

# 1 **Biogeosciences Supplemental Information**

## 2 **Will community calcification reflect reef accretion on future, degraded coral reefs?**

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### 5 **Methods**

#### 6 **S.1 Benthic Community Surveys**

##### 7 **S.1.1 Point-Contact Surveys**

8 A transect tape was laid along each 200 m transect length and the occupier of benthic space was  
9 recorded underneath each 1 m interval ( $n = 200 \text{ transect}^{-1}$ ). Categories were divided between coral  
10 (hermatypic), coral (soft), algae (fleshy, non-calcifying), other calcifier (e.g., clams, *Halimeda* spp.,  
11 coralline algae), rubble, and sediment. These surveys were repeated twice per transect at the beginning  
12 of the study (Jan 18-20 2020) to provide an initial understanding of the community structure prior to  
13 flow-metabolism measurements. Data are presented as relative % cover.

##### 14 **S.1.2 Photo Quadrat Surveys**

15 A transect tape was laid along each 200 m transect length and a 1 m<sup>2</sup> PVC quadrat was placed next to  
16 the tape at each 5 m interval ( $n = 40 \text{ transect}^{-1}$ ). A photo was taken of the quadrat and analysed using  
17 ImageJ (Rueden et al., 2017) to quantify the relative areal coverage per 1 m<sup>2</sup> for the following  
18 categories: coral (healthy), coral (unhealthy; paling/bleached), coral (soft) algae (fleshy, non-  
19 calcifying), other calcifier (e.g., clams, *Halimeda* spp., coralline algae), rubble, sediment (clean),  
20 sediment (red with cyanobacteria growth), and sediment (green with Chlorophyta growth). These  
21 surveys were repeated three times throughout the study, at the beginning prior to any observed  
22 bleaching (Jan 24 2020), in the middle after the first observed bleaching event (Feb 6 2020), and at the  
23 end of the study after several more observed bleaching incidents (Feb 13 2020). Data are presented as  
24 relative % cover through time.

### 25 **S.1.3 Mobile Invertebrate Surveys**

26 A transect tape was laid along each 200 m transect length relatively large, easily visible mobile  
27 invertebrates (e.g., sea cucumbers, sea hares, sea urchins) located 1 meter to the left or right along the  
28 transect were counted. Surveys were conducted at dawn to ensure a balance of visibility and  
29 invertebrate activity and repeated 3 times along each transect ( $n = 9 \text{ site}^{-1}$ ). Data are presented as  
30 abundance counts per  $\text{m}^2$  (individuals  $\text{m}^{-2}$ ). Individuals present at less than  $0.1 \text{ m}^{-2}$  were excluded from  
31 the final data reported but were included as part of the invertebrate taxonomy described below.

### 32 **S.1.4 Invertebrate Taxonomy**

33 While conducting the survey approaches detailed above, each time a new invertebrate morphospecies  
34 was encountered, photographs were taken and uploaded to iNaturalist, a biodiversity citizen science  
35 platform where identifications are contributed in real time by both amateur naturalists and professional  
36 taxonomists as part of a consensus system ([www.inaturalist.org](http://www.inaturalist.org)). Using a combination of taxonomic  
37 keys and crowdsourcing via iNaturalist, algae, corals, and other sampled marine invertebrates were  
38 identified to as fine a taxonomic level as possible. These data are presented as presence/absence across  
39 the entire 200 m x 400 m study area. Because sampling was conducted at low tide, most fish usually  
40 present in the lagoon were absent and excluded from benthic survey data.

## 41 **S.2 Lagoon Community Metabolism Measurements**

### 42 **S.2.1 Flow-Respirometry Approach**

43 Flow metabolism transects were established along a reef area previously characterised as degraded,  
44 where there is less than 10 % coral cover (Roelfsema et al., 2018). The flow-respirometry  
45 measurements were conducted within two designated reef areas (100 m x 200 m;  $0.02 \text{ km}^2$ ) which  
46 significantly differed in coral cover. The defined study area was determined based on the necessary  
47 transect length to achieve measurable differences in seawater dissolved oxygen ( $\Delta\text{DO} = \pm 4 - 7 \text{ mg L}^{-1}$ )  
48 <sup>1</sup>) between upstream and downstream locations ( $\sim 200 \text{ m}$ ; Langdon et al., 2010). Repeated

49 deployments of fluorescein dye packets across the research zone at differing tidal periods determined  
50 a specific 400 m x 100 m area of the reef where flow was unidirectional from east to west during a  
51 period spanning from 2 hours before to 1 hour after peak low tide (3 hours total). Outside of this period,  
52 the reef lagoon was no longer physically separated from the open ocean, flow became multidirectional,  
53 and the defined lagoon area became too deep and diluted with open ocean water to measure significant  
54 changes in seawater chemistry. The 400 m x 100 m area was then designated as two,. The spread of  
55 the dye path varied  $\pm 25$  m in a north/south direction and triplicate 200 m transects were spaced 50 m  
56 apart in parallel at each site so that NEC and NEP was averaged across the three downstream locations,  
57 representing all potential water flow paths of the overall study site area. Within each area, three 200m  
58 transects were established in parallel, 50 m distance from one another (Fig. 1). Water samples were  
59 collected as close in time as possible at these fixed upstream and downstream locations ( $n = 3 \text{ area}^{-1}$ )  
60 at peak low tide while lagoon currents were unidirectional, running east to west.

61 
$$\text{Equation 1: NEP} = \frac{3600}{100} \times \frac{\Delta DO \times \rho \times u \times d}{l}$$

62 
$$\text{Equation 2: NEC} = \frac{3600}{100} \times \frac{0.5 \times \Delta TA \times \rho \times u \times d}{l}$$

63 The flow-respirometry approach requires the following measurements: The change in DO and  $A_T$   
64 ( $\Delta DO$  and  $\Delta A_T$ ;  $\text{mmol kg}^{-1}$ ), the mean seawater density ( $\rho$ ;  $\text{kg m}^{-3}$ ), the mean current speed ( $\text{cm s}^{-1}$ ),  
65 the mean depth over the transect ( $d$ ; meters), and the length of the transect ( $l$ ; meters).

66 Salinity (psu) and dissolved oxygen (DO:  $\text{mg L}^{-1}$ ) was measured with a Hanna HI98194 multimeter  
67 and DO converted to  $\mu\text{mol kg}^{-1}$  using seawater density. DO probe calibration was performed weekly  
68 using a two-point calibration at 0% (sodium thiosulfate) and 100% saturated seawater equilibrated  
69 with the atmosphere. Samples for  $A_T$  were collected in 60 ml sample polycarbonate sample bottles,  
70 preserved with saturated Mercuric Chloride according to  $\text{CO}_2$  best practices (Dickson, 2007), and  
71 sealed with a screw top lid and parafilm. Seawater  $A_T$  was analysed by potentiometric titration using a

72 Metrohm 848 Titrino plus automatic titrator (~ 40 ml of seawater per sample) in duplicates (SD  
73 uncertainty < 2  $\mu\text{mol kg}^{-1}$ ). Overall analytical uncertainty for  $A_T$  (SD =  $\pm 2.4 \mu\text{mol kg}^{-1}$ ) measurements  
74 was estimated from repeated measurements of certified reference materials from the Scripps Institute  
75 of Oceanography (CRM; Batch 161).

### 76 **S.2.2 Slack Water Approach**

77 The slack-water approach was used to estimate rates of NEP and NEC over a relatively larger area of  
78 reef (~ 0.3  $\text{km}^2$ ) during a period of three hours around low tide. This period was chosen based on initial  
79 observations of current speed and direction. Starting two hours before peak low tide, the lagoon  
80 becomes separated from the open ocean and the current begins flowing unidirectionally toward the  
81 lagoon outlet to the west. This unidirectional flow behaviour continues until roughly 2 hours after peak  
82 low tide, at which time the flow begins to reverse as the tide fills back in over the reef crest. To avoid  
83 dilution with the open ocean and changing current vector directions confounding residence time  
84 estimates, water samples were collected from the same three locations ( $n = 3 \text{ day}^{-1}$ ) two hours before  
85 peak low tide and one hour following.

$$86 \quad \text{Equation 1: } NEP = \frac{\Delta DO \times \rho \times d}{\Delta t}$$

$$87 \quad \text{Equation 2: } NEC = \frac{0.5 \times \Delta A_T \times \rho \times d}{\Delta t}$$

88 The slack-water approach requires the following measurements: The change in DO and  $A_T$  ( $\Delta DO$  and  
89  $\Delta A_T$ ;  $\text{mmol kg}^{-1}$ ), the mean seawater density ( $\rho$ ;  $\text{kg m}^{-3}$ ), mean depth over the transect ( $d$ ; meters), and  
90 time between sampling ( $\Delta t$ ; hours). Given the time between samples (~ 3 h) and mean current speeds  
91 (~ 20  $\text{cm s}^{-1}$ ), these measurements represent a transect length of roughly 2.5 – 3km of reef.

### 92 **S.2.3 Approach Comparison**

93 Both approaches to estimate benthic community NEP and NEC provide limitations and advantages  
94 with respect to each other (see Langdon et al., 2010). In the flow-respirometry approach, the exact

95 benthic area contributing to measured changes in seawater chemistry is known and its constituents can  
96 be quantified and related to the calculated rates of benthic metabolism. This approach, however,  
97 measures change in alkalinity over a relatively smaller area and time-period. Resulting fluxes in  $A_T$  ( $\pm$   
98  $30 - 60 \mu\text{mol kg}^{-1}$ ) and DO ( $\pm 20 - 50 \mu\text{mol kg}^{-1}$ ) are relatively small compared to the slack-water  
99 approach, thereby providing less confidence in calculated rates of benthic metabolism.

100 In contrast, the slack-water approach benefits from the relatively large changes in total alkalinity ( $A_T$ :  
101  $\pm 100 - 200 \mu\text{mol kg}^{-1}$ ) and dissolved oxygen (DO:  $\pm 80 - 150 \mu\text{mol kg}^{-1}$ ), which provides more  
102 confidence in  $A_T$  anomaly calculations and represent a large area of the reef flat relative to this study's  
103 flow-respirometry estimates. This approach, however, lacks specificity of the exact area of reef  
104 affecting changes in chemistry and DO fluxes are more vulnerable to gas exchange anomalies. As  
105 such, relating metabolic rates to the benthic community provides uncertainties given daily changes in  
106 mean current speed and, subsequently, the area of benthos reflected in the  $A_T$  and DO anomaly.

107 Overall, the combination of both approaches can work in tandem to compensate for their respective  
108 weaknesses. However, neither approach can accommodate dilution with the open ocean and generally  
109 need to be conducted in full sunlight or darkness so that community metabolism does not transition  
110 between autotrophy and heterotrophy in the middle of the measurements. For this reason, community  
111 metabolism estimates were paused from Jan 27 – Feb 2 when peak low tide occurred around dawn and  
112 dusk and changes in DO and  $A_T$  were negligible.

#### 113 **S.2.4 Air-Sea Gas Exchange Corrections**

114 NEP estimates were corrected for the air-sea gas exchange ( $F_{O_2}$ ) of oxygen using the gas-transfer  
115 velocity relationships outlined by Wanninkhof (1992) and Wanninkhof et al., (2009).  $F_{O_2}$  was  
116 calculated with the following equation.

$$117 \quad F_{O_2} = k K_0 (fO_{2_{water}} - fO_{2_{air}})$$

118 where  $k$  is the gas transfer velocity (calculated using and averaged daily wind speed from BOM  
119 data),  $K_0$  is the gas transfer coefficient,  $fO_{2\text{water}}$  is the concentration of seawater dissolved oxygen  
120 ( $\text{mg L}^{-1}$ ) at the time of the downstream measurement,  $fO_{2\text{air}}$  ( $\text{mg L}^{-1}$ ) was assumed to be 100%  
121 saturation at the air temperature over the 3-h measurement period ( $\sim 8.10 \text{ mg L}^{-1}$ ).

### 122 **S.3 Statistical Analyses**

123 All statistical analyses were performed with the SPSS statistics software (SPSS Inc. 2013 Version  
124 26.0). To compare measured differences in benthic cover (percent coral, percent algae, percent  
125 bleached coral tissue, sediment overgrowth) and community metabolism (Net ecosystem production  
126 [NEP] and net ecosystem calcification [NEC]) between triplicate transects, measurement days ( $n =$   
127 12), and Lagoon sites (Lagoon site 1, Lagoon site 2, and Slack Water), a one-way analysis of variance  
128 (ANOVA) model was used in which transect, day, or site was a fixed effect and measured values for  
129 percent cover, NEP, and NEC were treated as the response variable. Results for percent cover  
130 compared among triplicate transects and Lagoon sites are displayed in Tables S1 and S2, respectively.  
131 Before community metabolism measurements were compared, assumptions of normality and equality  
132 of variance were evaluated with a Shapiro Wilk test (Table S4). Results for community metabolism  
133 compared among triplicate transects, measurement days, and Lagoon sites are displayed in Tables S5,  
134 S6, and S7, respectively. A Tukey HSD post-hoc test was used to perform pairwise comparisons for  
135 measured community NEC between Lagoon site 1, Lagoon site 2, and the slack-water approach (Table  
136 S7). To explore relationships between NEC as a function of NEP, Model II regression techniques were  
137 used to test for significant linear relationships (cutoff value  $p < 0.1$ ) and an ANCOVA was used to test  
138 for differences in NEC vs. NEP slope categorized by Lagoon site (Lagoon site 1 and Lagoon site 2).

## 139 **Results**

### 140 **S.4 Invertebrate Taxonomy Results**

141 Overall, we found 25 coral species in the lagoonal reef study area, 22 of which were hard corals and  
142 three soft corals (Fig. 2; Table S8). Thirteen algae morphospecies were observed, with one identified  
143 as species *Valonia ventricosa* and the rest unidentified. Across all other invertebrate taxa, 19 species  
144 of echinoderms, bivalves, and polychaetes, and 24 species of crustaceans and gastropods were  
145 observed. Of the 43 non-coral invertebrate species, 15 were associated with colonies of *Pocillopora*  
146 corals. Sea cucumbers (e.g., *Holothuria* spp., *Stichopus* spp.) were the dominant mobile invertebrate,  
147 the Lollyfish sea cucumber (*Holothuria atra*) was the most common across both Lagoon sites ( $1.2 \pm$   
148  $0.2$  individuals  $m^{-2}$ ). Second in abundance was the Hermann's Sea Cucumber (*Stichopus hermanni*)  
149 ( $0.4 \pm 0.1$  individuals  $m^{-2}$ ). Other notable invertebrates included Linckia sea stars (*Linckia guildingia*,  
150 *Linckia laevigata*) and white-speckled sea hares (*Aplysia argus*) (all found in abundances  $< 0.1$   
151 individuals  $m^{-2}$ ). The largest mobile invertebrates observed were Bailer Shell snails (*Melo amphora*)  
152 at 30 cm in length and white-spotted hermit crabs (*Dardanus megistos*) occupying Bailer shells ( $< 0.1$   
153 individuals  $m^{-2}$ ).

154 Our observations included 8 species with a conservation status of near threatened or higher, including  
155 the small giant clam *Tridacna maxima*, Herrmann's sea cucumber (*Stichopus hermanni*), and 6 coral  
156 species (*Porites attenuata*, *Acropora secale*, *Isopora palifera*, *Stylophora pistillata*, *Favites halicora*,  
157 *Favites rotundata*). Notably, our observation of the aglajid slug *Tubulophilinopsis gardineri* is one of  
158 just 5 from Heron Island, representing the southernmost limit of its eastern coast distribution. We also  
159 observed an undescribed nudibranch species, a yellow-brown *Gymnodoris* (Figure 5). A complete list  
160 of all species described can be found in the Supplemental Material (Table S8).

## 161 **S.5 Lagoon Temperature and Light**

162 Temperature across the Lagoon site 1 exhibited a mean value of  $28.6 \pm 1.5$  °C and varied between a  
163 minimum of 25.8 °C and a maximum of 34.8 °C (Table 2). Light at Lagoon site 1 exhibited a mean  
164 value of  $328 \pm 247$   $\mu\text{mol quanta m}^{-2} \text{ s}^{-1}$  and maximum values of  $1001 \mu\text{mol quanta m}^{-2} \text{ s}^{-1}$  (Fig. 1).  
165 Temperature across Lagoon site 2 exhibited a mean value of  $28.6 \pm 1.5$  °C and varied between a

166 minimum of 25.9 °C and a maximum of 34.6 °C. Light at Lagoon site 2 exhibited a mean value of 336  
167  $\pm 254 \mu\text{mol quanta m}^{-2} \text{ s}^{-1}$  and maximum values of  $969 \mu\text{mol quanta m}^{-2} \text{ s}^{-1}$ .

## 168 **S.6 Lagoon Community Bleaching Extent**

169 Dark-adapted yield was  $0.662 \pm 0.010$  for *Acropora* spp. fragments and  $0.576 \pm 0.020$  for “Other”  
170 fragments (mean  $\pm$  SE, n = 35) on Feb 4<sup>th</sup>. On Feb 9<sup>th</sup>, yield declined 35% for *Acropora* spp. to  $0.430$   
171  $\pm 0.014$  (n = 15) and 25% for “Other” fragments to  $0.434 \pm 0.018$  (n = 20). Symbiodiniaceae densities  
172 were  $0.976 \pm 0.135 \times 10^6 \text{ cm}^{-2}$  for *Acropora* spp. (n = 15) and  $0.507 \pm 0.160 \times 10^6 \text{ cm}^{-2}$  for “Other”  
173 fragments (n = 10) on Jan 30<sup>th</sup>. On Feb 12<sup>th</sup>, *Acropora* spp. densities had declined by 48% to  $0.504 \pm$   
174  $0.0849 \times 10^6 \text{ cm}^{-2}$  (n = 15) and by 18% for “Other” fragments to  $0.414 \pm 0.094 \times 10^6 \text{ cm}^{-2}$  (n = 15) (Fig.  
175 3).

## 176 **Discussion**

### 177 **S.7 Estimated Organism Contribution to NEC at Elevated Temperatures**

178 To estimate the potential effect of a +1.1 °C change in seawater temperature on coral calcification for  
179 corals observed within the lagoon study sites the following aquaria manipulation studies were  
180 reviewed: Edmunds, 2005; Anthony et al., 2008; Cantin et al., 2010; Comeau et al., 2013, 2016; and  
181 the following meta-analysis and modeling studies were reviewed: Lough and Barnes, 2000; McNeil et  
182 al., 2004; Evenhuis et al., 2015; Kornder et al., 2018; Bove et al., 2020.



183 **Tables**

184 Table S1: One-way ANOVA results (p-values) comparing measured percent coral and algae cover  
 185 between triplicate transects within each Lagoon site (Lagoon site 1, Lagoon site 2). Data were pooled  
 186 among replicate point-contact survey efforts (n = 2 transect<sup>-1</sup>). A **bolded** value (p-value < 0.05)  
 187 indicates that the percent cover significantly differed between transects within each Lagoon site.

<b>Point-Contact Survey Method</b>				
<b>Cover</b>	<b>Lagoon site 1</b>		<b>Lagoon site 2</b>	
	<b>df</b>	<b>p-value</b>	<b>df</b>	<b>p-value</b>
% Coral Cover	2	0.791	2	0.959
% Algae Cover	2	0.256	2	0.214
% Sediment Cover	2	0.421	2	0.956

188

189

190 Table S2: One-way ANOVA results (p-values) comparing measured percent coral and algae cover  
 191 between Lagoon site 1 and Lagoon site 2. Data were pooled among replicate point-contact survey  
 192 efforts and triplicate transects within each Lagoon site (n = 6 site<sup>-1</sup>). A **bolded** value (p-value < 0.05)  
 193 indicates that the percent cover significantly differed between Lagoon sites.

<b>Point-Contact Survey Method</b>		
<b>Cover</b>	<b>df</b>	<b>p - value</b>
% Coral Cover	<b>1</b>	<b>0.001</b>
% Algae Cover	<b>1</b>	<b>0.011</b>
% Sediment Cover	1	0.122

194

195 Table S3: One-way ANOVA results (p-values) comparing measured percent coral and algae cover  
 196 between triplicate transects within each Lagoon site (Lagoon site 1, Lagoon site 2). Data were pooled  
 197 among triplicate photo-quadrat survey efforts over time (n = 120 transect<sup>-1</sup>). A **bolded** value (p-value  
 198 < 0.05) indicates that the percent cover significantly differed between transects.

<b>Photo-Quadrat Survey Method</b>				
<b>Cover</b>	<b>Lagoon site 1</b>		<b>Lagoon site 2</b>	
	<b>df</b>	<b>p-value</b>	<b>df</b>	<b>p-value</b>
% Coral Cover	2	0.469	2	0.818
% Algae Cover	2	0.721	2	0.796
% Sediment Cover	2	0.859	2	0.403

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200

201

202

203

204

205 Table S4: One-way ANOVA results (p-values) comparing measured percent coral and algae cover  
 206 between Lagoon site 1 and Lagoon site 2. Data were pooled among triplicate photo-quadrat survey  
 207 efforts and triplicate transects within each Lagoon site (n = 360 site<sup>-1</sup>). A **bolded** value (p-value < 0.05)  
 208 indicates that the percent cover significantly differed between Lagoon site 1 and Lagoon site 2.

Photo-Quadrat Survey Method		
Cover	df	p - value
% Coral Cover	<b>1</b>	<b>0.000</b>
% Algae Cover	1	0.273
% Sediment Cover	1	0.140

209

210 Table S5: One-way ANOVA results for percent bleached coral tissue (Coral Bleaching) and percent  
 211 sediment exhibiting overgrowth (Sediment Overgrowth) compared over the three survey efforts  
 212 through time (Jan 24, Feb 6, and Feb12 2020) at Lagoon site 1. Data were pooled among all triplicate  
 213 transects. Tukey HSD post-hoc test results are to compare differences between each survey effort (n =  
 214 3). A **bolded** value (p-value < 0.05) indicates that the difference was significant between time points.

Photo-Quadrat Survey Method: Lagoon site 1							
Lagoon site 1		df	F-value	p - value			
Coral Bleaching		<b>2</b>	<b>67.2</b>	<b>0.000</b>			
Sediment Overgrowth		<b>2</b>	<b>18.3</b>	<b>0.003</b>			
Tukey HSD							
Dependent Variable	(I) Time	(J) Time	Mean	Std. Error	Sig.	95% Confidence Interval	
			Difference (I-J)			Lower Bound	Upper Bound
Coral Bleaching	Jan 24	Feb 6	<b>-16.33</b>	<b>4.93</b>	<b>.037</b>	<b>-31.48</b>	<b>-1.18</b>
		Feb 12	<b>-55.66</b>	<b>4.93</b>	<b>.000</b>	<b>-70.81</b>	<b>-40.51</b>
	Feb 6	Jan 24	<b>16.33</b>	<b>4.93</b>	<b>.037</b>	<b>1.18</b>	<b>31.48</b>
		Feb 12	<b>-39.33</b>	<b>4.93</b>	<b>.001</b>	<b>-54.48</b>	<b>-24.18</b>
	Jan 24	Feb 6	<b>55.66</b>	<b>4.93</b>	<b>.000</b>	<b>40.51</b>	<b>70.81</b>
		Feb 12	<b>39.33</b>	<b>4.93</b>	<b>.001</b>	<b>24.18</b>	<b>54.48</b>
Sediment Overgrowth	Jan 24	Feb 6	-2.33	1.36	.275	-6.50	1.84
		Feb 12	<b>-8.00</b>	<b>1.36</b>	<b>.003</b>	<b>-12.17</b>	<b>-3.82</b>
	Feb 6	Jan 24	2.33	1.36	.275	-1.84	6.50
		Feb 12	<b>-5.66</b>	<b>1.36</b>	<b>.014</b>	<b>-9.84</b>	<b>-1.49</b>
	Jan 24	Feb 6	<b>8.00</b>	<b>1.36</b>	<b>.003</b>	<b>3.82</b>	<b>12.17</b>
		Feb 12	<b>5.66</b>	<b>1.36</b>	<b>.014</b>	<b>1.49</b>	<b>9.84</b>

215

216 Table S6: One-way ANOVA results for percent bleached coral tissue (Coral Bleaching) and percent  
 217 sediment exhibiting overgrowth (Sediment Overgrowth) compared over the three survey efforts  
 218 through time (Jan 24, Feb 6, and Feb12 2020) at Lagoon site 2. Data were pooled among all triplicate  
 219 transects. Tukey HSD post-hoc test results are to compare differences between each survey effort (n =  
 220 3). A **bolded** value (p-value < 0.05) indicates that the difference was significant between time points.

<b>Photo-Quadrat Survey Method: Lagoon site 2</b>							
<b>Lagoon site 2</b>		<b>df</b>	<b>F-value</b>	<b>p - value</b>			
Coral Bleaching		<b>2</b>	<b>142.9</b>	<b>.000</b>			
Sediment Overgrowth		<b>2</b>	<b>10.5</b>	<b>.011</b>			
<b>Tukey HSD</b>							
Dependent Variable (I)	Time (J)	Time (I-J)	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
						Lower Bound	Upper Bound
Coral Bleaching	Jan 24	Feb 6	<b>-24.00</b>	<b>3.88</b>	<b>.002</b>	<b>-35.92</b>	<b>-12.07</b>
		Feb 12	<b>-65.00</b>	<b>3.88</b>	<b>.000</b>	<b>-76.92</b>	<b>-53.07</b>
	Feb 6	Jan 24	<b>24.00</b>	<b>3.88</b>	<b>.002</b>	<b>12.07</b>	<b>35.92</b>
		Feb 12	<b>-41.00</b>	<b>3.88</b>	<b>.000</b>	<b>-52.92</b>	<b>-29.07</b>
	Jan 24	Feb 6	<b>65.00</b>	<b>3.88</b>	<b>.000</b>	<b>53.07</b>	<b>76.92</b>
		Feb 12	<b>41.00</b>	<b>3.88</b>	<b>.000</b>	<b>29.07</b>	<b>52.92</b>
Sediment Overgrowth	Jan 24	Feb 6	-3.00	2.8	.564	-11.59	5.59
		Feb 12	<b>-12.33</b>	<b>2.80</b>	<b>.011</b>	<b>-20.93</b>	<b>-3.73</b>
	Feb 6	Jan 24	3.00	2.80	.564	-5.59	11.59
		Feb 12	<b>-9.33</b>	<b>2.80</b>	<b>.036</b>	<b>-17.93</b>	<b>-.73</b>
	Jan 24	Feb 6	<b>12.33</b>	<b>2.80</b>	<b>.011</b>	<b>3.73</b>	<b>20.93</b>
		Feb 12	<b>9.33</b>	<b>2.80</b>	<b>.036</b>	<b>.73</b>	<b>17.93</b>

221 Table S7: One-way ANOVA results for percent bleached coral tissue (Coral Bleaching) and percent  
 222 sediment exhibiting overgrowth (Sediment Overgrowth) compared over the three survey efforts  
 223 through time (Jan 24, Feb 6, and Feb12 2020) between Lagoon site 1 and Lagoon site 2. Data were  
 224 pooled among all triplicate transects. A **bolded** value (p-value < 0.05) indicates that the difference was  
 225 significant between Lagoon sites.

Photo-Quadrat Survey Method				
	Coral Bleaching		Sediment Overgrowth	
Date	df	p - value	df	p - value
Jan 24 2020	1	1.00	1	0.899
Feb 6 2020	1	0.067	1	0.692
Feb 12 2020	1	0.256	1	0.231

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227 Table S8: List of invertebrate taxonomy described in section 3.2.4.

Group	Taxon	Common name
Algae	<i>Caulerpa</i> spp.	
	Chlorophyta spp.	Green algae
	<i>Halimeda</i> spp.	
	<i>Laurencia</i> spp.	
	<i>Padina</i> sp.	
Corals	Rhodophyta spp.	Red algae
	<i>Valonia ventricosa</i>	Sailor's eyeball alga
	<i>Acropora secale</i>	
	<i>Acropora millepora</i>	
	<i>Acropora muricata</i>	
	<i>Acropora</i> spp.	Staghorn corals
	<i>Astrea curta</i>	
	<i>Cyphastrea chalcidicum</i>	
	<i>Dipsastraea</i> sp.	
	<i>Favites halicora</i>	
<i>Favites rotundata</i>		
<i>Goniastrea edwardsi</i>	Honeycomb coral	
<i>Goniopora</i> sp.	Flowerpot coral	
<i>Isopora palifera</i>		

	<i>Klyxum</i> sp.	
	<i>Lobophyllia agaricia</i>	
	<i>Montipora digitata</i>	
	<i>Montipora grisea</i>	
	<i>Montipora hispida</i>	
	<i>Montipora</i> sp.	
	<i>Platygyra daedalea</i>	Lesser valley coral
	<i>Platygyra</i> spp.	
	<i>Pocillopora damicornis</i>	
	<i>Pocillopora</i> sp.	Cauliflower coral
	<i>Porites attenuate</i>	
	<i>Porites cylindrica</i>	Yellow finger coral
	<i>Porites</i> sp.	Pore coral
	<i>Sarcophyton</i> spp.	Toadstool leather corals
	<i>Stylophora pistillata</i>	Hood coral
Crustaceans	Alpheidae sp.	Snapping shrimp
	<i>Alpheus</i> sp.	Snapping shrimp
	Brachyura spp.	Crabs
	<i>Calcinus latens</i>	Hidden hermit crab
	Caridea sp.	Caridean shrimp
	<i>Clibanarius corallinus</i>	Coral hermit crab
	<i>Dardanus megistos</i>	White-spotted hermit crab
	Majidae sp.	Spider crab
	Stomatopoda spp.	Mantis shrimps
	<i>Thalamita</i> sp.	
	<i>Trapezia serenei</i>	Coral crab
	<i>Zenopontonia soror</i>	Seastar shrimp
Echinoderms	<i>Culcita novaeguineae</i>	Pillow cushion star
	<i>Holothuria atra</i>	Lollyfish sea cucumber
	<i>Holothuria edulis</i>	Pinkfish sea cucumber
	<i>Holothuria leucospilota</i>	Black sea cucumber

	<i>Holothuria</i> sp.	
	<i>Linckia guildingi</i>	Guilding's sea star
	<i>Linckia laevigata</i>	Blue linckia
	<i>Nardoa novaecaledoniae</i>	Yellow mesh sea star
	<i>Stichopus herrmanni</i>	Herrmann's sea cucumber
	<i>Stichopus chloronotus</i>	Greenfish sea cucumber
Molluscs	<i>Aplysia argus</i>	White-speckled seahare
	<i>Atactodea striata</i>	Striate beach clam
	<i>Codakia paytenorum</i>	Payten's codakia
	<i>Chrysostoma paradoxum</i>	Orange-mouthed top shell
	<i>Clypeomorus bifasciata</i>	Double-banded creeper
	<i>Coralliophila</i> sp.	
	Ergalataxinae	
	<i>Gymnodoris</i> sp.	
	<i>Melo amphora</i>	Giant baler
	<i>Pitar</i> sp.	
	<i>Spondylus</i> sp.	Thorny oyster
	<i>Tectus fenestratus</i>	Latticed top shell
	<i>Tonna chinensis</i>	China tun
	<i>Tridacna maxima</i>	Small giant clam
	<i>Tubulophilinopsis gardineri</i>	Gardiner's headshield slug
	<i>Turbo argyrostomus</i>	Silvermouth turban
Polychaetes	<i>Perinereis</i> sp.	
	<i>Spirobranchus</i> sp.	Christmas tree worm
	Terebellidae sp.	Spaghetti worm
Sponges	Porifera sp.	

228 Table S9: Shapiro-Wilk test for normality in reef metabolism. Data are organized by rates of NEP and  
 229 NEC measured at Lagoon site 1, Lagoon site 2, and the larger lagoon area (Slack Water). Data for each  
 230 Lagoon site were pooled among triplicate parallel transects. NEP data were not included for the slack-  
 231 water method. If the significant value (Sig.) of the test is > 0.05 the data exhibit a normal distribution.

		Shapiro-Wilk		
	Site	Statistic	df	Sig.
NEP	Lagoon site 1	.951	36	.112
	Lagoon site 2	.984	36	.857
	Slack Water			
NEC	Lagoon site 1	.967	36	.356
	Lagoon site 2	.952	36	.117
	Slack Water	.962	33	.287

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238 Table S10: One-way ANOVA results (p-values) comparing measured reef metabolism (NEP and NEC)  
 239 between triplicate transects within each Lagoon site (Lagoon site 1, Lagoon site 2, and Slack Water).  
 240 Data were pooled among all 11 (Slack water) and 12 (Lagoon site 1 and Lagoon site 2) days of  
 241 measurements (3 days for Night NEC). A **bolded** value (p-value < 0.05) indicates that the measured  
 242 response in that specific metabolic parameter significantly differed between triplicate transects.

Metabolism	Lagoon site 1		Lagoon site 2		Slack Water	
	df	p-value	df	p-value	df	p-value
NEP	2	.471	2	.917		
NEC	2	.169	2	.489	2	.581
Night NEC					2	.617

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247 Table S11: One-way ANOVA results (p-values) comparing measured reef metabolism (NEP and NEC)  
 248 between measurement days within each Lagoon site (Lagoon site 1 and Lagoon site 2 = 12; Slack  
 249 Water = 11; Night NEC = 3). Data were pooled among all triplicate transects. A **bolded** value (p-value  
 250 < 0.05) indicates that the measured response in that specific metabolic parameter significantly differed  
 251 between triplicate transects.

Metabolism	Lagoon site 1		Lagoon site 2		Slack Water	
	df	p-value	df	p-value	df	p-value
NEP	11	.181	11	.099		
NEC	11	.506	11	.365	10	.073
Night NEC					2	.083

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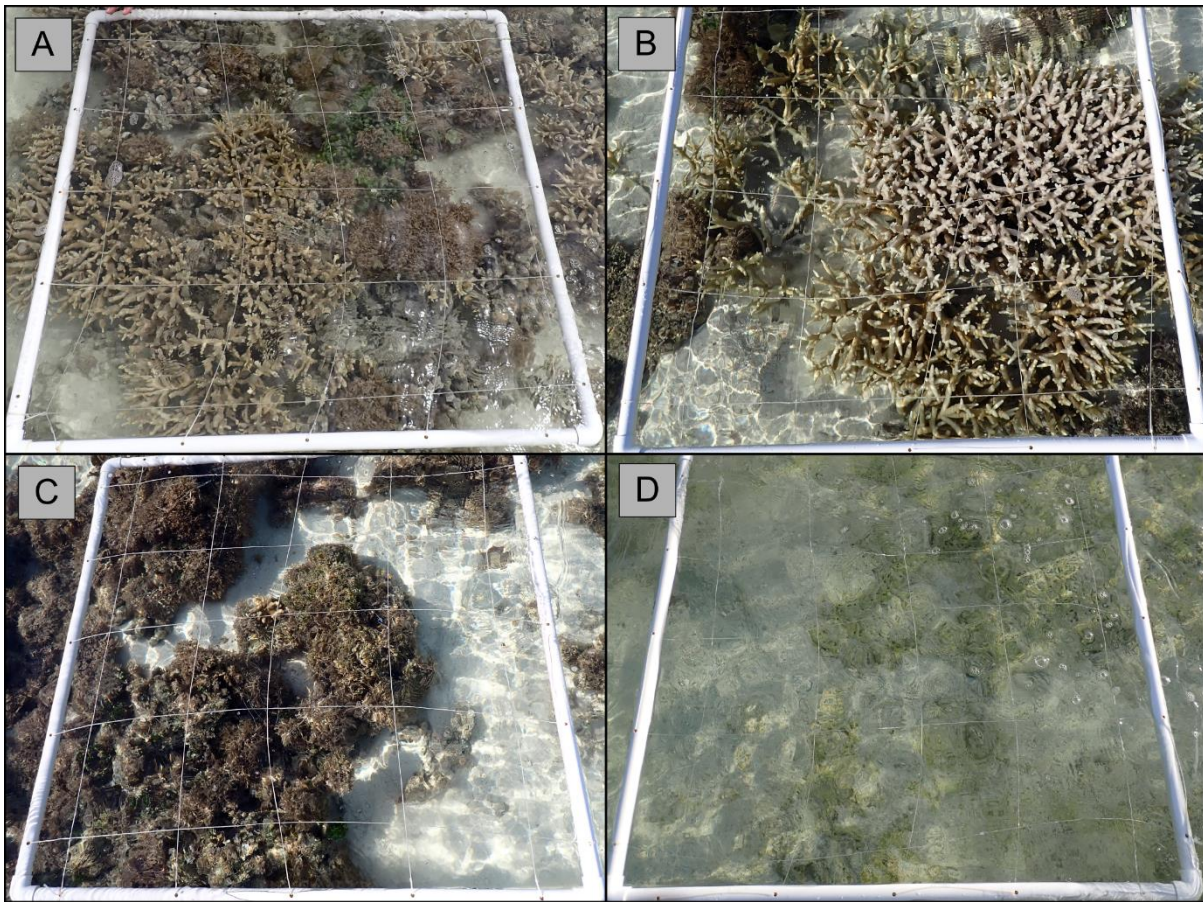


253 Table S12: One-way ANOVA results for NEP compared amongst Lagoon site 1 and Lagoon site 2 and  
 254 for NEC compared amongst Lagoon site 1, Lagoon site 2, and Slack Water. Data were pooled among  
 255 all triplicate transects and measurements days. Tukey HSD post-hoc test results are displayed for NEC  
 256 (n = 3). A **bolded** value (p-value < 0.05) indicates that the difference was significant between Lagoon  
 257 sites.

Metabolism		df	F-value	p - value
NEP		1	3.47	.067
NEC		<b>2</b>	<b>8.17</b>	<b>.001</b>

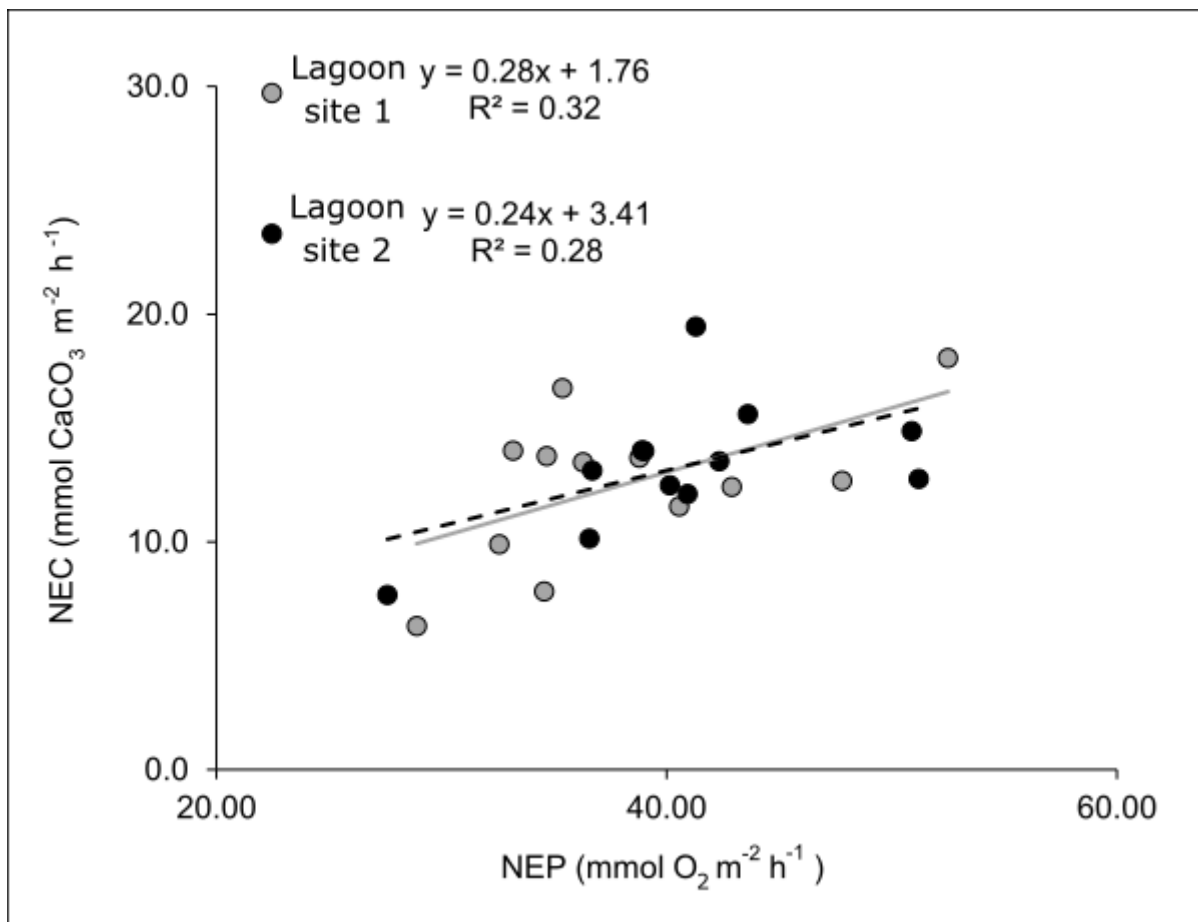
  

Tukey HSD						
(I) Site (J) Site		Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
Lagoon site 1	Lagoon site 2	-.8742	1.015	.666	-3.2916	1.5431
	Slack Water	<b>3.0361*</b>	<b>1.015</b>	<b>.010</b>	<b>.6187</b>	<b>5.4534</b>
Lagoon site 2	Lagoon site 1	.8742	1.015	.666	-1.5431	3.2916
	Slack Water	<b>3.9103*</b>	<b>1.015</b>	<b>.001</b>	<b>1.4929</b>	<b>6.3277</b>
Slack Water	Lagoon site 1	<b>-3.0361*</b>	<b>1.015</b>	<b>.010</b>	<b>-5.4534</b>	<b>-.6187</b>
	Lagoon site 2	<b>-3.9103*</b>	<b>1.01544</b>	<b>.001</b>	<b>-6.3277</b>	<b>-1.4929</b>



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260 Figure S.1: Photo-quadrat examples of various reef health. A) Healthy *Acropora* spp. coral observed  
261 during the first survey effort. B) Bleached *Acropora* spp. observed during the final survey effort. C)  
262 Example of fleshy algal growth as the dominant benthic organism D) Example of Chlorophyta  
263 overgrowth on the sediment.



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265 Figure S.2: Rates of net ecosystem calcification (NEC) as a function of net ecosystem production  
 266 (NEP) separated between study Lagoon site 1 (grey) and Lagoon site 2 (black).

267 **Supplemental Information References**

- 268 Anthony, K. R. N., Kline, D. I., Diaz-Pulido, G., Dove, S. and Hoegh-Guldberg, O.: Ocean  
269 acidification causes bleaching and productivity loss in coral reef builders, *Proc. Natl. Acad. Sci.*  
270 *U. S. A.*, 105(45), 17442–17446, doi:10.1073/pnas.0804478105, 2008.
- 271 Bove, C. B., Umbanhowar, J. and Castillo, K. D.: Meta-Analysis Reveals Reduced Coral Calcification  
272 Under Projected Ocean Warming but Not Under Acidification Across the Caribbean Sea, *Front.*  
273 *Mar. Sci.*, 7, 127, doi:10.3389/fmars.2020.00127, 2020.
- 274 Cantin, N. E., Cohen, A. L., Karnauskas, K. B., Tarrant, A. M. and McCorkle, D. C.: Ocean warming  
275 slows coral growth in the central Red Sea, *Science* (80-. ), 329(5989), 322–325,  
276 doi:10.1126/science.1190182, 2010.
- 277 Comeau, S., Edmunds, P. J., Spindel, N. B. and Carpenter, R. C.: The responses of eight coral reef  
278 calcifiers to increasing partial pressure of CO<sub>2</sub> do not exhibit a tipping point, *Limnol.*  
279 *Oceanogr.*, 58(1), 388–398, doi:10.4319/lo.2013.58.1.0388, 2013.
- 280 Comeau, S., Carpenter, R. C., Lantz, C. A. and Edmunds, P. J.: Parameterization of the response of  
281 calcification to temperature and pCO<sub>2</sub> in the coral *Acropora pulchra* and the alga *Lithophyllum*  
282 *kotschyianum*, *Coral Reefs*, 35(3), 929–939, doi:10.1007/s00338-016-1425-0, 2016.
- 283 Dickson, A. G., Sabine, C. L. and Christian, J. R.: Guide to best practices for ocean CO<sub>2</sub>  
284 measurements, North Pacific Marine Science Organization., 2007.
- 285 Edmunds, P. J.: The effect of sub-lethal increases in temperature on the growth and population  
286 trajectories of three scleractinian corals on the southern Great Barrier Reef, *Oecologia*, 146(3),  
287 350–364, doi:10.1007/s00442-005-0210-5, 2005.
- 288 Evenhuis, C., Lenton, A., Cantin, N. E. and Lough, J. M.: Modelling coral calcification accounting  
289 for the impacts of coral bleaching and ocean acidification, *Biogeosciences*, 12(9), 2607–2630,  
290 doi:10.5194/bg-12-2607-2015, 2015.
- 291 Kornder, N. A., Riegl, B. M. and Figueiredo, J.: Thresholds and drivers of coral calcification  
292 responses to climate change, *Glob. Chang. Biol.*, 24(11), 5084–5095, doi:10.1111/gcb.14431,  
293 2018.
- 294 Langdon, C., Gattuso, J.-P., Andersson, A., Océanologique, O. and Pierre, U.: Part 3 : Measurements  
295 of CO<sub>2</sub> - sensitive processes 13 Measurements of calcifi cation and dissolution of benthic  
296 organisms and communities Part 3 : Measurements of CO<sub>2</sub> - sensitive processes, , 213–232,  
297 2010.
- 298 Lough, J. M. and Barnes, D. J.: Environmental controls on growth of the massive coral *Porites*, J.  
299 *Exp. Mar. Bio. Ecol.*, 245(2), 225–243, doi:10.1016/S0022-0981(99)00168-9, 2000.
- 300 McNeil, B. I., Matear, R. J. and Barnes, D. J.: Coral reef calcification and climate change: The effect  
301 of ocean warming, *Geophys. Res. Lett.*, 31(22), 1–4, doi:10.1029/2004GL021541, 2004.
- 302 Roelfsema, C., Kovacs, E., Ortiz, J. C., Wolff, N. H., Callaghan, D., Wettle, M., Ronan, M.,  
303 Hamylton, S. M., Mumby, P. J. and Phinn, S.: Coral reef habitat mapping: A combination of  
304 object-based image analysis and ecological modelling, *Remote Sens. Environ.*, 208, 27–41,  
305 doi:10.1016/j.rse.2018.02.005, 2018.

- 306 Wanninkhof, R.: Relationship between wind speed and gas exchange over the ocean, *J. Geophys.*  
307 *Res.*, 97(C5), 7373–7382, doi:10.1029/92JC00188, 1992.
- 308 Wanninkhof, R., Asher, W. E., Ho, D. T., Sweeney, C. and McGillis, W. R.: Advances in  
309 Quantifying Air-Sea Gas Exchange and Environmental Forcing, *Ann. Rev. Mar. Sci.*, 1(1), 213–  
310 244, doi:10.1146/annurev.marine.010908.163742, 2009.