



Contrasting responses of phytoplankton productivity between coastal and offshore 1 surface waters in the Taiwan Strait and the South China Sea to future CO2-induced 2 acidification 3 4 Guang Gao¹, Tifeng Wang¹, Jiazhen Sun¹, Xin Zhao¹, Lifang Wang¹, Xianghui Guo¹, Kunshan Gao^{1,2}* 6 ¹State Key Laboratory of Marine Environmental Science & College of Ocean and Earth 7 Sciences, Xiamen University, Xiamen 361005, China 8 ²Co-Innovation Center of Jiangsu Marine Bio-industry Technology, Jiangsu Ocean University, Lianyungang 222005, China 10 11 *Corresponding author: ksgao@xmu.edu.cn 12





Abstract

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Future CO₂-induced ocean acidification (OA) has been documented to either inhibit or 15 enhance or result in no effect on marine primary productivity (PP). In order to examine 16 17 effects of OA under multiple drivers, we investigated the influences of OA (a decrease of 0.4 pH_{total} units with corresponding CO₂ concentrations ranged 22.0-39.7 μM) on PP 18 19 through deck-incubation experiments at 101 stations in the Taiwan Strait and the South China Sea (SCS), including the coastal zone, the continental shelf and slope, as well as 20 21 deep-water basin. The daily net primary productivities in surface seawater under incident solar radiation ranged from 17-306 μ g C (μ g Chl a)⁻¹ d⁻¹, with the responses of PP to OA 22 being region-dependent and the OA-induced changes varying from -88.03% (inhibition) 23 to 56.87% (enhancement). The OA-treatment stimulated PP in surface waters of coastal, 24 25 estuarine and shelf waters, but suppressed it in the South China Sea basin. Such 26 OA-induced changes in PP were significantly related to NO_X (the sum of NO₃⁻ and NO₂⁻) 27 availability, in situ pH and solar radiation in surface seawater, but negatively related to 28 salinity changes. Our results indicate that phytoplankton cells are more vulnerable to pH drop in oligotrophic waters. Considering high nutrient and low salinity in coastal waters 29 30 and reduced nutrient availability in pelagic zones with the progressive stratification associated with ocean warming, our results imply that future OA will enhance PP in 31 32 coastal waters but decrease it in pelagic oligotrophic zones. Keywords: CO₂; Taiwan Strait; ocean acidification; photosynthesis; primary productivity; 33





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South China Sea

1 Introduction

increased dissolved CO₂ and decreased pH of seawater (Gattuso et al., 2015), leading to 37 ocean acidification (OA). OA has been shown to result in profound influences on marine 38 39 ecosystems (see the reviews and literature therein, Mostofa et al., 2016; Doney et al., 2020). Marine photosynthetic organisms, which contribute about half of the global 40 primary production, are also being affected by OA (see the reviews and literatures therein, 41 Riebesell et al., 2018; Gao et al., 2019a). It is of general concern that the oceans are going 42 43 to take more or less CO₂ with progressive OA, since the amount of CO₂ uptake by the oceans is essential to predict global and ocean warming trends. Therefore, it is important 44 45 to understand the responses of the key players of marine biological CO₂ pump, the 46 phytoplankton, to OA and other climate change drivers. 47 Elevated CO₂ is well recognized to lessen the dependence of algae and cyanobacteria on energy-consuming CO₂ concentrating mechanisms (CCMs) which 48 concentrate CO₂ around Rubisco, the key site for photosynthetic carbon fixation (Raven 49 50 & Beardall, 2014 and references therein; Hennon et al., 2015). The energy freed up from 51 the down-regulated CCMs under increased CO₂ concentrations can be applied to other 52 metabolic processes, resulting in a modest increase in algal growth (Wu et al., 2010; Hopkinson et al., 2011; Xu et al., 2017). Accordingly, elevated CO₂ availability could 53

The oceans have absorbed about one-third of anthropogenically released CO₂, which





54 potentially enhance marine primary productivity (Schippers et al., 2004). For instance, across 18 stations in the central Atlantic Ocean primary productivity was stimulated by 55 15-19% under elevated dissolved CO₂ concentrations up to 36 µM (Hein and 56 57 Sand-Jensen 1997). On the other hand, neutral effects of OA on growth rates of phytoplankton communities were reported in five of six CO₂ manipulation experiments in 58 59 the coastal Pacific (Tortell et al., 2000). Furthermore, simulated future OA reduced surface PP in pelagic surface waters of Northern SCS and East China Sea (Gao et al., 60 2012). It seems that the impacts of OA on PP could be region-dependent. The varying 61 effects of OA may be related to the regulation of other factors such as light intensity (Gao 62 63 et al., 2012), temperature (Holding et al., 2015), nutrients (Tremblay et al., 2006) and community structure (Dutkiewicz et al., 2015). 64 65 Taiwan Strait of the East China Sea, located between southeast Mainland China and 66 the Taiwan Island, is an important channel in transporting water and biogenic elements 67 between the East China Sea (ECS) and the South China Sea (SCS). Among the Chinese coastal areas, the Taiwan Strait is distinguished by its unique location. In addition to 68 riverine inputs, it also receives nutrients from upwelling (Hong et al., 2011). Primary 69 70 productivity is much higher in coastal waters than that in pelagic zones due to increased 71 supply of nutrients through river runoff and upwelling (Chen, 2003; Cloern et al., 2014). The South China Sea (SCS), located from the equator to 23.8 N, from 99.1 to 121.1 E 72 and encompassing an area of about 3.5×10^6 km², is one of the largest marginal seas in 73





74 the world. As a marginal sea of the Western Pacific Ocean, it has a deep semi-closed basin (with depths > 5000 m) and wide continental shelves, characterized by a tropical 75 76 and subtropical climate (Jin et al., 2016). Approximately 80% of ocean organic carbon is 77 buried in the Earth's continental shelves and therefore continental margins play an essential role in the ocean carbon cycle (Hedges & Keil, 1995). Investigating how ocean 78 79 acidification affects primary productivity in the Taiwan Strait and the SCS could help us to understand the contribution of marginal seas to carbon sink under the future 80 81 CO₂-increased scenarios. Although small-scale studies on OA impacts have been conducted in the ESC and the SCS (Gao et al., 2012, 2017), our understanding of how 82 OA affects PP in marginal seas is still fragmentary and superficial. In this study, we 83 conducted three cruises in the Taiwan Strait and the SCS, covering an area of 8.3×10^5 84 km², and aimed to provide in-depth insight into how OA and/or episodic pCO₂ rise 85 86 affects PP in marginal seas with comparisons to other types of waters. 2 Materials and Methods 87 2.1 Investigation areas 88 To study the impacts of projected OA (dropping by ~0.4 pH) on marine primary 89 90 productivity in different areas, we carried out deck-based experiments during the 3 91 cruises supported by National Natural Science Foundation of China (NSFC), which took place in the Taiwan Strait (Jul 14th-25th, 2016), the South China Sea basin (Sep 6-24th, 92 2016), and the West South China Sea (Sep 14th to Oct 24th, 2017), respectively. The 93





experiments were conducted at 101 stations with coverage of 12 °N-26 °N and 110 94 ^oE-120 ^oE (Fig. 1). Investigation areas include the coastal zone (< 50 m), the continental 95 shelf (50-200 m) and the slope (200-1000 m), and the vast deep-water basin (> 1000 m). 96 97 2.2 Measurements of temperature and carbonate chemistry parameters The temperature and salinity of surface seawater at each station were monitored with 98 99 an onboard CTD (Seabird, USA). pH_{NBS} was measured with an Orion 2-Star pH meter (Thermo scientific, USA) that was calibrated with standard National Bureau of Standards 100 (NBS) buffers (pH=4.01, 7.00, and 10.01 at 25.0 °C; Thermo Fisher Scientific Inc., USA). 101 The analytical precision was ±0.001. Total alkalinity (TAIK) was determined using Gran 102 103 titration on a 25-mL sample with a TA analyzer (AS-ALK1, Apollo SciTech, USA) that 104 was regularly calibrated with certified reference materials supplied by A. G. Dickson at the Scripps Institution of Oceanography (Gao et al., 2018a). The analytical precision was 105 $\pm 2 \,\mu\text{mol kg}^{-1}$. CO₂ concentration in seawater and the pH_{Total} (pH_T) values was calculated 106 107 by using CO2SYS (Pierrot et al., 2006) with the input of pH_{NBS} and TAIK data. 2.3 Nutrient measurement 108 Surface seawater was collected from the Conductivity Temperature Depth (CTD) 109 110 rosette/Niskin bottles with a clean 125 mL HDPE (High-Density Polyethylene) sample 111 container. The nitrate and nitrite concentrations in seawater were then measured with a Technicon AA3 Auto-Analyzer (Bran-Lube, GmbH, Germany). The quantitative limits 112 for nitrate and nitrite were $0.1 \mu mol L^{-1}$ and $0.04 \mu mol L^{-1}$, respectively. We used 113





certified reference materials (CRMS) (https://www.jamstec.go.jp/scor/) as external 114 quality checks, and the analytical precision was better than ±1% during the whole cruise. 115 Nutrient measurement was conducted in the cruise of the South China Sea basin. Due to 116 117 the limit of human resources, it was not conducted in the other two cruises. 2.4 Solar radiation 118 119 The incident solar radiation intensity during the cruises was recorded with an 120 Eldonet broadband filter radiometer (Eldonet XP, Real Time Computer, Germany). This device has three channels for PAR (400-700 nm), UV-A (315-400 nm) and UV-B (280-121 315 nm) irradiance, respectively, which records the means of solar radiations over each 122 123 minute. The instrument was fixed at the top layer of the ship to avoid shading. 2.5 Determination of primary productivity 124 Surface seawater (0-1m) was collected a 10 L acid-cleaned (1 M HCl) plastic bucket 125 126 and pre-filtered (200 µm mesh size) to remove large grazers. To prepare high CO₂ (HC) 127 seawater, CO₂-saturated seawater was added into pre-filtered seawater until a decrease of ~0.4 units in pH (corresponding CO₂ concentrations being 22.0-39.7 µM) was 128 approached (Gattuso et al., 2010). The same amount of filtered seawater (0.22 µm) was 129 130 added into the pre-filtered seawater setting as ambient CO₂ (AC) control. Prepared AC 131 and HC seawater was allocated into 50-mL quartz tubes in triplicate, inoculated with 5 μCi (0.185 MBq) NaH¹⁴CO₃ (ICN Radiochemicals, USA), and then incubated for 24 h 132 (over a day-night cycle) under 100 % incident solar irradiances in a water bath for





134 temperature control by running through surface seawater. After the incubation, the cells were filtered onto GF/F filters (Whatman) and immediately frozen at -20 °C for later 135 analysis. In the laboratory, the frozen filters were transferred to 20 mL scintillation vials, 136 thawed and exposed to HCl fumes for 12 h, and dried (55 °C, 6 h) to expel non-fixed ¹⁴C, 137 as previously reported (Gao et al., 2017). Then 3 mL scintillation cocktail (Perkin 138 139 Elmer®, OptiPhase HiSafe) was added to each vial. After 2 h of reaction, the incorporated radioactivity was counted by a liquid scintillation counting (LS 6500, 140 141 Beckman Coulter, USA). The carbon fixation for 24 h incubation was taken as chlorophyll (Chl) a-normalized daily net primary productivity (PP, µg C (µg Chl a)⁻¹) 142 143 (Gao et al., 2017). The changes (%) of PP induced by ocean acidification were expressed 144 as (PP_{HC}-PP_{AC})/PP_{AC}×100, where PP_{HC} and PP_{AC} are the net daily primary productivity under HC and AC, respectively. 145 146 2.6 Chl a measurement 147 Pre-filtered (200 µm mesh size) surface seawater (500-2000 mL) at each station was filtered onto GF/F filter (25 mm, Whatman) and then stored at -80 °C. After returning to 148 laboratory, phytoplankton cells on the GF/F filter were extracted overnight in absolute 149 methanol at 4 °C in darkness. After centrifugation (5000 g for 10 min), the absorption 150 151 values of the supernatants were analyzed by a UV-VIS spectrophotometer (DU800, Beckman, Fullerton, California, USA). The concentration of chlorophylls a (Chl a) was 152 calculated according to Porra (2002). 153





2.7 Data analysis

The data of environmental parameters were expressed in raw and the data of PP were the means of triplicate incubations. Two-way analysis of variance (ANOVA) was used to analyze the effects of OA and location on PP. Least significant difference (LSD) was used to for *post hoc* analysis. Linear fitting analysis was conducted with Pearson correlation analysis to assess the relationship between PP and environmental factors. A 95% confidence level was used in all analyses.

3 Results

During the cruises, surface temperature ranged from 25.0 to 29.9 $^{\circ}$ C in the Taiwan Strait and from 27.1 to 30.2 $^{\circ}$ C in the South China Sea (Fig. 2a). Surface salinity ranged from 30.0 to 34.0 in the Taiwan Strait and from 31.0 to 34.3 in the South China Sea (Fig. 2b). The lower salinities were found in the estuaries of Minjiang and Jiulong Rivers as well as Mekong River-induced Rip current. High salinities were found in the SCS basin. Surface pH_T changed between 7.91-8.20 in the Taiwan Strait with the higher values in the estuary of Minjiang River (Fig. 2c). On the contrary, surface pH had a narrower range (8.06-8.23) in the South China Sea and the lower values occurred near the islands in the Philippines. TAIK ranged from 2100 to 2359 μ mol L⁻¹ in the Taiwan Strait and 2126 to 2369 μ mol L⁻¹ in the South China Sea (Fig. 2d). The lowest value occurred in the estuary of Minjiang River. CO₂ concentration in surface seawater changed from 6.4-15.9 μ M kg⁻¹ SW in the Taiwan Strait, and 9.3-14.3 μ M kg⁻¹ SW in the SCS (Fig. 1e). It showed an





opposite pattern to surface pH, with the lowest value in the estuary of Minjiang River in 174 the Taiwan Strait and highest value in near the islands in the Philippines in the South 175 China Sea. During the PP investigation period, the daytime mean PAR intensity ranged 176 from 126.6 to 145.2 W m⁻² s⁻¹ in the Taiwan Strait and 37.3 to 150.0 W m⁻² s⁻¹ in the SCS 177 (Fig. 2f). 178 The concentration of Chl a ranged from 0.11 to 12.13 µg L⁻¹ in the Taiwan Strait (Fig. 179 3). The highest concentration occurred in the estuary of the Minjiang River. The 180 concentration of Chl a in the SCS ranged from 0.037 to 7.43 µg L⁻¹. The highest 181 concentration was found in the coastal areas of Guangdong province in China. For both 182 the Taiwan Strait and the SCS, there were high Chl a concentrations (> 1.0 µg L⁻¹) in 183 coastal areas, particularly in the estuaries of the Minjing River, Jiulong River and Pearl 184 River. On the contrary, Chl a concentrations in offshore areas were lower than 0.2 μ g L⁻¹. 185 Surface primary productivity changed from 99-302 μ g C (μ g Chl a)⁻¹ d⁻¹ in the 186 Taiwan Strait, and from 17-306 μ g C (μ g Chl a)⁻¹ d⁻¹ in the South China Sea (Fig. 4). 187 High surface primary productivity (> 200 μ g C (μ g Chl a)⁻¹ d⁻¹) was found in the 188 estuaries of the Minjing River, Jiulong River, and Pearl River and areas near the East of 189 Vietnam. In pelagic zones, the surface primary productivity was usually lower than 100 190 $\mu g C (\mu g Chl a)^{-1} d^{-1}$. 191 Through a series of onboard CO₂-enrich experiments we observed that effects of the 192 elevated pCO₂ on primary productivity of surface phytoplankton community ranged from 193





194	-88.03% (inhibition) to 56.87% (promotion), revealing significant regional differences
195	(ANOVA, $F_{(100, 404)} = 4.103$, $p < 0.001$, Fig. 5). Among 101 stations, 70 stations showed
196	insignificant OA effects. OA increased PP at 6 stations and reduced PP at 25 stations.
197	Positive effects of OA on surface primary productivity was observed in the Taiwan Strait
198	and the western SCS (Fig. 5, red-yellow shading areas), with the maximal enhancement
199	of 56.9% in the station approaching Mekong River plume (LSD, $p < 0.001$). Reduction in
200	PP induced by the elevated CO ₂ was mainly found in the central SCS basin within the
201	latitudes of 10 $^{\rm o}{\rm N}$ to 14 $^{\rm o}{\rm N}$ and the longitudes of 114.5 $^{\rm o}{\rm E}$ to 118 $^{\rm o}{\rm E}$ (Fig. 5, blue-purple
202	shading areas), with inhibition rates ranging from 24.02% to 88.03% (Fig. 5, LSD, $p <$
203	0.05). These results showed a region-related effect of OA on photosynthetic carbon
204	fixation of surface phytoplankton assemblages. Overall, the elevated pCO ₂ had neutral or
205	positive effects on primary productivity in nearshore waters, while having adverse effects
206	in pelagic waters.
207	By analyzing the correlations between OA-induced PP changes and regional
208	environmental parameters, we found that OA-induced changes in phytoplankton primary
209	productivity was significantly positively related with in situ pH ($p < 0.001$, $r = 0.379$),
210	NOx availability (the concentrations of $NO_3^- + NO_2^-$ at the bottom of upper mixing layers
211	as they were unmeasurable in the surface water, $p = 0.002$, $r = 0.727$), PAR density ($p = 0.002$)
212	0.002, $r = 0.311$) and primary productivity ($p = 0.004$, $r = 0.284$) (Fig. 6 and Table S1).
213	On the other hand, the influence induced by OA was negatively related to salinity that





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ranged from 30.00 to 34.28 (p < 0.001, r = -0.418).

4 Discussion

increased or did not affect PP in coastal waters but reduced it in pelagic waters. Our results suggested that the enhanced effects of the OA treatment on photosynthetic carbon 218 219 fixation depend on regions of different physicochemical conditions. Higher levels of 220 nutrients due to runoffs or upwellings should be mainly responsible for the enhancement. On the other hand, such stimulation could be related to higher UV-attenuation in these coastal waters that contain more organic matters (Hader et al., 2015), since we employed 222 UV-transparent vessels for the incubations. In addition, coastal diatoms appear to benefit more from OA than pelagic ones (Li et al., 2016). Therefore, community structure differences might also be responsible for the differences of the short-term high CO₂-induced acidification between coastal and pelagic waters. OA is deemed to have two kinds of effects at least (Xu et al., 2017; Shi et al., 2019). The first one is the enrichment of CO₂, which is usually beneficial for photosynthetic carbon fixation and growth of algae because insufficient ambient CO₂ limits algal 230 photosynthesis (Hein & Sand-Jensen, 1997; Bach & Taucher, 2019). The other effect is the decreased pH which could be harmful because it disturbs the acid-base balance between extracellular and intracellular environments. For instance, the decreased pH projected for future OA was shown to reduce the growth of the diazotroph 233

In the present study, we found that the elevated pCO₂ and associated pH drop





Trichodesmium (Hong et al., 2017), decrease PSII activity by reducing removal rate of 234 PsbD (D2) (Gao et al., 2018b) and increase mitochondrial and photo-respirations in 235 diatoms and phytoplankton assemblages (Yang and Gao 2012, Jin et al., 2015). In 236 237 addition, OA could reduce the RuBisCO transcription of diatoms, which also contributed to the decreased growth (Endo et al., 2015). Therefore, the net impact of OA depends on 238 239 the balance between its positive and negative effects, leading to enhanced, inhibited or neutral influences, as reported in diatoms (Gao et al., 2012, Li et al., 2021) and 240 241 phytoplankton assemblages in the Arctic and subarctic shelf seas (Hoppe et al., 2018), the North Sea (Eberlein et al., 2017) and the South China Sea (Wu and Gao 2010, Gao et al., 242 243 2012). 244 In the present study, OA increased or did not affect PP in coastal waters but reduced 245 it in offshore waters. This is significantly related to nutrient availability (Fig. 6d), with 246 that the inhibitory effect was minimized when NOx availability increased. Riverine 247 inputs, including the Minjiang River, Jiulong River, Pearl River, and Mekong River, are 248 the primary source of nutrients in the coastal and shelf zones, resulting in higher concentrations of nutrients and lower salinity in these waters (Xiao et al., 2018). It was 249 250 reported that elevated pCO₂ decreased net organic carbon production of 251 natural plankton community in nutrient-depleted waters (Yoshimura et al., 2010). Furthermore, OA did not affect the specific growth rate of a diatom under N-replete 252 condition but reduced it under N-limited condition (Li et al., 2018). The alleviating effect 253





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of nutrient enrichment on OA-induced stress could be multifaceted. Firstly, algae could cope with the acid-base perturbation caused by OA through active proton pumps (McNicholl et al., 2019). The operation of such proton pumps need some essential proteins, such as plasma membrane H⁺-ATPase, whose synthesis is nutrient-dependent (Taylor et al., 2012; Xu et al., 2017). Secondly, it has been shown that nutrient enrichment could accelerate the repair rate of PSII via synthesizing the key proteins such as PsbA (D1), and PsbD (D2) (Geider et al., 1993; Li et al., 2015). Thirdly, nitrogen enrichment could significantly increase the synthesis and content of photosynthetic pigments including Chl a, phycocyanin, and phycoerythrin (Johnson & Carpenter, 2018; Gao et al., 2019b), contributing to high photosynthetic activity under stressful environmental conditions. Negative correlation between OA-induced changes of PP and salinity was found in this study. While little has been documented on the relationship between salinity and OA (Wulff et al., 2018; Sugie et al., 2020; Xu et al., 2020), lowered salinity has been shown to alleviate the impact of OA on a coccolithorphorid (Xu et al., 2020). Nevertheless, we presume the enhanced PP could mainly be related to nutrient availability because lower salinity in coastal waters usually companies with high nutrient levels (Li et al., 2011). In addition, local pH may be another factor that affects the impacts of OA. There are diurnal and seasonal fluctuations of pH in coastal waters and phytoplankton that adapt well to the fluctuant pH environments would be tolerant to the decreased pH caused by OA (Flynn et al., 2012, Li et al., 2016). On the other hand, the





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surface pH in the ocean basin is relatively stable, with a varied range of only ~0.024 over a month (Hofmann et al., 2011). Therefore, the phytoplankton cells living in these environments could be more sensitive to pH drop due to elevated pCO₂ (Li et al., 2016). The specific environmental conditions have profound effects on shaping diverse dominant phytoplankton groups (Boyd et al., 2010). Larger eukaryotic groups (especially diatoms) usually dominate the complex coastal regions, while picophytoplankton (Prochlorococcus and Synechococcus), characterizing with more efficient nutrients uptake, dominate the relatively stable offshore waters (Dutkiewicz et al., 2015). In summer and early autumn, previous investigations demonstrated that diatoms dominated in the northern waters and the Taiwan Strait (coastal and shelf regions) with the high abundance of phytoplankton, which are consistent with our Chl a data; Prochlorococcus and Synechococcus dominated in the SCS basin and the north of SCS (slope and basin regions) (Xiao et al., 2018, Zhong et al., 2020). In addition, it has been reported that larger cells benefit more from OA because a thicker diffusion layer around the cells limits the transport of CO₂ (Feng et al., 2010; Wu et al., 2014). In contrast, a thinner diffusion layer and higher surface to volume ratio in smaller phytoplankton cells can make them easier to transport CO₂ near the cell surface and within the cells, and therefore picophytoplankton species are less CO₂-limited (Bao and Gao, 2021). Therefore, different community structures between coastal and pelagic areas could also be responsible for the enhanced and inhibitory effects of OA.





Conclusions

By investigating the impacts of the elevated pCO₂ on PP in the Taiwan Strait and the 295 SCS, we demonstrated that such short OA-treatments induced changes in PP were mainly 296 297 related to NOx availability based on Pearson correlation coefficients, supporting the hypothesis that negative impacts of OA on PP increase from coastal to pelagic waters 298 299 (Gao et al., 2019a). In view of ocean climate changes, strengthened stratification due to 300 global warming would reduce the upward transports of nutrients and further reduce 301 nutrient availability, consequently, leading to exacerbating impacts of OA on PP in pelagic zones. Meanwhile, PP in coastal and/or upwelled waters would be stimulated or 302 303 non-affected by OA with continuous discharges of nutrients from terrestrial environments, 304 which may imply higher PP and enhance frequency of harmful algal blooms in future 305 oceans. 306 Data availability. All data are included in the article or Supplement. 307 Author contributions. KG and TW developed the original idea and designed research. 308 TW and JS carried out fieldwork. GG provided statistical analyses and prepared figures. GG, KG, and XZ wrote the manuscript. All contributed to revising the paper. 309 310 Competing interests. The contact author has declared that neither they nor their 311 co-authors have any competing interests. Disclaimer. Publisher's note: Copernicus Publications remains neutral with regard to 312 jurisdictional claims in published maps and institutional affiliations. 313





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334 acidification on marine ecosystems and reliant human communities, Annu. Rev. Env. Resour., 45, 83-112, 2020. 335 Dutkiewicz, S., Morris, J. J., Follows, M. J., Scott, J., Levitan, O., Dyhrman, S. T., and 336 Berman-Frank, I.: Impact of ocean acidification on the structure of future 337 phytoplankton communities, Nat. Clim. Change, 5, 1002-1006, 2015. 338 339 Eberlein, T., Wohlrab, S., Rost, B., John, U., Bach, L. T., Riebesell, U., and Van de Waal, D. B.: Effects of ocean acidification on primary production in a coastal North Sea 340 phytoplankton community, Plos One, 12, 1-15, 2017. 341 Endo, H., Sugie, K., Yoshimura, T., and Suzuki, K.: Effects of CO₂ and iron availability 342 343 on rbcL gene expression in Bering Sea diatoms, Biogeosciences, 12, 2247-2259, 2015. 344 Feng, Y., Hare, C. E., Rose, J. M., Handy, S. M., DiTullio, G. R., Lee, P. A., Smith, W. O., 345 346 Peloquin, J., Tozzi, S., Sun, J., Zhang, Y., Dunbar, R. B., Long, M. C., Sohst, B., 347 Lohan, M., and Hutchins, D. A.: Interactive effects of iron, irradiance and CO₂ on Ross Sea phytoplankton, Deep-Sea Res. PT. I, 57, 368-383, 2010. 348 Flynn, K. J., Blackford, J. C., Baird, M. E., Raven, J. A., Clark, D. R., Beardall, J., 349 350 Brownlee, C., Fabian, H., and Wheeler, G. L.: Changes in pH at the exterior surface of plankton with ocean acidification, Nat. Clim. Change, 2, 510-513, 2012. 351 Gao, G., Xu, Z. G., Shi, Q., and Wu, H. Y.: Increased CO2 exacerbates the stress of 352 ultraviolet radiation on photosystem II function in the diatom Thalassiosira 353





weissflogii, Environ. Exp. Bot., 156, 96-105, 2018b. 354 Gao, G., Gao, Q., Bao, M. L., Xu, J. T., and Li, X. S.: Nitrogen availability modulates the 355 effects of ocean acidification on biomass yield and food quality of a marine crop 356 357 Pyropia yezoensis, Food Chem, 271, 623-629, 2019b. Gao, G., Jin, P., Liu, N., Li, F. T., Tong, S. Y., Hutchins, D. A., and Gao, K. S.: The 358 359 acclimation process of phytoplankton biomass, carbon fixation and respiration to the combined effects of elevated temperature and pCO₂ in the northern South China Sea, 360 361 Mar. Pollut. Bull., 118, 213-220, 2017. Gao, G., Xia, J. R., Yu, J. L., Fan, J. L., and Zeng, X. P.: Regulation of inorganic carbon 362 363 acquisition in a red tide alga (Skeletonema costatum): The importance of phosphorus availability, Biogeosciences, 15, 4871-4882, 2018a. 364 Gao, K. S., Beardall, J., Häder, D. P., Hall-Spencer, J. M., Gao, G., and Hutchins, D. A.: 365 366 Effects of ocean acidification on marine photosynthetic organisms under the 367 concurrent influences of warming, UV radiation, and deoxygenation, Front. Mar. Sci., 6, 1-18, 2019a. 368 Gao, K. S., Xu, J. T., Gao, G., Li, Y. H., Hutchins, D. A., Huang, B. Q., Wang, L., Zheng, 369 370 Y., Jin, P., Cai, X. N., Hader, D. P., Li, W., Xu, K., Liu, N. N., and Riebesell, U.: 371 Rising CO₂ and increased light exposure synergistically reduce marine primary productivity, Nat. Clim. Change, 2, 519-523, 2012. 372 Gattuso, J. P., Gao, K. S., Lee, K., Rost, B., and Schulz, K. G.: Approaches and tools to 373





manipulate the carbonate chemistry, pp 41-52. Guide to best practices for ocean 374 acidification research and data reporting, edited by: Riebesell, U., Fabry, V. J., 375 Hansson, L., and Gattuso J.-P., Luxembourg: Publications Office of the European 376 377 Union, 2010. Gattuso, J. P., Magnan, A., Bill & R., Cheung, W. W. L., Howes, E. L., Joos, F., Allemand, 378 379 D., Bopp, L., Cooley, S. R., Eakin, C. M., Hoegh-Guldberg, O., Kelly, R. P., Portner, H. O., Rogers, A. D., Baxter, J. M., Laffoley, D., Osborn, D., Rankovic, A., Rochette, 380 J., Sumaila, U. R., Treyer, S., and Turley, C.: Contrasting futures for ocean and 381 society from different anthropogenic CO₂ emissions scenarios, Science, 349, 382 383 aac4722, 2015. Geider, R. J., La Roche, J., Greene, R. M., and Olaizola, M.: Response of the 384 photosynthetic apparatus of Phaeodactylum tricornutum (Bacillariophyceae) to 385 386 nitrate, phosphate, or iron starvation, J. Phycol., 29, 755-766, 1993. 387 Häder, D. P., Williamson, C. E., Wängberg, S. A., Rautio, M., Rose, K. C., Gao, K. S., Helbling, E. W., Sinha, R. P., and Worrest, R.: Effects of UV radiation on aquatic 388 ecosystems and interactions with other environmental factors, Photoch. Photobio. 389 390 Sci., 14, 108-126, 2015. 391 Hedges, J. I., and Keil, R. G.: Sedimentary organic matter preservation: an assessment and speculative synthesis, Mar. Chem., 49, 81-115, 1995. 392 Hein, M., and Sand-Jensen, K.: CO₂ increases oceanic primary production, Nature, 388, 393





394 526-527, 1997. Hennon, G. M. M., Ashworth, J., Groussman, R. D., Berthiaume, C., Morales, R. L., 395 Baliga, N. S., Orellana, M. V., and Armbrust, E. V.: Diatom acclimation to elevated 396 CO₂ via cAMP signalling and coordinated gene expression, Nat. Clim. Change, 5, 397 761-765, 2015. 398 399 Hofmann, G. E., Smith, J. E., Johnson, K. S., Send, U., Levin, L. A., Micheli, F., Paytan, A., Price, N. N., Peterson, B., Takeshita, Y., Matson, P. G., Crook, E. D., Kroeker, K. 400 J., Gambi, M. C., Rivest, E. B., Frieder, C. A., Yu, P. C., and Martz, T. R.: 401 High-frequency dynamics of ocean pH: A multi-ecosystem comparison, Plos One, 6, 402 403 1-11, 2011. Holding, J. M., Duarte, C. M., Sanz-Mart n, M., Mesa, E., Arrieta, J. M., Chierici, M., 404 Hendriks, I. E., Garcia-Corral, L. S., Regaudie-de-Gioux, A., Delgado, A., Reigstad, 405 406 M., Wassmann, P., and Agusti, S.: Temperature dependence of CO₂-enhanced 407 primary production in the European Arctic Ocean, Nat. Clim. Change, 5, 1079-1082, 2015. 408 Hong, H. S., Chai, F., Zhang, C. Y., Huang, B. Q., Jiang, Y. W., and Hu, J. Y.: An 409 410 overview of physical and biogeochemical processes and ecosystem dynamics in the 411 Taiwan Strait, Cont. Shelf Res., 31, S3-S12, 2011. Hong, H. Z., Shen, R., Zhang, F. T., Wen, Z. Z., Chang, S. W., Lin, W. F., Kranz, S. A., 412 Luo, Y. W., Kao, S. J., Morel, F. M. M. and Shi, D. L.: The complex effects of ocean 413





433

414 acidification on the prominent N₂-fixing cyanobacterium *Trichodesmium*. Science, 356, 527-530, 2017. 415 Hopkinson, B. M., Dupont, C. L., Allen, A. E., and Morel, F. M.: Efficiency of the 416 417 CO₂-concentrating mechanism of diatoms, P. Natl. Acad. Sci. USA., 108, 3830-3837, 2011. 418 419 Hoppe, C. J. M., Wolf, K. K. E., Schuback, N., Tortell, P. D., and Rost, B.: Compensation of ocean acidification effects in Arctic phytoplankton assemblages. Nat. Clim. 420 Change, 8, 529-533, 2018. 421 Jin, P., Gao, G., Liu, X., Li, F. T., Tong, S. Y., Ding, J. C., Zhong, Z. H., Liu, N. N., and 422 423 Gao, K. S.: Contrasting photophysiological characteristics of phytoplankton assemblages in the Northern South China Sea, Plos One, 11, 1-16, 2016. 424 Jin, P., Wang, T. F., Liu, N. N., Dupont, S., Beardall, J., Boyd, P. W., Riebesell, U., and 425 426 Gao, K. S.: Ocean acidification increases the accumulation of toxic phenolic 427 compounds across trophic levels, Nat. Commun., 6, 1-6, 2015. Johnson, M. D., and Carpenter, R. C.: Nitrogen enrichment offsets direct negative effects 428 of ocean acidification on a reef-building crustose coralline alga, Biol. Letters, 14, 429 430 1-5, 2018. 431 Lan, J., Hong, J. and Li, P.: Seasonal variability of cool-core eddy in the Western South

Li, F. T., Wu, Y. P., Hutchins, D. A., Fu, F. X., and Gao, K. S.: Physiological responses of

China Sea, Adv. Earth Sci., 21, 1145-1152, 2006.





- coastal and oceanic diatoms to diurnal fluctuations in seawater carbonate chemistry
- under two CO₂ concentrations, Biogeosciences, 13, 6247-6259, 2016.
- 436 Li, F. T., Beardall, J., and Gao, K. S.: Diatom performance in a future ocean: interactions
- between nitrogen limitation, temperature, and CO₂-induced seawater acidification,
- 438 ICES J. Mar. Sci., 75, 1451-1464, 2018.
- 439 Li, G., Gao, K. S., Yuan, D. X., Zheng, Y., and Yang, G. Y.: Relationship of
- 440 photosynthetic carbon fixation with environmental changes in the Jiulong River
- estuary of the South China Sea, with special reference to the effects of solar UV
- radiation, Mar. Pollut. Bull., 62, 1852-1858, 2011.
- 443 Li, H. X., Xu, T. P., Ma, J., Li, F. T., and Xu, J. T.: Physiological responses of
- 444 Skeletonema costatum to the interactions of seawater acidification and the
- combination of photoperiod and temperature, Biogeosciences, 18, 1439-1449, 2021.
- 446 Li, W., Gao, K., and Beardall, J.: Nitrate limitation and ocean acidification interact with
- 447 UV-B to reduce photosynthetic performance in the diatom *Phaeodactylum*
- *tricornutum*, Biogeosciences, 12, 2383-2393, 2015.
- 449 McNicholl, C., Koch, M. S., and Hofmann, L. C.: Photosynthesis and light-dependent
- 450 proton pumps increase boundary layer pH in tropical macroalgae: A proposed
- mechanism to sustain calcification under ocean acidification, J. Exp. Mar. Biol.
- 452 Ecol., 521, 1-12, 2019.
- 453 Mostofa, K.M., Liu, C.Q., Zhai, W., Minella, M., Vione, D., Gao, K., Minakata, D.,





454 Arakaki, T., Yoshioka, T., Hayakawa, K. and Konohira, E.: Reviews and Syntheses: Ocean acidification and its potential impacts on marine ecosystems, Biogeosciences, 455 13, 1767-1786, 2016. 456 Pierrot, D., Wallace, D.W. R., and Lewis, E.: MS Excel program developed for CO₂ 457 system calculations. ORNL/CDIAC-105a, Carbon Dioxide Information Analysis 458 459 Center, Oak Ridge National Laboratory, US Department of Energy, Oak Ridge, Tennessee, USA., 2006. 460 Porra, R. J.: The chequered history of the development and use of simultaneous equations 461 for the accurate determination of chlorophylls a and b, Photosynth. Res., 73, 462 463 149-156, 2002. Raven, J. A., and Beardall, J.: CO₂ concentrating mechanisms and environmental change, 464 Aquat. Bot., 118, 24-37, 2014. 465 Schippers, P., Lürling, M., and Scheffer, M.: Increase of atmospheric CO₂ promotes 466 467 phytoplankton productivity, Ecol. Lett., 7, 446-451, 2004. Shi, D. L., Hong, H. Z., Su, X., Liao, L. R., Chang, S. W., and Lin, W. F.: The 468 physiological response of marine diatoms to ocean acidification: Differential roles of 469 470 seawater *p*CO₂ and pH, J. Phycol., 55, 521-533, 2019. 471 Sugie, K., Fujiwara, A., Nishino, S., Kameyama, S. and Harada, N.: Impacts of 472 temperature, CO₂, and salinity on phytoplankton community composition in the Western Arctic Ocean, Front. Mar. Sci., 6, 1-17, 2020. 473





- 474 Taylor, A. R., Brownlee, C., and Wheeler, G. L.: Proton channels in algae: Reasons to be
- 475 excited, Trends Plant Sci., 17, 675-684, 2012.
- 476 Tortell, P. D., Rau, G. H., and Morel, F. M. M.: Inorganic carbon acquisition in coastal
- 477 Pacific phytoplankton communities, Limnol. Oceanogr., 45, 1485-1500, 2000.
- 478 Tremblay, J. E., Michel, C., Hobson, K. A., Gosselin, M., and Price, N. M.: Bloom
- dynamics in early opening waters of the Arctic Ocean. Limnol. Oceanogr., 51,
- 480 900-912, 2006.
- 481 Riebesell, U., Aberle-Malzahn, N., Achterberg, E. P., Alguer ó-Mu ñiz, M.,
- 482 Alvarez-Fernandez, S., Ar stegui, J., Bach, L. T., Boersma, M., Boxhammer, T.,
- 483 Guan, W. C., Haunost, M., Horn, H. G., Loscher, C. R., Ludwig, A., Spisla, C.,
- 484 Sswat, M., Stange, P., and Taucher, J.: Toxic algal bloom induced by ocean
- acidification disrupts the pelagic food web, Nat. Clim. Change, 8, 1082-1086, 2018.
- 486 Wu, Y., Gao, K., and Riebesell, U.: CO2-induced seawater acidification affects
- physiological performance of the marine diatom *Phaeodactylum tricornutum*,
- 488 Biogeosciences, 7, 2915-2923, 2010.
- 489 Wu, Y., Campbell, D. A., Irwin, A. J., Suggett, D. J., and Finkel, Z. V.: Ocean
- 490 acidification enhances the growth rate of larger diatoms. Limnol. Oceanogr., 59,
- 491 1027-1034, 2014.
- 492 Wu, Y. P., and Gao, K. S.: Combined effects of solar UV radiation and CO2-induced
- 493 seawater acidification on photosynthetic carbon fixation of phytoplankton





494 assemblages in the South China Sea. Chinese Sci. Bull., 55, 3680-3686, 2010. Wulff, A., Karlberg, M., Olofsson, M., Torstensson, A., Riemann, L., Steinhoff, F. S., 495 Mohlin, M., Ekstrand, N., and Chierici, M.: Ocean acidification and desalination: 496 497 Climate-driven change in a Baltic Sea summer microplanktonic community, Mar. Biol., 165, 1-15, 2018. 498 499 Xiao, W. P., Wang, L., Laws, E., Xie, Y. Y., Chen, J. X., Liu, X., Chen, B. Z., and Huang, B. Q.: Realized niches explain spatial gradients in seasonal abundance of 500 501 phytoplankton groups in the South China Sea, Prog. Oceanogr., 162, 223-239, 2018. Xu, J. K., Sun, J. Z., Beardall, J., and Gao, K. S.: Lower salinity leads to improved 502 503 physiological performance in the coccolithophorid *Emiliania huxleyi*, which partly ameliorates the effects of ocean acidification, Front. Mar. Sci., 7, 1-18, 2020. 504 Xu, Z. G., Gao, G., Xu, J. T., and Wu, H. Y.: Physiological response of a golden tide alga 505 506 (Sargassum muticum) to the interaction of ocean acidification and phosphorus 507 enrichment, Biogeosciences, 14, 671-681, 2017. Yang, G. Y., and Gao, K. S.: Physiological responses of the marine diatom *Thalassiosira* 508 pseudonana to increased pCO₂ and seawater acidity, Mar. Environ. Res., 79, 509 510 142-151, 2012. 511 Yoshimura, T., Nishioka, J., Suzuki, K., Hattori, H., Kiyosawa, H. and Watanabe, Y. W.: Impacts of elevated CO₂ on organic carbon dynamics in nutrient depleted Okhotsk 512 Sea surface waters. J. Exp. Mar. Biol. Ecol., 395, 191-198, 2010. 513





Zhong, Y. P., Liu, X., Xiao, W. P., Laws, E. A., Chen, J. X., Wang, L., Liu, S. G., Zhang,
F., and Huang, B. Q.: Phytoplankton community patterns in the Taiwan Strait match

the characteristics of their realized niches, Prog. Oceanogr., 186, 1-15, 2020.





517	Figure captions
518	Fig. 1 Sampling stations for the incubation experiments in the Taiwan Strait and the
519	South China Sea during three cruises. Taiwan Strait cruise was conducted in July 2016
520	(red dots), South China Sea Basin cruise were conducted in September 2016 (blue dots)
521	and Western South China Sea cruise was conducted in September 2017 (black dots). The
522	arrows represent surface circulation fields in summer in the vicinity of Vietnam coast
523	based on Lan et al. (2006).
524	Fig. 2 Temperature (°C, panel a), salinity (panel b), pH (panel c), total alkalinity (μmol
525	L^{-1} , panel d), and CO_2 ($\mu mol\ kg^{-1}\ SW$, panel e) in surface seawater and mean PAR
526	intensity (W m ⁻² s ⁻¹ , panel f) during the PP incubation experiments.
527	Fig. 3 Chl a concentration ($\mu g L^{-1}$) in the Taiwan Strait and the South China Sea during
528	research cruises.
529	Fig. 4 Surface primary productivity ($\mu g \ C \ (\mu g \ Chl \ a)^{-1} \ d^{-1}$) in the Taiwan Strait and the
530	South China Sea during research cruises.
531	Fig. 5 Ocean acidification (pH decreases of 0.4 units) induced changes (%) of surface
532	primary productivity in the Taiwan Strait and the South China Sea. Red-yellow shading
533	represents a positive effect on PP and blue-purple shading represents a negative effect.
534	Positive effect was found in coastal waters and estuary affected waters, such as the
535	Taiwan Strait, the Pearl River plume, Mekong River induced Rip current in West China
536	Sea. Negative effect was found in surface of oligotrophic waters like SCS Basin.





Fig. 6 Ocean acidification (pH decreases of 0.4 units) induced changes (%) on surface primary productivity in the South China Sea as a function of salinity (a), PAR (b), ambient pH (c), and nitrate plus nitrite concentration (d). The dotted lines represent 95% confidence intervals.





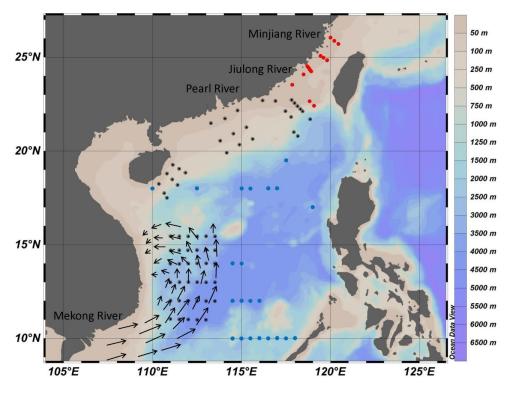


Fig. 1



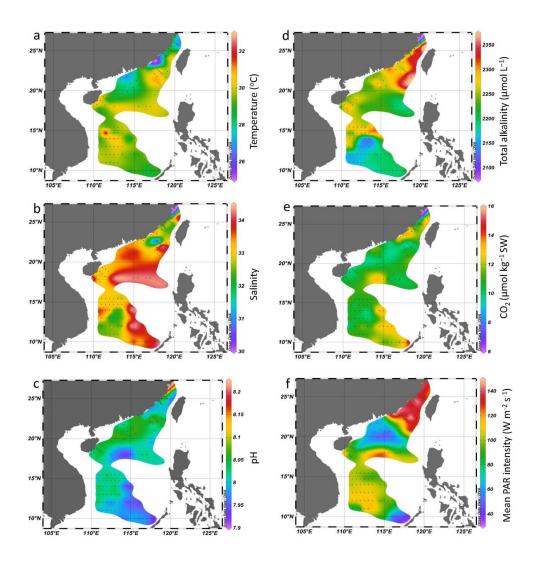


Fig. 2





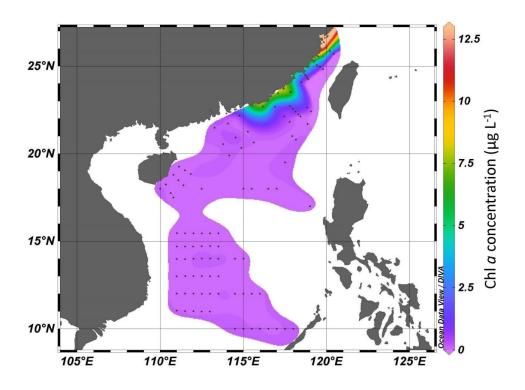


Fig. 3





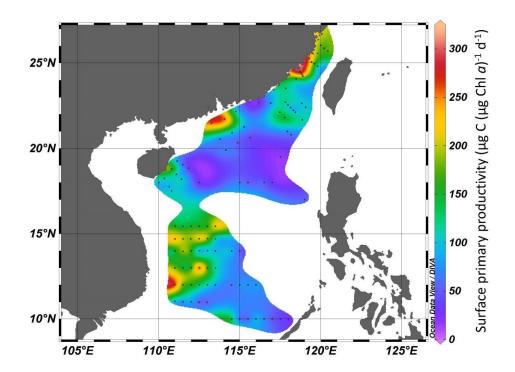


Fig. 4





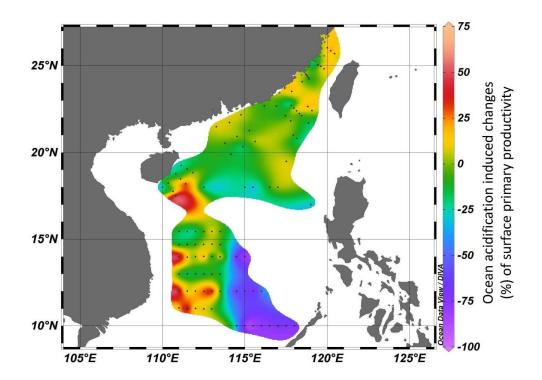


Fig. 5





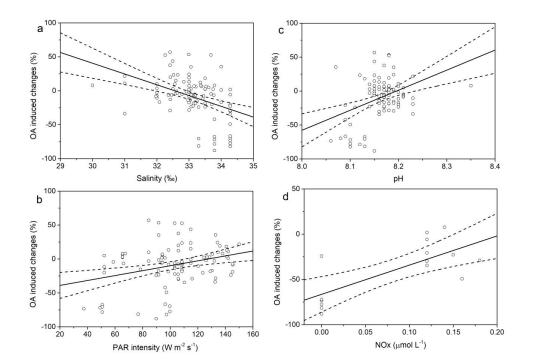


Fig. 6