

1 **Biogeosciences Supplemental Information**

2 **Will community calcification reflect reef accretion on future, degraded coral reefs?**

3 Coulson A. Lantz^{1,2}, William Leggat², Jessica L. Bergman¹, Alexander Fordyce², Charlotte Page¹,
4 Thomas Mesaglio¹, Tracy D. Ainsworth¹

5 **Methods**

6 **S.1 Benthic Community Surveys**

7 **S.1.1 Point-Contact Surveys**

8 A transect tape was laid along each 200 m transect length and the occupier of benthic space was
9 recorded underneath each 1 m interval ($n = 200 \text{ transect}^{-1}$). Categories were divided between coral
10 (hermatypic), coral (soft), algae (fleshy, non-calcifying), other calcifier (e.g., clams, *Halimeda* spp.,
11 coralline algae), rubble, and sediment. These surveys were repeated twice per transect at the beginning
12 of the study (Jan 18-20 2020) to provide an initial understanding of the community structure prior to
13 flow-metabolism measurements. Data are presented as relative % cover.

14 **S.1.2 Photo Quadrat Surveys**

15 A transect tape was laid along each 200 m transect length and a 1 m² PVC quadrat was placed next to
16 the tape at each 5 m interval ($n = 40 \text{ transect}^{-1}$). A photo was taken of the quadrat and analysed using
17 ImageJ (Rueden et al., 2017) to quantify the relative areal coverage per 1 m² for the following
18 categories: coral (healthy), coral (unhealthy; paling/bleached), coral (soft) algae (fleshy, non-
19 calcifying), other calcifier (e.g., clams, *Halimeda* spp., coralline algae), rubble, sediment (clean),
20 sediment (red with cyanobacteria growth), and sediment (green with Chlorophyta growth). These
21 surveys were repeated three times throughout the study, at the beginning prior to any observed
22 bleaching (Jan 24 2020), in the middle after the first observed bleaching event (Feb 6 2020), and at the
23 end of the study after several more observed bleaching incidents (Feb 13 2020). Data are presented as
24 relative % cover through time.

25 **S.1.3 Mobile Invertebrate Surveys**

26 A transect tape was laid along each 200 m transect length relatively large, easily visible mobile
27 invertebrates (e.g., sea cucumbers, sea hares, sea urchins) located 1 meter to the left or right along the
28 transect were counted. Surveys were conducted at dawn to ensure a balance of visibility and
29 invertebrate activity and repeated 3 times along each transect ($n = 9 \text{ site}^{-1}$). Data are presented as
30 abundance counts per m^2 (individuals m^{-2}). Individuals present at less than 0.1 m^{-2} were excluded from
31 the final data reported but were included as part of the invertebrate taxonomy described below.

32 **S.1.4 Invertebrate Taxonomy**

33 While conducting the survey approaches detailed above, each time a new invertebrate morphospecies
34 was encountered, photographs were taken and uploaded to iNaturalist, a biodiversity citizen science
35 platform where identifications are contributed in real time by both amateur naturalists and professional
36 taxonomists as part of a consensus system (www.inaturalist.org). Using a combination of taxonomic
37 keys and crowdsourcing via iNaturalist, algae, corals, and other sampled marine invertebrates were
38 identified to as fine a taxonomic level as possible. These data are presented as presence/absence across
39 the entire 200 m x 400 m study area. Because sampling was conducted at low tide, most fish usually
40 present in the lagoon were absent and excluded from benthic survey data.

41 **S.2 Lagoon Community Metabolism Measurements**

42 **S.2.1 Flow-Respirometry Approach**

43 Flow metabolism transects were established along a reef area previously characterised as degraded,
44 where there is less than 10 % coral cover (Roelfsema et al., 2018). The flow-respirometry
45 measurements were conducted within two designated reef areas (100 m x 200 m; 0.02 km^2) which
46 significantly differed in coral cover. The defined study area was determined based on the necessary
47 transect length to achieve measurable differences in seawater dissolved oxygen ($\Delta\text{DO} = \pm 4 - 7 \text{ mg L}^{-1}$)
48 ¹) between upstream and downstream locations ($\sim 200 \text{ m}$; Langdon et al., 2010). Repeated

49 deployments of fluorescein dye packets across the research zone at differing tidal periods determined
50 a specific 400 m x 100 m area of the reef where flow was unidirectional from east to west during a
51 period spanning from 2 hours before to 1 hour after peak low tide (3 hours total). Outside of this period,
52 the reef lagoon was no longer physically separated from the open ocean, flow became multidirectional,
53 and the defined lagoon area became too deep and diluted with open ocean water to measure significant
54 changes in seawater chemistry. The 400 m x 100 m area was then designated as two,. The spread of
55 the dye path varied ± 25 m in a north/south direction and triplicate 200 m transects were spaced 50 m
56 apart in parallel at each site so that NEC and NEP was averaged across the three downstream locations,
57 representing all potential water flow paths of the overall study site area. Within each area, three 200m
58 transects were established in parallel, 50 m distance from one another (Fig. 1). Water samples were
59 collected as close in time as possible at these fixed upstream and downstream locations ($n = 3 \text{ area}^{-1}$)
60 at peak low tide while lagoon currents were unidirectional, running east to west.

61
$$\text{Equation 1: NEP} = \frac{3600}{100} \times \frac{\Delta DO \times \rho \times u \times d}{l}$$

62
$$\text{Equation 2: NEC} = \frac{3600}{100} \times \frac{0.5 \times \Delta TA \times \rho \times u \times d}{l}$$

63 The flow-respirometry approach requires the following measurements: The change in DO and A_T
64 (ΔDO and ΔA_T ; mmol kg^{-1}), the mean seawater density (ρ ; kg m^{-3}), the mean current speed (cm s^{-1}),
65 the mean depth over the transect (d ; meters), and the length of the transect (l ; meters).

66 Salinity (psu) and dissolved oxygen (DO: mg L^{-1}) was measured with a Hanna HI98194 multimeter
67 and DO converted to $\mu\text{mol kg}^{-1}$ using seawater density. DO probe calibration was performed weekly
68 using a two-point calibration at 0% (sodium thiosulfate) and 100% saturated seawater equilibrated
69 with the atmosphere. Samples for A_T were collected in 60 ml sample polycarbonate sample bottles,
70 preserved with saturated Mercuric Chloride according to CO_2 best practices (Dickson, 2007), and
71 sealed with a screw top lid and parafilm. Seawater A_T was analysed by potentiometric titration using a

72 Metrohm 848 Titrino plus automatic titrator (~ 40 ml of seawater per sample) in duplicates (SD
73 uncertainty < 2 $\mu\text{mol kg}^{-1}$). Overall analytical uncertainty for A_T (SD = $\pm 2.4 \mu\text{mol kg}^{-1}$) measurements
74 was estimated from repeated measurements of certified reference materials from the Scripps Institute
75 of Oceanography (CRM; Batch 161).

76 **S.2.2 Slack Water Approach**

77 The slack-water approach was used to estimate rates of NEP and NEC over a relatively larger area of
78 reef (~ 0.3 km^2) during a period of three hours around low tide. This period was chosen based on initial
79 observations of current speed and direction. Starting two hours before peak low tide, the lagoon
80 becomes separated from the open ocean and the current begins flowing unidirectionally toward the
81 lagoon outlet to the west. This unidirectional flow behaviour continues until roughly 2 hours after peak
82 low tide, at which time the flow begins to reverse as the tide fills back in over the reef crest. To avoid
83 dilution with the open ocean and changing current vector directions confounding residence time
84 estimates, water samples were collected from the same three locations ($n = 3 \text{ day}^{-1}$) two hours before
85 peak low tide and one hour following.

$$86 \quad \text{Equation 1: } NEP = \frac{\Delta DO \times \rho \times d}{\Delta t}$$

$$87 \quad \text{Equation 2: } NEC = \frac{0.5 \times \Delta A_T \times \rho \times d}{\Delta t}$$

88 The slack-water approach requires the following measurements: The change in DO and A_T (ΔDO and
89 ΔA_T ; mmol kg^{-1}), the mean seawater density (ρ ; kg m^{-3}), mean depth over the transect (d ; meters), and
90 time between sampling (Δt ; hours). Given the time between samples (~ 3 h) and mean current speeds
91 (~ 20 cm s^{-1}), these measurements represent a transect length of roughly 2.5 – 3km of reef.

92 **S.2.3 Approach Comparison**

93 Both approaches to estimate benthic community NEP and NEC provide limitations and advantages
94 with respect to each other (see Langdon et al., 2010). In the flow-respirometry approach, the exact

95 benthic area contributing to measured changes in seawater chemistry is known and its constituents can
96 be quantified and related to the calculated rates of benthic metabolism. This approach, however,
97 measures change in alkalinity over a relatively smaller area and time-period. Resulting fluxes in A_T (\pm
98 $30 - 60 \mu\text{mol kg}^{-1}$) and DO ($\pm 20 - 50 \mu\text{mol kg}^{-1}$) are relatively small compared to the slack-water
99 approach, thereby providing less confidence in calculated rates of benthic metabolism.

100 In contrast, the slack-water approach benefits from the relatively large changes in total alkalinity (A_T :
101 $\pm 100 - 200 \mu\text{mol kg}^{-1}$) and dissolved oxygen (DO: $\pm 80 - 150 \mu\text{mol kg}^{-1}$), which provides more
102 confidence in A_T anomaly calculations and represent a large area of the reef flat relative to this study's
103 flow-respirometry estimates. This approach, however, lacks specificity of the exact area of reef
104 affecting changes in chemistry and DO fluxes are more vulnerable to gas exchange anomalies. As
105 such, relating metabolic rates to the benthic community provides uncertainties given daily changes in
106 mean current speed and, subsequently, the area of benthos reflected in the A_T and DO anomaly.

107 Overall, the combination of both approaches can work in tandem to compensate for their respective
108 weaknesses. However, neither approach can accommodate dilution with the open ocean and generally
109 need to be conducted in full sunlight or darkness so that community metabolism does not transition
110 between autotrophy and heterotrophy in the middle of the measurements. For this reason, community
111 metabolism estimates were paused from Jan 27 – Feb 2 when peak low tide occurred around dawn and
112 dusk and changes in DO and A_T were negligible.

113 **S.2.4 Air-Sea Gas Exchange Corrections**

114 NEP estimates were corrected for the air-sea gas exchange (F_{O_2}) of oxygen using the gas-transfer
115 velocity relationships outlined by Wanninkhof (1992) and Wanninkhof et al., (2009). F_{O_2} was
116 calculated with the following equation.

$$117 \quad F_{O_2} = k K_0 (fO_{2_{water}} - fO_{2_{air}})$$

118 where k is the gas transfer velocity (calculated using and averaged daily wind speed from BOM
119 data), K_0 is the gas transfer coefficient, $fO_{2\text{water}}$ is the concentration of seawater dissolved oxygen
120 (mg L^{-1}) at the time of the downstream measurement, $fO_{2\text{air}}$ (mg L^{-1}) was assumed to be 100%
121 saturation at the air temperature over the 3-h measurement period ($\sim 8.10 \text{ mg L}^{-1}$).

122 **S.3 Statistical Analyses**

123 All statistical analyses were performed with the SPSS statistics software (SPSS Inc. 2013 Version
124 26.0). To compare measured differences in benthic cover (percent coral, percent algae, percent
125 bleached coral tissue, sediment overgrowth) and community metabolism (Net ecosystem production
126 [NEP] and net ecosystem calcification [NEC]) between triplicate transects, measurement days ($n =$
127 12), and Lagoon sites (Lagoon site 1, Lagoon site 2, and Slack Water), a one-way analysis of variance
128 (ANOVA) model was used in which transect, day, or site was a fixed effect and measured values for
129 percent cover, NEP, and NEC were treated as the response variable. Results for percent cover
130 compared among triplicate transects and Lagoon sites are displayed in Tables S1 and S2, respectively.
131 Before community metabolism measurements were compared, assumptions of normality and equality
132 of variance were evaluated with a Shapiro Wilk test (Table S4). Results for community metabolism
133 compared among triplicate transects, measurement days, and Lagoon sites are displayed in Tables S5,
134 S6, and S7, respectively. A Tukey HSD post-hoc test was used to perform pairwise comparisons for
135 measured community NEC between Lagoon site 1, Lagoon site 2, and the slack-water approach (Table
136 S7). To explore relationships between NEC as a function of NEP, Model II regression techniques were
137 used to test for significant linear relationships (cutoff value $p < 0.1$) and an ANCOVA was used to test
138 for differences in NEC vs. NEP slope categorized by Lagoon site (Lagoon site 1 and Lagoon site 2).

139 **Results**

140 **S.4 Invertebrate Taxonomy Results**

141 Overall, we found 25 coral species in the lagoonal reef study area, 22 of which were hard corals and
142 three soft corals (Fig. 2; Table S8). Thirteen algae morphospecies were observed, with one identified
143 as species *Valonia ventricosa* and the rest unidentified. Across all other invertebrate taxa, 19 species
144 of echinoderms, bivalves, and polychaetes, and 24 species of crustaceans and gastropods were
145 observed. Of the 43 non-coral invertebrate species, 15 were associated with colonies of *Pocillopora*
146 corals. Sea cucumbers (e.g., *Holothuria* spp., *Stichopus* spp.) were the dominant mobile invertebrate,
147 the Lollyfish sea cucumber (*Holothuria atra*) was the most common across both Lagoon sites ($1.2 \pm$
148 0.2 individuals m^{-2}). Second in abundance was the Hermann's Sea Cucumber (*Stichopus hermanni*)
149 (0.4 ± 0.1 individuals m^{-2}). Other notable invertebrates included Linckia sea stars (*Linckia guildingia*,
150 *Linckia laevigata*) and white-speckled sea hares (*Aplysia argus*) (all found in abundances < 0.1
151 individuals m^{-2}). The largest mobile invertebrates observed were Bailer Shell snails (*Melo amphora*)
152 at 30 cm in length and white-spotted hermit crabs (*Dardanus megistos*) occupying Bailer shells (< 0.1
153 individuals m^{-2}).

154 Our observations included 8 species with a conservation status of near threatened or higher, including
155 the small giant clam *Tridacna maxima*, Herrmann's sea cucumber (*Stichopus hermanni*), and 6 coral
156 species (*Porites attenuata*, *Acropora secale*, *Isopora palifera*, *Stylophora pistillata*, *Favites halicora*,
157 *Favites rotundata*). Notably, our observation of the aglajid slug *Tubulophilinopsis gardineri* is one of
158 just 5 from Heron Island, representing the southernmost limit of its eastern coast distribution. We also
159 observed an undescribed nudibranch species, a yellow-brown *Gymnodoris* (Figure 5). A complete list
160 of all species described can be found in the Supplemental Material (Table S8).

161 **S.5 Lagoon Temperature and Light**

162 Temperature across the Lagoon site 1 exhibited a mean value of 28.6 ± 1.5 °C and varied between a
163 minimum of 25.8 °C and a maximum of 34.8 °C (Table 2). Light at Lagoon site 1 exhibited a mean
164 value of 328 ± 247 $\mu\text{mol quanta m}^{-2} \text{ s}^{-1}$ and maximum values of $1001 \mu\text{mol quanta m}^{-2} \text{ s}^{-1}$ (Fig. 1).
165 Temperature across Lagoon site 2 exhibited a mean value of 28.6 ± 1.5 °C and varied between a

166 minimum of 25.9 °C and a maximum of 34.6 °C. Light at Lagoon site 2 exhibited a mean value of 336
167 $\pm 254 \mu\text{mol quanta m}^{-2} \text{ s}^{-1}$ and maximum values of $969 \mu\text{mol quanta m}^{-2} \text{ s}^{-1}$.

168 **S.6 Lagoon Community Bleaching Extent**

169 Dark-adapted yield was 0.662 ± 0.010 for *Acropora* spp. fragments and 0.576 ± 0.020 for “Other”
170 fragments (mean \pm SE, n = 35) on Feb 4th. On Feb 9th, yield declined 35% for *Acropora* spp. to 0.430
171 ± 0.014 (n = 15) and 25% for “Other” fragments to 0.434 ± 0.018 (n = 20). Symbiodiniaceae densities
172 were $0.976 \pm 0.135 \times 10^6 \text{ cm}^{-2}$ for *Acropora* spp. (n = 15) and $0.507 \pm 0.160 \times 10^6 \text{ cm}^{-2}$ for “Other”
173 fragments (n = 10) on Jan 30th. On Feb 12th, *Acropora* spp. densities had declined by 48% to $0.504 \pm$
174 $0.0849 \times 10^6 \text{ cm}^{-2}$ (n = 15) and by 18% for “Other” fragments to $0.414 \pm 0.094 \times 10^6 \text{ cm}^{-2}$ (n = 15) (Fig.
175 3).

176 **Discussion**

177 **S.7 Estimated Organism Contribution to NEC at Elevated Temperatures**

178 To estimate the potential effect of a +1.1 °C change in seawater temperature on coral calcification for
179 corals observed within the lagoon study sites the following aquaria manipulation studies were
180 reviewed: Edmunds, 2005; Anthony et al., 2008; Cantin et al., 2010; Comeau et al., 2013, 2016; and
181 the following meta-analysis and modeling studies were reviewed: Lough and Barnes, 2000; McNeil et
182 al., 2004; Evenhuis et al., 2015; Kornder et al., 2018; Bove et al., 2020.

183 **Tables**

184 Table S1: One-way ANOVA results (p-values) comparing measured percent coral and algae cover
 185 between triplicate transects within each Lagoon site (Lagoon site 1, Lagoon site 2). Data were pooled
 186 among replicate point-contact survey efforts ($n = 2 \text{ transect}^{-1}$). A **bolded** value (p-value < 0.05)
 187 indicates that the percent cover significantly differed between transects within each Lagoon site.

Point-Contact Survey Method				
Cover	Lagoon site 1		Lagoon site 2	
	df	p-value	df	p-value
% Coral Cover	2	0.791	2	0.959
% Algae Cover	2	0.256	2	0.214
% Sediment Cover	2	0.421	2	0.956

188

189

190 Table S2: One-way ANOVA results (p-values) comparing measured percent coral and algae cover
 191 between Lagoon site 1 and Lagoon site 2. Data were pooled among replicate point-contact survey
 192 efforts and triplicate transects within each Lagoon site ($n = 6 \text{ site}^{-1}$). A **bolded** value (p-value < 0.05)
 193 indicates that the percent cover significantly differed between Lagoon sites.

Point-Contact Survey Method		
Cover	df	p - value
% Coral Cover	1	0.001
% Algae Cover	1	0.011
% Sediment Cover	1	0.122

194

195 Table S3: One-way ANOVA results (p-values) comparing measured percent coral and algae cover
 196 between triplicate transects within each Lagoon site (Lagoon site 1, Lagoon site 2). Data were pooled
 197 among triplicate photo-quadrat survey efforts over time ($n = 120 \text{ transect}^{-1}$). A **bolded** value (p-value
 198 < 0.05) indicates that the percent cover significantly differed between transects.

Photo-Quadrat Survey Method				
Cover	Lagoon site 1		Lagoon site 2	
	df	p-value	df	p-value
% Coral Cover	2	0.469	2	0.818
% Algae Cover	2	0.721	2	0.796
% Sediment Cover	2	0.859	2	0.403

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205 Table S4: One-way ANOVA results (p-values) comparing measured percent coral and algae cover
 206 between Lagoon site 1 and Lagoon site 2. Data were pooled among triplicate photo-quadrat survey
 207 efforts and triplicate transects within each Lagoon site (n = 360 site⁻¹). A **bolded** value (p-value < 0.05)
 208 indicates that the percent cover significantly differed between Lagoon site 1 and Lagoon site 2.

Photo-Quadrat Survey Method		
Cover	df	p - value
% Coral Cover	1	0.000
% Algae Cover	1	0.273
% Sediment Cover	1	0.140

209

210 Table S5: One-way ANOVA results for percent bleached coral tissue (Coral Bleaching) and percent
 211 sediment exhibiting overgrowth (Sediment Overgrowth) compared over the three survey efforts
 212 through time (Jan 24, Feb 6, and Feb12 2020) at Lagoon site 1. Data were pooled among all triplicate
 213 transects. Tukey HSD post-hoc test results are to compare differences between each survey effort (n =
 214 3). A **bolded** value (p-value < 0.05) indicates that the difference was significant between time points.

Photo-Quadrat Survey Method: Lagoon site 1							
Lagoon site 1		df	F-value	p - value			
Coral Bleaching		2	67.2	0.000			
Sediment Overgrowth		2	18.3	0.003			
Tukey HSD							
Dependent Variable	(I) Time	(J) Time	Mean	Std. Error	Sig.	95% Confidence Interval	
			Difference (I-J)			Lower Bound	Upper Bound
Coral Bleaching	Jan 24	Feb 6	-16.33	4.93	.037	-31.48	-1.18
		Feb 12	-55.66	4.93	.000	-70.81	-40.51
	Feb 6	Jan 24	16.33	4.93	.037	1.18	31.48
		Feb 12	-39.33	4.93	.001	-54.48	-24.18
	Jan 24	Feb 6	55.66	4.93	.000	40.51	70.81
		Feb 12	39.33	4.93	.001	24.18	54.48
Sediment Overgrowth	Jan 24	Feb 6	-2.33	1.36	.275	-6.50	1.84
		Feb 12	-8.00	1.36	.003	-12.17	-3.82
	Feb 6	Jan 24	2.33	1.36	.275	-1.84	6.50
		Feb 12	-5.66	1.36	.014	-9.84	-1.49
	Jan 24	Feb 6	8.00	1.36	.003	3.82	12.17
		Feb 12	5.66	1.36	.014	1.49	9.84

216 Table S6: One-way ANOVA results for percent bleached coral tissue (Coral Bleaching) and percent
 217 sediment exhibiting overgrowth (Sediment Overgrowth) compared over the three survey efforts
 218 through time (Jan 24, Feb 6, and Feb12 2020) at Lagoon site 2. Data were pooled among all triplicate
 219 transects. Tukey HSD post-hoc test results are to compare differences between each survey effort (n =
 220 3). A **bolded** value (p-value < 0.05) indicates that the difference was significant between time points.

Photo-Quadrat Survey Method: Lagoon site 2							
Lagoon site 2		df	F-value	p - value			
Coral Bleaching		2	142.9	.000			
Sediment Overgrowth		2	10.5	.011			
Tukey HSD							
Dependent Variable	(I) Time	(J) Time	Mean	Std. Error	Sig.	95% Confidence Interval	
			Difference (I-J)			Lower Bound	Upper Bound
Coral Bleaching	Jan 24	Feb 6	-24.00	3.88	.002	-35.92	-12.07
		Feb 12	-65.00	3.88	.000	-76.92	-53.07
	Feb 6	Jan 24	24.00	3.88	.002	12.07	35.92
		Feb 12	-41.00	3.88	.000	-52.92	-29.07
	Jan 24	Feb 6	65.00	3.88	.000	53.07	76.92
		Feb 12	41.00	3.88	.000	29.07	52.92
Sediment Overgrowth	Jan 24	Feb 6	-3.00	2.8	.564	-11.59	5.59
		Feb 12	-12.33	2.80	.011	-20.93	-3.73
	Feb 6	Jan 24	3.00	2.80	.564	-5.59	11.59
		Feb 12	-9.33	2.80	.036	-17.93	-.73
	Jan 24	Feb 6	12.33	2.80	.011	3.73	20.93
		Feb 12	9.33	2.80	.036	.73	17.93

221 Table S7: One-way ANOVA results for percent bleached coral tissue (Coral Bleaching) and percent
 222 sediment exhibiting overgrowth (Sediment Overgrowth) compared over the three survey efforts
 223 through time (Jan 24, Feb 6, and Feb12 2020) between Lagoon site 1 and Lagoon site 2. Data were
 224 pooled among all triplicate transects. A **bolded** value (p-value < 0.05) indicates that the difference was
 225 significant between Lagoon sites.

Photo-Quadrat Survey Method				
	Coral Bleaching		Sediment Overgrowth	
Date	df	p - value	df	p - value
Jan 24 2020	1	1.00	1	0.899
Feb 6 2020	1	0.067	1	0.692
Feb 12 2020	1	0.256	1	0.231

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227 Table S8: List of invertebrate taxonomy described in section 3.2.4.

Group	Taxon	Common name
Algae	<i>Caulerpa</i> spp.	
	Chlorophyta spp.	Green algae
	<i>Halimeda</i> spp.	
	<i>Laurencia</i> spp.	
	<i>Padina</i> sp.	
Corals	Rhodophyta spp.	Red algae
	<i>Valonia ventricosa</i>	Sailor's eyeball alga
	<i>Acropora secale</i>	
	<i>Acropora millepora</i>	
	<i>Acropora muricata</i>	
	<i>Acropora</i> spp.	Staghorn corals
	<i>Astrea curta</i>	
	<i>Cyphastrea chalcidicum</i>	
	<i>Dipsastraea</i> sp.	
	<i>Favites halicora</i>	
<i>Favites rotundata</i>		
<i>Goniastrea edwardsi</i>	Honeycomb coral	
<i>Goniopora</i> sp.	Flowerpot coral	
<i>Isopora palifera</i>		

	<i>Klyxum</i> sp.	
	<i>Lobophyllia agaricia</i>	
	<i>Montipora digitata</i>	
	<i>Montipora grisea</i>	
	<i>Montipora hispida</i>	
	<i>Montipora</i> sp.	
	<i>Platygyra daedalea</i>	Lesser valley coral
	<i>Platygyra</i> spp.	
	<i>Pocillopora damicornis</i>	
	<i>Pocillopora</i> sp.	Cauliflower coral
	<i>Porites attenuate</i>	
	<i>Porites cylindrica</i>	Yellow finger coral
	<i>Porites</i> sp.	Pore coral
	<i>Sarcophyton</i> spp.	Toadstool leather corals
	<i>Stylophora pistillata</i>	Hood coral
Crustaceans	Alpheidae sp.	Snapping shrimp
	<i>Alpheus</i> sp.	Snapping shrimp
	Brachyura spp.	Crabs
	<i>Calcinus latens</i>	Hidden hermit crab
	Caridea sp.	Caridean shrimp
	<i>Clibanarius corallinus</i>	Coral hermit crab
	<i>Dardanus megistos</i>	White-spotted hermit crab
	Majidae sp.	Spider crab
	Stomatopoda spp.	Mantis shrimps
	<i>Thalamita</i> sp.	
	<i>Trapezia serenei</i>	Coral crab
	<i>Zenopontonia soror</i>	Seastar shrimp
Echinoderms	<i>Culcita novaeguineae</i>	Pillow cushion star
	<i>Holothuria atra</i>	Lollyfish sea cucumber
	<i>Holothuria edulis</i>	Pinkfish sea cucumber
	<i>Holothuria leucospilota</i>	Black sea cucumber

	<i>Holothuria</i> sp.	
	<i>Linckia guildingi</i>	Guilding's sea star
	<i>Linckia laevigata</i>	Blue linckia
	<i>Nardoa novaecaledoniae</i>	Yellow mesh sea star
	<i>Stichopus herrmanni</i>	Herrmann's sea cucumber
	<i>Stichopus chloronotus</i>	Greenfish sea cucumber
Molluscs	<i>Aplysia argus</i>	White-speckled seahare
	<i>Atactodea striata</i>	Striate beach clam
	<i>Codakia paytenorum</i>	Payten's codakia
	<i>Chrysostoma paradoxum</i>	Orange-mouthed top shell
	<i>Clypeomorus bifasciata</i>	Double-banded creeper
	<i>Coralliophila</i> sp.	
	Ergalataxinae	
	<i>Gymnodoris</i> sp.	
	<i>Melo amphora</i>	Giant baler
	<i>Pitar</i> sp.	
	<i>Spondylus</i> sp.	Thorny oyster
	<i>Tectus fenestratus</i>	Latticed top shell
	<i>Tonna chinensis</i>	China tun
	<i>Tridacna maxima</i>	Small giant clam
	<i>Tubulophilinopsis gardineri</i>	Gardiner's headshield slug
	<i>Turbo argyrostomus</i>	Silvermouth turban
Polychaetes	<i>Perinereis</i> sp.	
	<i>Spirobranchus</i> sp.	Christmas tree worm
	Terebellidae sp.	Spaghetti worm
Sponges	Porifera sp.	

228 Table S9: Shapiro-Wilk test for normality in reef metabolism. Data are organized by rates of NEP and
 229 NEC measured at Lagoon site 1, Lagoon site 2, and the larger lagoon area (Slack Water). Data for each
 230 Lagoon site were pooled among triplicate parallel transects. NEP data were not included for the slack-
 231 water method. If the significant value (Sig.) of the test is > 0.05 the data exhibit a normal distribution.

		Shapiro-Wilk		
	Site	Statistic	df	Sig.
NEP	Lagoon site 1	.951	36	.112
	Lagoon site 2	.984	36	.857
	Slack Water			
NEC	Lagoon site 1	.967	36	.356
	Lagoon site 2	.952	36	.117
	Slack Water	.962	33	.287

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238 Table S10: One-way ANOVA results (p-values) comparing measured reef metabolism (NEP and NEC)
 239 between triplicate transects within each Lagoon site (Lagoon site 1, Lagoon site 2, and Slack Water).
 240 Data were pooled among all 11 (Slack water) and 12 (Lagoon site 1 and Lagoon site 2) days of
 241 measurements (3 days for Night NEC). A **bolded** value (p-value < 0.05) indicates that the measured
 242 response in that specific metabolic parameter significantly differed between triplicate transects.

Metabolism	Lagoon site 1		Lagoon site 2		Slack Water	
	df	p-value	df	p-value	df	p-value
NEP	2	.471	2	.917		
NEC	2	.169	2	.489	2	.581
Night NEC					2	.617

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247 Table S11: One-way ANOVA results (p-values) comparing measured reef metabolism (NEP and NEC)
 248 between measurement days within each Lagoon site (Lagoon site 1 and Lagoon site 2 = 12; Slack
 249 Water = 11; Night NEC = 3). Data were pooled among all triplicate transects. A **bolded** value (p-value
 250 < 0.05) indicates that the measured response in that specific metabolic parameter significantly differed
 251 between triplicate transects.

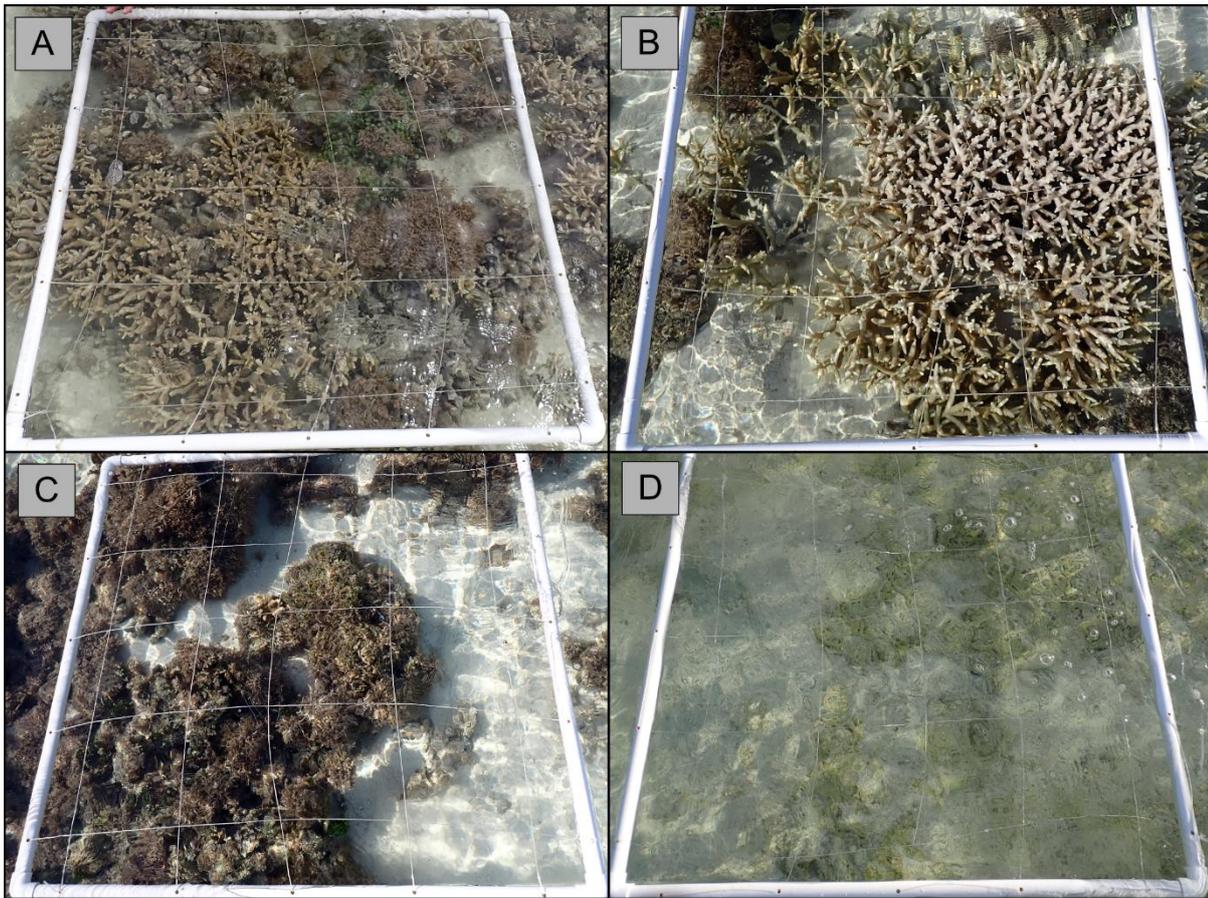
Metabolism	Lagoon site 1		Lagoon site 2		Slack Water	
	df	p-value	df	p-value	df	p-value
NEP	11	.181	11	.099		
NEC	11	.506	11	.365	10	.073
Night NEC					2	.083

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253 Table S12: One-way ANOVA results for NEP compared amongst Lagoon site 1 and Lagoon site 2 and
 254 for NEC compared amongst Lagoon site 1, Lagoon site 2, and Slack Water. Data were pooled among
 255 all triplicate transects and measurements days. Tukey HSD post-hoc test results are displayed for NEC
 256 (n = 3). A **bolded** value (p-value < 0.05) indicates that the difference was significant between Lagoon
 257 sites.

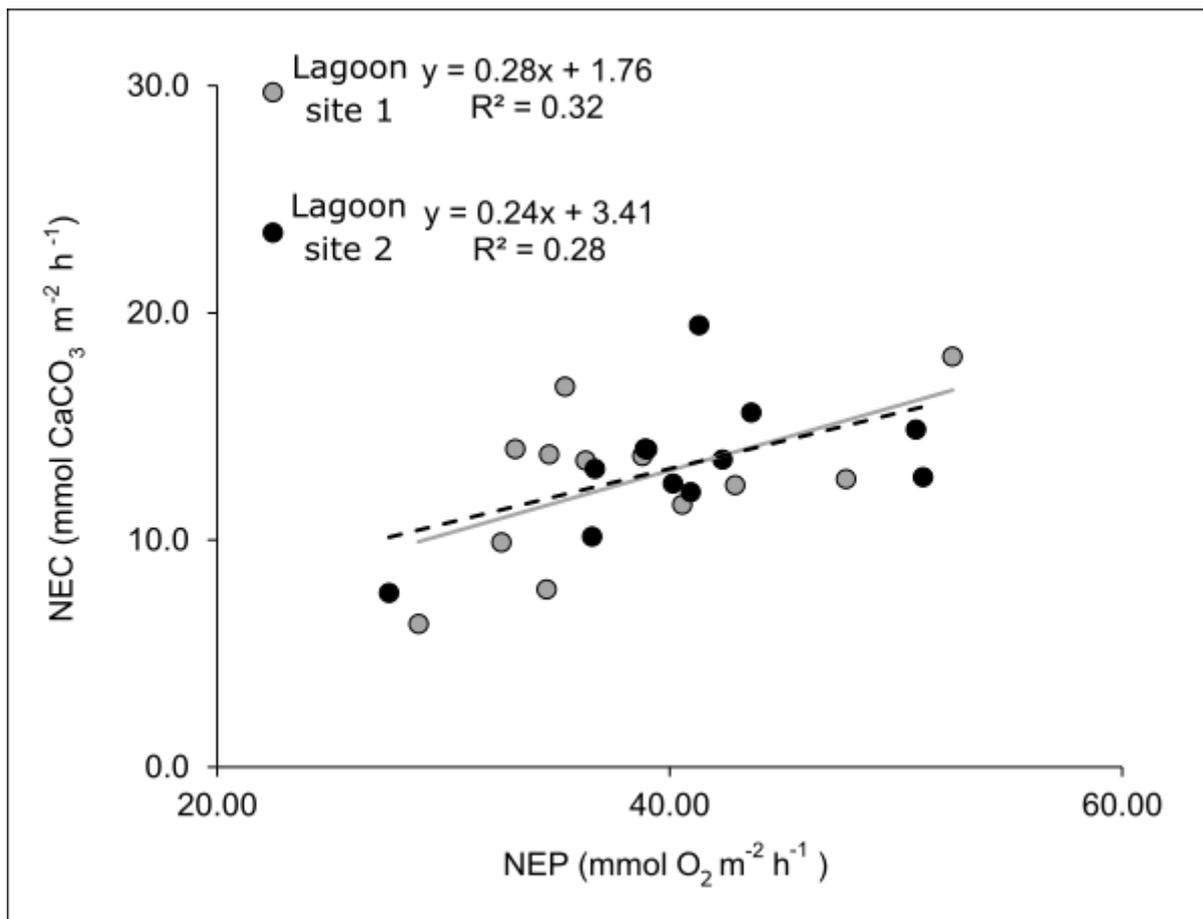
Metabolism		df	F-value	p - value
NEP		1	3.47	.067
NEC		2	8.17	.001

Tukey HSD						
(I) Site (J) Site		Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
Lagoon site 1	Lagoon site 2	-.8742	1.015	.666	-3.2916	1.5431
	Slack Water	3.0361*	1.015	.010	.6187	5.4534
Lagoon site 2	Lagoon site 1	.8742	1.015	.666	-1.5431	3.2916
	Slack Water	3.9103*	1.015	.001	1.4929	6.3277
Slack Water	Lagoon site 1	-3.0361*	1.015	.010	-5.4534	-.6187
	Lagoon site 2	-3.9103*	1.01544	.001	-6.3277	-1.4929



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260 Figure S.1: Photo-quadrat examples of various reef health. A) Healthy *Acropora* spp. coral observed
261 during the first survey effort. B) Bleached *Acropora* spp. observed during the final survey effort. C)
262 Example of fleshy algal growth as the dominant benthic organism D) Example of Chlorophyta
263 overgrowth on the sediment.



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265 Figure S.2: Rates of net ecosystem calcification (NEC) as a function of net ecosystem production
 266 (NEP) separated between study Lagoon site 1 (grey) and Lagoon site 2 (black).

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