



# Deep-water inflow event increases sedimentary phosphorus release on a multi-year scale

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**Abstract.** Phosphorus fertilisation (eutrophication) is expanding oxygen depletion in coastal systems worldwide. Under low-oxygen bottom-water conditions, phosphorus release from the sediment is elevated which further stimulates primary production. It is commonly assumed that re-oxygenation could break this ‘vicious cycle’ by increasing sedimentary phosphorus retention. Recently, a deep-water inflow into the Baltic Sea created a natural in-situ experiment that allowed us to investigate if temporary re-oxygenation stimulates sedimentary retention of dissolved inorganic phosphorus (DIP). Surprisingly, during this three-year-long study, we observed a transient but considerable increase, rather than a decrease, in the sediment efflux of DIP and other dissolved biogenic compounds. This suggested that the oxygenated inflow elevated the organic matter degradation in the sediment. As a result, the net sedimentary DIP release per m<sup>2</sup> was 35-70% higher over the years following the re-oxygenation than before. In contrast to previous assumptions, our results show that inflows of oxygenated water to anoxic bottom waters can increase the sedimentary phosphorus release.

## 1 Introduction

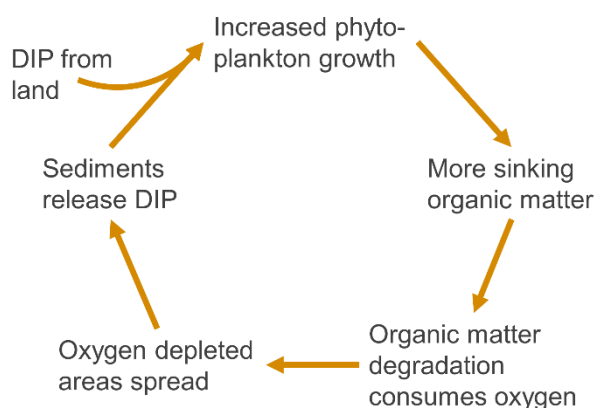
25 Eutrophication is one of the main causes of oxygen depletion in coastal systems worldwide (Breitburg et al., 2018; Diaz and Rosenberg, 2008). Excess input of nutrients from land stimulates primary production, resulting in a higher delivery of organic matter to deeper coastal waters and the seafloor (Breitburg et al., 2018; Middelburg and Levin, 2009). Oxygen is consumed as this organic matter is degraded and in severe cases, anoxia can develop. Depletion of oxygen can lead to decreasing biodiversity and thereby a loss of important ecosystem functions and services (Breitburg et al., 2018; Middelburg and Levin, 2009). One



30 important example is food supply, since many coastal areas are key sites for fishing and aquaculture (Diaz and Rosenberg, 2008).

The Baltic Sea is strongly affected by eutrophication and oxygen depletion. The area has been argued to be a “time machine” for future coastal areas worldwide, due to its long history of anthropogenic disturbances and further impacts of climate change (Reusch et al., 2018). As such, understanding the feedback mechanisms between eutrophication and oxygen depletion in the  
35 Baltic Sea can help us understand the future of our coastal zones. The central parts of the Baltic Sea have been naturally oxygen-depleted for thousands of years due to a strong water column stratification, long residence time and limited deep-water renewal (Zillén et al., 2008). However, high inputs of nutrients from land have severely worsened the situation and the areal extent of benthic oxygen depletion has increased 10 times in size over the last 115 years (Carstensen et al., 2014). Despite improvements in wastewater treatment and decreased nutrient input from agriculture, the oxygen-depleted area in the Baltic  
40 Sea has not decreased in size (Reusch et al., 2018).

These low-oxygen conditions in the Baltic Sea are largely sustained by enhanced recycling of dissolved inorganic phosphorus (DIP, mainly phosphate) from sediments with oxygen-depleted overlying water (Vahtera et al., 2007). This ‘vicious cycle’ (Fig. 1; Vahtera et al., 2007) has been suggested to drive enhanced recycling of DIP; high sedimentary release of DIP fuels extensive growth of cyanobacteria and other phytoplankton which consume oxygen as they are degraded, resulting in increased  
45 oxygen depletion and further release of DIP. The elevated release of DIP from oxygen-depleted sediments is mainly a consequence of two factors. Firstly, under low-oxygen conditions there is a preferential remineralisation of phosphorus (P) in organic matter relative to nitrogen and carbon (Jilbert et al., 2011). Secondly, iron and manganese oxides, which adsorb and retain DIP in the sediment under oxic conditions, are reduced and solubilised in anoxic environments (Jilbert and Slomp, 2013). The DIP is then released back to the water column, instead of being retained in the sediment where it eventually could  
50 be buried as authigenic P minerals (Slomp et al., 1996). Due to high sedimentary DIP concentrations there is some formation of authigenic P phases also in the anoxic areas of the Baltic Sea (Jilbert and Slomp, 2013), but the overall burial efficiency of P in this environment is low (0.2 – 12 %, compared to > 14 % in oxic parts of the Baltic Sea; Viktorsson et al., 2013).





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**Figure 1.** A simplified version of the 'vicious cycle' in the Baltic Sea. The figure, based on Vahtera et al. (2007), describes the feedback mechanism between the release of dissolved inorganic phosphorus (DIP) and oxygen depletion. Briefly, DIP from land fuels phytoplankton growth and thereby elevates the export of organic matter to the seafloor. When the organic matter is degraded, oxygen is depleted. The release of DIP is elevated from oxygen-depleted sediment and further fuels phytoplankton growth.

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It is commonly assumed that re-oxygenation of anoxic sediments would reduce the release of DIP (Jilbert and Slomp, 2013; Ruttenberg, 2003), which could break the vicious cycle. Intrusions of large volumes of salty, oxygenated water from the North Sea (Major Baltic Inflows; MBIs), can temporarily bring new oxygen to the anoxic areas of the Baltic Sea (Liblik et al., 2018). In the winter of 2014-2015, the largest MBI since 1951 occurred and previously long-term anoxic areas were oxygenated (Liblik et al., 2018). Studies conducted shortly after the inflow did indeed show a lowered DIP release from the sediment, but the total DIP release only decreased with 5-23 % on a basin-wide scale (Hall et al., 2017; Sommer et al., 2017) and sequestration of P in the surface sediment was low (Hermans et al., 2019). It has so far been unknown how the sedimentary P dynamics evolved after anoxia was re-established.

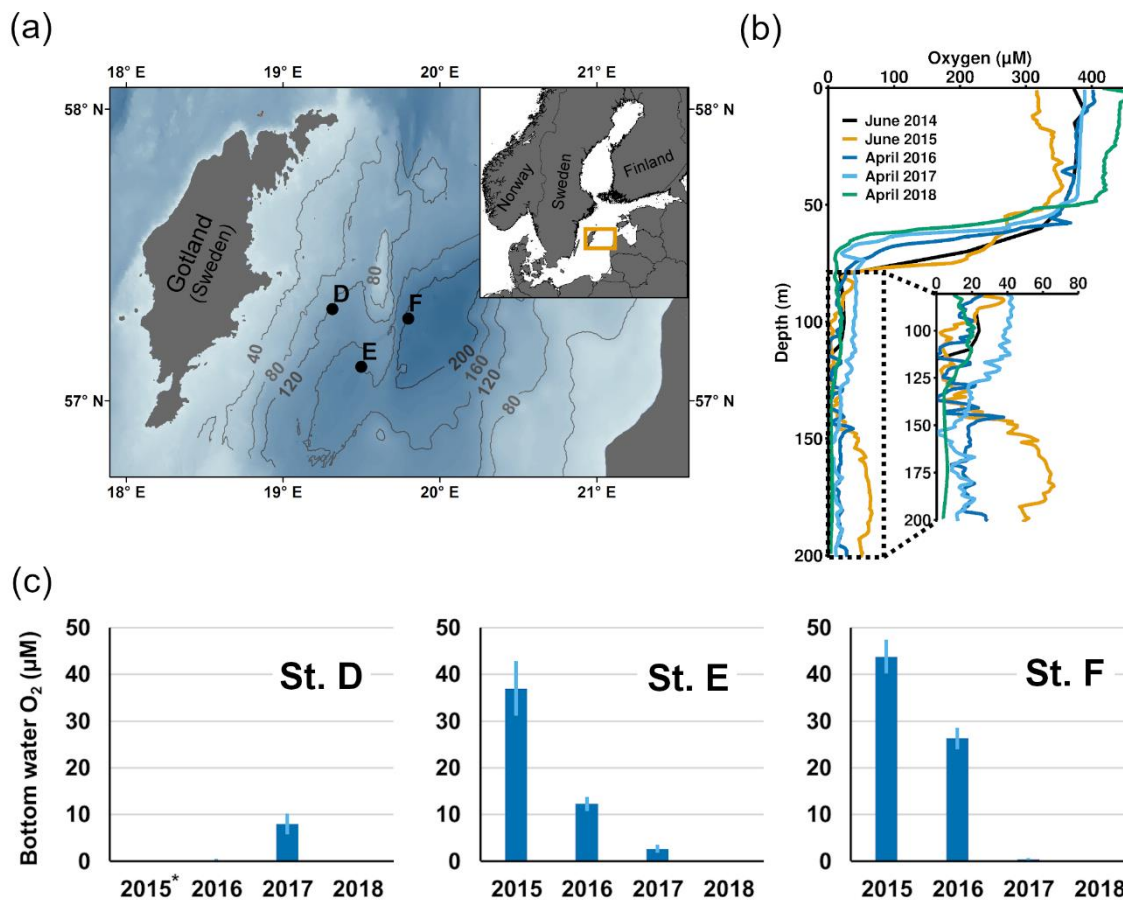
Here we present a time series study from the Eastern Gotland Basin (EGB, Fig. 2a) in the Baltic Sea, a long-term anoxic basin that was temporarily oxygenated by the 2014-2015 MBI. To investigate the effect of the oxygenation event on the sedimentary P cycle, we conducted in situ measurements of sediment-water fluxes and collected pore-water and solid-phase samples during three years (2016-2018). By combining our results with data from 2008 (Nilsson et al., 2019; Viktorsson et al., 2013), 2010 (Nilsson et al., 2019; Viktorsson et al., 2013) and 2015 (Hall et al., 2017), we construct a unique time series from prior to until after the inflow and show that contrary to previous assumptions, the inflow of oxygenated water to this anoxic area increased the sedimentary phosphorus release on a multi-year scale.

## 2 Materials and Methods

### 2.1 Site characteristics

The EGB is the largest sub-basin of the Baltic Sea, situated between Latvia and the Swedish island Gotland (Fig. 2a). The water column is normally hypoxic below the halocline at 60-80 m depth and anoxic at the seafloor (maximum depth 249 m). The MBI that occurred in the winter of 2014-2015 reached the EGB in March 2015, resulting in a more oxygenated but still hypoxic (40-60  $\mu\text{M}$ ) water layer below 130-140 m depth (Fig. 2b,c; Hall et al., 2017; Liblik et al., 2018). The oxygen had been consumed by the end of 2015, but a new inflow oxygenated the area again between January and May 2016 (Liblik et al., 2018). In 2017, several smaller inflows also brought oxygen to the 80-170 m depth layer (Liblik et al., 2018).

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**Figure 2.** Description of sampling sites and oxygen conditions. (a) Map of the sampling sites in the Eastern Gotland Basin, Baltic Sea. Bathymetric data from the Baltic Sea Bathymetry Database (v. 0.9.3), the map was created using the software ArcMap™ (v. 10.6) by Esri. (b) Water-column profiles of oxygen. Data from SMHI (SMHI, 2019; year 2014, station BY15), Hall et al. (2017; year 2015, station F) and van de Velde et al. (2020b; years 2016–2018, station F). (c) Bottom-water oxygen ( $\text{O}_2$ ) concentrations measured by the benthic chamber lander (average  $\pm$  standard deviation). The detection limit was  $0.5 \mu\text{M}$ . Data from Hall et al. (2017) and van de Velde et al. (2020b). \*no measurement.

During yearly expeditions to the EGB in April 2016–2018, we visited three stations along a depth transect (Fig. 2a, Table A1). The stations are situated below the halocline and were oxygenated by inflows at different points during the sampling period. Station D (130 m) is situated at the top boundary of the oxygenated layer. It was anoxic during the sampling in 2016 (Fig. 2c), but monitoring data from the Swedish Meteorological and Hydrological Institute (SMHI; Fig. A1; SMHI, 2019) and previous studies (Dellwig et al., 2018; Hermans et al., 2019; Sommer et al., 2017) from the eastern part of the EGB suggest that this station might have been temporarily oxygenated by the MBI prior to our visit. Station D received some oxygen with the smaller inflow in 2017, but was anoxic again in 2018. Station E (170 m) and F (210 m) were oxygenated by the MBI (Fig. 2c). The



oxygen at station F had been consumed already in 2017, while it decreased stepwise at station E. Hydrogen sulphide was only detected in the bottom water at station D in 2016 (Broman et al., 2020; Marzocchi et al., 2018). For clarity, the remainder of this paper will focus on the results from station D and F. These stations represent the least and most affected sites, respectively. The results from station E, which are consistent with the results from station F, are presented in Figs. A2-A4.

## 2.2 Water-column and benthic in-situ flux measurements

Upon arrival at the stations, a CTD (SBE 911, Sea-Bird Scientific) with a high-accuracy oxygen sensor (SBE 43, Sea-Bird Scientific) was deployed to record water-column profiles of salinity, temperature and oxygen (van de Velde et al., 2020b). Sediment-water fluxes of nutrients and dissolved inorganic carbon (DIC) were subsequently measured using the Gothenburg benthic chamber lander (Kononets et al., 2020). Deployment of the lander during these samplings has been described in detail elsewhere (Hall et al., 2017; van de Velde et al., 2020b). Briefly, the lander carried four incubation chambers incubating 400 cm<sup>2</sup> of sediment with overlying water each. At pre-set times; nine syringes per chamber sampled the overlying water. Sensors monitored conductivity, temperature and oxygen inside and outside the chambers. Chambers that initially contained oxygen did not become anoxic over the course of the incubations. Chamber volumes were calculated from the decrease in salinity following the injection of a small, known volume of Milli-Q water. Sediment and water were incubated for 14 hours. Immediately after lander recovery, the syringes were emptied and samples were filtered through pre-rinsed 0.45 µm cellulose acetate filters (Sartorius). The lander was deployed twice per station except for two occasions (F 2017, E 2018) when time constraints only allowed one deployment. Calculation of benthic nutrient and DIC fluxes are described in van de Velde et al. (2020b). In short, a linear regression model, simple or quadratic, was fitted to data of the concentration change over time in the chamber water. After verifying model and data quality, the flux was calculated by multiplying the chamber water height with the initial slope of the regression line. Fluxes were considered significant if the slope of the regression line had a p-value lower than 0.05. The fluxes from 2008, 2010 and 2015 (Hall et al., 2017; Nilsson et al., 2019; Viktorsson et al., 2013) have been re-evaluated to assure that the same method is used and thus the results are comparable.

## 2.3 Sediment sampling

Pore water was extracted from sediment cores (9.9 cm inner diameter) collected with a multiple corer. At each station, one core was sectioned in a portable glove bag (N<sub>2</sub> atmosphere; Aldrich AtmosBag, Sigma-Aldrich). The cores were sliced at 0.5 cm resolution from 0 to 2 cm depth, at 1 cm resolution between 2 and 6 cm depth, and in 2 cm slices from 6 to 20 cm depth. The sediment was transferred into centrifuge tubes and pore water was collected using Rhizon samplers (Rhizosphere research products).



135 Samples for solid-phase speciation of phosphorus were collected at station D and F in 2016 and 2017. Due to logistical constraints, samples were not collected on the other occasions. Sediment was collected using a modified boxcorer (Blomqvist et al., 2015) from which 6 cores were taken using transparent PVC core liners (6 cm inner diameter; 30 cm long). Two cores per station were sectioned in a portable glove bag ( $N_2$  atmosphere; Captair Field pyramid, Erlab) under constant oxygen monitoring (Portable oxygen analyser 3110 with trace oxygen sensor, Teledyne). The cores were sliced at 0.5 cm resolution from 0 to 3 cm depth, at 1 cm resolution between 3 and 10 cm depth, and in 2 cm slices from 10 to 14 cm depth. The sediment was transferred into polypropylene centrifuge tubes (TPP), centrifuged at 2500g for 10 min (Sigma 3-16L, Sigma) and was  
140 returned to the anoxic glove bag. After removal of the overlying pore water, the solid phase was freeze-dried and stored in an aluminium bag under  $N_2$  atmosphere for later speciation of phosphorus. In 2016, the pore water was filtered (0.42  $\mu m$  cellulose acetate filters, Chromafil Xtra) and acidified with 100  $\mu L mL^{-1}$   $H_2SO_4$  (1 M) for later analysis as a control of our pore-water sampling procedure.

### 145 2.3 Bottom-water and pore-water analysis

Nutrient samples from the lander and multiple corer pore-water samples were determined by segmented flow colorimetric analysis (Alpkem Flow Solution IV, OI Analytical; Koreleff, 1983). The analytical precisions were 3 % for DIP, 3 % for dissolved inorganic nitrogen (DIN) and 2.5 % for dissolved silica (DSi). In 2016, pore-water DIP samples from the box corer were determined by segmented flow colorimetric analysis (QuAAtro, SEAL Analytical; Aminot et al., 2009). The precision  
150 was < 4 %.

DIC concentrations in the lander samples were determined on-board the ship by non-dispersive infrared spectrometry (LI-COR 6262) after acidification with phosphoric acid (Nilsson et al., 2019). Repeated measurements of certified reference material (CRM from Dickson Laboratory, Scripps Inst. of Oceanography) were conducted to obtain a two-point calibration and correction for potential drift in the system. The analytical precision was 0.2 % (relative standard deviation).  
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### 2.4 Sequential phosphorus extractions

A subsample of ~100 mg sediment from each depth was used for sequential P extraction (SEDEX; Ruttenberg, 1992; Slomp et al., 1996). The SEDEX procedure separates total sediment P into five fractions; exchangeable or loosely sorbed P ( $P_{ex}$ ), easily reduced or reactive iron bound P ( $P_{Fe}$ ), authigenic P ( $P_{auth}$ ), detrital P ( $P_{det}$ ) and P associated with organic matter ( $P_{org}$ ).  
160 All extractions were performed at room temperature and under constantly agitated conditions; extraction solutions and times are shown in Table 1. After each extraction, the sample was centrifuged (2500 g for 10 min) and the supernatant was filtered (0.45  $\mu m$  cellulose acetate) and subsequently analysed spectrophotometrically. The first two extraction steps were performed under oxygen-free conditions. The  $P_{Fe}$  fraction was analysed by inductively coupled plasma – optical emission spectroscopy (ICP-OES), since the reagent interferes with the spectrophotometric colour reagent. Performance of the method was monitored



165 by analysing one sample in triplicate during each batch of extractions (to assess the repeatability by calculating the relative standard deviation), and by analysing subsamples of one sample with every batch of extractions (to assess reproducibility by calculating the relative standard deviation). For each extraction step, repeatability was around 5% and reproducibility was around 10%. Calculation of sedimentary inventories of different P fractions are described in appendix B.

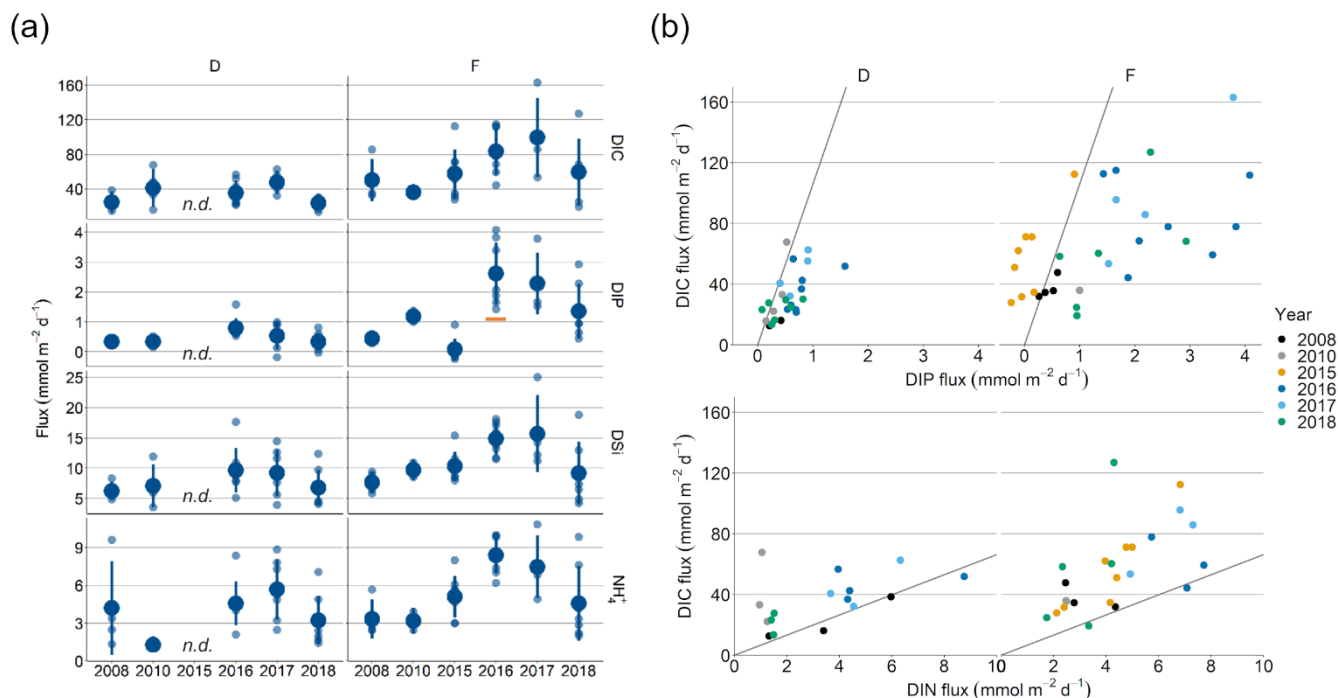
170 **Table 1.** Specifics for the sequential extractions of phosphorus. Brief summary of the reagents, methods and times used for SEDEX. Each step was followed by a wash with 1M MgCl<sub>2</sub> (pH 8) for 30 minutes, to avoid re-adsorption of DIP.

Fraction	Extraction solution	Time
P <sub>ex</sub>	1 M MgCl <sub>2</sub>	30 min
P <sub>Fe</sub>	12 g sodium dithionite in 480 mL sodium acetate (1 M) + 60 mL sodium bicarbonate (1 M)	8 h
P <sub>auth</sub>	300 mL sodium acetate (1 M) + 1700 mL acetic acid (1 M)	6 h
P <sub>det</sub>	1 M HCl	24 h
P <sub>org</sub>	Combustion at 550°C	2 h
	1 M HCl	24 h

### 3 Results and Discussion

#### 175 3.1 Enhanced sedimentary P release after the inflow

Sediment-water fluxes measured in situ showed a clear pattern at station F following the MBI, for DIP as well as for several other biogenic solutes (DIC, DSi and DIN; Fig. 3a). In 2015, the DIP efflux decreased markedly at station F (by 88 %) compared to the 2008 and 2010 (pre-inflow) values, and even turned into an uptake in about half of the chambers (Hall et al., 2017). In 2016, however, the DIP flux was about 2.5 times higher than before the inflow, after which it decreased stepwise in  
180 2017 and 2018, returning to pre-inflow values. Although there was substantial variation in the fluxes during each measurement campaign, the standard deviations for DIP, DSi and DIN in 2016 did not overlap with standard deviations for data from previous years, which suggests that the fluxes were indeed elevated after the inflow.



185 **Figure 3.** Sediment-water fluxes measured with the benthic chamber lander. Flux data are from before the inflow (2008, 2010; Hall et al.,  
 2017; Nilsson et al., 2019; Viktorsson et al., 2013)), right after the inflow (2015; Hall et al., 2017)) and post inflow (2016-2018; this study).  
 (a) Changes in fluxes with time. Small circles mark individual chamber measurements, large circles mark station averages and lines represent  
 station standard deviations. The orange line in the DIP-graph for station F marks the expected flux for 2017, based on the decrease in solid phase  
 inventory. n.d. = no data. (b) Fluxes of DIC versus DIP and DIN. The grey line represents the Redfield ratios of 106:1 for DIC:DIP  
 190 and 106:16 for DIC:DIN.

The decrease in DIP flux at station F in 2015 temporarily raised the DIC:DIP flux ratio above the Redfield ratio (Redfield,  
 1958) of 106:1 (Fig. 3b; Hall et al., 2017). As the DIC:DIN ratio did not change, the lowered DIP flux was most likely due to  
 trapping of P by iron and manganese oxides, formed at the sediment surface as a result of the oxygenation. When anoxia  
 195 returned in 2016,  $\text{P}_{\text{Fe}}$  that had been temporarily trapped during oxic conditions was expected to be released and increase the  
 sedimentary DIP efflux. However, the DIC:DIP flux ratios at station F in the years following the inflow suggest that release  
 of  $\text{P}_{\text{Fe}}$  had a negligible effect on the DIP fluxes. In 2016-2018, the DIC:DIP ratio decreased to ~40, which is within the range  
 of what is normally observed in the anoxic parts of the EGB (~20-70; Sommer et al., 2017; Viktorsson et al., 2013). The  
 consistent DIC:DIP ratio implies that other factors than release of  $\text{P}_{\text{Fe}}$  contributed to the increased benthic DIP efflux at station  
 200 F in 2016-2017. An intriguing clue to what could have caused the elevated DIP fluxes was given by the fluxes of DIC, DSi  
 and DIN, which followed the same pattern of temporarily increased fluxes. The co-variation suggests that a common factor  
 affected all solute fluxes, hinting towards an increase in the organic matter mineralisation rate and/or organic matter supply.





The flux pattern observed at station F was not seen at station D, where the fluxes were relatively constant over the years. A sedimentary uptake of DIP was measured in one chamber in 2017, suggesting that the oxygenation of the sediment was weak and heterogeneous. Unlike station F, the DIC:DIP ratio was not affected by the oxygenation (Fig 3b). Station D is situated at the edge of the area that was affected by the MBI and the oxygenation there was less strong (Fig. 2c). The conditions at station D can thereby be assumed to represent a lower boundary for effects from the inflow.

### 3.2 Sedimentary phosphorus dynamics after the inflow

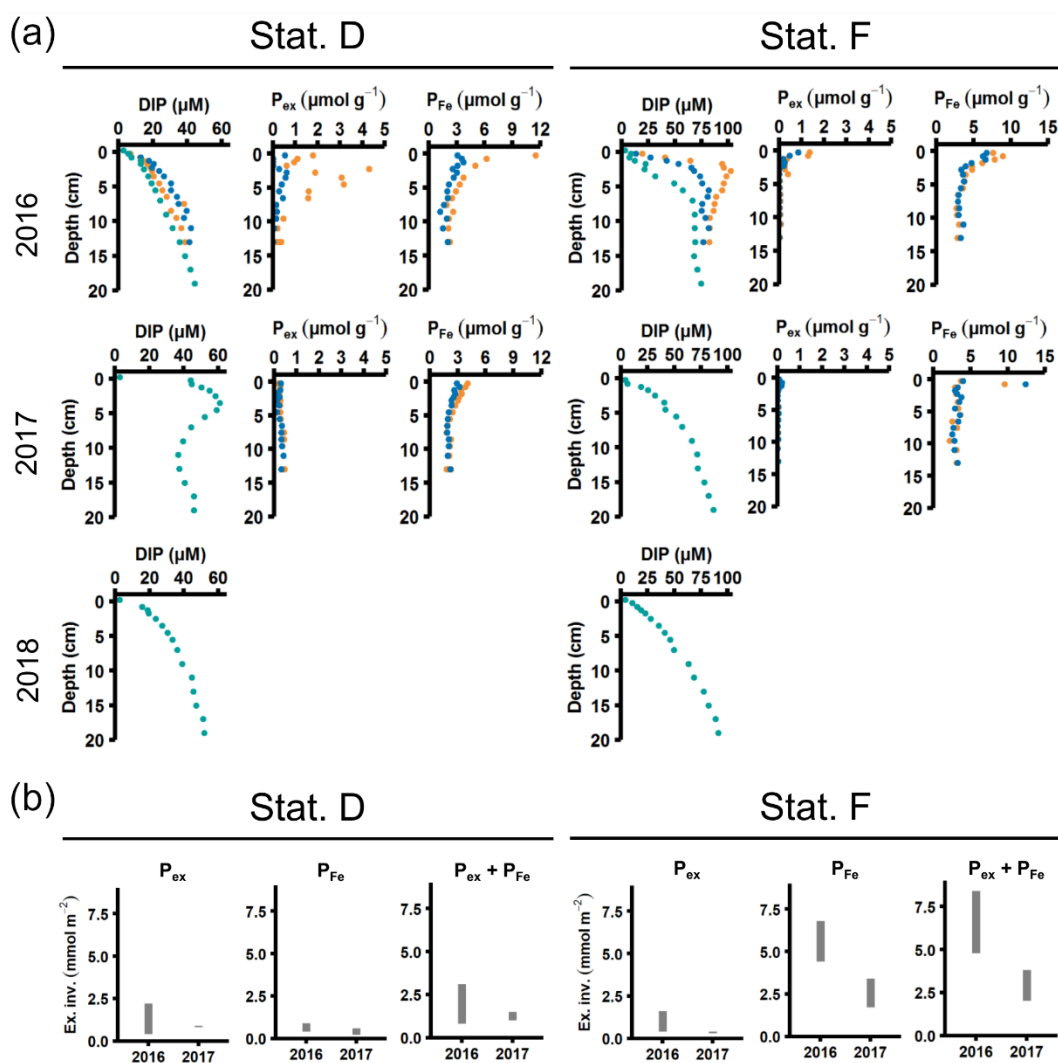
Sediment profiles from all stations showed clear signs of oxygenation after inflows (Fig. 4a, A3 and A4). At station F, the pore-water DIP (Fig. 4a) displayed distinct subsurface peaks in 2016. This type of profile suggests production of DIP in the upper sediment layers (Ruttenberg, 2003), probably caused by a sudden influx of P in the form of reactive organic matter or P adsorbed to iron and manganese oxides. In contrast, pore-water DIP gradually accumulated with depth in 2017 and 2018. Both the  $P_{\text{exch}}$  and  $P_{\text{Fe}}$  concentrations were higher in 2016 than in 2017 (Fig. 4a; other P fractions in Fig. A4). This indicates that there was a transient enrichment of these fractions in 2016 that had disappeared in 2017, which would explain the DIP peak. The sequestration of P after oxygenation was calculated as the excessive inventories (Fig. B1) of  $P_{\text{ex}}$  and  $P_{\text{Fe}}$ , i.e. the integrated enrichment of these fractions compared to their background concentrations. The excess inventory of  $P_{\text{exch}}$  and  $P_{\text{Fe}}$  suggest that the net P retention at station F by the sampling occasion in 2016 was 4.8 – 8.4 mmol P m<sup>-2</sup> (Fig. 4b), in consistency with a study from the eastern part of the EGB where 12 mmol P m<sup>-2</sup> was estimated to have been retained due to the MBI (Hermans et al., 2019). By the sampling occasion in 2017, the net excess inventory had decreased to 2.0 – 3.8 mmol P m<sup>-2</sup>. This means that 1.0 – 6.4 mmol P m<sup>-2</sup> was released from temporary storage in the sediment between the 2016 and 2017 samplings. As no increase was seen in the  $P_{\text{auth}}$  fraction (Fig. A4), it appears that most of this P was lost to the overlying water. A release of 6.4 mmol P m<sup>-2</sup> in one year would lead to an average sedimentary efflux of 0.02 mmol m<sup>-2</sup> d<sup>-1</sup>. However, the benthic flux measured in 2017 was more than 2 mmol m<sup>-2</sup> d<sup>-1</sup> higher than before the inflow (Fig. 3a). As suggested by the DIC:DIP ratio, another source of P must have contributed to the DIP flux at station F in 2016-2018.

Signs of oxygenation were also seen at station D. In 2016 and 2018, the pore-water DIP accumulated with sediment depth. However, the profile was very different in 2017 and displayed a peak at 4 cm depth, suggesting an increased release of reactive P (Ruttenberg, 2003). Very little  $P_{\text{ex}}$  and  $P_{\text{Fe}}$  was present at station D (Fig. 4a), which is expected for sediments deposited in anoxic waters (Ruttenberg, 2003). However, subsurface peaks of  $P_{\text{ex}}$  and  $P_{\text{Fe}}$  at station D were visible in one of the cores from 2016. The variation between replicate cores in 2016 raises the question whether changes in pore-water DIP,  $P_{\text{ex}}$  and  $P_{\text{Fe}}$  at station D were caused by spatial variability, or if there was a change over time in response to external factors. However, the changes in sedimentary P profiles and fractionation follow what would be expected if station D had been partially affected by the MBI; a likely scenario based on monitoring oxygen data (SMHI, 2019).

On the other hand, the 2017 inflow had a smaller effect on the sediment geochemistry at station D, since no peaks were present in the cores collected that year. At station D, the  $P_{\text{ex}} + P_{\text{Fe}}$  excess inventory change between the 2016 and 2017 samplings



240 ranged from +0.2 to -2.1 mmol m<sup>-2</sup> (Fig. 4b). As there was no increase in the authigenic P fraction (Fig. A4), it is likely that this P was released to the water column as DIP. The excess P release at station D would have had a negligible contribution to the DIP flux, which indeed did not change markedly over the sampling (Fig 3). Diffusive fluxes calculated from the pore-water profiles were considerably lower than the in situ measured fluxes (Table C1). This is not surprising, however, since fluxes calculated from pore-water gradients can underestimate the actual flux e.g. due to too coarse vertical resolution (Nilsson et al., 2019; Noffke et al., 2012; Sundby et al., 1986). The mismatch between in situ measured and calculated diffusive fluxes further suggests that the change in fluxes after the inflow was caused by processes occurring close to the sediment-water interface.



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**Figure 4.** Pore-water and sediment solid phase data. (a) Pore-water DIP profiles, solid-phase exchangeable or loosely-sorbed P ( $P_{ex}$ ) and easily-reduced or reactive iron-bound P ( $P_{Fe}$ ) profiles from station D and F in the Eastern Gotland Basin, sampled in April 2016, 2017 and 2018. Note the differences in x axes. Profiles with green markers are from big sediment cores (inner diameter 9.9 cm); profiles with blue and orange markers are from replicate small sediment cores (inner diameter 6 cm). (b) Excess inventories (min – max range) of  $P_{ex}$ ,  $P_{Fe}$  and the sum of the two fractions at stations D and F in 2016 and 2017.

### 3.3 Increased mineralisation after the inflow

Our data suggest that the elevated fluxes of DIP and other dissolved biogenic elements were caused by increased organic matter mineralisation. Changes in sedimentary mineralisation rates can be caused by an enhanced input of reactive organic matter to the sediment (for example after a spring bloom or by enhanced organic matter export out of the water column) or by an increase of the intrinsic mineralisation rate of the organic matter already present in the sediment. We will discuss these different possibilities below.

Data for this study were collected during different seasons (2008, 2010: August/September; 2015: early July; 2016-2018: April), so some variation in fluxes due to seasonality is expected. However, even though DIC and nutrient effluxes should be highest in 2008, 2010 and 2015 due to sedimentation of fresh phytoplankton material (Berelson et al., 2003), similar flux values were measured in 2018. Instead, the highest fluxes were recorded at station F during the spring samplings (2016-2018, Fig. 3a). The differences in timing of the sampling relative to the spring bloom could still be a potential cause. However, both the lack of clear surficial peaks in the chlorophyll a sediment profiles (Fig. D1) and monthly water-column monitoring data of phytoplankton and nutrient concentrations from monitoring data (Fig. A5; SMHI, 2019) suggest that all samplings in 2016-2018 were conducted before the spring blooms. In fact, the year when the sampling was most likely to coincide with the spring bloom according to the SMHI data was in 2018, which was the year with the lowest fluxes of all spring samplings. It thus seems highly unlikely that seasonality was driving the observed flux pattern.

The MBI could also have pushed up deep water to the surface, thereby providing nutrients to phytoplankton blooms. No increase in primary production was observed, however, during either the spring bloom or summer bloom in the years after the MBI (Johansen and Skjjevik, 2016-2017). Instead, the variation of fluxes closely follows the pattern of oxygenation, with higher fluxes just after the oxygenation and a gradual decrease to pre-MBI values once all oxygen had been consumed in the bottom waters (Fig. 2c, Fig. 3).

The oxygenation of the bottom waters following a MBI could potentially lead to enhanced reactive particulate organic matter input to the sediment. In oxygen minimum zones, particle fluxes are normally higher than in surrounding oxic waters, possibly because of lower particle mineralisation rates as a result of less energetic electron acceptors or loss of zooplankton that consume particles (Le Moigne et al., 2017). However, a study conducted in the EGB in the summer of 2015 suggested that the particle sedimentation rate and freshness of the organic material reaching the sediment increased due to the MBI (Cisternas-Novoa et al., 2019). In the newly oxygenated EGB, the flux of particulate organic carbon (POC) increased by 18 % between 40 m and 180 m depth, while it decreased by 28 % at a permanently anoxic site nearby (Cisternas-Novoa et al., 2019). The cause of the

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increased POC transport in the EGB was thought to be the formation of manganese oxides (Mn-oxides) in the oxygenated water column, scavenging organic matter while sinking (Cisternas-Novoa et al., 2019). Several other studies have confirmed that the MBI led to a considerable formation and sedimentation of Mn-oxides in the water column of the EGB (Dellwig et al., 2018; Hermans et al., 2019; van de Velde et al., 2020b). Enhanced transport of organic matter to the sediment via Mn-oxides could thus have led to a higher deposition and increased benthic organic matter degradation rates. According to the numbers presented by Cisternas-Novoa et al. (2019), however, the increased vertical POC flux after oxygenation would still have remained relatively low. In contrast, lateral transport could have been a substantial source of POC and associated elements after the inflow. Increased turbidity was measured close to the seafloor below 140 m depth in March 2015, as a gravity current generated by the inflow moved along the slope (Schmale et al., 2016). The inflow itself could thus have enhanced the constantly ongoing lateral transport of relatively fresh organic matter from shallower to deeper areas (Nilsson et al., 2020).

Oxygenation can also allow re-colonisation of previously anoxic sediments by benthic animals. Their presence would result in sediment reworking (bio-mixing) and bio-irrigation, which are believed to stimulate organic matter degradation (van de Velde et al., 2020a). Two individuals of *Bylgide sarsi* were observed in the area after the inflow (Hall et al., 2017; Stigebrandt et al., 2018). There were no signs of colonisation by animals in the newly oxygenated area, however, and it is likely that the two specimens had arrived with the inflowing water (Stigebrandt et al., 2018). We therefore exclude animal activity as an explanation for the enhanced sediment-water fluxes after the MBI.

Oxygen can also directly contribute to an enhancement of the organic matter mineralisation rate. Increased mineralisation rates under oxic conditions, compared to anoxic conditions, have been suggested for refractory (or aged) and some types of fresh organic matter (Arndt et al., 2013; Bianchi et al., 2018). The reason for this is debated, but factors such as production of different enzymes and higher energy yield during degradation of organic matter under oxic conditions have been proposed (LaRowe et al., 2020). Although oxygen can affect the mineralisation of organic matter, the connection between oxygen and increased DSi fluxes is less clear. It is possible that oxygen promoted the degradation of the organic coating covering diatom frustules, which in turn could enhance the dissolution of biogenic silica (Bidle and Azam, 1999). The elevated DSi fluxes observed here could also indicate that an increased input of organic matter (to which biogenic silica is associated), rather than the presence of oxygen, was causing the change in fluxes.

Interestingly, a study from the EGB conducted shortly after the MBI found that while the intrusion of oxygen in itself did not enhance the heterotrophic bacterial production in the water column, the removal of hydrogen sulphide did (Piontek et al., 2019). The same mechanism could be relevant for the surface layer of the sediment, since the MBI also resulted in the removal of hydrogen sulphide right below the sediment-water interface (Marzocchi et al., 2018).

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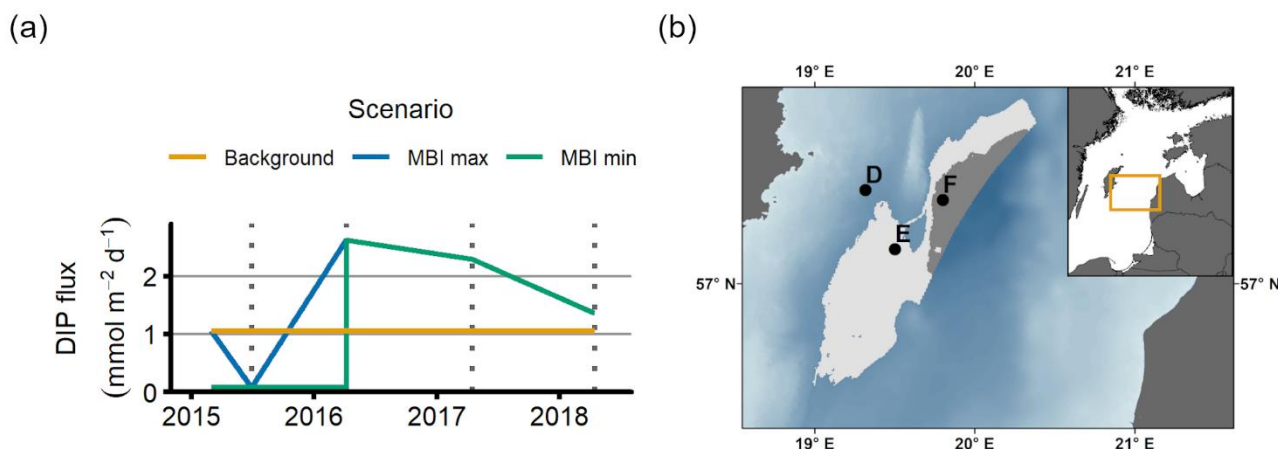
### 3.4 Implications for coastal oxygen depletion

Following the MBI, there was a substantial elevation of the DIP fluxes in the EGB. To estimate the impact on the P inventory in the area, we propose three scenarios for the total DIP release from the sediment between March 1 2015 and April 16 2018



(Fig. 5a). The Background scenario describes a situation without the MBI, assuming that the flux equals the average of the  
315 fluxes in 2008, 2010 and 2018. The MBI min scenario describes a situation where the flux decreased to the value measured in  
2015 as soon as oxygen reached the EGB, stayed at that level until oxygen disappeared and then changed linearly between the  
measured values. The MBI max scenario describes a situation where the flux begins at the Background value and then changes  
linearly between the measured values. At station F, the Background scenario gives a total DIP release of  $1.20 \text{ mol m}^{-2}$ , while  
the MBI min scenario results in a total release of  $1.62 \text{ mol DIP m}^{-2}$  (35 % increase) and the MBI max scenario gives a total  
320 release of  $2.04 \text{ mol DIP m}^{-2}$  (70 % increase). At station E, the total release of DIP is increased by 62 % and 96 % (Table E1).  
Upscaling of these results (appendix E) further shows the importance of the increased DIP release. The calculation is limited  
to the western flank of the EGB, since previous studies have shown that the sediment-water fluxes generally are lower on the  
eastern side (Hall et al., 2017; Nilsson et al., 2019; Sommer et al., 2017; Viktorsson et al., 2013). We assume that the  
sedimentary DIP release per  $\text{m}^2$  at 150-190 m depth equalled that at station E, while the release at sediments deeper than 190  
325 m were represented by station F (Fig. 5b). The MBI scenarios then predict an extra release of 9.28-15.54  $\text{kton y}^{-1}$ . These  
numbers can be put in relation to the total release of DIP from oxygen-depleted sediments in the central Baltic Sea (Baltic  
Proper), which has been estimated to 60 – 150  $\text{kton y}^{-1}$  (Almroth-Rosell et al., 2015; Hall et al., 2017; Sommer et al., 2017;  
Viktorsson et al., 2013). Even our conservative estimates suggest a substantial increase in sedimentary DIP release following  
the MBI.

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**Figure 5.** Upscaling of DIP release. (a) Total DIP release at station F according to three different scenarios: Background, MBI max and MBI min. (b) Sediment areas (western flank of the EGB) used for upscaling calculations. The DIP flux per  $\text{m}^2$  in the light grey area (150-190 m) is assumed to be the same as at station E, while the dark grey area ( $> 190 \text{ m}$ ) is represented by station F. Bathymetric data from the Baltic Sea Bathymetry Database (version 0.9.3), the map was created using the software ArcMap<sup>TM</sup> (v. 10.6) by Esri.

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#### 4 Conclusions

In this paper, we show that an oxygenated inflow to long-term anoxic sediments temporarily elevated organic matter degradation, thereby increasing the sedimentary release of DIP. Oxygen is generally believed to be able to break the positive feedback between anoxia and eutrophication (Jilbert and Slomp, 2013; Ruttenberg, 2003), but that was not the case with the short and moderate oxygenation in this study. Instead, our study points to three ways in which these oxygenation events could increase the return of nutrients to the water column: enhanced vertical and lateral transport of organic matter and increased mineralisation rates under oxic conditions. Most likely, a combination of these factors caused the increase in sedimentary nutrient and DIC efflux observed here. Regardless of the mechanism, enhanced mineralisation of organic matter after oxygenated inflows is a previously unrecognised feedback mechanism in eutrophic coastal systems (Breitburg et al., 2018; Middelburg and Levin, 2009).

In estuaries and coastal seas around the world, climate change and land use are predicted to both strengthen and expand oxygen-depleted zones (Altieri and Gedan, 2015). While oxygenated inflows have been thought to mitigate eutrophication feedbacks by enhancing P burial, our results show that they also can lead to increased mineralisation of organic matter. In long-term anoxic areas, irregular inflows might not bring enough oxygen to oxidise reduced chemical species and promote removal of nutrients. In those systems, enhanced degradation of organic matter caused by oxygenated inflows could instead increase the availability of phosphorus and other nutrients, thereby strengthening eutrophication.

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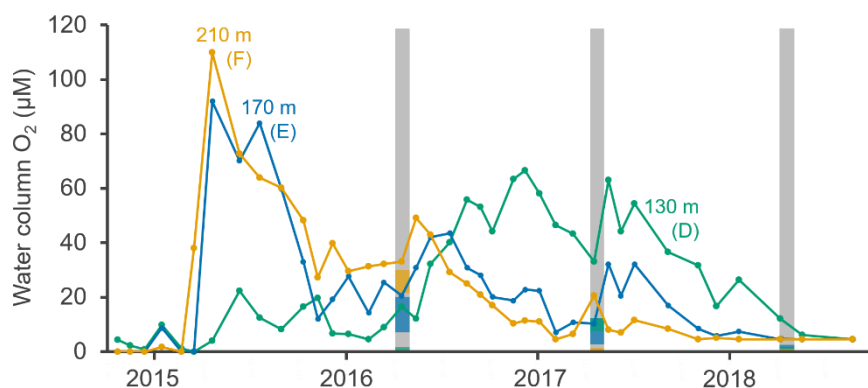


370 **Appendix A: Supplementary figures and tables**

**Table A1.** Description of stations. Bottom water salinity, temperature and oxygen concentration as measured by the benthic chamber lander or CTD. Limit of detection for oxygen was 0.5  $\mu\text{M}$ .

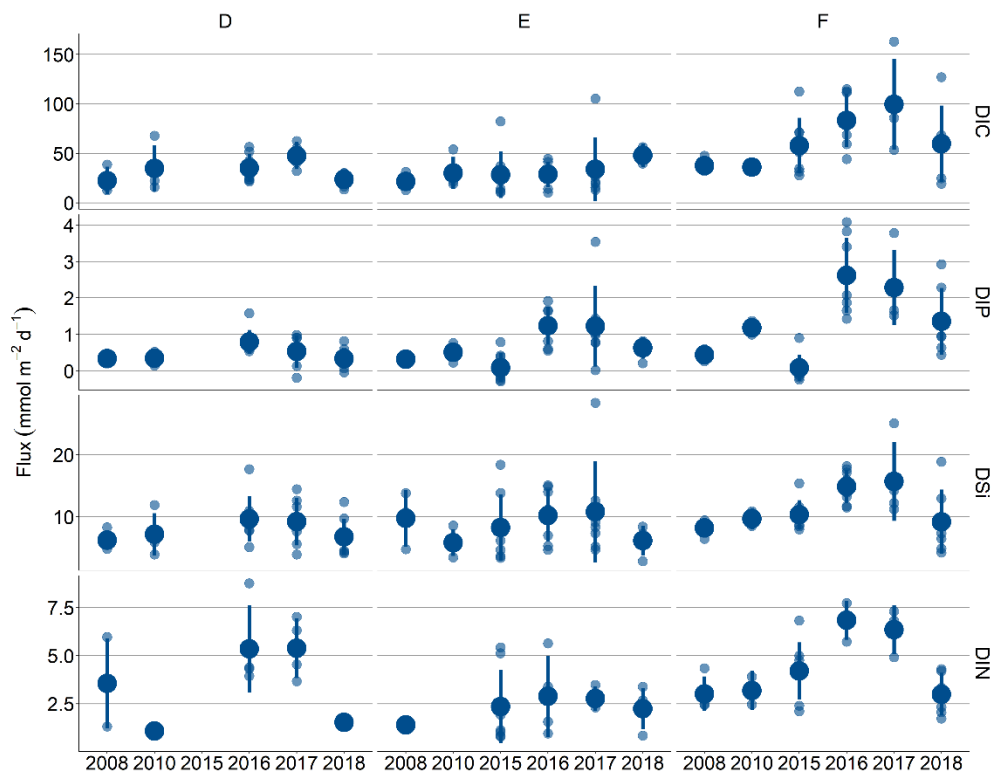
Station	Depth	Year	Salinity	Temp. ( $^{\circ}\text{C}$ )	Oxygen ( $\mu\text{M}$ )
<b>D</b>	130 m	2016	12.5-12.7	6.5-6.6	<0.5
		2017	12.7	6.7-6.8	8-13
		2018	12.4-12.6	6.6-6.9	<0.5
<b>E</b>	170 m	2016	12.9-13.1	6.8-6.9	7-20
		2017	13.1	6.9-7.2	3-13
		2018	13.1	6.8-7.1	<0.5
<b>F</b>	210 m	2016	13.8-13.9	7.3-7.4	22-30
		2017	13.5	7.2-7.4	<0.5
		2018	13.1-13.3	6.9-7.15	<0.5

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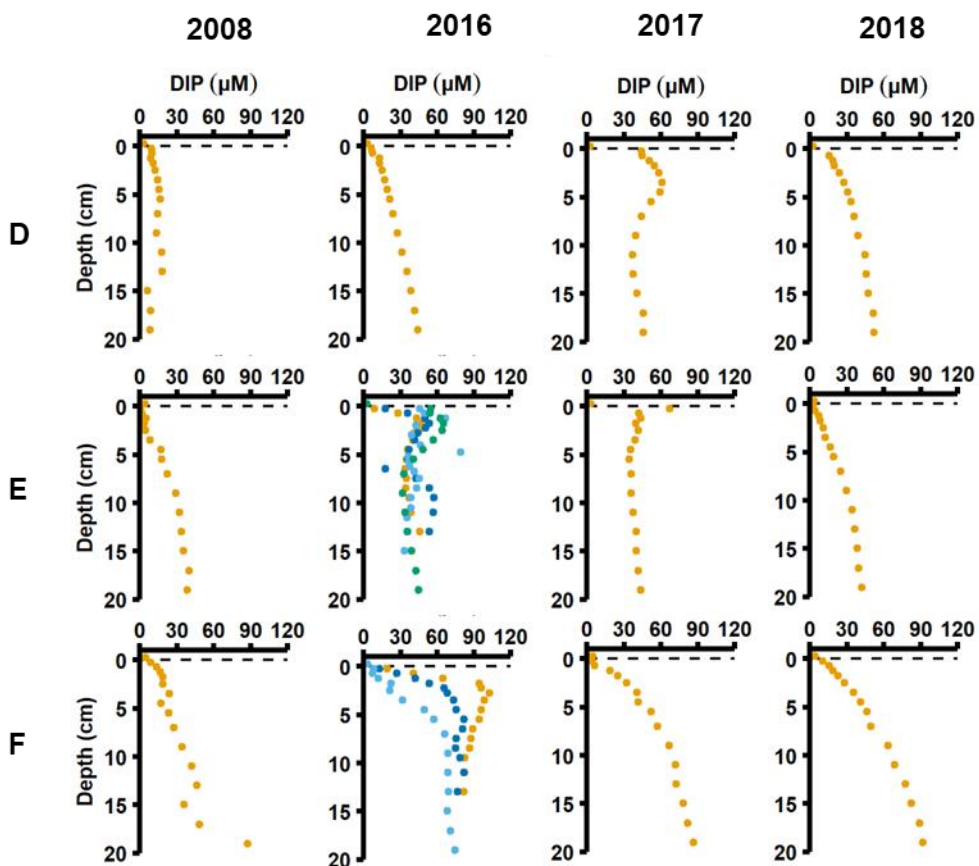
**Figure A1.** Oxygen monitoring data. Water column oxygen concentrations at depths corresponding to stations D-F at the SMHI (Swedish Meteorological and Hydrological Institute) monitoring station BY15 (SMHI, 2019). The grey lines indicate when samplings for this study were conducted. Colours on the lines show the bottom water concentration measured by the benthic chamber lander at each station.

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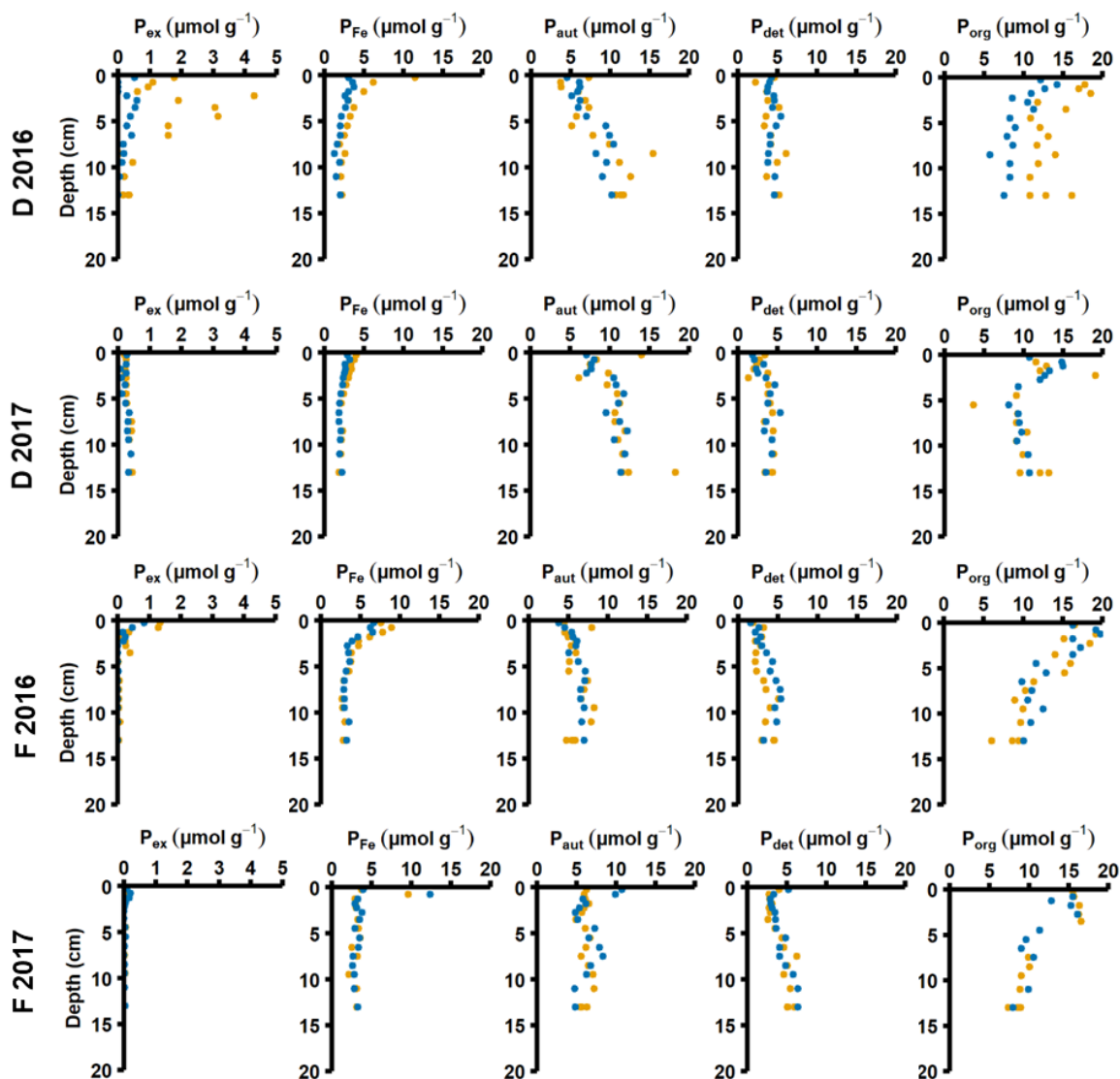


385 **Figure A2.** Sediment-water fluxes measured with the benthic chamber lander. Flux data are from before the inflow (2008, 2010; Hall et al., 2017; Nilsson et al., 2019; Viktorsson et al., 2013), right after the inflow (2015; Hall et al., 2017)) and post inflow (2016-2018; this study). There are no data from station D in 2015.

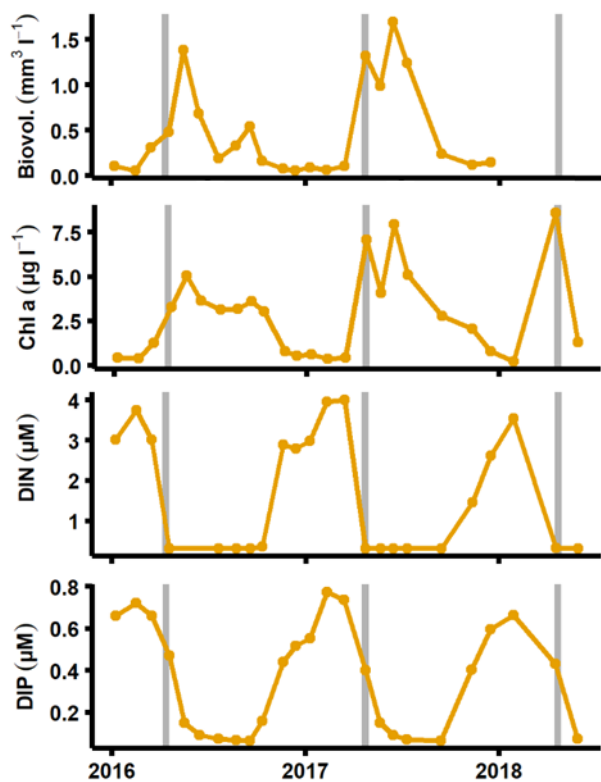




390 **Figure A3.** Pore-water profiles of dissolved inorganic phosphorus (DIP). Profiles from 2008 are from ref. (Viktorsson et al., 2013). Different colors show replicate cores: orange markers – core 1, dark blue markers – core 2, light blue markers – core 3, green markers – core 4.



395 **Figure A4.** Solid phase phosphorus speciation.  $P_{ex}$  – exchangeable or loosely sorbed P,  $P_{Fe}$  – easily reduced or reactive iron bound P,  $P_{auth}$  – authigenic P,  $P_{det}$  – detrital P,  $P_{org}$  – P associated with organic matter. The blue and orange markers represent replicate small sediment cores (inner diameter 6 cm).



**Figure A5.** Primary production and nutrient monitoring data. Data from SMHI station BY15, top 10 m of the water column (SMHI, 2019). The graphs show (from top to bottom) phytoplankton biovolume, chlorophyll a concentrations, DIN (nitrate + nitrite + ammonium) concentrations and dissolved inorganic phosphorus (DIP) concentrations. The grey lines indicate when samplings for this study were conducted.

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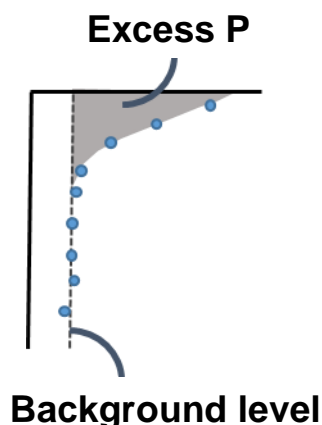


## Appendix B: Calculation of solid phosphorus inventories

The solid phase inventory between the sediment surface and sediment depth  $z$  was calculated by multiplying the solid phase density ( $\sigma_{\text{sed}}$ ) with the depth-integrated porosity ( $\phi$ ) and concentration of the phosphorus fraction ( $C_P$ ) of interest:

$$415 \quad \text{Inventory} = \sigma_{\text{sed}} \int_0^{z_{\text{end}}} (1-\phi(z)) C_P(z) dz \quad (\text{B1})$$

Values for  $\phi$  and  $\sigma_{\text{sed}}$  were taken from van de Velde et al. (2020b). We were interested in the ‘excess’ P inventory (Fig. B1), which is the transiently increased P fraction. For  $P_{\text{ex}}$ , we consider the excess P inventory to be the inventory as calculated by equation 1, as the concentration of  $P_{\text{ex}}$  decreased to 0 below 5 cm in both 2016 and 2017. The  $P_{\text{Fe}}$  profile had a background concentration of  $\sim 3 \mu\text{mol g}^{-1}$  below 5 cm, which was consistent between sampling years. The concentration at the water interface in 2017 (i.e. the concentration in the settling particles that had not been exposed to the oxygen-rich water of the MBI) was also close to  $3 \mu\text{mol g}^{-1}$ . Therefore, we determined the ‘excess’  $P_{\text{Fe}}$  concentration as the inventory as calculated by equation 1, subtracted by a background inventory calculated by replacing  $C_P$  in equation 1 with the average concentration below 5 cm.



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**Figure B1.** Definition of ‘excess’ phosphorus (P). Schematic figure showing the definition of ‘excess’ phosphorus (P), which is the integrated enrichment of a P fraction compared to its background concentration.

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### Appendix C: Calculation of diffusive fluxes

Diffusive fluxes ( $J$ ) were calculated from the pore water profiles of DIP (Fig. A3) according to Eq. C1:

$$435 \quad J = -\varphi D_s \frac{\partial C}{\partial z} \quad (C1)$$

where  $D_s$  is the sediment diffusion coefficient and  $C$  is the concentration (Boudreau, 1997).  $D_s$  was calculated according to Eq. C2:

$$440 \quad D_s = \frac{D}{\theta^2} = \frac{D}{1 - \ln(\varphi^2)} \quad (C2)$$

where  $D$  is the diffusion coefficient in free solution and  $\theta$  is the tortuosity (Boudreau, 1997). The R package ‘marelac’ (Soetaert et al., 2020) was used to calculate  $D$ .

445 **Table C1.** Diffusive fluxes of dissolved inorganic phosphorus. The fluxes are calculated from pore water profiles, values are given in  $\text{mmol m}^{-2} \text{d}^{-1}$ . The 2008 values are calculated from profiles from ref. (Viktorsson et al., 2013). When there are results from multiple cores, the core number is given in brackets. For profiles that displayed production peaks, both the upward flux ( $J_{\text{up}}$ ) and the downward flux ( $J_{\text{down}}$ ) from that peak are presented.

	2008	2016	2017	2018
<b>D</b>	$J_{\text{up}} = 0.023$ $J_{\text{down}} = 0$ $J_{\text{sum}} = 0.023$	$J_{\text{up}} = 0.015$ $J_{\text{down}} = 0$ $J_{\text{sum}} = 0.015$	$J_{\text{up}} = 0.276$ $J_{\text{down}} = 0.016$ $J_{\text{sum}} = 0.292$	$J_{\text{up}} = 0.029$ $J_{\text{down}} = 0$ $J_{\text{sum}} = 0.029$
<b>E</b>	$J_{\text{up}} = 0.008$ $J_{\text{down}} = 0$ $J_{\text{sum}} = 0.008$	$J_{\text{up}} = 0.100$ (1), 0.115 (2), 0.311 (3), 0.373 (4) $J_{\text{down}} = 0.047$ (1), 0.034 (2), 0.007 (3), 0.024 (4) $J_{\text{sum}} = 0.147$ (1), 0.149, 0.318, 0.397	$J_{\text{up}} = 0.459$ $J_{\text{down}} = 0.178$ $J_{\text{sum}} = 0.636$	$J_{\text{up}} = 0.012$ $J_{\text{down}} = 0$ $J_{\text{sum}} = 0.012$
<b>F</b>	$J_{\text{up}} = 0.026$ $J_{\text{down}} = 0$ $J_{\text{sum}} = 0.026$	$J_{\text{up}} = 0.165$ (1), 0.092 (2), 0.030 (3) $J_{\text{down}} = 0.010$ (1), 0 (2), 0 (3) $J_{\text{sum}} = 0.175$ (1), 0.092 (2), 0.030 (3)	$J_{\text{up}} = 0.053$ $J_{\text{down}} = 0$ $J_{\text{sum}} = 0.053$	$J_{\text{up}} = 0.041$ $J_{\text{down}} = 0$ $J_{\text{sum}} = 0.041$



#### Appendix D: Pigment extractions

450 Pigment samples were collected in 2008, 2017 and 2018. Sediment cores were collected with a multiple corer and were sliced at 0.5 cm resolution from 0 to 2 cm depth. From each slice, 1 mL of sediment was transferred to 15 mL centrifuge tubes and was subsequently frozen in liquid nitrogen. The samples were extracted in 10 mL of an acetone:methanol (80:20) solution at -20°C for 24 h before they were sonicated for 60 s and filtered through 0.45 µm nylon filters. Extracts were separated and identified by high performance liquid chromatography (HPCL, Shimadzu LCsolutions System) with photodiode array detection (SPD-M20A). The relative concentration of sample chlorophyll a was determined from calibration curves acquired from analysis of an external standard (DHI LAB).

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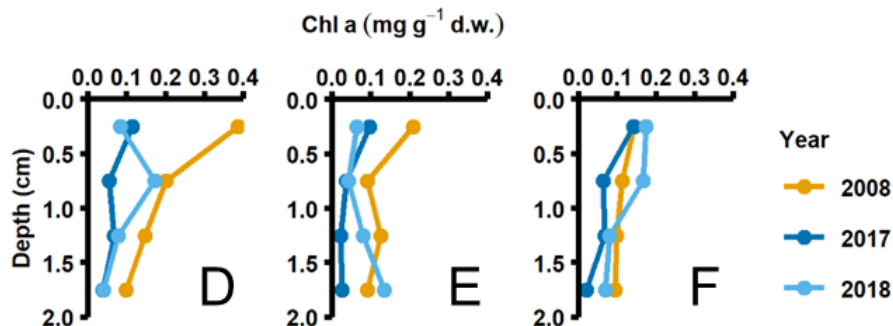


Figure D1. Chlorophyll a profiles. Profiles of chlorophyll a (Chl a) in the top 2 cm of the sediment.

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## Appendix E: Upscaling calculations

Bathymetry data from the Baltic Sea Bathymetry Database (version 0.9.3) were used for the upscaling calculations. Sediment surface areas were extracted using the tool ‘Surface Volume’ in the software ArcMap™ (v. 10.6) from Esri. The area at 150-  
 475 190 m depth was represented by station E, while sediments deeper than 190 m were represented by station F. The extracted areas were multiplied with the total DIP release calculated for the Background, MBI min and MBI max scenarios, and were then divided by the time used in the total release scenarios (3.13 years) to obtain an areal release per year.

480 **Table E1.** Total release of dissolved inorganic phosphorus (DIP). The total DIP release represent the entire time period from the arrival of oxygen until the end of the study (3.13 years).

<b>Total DIP release</b>			
	<b>Background (mol m<sup>-2</sup>)</b>	<b>Min release (mol m<sup>-2</sup>)</b>	<b>Max release (mol m<sup>-2</sup>)</b>
<b>Station E</b>	0.52	0.84	1.02
<b>Station F</b>	1.2	1.62	2.04
<b>Upscaling</b>			
	<b>Background (kton y<sup>-1</sup>)</b>	<b>Min release (kton y<sup>-1</sup>)</b>	<b>Max release (kton y<sup>-1</sup>)</b>
<b>150 – 190 m</b> <b>(2179.518 km<sup>2</sup>)</b>	11.21	18.11	22.00
<b>&gt; 190 m</b> <b>(571.7793 km<sup>2</sup>)</b>	6.79	9.17	11.54
<b>Sum</b>	18.00	27.28	33.54
<b>Excess release</b>		9.28	15.54

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*Data availability* Data presented in this manuscript are available from the VLIZ data repository (<https://doi.org/10.14284/442>).

*Author contribution* AH, SV, MK, EAR and PH designed the study. AH, SV, EAR and PH conducted sediment sampling, MK coordinated lander deployments. AH, SV and ML carried out SEDEX. AH and SV performed statistical analyses, inventory  
510 calculations and upscaling calculations. AH and SV interpreted the data and wrote the manuscript with input from all authors.

*Competing interests* The authors declare that they have no conflict of interest.

*Acknowledgements* We thank the crew of the University of Gothenburg R/V *Skagerak* for support at sea, Martine Leermakers  
515 for assistance with the ICP-OES analysis and Elizabeth Robertson for proofreading the manuscript.

*Financial support* This work was supported by the Swedish Research Council (VR grant 2015-03717 to PH). SVDV was supported by an appointment to the NASA Postdoctoral Program at the University of California, Riverside, administered by Universities Space Research Association under contract with NASA.

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