carbon and nitrogen, respectively. Particle size analysis was performed separately on four soil samples collected at each flux plot, in the two seasons (October 2017 and March 2018), according to EMBRAPA (1997).

At each gas flux measurement, environmental variables such as air temperature (T_{air}, °C), relative humidity (RH, %), and wind speed (W_s, m s^{-1}) were quantified with a portable thermo-hygrometer (model AK821) at the height of 2.0 m above the soil surface. Soil temperature (T_s, °C) was measured with a portable digital thermometer (model TP101) after each gas flux measurement. Daily precipitation was obtained from an automatic precipitation station installed at a pier on the banks of the Mojuim River in São Caetano das Odivelas (coordinates: -0.738333 latitude; -48.013056 longitude).

2.5 Statistical analyses

On the Macaca Island, two treatments were allocated (low and high topography), with two plots in either. In each plot, eight chambers were randomly distributed, which were considered sample repetitions. The normality of the data of FCH_4, FCO_2, and soil physicochemical parameters was evaluated using the Shapiro-Wilks method. The soil CO_2 and CH_4 flux showed a non-normal distribution. Therefore, we used the non-parametric ANOVA (Kruskal-Wallis, $p < 0.05$) to test the differences between the two treatments among months and seasons. The physicochemical parameters were normally distributed. Therefore, a parametric ANOVA was used to test the statistical differences ($p < 0.05$) between the two treatments among months and seasons. Pearson correlation coefficients were calculated to determine the relationships between soil properties and gas fluxes in the months (dry and wet season) when the chemical properties of the soil were analyzed at the same time as gas fluxes were measured. Statistical analyses were performed with the free statistical software Infostat 2015®.

3 Results

3.1 Carbon dioxide and methane fluxes

CO_2 fluxes differed significantly between topographies only in January ($H = 3.915$; $p = 0.048$), July ($H = 9.091$; $p = 0.003$), and November ($H = 11.294$; $p < 0.000$) (Figure 2;

Supplementary Information, SI 1), with generally higher fluxes at the high topography than at the low topography. At the high topography, CO_2 fluxes were significantly higher ($H = 24.510$; $p = 0.011$) in July compared to August and December, March, October, and May, not differing from the other months of the year. Similarly, at the low topography, CO_2 fluxes were statistically higher ($H = 19.912$; $p = 0.046$) in September and February than in January and November, not differing from the other months. We found a mean monthly flux of 327.9 ± 78.0 mg CO_2 m^{-2} h^{-1} (mean ± standard error) and 217.2 ± 51.0 mg CO_2 m^{-2} h^{-1} at the high and low topographies, respectively.

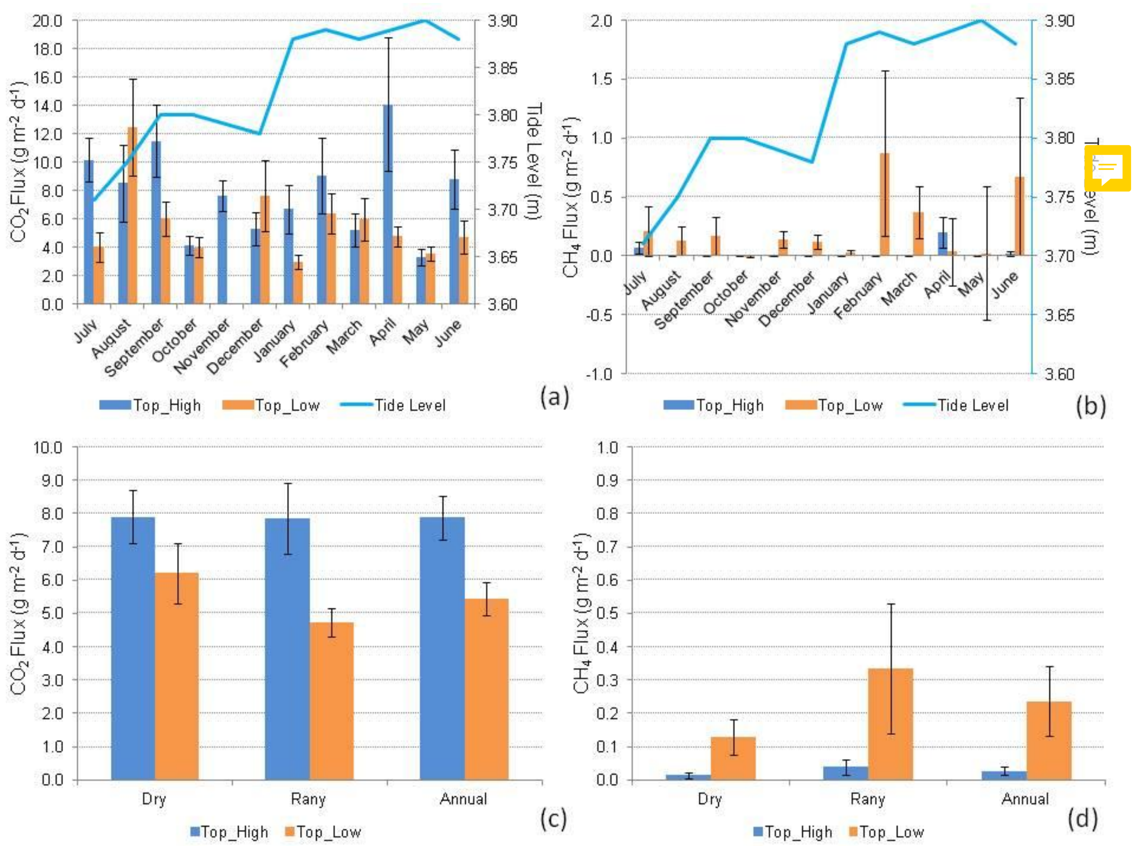

Figure 2. CO_2 (a) and CH_4 (b) fluxes (g CO_2 or CH_4 m^{-2} d^{-1}) monthly (July 2018 to June 2019) ($n = 16$). Seasonal (Dry and Rainy) and annual fluxes of CO_2 (c) and CH_4 (d), at high (Top_High) and low (Top_Low) topographies ($n = 96$), in a mangrove forest soil compared to tide level (Tide Level). The bars represent the standard error of the mean.

The CH_4 fluxes were statistically different between topographies only in November ($H = 9.276$; $p = 0.002$) and December ($H = 4.945$; $p = 0.005$), with higher fluxes at the low topography (Figure 2; SI 1). At the high topography, CH_4 fluxes were significantly ($H = 40.073$; $p < 0.001$) higher in April and July compared to the other months studied, and in November CH_4 was consumed from the atmosphere (Figure 2; SI 1). Similarly, CH_4

fluxes at the low topography did not vary significantly among months ($H = 10.114$; $p = 0.407$).

Greenhouse gas fluxes (Figure 2) were only significantly different between topographies in the dry season (Figure 3), period when CO_2 fluxes were higher ($H = 7.378$; $p = 0.006$) at the high topography and CH_4 fluxes at the low topography ($H = 8.229$; $p < 0.001$). In the Macaca Island, the mean annual fluxes of CO_2 and CH_4 were 6.659 ± 0.419 g CO_2 m^{-2} d^{-1} and 0.132 ± 0.053 g CH_4 m^{-2} d^{-1}, respectively.

3.2 Weather data

There was a marked seasonality during the study period (Figure 2), with 2,155.0 mm of precipitation during the rainy period and 1,016.5 mm during the dry period. The highest tides occurred in the period of greater precipitation (Figure 3) due to the rains. However, the rainfall distribution was different from the climatological normal (Figure 3). The precipitation in the rainy season was 553.2 mm below and in the dry season was 589.1 mm above the climatological normal. Thus, in the period studied, the dry season was rainier and the rainy season drier than the climatological normal, which may be a consequence of global climate change.

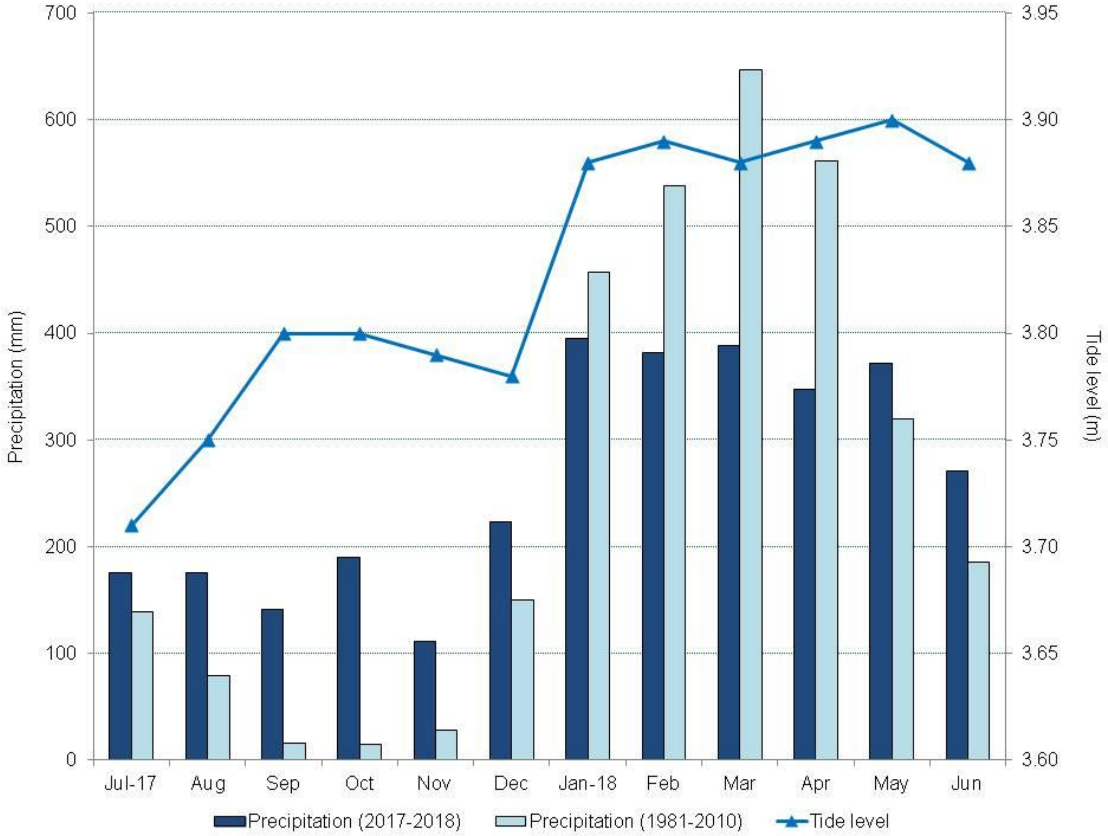

Figure 3. Monthly climatological normal in the municipality of Soure (1981-2010, mm), monthly precipitation (mm), and maximum tide height (m) from 2017 to 2018, in the municipality of São Caetano de Odivelas (PA).

T_{air} was significantly higher (LSD = 0.72, p = 0.01) at the high (31.24 ± 0.26 °C) than at the low topography (30.30 ± 0.25 °C) only in the rainy season (Figure 4a). No significant variation in T_s was found between topographies in either season (Figure 4b). RH was significantly higher (LSD = 2.55, p = 0.01) at the high topography (70.54 ± 0.97%) than at the low topography (66.85 ± 0.87%) only in the rainy season (Figure 4c). W_s (Figure 4d) was significantly higher (LSD = 0.15, p < 0.00) at the low (0.54 ± 0.06 m s^{-1}) than at the high topography (0.24 ± 0.04 m s^{-1}) also in the rainy season.

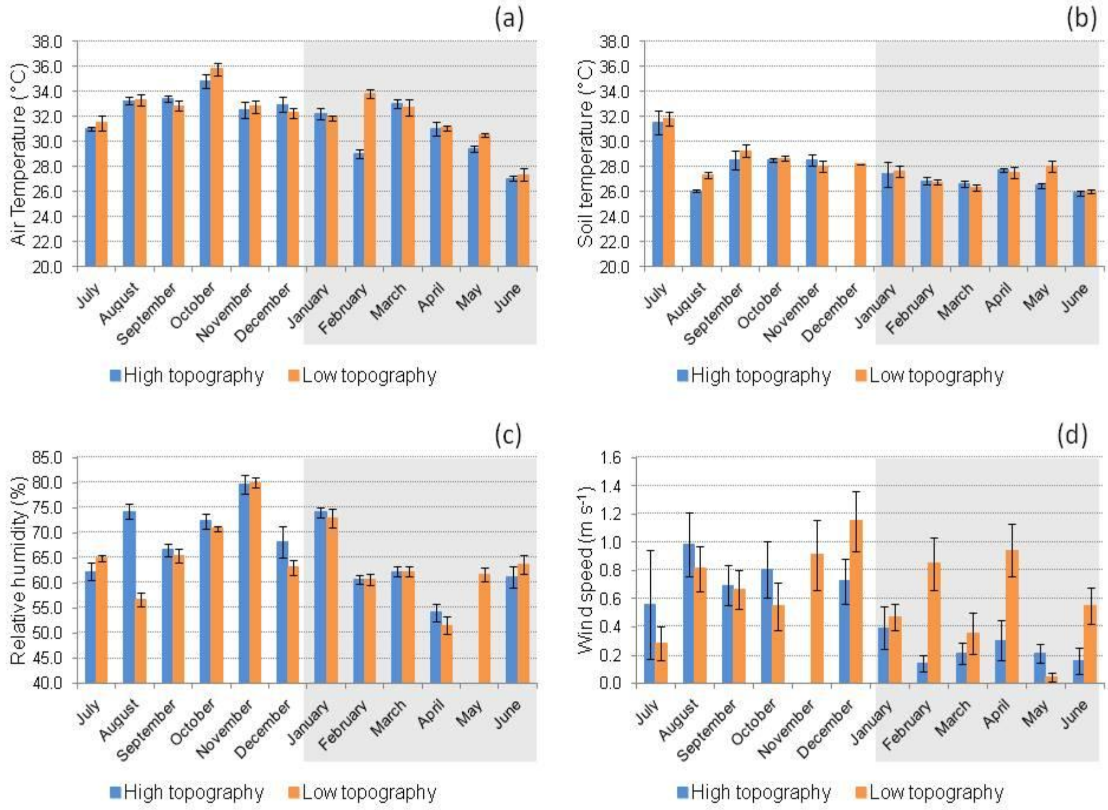

Figure 4. a) Air temperature (°C), b) soil temperature (°C), c) relative humidity (%), and d) wind speed (m s^{-1}) at high and low topographies, from July 2017 to June 2018 in a mangrove area in the Mojuim River estuary. Bars highlighted in grey correspond to the rainy season (n = 16). The bars represent the standard error.

3.3 Soil characteristics

Silt concentration was higher at the low topography (LSD: 14.763; p= 0.007) and clay concentration was higher at the high topography plots (LSD: 12.463; p= 0.005), in both seasons studied (Table 1). Soil particle size analysis did not differ statistically ($p > 0.05$) between the two seasons (Table 1). Soil moisture did not vary significantly ($p > 0.05$) between topographies at each season, or between seasonal periods at the same topography (Table 1). The pH varied statistically (LSD: 5.950; p= 0.006) only at the low topography when the two seasons were compared, being more acidic in the dry period (Table 1). The pH values were significantly (LSD: 0.559; p= 0.008) higher in the dry season (Table 1). No variation in Eh was identified between topographies and seasons (Table 1), although it was higher in the dry season than in the rainy season. However, Sal values were higher (LSD: 3.444; p = 0.010) at the high topography than at the low topography in the dry season (Table 1). In addition, Sal was significantly higher in the dry season than in the rainy season, in both high (LSD: 2.916; $p < 0.001$) and low (LSD: 3.003; $p < 0.001$) topographies (Table 1). (Table 1).

Table 1. Analysis of Sand (%), Silt (%), Clay (%), Moisture (%), pH, Redox Potential (Eh, mV) and salinity (Sal; ppt) in the mangrove soil of high and low topographies, and in the rainy and dry seasons (Macaca island, São Caetano das Odivelas). Numbers represent the mean ± standard error of the mean. Lower case letters compare topographies in each seasonal period and upper-case letters compare the same topography between seasonal periods. Different letters indicate statistical difference (LSD, $p < 0.05$).

Season	Topography	Sand (%)	Silt (%)	Clay (%)	Moisture (%)	pH	Eh (mV)	Sal (ppt)
Dry	High	12.1±1.4^{aA}	41.8±3.3^{bA}	46.1±2.6^{aA}	73.1±6.6^{aA}	5.5±0.2^{aA}	190.25±45.53^{aA}	35.25±1.11^{aA}
	Low	9.7±2.5^{aA}	63.6±6.1^{aA}	26.6±5.2^{bA}	86.9±3.4^{aA}	5.3±0.3^{aA}	106.38±53.76^{aA}	30.13±1.16^{bA}
	Mean	10.9±1.4^{A}	52.7±4.4^{A}	36.4±3.8^{A}	80.0±4.0^{A}	5.4±0.2^{A}	148.31±35.71^{A}	32.69±1.02^{A}
Rainy	High	12.1±1.4^{aA}	41.8±3.3^{bA}	46.1±2.6^{aA}	88.9±3.5^{aA}	4.9±0.4^{aA}	92.50±56.20^{aA}	7.50±0.78^{aB}
	Low	9.7±2.5^{aA}	63.6±6.1^{aA}	26.6±5.2^{bA}	88.6±3.7^{aA}	4.4±0.1^{aB}	36.25±49.97^{aA}	8.13±0.79^{aB}
	Mean	10.9±1.4^{A}	52.7±4.4^{A}	36.4±3.8^{A}	88.7±2.5^{A}	4.6±0.2^{B}	64.38±37.04^{A}	7.81±0.54^{B}

The C_{mic} did not differ between topographies in the two seasons (Table 2). However, T_C was significantly higher in the low topography in the dry season (LSD: 5.589; $p < 0.000$) and in the rainy season (LSD: 5.777; $p = 0.024$). In addition, C_{mic} was higher in the dry season in both the high (LSD: 11.325; $p < 0.010$) and low (LSD: 9.345; $p < 0.000$) topographies (Table 2). N_{mic} did not vary between topographies seasonally. However, N_{mic} in the high (LSD: 9.059; $p = 0.013$) and low topographies (LSD: 4.447; $p = 0.001$) was higher during the dry season (Table 2). The C/N ratio (Table 2) was higher in the low than in the high topography in both the dry (LSD: 3.142; $p < 0.000$) and rainy seasons (LSD: 3.675; $p = 0.033$). However, only in the low topography was the C/N ratio higher (LSD: 1.863; $p < 0.000$) in the dry season than in the rainy season (Table 2). Soil OM was higher at the low topography in the rainy (LSD: 9.950; $p = 0.024$) and in the dry seasons (LSD: 9.630; $p < 0.000$). However, only in the lowland topography was the OM concentration higher in the dry season than in the rainy season (Table 2).

Table 2. Seasonal and topographic variation in microbial Carbon (C_{mic}; mg kg^{-1}), microbial Nitrogen (N_{mic}, mg kg^{-1}), Total Carbon (T_C; g kg^{-1}), Total Nitrogen (N_T; g kg^{-1}), Carbon/Nitrogen ratio (C/N) and Soil Organic Matter (OM; g kg^{-1}). Numbers represent the mean (±standard error). Lower case letters compare topographies at each season, and upper-case letters compare the topography between seasons.

Season	Topography	C_{mic} mg kg^{-1}	N_{mic} mg kg^{-1}	T_C g kg^{-1}	T_N g kg^{-1}	C/N	OM g kg^{-1}
Dry	High	22.12±5.22^{aA}	12.76±4.20^{aA}	14.12±2.23^{bA}	1.43±0.06^{aA}	9.60±1.20^{bA}	24.35±3.84^{bA}
	Low	26.34±4.23^{aA}	10.34±2.05^{aA}	26.44±1.35^{aA}	1.56±0.04^{aA}	16.98±0.84^{aA}	45.59±2.32^{aA}
	Mean	24.23±3.29 ^{A}	11.55±2.28 ^{A}	20.28 ±2.03 ^{A}	1.49±0.04 ^{A}	13.29±1.19^{A}	34.97±3.50 ^{A}
Rainy	High	7.40±0.79^{aB}	0.75±0.41^{aB}	11.46±2.48^{bA}	1.32±0.04^{aA}	8.42±1.70^{bA}	19.75±4.27^{bA}
	Low	5.95±1.06^{aB}	1.23±0.28^{aB}	18.27±1.06^{aB}	1.46±0.06^{aA}	12.47±0.22^{aB}	31.51±1.83^{aB}
	Mean	6.68±0.67 ^{B}	0.99±0.25 ^{B}	14.86 ±1.57 ^{B}	1.39±0.04 ^{A}	10.44±0.98^{A}	25.63±2.71 ^{B}

3.4 Vegetation structure and biomass

Only the species *R. mangle* and *A. germinans* were found in the floristic survey carried out. The DBH did not vary significantly between the topographies for either species (Table 3). However, *R. mangle* had a higher DBH than *A. germinaris* at both high (LSD: 139.304; p = 0.037) and low topographies (LSD: 131.307; p = 0.001). The basal area (BA) and AGB did not show significant variation (Table 3). A total aboveground biomass of 322.1 ± 49.6 Mg ha^{-1} was estimated.

Table 3: Summed Diameter at Breast Height (DBH; cm), Basal Area (BA; m^2 ha^{-1}) and Aboveground Biomass (AGB; Mg ha^{-1}) at high and low topographies in the mangrove forest of the Mojuim River estuary. Numbers represent the mean ± standard error of the mean. Lower case letters compare topographic height for each species, and upper-case letters compare species at each topographic height, using Tukey's test ($p < 0.05$).

Specie	Topography	N ha^{-1}	DBH (cm)	BA (m^2 ha^{-1})	AGB (Mg ha^{-1})
Rhizophora mangle	High	302.4±20.5	238.8±24.9^{aA}	17.3±2.0^{aA}	219.3±25.7^{aA}
	Low	310.4±37.6	283.5±45.0^{aA}	24.2±4.3^{aA}	338.7±62.9^{aA}
Avicennia germinans	High	47.7±20.5	86.8±51.2^{aB}	13.8±9.2^{aA}	135.3±94.7^{aA}
	Low	15.9±9.2	46.1±29.3^{aB}	11.8±8.8^{aA}	136.0±108.3^{aA}
Total	High	350.2±18.4	325.6±33.6^{a}	31.1±7.5^{a}	304.5±99.8^{a}
	Low	346.2±41.0	296.0±23.7^{a}	30.0±4.1^{a}	330.8±60.4^{a}

The equations for biomass estimates (AGB) were: *R. mangle* = 0.1282*$DBH^{2.6}$; *A. germinans* = 0.14*$DBH^{2.4}$; and Total = 0.168*ρ*$DBH^{2.47}$, where $\rho_{R.\ mangle}$ = 0.87; $\rho_{A.\ germinans}$ = 0.72 (Howard et al., 2014).

3.5 Drivers of greenhouse gas fluxes

In the rainy season, CO_2 efflux was correlated with T_{air} (Pearson = 0.23, p = 0.03), RH (Pearson = -0.32, p < 0.00) and T_s (Pearson = 0.21, p = 0.04) only at the low topography. In the dry season CO_2 flux was correlated with T_s (Pearson = 0.39, p < 0.00) at the low topography. The dry season was the period in which we found the greatest amount of significant correlations between CO_2 efflux and soil chemical parameters, while the C:N ratio, OM, and Eh were correlated with CO_2 efflux in both seasons (Table 4). The negative correlation between T_C, N_T, C/N, and OM, along with the positive correlation of N_{mic} with soil CO_2 flux, in the dry period, indicates that microbial activity is a decisive factor for CO_2 efflux (Table 4). Soil moisture in the Mojuim River mangrove forest negatively influenced CO_2 flux in both seasons (Table 4). However, soil moisture was not correlated with CH_4 flux. No significant correlations were found between CH_4 efflux and the chemical properties of the soil in the mangrove of the Mojuim River estuary (Table 4). However, more detailed studies on CH_4 efflux and on its relationship with methanotrophic bacteria and abiotic factors (mainly ammonia and sulfate) are needed due to the average flux of 4.70 mg C m^{-2} h^{-1} and the extreme monthly and seasonal variations.

Table 4. Correlation coefficient (Pearson) of CO_2 and CH_4 fluxes with chemical parameters of the soil in a mangrove area in the Mojuim River estuary.

Gas Flux ($g\ m^{-2}\ d^{-1}$)	Season	T_C ($g\ kg^{-1}$)	T_N ($g\ kg^{-1}$)	C_{mic} ($mg\ kg^{-1}$)	N_{mic} ($mg\ kg^{-1}$)	C/N	OM ($g\ kg^{-1}$)	Sal (ppt)	Eh (mV)	pH	Moisture (%)
	Dry	-0.68^{**}	-0.59^{*}	0.18^{NS}	0.61^{**}	-0.66^{**}	-0.67^{**}	-0.07^{NS}	0.51^{*}	0.21^{NS}	-0.49^{*}
CO_2	Rainy	-0.44^{NS}	-0.20^{NS}	-0.15^{NS}	-0.32^{NS}	-0.50^{*}	-0.63^{**}	-0.54^{*}	0.53^{*}	0.47^{NS}	-0.54^{*}
	Annual	-0.50^{**}	-0.35^{*}	-0.18^{NS}	0.00^{NS}	-0.53^{**}	-0.48^{**}	-0.30^{NS}	0.39^{*}	0.23^{NS}	-0.56^{**}
	Dry	0.30^{NS}	0.07^{NS}	-0.14^{NS}	-0.24^{NS}	0.34^{NS}	0.02^{NS}	-0.04^{NS}	-0.38^{NS}	0.26^{NS}	0.26^{NS}
CH_4	Rainy	0.05^{NS}	-0.09^{NS}	0.44^{NS}	-0.27^{NS}	0.09^{NS}	-0.11^{NS}	-0.04^{NS}	-0.13^{NS}	-0.07^{NS}	0.04^{NS}
	Annual	0.04^{NS}	-0.10^{NS}	-0.01^{NS}	-0.18^{NS}	0.08^{NS}	-0.01^{NS}	-0.17^{NS}	-0.21^{NS}	-0.08^{NS}	0.02^{NS}

Total Carbon (T_C; $g\ kg^{-1}$); Total Nitrogen (T_N; $g\ kg^{-1}$); Microbial Carbon (Cmic, $g\ kg^{-1}$); Microbial Nitrogen (N_{mic}, $g\ kg^{-1}$); Carbon and Nitrogen ratio (C/N); Organic Matter (OM; $g\ kg^{-1}$); Salinity (Sal; ppt); Redox Potential (Eh; mV); Soil Moisture (Moisture, %).

NS= not significant; * significant effects at $p \leq 0.05$; ** significant effects at $p \leq 0.01$

4 Discussion

4.1 Carbon dioxide and methane flux

It is important to consider that the year under study was rainier in the dry season (2017) and less rainy in the wet season (2018) when the climatological average is concerned (1981-2010) (Figure 3). Perhaps this variation is related to the effects of global climate changes. Under these conditions, negative and positive flows of the two greenhouse gases were found (negative values represent gas consumption) Under these conditions, the CO_2 flux from the mangrove soil ranged from -5.06 to 68.96 g CO_2 m^{-2} d^{-1} (mean 6.66 g CO_2 m^{-2} d^{-1}), while the CH_4 flux ranged from -5.07 to 11.08 g CH_4 m^{-2} d^{-1} (mean 0.13 g CH_4 m^{-2} d^{-1}), resulting in a total carbon rate of 1.92 g C m^{-2} d^{-1} or 7.00 Mg C ha^{-1} y^{-1} (Figure 2). The negative CO_2 flux is apparently a consequence of the increased CO_2 solubility in tidal waters or of the increased sulfate reduction, as described in the literature (Borges et al., 2018; Chowdhury et al., 2018; Nóbrega et al., 2016). Fluctuations in redox potential altered the availability of the terminal electron acceptor and donor, and the forces of recovery of their concentrations in the soil, such that a disproportionate release of CO_2 can result from the alternative anaerobic degradation processes such as sulfate and iron reduction (Chowdhury et al., 2018). The soil carbon flux in the mangrove area in the Amazon region was within the range of findings for other tropical mangrove areas (2.57 to 11.00 g CO_2 m^{-2} d^{-1}; Shiau and Chiu, 2020). However, the mean flux of 6.2 mmol CO_2 m^{-2} h^{-1} recorded in this Amazonian mangrove was much higher than the mean efflux of 2.9 mmol CO_2 m^{-2} h^{-1} recorded in 75 mangroves during low tide periods (Alongi, 2009).

An emission of 0.010 Tg CH_4 y^{-1}, 0.64 g CH_4 m^{-2} d^{-1} (Rosentreter et al., 2018a), or 26.7 mg CH_4 m^{-2} h^{-1} has been reported for tropical latitudes (0 and 5°). In our study, the monthly average of CH_4 flux was higher at the low (7.3 ± 8.0 mg CH_4 m^{-2} h^{-1}) than at the high topography (0.9 ± 0.6 mg C m^{-2} h^{-1}), resulting in 0.13 g CH_4 m^{-2} d^{-1} or 0.48 Mg CH_4 ha^{-1} y^{-1} (Figure 2). Therefore, the CH_4-C fluxes from the mangrove soil in the Mojuim River estuary were much lower than expected. It is known that there is a microbial functional module for CH_4 production and consumption (Xu et al., 2015) and diffusibility (Sihi et al., 2018), and considers three key mechanisms: aceticlastic methanogenesis (acetate production), hydrogenotrophic methanogenesis (H_2 and CO_2 production), and aerobic methanotrophy (CH_4 oxidation and O_2 reduction). The average emission from the soil of 8.4 mmol CH_4 m^{-2} d^{-1} was well below the fluxes recorded in the Bay of Bengal, with 18.4 mmol CH_4 m^{-2} d^{-1} (Biswas et al., 2007). In the Amazonian mangrove studied the mean annual carbon equivalent efflux was 429.6 mg $CO_{2\text{-eq}}$ m^{-2} h^{-1}. This value is 0.00004% of the erosion losses of 103.5 Tg $CO_{2\text{-eq}}$ ha^{-1} y^{-1} projected for the next century in tropical mangrove forests (Adame et al., 2021). These higher CO_2 flux concomitantly with lower CH_4 flux in this Amazonian estuary are probably a consequence of changes in the rainfall pattern already underway, where the dry season was wetter and the rainy season drier when compared to the climatological normal.

4.2 Drivers of greenhouse gas fluxes

Mangrove areas are periodically flooded, with a larger flood volume during the syzygy tides, especially in the rainy season. The hydrological condition of the soil is determined by the microtopography and can regulate the respiration of microorganisms (aerobic or anaerobic), being a decisive factor in controlling the CO_2 efflux (Dai et al., 2012; Davidson et al., 2000; Ehrenfeld, 1995). In the two climatic periods of the year, the high topography produced more CO_2 (7.869 ± 1.873 g CO_2 m^{-2} d^{-1}) than the low topography (5.212 ± 1.225 g CO_2 m^{-2} d^{-1}) (Figure 2; SI 1). No significant influence on CO_2 flux was observed due to the low variation in high tide level throughout the year (0.19 m) (Figure 2), although it was numerically higher at the high topography. However, tidal height and the rainy season resulted in a higher CO_2 flux (rate high/low =1.7) at the high topography (7.858 ± 0.039 g CO_2 m^{-2} d^{-1}) than at the low topography (4.734 ± 0.335 g CO_2 m^{-2} d^{-1}) (Figure 2; SI 1). This result is because the root systems of most flood-tolerant plants remain active when flooded (Angelov et al., 1996). Still, the high topography has longer flood-free periods, which only happens when the tides are syzygy or when the rains are torrential.

CO_2 efflux was higher in the high topography than in the low topography in the rainy season (when soils are more subject to inundation), i.e., 39.8% lower in the forest soil exposed to the atmosphere for less time. Measurements performed on 62 mangrove forest soils showed an average flux of 2.87 mmol CO_2 m^{-2} h^{-1} when the soil was exposed to the atmosphere, while 75 results on flooded mangrove forest soils showed an average emission of 2.06 mmol CO_2 m^{-2} h^{-1} (Alongi, 2007, 2009), i.e., 28.2% less than for the dry soil. This reflects the increased facility gases have for molecular diffusion than fluids, and the increased surface area available for aerobic respiration and chemical oxidation during air exposure (Chen et al., 2010). Some studies attribute this variation to the temperature of the soil when it is exposed to tropical air (Alongi, 2009), which increases the export of dissolved inorganic carbon (Maher et al., 2018). However, although despite the lack of significant variation in soil temperature between topographies at each time of year (Figure 4b), there was a positive correlation (Pearson = 0.15, p = 0.05) between CO_2 efflux and soil temperature at the low topography.

Some studies show that CH_4 efflux is a consequence of the seasonal temperature variation in mangrove forest under temperate/monsoon climates (Chauhan et al., 2015; Purvaja and Ramesh, 2001; Whalen, 2005). However, in your study CH_4 efflux was correlated with Ta (Pearson = -0.33, $p < 0.00$) and RH (Pearson = 0.28, p = 0.01) only in the dry season and at the low topography. The results show that the physical parameters do not affect the fluxes in a standardized way, and their greater or lesser influence depends on the topography and seasonality.

A compilation of several studies showed that the total CH_4 emissions from the soil in a mangrove ecosystem range from 0 to 23.68 mg C m^{-2} h^{-1} (Shiau and Chiu, 2020), and our study showed a range of -0.01 to 31.88 mg C m^{-2} h^{-1} (mean of 4.70 ± 5.00 mg C m^{-2} h^{-1}). The monthly CH_4 fluxes were generally higher at the low (0.232 ± 0.256) than at the high (0.026 ± 0.018) topography, especially during the rainy season when the tides were higher (Figure 2). Only in the dry season was there a significantly higher production at the low than at the high topography (Figure 2; SI 1). The low topography produced 0.0249 g C m^{-2} h^{-1} more to the atmosphere in the rainy season than in the dry season (Figure 2), and a similar seasonal pattern was recorded in other studies (Cameron et al., 2021).

The mangrove soil in the Mojuim River estuary is rich in silt and clay (Table 1), which reduces sediment porosity and fosters the formation and maintenance of anoxic conditions (Dutta et al., 2013). In addition, the lack of oxygen in the flooded mangrove soil favors microbial processes such as denitrification, sulfate reduction, methanogenesis, and redox reactions (Alongi and Christoffersen, 1992). A significant amount of CH_4 produced in wetlands is dissolved in the pore water due to high pressure, causing supersaturation, which allows CH_4 to be released by diffusion from the sediment to the atmosphere and by boiling through the formation of bubbles.

Studies show that the CO_2 flux tends to be lower with high soil saturation (Chanda et al., 2014; Kristensen et al., 2008). A total of 395 Mg C ha^{-1} was found at the soil surface (0.15 m) in the mangrove of the Mojuim River estuary, which was slightly higher than the 340 Mg C ha^{-1} found in other mangroves in the Amazon (Kauffman et al., 2018), however being significantly 1.8 times greater at the low topography (Table 2). The finer soil texture at the low topography (Table 1) reduces groundwater drainage which facilitates the accumulation of C in the soil (Schmidt et al., 2011).

4.3 Mangrove biomass

Only the species *R. mangle* and *A. germinans* were found in the floristic survey carried out, which is aligned with the results of other studies in the same region (Menezes et al., 2008). Thus, the variations found in the flux between the topographies in the Mojuim River estuary are not related to the mangrove forest structure, because there was no difference in the aboveground biomass. Since there was no difference in the species composition, the belowground biomass is not expected to differ either (Table 3).

Assuming that the amount of carbon stored is 0.42 of the total biomass (Sahu and Kathiresan, 2019), the mangrove forest biomass of the Mojuim River estuary stores 127.9 and 138.9 Mg C ha^{-1} at the high and low topographies, respectively. This result is well below the 507.8 Mg C ha^{-1} estimated for Brazilian mangroves (Hamilton and Friess, 2018), but are near the 103.7 Mg C ha^{-1} estimated for a mangrove at Guará's island (Salum et al., 2020), 108.4 Mg C ha^{-1} for the Bragantina region (Gardunho, 2017), and 132.3 Mg C ha^{-1} in French Guiana (Fromard et al., 1998). The biomass found in the Mojuim estuary does not differ from the biomass found in other Amazonian mangroves, despite being much lower than that found in other Brazilian mangroves. The estimated primary production for tropical mangrove forests is 218 ± 72 Tg C y (Bouillon et al., 2008).

4.4 Biogeochemical parameters

During the seasonal and annual periods, CH_4 efflux was not significantly correlated with chemical parameters (Table 5), which is similar to the observed in another study (Chen et al., 2010). Flooded soils present reduced gas diffusion rates, which directly affects the physiological state and activity of microbes, by limiting the supply of the dominant electron acceptors (e.g., oxygen), and gases (e.g., CH_4) (Blagodatsky and Smith, 2012). The importance of soil moisture was evident in the richness and diversity of bacterial communities in a study that compared the different pore spaces filled with water (Banerjee et al., 2016). Furthermore, sulfate reduction in flooded soils (another pathway of organic matter metabolism) is dependent on the redox potential of the soil. However, no sulfate reduction occurs when the redox potential has values are above -

150 mv (Connell and Patrick, 1968). In our study, Eh was above 36.0 mV indicating that sulfate reduction probably did not influence the OM metabolism.

On the other hand, increasing soil moisture provides the microorganisms with essential substrates such as ammonium, nitrate, and soluble organic carbon, and increases gas diffusion rates in the water (Blagodatsky and Smith, 2012). Biologically available nitrogen often limit marine productivity (Bertics et al., 2010), and thus can affect CO_2 fluxes to the atmosphere. However, a mangrove fertilization experiment showed that CH_4 emission rates were not affected by N addition (Kreuzwieser et al., 2003). A higher concentration of C_{mic} and N_{mic} in the dry period (Table 2), both in the high and low topographies, indicated that microorganisms are more active when the soil spends more time aerated in the dry period (Table 2), period when only the high tides produce anoxia in the mangrove soil mainly in the low topography. Under reduced oxygen conditions, in a laboratory incubated mangrove soil, the addition of nitrogen resulted in a significant increase in the microbial metabolic quotient, showing no concomitant change in microbial respiration, which was explained by a decrease in microbial biomass (Craig et al., 2021).

The high OM concentration at the two topographic heights (Table 2), at the two seasons studied, and the respective negative correlation with CO_2 flux (Table 5) confirm the importance of microbial activity in mangrove soils (Gao et al., 2020). Also, CH_4 produced in flooded soils can be converted mainly to CO_2 by the anaerobic oxidation of CH_4 (Boetius et al., 2000; Milucka et al., 2015; Xu et al., 2015) which may contribute to the higher CO_2 efflux in the Mojuim River estuary compared to other tropical mangroves (Rosentreter et al., 2018b). The belowground C stock is considered the largest C reservoir in a mangrove ecosystem, and it results from the low OM decomposition rate due to flooding (Marchand, 2017).

The higher water salinity influenced by the tidal movement in the dry season (Table 1) seems to result in a lower CH_4 flux at the low topography (Dutta et al., 2013; Lekphet et al., 2005; Shiau and Chiu, 2020). Sulfate (SO_4^{2-}) in the brine affects the competition between SO_4^{2-} reduction and methanogenic fermentation, as sulfate-reducing bacteria are more efficient at using hydrogen than methanotrophic bacteria (Abram and Nedwell, 1978; Kristjansson et al., 1982), a key factor fostering reduced CH_4 emissions. At high SO_4^{2-} concentrations methanotrophic bacteria use CH_4 as an energy source and oxidize it to CO_2 (Coyne, 1999; Segarra et al., 2015), increasing the efflux of CO_2 and reduced

CH_4 (Megonigal and Schlesinger, 2002; Roslev and King, 1996). This may explain the high CO_2 efflux found throughout the year at the high and, especially, at the low topographies (Figure 3).

Studies in coastal ecosystems in Taiwan have reported that methanotrophic bacteria can be sensitive to soil pH, and reported an optimal growth at pH ranging from 6.5 to 7.5 (Shiau et al., 2018). The higher soil acidity in the Mojuim River wetland (Table 1) may be inhibiting the activity of methanogenic bacteria by increasing the population of methanotrophic bacteria, which are efficient in CH_4 consumption (Chen et al., 2010; Hegde et al., 2003; Shiau and Chiu, 2020). In addition, the pneumatophores present in *R. mangle* increase soil aeration and reduce CH_4 emissions (Allen et al., 2011; He et al., 2019). Spatial differences (topography) in CH_4 emissions in the soil can be attributed to substrate heterogeneity, salinity, and the abundance of methanogenic and methanotrophic bacteria (Gao et al., 2020). The high Eh values found in both topographies, mainly in the dry period (Table 1), hinder CH_4 emission. Soil Eh above -150 mV has been considered limiting for CH_4 production (Yang and Chang, 1998). Increases in CH_4 efflux with reduced salinity were found as a consequence of intense oxidation or reduced competition from the more energetically efficient SO_4^{2-} and NO^{3-} reducing bacteria when compared to the methanogenic bacteria (Biswas et al., 2007). This fact can be observed in the CH_4 efflux in the mangrove of the Mojuim River, because there was an increased CH_4 production especially in the low topography in the rainy season (Figure 3), when water salinity is reduced (Table 1) due to the increased precipitation. However, we did not find a correlation between CH_4 efflux and salinity, as previously reported (Purvaja and Ramesh, 2001)

5 Conclusions

The most recent estimate between latitude 0° to 23.5° S shows an emission of 2.3 g CO_2 m^{-2} d^{-1} (Rosentreter et al., 2018b). However, the efflux in the mangrove of the Mojuim River estuary was 6.7 g CO_2 m^{-2} d^{-1}. For the same latitudinal range, Rosentreter et al. (2018c) estimated an emission of 0.64 g CH_4 m^{-2} d^{-1}, and we found an efflux of 0.13 g CH_4 m^{-2} d^{-1}. Seasonality was important for CH_4 efflux but did not influence CO_2 efflux. The differences in fluxes may be an effect of global climate changes on the terrestrial biogeochemistry at the plant-soil-atmosphere interface, as indicated by the deviation in precipitation values from the climatology normal, making it necessary to extend this study for more years. Using the factor of 23 to convert the global warming potential of

CH_4 to CO_2 (IPCC, 2001), the CO_2 equivalent emission was 35.4 Mg $CO_{2\text{-eq}}$ ha^{-1} yr^{-1}. Over a 100-year time period, a radiative forcing due to the continuous emission of 0.05 kg CH_4 m^{-2} y^{-1}found in this study, would be offset if CO_2 sequestration rates were 2.16 kg CO_2 m^{-2} y^{-1} (Neubauer and Megonigal, 2015).

Microtopography should be considered when determining the efflux of CO_2 and CH_4 in mangrove forests in an Amazon estuary. The low topography in the mangrove forest of Mojuim River had a higher concentration of organic carbon in the soil. However, it did not produce a higher CO_2 efflux because it was negatively influenced by soil moisture, which was indifferent to CH_4 efflux. MO, C/N ratio, and Eh were critical in soil microbial activity, which resulted in a variation in CO_2 flux during the year and seasonal periods. Thus, the physicochemical properties of the soil are important for CO_2 flux, especially in the rainy season. Still, they did not influence CH_4 fluxes.

Data availability: The data used in this article belong to the doctoral thesis of Saul Castellón, within the Postgraduate Program in Environmental Sciences, at the Federal University of Pará. Access to the data can be requested from Dr. Castellón (saulmarz22@gmail.com), which holds the set of all data used in this paper.

Author contributions: SEMC and JHC designed the study and wrote the article with the help of JFB, MR, MLR, and CN. JFB assisted in the field experiment. MR provided logistical support in field activities.

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6 References

Abram, J. W. and Nedwell, D. B.: Inhibition of methanogenesis by sulphate reducing bacteria competing for transferred hydrogen, Arch. Microbiol., 117(1), 89–92, doi:10.1007/BF00689356, 1978.

Adame, M. F., Connolly, R. M., Turschwell, M. P., Lovelock, C. E., Fatoyinbo, T., Lagomasino, D., Goldberg, L. A., Holdorf, J., Friess, D. A., Sasmito, S. D., Sanderman, J., Sievers, M., Buelow, C., Kauffman, J. B., Bryan-Brown, D. and Brown, C. J.: Future carbon emissions from global mangrove forest loss, Glob. Chang. Biol., 27(12), 2856–2866, doi:10.1111/gcb.15571, 2021.

Allen, D., Dalal, R. C., Rennenberg, H. and Schmidt, S.: Seasonal variation in nitrous oxide and methane emissions from subtropical estuary and coastal mangrove sediments, Australia, Plant Biol., 13(1), 126–133, doi:10.1111/j.1438-8677.2010.00331.x, 2011.

Almeida, R. F. de, Mikhael, J. E. R., Franco, F. O., Santana, L. M. F. and Wendling, B.: Measuring the labile and recalcitrant pools of carbon and nitrogen in forested and agricultural soils: A study under tropical conditions, Forests, 10(7), 544, doi:10.3390/f10070544, 2019.

Alongi, D. M.: The contribution of mangrove ecosystems to global carbon cycling and greenhouse gas emissions, in Greenhouse gas and carbon balances in mangrove coastal ecosystems, edited by Y. Tateda, R. Upstill-Goddard, T. Goreau, D. M. Alongi, A. Nose, E. Kristensen, and G. Wattayakorn, pp. 1–10, Gendai Tosho, Kanagawa, Japan., 2007.

Alongi, D. M.: The energetics of mangrove forests, Springer., 2009.

Alongi, D. M. and Christoffersen, P.: Benthic infauna and organism-sediment relations in a shallow, tropical coastal area: influence of outwelled mangrove detritus and physical disturbance, Mar. Ecol. Prog. Ser., 81(3), 229–245, doi:10.3354/meps081229, 1992.

Alongi, D. M. and Mukhopadhyay, S. K.: Contribution of mangroves to coastal carbon cycling in low latitude seas, Agric. For. Meteorol., 213, 266–272, doi:10.1016/j.agrformet.2014.10.005, 2015.

Angelov, M. N., Sung, S. J. S., Doong, R. Lou, Harms, W. R., Kormanik, P. P. and Black, C. C.: Long-and short-term flooding effects on survival and sink-source relationships of swamp-adapted tree species, Tree Physiol., 16(4), 477–484, doi:10.1093/treephys/16.5.477, 1996.

Araujo, A. S. F. de: Is the microwave irradiation a suitable method for measuring soil microbial biomass?, Rev. Environ. Sci. Biotechnol., 9(4), 317–321, doi:10.1007/s11157-010-9210-y, 2010.

Banerjee, S., Helgason, B., Wang, L., Winsley, T., Ferrari, B. C. and Siciliano, S. D.: Legacy effects of soil moisture on microbial community structure and N2O emissions, Soil Biol. Biochem., 95, 40–50, doi:10.1016/j.soilbio.2015.12.004, 2016.

Bastviken, D., Tranvik, L. J., Downing, J. A., Crill, P. M. and Enrich-Prast, A.: Freshwater Methane Emissions Offset the Continental Carbon Sink, Science (80-.)., 331(6013), 50–50, doi:10.1126/science.1196808, 2011.

Bauza, J. F., Morell, J. M. and Corredor, J. E.: Biogeochemistry of Nitrous Oxide Production in the Red Mangrove (Rhizophora mangle) Forest Sediments, Estuar. Coast. Shelf Sci., 55(5), 697–704, doi:10.1006/ECSS.2001.0913, 2002.

Bertics, V. J., Sohm, J. A., Treude, T., Chow, C. E. T., Capone, D. G., Fuhrman, J. A. and Ziebis, W.: Burrowing deeper into benthic nitrogen cycling: The impact of Bioturbation on nitrogen fixation coupled to sulfate reduction, Mar. Ecol. Prog. Ser., 409, 1–15, doi:10.3354/meps08639, 2010.

Biswas, H., Mukhopadhyay, S. K., Sen, S. and Jana, T. K.: Spatial and temporal patterns of methane dynamics in the tropical mangrove dominated estuary, NE coast of Bay of Bengal, India, J. Mar. Syst., 68(1–2), 55–64, doi:10.1016/j.jmarsys.2006.11.001, 2007.

Blagodatsky, S. and Smith, P.: Soil physics meets soil biology: Towards better mechanistic prediction of greenhouse gas emissions from soil, Soil Biol. Biochem., 47, 78–92, doi:10.1016/J.SOILBIO.2011.12.015, 2012.

Boetius, A., Ravenschlag, K., Schubert, C. J., Rickert, D., Widdel, F., Gleseke, A., Amann, R., Jørgensen, B. B., Witte, U. and Pfannkuche, O.: A marine microbial consortium apparently mediating anaerobic oxidation methane, Nature, 407(6804), 623–626, doi:10.1038/35036572, 2000.

Borges, A. V., Abril, G., Darchambeau, F., Teodoru, C. R., Deborde, J., Vidal, L. O., Lambert, T. and Bouillon, S.: Divergent biophysical controls of aquatic CO2 and CH4 in the World’s two largest rivers, Sci. Rep., 5, doi:10.1038/srep15614, 2015.

Borges, A. V., Abril, G. and Bouillon, S.: Carbon dynamics and CO2 and CH4 outgassing in the Mekong delta, Biogeosciences, 15(4), doi:10.5194/bg-15-1093-2018, 2018.

Bouillon, S., Borges, A. V., Castañeda-Moya, E., Diele, K., Dittmar, T., Duke, N. C., Kristensen, E., Lee, S. Y., Marchand, C., Middelburg, J. J., Rivera-Monroy, V. H., Smith, T. J. and Twilley, R. R.: Mangrove production and carbon sinks: A revision of global budget estimates, Global Biogeochem. Cycles, 22(2), doi:10.1029/2007GB003052, 2008.

Brookes, P. C., Landman, A., Pruden, G. and Jenkinson, D. S.: Chloroform fumigation and the release of soil nitrogen: A rapid direct extraction method to measure microbial biomass nitrogen in soil, Soil Biol. Biochem., 17(6), 837–842, doi:10.1016/0038-0717(85)90144-0, 1985.

Cameron, C., Hutley, L. B., Munksgaard, N. C., Phan, S., Aung, T., Thinn, T., Aye, W. M. and Lovelock, C. E.: Impact of an extreme monsoon on CO2 and CH4 fluxes from mangrove soils of the Ayeyarwady Delta, Myanmar, Sci. Total Environ., 760, 143422, doi:10.1016/j.scitotenv.2020.143422, 2021.

Castillo, J. A. A., Apan, A. A., Maraseni, T. N. and Salmo, S. G.: Soil greenhouse gas fluxes in tropical mangrove forests and in land uses on deforested mangrove lands, Catena, 159, 60–69, doi:10.1016/j.catena.2017.08.005, 2017.

Chanda, A., Akhand, A., Manna, S., Dutta, S., Das, I., Hazra, S., Rao, K. H. and Dadhwal, V. K.: Measuring daytime CO2 fluxes from the inter-tidal mangrove soils of Indian Sundarbans, Environ. Earth Sci., 72(2), 417–427, doi:10.1007/s12665-013-2962-2, 2014.

Chauhan, R., Datta, A., Ramanathan, A. and Adhya, T. K.: Factors influencing spatio-temporal variation of methane and nitrous oxide emission from a tropical mangrove of eastern coast of India, Atmos. Environ., 107, 95–106, doi:10.1016/j.atmosenv.2015.02.006, 2015.

Chen, G. C., Tam, N. F. Y. and Ye, Y.: Spatial and seasonal variations of atmospheric N2O and CO2 fluxes from a subtropical mangrove swamp and their relationships with soil characteristics, Soil Biol. Biochem., 48, 175–181, doi:10.1016/j.soilbio.2012.01.029, 2012.

Chen, G. C., Ulumuddin, Y. I., Pramudji, S., Chen, S. Y., Chen, B., Ye, Y., Ou, D. Y., Ma, Z. Y., Huang, H. and Wang, J. K.: Rich soil carbon and nitrogen but low atmospheric greenhouse gas fluxes from North Sulawesi mangrove swamps in Indonesia, Sci. Total Environ., 487(1), 91–96, doi:10.1016/j.scitotenv.2014.03.140, 2014.

Chen, G. C. C., Tam, N. F. Y. F. Y. and Ye, Y.: Summer fluxes of atmospheric greenhouse gases N2O, CH4 and CO2 from mangrove soil in South China, Sci. Total Environ., 408(13), 2761–2767, doi:10.1016/j.scitotenv.2010.03.007, 2010.

Chowdhury, T. R., Bramer, L., Hoyt, D. W., Kim, Y. M., Metz, T. O., McCue, L. A., Diefenderfer, H. L., Jansson, J. K. and Bailey, V.: Temporal dynamics of CO2 and CH4 loss potentials in response to rapid hydrological shifts in tidal freshwater wetland soils, Ecol. Eng., 114, 104–114, doi:10.1016/j.ecoleng.2017.06.041, 2018.

Chuang, P. C., Young, M. B., Dale, A. W., Miller, L. G., Herrera-Silveira, J. A. and Paytan, A.: Methane and sulfate dynamics in sediments from mangrove-dominated tropical coastal lagoons, Yucatan, Mexico, Biogeosciences, 13(10), 2981–3001, 2016.

Connell, W. E. and Patrick, W. H.: Sulfate Reduction in Soil: Effects of Redox Potential and p H, Science (80-.)., 159(3810), 86–87, doi:10.1126/science.159.3810.86, 1968.

Coyne, M.: Soil Microbiology: An Exploratory Approach, Delmar Publishers, New York, NY, USA., 1999.

Craig, H., Antwis, R. E., Cordero, I., Ashworth, D., Robinson, C. H., Osborne, T. Z., Bardgett, R. D., Rowntree, J. K. and Simpson, L. T.: Nitrogen addition alters composition, diversity, and functioning of microbial communities in mangrove soils: An incubation experiment, Soil Biol. Biochem., 153, 108076, doi:10.1016/j.soilbio.2020.108076, 2021.

Dai, Z., Trettin, C. C., Li, C., Li, H., Sun, G. and Amatya, D. M.: Effect of Assessment Scale on Spatial and Temporal Variations in CH4, CO2, and N2O Fluxes in a Forested Wetland, Water, Air, Soil Pollut., 223(1), 253–265, doi:10.1007/s11270-011-0855-0, 2012.

Davidson, E. A., Verchot, L. V., Cattanio, J. H., Ackerman, I. L. and Carvalho, J. E. M.: Effects of soil water content on soil respiration in forests and cattle pastures of eastern Amazonia, Biogeochemistry, 48(1), 53–69, doi:10.1023/a:1006204113917, 2000.

Donato, D. C., Kauffman, J. B., Murdiyarso, D., Kurnianto, S., Stidham, M. and Kanninen, M.: Mangroves among the most carbon-rich forests in the tropics, Nat. Geosci., 4(5), 293–297, doi:10.1038/ngeo1123, 2011.

Dutta, M. K., Chowdhury, C., Jana, T. K. and Mukhopadhyay, S. K.: Dynamics and exchange fluxes of methane in the estuarine mangrove environment of the Sundarbans, NE coast of India, Atmos. Environ., 77, 631–639, doi:10.1016/j.atmosenv.2013.05.050, 2013.

Ehrenfeld, J. G.: Microsite differences in surface substrate characteristics in Chamaecyparis swamps of the New Jersey Pinelands, Wetlands, 15(2), 183–189, doi:10.1007/BF03160672, 1995.

El-Robrini, M., Alves, M. A. M. S., Souza Filho, P. W. M., El-Robrini M. H. S., Silva Júnior, O. G. and França, C. F.: Atlas de Erosão e Progradação da zona costeira do Estado do Pará – Região Amazônica: Áreas oceânica e estuarina, in Atlas de Erosão e Progradação da Zona Costeira Brasileira, edited by D. Muehe, pp. 1–34, São Paulo., 2006.

EPA, E. P. A.: Inventory of U.S. Greenhouse Gas Emissions and Sinks: 1990–2015., 2017.

Fernandes, W. A. A. and Pimentel, M. A. da S.: Dinâmica da paisagem no entorno da RESEX marinha de São João da Ponta/PA: utilização de métricas e geoprocessamento, Caminhos Geogr., 20(72), 326–344, doi:10.14393/RCG207247140, 2019.

Ferreira, A. S., Camargo, F. A. O. and Vidor, C.: Utilização de microondas na avaliação da biomassa microbiana do solo, Rev. Bras. Ciência do Solo, 23(4), 991–996, doi:10.1590/S0100-06831999000400026, 1999.

Ferreira, S. da S.: Entre marés e mangues: paisagens territorializadas por pescadores da resex marinha de São João da Ponta/PA /, Universidade Federal do Pará., 2017.

França, C. F. de, Pimentel, M. A. D. S. and Neves, S. C. R.: Estrutura Paisagística De São João Da Ponta, Nordeste Do Pará, Geogr. Ensino Pesqui., 20(1), 130–142, doi:10.5902/2236499418331, 2016.

Frankignoulle, M.: Field measurements of air-sea CO2 exchange, Limnol. Oceanogr., 33(3), 313–322, 1988.

Friesen, S. D., Dunn, C. and Freeman, C.: Decomposition as a regulator of carbon accretion in mangroves: a review, Ecol. Eng., 114, 173–178, doi:10.1016/j.ecoleng.2017.06.069, 2018.

Fromard, F., Puig, H., Cadamuro, L., Marty, G., Betoulle, J. L. and Mougin, E.: Structure, above-ground biomass and dynamics of mangrove ecosystems: new data from French Guiana, Oecologia, 115(1), 39–53, doi:10.1007/s004420050489, 1998.

Gao, G. F., Zhang, X. M., Li, P. F., Simon, M., Shen, Z. J., Chen, J., Gao, C. H. and Zheng, H. L.: Examining Soil Carbon Gas (CO2, CH4) Emissions and the Effect on Functional Microbial Abundances in the Zhangjiang Estuary Mangrove Reserve, J. Coast. Res., 36(1), 54–62, doi:10.2112/JCOASTRES-D-18-00107.1, 2020.

Gardunho, D. C. L.: Estimativas de biomassa acima do solo da floresta de mangue na península de Ajuruteua, Bragança – PA, Federal University of Pará, Belém, Brazil., 2017.

Hamilton, S. E. and Friess, D. A.: Global carbon stocks and potential emissions due to mangrove deforestation from 2000 to 2012, Nat. Clim. Chang., 8(3), 240–244, doi:10.1038/s41558-018-0090-4, 2018.

He, Y., Guan, W., Xue, D., Liu, L., Peng, C., Liao, B., Hu, J., Zhu, Q., Yang, Y., Wang, X., Zhou, G., Wu, Z. and Chen, H.: Comparison of methane emissions among invasive and native mangrove species in Dongzhaigang, Hainan Island, Sci. Total Environ., 697, 133945, doi:10.1016/j.scitotenv.2019.133945, 2019.

Hegde, U., Chang, T.-C. and Yang, S.-S.: Methane and carbon dioxide emissions from Shan-Chu-Ku landfill site in northern Taiwan., Chemosphere, 52(8), 1275–1285, doi:10.1016/S0045-6535(03)00352-7, 2003.

Herz, R.: Manguezais do Brasil, Instituto Oceanografico da Usp/Cirm, São Paulo, Brazil., 1991.

Howard, J., Hoyt, S., Isensee, K., Telszewski, M. and Pidgeon, E.: Coastal Blue Carbon: Methods for Assessing Carbon Stocks and Emissions Factors in Mangroves, Tidal Salt Marshes, and Seagrasses, edited by J. Howard, S. Hoyt, K. Isensee, M. Telszewski, and E. Pidgeon, International Union for Conservation of Nature, Arlington, Virginia, USA. [online] Available from: http://www.unesco.org/new/en/natural-sciences/ioc-oceans/sections-and-programmes/ocean-sciences/ocean-carbon/coastal-blue-carbon/ (Accessed 11 September 2019), 2014.

IPCC: Climate Change 2001: Third Assessment Report of the IPCC, Cambridge., 2001.

Islam, K. R. and Weil, R. R.: Microwave irradiation of soil for routine measurement of microbial biomass carbon, Biol. Fertil. Soils, 27(4), 408–416, doi:10.1007/s003740050451, 1998.

Kalembasa, S. J. and Jenkinson, D. S.: A comparative study of titrimetric and gavimetric methods for determination of organic carbon in soil, J. Sci. Food Agric., 24, 1085–1090, 1973.

Kauffman, B. J., Donato, D. and Adame, M. F.: Protocolo para la medición, monitoreo y reporte de la estructura, biomasa y reservas de carbono de los manglares, Bogor, Indonesia., 2013.

Kauffman, J. B., Bernardino, A. F., Ferreira, T. O., Giovannoni, L. R., De Gomes, L. E. O., Romero, D. J., Jimenez, L. C. Z. and Ruiz, F.: Carbon stocks of mangroves and salt marshes of the Amazon region, Brazil, Biol. Lett., 14(9), doi:10.1098/rsbl.2018.0208, 2018.

Kreuzwieser, J., Buchholz, J. and Rennenberg, H.: Emission of Methane and Nitrous Oxide by Australian Mangrove Ecosystems, Plant Biol., 5(4), 423–431, doi:10.1055/s-2003-42712, 2003.

Kristensen, E., Bouillon, S., Dittmar, T. and Marchand, C.: Organic carbon dynamics in mangrove ecosystems: A review, Aquat. Bot., 89(2), 201–219, doi:10.1016/J.AQUABOT.2007.12.005, 2008.

Kristjansson, J. K., Schönheit, P. and Thauer, R. K.: Different Ks values for hydrogen of methanogenic bacteria and sulfate reducing bacteria: An explanation for the apparent inhibition of methanogenesis by sulfate, Arch. Microbiol., 131(3), 278–282, doi:10.1007/BF00405893, 1982.

Lekphet, S., Nitisoravut, S. and Adsavakulchai, S.: Estimating methane emissions from mangrove area in Ranong Province, Thailand, Songklanakarin J. Sci. Technol. ·, 27(1), 153–163 [online] Available from: https://www.researchgate.net/publication/26473398 (Accessed 29 January 2019), 2005.

Maher, D. T., Call, M., Santos, I. R. and Sanders, C. J.: Beyond burial: Lateral exchange is a significant atmospheric carbon sink in mangrove forests, Biol. Lett., 14(7), 1–4, doi:10.1098/rsbl.2018.0200, 2018.

Mahesh, P., Sreenivas, G., Rao, P. V. N. N., Dadhwal, V. K., Sai Krishna, S. V. S. S. and Mallikarjun, K.: High-precision surface-level CO2 and CH4 using off-axis integrated cavity output spectroscopy (OA-ICOS) over Shadnagar, India, Int. J. Remote Sens., 36(22), 5754–5765, doi:10.1080/01431161.2015.1104744, 2015.

Marchand, C.: Soil carbon stocks and burial rates along a mangrove forest chronosequence (French Guiana), For. Ecol. Manage., 384, 92–99, doi:10.1016/j.foreco.2016.10.030, 2017.

McEwing, K. R., Fisher, J. P. and Zona, D.: Environmental and vegetation controls on the spatial variability of CH4 emission from wet-sedge and tussock tundra ecosystems in the Arctic, Plant Soil, 388(1–2), 37–52, doi:10.1007/s11104-014-2377-1, 2015.

Megonigal, J. P. and Schlesinger, W. H.: Methane-limited methanotrophy in tidal freshwater swamps, Global Biogeochem. Cycles, 16(4), 35-1-35–10, doi:10.1029/2001GB001594, 2002.

Menezes, M. P. M. de, Berger, U. and Mehlig, U.: Mangrove vegetation in Amazonia : a review of studies from the coast of Pará and Maranhão States , north Brazil, Acta Amaz., 38(3), 403–420, doi:10.1590/S0044-59672008000300004, 2008.

Milucka, J., Kirf, M., Lu, L., Krupke, A., Lam, P., Littmann, S., Kuypers, M. M. M. and Schubert, C. J.: Methane oxidation coupled to oxygenic photosynthesis in anoxic waters, ISME J., 9(9), 1991–2002, doi:10.1038/ismej.2015.12, 2015.

Monz, C. A., Reuss, D. E. and Elliott, E. T.: Soil microbial biomass carbon and nitrogen estimates using 2450 MHz microwave irradiation or chloroform fumigation followed by direct extraction, Agric. Ecosyst. Environ., 34(1–4), 55–63, doi:10.1016/0167-8809(91)90093-D, 1991.

Neubauer, S. C. and Megonigal, J. P.: Moving Beyond Global Warming Potentials to Quantify the Climatic Role of Ecosystems, Ecosystems, 18(6), 1000–1013, doi:10.1007/S10021-015-9879-4/TABLES/2, 2015.

Nóbrega, G. N., Ferreira, T. O., Siqueira Neto, M., Queiroz, H. M., Artur, A. G., Mendonça, E. D. S., Silva, E. D. O. and Otero, X. L.: Edaphic factors controlling summer (rainy season) greenhouse gas emissions (CO2 and CH4) from semiarid mangrove soils (NE-Brazil), Sci. Total Environ., 542, 685–693, doi:10.1016/j.scitotenv.2015.10.108, 2016.

Norman, J. M., Kucharik, C. J., Gower, S. T., Baldocchi, D. D., Crill, P. M., Rayment, M., Savage, K. and Striegl, R. G.: A comparison of six methods for measuring soil-surface carbon dioxide fluxes, J. Geophys. Res. Atmos., 102(D24), 28771–28777, doi:10.1029/97JD01440, 1997.

Peel, M. C., Finlayson, B. L. and McMahon, T. A.: Updated world map of the Köppen-Geiger climate classification, Hydrol. Earth Syst. Sci., 11(5), 1633–1644, doi:10.1002/ppp.421, 2007.

Poffenbarger, H. J., Needelman, B. A. and Megonigal, J. P.: Salinity Influence on Methane Emissions from Tidal Marshes, Wetlands, 31(5), 831–842, doi:10.1007/s13157-011-0197-0, 2011.

Prost, M. T., Mendes, A. C., Faure, J. F., Berredo, J. F., Sales, M. E. ., Furtado, L. G., Santana, M. G., Silva, C. A., Nascimento, I. ., Gorayeb, I., Secco, M. F. and Luz, L.: Manguezais e estuários da costa paraense: exemplo de estudo multidisciplinar integrado (Marapanim e São Caetano de Odivelas), in Ecossistemas costeiros: impactos e gestão ambiental, edited by M. T. Prost and A. Mendes, pp. 25–52, FUNTEC and Museu Paraense Emílio Goeldi, Belém, Brazil., 2001.

Purvaja, R. and Ramesh, R.: Natural and Anthropogenic Methane Emission from Coastal Wetlands of South India, Environ. Manage., 27(4), 547–557, doi:10.1007/s002670010169, 2001.

Purvaja, R., Ramesh, R. and Frenzel, P.: Plant-mediated methane emission from an Indian mangrove, Glob. Chang. Biol., 10(11), 1825–1834, doi:10.1111/j.1365-2486.2004.00834.x, 2004.

Reeburgh, W. S.: Oceanic Methane Biogeochemistry, Chem. Rev., 2, 486–513, doi:10.1021/cr050362v, 2007.

Robertson, A. I., Alongi, D. M. and Boto, K. G.: Food chains and carbon fluxes, in Coastal and Estuarine Studies, edited by A. I. Robertson and D. M. Alongi, pp. 293–326, American Geophysical Union., 1992.

Rocha, A. S.: Caracterização física do estuário do rio Mojuim em São Caetano de Odivelas - Pa, Universidade Federal do Pará., 2015.

Rollnic, M., Costa, M. S., Medeiros, P. R. L. and Monteiro, S. M.: Tide Influence on Suspended Matter Transport in an Amazonian Estuary, in Journal of Coastal Research, vol. 85, pp. 121–125, Allen Press., 2018.

Rosentreter, J. A., Maher, D. T., Erler, D. V., Murray, R. H. and Eyre, B. D.: Methane emissions partially offset “blue carbon” burial in mangroves, Sci. Adv., 4(6), eaao4985, doi:10.1126/sciadv.aao4985, 2018a.

Rosentreter, J. A., Maher, D. . T., Erler, D. V. V., Murray, R. and Eyre, B. D. D.: Seasonal and temporal CO2 dynamics in three tropical mangrove creeks – A revision of global mangrove CO2 emissions, Geochim. Cosmochim. Acta, 222, 729–745, doi:10.1016/j.gca.2017.11.026, 2018b.

Roslev, P. and King, G. M.: Regulation of methane oxidation in a freshwater wetland by water table changes and anoxia, FEMS Microbiol. Ecol., 19(2), 105–115, doi:10.1111/j.1574-6941.1996.tb00203.x, 1996.

Sahu, S. K. and Kathiresan, K.: The age and species composition of mangrove forest directly influence the net primary productivity and carbon sequestration potential, Biocatal. Agric. Biotechnol., 20, 101235, doi:10.1016/j.bcab.2019.101235, 2019.

Salum, R. B., Souza-Filho, P. W. M., Simard, M., Silva, C. A., Fernandes, M. E. B., Cougo, M. F., do Nascimento, W. and Rogers, K.: Improving mangrove above-ground biomass estimates using LiDAR, Estuar. Coast. Shelf Sci., 236, 106585, doi:10.1016/j.ecss.2020.106585, 2020.

Schmidt, M. W. I., Torn, M. S., Abiven, S., Dittmar, T., Guggenberger, G., Janssens, I. A., Kleber, M., Kögel-Knabner, I., Lehmann, J., Manning, D. A. C., Nannipieri, P., Rasse, D. P., Weiner, S. and Trumbore, S. E.: Persistence of soil organic matter as an ecosystem property, Nature, 478(7367), 49–56, doi:10.1038/nature10386, 2011.

Segarra, K. E. A., Schubotz, F., Samarkin, V., Yoshinaga, M. Y., Hinrichs, K. U. and Joye, S. B.: High rates of anaerobic methane oxidation in freshwater wetlands reduce potential atmospheric methane emissions, Nat. Commun., 6(1), 1–8, doi:10.1038/ncomms8477, 2015.

Shiau, Y.-J. and Chiu, C.-Y.: Biogeochemical Processes of C and N in the Soil of Mangrove Forest Ecosystems, Forests, 11(5), 492, doi:10.3390/f11050492, 2020.

Shiau, Y. J., Cai, Y., Lin, Y. Te, Jia, Z. and Chiu, C. Y.: Community Structure of Active Aerobic Methanotrophs in Red Mangrove (Kandelia obovata) Soils Under Different Frequency of Tides, Microb. Ecol., 75(3), 761–770, doi:10.1007/s00248-017-1080-1, 2018.

Sihi, D., Davidson, E. A., Chen, M., Savage, K. E., Richardson, A. D., Keenan, T. F. and Hollinger, D. Y.: Merging a mechanistic enzymatic model of soil heterotrophic respiration into an ecosystem model in two AmeriFlux sites of northeastern USA, Agric. For. Meteorol., 252, 155–166, doi:10.1016/J.AGRFORMET.2018.01.026, 2018.

Souza Filho, P. W. M.: Costa de manguezais de macromaré da Amazônia: cenários morfológicos, mapeamento e quantificação de áreas usando dados de sensores remotos, Rev. Bras. Geofísica, 23(4), 427–435, doi:10.1590/S0102-261X2005000400006, 2005.

Sparling, G. P. and West, A. W.: A direct extraction method to estimate soil microbial C: calibration in situ using microbial respiration and 14C labelled cells, Soil Biol. Biochem., 20(3), 337–343, doi:10.1016/0038-0717(88)90014-4, 1988.

Sundqvist, E., Vestin, P., Crill, P., Persson, T. and Lindroth, A.: Short-term effects of thinning, clear-cutting and stump harvesting on methane exchange in a boreal forest, Biogeosciences, 11(21), 6095–6105, doi:10.5194/bg-11-6095-2014, 2014.

Valentim, M., Monteiro, S. and Rollnic, M.: The Influence of Seasonality on Haline Zones in An Amazonian Estuary, J. Coast. Res., 85, 76–80, doi:10.2112/SI85-016.1, 2018.

Valentine, D. L.: Emerging Topics in Marine Methane Biogeochemistry, Ann. Rev. Mar. Sci., 3(1), 147–171, doi:10.1146/annurev-marine-120709-142734, 2011.

Vance, E. D., Brookes, P. C. and Jenkinson, D. S.: An extraction method for measuring soil microbial biomass C, Soil Biol. Biochem., 19(6), 703–707, doi:10.1016/0038-0717(87)90052-6, 1987.

Verchot, L. V., Davidson, E. A., Cattânio, J. H. and Ackerman, I. L.: Land-use change and biogeochemical controls of methane fluxes in soils of eastern Amazonia, Ecosystems, 3(1), 41–56, doi:10.1007/s100210000009, 2000.

Whalen, S. C.: Biogeochemistry of Methane Exchange between Natural Wetlands and the Atmosphere, Environ. Eng. Sci., 22(1), 73–94, doi:10.1089/ees.2005.22.73, 2005.

Xu, X., Elias, D. A., Graham, D. E., Phelps, T. J., Carroll, S. L., Wullschleger, S. D. and Thornton, P. E.: A microbial functional group-based module for simulating methane production and consumption: Application to an incubated permafrost soil, J. Geophys. Res. Biogeosciences, 120(7), 1315–1333, doi:10.1002/2015JG002935, 2015.

Yang, S. S. and Chang, H. L.: Effect of environmental conditions on methane production and emission from paddy soil, Agric. Ecosyst. Environ., 69(1), 69–80, doi:10.1016/S0167-8809(98)00098-X, 1998.

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