Spruce bark beetles (*Ips typographus*) cause up to 700 times higher bark BVOC emission rates <u>compareds to healthy</u> from Norway spruce (*Picea abies*)

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10 Abstract

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Emissions of biogenic volatile organic compound (BVOC) from the bark of Norway spruce (Picea abies) trees can be affected by stress, such as infestation of spruce bark beetles (*Ips typographus*). We studied the The in emission rates betweenfrom healthy spruce bark and infested Norway spruce bark was studied, as well as , the influence of time since spruce bark beetle infestation started and the difference in emission rates from bark beetle drilled entry holes and exit holesentry and exit holes. Bark chamber measurements on both healthy trees and infested treeshealthy and infested trees were performed during the summer of 2019 at Hyltemossa and Norunda research stationat two sites in Sweden. To consider the seasonal pattern of the spruce bark beetle, twe divided the emission rates from infested trees were divided into two seasons, an early season dominated by entry holes and a late season with mainly exit holes. The Our findings results showed a significant difference in emission rates from healthy and infested trees, independent of season. The seasonal average standardized BVOC emission rates of from healthy trees was $32\frac{1.89}{1.89} \pm 52\frac{1.67}{1.89} \mu \text{g m}^{-2} \text{h}^{-1}$ (mean \pm standard deviation), while the average standardized BVOC emission rates offrom infested trees were 6700385 µg m⁻² h⁻¹ and 2000102 µg m⁻² h⁻¹ during early and late season respectively. AWe also found an exponentially decreasing relationship was found with BVOC emission rates decreased exponentially with time and time since infestation started where the emission rates and indicated induced emission rates for about one year after which the emission rates were similar to those from reached the same level as the constitutive BVOC emission rates from healthy bark after around one year. When comparing bark monoterpene BVOC emission rates with emission rates from needles, we found that the constitutive constitutive needle emission rates were found to be were 11 times higher than the healthy constitutive bark emissions. However, the emission rates from infested Norway spruce tree bark were instead 6 to 20 times higher than the constitutive needle emissions, causing substantial increases in the total tree BVOC emission rate. (550 % to 1900 % increase). This study adds evidence that spruce bark beetle induced bark BVOC emissions are higher than previously thought and highlights the need for further research with more samples more frequently throughout the season to fully understand the impact, which is required to quantify spruce bark beetle infestations impacts on the atmospheric chemistry and climate change.

35 1 Introduction

In Europe, forest damage caused by outbreaks of the European spruce bark beetle (*Ips typographus*) is the third largest disturbance after storm-felling and forest fires (Jönsson et al., 2012; Schelhaas et al., 2003). In Sweden, the drought in the summer of 2018 led to increased bark beetle outbreaks which, in 2020_7 were estimated to affect

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about 8 million m³ (standing timber volume) Norway spruce (*Picea abies*) forest (Wulff and Roberge, 2020). This is the largest stock of forest volume killed by spruce bark beetles recorded in a single year in Sweden; in the period of 1990-2010, around 150₂-000 m³ forest in southern Sweden was damaged on average per year (Wulff and Roberge, 2020). Climate change amplifies the risk of bark beetle outbreaks as the elevated risk of storm felling and drought favor the bark beetles with easier access to weakened trees (Jönsson et al., 2012). Higher temperatures and a longer growing season can also lead to an additional generation of spruce bark beetles per year (Jakoby et al., 2019; Jönsson et al., 2012). A larger bark beetle population, triggered by weather extremes, is associated with an increased risk of attacks on healthy spruce trees; with outbreaks leading to extensive damage to the forests (Jakoby et al., 2019; Seidl et al., 2014).

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Biogenic volatile organic compounds (BVOCs) emitted from trees function for example as a defense system against heat and oxidative stress (Loreto and Schnitzler, 2010). They are highly volatile and chemically reactive and can react directly with oxidizing species or act as membrane stabilizers (Brilli et al., 2009; Kleist et al., 2012; Sharkey et al., 2001). The efficiency of BVOCs to form oxidation products depends on the specific BVOC's molecular structure (Bonn and Moortgat, 2002; Roldin et al., 2019; Thomsen et al., 2021). As BVOCs are emitted, they can enhance chemical reactions which in turn can lead to increased tropospheric ozone concentration, or BVOCs can get oxidized and may foster the formation of secondary organic aerosol (SOA; Kulmala et al., 2003). Stress-induced BVOC emissions alter the oxidation capacity as some BVOC species are more efficient to act as secondary organic aerosol (SOA)SOA precursors and foster particle growth (Roldin et al., 2019; Thomsen et al., 2021). Boreal forests experiencing abiotic or biotic stress due to large-scale forest disturbances might thus increase the production of BVOC species highly efficient as precursors of SOA to the atmosphere. As BVOCs are emitted, they can enhance chemical reactions that in turn can lead to increased tropospheric ozone concentration or be oxidized and foster the formation of SOA_(KulKulmalamala et al., 2003). This results in high uncertainties regarding the contribution to either a negative feedback loop (cloud formation and radiation scattering (Paasonen et al., 2013)) or a positive feedback loop (increased tropospheric ozone) for the climate (Arneth et al., 2010; Jia et al., 2019). The impact of aerosol formation including the BVOC SOA feedback still remains the largest uncertainty in our understanding of their radiative forcing (IPCC, 2014; Jia et al., 2019). Emission of BVOCs due to plant stress further increase the uncertainties of their impact in a changing climate.

Increased BVOC concentrations in plant tissue can also fight off predators (Laothawornkitkul et al., 2009; Li et al., 2019; Rieksta et al., 2020), and conifer trees use BVOCs as a defense mechanism against spruce bark beetles (Celedon and Bohlmann, 2019; Krokene, 2015; Raffa and Berryman, 1982). The parental bark beetles attack spruce Norway spruce trees by drilling entry holes into the bark and form eggs galleries in the phloem. After about eight weeks, the new generation starts to leave the tree by boringdrilling exit holes (Öhrn et al., 2014). To prevent a successful attack, the spruceNorway spruce increase the resin flow which submerges the parental bark beetles and egg galleries, potentially killing beetles or pushing them out of the entry hole (Raffa, 1991). The resin serves as a storage pool containing BVOCs that volatilize when the resin is flowing out of the tree, making the resin harden and close the wound. BVOCs are emitted constitutively from the trunk of the spruceNorway spruce, but when the emissions are induced as a stress-response, they have been shown to be toxic to spruce bark beetles (Celedon and Bohlmann, 2019; Krokene, 2015), especially certain compounds like myrcene and egg-terpinene (Everaerts et al., 1988). Studies on conifers attacked by bark beetles found evidence of increased monoterpene (MT) content at the attacked location (Amin et al., 2013; Ghimire et al., 2016; Zhao et al., 2011b). Quand occurrence of the oxygenated MT eucalyptol hase been found to indicate induced defense and higher survival rates from

Norway spruce attacked by spruce bark beetles (Schiebe et al., 2012). When c Comparing BVOC emission sources from different parts on conifer trees, trunk emissions are suggested to potentially contribute a lot more to the whole tree emissions than previously thought, even when not attacked by bark beetles (Greenberg et al., 2012).

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The defense mechanism of Norway spruce is poorly understood There is still a lot to learn about the defense mechanism of Norway spruce, only as few studies have analyzed the induced BVOC emission from the trunk following an attack of the European spruce bark beetle (Ghimire et al., 2016; Zhao et al., 2011b). The aim of this study was to (i) investigatestudy the BVOC emissions from Norway spruce trunks and the impact of spruce bark beetles and to (ii) studyinvestigate the relation connections-betweenof BVOC emission rates, number of bark beetle drilled holes and time. The aim was also to (iii) put our study in a broader perspective by compare and connectonnect ing the spruce bark beetle induced BVOC emission rates to needle emissions and other stresses like heat stress. Based on previous findings, we formulated three hypotheses were formulated: (H1). Our first hypothesis was that infested trees have higher bark emission rates than healthy trees, and that infestation changes the emission blend, (H2). Our second hypothesis was that BVOC emission rates are would be highest at the start of infestation, and decrease over time in response to declining tree vitality and eventual death of the tree and (H3)-We also wanted to investigate the connections of emission rates and number and type of bark beetle holes, where our third hypothesis was that the type of the bark beetle drilled holes matter for influence the BVOC emission rates where there is a relationship between the number of entry holes and emission rates rather than a highthe total amount number of holes. The We reasoning behind H3 wased that after a successful infestation, the number of holes increase after some weeks when the new generation leave the tree through exit holes, but the emission rates would decrease as the tree is dying due to bark damage and blue stain fungi preventing transport of water and nutrients. In a sub-study, two treestree individuals with different initial health status were selected and followed throughout the growing season by repeating the measurements during athe successful attack and infestation of bark beetles. From The aim (iv) of this sub-study we were hopingwas to see if the difference in initial health status of the trees would result in different emission rates and emission blends, and toalso analyze how the individual emission blend changed over time after a successful infestation.

2 Methods

2.1 Site description

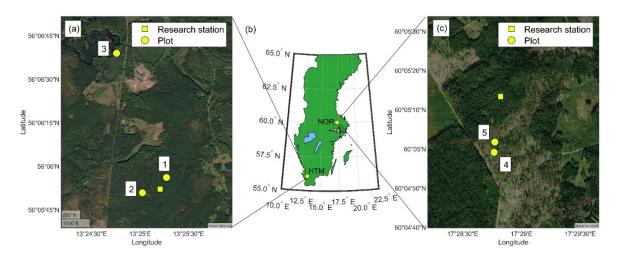
SixFive measurement campaigns were carried out from May to August 2019 (Table 1), where five campaigns were located at the ICOS (Integrated Carbon Observation System, ICOS-Sweden.se) research station in Hyltemossa (HTM, 56°06′N, 13°25′E; Fig. 1b) and one-additional campaign at the ICOS research station in Norunda (NOR, 60°05′N, 17°29′E; Fig. 1b). The forest in HTM is dominated (>97% of the species composition) by Norway spruce (*Picea abies*) with a small fraction (<3%) of Scots pine (*Pinus sylvestris*) and deciduous trees. The understory vegetation is sparse, containing mostly mosses (Heliasz et al., n.d.). The forest in NOR is dominated by Norway spruce (54%) and Scots pine (37%) with a small fraction (9%) of deciduous trees and an understory vegetation with shrubs of mostly blueberries, cranberries, mosses and flowers (Mölder et al., n.d.). Both facilities are located inside managed forests, but the age and height of the trees differ. In HTM the trees are around 40 years old with an average height of 19 m in 2019 and NOR has a forest stand of mixed ages around 60-80 years and up to 110 years with a height of around 25 m for the dominating trees in 2019 (Heliasz et al., 2021; Mölder et al., 2021).

Table 1. Time table of the six campaigns conducted during 2019 at the sites Hyltemossa (HATM) and Norunda (NOR). Indicated is also the number of collected samples during each campaign at each plot and the status of the Norway spruce when measurements were collected (healthy or infested).

ţ -			Number of collected samples										
Month	<u>Date</u>	<u>Site</u>	Plo	Plot 1		Plot 2		Plot 3		ot 4	Plot 5		<u>Total</u>
			Healthy	Infested	Healthy	Infested	Healthy	Infested	Healthy	Infested	Healthy	Infested	=
May	4th-6th	HTM	<u>12</u>	<u>3</u>	<u>12</u>	Ξ	<u>12</u>	Ξ					<u>39</u>
<u>June</u>	4th-6th & 13th	HTM	9	1	<u>12</u>	Ξ	<u>6</u>	<u>6</u>					<u>34</u>
<u>July</u>	2nd-4th	HTM	<u>12</u>	Ξ	<u>12</u>	Ξ	<u>6</u>	<u>6</u>		-	Ξ.		<u>36</u>
<u>July-</u> <u>August</u>	<u>30th-1st</u>	HTM	<u>12</u>	=	<u>12</u>	=	<u>6</u>	<u>6</u>					<u>36</u>
August	21st-22nd	<u>NOR</u>			=				Ξ	<u>9</u>	Ξ	<u>9</u>	<u>18</u>
August	26th-27th	HTM	<u>12</u>	2	<u>12</u>	Ξ		_			_		<u>24</u>

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130 Figure 1: The location of the study sites in Sweden (b), with Hyltemossa (HTM; a) displayed to the leftright, the location in Sweden in the middle (b) and Norunda (NOR; c) displayed to the rightleft. Measurement plots (yellow circle) at HTM (1-3) and NOR (4-5) are shown in the site-specific maps and their location relative to the ICOS station (yellow square). Healthy Norway spruce trees were measured in plot 1-3 and infested spruce trees were measured in plot 1, 3, 4 & 5. The sub-study was conducted in plot 3. The fFigure is created in MATLAB and Mapping Toolbox release 2021a (The MathWorks, Inc., Natick, MA, USA).

Three plots in HTM and two plots in NOR were selected for the study (Fig. 1a,c), and two plots in NOR (Fig. 1e). Two plots in HTM were located inside the Norway spruce plantation used by ICOS, while the third plot was located around 1.6 km north of the ICOS station in an older (about 100 years) forest stand. In NOR the locations were chosen based on availability of bark beetle-affected trees inside the forest plantation. Four Norway spruce trees were selected at each plot in HTM, and three trees at each plot in NOR. A total of 18 trees were measured, whereof 12 were measured repeatedly during the growing season in HTM. For all plots in HTM healthy Healthy trees were selected where the tree health was determined by visual examination in close contact with the forest manager employed by the forest owner, at Gustafsborgs Säteri AB, in May 2019. Trees potentiallythat could have been stressed by forestry machinery or pests were not selected for the study. The infested trees were selected based on the signs of spruceif bark beetle infestation. In addition, Ttwo trees seemingly stressed from Signs of late bark beetle infestation from the previous year, in 2018, were found on two trees, were chosen, one at plot 1 only measured in the early season and one at plot 3 which later got infested again and used for the comparison of healthy

and stressed trees when infested. These trees were selected for the study to analyze long-term infestation effects.

Only infested trees were selected in NOR, Aasn active spruce bark beetle outbreaks wasere occurring in NOR during 2019 and only infested trees were selected at that site.

The sub-study was conducted at plot 3 in HTM where two trees were selected, one already stressed from-previous infestation in the previous year and one healthy (Fig. 1a).

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To ensure bark beetle infestation. To enable the measurements of healthy trees which later got infested, a The healthy a tree at plot 3 in HTM (Fig. 1a) was baited using a bark beetle slit trap with pheromones to facilitate bark beetle infestation. The trap was installed between the 1st and 2st and 2st and June (Table 1). One bage of biological attractant was used containing the pheromone sused in the trap were a combination of 2,3,2 Methylbutenol, eis Verbenol and Ipsdienol (Typosan P306, Plantskydd AB, Ljungbyhed, Sweden PS), and one standard bag was inserted in the slit trap. Two trees were successfully infested and measured repeatedly to track the emission pattern after infestation. Both of the trees were successfully infested and measurements were repeated throughout the season to see the effects of infestation over time.

The weather during the measurement periods varied from cold and humid to warm and dry conditions. The average temperature during the growing season (May-August) was 14.6 °C (± 4.6 °C) in HTM and 14.2 °C (± 4.2 °C) in NOR with the sum of the precipitation over the growing season being 168 mm in HTM and 151 mm in NOR (Fig. 2). The daily average temperatures during the measurement periods ranged from 5 °C-C tto 22 °C for both sites, with a daily total rainfall up to 3 mm (Fig. 2). In HTM, it was coldest (1 °C to 11 °C) during the measurement campaign in May and warmest (12 °C to -28 °C) during the measurement campaign 2-in June-and coldest (1 11 °C) during the 1st-campaign in May.

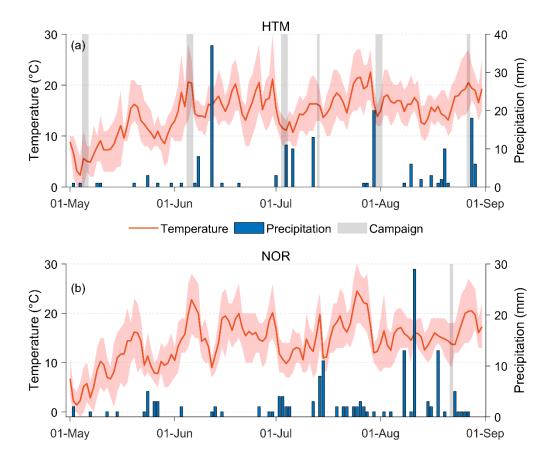


Figure 2: The total daily precipitation (blue bars) and daily average temperature (red line) with daily minimum and maximum temperature (red shade) and total daily precipitation (blue bars) with daily minimum and maximum temperature (red shade) for the study sites in (a) Hyltemossa and (b) Norunda. The times for the campaigns are marked in grey. The weather data was acquired from the Data: ICOS research stations at the study sites (Heliasz, 2020; Mölder, 2021).

2.2 Experimental design

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The bark emissions from the trees were measured using a tree trunk chamber connected with PTFE tubing (Teflon, Swagelok, Solon, OH, USA) to a pump box system consisting of a diaphragm pump (1420 VPD, Gardner Denver Thomas GmbH, Memmingen, Germany) and a flow meter (GFM mass flow meter, Aalborg Instruments & Controls Inc., USA). The pump box system was used to provide purge air with a flowrate of 0.7 lpm (liters per minute) to the trunk chamber with a volume of 0.6-0.9 Llitres (Fig. 3). Adsorbent tubes were used to take air samples from the chamber. A hydrocarbon-BVOC filtertrap_(Hydrocarbon trap, Alltech, Associates Inc., USA) containing activated carbon and MnO2-coated copper nets was mounted between the pump box and chamber to scrub the purge air of remove VOCs BVOCs and O3 to ensure that only clean air entered the chamber. The chamber consisted of a metal frame and a flexible polyethylene foam base to fit the tree trunk and was fastened with straps around the tree trunk. The inside of the chamber had been carefully wrapped with pre-conditioned (oven-cleaned 40°C, 3 hours) polyamid bags (Toppits, Cofresco Frischhalteprodukte GmbH, Germany) to avoid contamination with BVOC from the chamber foam base. During measurements, the chamber was closed with Aa metal lid with in- and outgoing PTFE-tubinglines (\(\tilde{\pi} \) 6.35 mm^{1/4}") for purge air and sample collection was used to close the chamber during the measurement. for purge and sample flow. Temperature of the air Air temperature within the chamber was measured with a temperature probe (HI 145, Hanna Instruments, RI, USA) during samplingBVOC collection, and the bark surface temperature was measured with an infrared thermometer (IRT260, Biltema, Sweden) was used to take bark surface temperature_before and after each sample from bark inside the chamber before and after each BVOC collection.

For each campaign, trees from one plot were sampled per day with BVOC collection, typically starting aroundt 08:00 (LT) and ending aroundt 19:00 (LT) alternating sampling between the trees. The chamber bases were secured in place in the North or East orientation of the trunk onto the tree trunks every morning. The chambers were and were left open during the day to avoid built up concentrations of BVOCs inside the chambers. Prior to the sampling the The bark temperature was measured at four different points inside the base prior to the sampling after which the lid was fastened and the chamber was flushed for 15 minutes before sampling started. Air temperature inside the chamber was measured at the start and at the end of the BVOC collection to note potential temperature differences during the sample period. After sampling, the lid was removed and the bark temperature was measured again.

The start of the infestation was determined by the beetles' swarming time in relation to when the tree infestation was detected. For plot 3 in HTM₇ the start of the infestation was seen during the campaign in June (Table 1)witnessed on site. Ffor the other plots the swarming time was retrieved using data from Skogsstyrelsen Statistical Database (Skogsstyrelsen, n.d.) taken at the plots closest to the measurement sites (Supplementary material, Table S1, Table S2). which showed a A late swarm in HTM was detected around week 25-28 for 2018 (Fig. S1), and a main swarm in NOR around week 20-21 for 2019- (Fig. S2)(Skogsstyrelsen, n.d.), S1 in the Supplement).

The bark inside the chambers was controlled visually before each measurement to count the number of bark beetle holes and to assess potential lichen and algal cover as seen in Table 1. By looking at bark photosgraphs, the holes were later separated into entry or exit holes for the infested trees (Fig. 4). The separation depended on the characteristics of the hole where entry holes were determined to have more resin bleed compared to exit holes, which also had a rounder shape as seen in Fig. 4. Table 1The number of holes counted inside the chamber area is listed in Table 2 along with an extrapolation of the counted holes from the chamber area to square meter bark area. Entry holes were found for measurements taken up to 100 days after infestation and exit holes were predominantly found for measurements taken after 100 days of infestation, with the latest measurement occurring 350 days after estimated infestation start. As the two hole types were consistently occurring before or after 100 days since infestation start, the measurements with mainly entry holes is referred to as the early season and the measurements with mainly exit holes as the late season. describes the number of visual holes inside the chamber and the extrapolated number per square meter of bark area.

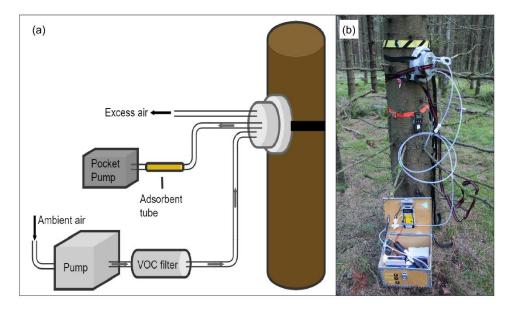
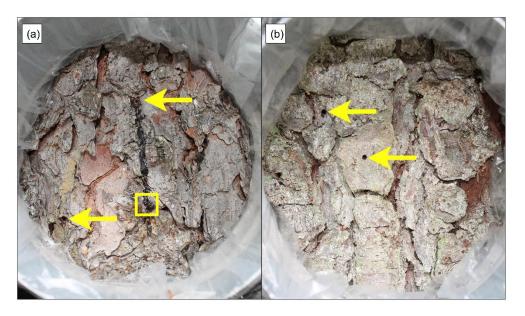


Figure 3: The experimental setupehematics (a) and a field photo (b) contained and the tree trunk chamber mounted on a Norway spruce trunk. The chamber istree, attached connected to a pump box used to provide BVOC- and O₃-free purge air. The BVOC samples were collected with adsorbent tubes connected to the chamber by extracting the air from the chamber using a pocket pump. A photograph of how the setup looked in the field is displayed to the right (b).



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Figure 4: Examples of infested Norway spruce trees with (a) entry-holes and (b) exit holes, where tThe arrows points towards indicate examples of bark beetle drilled holes and the square box frames shows a bark beetle. There are more holes in the pictures than indicated by the arrows. More holes can be found in the picture than pointed to.

Table 1Table 2. The infested Norway spruce trees indicated by their tree ID, and their location by their site, and plot and their respective and the number of holes counted inside the respective chamber at the given date for each tree. The counted holes were upscaled extrapolated to holes per square meter of bark surface and the majority of the hole type was determined to either mostly entry holes or mostly exitntry holes. The Norway spruce tree with the ID S1S1 was infested during the in-late season 2018 and had thus a already a majority of exit holes early in 2019.

Tree ID	Site	<u>Plot</u>	Date	Number of holes inside chamber	Upscaled to holes per m ²	Hole tType majority
S1S1	<u>HTM</u>	<u>1</u>	2019-05-04	12	1062	exit
S1S1	<u>HTM</u>	<u>1</u>	2019-06-05	8	708	exit
S3S2	<u>HTM</u>	<u>3</u>	2019-06-04	4	354	entry

S3S3	<u>HTM</u>	<u>3</u>	2019-06-04	5	442	entry
S3S2	<u>HTM</u>	<u>3</u>	2019-07-03	3	286	entry
S3S3	<u>HTM</u>	<u>3</u>	2019-07-03	5	465	entry
S3S2	<u>HTM</u>	<u>3</u>	2019-08-01	6	531	entry
S3S3	<u>HTM</u>	<u>3</u>	2019-08-01	15	1273	entry
S4S1	<u>NOR</u>	<u>4</u>	2019-08-21	4	354	exit
S4S2	<u>NOR</u>	<u>4</u>	2019-08-21	5	442	exit
S4S3	<u>NOR</u>	<u>4</u>	2019-08-21	5	442	exit
S5S1	<u>NOR</u>	<u>5</u>	2019-08-22	5	442	exit
S5S2	<u>NOR</u>	<u>5</u>	2019-08-22	4	340	exit
S5S3	<u>NOR</u>	<u>5</u>	2019-08-22	7	619	exit

2.3 BVOC sampling and analysis

A total of 187 samples were taken, where 147 samples were from healthy Norway spruce and 40 were from infested Norway spruce (Table 1). Stainless steel cartridges (Markes International Limited, Llantrisant, UK) packed with adsorbents Tenax TA (a porous organic polymer) and Carbograph 1TD (graphitized carbon black) were used to sample BVOCs. The BVOCs were sampled from the chambers using flow-controlled pocket pumps (Pocket Pump, SKC Ltd., Dorset, UK) which extracted by extracting the air through the steel cartridges at a flow rate of 200 ml min⁻¹ and a sampling time of ca. 30 minutes. The collected volume for each sample was between 5 to 6 Liter. Blank samples were collected from air entering the chamber inlet air twice per day, once once before the first sample and once once after the last sample of the day to capture from air entering the chamber twice a day to capture possible background contamination of the filtered purge air. Thir temperature was measured inside the chamber was measured at the start of the BVOC sampling and at the end of the BVOC collection to note potential temperature differences during the sample period. After sampling was finished When sampling was done, the chamber lid was removed and, the bark temperature was measured again following the same procedure as mentioned above. The same method was repeated throughout the day until all trees of that plot were measured three times.

After collecting the BVOC samples, the adsorbent cartridges were capped and stored in a refrigerator (at ~3 °C) before being analyzsed using a two-stage automated thermal desorption apparatus coupled to a gas-chromatograph mass-spectrometer. Desorption was done on a Turbomatrix ATD 650 (PerkinElmer, Waltham, MA, USA) by - Cartridges were initiallyprimary heating the cartridgested to 280 °C in a flow of purified helium (He, ALPHAGAZ 1, Air Liquide Gas AB, Sweden) for 10 minutes, in order for the VOCs-BVOCs to volatilize. After the primary desorption, VOCs-BVOCs were cryo-focused downstream on a Tenax TA cold trap maintained at -30 °C. The cold trap was flash-heated (40 °C sec-1) to 300 °C for 6 minutes to perform a second desorption. The volatilized VOCs-BVOCs were passed via a heated transfer line using He as carrier gas, to a gas chromatograph-mass spectrometry system (GC-MS, Shimadzu QP2010 Plus, Shimadzu Corporation, Japan). The BVOCs were separated using a BPX5 capillary column (50 m, I.D. 0.32 mm, film thickness 1.0 μm, Trajan Scientific, Australia) and the oven temperature was initially held at 40 °C for 1 minute, raised to 210 °C at a rate of 5 °C min-1 and further increased to 250 °C at a rate of 20 °C min-1 and lastly held for 2 minutes. Pure standard solutions of isoprene, egpinene, ββ-pinene, p-cymene, eucalyptol, limonene, 3-carene, linalool, eg-humulene, ββ-caryophyllene, longifolene and myrcene were pre-prepared in methanol (Merck KGaA, Darmstadt, Germany) and injected onto adsorbent cartridges in a stream of Hehelium and analyzed with the same conditions as samples. When quantifying

BVOCs for which no standards were available, αq-pinene was used for MTs, and αq-humulene for sesquiterpenes (SQT). For other BVOCs which could not be quantified during the studydid not match any standard, the amount of the compound present on the sample was calculated as a percentage of the total amount on the sample using the chromatogram peak area. The peaks of longifolene and ββ-caryophyllene where coeluted in the chromatography and are therefore presented together as a sum of two compounds in this study. The chromatogram peaks were identified based on comparison with retention times and mass spectra of standards and the mass spectra in the NISTOSTOS library. LabSolutions GCMS post run analysis program was used for data processing (Version 4.30, Shimadzu Corporation, Japan). Detection limit was set to 0.4 ng in the analysis software based on the analysis of blank samples.

Two outliers -in the BVOC samples were found from two Norway spruce trees located at plot 1 in HTM (Fig. 1a). The bark was examined with bark photos and it was revealed was detected that the chamber in both cases had been placed upon a small emerging branch with some spots of resin as well as with one single needle stuck on the bark. This was believed to have caused the outliers and these samples were considered unusable and excluded from further analysis. All samples from one Norway spruce at plot 2 were also excluded from the analysis after discovering placements on top of a bark hole likely not originating from spruce bark beetles, and thus not suitable in this study.

2.4 Emission rate calculation and standardization

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295 The BVOC concentrations obtained from the sample—air analysis were converted to emission rate (ER) (μg m⁻² h⁻¹) according to Eq. (1), following Ortega & Helmig (2008):

$$ER = \frac{[c_{out} - c_{in}]Q}{A},\tag{1}$$

where C_{out} (µg l⁻¹) is the concentration of each compound within the chamber, and C_{in} (µg l⁻¹) is the concentration of the compound in the filtered inlet air, Q is the flow rate through the chamber (l min⁻¹) and A is the bark surface area (m⁻²) covered byinside the chamber.

The ER per hole was calculated as the average by dividing the ER derived from Eq. (1) for the respective sample with the number of <u>counted</u> holes (the number of holes can be found in Table 1 Table 2) according to Eq. (2):

$$ER \ per \ hole = \frac{ER}{\# \text{ holes}},\tag{2}$$

Finally, the ER for one square meter of bark surface, ER_{sqm} , was extrapolated based on the number of holes within the chamber and the chamber's bark area according to Eq. (3):

$$ER_{sqm} = ER \cdot \#holes \cdot \frac{1}{4},$$
 (3)

The emission rates of the infested Norway spruce trees were scaled with an average of the holes per square meter found for the measured trees in HTM and NOR (Table 2). By doing this, any variation in emission rate caused by a difference in amount of holes was removed which enabled more accurate comparison between the infested trees. To remove the influence from the variations of the emission rates due to the difference in amount of bark beetle holes, the emission rates were scaled to represent the same number of holes using an average number of bark beetle holes per square meter calculated using the counts from this study in Table 1 Table 2. The average holes per

315 square meter was applied to the emission rates for the infested trees, enabling comparison of emission rates from all infested trees.

The hole type majority was determined from bark photos and used tTo separate entry and exit holes in the data set. A majority of entry holes were found for, measurements taken up to 100 days after infestation was determined to have and a majority of exitntry holes for and measurements later than 100 days after since infestation, with the latest measurement occurring 350 days after infestation start, had a majority of exit holes. The timeframe of 100 days was determined by confirming the hole type majority from bark photographs. As the two hole types were consistently occurring before or after 100 days since infestation start, the measurements with mainly entry holes is referred to as the early season and the measurements with mainly exit holes as the late season.

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As the bark surface temperature varied over the season and between the days, the emission rates were standardized using the algorithm for stored, temperature dependent BVOCs (G93) by Guenther et al., (1993; G93) according to Eq. (4):

$$330 \quad M = M_S \cdot e^{(\beta(T - T_S))}, \tag{4}$$

where M is the emission rate (µg m⁻² h⁻¹) at a given bark temperature, T, and $\beta \underline{\beta}$ (0.09 K⁻¹) is an empirical coefficient establishing the temperature dependency (Guenther et al., 1993). M_s is the emission rate at standard temperature T_s of 30 °C.

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The temperature sensitivity of compound emission rates was calculated using a Q_{10} relationship (Lloyd and Taylor, 1994) following Seco et al. (2020) where the Q_{10} coefficient represents the factor by which the compound emission rate increases for every 10 °C temperature increase from a reference emission rate, F_0 . Only compounds appearing in more than three individual samples were selected for further analysis <u>using this method</u>. Log transformed emission rates were binned into 1 °C bins and the mean emission rate per bin was calculated except for bins with only one value. An orthogonal distance regression was applied to the binned mean emission rates weighed by their standard deviation to determine Q_{10} and F_0 using Eq. (5):

$$F = F_0 \cdot Q_{10}^{(T-T_0)/10},\tag{5}$$

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where F_0 is the reference emission rate at temperature T_0 (=30 °C), F is the flux rate at bark surface temperature T (°C), and Q_{I0} is the temperature coefficient.

Based on the Guenther algorithm (G93, Guenther et al., 1993; Eq. (4)) and the Q_{10} temperature dependency calculation (Q_{10} , Lloyd and Taylor, 1994; Eq. (5)), an estimation of the total BVOC constitutive total bark VOC emission rate from healthy Norway spruce bark and the needleleaf VOC BVOC emission rate throughout the season was calculated. Both algorithms were used to calculate bark BVOC emissions, while only G93 was used to calculate the needleleaf emission, as well as bark SQT emission rate. The calculated modelled emissions for bark was based on the measured tree trunk temperature from the ICOS ecosystem data in HTM (Heliasz, 2020), taken at 3 meter height. An average of the trunk temperature measurements was taken in the Neorth and Eeast orientation of the trunk was used for this. because the trunk BVOC sampling was taken in these orientations.

The <u>needleleaf</u> emissions for MT and SQT were calculated according to Eq. (4) using the standardized seasonal average emission rate (M_s), 1.25 μg g(dw)⁻¹ h⁻¹ for MT and 0.34 for SQT μg g(dw)⁻¹ h⁻¹, taken from van Meeningen et al. (2017) measured in HTM during 2016. The temperature input was the canopy-level air temperature measured at 24 meter agl. taken from the HTM ICOS station (Heliasz, 2020). The output of Eq. (4) was scaled from g(dw) to m² by using a specific leaf area (SLA) of 38.4 cm⁻² g⁻¹ calculated from Wang et al. (2017).

Two outliers in the BVOC samples were found and after examination with bark photography, it was discovered that
the chamber in both cases had been placed upon a small emerging branch with some spots of resin as well as with
one single needle stuck on the bark. These samples were considered unusable and excluded from further analysis.

All samples from one spruce at plot 2 were also excluded from the analysis after discovering placements on top of
a bark hole likely not originating from spruce bark beetles, and thus not suitable in this study.

2.5 Statistical analysis

All samples were tested for normality by creating normal probability curves (normplot, MATLAB R2021a, The MathWorks, Inc., MA, USA) which indicated ing no normal distribution in the data. Statistical analysis of all measurements were thus performed using a Kruskall-Wallis test (MATLAB R2021a, The MathWorks, Inc., MA, USA) with a level of significance set to *P*<0.05. To assure there werethat no deviation between the plots of the healthy Norway spruce trees in HTM occurred, the we tested the following scenario was tested: 1) the difference in emission rates from the healthy trees at plot 1, 2 and 3 in HTM. To test the studyour aim and hypotheses, t-we tested the following scenarios were tested: 2) the difference in emission rates from healthy and infested trees from all plots and sites (H1), 3) the difference in emission rates from one initially healthy spruce and one initially stressed spruce (aim (iv)) and 4) the difference between the calculated *Q*₁₀ coefficient and *F*₀ for the healthy and infested trees (aim (iii)).

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To test H2 and H3, an exponential function (Curve Fitting Toolbox, MATLAB R2021a, The MathWorks, Inc., MA, USA) was fitted applied to the data using (Eq. 6):

$$f(x) = a \cdot \times e^{b \cdot \times x},$$

$$(6)$$

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 $f(x) = a \times e^{b \times x}$, where x is the emission rate in μ g m⁻² h⁻¹. The following scenarios were compared:)

1) emission rates from infested Norway spruce and the evolvement over time (H2) and 2) emission rates from infested Norway spruce and the number and type of bark beetle holes (H3).

390 The significance of the difference between the healthy tree sites, the emission rates from control trees and infested trees, the difference in emission rates from the two infested trees S3S2 and S3S3, and the difference between Q_{10} and F_0 for healthy and infested trees was analyzed using a Kruskal Wallis test. The level of significance was p < 0.05.

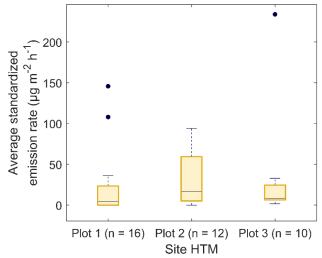
3 Results

395 3.1 Constitutive and induced Bark BVOC emissions from healthy and infested Norway spruce bark

For the healthy spruce Norway spruce trees in HTM, the average total temperature standardized temperature standardized bark BVOC emission rate from all samples (n=113) was $32\frac{1.89}{1.672} \pm 5\frac{1.672}{1.672} \mu g \text{ m}^{-2} \text{ h}^{-1}$ (mean ± standard

deviation: Table 3). The most dominant BVOC group was MTs $(29.37 \pm 51.01 \, \mu g \, m^{-2} \, h^{-1}$: Table 3)) followed by SQTs $(2.12 \pm 3.217 \, \mu g \, m^{-2} \, h^{-1}$: Table 3)). Isoprene emissions were detected in 58% of total samples from the healthy sprucetree bark with an average emission rate of $0.40 \pm 0.985 \, \mu g \, m^{-2} \, h^{-1}$ (Table 3).

The variability of the emission rates differed little between the <u>plots in HTMsites.</u> where plot 1 had a daily average total temperature standardized bark emission rate of $23.26 \pm 42.53 \, \mu g \, m^2 \, h^4$, plot 2 of $30.38 \pm 32.35 \, \mu g \, m^2 \, h^4$ and plot 3 of $47.54 \pm 79.22 \, \mu g \, m^2 \, h^4$. The standardized emission rates were ranging from 0-145.7 $\mu g \, m^2 \, h^4$, for plot 1, 0-94.2 $\mu g \, m^2 \, h^{-1}$ for plot 2 and 1.5-2354 $\mu g \, m^2 \, h^{-1}$ for plot 3 where the median <u>emission rates wereare</u> 4.1, 17 and <u>87.9</u> $\mu g \, m^2 \, h^{-1}$ respectively (Fig. 5). No statistically significant difference (<u>p value > P0>0</u>.3) was found for the <u>in standardized</u> emission rates between the <u>sites-plots</u> and no clear pattern of diurnal variation was found in the samples.



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410 Figure 5: Boxplots of tThe temperature-standardized temperature standardized emission rates of the control-healthy trees for-the plot 1-3 in Hyltemossa, where plot_3 is located furthest away from the station in an older forest stand.

The number of samples taken at each plot is indicated by n, the black dots indicates outliers. the whiskers indicate the minimum and maximum value and the line indicates the median. There is no The difference in emission rates was tested using a Kruskall-Wallis test (MATLAB R2021a, The MathWorks, Inc., MA, USA) and indicated eating nostatistically significant difference for the daily average of the total temperature-standardized temperature standardized emission rates (p>P0>0.3).

For the bark beetle infested trees <u>located in both sites (HTM and NOR)</u>, the the calculations of seasonal average emission rate was separated into early season and late season as the bark beetle typically drill entry holes earlier, and exit holes later. The seasons were separated based on infestation start, where the early season was less than 100 days since infestation start and the late season after more than 100 days of infestation. The average total temperature standardized_bark_emission rate for Norway spruce infested in_from the early season bark beetle infested spruceNorway spruce trees (n = 6) was 6,70690 ± 6,90860_µg m⁻²-h⁻¹ (mean ± standard deviation; Table 3). T, while the average for trees infested in the late season (n = 8) was 2,001970 ± 1,3100 µg m⁻² h⁻¹ (Table 3). MTs was the most dominant BVOC group throughout the season with an average of 6,6030 ± 6,7040 µg m⁻² h⁻¹ for the late season, followed by SQTs (early: 53 ± 74 µg m⁻² h⁻¹, late: 18 ± 24 µg m⁻² h⁻¹; Table 3⁻). Throughout the season, isoprene was also found in 42_% of the samples with an average emission rate of 3.436 ± 6.769 µg m⁻² h⁻¹ during the early season and 0.14 ± 0.20 µg m⁻² h⁻¹ for the late season (Table 3).

For all measured Norway spruce trees at both sites, a total of 74 individual VOCs-BVOCs were found throughout the measurement period for all samples (n = 151) whereof 32 were MTs, 5 were SQTs and 37 were classified as other BVOCs including isoprene. For the healthy spruceNorway spruce tree samples in HTM (n = 113), 44 individual compounds were found in total, where 12 were MTs, 2 were SQTs and 30 other BVOCs including isoprene. The Despite the lower sample count for the infested spruceNorway spruce trees measured at both sites had less samples (n = 38) compared to the healthy tree samples, but a higher number of individual compounds was found with 52 compounds in total where the majority of the compounds were MTs (n = 30) which was more than the double compared to the healthy trees. There were also more SQTs (n = 5) found in the infested tree samples, but less other BVOCs including isoprene; (n = 17) compared to the healthy tree samples. For the infested trees, there was also a difference in how many compounds were found early in the season compared to later, in total 40 and 33 individual compounds were found for the early and 33 compounds for the late season respectively (Table 3). For MTs and SQTs, more individual compounds were found in the early season (27 MTs & 5 SQTs) compared to the late season (17 MTs & 2 SQTs), but for the other BVOCs more were found in the later season which had 14 individual compounds identified compared to 8 in the early season.

A significant difference was found for the daily average of the total standardized bark BVOC emission rate when comparing healthy trees and infested trees healthy and infested trees from all plots and sites, for both early and late season (p < P0 < .00;.001; Fig. 6). During the early season, the infested trees had a median emission rate of 6,400 $\mu g m^{-2} h^{-1}$ and 2,100 $\mu g m^{-2} h^{-1}$ during the late season (Fig. 6). The emission rates for infested trees during the early and late season were around This is around 740- and 240-fold0 times higher for the infested trees during early and late season-compared to the median of the healthy trees (The median of the daily average emission rates is 8.6 $\mu g m^{-2} h^{-1}$; Fig. 6). for the healthy control trees, and 6385 $\mu g m^{-2} h^{-1}$ and 2102 $\mu g m^{-2} h^{-1}$ for the infested trees during early and late season respectively, i.e. around 740 and 240 times higher for the infested individuals early and late in the season, respectively.

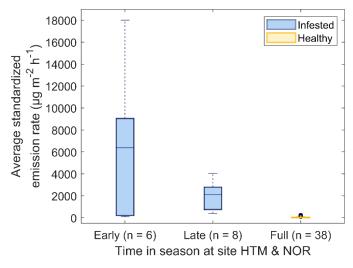


Figure 6: Boxplots of the The temperature standardized emission rates of the infested Norway spruce trees (blue) during the early and late season and the healthy Norway spruce trees (yellow) during the full season. The healthy Comparison between the daily average total standardized emission rate per measured tree and time for healthy trees were measured in Hyltemossa and infested trees were measured both in Hyltemossa and Norunda. To consider the seasonal pattern of the spruce bark beetle, the infested trees were divided into early and late season, where the early season is dominated by entry holes and the late season by exit holes.—The number of samples taken at each plot is indicated by n and the black dots indicates outliers, the whiskers indicate the minimum and maximum value and the line indicates the median. The difference in emission rates for the infested and healthy trees was tested using a Kruskall-Wallis test (MATLAB R2021a, The MathWorks, Inc., MA, USA) which indicates significantly higher emission rates from infested trees during both seasons compared to the healthy trees (P<.001 for both). The emission

470 A difference between the healthy trees and infested trees healthy and infested trees from both sites was also apparent in the occurrence of the compounds throughout the samples (Table 2 Table 3). The most common MT compounds among the healthy spruce Norway spruce trees were equipinene (76 %, relative occurrence in all samples), \(\beta \beta \), 3-carene (48 %) and limonene (44 %). For infested \(\frac{\text{spruce}}{\text{Norway}} \) spruce trees in both seasons, the mentioned MTs were also the most occurring compounds but they also, however, they occurred 475 in more samples (88-100 %). The late season also had 100 % occurrence of (1S)-camphene while that compound only occurred in 75 % of the samples for the early season. For SQTs, eq. humulene occurred most among the healthy trees (47 %) followed by longifolene+ββ-caryophyllene (17 -%). For the infested trees in the earlylate season the pattern was reversed, the SQTs occurring most were longifolene+ββ-caryophyllene (565-%) followed by α-humulene (3148 %). For the early season, The late season showed a similar pattern -where the occurrence 480 was similar for longifolene + $\beta \underline{\beta}$ -caryophyllene occurred most (556 %) followed by but $\alpha \underline{\alpha}$ -humulene occurred in more samples (1831 %) which occurred in fewer samples compared to the early season compared to the late season. The SQTs germeacrene D, isoledene and ββ-cubebene were found to be emitted from one of the infested trees duringing the early season, but was not discovered d at any other time. Isoprene was found to be mostly occurring among the other BVOCs for both the healthy (58 %) and infested spruce Norway spruce during the early and late 485 season (63 % and 27 % respectively). After isoprene, decanal (45 %), benzene (45 %), nonanal (38 %) and toluene (21 %) were occurring most for the healthy spruceNorway spruce. For the early season infested spruceNorway spruce trees, 2-methyl-1-phenylpropene (38 %) and 2-methyl-3-buten-2-ol (19 %) were occurring most after isoprene, however, this was not the case for the infested trees in the late season where 2-methyl-3-buten-2-ol was not emitted and 2-methyl-1-phenylpropene occurred in 9 % of the samples.

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Comparinged to the constitutive emission rates from healthy infested trees to, the emission rates of healthy infested trees, the emission rates were shownshowed _to_increases for all individual compounds with increases ranging from a 200 %3-fold increase to a 250,000 %2580-fold increase for both early and late season (Table 2Table 3). The group of MTs had the highest (230-fold) increase of 22,467800 % during the early season compared towhile SQTs (increased by 2,400 %25-fold increase) and isoprene (8-fold increase) e increased with 740 %. The emission rates during the late season also contributed showed to aa 65-foldn increase of 6,6008 % for the MTs and 70049 % 8-fold for the SQTs, however, isoprene was found to have a 0.4-fold be decrease sing from the infested tree emission rates in the late season compared to healthy tree emission rates. (65 %). The compound (+)-sabinene had the highest increase of all individual compounds (2580-fold(2579007863 %) during the early season when comparing healthy and infested tree emission rates (Table A1). The compounds: tricyclene, eucalyptol, 4-carene, zeta (fenchene, eq-phellandrene, trans,trans(4E.6E)-alloocimene, norbornane, gammay-terpinene, eq-fenchene, 2-carene, eq-thujene, eq-terpinene wereas only found only occurred emitted from in_infested trees and indicate a change in the chemical composition of the emitted BVOCs when a tree is infested (full table is found in Appendix Table A1).

Table 2. The compounds occurring most from all samples throughout the season for healthy trees and infested trees. Presented is the temperature standardized seasonal average emission rate (µg m⁻²·h⁻¹ ± one standard deviation) for each compound and the groups of MT, SQT and other BVOCs and the occurrence (%) of each compound in all samples. The increase (%) is presented for the infested trees as an increase from healthy to infested. The compounds that were

510 identified but unable to quantify is presented as n.q. (no quantification). A full list of all identified compounds is found in Table A1.

_	Hea	lth y	Infes	ted early so	eason	Infe	sted late se	ason
Compound name	$\frac{average \pm}{std}$ $\frac{(\mu g \ m^2 \ h^4)}{}$	occurrence (%)	average ± std (μg m⁻² h⁻¹)	increase (%)	occurrence (%)	average ± std (μg m² h¹)	increase (%)	occurrence (%)
Monoterpenes	29.37 ± 51.01	-	6630 ± 6740	22678	-	1950 ± 1350	6608	-
alpha Pinene	11.49	76.11	911.14	7830	100	824.64	7077	100
beta Pinene	8.22	55.75	954.17	11508	100	225.28	2641	100
3-Carene	2.48	48.67	285.16	11398	100	33.07	1233	95
Limonene	1.89	44.25	320.82	16875	88	85.43	4420	100
p-Cymene	0.49	39.82	241.45	49176	63	53	10716	77
beta-Myrcene	0.32	17.70	159.76	49825	79	6.3	1869	86
beta Phellandrene	2.7	10.62	673.13	24831	44	189.47	6917	68
(1S)-Camphene	1.7	6.19	1516	89076	75	388.82	22772	100
(+)-Sabinene	0.08	0.88	206.37	257863	44	2.93	3563	5
Sesquiterpenes	2.12 ± 3.17	-	53.0 ± 74	2400	-	18 ± 24	749	-
Longifolene+bet a Caryophyllene	0.7	17.70	37.65	5279	56	13.6	1843	55
alpha Humulene	1.42	47.79	4.86	242	31	4.52	218	18
Germacrene D	-	-	3.86	-	19	-	-	-
Isoledene	-	-	3.25	-	19	-	-	-
beta Cubebene	-	-	3.2	-	19	_	-	_
Other BVOCs	0.40 ± 0.85	-	3.36 ± 6.69		-	0.14 ± 0.20	-	-
Isoprene	0.4	58.41	3.36	740	63	0.14	-65	27
Decanal	n.q.	45.13	n.q.	-	13	n.q.	-	14
Benzene	n.q.	45.13	-	-	-	n.q.	-	14
Nonanal	n.q.	38.94	n.q.	-	6	n.q.	-	14
Toluene	n.q.	21.24	n.q.	-	6	n.q.	-	9
2 Methyl 1 phenylpropene 2 methyl 3	-	-	n.q.	-	38	n.q.	-	9
buten 2 ol	-	-	n.q.	-	19		-	

Table 3. Seasonal average temperature standardized emission rate (μg m² h¹ ± one standard deviation) from all Norway spruce trees located in Hyltemossa and Norunda. Presented areis the frequently occurring unique compounds, compound groups (Monoterpenes, sesquiterpenes and other BVOCs) and total BVOCsemission emitted from healthy and infested (early and late season) Norway spruce bark. The increase or decrease (%) is presented for the infested trees as a change in emission rate from healthy to infested. The occurrence (%)- is presented to indicates how often much each compound appeared in the samples throughout the growing season. The compounds that were identified but unable to quantify areis presented as n.q. (no quantification). A full list of all identified compounds is found in the Appendix (Table A1).

-	<u>Hea</u>	<u>lthy</u>	Infest	ted early se	<u>eason</u>	<u>Infested late season</u>			
Compound name	$\frac{average \pm}{std}$ $(\mu g m^{-2} h^{-1})$	occurrence (%)	$\frac{average \pm}{std}$ $(\mu g m^{-2} h^{-1})$	increase (%)	occurrence (%)	$\frac{average \pm}{std}$ $(\mu g m^{-2} h^{-1})$	increase (%)	occurrence (%)	
Monoterpenes	<u>29 ± 51</u>	-	6,600 ± 6,700	22,400	1	1,900 ± 1,300	<u>6,500</u>	-	
<u>α-Pinene</u>	<u>12 ± 20</u>	<u>76</u>	910 ± 1030	<u>7,800</u>	<u>100</u>	820 ± 890	<u>7,100</u>	<u>100</u>	
<u>β-Pinene</u>	<u>8 ± 19</u>	<u>56</u>	950 ± 960	11,500	<u>100</u>	230 ± 170	2,600	<u>100</u>	
3-Carene	<u>3 ± 5</u>	<u>49</u>	290 ± 420	<u>11,400</u>	<u>100</u>	<u>30 ± 40</u>	<u>1,200</u>	<u>95</u>	
<u>Limonene</u>	<u>2 ± 3</u>	<u>44</u>	320 ± 320	16,900	<u>88</u>	<u>90 ± 80</u>	<u>4,400</u>	<u>100</u>	
p-Cymene	<u>1 ± 1</u>	<u>40</u>	<u>240 ± 320</u>	<u>49,200</u>	<u>63</u>	50 ± 40	10,700	<u>77</u>	

<u>β-Myrcene</u>	0.3 ± 0.8	<u>18</u>	160 ± 170	49,800	<u>79</u>	10 ± 6	<u>1,900</u>	<u>86</u>
<u>β-Phellandrene</u>	<u>3 ± 8</u>	<u>11</u>	670 ± 660	<u>24,800</u>	<u>44</u>	190 ± 160	<u>6,900</u>	<u>68</u>
(1S)-Camphene	<u>2 ± 6</u>	<u>6</u>	$\frac{1,520 \pm}{1,970}$	89,100	<u>75</u>	390 ± 230	22,800	<u>100</u>
(+)-Sabinene	0.1 ± 0	<u>1</u>	<u>210 ± 200</u>	257,900	<u>44</u>	<u>3 ± 0</u>	<u>3,600</u>	<u>5</u>
<u>Sesquiterpenes</u>	2.1 ± 3.2	-	53 ± 74	<u>2,400</u>	-	<u>18 ± 24</u>	<u>700</u>	1
Longifolene+β- Caryophyllene	0.7 ± 1.4	<u>18</u>	38 ± 70	<u>5,300</u>	<u>56</u>	<u>14 ± 24</u>	<u>1,800</u>	<u>55</u>
<u>α-Humulene</u>	<u>1.4 ± 3</u>	<u>48</u>	<u>5 ± 9</u>	<u>200</u>	<u>31</u>	<u>4 ± 13</u>	<u>200</u>	<u>18</u>
Germacrene D	Ξ	Ξ	<u>4</u>	Ξ	<u>19</u>	Ξ	Ξ	Ξ
<u>Isoledene</u>	=	Ξ	<u>3</u>	Ξ	<u>19</u>	Ξ	Ξ	Ξ
<u>β-Cubebene</u>	Ξ	Ξ	<u>3</u>	Ξ	<u>19</u>	-1	Ξ	Ξ
Other BVOCs	0.4 ± 0.9		3.4 ± 6.7		_	0.1 ± 0.2		
	<u>0:1 = 0:9</u>	-			-		-	-
Isoprene	0.4 ± 0.9	<u>58</u>	3 ± 7	<u>700</u>	<u>63</u>	0.1 ± 0.2	<u>-65</u>	<u>-</u> <u>27</u>
				<u>700</u> <u>-</u>				
Isoprene	0.4 ± 0.9	<u>58</u>	<u>3 ± 7</u>		<u>63</u>	0.1 ± 0.2	<u>-65</u>	<u>27</u>
Isoprene Decanal	0.4 ± 0.9 n.q	<u>58</u> <u>45</u>	3 ± 7 n.q		63 13	0.1 ± 0.2 n.q	<u>-65</u>	27 14
Isoprene Decanal Benzene	0.4 ± 0.9 n.q n.q	58 45 45	3 ± 7 n.q -		63 13 =	0.1 ± 0.2 n.q n.q	<u>-65</u>	27 14 14
Isoprene Decanal Benzene Nonanal Toluene 2-Methyl-1- phenylpropene	0.4 ± 0.9 n.q n.q n.q	58 45 45 39	3 ± 7 n.q = n.q	2 2 2	63 13 = 6	0.1 ± 0.2 n.q n.q n.q	<u>-65</u>	27 14 14 14
Isoprene Decanal Benzene Nonanal Toluene 2-Methyl-1-	0.4 ± 0.9 n.q n.q n.q n.q	58 45 45 39 21	3 ± 7 n.q = n.q n.q	2 2 2	63 13 = 6 6	0.1 ± 0.2 n.q n.q n.q n.q	<u>-65</u>	27 14 14 14 9

=	Hea	<u>lthy</u>	<u>Infes</u>	ted early so	eason	<u>Infe</u>	sted late se	ason
Compound name	$\frac{average \pm}{std}$ $\frac{std}{(\mu g m^{-2} h^{-1})}$	occurrence (%)	$\frac{average \pm}{std}$ $\frac{std}{(\mu g m^2 h^{-1})}$	increase (%)	occurrence (%)	$\frac{average \pm}{std}$ $\frac{std}{(\mu g m^2 h^{-1})}$	increase (%)	occurrence
Monoterpenes	29 ± 51	П	<u>6600 ±</u> <u>6700</u>	<u>22400</u>	П	<u>1900 ±</u> <u>1300</u>	<u>6500</u>	=
<u>α-Pinene</u>	<u>11.5</u>	76	910	7800	100	820	7100	<u>100</u>
<u>β-Pinene</u>	<u>8.2</u>	<u>56</u>	<u>950</u>	<u>11500</u>	<u>100</u>	230	2600	<u>100</u>
3-Carene	<u>2.5</u>	<u>49</u>	290	<u>11400</u>	<u>100</u>	<u>30</u>	<u>1200</u>	<u>95</u>
<u>Limonene</u>	<u>1.9</u>	<u>44</u>	320	<u>16900</u>	<u>88</u>	90	<u>4400</u>	<u>100</u>
p-Cymene	<u>0.5</u>	<u>40</u>	240	<u>49200</u>	<u>63</u>	<u>50</u>	<u>10700</u>	77
<u>β-Myrcene</u>	<u>0.3</u>	<u>18</u>	<u>160</u>	<u>49800</u>	79	<u>10</u>	<u>1900</u>	<u>86</u>
<u>β-Phellandrene</u>	<u>2.7</u>	<u>11</u>	<u>670</u>	<u>24800</u>	<u>44</u>	<u>190</u>	<u>6900</u>	<u>68</u>
(1S)-Camphene	<u>1.7</u>	<u>6</u>	<u>1520</u>	89100	75	390	22800	<u>100</u>
(+)-Sabinene	<u>0.1</u>	<u>±</u>	210	<u>257900</u>	<u>44</u>	<u>0</u>	<u>3600</u>	<u>5</u>
<u>Sesquiterpenes</u>	$\frac{2.1 \pm 3.2}{}$	П	$\frac{53 \pm 74}{}$	2400	Ξ	$\frac{18 \pm 24}{1}$	700	=
Longifolene+β- Caryophyllene	<u>0.7</u>	<u>18</u>	<u>40</u>	<u>5300</u>	<u>56</u>	<u>10</u>	<u>1800</u>	<u>55</u>
<u>α-Humulene</u>	<u>1.4</u>	<u>48</u>	<u>4.9</u>	200	<u>31</u>	<u>4.5</u>	200	<u>18</u>
Germacrene D	Ξ	Ξ	<u>3.9</u>	Ξ	<u>19</u>	Ξ	Ξ	Ξ
<u>Isoledene</u>	=	Ξ	<u>3.3</u>	Ξ	<u>19</u>	Ξ	Ξ	Ξ
<u> </u>	Ξ	Ξ	<u>3.2</u>	Ξ	<u> 19</u>	Ξ	Ξ	Ξ
Other BVOCs	0.4 ± 0.9	Ξ	3.4 ± 6.7		Ξ	0.1 ± 0.2	Ξ	Ξ
<u>Isoprene</u>	<u>0.4</u>	<u>58</u>	<u>3.4</u>	700	<u>63</u>	<u>0.1</u>	<u>-65</u>	27
<u>Decanal</u>	=	<u>45</u>	=	Ξ	<u>13</u>	=	=	<u>14</u>
Benzene	Ξ	<u>45</u>	Ξ	Ξ	Ξ	Ξ	Ξ	<u>14</u>
Nonanal	Ξ	39	Ξ	=	<u>6</u>	Ξ	=	<u>14</u>
<u>Toluene</u>	=	<u>21</u>	Ξ	Ξ	<u>6</u>	=	Ξ	<u>9</u>

2 Methyl 1 phenylpropene	=	Ξ	=	Ξ	38	Ξ	Ξ	9
2 Methyl 3 buten 2 ol	=	=	=	Ξ	<u>19</u>	п	Ξ	=
Total	32 ± 52	Ξ	6700 ± 6900	20900	Ξ	<u>2000 ±</u> 1300	<u>6000</u>	=

3.1.1 Scaling the infested tree bark emission with number of bark beetle holes

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The temperature standardized emission rates (TS) for the total BVOCs from the bark beetle infested Norway spruce trees from both sites were ranging had seasonal averages ranging from aroundbout 500 to 13,000 µg m⁻² h⁻¹ for all trees throughout the measurement period (Fig. 7). The daily average TS emission rate per bark beetle hole for infested trees during the early season was 22.04_-± 298.69 µg hole-1 h-1 which is emissions from mainly entrance holes and. The bark beetle holes from infested trees during the late season and with mainly exit holes had a daily average temperature standardized emission rate of 4.41 ± 3.51 µg hole-1 h-1 for the late season. The average number of holes per square meter bark area found in this study was 554 based on the values in Table 1. When applying scaling the TS emission rates with the average number of bark beetle holes to emission rates with bark beetle holes (BBH), a comparison with only the TS emission rates showed that the total average of BBH emission rates were higher (Fig. 7). During the early season for the trees measured in the early season the BBH emission rates were about 2,500 μ g m⁻² h⁻¹ higher (9,200 \pm 6,2030 μ g m⁻² h⁻¹) compared to TS emission rates (6,70690 \pm 5,140000 μg m⁻²h⁻¹) and for. For the trees measured in the late season the BBH emission rates were 150 μg m⁻²h⁻¹ higher $(900 \pm 650 \,\mu g \,m^{-2} \,h^{-1})$ compared to TS emission rates $(750 \pm 400 \,\mu g \,m^{-2} \,h^{-1})$. For the individual trees and for the group of MT, the BBH emission rates of MT increased compared to TS emission rates for all trees but two (tree S1S1 and tree S5S3) where the TS emission rates were about 300 µg m⁻² h⁻¹ higher (Fig. 7). The inconsistent variation in emission rates scaled with BBH or TS can be explained by the difference in number of bark beetle holes found per tree (Table 1 able 2). The TS emission rates only consider the bark beetle holes inside the bark chamber while the BBH emission rates are calculated based on holes extrapolated to average holes per square meter., By applying and an average of the bark beetle holes found in this studys the same average of bark beetle holes were used for all to all trees, any variations in emission rates caused by a different due to amount of holes can be disregarded can be disregarded. The results from the infested trees are were thus from here on presented as BBH emission unless stated otherwise.

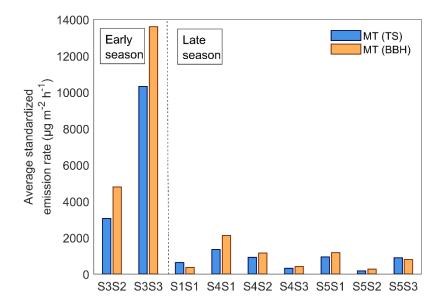


Figure 7: The seasonal average temperature standardized emission rate for the group of monoterpenes (MT) fromor all infested Norway spruce trees located in Hyltemossa and Norunda, for the group of monoterpenes (MT The tree ID is presented at the x-axis and is), separated into early season (< 100 days since infestation start) and late season (less or more than 100 days since infestation start). The temperature standardized emission rates (TS) are presented in blue, while the emission rates also scaled to the same by the average number of number of bark beetle holes (BBH) is presented in orange. The emission rates are a lot higher in the early season where the emission occur mainly from entry holes compared to the late season with mainly exit holes. This is evident even if the emission rates were re-calculated to represent equal number of holes.

3.2 The influence of time since infestation on emission rate from infested trees

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To study the influence of time on the emission rates after a-Norway spruce trees were infested by spruce bark beetles, measurements were done as differenttaken at different occasions in relation-times to-after infestation start in both HTM and NORfor both sites. The earliest After a tree was infested by bark beetles, its emission rates were found to decrease with time passed since the start of the infestation. When the measurements occurred started_after 12 days since the start of after infestation and showed an, the average emission rate for for the total-all VOCs BVOCs of where around 1.90850 μg m⁻² h⁻¹, when excluding athe tree with lowered defence (as presented in Sect. 3.43.2.3; Fig. 8a; excluded tree is marked in yellow). An exponential function $(f(x) = a \times e^{b \times x}, \text{ where } x \text{ is the emission rate in } \mu \text{g m}^{-2} \text{h}^{-1})$ was fitted to all data points according to Eq. 6. Three compounds were selected for further analysis, ββ-phellandrene, eucalyptol and ββ-pinene, using the same exponential function. The emission rates after 12 days were different for the individual compounds compared to the total average, ββ-phellandrene and ββ-pinene have emission rates of around 3.000-3.500 μg m⁻² h⁻¹ and 2.000-2.500 μg m⁻² h⁻¹ respectively (Fig. 8b_x-and-d). Eucalyptol was emitted at slightly lower rates of around 4 and 25 μg m⁻² h⁻¹, depending on if the tree had lowered defence or not, where the low emission rates came from the lowered defence tree (Fig. 8c). Some compounds were not emitted from all infested trees: eucalyptol was only observed from 4 individual trees, and ββ-phellandrene from 7 trees, while ββ-pinene was emitted from all infested trees (n = 9).

After about 100 days since start of infestation, the trees were showing signs of browning and loss of needles—and The emission rates for the total BVOCs had decreased with about 80 %showed a 5-fold decrease on average, emitting around from the start to around 300 μg m⁻² h⁻¹, The emission rates were but still at levels higher than the seasonal—constitutive emissions from healthy Norway spruce trees in HTM (around 30 μg m⁻² h⁻¹; Fig. 8). Compared to the emission rate of the total BVOCs after 100 days since infestation, the emission rates from the compounds ββ-phellandrene and ββ-pinene were at about the same level, however, the decrease on average since

the start of infestation was higher (88 % and 89 %, respectively around 9-fold). Eucalyptol did not have an as distinct decrease but had only decreased with, after 100 days the emission rates had only decreased with 10 %1-fold on average on average. When tThe Norway spruce with ID S1S1 located in plot 1 in HTM was tree memeasured after more than 300 days since the start of infestation start it had lost almost had lost almost all of its needles and some bark. At that time, the, and the emission rates of the total BVOC emission rates from that trees were down to around 40 μg m⁻² h⁻¹, which was at the same level as the constitutive emissions from healthy trees at the same at time (on average 38 μg m⁻² h⁻¹; Fig. 8). No emissions of eucalyptol were found after more than 300 days, but the emission rates of ββ-phellandrene and ββ-pinene after 315 days post infestation were around 70 and 27068 μg m⁻² h⁻¹ respectively, however, after 350 days the emission rates went down to around 32 and 58 μg m⁻² h⁻¹ respectively, also comparable with the constitutive emissions from healthy trees at that time (around 45 μg m⁻² h⁻¹).

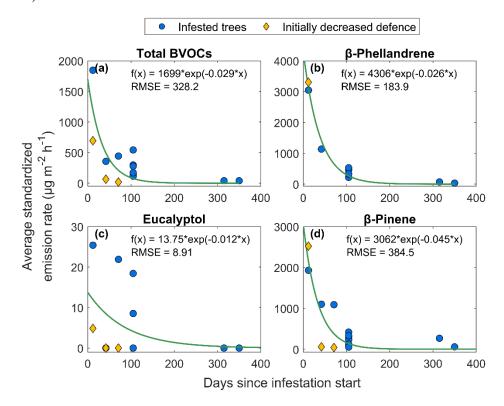


Figure 8: The relationship between average temperature standardized emission rate from all infested Norway spruce trees (blue circle) in Hyltemossa and Norunda and the number of days sinceafter passed since start of infestation start for (a) totalall BVOCs and the compounds; (b) beta per phellandrene, (c) eucalyptol and (d) beta-pinene per pinene. An exponential curve was fitted to the data according to Eq. 6 (green line). All trees are included in the exponential fitted curve, however one tree had initially lowered defence and from a late bark beetle attack previous season and is in the figure mm arked specifically in the figure (yellow diamond) in yellow and diamond-shape for visualization.

3.3 The difference in BVOC emission rates from bark beetle entry holes and exit holesentry and exit holes

No clear relationship was found between the total number of holes and emission rates, likely due to a mixed signal from the type (entry or exit) and time since infestation. The total BVOC temperature standardized emission rates were generally lower from exit holes compared to entry holes when the Norway spruce number of holes were trees had similar amounts of holes (Fig. 9a). The individual compounds emitted from both the entry and exit holes were dominated by $\beta\beta$ -phellandrene, $\beta\beta$ -pinene, $\alpha\alpha$ -pinene and (1S)-camphene (Table 3). The compounds found from entry holes but not from exit holes were: 2-carene, 4-carene, $\alpha\alpha$ -fenchene, $\alpha\alpha$ -phellandrene, $\alpha\alpha$ -terpinene, $\alpha\alpha$ -terpi

However, for the compound eucalyptol (Fig. 9c) emissions were only found from four individuals, which had similar emission rates regardless of entry or exit holes. The oxygenated <u>compounds-monoterpenes</u> myrtenal and bornyl acetate were only found in entry holes but could not be quantified <u>(Table 3)</u>.

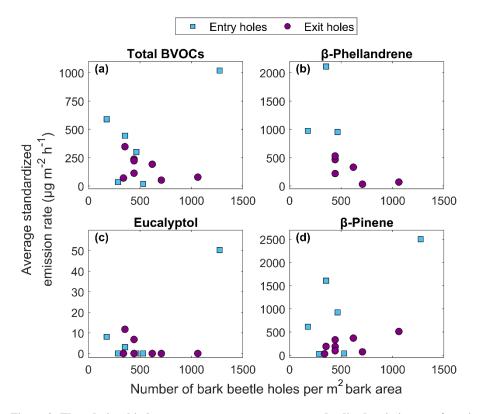


Figure 9: The relationship between average temperature standardized emission rate from infested Norway spruce trees in Hyltemossa and Norunda and the number of bark beetle holes per m² bark area for (a) totalall BVOCs, and the compounds: (b) betaβ-phellandrene, (c) eucalyptol and (d) beta-pineneβ-pinene. There is a distinction between the entry The bark beetle holes are separated into entry holes (bluegreen squares) and the exit holes (purple circles). The exit holes were appearing later during the season (> 100 days since infestation start) which could explain the lower emission rates as the vitality of the trees decrease with higher number of holes. High emission rates from exit holes could indicate signals from blue stain fungi.

3.4 Bark beetle infestation impact over time from two trees with different initial health status

As a part of the sub-study, athe At plot 3, where the bark beetle trap was installed at plot 3 in HTM (Fig. 1). This resulted in successful bark beetle infestation of two Norway spruce trees-with different initial health status, one healthy (tree ID S3S3) and one stressed (tree ID S3S2) were infested by bark beetles. One of the trees had no visual stress signs and was assumed to be healthy (S3S3). The other tree (S3S2) had visual signs of stress already before the beetle attack with old resin flow located high on the stem, most likely due to a late summer attack the previous season. The different status of the trees can be identified in Fig. 10 (a,-b) where S3S2 had significantly higher ($p \rightarrow P0 > 0.02$) total emission rate of bark BVOCs in May, before the infestation, compared to the healthy spruce Norway spruce (S3S3). The remaining two Norway spruce trees on plot 3 were not infested and aAn average of the emission rates for two control healthy trees at plot 3 were taken from these trees were taken to compare with the infested trees is also presented in Figure 8, however, onlyOnly four compounds for the healthy trees were found in May, longifolene+ $\beta\beta$ -caryophyllene, $\alpha\alpha$ -humulene and isoprene (Fig. 10a,b), which had similar emission rates as the other trees and t. The control healthy trees remained at low emission rates for the remaining months (Fig. 10a h) and are thus not included in further comparison of the trees_(Fig. 10). The total emission rates for S3S3 and S3S2-both_trees were induced in June when the bark beetle infestation started (Fig. 10c-2d). There was

no but there was no significant difference (p > P0 > 0.2) in emission rates between the trees, but the compound blend differed between the S3S3 and S3S2. The tree However, a difference was seen in the compound blend. S3S3 had a higher emission rate from the compound 1S camphene(1S)-Camphene (about 800 μg m²h¹) and was also emitting zetaζ-fenchene, trans,trans(4E,6E)-alloocimene, norbornane and αα-thujene compounds which were not emitted from S3S2. The samples from S3S2 in June (Fig.10c,-d) did however contain the bark beetle pheromones germacrene D, isoledene and ββ-cubebene which were not found in the compound blend from S3S3. In July, the emission rate from S3S2 was significantly lower (p < P0 < 0.0325) and close to zero compared to S3S3, which still had high emission rates of ββ-phellandrene, αα-pinene, ββ-pinene and 1S camphene(1S)-Camphene (Fig. 10e,f). A similar difference between the trees was apparent in August as well (Fig. 10g,h). The spruceNorway spruce S3S3 was also found to emit the compound verbenone in August, a compound which could not be quantified in the study (Table 3), which was not found in S3S2.

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The individual compound blend was also found to change over time for the healthy tree, \$383 as it was3, when it 650 got infested and whenas the infestation continued (Fig. A1). In May, when the tree was healthy, Before infestation, in total in total 10 compounds in total were identified, with dominant BVOC being dominated by other BVOCs: decanal (28 %), nonanal (20 %), toluene (16 %) and 1,3,5-triflourobenzene (15 %) where the percentage represents the amount of the compound found in the sample relative to the total amount (Fig. A1) by mass. After bark beetle infestation started in June, the number of detected compounds increased to 27 and was now, dominated by 655 MTs (1S camphene(1S)-Camphene (18%), 5-vinyl-m-xylene (15%) and ββ-phellandrene (9%); Fig. A1)-but also other BVOCs (5 vinyl m xylene (15%)). The emissions during the campaign in July also consisted mainly of MTs with largest contributions from $\beta\beta$ -phellandrene (23 %) and $\beta\beta$ -pinene (23 % and (22 % respectively) followed by 1S camphene (1S)-Camphene (19 %) and αα-pinene (19 % and 16 %; Fig A1-respectively). The compound composition in August was similar as in June, with the majority of the blend consisting of MTs dominated by 1S camphene(1S)-Camphene, αα-pinene and ββ-pinene (22 %, 13 % & 11 % respectively) and other 660 BVOCs (2-methyl-1-phenylpropene (6%); Fig. A1).

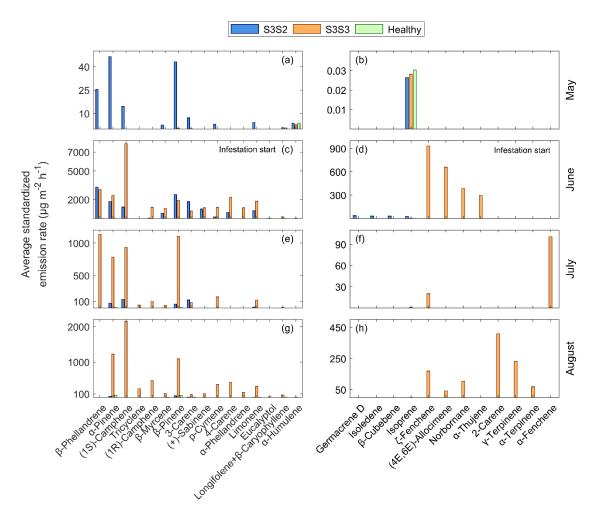


Figure 10: The average temperature standardized BVOC emission rates for all compounds from Norway spruce at plot 3 in Hyltemossa: healthy trees (green), infested spruce with ID S3S2 (blue), and infested spruce with ID S3S3 (orange) and healthy trees (green). Measurements were taken in 2019 during (a,b) May, (c,d) June, (e,f) July and (g,h) August. The graphs are horizontally separated for visibility due to large differences in scale. The healthy trees are included in all graphs but the emission rates are not visible on the same scale as the infested trees in (c,d) June or (e,f) July. The bark beetle infestation had not started in (a,b) May, however, the spruce S3S2 was already subjected to stress from late bark beetle attacks during the previous season before the bark beetle infestation started again in (c,d) June, leading to higher emission rates already in MayThe average standardized emission rates for all compounds for the two spruces S3S2 (blue), S3S3 (orange) and control trees at site 3 (green) for May (a-b), June (c-d), July (e-f) and August (g-h). The graphs are horizontally separated for visibility due to large differences in scale. The control trees are included in all graphs but the emission rates are not visible on the same scale as the infested trees in June (c-d) or July (e-f). The bark beetle infestation had not started in May (a-b), however, the spruce S3S2 was already subjected to stress from late bark beetle attacks previous season before the bark beetle infestation started again in June (e-d), leading to higher emission rates in May.

3.5 Reference emission rate at 30°C and calculated Q_{10} coefficient

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From Eq. (5), the reference emission rate at 30°C (F_0) and the increase in emission rate with every 10°C (Q_{10} coefficient) were calculated for both-the healthy and infested spruce Norway spruce trees as well as for the infested treesfrom both sites. Only compounds found in at least three samples were included in the calculation (Table A2). The result for emitted compounds from both healthy trees and infested trees shows a Q_{10} coefficient rangening spread from 0.1 to 576 for healthy trees and 1.3 to 9804 for infested trees where the Q_{10} coefficient was higher when the trees where infested increased for infested trees for all compounds but one, p-cymene (Table A2). The F_0 value, which indicates the emission rate that compound would have at 30°C for each specific compound, and for this value there was also showed a difference between the healthy and infested trees. The spread of F_0 for healthy trees was ranging from $0.0\underline{105}$ to $9\underline{32}$ µg m⁻² h⁻¹ and from compared to 0.5 to $34\underline{,}900$ µg m⁻² h⁻¹ for the

infested trees, however, compared to the Q_{10} coefficient, the but in this case the F_0 value was higher for all compounds from the infested trees compared to the healthy (Table A2). The average Q_{10} coefficient for all compounds for healthy trees was 13 while it was 96 for infested trees, indicating a 7-fold increase leading to an increase of the Q_{10} coefficient by 60034 %... Thee same average for F_0 was 210.8 μ g m⁻² h⁻¹ for healthy trees and 2.65048 µg m⁻² h⁻¹ for infested trees, a 127-foldn increase, an increase of 12.600 %, which is in line with the increased emission rates when standardized according to G93 (Table A1). The highest increase in both Q_{10} and F_0 was seen in the compounds $\beta\beta$ -pinene and longifolene+ $\beta\beta$ -caryophyllene (two SQTs quantified together) with Q_{I0} increasing $\underline{125\text{-fold}}$ with $\underline{12,500}$ % and $\underline{22,400}$ % $\underline{225\text{-fold}}$ respectively, and F_0 increasing with $\underline{3,160\text{-fold}}$ 316,000 $\frac{4}{8}$ and $\frac{209{,}800 \, \%2{,}100\text{-fold}}{200 \, \text{m}}$ respectively. The lowest change for Q_{10} was seen in $\frac{6}{8}$ -pinene and p-cymene, where $\underline{\alpha}$ -pinene <u>had a 1-fold increase</u> was increasing with 0.7 % from healthy to infested and the \underline{Q}_{10} coefficient for pcymene -was-actually decreasing with 34 %had a 0.6-fold decrease for infested trees compared to healthy. Despite the lower Q_{10} coefficient for p-cymene in infested trees, its F_0 value was still higher for the infested trees, however, it had the lowest increase with the increase of 45049 %5-fold was low compared to the other MT compounds. Isoprene was seen to have the overall lowest increase in F_0 , increasing with 89 % with 0.1-fold from healthy to infested. A significant difference was found for the Q_{10} coefficients for healthy and infested trees (p < P_0 <0.032) as well as for F_0 ($\mathbf{p} \leftarrow \underline{P0} \leq 0.0\underline{106}$).

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There were four compounds for which the requirements for the calculations where only fulfilled for infested trees. Those where eucalyptol, tricyclene, (1R)-camphene and (+)-sabinene, for which an increase or comparison between healthy and infested <u>trees</u> cannot be made, but this might indicate that these compounds could be limited to emissions from infested trees only.

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$3.6\ Calculated \underline{seasonal}\underline{-constitutive}\ BVOC\ emissions\ from\underline{-healthy\ Norway\ spruce}\ bark\ \underline{-over\ the\ season}\ and\ \underline{-comparison\ with\ \underline{-needles}\underline{-leaf\ emissions}}$

The calculated Leafneedle emissions for -over-the growing season of 2019 in Hyltemossa varied betweenwereas found to vary an average with an average of around 60 to 170 μ g m⁻² h⁻¹ for leafneedle MT in July and August and an average of around 25 to 100 in May, and 50 to 120 μ g m⁻² h⁻¹ in May and September, respectively (Fig. 11b). Bark emissions have beenemissions were based on measured tree trunk temperature at 3m agl, averaged from 2 directions (North and East) and the average standardized emissions (*Ms*), 29 μ g m⁻² h⁻¹ for MT and 2 μ g m⁻² h⁻¹ for SQT, and the Q_{10} approach for healthy trees. The calculated emission rates from bark reached a maximum around 16 μ g m⁻² h⁻¹ in July, which is ten times lower than the calculated leafneedle emissions at the same time (Fig. 11c). The bark emission rates remained below 10 μ g m⁻² h⁻¹ for most of the growing season. The estimated bark emission rates from healthy trees using the Q_{10} approach were generally about 5 μ g m⁻² h⁻¹ lower than the calculated emission rates using the G93 approach, but steeply increased during the warmest days to match the G93-emissions (Fig. 11c).

For SQTs, the <u>leafneedle</u> emissions peaked at 30 µg m⁻² h⁻¹ in late July at the same time <u>as</u> when MT emissions were high, and also showed emissions up to 20 µg m⁻² h⁻¹ earlier in June <u>(Fig. A2b)</u>. For most of May and September, SQT emissions from <u>needlesleaves</u> were calculated to be below 5 µg m⁻² h⁻¹. Bark emissions of SQT for healthy trees estimated with G93 were well below 1 µg m⁻² h⁻¹ throughout the season (maximum 0.75 µg m⁻² h⁻¹ in late July), and below 0.3 µg m⁻² h⁻¹ most of the time <u>(Fig. A2c)</u>.

However, when comparing estimated bark emission from healthy trees with actual measurements of infested trees, bark emissions from infested trees were much higher (Fig. 11a). The measured bark MT emission rate from the infested tree reached up to around 18₂000 μg m⁻² h⁻¹ as a daily average for one day, making the total MT emission rate (including leafneedle emissions) increase by almost a 100-fold when the tree was infested. The lowest measured infested tree emission rate (around 3₂900 μg m⁻² h⁻¹) for MT was found during the July campaign, however, despite it being the lowest, itthis was still considerably higher than the constitutive MT emission rate from healthy trees of that day (70 μg m⁻² h⁻¹ including both needle and bark; Fig. 11), which was calculated to be around 70 μg m⁻² h⁻¹ including both leafneedle and bark VOC BVOC emission. Bark MT eEmission rates from the bark of the infested trees were at least around 55 times higher than the total MT emission rate from both needlesleaves and bark of a healthy tree.

. Emission rates from the bark of the infested trees were about 55 times higher than the total constitutive MT emission rate from both leaves and bark, even at the lowest measured emission rates.

For the SQT emission rates, the difference was not as distinct. The SQT emission rate reached maximum emission rate of around 0.75 μ g m⁻² h⁻¹, for bark emissions (Fig. A2b) and around 30 μ g m⁻² h⁻¹ for leafneedle emission (Fig. A2a) whileen the measured bark emission rate from the infested tree peaked around 40 μ g m⁻² h⁻¹ as a daily average for one day, indicating an 1.3-fold increase when a tree is infested. The lowest measured infested tree emission rate was also in July for the SQTs, at around 1.4 μ g m⁻² h⁻¹, which was still higher than the calculated healthyconstitutive bark emission rate at around 0.2 μ g m⁻² h⁻¹, but lower than the constitutive leafneedle emission rate of about 5 μ g m⁻² h⁻¹.

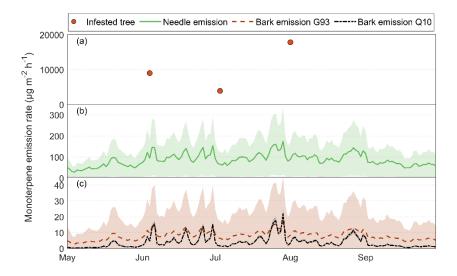


Figure 11. The measured and calculated BVOC emission rates for the group monoterpenes from Norway spruce in Hyltemossa; (a) the actual measured emission rates from infested Norway spruce bark (red dot), (b) calculated needle emission rates (green line) eonstitutiveand—(c) calculated healthy bark—VOC—emission. The needle emissions wereare calculated based on the Guenther algorithm (Guenther et al., 1993)—and the measured emission rates wereare taken from van Meeningen et al., (2017), —and—specific needle area (SLA) was taken from Wang et al., (2017) and—using the air temperature at 24 m was taken from the HTM ICOS station (Heliasz, 2020). The healthy bark emission is calculated emission rates from the group monoterpenes—based on the tree temperature taken at 3 meters height in the north—North and eastEast orientation, data taken from the HTM ICOS station (Heliasz, 2020). The bark emissions are calculated using (black) is calculated based on the Guenther algorithm (Guenther et al., 1993); orange) and the Q10 temperature

dependency (black) (orange) based on measured emission rates in this study (black). The shaded areas (green, shade and orange and black shade) represent the standard deviation from the mean for the respective calculation method. The (b) leaf emission rates (green) are calculated based on the Guenther algorithm and the measured emission rates are taken from van Meeningen et al., (2017) and specific leaf area (SLA) was taken from Wang et al., (2017), using the air temperature at 24 meters taken from the HTM ICOS station (Heliasz, 2020). For comparison, (a) the actual measured bark VOC emission rates from one infested tree over time, from this study, is included in the Figure (red dot).

4 Discussion

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4.1 Constitutive and induced bark Bark BVOC emissions from healthy or infested from Norway spruce

Both emission rates and composition blend of bark BVOCs from Norway spruce trees were found to change when infested by spruce bark beetles, which is in line with previous studies on bark beetle infestation of conifer trees (Amin et al., 2013; Birgersson and Bergström, 1989; Ghimire et al., 2016; Heijari et al., 2011). In this study, we identified 29 compounds unique to infested trees were identified from which where the majority were MTs (n = 19; Table A1). Several of the identified compounds were only emitted from infested trees trees, for example, eucalyptol, isoledene, (+) camphor, tricyclene, α phellandrene, which is were consistent with the findings of Ghimire et al. (2016), for example, eucalyptol, isoledene, (+)-camphor, tricyclene, α-phellandrene (Table A1). The findings in this study also show ilsoprene emission was also found to be emitted from both healthy trees and infested treeshealthy and infested tree barks which initially, which was believed to originate from potential lichen cover as a study by (Zhang-Turpeinen et al., (2021), however, found a positive correlation between isoprene and lichen cover. However, when visually evaluating bark photos for algae and lichen coverage thereis was no clear indication that higher coverage coincided with isoprene emissions ere seemed not to be a relationship when assessing the lichen cover from bark photographs, making the origin of the it uncertain if the isoprene emission did originate from the bark or not.emission uncertain. This is also consistent with the study by Ghimire et al. (2016) also assessed the lichen coverage and isoprene emission and their results are consistent with this study, in which they did not find any-statistically significant relationship with isoprene emission from bark and lichen or algal cover.

With regard to the quantity of BVOC emission however, tThe results of this study indicate a much greater difference—increase in BVOC between emission rates from healthy trees and infested treeshealthy and insect infested conifer trees compared tothan previous findings (Amin et al., 2013; Ghimire et al., 2016; Heijari et al., 2011). In this study, We found the total Norway spruce constitutive—bark BVOC emission rates was found to be from healthy trees to have a seasonal average of 31.89 ± 51.67 μg m²h² (mean ± standard deviation), while the infested trees had an average of 6630 ± 6740 μg m²h² (mean ± standard deviation) for the early season and 1950 ± 1350 μg m²h² for the late season. This implies that bark from infested trees emits—63 to 215-fold times higher when a Norway spruce was infested more BVOC to the atmosphere than compared to healthy trees, depending on how long the infestation hads been ongoing (Fig. 6), where the emissions are higher earlier and decrease with time. Previous findings reported have found increases in emission rates up to 3-fold when Engelmann spruce (Picea engelmannii) was infested by spruce beetles (Dendtoctunus rufipennis;) (Amin et al., 2013), up to 10-fold when Scots pine (Pinus sylvestris) was infested by weevils (Hylobius abietis;) (Heijari et al., 2011) and The measured emission rates in this study are 3 to 9 times higher than the emission rates in the study by Ghimire et al. (2016) in which they found up to 15-fold increase ofd emission rates from all BVOCs when comparing healthy Norway

spruce trees with trees infested by the spruce bark beetle (Ghimire et al., 2016). The measured emission rates in this study are still 3 to 9 times higher than the emission rates in the study by Ghimire et al. (2016) which had ith the highest increase comparing healthy and infested Norway spruce bark. As the emission rates in the study by Ghimire et al. (2016) are is also standardized according to G93, temperature is not an-should impacting factor on the emission rates to create this large difference. -A possible reason might however be for the difference in measured emission rates might be the time since of the infestation start in relation to the measurements. In our study we found an An exponential decay in emission rates over time after infestation were found in this study (Fig. 8), suggesting that if measurements were taken early during the infestation the emission rates would be higher. However, the measured emission rates in this study from exit holes (measured after 100 days) during the late seasonin this study areis still higher than the emission rates found in June for Ghimire et al. (2016) originating from unspecified hole type. As they did not specify how long time the infestation had been ongoing it makes a comparison difficult as their June measurement could have originated from older infestations. -In thisour study, we conducted measurements were taken throughout the growing season, from the same spruce, starting before the bark beetle infestations and. This allowed us to capture from the emission rates from very early infestation to later stages, something that, to our knowledge, has not been done before. Birgersson and Bergström (1989) did measure volatiles emitted from entry holes in bark beetle infested Norway spruce during the first week of infestation, but not longer. This finding makes it very important to consider the influence of time since infestation when modelling emission rates of BVOCs from infested Norway spruce. We could see a trend with exponential decrease in emission rates with time for both the total BVOCs as well as selected compounds. The spruce bark beetles typically have a first swarm in May, followed by a sister brood in June and the initiation of a second generation in July (Jönsson et al., 2009). The trend with decreased emission rates we found is only related to the start of infestation, regardless of the time in the season and from which swarming period the infestation started.

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The results from this study found eEmission rates from infested Norway spruce bark were found to decrease with time, similarly to what (Eller et al., (2013) found when piercing holes in needles from Ponderosa pine (Pinus ponderosa). They found MT emission rates to increase by four orders of magnitude when the needle was pierced compared to undamaged needles and as the exposed resin hardened, the emission rates decreased exponentially until they reached similar levels as undamaged needles already after 30 days. In this study, induced we found increased emission rates from infested Norway spruce bark were seen to last up to 300 days before reaching similar levels as healthy bark, indicating the death of the tree (Fig. 8). This would suggest that the increased emission rates and their exponential decay over time is not only due to exposure of resin occurring as the spruce bark beetle first drill a hole, but also due to the ecological impact developmental process of the beetle and indicated by the visibility of the different hole types created. This is supported by the fact that The timing of the infestation plays an important role in the increased emission rates. wWhen comparing the total number of holes to emission rates for the individual spruceNorway spruce trees, there was no distinct pattern was found (Fig. 9). The hypothesis was that emission rates would be higher at the start of infestation and decrease over time with declining vitality of the trees, which could be explained by a relationship between emission rates and the type of bark beetle hole rather than the total number of holes. However, when the holes were separated When separating the holes into entry and exit, it was apparent that entry holes generally have higher emission rates compared to exit holes (Fig. 9)₅. which This can also be supported by the result indicating higher emission rates at the start of an infestation the relation to time since infestation where the emission rates were highest in the beginning (Fig. 8) when there is a majority of entry holes (Table 2) and entry holes generally appear at the start of an infestation. For the total VOCs BVOCs there is a large spread in emission rates, and the second highest emission rate came from an individual (tree ID S3S2) at plot 3 in HTM that withhad less than 3200 holes per square meter (Table 2), and which was determined to have mainly entry holes (Fig. 9). Some of the lowest emission rates came from an individual an individual (tree ID S1S1) with more than 1000 holes per square meter with mainly exit holes (Table 2). The same is true when looking at the compounds ββ-phellandrene, eucalyptol and ββ-pinene (Fig. 9). The importance of hole type could be further supported when the emission rates were scaled with temperature only and with additional scaling from the averaged bark beetle holes from our study (Fig. 7). Comparing all trees, there is a larger difference in emission rates comparing the two scaling approaches from tree S3S2 and S3S3, which both had a majority of entry holes. Based on tThis result supports, the speculation is that the high emission rates are linked to the time since infestation start. rather explained by the hole type than the total amount of holes. This indicates that when only taking temperature into account, the emission rates were lower, but when all trees had been scaled to have the same number of bark beetle holes, the difference in emission rates is more distinct—especially from the two trees with a majority of entry holes. This supports the conclusion that high emissions are rather explained by the type of holes, where entrance holes are more relevant than exit holes, than the total amount of holes.

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The number of identified BVOC compounds found to be emitted from entry holes were higher compared to exit holes (Table 3), which further supports the assumption that the emission rates are due to ecological impacts of the spruce bark beetles and not only exposed resin. To further support the speculation that the increased emission rates are due to ecological impacts of the spruce bark beetles and not only exposed resin, the number of identified BVOC compounds found emitted from entry holes were higher compared to exit hole (Table 3) The number of identified compounds was also higher in emissions from entry holes. This is consistent with Birgersson and Bergström (1989) who looked at volatiles emitted from entry holes in bark beetle infested spruceNorway spruce. They did however not look at exit holes, but their findings show that during the early stage of an attack, the MT emissions are high and the concentration of the collected MTs during the first day is consistent with what wase found in thisour study 12 days after infestation. Two oxygenated MTs compounds were found only from the entry holes which is consistent with the findings of Birgersson and Bergström (1989) and indicates emissions from the phloem. The bark beetle pheromones: germacrene D, isoledene, ββ-cubebene and 2-methyl-3-buten-2-ol (MB, not quantified) were also only found emitted from entry holes in this study – however, only from one tree (tree ID S3S2) and one day throughout the studyonly during the June campaign (Fig. 10d). The presence of MBThis could be indications <u>indicative</u> of beetlespresent in the galleries during the measurements of that individual tree_(Birgersson et al., 1988; Zhao et al., 2011a). The high increase in emission rates couldan also be an impact a result from of bark beetle associated blue-stain fungus (E. polonica) as a study by Mageroy et al., (2020) found that inoculation withof the fungus in Norway spruce bark was shown to increase the concentration of total terpenes 91-fold after 35 days compared to the concentration in healthy bark. They did however not measure the emission rates from the bark as done in this study, but their finding of high terpene concentration coincides with the high emission rates from infested Norway spruce bark found in this study.-

4.1.1 Indications of differences in emissions from healthy or stressed trees during infestation

At plot 3 in HTM we selected trees Two Norway spruce trees with various different health status were selected at plot 3 in HTM to study if the health status might have an impact on induced tree emissions from bark beetle infestations. Prior to the infestation, we found that the emission rates were different; one tree (ID S3S3) had lower emission rates compared to the other (ID S3S2) which had significantly higher emission rates (Fig. 10a,b). The higher emission rates from S23S23 can be a sign of stress (Loreto and Schnitzler, 2010) and was not only visible in the BVOC emissions, but also from resin flow on the bark, supporting a theory that the high emission rates were

caused by a late summer attack from spruce bark beetles during the previous season. In June, the two trees were subjected to spruce bark beetle infestation, S3S3 foir the first time and S3S2 for the second time and Prior to the infestation, we found that emission rates for two spruces were different; one emitted significantly more BVOCs compared to the other, a state which was indicating stress (Fig. 10). This was not only visible in the BVOC emissions, but also from resin flow on the bark, supporting the claim that the high emission rates were from stress, something which was caused by a late summer attack from spruce bark beetles during the previous season. Despite the old infestation, the stressed tree was attacked by spruce bark beetles again during our measurement period. When both trees had been infested by bark beetles, and were 12 days into the infestation they both trees showed induced emission rates (Fig. 10c,d). There was no significant difference in their respective emission rates; however, the trees were emitting slightly different compound blends, which might have been a caused by from the initial status of the trees. Tree S3S3 he initially healthy tree showehad induced emissions of mostly MTs-mostly induced MTs where of mainly (1S)-camphene, ββ-phellandrene, αα-pinene and 4-carene were dominating the emissions, but there were also emissions of ζ -fenchene, trans, trans(4E, 6E)-alloocimene, norbornane and $\alpha\alpha$ thujene which were not found to be emitted from the initially stressed treetree S3S2. The initially stressed treeTree <u>S3S2</u> was emitting several of the same MTs as S3S3 with the majority being emissions of $\beta\beta$ -phellandrene and ββ-pinene. But in addition to this, the the previously mentioned bark beetle pheromones were found in the samples taken 12 days after infestation from this treepreviously mentioned being emitted once came from this initially stressed tree, and was only found 12 days into the infestation. The pheromones could indicate that there was ongoing blue stain fungi infection caused from the infestation that happened in the previous season or that the tree already had successful bark beetle infestation again (Birgersson et al., 1988; Zhao et al., 2011a). Previous studies have found that priming Norway spruce bark with methyl jasmonate (MeJA) as well as inoculating with blue-stain fungi have increased the spruce defense towards spruce bark beetles (Mageroy et al., 2020; Zhao et al., 2011a). As tree S3S2 had survived a previous infestation, a speculation can be that the tree also had tungi present during the new attack. However, the tree defensedefense for this tree was not found to be increased but it was rather found that the spruce bark beetles easily overtook it evident by as there was visible entry holesentry holes present, something which was found to be lower for trees previous studies have indicated should be lower compared to healthy trees when they are primed with MeJA or inoculated with fungi compared to healthy treesor inoculated beetles (Mageroy et al., 2020; Zhao et al., 2011a). Additional nother evidence for lowered defense of tree S3S2 is a comparison to the initially healthy tree's (S3S3) in regards to BVOC emission rates (Fig. 10):- 7 something that could be supported as the tree was cut down at the end of the measurement period. aAs the infestation continued, the emission rates from the already stressed sprucetree S3S2 weregot significantly lower than from compared to the initially healthy spruce S3S3, an indication of decreased vitality of spruce S3S2 while tree S3S3 had induced emission rates until August indicating ongoing defense, which continued at high emission rates until August. The However, the last measurement in August did however revealled occurrence of verbenone from the initially healthy treetree S3S3, which have been found to be emitted with successful fungal establishment and has been shown to repel bark beetles (Bakke, 2009; Cale et al., 2019). The findings of verbenone could indicate that the bark beetles had successfully overtaken the spruce Norway spruce S3S3 in August, however, this could not be confirmed as the forest owner had to take down the trees which made further measurements impossible. As bark beetle outbreaks have been seen to increase in number, there might be an increase in the number of healthy trees being attacked and killed in addition to the typical attacks on already stressed trees (Jakoby et al., 2019). The results from this study revealed different blends of compounds when a tree was already stressed from previous infestation and attacked again compared to when the tree was healthy before the attack. Another important note on this is that the previously attacked tree (S3S2) indicated In addition to this, we could also see that the one with

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previous infestation indicated induced emission rates until the start of the next season (Fig. 10a,b), during which the tree was infested again with further induced emissions. The healthy tree (S3S3) did however have higher induced emission rates for longer when it was infested compared to the stressed one healthy compared to when tree S3S2 was infested again. These results indicate that the second generation of spruce bark beetles, attacking late during the growing season, might lead to induced emission rates continuing until the next season year. If a tree attacked late during the growing season survives, it can be attacked again the next season along with attacks on healthy trees. This might indicate that when new attacks can occur on the same tree, as well as on healthy trees. The increased attacks on healthy trees as well as the initiation of a second generation of spruce bark beetles and the attack on healthy trees might have a larger impact on the total bark BVOC emission rates from Norway spruce where they are induced for longer.

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The high BVOC emission rates from infested trees does not only affect the trees themselves, but the emissions ultimately theyit also impacts atmospheric processes. Induced emission rates fromef BVOCs due to insect herbivory have been found to potentially increase SOA yields when modelling an increase in emission rates (Bergström et al., 2014). This which would support the speculation that bark beetle induced BVOC emission rates could beareis important to consider when modelling or measuring SOA formation. Taking not only the quantitative aspects of bark beetle induced emission rates into account, but also the qualitative effects into account, the SQTs αα-humulene, longifolene and ββ-caryophyllene and MT α-pinene have been found to have highest SOA yields compared to 16 other BVOCs, where α pinene had the 9th highest SOA yield (Lee et al., 2006). This study found In our study we foundil increased emission rates from of longifolene+ββ-caryophyllene (quantified together) of around 54- and 20-fold from emission rates at around 5300 % and 1850 % from infested trees depending on the time since infestation in the early and late season was found in this study (Table 2Table 3). This could lead to potentially increased SOA yields when forests are subjected to bark beetle outbreaks. The MTs limonene and myrcene were slightly below the SQTs in ranking of SOA yield, and according to theour findings in this study, they were seen to increase with an average of 4400 to 16900 %50- to 170-fold and 1900 to 49900 %30to 530-fold respectively, depending on time since infestation the season, where the highest increase was in the early season. Myrcene had the third highest percentage increase for the seasonal average of all compounds, and appeared in around 80 % of all samples from infested trees, compared to about 20 % from the healthy tree samples. This change in compound blend could potentially lead to large impacts on SOA yield from bark beetle infested trees overall and highlights the importance of measuring and accounting for bark BVOC emissions.

In the comparison of the initially stressed tree and the initially healthy tree, it was apparent that there originally was no significant difference in the total emission rate, but different compound blends were emitted. Linking this to SOA, the higher emissions of limonene and myrcene from the initially healthy tree early during the infestation indicates that higher SOA yields might come from healthy spruce Norway spruce trees when infested. The high emission rates were also seen to continue until the trees were taken down in August which implies implying that potential increase in SOA yield might continue for longer. Compounds unique to both infested trees were emitted as well, where the stressed tree emitted bark beetle related pheromones and the initially healthy tree emitted a broader blend of MTs, these individual compounds might play a role in the SOA yield as well. An increase of As the bark beetle infested trees generally would impact SOA yields, more attacks on healthy trees might further affect the atmospheric processes, specifically production of SOA.

4.2 Bark beetle induced BVOC emissions in relation to other stresses, leafneedle emissions and modelling

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Significant increases of the temperature standardized BVOC emissions of Norway spruce bark of up to around 22000 % for the total group of MTs-were seen when trees were infested by bark beetles early in the season. Around a 230-fold increase in emission rate was seen for the group of MTs. This high increase in emission rate from insect stress for Norway spruce has not previously been observed according to the review by Yu et al. (2021), in which the highest recorded increase was around 2,000_%, including previous studies on spruce bark beetles. Ips typographus. Heat stress from higher temperatures was also identified as a stressor but it did-did also not increase BVOC emissions as much as stress from bark beetles (Yu et al., 2021). A study on Norway spruce with higher air temperatures of 40 °C found that BVOCs increased by 175 % compared to emission rates at air temperature of 30°C (Esposito et al., 2016). This is not close, notto comparable much lower than to the increase found in emission rates from bark beetle infestation found in this study. However, the impact of combined stresses from temperature and insect attacks might further increase the BVOC emissions. This is illustrated by the increase of reference emission rates after infestation across all BVOCs (F₀, Table A2) which are standardized at a reference temperature and therefore the difference between F_0 from healthy to infested trees could be interpreted as the change in stress induced BVOC emissions due to bark beetle attacks without the temperature effect. In our study we found the The temperature sensitivity as expressed by the Q10 coefficient was also found to increase for all compounds but onechange when trees become infested. The Our analysis of the Q_H coefficient showed that the coefficient increased for all compounds but one (of 15, namely p-cymene, Table A2) when trees became infested, indicating that the emission rates were accelerated by both a higher reference emission rate and an accelerated temperature response compared to healthy trees, as temperatures increased, indicating that the emission rates would increase if the temperature increased. The combination of bark beetle stress with increased temperature might thus lead to even higher increase in emission rates. This is however not the focus of this study, but as we found the BVOC compounds temperature sensitivity was found to increase when trees were infested, and as we found bark beetle infestations -to-increased bark BVOC emissions more than any other comparable stress, there might be high influences of BVOC emissions from combined stress, making it important to account for when modelling the emissions.

The increase of bark emission rates we seen found in this study is are high enough to considerably add to the emission rates of a full tree when comparing with emission rates from needles - which is considered the part of the tree with the highest emission rates. When modelling the emission rates, two approaches were used for the bark MT emissions, the G93 algorithm and the Q_{10} approach. The results showed similarities in pattern but the Q_{10} approach had larger increases in emission rates with higher temperature increase, something that was expected. The G93 modelled emission rates were constantly higher than the Q_{10} , and had less variability, which might be explained by the empirical coefficients used in the G93 compared to only taking temperature into account fitting $\underline{F_0}$ and $\underline{O_{10}}$ for each compound separately. For the needle emission rates, only G93 was used because of the light dependent nature of some BVOCs emitted from the needles that could not be explained by the temperature in the Q_{10} approach only. The seasonal average emission rates from spruce Norway spruce needles were measured during 20176 in Hyltemossa in a study by van Meeningen et al. (2017). As the study was conducted at the same site, their results were applied to thiseour study as a comparison of bark BVOC emission to needle emission. It was clear that constitutive MT emissions from healthy bark does not compare to the needle emission (Fig. 11), where the seasonal average of the MT emissions where 11 times higher from needles than bark. However, when comparing seasonal average bark MT emission from infested trees with needle emissions; it was the other way around; the bark emissions from infested trees where 6 to 20 times higher than the needle emissions depending on the time of season. The <u>bark constitutive</u> MT emission from <u>bark of</u> healthy <u>spruce Norway spruce</u> trees accounted for 8 % of the total emission rates from bark and needles. However, if there <u>was were</u> an ongoing infestation from bark beetles, the bark emission rates would account for 95 % of the total emission rates during the early season, and 85 % during the late season. When comparing with the seasonal average of the emission rates from healthy trees, spruce bark beetle infestation could The infestation would lead to a 6- to 20-fold increase in the total emission rates from bark and needles <u>by 550 %</u> to 1,900 %, depending on the time in the season, when comparing with the seasonal average of the constitutive emission rates from healthy trees.

When a tree is infested, the emission rates increase significantly which can cause large local effects both for tree health but also and SOA production. The BVOC emission increase can also cause more widespread effects, -if the outbreaks are sustained at high levels; there would be large impacts regionally. During 2020 in Sweden, 8 million m³ forest was affected by spruce bark beetles (Wulff and Roberge, 2020). This represents about 0.7 % of the total volume of Norway spruce trees with a diameter larger than 15 cm in Sweden (Skogsstyrelsen, n.d.). Using the seasonal average from the early season and late season of the bark beetle infested emission rates of MT found in this study and the needle emission rates from van Meeningen et al. (2017), the infested trees during 2020 would contribute to an increase of about 4 to 13 % of total MT emission rate from Norway spruce trees in all of Sweden, including emissions from canopy and stem. The effects from insect herbivory and specifically spruce bark beetles might thus be underestimated both in emission and vegetation models (MEGAN, LPJ-GUESS; Guenther et al., 2006; Schurgers et al., 2009) and atmospheric chemistry models estimating BVOC impacts on oxidation capacity and SOA formation (ADCHEM; Roldin et al., 2011). Evident by the difference in emission rates during the early or late season, it is also important to consider the influence of time after infestation when modelling emission rates of BVOCs from infested Norway spruce to get a correct estimation of the spruce bark beetle impact.

5 Conclusion

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Norway spruce trees are emitting BVOCs from the bark as a stress response to spruce bark beetles, and as the number of spruce bark beetle outbreaks increase; it will impact the total emission of BVOCs. The aim of theour study was to examine how spruce bark beetles affect the BVOC emission rates from Norway spruce bark by looking at the difference between healthy and infested trees, the time passed after since infestation start and the difference in emissions from different bark beetle drilled holes types. We also wanted One aim was also to provide an insight into how the BVOC emissions change from non-infested to infested, and following the infestation over time.-TheOur results study shows that there is a significant difference in BVOC emission rates from healthy spruce bark and infested spruce Norway spruce bark, but also a relationship between BVOC emissions from infested trees and the time passed since after infestation start, which can be supported by theour results indicating that indicated a difference in emissions from bark beetle drilled entry holes and exit holes entry and exit holes. We also saw that The initiation of a second generation of bark beetles, which can lead to late summer attacks, can potentially have prolonged impacts on the BVOC emissions as we found emission rates were found to be induced until the start of the next season. When the tree was infested again, the emission rates wereas further induced to reach the same levels as the induced emissions of a tree that was healthy before infestation. As the infestation proceeded, there was we saw a difference in the emission rate and compound blend when comparing the initially stressed tree with the initially healthy tree, where the emission rates were induced to high levels until August for the initially healthy tree, but not for the initially stressed tree. Further studies are needed to support the findings and speculations of this study but also to analyze tThe entire impact of spruce bark beetles on Norway spruce trees. would require

further studies, T-and the importance of further such studies is supported by theour findings that the bark beetle induced BVOC emission rates can be considerably higher than previously thought and could potentially increase thelead to a 1.1-fold increase of total MT emissions from Norway spruce in Sweden-with 4 to 13 %. Even further work would be needed in investigating the impact of coupled stress factors. We found a potential link between temperature stress and bark beetle stress was identified in this study, where trees seems to become more sensitive to temperature leading towith a potential to have even potentially higher emission rates when temperatures increase in conjunction with bark beetle infestations. We believe that Based on the findings of this study, bark beetle infestations are believed town have higher impacts on the atmosphere and climate change than previously thought and samples from more trees and more frequently throughout the season are is needed in order to fully understand the impact.

Appendix A

Table A1. Seasonal average temperature standardized emission rate (μg m⁻² h⁻¹ ± one standard deviation) from all Norway spruce trees located in Hyltemossa and Norunda. Presented areis the all the identified compounds, compound groups (Monoterpenes, sesquiterpenes and other BVOCs) and total emission emitted from healthy and infested (early and late season) Norway spruce bark. The increase or decrease (%) is presented for the infested trees as a change in emission rate from healthy to infested. The occurrence (%) is presented to indicate how much indicates how often each compound appeared in the samples. The compounds that were identified but unable to quantify is presented as n.q. (no quantification). All identified compounds from all samples throughout the season separated into healthy trees and infested trees. Presented is the seasonal average emission rate (μg m⁻² h⁻¹ ± one standard deviation) for each compound and the groups of MT, SQT and other BVOCs and the occurrence (%) of each compounds for all samples. The increase (%) is presented for the infested trees as an increase from healthy to infested. The compounds that where identified but unable to quantify is presented as n.q. (no quantification).

-	Heal	lth <u>y</u>	Infes	ted early se	eason	Infe	sted late se	ason_
Compound name	<u>average ±</u> <u>std</u> (μg m ⁻² h ⁻¹)	occurrence (%)	$\frac{average \pm}{std}$ $(\mu g m^{-2} h^{-1})$	increase (%)	occurrence (%)	$\frac{average \pm}{std}$ $(\mu g m^{-2} h^{-1})$	increase (%)	occurrence (%)
Monoterpenes	<u>29 ± 51</u>	-	6,600 ± 6,700	22,400	-	1,900 ± 1,300	6,500	-
<u>α-pinene</u>	<u>12 ± 20</u>	<u>76</u>	910 ± 1030	<u>7,800</u>	<u>100</u>	820 ± 890	7,100	<u>100</u>
<u>β-pinene</u>	<u>8 ± 19</u>	<u>56</u>	950 ± 960	11,500	<u>100</u>	230 ± 170	<u>2,600</u>	<u>100</u>
<u>3-Carene</u>	<u>3 ± 5</u>	<u>49</u>	290 ± 420	11,400	<u>100</u>	30 ± 40	<u>1,200</u>	<u>95</u>
<u>Limonene</u>	<u>2 ± 3</u>	<u>44</u>	320 ± 320	<u>16,900</u>	<u>88</u>	90 ± 80	<u>4,400</u>	<u>100</u>
<u>p-Cymene</u>	<u>1 ± 1</u>	<u>40</u>	240 ± 320	49,200	<u>63</u>	50 ± 40	10,700	<u>77</u>
<u>β-Myrcene</u>	0.3 ± 0.8	<u>18</u>	160 ± 170	49,800	<u>79</u>	<u>10 ± 6</u>	<u>1,900</u>	<u>86</u>
<u>β-Phellandrene</u>	<u>3 ± 8</u>	<u>11</u>	670 ± 660	24,800	<u>44</u>	<u>190 ± 160</u>	<u>6,900</u>	<u>68</u>
(1S)-Camphene	<u>2 ± 6</u>	<u>6</u>	1,520 ± 1,970	89,100	<u>75</u>	390 ± 230	22,800	<u>100</u>
2- Cyclopentylcyclo pentanone	<u>n.q-</u>	<u>4</u>	=	Ξ	=	<u>n.q</u>	Ξ	<u>5</u>
<u>α-Terpineol</u>	<u>n.q-</u>	<u>3</u>	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ.
5-Ethyl-m- xylene	<u>n.q-</u>	<u>1</u>	=	Ξ	Ξ	=	Ξ	=
(+)-Sabinene	0.1 ± 0	<u>1</u>	210 ± 200	257,900	<u>44</u>	<u>3 ± 0</u>	3,600	<u>5</u>
(1R)-Camphene	<u>=</u>	Ξ	190 ± 740	<u>=</u>	<u>19</u>	<u>110 ± 80</u>	<u>=</u>	<u>77</u>
Tricyclene	Ξ	Ξ	170 ± 250	<u>=</u>	<u>50</u>	<u>20 ± 20</u>	Ξ	<u>36</u>
<u>Eucalyptol</u>	Ξ	Ξ	10 ± 20	<u>=</u>	<u>44</u>	<u>2 ± 5</u>	Ξ	<u>27</u>
(+)-Camphor	Ξ	Ξ	<u>=</u>	<u>=</u>	Ξ	<u>n.q-</u>	<u>=</u>	<u>46</u>
<u>Pinocarvone</u>	Ξ	Ξ	<u>n.q</u>	Ξ	<u>44</u>	<u>n.q-</u>	Ξ	<u>5</u>
<u>4-Carene</u>	Ξ	Ξ	350 ± 280	Ξ	<u>38</u>	Ξ	Ξ	Ξ
<u>ζ-Fenchene</u>	Ξ	Ξ	120 ± 190	Ξ	<u>25</u>	<u>1 ± 0</u>	Ξ	<u>5</u>
<u>α-Phellandrene</u>	Ξ	Ξ	110 ± 200	Ξ	<u>31</u>	Ξ	Ξ	Ξ
(1R)-(-)- Myrtenal	=	Ξ	<u>n.q</u>	Ξ	<u>31</u>	Ξ	Ξ	=
(4E,6E)- Allocimene	Ξ	Ξ	50 ± 80	Ξ	<u>25</u>	=	Ξ	=
5-Vinyl-m- xylene	Ξ	Ξ	<u>n.q</u>	Ξ	<u>25</u>	=	Ξ	=
<u>3-Pinanone</u>	Ξ	Ξ	Ξ	Ξ	Ξ	<u>n.q</u>	Ξ	<u>14</u>
Norbornane	Ξ	Ξ	60 ± 80	Ξ	<u>19</u>	Ξ	Ξ	Ξ
<u>γ-Terpinene</u>	Ξ	Ξ	90 ± 0	Ξ	<u>13</u>	Ξ	Ξ	Ξ
<u>α-Fenchene</u>	Ξ	Ξ	10 ± 0	Ξ	<u>13</u>	Ξ	Ξ	Ξ
2-Carene	Ξ	Ξ	160 ± 0	Ξ	<u>13</u>	=	Ξ	Ξ

<u>α-Thujene</u>			<u>20 ± 0</u>		<u>13</u>			_ 1
Verbenone	<u>-</u>		<u>n.q-</u>	<u> </u>	13 13	=	Ξ.	_
Myrtenal	-	=	<u>n.q-</u>	_ _	<u>15</u> <u>6</u>	_	=	=
α-Terpinene	<u>-</u>	=	30 ± 0	_ _	<u>o</u> 6	_	=	=
Sesquiterpenes	2.1 ± 3.2		$\frac{30 \pm 0}{53 \pm 74}$	2,400		- 18 ± 24	<u>=</u> <u>700</u>	Ξ
Longifolene+β-		_			-			-
<u>Caryophyllene</u>	0.7 ± 1.4	<u>18</u>	38 ± 70	<u>5,300</u>	<u>56</u>	14 ± 24	<u>1,800</u>	<u>55</u>
<u>α-Humulene</u>	<u>1.4 ± 3</u>	<u>48</u>	<u>5 ± 9</u>	<u>200</u>	<u>31</u>	<u>4 ± 13</u>	<u>200</u>	<u>18</u>
Germacrene D	Ξ	Ξ	<u>4 ± 0</u>	Ξ	<u>19</u>	Ξ	Ξ	Ξ
<u>Isoledene</u>	Ξ	Ξ	<u>3 ± 0</u>	Ξ	<u>19</u>	Ξ	Ξ	Ξ
<u>β-Cubebene</u>	Ξ.	Ξ	3 ± 0	Ξ	<u>19</u>	Ξ	Ξ	Ξ
Other BVOCs	0.4 ± 0.9	_	3.4 ± 6.7		_	0.1 ± 0.2	_	_
<u>Isoprene</u>	0.4 ± 0.9	<u>58</u>	<u>3 ± 7</u>	<u>700</u>	<u>63</u>	<u>0.1 ± 0.2</u>	<u>-65</u>	<u>27</u>
<u>Decanal</u>	<u>n.q-</u>	<u>45</u>	<u>n.q</u>	Ξ	<u>13</u>	<u>n.q-</u>	<u>=</u>	<u>14</u>
Benzene	<u>n.q-</u>	<u>45</u>	<u>=</u>	Ξ	Ξ	<u>n.q-</u>	<u>=</u>	<u>14</u>
Nonanal	<u>n.q-</u>	<u>39</u>	<u>n.q-</u>	Ξ	<u>6</u>	<u>n.q-</u>	Ξ	<u>14</u>
<u>Toluene</u>	<u>n.q-</u>	<u>21</u>	<u>n.q-</u>	Ξ	<u>6</u>	<u>n.q-</u>	Ξ	<u>9</u>
1,3,5-	<u>n.q-</u>	<u>14</u>	=	Ξ	Ξ	Ξ	<u>=</u>	Ξ
<u>Trifluorobenzene</u> <u>Benzaldehyde</u>	<u>n.q-</u>	<u>12</u>				<u>n.q-</u>	_	<u>9</u>
Butyl formate	<u>n.q-</u>	<u>8</u>	=	=	=	<u>n.q-</u>		<u>5</u>
<u>Caprolactam</u>	<u>n.q-</u>	<u>5</u> 7	_	_	=	_	_	
Cyclopentanone	<u>n.q-</u>	<u>5</u>	=	_	= =	<u>-</u> <u>n.q-</u>	_	<u>-</u> <u>9</u>
Methanesulfonic	<u>n.q-</u>		Ξ	Ξ		<u>n.q-</u>	_	
<u>anhydride</u>		<u>5</u>	Ξ	Ξ	Ξ		=	<u>5</u>
Trimethylbenzol	<u>n.q-</u>	<u>2</u>	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ.
<u>m-Xylene</u>	<u>n.q-</u>	<u>2</u>	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ
<u>Ethylhexanol</u>	<u>n.q-</u>	<u>2</u>	=	Ξ	Ξ	Ξ	Ξ	Ξ
Acetic acid	<u>n.q-</u>	<u>2</u>	=	Ξ	Ξ	Ξ	Ξ	Ξ
tert-Butylamine	<u>n.q-</u>	<u>1</u>	=	Ξ	Ξ	<u>n.q</u>	Ξ	9
<u>m-Ethyltoluene</u>	<u>n.q-</u>	<u>1</u>	=	Ξ	Ξ	Ξ	Ξ	Ξ
o-Ethyltoluene	<u>n.q-</u>	<u>1</u>	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ
Methyl 3- hydroxy-2,2-	<u>n.q-</u>							
dimethylpropano		<u>1</u>	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ
ate	<u>n.q-</u>	1						
1-Pentene Butanal	<u>n.q-</u>	1	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ.
1-Nonene	<u>n.q-</u>	<u>1</u>	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ
Isobutenyl	<u>n.q-</u>	<u>1</u>	Ξ	Ξ	=	Ξ	Ξ	Ξ.
methyl ketone	<u> </u>	<u>1</u>	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ
<u>Diacetone</u> <u>alcohol</u>	<u>n.q-</u>	<u>1</u>	Ξ	Ξ	<u>=</u>	Ξ	Ξ	Ξ
Furfural	<u>n.q-</u>	<u>1</u>	Ξ.	Ξ	<u>=</u>	<u>=</u>	_	Ξ
1,6-Anhydro-β-	<u>n.q-</u>						_	
d-talopyranose		<u>1</u>	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ
dl-3,4- Dehydroproline	<u>n.q-</u>	<u>1</u>	Ξ.	Ξ	<u>=</u>	Ξ	<u>=</u>	Ξ
methyl ester		_	_	-	_	_	_	_
6,10,14- Trimethyl-2-	<u>n.q-</u>	<u>1</u>	_	_	Ξ	_	_	Ξ
<u>pentadecanone</u>		<u> </u>	Ξ	Ξ	_	Ξ	_	-
Undecanal	<u>n.q-</u>	<u>1</u>	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ
Carbon disulfide	<u>n.q-</u>	<u>1</u>	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ
2-Methyl-1- phenylpropene	Ξ	Ξ	<u>n.q-</u>	Ξ	<u>38</u>	<u>n.q</u>	Ξ	<u>9</u>
2-Methyl-3-			<u>n.q-</u>		10			
buten-2-ol	Ξ	Ξ		Ξ	<u>19</u>	Ξ.	Ξ	Ξ

Benzoic acid	<u>=</u>	<u>=</u>	<u>=</u>	Ξ	Ξ	<u>n.q-</u>	Ξ	9
<u>Acetophenone</u>	Ξ	Ξ	Ξ	Ξ	Ξ	<u>n.q-</u>	Ξ	<u>9</u>
Methyl acetate	Ξ	Ξ	Ξ	Ξ	Ξ	<u>n.q-</u>	Ξ	9
(-)-Bornyl acetate	=	Ξ	<u>n.q</u>	Ξ	<u>13</u>	=	Ξ	Ξ
Bornyl acetate	Ξ	Ξ	<u>n.q</u>	Ξ	<u>13</u>	Ξ	Ξ	Ξ

=	<u>Health</u>	<u> </u>	Infested	early sea	son	Infested	late seas	ion
Compound name	average ± std	<u>occurre</u>	average ± std	<u>increa</u>	<u>occurre</u>	average ± std	<u>increa</u>	<u>occurre</u>
	$\frac{(\mu g m^{-2} h^{-1})}{(\mu g m^{-2} h^{-1})}$	<u>nce (%)</u>	$\frac{(\mu g m^2 h^4)}{(\mu g m^2 h^2)}$	<u>se (%)</u>	<u>nce (%)</u>	$\frac{(\mu g m^{-2} h^{-1})}{(\mu g m^{-2} h^{-1})}$	<u>se (%)</u>	<u>nce (%)</u>
Monoterpenes	29 ± 51	=	$6,600 \pm 6,700$	22,400	Ξ	1,900 ± 1,300	6,500	=
α-pinene	<u>12</u>	76	910	7,800	100	820	7,100	100
<u>β pinene</u>	<u>&</u>	56	<u>950</u>	<u>11,500</u>	100	230	2,600	100
3-Carene	<u>3</u>	<u>49</u>	290	<u>11,400</u>	100	30	1,200	<u>95</u>
<u>Limonene</u>	<u>2</u>	<u>44</u>	<u>320</u>	16,900	<u>88</u>	90	<u>4,400</u>	100
p-Cymene	<u>±</u>	<u>40</u>	<u>240</u>	<u>49,200</u>	<u>63</u>	<u>50</u>	<u>10,700</u>	77
<u>β-Myrcene</u>	<u>0</u>	<u>18</u>	160	<u>49,800</u>	79	10	1,900	86
<u>β-Phellandrene</u>	<u>3</u>	11	<u>670</u>	24,800	<u>44</u>	190	6,900	68
(1S) Camphene	<u>2</u>	<u>6</u>	<u>1,520</u>	89,100	<u>75</u>	<u>390</u>	22,800	<u>100</u>
2- Cyclopentylcyclopentan one	Ξ	<u>4</u>	Ξ	Ξ	Ξ	Ξ	Ξ	<u>5</u>
α-Terpineol	Ξ	<u>3</u>	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ
5-Ethyl-m-xylene	=	<u>±</u>	=	=	Ξ	=	=	=
(+)-Sabinene	<u> </u>	<u>±</u>	<u>210</u>	257,90 0	<u>44</u>	<u>3</u>	<u>3,600</u>	<u>5</u>
(1R) Camphene	=	Ξ	<u>190</u>	=	<u>19</u>	<u>110</u>	=	77
<u>Tricyclene</u>	=	=	170	=	<u>50</u>	<u>20</u>	=	<u>36</u>
Eucalyptol	=	Ξ	<u>10</u>	Ξ	<u>44</u>	<u>2</u>	Ξ	27
(+) Camphor	=	Ξ	=	=	Ξ	=	=	<u>46</u>
<u>Pinocarvone</u>	=	=	=	=	<u>44</u>	=	=	<u>5</u>
4-Carene	=	=	<u>350</u>	=	<u>38</u>	=	=	=
<u>ζ-Fenchene</u>	=	Ξ	<u>120</u>	=	<u>25</u>	<u>1</u>	=	<u>5</u>
<u>α-Phellandrene</u>	=	=	<u>110</u>	=	<u>31</u>	=	=	=
(1R) (-) Myrtenal	=	Ξ	=	Ξ	<u>31</u>	=	Ξ	Ξ
(4E,6E)-Allocimene	=	Ξ	<u>50</u>	Ξ	<u>25</u>	=	Ξ	Ξ
5 Vinyl m xylene	=	Ξ	=	=	<u>25</u>	=	=	=
<u>3-Pinanone</u>	=	Ξ	=	=	=	=	=	<u>14</u>
Norbornane	=	Ξ	<u>60</u>	Ξ	<u>19</u>	=	Ξ	Ξ
<u>y Terpinene</u>	=	Ξ	<u>90</u>	=	<u>13</u>	=	=	=
<u>α-Fenchene</u>	=	Ξ	<u>10</u>	=	<u>13</u>	=	=	=
2-Carene	Ξ	Ξ	<u>160</u>	Ξ	<u>13</u>	Ξ	Ξ	Ξ
<u>α-Thujene</u>	=	Ξ	<u>20</u>	Ξ	<u>13</u>	=	Ξ	Ξ
<u>Verbenone</u>	=	=	=	=	<u>13</u>	=	=	=
<u>Myrtenal</u>	Ξ	Ξ	Ξ	Ξ	<u>6</u>	Ξ	Ξ	Ξ
<u>α-Terpinene</u>	П	Ξ	<u>30</u>	Ξ	<u>6</u>	П	Ξ	=
Sesquiterpenes	$\frac{2.1 \pm 3.2}{}$	=	53 ± 74	2,400	=	18 ± 24	700	=
Longifolene+β	<u>±</u>	<u>18</u>	<u>38</u>	5,300	<u>56</u>	<u>14</u>	1,800	<u>55</u>
Caryophyllene α Humulene	<u>+</u>	<u>48</u>	<u></u> <u>5</u>	200	31	<u>—</u> <u>5</u>	200	18
Germacrene D			<u>≠</u> <u>4</u>		21 19			
Isoledene	=	=	<u>4</u> <u>3</u>	Ξ	19 <u>19</u>	=	Ξ	=
	Ξ	Ξ		Ξ		Ξ	Ξ	=
<u>β-Cubebene</u>	=	Ξ	<u>3</u>	=	<u>19</u>	=	=	Ξ.

Other BVOCs	0.4 ± 0.9	=	3.4 ± 6.7	-	Ξ.	0.1 ± 0.2	=	=
<u>Isoprene</u>	<u>0</u>	<u>58</u>	<u>3</u>	700	<u>63</u>	<u>0</u>	<u>-65</u>	27
Decanal	Ξ	<u>45</u>	=	Ξ	<u>13</u>	=	Ξ	<u>14</u>
Benzene	Ξ	<u>45</u>	=	=	Ξ	=	=	<u>14</u>
Nonanal Nonanal	=	<u>39</u>	=	=	<u>6</u>	=	=	<u>14</u>
Toluene	=	21	=	Ξ	<u>6</u>	=	Ξ	<u>9</u>
1,3,5 Trifluorobenzene	=	<u>14</u>	=	=	Ξ	=	=	Ξ
Benzaldehyde	Ξ	<u>12</u>	=	=	=	=	=	<u>9</u>
Butyl formate	Ξ	<u>8</u>	=	Ξ	Ξ	=	Ξ	<u>5</u>
<u>Caprolactam</u>	Ξ	7	=	Ξ	Ξ	=	Ξ	Ξ
<u>Cyclopentanone</u>	=	<u>5</u>	=	=	=	=	=	<u>9</u>
Methanesulfonic anhydride	Ξ	<u>5</u>	Ξ	=	=	=	=	<u>5</u>
<u>Trimethylbenzol</u>	=	<u>2</u>	=	=	=	=	=	=
m-Xylene	Ξ	<u>2</u>	=	Ξ	Ξ	=	Ξ	Ξ
Ethylhexanol	Ξ	<u>2</u>	=	=	Ξ	=	=	Ξ
Acetic acid	=	<u>2</u>	=	Ξ	=	=	Ξ	=
tert-Butylamine	Ξ	<u>±</u>	=	Ξ	Ξ	=	Ξ	<u>9</u>
m-Ethyltoluene	=	±	=	=	Ξ	=	Ξ	Ξ
o-Ethyltoluene	Ξ	<u>±</u>	=	=	=	=	=	Ξ
Methyl 3 hydroxy 2,2 dimethylpropanoate	Ξ	<u>1</u>	=	=	Ξ	=	=	Ξ
<u>1 Pentene</u>	=	<u>±</u>	=	=	=	=	=	=
Butanal	=	<u>±</u>	=	Ξ	Ξ	=	Ξ	Ξ
1-Nonene	=	±	=	=	Ξ	=	Ξ	Ξ
<u>Isobutenyl methyl</u> <u>ketone</u>	Ξ	<u>±</u>	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ
Diacetone alcohol	=	<u>1</u>	=	=	Ξ	=	=	=
<u>Furfural</u>	=	<u>±</u>	Ξ	Ξ	Ξ	=	Ξ	Ξ
1,6 Anhydro β d talopyranose	=	<u>±</u>	Ξ	=	Ξ	=	=	Ξ
dl 3,4 Dehydroproline methyl ester	=	<u>±</u>	Ξ	=	Ξ	=	=	=
6,10,14-Trimethyl-2- pentadecanone	Ξ	<u>±</u>	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ
<u>Undecanal</u>	Ξ	<u>1</u>	=	=	Ξ	<u> </u>	=	Ξ
Carbon disulfide	=	<u>1</u>	_	=	=	<u>-</u>	=	=
2-Methyl-1-	=	=	=	_	<u>38</u>	=		<u>9</u>
phenylpropene			_	_			=	
2-Methyl-3-buten-2-ol	=	Ξ	Ξ	=	<u>19</u>	Ξ	=	=
Benzoic acid	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ	Ξ	<u>9</u>
<u>Acetophenone</u>	Ξ	Ξ	=	Ξ	=	=	=	<u>9</u>
Methyl acetate	Ξ	Ξ	=	Ξ	=	=	=	<u>9</u>
(-)-Bornyl acetate	Ξ	Ξ	Ξ	Ξ	13	Ξ	Ξ	Ξ
Bornyl acetate	Ξ	=	=	=	<u>13</u>	=	=	=
Total	$\frac{32 \pm 52}{1}$	=	$6,700 \pm 6,900$	20,900	Ξ	$2,000 \pm 1,300$	<u>6,000</u>	=

-	Healthy		Infested early season			Infested late season		
Compound name	$\frac{average \pm std}{(\mu g \ m^{-2} \ h^{-1})}$	occurre nce (%)	$\frac{average \pm std}{(\mu g \ m^{-2} \ h^{-1})}$	increa se (%)	occurre nce (%)	$\frac{average \pm std}{(\mu g \ m^{-2} \ h^{-1})}$	increa se (%)	occurre nce (%)
Monoterpenes	29.37 ± 51.01	-	6690 ± 6860	22678. 35	-	1970 ± 1310	6607.5 2	-
alpha-Pinene	11.49	76.11	911.14	7829.8 5	100.00	824.64	7077.0 2	100.00

	I		I	11507.		I	2640.6	
beta-Pinene	8.22	55.75	954.17	91	100.00	225.28	3	100.00
3 Carene	2.48	48.67	285.16	11398. 39	100.00	33.07	1233.4 7	95.45
Limonene	1.89	44.25	320.82	16874. 60	87.70	85.43	4420.1 1	100.00
p-Cymene	0.49	39.82	241.45	49175. 51	62.50	53	10716. 33	77.27
beta Myrcene	0.32	17.70	159.76	4 9825. 00	78.95	6.3	1868.7 5	86.36
beta-Phellandrene	2.7	10.62	673.13	24830. 74	43.75	189.47	6917.4 1	68.18
(1S) Camphene	1.7	6.19	1516	89076. 4 7	75.00	388.82	22771. 76	100.00
2- Cyclopentylcyclopenta	n.q.	4.40	-	_	_	n.q.	_	4.54
none	n a	4.42			_			
alpha Terpineol	n.q.	2.65	_	_	_	_	_	_
5-Ethyl-m-xylene	n.q.	0.88		- 25786	<u>-</u>	_	- 3562.5	<u>-</u>
(+) Sabinene	0.08	0.88	206.37	2.50	43.75	2.93	0	4.54
(1R) Camphene	-	-	186.16	-	18.75	112.32	-	77.27
Tricyclene	_	-	173.42	_	50.00	24.52	-	36.36
Eucalyptol	-	-	10.25	-	43.75	2.32	-	27.27
(+) Camphor	-	_	_	-	-	n.q.	_	45.45
Pinocarvone	-	-	n.q.	-	43.75	n.q.	-	4.54
4-Carene	-	-	348.15	_	37.50	-	-	_
zeta Fenchene	_	_	116.96	_	25.00	0.92	_	4.54
alpha Phellandrene	_	_	114.43	_	31.25	_	_	_
(1R) () Myrtenal	_	_	n.q.	_	31.25	_	_	_
trans,trans-Allocimene	_	_	50.9	_	25.00	_	_	_
5 Vinyl m xylene	_	_	n.q.	_	25.00	_	_	_
3 Pinanone	_	_	_	_	_	n.q.	_	13.63
Norbornane	_	_	60.27	_	18.75	_	_	_
gamma Terpinene	_	_	88.84	_	12.50	_	_	_
alpha Fenchene	_	_	14.07	_	12.50	_	_	_
2-Carene	_	_	156.65	_	12.50	_	_	_
alpha Thujene	_	_	15.83	_	12.50	_	_	_
Verbenone	_	_	n.q.	_	12.50	_	_	_
Myrtenal	_	_	n.q.	_	6.25	_	_	_
	_	_	26.06	_	6.25	_	_	_
alpha-Terpinene				2400.0	0.20	10.01	- 40.04	
Sesquiterpenes	2.12 ± 3.17	_	53.0 ± 74	0	_	18 ± 24	749.06	
Longifolene+beta- Caryophyllene	0.7	17.70	37.65	5278.5 7	56.25	13.6	1842.8 6	54.54
alpha Humulene	1.42	47.79	4.86	242.25	31.25	4.52	218.31	18.18
Germacrene D	-	-	3.86	-	18.75	-	-	-
Isoledene	-	-	3.25	-	18.75	-	-	-
beta-Cubebene		<u> </u>	3.2		18.75			<u> </u>
Other BVOCs	0.40 ± 0.85	-	3.36 ± 6.69	740.00	-	0.14 ± 0.20	-65.00	-
Isoprene	0.4	58.41	3.36	740.00	62.50	0.14	-65.00	27.27
Decanal	n.q.	45.13	n.q.	-	12.50	n.q.	-	13.63
Benzene	n.q.	45.13	_	_	_	n.q.	-	13.63
Nonanal	n.q.	38.94	n.q.	_	6.25	n.q.	_	13.63
Toluene		21.24		_	6.25		_	9.09
	II.Q.	21.27	11.q.		0.23	11.4.	_	9.09
1,3,5 Trifluorobenzene	n.q. n.q.	14.16	n.q. -	_	0.23 -	n.q. -	_	9.09 -

	i		1		1	1		
Benzaldehyde	n.q.	11.50	-	-	_	n.q.	-	9.09
Butyl formate	n.q.	7.96	-	-	_	n.q.	-	4.54
Caprolactam	n.q.	7.08	-	-	_	-	-	_
Cyclopentanone Methanesulfonic	n.q.	5.31	-	-	-	n.q.	-	9.09
anhydride	n.q.	5.31	-	-	-	n.q.	-	4.54
Trimethylbenzol	n.q.	1.77	-	-	_	-	-	_
m-Xylene	n.q.	1.77	-	-	_	-	-	_
Ethylhexanol	n.q.	1.77	-	-	_	-	-	-
Acetic acid	n.q.	1.77	-	-	-	-	-	_
tert-Butylamine	n.q.	0.88	-	-	-	n.q.	-	9.09
m-Ethyltoluene	n.q.	0.88	-	_	-	-	-	-
o Ethyltoluene Methyl 3 hydroxy 2,2	n.q.	0.88	-	-	-	-	-	_
dimethylpropanoate	n.q.	0.88	-	-	-	-	-	-
1 Pentene	n.q.	0.88	-	-	-	-	-	-
Butanal	n.q.	0.88	-	-	-	-	-	-
1 Nonene Isobutenyl methyl	n.q.	0.88	-	-	_	-	-	-
ketone	n.q.	0.88	-	-	-	-	-	-
Diacetone alcohol	n.q.	0.88	-	-	-	-	-	-
Furfural 1,6 Anhydro beta d	n.q.	0.88	-	-	-	-	-	-
talopyranose dl-3,4-Dehydroproline	n.q.	0.88	-	-	-	-	-	_
methyl ester 6,10,14 Trimethyl 2	n.q.	0.88	-	-	-	-	-	-
pentadecanone	n.q.	0.88	-	-	-	-	-	-
Undecanal	n.q.	0.88	-	-	-	-		-
Carbon disulfide 2-Methyl-1-	n.q.	0.88	-	-	-	-	-	-
phenylpropene alpha,alpha-	-	-	n.q.	-	37.50	n.q.	-	9.09
Dimethylallyl alcohol	-	-	n.q.	-	18.75	-	-	-
Benzoic acid	-	-	-	-	-	n.q.	-	9.09
Acetophenone	-	-	-	-	-	n.q.	-	9.09
Methyl acetate	-	-	-	-	-	n.q.	-	9.09
(-)-Bornyl acetate	-	-	n.q.	-	12.50	-	-	-
Bornyl acetate	-	_	n.q.	-	12.50	i	-	_

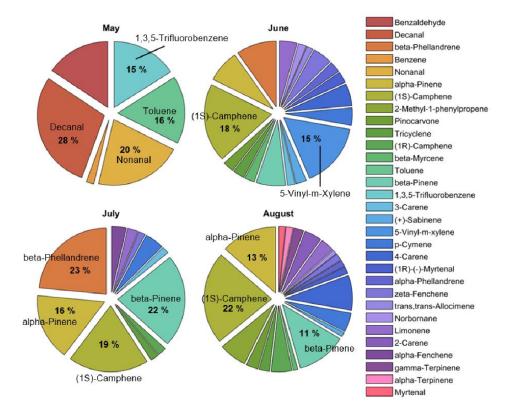


Figure A1: The daily average blend from the spruceNorway spruce S3S3 and percentage contribution in mass throughout the summer (May, June, July and August), showing only compounds with a mass contribution of at least 1 %.

Table A2. The difference between healthy and infested trees when applying the calculations for the Q₁₀ temperature dependency. The reference emission rate at 30°C (F₀ is the reference emission rate standardized) entails higher emission rates at 30°C with a higher number. The Q₁₀ coefficient indicates the emission rate change for every 10°C temperature difference and is therefore a measure of temperature sensitivity. to temperature, leading to higher emission rates at higher temperatures when the coefficient is larger.

	F ₀ (μg m ⁻² h ⁻¹)	Q10		
Compound name	Healthy Infested H		Healthy	Infested	
Monoterpenes					
beta-Pinene β-pinene	<u>11</u> 11.0	<u>34,900</u> 34901.4	<u>8</u> 7.8	<u>980</u> 981.6	
(1R)-Camphene	<u></u>	<u>1,500</u> 1503.0	==	<u>80</u> 80.2	
beta β-Phellandrene	<u>21</u> 20.6	<u>1,240</u> 1240.0	<u>3</u> 2.8	<u>38</u> 37.8	
alpha-Pinene α-pinene	<u>55</u> 55.0	<u>880</u> 8 79.5	<u>19</u> 18.7	<u>19</u> 18.8	
(1S)-Camphene	<u>93</u> 92.6	<u>470</u> 4 70.1	<u>6</u> 6.3	<u>1414.3</u>	
beta β-Myrcene	<u>16</u> 15.8	<u>250</u> 248.2	<u>57</u> 56.7	<u>170</u> 167.6	
Limonene	<u>13</u> 12.7	<u>120</u> 116.4	<u>12</u> 11.6	<u>1716.9</u>	
3-Carene	<u>5</u> 4.6	<u>110</u> 111.4	<u>76.6</u>	<u>2625.8</u>	
p-Cymene	<u>16</u> 15.7	<u>90</u> 8 6.3	<u>22</u> 22.4	<u>15</u> 14.6	
Tricyclene	=	<u>80</u> 76.3	=	<u>1413.6</u>	
(+)-Sabinene	=	<u>70</u> 66.3	=	<u>87.5</u>	
Eucalyptol	=	<u>3</u> 2.5	=	<u>3</u> 3.5	
Sesquiterpenes					
Longifolene+ beta β-Caryophyllene	0.010.01	<u>24</u> 23.6	<u>0.1</u> 0.1	<u>33</u> 33.1	
alpha α-Humulene	<u>0.01</u> 0.01	<u>0.5</u> 0.5	<u>0.7</u> 0.7	<u>1</u> 1.3	

1115

Isoprene	<u>1</u> 1.3	<u>2</u> 2.4	<u>1010.2</u>	<u>2424.4</u>	
130010110	_	_			

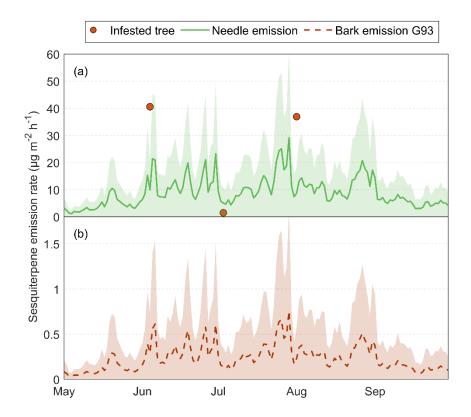


Figure A2. The measured and calculated BVOC emission rates for the group sesquiterpenes from Norway spruce in

Hyltemossa: (a) the actual measured emission rates from infested Norway spruce bark (red dot) and calculated needle
emission rates (green line) and (b) calculated healthy bark emission. The needle emissions are calculated based on the
Guenther algorithm (Guenther et al., 1993) and the measured emission rates are taken from van Meeningen et al.,
(2017) and specific needle area (SLA) was taken from Wang et al., (2017), using the air temperature at 24 m taken from
the HTM ICOS station (Heliasz, 2020). The healthy bark emission is calculated with the Guenther algorithm (orange)
based on the tree temperature taken at 3 m height in the North and East orientation using data taken from the HTM
ICOS station (Heliasz, 2020). The shaded areas (green, orange and black shade) represent the standard deviation from
the mean for the respective calculation method.

The modelled calculated emission rates for the group sesquiterpenes from (b) constitutive healthy bark VOC BVOC emission rates from the group sesquiterpenes based on the tree temperature taken at 3 meters height in the northNorth and eastEast orientation, data taken from the HTM ICOS station (Heliasz, 2020). The bark emission (black) is calculated based on the Guenther algorithm (Guenther et al., 1993) based on measured emission rates in this study. The (a) leafneedle emission rates (green) are calculated based on the Guenther algorithm and the measured emission rates are taken from van Meeningen et al., (2017) and specific leafneedle area (SLA) was taken from Wang et al., (2017) using the air temperature at 24 meters taken from the HTM ICOS station (Heliasz, 2020). For comparison, the actual measured bark VOC BVOC emission rates from one infested tree over time, from this study, is included in the Figure (red-dot).

1120 Author contribution

EJ and TH designed and planned the campaigns. EJ performed the measurements. EJ performed the data analysis with contributions from KL, AG and AMJ. Funding was acquired by TH. EJ prepared the manuscript draft with contributions from all co-authors.

Competing interests

The authors declare that they have no conflict of interest.

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