Stable isotope ratios in seawater nitrate reflect the influence of Pacific water along the Northwest Atlantic margin

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Abstract. The flow of Pacific water to the North Atlantic exerts a globally significant control on nutrient balances between the two ocean basins, and strongly influences biological productivity in the Northwest Atlantic. Nutrient ratios of nitrate (NO_3^-) versus phosphate (PO_4^{3-}) have previously been used to complement salinity characteristics in tracing the distribution of Pacific water in the North Atlantic. We expand on this premise and demonstrate that the fraction of Pacific water as determined by $NO_3^-:PO_4^{3-}$ ratios can be quantitatively predicted from the isotopic composition of sub-euphotic nitrate in the Northwest Atlantic. Our linear model thus provides a critically important framework for interpreting $\delta^{15}N$ signatures incorporated into both modern marine biomass, as well as organic material in historical and paleoceanographic archives along the Northwest Atlantic margin.

1 Introduction

Pacific water from the Bering Strait constitutes a major fraction of the polar outflow to the Northwest Atlantic Ocean (McLaughlin et al., 1996; Jones et al., 2003; Aksenov et al., 2010). Besides redistributing heat and freshwater (Tang et al., 2004; Carmack et al., 2016), it also plays a critical role in the transport of nutrients between the two ocean basins (Tremblay et al., 2015; Lehmann et al., 2019). Pacific water has relatively high nutrient concentrations (Macdonald et al. 2010). These nutrients support high productivity on the Bering and Chukchi shelves (Arrigo and van Dijken, 2011), which in turn fuels high rates of sedimentary denitrification both in the shelf regions and along the Bering continental slope (Devol et al., 1997; Lehmann et al., 2005, 2007; Chang and Devol, 2009; Granger et al., 2011; Brown et al., 2015). The resulting excess in silicate ($Si(OH)_4$) and phosphate (PO_4^{3-}) (relative to nitrate (NO_3^-)) is a significant source of these nutrients to the Atlantic (Torres-

Valdés et al., 2013). In particular, the excess PO_4^{3-} supports N_2-fixation in the Atlantic, thereby helping to balance the global oceanic nitrogen budget (Yamamoto-Kawai et al., 2006).

Pacific-derived nutrients also influence biological productivity along the Northwest Atlantic shelf complex. The NO_3^- deficit in Pacific water sets an upper limit on productivity, which otherwise would be higher in the presence of more NO_3^--enriched Atlantic water (Harrison and Li, 2008). The $Si(OH)_4$ and PO_4^{3-} excess influences plankton community composition (Harrison et al., 2013; Fragoso et al., 2017). Interannual and decadal-scale variability in the circulation of Pacific water into the Northwest Atlantic may help to explain recent observed changes in the magnitude and composition of primary productivity with potential bottom-up effects on ecosystem functioning (Drinkwater et al., 2003; Greene et al., 2013; Townsend et al., 2015).

Given its importance to downstream circulation, nutrient budgets, and productivity, it is useful to track the distribution of Pacific water using chemical tracers. Jones et al. (1998) characterized $NO_3^-:PO_4^{3-}$ relationships for "pure" Pacific and Atlantic endmember waters. They further demonstrated that the concentrations of NO_3^- and PO_4^{3-} in a water sample relative to the endmember relationships may be used to quantify the contribution of Pacific water (i.e., "fraction Pacific water", or fPW). With this approach, the spatial and depth distributions of fPW were used to map the flow of Pacific water through the Arctic and North Atlantic Oceans (Jones et al., 1998, 2003). The same approach has also been used to deconvolute fluxes of freshwater originating from Pacific water from that of sea ice meltwater and meteoric water (Yamamoto-Kawai et al., 2008; Sutherland et al., 2009; Azetsu-Scott et al., 2012; Benetti et al., 2016). In another study, time-series nutrient data were used to track fPW and thereby infer changes in circulation patterns over a thirty-year period in Disko Bay, Greenland (Hanson et al., 2012). The use of $NO_3^-:PO_4^{3-}$ as a proxy for Pacific water has, however, important limitations. For example, the approach requires an assumption of constant stoichiometry associated with the uptake and recycling of nutrients, which may not hold in all regions (Michel et al., 2002; Mills et al., 2015). Moreover, sensitivity to NO_3^- source and sink processes such as N_2 fixation and denitrification may lead to an under-, or overestimation of fPW, respectively. Finally, seawater $NO_3^-:PO_4^{3-}$ ratios are not preserved in organisms or sedimentary archives, thus limiting their use in establishing changing baselines in an ecological or paleoceanographic context.

The nitrogen ($^{15}N/^{14}N$) and oxygen ($^{18}O/^{16}O$) isotope ratios in NO_3^- (expressed as $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$) represent a complementary tool to trace the distribution and modification of Pacific water, possibly addressing shortcomings related to the use of stoichiometric nutrient tracers. Coupled N and O isotope ratios provide insights into the internal cycling of NO_3^-, as well as input and removal processes. The preferential reaction of the lighter ^{14}N and ^{16}O during both phytoplankton uptake and denitrification results in an enrichment of $\delta^{15}N$ and $\delta^{18}O$ of the dissolved NO_3^- pool with a ratio of ~1 (Casciotti et al., 2002; Granger et al., 2004, 2008; Sigman et al., 2005). Conversely, the recycling or regeneration of NO_3^- via nitrification (the oxidation of ammonium (NH_4^+) to nitrite (NO_2^-) and NO_3^-) leads to a decoupling of the N and O isotopic signature of NO_3^- (Sigman et al., 2005; Lehmann et al., 2005; Granger and Wankel, 2016). The $\delta^{15}N$ of newly nitrified NO_3^- depends on the N isotopic composition of its source substrate and hence mirrors the isotopic signature of the organic matter exported from the

surface. In contrast, the $\delta^{18}O$ of newly nitrified NO_3^- remains independent from its N source and approaches the $\delta^{18}O$ signature of seawater ($\delta^{18}O_{H2O}$ +1.1‰; Casciotti et al., 2008; Sigman et al., 2009; Buchwald et al., 2012). The resulting NO_3^- isotope fingerprints of particular water masses have led to their increasing use as unique water mass tracers (e.g., Granger et al. 2018, Lehmann et al. 2018)

70   The goal of this paper is to establish the use of $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$ as a new chemical oceanographic tracer for tracking the distribution of Pacific water to the Northwest Atlantic. We present new data from Baffin Bay, Davis Strait, and the Labrador Sea (Fig. 1), highlighting differences in NO_3^- isotopic ratios among the different water masses found in those regions. We evaluate the preservation of $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$ signatures during southward advection of Pacific water from the Arctic Archipelago to the Labrador Shelf and present a linear relationship between $\delta^{15}N_{NO3}$ and fPW for the Northwest Atlantic

75   margin. Lastly, we entertain implications of our findings for regional isotope ecological and paleoceanographic studies.

2 Materials and Methods

2.1 Sample collection and nutrient measurements

Seawater samples were collected opportunistically during four different expeditions that sampled 25 stations along the NW

80   Atlantic margin from the mid-Labrador Shelf to northern Baffin Bay between the years 2005 and 2016 (Fig. 1). New and previously published data are presented. New data were collected during 1) expedition MSM45 of the *Maria S. Merian* in August 2015 and 2) an ArcticNet expedition (AMD-2016-002a) of the Canadian Coast Guard Ship (CCGS) *Amundsen* in July through September 2016. Previously published data are from 3) expedition HUD-2005-016 of the CCGS *Hudson* in June 2005; 4) and a GEOTRACES (GN02) expedition aboard the CCGS *Amundsen* in July/August 2015. Stations associated with each

85   expedition are indicated in the Fig. 1 stations legend. Sample collection and analytical protocols for the MSM45 and AMD-2016-002a expeditions are given below. Protocols for the HUD-2005-016 expedition are provided in Sherwood et al. (2011) and for the GEOTRACES expedition in Lehmann et al. (2019). Samples were collected under ice-free conditions during all expeditions. Station and bottle data are provided as a supplementary data file.

90   Samples from the MSM45 and AMD-2016-002a expeditions were collected with a rosette water sampler holding 24 x 10L Niskin bottles mounted with a conductivity-temperature-depth (CTD) profiler equipped with sensors for dissolved oxygen and fluorescence. During the MSM45 expedition, samples for nutrient and NO_3^- isotope analysis were collected from the Niskin bottles into separate, triple-rinsed, 60 ml high-density polyethylene (HDPE) bottles with pre-filtration through 0.45 μm
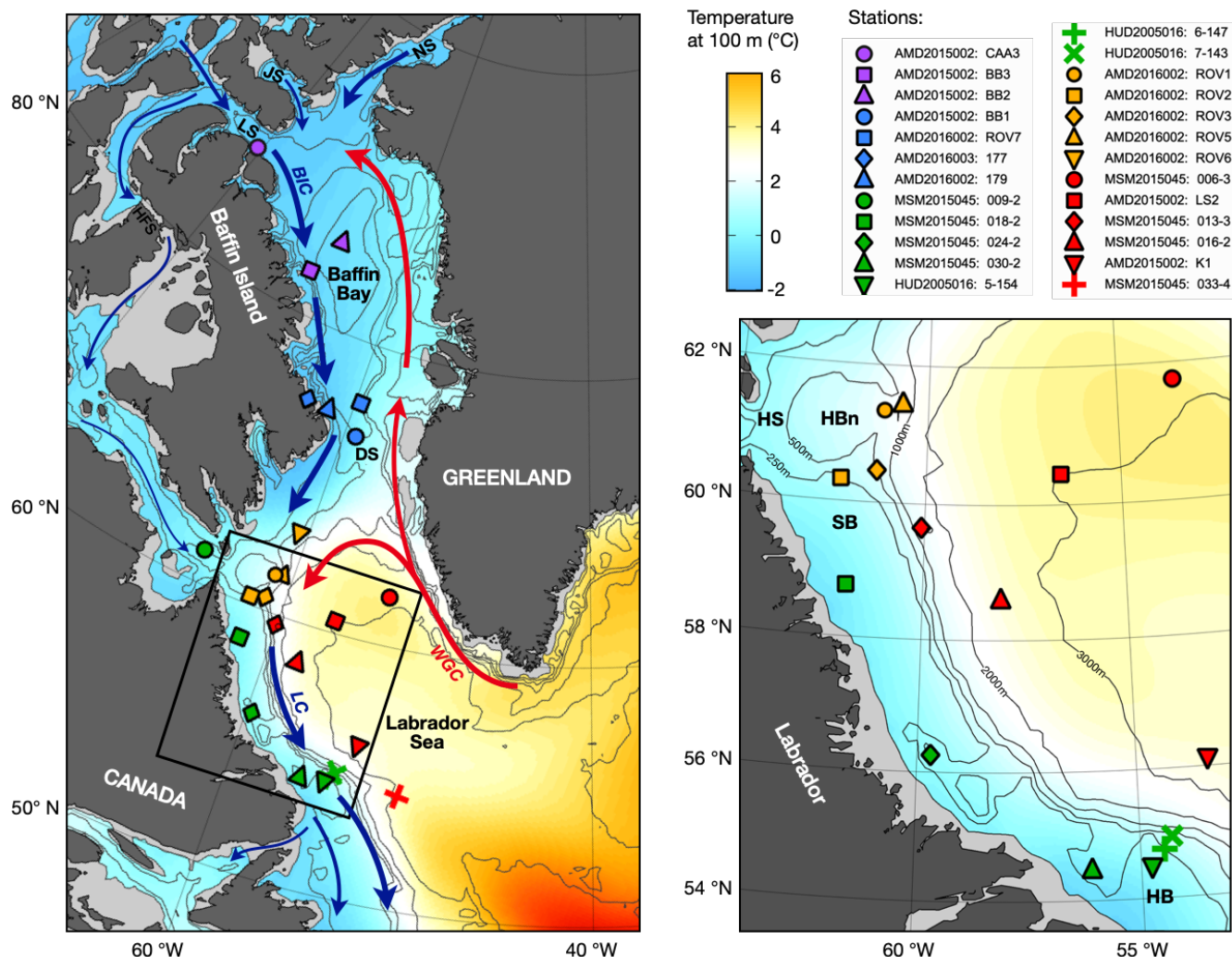
Figure 1: **Map and inset detail of study region with World Ocean Atlas (WOA18) climatological mean temperature at 100 m water depth (colour shading), bathymetry (black contours), major surface currents (arrows) and sampling stations. Abbreviations: West Greenland Current (WGC), Baffin Island Current (BIC), Labrador Current (LC), Nares Strait (NS), Jones Sound (JS), Lancaster Sound (LS), Hecla and Fury Straits (HFS), Hudson Strait (HS), Hatton Basin (HBn), Saglek Bank (SB), Hamilton Bank (HB).**

surfactant-free cellulose acetate (SFCA) membrane filters and stored at -20 °C. Concentrations of NO_3^-, NH_4^+, and $Si(OH)_4$ were measured post-cruise at Dalhousie University according to standard protocols (Grasshoff, 1969) using a Bran and Luebbe autoanalyzer III. During the AMD-2016-002a expedition, samples for nutrient analysis were pre-filtered (0.2 μm) into 15 ml acid-rinsed centrifuge tubes. The samples were analyzed onboard the ship for NO_3^-, NO_2^-, PO_4^{3-}, and $Si(OH)_4$ concentrations using a Bran and Luebbe autoanalyzer III following standard protocols (Grasshoff, 1969). Samples for NO_3^- isotope analysis were collected into acid-cleaned and triple-rinsed 60 ml HDPE bottles without pre-filtration, and stored at -20 °C.

2.2 NO₃⁻ isotope analyses

Seawater samples were prepared for the measurement of dual N and O isotope ratios in NO_3^- following the denitrifier method (Sigman et al., 2001; Casciotti et al., 2002). This method quantitatively converts NO_3^- present in the water samples to nitrous oxide (N_2O) by introducing cultured denitrifying bacteria (*Pseudomonas chlororaphis* f. sp. *aureofaciens,* ATCC# 13985) that lack N_2O reductase activity. The resulting N_2O gas is then analyzed by isotope ratio mass spectrometry (IRMS). Isotopic ratios are reported in delta notation following Eq. (1):

$$\delta^{15}N \text{ or } \delta^{18}O = [R_{sample}/R_{standard} - 1] \times 1000, \qquad\qquad (1)$$

where R represents either $^{15}N/^{14}N$ or $^{18}O/^{16}O$, the standard is the N_2 in the atmosphere (air) or the oxygen in Vienna Standard Mean Ocean Water (VSMOW), and the units are reported as per mille (‰) deviation from the standard ratios. Sample $\delta^{18}O_{NO3}$ data were not corrected for the $\delta^{18}O$ of seawater, because the latter varies minimally, from -2.2 – 0.2 ‰ (Lehmann et al., 2019).

Samples from the MSM45 expedition were analyzed at Dalhousie University using a Thermo Fisher Delta V Plus IRMS interfaced with a Thermo Gas Bench inlet. Data were calibrated using seawater-based reference material USGS-32 ($\delta^{15}N$ = +180 ‰ vs. air, $\delta^{18}O$ = +25.7 ‰ vs. VSMOW), USGS-34 ($\delta^{15}N$ = -1.8 ‰ vs. air, $\delta^{18}O$ = -27.9 ‰ vs. VSMOW) and IAEA-N3 ($\delta^{15}N$ = +4.7 ‰ vs. air, $\delta^{18}O$ = +25.6 ‰ vs. VSMOW) (Böhlke et al., 2003; Gonfiantini, 1984). NO_2^- concentrations were below the detection limit of 0.2 µM, so no prior NO_2^- removal was performed. The blank size constituted < 2 % to the overall sample size for the standard 20 nmol target N_2O concentrations, and < 5 % for the low concentration samples from the biologically productive zone. Analytical reproducibility based on replicate measurements averaged 0.2 ‰ for $\delta^{15}N$ and 0.4 ‰ for $\delta^{18}O$.

Samples from the AMD-2016-002a expedition were analyzed at the University of Basel using a ThermoFisher Delta V Plus IRMS with a customized purge and trap system (modified after McIlvin and Casciotti, 2010, 2011). NO_3^- samples were analyzed in parallel to isotope reference material USGS-34 and IAEA-N3. NO_2^- concentrations were < 0.36 µM, which is < 4 % of the corresponding NO_3^- concentrations, so no prior NO_2^- removal was performed. Blanks constituted < 1 % to the overall sample size. Analytical reproducibility based on replicate measurements averaged 0.2 ‰ for $\delta^{15}N$ and 0.3 ‰ for $\delta^{18}O$.

2.3 Definitions and Calculations

Seawater potential temperature (θ) and potential density anomaly referenced to surface pressure (σ_θ) were calculated from CTD data using the "oce" package in the R computing platform (Kelley and Richards, 2017). Water masses (Table 1) were operationally defined on the basis of θ and/or σ_θ thresholds following published conventions (Stramma et al., 2004; Fratantoni and Pickart, 2007; Azetsu-Scott et al., 2012). Note that Halocline Water (HW) and Labrador Shelf Water (LShW) are distinct water masses, despite overlapping θ and σ_θ characteristics. The latter is formed in the Hudson Strait area from mixing of HW with Irminger Water (IW) and Hudson Bay outflow water (Sutcliffe et al., 1983; Straneo and Saucier, 2008). The base of the

biologically productive zone (z_P) was determined from CTD profiles of chlorophyll fluorescence as the shallowest depth below the subsurface chlorophyll maximum where values < 0.1 mg/m^3 were encountered. Apparent Oxygen Utilization (AOU) was

140   calculated from CTD *in situ* dissolved oxygen profiles using equations in Weiss et al. (1970). The N* parameter, quantifying NO_3^- to PO_4^{3-} imbalances relative to Redfield stoichiometry, was calculated following Eq. (2):

$$N^* = NO_3^- - 16PO_4^{3-} + 2.95, \tag{2}$$

where the constant 2.95 forces a global mean N* of zero (Gruber and Sarmiento, 1997; Deutsch et al., 2001). Regenerated PO_4 (P_{reg}) was calculated from AOU and the stoichiometric constant of Anderson and Sarmiento (1994) from Eq. (3):

145     $$P_{reg} = AOU/170. \tag{3}$$

The ratio of regenerated to measured PO_4^{3-} is reported as $P_{reg/meas}$.

Fraction Pacific water (fPW) was calculated from N and P concentration data in relation to N:P relationships for Pacific and Atlantic endmembers, following Jones et al. (1998). For the purposes of data representativity, accessibility, and propagation

150   of error calculations, we derived new equations for Atlantic and Pacific waters using public domain data from the 2019 version of the Global Ocean Data Analysis Project (GLODAP; Olsen et al., 2020). The N:P relationships are sensitive to the choice of dissolved inorganic nitrogen (DIN) species (NO_3^-, NO_2^-, NH_4^+), particularly in highly productive shelf regions where a significant fraction of the total dissolved N may not be fully nitrified (Yamamoto-Kawai et al., 2008; Mills et al., 2015). The equations reported here use only NO_3^- concentrations, as the inclusion of NO_2^- and NH_4^+ was found to have a negligible impact

155   on fPW calculations (less than calculated uncertainties, see below).

Pacific endmember data were selected from a region encompassing the Canada Basin of the Beaufort Sea (Fig. S1a). The data were filtered (bottom depths > 500m), to exclude shelf waters where NH_4^+ accounts for a significant fraction of the DIN. Data were further filtered ($S \leq 33.5$; $NO_3^- > 2.5$ μM) to exclude waters of Atlantic origin, as well as data from below a kink in the

160   NO_3^- vs. PO_4^{3-} relationship where NO_3^- is exhausted before PO_4^{3-} (Yamamoto-Kawai et al., 2008). The resulting relationship for Pacific water (PW) was calculated following Eq. (4):

$$NO_3^{PW} = 14.07 \pm 0.09 \times PO_4^{PW} - 11.53 \pm 0.15, (r^2 = 0.85, n = 4109). \tag{4}$$

This relationship is within error of the one reported in Yamamoto-Kawai et al. (2008), which was based on total DIN vs. PO_4^{3-} data for a region also encompassing the Chukchi Sea (Fig. S1b). This indicates that the NH_4^+ that accumulates on the Bering

165   and Chukchi shelves in summer is largely nitrified by the time the Pacific-origin shelf waters reach the Canada Basin and Amundsen Gulf (Brown et al. 2015; Granger et al., 2018). In other words, the use of total DIN (instead of just NO_3^-) to define the Pacific N:P relationship, would have a negligible effect on the derived Pacific endmember relationship in Eq. (4).

Atlantic endmember data were obtained from the Irminger Sea (Fig. S1a), which, based on drifter trajectories, represents the

170   source region for waters entering the Labrador Sea via the Irminger Current (Cuny et al., 2002; Jakobsen et al., 2003). Data from the region were filtered ($S \geq 35$, $NO_3^- > 2.5$ μM) to exclude polar waters entering the area through Fram Strait (Sutherland

et al., 2009), and waters affected by nutrient drawdown. The resulting relationship for Atlantic water (AW) entering the Labrador Sea was calculated following Eq. (5):

$$NO_3^{AW} = 15.54 \pm 0.10 \times PO_4^{AW} - 0.26 \pm 0.10, (r^2 = 0.89, n = 2669). \tag{5}$$

175 For any given sample PO_4^{3-} concentration, Eq. (4) and (5) define theoretical endmember NO_3^{PW} and NO_3^{AW} concentrations, respectively. The fPW was then calculated from the sample NO_3^- concentration in relation to NO_3^{PW} and NO_3^{AW}, following Eq. (6):

$$fPW = (NO_3^{sample} - NO_3^{AW}) / (NO_3^{PW} - NO_3^{AW}). \tag{6}$$

Negative values were considered to be devoid of Pacific water and were set to zero. Analytical error in NO_3^- and PO_4^{3-}
180 measurements averaged 1 % and 2 %, respectively. Errors propagated through Eq. (4), (5), and (6) with statistical bootstrapping (n = 10,000, with uniform distributions) resulted in uncertainties of ±3 % (at the 95 % confidence level) on fPW estimates.

Table 1: Water mass definitions, depth ranges, and statistical summaries (mean ± 1 st. dev. (or absolute difference where n < 3), and
185 **number of samples in parentheses) of physical and chemical properties by water mass. HW: Halocline Water; BBW: Baffin Bay Water; LShW: Labrador Shelf Water; IW: Irminger Water; LSW: Labrador Sea Water; NEADW: Northeast Atlantic Deep Water; DSOW: Denmark Strait Overflow Water. All properties were calculated for waters below the biologically productive zone (z_p).**

Water Mass	Operational Definition	Depth Range (m)	S	θ (°C)	AOU (µM)	NO_3^- (µM)	PO_4^{3-} (µM)	$Si(OH)_4$ (µM)	$\delta^{15}N_{NO3}$ (µM)	$\delta^{18}O_{NO3}$ (µM)	N* (µM)	$P_{reg/meas}$	fPW
HW	$\sigma_\theta \leq 27.3$, $\theta \leq 0$	71 – 304	33.30 ± 0.45 (29)	-1.13 ± 0.50 (29)	60 ± 22 (29)	10.1 ± 2.0 (29)	0.98 ± 0.10 (29)	12.6 ± 3.0 (29)	6.0 ± 0.5 (21)	1.3 ± 1.2 (21)	-2.6 ± 1.6 (29)	0.36 ± 0.12 (29)	0.39 ± 0.13 (29)
BBW	$27.5 > \sigma_\theta \leq 27.8$, $\theta \leq 2$	300 – 2286	34.49 ± 0.04 (31)	1.06 ± 0.75 (31)	143 ± 49 (31)	18.5 ± 2.6 (31)	1.40 ± 0.26 (31)	41.6 ± 25.5 (31)	6.1 ± 0.5 (31)	1.1 ± 0.2 (31)	-0.7 ± 1.7 (31)	0.58 ± 0.10 (31)	NA[a]
LShW	$\sigma_\theta \leq 27.4$, $\theta \leq 2$	49 – 188	33.63 ± 0.46 (32)	0.10 ± 0.97 (32)	44 ± 8 (32)	10.2 ± 1.8 (32)	0.88 ± 0.06 (28)	8.8 ± 2.0 (31)	5.4 ± 0.7 (18)	2.1 ± 0.5 (11)	-0.4 ± 1.6 (28)	0.29 ± 0.04 (28)	0.22 ± 0.12 (28)
IW	$27.3 > \sigma_\theta \leq 27.68$, $\theta > 2$	71 – 826	34.61 ± 0.19 (54)	3.20 ± 0.80 (54)	52 ± 19 (53)	14.5 ± 1.6 (46)	1.00 ± 0.07 (36)	10.2 ± 3.8 (44)	4.9 ± 0.3 (39)	2.1 ± 0.5 (37)	1.9 ± 0.8 (36)	0.31 ± 0.11 (36)	0.04 ± 0.05 (36)
LSW	$27.68 > \sigma_\theta \leq 27.80$, $\theta > 2$	205 – 1644	34.88 ± 0.03 (34)	3.54 ± 0.25 (34)	36 ± 11 (32)	16.1 ± 0.8 (31)	1.02 ± 0.05 (23)	8.5 ± 1.8 (32)	4.8 ± 0.3 (34)	2.1 ± 0.4 (34)	2.6 ± 0.4 (23)	0.18 ± 0.05 (23)	0.00 ± 0.02 (23)
NEADW	$27.80 > \sigma_\theta > 27.88$	1825 – 2679	34.92 ± 0.00 (10)	2.83 ± 0.28 (10)	51 ± 5 (8)	16.0 ± 0.5 (8)	1.04 ± 0.02 (3)	10.8 ± 0.8 (5)	4.7 ± 0.3 (10)	1.8 ± 0.3 (11)	2.3 ± 0.1 (3)	0.28 ± 0.00 (3)	0.00 ± 0.00 (3)
DSOW	$27.88 > \sigma_\theta$	2635 – 3014	34.91 ± 0.01 (5)	1.95 ± 0.26 (5)	53 ± 6 (3)	15.2 ± 0.4 (5)	1.01 ± 0.02 (2)	12.2 ± 1.3 (4)	4.9 ± 0.2 (7)	2.0 ± 0.3 (7)	2.0 ± 0.12 (2)	NA	0.02 ± 0.01 (2)

[a]It is not possible to calculate fPW for BBW. See section 3.2.4 for details

3 Results

3.1 Hydrographic summary

Pacific water propagates as a halocline layer through the Canadian Arctic Archipelago, then through Baffin Bay, and into the Labrador Sea via Davis and Hudson Straits (Fig. 1; Tang et al., 2004). The 25 stations that were sampled for this study are distributed along this net transport pathway of Pacific water and are therefore ideally situated for investigating the distribution

 of NO_3^- isotopic ratios with respect to fPW. The stations are grouped into five hydrographic regimes on the basis of common water column properties. A summary of hydrographic properties by regime follows below. To help with visualization, the data are colour-coded by hydrographic regime consistently throughout the subsequent figures. A diagram of θ-S data is shown in Fig. 2, with delineations for the different water masses. Depth profiles are shown for all stations in Fig. 3 and separately for each hydrographic regime in Fig. S1-S5.
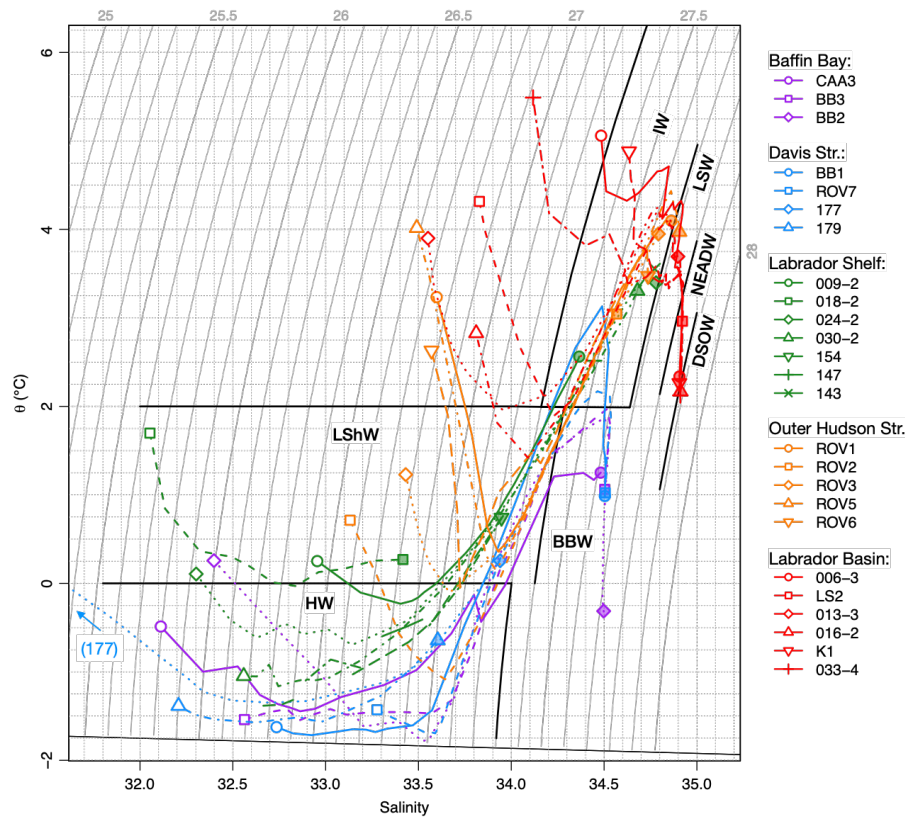
Figure 2: Temperature - salinity diagram for individual stations. For each profile, the 30m depth level is indicated by open symbols; the bottom level is indicated by shaded symbols. Data from < 30 m omitted for clarity. Bold black lines indicate temperature and σ_T limits for water masses discussed in text: HW, Halocline Water, LShW, Labrador Shelf Water; BBW, Baffin Bay Water; IW, Irminger Water; LSW, Labrador Sea Water; NEADW, Northeast Atlantic Deep Water; DSOW, Denmark Strait Overflow Water.

The Baffin Bay regime was represented by three stations (BB2, BB3, CAA3). Station BB2 was located inside the central Baffin gyre at a depth of 2300 m. BB3 was located along the path of the Baffin Island Current at a depth of 1243 m. CAA3 was located at the southern side of Lancaster Sound at a depth of 690 m. The three stations displayed similar θ and S profiles, characteristic of Baffin Bay more broadly (Fig. 3; Tang et al., 2004). A surface layer, formed by summer warming and melting, extended down to 20-30 m. Below this, a layer of almost isothermal, cold ($\theta < -1.5$ °C) water, increasing in salinity from < 32 to 34, extended down to 200 m. This layer represents the Pacific-sourced HW, which is formed in the Beaufort Sea and adjacent shelves, and then modified by regional winter cooling and sea ice formation in northern Baffin Bay and along the northwestern Greenland coast (Bourke et al., 1989; Münchow et al., 2015; Rysgaard et al. 2020). Below the HW, a warmer (θ approaching +2 °C) and saltier (S > 34) layer extended down to 700 m. This layer represents the diluted remnants of Atlantic-sourced IW, often referred to as West Greenland Intermediate Water, which flows northward via the West Greenland Current and spreads throughout the entire Baffin Bay (Tang et al., 2004; Münchow et al., 2015). The θ and S data over this depth interval at CAA3 were more variable, reflecting the interleaving of different water masses by the complex tidal currents in Lancaster Sound (Fig. S3; Prinsenberg and Hamilton, 2005). At stations BB2 and BB3, the waters below 700 m form a distinct tail on the θ-S diagram (Fig. 2). We refer to this as "Baffin Bay Water" (BBW; $\theta < 2$ °C, $27.5 > \sigma_\theta \leq 27.8$), which, for convenience, groups waters generally referred to as Baffin Bay Deep Water for 1200 < z < 1800 m and Baffin Bay Bottom Water for z > 1800 m (Tang et al., 2004), as well as the shallower waters from 700 < z < 1200 m. The BBW is also distinguished by the rapid increase in AOU with depth (Fig. 3c).

The northern Davis Strait regime was represented by four stations (177, 179, BB1, ROV7). Station 177 was located within 2 km of coastal Baffin Island at a depth of 376 m. Despite the coastal location of station 177, it is hydrographically connected to more open water via a deep, northeast-trending cross-shelf trough (Broughton Trough). Station 179 was located on the Baffin shelf break at 186 m. Station BB1 was located on the northern flank of the Davis Strait sill at a depth of 1042 m. Station ROV7 was located over the Greenland slope (Disko Fan) at a depth of 932 m. Hydrographic profiles at these four stations were similar to those of the Baffin Bay regime, with the characteristic HW and IW layers (Fig. 3). A seemingly thicker surface layer extending down to > 50 m at station 177 is a result of the later sampling date (late September) than at the other three stations, which were sampled in late July/early August. The HW layer was thicker at stations 177 and 179, which are located in the path of the Baffin Island Current and thins out toward the more centrally located BB1 and ROV7, also evident from the shallowing isopycnals (Fig. S4 σ_θ profiles; Tang et al., 2004; Azetsu-Scott et al., 2012). Stations BB1 and ROV7 sampled the IW ($\theta > 2$ °C, S > 34.4) from 300-500 m, and BBW below about 700 m.
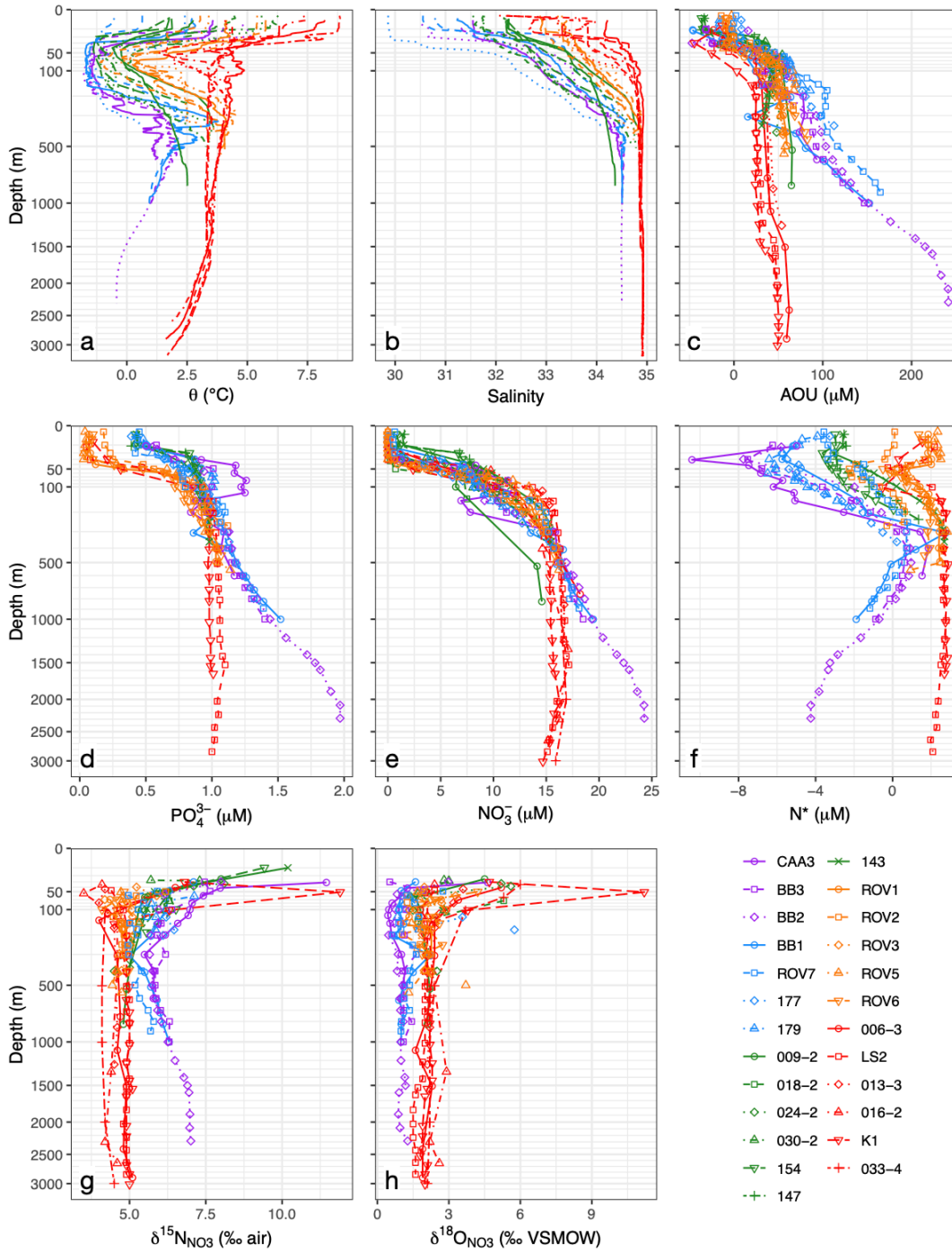
Figure 3: Depth profiles for (a) potential temperature, (b) salinity, (c) apparent oxygen utilization (AOU), (d) PO$_4^{3-}$, (e) NO$_3^-$, (f) N*, (g) δ^{15}N of NO$_3^-$, and (h) δ^{18}O of NO$_3^-$. Points not connected by lines in (h) are suspected analytical outliers.

The Labrador Shelf regime comprised seven stations (009, 018, 024, 030, 154, 147, 143). Station 009 was located over a > 900 m deep basin in the main channel of the Hudson Strait but had a similar hydrographic profile to the other stations on the Labrador Shelf. Stations 018, 024, and 030 were located on an along-shelf transect, located at depths of 200 m, 534 m, and 535 m, respectively. Stations 154, 147, and 143 were located along an outer cross-shelf transect of Hamilton Bank at depths of 202 m, 245 m, and 344 m, respectively. A surface layer extended down to about 30 m at all stations, underlain by the remnants of the HW, modified by tidal mixing and warming southward of Davis Strait (Fig. 3; Tang et al., 2004). This layer is often called the "Cold Intermediate Layer" (Colbourne et al., 2016), but herein is referred to as "Labrador Shelf Water" (LShW; $\theta < 2$ °C, S < 34.2) for ease of reference. The LShW extended down to between 150-300 m and was underlain by IW where the bottom depth exceeded 300 m. The influence of IW increased from west to east, as becomes apparent from the cross-shelf increase in θ and S at stations 030, 154, 147, and 143 (Fig. S5; Fratantoni and Pickart, 2007).

The outer Hudson Strait regime comprised five stations (ROV1, ROV2, ROV3, ROV5, ROV6) concentrated around an area seaward of the Hudson Strait, around the shelf break (Fig. 1 inset). Stations ROV1 and ROV5 were located on the sill of an outer shelf bathymetric depression (Hatton Basin) at approximately 500 m water depth. Stations ROV2 and ROV3 were located along the northern flank of Saglek Bank at 279 and 436 m, respectively. Station ROV6 was located further north, at 456 m depth, but had similar hydrography as the other four stations (Fig. 3). The surface and bottom currents in these areas are quite strong, up to 0.60 m s^{-1} at station ROV3, generally flowing NW to SE, but with a strong tidal influence linked to the macrotidal oscillation in Frobisher Bay (Zedel et al., unpublished bottom current meter data from NE Saglek Bank). The surface layer extended down to about 30 m at all stations. Nutrient data, specifically PO$_4^{3-}$ concentrations and N* values, discussed below, clearly distinguished the surface waters of this regime from the other regimes presented above. The water mass structure was overall similar to that of the Labrador Shelf regime, but with a thinner, warmer and saltier layer of LShW underlain by warmer and saltier IW (Fig. S6).

Finally, the Labrador Basin regime comprised six stations (006, 013, 016, 033, LS2, K1) located in the deep waters of the Labrador Sea, at depths of 1280 m (station 013) to 3292 m (station 033). Hydrographic profiles at these stations (Fig. 3) reflect the well-known water mass structure in the Labrador Sea (e.g. Yashayaev and Loder, 2016). Doming of isopycnals leads to the thinning and shoaling of the IW layer from the margins (e.g. Station 013, IW: 70-500 m) to the center of the basin (e.g. Station K1, IW: 30-150 m) (Fig. S7). Below the IW, a thick layer of Labrador Sea Water (LSW: $27.68 > \sigma_\theta \leq 27.80$, $\theta > 2$ °C), extended down to 1500-2000 m, underlain by Northeast Atlantic Deep Water (NEADW: $27.80 > \sigma_\theta > 27.88$) to 2400-2700 m, then Denmark Strait Overflow Water (DSOW: $\sigma_\theta > 27.88$).

3.2 Nutrients

3.2.1 Nutrient concentrations in the biologically productive zone

275   Sampling was conducted in the months of June – August, which follows the spring bloom throughout most of the study region (Tremblay et al., 2006; Frajka-Williams et al., 2010). Complete or near-complete utilization of NO_3^-, PO_4^{3-}, and $Si(OH)_4$ was observed in the upper 30 m of the water column at all sites, with evidence of partial nutrient utilization to < 120 m. Minima in NO_3^- in the surface waters averaged < 1 μM and did not vary by hydrographic region (Fig. 3e). Minima in PO_4^{3-}, by contrast, exhibited a striking bimodal distribution with respect to region, with concentrations < 0.1 μM for most of the outer Hudson

280   Strait and Labrador Basin stations, and > 0.4 μM for all of the Baffin Bay, Davis Strait, and Labrador Shelf stations (Fig. 3d). Minima in $Si(OH)_4$ exhibited a similar bimodality (< 1 μM and > 5 μM) with respect to hydrographic regions (Fig. S1-S5). Thus, NO_3^- was relatively more limiting to primary production than either PO_4^{3-} or $Si(OH)_4$ in the colder/fresher hydrographic regions, as observed previously (Tremblay et al., 2006; Harrison and Li, 2008; Martin et al., 2010; Ferland et al., 2011; Fragoso et al., 2017).

285   ### 3.2.2 Nutrient concentrations below the biologically productive zone

Nutrient concentrations generally stabilized below the biologically productive zone (depth > z_P), with the exception of BBW, in which concentrations increased rapidly with depth (Fig. 3d, e). The elevated concentrations result from in situ nutrient regeneration in Baffin Bay Deep and Bottom Water (Jones et al., 1984; Tremblay et al., 2002; Lehmann et al. 2019). Baffin Bay is a 2300 m deep basin enclosed by < 700 m deep sills. The enclosed bathymetry and permanent halocline restrict

290   circulation, thereby trapping particulate organic matter (POM) and remineralized nutrients. More precisely, given the long residence time of the deep and bottom waters (77-1450 years; Top et al., 1980; Wallace et al., 1985), high fluxes of POM originating from the productive northern Baffin Bay (Klein et al., 2002; Tremblay et al., 2002; Lalande et al., 2009) accumulate at depth. The subsequent *in situ* remineralization of this sinking POM leads to the observed increase in nutrients, seen as an increase in $P_{reg/meas}$, and drawdown of oxygen (increase in AOU) in the deep basin (Fig. 3c-e). While O_2 concentrations remain

295   too high to support denitrification in the water column, dissimilatory NO_3^- consumption in the sediments is supported by the low oxygen concentrations in the water above, and acts as a potential sink for dissolved NO_3^- in the lower water column (Lehmann et al., 2019). Indeed, BBW had two-fold higher PO_4^{3-} (1.4 ± 0.3 μM) and three-fold higher $Si(OH)_4$ concentrations (41 ± 25 μM) than any of the other water masses (Table 1), but only somewhat higher NO_3^- (19 ± 3 μM). For water masses other than BBW, there were significant differences in NO_3^- and PO_4^{3-}, but not $Si(OH)_4$ (1-way ANOVA). HW and LShW had

300   lower NO_3^- concentrations (~10 μM) than IW, LSW, NEADW (~15 μM). The distribution of PO_4^{3-} by water mass was similar, except that the concentration in HW (0.98 μM) was closer to that of IW, LSW, and NEADW (> 1 μM) than LShW (0.88 μM).
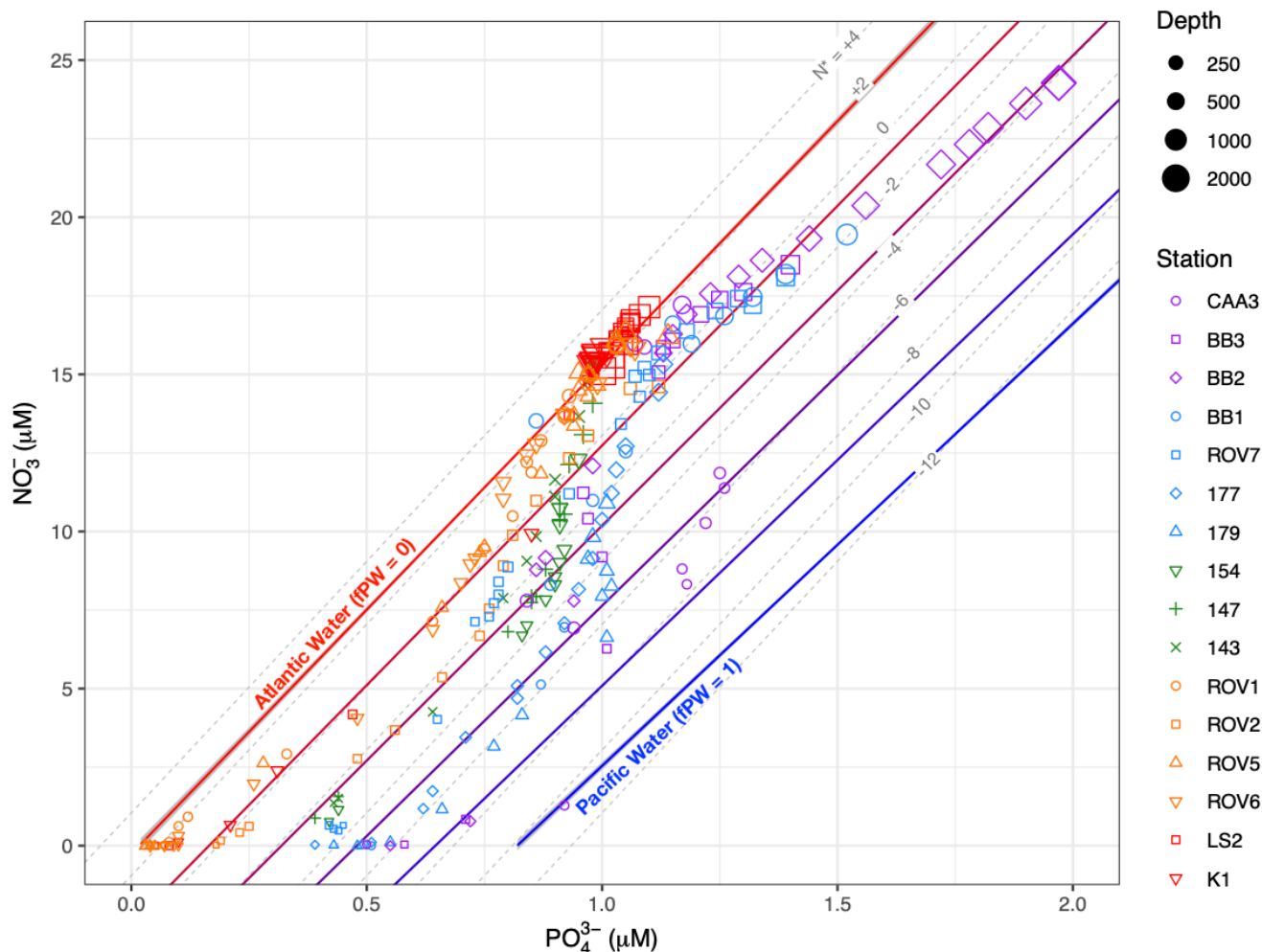
Figure 4: NO₃⁻ vs. PO₄³⁻ data for samples from all water depths in this study, with lines representing empirically-derived Atlantic and Pacific water endmember N:P relationships. Endmember lines are enclosed by 95 % confidence intervals. Coloured red to blue lines between Atlantic and Pacific endmembers represent lines of constant fraction Pacific water (fPW), in increments of 0.2. Also shown are grey dashed lines of constant N*.

3.2.3 Nutrient ratios

NO₃⁻ to PO₄³⁻ stoichiometry is expressed in profiles of N*, which were available for 19 of the 25 stations with paired NO₃⁻ and PO₄³⁻ concentration data (Fig. 3f). Positive N* reflects a water mass history of excess fixed NO₃⁻, e.g., by net N₂-fixation; negative N* reflects a NO₃⁻ deficit, relative to the mean global ocean (Gruber and Sarmiento, 1997), induced by denitrification in the broadest sense (i.e., including other modes of suboxic DIN transformations to N such as anammox). N* signatures can be imported from other ocean regions, or can be generated within a given water mass or region, depending on biogeochemical conditions. Positive N* occurred throughout most of the outer Hudson Strait and Labrador Basin profiles. The water masses

LSW, NEADW, and DSOW (z > 200 m) all showed mean N* > 2 µM (Table 1), consistent with an Atlantic origin (Gruber and Sarmiento, 1997; Jenkins et al., 2015). The deflections to lower N* at z ~ 100 m (Fig. 3f) correspond to IW, which has lower N* due to mixing at the shelf-slope front (Cuny et al., 2002; Fratantoni and Pickart, 2007). Negative N* occurred through most of the Baffin Bay, Davis Strait, and Labrador Shelf profiles (Fig. 3f). As noted earlier, HW partially originates from the Bering and Chukchi shelf areas, where sedimentary denitrification fueled by high water column productivity acts as a sink for dissolved NO_3^-. The resulting pronounced minimum in N* (< -10 µM; Yamamoto-Kawai et al., 2008; Mills et al., 2015) propagates via HW through the Canadian Arctic Archipelago and into Baffin Bay (Carmack and McLaughlin, 2011; Tremblay et al., 2015). This import of fixed-N deficient waters explains the lowest N* values in HW (Table 1), with minima < 6 µM at stations CAA3, BB3, BB2, BB1, 177, and 179 (Fig. 3f). BBW had the next most negative N*, likely due to the upward propagation of partially denitrified bottom water nutrients (Tremblay et al., 2002; Lehmann et al., 2019). LShW had the most variable N* signatures, resulting from the mixing of HW and IW. The effect of this mixing is clearly evident in the cross-shelf increase in N* on the Labrador Shelf, from < -3 µM at station 154 to > 2 µM at station 143 (Fig. S5).

3.2.4 Fraction Pacific water

A cross plot of NO_3^- vs. PO_4^{3-} concentrations helps to visualize the fraction of Pacific water among individual water samples relative to lines representing 'pure' Atlantic and Pacific waters (Fig. 4). Note that the Atlantic line (fPW = 0) coincides with a line of constant N* = 2 µM (Fig. 4), which is also the average N* value for North Atlantic intermediate waters (Gruber and Sarmiento, 1997). The Pacific line (fPW = 1) falls between the lines where N* = -10 to -12 µM. Thus, depth profiles of fPW mirror those of N* (Fig. S3-S7). Data from the Labrador Basin regime fall on or close to the Atlantic line because the water masses at these stations (IW, LSW, NEADW, DSOW) are mostly Atlantic-sourced. Data from the other regimes plot increasingly towards the Pacific line in the order of Hudson Strait, Labrador Shelf, Davis Strait, and Baffin Bay. Maxima in fPW (> 0.6) were found within the core of HW sampled at Lancaster Sound (CAA3) and along the path of the Baffin Island Current (BB2, BB3, BB1, 177 179) (Fig. S3, S4).

Water samples with PO_4^{3-} > 1.25 µM and NO_3^- > 17.5 µM correspond to BBW (Fig. 4). The data fall along a N:P trajectory with a slope of 9.6 +/- 0.3. This slope is considerably lower than the slopes of either Atlantic or Pacific endmember waters. It arises from *in situ* remineralization of POM, as indicated by $P_{reg/meas}$ values > 0.5 (Fig. S3, S4), with a partial loss of NO_3^- via sedimentary denitrification (Lehmann et al., 2019). The denitrification generates N* values as low as -4.3 µM. Thus, the process that leads to low N* in BBW is separate and distinct from the processes that generate low N* in HW. As a result, it is not possible to calculate fPW for BBW, because remineralization and denitrification overprint the pre-formed N:P signatures (Jones et al., 2003).

Another complication with fPW estimates, as noted in section 2.3, is that elevated concentrations of NO_2^- and NH_4^+ may alter apparent N:P ratios with respect to the derived endmember relationships. Within the overall study region, NO_2^- and NH_4^+

concentrations below the euphotic zone are generally < 1 μM (Harrison and Li, 2008; Martin et al., 2010; Azetsu-Scott et al., 2012). Where measured in the present study, NO_2^- concentrations were < 0.36 μM and NH_4^+ concentrations were < 1 μM, except for six samples from the Labrador Basin regime, with NH_4^+ concentrations up to 2.5 μM (Fig. S3-S7). Thus, with the exception of those few samples, the overall low NO_2^- and NH_4^+ concentrations should have little impact on fPW estimates.
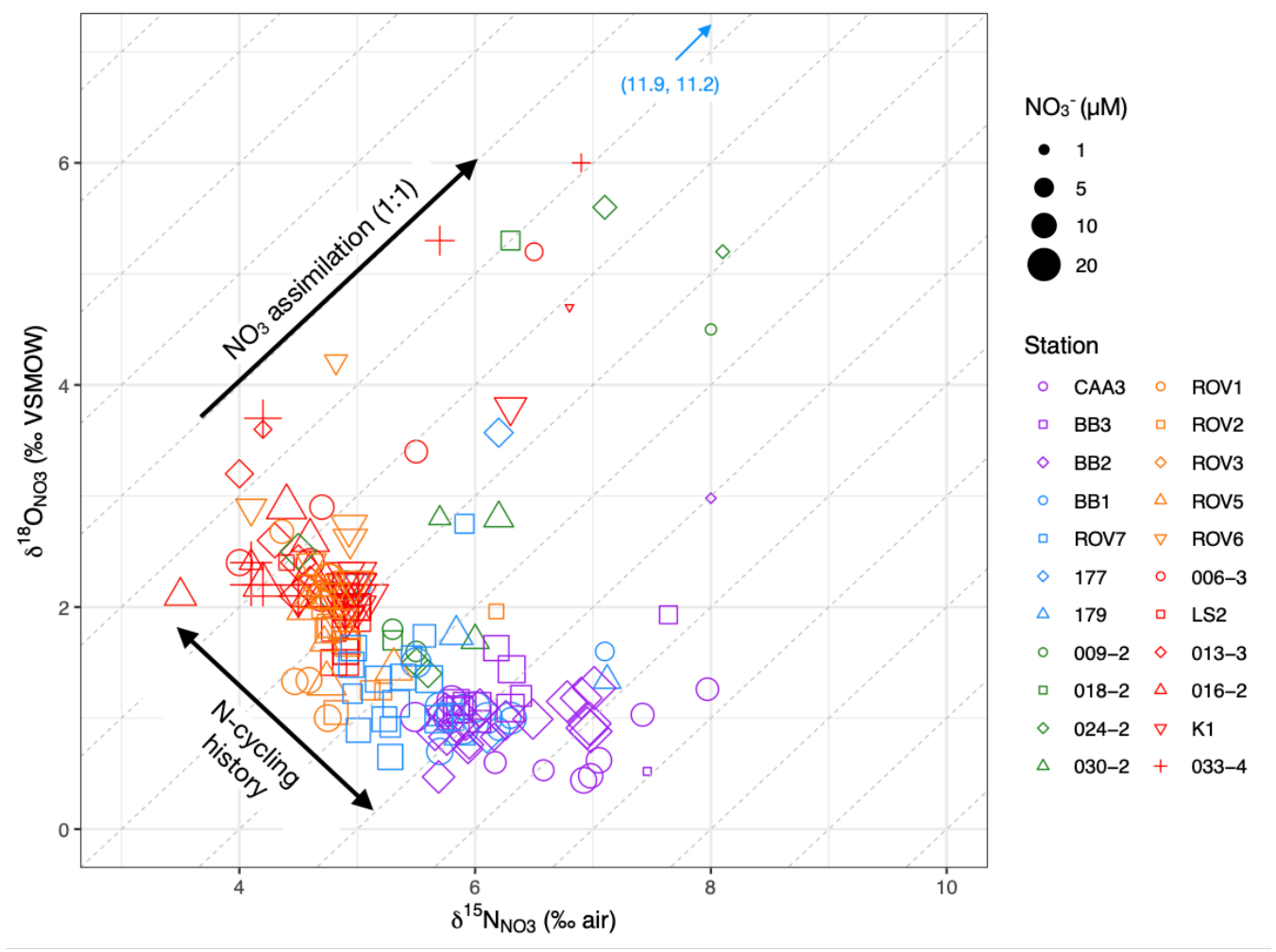
Figure 5: $\delta^{18}O_{NO3}$ vs. $\delta^{15}N_{NO3}$ by station and NO_3^- concentration. The dense cluster of data centered at bottom left represents deep waters ($> z_p$). Diagonal lines represent 1:1 isotopic fraction of ^{18}O and ^{15}N. Arrows denote isotopic fractionation associated with NO_3^- assimilation in the biologically productive zone, and differences in water mass N cycling histories.

3.3 NO$_3^-$ isotope ratio variability

Isotope ratios of NO$_3^-$ were measured at all 25 stations for $\delta^{15}N_{NO3}$ and all but the three stations from the HUD-2005-016 expedition for $\delta^{18}O_{NO3}$. Patterns of isotopic variability are presented separately for waters in and below the base of biologically productive zone (z_P) in the following sub-sections.

3.3.1 Isotope ratios of NO$_3^-$ in the biologically productive zone

For waters above z_P, $\delta^{15}N_{NO3}$ increased from values of around 5-6 ‰ to maxima of 12 ‰ toward the surface (Fig. 3g). The $\delta^{18}O_{NO3}$ similarly increased from < 2 ‰ to 11 ‰ (Fig. 3h). In a cross plot of $\delta^{18}O_{NO3}$ vs. $\delta^{15}N_{NO3}$, these isotopic enrichments extend approximately along lines of 1:1 (Fig. 5). The increase in isotopic ratios coincides with a decrease in NO$_3^-$ concentrations, and an increase in chlorophyll, as interpreted from fluorescence profiles (Fig S1-S5). Together, these patterns are consistent with coupled (identical) fractionation of ^{15}N and ^{18}O during NO$_3^-$ assimilation (Granger et al., 2004; Sigman et al., 2005).

To further demonstrate the effect of NO$_3^-$ assimilation on isotopic ratios, $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$ are plotted against the natural logarithm of NO$_3^-$ concentrations, where the "kinks" in the relationships represent the base of the NO$_3^-$ assimilation zone (Fig. 6). To the left of the kinks, both $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$ increase with decreasing NO$_3^-$, again consistent with coupled fractionation of ^{15}N and ^{18}O during NO$_3^-$ assimilation. Moreover, assuming a mainly vertical supply of nutrients to the euphotic zone, the isotopic composition of the NO$_3^-$ used in assimilation may be approximated by the minima in $\delta^{15}N$ and $\delta^{18}O$ at the kinks (Fig. 6; Rafter and Sigman, 2016; Peters et al., 2018). In this respect, the $\delta^{15}N$ of the assimilated NO$_3^-$ increases from lowest values at the Labrador Basin stations to highest values at the Baffin Bay stations. The minima in $\delta^{18}O_{NO3}$ data show the opposite trend, with the lowest values at the Baffin Bay stations and highest values in the Labrador Basin.

3.3.2 Isotope ratios of NO$_3^-$ below the biologically productive zone

Below z_P, $\delta^{15}N_{NO3}$ ranged from 4.1-6.5 ‰ (Fig. 3) and varied significantly by water mass (1-way ANOVA; Table 1). The Atlantic-derived water masses (IW, LSW, NEADW, DSOW) sampled in the Hudson Strait and Labrador Basin regimes had the lowest mean $\delta^{15}N_{NO3}$ (4.8 ± 0.3‰). This value is identical to the $\delta^{15}N_{NO3}$ of North Atlantic intermediate-depth waters; it represents the basin-scale N isotopic mass balance between relatively ^{15}N-depleted in Atlantic subtropical thermocline water and Mediterranean Overflow Water, and relatively ^{15}N-enriched NO$_3^-$ in Antarctic Intermediate Water (Marconi et al., 2015). The Pacific-influenced HW, as well as BBW sampled in Baffin Bay and Davis Strait, displayed the highest mean $\delta^{15}N_{NO3}$ (> 6 ‰). The elevated $\delta^{15}N_{NO3}$ in HW reflects its predominant origin in the western Arctic. At the entrance to the western Arctic, Pacific-origin NO$_3^-$ propagating onto the Bering Shelf has an already high $\delta^{15}N_{NO3}$ (6.3 ‰; Lehmann et al., 2005). As Pacific waters flow across the productive Bering and Chukchi shelves, NO$_3^-$ becomes further isotopically enriched due to benthic coupled nitrification-denitrification (CPND), which results in the removal of isotopically light NH$_4^+$ from the system and the

efflux of heavy NH_4^+ into the overlying water column (Granger et al., 2011; Brown et al., 2015). Subsequent water column nitrification leads to the characteristically high $\delta^{15}N_{NO3}$ signature of the western Arctic upper halocline (~ 8.0 ‰, Brown et al., 2015; Granger et al., 2018; Fripiat et al., 2018). The $\delta^{15}N_{NO3}$ signature in BBW, on the other hand, is consistent with *in situ* remineralization in deep Baffin Bay, as indicated by high AOU and nutrient concentrations. The high $\delta^{15}N_{NO3}$ (> 7.0 ‰; Fig. 3g) indicates that the POM exported to the deep Baffin Bay is largely fueled by Pacific-derived nutrients in northern Baffin Bay (Lehmann et al., 2019), given that the N isotopic composition of newly nitrified NO_3^- largely reflects the signature of its source substrate. The LShW sampled on the Labrador shelf exhibited intermediate and more variable $\delta^{15}N_{NO3}$ signatures (5.4 ± 0.7 ‰), which, as with the corresponding N* data, was consistent with mixing of HW and IW across the Labrador Shelf.
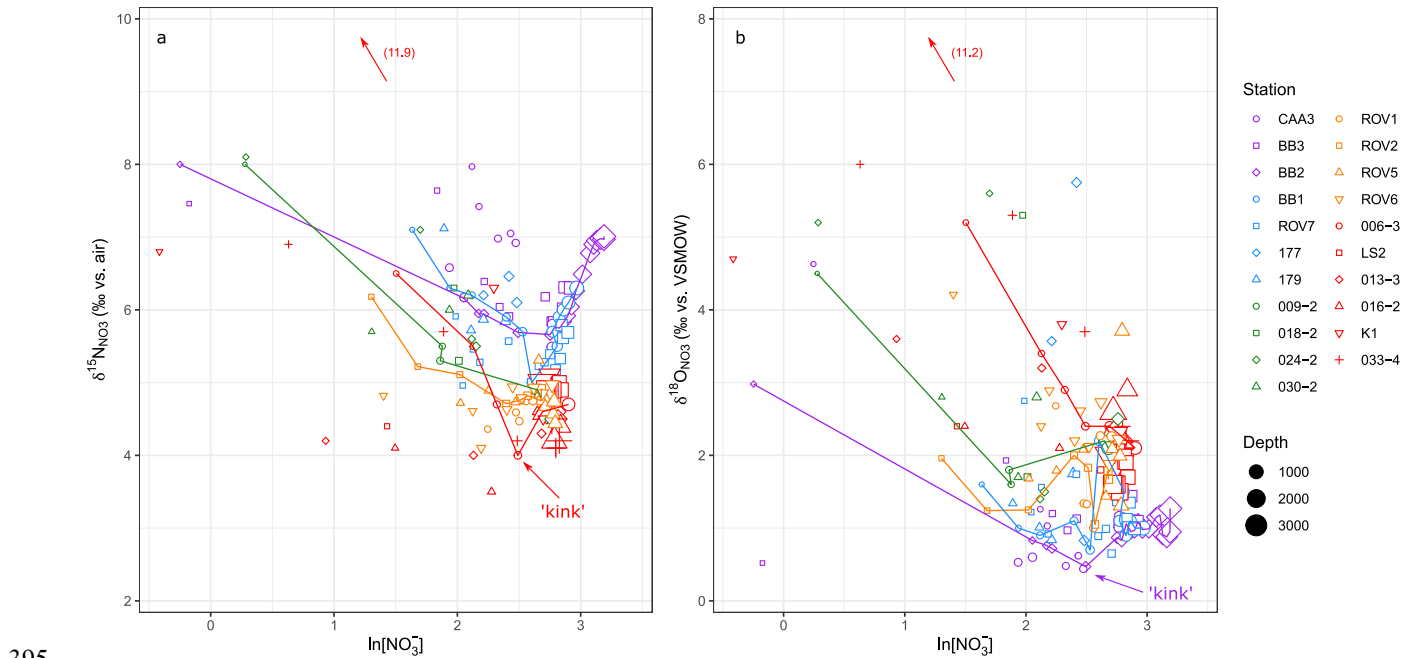
Figure 6: (a) $\delta^{15}N_{NO3}$ and (b) $\delta^{18}O_{NO3}$ plotted against the natural logarithm of NO_3^- concentrations. The 'kink' represents the base of the NO_3^- assimilation zone. For clarity, only some of the station data are connected by lines.

Values of $\delta^{18}O_{NO3}$ also varied significantly by water mass (1-way ANOVA) but in mirror image to $\delta^{15}N_{NO3}$ (Table 1). The LSW, NEADW, and DSOW exhibited higher $\delta^{18}O_{NO3}$ (> 1.8 ‰); HW and BBW had lower values (> 0.3 ‰). The low $\delta^{18}O_{NO3}$ in HW is within the range of values previously reported for the western Arctic upper halocline layer (Brown et al., 2015; Granger et al., 2018; Fripiat et al., 2018), where values close to 0 ‰ are indicative of the highly remineralized NO_3^- pool, as nitrification introduces a low $\delta^{18}O$ close to a value of ambient seawater (+1.1 ‰; Casciotti et al., 2008; Sigman et al., 2009; Buchwald et al., 2012). Low $\delta^{18}O_{NO3}$ values associated with BBW similarly reflect the high proportion of remineralized nutrients in deep Baffin Bay (Lehmann et al., 2019). The significantly higher $\delta^{18}O_{NO3}$ in the Labrador Sea subsurface layer

reflects the remote signal of partial assimilation in the Southern Ocean, as well as a higher $\delta^{18}O$ of water oxygen atoms that are incorporated during remineralization in transit in the Atlantic versus the Arctic (Marconi et al., 2015; Granger et al., 2018). The differential NO_3^- isotope tagging of the various sub-euphotic water masses, which is a function of their different origin and N-cycling history, holds great potential to trace the distribution of these water masses in the Northwest Atlantic and thus

410   to assess the contribution from Pacific sources.

To explore the biogeochemical drivers of $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$ in the different water masses further, a correlation matrix of physical and chemical variables was constructed (Fig. S8). The strongest covariates of $\delta^{15}N_{NO3}$ were fPW (r = 0.89) and N* (r = -0.86), followed by θ (r = -0.75), then variables associated with diatom and POM remineralization in BBW: $Si(OH)_4$ (r =

415   0.65), AOU (r = 0.64), and PO_4^{3-} (0.61). Upon recalculating the correlation matrix *without* BBW (see below), the correlations with fPW (r = 0.91) and N* (r = -0.89) became even stronger, followed by salinity (r = -0.82), θ (r = -0.81) and NO_3^- (r = -0.65). All five of these parameters exhibit multicollinearity, that is, waters with high fPW also have low N*, and are colder, fresher, and have less preformed NO_3^- than waters with low fPW. The same parameters were also correlated with $\delta^{18}O_{NO3}$ but opposite in sign (Fig. S9).

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In a plot of $\delta^{15}N_{NO3}$ versus N*, the regression line through the main group of data is highly significant (p << 0.001) with an r^2 value of 0.78 (Fig. 7a). Note that BBW data plot above and to the right of the rest of the data. We hypothesize that this shift arises from remineralization of PO_4^{3-} and NO_3^-, followed by loss of the NO_3^- via sedimentary denitrification (Lehmann et al., 2019). A conceptual model of this two-step process is shown in the Fig. 7a inset. The source of preformed nutrients in the deep

425   Baffin Bay is still debated (Tang et al., 2004), but, assuming a dominantly Atlantic source (Azetsu-Scott et al., 2012), the preformed nutrients would plot near the other Atlantic waters with N* values > 0 µM and $\delta^{15}N_{NO3}$ ~ +5 ‰ (e.g., Marconi et al., 2015; Granger et al., 2018; Fripiat et al., 2018). Under a simplifying assumption of near-Redfield stoichiometry, the N* would remain unchanged during remineralization. (We note, however, that lower than Redfield N:P uptake has been documented in Baffin Bay (Harrison et al., 1982) which would shift the N* to lower values). The POM originates in the

430   overlying $^{15}N_{NO3}$-enriched HW, which would generate remineralized NO_3^- with relatively high $\delta^{15}N$ in BBW. Subsequent sedimentary denitrification would shift the N* to lower values because the process acts as a sink for NO_3^- but not PO_4^{3-}. This process has negligible effect on water column $\delta^{15}N_{NO3}$, as the NO_3^- is completely reduced in the sediments such that there is effectively no isotopically modified NO_3^- to diffuse back into the overlying water (Brandes and Devol, 1997; Lehmann et al., 2004; 2007). This again highlights that the processes affecting N* and $\delta^{15}N_{NO3}$ in BBW are separate and distinct from those

435   influencing HW or the other water masses (Lehmann et al., 2019).

The relationship between $\delta^{15}N_{NO3}$ and fPW is also highly significant (p << 0.001) with an r^2 of 0.80 (Fig. 7b). The intercept, corresponding to 100 % Atlantic water, is 4.8 ± 0.04 ‰. This value coincides exactly with previous estimates of the mean $\delta^{15}N_{NO3}$ (4.8 ‰) in North Atlantic intermediate and deep waters (Marconi et al., 2015). This is not surprising given that

440   samples of 100 % Atlantic water are represented in the data distribution (Fig. 4). The regression also predicts the $\delta^{15}N_{NO3}$ for

100 % Pacific water at 8.3 ± 0.2 ‰. This value likewise matches with previous measurements of the Pacific halocline water

measured in the eastern Beaufort Sea, downstream of the centers of CPND ($\delta^{15}N_{NO3}$ = 8.0 ± 0.1 ‰; Brown et al., 2015; Granger

et al., 2018). Such accurate prediction of the Pacific endmember is remarkable, considering the degree of extrapolation beyond

the limit (fPW ≤ 0.6) of sample data.

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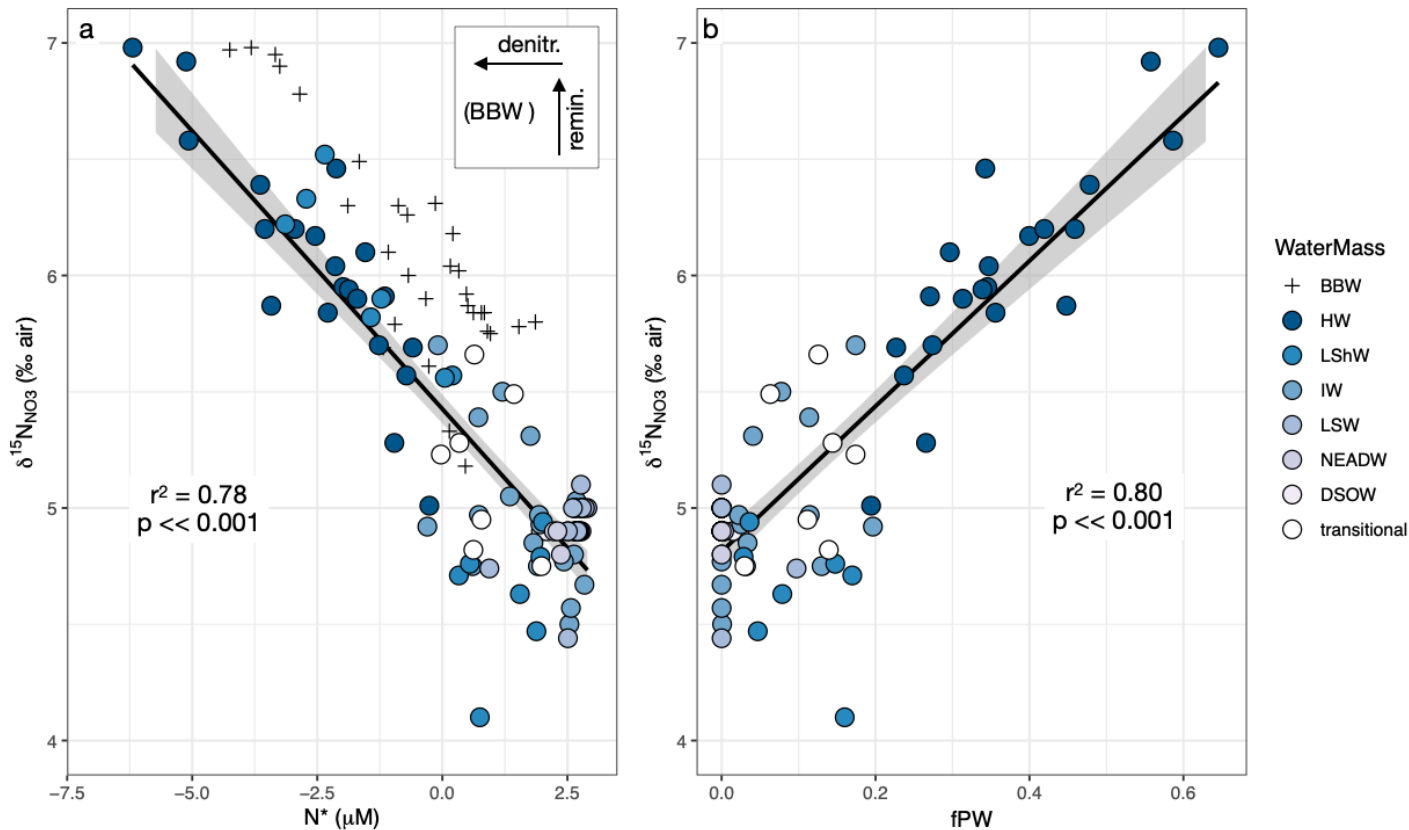
Figure 7: (a) $\delta^{15}N_{NO3}$ vs. N*. Regression line excludes BBW because these waters are affected by remineralization and denitrification as indicated by inset conceptual schematic. See text for explanation. (b) $\delta^{15}N_{NO3}$ vs. fPW, with BBW excluded because it is not

450   **possible to calculate fPW when NO_3^- and PO_4^{3-} ratios are not conserved. Data in both plots are for depths > z_p to avoid the effects of NO_3^- assimilation. Regression lines in both plots are bounded by 95 % confidence intervals.**

455 Relationships for $\delta^{18}O_{NO3}$ vs. N* and $\delta^{18}O_{NO3}$ vs. fPW (Fig. S9) were also highly significant (p << 0.001), but weaker, with r^2 values of 0.38 and 0.35, respectively. The $\delta^{18}O_{NO3}$ vs. fPW relationship predicts $\delta^{18}O_{NO3} = 1.9 \pm 0.1$ ‰ for 100 % Atlantic water and $\delta^{18}O_{NO3} = -0.4 \pm 0.3$ ‰ for 100 % Pacific water, close to previous direct measurements of Atlantic and Pacific water in their respective source regions (Marconi et al., 2015; Brown et al., 2015; Granger et al., 2018).

4 Discussion

4.1 Preservation of NO_3^- isotope signatures

One of the objectives of this paper is to assess the preservation of NO_3^- isotope signatures during transit of Pacific water from the Canadian Arctic Archipelago southward into the Northwest Atlantic. Accurate prediction of $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$ in Atlantic and Pacific source waters based on the relationships in Figs. 7(b) and S9(b) is perhaps the strongest indication that the signatures are well preserved. This preservation likely is facilitated by the oxic conditions in all water masses, as well as the extreme vertical density gradient, which isolates the HW from vertical mixing as it propagates downstream from the Arctic (Tremblay et al., 2015). We also consider that N_2-fixation would act to increase N* while decreasing $\delta^{15}N_{NO3}$, and while there is some evidence of diazotrophy in northern waters (Blais et al., 2012; Sipler et al., 2017; Harding et al., 2018) reported rates are small to negligible, and are therefore unlikely to impact water column $\delta^{15}N_{NO3}$ signatures, as suggested by the data in Fig. 7a.

4.2 Prediction of fraction Pacific water from $\delta^{15}N_{NO3}$

Another objective of this study is to evaluate the use of $\delta^{15}N_{NO3,}$ specifically, as a water mass tracer because it is potentially reflected in the $\delta^{15}N$ of living and detrital biomass. Based on the preceding results, sub-euphotic zone $\delta^{15}N_{NO3}$ may be considered as the product of two-endmember mixing of Atlantic and Pacific water (Fig. 7b). Thus, by inversion of the linear regression in Fig. 7b, fPW may be derived. To achieve normality of model residuals it was necessary to remove data where fPW = 0, as well as three outliers identified in quantile-quantile plots. The resulting final model follows Eq. (7):

$$fPW = 0.23 \pm 0.02 \times \delta^{15}N_{NO3} - 1.06 \pm 0.08, (r^2 = 0.83, p << 0.001, n = 46). \tag{7}$$

The 95 % confidence intervals on fPW predictions range from 0.02 to 0.1, which is roughly equal to the error associated with estimating fPW from NO_3^- vs. PO_4^{3-} data (section 3.3; see also Jones et al., 2003). In other words, $\delta^{15}N_{NO3}$ may be used to estimate fPW about as well as nutrient data. Of course, it is less labour intensive to analyze and use N:P concentrations to estimate fPW, but there are scenarios in which a proxy $\delta^{15}N_{NO3}$ approach is complementary or even essential. For example, $\delta^{15}N_{NO3}$ of a water mass integrates the overall history of nutrient cycling, thereby smoothing out short-lived contingencies inherent to seawater chemistry data. N:P concentration data alone also cannot identify and/or distinguish between the various processes (i.e., remineralization, nitrification, denitrification, diazotrophy) affecting nutrient concentrations and stoichiometric ratios. As a case in point, the two-step process of remineralization followed by sedimentary denitrification in BBW would not be obvious without paired N:P and $\delta^{15}N_{NO3}$ data (Fig. 7a), and invites further investigations to help clarify their proportional

effects on bottom water N:P ratios (Lehmann et al., 2019). We also suggest that isotope data can be used to screen samples that deviate from a two-end member mixing model for the calculation of fPW. Finally, the relationship in equation 7 provides, for the first time, a coherent framework for interpreting $\delta^{15}N$ signatures incorporated into living and paleo-organic materials in the hydrographically complex Northwest Atlantic marine ecosystem.

4.3 Incorporation of $\delta^{15}N_{NO3}$ into "baseline" $\delta^{15}N$ for foodweb and paleoceanographic studies

In isotope ecology and paleoceanography contexts, the term "baseline" $\delta^{15}N$ usually refers to the $\delta^{15}N$ of primary producer (phytoplankton) biomass. This baseline $\delta^{15}N$ signature is propagated to organisms higher up in the food web, overprinted by trophic fractionation, which is often assumed to be about +3.4 ‰ per trophic level but in fact widely variable (Minagawa & Wada, 1984; Vander Zanden and Rasmussen, 2001). The baseline signature may also be altered by bacterial degradation during sinking and sedimentation of particulate organic material (Lehmann et al., 2002; Robinson et al., 2012). In either instance, it is critical to know, or be able to approximate, baseline $\delta^{15}N$ in order to interpret the environmental significance of $\delta^{15}N$ variability recorded in organisms or sediments.

Phytoplankton fractionate against the heavier isotopes of N and O during growth on NO_3^-. Thus, under open-system conditions, the $\delta^{15}N$ of the phytoplankton will be lower than that of the NO_3^-. The isotopic fractionation varies from about 2 - 10 ‰, depending on phytoplankton species and growth conditions (Needoba et al., 2003). Under the semi-closed conditions of NO_3^- drawdown in the ocean euphotic zone, the $\delta^{15}N$ of NO_3^- and phytoplankton both increase, following Rayleigh fractionation kinetics (Fig. 6). If the NO_3^- is exhausted the $\delta^{15}N$ of phytoplankton will, by isotope mass balance, converge on that of the original, unassimilated NO_3^-. Hence, baseline $\delta^{15}N$ reflects the combined influences of $\delta^{15}N_{NO3}$ and the degree of NO_3^- utilization (Altabet et al., 1999; Trull et al., 2008). It is difficult to distinguish between these influences unless $\delta^{15}N$ is measured on paired samples of phytoplankton and NO_3^-. This has not been done anywhere in our study region, apart from studies located in more inshore areas (Ostrom et al., 1997). Nevertheless, it is possible to make general inferences about nutrient drawdown and its effect on baseline $\delta^{15}N$ from a consideration of regional nutrient-plankton bloom dynamics. In the Labrador Sea and Baffin Bay, light is the principal limiting factor to phytoplankton growth for most of the year; however, during the peak summer growth period, NO_3^- becomes co-limiting or limiting as concentrations within the mixed layer are depleted (Harrison and Li, 2008). This applies even in the more light-limited Arctic, where productivity is tightly coupled to NO_3^- availability (Tremblay et al., 2006; Martin et al., 2010). Therefore, the $\delta^{15}N$ of the accumulated phytoplankton biomass should approach that of the pre-assimilated NO_3^-, as identified by the "kink" in Fig. 6. This was confirmed in a study of spring bloom dynamics in the North Water Polynya in northern Baffin Bay, where the $\delta^{15}N$ of phytoplankton converged on the $\delta^{15}N_{NO3}$ of Arctic halocline water (8.3 ‰) as the fraction of unassimilated NO_3^- was drawn down to < 10 % of the pre-bloom concentrations (Tremblay et al., 2006). Considering that northern Baffin Bay is located at a latitude of maximum light limitation, we would predict that the patterns observed there also apply to Pacific-influenced waters of the more southerly Baffin Bay and continental shelves of eastern Canada, except perhaps for inshore and upwelling regions, where NO_3^- would be less limiting. For Atlantic-

influenced waters, NO_3^- is already relatively less limiting than P and Si (Fig. 3). Under these conditions, phytoplankton will be more likely to fractionate against ^{15}N (becoming "lighter"), thereby amplifying the existing differences in $\delta^{15}N_{NO3}$ between the Atlantic and Pacific derived water masses (Fig. 6). Additional studies are needed to determine the effective fractionation, if any, over seasonal and longer time scales.

4.4 Implications for isotope ecology

Results presented here may help to explain previously documented spatial variability in organism $\delta^{15}N$ in the Northwest Atlantic and Baffin Bay regions. For example, Sherwood and Rose (2005) examined bulk $\delta^{15}N$ in invertebrates and fish in waters off Newfoundland and Labrador. Organism $\delta^{15}N$ within each feeding guild was consistently higher, by up to 2.7 ‰, at coastal sites compared to shelf break sites. Part of this offset may be explained by the cross-shelf gradient in fPW, which increases from < 0.02 at the shelf break to > 0.5 at the coast (Pepin et al., 2013; see also the fPW and $\delta^{15}N_{NO3}$ profiles for stations 143, 147, and 154, Fig. S5), and corresponds to an increase of 2.3 ‰ in $\delta^{15}N_{NO3}$ based on Eq. (7). Similarly, in studies located off western Greenland, Hansen, J.H. et al. (2012) and Hedeholm et al. (2012) reported that $\delta^{15}N$ in primary consumers increased by 2 - 3 ‰ over a latitudinal gradient from 60 - 72°N. Subsurface fPW off southern Greenland is essentially zero (Sutherland et al., 2009; Azetsu-Scott et al., 2012), and increases northward as IW and West Greenland Shelf Water mixes with HW, reaching values > 0.4 based on sparse nearby data (e.g., Station ROV7 profile, Fig. S4; Hansen et al., 2012). The corresponding increase in $\delta^{15}N_{NO3}$ is > 2 ‰. This suggests that, in the examples above, the spatial variability in organism $\delta^{15}N$ may be attributed largely to the differential water mass partitioning, rather than spatial variations in the degree of NO_3^- utilization directly at the respective study sites. Finally, Sherwood et al. (2008) examined the bulk $\delta^{15}N$ in the tissues of deep-sea corals collected along the continental slope from Hudson Strait (62°N) to the southwest Grand Banks of Newfoundland (43°N). They found *no* overall change in $\delta^{15}N$ with respect to latitude, but this is consistent with the minimal latitudinal change in fPW (< 0.1) along the path of the slope component of the Labrador Current (Jones et al., 2003). Overall, these examples reiterate the fundamental importance of accounting for variability in baseline $\delta^{15}N$ in isotope ecology studies (e.g., de la Vega et al., 2021). It is not always feasible to measure $\delta^{15}N$ in NO_3^- or primary producers directly, thus we suggest that baseline $\delta^{15}N$ for Canadian Arctic and Northwest Atlantic region may be approximated, to a first degree, from nutrient concentrations and either of the N* or fPW relationships presented in Fig. 7.

4.5 Implications for paleoceanography

Our results also have important implications for regional paleoceanographic interpretations of $\delta^{15}N$ as recorded in sedimentary organic matter and in long-lived biological archives. With respect to sediments, $\delta^{15}N$ is confounded by site-to-site differences in sedimentation rates and diagenetic effects (Robinson et al., 2012). Nevertheless, known spatial patterns track the expected distribution of fPW, with lower values of 4 - 6 ‰ in the central Labrador Sea and Southwest Greenland margin, and higher values 6 - 9 ‰ on the Labrador shelf and northern Baffin Bay (Muzuka & Hillaire-Marcel, 2000; Cormier et al., 2016; Kienast et al., 2020; Limoges et al., 2020). Thus, by extension, downcore trends in $\delta^{15}N$ should reflect advection-related temporal

changes in fPW. Based on arguments in section 3.6, this advection influence is likely to exceed the influence of surface water NO_3^- utilization, particularly where NO_3^- is limiting. This may help to explain why downcore variations in $\delta^{15}N$ are positively correlated with other, biomarker and micropaleontological proxies of Arctic throughflow to Baffin Bay (Cormier et al. 2016; Limoges et al. 2020), confirming the potential of sedimentary $\delta^{15}N$ to quantitatively reconstruct changes in fPW in the past, at least in areas where local changes in nutrient utilization did not play a greater role. This also applies to records of $\delta^{15}N$ recorded in biological archives such as deep-sea corals, which have been shown to track changes in the southward advection of the Labrador Current over the 20th century (Sherwood et al., 2011). We note that, as the N-cycling regimes in the source region and/or in the North Atlantic may have shifted in the past, long term changes in downcore or archival $\delta^{15}N$ may also be influenced by variability in endmember $\delta^{15}N_{NO3}$ signatures (i.e. NO_3^- "inventory-altering" effects; Galbraith et al. 2013), particularly for the Pacific water endmember which is sensitive to primary productivity via sedimentary CPND on the western Arctic shelves. Thus, long term variability in $\delta^{15}N$ should be carefully interpreted in the context of all three influences - nutrient utilization, advection, and changes to endmember $\delta^{15}N_{NO3}$ signatures - together with other lines of paleoceanographic evidence.

5. Conclusions

The flow of Pacific water through the Canadian Arctic Archipelago and into the Northwest Atlantic plays a key role in global thermohaline circulation and biogeochemical cycling. The isotopic composition of NO_3^- presents a new way to track this influence, expanding on the existing N:P stoichiometry approach. Isotopically distinct Pacific water ($\delta^{15}N_{NO3}$ = 8.3 ‰; $\delta^{18}O_{NO3}$ = 0 ‰) travels as a subsurface halocline layer through the Canadian Arctic Archipelago, and onward to Labrador Shelf, with little apparent alteration other than mixing with Atlantic water ($\delta^{15}N_{NO3}$ = 4.8 ‰; $\delta^{18}O_{NO3}$ = 2.0 ‰). The resulting two-endmember mixing of Pacific and Atlantic water is described by a new empirical relationship that may be used to estimate the fraction of Pacific water from $\delta^{15}N_{NO3}$. The deep waters of Baffin Bay are distinctly different, with nutrient inventories showing an imprint of both *in situ* remineralization and sedimentary denitrification. These deep waters are isolated below 500 m and therefore do not influence baseline $\delta^{15}N$ incorporated into primary producer biomass. Rather, baseline $\delta^{15}N$ throughout the Labrador-Baffin region should primarily reflect the fraction of Pacific water, particularly where NO_3^- is the limiting nutrient. Overall, these results provide a new framework for interpreting spatial and temporal patterns of $\delta^{15}N$ in isotope ecology and paleoceanography contexts. In particular they highlight the potential of $\delta^{15}N$ recorded in sedimentary and organic paleo-archives to quantitatively reconstruct changes in fraction of Pacific water in the past.

Supplement link

Supplementary information accompanies this manuscript.

Author contributions

580 OS and SD and MK conceptualized the research with input from all authors. OS, SD and MK collected the samples. SD and NL carried out nitrate isotope analyses. OS, SD, NL, CB and MK analyzed the data. OS, SD and NL prepared the manuscript with contributions from all authors.

Competing interests

The authors declare that they have no conflicts of interest

Acknowledgements

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We thank the captain, crew and science staff of the R/V *Maria S. Merian* MSM45 and CCGS *Amundsen* AMD2016 expeditions. We also thank ArcticNet (a Canadian Network of Centres of Excellence), and Amundsen Science for their in-kind contributions to expedition logistics and scientific equipment. McKenzie Mandich (Dalhousie University) analyzed nutrient concentrations for the MSM45 samples. Jean-Éric Tremblay facilitated sample collection and nutrient analysis during 590 the AMD2016 expedition. Thomas Kuhn (University of Basel) analyzed nitrate isotopes for the AMD2006 samples. Claude Hillaire-Marcel provided input on earlier drafts of the manuscript. Funding for this project was provided by the Canadian Foundation for Innovation, and the Natural Sciences and Engineering Research Council (NSERC) of Canada through a Climate Change and Atmospheric Research (CCAR) grant to Paul Myers et al. (2013-2018), a ship time (STAC) grant to EE et al., a Strategic Projects grant to MK, OS, et al., and Discovery Grants to OS, CB, EE, MK and Claude Hillaire-Marcel.

References

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Aksenov, Y., Bacon, S., Coward, A. C., and Holliday, P.: Polar outflow from the Arctic Ocean: A high resolution model study, J. Mar. Syst. 83, 14-37, https://doi.org/10.1016/j.jmarsys.2010.06.007, 2010.

Altabet, M. A., and Francois, R.: Sedimentary nitrogen isotopic ratio as a recorder for surface ocean nitrate utilization. Glob. Biogeochem. Cy., 8(1), 103-116, https://doi.org/10.1029/93GB03396, 1994.

600 Altabet, M. A., Pilskaln, C., Thunell, R., Pride, C., Sigman, D. M., Chavez, F. and Francois, R.: The nitrogen isotope biogeochemistry of sinking particles from the margin of the eastern North Pacific, Deep. Sea Res. I, 46(4), 655-679, https://doi.org/10.1016/S0967-0637(98)00084-3, 1999.

Anderson, L. A. and Sarmiento, J. L.: Redfield ratios of remineralization determined by nutrient data analysis, Glob. Biogeochem. Cy., 8(1), 65-80, https://doi.org/10.1029/93GB03318, 1994.

605 Arrigo, K., and van Dijken, G. L.: Secular trends in Arctic Ocean net primary production. J. Geophys. Res., 116, C09011, https://doi.org/10.1029/2011JC007151, 2011.

Azetsu-Scott, K., Petrie, B., Yeats, P. and Lee, C.: Composition and fluxes of freshwater through Davis Strait using multiple chemical tracers, J. Geophys. Res., 117, C12011, https://doi.org/10.1029/2012JC008172, 2012.

610   Benetti, M., Reverdin, G., Lique, C., Yashayaev, I., Holliday, N. P., Tynan, E., Torres-Valdes, S., Lherminier, P., Treguer, P., and Sarthou, G.: Composition of freshwater in the spring of 2014 on the southern Labrador shelf and slope, J. Geophys. Res. Oceans, 122, 1102-1121, https://doi.org/10.1002/2016JC012244, 2017.

Blais, M., Tremblay, J.-É., Jungblut, A. D., Gagnon, J., Martin, J., Thaler, M. and Lovejoy, C.: Nitrogen fixation and identification of potential diazotrophs in the Canadian Arctic, Glob. Biogeochem. Cy., 26(3), https://doi.org/10.1029/2011GB004096, 2012.

615   Böhlke, J. K., Mroczkowski, S. J. and Coplen, T. B.: Oxygen isotopes in nitrate: new reference materials for ^{18}O:^{17}O:^{16}O measurements and observations on nitrate-water equilibration, Rapid Commun. Mass Spectrom., 17(16), 1835-1846, https://doi.org/10.1002/rcm.1123, 2003.

Bourke, R. H., Addison, V. G. and Paquette, R. G.: Oceanography of Nares Strait and northern Baffin Bay in 1986 with emphasis on deep and bottom water formation, J. Geophys. Res., 94(C6), 8289-8302, 620   https://doi.org/10.1029/JC094iC06p08289, 1989.

Brandes, J. A. and Devol, A. H.: Isotopic fractionation of oxygen and nitrogen in coastal marine sediments, Geochim. Cosmochim. Acta, 61(9), 1793-1801, https://doi.org/10.1016/S0016-7037(97)00041-0, 1997.

Brown, Z. W., Casciotti, K. L., Pickart, R. S., Swift, J. H. and Arrigo, K. R.: Aspects of the marine nitrogen cycle of the Chukchi Sea shelf and Canada Basin, Deep Sea Res. II, 118, 73-87, https://doi.org/10.1016/j.dsr2.2015.02.009, 2015.

625   Buchwald, C., Santoro, A. E., McIlvin, M. R. and Casciotti, K. L.: Oxygen isotopic composition of nitrate and nitrite produced by nitrifying cocultures and natural marine assemblages, Limnol. Oceanogr., 57(5), 1361-1375, https://doi.org/10.4319/lo.2012.57.5.1361, 2012.

Carmack, E. C. and McLaughlin, F. A.: Towards recognition of physical and geochemical change in Subarctic and Arctic Seas, Prog. Oceanogr., 90(1-4), 90-104, https://doi.org/10.1016/j.pocean.2011.02.007, 2011.

630   Carmack, E. C., Yamamoto-Kawai, M., Haine, T. W. N., Bacon, S., Bluhm, B. A., Lique, C., Melling, H., Polyakov, I. V., Straneo, F., Timmermans, M. L. and Williams, W. J.: Freshwater and its role in the Arctic Marine System: Sources, disposition, storage, export, and physical and biogeochemical consequences in the Arctic and global oceans, J. Geophys. Res. Biogeosci., 121(3), 675–717, https://doi.org/10.1002/2015JG003140, 2016.

Casciotti, K. L., Trull, T. W., Glover, D. M. and Davies, D.: Constraints on nitrogen cycling at the subtropical North Pacific 635   Station ALOHA from isotopic measurements of nitrate and particulate nitrogen, Deep Sea Res. II, 55(14), 1661-1672, https://doi.org/10.1016/j.dsr2.2008.04.017, 2008.

Casciotti, K. L., Sigman, D. M., Hastings, M. G., Böhlke, J. K. and Hilkert, A.: Measurement of the oxygen isotopic composition of nitrate in seawater and freshwater using the denitrifier method, Anal. Chem., 74(19), 4905-4912, https://doi.org/10.1021/ac020113w, 2002.

Chang, B. X. and Devol, A. H.: Seasonal and spatial patterns of sedimentary denitrification rates in the Chukchi sea, Deep Sea Res. II, 56(17), 1339-1350, https://doi.org/10.1016/j.dsr2.2008.10.024, 2009.

Colbourne, E., Holden, J., Senciall, D., Bailey, W., S. Snook and Higdon, J.: Physical Oceanographic Conditions on the Newfoundland and Labrador Shelf during 2015. DFO Can. Sci. Advis. Sec. Res. Doc. 2016/079. v +40 p., 2016.

Cormier. M.-A., Rochon, A., de Vernal, A., and Gélinas, Y.: Multi-proxy study of primary production and paleoceanographical conditions in northern Baffin Bay during the last centuries, Mar. Micropaleontol., 127, 1-10, https://doi.org/10.1016/j.marmicro.2016.07.001, 2020.

Cuny, J., Rhines, P. B., Niiler, P. P. and Bacon, S.: Labrador Sea Boundary Currents and the Fate of the Irminger Sea Water, J. Phys. Oceanogr., 32(2), 627-647, https://doi.org/10.1175/1520-0485(2002)032%3C0627:LSBCAT%3E2.0.CO;2, 2002.

de la Vega, C., Mahaffey, C., Tuerena, R. E., Yurkowski, D. J., Ferguson, S. H., Stenson, G. B., Nordøy, E. S., Haug, T., Biuw, M., Smout, S., Hopkins, J., Tagliabue, A. and Jeffreys, R. M.: Arctic seals as tracers of environmental and ecological change, Limnol. Oceanogr. Lett., 6, 24-32 https://doi.org/10.5285/66AAF3C8-FA58-41DE-8EF1-480A2E125408, 2021.

Deutsch, C., Gruber, N., Key, R. M., Sarmiento, J. L., and Ganachaud, A.: Denitrification and N_2 fixation in the Pacific Ocean. Global Biogeochem. Cy., 15(2), 483-506, https://doi.org/10.1029/2000GB001291, 2001.

Devol, A. H., Codispoti, L. A. and Christensen, J. P.: Summer and winter denitrification rates in western Arctic shelf sediments, Cont. Shelf Res., 17(9), 1029-1050, https://doi.org/10.1016/S0278-4343(97)00003-4, 1997.

Drinkwater, K. F., Belgrano, A., Borja, A., Conversi, A., Edwards, M., Greene, C. H., Otterson, G., Pershing, A. J., and Walker, H.: The Response of Marine Ecosystems to Climate Variability Associated With the North Atlantic Oscillation. Geophys. Monog. - Am. Geophys. Union, 134, 211-234, https://doi.org/10.1029/134GM10, 2003.

Ferland, J., Gosselin, M., and Starr, M.: Environmental control of summer primary production in the Hudson Bay system: The role of stratification. J. Marine Syst. 88, 385–400, https://doi.org/10.1016/j.jmarsys.2011.03.015, 2011.

Fragoso, G. M., Poulton, A. J., Yashayaev, I. M., Head, E. J. H., and Purdie, D. A.: Spring phytoplankton communities of the Labrador Sea (2005-2014): pigment signatures, photophysiology and elemental ratios, Biogeosciences, 14, 1235-1259, https://doi.org/10.5194/bg-14-1235-2017, 2017.

Frajka-Williams, E., and Rhines, P. B.: Physical controls and interannual variability of the Labrador Sea spring phytoplankton bloom in distinct regions, Deep-Sea Res. I, 57, 541-552, https://doi.org/10.1016/j.dsr.2010.01.003, 2010.

Fratantoni, P. S. and Pickart, R. S.: The Western North Atlantic Shelfbreak Current System in Summer, J. Phys. Oceanogr., 37(10), 2509-2533, https://doi.org/10.1175/JPO3123.1, 2007.

Fripiat, F., Declercq, M., Sapart, C. J., Anderson, L. G., Bruechert, V., Deman, F., Fonseca-Batista, D., Humborg, C., Roukaerts, A., Semiletov, I. P. and Dehairs, F.: Influence of the bordering shelves on nutrient distribution in the Arctic halocline inferred from water column nitrate isotopes, Limnol. Oceanogr., 63, 2154-2170, https://doi.org/10.1002/lno.10930, 2018.

Galbraith, E. D., and Kienast, M.: The acceleration of oceanic denitrification during deglacial warming. Nat. Geosci., 6(7), 579-584, https://doi.org/10.1038/ngeo1832, 2013.

Gonfiantini, R.: Stable Isotope Reference Samples for Geochemical and Hydrological Investigations, Int. J. Appl. Radiat. Is., 35(5), 426, https://doi.org/10.1016/0020-708X(84)90059-0, 1984.

Granger, J., and Wankel, S. D.: Isotopic overprinting of nitrification on denitrification as a ubiquitous and unifying feature of environmental nitrogen cycling. Proc. Nat. Acad. Sci., 113(42), E6391-E6400. https://doi.org/10.1073/pnas.1601383113, 2016

Granger, J., Sigman, D. M., Needoba, J. A. and Harrison, P. J.: Coupled nitrogen and oxygen isotope fractionation of nitrate during assimilation by cultures of marine phytoplankton, Limnol. Oceanogr., 49(5), 1763-1773, https://doi.org/10.4319/lo.2004.49.5.1763, 2004.

Granger, J., Sigman, D. M., Lehmann, M. F. and Tortell, P. D.: Nitrogen and oxygen isotope fractionation during dissimilatory nitrate reduction by denitrifying bacteria, Limnol. Oceanogr., 53(6), 2533-2545, https://doi.org/10.4319/lo.2008.53.6.2533, 2008.

Granger, J., Prokopenko, M. G., Sigman, D. M., Mordy, C. W., Morse, Z. M., Morales, L. V., Sambrotto, R. N. and Plessen, B.: Coupled nitrification-denitrification in sediment of the eastern Bering Sea shelf leads to 15N enrichment of fixed N in shelf waters, J. Geophys. Res. Ocean., 116(11), https://doi.org/10.1029/2010JC006751, 2011.

Granger, J., Sigman, D. M., Gagnon, J., Tremblay, J.-E. and Mucci, A.: On the Properties of the Arctic Halocline and Deep Water Masses of the Canada Basin from Nitrate Isotope Ratios, J. Geophys. Res. Ocean., 123(8), 5443–5458, https://doi.org/10.1029/2018JC014110, 2018.

Grasshoff, K.: A simultaneous multiple channel system for nutrient analysis in seawater with analog and digital analog record, Adv. Autom. Anal. Tech. Int. Conf., 11, 133-145, 1969.

Greene, C. H., Meyer-Gutbrod, E., Monger, B. C., McGarry, L. P., Pershing, A. J., Belkin, I. M., Fratantoni, P. S., Mountain, D. G., Pickart, R. S., Proshutinsky, A., Ji, R., Bisagni, J. J., Hakkinen, S. M. A., Haidvogel, D. B., Wang, J., Head, E., Smith, P., Reid, P. C. and Conversi, A.: Remote climate forcing of decadal-scale regime shifts in Northwest Atlantic shelf ecosystems, Limnol. Oceanogr., 58(3), 803-816, https://doi.org/https://doi.org/10.4319/lo.2013.58.3.0803, 2013.

Gruber, N. and Sarmiento, J. L.: Global patterns of marine nitrogen fixation and denitrification, Glob. Biogeochem. Cy., 11(2), 235-266, https://doi.org/10.1029/97GB00077, 1997.

Hansen, J., H., Hedeholm, R. B., Sünksen, K., Christensen, J. T., and Grønkjær, P: Spatial variability of carbon (δ^{13}C) and nitrogen (δ^{15}N) stable isotope ratios in an Arctic marine food web, Mar. Ecol. Prog. Ser. 467, 47-57, https://doi.org/10.3354/meps09945, 2012.

Hansen, M. O., Nielsen, T. G., Stedmon, C. A., and Munkm, P.: Oceanographic regime shift during 1997 in Disko Bay, Western Greenland, Limnol. Oceanogr., 57(2), 634-644, https://doi.org/10.4319/lo.2012.57.2.0634, 2012.

Harding, K., Turk-Kubo, K. A., Sipler, R. E., Mills, M. M., Bronk, D. A. and Zehr, J. P.: Symbiotic unicellular cyanobacteria fix nitrogen in the Arctic Ocean, Proc. Natl. Acad. Sci., 115(52), 13371-13375, https://doi.org/10.1073/pnas.1813658115, 2018.

Harrison, W. G., and Li, W. K. W.: Phytoplankton Growth and Regulation in the Labrador Sea: Light and Nutrient Limitation, J. Northw. Atl. Fish. Sci., Vol. 39: 71-82, https://doi.org/10.2960/J.v39.m592, 2008.

Harrison, W. G., Platt, T., and Irwin, B.: Primary Production and Nutrient Assimilation by Natural Phytoplankton Populations
710 of the Eastern Canadian Arctic, Can. J. Fish. Aquat. Sci., 39, 335-345, https://doi.org/10.1139/f82-046, 1982.

Harrison, W. G., Børsheim, K. Y., Li, W. K., Maillet, G. L., Pepin, P., Sakshaug, E., Skogen, M. D., and Yeats, P. A.: Phytoplankton production and growth regulation in the Subarctic North Atlantic: A comparative study of the Labrador Sea-Labrador/Newfoundland shelves and Barents/Norwegian/Greenland seas and shelves. Prog. Oceanogr., 114, 26-45, https://doi.org/10.1016/j.pocean.2013.05.003, 2013.

715 Hedeholm, R., Grønkjær, P., and Rysgaard, S.: Feeding ecology of capelin (*Mallotus villosus* Muller) in West Greenland waters, Polar Biol., 35, 1533-1543, https://doi.org/10.1007/s00300-012-1193-4, 2012.

Jakobsen, P. K., Ribergaard, M. H., Quadfasel, D., Schmith, T., and Hughes, C. W.: Near-surface circulation in the northern North Atlantic as inferred from Lagrangian drifters: Variability from the mesoscale to interannual, J. Geophys. Res., 108(C8), 3251, https://doi.org/10.1029/2002JC001554, 2003.

720 Jenkins, W. J., Smethie, W. M., Boyle, E. A. and Cutter, G. A.: Water mass analysis for the U.S. GEOTRACES (GA03) North Atlantic sections, Deep Sea Res. II, 116, 6-20, https://doi.org/https://doi.org/10.1016/j.dsr2.2014.11.018, 2015.

Jones, E. P., Anderson, L. G. and Swift, J. H.: Distribution of Atlantic and Pacific waters in the upper Arctic Ocean: Implications for circulation, Geophys. Res. Lett., 25(6), 765-768, https://doi.org/10.1029/98GL00464, 1998.

Jones, E. P.: Tracing Pacific water in the North Atlantic Ocean, J. Geophys. Res., 108(C4), 3116,
725 https://doi.org/10.1029/2001JC001141, 2003.

Kelley, D., and Richards, C.: oce: Analysis of Oceanographic Data. R package version 0.9-22, 2017.

Kienast, M., Davin, S., Doering, K., Hebbeln, D., Kienast, S., Lehmann, N., Schneider, R., Sherwood, O., and Weiser, J.: Isotopic evidence for changes in the origin and cycling of nitrogen in the Labrador Sea during the last 8,000 years, EGU General Assembly Online, 4-8 May 2020, EGU2020-12134, https://doi.org/10.5194/egusphere-egu2020-12134, 2020.

730 Klein, B., LeBlanc, B., Mei, Z.-P., Beret, R., Michaud, J., Mundy, C.-J., von Quillfeldt, C. H., Garneau, M.-È., Roy, S. and Gratton, Y.: Phytoplankton biomass, production and potential export in the North Water, Deep Sea Res. II, 49(22-23), 4983-5002, https://doi.org/10.1016/S0967-0645(02)00174-1, 2002.

Lalande, C., Forest, A., Barber, D. G., Gratton, Y. and Fortier, L.: Variability in the annual cycle of vertical particulate organic carbon export on Arctic shelves: Contrasting the Laptev Sea, Northern Baffin Bay and the Beaufort Sea, Cont. Shelf Res.,
735 29(17), 2157-2165, https://doi.org/10.1016/j.csr.2009.08.009, 2009.

Lehmann, M. F., Bernasconi, S. F., Barbieri, A., and McKenzie, J. A.: Preservation of organic matter and alteration of its carbon and nitrogen isotope composition during simulated and in situ early sedimentary diagenesis, Geochim. Cosmochim. Acta, 66(20), 3573-3584, https://doi.org/10.1016/S0016-7037(02)00968-7, 2002.

Lehmann, M. F., Sigman, D. M., McCorkle, D. C., Brunelle, B. G., Hoffmann, S., Kienast, M., Cane, G. and Clement, J.:

740 Origin of the deep Bering Sea nitrate deficit: Constraints from the nitrogen and oxygen isotopic composition of water column nitrate and benthic nitrate fluxes, Glob. Biogeochem. Cy., 19(4), GB4005, https://doi.org/10.1029/2005GB002508, 2005.

Lehmann, M. F., Sigman, D. M., McCorkle, D. C., Granger, J., Hoffmann, S., Cane, G., and Brunelle, B. B.: The distribution of nitrate $^{15}N/^{14}N$ in marine sediments and the impact of benthic nitrogen loss on the isotopic composition of oceanic nitrate, Geochim. Cosmochim. Acta, 71(22), 5384-5404, https://doi.org/10.1016/j.gca.2007.07.025, 2007.

745 Lehmann, N., Kienast, M., Granger, J., Bourbonnais, A., Altabet, M. A. and Tremblay, J.-É.: Remote western Arctic nutrients fuel remineralization in deep Baffin Bay, Global Biogeochem. Cy., 33, 649-667, https://doi.org/10.1029/2018GB006134, 2019.

Limoges, A., Weckström, K., Ribeiro, S., Georgiadis, E., Hansen, K. E., Martinez, P., Seidenkrantz, M.-S., Giraudeau, J., Crosta, X, and Massé, G.: Learning from the past: Impact of the Arctic Oscillation on sea ice and marine productivity off

750 northwest Greenland over the last 9,000 years, Glob. Change Biol., 26, 6767-6786, https://doi.org/10.1111/gcb.15334, 2020.

Marconi, D., Weigand, M. A., Rafter, P. A., McIlvin, M. R., Forbes, M., Casciotti, K. L. and Sigman, D. M.: Nitrate isotope distributions on the US GEOTRACES North Atlantic cross-basin section: Signals of polar nitrate sources and low latitude nitrogen cycling, Mar. Chem., 177, 143-156, https://doi.org/10.1016/j.marchem.2015.06.007, 2015.

Martin, J., Tremblay, J.-É., Gagnon, J., Tremblay, G., Lapoussière, A., Jose, C., Poulin, M., Gosselin, M., Gratton, Y. and

755 Michel, C.: Prevalence, structure and properties of subsurface chlorophyll maxima in Canadian Arctic waters, Mar. Ecol. Prog. Ser., 412, 69-84, https://doi.org/10.3354/meps08666, 2010.

McIlvin, M. R. and Casciotti, K. L.: Fully automated system for stable isotopic analyses of dissolved nitrous oxide at natural abundance levels, Limnol. Oceanogr. Methods, 8, 54-66, https://doi.org/10.4319/lom.2010.8.54, 2010.

McIlvin, M. R. and Casciotti, K. L.: Technical Updates to the Bacterial Method for Nitrate Isotopic Analyses, Anal. Chem.,

760 83(5), 1850-1856, https://doi.org/10.1021/ac1028984, 2011.

McLaughlin, F. A., Carmack, E. C., Macdonald, R. W. and Bishop, J. K. B.: Physical and geochemical properties across the Atlantic/Pacific water mass front in the southern Canadian Basin, J. Geophys. Res. Ocean., 101(C1), 1183-1197, https://doi.org/10.1029/95JC02634, 1996.

McLaughlin, F. A., Carmack, E. C., Macdonald, R. W., Melling, H., Swift, J. H., Wheeler, P. A., Sherr, B. F., and Sherr, E.

765 B.: The joint roles of Pacific and Atlantic-origin waters in the Canada Basin, 1997–1998, Deep Sea Res. I, 51(1), 107-128, https://doi.org/10.1016/j.dsr.2003.09.010, 2004.

Michel, C., Gosselin, M., and Nozais, C.: Preferential sinking export of biogenic silica during the spring and summer in the North Water Polynya (northern Baffin Bay): Temperature or biological control? J. Geophys. Res., 107(C7), 3064, https://doi.org/10.1029/2000JC000408, 2002.

770 Mills, M. M., Brown, Z. W., Lowry, K. E., van Dijken, G. L., Becker, S., Pal,S., Benitez-Nelson, C. R., Downer, M. M., Strong, A. L., Swift, J. H., Pickart, R. S., and Arrigo, K. R.: Impacts of low phytoplankton NO_3^-:PO_4^{3-} utilization ratios over the Chukchi Shelf, Arctic Ocean, Deep Sea Res. II, 118(A), 105-121, https://doi.org/10.1016/j.dsr2.2015.02.007, 2015.

Minagawa, M., and Wada, E.: Stepwise enrichment of ^{15}N along food chains: Further evidence and the relation between $\delta^{15}N$ and animal age, Geochim. Cosmochim. Acta, 48(5), 1135-1140, https://doi.org/10.1016/0016-7037(84)90204-7, 1984.

775 Münchow, A., Falkner, K. K. and Melling, H.: Baffin island and west Greenland current systems in northern Baffin bay, Prog. Oceanogr., 132, 305-317, https://doi.org/10.1016/j.pocean.2014.04.001, 2015.

Muzuka, A. N., and Hillaire-Marcel, C.: Burial rates of organic matter along the eastern Canadian margin and stable isotope constraints on its origin and diagenetic evolution. *Mar. Geol.*, 160(3-4), 251-270, https://doi.org/10.1016/S0025-3227(99)00022-5, 1999.

780 Needoba, J. A., Waser, N. A., Harrison, P. J., and Calvert, S. E.: Nitrogen isotope fractionation in 12 species of marine phytoplankton during growth on nitrate, Mar. Ecol. Prog. Ser., 255, 81-91 https://doi.org/10.3354/meps255081, 2003.

Olsen, A., Lange, N., Key, R., Tanhua, T., Alvarez M., et al.: GLODAPv2.2019 - an update of GLODAPv2. Earth Syst. Sci. Data, 2019, 11(3), 1437-1461. https://doi.org/10.5194/essd-11-1437-2019, 2020

Ostrom, N. E., Macko, S. A., Deibel, D. and Thompson, R. J.: Seasonal variation in the stable carbon and nitrogen isotope

785 biogeochemistry of a coastal cold ocean environment, Geochim. Cosmochim. Acta, 61(14), 2929-2942, https://doi.org/10.1016/S0016-7037(97)00131-2, 1997.

Pepin, P., Maillet, G., Fraser, S., Shears, T., and Redmond, G.: Optical, chemical, and biological oceanographic conditions on the Newfoundland and Labrador Shelf during 2011-12. DFO Can. Sci. Advis. Sec. Res. Doc. 2013/051. v + 38 p, 2013.

Peters, B. D., Lam, P. J., and Casciotti, K. L.: Nitrogen and oxygen isotope measurements of nitrate along the US

790 GEOTRACES Eastern Pacific Zonal Transect (GP16) yield insights into nitrate supply, remineralization, and water mass transport, Mar. Chem., 201, 137-150, https://doi.org/10.1016/j.marchem.2017.09.009, 2018.

Prinsenberg, S. J. and Hamilton, J.: Monitoring the volume, freshwater and heat fluxes passing through Lancaster sound in the Canadian arctic archipelago, Atmos. Ocean, 43(1), 1-22, https://doi.org/10.3137/ao.430101, 2005.

Rafter, P. A., and Sigman, D. M.: Spatial distribution and temporal variation of nitrate nitrogen and oxygen isotopes in the

795 upper equatorial Pacific Ocean, Limnol. Oceanogr. 61, 14-31, https://doi.org/10.1002/lno.10152, 2016.

Redfield, A. C., Ketchum, B. H. and Richard, F. A.: The influence of organisms on the composition of seawater, in The Sea, edited by M. N. Hill, pp. 26-77, Wiley-Interscience, New York, 1963.

Robinson, R. S., Kienast, M., Luiza Albuquerque, A., Altabet, M., Contreras, S., De Pol Holz, R., Dubois, N., Francois, R., Galbraith, E., Hsu, T.-C., Ivanochko, T., Jaccard, S., Kao, S.-J., Kiefer, T., Kienast, S., Lehmann, M., Martinez, P., McCarthy,

800 M., Möbius, J., Pedersen, T., Quan, T. M., Ryabenko, E., Schmittner, A., Schneider, R., Schneider-Mor, A., Shigemitsu, M., Sinclair, D., Somes, C., Studer, A., Thunell, R., and Yang, J.-Y: A review of nitrogen isotopic alteration in marine sediments, Paleoceanography, 27, PA4203, https://doi.org/10.1029/2012PA002321, 2012.

Rysgaard, S., Boone, W., Carlson, D., Sejr, M. K., Bendtsen, J., Juul-Pedersen, T., Lund, H., Meire, L. and Mortensen, J.: An Updated View on Water Masses on the pan-West Greenland Continental Shelf and Their Link to Proglacial Fjords, J. Geophys.

805 Res. Ocean., 125(2), https://doi.org/10.1029/2019JC015564, 2020.

Sherwood, O. A., Lehmann, M. F., Schubert, C. J., Scott, D. B., and McCarthy, M. D.: Nutrient regime shift in the western North Atlantic indicated by compound-specific δ^{15}N of deep-sea gorgonian corals, Proc. Nat. Acad. Sci., 108(3), 1011-1015, https://doi.org/10.1073/pnas.1004904108, 2011.

Sherwood, O. A., Jamieson, R. E., Edinger, E. N., and Wareham, V. E.: Stable C and N isotopic composition of cold-water corals from the Newfoundland and Labrador continental slope: Examination of trophic, depth and spatial effects, Deep Sea Res. I, 55(10), 1392-1402, https://doi.org/10.1016/j.dsr.2008.05.013, 2008.

Sherwood, G. D., and Rose, G. A.: Stable isotope analysis of some representative fish and invertebrates of the Newfoundland and Labrador continental shelf food web, Estuar. Coast. Shelf S., 63(4), 537-549, https://doi.org/10.1016/j.ecss.2004.12.010. 2005.

Sigman, D. M., Casciotti, K. L., Andreani, M., Barford, C., Galanter, M. and Böhlke, J. K.: A bacterial method for the nitrogen isotopic analysis of nitrate in seawater and freshwater, Anal. Chem., 73(17), 4145-4153, https://doi.org/10.1021/ac010088e, 2001.

Sigman, D. M., Granger, J., DiFiore, P. J., Lehmann, M. M., Ho, R., Cane, G. and van Geen, A.: Coupled nitrogen and oxygen isotope measurements of nitrate along the eastern North Pacific margin, Glob. Biogeochem. Cy., 19(4), GB4022, https://doi.org/10.1029/2005GB002458, 2005.

Sigman, D. M., DiFiore, P. J., Hain, M. P., Deutsch, C., Wang, Y., Karl, D. M., Knapp, A. N., Lehmann, M. F. and Pantoja, S.: The dual isotopes of deep nitrate as a constraint on the cycle and budget of oceanic fixed nitrogen, Deep Sea Res. I, 56(9), 1419-1439, https://doi.org/10.1016/j.dsr.2009.04.007, 2009.

Sipler, R. E., Gong, D., Baer, S. E., Sanderson, M. P., Roberts, Q. N., Mulholland, M. R. and Bronk, D. A.: Preliminary estimates of the contribution of Arctic nitrogen fixation to the global nitrogen budget, Limnol. Oceanogr. Lett., 2(5), 159-166, https://doi.org/10.1002/lol2.10046, 2017.

Steele, M., Morison, J., Ermold, W., Rigor, I., Ortmeyer, M. and Shimada, K.: Circulation of summer Pacific halocline water in the Arctic Ocean, J. Geophys. Res. Ocean., 109(C2), C02027, https://doi.org/10.1029/2003JC002009, 2004.

Stramma, L., Kieke, D., Rhein, M., Schott, F., Yashayaev, I., and Koltermann, K. P.: Deep water changes at the western boundary of the subpolar North Atlantic during 1996 to 2001, Deep Sea Res. I, 51(8), 1033-1056, https://doi.org/10.1016/j.dsr.2004.04.001, 2004.

Straneo, F. and Saucier, F.: The outflow from Hudson Strait and its contribution to the Labrador Current, Deep Sea Res. I, 55(8), 926-946, https://doi.org/10.1016/j.dsr.2008.03.012, 2008.

Sutcliffe Jr., W. H., Loucks, R. H., Drinkwater, K. F. and Coote, A. R.: Nutrient Flex onto the Labrador Shelf from Hudson Strait and its Biological Consequences, Can. J. Fish. Aquat. Sci., 40(10), 1692-1701, https://doi.org/10.1139/f83-196, 1983.

Sutherland, D. A., Pickart, R. S., Peter Jones, E., Azetsu-Scott, K., Jane Eert, A. and Ólafsson, J.: Freshwater composition of the waters off southeast Greenland and their link to the Arctic Ocean, J. Geophys. Res., 114, C05020, https://doi.org/10.1029/2008JC004808, 2009.

Top, Z., Clarke, W. B., Eismont, W. C. and Jones, E. P.: Radiogenic helium in Baffin Bay bottom water, J. Mar. Res., 38(3),

840   435-452, 1980.

Tang, C. C., Ross, C. K., Yao, T., Petrie, B., DeTracey, B. M., and Dunlap, E.: The circulation, water masses and sea-ice of Baffin Bay. Prog. Oceanogr., 63(4), 183-228, https://doi.org/10.1016/j.pocean.2004.09.005, 2004.

Torres-Valdés, S., Tsubouchi, T., Bacon, S., Naveira-Garabato, A. C., Sanders, R., McLaughlin, F. A., Petrie, B., Kattner, G., Azetsu-Scott, K. and Whitledge, T. E.: Export of nutrients from the Arctic Ocean, J. Geophys. Res. Oceans, 118, 1625-1644,

845   https://doi.org/10.1002/jgrc.20063, 2013.

Townsend, D. W., Pettigrew, N. R., Thomas, M. A., Neary, M. G., McGillicuddy, D. J. and Donnell, J. O.: Water masses and nutrient sources to the Gulf of Maine, J. Mar. Res., 141(73), 93-122, https://doi.org/10.1357/002224015815848811, 2015.

Tremblay, J.-É., Anderson, L. G., Matrai, P., Coupel, P., Bélanger, S., Michel, C. and Reigstad, M.: Global and regional drivers of nutrient supply, primary production and CO2 drawdown in the changing Arctic Ocean, Prog. Oceanogr., 139, 171-196,

850   https://doi.org/10.1016/j.pocean.2015.08.009, 2015.

Tremblay, J.-É., Gratton, Y., Carmack, E. C., Payne, C. D. and Price, N. M.: Impact of the large-scale Arctic circulation and the North Water Polynya on nutrient inventories in Baffin Bay, J. Geophys. Res., 107(C8), https://doi.org/10.1029/2000JC000595, 2002.

Tremblay, J.-É., Hattori, H., Michel, C., Ringuette, M., Mei, Z. P., Lovejoy, C., Fortier, L., Hobson, K. A., Amiel, D. and

855   Cochran, K.: Trophic structure and pathways of biogenic carbon flow in the eastern North Water Polynya, Prog. Oceanogr., 71(2-4), 402-425, https://doi.org/10.1016/j.pocean.2006.10.006, 2006.

Trull, T. W., Davies, D., and Casciotti, K.: Insights into nutrient assimilation and export in naturally iron-fertilized waters of the Southern Ocean from nitrogen, carbon and oxygen isotopes. Deep-Sea Res. II, 55, 820-840, https://doi.org/10.1016/j.dsr2.2007.12.035, 2008.

860   Vander Zanden, M. J.. and Rasmussen, J. B.: Variation in $\delta^{15}N$ and $\delta^{13}C$ trophic fractionation: Implications for aquatic food web studies, Limnol. Oceanogr. 46(8), 2061-2066, https://doi.org/10.4319/lo.2001.46.8.2061, 2001.

Wallace, D. W. R.: A study of the ventilation of Arctic waters using chlorofluoromethanes as tracers, Dalhousie University, Halifax NS, Canada., 1985.

Weiss, R. F.: The solubility of nitrogen, oxygen and argon in water and seawater, Deep Sea Res. 17(4), 721-735,

865   https://doi.org/10.1016/0011-7471(70)90037-9, 1970.

Woodgate, R. A., Weingartner, T. J., and Lindsay, R.: Observed increases in Bering Strait oceanic fluxes from the Pacific to the Arctic from 2001 to 2011 and their impacts on the Arctic Ocean water column, Geophys. Res. Lett., 39(24), https://doi.org/10.1029/2012GL054092, 2012.

Yamamoto-Kawai, M., Carmack, E. and McLaughlin, F.: Nitrogen balance and Arctic throughflow, Nature, 443(7107), 43,

870   https://doi.org/10.1038/443043a, 2006.

Yamamoto-Kawai, M., McLaughlin, F. A., Carmack, E. C., Nishino, S. and Shimada, K.: Freshwater budget of the Canada Basin, Arctic Ocean, from salinity, δ^{18}O, and nutrients, J. Geophys. Res. Ocean., 113(1), C01007, https://doi.org/10.1029/2006JC003858, 2008.

Yashayaev, I. and Loder, J. W.: Recurrent replenishment of Labrador Sea Water and associated decadal-scale variability, J. Geophys. Res. Ocean., 121(11), 8095-8114, https://doi.org/10.1002/2016JC012046, 2016.