Dissolved organic matter signatures in urban surface waters: spatio-temporal patterns and drivers

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Abstract

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Advances in analytical chemistry have facilitated the characterization of dissolved organic matter (DOM), which has improved understanding of DOM sources and transformations in surface waters. For urban waters, however, where DOM diversity is likely to be high, the interpretation of DOM signatures is hampered by a lack of basic information on the influence of land cover and anthropogenic factors such as nutrient enrichment and release of organic contaminants. Here we explored the spatiotemporal variation of DOM composition in contrasting urban water bodies, based on spectrophometry spectrophotometry and fluorometry, size-exclusion chromatography and ultrahighresolution mass spectrometry, to identify linkages between DOM signatures and potential drivers. The highly diverse DOM we observed distinguished lakes and ponds, which are characterized by a high proportion of autochthonous DOM, from rivers and streams with more allochthonous where allochthonous DOM is more prevalent. Seasonal variation in DOM composition was apparent in all types of water bodies, driven by apparently due to the interactions between phenology and urban influences. Specifically, such as, nutrient supply, the percentage of green space adjacent surrounding to the water bodies and point source pollution-emerged as major urban drivers of DOM composition. Optical DOM properties also revealed the influence of effluents from waste waterwastewater treatment plants, suggesting their that simple optical measurements can be useful in water-quality assessment and monitoring. Furthermore, optical measurements, informing about processes both within water bodies and in their surroundings, catchments. which could improve the assessment of ecosystem functioning and integrity.

1 Introduction

Urban freshwaters typically receive high loads of organic carbon, nutrients and micropollutants, ranging from pharmaceuticals and personal care products to industrial chemicals and more (Schwarzenbach et al., 2006). Although routine wastewater treatment is increasingly effective, chemical stressors in urban freshwaters remain widespread. Prominent reasons are pollution legacies (Ladwig et al., 2017; Baume and Marcinek, 1993) and continued uncontrolled inputs, particularly by stormwater runoff (Council, 2009). In addition, urban surface waters tend to suffer from severe hydromorphological modifications. This includes the lateral and vertical disconnection from floodplains and aquifers and results in large impacts on the extent and complexity of riverine habitat (White and Walsh, 2020). Furthermore, the disruption of connectivity limits the self-purification capacity of urban surface waters (D'arcy et al., 2007), which can lead to turbid water and visually unpleasant and potentially harmful algal blooms(Carpenter et al., 1998). This and the resulting failure of citizens to recognize limited recognition of urban freshwaters as providers of ecosystem services (Huser et al., 2016) calls for improved water management strategies that consider ecological in addition to hygienic and chemical criteria (Gessner et al., 2014).

The concentration and chemical composition of dissolved organic matter (DOM), typicallygenerally quantified as dissolved organic carbon (DOC), are key characteristics of aquatic ecosystems. Both concentration and composition are governed by allochthonous inputs and internal biological production and transformation processes (Williams et al., 2016). Typically, however, water quality monitoring only considers concentration and bulk quality properties (e.g. biological oxygen demand, BOD) as measures of DOM availability to, and degradation by, heterotrophic microbes (Jouanneau et al., 2014). This narrow-focus is at odds with the extreme diversity of DOM observed in freshwaters, where thousands of compounds can be chemically distinguished (Kellerman et al., 2014; Peter et al., 2020; Stanley et al., 2012). This high diversity and its the strong spatio-temporal variation of DOM composition suggestsuggests much potential for DOM characteristics to provide insights into the state of freshwater ecosystems in water quality assessment and monitoring. In fact, additional insights into freshwater ecosystems may be gained if the very high diversity of DOM can be used to inform about water quality for ecosystem assessment and monitoring purposes.

Recent progress in analytical methods has increasingly enabled the detailed characterization of DOM to elucidate the sources and fates in surface waters (Xenopoulos et al., 2021). Optical properties can inform not only about the chemical characteristics of DOM but also, for example, about large-scale gradients in aquatic networks (Creed et al., 2015) or the degree of aquatic-terrestrial ecosystem coupling (Sankar et al., 2020; Lambert et al., 2015; Yamashita et al., 2010; Catalán et al., 2013). Fluorescence excitation-emission matrices (EEM) can be processed by parallel factor analysis (PARAFAC) to identify independently fluorescing DOM components (Cory and Mcknight, 2005). Size-exclusion chromatography partitions bulk DOM into molecular size fractions, which also tend to differ in origin and bioavailability (Huber et al., 2011). Finally, the advent of ultrahigh-resolution mass spectrometry (FT-ICR-MS or Orbitrap-MS) has greatly refined the characterization of DOM, revealing associations between compositional turnover of DOM differing in molecular diversity and landscape-scale environmental gradients in lakes (Kellerman et al., 2014) and rivers (Peter et al., 2020).

In the present study we explored variation in the chemical composition of DOM over time and space in contrasting urban surface waters, hypothesizing that a detailed chemical characterization of DOM yields signatures of various human influences. To this end, we explored linkages between chemical DOM-composition of DOM and potential drivers determining DOM signatures, including land cover, eutrophication and chemical pollution, which we captured by using a suite of proxies. Our specific goals were to: (i) describe spatio-temporal patterns of DOM composition across a range of urban freshwaters encompassing streams, rivers, ponds and lakes; (ii) identify environmental factors accounting for the observed patterns; and thereby (iii) explore how information on DOM composition could be included in urban freshwater assessment and monitoring, complementing approaches and metrics currently used.

2 Methods

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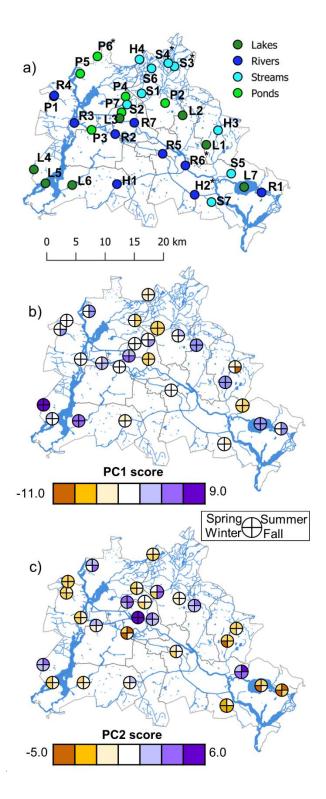
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2.1 Study sites

The study was conducted in 32 freshwater sites located in the city of Berlin, Germany. Nearly 6.5% of the municipal area (889 km²) is covered by freshwaters. These comprise 60 lakes (>1 ha), about 500 ponds, the two slow-flowing lowland rivers, the Rivers Spree and Havel, and numerous streams, ditches and canals. Selection of the 32 study sites followed a stratified random sampling design (Fig. 1a, Supporting Information Table S1A1). Based on geographical information for Berlin's water bodies, we randomly selected 7 sites in each of 4 strata: lakes, ponds, rivers and streams. Rivers and streams were classified according to a width cutoff of 5 m. Monitoring data on water chemistry (Berlin city administration, SenUVK 2009-2014) were used in a cluster analysis to identify highly polluted sites. These were excluded from the pool used for randomly selecting study sites. Instead, two such rivers (H1 and H2) and two streams (H3 and H4) were deliberately added as polluted sites to lengthen the environmental gradient.

Instead, two organically polluted rivers (H1 and H2) and streams (H3 and H4) were deliberately added to lengthen the environmental gradient. Sites H1 and H2 received WWTP effluents (Fig. 1a, Table A2) and sites H3 and H4 presented high levels of diffused pollution. Other sites for, for some of which monitoring data were unavailable (streams and ponds), were also affected by pollution (Table A1): Pond P4 was formerly connected to an old waste water treatment plant and still receives stormwater inflow during heavy rain events; -S5 is located immediately downstream of a WWTP; and R7 became a receiving stream in 2015 (Nega et al., 2019), which was too recent for the site to become classified as polluted based on the monitoring data.-Land use data obtained from the Berlin city administration (Senate Administration for Environment, 2017) were used to calculate the proportion of paved areas and green spaces within a 50-m perimeter around the selected water bodies using open-source geoinformation software (Qgis Development Team, 2017). The 50-m perimeter was chosen to capture influences in the immediate vicinity of the sites, such as of the riparian zone and slightly beyond, but not of the whole catchments, which are highly variable in size and tend to be difficult to define in urban areas. Delineation of the 50-m perimeter enabled us to distinguish particularly between urban sites adjacent to paved surfaces vs. green spaces. Tufekcioglu (2020) and Johnson (2005) used buffer zones of similar size and a study on ponds using perimeters of up to 3200 m found 50 and 100 m to be most appropriate to assess land-cover effects (Declerck, 2006). We repeatedly sampled all 32 sites in each of four campaigns conducted over an annual cycle, first in spring (April-May 2016), then in summer (July-August 2016), autumn (SeptemberOctober 2016) and winter (February March 2017). We also obtained data on land use from the Berlin city administration(Senate Administration for Environment, 2017), which we used to calculate the proportion of paved and green areas within 50 m buffer strips adjacent to each of the selected water bodies using open source geoinformation software (QGIS Development Team, 2017, Open Source Geospatial Foundation Project. http://qgis.osgeo.org). All samples were taken during base flow conditions (Fig. E1).



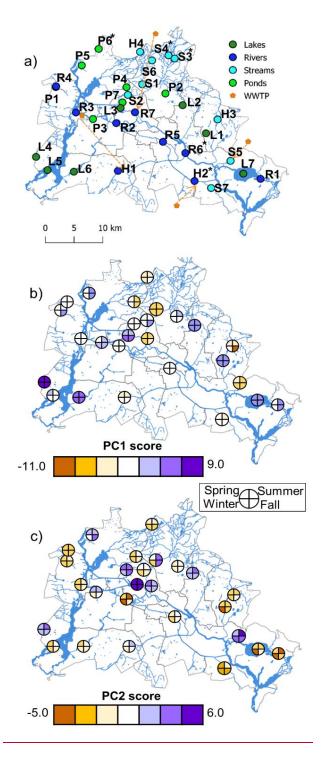


Figure 1 Map of 32 sampling sites in the city of Berlin, including 7 lakes (dark green), 7 ponds (light green), 9 streams (light blue), and 9 rivers (dark blue), including two heavily polluted stream sites and two heavily polluted river sites (a), and PCA scores for these sites in four different seasons (b, c). Site codes are given in Table S1. Wastewater Treatment Plants (WWTP) are shown in orange, arrows point to locations where the effluents are discharged (a). Scores of a Principal Component Analysis (PCA) of DOM characteristics are shown as color gradients for all sites sampled in four seasons (b, c). The PCA is based on DOC concentrations, all absorbance and fluorescence data, absolute component-specific fluorescence intensities from PARAFAC, and data from size-exclusion chromatography. Different colours indicate differences in DOM composition. Site codes are given in Table A1. Sites marked by asterisks (*) were restricted to 3 seasons and hence excluded from the PCA.

2.2 Physico-chemical field measurements and water sampling

We repeatedly sampled all 32 sites in each of four campaigns conducted over an annual cycle, first in spring (April-May 2016), then in summer (July-August 2016), autumn (September-October 2016) and winter (February-March 2017). All field visits occurred during base flow conditions (Fig. E1), During each field visit, wWe measured water temperature, pH, the dissolved oxygen (DO) concentration and electrical conductivity using a hand-held WTW Multiprobe 3320 (pH320, OxiCal-SL, Cond340i, Weilheim, Germany) or a smarTROLL probe (In-Situ, Fort Collins, CO, USA). We also collected integrative water samples (2 L) from the upper 0.5 m water layer for chlorophyll-a and DOM analyses. The water was kept cool in acid-washed polycarbonate Nalgene bottles placed in a cooling box pending filtration in the laboratory (GF75, 0.3 µm average pore size; Advantec, Tokyo, Japan) within 6 hours after sampling. Additional volumes of surface water were filtered through pre-combusted glass fiber filters (GF75) directly in the field. These filters were placed intoin acid-washed, pre-combusted (450 °C, 4h4 h) glass vials (15-20 mL) sealed with a PTFE septum in a screw-cap for later measurements of dissolved organic carbon (DOC) concentrations, DOM fluorescence and absorbance, and DOM molecular size distribution. The water passed through the filter was collected in acid-washed polyethylene tubes for analyses of soluble reactive phosphorus (SRP), nitrate (NO₃-), nitrite (NO₂-), ammonium (NH₄⁺) and trace organic compounds (TrOCs). We also took unfiltered water samples for total phosphorus (TP) analysis. For each variable, we collected three replicate samples at each site in each season. We stored all samples in the dark in a cooling box during transport. To preserve samples and remove all inorganic carbon, we acidified (pH 2) the water for DOC, NO₃-, NO₂- and NH₄+ analyses with 2 M HCl within 6 hours after sample collection. DOC concentrations and DOM fluorescence and absorbance were measured within 24 h. Filtered water for analyses of SRP, NO₃-, NO₂-, NH₄⁺ and TrOCs was frozen at -20 °C.

2.3 DOM characterization

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We determined total DOC concentrations by high-temperature catalytic combustion and infrared spectrometry on a TOC-V Analyzer (Shimadzu, Kyoto, Japan), with a 0.5 mg L⁻¹ limit of quantification and a typical analytical precision of 3%. DOM absorbance and fluorescence were simultaneously determined on an Aqualog instrument (Horiba Ltd, Kyoto, Japan). We used the absorbance spectra to calculate several indexes (Table A2) using ultra-pure water as a blank. –From each site and season, we measured three analytical replicates of each of the three independent samples.
**We generated nine measurements (each of three process replicates was measured 3 times) immediately after 3 blanks.
The high level of replication allowed identification of artefactual measurements and outlier removal following a visual check of absorbance spectra and fluorescence excitation-emission matrices (EEMs). The Ffluorescence data was expressed in Raman units, removing the need for an external quantification standard. **

Iron can form stable complexes with DOC and interfere with optical DOC measurements, so that the two variables are not independent (Maranger et al., 2003). We found that the quotient of light absorbance at 420 nm (a_{420}) and DOC concentration, a measure of the optical signal returned per unit DOC (Weyhenmeyer et al., 2014), was indeed significantly correlated (p = 0.002) with the Fe concentration measured in the monitoring program of the Senate of Berlin in two of our lakes (L5 and L7: 0.06 ± 0.03 mg/L), four of the rivers (H1, R1, R6 and R7: 0.30 ± 0.14) and two of the streams (H3 and H4: 0.31 ± 0.14). The relationship explained 31% of the overall variation (Fig. E2).

Consequently, Fe could have influenced our optical estimates of DOC concentration. However, because our analysis rests on differences in DOM composition as opposed to concentration (see below), it is unlikely that the presence of Fe notably influenced the spatial and temporal patterns observed in our study.

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We calculated several indices from the absorbance spectra (Table B2): the specific UV absorption (SUVA₂₅₄) as a proxy for DOM aromaticity (Weishaar et al., 2003), the ratio of absorbance at 250 and 365 nm (E2:E3) as an (inverse) indicator of molecular size (Peuravuori and Pihlaja, 1997), the ratio of E4:E6 as an indicator of humification (Chen et al., 1977), the short wavelength slope within the wavelength region of 275 295 nm (Helms et al., 2008) as an inverse correlate with molecular weight and aromaticity, and the ratio of slopes (SR) computed from short and long wavelength regions (Loiselle et al., 2009) as another negative correlate with DOM molecular weight. We used the fluorescence data to compute the freshness index β/α (Table A2B2) (Wilson and Xenopoulos, 2009), which indicates the relative importance of recently produced DOM (Parlanti et al., 2000). Furthermore, we calculated the fluorescence index (FIXFI) as the ratio of fluorescence intensities at the emission wavelengths of 470 and 520 nm (obtained at an excitation wavelength of 370 nm), which has proved useful to distinguish the relative contributions of terrestrial (FIX-1.4) and aquatic (FIX-1.9) sources of DOM (Mcknight et al., 2001) plants (FI~1.2) and microbes or algae (FI~1.4) as sources of DOM (Cory et al., 2010; Cory and Mcknight, 2005; Jaffé et al., 2008; Fellman et al., 2010). Finally, we computed the humification index (HIX) as a proxy for humic substances (Ohno, 2002). Fluorescence excitation emission matrices (EEMs) EEMs were used for PARAFAC analysis, a multivariate three-way modeling approach decomposing EEMs into individual fluorophores (Bro, 1997; Stedmon and Bro, 2008). We derived 87 components from a total of 116 EEMs and compared their loading spectra with the OpenChrom/OpenFluor database (http://www.openfluor.org) (Murphy et al., 2014). See Supporting Information for details of data processing, including PARAFAC analysis.

Prior to PARAFAC, we interpolated missing data in Rayleigh scatter regions to expedite the modeling process (Bro, 1997). The calculations for PARAFAC were performed using Matlab (version 7.11.0, MathWorks) and the DOMFluor Toolbox (1.7) following Stedmon & Bro (2008). We limited the number of components to 10, rigorously checked residual EEM plots, and assessed the final models by split-half validation (Fig. B1) as recommended by Stedmon and Bro (2008).

-The molecular size distribution of DOM was analyzed by liquid size-exclusion chromatography in combination with UV and IR detection of organic carbon and UV detection of organic nitrogen (LC-OCD-OND) (Huber et al., 2011). The instrument was calibrated with IHSS Suwannee River I Humic Acid and Fulvic Acid standards (International Humic Substance Society, St Paul, MN, USA). Carbon and nitrogen detectors were calibrated with potassium hydrogen phthalate (C) and sodium nitrate (N). Limits of quantification were 0.1 mg C L⁻¹, and 0.01 mg N L⁻¹, analytical precision of based on repeated standard measurements was better than 3%. We determined concentrations of three molecular size fractions: humic-like substances (HS-C and HS-N reported in mg C L⁻¹ and mg N L⁻¹, respectively), high-molecular weight non-humic substances (reported as HMWS-C and HMWS-N, in mg C L⁻¹ and mg N L⁻¹) and low-molecular weight substances (LMWS, in mg C L⁻¹).

To examine the molecular composition of DOM, we used ultrahigh-resolution Fourier-Transform Ion Cyclotron Mass Spectrometry (FT-ICR-MS). We extracted DOM on Agilent Bond Elut PPL solid phase columns (Dittmar et al., 2008) from 1 L of filtered water acidified to pH 2.We extracted DOM on Agilent Bond Elut PPL solid-phase columns (Dittmar et al., 2008) from 1 L of filtered water acidified to pH 2. We then diluted extracts to 10 µg L⁻¹ C in 1/1 ultrapure water/methanol before broadband mass spectrometry on a 15 Tesla Solarix FT-ICR-MS (Bruker Daltonics, Bremen, Germany) in electrospray ionization negative mode (300 accumulated scans, ion accumulation time of 0.1 s, flow rate of 240 µL/h). We performed internal mass calibration and exported the raw mass lists from 150 to 1000 Da for further data processing using previously established R code (Del Campo et al., 2019). Briefly, we first applied a method detection limit similar to Riedel & Dittmar (Riedel and Dittmar, 2014) before aligning m/z values across samples (Del Campo et al., 2019). Briefly, we first applied a method detection limit similar to Riedel & Dittmar (2014) before aligning m/z values across samples (Del Campo et al., 2019). Subsequently, we assigned chemical formulas to mean m/z values assuming single-charged deprotonated molecular ions and Cl-adducts for a maximum elemental combination of C₁₀₀H₂₅₀O₈₀N₄P₂S₂, respecting chemical constraints and using rigorous mass error assessments, stable isotope confirmation and homologous series assessment (Del Campo et al., 2019). More detail of the FT ICR MS methods can be found in the Supplement material. To condense the mass spectrometric information, we derived 12 molecular groups. To eliminate doubtful formula assignments, we performed (i) an accurate assessment of mass error including its partitioning into random and systematic components (Savory et al., 2011): (ii) an exploration for stable isotope validation by daughter peaks (Koch et al., 2007), and (iii) a homologous series assessment based on CH₂, CO₂ and H₂O as chemical building blocks for aliphatic, acid-based and alcohol-based elongation (Koch et al., 2007). To condense the mass-spectrometric data, we grouped formulas into 12 non-overlapping molecular groups (Lesaulnier et al., 2017) based on elemental composition and calculated the average molecular mass, number of formulas (molecular richness) and total intensity for each of them. In addition, we computed the double-bond equivalents (DBE) and the aromaticity index (AI) as indicators of unsaturated compounds (Koch and Dittmar, 2006), and the molecular lability boundary (MLB) as a measure of lability (D'andrilli et al., 2015). Finally, we used van Krevelen plots to present the sum formulas derived from the FT-ICR-MS data in a space defined by O:C (oxygen richness) and H:C (saturation) ratios. We used random order of plotting to avoid bias due to systematic overplotting of thousands of compounds with identical O:C and H:C ratios.

2.4 Additional water-chemical analyses

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We determined total DOC concentrations by high-temperature catalytic combustion and infrared spectrometry on a TOC V Analyzer (Shimadzu, Kyoto, Japan). NO₃, NO₂ and NH₄ were analyzed on a FIAcompact (MLE GmbH, Dresden, Germany). TP was measured using the same technique with unfiltered water samples that were digested with K₂S₂O₈ (30 min at 134 °C). We measured chlorophyll-*a* concentrations spectrophotometrically (HITACHI U2900; Tokyo, Japan) following hot ethanol extraction (Jespersen and Christoffersen, 1987). Tokyo, Japan) following hot ethanol extraction (Jespersen and Christoffersen, 1987) of three GF75 filters from each individual water sample. Concentrations of 18 trace organic compounds (TrOCs) were determined by HPLC-MS/MS (Shimadzu, Kyoto, Japan)

(Zietzschmann et al., 2016). These included chemicals such as acesulfame (a sweetener), benzotriazole (a corrosion inhibitor), and drug residues like carbamazepine and gabapentin (Table B4C1).

2.5 Data analysis

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We used repeated-measures ANOVA to test for differences among types of water bodies and seasonal-sampling periods (referred to as seasons hereafter) for a variety of response variables; there was non significant as the interaction between water body type and season. Further was not significant we recomputed models including main effects only. Furthermore, we assessed the importance of seasonal variation in each water body type by computing a respective variance component using a type-II ANOVA (aka variance component analysis) for data from each water body type with season and site -ID as random factors; this approach assesses facilitates the assessment of temporal variation as a fraction of total variation within each water body type. Normal distribution was assessed graphically by quantile plots and histograms. For ANOVA, data were $\log(x)$ or \sqrt{x} -transformed to achieve conditions of normality and variance homogeneity of the residuals.

For constrained multivariate analyses we considered land cover adjacent to the water bodies, trophic state and micropollutant load as drivers of variation in DOM chemical composition. We used the percentages of urban green space and paved areas as a proxyproxies for land cover—and assessed trophic state based on, concentrations of TP, NH₄+, NO₃- and chlorophyll a. Finally, as a measure of trophic state; and the first axis of a principal component analysis (PCA) based on the mean TrOC dataset was used concentration—as a proxy for micropollutant load. We also performed a principal component analysis with all the TrOCs.

We followed a three-step approach to analyze the spatio-temporal patterns of DOM composition: First we identified major axes of variation in DOM composition by a PCA based on quantitative indicators of DOM, analytically accessible fractions thereof or quantitative proxies: DOC concentration, all absorbance and fluorescence data, absoluteindices, component-specific fluorescence intensities from PARAFAC normalized to DOC, and the results from-size-exclusion chromatography data. Only the 27 sites sampled in all four seasons were included in this analysis. All variables were standardized to a mean of zero with a variance of 1 to ensure equal weightingsweighting, and projected onto the ordination space using Pearson correlations of the variables with PCA axes in a distance biplot (sensu Legendre and Legendre-(, 2012). To explore spatial patterns, we mapped PC1 and PC2 scores onto Berlin's landscape using QGIS (QGIS Development Team, 2017, Open Source Geospatial Foundation Project; http://qgis.osgeo.org).).

Second, we used the same dataset as <u>the</u> dependent matrix in a redundancy analysis (RDA) with the set of potential drivers described above <u>used</u> as predictor variables. <u>The goal of the RDA was to identify potential drivers of DOM composition and thereby assess, reciprocally, whether various DOM descriptors are ecologically informative. We started with the full RDA model and forward-selected drivers (Legendre and Legendre, 2012). For hypothesis tests in the RDA, permutations were restricted to account for repeated measurements at the same sites across seasons by first permuting sets of four seasonal measurements across sites and then permuting across seasons within each site. To check our ability to identify drivers behind major variation observed in DOM composition, we used Procrustes analysis</u>

to assess the similarity of PCA and RDA ordinations, including a permutation-based test of the non-randomness of the achieved superimposition (Mardia, 1979; Peres-Neto and Jackson, 2001).

Third, we exploited results of the FT-ICR-MS to facilitate interpretation of the two major axes of variation in DOM chemical composition resulting from the PCA. The FT-ICR-MS data were only available for three seasons and were purely compositional (relative intensities), as the many thousands of compounds contained in the spectra cannot be calibrated to yield concentrations. To link the quantitative and compositional datasets, we correlated PCA scores—of PCA—axes with compound-specific relative intensities of the mass spectra. The compound-specific correlation coefficients were then used as eolorcolour codes in van Krevelen plots, which locate chemical formulae identified by FT ICR MS in a space defined by oxygen richness (O:C) and saturation (H:C). FT-ICR-MS-derived information such as the richness or average weight of specific molecular groups werewas also projected onto the PCA ordination space as arrows, provided correlation coefficients were >0.2. All statistical analyses and graphs were made with R 3.2.4 (R Core Team, 2016). (R Core Team, 2016).

275 3. Results

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3.1 Physico-chemical characteristics

Among all physico-chemical variables, only DOC concentration and temperature differed significantly among types of water bodies (p<0.05 and p<0.001, respectively). Temperature varied strongly across seasons, but still proved significantly different among water body types, with lakes and rivers being warmer than ponds and streams. DOC concentrations did not vary across seasons, but were significantly higher in ponds and streams than in lakes and rivers. Ponds also showed the highest chlorophyll-*a* concentrations and rivers the lowest, but these differences were not significant.

Separate ANOVAs for each water body type showed that seasonal variation in TP and NH_4^+ concentrations was highest in rivers and streams- (Table B1). Seasonal variation in NO_3^- concentrations was generally high, but systematic differences were neither detected among seasons nor sites (Table A1B1). Seasonal variation of chlorophyll-a concentrations was also high and similar across types of water bodies.

The analysis of TrOCs identified acesulfame, a widely used artificial sweetener -(Buerge et al., 2009), in 72 out of a total of 120 samples taken at 32 sites across all seasons (Table B1). Similarly, two corrosion inhibitors included in the analysis(Buerge et al., 2009), in 72 out of a total of 120 samples taken at 32 sites across all seasons (Table C1). Similarly, two corrosion inhibitors, benzotriazole and methylbenzotriazole (Cotton and Scholes, 1967; Tamil Selvi et al., 2003), occurred in 68 and 63 samples, respectively. Fifteen other TrOCs were detected in at least 2 and up to 62 samples (Table B4C1). Rivers showed the highest concentrations throughout the year. The first principal component of the PCA considering all TrOCs explained 61% of the total variance (Fig. B4C1) and separated streams and rivers with higher concentrations from ponds and lakes where concentrations of TrOCs were lower and often undetectable, particularly in ponds (Table B2):C2). The strong positive correlations between most of the TrOCs suggested the applicability of a simple average TrOC concentration as a proxy for micropollutant load in further analysis; this mean was computed across all TrOCs after z-standardization of each TrOC for equal weighting.

3.2 DOM composition

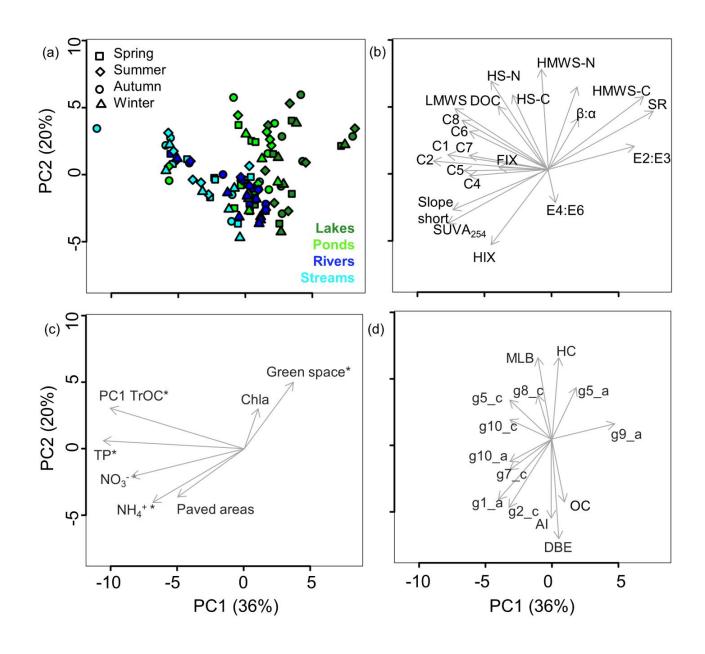
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PARAFAC modeling resulted in <u>87</u> components referred to as C1-<u>C8C7</u> (Table <u>A3B3</u>, Fig. <u>A1B1</u>). Components C6 and <u>C8C7</u> were previously found to be protein-like, whereas all other components have been reported as humic-like (Table <u>A3B3</u>). In contrast to the standard physico-chemical variables <u>we measured</u> and <u>the results from</u> size-exclusion chromatography (Table <u>A6B6</u>), the PARAFAC components and absorbance and fluorescence indices generally showed significant differences among water body types (Table <u>A4B4</u> and <u>A5B5</u>).

The first axis of the PCA analyzing spatio-temporal patterns of DOM chemical composition explained 3634% of the total variance (Fig. 2). PC1 was largely defined by the negative loadings for C2C1 and C1C2 (representing humic substances originating from waste waterwastewater treatment), the short wavelength slope, SUVA₂₅₄ and LMWS (Fig. 2b). Furthermore, PC1 correlated positively with the absorption slope ratio, E2:E3 (molecular size), β/α and HMWS-C. This axis separated water body types, from lakes on the right to ponds, rivers, and finally streams on the left. The optical proxies identified PC1 as a gradient spanning from lakes, where DOM had lower aromaticity and contained more freshly produced material, to streams, which showed high aromaticity and low proportions of fresh DOM. Pond P4, which was identified as an outlier because of particularly high NH₄⁺ concentrations, also showed a rather distinct DOM composition.



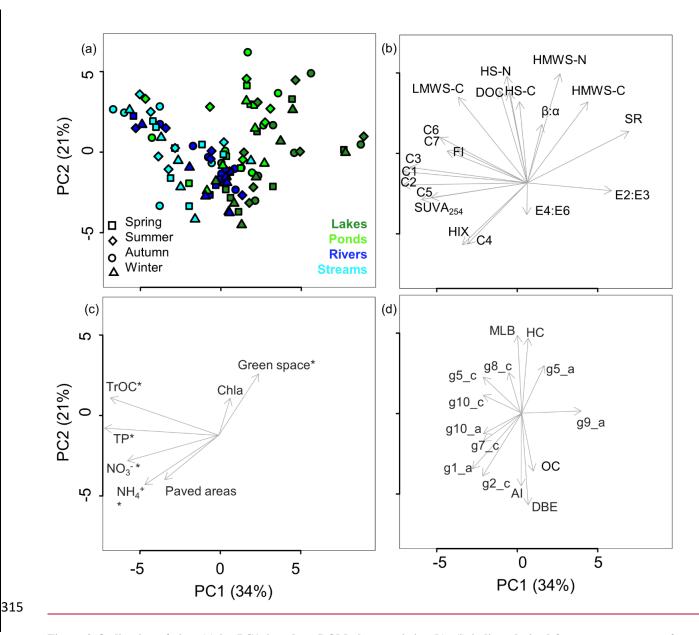
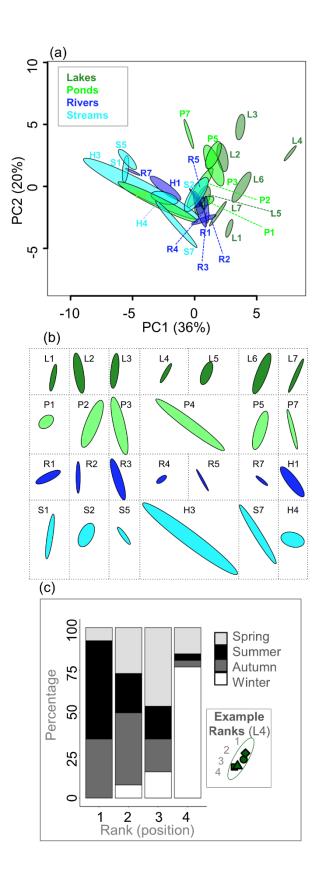


Figure 2 Ordination of sites (a) by PCA based on DOM characteristics (b): (i) indices derived from measurements of absorbance (E2:E3 indicating molecular size, E4:E6 representing the humification ratio, the slope ratio SR, and SUVA₂₅₄) and fluorescence (freshness index β:α, fluorescence index FIX, short-wavelength slope FI, and humification index HIX), (ii) PARAFAC components C1 to C8C7, and (iii) data from size exclusion chromatography (humic-like substances HS, high-molecular weight non-humic substances HMWS, low-molecular weight substances LMWS). (c) RDAPotential drivers of DOM composition, that were used as constraints in the RDA, were mapped onto the PCA ordination, with the significant constraints marked by an asterisk (*). (d) FT-ICR-MS-derived indices and molecular groups mapped onto the PCA ordination representing only groups correlated with PC1 or PC2 (r>0.2; oxygen richness O:C, saturation level indicated by H:C, double-bond equivalents DBE, aromaticity index AI, molecular lability boundary MLB, molecular groups g1 and g2 indicating black carbon without and with heteroatoms, g5 consisting of unsaturated aliphatics, g7 representing saturated fatty acids, g8 and g9 denoting carbohydrates without and with heteroatoms N, S or P, and g10 comprising peptides). The molecular group measures are either average masses (marked by 'mama') or counts of molecules (marked by 'c'eeg).

PC2 explained an additional $\frac{2021}{9}$ % of the total variance and correlated positively with HMWS (mg N/L) and β/α , and negatively with HIX. An exploration of spatio-temporal variation by plotting site-specific PC scores (Fig. 3) identified PC2 as the axis capturing temporal variation, with the four seasons aligning vertically at most sites. Winter and summer had the lowest and highest PC2 scores, respectively, with transitional seasons located in between. Thus, higher proportions of humic substances in winter contrast with more labile DOM in summer. In agreement with the variable-specific seasonal variance components, the degree of seasonal differentiation differed among water body types also in multivariate space, being higher in streams and ponds than in the larger lakes and rivers (Fig. 3b). Except for sites P4 andsite H3, two water bodies behaving exceptionally also in many other respects, seasonal variability was poorly reflected by PC1, which largely captured variation among individual water bodies or water body types, separating flowing from standing waters. Visual inspection of PCA scores mapped across Berlin (Fig. 1b,c) did not reveal a spatial signature transcending types of water bodies. RDA identified the areal percentage of green space adjacent to the water bodies, TP, NH₄+, NO₃- and the first axis of the PCA based on TrOCsmean TrOC concentration as significantly predictors of DOM composition (Fig. C1D1). The resulting PCA and RDA ordinations for DOM were significantly strongly correlated (Procrustes rotation 0.73, p<0.001), suggesting that the considered predictors were indeed major drivers of variation in DOM chemical composition.



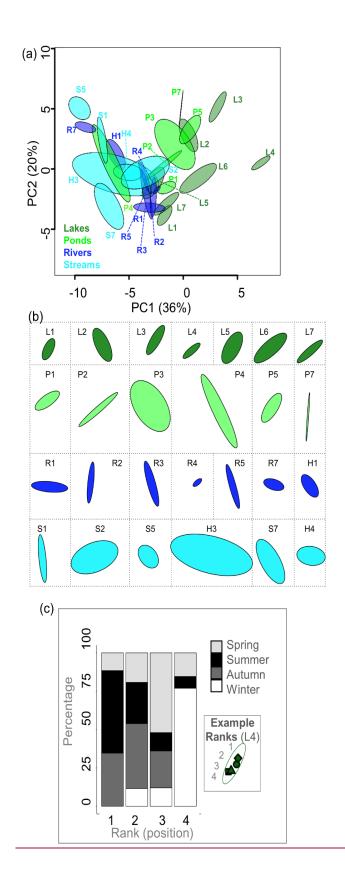


Figure 3 (a) PCA biplot based on DOM absorbance and fluorescence indices, PARAFAC components and size exclusion chromatography data from 4 contrasting types of urban freshwater bodies, including lakes, ponds, rivers, and streams, in addition to two streams and two rivers specifically selected as highly polluted sites. Each of the ellipses represents one sampling site that was visited 4 times, once in each season. Site codes are given in Table \$141. (b) Visual comparison of site-specific seasonal variation based on the size, shape and orientation of ellipses, plotted separately per site. (c) Seasonal variation across sites illustrated by ranking the sampling dates at each site according to the PC2 scores, as shown in the inset. The stacked histograms show frequencies of the seasons across the four ranks. Summer samples tend to produce high scores at most sampling sites, whereas winter samples tend to score low.

High-resolution mass spectrometric analyses of samples from three seasons provided additional insights into the chemical composition of DOM. Overall, we detected 6446 molecular formulas, most of them representing molecular groups typical of humic material derived from soils. This includes highly unsaturated O-rich compounds, polyphenols and other aromatic structures, followed by unsaturated aliphatic polyphenols, and polycyclic aromatic compoundses with aliphatic chains. The van Krevelen plots revealed a positive correlation of lignin-like molecules and carbohydrates with PC1 of DOM and identified these molecules as abundant in lakes (Fig. A2B2). In contrast, the negative association of proteins with PC1 was typical of streams. Information on the molecular groups identified by FT-ICR-MS and projected on the PCA space (Fig. 2d) showed carbohydrates and sugars containing N, S or P to be positively related to PC1. Furthermore, PC1 was negatively related to black carbon, polyphenols and polycyclic aromatic compounds with aliphatic chains, which are all typical of soil-derived humic material, as well as with unsaturated aliphatics, saturated fatty acids and peptides, indicating that all of these molecular groups were more important in streams. Lastly, the computed molecular lability boundary (MLB), carbohydrates, sugars without heteroatoms (N, S or P) and unsaturated aliphatics were positively related to PC2, while AI, DBE, black carbon and polyphenols were negatively related to PC2.

4. Discussion

4.1 Spatial patterns and drivers of DOM signatures

Our results show that the chemical composition of DOM in contrasting surface waters of the metropolitan area of Berlin, Germany, is highly diverse. This reflects both aquatic-terrestrial linkages and DOM transformations within the aquatic systems (Fonvielle et al., 2021) and suggests a high ecosystem level functional diversity across the urban aquatic network. Clear differences among the four types of water bodies we investigated were due to distinct signatures of streams and rivers vs. ponds and lakes. This was revealed especially by the first principal component (PC1) of a PCA (Fig. 2), which reflects the dominant gradient defined by variation in DOM composition across the 32 urban sites included in the study. Since optical measurements play an important role in our analysis of DOM, it is important to consider potential interference by iron. Elevated iron concentrations lead to brownification, similar to effects of allochthonous DOM, and Fe and DOC also form stable complexes, so that the two variables are not independent (Maranger et al, 2003). However, Fe data available from the Senate of Berlin for two of our lakes (L5 and L7: 0.06 ± 0.03 mg/L), four of the rivers (H1, R1, R6 and R7: 0.30 ± 0.14) and two of the streams (H3 and H4: 0.31 ± 0.14), all concentrations were below the threshold of 1 mg/L, suggesting that Fe increases may result in a420/DOC increases (Weyhenmeyer et al, 2014).

Stream DOM exhibited higher aromaticity (as indicated by SUVA₂₅₄) and lower amounts of recently produced, low-molecular DOM (as indicated by the freshness index or the slope ratio) than lakes at the opposite end of the gradient. This pattern matches results from agricultural streams near Berlin, where SUVA₂₅₄ values up to 3 L m⁻¹ mg⁻¹ have beenwere reported (Graeber et al., 2012) and from an urban river in southwestern Korea (SUVA₂₅₄ values of 2.5 L m⁻¹ mg⁻¹) (Park, 2009). The distinct signature is also reflected in other DOM components, such as the fluorophore C2, which was more important in streams and identified as terrestrial humic material (Murphy et al., 2011). Streams also showed higher levels of humic-like (C1) and protein-like (C8C7) compounds, whereas higher values of the freshness index characterized lakes. These patterns consistently indicate that the arrangement of sites along PC1 reflects a gradient of allochthonous vs autochthonous sources of DOM. A corollary of this finding is that despite the potentially pervasive influence of the urbanized surroundings, urban streams in particular are more tightly linked to the terrestrial environment than urban lakes, just as is the case for flowing and standing waters in natural landscapes (Larson et al., 2014).

In contrast to natural landscapes, however, the linkage of urban waters with their terrestrial surroundings is mediated by paved surfaces and engineered flow paths, including roof run-off into rain gutters, extensive (partially leaky) sanitation networks and sewage overflows in WWTPs that are activated following heavy rainfall or snowmelt. The urban gradient from allochthonous to autochthonous DOM sources we document could thus be driven by surface run-off rather than soil seepage and subsequent delivery of DOM to surface waters via groundwater Although we did not sample after major storms (Fig. E1), we would expect legacy effects of past runoff events to differ among sites, depending on the extent of green space and impervious surface area in the surroundings of the sites. — This interpretation is supported by higher levels of proteins (Fig. 2) characterizing the urban streams and rivers, as opposed to soil-derived humic DOM signatures typical of unimpacted streams and rivers (Hutchins et al., 2017). The proteins could originate from surface runoff integrating various sources of urban pollution but they might also derive from WWTPs, as implied by the nature of some of the PARAFAC components we identified (Table A3B3). For instance, the humic fluorophore C2 has been reported in WWTP effluents that may be discharged into urban surface waters (Murphy et al., 2011). Point-source inputs were also identified as drivers of DOM composition by the influence of TrOCs in our RDA and their correlations with C2 and C8C7, all of which are components of WWTP effluents.

Lakes differ from streams by a typically greater importance of autochthonous production. Since this production is fostered by abundant nutrients. Elevated nutrient supply (given sufficient light), elevated nutrient concentrations should hence coincide with DOM signatures indicative of autochthonous carbon sources, assources. This pattern has been found in agricultural streams, where the freshness index β : α indicating autotrophic activity was related to high nitrogen concentrations (Wilson and Xenopoulos, 2009). This patternHowever, it contrasts with the negative relation between nitrogen concentration and the proportion of fresh DOM found across our study sites, where high nutrient concentrations were instead strongly related to DOM components of WWTP effluents. This typically resulted in an allochthonous DOM character at high-nitrogen sites. Notably, signatures like lower β/α in WWTP-impacted sites may also be a consequence of the highly processed nature of DOM that underwent degradation in a WWTP.

Similarly, the TP concentration was significantly related to DOM composition in our RDA, where phosphorus-rich water bodies also proved to have more allochthonous than autochthonous DOM. This points to inputs from urban surface runoff rather than groundwater inflow where long flow paths and residence times provide ample opportunities for phosphorus immobilization. As with N, additional phosphorus may derive from WWTP effluents, as suggested by the positive relationship between TP concentration and the fluorophore C2 as a putative tracer of WWTP effluents (Murphy et al., 2011). Overall, the negative relationships between nutrient availability and the importance of autochthonous components in the DOM pool suggests that while streams and rivers may efficiently collect N and P from the urban environment; lakes are more efficient at channeling nutrients into autochthonous production. Thus, the autochthonous DOM signature in urban lakes appears to be largely independent of nutrient supply and rather be facilitated by longer water residence times, higher water temperature and favorable light conditions.

Our results on urban surfaces driving urban allochthonous DOM composition meet our expectation that land cover notably influences the composition of DOM in urban surface waters (Williams et al., 2016; Sankar et al., 2020). This conclusion is supported by results fromof our RDA, which identified the presence of green spaces in the perimeter of the water bodies as a significant influence. However, the relationship between land cover and DOM composition must be interpreted with caution because all lakes were situated in areas with green spaces in their surroundings, whereas streams ran through areas dominated by buildings and paved surfaces. The urban running waters, more than lakes and ponds, thus received high surface runoff during rain events, including high inputs of pollutants and allochthonous DOM. Given the evident negative relationship between green space and paved surface areas (R= 0.47, p<0.001), green spaces might be used as an inverse proxy for paved surfaces influencing DOM signatures in urban surface waters. However, since paved surface area per se did not emerge as a significant predictor in our RDA, land cover can at best partly account for the observed variation in DOM composition across the contrasting urban sites we investigated.

Except for ponds and some lakes, all investigated water bodies had direct surface water connections, which could result in spatial autocorrelation-(Peterson et al., 2006b; Peterson et al., 2006a). In addition, spatial patterns may arise from the prominent land cover gradients in Berlin, ranging from forested areas to densely populated urban centers. Since the sampling design of our study does not lend itself to a formal analysis of spatial autocorrelation, we explored spatial patterns with DOM proxies in maps (Fig. 1b,c) but found no obvious relationships. Instead, type-specific characteristics of the water bodies were pronounced, largely independent of hydrological connections. Factors potentially contributing to the resulting heterogeneity across the surface waters in the city include specific local stressors such as point-source inputs of pollutants, spatially variable urban surface runoff delivering allochthonous DOM, and hydraulic-engineering structures such as sluices. Thus, our map of DOM composition (Fig. 1b,c) could be interpreted as visualizing urban-heterogeneity in aquatic ecosystem diversity and condition the conditions of urban surface freshwaters.

4.2 Seasonal patterns and drivers of DOM signatures

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Seasonal variation in DOM signatures occurred in all types of water bodies mostly independent from variation among the four water body types. With a few exceptions, P4 and H3 being the most prominent examples example, seasonal variation of DOM composition was consistent across all water body types. (Fig. 3a,b), Assessed separately at each site

(Fig. 3b), DOM was generally fresher in summer and autumn than in winter and spring, as indicated by higher ratios of β : α and more HMWS-N as indicators of polysaccharides and proteins (Thurman, 1985), whereas humic matter was more abundant in winter, and the pattern in spring was not clear-cut. Our rank-based analysis of PC2 scores (Fig. 3c) suggests a consistent seasonal pattern of changes in DOM composition across sites, which emerged even though the variation within individual sites was limited along PC2.

At least four potential processes could account for the observed seasonal turnover in DOM composition: exudates of aquatic primary producers, microbial and sunlight-induced transformation of DOM, and terrestrial inputs from riparian vegetation (Spencer et al., 2009; Cory et al., 2015), all of which could be influenced by the urban environment. Seasonal variation in light conditions could be important in influencing DOM composition by primary producers, independent of nutrient supply (see above), and temperature changes might also play a role, especially in determining rates of microbial DOM transformations. Pulses of leaf litter falling or swept or blown into urban water bodies could be an additional source of DOM varying with season (Gessner et al., 1999). This holds particularly for urban green spaces and water courses lined by woody riparian vegetation. However, quantification of the relative importance of different drivers of seasonal patterns remains difficult based on the data currently available for urban settings.

The ponds and streams included in our study showed higher and less predictable seasonal turnoverchanges in DOM composition than the lakes and rivers, as revealed by the pattern along PC2 (Fig. 3). This indicates indicates that the nature and degree of aquatic-terrestrial coupling in urban settings leaves an imprint on seasonal changes in DOM composition. Therefore, the turnover of DOM. Surveysmore extensive the time series data from surveys of DOM dynamics should hence be more informative, the better can they inform about ecosystem conditions than assessments based on single grab samples or averaged data; complementing established procedures in water quality assessment and monitoring. Inputs of DOM from WWTP effluents may also be captured by the seasonal patterns, although that influence is likely variable, as indicated by considerable seasonal turnover of DOM at site H3contrasting with a minimal turnover at sites S5 and R7 (small ellipses in Fig. 3b), despite the influence of WWTP effluents at those sites.

4.3 DOM composition as a potential basis for urban surface water monitoring

The fact that our analysis of DOM composition revealed behaviorspecific characteristics of individual water bodies underlines the potential usefulnessyalue of DOM descriptors as ecosystem scale functional indicators that could be included in regular water-quality assessment and monitoring. Some sites deviated from the general pattern observed for water bodies of the same type. P4, for example, was formerly connected to an sewage farmold waste water treatment plant and appeared to be influenced by previously unrecognized storm waterstormwater runoff-that likely delivered inputs during heavy rain. The site was characterized by. This legacy matches the particularly high levels of nutrients characterizing this site, especially NH₄⁺, andcombined with a distinct DOM composition. Similarly, S5, located immediately downstream of a WWTP, although not specifically selected as a highly polluted site, also showed a distinct DOM composition as reflected by its highly negative PC1 score (Fig. 2a), indicating that the allochthonous influence was likely the strongest among all sites. Site R7 showed the same pattern as S5, and although not initially recognized as being affected by a WWTP, its DOM composition revealed that it had actually received WWTP effluents, which happened since the end 2015 (Nega et al., 2019)-received WWTP effluents, which has actually

happened since the end of 2015 (Nega et al., 2019). The distinct signatures at these individual sites are thus a promising starting point for incorporating information on DOM composition in water-quality assessment and monitoring. Notably, DOM optical indicexes are would be highly cost-effective to apply and yield information that is not easily obtained by classic approaches. Robustness of such assessments would further increase when they are based on continuous time series. This could strengthen the implementation of current legal frameworks such as the EU Water Framework Directive aiming at an integrative water-quality assessment, including of urban water bodies.

5. Conclusion

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The composition of DOM collected analyzed in a suite of contrasting water bodies in a large metropolitan area, the city of Berlin in Germany, is diverse, varying widely in molecular size and other features related to the degree of allochthonous inputs and conveying a distinct urban character. DOM features clearly differentiated water body types, from lakes with highly abundant autochthonous DOM to streams with more allochthonous DOM. Seasonal variation of DOM was prevalent in all water body types but likely to be driven not only by phenology but also by distinctly urban driversinfluences such as nutrient supply, WWTP-inputs effluents, reduced leaf litter input or flashy runoff resulting from sealed surfaces. Nutrient supply, the percentage of green space and concentrations of trace organic pollutants (as proxies for point source influences) were identified as major drivers of DOM composition. Notably, easily measured In particular, simple optical data on DOM measurements of DOM characteristics were sufficient to detect WWTP effluents, a result that was confirmed corroborated by our data on TrOCTrOCs. This suggests that optical analysis of DOM analyses could be a useful starting point in approach to complement current water-quality assessments and monitoring. Optical Such analyses of DOM are fast, inexpensive and easily implemented, and could be complemented further supported by more sophisticated and, potentially automated analyses such as the massspectrometric quantification of TrOCs. DOM composition can inform about processes both within water bodies and in the terrestrial surroundings; therefore, water-quality assessments could benefit from integrating information on DOM composition. Robustness of the assessments approach would increase if the DOM assessments were based on time series or even continuous monitoring, for which knowledge and technology for which are readilyalready available. This could; this could; indeed and thus-strengthen current assessments as implemented in legal frameworks such as the EU Water Framework Directive, which aims at an integrative assessment of the "ecological status" of water bodies.

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Appendix A includes a table showing site coordinates land cover and special features

Table A1: Coordinates, land cover, origin and special features. Longitude is given in decimal degrees East and latitude in decimal degrees North.

Site ID	Site name	Water body type	<u>Latitude</u>	Longitude	Agri- culture (%)	Forest (%)	Urban pave- ment (%)	Urban green space (%)	Origin
<u>H1</u>	<u>Teltowkanal</u>	River	52.44239	13.32454	0	<u>0</u>	<u>60</u>	<u>30</u>	<u>Artificial</u>
<u>H2</u>	<u>Teltowkanal</u>	River	52.42642	13.52039	<u>0</u>	<u>0</u>	<u>100</u>	<u>0</u>	<u>Artificial</u>
<u>H3</u>	Wuhle	<u>Stream</u>	52.52562	13.57913	<u>50</u>	<u>0</u>	<u>50</u>	<u>0</u>	<u>Natural</u>
<u>H4</u>	Tegeler Fliess	Stream	52.63442	13.38013	<u>50</u>	<u>0</u>	<u>10</u>	<u>40</u>	<u>Natural</u>
<u>L1</u>	Biesdorfer See	<u>Lake</u>	52.50331	13.5497	<u>0</u>	<u>0</u>	<u>50</u>	<u>50</u>	<u>Artificial</u>
<u>L2</u>	<u>Obersee</u>	<u>Lake</u>	<u>52.54856</u>	13.48972	<u>0</u>	<u>0</u>	<u>50</u>	<u>50</u>	Artificial
<u>L3</u>	Ploetzensee	<u>Lake</u>	<u>52.5438</u>	13.33049	<u>0</u>	<u>0</u>	<u>0</u>	<u>100</u>	<u>Natural</u>
<u>L4</u>	Gross Glienicker	<u>Lake</u>	<u>52.46417</u>	13.11489	<u>0</u>	<u>10</u>	<u>0</u>	<u>90</u>	<u>Natural</u>
<u>L5</u>	<u>Havel</u>	<u>Lake</u>	<u>52.4431</u>	13.14453	<u>0</u>	<u>0</u>	<u>100</u>	<u>0</u>	<u>Natural</u>
<u>L6</u>	Schlachtensee	<u>Lake</u>	52.44066	<u>13.21183</u>	<u>0</u>	<u>60</u>	<u>30</u>	<u>10</u>	<u>Natural</u>
<u>L7</u>	Müggelsee	<u>Lake</u>	<u>52.43837</u>	13.6451	<u>0</u>	<u>70</u>	<u>30</u>	<u>0</u>	<u>Natural</u>
<u>P1</u>	<u>Hoheheideteich</u>	Pond	52.57694	13.16428	<u>0</u>	<u>100</u>	<u>0</u>	<u>0</u>	<u>Natural</u>
<u>P2</u>	Hamburger Teich	<u>Pond</u>	<u>52.56738</u>	13.44549	<u>0</u>	<u>0</u>	<u>30</u>	<u>70</u>	Artificial
<u>P3</u>	Ruhwaldteich	<u>Pond</u>	<u>52.52573</u>	13.25998	<u>0</u>	<u>0</u>	<u>50</u>	<u>50</u>	Artificial
<u>P4</u>	Kienhorstbecken	<u>Pond</u>	52.57724	13.34556	<u>0</u>	<u>0</u>	<u>0</u>	<u>100</u>	<u>Artificial</u>
<u>P5</u>	Mittelfeldteich	<u>Pond</u>	<u>52.61208</u>	13.23045	<u>0</u>	<u>100</u>	<u>0</u>	0	Artificial
<u>P6</u>	<u>Neurandteich</u>	Pond	52.63883	13.27377	<u>0</u>	<u>0</u>	<u>65</u>	<u>35</u>	<u>Artificial</u>
<u>P7</u>	<u>Möwensee</u>	<u>Pond</u>	52.55282	13.33545	<u>0</u>	<u>0</u>	<u>30</u>	<u>70</u>	<u>Artificial</u>

<u>R1</u>	<u>Müggelspree</u>	River	<u>52.42985</u>	13.68912	<u>0</u>	<u>0</u>	<u>100</u>	<u>0</u>	<u>Natural</u>
<u>R2</u>	<u>Landwehrkanal</u>	River	<u>52.51935</u>	13.31959	<u>0</u>	<u>0</u>	<u>80</u>	<u>20</u>	<u>Artificial</u>
<u>R3</u>	<u>Spree</u>	River	52.53613	13.21622	<u>0</u>	<u>0</u>	<u>100</u>	<u>0</u>	<u>Natural</u>
<u>R4</u>	Kuhlake	River	52.57817	13.16509	<u>0</u>	<u>100</u>	<u>0</u>	<u>0</u>	<u>Natural</u>
<u>R5</u>	Neukölln Canal	River	<u>52.48936</u>	13.43949	<u>0</u>	<u>0</u>	<u>30</u>	<u>70</u>	Artificial
<u>R6</u>	Spree	River	<u>52.47137</u>	13.49683	<u>0</u>	<u>0</u>	<u>100</u>	<u>0</u>	<u>Natural</u>
<u>R7</u>	<u>Panke</u>	River	52.5369	13.36759	<u>0</u>	<u>0</u>	<u>60</u>	<u>40</u>	<u>Natural</u>
<u>S1</u>	Zingergraben	<u>Stream</u>	<u>52.58209</u>	13.38594	<u>0</u>	<u>0</u>	<u>95</u>	<u>5</u>	<u>Artificial</u>
<u>S2</u>	<u>Schwarzer</u> <u>Graben</u>	Stream	52.56488	13.34918	<u>0</u>	<u>0</u>	<u>50</u>	<u>50</u>	<u>Natural</u>
<u>S3</u>	Graben 1 Buch	<u>Stream</u>	52.62384	13.46883	<u>0</u>	<u>100</u>	<u>0</u>	<u>0</u>	<u>Artificial</u>
<u>\$4</u>	Graben 73 Buchholz	<u>Stream</u>	52.62881	<u>13.45315</u>	100	<u>0</u>	<u>0</u>	<u>0</u>	<u>Artificial</u>
<u>S5</u>	<u>Erpe</u>	Stream	<u>52.45888</u>	13.61245	<u>0</u>	<u>50</u>	<u>50</u>	<u>0</u>	<u>Natural</u>
<u>S6</u>	Koppelgraben	Stream	52.62065	13.41089	<u>50</u>	<u>0</u>	<u>30</u>	<u>20</u>	<u>Unknown</u>
<u>87</u>	Plumpengraben	Stream	<u>52.41513</u>	13.5628	<u>0</u>	<u>0</u>	<u>100</u>	<u>0</u>	<u>Natural</u>

Appendix B includes that complement the physico-chemical and dissolved organic composition information.

Table A1B1: Physico-chemical characteristics (mean \pm SD and % variance explained) of four contrasting types of water bodies in the city of Berlin. Means and standard deviations were computed across all seasons and sites. The percentages of variance explained (% Var) refer to the effect of season within each water body type, calculated by type-II ANOVA (aka variance component analysis), with season treated as a random factor. F-values refer to results of repeated-measures ANOVAs testing for differences among water body types (*** p<0.001, * p<0.05, ns = not significant).

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Water body	Temper	ature	DO	C	TP		NH ₄	+	NO ₃	-	Chloro a	phyll
	(0.6%)	%	(mg/	%		%		%		%		%
type	(°C)	Var	L)	Var	(mg/L)	Var	(mg/L)	Var	(mg/L)	Var	(μg/L)	Var
Lakes	14.6 ± 6.9	94	7.5 ± 2.7	23	0.05 ± 0.05	34	0.07 ± 0.07	26	0.22 ± 0.36	48	6.2 ± 14.2	61
Ponds	13.7 ± 5.3	94	10.3 ± 3.0	32	0.09 ± 0.07	30	0.27 ± 0.67	30	0.03 ± 0.06	52	7.3 ± 7.7	55
Rivers	15.2 ± 6.2	92	8.0 ± 1.6	13	0.10 ± 0.08	43	0.15 ± 0.14	68	1.12 ± 1.66	46	2.1 ±2.9	51
Streams	11.3 ± 4.9	87	11.7 ± 5.5	42	0.26 ± 0.31	21	0.36 ± 0.65	76	0.91 ± 1.53	43	5.3 ± 9.7	53
$F_{\text{water body}}$	9.4***		3.8*		2.8 ^{ns}		1.3 ^{ns}		2.5 ^{ns}		1.0 ^{ns}	

Variable	Definition Description
SUVA ₂₅₄	Proxy for DOM aromaticity (Weishaar et al., 2003)
E2:E3	Ratio of absorbance at 250 and 365 nm, as an (inverse) indicator of molecular size (Peuravuori and Pihlaja, 1997) (Chen et al., 1977)
E4:E6	Indicator of humification (Chen et al., 1977)
Slope shortSR	Short wavelength slope within the wavelength region of 275 295 nm as an inverse correlate with molecular weight and aromaticity (Helms et al., 2008)Ratio of slopes (SR) computed from short and long wavelength regions as another negative correlate with DOM molecular weight (Loiselle et al., 2009)
<u>SRFI</u>	Ratio of slopes (SR) computed from short and long wavelength regions as another negative correlate with DOM molecular weight (Loiselle et al., 2009) Fluorescence index (FI) Ratio of the fluorescence intensities at the emissions 470 and 520 (obtained at excitation wavelength of 370nm). Indicator of DOM derived from terrestrial plants (FI around 1.2) or from microbes or algae (FI around 1.4) (Fellman et al., 2010; Cory and Mcknight, 2005; Cory et al., 2010; Jaffé et al., 2008)
FIX- HIX	Fluorescence index (FIX) as the ratio of fluorescence intensities at the emission wavelengths of 470 and 520 nm (obtained at excitation wavelength of 370 nm), which has proved useful to distinguish the relative contributions of terrestrial (FIX~1.4) and aquatic (FIX~1.9) sources of DOM (Meknight et al., 2001) Humification index (HIX) as a proxy for humic substances (Ohno, 2002)
ΗΙΧ β/α	Humification index (HIX) as a proxy for humic substances (Ohno, 2002) Freshness index β/α (Wilson and Xenopoulos, 2009), which indicates the relative importance of recently produced DOM (Parlanti et al., 2000)
β/α	Freshness index β/α (Wilson and Xenopoulos, 2009), which indicates the relative importance of recently produced DOM (Parlanti et al., 2000)

Table <u>A3B3</u>: Designation, excitation (Ex) and emission (Em) wavelengths of PARAFAC components, and the number of studies with matching components reported in OpenFluor (checked on the 28th March 2022) (Murphy et al., 2014).

	PARAFAC component	Ex	Em	OpenFluor reference matches (0.95)	Explanation and selected references
•	C1	250	446	128	Humic-like, peak A (Coble, 1996); humic-like and recalcitrant (C1) (Hansen et al., 2016)
	C2	250	500	70 22	Terrestial humic-like in waste water treatment impacted water, (G1) (Murphy et al., 2011); ubiquitous and recalcitrant humic (C2) (Chen et al., 2017)
	C3	306	408	20 13	Humic-like, peak M (Coble, 1996); humic-like (C3) (Stedmon and Markager, 2005)
	C4	256	444	8 9	Terrestrial humic-like, suggested as photo-refractory (C2) (Yamashita et al., 2010); terrestrial humic-like (C3) (Williams et al., 2013)
	C5	250	382	12 2	Anthropogenic, microbial humic-like (C6) (Williams et al., 2016)
	C6	294	352	33 13	Similar to tryptophan (C3) (Catalán et al., 2015); protein- like, linked to autochthonous production (C3) (Amaral et al., 2016)
	C7	320	396	8	Humic like, peak M (Coble, 1996); microbially transformed autochthonous DOM (C2) (Osburn et al., 2011)
_	C7 <mark>8</mark>	276	326	68 15	Protein-like, peak B (Coble, 1996); waste water treatment protein (C2) (Teymouri, 2007)

Table A4: Absorbance B4: Variables of absorbance and fluorescence analysis variables analyses (mean ± SD and % variance explained) in contrasting types of urban surface water bodywaters. Means and standard deviations were computed across all seasons and sites. The percentages of variance explained (% Var) refer to the effect of season within each water body type, calculated by a type-II ANOVA (aka variance component analysis), with season treated as a random factor. F-values refer to results of repeated-measures ANOVA testing for differences among water body types (***p<0.001, **p<0.01, *p<0.05, ns = not significant). Abbreviations explained in Table A2B2.

Water	SUVA	254	E2:E	3	E4:I	Ε6	Slope sh	ort	SR		FI	E	HIX	[β/ο	ı
body type		% Var		% Var		% Var	-	% Var		% Var		% Var		% Var		% Var
Lakes	1.55 ± 0.39	20	8.99 ± 2.14	6	3.02± 1.34	67	-0.024 ± 0.004	8	1.38 ± 0.27	12	1.61 ± 0.08	69	0.77 ± 0.08	9	0.86 ± 0.09	12
Ponds	2.14 ± 0.51	37	6.65 ± 1.24	14	3.12± 0.74	46	0.019 ± 0.003	13	1.20 ± 0.19	29	1.52 ± 0.06	61	0.83 ± 0.04	47	0.70 ± 0.04	35
Rivers	2.25 ± 0.15	55	7.04 ± 0.98	12	4.40± 14.27	80	-0.019 ± 0.002	22	0.017 ± 0.002	76	0.68 ±0.11	10	0.85 ± 0.03	24	0.79 ± 0.09	19
Streams	2.50 ±0.52	65	6.328 ± 0.876	54	3.53± 2.31	75	$\frac{-0.017 \pm}{0.001}$	57	0.97 ± 0.13	22	1.63 ± 0.14	11	0.86 ± 0.05	21	0.73 ± 0.10	24
F _{water}	11.8***		5.8**		2.6 ns		8.0***	_	9.2***		3.5*		4.9**		5.5**	

Table A5B5: PARAFAC components results (mean ± SD and % variance explained) in contrasting types of urban surface water bodywaters. Means and standard deviations were computed across all seasons and sites. The percentages of variance explained (% Var) refer to the effect of season within each water body type, calculated by a type-II ANOVA (aka variance component analysis), with season treated as a random factor. F-values refer to results of repeated-measures ANOVA testing for differences among water body types (***p<0.001, **p<0.01, *p<0.05, ns = not significant).

5	6	n
J	v	U

Water	C1		C2		C3		C4		C5		C6		C7	<u>.</u>	C8	
body type	-	% Var	-	% Var	-	% Var	-	% Var	-	% Var	-	% Var	-	% Var	-	% Var
Lakes	0.17 ± 0.09	20	0.12 ± 0.06	14	0.17 ± 0.08	27	0.23 ± 0.10	11	0.23 ± 0.16	10	0.20 ± 0.10	15	0.09 ± 0.05	27	0.17 ± 0.10	9
Ponds	$\frac{0.27 \pm}{0.07}$	60	0.22 ± 0.09	65	0.34 ± 0.17	57	0.32 ± 0.15	48	0.36 ± 0.29	37	0.16 ± 0.07	5 4	0.08 ± 0.05	67	0.22 ± 0.11	48
Rivers	0.59 ± 0.41	8	0.34 ± 0.17	12	0.28 ± 0.10	56	0.37 ± 0.09	38	0.42 ± 0.19	41	0.35 ± 0.25	12	0.37 ± 0.33	7	$\frac{0.23 \pm}{0.12}$	15
Streams	0.88 ± 0.55	18	0.56 ± 0.29	32	0.55 ± 0.48	65	0.67 ± 0.47	44	0.82 ± 0.62	64	0.42 ± 0.32	2 4	0.46 ± 0.43	16	0.33 ± 0.17	47
Fwater body	6.7***	-	10.5***	-	7.4***	_	6.8***	-	8.0***	-	2.7 ^{ns}	-	3.8*	-	2.8 ^{n.s.}	_

Water	<u>C1</u>		<u>C2</u>		<u>C3</u>		<u>C4</u>		<u>C5</u>		<u>C6</u>		<u>C7</u>	
<u>body</u> <u>type</u>	-	<u>%</u> <u>Var</u>	-	<u>%</u> <u>Var</u>	_	<u>%</u> <u>Var</u>	-	<u>%</u> <u>Var</u>	-	<u>%</u> <u>Var</u>	-	<u>%</u> <u>Var</u>	-	<u>%</u> <u>Var</u>
<u>Lakes</u>	0.17 ± 0.10	<u>17</u>	0.13 ± 0.06	<u>14</u>	0.24 ± 0.12	<u>20</u>	0.28 ± 0.12	<u>14</u>	0.31 ± 0.18	<u>11</u>	0.19 ± 0.10	<u>15</u>	0.18 ± 0.11	9
<u>Ponds</u>	0.24 ± 0.14	<u>73</u>	0.24 ± 0.11	<u>80</u>	0.40 ± 0.22	<u>63</u>	0.46 ± 0.21	<u>63</u>	0.52 ± 0.38	<u>73</u>	0.16 ± 0.11	<u>65</u>	0.25 ± 0.14	<u>45</u>
Rivers	0.51 ± 0.34	<u>8</u>	0.34 ± 0.17	<u>12</u>	0.66 ± 0.37	<u>10</u>	0.44 ± 0.12	<u>32</u>	0.61 ± 0.26	<u>30</u>	0.32 ± 0.22	<u>14</u>	0.25 ± 0.14	<u>14</u>
Streams	0.76 ± 0.47	<u>16</u>	0.57 ± 0.30	<u>53</u>	1.00 ± 0.58	<u>30</u>	0.85 ± 0.63	<u>64</u>	1.08 ± 0.73	<u>16</u>	0.37 ± 0.29	<u>25</u>	0.35 ± 0.18	<u>45</u>
Fwater body	6.3**		10.4***		7.6***		7.2***		8.4***		<u>2.2n.s</u>		2.6n.s.	

Table A6: SizeB6: Results of size exclusion chromatography results (mean ± SD and % variance explained) inof samples from contrasting types of urban surface water bodywaters. Means and standard deviations were computed across all seasons and sites. The percentages of variance explained (% Var) refer to the effect of season within each water body type, calculated by a type-II ANOVA (aka variance component analysis), with season treated as a random factor. F-values refer to results of repeated-measures ANOVA testing for differences among water body types (***p<0.001, **p<0.01, *p<0.05, ns = not significant). HS, humic-like substances; HMWS, high-molecular weight non-humic substances; and LMWS, low-molecular weight substances.

Water	HMSW	7	HMSW		HS		HS	LMW		Val
body type	(mg C/L)	% Var	(mg N/L)	% Va r	(mg C/L)	% Var	(mg N/L)	% Var	(mg C/L)	
Lakes	0.96 ± 0.74	27	0.11 ± 0.07	9	4.14 ± 1.49	19	0.25 ± 0.11	12	0.83 ± 0.26	18
Ponds	1.32 ± 0.60	41	0.16 ± 0.06	36	6.29 ± 2.42	26	0.33 ± 0.13	17	1.19 ± 0.46	45
Rivers	0.59 ± 0.20	60	0.09 ± 0.03	24	5.21 ± 0.97	33	0.31 ± 0.09	31	1.10 ± 0.41	22
Streams	0.73 ± 0.45	52	0.10 ± 0.05	41	7.21 ± 3.75	25	0.41 ± 0.27	9	1.48 ± 0.62	39
Fwater body	4.2*		2.9 ^{ns}		2.9 ^{ns}		1.3 ^{ns.}		3.7*	

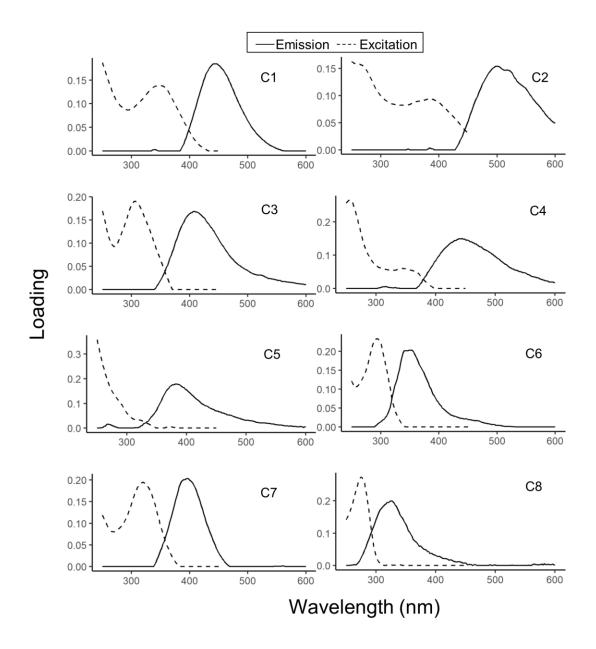
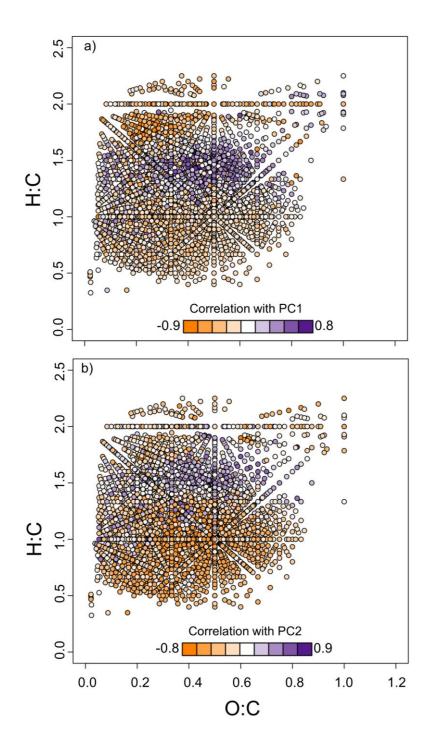
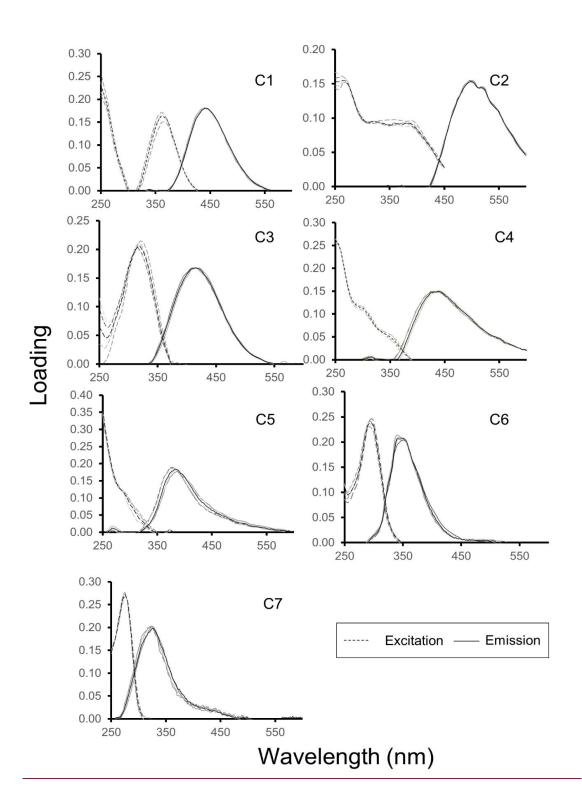


Figure A1 PARAFAC components emission and excitation wavelength.





<u>Figure B1 Emission and excitation wavelengths of PARAFAC components. Solid lines represent emission spectra, dashed lines excitation spectra. Lines in different shades of grey refer to models using different sample sub-sets of a split-half validation analysis.</u>

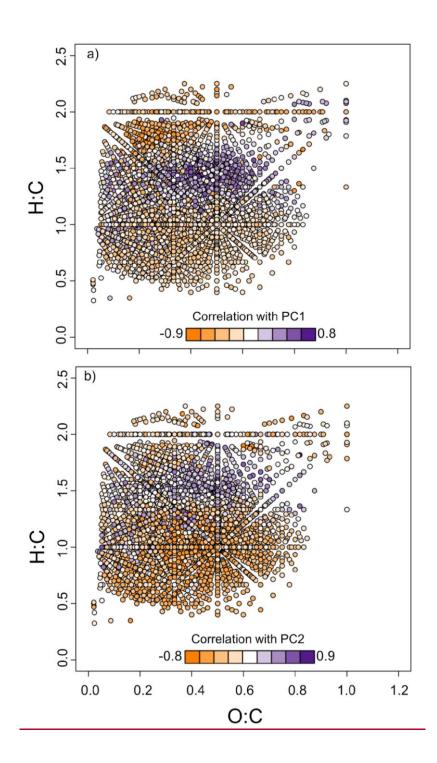


Figure A2B2 Van Krevelen plots showing all molecules (sum formulas) identified by FT-ICR-MS analysis of DOM samples collected at 32 urban sites over three seasons (summer, autumn and winter). ColorColour indicates molecule-specific Spearman correlation coefficients of the relative intensities of each compound with the first (a) and second (b) axis of the PCA shown in Figures 2 and 3. The data points were plotted in random order to avoid bias resulting from identical O:C and H:C ratios for many sum formulas.

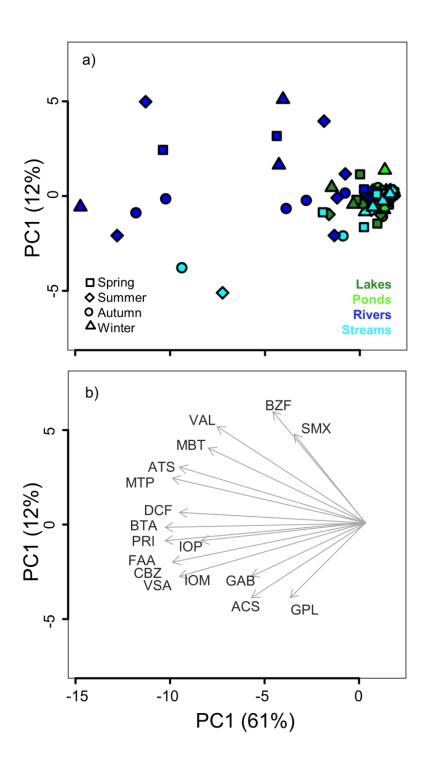
Appendix BC includes tables and figures that complement Trace Organic Compounds analysis

Table B1C1: Trace organic compounds (TrOCs) analyzed in samples collected in urban surface waters. LLoQ = Limit of Quantification. Frequency refers to the number of occasions where concentrations exceeded the LLoQ.

Acronym	LLoQ (µg/L)	Frequency	Name	Description
ACS	0.1	72	Acesulfame	Sweetener
ATS	0.05	42	Amidrotrizoic	Radiocontrast agent
BTA	0.1	68	Benzotriazole	Corrosion inhibitor
BZF	0.1	6	Benzafibrate	Lipid-lowering agent
CBZ	0.05	44	Carbamazepine	Anticonvulsant
DCF	0.05	30	Diclofenac	Analgesic/anti-inflammatory agent
FAA	0.1	46	4-formylamin metabolite of metamizol	Analgesic
GAB	0.1	62	Gabapentin	Drug for epilepsy treatment/pain killer
GPL	0.05	39	Gabapentin-lactam	Derivate of gabapentin
IOM	0.1	40	Iomeprol	Radiocontrast agent
IOP	0.01	52	Iopromide	Radiocontrast agent
MBT	0.1	63	Methylbenzotriazole	Corrosion inhibitor
MTP	0.1	31	Metoprolol	Beta blocker
PRI	0.05	31	Primidone	Anticonvulsant
SMX	0.1	2	Sulfamethoxazole	Antibiotic
VAL	0.1	30	Valsartan	At1-receptor antagonist
VLX	0.1	3	Venlafaxine	Antidepressant
VSA	0.1	62	Valsartan acid	Antihypertensive agent

Table <u>B2C2</u>: Mean concentrations and standard deviations of Trace Organic Compound (TrOC) per water body type. See Table <u>B4C1</u> for full names. BZF, SMX and VLX were always below the limit of quantification (LLoQ) and are hence omitted from the table.

	TrOC concentration (μg/L)			
Acronym	Lakes	Ponds	Rivers	Streams
ACS	0.23 ± 0.17	0.15 ± 0.16	0.28 ± 0.17	0.78 ± 1.35
ATS	0.08 ± 0.12	<lloq< td=""><td>0.74 ± 1.12</td><td>0.43 ± 0.88</td></lloq<>	0.74 ± 1.12	0.43 ± 0.88
BTA	0.34 ± 0.51	0.38 ± 0.91	2.37 ± 3.31	2.16 ± 3.61
CBZ	0.07 ± 0.08	<lloq< td=""><td>0.37 ± 0.48</td><td>0.41 ± 0.66</td></lloq<>	0.37 ± 0.48	0.41 ± 0.66
DCF	<lloq< td=""><td><lloq< td=""><td>0.97 ± 1.38</td><td>0.88 ± 2.14</td></lloq<></td></lloq<>	<lloq< td=""><td>0.97 ± 1.38</td><td>0.88 ± 2.14</td></lloq<>	0.97 ± 1.38	0.88 ± 2.14
FAA	0.15 ± 0.23	<lloq< td=""><td>1.25 ± 1.66</td><td>2.10 ± 4.25</td></lloq<>	1.25 ± 1.66	2.10 ± 4.25
GAB	0.27 ± 0.36	<lloq< td=""><td>0.42 ± 0.43</td><td>0.74 ± 1.12</td></lloq<>	0.42 ± 0.43	0.74 ± 1.12
GPL	0.10 ± 0.17	<lloq< td=""><td>0.19 ± 0.42</td><td>0.13 ± 0.18</td></lloq<>	0.19 ± 0.42	0.13 ± 0.18
IOM	0.18 ± 0.28	<lloq< td=""><td>1.18 ± 2.36</td><td>1.44 ± 2.91</td></lloq<>	1.18 ± 2.36	1.44 ± 2.91
IOP	0.09 ± 0.19	0.01 ± 0.02	0.27 ± 0.32	1.46 ± 3.79
MBT	0.27 ± 0.39	0.11 ± 0.24	0.91 ± 1.01	0.69 ± 1.27
MTP	<lloq< td=""><td><lloq< td=""><td>0.47 ± 0.58</td><td>0.63 ± 1.55</td></lloq<></td></lloq<>	<lloq< td=""><td>0.47 ± 0.58</td><td>0.63 ± 1.55</td></lloq<>	0.47 ± 0.58	0.63 ± 1.55
PRI	0.03 ± 0.02	<lloq< td=""><td>0.16 ± 0.22</td><td>0.26 ± 0.52</td></lloq<>	0.16 ± 0.22	0.26 ± 0.52
VAL	<lloq< td=""><td><lloq< td=""><td>0.39 ± 0.44</td><td>0.97 ± 3.65</td></lloq<></td></lloq<>	<lloq< td=""><td>0.39 ± 0.44</td><td>0.97 ± 3.65</td></lloq<>	0.39 ± 0.44	0.97 ± 3.65
VSA	0.70 ± 1.02	<lloq< td=""><td>3.22 ± 3.84</td><td>3.33 ± 5.72</td></lloq<>	3.22 ± 3.84	3.33 ± 5.72



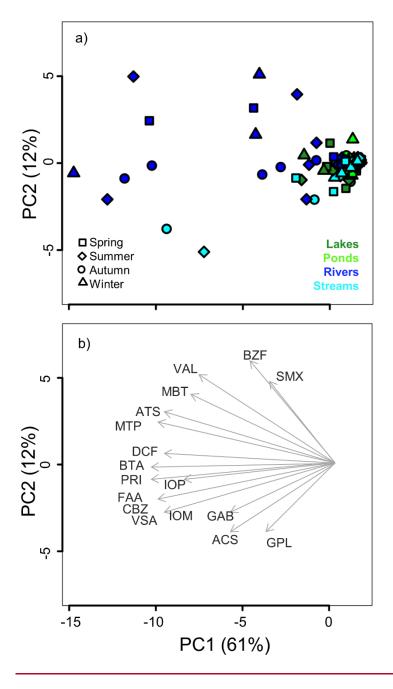
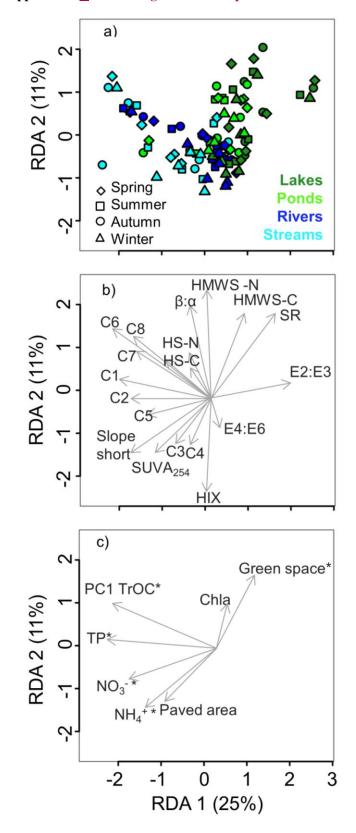


Figure <u>B1C1</u> Principal Component Analysis (PCA) of <u>32</u> urban <u>sampling</u> sites <u>in the city of Berlin over four seasons</u> (a) and Trace Organic Compounds (TrOCs) (b). <u>ExtremeSite S5 had extreme PC1 and PC2 scores; the</u> site <u>S5</u> was included in the analysis but is not presented in the biplot to better visualize variability among the other sites. Abbreviations of the TrOCs (B) are explained in Table <u>B1C1</u>.

Appendix CD includes figures that complement the statistical RDA analysis



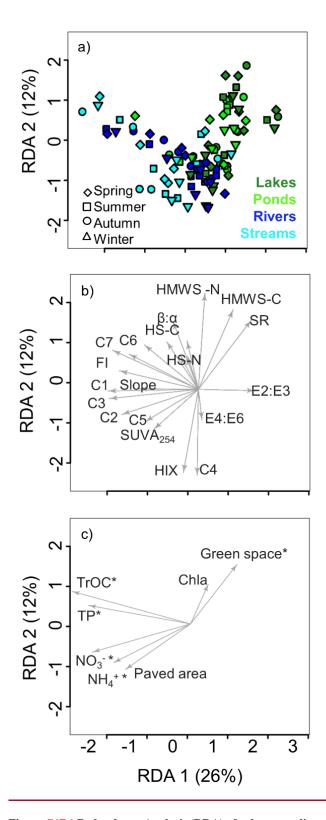


Figure C1D1 Redundancy Analysis (RDA) of urban sampling sites (a) visited 4 times over one year, the DOM characteristics included in the analysis (b) and the predictor variables (c), the last marked by an asterisk (*) when significant. DOM characteristics include (i) absorbance and fluorescence indexes (E2:E3, molecular size, E4:E6, indicator of humification, SR, slope ratio, β : α , freshness index, slope short, SUVA₂₅₄ and HIX, humification index), (ii) PARAFAC components (C1 to

Supplement. The supplement related to this article is available online. Appendix E Includes precipitation and flow in the city of Berlin during the study and the relation of Iron and absorbance at 420 relative to DOC.

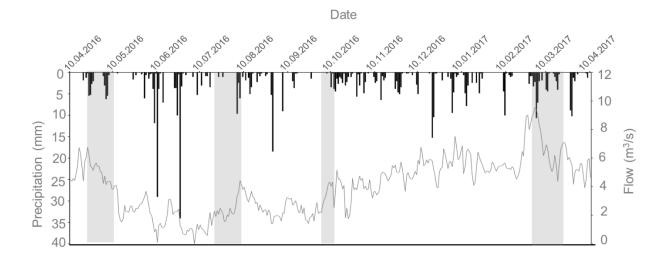


Figure E1 Precipitation and flow at a site within the city of Berlin during the study period, with the grey boxes indicating the four sampling periods.

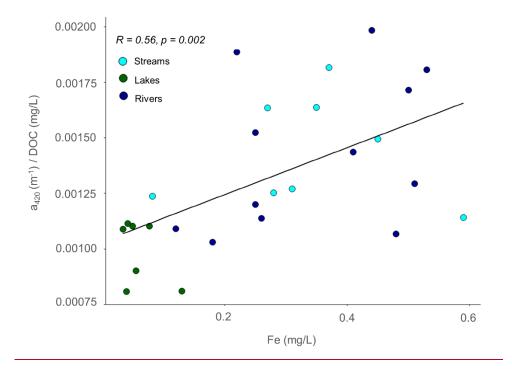


Figure E2 Relationship between iron (Fe) and the absorbance at 420nm relative to DOC(a420/DOC)

Author contributions. All authors contributed to designing the study. CR and SH collected the data. CR did the optical analysis and the PARAFAC modelling, GS carried out the FT-ICR-MS analysis. CR and GS conducted the statistical analysis. CR led the manuscript writing, jointly with GS. All authors discussed results and edited the manuscript.

Competing interests. The authors declare that they not to have no a conflict of interest.

Data availability. The data will be <u>made</u> available at a <u>suitable repository at</u> ttps://www.pangaea.de/https://www.re3data.org/.

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