1 Partitioning carbon sources between wetland and well-drained

2 ecosystems to a tropical first-order stream - Implications to carbon

3 cycling in the whole watershedat the watershed scale (Nyong,

- 4 Cameroon)
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20 Abstract

- 21 Tropical rivers emit large amounts of carbon (C) dioxide (CO₂) to the atmosphere, in particular due to largegreat wetland to
- 22 river earbon (C)C inputs. Yet, tropical African rivers remain largely understudied and little is known about the partitioning of
- 23 C sources between wetland and well-drained ecosystems to rivers. In the Nyong watershed (Cameroon, 27 800 km²), we
- 24 fortnightly measured total alkalinity, dissolved inorganic C used together with pH to compute the water CO₂ partial pressure
- 25 (pCO₂), dissolved and particulate organic C (DOC and POC) and total suspended matter, in groundwater located in a well-
- drained forest (hereafter referred as non-flooded forest groundwater) and in stream orders 1 to 6_3 , total alkalinity, dissolved inorganic C (DIC) used together with pH to compute the partial pressure of CO₂ (pCO₂), dissolved and particulate organic C
- 27 inorganic C (DIC) used together with pH to compute the partial pressure of CO_2 (pCO₂), dissolved and particulate organic C 28 (DOC and POC) and total suspended matter. In addition, we occasionally measured supplemented C measurements with
- 29 measures of heterotrophic respiration in the river in stream orders 1 and 5. In the first-order stream, DOC and POC
- 30 concentrations increased during rainy seasons when the hydrological connectivity with the riparian wetland increased whereas
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31 the concentrations of the same parameters decreased during dry seasons when the wetland was shrinking. In larger streams 32 (order > s higher than 1), the same seasonality was observed showing that wetland in headwaters were significant sources of 33 organic C for these rivers, even though higher POC concentration evidenced an additional source of POC in these riverslarger 34 streams during rainy seasons that was most likely POC originating from floating macrophytes. During rainy seasons, Ftheis 35 seasonal flush of organic matter from the wetland in the first order catchment and from the macrophytes in higher-order rivers 36 during rainy seasons significantly affected downstream metabolism, as evidenced by lower oxygen saturation together with 37 higher respiration rates pCO2 in stream orders 5 -(756±333 gC-CO2 m⁻² yr⁻¹) and 6 compared to 1 (286±228 gC-CO2 m⁻² yr⁻¹) 38 1). In the first-order catchment, the sum of the hydrological export of C hydrologically exported from non-flooded forest 39 groundwater (6.23±3.0 tMgC yr⁻¹) and wetland (4.0±1.5 MgtC yr⁻¹) to the stream represented 3-5% of the local catchment net 40 C sink. In the first-order catchment, non-flooded forest groundwater exporteds 1.6 times more C than wetland, however, when 41 weighed by surface area, C inputs from non-flooded forest groundwater and wetland to the stream contributed to 27% 42 $(13.\pm 0.2 \text{ tMgC yr}^{-1})$ and 73% $(33.03\pm 12.45 \text{ tMgC yr}^{-1})$ of the total hydrological C inputs, respectively. At the scale of the 43 Nyong watershed scale, the yearly integrated CO₂ degassing from the entire river network was 6520±1610 10³ +GgC-CO₂ yr⁻ 44 ¹ (23.45±5.86 tMgC_CO₂ km⁻² yr⁻¹ when weighed by the Nyong watershed surface area) whereas average heterotrophic 45 respiration in the river and CO₂ degassing rates were 52199 ± 40327 and $11605\ 085\pm580\ 2\ 544\ gC-CO_2\ m^2\ yr^1$ mmol m²-d⁺, 46 which implied that only $\sim \frac{8.510}{9}$ of the CO₂ degassing at the water-air interface was supported by heterotrophic respiration in the river. In addition, the total fluvial C export of $19\underline{10}\pm10\underline{80}$ $\pm0^3\pm\underline{6g}C$ yr⁻¹ (10.3 $\pm5.\underline{89}$ $\pm\underline{Mg}C$ km⁻² yr⁻¹ when weighed by 47 48 the Nyong watershed surface area) plus the yearly integrated CO₂ degassing from the entire river network represented $\sim 1011\%$ 49 of the net C sink estimated for the whole Nyong watershed. FinallyIn the tropics, we highlight that attributing to a unique 50 terrestrial source (well-drained ecosystems) the whole amount of riverine C emitted to the atmosphere and hydrologically 51 exported at the outlet and ignoring the river-wetland connectivity might lead to the misrepresentation of C dynamics in 52 headwaters, and thereby in the whole watershed.

53 1. Introduction

54 Despite their small surface area worldwide (Allen and Pavelsky, 2018), inland waters (rivers, lakes and reservoirs) have a 55 critical role in the global carbon (C) cycle. -as-Inland watersthey receive large amount of C from the drainage of land, i.e., 56 (terra firmefrom well-drained ecosystems as non-flooded soils_via-and groundwater and overland flow), and wetlanda-(i.e., 57 from flooded soils) (Abril and Borges, 2019; Cole and Caraco, 2001). The C entering inland waters, which is processed and 58 subsequently transferred to the atmosphere and the ocean (Cole et al., 2007; Ludwig et al., 1996; Meybeck, 1982; Tank et al., 59 2016) (Abril and Borges, 2019; Allen and Pavelsky, 2018; Cole et al., 2007; Ludwig et al., 1996; Meybeck, 1982). Besides, 60 inland waters are significant hotspots of C dioxide (CO2) degassing (e.g., Raymond et al., 2013) as they are usually 61 supersaturated with CO2 compared to the overlying atmosphere. Since the seminal paper by Cole et al. (2007) who estimated 62 that 0.75 PgC-CO2 was emitted annually to the atmosphere from global inland waters, global emissions estimates have

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increased substantially. In the most spatially explicit scaling study, degassing estimate from global inland waters was 2.1 PgC-CO₂ yr⁻¹ of CO₂ (Raymond et al., 2013). Later, this estimate has been updated with more accurate CO₂ emissions estimates from African and Amazonian rivers and from small ponds, resulting in the latest estimate of 3.9 PgC-CO₂ yr⁻¹ to which 0.2-

66 0.55 PgC-CO₂ yr⁻¹ might be still added as CO₂ emissions estimates for from rivers are usually not integrated over a full day
67 (Borges et al., 2015a; Drake et al., 2018; Gómez-Gener et al., 2021; Holgerson and Raymond, 2016; Raymond et al., 2013;
68 Sawakuchi et al., 2017) Globally, the latest estimate of the CO₂ degassing of CO₂-from inland waters was in the same order
69 of magnitude as the net terrestrial C sink (3.4 PgC yr⁻¹; Friedlingstein et al., 2020).

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- 71 Raymond et al. (2013) showed that CO₂ emissions from global rivers (1.8 PgC-CO₂ yr⁻¹) mainly depends on emissions in 72 tropical rivers, since tropical riversthese account for ~80% of the global emissions. However, the magnitude of CO₂ emissions 73 from tropical rivers was poorly constrained because its estimation was based on very few data from the tropics and probably 74 biased by the overwhelming dominance of data from the Amazon basin over other tropical basins, resulting in uncertain 75 interpolation and scaling. Indeed, based on CO2 emissions measurements in African and Amazonian rivers including the 76 Amazon and the Congo, Borges et al. (2015a) estimated that tropical rivers could emit alone 1.8±0.4 PgC-CO₂ yr⁻¹. This 77 significant flux at the global scale, estimated from direct measurements, demonstrates the importance of $C\underline{O}_2$ emissions from 78 tropical rivers, calling for attention to tropical systems, in particular to Africa, where very few data on C stock and C cycle are 79 available. These data are crucial to refine the global CO2 budget since tropical rivers have been identified in global earth 80 modelling approaches as systems exhibiting higher CO2 emission rates per unit area than those in the temperate and boreal 81 regions (Lauerwald et al., 2015; Raymond et al., 2013). In addition, in these modelling studies the CO₂ emission upscaling 82 was done using the GLORICH dataset, in which the water CO2 partial pressure (pCO2) is actually estimated from pH and total 83 alkalinity (TA). This calculation method leads to overestimate pCO2 up to 75 times, notably in low buffered and high organic 84 waters, which that are representative for boreal and tropical rivers (Abril et al., 2015). In contrast, pCO2 estimated from pH 85 and dissolved inorganic C (DIC) measurements is relatively robust (Åberg and Wallin, 2014). Thus, empirically measuring 86 pCO2 and to a lesser extent DIC, rather than relying on pCO2 estimated from pH and TA that is prone to large error are key to 87 improving CO2 emissions estimates from inland waters. 88

In tropical watersheds, CO₂-enriched wetland waters directly contribute to the CO₂ dissolved in riverine waters, in particular
 during high water periods when wetland-river connectivity is increasing (Abril et al., 2014; Borges et al., 2015a, 2015b, 2019).
 Indeed, tropical wetlands are productivity hotspots and a large fraction of their biomass is released to the water through litter fall and roots exudation, which fuels heterotrophic respiration in the wetland and enrich the water in CO₂ (Abril et al., 2014;
 Abril and Borges, 2019). In addition, during high water periods, the drainage of tropical wetlands releases large amounts of C
 and organic matter (OM) to the rivers that might enhance heterotrophic respiration in downstream rivers, indirectly increasing

95 <u>CO2 concentration in tropical rivers (Borges et al., 2019; Engle et al., 2008; Lambert et al., 2016a; Richey et al., 2002</u>

96 Nonetheless, in large tropical rivers, heterotrophic respiration in the river is usually a small component of the riverine CO₂

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97	budget because of the large dominance of the drainage of wetland in the overall budget (Abril et al., 2014; Borges et al., 2019).	
98	Large tropical rivers have the ability to transport CO2-enriched wetland waters far enough from the point source because of	
99	faster water movement relative to gas exchange (Abril et al., 2014). In the Amazon and the Congo watersheds, the intensity of	
100	the CO2 degassing from the rivers has been thus related to the percentage of the wetland cover (Abril et al., 2014; Borges et	
101	al., 2019, 2015b), showing that wetlands are the main source of OM fuelling CO2 production in tropical watersheds. However,	
102	as in temperate rivers, the CO ₂ dissolved in tropical rivers also originates from well-drained ecosystems (non-flooded soils	
103	and groundwater) in which CO2 comes from plant root and microbial respiration (Johnson et al., 2006, 2008).	Mis en forme : Anglais (Royaume-Uni)
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105	In tropical watersheds, considering the importance of lateral inputs in sustaining riverine C fluxes, quantifying hydrological <u>C</u>	Mis en forme : Anglais (Royaume-Uni)
106	fluxes resulting from the drainage of well-drained ecosystems and wetlands is thus fundamental to close the riverine C budget.	
107	Still, in tropical watersheds, questions remain about the quantification and partitioning of hydrological C fluxes resulting from	
108	the drainage of well-drained ecosystems and wetland and their significance in comparison to the local net terrestrial C sink	
109	(Duvert et al., 2020a). At the plot scale and in temperate climate, the very few studies that compare the local net terrestrial C	Mis en forme : Anglais (Royaume-Uni)
110	sink with direct measurements of the hydrological export of C from well-drained ecosystems showed less than 3% of the local	
111	net terrestrial C sink is actually exported to the aquatic environment (Deirmendjian et al., 2018; Kindler et al., 2011). In a	Mis en forme : Anglais (Royaume-Uni)
112	small tropical catchment (140 km²) in Australia, in which the land use was shared between dry savanna and wetland, the	
113	contribution of the total hydrological export of C to the stream relative to the local net terrestrial C sink was 7% (Duvert et al.,	Mis en forme : Anglais (Royaume-Uni)
114	2020a). However, Duvert et al. (2020a) did not partition the hydrological export of C to the river between dry savanna and	Mis en forme : Anglais (Royaume-Uni)
115	wetland. Furthermore, to the best of our knowledge, partitioning the hydrological export of C to rivers between well-drained	
116	ecosystems and wetland has never been done in tropical Africa. As the hotter warmer and wetter conditions expected in tropical	Mis en forme : Anglais (Royaume-Uni)
117	Africa in a near future will likely modify C fluxes at the watershed scale, integrative studies on C cycling in tropical watersheds	
118	are required to get a better grasp of the present drivers of riverine CO2 emissions and thus to better predict future changes	
119	<u>(Duvert et al., 2020a)</u>	Mis en forme : Anglais (Royaume-Uni)
120	The dissolved CO2 in riverine waters originates concomitantly from heterotrophic respiration in the river, i.e., from the	Mis en forme : Anglais (Royaume-Uni)
121	decomposition of organic matter (OM) in the aquatic system itself, and from the drainage of land and wetland (Abril and	
122	Borges, 2019; Borges et al., 2015; Hotchkiss et al., 2015), In tropical watersheds, riverine respiration is usually a small_	Mis en forme : Anglais (Royaume-Uni)
123	component of the riverine CO ₂ budget because of the large dominance of the drainage of land and wetland in the overall budget	
124	(e.g., Borges et al., 2019), The quantification of hydrological C fluxes originating from the drainage of land and wetland is	Mis en forme : Anglais (Royaume-Uni)
125	thus fundamental to close the riverine C budget in tropical watersheds. In the Amazon and the Congo watersheds, the intensity	
126	of the CO2-degassing from the rivers has been related to the percentage of the wetland cover (Abril et al., 2014; Borges et al.,	
127	2019, 2015b), showing that wetlands are the main source of OM fuelling CO2 production in tropical watersheds. Indeed,	Mis en forme : Anglais (Royaume-Uni)
128	tropical wetlands are productivity hotspots and a large fraction of their biomass is released to the water through litter-fall and	
129	roots exudation, which fuels heterotrophic respiration in the wetland and enrich the water in CO ₂ (Abril et al., 2014; Abril and	
130	Borges, 2019). The drainage of wetlands also releases large amounts of OM directly to the rivers, enhancing heterotrophic	Mis en forme : Anglais (Royaume-Uni)
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131	respiration in the river and therefore supports CO2 degassing from the rivers (Abril et al., 2014; Abril and Borges, 2019). Still,	Mis en forme : Anglais (Royaume-Uni)
132	in tropical watersheds, questions remain about the quantification and partitioning of hydrological C fluxes originating from	
133	the drainage of land and wetland at the plot scale and their significance in comparison to the local net terrestrial C sink (Duvert	
134	et al., 2020a). At smaller scale, the very few studies that compare the local net terrestrial C sink with direct measurements of	
135	the hydrologic export of C from land showed that only a small fraction (between 3 and 7%) of the net terrestrial C sink is	
136	actually exported to the aquatic environment, whether in temperate or tropical ecosystems (Deirmendjian et al., 2018; Duvert	
137	et al., 2020a; Kindler et al., 2011) but to the best of our knowledge this kind of work has never been done in tropical Africa.	Mis en forme : Anglais (Royaume-Uni)
138	As the hotter and wetter conditions expected in tropical Africa in a near future will likely modify C fluxes at the watershed	
139	scale, integrative studies on C cycling in tropical watersheds are required to understand the present conditions and thus to	
140	better predict future changes (Duvert et al., 2020a).	
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142	The Nyong River basin (South Cameroon) belongs to the Critical Zone Observatories' (CZOs; Gaillardet et al., 2018) network	Anglais (Royaume-Uni), Contour du texte
143	named Multiscale TROPIcal CatchmentS (M-TROPICS; https://mtropics.obs-mip.fr; Audry et al., 2021) and is a long-term	Mis en forme : Anglais (Royaume-Uni)
144	monitoring program of hydrological and environmental parameters in the tropics. In this study, we used rainfall, water table	
145	level and river discharges measured in the framework of the M-TROPICS observatory. The first objective of this study is to	
146	estimate the riverine C budget of a first-order catchment, the Mengong catchment, a nested sub-catchment of the Nyong	
147	watershed. The hydrological inputs of C from the drainage of land (i.e., from groundwater located-in a well-drained forest;	
148	hereafter referred as non-flooded forest groundwater) and from wetland to the stream, the heterotrophic respiration in the river,	
149	the CO2 degassed to the atmosphere, and the C hydrologically exported at the stream outlet are estimated and compared with	
150	the local net terrestrial C sink, and will be discussed. In lines with recent studies in large tropical watersheds (Abril et al., 2014;	
151	Borges et al., 2015; 2019), we expect that lateral inputs of C from wetland to the stream are significant in comparison with	
152	lateral inputs of C from non-flooded forest groundwater. The second objective of this study is to evaluate the changes in	
153	organic and inorganic C concentration over the seasons in the riverine continuum, from non-flooded forest groundwater to the	
154	different stream orders (order 1 to 6). In the Nyong watershed, downstream (order > 1) riverine C concentrations throughout	
155	a water cycle will be compared with those observed upstream in the Mengong stream (order 1) in order to evaluate how the	
156	biogeochemical cycle of C and its resulting atmospheric CO2 emissions is affected by the connectivity with the wetland	
57	domain. Ultimately, the variations of the C concentrations in the Nyong watershed throughout a water cycle will be compared	
58	with those observed in the Mengong sub-catchment in order to evaluate how the biogeochemical cycle of C and its resulting	
59	CO2 emissions to the atmosphere in a large tropical watershed is affected by the connectivity with the wetland domain.	Mis en forme : Anglais (Royaume-Uni)

160 2. Materials and Methods

161 2.1. Study site

162 2.1.1. The Nyong watershed

163 The Nyong watershed (27 800 km², Cameroon) is located between 2.8 and 4.5° N and 9.5 and 13.3° E, mainly in the Southern 164 Cameroon Plateau (600-900 m high) (Fig. 1). The landscape of the Southern Cameroon Plateau mostly consists in a succession 165 of convex rounded hills separated by flat wetlands of variable sizes (Olivry, 1986). We adopt the common definition of 166 wetlands as habitats with continuous, seasonal, or periodic standing water or saturated soils (Mitsch et al., 2012). The main 167 stem (the Nyong River, stream order 6) is 690 km long and is flowingflows west to the Atlantic Ocean (Fig.1). In the eastern 68 part of the watershed (from Abong Mbang to Akonolinga; Fig. 1), the Nyong River is flowingflows through large riparian 69 wetlands of variable size that laterally extended from the river up to 2-3 km according to seasons according to seasons, up to 70 2-3 km width (Olivry, 1986). In the western part of the basin (downstream to Akonolinga; Fig. 1), riparian wetlands extent is 71 less pronounced and the Nyong river is flowingflows through mature forest in a well-chanelled river bed (Olivry, 1986). İ72

173 The Nyong watershed experiences an equatorial climate with four seasons of unequal importance with two maxima and 174 minima: a short rainy season (SRS: Apr-June), a short dry season (SDS: July-August), a long rainy season (LRS: Sept.-Nov) 175 and a long dry season (LDS: Dec-March) (Suchel, 1987). The catchment lithology is composed of metamorphic and plutonic 176 rocks with the absence of carbonate rocks and minerals (Viers et al., 2000). Slopes and hills are recovered by a thick lateritic 177 profile (20-40 m) poor in C, whereas in the wetlands (i.e., in the depressions) the upper part of the hydromorphic soils shows 178 an enrichment in OM (Boeglin et al., 2003; Nyeck et al., 1999). Ferrealitic soils covers about 80% of the Nyong watershed, 179 and this soil cover can reach 40 m thick (Braun et al., 2005). On hills and hillsides, the vegetation cover is dominated by semi 180 deciduous-forest whereas in the wetlands Raffia palm trees usually dominates.

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182 In the Nyong watershed, six sites were sampling fortnightly from January to December 2016 (22 times during the sampling 183 period), namely from upstream to downstream: the small first-order Mengong catchment (at the source and the outlet of the 184 catchment), the Awout River (order 3), the So'o River (order 4), and the Nyong River at Mbalmayo (order 5) and Olama (order 185 6); all sampling sites were located in the western part of the watershed (Table 1; Fig. 1). Noteworthy, the Mengong catchment 186 is described in detail in the next section 2.1.2. The Awout River is flowingflows for about 30 km in a partially marshy river 187 bed. The So'o River is the southern forest extension of the Nyong Watershed and is the main tributary on the left bank of the 188 Nyong River. The Mbalmayo sampling station is located on the Nyong River upstream the confluence with the So'o, while 189 the Olama sampling station is located downstream theis confluence with the So'o. Each sampling site (except the Mengong 190 source) are gauging stations calibrated for discharges measurements, monitored daily since 1998 and are publicly available at

191 https://doi.org/10.6096/BVET.CMR.HYDRO (Audry et al., 2021). The yearly average discharge of the Nyong River at Olama

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was ~195 m³ s⁻¹ for both the 1998-2020 period (long-term average) and the year of sampling 2016 (Fig. 2). Also, the average

i93 monthly discharges during the year 2016 did not differ significantly from the average monthly discharges from the 1998-2020

194 period (Fig. 2). The annual rainfall in the Nyong watershed was 1986 mm in 2016 which is in the upper range of rainfall

195 (1600±290 mm) for the 1998-2020 period (Fig. 2). Altogether, this shows that hydrological fluxes occurring during the

196 sampling year 2016 were typical of the hydrological fluxes usually occurring in the Nyong watershed.

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In addition, the C exported at the most downstream station (Nyong <u>River</u> at Olama) is considered as representative as the C exported to the Atlantic Ocean by the whole Nyong watershed because the contribution of the tributaries downstream from this station is negligible (Nkoue-ndondo, 2008). Indeed, Brunet et al. (2009) measured both hydrological export of DIC and DOC from the Nyong River at Olama and also more downstream near Déhané (very close to the Nyong river outlet; Fig. 1) and they showed that these fluxes (in tMgC km⁻² yr⁻¹, weighed by the catchment surface area drained at Olama or Edea) were similar at Olama (4.2±0.1 and 0.8±0.1 tMgC km⁻² yr⁻¹, for DOC and DIC, respectively) and Déhané (3.9±0.2 and 1.1±0.1 tMgC km⁻² yr⁻¹).

205 2.1.2. The first-order Mengong catchment

206 The Mengong Catchment is 0.6 km² and consists of a convexo-concave landscape, ranging from 669 m at the river outlet to 207 703 m at the top of the hill, separated by a flat wetland that covers 0.12 km² (Fig. 3). Semi-deciduous rainforest (Sterculiaceae-208 Ulmaceae, C3 plant) covers most of the hills and hillsides, whereas most of the wetland vegetation comprises semi-aquatic 209 plants of the Araceae family (C4) and tree populations of Gilbertiodendron deweverei (Caesalpiniaceae, C3) and Raffia 210 monbuttorum (raffia palm trees, C3) (Braun et al., 2005, 2012). The hillside soil cover is a thick lateritic soil that consists of a 211 succession of four main horizons, namely from the bottom to the top, the saprolitic horizon, the mottled clay horizon, the 212 ferruginous horizon, and the soft clayey topsoil; the thickness and distribution of these soil layers depend on the topographic 213 position (Fig. \$13). The groundwater floods the fractured bedrock, the entire saprolite, and partly the mottled clay horizon 214 (Braun et al. 2005; Fig. 3). The soil cover is 15 m thick at the top of the north hill (piezometer 1); the depth however, decreases 215 progressively towards the flat wetland (Fig. 3). The roots of the hillside vegetation are essentially located in the topsoil horizon, 216 which has a depth of 5 to 6 m at the top of the hill (at piezometer 1) and has a depth of 3 to 4 m (at piezometer 2) at the mid-217 slope (Braun et al. 2005; Fig. 3). In the wetland, a dark-brown organic-rich sandy material with a thickness ranging from 0.1 218 to 1 m tops the hydromorphic soil. In this organic horizon, OM can reach up to 20% by weight, and it is composed of a thick 219 mat of dead and living roots and tubers originating from the wetland vegetation (Braun et al. 2005; Fig. 3). Noteworthy, the 220 first-order Mengong catchment is considered representative of the South Cameroon plateau (and thus of the Nyong watershed) 221 that also consists itself in multiconvex land form developed on granitic terrains separated by flat wetland (Braun et al., 2012). 222 Moreover, the same soil cover and plant species are observed in the Mengong catchment and in the Nyong watershed but it 223 should be noted that the wetland extent is larger in the Mengong Catchment (20%) than in the whole watershed (~5%) (Table.

225 map by Gumbricht et al., (2017) (Table 1). 226 227 Groundwater draining the hillside emerges at two sources (Qhill) in the watershed catchment head and at specific seepage points 228 (Qbase) along the hillside/wetland boundaries (Fig. 3). Only one of these two sources is perennial, the other dries up during dry 229 periods (Fig. 3; Braun et al., 2005; Maréchal et al., 2011). Note that groundwater that emerges at sources and at specific 230 seepage points will be further referred as non-flooded forest groundwater. Qhill is conveyed to the stream with negligible 231 interaction with the wetland, while Q_{base} fed the wetland, which is flooded all year long (Maréchal et al., 2011). In addition, 232 according to observations made in the Mengong catchment during most of the rainfall events by Maréchal et al. (2011), it is 233 assumed that the overland flow can be neglected on the forested hillside as the porous soil have a high infiltration capacity. 234 Therefore, the water budget of the hillside aquifer system, as shown in Fig. 3, is the following: 235 $R_{hill} = Q_{hill} + Q_{base}$ (Eq. 1) 236 where. 237 Rhill is the recharge rate of the hillside by infiltration of rain water. Maréchal et al. (2011) estimated Rhill at 20% of the yearly 238 rainfall occurring in the Mengong catchment, based on a hydrological model related to chloride mass balance at the catchment 239 scale. Q_{hill} and Q_{base} represents 90 and 10% of $R_{hill},$ respectively. 240 241 The total streamflow at the outlet of the Mengong catchment (Q_{ST}), as shown in Fig. 3, is the sum of the contributions of Q_{hill}, 242 the exchange flow between the wetland and the stream (Q_{WLST}) and the overland flow on the wetland surface (OF_{WL}), as the 243 following: 244 $Q_{ST} = Q_{hill} + Q_{WL/ST} + OF_{WL}$ (Eq. 2) 245 where, 246 OFwL represents 35% of the of the yearly rainfall in the Mengong catchment (Maréchal et al. 2011). Note that both Qhill and 247 OF_{WL} can be estimated from the yearly rainfall over the Mengong catchment and Q_{ST} is measured. Q_{WLST} can be thus obtained 248 by difference, but only on a yearly basis.

1). Note that wetland extent in larger catchment higher than 1 were was estimated from GIS analysis using the global wetland

249 2.2. Sampling and laboratory work

The water samples in the Nyong, So'o and Awout Rivers were collected from bridges using a Niskin Bottle (3L) attached to a rope. At the Mengong source, the water samples were taken directly from the source where non-flooded forest groundwater seeps out from a polyvinyl chloride pipe. Note that the pipe is only a few centimetres long, thus limiting considerably the contact time between water and atmospheric air. Additionally, each sampling bottle was left to overflow to avoid catching air bubbles. At the Mengong outlet, the shallow depth permitted retrieving water samples directly from the stream.

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Dissolved inorganic C (DIC), TA, dissolved and particulate organic C (DOC and POC) dissolved organic (DOC) and 256 257 particulate organic (POC) C, total suspended matter (TSM) and the POC content of the TSM (POC%) were measured in 258 replicates from one offsingle samples. At each sampling site, we measured the physico-chemical parameters of the water 259 (temperature, pH, oxygen saturation, and specific conductivity). In addition, we carried out 14 measurements of heterotrophic 260 respiration in the river at two sampling sites (in the Mengong stream and in the Nyong River at Mbalmayo). The water 261 temperature, pH, oxygen saturation and specific conductivity were measured in-situ using portable probes (WTW®) between 262 January and March 2016 and using an YSI® ProDSS Multiparameter Digital Water Quality Meter between April and 263 December 2016. Calibration of sensors was carried out prior to sampling campaigns and regularly checked during the 264 campaigns. For the WTW® probes, the conductivity cell was calibrated with a 1_000 µS cm⁻¹ (25°C) standard and the pH 265 probe was calibrated using NBS buffer solutions (4 and 7). The YSI® ProDSS was calibrated using the protocols recommended 266 by the manufacturer. The conditioning of water samples was done directly after the field trips in Cameroon at the Institut de 267 Recherches Géologiques et Minières (IRGM) of Yaounde, while chemical analyses were done in France at Toulouse in the 268 laboratory of Géosciences Et Environnement (GET). For TSM, POC and POC%, a filtration (0.5-1.5 L) was carried out on 269 pre-weighed and pre-combusted GF/F glass fiberfibre filters (porosity of 0.7 µm). The filters were then dried at 60 °C and 270 stored in the dark at room temperature for subsequent analysis. TSM was determined by gravimetry with a Sartorius scale 271 (precision of the scale was ±0.1 mg). The filters were acidified in crucibles with 2N HCl to remove carbonates and were then 272 dried at 60 °C to remove inorganic C and the remaining acid and water and then analysed by the Rock Eval pyrolysis method 273 to measure POC and POC% (Lafargue et al., 1998). For DOC, a portion of the POC filtrate was kept in glass bottles (60 mL) 274 pyrolyzed beforehand, in which 3 drops of phosphoric acid (85% H₃PO₄) were added to convert all DIC species to CO₂. The 275 glass bottles were sealed with septa made of polytetrafluoroethylene (PTFE). DOC samples were stored at 3-5°C and DOC 276 concentrations were measured by thermal oxidation after a DIC removal step with a SHIMADZU TOC 500 analyser in TOC-277 IC mode (Sharp, 1993).

279 We stored TA samples at 20°C in polypropylene bottles after filtration using a syringe equipped with acetate cellulose filters 280 (porosity of 0.22 µm). TA was then analysed by automated electro-titration (Titrino Metrohm®) on 50 mL-samples with 0.1 N HCl as the titrant. The equivalence point was determined from pH between 4 and 3 with the Gran method (Gran, 1952). 281 282 DIC samples were collected in 70 mL glass serum bottles sealed with a butyl stopper and treated with 0.3 mL of HgCl2 at 20 283 g L⁻¹ to avoid microbial respiration during storage. Vials were carefully sealed such that no air remained in contact with 284 samples and were stored in the dark to prevent photo-oxidation. DIC was measured with the headspace technique. The 285 headspace was created with 15 mL of N_2 gas, and 100 μ L of 85% H₃PO₄ was added in the serum bottles to convert all DIC 286 species to CO2. After strong shaking and overnight equilibration at constant room temperature, a subsample of the headspace 287 (1 mL) was injected with a gastight syringe into a gas chromatograph equipped with a flame ionization detector (SRI 8610C 288 GC-FID). The gas chromatograph was calibrated with CO₂ standards of 400, 1 000 and 3 000 ppm (Air Liquide® France). In 289 addition, we estimated the water pCO2 from the CO2SYS software (Lewis and Wallace Upton, 1998), using DIC, pH, water

278

temperature measurements, and the carbonic acid dissociation constants of Millero (1979) and the CO₂ solubility from Weiss (1974).

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293 In addition, we carried out 14 measurements of heterotrophic respiration in the river at two sampling sites (in the Mengong 294 stream and in the Nyong River at Mbalmayo). In the Mengong stream and in the Nyong River at Mbalmayo, sixFor each 295 sampling, six 70-mL serum bottles collected similarly as for DIC samples, were used for the determination of heterotrophic 296 respiration in the river. Three serum bottles were directly poisoned in the field with 0.3 mL of HgCl₂ at 20 g L⁻¹. The three 297 other serum bottles were incubated in a cool-dark box during 24 hours. The cool-dark box was protected from light and filled 298 with water from the river to maintain inside the cool-dark box a water temperature similar to the one-water temperature 299 observed in the river. At the end of the incubations, the serum bottles were poisoned with 0.3 mL of HgCl2 and stored in the 300 dark and at room temperature. To estimate volumetric rates of heterotrophic respiration in the river, we measured the increase 301 in CO2 in the incubated serum bottles compared to those poisoned directly in the field. CO2 was measured similarly as for 302 DIC, using the headspace technique but without a prior acidification with H₃PO₄. Subsequently, volumetric rates of 303 heterotrophic respiration in the river were depth-integrated with the river depth at the day of sampling. The river depth was 304 retrieved from discharge-depth relationship established in the framework of the M-TROPICS observatory. Noteworthy, our 305 method does not represent total heterotrophic respiration in the river since it does not include benthic respiration. A mean 306 benthic respiration measured in various tropical rivers of 21-222 gCmmol m⁻² yrd⁻¹ by Cardoso et al. (2014) will be was 307 therefore added to estimate total heterotrophic respiration in the river.

308 2.3. Determination of catchments surface area, water surface area, slope and gas transfer velocity (k600)

10 In the Nyong watershed, Tthe sub-catchments surface areas and the determination of the different stream orders were estimated from the hydrological modelling tools available in QGIS3.16® and the digital elevation model (DEM, 15 sec resolution) conditioned for hydrology (HydroSHEDS; Lehner et al., 2008). In the Nyong watershed, the HydroSHEDS flowline dataset (15 sec resolution) enabled the precise determination of the total length of each stream order (1 to 6). To estimate the average monthly river width (W) in each stream order of the Nyong watershed, we used the average monthly discharges from the five gauging stations (located on stream-orders 1, 3, 4, 5 and 6) and the hydraulic equation described by Raymond et al. (2012), as follows:

- 316 W = 12.88Q_{monthly}^{0.42}
- 317 where,
- 318 Q_{monthy} is the average monthly discharge in 2016 in the stream orders 1, 3, 4, 5 or 6.

319 Since we did not measure discharge in stream order 2, the average width of stream order 2 was extrapolated from the best

320 exponential regression curve from the relationship between stream order and average monthly river width, as indeed, river

- 321 width within a basin scale exponentially with stream order for all river orders (Strahler, 1957). We used the average monthly
- 322 river width and the total length per stream order to estimate the monthly water surface area per stream order. We fused the

10

Mis en forme : Anglais (Royaume-Uni)

Mis en forme : Anglais (Royaume-Uni)

(Eq. 3)

323	HydroSHEDS DEM and flowline dataset to assign an altitude to each river point and thus to determine the average slope (S)
324	per stream order. To calculate the average monthly flow velocity (V) per stream order, we used the following hydraulic
325	equation described by Raymond et al. (2012), as follows:
326	$V = 0.19 Q_{monthly}^{0.29}$ (Eq. 4)
327	The average monthly flow velocity in stream order 2 was extrapolated from the best exponential regression curve from the
328	relationship between stream order and monthly average flow velocity. In each stream order, the monthly gas transfer velocity
329	normalized to a Schmidt number of 600 (k ₆₀₀ in m d ⁻¹) was derived from the parameterization as a function of S (unitless) and
330	V (m s ⁻¹) as in the Eq. 5 by Raymond et al. (2012):
331	$k_{600} = VS*2841+2.02$ (Eq. 5)
332	As described by Borges et al. (2019), we chose this parameterization because it is based on the most comprehensive
333	compilation of k values in streams which, in addition, was used in the global upscaling of CO2 emissions from rivers by both
334	Raymond et al. (2013) and Lauerwald et al. (2015).
335	2.4. C fluxes estimation at the Nyong watershed scale
336	2.4.1. CO2 degassing and heterotrophic respiration
337	In each stream order, monthly rate of CO ₂ degassing at the water-air interface (F _{degas} ; in mmol-gC-CO ₂ m ⁻² yrd ⁻¹) was estimated
338	as follows:
339	$F_{degas} = k_{600} K_0 (p_{CO2w} - p_{CO2a}) $ (Eq. 6)
340	where,
341	K_0 is the solubility coefficient of CO ₂ determined from the water temperature (Weiss, 1974), k_{600} is the monthly gas transfer
342	velocity of CO ₂ (section 2.3), pCO _{2w} and pCO _{2a} are the monthly partial pressures of CO ₂ in the surface waters of the different
343	stream orders and in the overlying atmosphere (set to 400 ppmv), respectively.
344	
345	In each stream order, we multiplied monthly F_{degas} in <u>gC-CO₂ m⁻² yr⁻¹mmol m⁻² d⁻¹</u> by the respective monthly water surface
346	area to estimate the monthly CO ₂ emissions (F_{degas} in tGgC-CO ₂ yr ⁻¹) integrated in each stream order. We summed F_{degas} in
347	tGgC-CO ₂ yr ⁻¹ in each stream order to estimate the total quantity of CO ₂ degassed from the Nyong watershed from the entire
348	river network and then normalized by the Nyong watershed surface area (tMgC-CO ₂ km ⁻² yr ⁻¹). Note, we did not measure
349	pCO ₂ in second-order streams but estimated the pCO ₂ by averaging the pCO ₂ measured in the first- and third-order streams.
350	
351	At the watershed scale, volumetric rates of heterotrophic respiration in the river were estimated from the increase in CO ₂ in
352	the incubated serum bottles over 24h in stream orders 1 and 5. The volumetric respiration rates in stream orders 1 and 5 were

³⁵³ 354 depth-integrated and subsequently averaged to estimate an average rate of heterotrophic respiration from the entire river

network of the Nyong watershed.

355 2.4.2. C export to the ocean

256		
356	The C hydrologically exported to the ocean (F_{ocean}) was calculated monthly at the most downstream station (Nyong at Ol	.ama)
357	as the following:	
358	$F_{ocean} = Q_{olama} [C]_{olama} $ (Eq	ı. 7)
359	where Qolama and [C]olama are the monthly average discharges and concentrations of POC, DIC or DOC at Olama, respectively of the second	ively.
360	F_{ocean} was estimated in $tGgC yr^{-1}$ and then normalized by the catchment surface area at Olama ($tMgC km^{-2} yr^{-1}$).	
361	2.5. Riverine-Stream C budget of the first-order Mengong catchment	
362	2.5.1. The different C fluxes	
363	At the Mengong catchment scale, as described above in the section 2.1.2, there are two sources fuelling the Mengong st	ream
364	with C, namely non-flooded forest groundwater (F_{GW}) and wetland (F_{WL}). The C entering the Mengong stream has two outputs the formula of the strength of the strengt	itputs
365	as this C is either degassed at the water-air interface (F_D) -in the form of CO ₂ (F_D) -or hydrologically exported at the st	tream
366	outlet (F _{OUT}). Noteworthy, heterotrophic respiration in the stream (F _{RH}) is considered as an C input for the DIC budget,	while
367	a C output for the DOC budget (assuming respiration occurs on DOC only). At the Mengong catchment scale, Rriverine	DIC,
368	DOC and POC budgets (DIC _{budget} , DOC _{budget} , POC _{budget}) are thus the difference between C inputs and outputs ₂ as follows	3:
369	$DIC_{budget} = F_{GW} + F_{WL} + F_{RH} - F_D - F_{OUT} $ (Eq	j. 8)
370	$DOC_{budget} = F_{GW} + F_{WL} - F_{RH} - F_{OUT} $ (Eq	<i>. 9)</i>
371	$POC_{budget} = F_{WL} - F_{OUT} $ (Eq	ı. 10)
372	Noteworthy, DIC _{budget} , DOC _{budget} and POC _{budget} these C budgets at the Mengong catchment scale cannot be estimated mo	nthly
373	as for F _{degas} or F _{ocean} at the Nyong watershed scale, because water fluxes described in Equations 1 and 2, in particular Q _{hi}	ill and
374	OF _{WL} , which are needed to estimate F _{GW} and F _{WL} (see section 2.5.2), can only be estimated yearly from yearly rainfall i	in the
375	Mengong catchment (see section 2.1.2).	
376	2.5.2. Hydrological C inputs to the stream from non-flooded forest groundwater and wetland	
377	According to equations 1 and 2, we can estimate the quantity of dissolved carbon leached from non-flooded forest ground	water
378	to the Mengong stream (F_{GW}) was estimated as the following:	
379	$F_{GW} = Q_{hill} [C]_{GW} $ (Eq	į. 11)

380 where,

381 [C]_{GW} is the yearly average concentration of DIC or DOC in the Mengong source. F_{GW} (tMgC yr⁻¹) iwas normalized by the

surface area of 0.48 km² drained by the hillside (tMgC km⁻² yr⁻¹). Noteworthy, F_{GW} represents hydrological input of C to the stream from the drainage of land (well-drained ecosystem).

384

385 a-A part of non-flooded forest groundwater fed the wetland (F_{GW-bis}) and can be estimated as the following:

386	$F_{GW-bis} = Q_{base} [C]_{GW} $ (Eq. 12)
387	F _{GW-bis} does not account to the stream C budget because Q _{base} is not feeding the stream, but does account to the total quantity
388	of C hydrologically leached from land. F _{GW-bis} (MgC yr ⁻¹) was normalized by the surface area drained by the hillside (MgC
389	<u>km⁻² yr⁻¹).</u>
390	
391	According to equations 1 and 2, we can estimate the quantity of dissolved C leached from the wetland to the Mengong stream
392	(F _{WL}) <u>was estimated</u> as the following:
393	$F_{WL} = (OF_{WL} + Q_{WLST}) * [C]_{WL} $ (Eq. 13)
394	where,
395	$[C]_{WL}$ are the concentrations of DOC or DIC in the topsoil solution (0.4 m) of the Mengong wetland, measured at 1 420±750
396	and 1 430±900 µmol L-1 by Braun et al. (2005) and Nkoue Ndondo et al. (2020), respectively. F _{WL} (tMgC yr-1) wasis
397	normalized by the surface area of 0.12 km ² drained by the wetland ($tMgC$ km ⁻² yr ⁻¹).
398	
399	In the Mengong catchment, as described in the section 2.1.2, overland flow on hillsides is negligible and there is no particulate
400	C in non-flooded forest groundwater. Therefore, it can be safely assumed that POC at the Mengong outlet should originates
401	mostly from the drainage and erosion of the wetland. Accordingly, it was assumed that the hydrological export of POC at the
402	Mengong outlet is similar to the POC hydrologically exported from the wetland (FwL). For POC, FwL can thus be estimated as
403	the following:
404	$F_{WL} = Q_{outlet} [POC]_{OUT} $ (Eq. 14)
405	where,
406	Qoutlet and [POC] _{OUT} are the yearly average discharge and POC concentration at the Mengong outlet, respectively. F _{WL} (tMgC
407	yr ⁻¹) wasis normalized by the surface area of the wetland ($tMgC$ km ⁻² yr ⁻¹).
408	2.5.3. CO2 degassing and heterotrophic respiration in the stream
409	It has been shown that a large fraction of C_{Ω_2} degassing in headwaters was actually missed by conventional stream sampling
410	because a large fraction of the degassing occurs as hotspots in the vicinity of groundwater resurgences (e.g., Deirmendjian and
411	Abril, 2018; Johnson et al., 2008). Therefore, we estimated F _D (MgC-CO ₂ yr ⁻¹) was estimated from a mass balance that
412	calculates the loss of the dissolved CO ₂ between non-flooded forest groundwater (F _{D-GW}) (or wetland; F _{D-WL}) and stream water,
413	using CO ₂ concentrations and drainage data, a method similar to Deirmendjian and Abril (2018) and Duvert et al. (2020a), as
414	the following:
415	$F_{D-GW} = ([CO_2]_{GW} - [CO_2]_{OUT}) * Q_{Hill}$ (Eq. 15)
416	$F_{D-WL} = ([CO_2]_{WL} - [CO_2]_{OUT}) * (OF_{WL} + Q_{WLST})$ (Eq. 16)
417	$F_{\rm D} = F_{\rm D-GW} + F_{\rm D-WL} \tag{Eq. 17}$
418	where,

419	[CO2]GW, [CO2]WL and [CO2]OUT are the yearly average CO2 concentrations in non-flooded forest groundwater, wetland, and
420	stream outlet, respectively, F _D , F _{D-GW} and F _{D-WI} (all three fluxes in MgC-CO ₂ yr ⁻¹) were then normalized by the surface area

stream outlet, respectively. F_D, F_{D-GW} and F_{D-WL} (all three fluxes in MgC-CO₂ yr⁻¹) were then normalized by the surface area 421 of the Mengong catchment (MgC-CO₂ km⁻² yr⁻¹), the surface area drained by the hillside (MgC-CO₂ km⁻² yr⁻¹), and the surface

422 area drained by the wetland (MgC-CO₂ km⁻² yr⁻¹), respectively.

2.5.4. C hydrologically exported at the Mengong stream outlet

423

424 Rates of heterotrophic respiration in the Mengong stream were estimated from the increase in CO2 in the incubated serum

425 bottles over 24h in the Mengong stream, which were subsequently depth-integrated. In the Mengong catchment, depth-

426 integrated rates of heterotrophic respiration in the river were multiplied by the Mengong stream surface area to obtain the

427 integrated contribution of heterotrophic respiration for the whole stream (F_{RH} in MgC-CO₂ yr⁻¹). To estimate the Mengong

- 428 stream surface area, stream width was estimated from equation 3 whereas stream length (750 m) was empirically determined 429 from field measurement by Maréchal et al. (2011). F_{RH} was then normalized by the surface area of the Mengong catchment
- 430 (MgC-CO₂ km⁻² yr⁻¹).

431

432	Based on equation 23, the quantity of C hydrologically exported at the outlet of the Mengong catchment (MgC yr ⁻¹) can be
433	estimated as the following:
434	$F_{OUT} = Q_{ST} [C]_{OUT} $ (Eq. 18)
435	where, [C] _{OUT} is the concentration of POC, DOC or DIC at the Mengong stream outlet, respectively. F _{OUT} (MgC yr ⁻¹) was ther
436	normalized by the surface area of the wetland (MgC km ⁻² yr ⁻¹).
437	2.5.5. Heterotrophic respiration in the stream

438 F_{RH} in mmol m³-d⁴ is the average heterotrophic respiration in the Mengong stream obtained from the increase in CO₂ in the 439 incubated serum bottles over 24h, FRH was converted in mmol m⁻² d⁻⁴ by multiplying by the average depth at the Mengong 440 stream outlet.

441 3. Results

442 3.1. Hydrology

443 In 2016, the discharges were 0.009±0.002 (range was 0-0.35), 3.9±4.8 (0-35), 35.6±40.6 (3.4-175), 146±112 (21-392) and 444 195±160 (8-640) m3 s⁻¹, in stream orders, 1, 3, 4, 5 and 6, respectively (Table 1, Fig. 2). All river discharges seasonally peaked 445 twice a year during the two rainy seasons, both separated by dry seasons; the groundwater water table followed the same trend 446 (Figs. 2, 4-5). Specifically, the beginning to middle of the rainy seasons corresponded to a period of increasing river discharge 447 and groundwater water table level, while the end of the rainy seasons and the dry seasons corresponded to a period of

448 decreasing river discharge and groundwater water table level (Figs. 2, 4-5). In each stream order, low-water period and lowest

449 discharges were observed during the long dry season (Figs. 2). The stream orders 1 and 3 were dried up during the long dry

450 seasons (from the 01st Jan. to the 15th Mar. 2016 and to the 28th Apr. 2016, for stream order 1 and 3, respectively) whereas the

451 streams with orders higher than 3 were never dried up (Fig. 2).

452 3.2. Seasonal variations of C and ancillary parameters in non-flooded forest groundwater

453 Yearly averages and ranges in C and ancillary parameters in non-flooded forest groundwater are detailed in Tables 2 and 3. 454 The coefficients of variation of groundwater temperature, pH and specific conductivity were lower than 5% showing a strong 455 stability for these parameters throughout the water cycle. Oxygen saturation in non-flooded forest groundwater increased 456 during the long dry season and peaked at the end of the same season (up to 68% the 30th Mar. 2016), then slowly decreased 457 towards the end of the long rainy season (down to 38% the 15th Nov. 2016) (Fig. 4). pCO₂ in non-flooded forest groundwater 458 concentration exhibited strong temporal variations (coefficient of variation was about 50%), and peaked in the middle of the 459 short (up to 100 000 ppmv the 16th Feb. 2016) and long (up to 200 000 ppmv the 01st Aug. 2016) dry seasons, while decreasing 460 during the two wet seasons (Fig. 4). All year long, DOC in non-flooded forest groundwater was below the detection limit of 1 461 mg L⁻¹ (<83 µmol L⁻¹); note we considered this threshold as the average DOC concentration in non-flooded forest groundwater. 462 Despite one peak of TA that was up to 138 µmol L⁻¹ the 29th Sep. 2016, TA in non-flooded forest groundwater was relatively 463 stable through the water cycle (Fig. 4).

464 3.3. Seasonal variations of C and ancillary parameters in surface waters

465 Yearly averages and ranges in C and ancillary parameters in surface waters are detailed in Tables 2 and 3. In streams orders 1 466 and 3 variations of pH, specific conductivity and oxygen saturation were weakly affected by the discharge as indicated by non-467 correlations between these parameters and the discharge in these streams (Table 4, Fig. 5). Nonetheless, in the stream order 3, 468 we observed an increased in oxygen saturation during dry periods (Fig. 5). On the contrary, in streams orders 4, 5 and 6, 469 variations of pH, specific conductivity and oxygen saturation as a function of river discharge were more pronounced as these 470 parameters peaked during dry seasons and decreased during rainy seasons as indicated by significant negative correlations 471 between these parameters and the discharge in these streams (Table 4, Fig. 5).

472

473 DOC concentration in stream order 1 increased at the beginning of the re-flowing period (i.e., at the beginning of the short 474 rainy season, up to 4 140 µmol L-1 the 14th Apr. 2016) (Fig. 5). In the other stream orderslarger streams (order > 1), a similar 475 DOC trend occurred but with a slight delay of about a couple of weeks in comparison to the one observed in stream order 1 476 (Fig. 5). In all stream orders, aAfter theis seasonal peak of DOC at the beginning of the short rainy season, DOC concentration 477 quickly decreased to reach minimum values during the following short dry season, then DOC concentration was rather stable 478 until the first rains fall again in the next short rainy season (Fig. 5). In stream order 1, POC and TSM concentrations also 479 peaked significantly at the beginning of the re-flowing period, driving the negative correlation of these two parameters with

the discharge in this stream order 1; we did not observe a similar increase in higher order streams (Table 4; Fig. 5). In addition, in stream order 1, POC%, POC and TSM concentrations increased during the two wet seasons, while decreased during the short dry season; a similar trend was observed in stream orders 5 and 6 as indicated by the positive correlations between POC and TSM and the discharge in <u>stream orders 5 and 6 these streams</u> (Table 4; Fig. 5). In contrast, in stream orders 3 and 4, TSM concentration did not follow this trend as it peaked during the short dry season and at the beginning of the long dry season (Fig. 5).

- In all stream orders, we observed an increase in TA concentration during the long rainy season followed by a quick decrease (Fig. 5). Overall, there was also a peak in TA concentration at the end of the long dry season followed by a decrease during the following short rainy and dry seasons, driving the significant negative correlations between discharge and TA concentration in stream orders 4, 5 and 6 (Table 4, Fig. 5). In the stream order 1, pCO₂ exhibited a similar trend to the POC, with values peaking during the two wet seasons (Fig. 5). In larger streams (order <u>>s higher than 1</u>), pCO₂ <u>also</u> seasonally peaked during the long rainy season, but more significantly in stream orders 5 and 6 as indicated by the positive correlations between pCO₂ and discharge in these streams <u>orders 5 and 6</u> (Table 4).
- 3.4. Spatial variations of C and ancillary parameters across non-flooded forest groundwater, and increasing stream
 orders

TSM and POC concentrations were not significantly different in streams orders 3, 5 and 6, but were significantly lower in stream order 1, while being significantly higher in stream order 4 (p<0.001, Kruskall-Wallis with Dunn's multiple comparisons tests) (Fig. 6). POC content of the TSM was significantly higher in stream order 1 in comparison to all other stream orders, while not being significantly different between stream orders 3 to 6 (p<0.05, Kruskall-Wallis with Dunn's multiple comparisons tests) (Fig. 6). DOC concentration was not significantly different between streams orders 1, 4, 5 and 6, but was significantly lower in non-flooded forest groundwater, while being significantly higher in stream order 3 (p<0.001, Kruskall-Wallis with Dunn's multiple comparisons tests) (Fig. 6).

503

486

The oxygen saturation was not significantly different between non-flooded forest groundwater and streams orders 1, 3 and 4, whereas it was significantly lower in the Nyong River (streams orders 5 and 6) (p<0.05, Kruskall-Wallis with Dunn's multiple comparisons tests) (Fig. 6). TA concentration was significantly higher in stream order 1 than in non-flooded forest groundwater (p>0.01, Mann-whitney test) (Fig. 6) (Fig. 5). In addition, TA concentration was significantly higher in streams orders 5 and 6 than in non-flooded forest groundwater and in streams orders 1, 3 and 4 (p<0.001, Kruskall-Wallis with Dunn's multiple comparisons tests) (Fig. 6). pCO₂ was significantly higher in non-flooded forest groundwater, while was similar in all other stream orders (p<0.001, Kruskall-Wallis with Dunn's multiple comparisons tests) (Fig. 6), even though pCO₂ decreased overall

511 from stream order 1 to 6 (Table 3).-



512 3.5. C budget at the Mengong catchment scale

513 The DIC_{budget} was well-balanced, showing inputs and outputs fluxes not statistically different (p>0.05; Mann-Whitney test) \$14 and differing only by 6% (Table 5),- This indicate indicating that all DIC fluxes have been considered and well constrained \$15 (Table 5). In contrast, the DOC_{budget} was not balanced, showing statistically different inputs and outputs fluxes (p<0.001; **\$**16 Mann-Whitney test) by 235240% (Table 5), This shows showing that unidentified DOC inputs were overlooked from the \$17 estimated budget (Table 5). The quantity of hydrologically exported C from non-flooded forest groundwater (F_{GW} + F_{GW-bis}) **5**18 was 6.8±3.0 tMgC yr⁻¹ (14.1±6.2 tMgC km⁻² yr⁻¹), DIC contributing for 97% (Fig. 7). Noteworthy, 10% of the C hydrologically **5**19 exported from non-flooded forest groundwater this export goes to the wetland (FGW-bis) rather than the stream (FGW); and 97% \$20 of this hydrological export of C occurred as DIC (Fig. 7). The annual flux of thequantity of hydrologically exported C from \$21 wetland to the stream (FwL) was 4.0±1.5 tMgC yr⁻¹ (33.0±12.4 tMgC km⁻² yr⁻¹); DOC, DIC and POC contributing for 45, 45 \$22 and 5%, respectively (Fig. 7). The annual flux of C degassed to the atmosphere as CO_2 (F_D) was 5.5±2.3 tMgC-CO₂ yr⁻¹, while \$23 the heterotrophic respiration in the stream (F_{RH}) was 0.3±0.4-3 tMgC-CO₂ yr⁻¹ (Fig. 7).

\$24 3.6. CO2 degassing and C export to the ocean at the Nyong watershed scale

525 Spatially, yearly averages of monthly k_{600} increased from stream order 1 (2.2±0.1 m d⁻¹) to 4 (3.0±0.3) and subsequently \$26 decreased downstream in stream orders 5 (2.3±0.1) and 6 (2.5±0.2), (Table 6). In contrast, monthly k600 did not exhibit much \$27 seasonal variations (Table 6; Fig. S24). Spatially, yearly averages of monthly CO2 degassing rates were similar in stream 528 orders 1, 2, 3 and 4 but significantly lower in stream orders 5 and 6 (p<0.001, Kruskall-Wallis with Dunn's multiple \$29 comparisons tests) (Table 6). Rates of heterotrophic respiration were 28646 ± 2282 and 756454 ± 3334 gC-CO₂ m⁻² yr⁻¹ mmol-m⁻² \$30 2 d⁺ in stream order 1 and 5, respectively, whereas CO₂ degassing rates were $5344\pm27731220\pm640$ and $3706\pm1540846\pm350$ \$31 $gC-CO_2$ m⁻² yr⁻¹mmol m⁻² d⁺¹ in the same stream orders, respectively (Table 6). Seasonally, considering all stream orders, the \$32 monthly average CO_2 degassing rates during rainy seasons wasere in average 20% higher in comparison to the average CO_2 \$33 degassing rates during dry seasons, explaining higher integrated CO2 degassing during rainy seasons at the Nyong watershed \$34 scale (Fig. 8). In addition, at the Nyong watershed scale, the yearly integrated CO₂ degassing (F_{degas}) was $650652\pm160\cdot161\cdot10^3$ \$35 tGgC-CO2 yr⁻¹ (23.54±5.6-8 tMgC-CO2 km⁻² yr⁻¹ when weighed by the Nyong watershed surface area); and the yearly \$36 integrated hydrological C export to the ocean (Focean) was 12±9-10 403-tGgC yr-1 (0.6±0.5 tMgC km-2 yr-1) for POC, \$37 $\frac{130134\pm90100-10^{3}+GgC}{10} \text{ yr}^{-1} (7.2\pm5.4 \text{ tMgC km}^{-2} \text{ yr}^{-1}) \text{ for DOC, and } 46\pm42 \pm \frac{10^{3}+GgC}{10} \text{ yr}^{-1} (2.5\pm2.3 \text{ tMgC km}^{-2} \text{ yr}^{-1}) \text{ for DIC};$ \$38 more than 50% of Focean these fluxes occurreding during the long rainy season (Tables 6-7; Fig. 8).

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539 4. Discussion

564

540 4.1. Non-flooded forest groundwater and wetland as C sources in a first-order catchment

541 The drainage of non-flooded forest groundwater (i.e., groundwater from the hillside lateritic system) and wetland (i.e., 542 hydromorphic system) fuels the Mengong stream with organic and inorganic C (Figs. 3, 7; Boeglin et al., 2005; Viers et al., 543 1997). In the hillside lateritic system, overland flow is negligible owing to limited soil erosion due to dense vegetation cover 544 and high soil porosity facilitating rainfall infiltration (Braun et al., 2005; Maréchal et al., 2011). Consequently, hydrological 545 export of soil C to the stream by overland flow from the hillside is considered as negligible. In contrast to the hillside lateritic 546 system, overland flow is a possible C pathway from the hydromorphic wetland system to the stream (Fig. 3; Maréchal et al., 547 2011). Thus, the stream POC shall originates mostly from the overland flow over the wetland, as also suggested by similar 548 δ^{13} C values of total organic carbon (TOC) in the wetland soil and in the POC observed in the stream outlet (range was -28 to 549 -31‰) of the Mengong catchment by Nkoue-Ndondo et al. (2020). The fact that POC and TSM concentrations in the Mengong 550 stream increased during rainy seasons, when the hydrological connectivity with the surrounding wetland is enhanced, is also 551 in a good agreement with the identification of wetland as the main (if not exclusive) source of POC and TSM. Furthermore, 552 Nkoue-Ndondo et al. (2020) did not observe seasonal variations of the δ^{13} C-POC signature in the Mengong stream. This 553 suggests that the additional POC source observed at the beginning of the reflowing period also originates from the erosion of 554 the wetland even though this hydrological period was characterized by a weaker hydrological connectivity with the wetland 555 compared to rainy seasons. In the Mengong wetland, litter-fall measurement by Nkoue-Ndondo (2008) was 116 t yr⁻¹ of wet \$56 OM with a mean C content of 22.5%, which is equivalent to 26 tMgC yr⁻¹, a flux 75-times higher than our conservative \$57 estimation of the POC leached from the wetland to the stream (0.3 tMgC yr⁻¹, Fig. 7). This implies that most of the wetland 558 litter-fall accumulates in the wetland soil rather than being hydrologically exported to the stream in the form of POC, in 559 particular due to limited overland flow in the wetland due to flat topography (Maréchal et al., 2011). However, the in-situ \$60 degradation of this highly labile OM_from litter-fall might contribute to the DOC and DIC fluxes from the wetland to the 561 stream. Indeed, tropical wetlands are recognized as productivity hotspots and a large fraction of the litter-fall is degraded in-562 situ by heterotrophic respiration in the water and sediment, enriching wetland waters in DOC and DIC (Abril et al., 2014; 563 Borges et al., 2015a).

In the Mengong catchment, waters originating from the drainage of non-flooded forest groundwater and wetland are considered as clear and coloured waters, respectively, the colour reflecting their DOC content (Boeglin et al., 2005; Viers et al., 1997). Indeed, DOC concentration was low in clear waters (<83 µmol L⁻¹) whereas <u>DOC concentration</u> # was high in coloured waters (1_420±750 µmol L⁻¹) (Table 3; Viers et al., 1997). The DOC in the soil solution has distinct sources that are litter leaching, root and microbial exudates, rainfall (throughfall and stemflow), and decaying fine roots (Bolan et al., 2011; Kalbitz et al., 2000). Once in the soil solution, DOC is however rapidly adsorbed onto soil minerals during its percolation through the soil column due to the soil capacity for DOC stabilization (Kothawala et al., 2009; Neff and Asner, 2001) by sorption on Fe (and

\$72 Al) oxides and hydroxides and clay minerals (Kaiser et al., 1996; Kothawala et al., 2009; Sauer et al., 2007). DOC sorption in \$73 soilsprocess significantly reduces DOC mineralisation rates in soils (Hagedorn et al., 2015; Kalbitz et al., 2005; Kalbitz and \$74 Kaiser, 2008) and DOC export from soils (Shen et al., 2015). DOC sorption in soils^{III} also partly explains the decreasing 575 gradient of DOC concentration with depth commonly observed in boreal (e.g., Moore, 2003), temperate (e.g., Deirmendjian 576 et al., 2018) and tropical (e.g., Johnson et al., 2006) soils. DOC sorption in soils is actually strongly related to the availability 577 of Fe (and Al) oxides and hydroxides, and clay minerals, which are present both in the hillside lateritic and in the hydromorphic \$78 wetland soils of the Mengong catchment (Fig. 3S1). In the hillside lateritic system, soil DOC is probably well stabilized in the 579 iron-rich and clay horizons preventing DOC leaching to the non-flooded forest groundwater (Braun et al., 2005, 2012). \$80 Furthermore, DOC must be desorbed from soil minerals in order to be exported to groundwater (Deirmendjian et al., 2018; 581 Sanderman and Amundson, 2008). Studies have shown that water saturation of the topsoil generates reducing conditions in \$82 the saturated soil (Camino-Serrano et al., 2014; Fang et al., 2016) and that this processwhich limits the retention of soil DOC 583 and thus enhances its export to groundwater (Deirmendjian et al., 2018). In the hillside lateritic system, the non-flooded forest 584 groundwater table never reaches the topsoil where soil DOC is high. Therefore, DOC adsorption in these soils might be \$85 enhanced. In the hydromorphic wetland system, the groundwater saturates the topsoil all year long (Fig. S13) which might 586 reduce DOC adsorption in this compartment. In addition, hydromorphic conditions occurring in the Mengong wetland soil 587 favour the solubilisation of Fe (Oliva et al., 1999), which is supposed to reduce DOC sorption. Altogether, this explains the 588 low and high DOC concentrations observed in the non-flooded groundwater and the wetland, respectively. In addition, the 589 results showed that stream DOC increased during the first wet season only. In the Mengong catchment, Nkounde-Ndondo 590 (2008) described the piston flow that occurs at the beginning of the short rainy season, which is caused by new infiltration of 591 water on the hills and hillsides that pushes the older soil water downstream (e.g., Huang et al. (2019) and references therein), \$92 allowing pressure on the aquifer and thus exfiltration at the bottom of the slope (i.e., in the wetland; Fig. 3). This implies **\$**93 thatConsequently, wetland DOC is quickly flushed during the first rains and originates from the subsurface horizons of the 594 wetland soil. Later in the season, the decrease of stream DOC is due to dilution with non-flooded forest groundwater with low 595 DOC content. Noteworthy, our stream DOC budget was not balanced (Table 5; Fig. 7), indicating that sources contributing to 596 the DOC content of the Mengong stream were overlooked. An additional DOC source that was quantified by Braun et al. 597 (2005) during 4 years in the Mengong catchment is DOC in the throughfall. These authors determined that the average DOC **\$**98 concentration in the throughfall was $3.6\pm3.5 \text{ mg L}^{-1}$. Applying this average DOC concentration in the throughfall to the rainfall **\$**99 in 2016 and the catchment surface area gives an additional DOC input from precipitation of 4.3±4.3 tMgC yr⁻¹, which allows 600 closing the DOC budget at the Mengong catchment scale. 601

602 Non-flooded forest groundwater and wetland exhibited high DIC concentrations, 2 940±1485 and 1 430±900 µmol L⁻¹,

603 respectively and, in both systems, DIC was mostly in the CO2 form (>90%) (Table 3). In the hillside system, non-flooded

604 forest groundwater was free of DOC. This result, along with the fact that mMicrobial activity has been shown to be limited in

605 many aquifers by the availability of DOC (e.g., Malard and Hervant (1999) and references therein). Thus, ass -non-flooded

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606 forest groundwater was free of DOC, suggest that CO2 in non-flooded forest groundwater likely_comes from soil respiration 607 in the overlaying non-saturated soil - rather than respiration in-within the groundwater-itself - and then is transported downward 608 by diffusion rather than percolation with rain water. Indeed, the thickness of the lateritic cover on hills and slopes of the 609 Mengong catchment considerably slows the water percolation in the bedrock (Boeglin et al., 2005). In the tropics, the soil 610 respiration rate is mostly affected by soil moisture as soil temperature exhibits low seasonal variations (Davidson et al., 2000). 611 Accordingly, soil respiration rates usually decrease from rainy to dry seasons in tropical ecosystems due to decreasing soil **6**12 moisture (Davidson et al., 2000; Schewendenmann and Veldkamp, 2006)(Davidson et al., 2000). Nevertheless, in the Mengong 613 $catchment, pCO_2 \ in \ non-flooded \ forest \ groundwater \ peaked \ during \ dry \ seasons \ and \ started \ to \ decrease \ later \ in \ the \ same \ season$ 614 and then during the following rainy season (Fig. 4). In mature forest of Amazonia, Johnson et al. (2008) observed a similar 615 trend in groundwater that they attributed to an increase in vegetation water uptake and roots activity in deep soils during the **6**16 onset of the dry seasons. Indeed, during dry seasons, tropical mature forest depends on deep root system to extract water from 617 the soil and deep root system also provide inorganic and organic C to the deep soil trough root respiration and exudation 618 (Nepstad et al., 1994), they also showed that groundwater pCO2-decreased later in the season because of losses due to drainage 619 and diffusional losses. Furthermore, during dry seasons, the diffusion of CO_2 in the porous soil is facilitated in tropical forest **6**20 (Adachi et al., 2006) because low soil water content increases air-filled pore space (Schewendenmann and Veldkamp, 2006), 621 very likely favouring the downward diffusion of soil CO2 and its subsequent dissolution in groundwater, as also observed in **6**22 temperate forests (Deirmendjian et al., 2018; Tsypin and Macpherson, 2012) (Deirmendjian et al., 2018). In the non-flooded 623 forest groundwater, oxygen saturation was about 40% but increased during dry seasons whereas decreasing during rainy **6**24 seasons (Fig. 4). This shows that a Atmospheric air can thus penetratespenetrate the soil atmosphere deeply, in particular during 625 dry seasons when the diffusion in the porous soil is facilitated, and can reach the non-flooded forest groundwater. In the 626 wetland hydromorphic system, the soil is permanently saturated which limits aerobic respiration of microbes in the soil and **6**27 leading to the accumulation of OM in the soil profile. This likely explains, likely explaining the lower CO2 concentration 628 observed in the wetland compared to non-flooded forest groundwater (Table 3). Nonetheless, it should be noted that wetland 629 vegetation can actively transport oxygen to the root zone via their aeremchyma (Haase and Rätsch, 2010), creating a complex 630 oxic-anoxic interface that promotes aerobic respiration but also supplies labile OM to anaerobic degradation (and 631 methanogenesis) fuelling CO₂ (and CH₄) production (Piedade et al., 2010). This is in a good agreement with δ^{13} C-DIC 632 signatures of -16‰ measured by Nkoue Ndondo et al. (2020) in the wetland soil, which are indeed close to the C4 signature 633 of aquatic grassland found in the Mengong wetland. In addition to drainages of non-flooded forest groundwater and wetland, **6**34 stream DIC can also originates from in-situ respiration of DOC. In-situ respiration of DOC. This process is corroborated by 635 our results of incubations (Table 6), and by the δ^{13} C-DIC at the Mengong stream outlet that was more depleted in 13 C than in 636 non-flooded forest groundwater and wetland (Nkoue Ndondo et al., 2020), which highlights in-stream respiration from an 637 organic ¹³C-depleted source.

638

639 Non-flooded forest groundwater and wetland both exhibited low TA concentrations, 53±26 and 122±46 µmol L⁻¹, respectively; 640 nonetheless TA concentration was significantly higher in wetland (Table 3; Fig. 6). Considering the granitic lithology (i.e., 641 absence of carbonate minerals) of the Nyong watershed, TA in non-flooded forest groundwater and wetland might originate 642 from the weathering of silicate minerals as dissolved CO2 can react with silicate minerals to produce bicarbonates (Meybeck, 643 1987). Applying TA concentration in non-flooded forest groundwater into Equations 11 and 12 results in a silicate weathering 644 rate in the overlaying lateritic soil of 0.2±0.1 tMgC km⁻² yr⁻¹, whereas applying TA concentration in wetland into Equation 13 645 results in a silicate weathering rate in wetland of 1.3±0.4 tMgC km⁻² yr⁻¹. The silicate weathering rate in the wetland soil is 646 thus 550% higher than in the non-flooded lateritic soil. Even though these two rates remain low compared to weathering rates **6**47 in carbonated environment, they are typical of silicate weathering rates which are in the range 0.1-5.2 tMgC km⁻² yr⁻¹ as 648 estimated from diverse worldwide basins by Amiotte Suchet et al. (2003). In non-flooded forest groundwater, the low TA 649 concentrations and silicate weathering rates, along with, the absence of significant seasonal variations of TA, are likely related 650 to the relatively inert mineralogy of the lateritic soil cover (Braun et al., 2005, 2012). In the Nyong watershed, these low 651 silicate weathering rates are in a good agreement with the low mineral dissolved load in the aquifer (Braun et al., 2002) and 652 by the dissolved silica fluxes in rivers that were significantly lower compared to the annual rainfall (White and Blum, 1995). **6**53 In addition, silicate weathering rates in the wetland might be enhanced by the leaching of humic acids from the vegetation to 654 the hydromorphic soils (Braun et al., 2005; Nkoue-ndondo, 2008).

455 4.2. Influence of wetland-river connectivity <u>on riverine C cycling</u> at the Nyong watershed scale

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656 The role of wetland on riverine C cycling in tropical watersheds is commonly explored using empirical relationships between 657 wetland extent and C concentrations in the stream water of the different sub-catchments of a given watershed. Establishing 658 such empirical relationships in the Nyong watershed is extremely challenging owing the similar wetland extent (about 5% of 659 the surface area; Table 1) in the sub-catchments, with the exception of the first-order Mengong catchment where the wetland 660 extent represents 20%. However, this role can be explored by comparing the seasonality of C concentrations in stream order **6**61 1 - in which the wetland dynamic as a riverine C source has been discussed in the above section - with respect to the other 662 larger streams (order > 1)s. Thus, for a given parameter, similar seasonality in stream order 1 and the other stream orders the 663 larger streams might suggest that C sources and processes are similar in both (sub)systems.

Similarly to what we observed in the Mengong catchment, wetlands might be also considered as the main source of POC for surface waters in the whole Nyong watershed based on (1) the low slopes in the watershed, (2) the high infiltration capacity of the soil, (3) the similar normalized export of POC from wetland to the Mengong stream (order 1) and from the Nyong watershed to the ocean (Tables 5 and 7), and (4) the probable low pelagic primary production in the surface waters of the Nyong watershed, as usually observed in tropical rivers with high DOC concentrations (>1_500 μ mol L⁻¹) where light attenuation caused by browning (coloured waters) strongly limits aquatic photosynthesis (Borges et al. 2019). Moreover, the seasonality of POC was similar in stream order 1 and in high-order streams, increasing during rainy seasons while decreasing

672 during dry seasons (Fig. 5). This suggests that in high-order streams Thus, the POC leached from wetlands from low-order 673 catchments might acts as an important POC source to high-order streams. However, during rainy seasons, the higher POC 674 concentration observed in high-order streams in comparison to the stream order 1 (Figs. 5-6) might also suggests an additional 675 POC source in in high-order streams these streams during rainy seasons. In high-order streams, given that POC% increased 676 during rainy seasons, river bed and banks erosion is not likely as this process would have exported more TSM than POC, as 677 observed in the tropical Tana River in Kenya by Tamooh et al. (2012). As pelagic primary production is also unlikely, POC 678 leached from wetlands riparian to high-order streams and POC leached from floating macrophytes that develops in the river 679 bed of high-order streams during the dry seasons anterior to the rainy seasons are more suitable hypotheses to explain the 680 additional POC source observed in high-order streams during rainy seasons. Indeed, as in the Amazonian basin (e.g., Abril et 681 al., 2014; Engle et al., 2008; Silva et al., 2013), we observed in high-order streams the development of floating macrophytes 682 during dry seasons. In high-order streams, the development of these floating macrophytes was accompanied by peaks of 683 oxygen saturation during dry seasons (Table 4; Fig. 5). This last feature is in line with the high photosynthesis capacity of 684 macrophytes that results in oxygen-enriched water during daylight (Sabater et al., 2000). According to the flood pulse concept 685 in tropical rivers by Junk et al. (1989), floating macrophytes might be hydrologically exported during rainy seasons when the **6**86 river discharge increased sufficiently. In high-order streams of the Nyong watershed, theis seasonal wetland and floating **6**87 macrophytes flush of \underline{C} and \underline{OM} is also supported by other evidences such as higher pCO_2 and POC% along with lower oxygen 688 saturation observed in these streams. On the one hand, these features might be attributed to enhanced heterotrophic respiration 689 in the river fuelled by export of freshly-produced and young OM (Engle et al., 2008; Mayorga et al., 2005; Tamooh et al., 690 2014). BesidesMoreover, OM leached from tropical wetland can be photodegraded downstream into more labile lower 691 molecular weight compounds that in turn also enhances heterotrophic respiration in the river, as observed in the Congo River 692 by Lambert et al. (2016). On the other hand, the drainage of wetland can also directly account for CO2 emissions from surface 693 waters as under flooded conditions, roots and microbial respiration occurring in wetland directly release CO2 to the water 694 (Abril et al., 2014; Moreira-Turcq et al., 2013). These two patterns usually explain the positive correlation between pCO2 and 695 river discharge in tropical systems (Table 4; Borges et al., 2019). On the contrary, during dry periods, the wetlands are 696 shrinking and the river become more hydrologically disconnected from wetlands, explaining the lower pCO2 in tropical rivers 697 during dry seasons (Abril and Borges, 2019). The importance of river-wetland connectivity was also evidenced by the first 698 POC increase at the beginning of the re-flowing period that was not observed downstream (Fig. 5). This suggests POC was 699 quickly oxidized in-situ, or did not reach downstream due to weak hydrological connectivity with high-order streams during 700 this period. Indeed, when the Mengong (stream (order 1) was flowing again, the downstream Awout River (stream (order 3) 701 was still dry. This highlights the complex deposition and remobilisation cycles of TSM and POC in tropical rivers (Geeraert 702 et al., 2017; Moreira-Turcq et al., 2013). Finally, in stream orders 3 and 4, we observed an additional increase of TSM during 703 dry seasons, while POC% decreased (Fig. 5). This suggests that more TSM than POC was leached into these streams during 704 dry seasons. We assume that river bed and banks erosion could drive this seasonal trend. In the tropical Tana River in Kenya, 705 based on radionuclide's ratio reflecting the age of TSM, Tamooh et al. (2014) showed that TSM was old and increased during

706 dry seasons. This was attributed to inputs of older sediments, with river banks erosion and/or resuspended sediments suggested 707 as the main sources.

708

709 In surface waters, in contrast to pCO₂ and POC data, we did not observe a positive correlation between DOC and the river 710 discharge, in agreement with Brunet et al. (2009) who showed that DOC in the Nyong watershed was only flushed during a 711 short period of time at the beginning of the short rainy season (Fig. 5). In contrast to POC, DOC did not peak a second time 712 during the long rainy season (Fig. 5). We have no explanation to this trend, except that possibly, the fact that this probable the 713 second flush of DOC was faster than our fortnightly sampling frequency. Nonetheless, DOC exhibited a similar seasonality in 714 stream order 1 and high-order streams, but with a slight lag time due to the time the water needs to flow from upstream to 715 downstream showing that wetland from low-order streams are significant sources of DOC for downstream rivers. In addition, 716 in the Awout River (order 3), a significant increase in DOC was observed at the beginning of the reflowing period indicating 717 an additional source of DOC (Fig. 5). Actually, before the reflowing period, the bed of the Awout River (order 3) was 718 completely vegetated by large macrophytes (up to 2 m tall) and many small pockets of stagnating water remained. DOC could 719 accumulate in these stagnating waters and be remobilized when the water flows again, as observed in temperate rivers 720 (Deirmendjian et al., 2019; Sanders et al., 2007). The seasonal wetland flush in high-order streams can be also evidenced by 721 peaks of TA during the long rainy seasons, while the increase in TA in streams orders 5 and 6 during the long dry seasons 722 could not be explained by wetland inputs to river. In stream orders 5 and 6, during the long dry season, surface waters are 723 likely fed by deeper groundwater, which are older and likely characterized by higher TA concentrations than shallower levels, 724 as observed in temperate (Deirmendjian and Abril, 2018) and tropical (Duvert et al., 2020b) catchments. Duvert et al. (2020b) 725 In the later study situated in a small tropical catchment in Australia, the authors gave additional evidence of a shift from 726 biogenic (wetlands) to geogenic C source during dry seasons caused by changing water sources.

727 4.3. C fluxes at the plot (first-order) and the watershed scales

728 At the first-order Mengong catchment scale, we estimated each fluxes of the stream C budget were estimated independently. 729 Hydrological C inputs from wetland (F_{WL}) and non-flooded forest groundwater (F_{GW}) to the stream contributed to 38% 730 (4.0±1.5 tMgC yr⁻¹) and 62% (6.32±3.0 tMgC yr⁻¹) of the total hydrological C inputs, respectively (Table 5; Fig. 7). However, 731 when the later fluxes are weighed by respective surface area, F_{WL} and F_{GW} contributed to 73% (33.30±12.5.4 MgtC yr⁻¹) and 732 27% (13.40±6.2 tMgC yr⁻¹) of the total hydrological C inputs to the stream, respectively (Fig. 7). In the first-order Mengong 733 catchment, 83% and 17% of the CO2 degassing (58% and 42% if weighed by surface area) from the stream are sustained by 734 inputs of DIC from non-flooded forest groundwater and wetland, respectively (Fig. 7). At the Nyong watershed scale; our 735 Fstudy design did not allow rom our study design it was not possible to estimate these tweed contributions of CO2 degassing 736 from non-flooded forest groundwater and wetland at the Nyong watershed seale. However, we might assume that the wetland 737 contribution to CO2 degassing become greater with increasing stream order, particularly considering larger riparian wetlands

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738 in high-order streams and the development of floating macrophytes in river bed during dry seasons (Olivry, 1986). 739 Nonetheless, our results are in line with the growing consensus that tropical wetlands contribute significantly to the C inputs 740 in tropical rivers (Abril et al., 2014; Borges et al., 2015a, 2019, 2015b; Duvert et al., 2020a, 2020b). In the Mengong catchment, 741 an important fraction (~50%) of the C entering the stream directly returns to the atmosphere through CO2 degassing at the 742 water-air interface (Fig. 7); the remaining C is transported, processed and further degassed downstream (Abril et al., 2014). In 743 the Nyong watershed, our estimated k₆₀₀ are typical of lowland tropical rivers (e.g., Alin et al., 2011; Borges et al., 2019), and 744 their The weak seasonality of $\rho ur k_{600}$ shows that higher CO₂ degassing rates during rainy seasons are rather a function of the 745 increase of CO2 water-air gradient during rainy seasons - which is due to seasonal flush of wetland and macrophytes - rather 746 than the increase in k600 usually observed during high water periods because of increasing water turbulence. In the Nyong 747 watershed, the average heterotrophic respiration in the river (pelagic plus benthic) rate was 99521 ± 40327 gC-CO₂ m⁻² yr⁻¹ 748 mmol m²-d⁺ on average-whereas the average CO₂ degassing rate was 5 085±2 544 gC-CO₂ m⁻² yr⁻¹ 1+60±580 mmol m⁻²-d⁺ 749 (Table 6). This implies that Consequently, only ~8.510% of the degassing at the water-air interface was supported by 750 heterotrophic respiration in the river. These rates are consistent with observations measurements by Borges et al. (2019), who 751 showed that, in the Congo basin, the heterotrophic respiration (pelagic only) in the river averaged 81-355 gC-CO2 m⁻² yr⁻¹ 752 mmol-m²-d⁺ and represented ~11% of the average $\underline{CO_2C}$ degassing rate of $\underline{3}$ 240740 $\underline{gC-CO_2}$ m⁻² yr⁻¹ yrmmol-m²-d⁺. In the 753 same way, heterotrophic respiration in the river accounts for less than 20% of the CO2 degassing flux from the Amazon Basin 754 (Abril et al., 2014). Moreover, in the Nyong watershed, the ratio between rates of CO₂C degassing rates and heterotrophic 755 respiration in the river decreased in the stream order 5 (ratio of 6.54.9) compared to stream order 1 (ratio of 4818.6) (Table 6). 756 This is in line with the recent findings by Hotchkiss et al. (2015) in temperate rivers, where they showed that the contribution 757 of internal metabolism to account for CO2 emissions increased from upstream to downstream, or with the more recent findings 758 by Borges et al. (2019) in the Congo basin who found a ratio of CO₂ degassing rates to heterotrophic respiration in the river 759 rates of 29-137 and 3-17 in low- and high-order streams, respectively. Borges et al. (2019) These authors attributed their ⁷⁶⁰ observations to the prevalence of lateral CO2 inputs in sustaining CO2 emissions. 761 762 In the Nyong watershed, about 6% ($0.6\pm0.5 \pm MgC \text{ km}^{-2} \text{ yr}^{-1}$), 69% ($7.2\pm5.4 \pm MgC \text{ km}^{-2} \text{ yr}^{-1}$) and 24% ($2.5\pm2.3 \pm MgC \text{ km}^{-2} \text{ yr}^{-1}$) 763 1) of the Focean occurs in the POC, DOC and DIC forms, respectively, (Table 7). These C exports to the ocean are consistent but

slightly different from those reported by Meybeck (1993) for rivers in tropical humid regions, as he estimated that 20% (1.9 tMgC km⁻² yr⁻¹), 53% (5.1 tMgC km⁻² yr⁻¹) and 27% (2.6 tMgC km⁻² yr⁻¹) occurs in the POC, DOC and DIC forms, respectively. Therefore, in the Nyong watershed, the export of DIC to the ocean was typical of humid tropical regions while the export of POC was lower and DOC was higher. In the Nyong watershed, lower POC export to the ocean might be explained by the low watershed slope and the negligible overland flow that limits soil erosion. In contrast, DOC concentration in the surface waters of the Nyong watershed was in the upper range of those reported for other African rivers (range is 50 to 4 270 µmol L⁻¹; Tamooh et al. 2014 and references therein), thereby driving the higher DOC export to the ocean, which might be explained by

771 higher wetland extent than in the other African rivers. Huang et al. (2012) estimated the quantity of C exported to the ocean

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772	from African tropical rivers (30°N-30°S) at 0.3, 1 and 0.6 tMgC km ⁻² yr ⁻¹ for the POC, DOC and DIC forms, respectively, but
773	they did not partition these tropical rivers in humid or dry climates; our estimations of C export to the ocean were significantly
774	higher for the tropical Nyong watershed located in humid climate region. This shows the importance to upscale C fluxes for
775	the same climatic regions, such as the widely used Koppen-Geiger climate classification system (Koppen, 1936) recently
776	updated by Peel et al. (2007), otherwise upscaling might be strongly biased. In the Nyong watershed, the ratio between the C
777	exported to the ocean and the CO2 emitted to the atmosphere is 1:0.3, in agreement with ratio of 1:0.2 measured by Borges et
778	al. (2015b) in the Congo River but contrasting with the global ratio of 1:1 estimated by Ciais et al. (2013) during the Fifth
779	Assessment Report of the Intergovernmental Panel on Climate Change (IPCC). It shows that, showing that, at least African
780	rivers but probably all tropical rivers are strong emitters of CO2, and therefore Therefore, biogeochemical data in Africanthese
781	rivers are urgently required to improve accuracy of regional and global CO2 emission estimates from inland waters, and
782	understand how they will respond to climate change (warming, change in hydrological cycle).
783	
784	The integration of the different C fluxes can be were done by comparing them with the terrestrial C budget. In the Mengong
785	catchment, the total hydrological export of C from land and wetland (F _{GW} , F _{GW-bis} , F _{WL}) represents ~3-5% of the catchment net
786	C sink (range 201-336 tMgC yr ⁻¹) (Fig. 7). This conclusion-low hydrological C export to the aquatic environment relative to
787	the catchment net C sink agrees with two plot studies in temperate ecosystems, which have shown that the hydrological export
788	of C from forest ecosystems is ~3% (Deirmendjian et al., 2018; Kindler et al., 2011). In the Nyong watershed, the sum of the
789	$yearly C\underline{O_2} \ degassed \ (F_{degas}) \ \underline{from \ the \ river \ network} \ and \ \underline{the \ C} \ hydrologically \ exported \ to \ the \ ocean \ (F_{ocean}) \ represented \ \underline{together} \ begin{picture}{llllllllllllllllllllllllllllllllllll$
790	~10% of the net terrestrial C sink estimated by Brunet et al. (2009) (Table 78). This conclusion agrees with Similarly, Duvert
791	et al. (2020a), which showedestimated that the C degassed to the atmosphere and hydrologically exported at the river outlet
792	represented \sim 7% of the local net terrestrial C sink in the small (140 km ²) tropical Howard catchment in Australia, \sim 20% if
793	accounting to C losses via fire. In contrast, from a modelling approach in the entire Amazon watershed, Hastie et al. (2019)
794	found that C degassed and hydrologically exported might represents 78% of the net terrestrial C sink. This is in line with
795	findings of Abril et al. (2014) and Borges and al. (2019) who respectively found that C degassed from the Amazon and Congo
796	watersheds was greater than the local net terrestrial C sink. Besides, Abril et al. (2014) attributed this C riverine CO ₂ degassing
797	from rivers to wetland C inputs as they showed that tropical wetland can-may hydrologically export 36-80% of their gross
798	primary production (GPP) while terrestrial landscapes hydrologically export few percent of their net C sink, between 3% for
799	forests and 13% from grasslands (Kindler et al., 2011). Altogether, this shows that in large watersheds such as the Amazon or
800	the Congo rivers, fluvial C losses could offset more significantly the local net terrestrial C sink compared to relatively small
801	tropical watersheds such as the Nyong or the Howard rivers, which is likely due to both more extensive wetland and greater
802	hydrological fluxes in the Amazon and the Congo.

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803 Conclusions

804 In a first-order catchment, we showed here by determining all the terms of the C mass balance independently that attributing \$05 the whole amount of the CO₂ emitted to the atmosphere and C exported to the stream outlet to a unique terrestrial source and 806 ignoring the river-wetland connectivity might leaded to the misrepresentation of C dynamics in small tropical catchments and 807 thus likely at larger scales. Indeed, in addition to the drainage of non-flooded forest groundwater to the stream, we highlighted 808 the drainage and erosion of wetland as an important C source for the stream. Non-flooded forest groundwater was a significant 809 source of C for surface waters, particularly for CO2, whereas in contrast, DOC and POC in surface waters were mainly provided 810 by the drainage and erosion of wetlands. The flush of C from wetland to first-order streams is seasonally enhanced during 811 rainy seasons when the connectivity with surface waters is greater, allowing the leaching of freshly and young OM to the 812 stream, and thus increasing heterotrophic respiration in the river downstream. Nonetheless, at the Nyong watershed scale, the \$13 CO_2 emissions from the entire river network remained largely sustained by <u>direct</u> inputs of CO_2 from land and wetland, as \$14 heterotrophic respiration in the river represents only ~8.510% of the CO2 degassing at the water-air interface. Moreover, at the \$15 Nyong watershed scale, we showed that the CO2 degassed from the entire river network and the C hydrologically exported to \$16 the ocean might offset ~ 1011 % of the net terrestrial C sink estimated from the watershed. This study supports the view that 817 African rivers are strong emitters of CO2 to the atmosphere, mostly sustained by wetland inputs, and this must be better 818 considered in global models.

819 Data availability

\$20	If accepted the database will be publicly available at zeonodo.orgThe dataset of C and ancillary parameters is available from
\$21	(Moustanha et al. (2021) at https://doi.org/10.5281/zenodo.5625039

822 Competing interests

823 The authors declare they have no conflict of interest

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Streams/Rivers	Mengong	Mengong	Awout	So'o	Nyong	Nyong
Stations	Source	Outlet	Messam	Pont So'o	Mbalmayo	Olama
Latitude	3.17°N	3.17°N	3.28°N	3.32°N	3.52°N	3.43°N
Longitude	11.83°E	11.83°E	11.78°E	11.48° E	11.5°E	11.28°E
Gauging station	No	Yes	Yes	Yes	Yes	Yes
Altitude (m)	680	669	647	634	634	628
Catchment area (km ²)	0.48	0.6	206	3 070	13 555	18 510
Wetlands (%)		20	5.7	5.3	4.6	4.4
Catchment slope (‰)	1.3	1.3	1.2	1.1	0.16	0.15
Stream order	groundwater	1	3	4	5	6
Averaged-annual river flow in 2016 (m $$ s $$)	0.00544 ^a	0.009±0.002	3.9±4.8	35.6±40.6	146±112	195±160
-1 Averaged-annual rainfall (mm yr)	1 986					

Table 1: <u>gG</u>eographical and hydrological <u>sub</u>-catchments characteristics. ^a represents Q_{hill} (Fig. 3) and it is estimated from <u>Eequation</u>-1.

Parameters	Т	рН	Specific conductivity	Oxygen saturation	TSM
Units	°C	Unitless	$\mu S \text{ cm}^{-1}$	%	mg L ⁻¹
Mengong wetland ^a	24.2±1.4	5.5±0.6			
	[21.9-26.3]	[4.9-6.6]			
Mengong source	23.2±0.1	5.0±0.1	15.1±0.8	50±8	
	[23~23.6]	[4.6~5.3]	[14.1~17.4]	[38~68]	
Mengong outlet (order 1)	22.9±0.7	5.6±0.2	16.7±4.5	52±7	5.3±2.1
	[21.9~24.4]	[5.3~6.0]	[5.2~24.7]	[39~62]	[1.8~11.1]
Awout (order 3)	22.5±0.5	5.6±0.2	21.6±5.5	47±9	10.4±6.1
	[22~23.5]	[5.0~6.1]	[16.5~40.3]	[37~67]	[4.9~27.5]
So'o (order 4)	23.9±1.3	6.1±0.2	23.4±5.0	57±6	14.4±3.8
	[22.4~27.6]	[5.7~6.6]	[18.3~35]	[46~69]	[8.2~22.9]
Nyong (Mbalamayo, order 5)	26.1±1.3	6.2±0.3	36.6±19	40±20	8.9±2.0
	[24.3~29.0]	[5.5~6.9]	[19.6~86.3]	[13~81]	[4.3~12.0]
Nyong (Olama, order 6)	25.7±1.4	6.2±0.3	31.4±12.8	43±12	9.7±3.2
	[24.1~28.8]	[5.5~6.6]	[20.1~69.3]	[24~67]	[3.7~14.8]

Table 2: Spatial distribution of physicochemical parameters (yearly average±standard deviation) in waters of the Nyong watershed during the sampling year 2016. The range is shown in square brackets. ^a was measured in the topsoil solution of the Mengong wetland at 0.4 m depth by Nkoue-Ndondo et al. (2020).

Parameters	pCO ₂	TA	DIC	DOC	POC	POC
Units	ppmv	µmol L ⁻¹	μmol L ⁻¹	μmol L ⁻¹	%	μmol L ⁻¹
Mengong wetland	36 840±23 190ª	122±46 ^a	1 430±900ª	1 420±750 ^b		
	[3 900-84 240]	[50-216]	[150-3 270]	[1 250 - 2 920]		
Mengong source	78 800±40 110	53±26	2 940±1 485	83		
	[12 700~209 000]	[15~138]	[500~7 560]			
Mengong outlet (order 1)	15 600±8 900	90± 36	670±360	1 925±970	23±5	101±44
	[3 980~41 000]	[20~156]	[170~1 710]	[1 090~4 150]	[14~26]	[14~213]
Awout (order 3)	15 400±7 300	67±39	670±315	3 200±1 840	16±3	130±50
	[5 760~26 710]	[11~166]	[260~1 170]	[2 000~7 550]	[11~21]	[72~243]
So'o (order 4)	12 700±5 100	74±34	670±260	2 170±980	18±4	210±60
	[4 900~23 200]	[10~145]	[300~1 320]	[1 100~5 320]	[12~29]	[125~360]
Nyong (Mbalamayo, order 5)	11 800±5 100	123±63	720±270	2 000±860	20±3	150±40
	[3 620~22 460]	[20~230]	[220~1 200]	[1 020~5 300]	[16~26]	[62~220]
Nyong (Olama, order 6)	11 000±5 550	134±70	640±330	1 860±440	18±2	150±50
	[3 000~21 700]	[10~265]	[170~1 240]	[1 100~2 880]	[15~23]	[55~235]

Table 3: Spatial distribution of C variables (yearly average±standard deviation) in waters of the Nyong watershed during the sampling year 2016. The range is shown in square brackets. ^a measured in the topsoil solution of the Mengong wetland at 0.4 m depth by Nkoue-Ndondo et al. (2020). ^b measured in the topsoil solution of the Mengong wetland at 0.4 m depth by Braun et et al. (2005).

	Mengong	Awout	So'o	Nyong at Mbalmayo	Nyong at Olama
	Discharge	Discharge	Discharge	Discharge	Discharge
Oxygen saturation	0.19	-0.32	-0.54	-0.85	-0.85
pH	-0.60	-0.17	-0.53	-0.76	-0.81
Specific conductivity	-0.02	0.11	-0.44	-0.63	-0.70
ТА	0.21	0.21	-0.39	-0.41	-0.37
pCO ₂	0.05	0.32	-0.03	0.46	0.38
DOC	0.32	-0.15	0.14	-0.14	0.28
TSM	-0.28	-0.56	-0.32	0.33	0.56
POC%	-0.50	-0.22	0.36	0.62	0.38
POC	-0.43	-0.62	0.27	0.46	0.70

Table 4: Correlations with p<0.05 (Pearson's eCorrelation test) between C or ancillary parameters and the discharge in the different stream orders. The Pearson's Ceorrelation coefficient is indicated and -significant correlations (p<0.05) are in bold.

Table 5: DOC, DIC and POC budgets in the first-order Mengong stream (Eqs. 8-10). Fluxes are in $\underline{\mathsf{fMgC}}$ yr⁻¹ and are described in details in the section 2.5. Briefly, F_{GW} is the quantity of dissolved carbon leached from non-flooded forest groundwater to the Mengong stream (Eq. 11), F_{WL} is the quantity of carbon leached from the Mengong wetland to the Mengong stream (Eqs. 13-14), F_D is the quantity of CO₂ degassed from the Mengong stream to the <u>overlying</u>-atmosphere, F_{RH} is the heterotrophic respiration in the Mengong stream, and F_{OUT} is the quantity of carbon hydrologically exported at the outlet of the Mengong <u>stream</u>eatchment.

	DOC _{budget}	$\operatorname{DIC}_{\operatorname{budget}}$	POC _{budget}
$F_{\rm GW}$	0.17±0.02	6.05 ± 2.98	
F _{WL}	1.80±0.95	1.82 ± 1.13	0.34±0.14
F _D		5.51 ± 2.30	
F _{RH}	0.32±0.30	0.32±0.30	
Fout	6.41±3.23	2.23 ± 1.20	0.34±0.14
Imbalance (inputs-outputs)	-4.76	0.45	0

Table 6: At the Nyong watershed scale, yearly averages with standard deviations (based on averaging monthly values in each stream order) of CO₂ degassing rates (F_{degas} in $GC-CO_2$ m⁻² yr⁻¹), k_{600} (m d⁻¹), water surface area (m²), and integrated CO₂ degassing flux (F_{degas} in $GgC-CO_2$ yr⁻¹), estimated in the different stream orders. Range (based on monthly values) is shown between brackets. In addition, rates of heterotrophic respiration (gC-CO₂ m⁻² yr⁻¹) in the stream orders 1 and 5 are indicated. ^a considering an additional benthic respiration in tropical rivers of 222 gC m⁻² yr⁻¹ by Cardoso et al. (2014). ^b calculated as the sum of the integrated CO₂ degassing flux in each stream order, this represents the CO₂ degassing flux from the entire river network (GgC-CO₂ yr⁻¹). ^c this represents the CO₂ degassing flux from the entire river network weighed by the surface area of the Nyong watershed (MgC-CO₂ km⁻² yr⁻¹). Note that measurements in second-order streams were extrapolated (see method).

Stream order	Respiration rates	k_{600}	F _{degas}	Water surface area	F _{degas}
	$gC\text{-}CO_2\ m^{\text{-}2}\ yr^{\text{-}1}$	m d ⁻¹	$gC-CO_2 m^{-2} yr^{-1}$	m²	GgC-CO ₂ yr ⁻¹
1	63.9±49.2 (286.4±227.9ª)	2.21±0.08	5 344±2 773	6 662±3340	42.5±26.7
		[2.04-2.31]	[2 436-12 089]	[337-11 5950]	[14.2-100.3]
		2.58±0.19	6 338±2 499	16 562±7 481	126.5±60.2
2		[2.18-2.83]	[2 936-10 739]	[2 794-27 216]	[49.8-220.9]
		2.94±0.29	6910±2661	17 149±7 361	137.0±57.0
3		[2.46-3.33]	[3 155-10 800]	[5 728-27 538]	[61.4-228.7]
		3.00±0.31	5 280±2 317	20 249±8 376	114.4±79.9
4		[2.55-3.45]	[363-9997]	[8117-34 143]	[3.9-318.7]
		2.35±0.10	3 706±1 540	23 180±9 389	90.3±58.0
5	533.4±534.3 (755.9±333.0ª)	[2.20-2.51]	[1 114-6 230]	[9 656-40 207]	[19.8-222.2]
		2.48±0.13	3 745±1 711	36 977±15 029	141.3±92.0
6		[2.28-2.71]	[957-6 354]	[15 741-65 121]	[35.9-317.8]
					651.9±160.6 ^b (<i>23.4</i> ±5.8 ^c)

Table 7: At the Nyong watershed scale, averages of monthly hydrological export of C to the ocean (Focean)
and of monthly CO ₂ degassing to the atmosphere (F _{degas}). ^a the net C sink estimated by Brunet et al. (2009)
for the entire Nyong watershed is also indicated.

	Focean	F _{degas}	F _{ocean}	F _{degas}	Watershed net C sink ^a
	GgC yr ⁻¹	GgC-CO ₂ yr ⁻¹	MgC km ⁻² yr ⁻¹	MgC-CO ₂ km ⁻² yr ⁻¹	MgC km ⁻² yr ⁻¹
DOC	134.0±99.8		7.2±5.		
DIC	45.5±42.4	651.9±160.6	2.4±2.3	23.4±5.8	
POC	11.9±9.9		0.6±0.5		
Total	191.4±108.9	651.9±160.6	10.3±5.8	23.4±5.8	300



Figure 1: Map of the Nyong watershed showing the river network, the wetland extent from Gumbricht et al_{τ} (2017) and the location of the sampling stations and some cities. Note, the Nyong River is displayed bolder than the other rivers. The background map is from Google Satellite®.

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Figure 2: (a-b) River discharges of the different gauging stations during the sampling year 2016, associated with rainfall measured at the Mengong catchment. (c) The box plots represent the variability of monthly Nyong River discharges from 1998 to 2020 and extreme box plots values represent minimum and maximum monthly discharges during the same period; whereas the green lines represent the average monthly discharges in 2016, and the red dashed line represents the yearly average discharge of 194.5 $m^3 s^{-1}$ for the 1998 to 2020 period (very close to the yearly average discharge of 195 $m^3 s^{-1}$ measured in 2016). (d) Yearly rainfall over the 1998-2020 period (1600±290 mm), and the red bar represents the yearly rainfall during the sampling year 2016. Hydrologic and rainfall data are from Audry et al. (2021).





Figure 3: (a) Map of the first-order Mengong catchment showing the wetland area and the hydrological fluxes that are partitioned between the main perennial source (Q_{hill} , blue arrows) of the non-flooded forest groundwater, specific seepage points all around the hillside/wetland boundaries (Q_{base} , orange arrows) of the non-flooded forest groundwater, and the discharge at the stream outlet (Q_{st} , white arrow). Note, A_{hill} is the surface area drained by the non-flooded forest groundwater. (b) Cross section of the dashed line from the map (a), showing the lithology of the hillside lateritic system and the hydromorphic wetland system, the recharge of the hillside system (R_{hill}); Q_{base} and Q_{hill} are also indicated. (c) Hydrological functioning of the first-order Mengong catchment. Note, $Q_{WL/ST}$ represents the groundwater flow exchanged between the wetland and the stream and OF_{WL} is the overland flow on the surface of the wetland. (d) Characteristic soil profiles at piezometers 1, 2 and 3, in which water table level. The figure 3 was adapted from Braun et al. (2005) and (2012) and from Maréchal et al. (2011).



Figure 4: In the first-order Mengong catchment temporal variations in the first-order Mengong catchment of rainfall, water-table level in piezometer 1 and 2 (see figure 3b) relative to sea level (elevation of the soil surface at piezometers 1 and 2 relative to sea level is also indicated by the horizontal lines); and pCO_2 , TA and ancillary parameters (oxygen saturation as O, pH, conductivity as specific conductivity) in non-flooded forest groundwater (measured at the perennial source). The temporal variations are separated into the four seasons that occurs in the Nyong watershed that are LDS as long dry season, SRS as short rainy season SDS as short dry season and LRS as long rainy season. Note, groundwater table level was retrieved from (Nkoue-ndondo et al.; (2020).

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Figure 5: temporal variations of river discharge, carbon (pCO₂, TA, DOC, POC) and ancillary parameters (<u>oxygen saturation as O₂</u>, pH, <u>conductivity as</u> specific conductivity, TSM) in surface waters of the Nyong watershed. The temporal variations are separated into the four seasons that occurs in the Nyong watershed that are LDS as long dry season, SRS as short rainy season SDS as short dry season and LRS as long rainy season.

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Figure 6: spatial variations of carbon parameters (pCO₂, TA, DOC, POC) and ancillary parameters (<u>oxygen</u> <u>saturation</u> Θ_2 , pH, specific conductivity, TSM) across non-flooded forest groundwater (GW) and streams orders 1, 3, 4, 5 and 6 in the Nyong watershed. <u>Note that the hashed and orange boxplots are for non-flooded</u> <u>forest groundwater and first-order stream, respectively. The boxplots represent the minimum, the first guartile, the median, the third quartile and the maximum.</u>



Figure 7: mass balance of C in the first_order Mengong catchment. All fluxes are in tMgC yr⁻¹, and in tMgC km⁻² yr⁻¹ when between brackets (weighed by the surface area of 0.48 km² drained by non-flooded forest groundwater for the net forest C sink, F_{GW} , F_{GW-bis} and F_{D-GW} , by the wetland surface area of 0.12 km² for the net wetland C sink, F_{WT} and F_{D-W} , and by the Mengong catchment area of 0.6 km² for F_{OUT} , F_D and F_{RH}), and they are associated with their corresponding equations as described in details in the section 2.5. Briefly, F_{GW} is the quantity of dissolved carbon leached from non-flooded forest groundwater to the Mengong stream (Eq. 11), F_{GW-bis} is the quantity of dissolved carbon leached from non-flooded forest groundwater to the Mengong stream (Eq. 13), F_{D} is the quantity of C degassed from the Mengong stream to the overlying atmosphere, F_{RH} is the heterotrophic respiration in the Mengong stream. In addition, net local-forest C sink of mature forest of the Mengong catchment estimated by Brunet et al. (2007) and Jones and Humphries (2002) are both indicated.



Figure 8: Monthly C fluxes at the Nyong watershed scale described in the section 2.4. For F_{occan} , the black dashed lines represent the yearly average of the different monthly C fluxes. For F_{decas} , the green dashed line is obtained by summing yearly integrated CO₂ deassing in each stream order, as in Table 6, which represents 651.9±160.6 GgC yr⁻¹. For F_{degas} , the black dashed line represents the yearly average of the different monthly CO₂ degassing fluxes from the entire river network, as in Figure 8, which represents 578.9±157.9 GgC yr⁻¹. The figures are separated into the four seasons that occurs in the Nyong watershed that are LDS as long dry season, SRS as short rainy season SDS as short dry season and LRS as long rainy season.

Figure S1: Monthly variations of k₆₀₀ (m d⁻¹) and water surface area (km²) in each stream order across seasons. The temporal variations are separated into the four seasons that occurs in the Nyong watershed that are LDS as long dry season, SRS as short rainy season SDS as short dry season and LRS as long rainy season.