



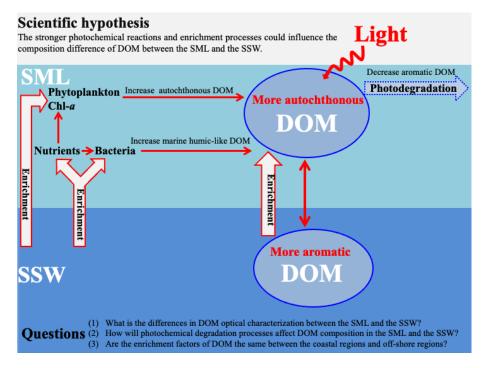
- Spatial-temporal distribution, photoreactivity and environmental
- 2 control of dissolved organic matter in the sea-surface microlayer of
- 3 the eastern marginal seas of China
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13 Graphical Abstract





#### Abstract

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As the boundary interface between the atmosphere and ocean, the sea-surface microlayer (SML) plays a significant role in the biogeochemical cycles of dissolved organic matter (DOM) and macronutrients in marine environments. In our study, chromophoric DOM (CDOM), fluorescent DOM, dissolved organic carbon, chlorophyll a, picoplankton, nutrients, and bacteria were frequently enriched in the SML. We focus specifically on the optical properties in the SML, and we find that the enrichment factors (EFs) of tryptophan-like component 4 was significantly higher than other fluorescence components; the longer wavelength absorption values of CDOM showed higher EFs in the SML, and the more significant relationship between CDOM and Chl-a in the SML, indicating that autochthonous DOM was more frequently enriched in the SML than the terrestrial DOM. We find that higher EFs were generally observed in the SML in the off-shore regions than in the coastal regions, and CDOM in the SML is photobleached less after relatively strong irradiation, as also indicated by the lower percentages of humic-like DOM and lower specific UV absorbance values (SUVA254) in the SML than the subsurface water (SSW). In combination with the SSW, the elevated nutrients may stimulate phytoplankton growth, biological activity and then production of abundant fresh autochthonous DOM in the SML. Our results revealed a general enrichment model and the more autochthonous properties of DOM in the SML than the SSW for exploring the oceanic air-sea layer environment.

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- 33 Keywords: Sea-surface microlayer; Dissolved organic matter; Photochemical degradation;
- 34 Enrichment processes; Eastern marginal seas of China

### 35 1 Introduction



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ocean, which covers about 70% of the Earth's surface. SML is physicochemically distinct from subsurface water (SSW, depth 3 ~ 5 m) and is characteristically enriched with phytoneuston, chlorophyll, particulate carbon, dissolved organic matter (Hardy 1982; Hardy and Apts, 1989), and biogenic organic compounds, such as lipids, proteins, and polysaccharides ((Liss and Duce, 1997; Liss and Duce, 2005). With a total thickness ranging between 1 µm and 1000 µm, the SML remains present in wind speeds of up to 6.6 m s<sup>-1</sup> (Wurl et al., 2011). A variety of processes contribute to the formation of the SML in aquatic systems, these include but are not limited to, scavenging by rising bubbles, atmospheric deposition, dissolved organic matter (DOM) photochemical degradation and transformation, secretion, and biodegradation by organisms living within the microlayer (Neuston), and migration of motile organisms into the SML (Aller et al., 2005; Wotton and Preston, 2005). The role of the microlayer in oceanic emissions is not well understood and fundamental advance in understanding its properties are needed. Because of its unique position at the air-sea interface, the biological and photochemical reactions of DOM in the SML could strongly impact the biogeochemical cycling of biologically important elements, for example, via the conversion of DOM into volatile species such as carbonyl sulfide (OCS), which influence the atmospheric chemistry and climate (Mopper et al., 2002). Air-sea gas exchanges of trace gases (e.g., CO, OCS, dimethylsulfide (DMS), and alkyl nitrates gases) can also be greatly influenced by biological and photochemical reactions at the sea surface (Blough, 1997). Optical measurements of absorbance and fluorescence have been applied to track DOM variability in aquatic ecosystems (McKnight et al., 2001; Zepp et al., 2004; Coble, 2007). The fraction of DOM that absorbs light in the ultraviolet and visible ranges of the electromagnetic spectrum and the

The sea-surface microlayer (SML) is the boundary interface between the atmosphere and the





58 fraction that exhibits a blue fluorescence are known as chromophoric DOM (CDOM) and fluorescence 59 DOM (FDOM), respectively, and their relative compositions can provide information differentiating between autochthonous and allochthonous sources (Coble, 1996; McKnight et al., 2001; Stedmon et 60 al., 2007). Photolysis of DOM promotes the formation of low-molecular-weight compounds, 61 62 increasing the bioavailability of biologically refractory materials and facilitating carbon uptake by 63 microbes (Kieber et al., 1989). Indices based on optical measurements of absorbance and fluorescence 64 are commonly used to track DOM composition and infer DOM processing due to their low analytical 65 cost and high throughput relative to molecular level analyses (Coble, 2007; Fellman et al., 2010; 66 Gabor et al., 2014). Recent studies have mainly focused on using the characteristics of CDOM as 67 indicators of the sources and degradation states of DOM (Massicotte et al., 2017) in the SSW, and its 68 vertical distribution in estuaries and open oceans (Yamashita et al., 2017; Margolin et al., 2018). 69 Even though there are many studies that have documented the enrichment in DOM (e.g. amino 70 acids; carbohydrates) and inorganic nutrients in the SML relative to the SSW (Orellana et al., 2011; 71 Chen et al., 2016), the differences in organic matter composition between the SML and SSW, the 72 different enrichment factors of DOM in the SML between the coastal regions and the off-shore regions, 73 and how do photochemical degradation activities regulate DOM concentration in the SML need more 74 thorough discussion. Here, we investigated the concentration and composition of DOM in the SML 75 relative to the SSW and the responses of DOM to photoexposure. We hypothesized that the 76 photochemical reactions and enrichment processes could influence the composition difference of 77 DOM between the SML and the SSW, and greater solar exposure in the SML than in the SSW would 78 enhance the mineralization of DOM. To test these hypotheses, our study was designed to answer the 79 questions: (1) What is the differences in optical characterization of DOM between the SML and the



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2.2 Sampling



81 off-shore regions? (3) How will photochemical degradation processes affect DOM composition in the 82 SML and the SSW? We, therefore, compared the optical properties of DOM between the SSW and the 83 SML, and EFs of CDOM, FDOM components, dissolved organic carbon (DOC), chlorophyll-a (Chl-a), 84 nutrients, and bacterial abundance from the coastal waters to open ocean in the eastern marginal seas 85 of China (including the East China Sea (ECS) and the Yellow Sea (YS)) during spring of 2017 and 86 2019, summer of 2018, and winter of 2019; discuss how the composition of accumulated DOM was 87 affected by environmental conditions (wind speed and salinity) within the SML; and conducted photoexposure experiments to compare photochemical degradation processes of DOM between the 88 89 SML and the SSW. 90 91 2 Materials and methods 92 2.1 Study Area 93 Five cruises were conducted during the four seasons, specifically, from: 27 March to 15 April 94 2017 (R/V "Dong Fang Hong 2"), 26 June to 19 July 2018 (R/V "Dong Fang Hong 2"), March 2019 95 (R/V "Zheyu No. 2"), and 28 December 2019 to 16 January 2020 (R/V "Dong Fang Hong 3"). The 96 station locations are shown in Fig. S1. In spring, summer, and winter, SML samples were collected in

SSW? (2) Are the enrichment factors (EFs) of DOM the same between the coastal regions and

depth using 24 × 10-L Niskin bottles mounted on a rosette equipped with a

the YS and the ECS, which are shallow seas located almost entirely on the continental shelf in the

We collected 220 paired SML and SSW water samples. SSW samples were collected at 2-5 m

western Pacific Ocean where there is strong interaction between land and sea.





conductivity-temperature-depth (CTD) profiler. The SML samples were collected using the screen sampling technique (Chen et al., 2016; Garrett, 1965) when conditions were calm. A screen sampler with a 1.6 mm mesh of stainless steel wire on a 40 cm × 40 cm stainless steel frame was used. The SML samples were collected in 500 mL brown sample bottles. The screen was held level and dipped into the sea surface, moved laterally in order to sample from an undisturbed film, and then withdrawn slowly from the surface. Repeated dipping was conducted until the desired volume was collected. The screen method is often applied during field studies because of its relatively short sampling time and large sample volume compared to other techniques (Momzikoff et al., 2004; Chen et al., 2016). Immediately after collection, samples were filtered using 0.7 µm glass fiber filters (GF/F, Whatmann) and the filtrates were transferred to 60 mL and 40 mL brown glass bottles (pre-cleaned and pre-combusted) for later CDOM and DOC analyses. All samples were frozen (-20°C) and protected from light, and upon arriving at the land laboratory, were analyzed as soon as possible.

# 2.3 Photoexposure experiment

SSW and SML water samples were collected in July 2018 at stations A3, BF, and H10 as well as D2 and F6 located in the YS and the ECS, respectively. Samples (SSW: 2L; SML: 500 mL) were immediately passed through 0.22 μm PES filters (Pall Corp. Port Washington, NY, USA) to remove the majority of bacteria, placed in acid-washed and pre-combusted brown glass bottles and stored at 4°C. Similarly, filtered samples from each site were placed in five 80 mL optically transparent quartz tubes (acid-washed and pre-combusted) and sealed without headspace or air bubbles to measure the effect of light exposure. The quartz tubes were positioned on their sides under the irradiation source to maximize the exposure of the sample; the water depth in each tube was 5 cm (i.e. the diameter of the tube). Both sets (SML and SSW) were irradiated for 6, 12, 24, 50, and 88 h (25°C) in a GLZ-C





- 124 Quantum Sensor (Top Cloud-Agri Instrument, Zhejiang, China) solar simulator. All samples for DOC
- 125 concentration measurements were acidified to approximately pH 2.0 with high purity HCl and
- 126 analyzed within 7 d, and absorbance spectra and fluorescence excitation emission matrices (EEMs)
- were run on non-acidified samples within 3 d of sampling (4°C and dark).
- 128 2.4 Analytical measurements
- 129 Determination of the CDOM absorption coefficient
- Absorption spectra were determined using a UV-visible spectrophotometer (UV-2550 bi-channel;
- 131 Shimadzu, Tokyo, Japan) equipped with two 10 cm path-length quartz cuvettes. Sample absorbance
- 132 was automatically corrected for the absorbance of Milli-Q water. Absorbance scans ranged from 200 to
- 133 800 nm, with a spectral resolution of 1 nm. The absorption coefficient of CDOM was calculated
- 134 according to equation (1):

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$$a(\gamma) = 2.303A(\gamma)/1$$
 (1)

- where,  $A(\lambda)$  is the absorbance at wavelength  $\lambda$ ; and r is the path length of the quartz cuvette in meters.
- 137 The spectral slope of the CDOM absorption curve (S) was calculated according to a non-linear
- regression over the 275–295 nm and 350–400 nm wavelength range, according to:

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$$a(\lambda) = a(\lambda_0) \exp[S(\lambda_0 - \lambda)] + K$$
 (2)

- 140 where,  $\alpha(\lambda)$  is the absorption coefficient at wavelength  $\lambda$ ;  $\alpha(\lambda_0)$  is the absorption at the reference
- wavelength  $\lambda_0$  of 440 nm; S is the spectral slope; and K is a background parameter that accounts for
- 142 baseline shifts or attenuation due to factors other than CDOM. S was measured in the wavelength
- 143 ranges of 275–295 nm (S<sub>275-295</sub>, nm<sup>-1</sup>) and 350–400 nm (S<sub>350-400</sub>, nm<sup>-1</sup>). S<sub>275-295</sub> is used to characterize
- 144 DOM, with high values generally indicative of low-molecular-weight DOM that are linked to
- photochemical modification (Helms et al., 2008; Ortega-Retuerta et al., 2009). The spectral slope ratio
- 146 (S<sub>R</sub>) was defined as the ratio of the two spectral slopes, S<sub>275-295</sub> to S<sub>350-400</sub>. S<sub>R</sub> is also a sensitive
- 147 indicator of photochemically induced changes in the molecular weight within the CDOM pool, with

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toolbox (Stedmon and Bro, 2008).





al., 2009). We used the absorption coefficient at 254 nm (a(254)) to determine the concentration and distribution of CDOM in the SML from the eastern marginal seas of China. The specific UV absorbance (SUVA254) can be used to measure aromaticity (Weishaar et al., 2003) and molecular weight (Chowdhury, 2013) of DOM, with higher values generally indicative of higher aromaticity. EEMs and determination of the CDOM fluorescence index EEMs were obtained using a F-4500 fluorescence spectrophotometer with a 1 cm quartz cuvette (Shimadzu) (Hoge et al., 1993). The emission spectra were scanned every 5 nm from 250 nm to 550 nm, and at the excitation wavelengths between 200-400 nm at 5 nm intervals, with 5 nm slit widths for the excitation and emission modes. The FL Toolbox, which was developed by Wade Sheldon (University of Georgia) for MATLAB, was used to remove the Rayleigh and Raman scattering peaks using the Delaunay triangulation method (Zepp et al., 2004). The fluorescence intensities of the samples were corrected with Milli-Q water blank EEMs and then normalized to the water Raman integrated area maximum fluorescence intensities (Ex/Em = 350 nm/365-430 nm, 5 nm bandpass) (Coble et al., 1998; Singh et al., 2010). Raman units (RU) (Stedmon et al., 2007; Singh et al., 2010) were used as the units for the Raman peak areas of water when the excitation wavelength of 350 nm was used for correction. EEMs were modeled using PARAFAC in MATLAB 7.5 with the DOMFluor

increases in S<sub>R</sub> suggesting stronger photochemical degradation (Helms et al., 2008; Ortega-Retuerta et

$$X_{ijk} = \sum_{n=1}^{F} a_{in} b_{jn} c_{kn} + \varepsilon_{ijk}$$
(3)

where  $X_{ijk}$  is the fluorescence intensity of the *i*th sample at the *k*th excitation and *j*th emission wavelengths;  $a_{in}$  is directly proportional to the concentration (scores) of the nth fluorophore in the *i*th sample;  $b_{jh}$  and  $c_{kn}$  are the estimates of the emission and excitation spectra (loadings) of the nth fluorophore at wavelengths *j* and *k*, respectively; *F* is the number of components (fluorophores); and  $\varepsilon_{ijk}$  represents the unexplained variability of the model (Singh et al., 2010). Split-half analysis validation was used to determine the number of fluorescent components. The fluorescence intensity of each fluorescent component was evaluated (Fig. S2, Supporting Information, Table 1).

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Determination of DOC, chlorophyll-a, heterotrophic bacterial abundance, dissolved oxygen, and other parameters

Concentrations of DOC were determined using the Shimadzu TOC-V<sub>CPH</sub> total organic carbon analyzer with an injection volume of 80 μL. The accuracy of the test was ensured by measuring a deep seawater reference (Hansell Laboratory, University of Miami) every 10 samples. The Chl-*a* concentration was determined by a fluorescence spectrophotometer (7200-000, Turner Designs, CA) after extraction in 90% acetone based on the procedure of Parsons et al. (1984). DO was determined by iodination using the Winkler titration method (Carpenter, 1964), the endpoint was determined using starch as a visual indicator. Salinity and temperature data were collected in situ by a conductivity-temperature-depth sensor. All phytoplankton samples were enumerated in triplicate according to Specification for Oceanographic Survey (State Bureau of Technical Supervision Bureau, 1992). Nutrient species concentrations were determined using an automatic analyzer (QuAAtro, Seal Analytical, Germany) (Grasshoff et al., 2007). Heterotrophic bacterial abundances were measured by flow cytometry (colorimetry, Beckman Coulter FC500-MPL) as described by Marie et al. (1997).

189 The enrichment factor (EF) in the SML is defined as follows

$$EF = C_M / C_S \tag{4}$$

where  $C_M$  is the concentration of any substance in the SML; and  $C_S$  is its concentration in the SSW. If the EF of a substance is greater than 1.0, that substance is considered enriched, if it is less than 1.0, it is considered depleted (Chen et al., 2016).

195 2.5 Statistical analyses

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The correlation coefficient (R) and probability (P) values were used to evaluate the goodness-of-fit. The correlation matrix, analysis of variance, and principal components analysis were conducted with SPSS version 18.0 (SPSS Inc., Chicago, IL, USA) to determine the possible relationships between the DOM parameters and environmental factors. A P-value ≤ 0.05 was considered significant. Regression analyses between the optical parameters of DOM and several biogeochemical parameters in the SSW and the SML samples were performed in the Table S1 and the Table S2, respectively. 3. Results and discussion 3.1 Distribution and chemical characterization of DOM in the SSW of the eastern marginal seas of China The surface distributions of salinity, temperature, CDOM, DOC, Chl-a, and several optical parameters in the study area during spring, summer and winter are shown in Fig. S3 (SSW)-S4 (SML) (Supporting Information). There was a strong south-to-north temperature gradient, with warmer waters in the ECS and cooler waters in the YS. Lower salinities were observed in the Changjiang Estuary and coastal waters. The lowest mean wind speed was observed in the summer of 2018 (Table 2). In spring and summer, the bacterial abundances were lower in the YS (spring mean concentration:  $2.26 \times 10^8$ cells/L; summer mean concentration:  $3.79 \times 10^8$  cells/L) than in the ECS (spring mean:  $2.98 \times 10^8$ cells/L; summer mean:  $7.64 \times 10^8$  cells/L), indicating that the warmer southern ECS had stronger biological activity in the SSW. The a(254) value ranged from 1.08 to 19.28 m<sup>-1</sup> in the SML and from 0.82 to 14.23 m<sup>-1</sup> in the SSW during these three seasons. CDOM absorption values and DOC concentrations were generally

decreased from the inshore to the offshore stations (Fig. S3 c)-d)). Higher a(254) values were



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generally observed in the Changjiang Estuary (spring: station D1 (4.13 m<sup>-1</sup>); summer: station D2 (3.98 m<sup>-1</sup>); winter: station D1 (3.14 m<sup>-1</sup>)) and in the northern YS (spring: station A2 (4.26 m<sup>-1</sup>); summer: station H11 (5.37 m<sup>-1</sup>); winter: station H12 (5.95 m<sup>-1</sup>)). There were significantly negative linear correlations between salinity and a(254) in all cruises in the SSW (p < 0.01, Fig. 2), especially in the ECS, implying that freshwater run-off and seawater mixing played a more important role in determining CDOM distributions in the SSW. The strongest negative linear relationship observed between salinity and a(254) was observed in winter when the influence of terrestrial input in this study region was maximal. In addition, SUVA<sub>254</sub> ranged from 0.51 to 8.39 L mg C<sup>-1</sup> m<sup>-1</sup> in the SML. In comparison with the SML, the SSW exhibited lower variability in SUVA254 values from 0.63 to 5.39 L mg C<sup>-1</sup> m<sup>-1</sup>, with higher values at the northern YS stations and Changjiang Estuary coastal stations (Fig. S3k)). According to the SUVA<sub>254</sub> trends observed by Massicotte et al. (2017), the DOM composition we observed in the SSW of the Changjiang Estuary ecosystem were more similar to the DOM measured in freshwater ecosystems than in the ocean. SUVA<sub>254</sub> underwent a sharp decrease from the Changjiang Estuary ecosystem to the southeastern ECS, suggesting that aromatic and/or highly conjugated DOM moieties were degraded along the aquatic continuum from the Changjiang Estuary to the open ocean. Higher S<sub>275-295</sub> values were also observed in some off-shore stations (Fig. S4i)). These comparisons showed that the DOM pools of the Changjiang Estuary contained molecules that were more HMW-DOM and contained more aromatic compounds, CDOM in the SSW of the southeastern ECS, which was derived predominantly from an autochthonous origin (phytoplankton production and bacterial activity), clearly showed the presence of organic matter freshly released into sea (Yang et al., 2020). The detail of mixing behavior, biological and photolytic degradation of dissolved organic matter in the East China Sea and the Yellow Sea were discussed in our previous paper (Yang et al.,





240 2020). 241 242 3.2 Fluorescence signature and factors controlling the composition of FDOM components in the SSW 243 and the SML 244 FDOM properties can be used as the sensitive indicator of DOM processing and water mass. Four 245 fluorescent components were identified by PARAFAC analysis with the DOM Fluor toolbox in 246 MATLAB 7.5 (Stedmon and Bro, 2008), hereafter named C1, C2, C3, and C4 (Fig. S2). The 247 humic-like C1 and C3 were categorized as two traditional types of humic-like fluorescent components 248 (Coble 1996). Component 1 had primary fluorescence excitation and emission peaks at 345 nm and 249 455 nm, respectively, which was similar to terrestrial humic-like fluorophores in the visible region 250 (peak C) (Osburn et al., 2012). Relative to C1, the fluorescence of C3 was blue-shifted and had 251 fluorescence peaks at 385 nm emission and 315 nm excitation. The microbial humic-like component 252 had a relatively shorter emission peak wavelength compared to the terrestrial humic-like PARAFAC 253 components previously identified in the open ocean (Catala et al., 2015). C2 exhibited Ex/Em maxima 254 at 255 nm/310 (375) nm, which could be considered tyrosine-like fluorescence (Stedmon et al., 2003) 255 and attributed to autochthonous and/or microbially consumed FDOM. C4 had an excitation range of 256 280 nm with an emission peak at 335 nm, which corresponded to peak T of the amino-acid-like 257 fluorescence of tryptophan, likely derived from in situ primary autochthonous substances and other 258 fresh biological sources (Coble, 1996). The tryptophan-like C4 and the humic-like C1 and C3 in the 259 SSW were all negatively correlated with salinity (P < 0.01, Table S1), but increased with the 260 increasing DO level. These suggested that water mixing and microbial activity were important factors in determining geographical distributions of FDOM in the SSW (Breitburg, et al., 2018; Yamashita et 261





al., 2017; Galgani and Engel, 2016). Moreover, the geographical distribution of humic-like C1 and 263 protein-like components were more similar to that of the Chl-a concentration in the SML (Fig. 3 a, b, 264 d). Such relationships suggested that the production of protein-like and humic-like FDOM with 265 phytoplankton production and decay in the SML. 266 FDOM enrichment in the SML of all stations ranged between 0.5 and 11 (n = 225) and FDOM was more frequently enriched (C1: 89.6%; C2: 73.2%; C3: 91.8%; C4: 93.4% of all samples) than 267 268 CDOM. The fluorescence intensity of the components in the SML samples decreased in the following 269 order: tryptophan-like > tyrosine-like > terrestrial humic-like > marine humic-like; whereas those in 270 the SSW samples decreased in the order: tyrosine-like > tryptophan-like > marine humic-like > 271 terrestrial humic-like. The tryptophan-like component (C4) was mostly enriched in the SML samples 272 with a median EF = 2.2 and a range from 0.2 and 8.0. The EF of C4 was clearly higher than other 273 components in all seasons (Fig. 5b)), especially in summer, and the FDOM composition in the SML 274 revealed a relatively higher proportion of autochthonous tryptophan-like FDOM than the SSW. It has 275 also been broadly recognized that tryptophan-like C4 in the particulate fraction is related to recent 276 primary production (Brym et al., 2014; Yamashita, 2014) and that phytoplankton excrete 277 tryptophan-like fluorophores (Romare-Castillo et al., 2010). Together, as already emphasized 278 previously, the variation observed for FDOM can be more related to that of Chl-a in the SML, these 279 observations suggested that the DOM enriched in the SML was made up of a relatively higher 280 proportion of marine autochthonous DOM than the SSW. 281 282 3.3 DOM and biogenic molecules accumulation in the SML 283 Up to 88% of our CDOM samples were enriched in the SML, with the median EF for a(254) of 284 1.3, ranging between 0.4 and 6.7. Concentrations of CDOM, FDOM, nutrients, bacterial abundance,

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and Chl-a in the SML were correlated with their respective SSW concentrations (Fig. 4), demonstrating that transport from the SSW to the SML is an important pathway. Furthermore, the relatively higher CDOM absorption enrichment value in the SML were found at longer wavelengths (Fig. 5a)) EF of a(355) > EF of a(254)). Marine production of DOM had the largest influence on the CDOM absorption properties in the longer wavelength range (Danhiez et al., 2017) (S<sub>320-412</sub>: DOM marine origin VS. S<sub>275-295</sub>: terrestrial DOM). Galgani and Engel (2016) also observed that amino acid-like fluorophores were highly enriched in the SML, not only due to their amphiphilic properties, but also due to their local production in the SML. Therefore, the marine local production might significantly affect the composition of DOM in the SML. Additionally, the nutrients showed significantly higher EFs (NO<sub>3</sub>:  $3.41 \pm 6.08$ , n = 41; NO<sub>2</sub>:  $3.57 \pm 5.54$ , n = 52; PO<sub>4</sub><sup>3</sup>:  $2.13 \pm 2.74$ , n =68; and  $SiO_3^{2-}$ : 6.53  $\pm$  13.67, n = 13) than biological and DOM parameters in the SML. The strong correlation between the SML and SSW concentrations of NO<sub>2</sub>, NO<sub>3</sub>, and SiO<sub>3</sub><sup>2</sup> (Fig. 4) showed that the similar fundamental drivers are probably at work in both compartments for these nutrients. For example, dissolved substances, particles, and microorganisms were brought to the interface by simple diffusion, rising bubbles (Jarvis, 1967), convection, and upwelling from sediments and subsurface water, and at the same time, the microlayer is also a sink for fallout from the atmosphere (Duce et al., 1976). In addition, we also observed the significant positive relationship between a(254) and Chl-a (R = 0.662, P < 0.01) in the SML during spring, and the positive relationship between the EF of PO<sub>4</sub><sup>3-</sup> and the EF of Chl-a (R = 0.319, P = 0.01, Table 3). These observations indicated that spatial variation of CDOM concentrations were related to Chl-a in the SML. The enrichment of inorganic nutrients would be an important factor influencing the production and composition of phytoplankton-produced DOM (Carlson and Hansell, 2003) in the SML. Therefore, phytoplankton growth, primary productivity rate,

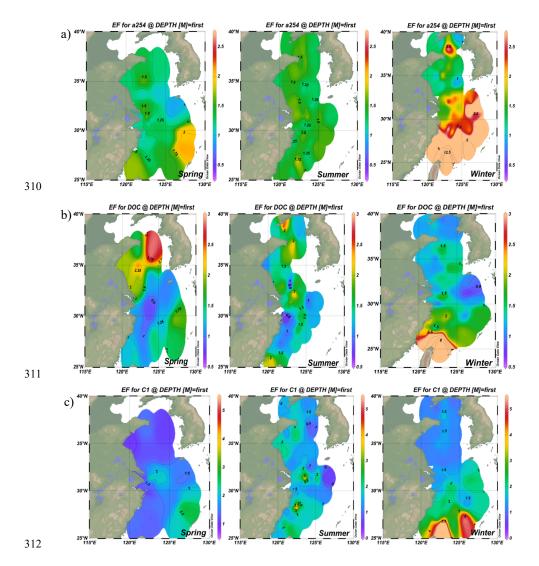




biological activity and marine autochthonous DOM production would all be enhanced by the enriched

308 nutrients in the SML.

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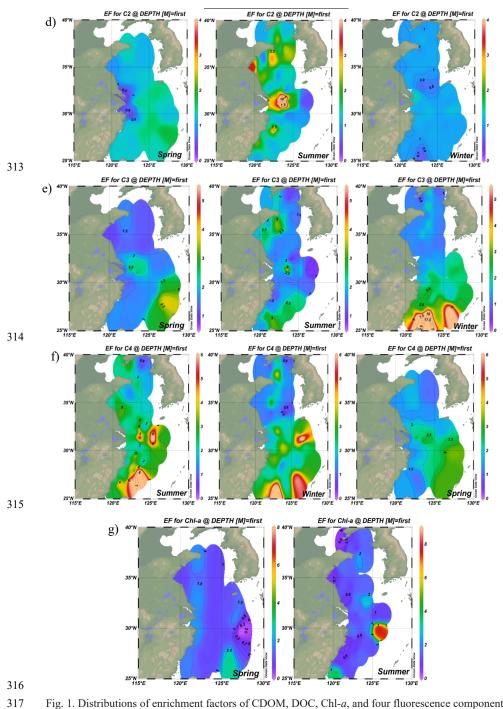


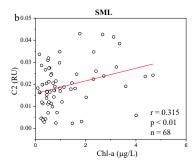
Fig. 1. Distributions of enrichment factors of CDOM, DOC, Chl-a, and four fluorescence components in the surface microlayer water during spring, summer, and winter. Increasing DOM yields were significant in coastal regions in all seasons, but the higher EFs were more pronounced in off-shore





320 regions. Summer SSW Spring SSW r = 0.0113 r = -0.794 p < 0.0001 n = 22 • ECS p = 0.95023n = 33 • ECS r = -0.797 p < 0.0001 n = 32 r = -0.844 p < 0.0001 n = 73 a(254) (m<sup>-1</sup>)  $a(254)^{3}(m^{-1})$ 321 Winter SSW Salinity 33 30 a(254) (m<sup>-1</sup>)

Fig. 2. Relationships between a(254) and salinity in the SSW in the YS and the ECS during spring, summer and winter.



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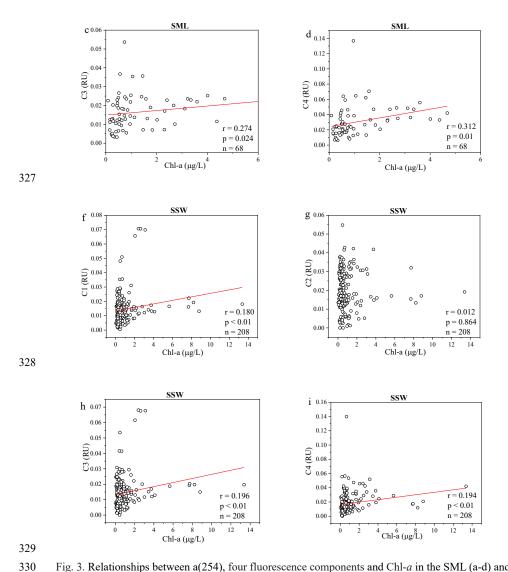
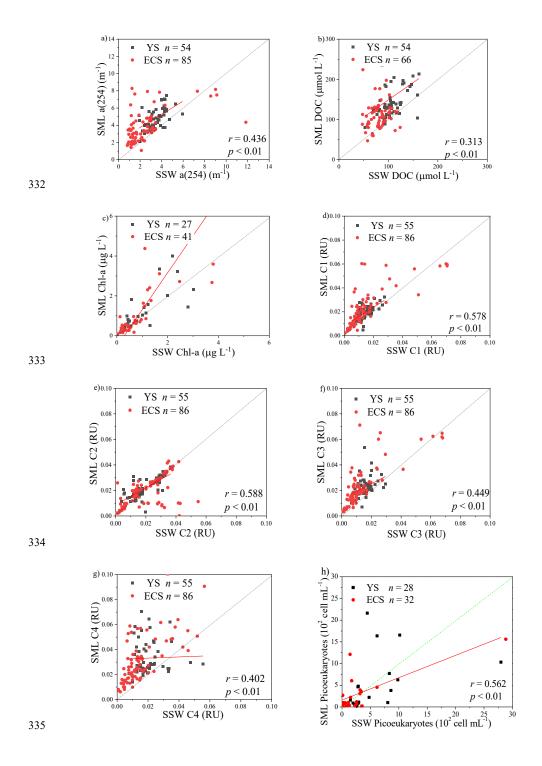


Fig. 3. Relationships between a(254), four fluorescence components and Chl-a in the SML (a-d) and in the SSW (f-i).







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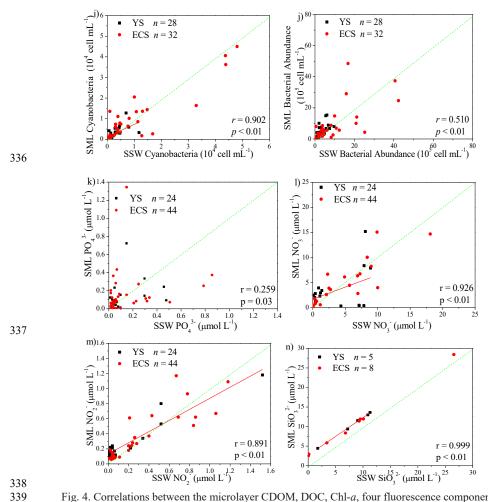


Fig. 4. Correlations between the microlayer CDOM, DOC, Chl-*a*, four fluorescence components concentrations, cyanobacteria, phytoplankton biomass, nutrients and bacterial abundance, and their subsurface water concentrations. The dashed lines correspond to the 1:1 lines, and the full lines are the regression models. (All DOM spectroscopic parameters sample were analyzed in spring, summer, autumn, and winter; Chl-*a* was determined in spring, summer, and summer; cyanobacteria, phytoplankton biomass, nutrients and bacterial abundance were determined in spring and summer.).





a)
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3 a(254)
a(355)
DOC
Chl-a
Cell
Po<sub>4</sub>
Po<sub>4</sub>
No<sub>3</sub>
No<sub>2</sub>
Spring
Summer
Winter

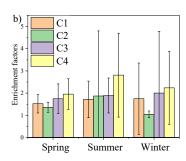


Fig. 5. Mean enrichment factor of a<sub>CDOM</sub> (254 nm and 355 nm), DOC, Chl-*a*, nutrients (PO<sub>4</sub><sup>3-</sup>; NO<sub>3</sub><sup>-</sup>; NO<sub>2</sub><sup>-</sup>, SiO<sub>3</sub><sup>2-</sup>), and four fluorescence components during spring, summer, and winter.

3.4 Wind speed influencing the enrichment of DOM optical properties

The wind speeds during our observations ranged from 0.2 to 14.9 m s<sup>-1</sup>. We divided them into three different wind regimes: low  $(0.0-2.0 \text{ m s}^{-1})$ , moderate  $(2.0-10.0 \text{ m s}^{-1})$ , and high  $(10.0-14.9 \text{ m s}^{-1})$ . Although the EFs of DOC and Chl-a were negatively correlated with wind speed (DOC: P = 0.002; Chl-a: P = 0.042), the EFs of CDOM and FDOM were not. During the low wind regime, no significant relationships were apparent between wind speed and either EFs of CDOM or FDOM, CDOM and FDOM were consistently enriched, with EFs ranging from 1.0 to 2.2, and a mean a(254) EF value of 1.3 (n = 20). However, the EFs during moderate winds had larger variability and ranged from 0.9 to 14.5, with a mean EF value of 1.6 (n = 143), and during high winds they ranged from 0.6 to 1.8, with a lower mean EF value of 1.1 (n = 18). In addition, depleted levels of CDOM (EF < 1) occurred at frequencies of 5.6%, 9.1%, and 20.0% during low, moderate, and high wind regimes, respectively. Therefore, although lower wind speeds and ascending bubbles might further promote the transportation of organic materials from the underlying waters, DOM enrichments were still observed at wind speeds up to > 10 m s<sup>-1</sup>. Reinthaler et al. (2008) also reported that higher enrichment was found at higher wind speeds. During moderate to high wind regimes, breaking waves not only can

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disrupt the surface film and physically drive DOM back into the bulk water, but also facilitate the formation of the SML as rising bubble plumes transported DOM to the surface, resulting in wider ranges of EFs (Frew et al., 2004). Higher wind speed does enhance mixing (Reinthaler et al., 2008), which can arguably favour transport of nutrients and DOM from the SSW equally (Wurl et al., 2011). Although wind speed appear to play an important role in the enrichment of surface-active DOM, the chemical composition of the SML influence its stability. For example, enrichments of sulphate half-ester groups in the SML (Wurl and Holmes, 2008) could increase stability because these groups can influence the intrinsic viscosity of marine polymers (Nichols et al., 2005) and sulphur-containing algal carbohydrates are less soluble and hydrolysable (Kok et al., 2000). Enrichment processes and biochemical processes of organic substances in the marine environment are all likely to be the more important contributors of DOM to the SML in our study regions. 3.5 Photochemical degradation of DOM in the SSW and the SML Photobleaching is one of the major mechanisms determining the geographical distributions of chromophoric and fluorescent DOM in the ocean (Helms et al., 2008; Brinkmann et al., 2003; Siegel et al., 2005). The average SUVA254 values in SSW were generally higher than those in the SML in our study regions (SSW:  $2.45 \pm 0.91$  L mg-C<sup>-1</sup> m<sup>-1</sup> vs. SML:  $2.39 \pm 1.34$  L mg-C<sup>-1</sup> m<sup>-1</sup>), and the most obvious distinction happened in summer (Table 2). These indicated that although CDOM concentration in the SSW was lower than that in the SML, CDOM in the SSW has a higher degree of aromaticity compared to the SML. Thus we performed photochemical incubation experiments to confirm whether photochemical reactions influenced the differentiated aromaticity and photo-reactive features of DOM between the SML and the SSW. After 88 h of exposure, the a(254) values were only 49.6%, 45.5%, 42.1%, 41.8% and 37.0% of



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the initial values at stations A3, BF, D2, F6, and H10 in the SSW, and 72.5%, 42.4%, 42.6%, 49.0% and 44.0% of the initial values at stations A3, BF, D2, F6, and H10 in the SML, respectively. Overall, a(254) and SUVA<sub>254</sub> decreased by  $49.9 \pm 12.8\%$  and  $43.0 \pm 15.5\%$ , respectively, in the SML, and by  $56.8 \pm 4.7\%$  and  $56.0 \pm 10.2\%$ , respectively in the SSW. Therefore, stimulated solar UV exposure caused a larger decrease in DOM absorbance in the SSW than the SML (Fig. 6). The relatively rapid decrease of SUVA254 in the SSW indicated a more rapid conversion of DOM to less humic-type materials than in the SML. Approximately 65% of FDOM was lost during the irradiation experiment, except in the case of the tyrosine-like component 2 from the SML at the station H10, which increased slightly. Photoproduction of tyrosine-like components has been previously reported by Zhu et al. (2017), who suggested that the photochemical degradation of CDOM contributed to the release small amounts of tyrosine-like fluorophores. The tryptophan-like C4, humic-like C1 and C3 were more photodegradated in the SSW than in the SML (Fig. 6c), e), f)). For example, C1, C3 and C4 show a marked decrease in the SSW at the off-shore station F6. Because of the origin of CDOM at the station F6 remote from the direct terrestrial influence, the majority of CDOM at the station F6 was thought to be a by-product of net primary production. The present results showed that a large fraction of the total CDOM in the SSW at the off-shore station F6 is still potentially sensitive to photooxidation. As already referred previously, CDOM in the SSW showed higher SUVA254 values, and higher percentages of humic-like DOM than in the SML. Therefore, the photochemically mediated shifts in DOM in the SSW were more pronounced than those in the SML in our incubation experiments, in terms of both absorption and fluorescence values. This heterogeneity in the EFs and photochemical reactivities of FDOM components can be related to the chemical and structural nature, such as molecular weight, aromaticity or humification of

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FDOM enrichment processes. Hydrophilic, carboxylic acid-bearing DOM moieties are preferentially degraded by simulated sunlight (Brinkmann et al., 2003). The largest fractions of photolabile DOM are made up of aromatic carbon rings or high double bond equivalent molecules (Kujawinski et al., 2004; Gonsior et al., 2009). The humic-like C1 and C3, all of which exhibited significantly positive relationships with SUVA<sub>254</sub> (< 0.001, Table 1) and shown higher aromaticity, were more prone to photochemical degradation (Fig. 6c), e), and f)). The tyrosine-like C2, as compared to other protein-like compounds, is generally considered more labile and susceptible to bacterial cycling and rapid consumption by microbiota (Medeiros et al., 2015). The SML experiences the most intense solar radiation, especially ultraviolet (UV) light (Obernosterer et al., 2005). Photochemical degradation may, therefore, be a sink for aromaticity fluorescent components in the SML, and be a source for the tyrosine-like C2. In addition, Blough (1997) discovered that photochemical production rates in the SML should lead to the more rapid oxidative turnover of materials at the interface and potentially to reactions and processes not observed in bulk waters. Therefore, differences in SUVA254 values and photoreaction behavior between the SSW and the SML may also reflect that DOM in the SML was already photobleached, which resulted in the decrease of DOM aromaticity, CDOM in the SSW appeared to be more susceptible to photochemical degradation than CDOM in the SML. Together, photo-irradiation have the significant influence on the accumulation of protein-like DOM and depletion of aromatic organic compounds in the SML, and organic carbon might have undergone a more rapid cycling in the SML than the SSW.





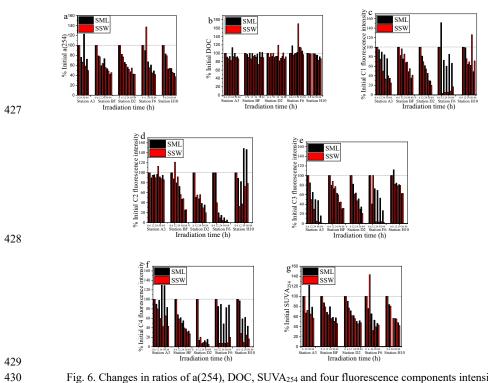


Fig. 6. Changes in ratios of a(254), DOC, SUVA $_{254}$  and four fluorescence components intensities to initial values for both SML and SSW sample.

3.6 Variations in the enrichment factors of CDOM, DOC, FDOM along the coastal regions to ocean

The concentrations of a(254) and DOC decreased from the coastal regions to the open ocean, and decreased from the northern part of the sampling area (the YS) to the southern part of the sampling area (the ECS) in both the SSW and the SML (Fig. S3 c)-d) and Fig. S4 a)-b)). However, CDOM and FDOM were more frequently enriched in the ECS (CDOM: 93% of all samples; FDOM: 72–94% of all samples) than that in the YS (CDOM: 86% of all samples; FDOM: 70–92% of all samples). The higher EF values for CDOM, FDOM, DOC, Chl-a, nutrients, and cell were generally observed in the ECS (Fig. 1). Lower EFs and EFs < 1, which indicate a depletion of CDOM in the SML, were usually observed at short distances from the coast (Fig. 1) with lower salinity. The salinity during our

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observations ranged from 23.6 to 35.1. Although CDOM and FDOM concentration negatively correlated with salinity, the EFs of CDOM and FDOM were weakly positive related with salinity (Fig. 7). The EFs of Chl-a and nutrient were also higher in the southeastern ECS (Fig. 1 and Fig. S5), where sufficient light and higher temperature combined to facilitate primary production and higher contributions of autochthonous materials to DOM. DOM in the SSW of the southern ECS was more dominated by marine autochthonous materials in our previous discussion (Yang et al., 2020). The Changjiang River discharges enormous amounts of N and P into the ECS (Liu et al., 2018), but phosphorus is generally the major limiting element for phytoplankton growth in the ECS (Liu et al., 2016). Thus the difference in EFs of CDOM and Chl-a between YS and the ECS, and between the coast and off-shore regions is likely due to the significantly nutrients enrichment in the SML in the off-shore regions. In winter, we observed especially higher EF values for CDOM and FDOM in the southern ECS (Fig. 1a)-f)). With wind from the northwest (Weng et al., 2011), biologically essential trace elements and anthropogenic emissions are carried from the land and can enter the ocean via the SML by wet or dry deposition. The EFs of humic-like C1 and C3 were relatively high in winter (Fig. 5b)), probably due to the input from atmospheric deposition during winter, and the relatively low CDOM concentrations in bulk water. Atmospheric deposition of organic carbon and nutrients were found a peak in winter over the coastal ECS (Wang et al., 2019). We suggested that the EFs of CDOM and FDOM increased from the coastal regions to the open ocean, and increased from the YS to the ECS were likely due to the enrichment of enough nutrients in the SML in the open ocean promote phytoplankton biomass and DOM production.





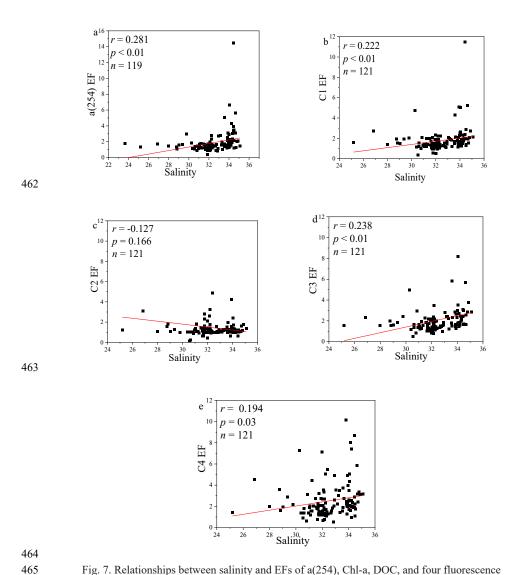


Fig. 7. Relationships between salinity and EFs of a(254), Chl-a, DOC, and four fluorescence components.

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The SML is an aggregate-enriched biofilm environment with distinct microbial communities, where the diversity of microorganisms can differ significantly from those of underlying waters (Liss and Duce, 2005; Cunliffe et al., 2013), the heterotrophic bacterial abundance in the SML was  $\sim 7.5$ fold greater than those in the SSW in the ECS during our spring cruise (Sun et al., 2020). Here, EFs showed a greater presence of bacteria and marine protein-like DOM in the SML than that in the SSW, while the protein-like DOM was linked to microbial utilization and degraded faster than the humic-like substances (Yang et al., 2017; Jørgensen et al., 2011). Therefore, compared to coastal waters that have larger terrestrial DOM and nutrients inputs, CDOM shown higher EFs in off-shore regions where DOM in the SSW is mostly of marine autochthonous origin with higher temperatures and stronger biological activity. The significantly higher abundance of cells, phytoplankton, nutrients, and protein-like DOM in the SML supported microbial activities, and further contributing to the local release of marine extracellular DOM directly from microbes in the SML in the off-shore regions. When exposed to higher light intensities (summer), obviously enhance mineralization of DOM in the SML, and relatively less photochemical degradation in SSW could result in lower percentage of aromatic DOM in SML than the SSW. We concluded that SML CDOM dynamics can be expressed as a simple balance among enrichment process, primary production and photochemical destruction. Thus, higher EF of DOM in the SML in off-shore regions are likely supported by a favorable combination of: 1) deposition and accumulation of amphiphilic compounds, 2) importance of bubble for upward transport of DOM and enrichment in SML, and 3) new production of DOM within the SML as a consequence of higher nutrients enrichment and the primary production.

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## 4. Conclusions





This study has provided the first data set that considers the distributions of CDOM, FDOM, DOC, Chl-a, nutrients, and bacterial abundances in the SML and SSW of the ECS and the YS during spring, summer, and winter. We have observed that the CDOM distribution related variability in primary production in the SML. Furthermore, we have demonstrated that localized and stronger photochemical oxidation may be responsible for the decrease in the aromaticity of the DOM in the SML, due to enhanced transformation or removal of terrestrial DOM, compared with the SSW. We also demonstrated that in off-shore seawaters away from terrigenous influence, the EFs of CDOM, DOC, FDOM and Chl-a in SML tend to be higher in off-shore regions than those in coastal regions, because of the relatively higher enrichment of nutrients which could enhance phytoplankton growth and promoted plant production and DOM production in the SML. Multiple observations of spatial distributions, seasonal variations, chemical compositions, and photochemical reactions of CDOM in the SML have supported the hypothesis that stronger enrichment and photochemical processes occur in the SML in ocean, resulting in relatively accelerated enrichment of more marine local production DOM in the SML than the SSW.

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513	References
514	Aller, J.Y., Kuznetsova, M., Jahns, C.J., Kemp, P.F., 2005. The sea surface microlayer as a source of
515	viral and bacterial enrichment in marine aerosols. J. Aerosol Sci. 36(5), 801-812.
516	https://doi.org/10.1016/j.jaerosci.2004.10.012
517	Breitburg, D., Levin, L.A., Oschlies, A., Grégoire, M., Chavez, F.P., Conley, D.J., Garçon, V., Gilbert,
518	D., Gutiérrez, D., Isensee, K., Jacinto, G.S., Limburg, K.E., Montes, I., Naqvi, S.W.A., Pitcher,
519	G.C., Rabalais, N.N., Roman, M.R., Rose, K.A., Seibel, B.A., Telszewski, M., Yasuhara, M.,
520	Zhang, J., Declining oxygen in the global ocean and coastal waters. Science 2018, 5, 359(6371),
521	eaam7240. https://doi.org/10.1126/science.aam7240
522	Brinkmann, T., Sartorius, D., Frimmel, F.H., 2003. Photobleaching of humic rich dissolved organic
523	matter. Aquat. Sci. 65(4), 415–424.
524	Brym, A., Paerl, H.W., Montgomery, M.T., Handsel, L.T., Ziervogel, K., Osburn, C.L., 2014. Optical
525	and chemical characterization of base-extracted particulate organic matter in coastal marine
526	environments. Mar. Chem. 162(6), 96–113. https://doi.org/10.1007/s00027-003-0670-9
527	Carlson, C.A., Hansell, D.A., 2003. The contribution of dissolved organic carbon and nitrogen to
528	biogeochemistry of the Ross Sea. In: DiTullio, G., Dunbar, R. (Eds.), Biogeochemical Cycles in
529	the Ross Sea. AGU Press, Washington DC, pp. 123–142.
530	Carpenter, J.H., 1964. The Chesapeake Bay Institute technique for the Winkler dissolved oxygen
531	method. Limnol. Oceanogr. 10(1964), 141–143. https://doi.org/10.4319/lo.1965.10.1.0141
532	Catala, T.S., Reche, I., Fuenteslema, A., Romeracastillo, C., Nietocid, M., Ortegaretuerta, E.,
533	Alvarezsalgado, X.A., 2015. Turnover time of fluorescent dissolved organic matter in the dark





534 global ocean. Nat. Com. 6(1), 5986-5993. https://doi.org/10.1038/ncomms6986 535 Chen, Y., Yang, G., Xia, Q., Wu, G., 2016. Enrichment and characterization of dissolved organic 536 matter in the surface microlayer and subsurface water of the South Yellow Sea. Mar. Chem. 537 182(Mar. 20), 1-13. https://doi.org/10.1016/j.marchem.2016.04.001 538 Chowdhury, S., 2013. Trihalomethanes in drinking water: Effect of natural organic matter distribution. Water SA 39(1), 1-7. https://doi.org/10.4314/wsa.v39i1.1 539 540 Coble, P.G., 1996. Characterization of marine and terrestrial DOM in seawater using excitation-emission 541 matrix spectroscopy. Mar. Chem. 51(4), 325-346. 542 https://doi.org/10.1016/0304-4203(95)00062-3 543 Coble, P.G., 2007. Marine optical biogeochemistry: the chemistry of ocean color. Chem. Rev. 107(2), 544 402-418. https://doi.org/10.1002/chin.200720265 545 Cunliffe, M., Engel, A., Frka, S., Gasparovic, B., Guitart, C., Murrell, J.C., Wurl, O., 2013. Sea surface 546 microlayers: A unified physicochemical and biological perspective of the air-ocean interface. 547 Prog. Oceanogr. 109(Feb.), 104-116. https://doi.org/10.1016/j.pocean.2012.08.004 548 Danhiez, F.P., Vantrepotte, V., Cauvin, A., Lebourg, E., Loisel, H., 2017. Optical properties of 549 chromophoric dissolved organic matter during a phytoplankton bloom. Implication for DOC 550 estimates from **CDOM** absorption. Limnol. Oceanogr. 62(4),1409-1425. https://doi.org/10.1002/lno.10507 551 552 Duce, R.A., Hoffman, G.L., Ray, B.J., Fletcher, I.S., Wallace, G.T., Fasching, J.L., Piotrowicz, S.R., 553 Walsh, P.R., Hoffman, E.J., Miller, J.M., Heffter, J.L., 1976. Trace metals in the marine atmosphere: Sources and fluxes, in: Marine Pollutant Transfer (H. L. Windom and R. A. Duce, 554 555 eds.), pp. 77-119, Lexington Books, Lexington. 556 Engel, A., Galgani, L., 2016. The organic sea-surface microlayer in the upwelling region off the coast





557 of Peru and potential implications for air-sea exchange processes. Biogeosciences 13(4), 558 989-1007. https://doi.org/10.5194/bg-13-989-2016 559 Frew, N.M., Bock, E.J., Schimpf, U., Hara, T., Hausecker, H., Edson, J.B., Jahne, B., 2004. Air-sea gas 560 transfer: Its dependence on wind stress, small-scale roughness, and surface films. J. Geophys. 561 Res.-Oceans 109(C8), S17. https://doi.org/10.1029/2003JC002131 562 Galgani, L., Engel, A., 2016. Changes in optical characteristics of surface microlayers hint to 563 photochemically and microbially mediated DOM turnover in the upwelling region off the coast of 564 Peru. Biogeosciences 13(8), 2453-2473. https://doi.org/10.5194/bg-13-2453-2016 Garrett, W.D., 1965. Collection of slick-forming materials from the sea surface. Limnol. Oceanogr. 565 566 10(1965), 602-605. https://doi.org/10.2307/2833459 Gonsior, M., Peake, B.M., Cooper, W.T., Podgorski, D., D'Andrilli, J., Cooper, W.J., 2009. 567 568 Photochemically induced changes in dissolved organic matter identified by ultrahigh resolution 569 fourier transform ion cyclotron resonance mass spectrometry. Environ. Sci. Technol. 43(3), 570 698-703. https://doi.org/10.1021/es8022804 571 Grasshoff, K., Kremling, K., Ehrhardt, M., 2007. Methods of Seawater Analysis, 3rd. Edition. pp. 572 407-420. 573 Guo, W., Yang, L., Zhai, W., Chen, W., Osburn, C.L., Huang, X., Li, Y., 2014. Runoff-mediated 574 seasonal oscillation in the dynamics of dissolved organic matter in different branches of a large bifurcated estuary-The Changjiang Estuary. J. Geophys. Res.-Biogeosciences 119, 776-793. 575 576 https://doi.org/10.1002/2013JG002540 577 Hardy, J.T., 1982. The sea surface microlayer: Biology, chemistry and anthropogenic enrichment. Prog. 578 Oceanogr. 11(4), 307-328. https://doi.org/10.1016/0079-6611(82)90001-5 579 Hardy, J.T., Apts, C.W., 1989. Photosynthetic carbon reduction: high rates in the sea-surface





580	microlayer. Mar. Biol. 101(3), 411-417. https://doi.org/10.1007/BF00428138										
581	Helms, J.R., Stubbins, A., Ritchie, J.D., Minor, E.C., Kieber, D.J., Mopper, K., 2008. Absorption										
582	spectral slopes and slope ratios as indicators of molecular weight, source, and photobleaching of										
583	chromophoric dissolved organic matter. Limnol. Oceanogr. 53(3), 955–969.										
584	https://doi.org/10.4319/lo.2008.53.3.0955										
585	Hoge, F.E., Vodacek, A., Blough, N.V., 1993. Inherent optical properties of the ocean: retrieval of the										
586	absorption coefficient of chromophoric dissolved organic matter from fluorescence measurements.										
587	Limnol. Oceanogr. 38(7), 1394–1402. https://doi.org/10.4319/lo.1993.38.7.1394										
588	Jarvis, N.L., 1967. Adsorption of surf ace-active material at the sea-air interface, Limnol. Oceanogr. 12,										
589	213–221. https://doi.org/10.4319/lo.1967.12.2.0213										
590	Jørgensen, L., Stedmon, C.A., Kragh, T., Markager, S., Middelboe, M., Søndergaard, M., 2011. Global										
591	trends in the fluorescence characteristics and distribution of marine dissolved organic matter. Mar.										
592	Chem. 126(1-4), 139-148. https://doi.org/10.1016/j.marchem.2011.05.002										
593	Kieber, D.J., Mcdaniel, J., Mopper, K., 1989. Photochemical source of biological substrates in sea										
594	water: implications for carbon cycling. Nature 341(6243), 637–639.										
595	https://doi.org/10.1038/341637a0										
596	Kok, M., Schouten, S., Sinninghe Damsté J.S., 2000. Formation of insoluble, nonhydrolyzable,										
597	sulfur-rich macromolecules via incorporation of inorganic sulfur species into algal carbohydrates.										
598	Geochim. Cosmochim. Acta 64(15), 2689–2699. https://doi.org/10.1016/S0016-7037(00)00382-3										
599	Liss, P.S., Duce, R.A., 1997. The Sea Surface and Global Change, Cambridge University Press, 519.										
600	Liss, P.S., Duce, R.A., 2005. The Sea Surface and Global Change. Cambridge University Press: UK.										





601 Liu, S.M., Qi, X.H., Li, X.N., Ye, H.R., Wu, Y., Ren, J.L., Zhang, J., Xu, W.Y., 2016. Nutrient 602 dynamics from the Changjiang (Yangtze River) estuary to the East China Sea. J. Mar. Syst. 603 154(Feb.), 15-27. https://doi.org/10.1016/j.jmarsys.2015.05.010 604 Liu, X., Beusen, A.H.W., Van Beek, L.P.H., Mogollón, J.M., Ran, X., Bouwman, A.F., 2018. Exploring 605 spatiotemporal changes of the Yangtze River (Changjiang) nitrogen and phosphorus sources, 606 retention and export to the East China Sea and Yellow Sea. Water Res. 142(Oct. 1), 246-255. 607 https://doi.org/10.1016/j.watres.2018.06.006 608 Margolin, A.R., Gonnelli, M., Hansell, D.A., Santinelli, C., 2018. Black sea dissolved organic matter 609 dynamics: insights from optical analyses. Limnol. Oceanogr. 63(3), 1425-1443. 610 https://doi.org/10.1002/lno.10791 Marie, D., Partensky, F., Jacquet, S., Vaulot, D., 1997. Enumeration and cell cycle analysis of natural 611 612 populations of marine picoplankton by flow cytometry using the nucleic acid stain SYBR Green I. 613 Appl. Environ. Microbiol. 63(1), 186–193. https://doi.org/10.1109/50.337494 Massicotte, P., Asmala, E., Stedmon, C., Markager, S., 2017. Global distribution of dissolved organic 614 615 matter along the aquatic continuum: Across rivers, lakes and oceans. Sci. Total Environ. 609(Dec. 616 31), 180-191. https://doi.org/10.1016/j.scitotenv.2017.07.076 617 Mcknight, D.M., Boyer, E.W., Westerhoff, P., Doran, P.T., Kulbe, T., Andersen, D.T., 2001. 618 Spectrofluorometric characterization of dissolved organic matter for indication of precursor 619 organic material and aromaticity. Limnol. Oceanogr. 46(1), 38-48. 620 https://doi.org/10.4319/lo.2001.46.1.0038 621 Medeiros, P.M., Seidel, M., Ward, N.D., Carpenter, E.J., Gomes, H.R., Niggemann, J., Dittmar, T., 622 2015. Fate of the Amazon River dissolved organic matter in the tropical Atlantic Ocean. Global 623 Biogeochem. Cycl. 29(5), 677-690. https://doi.org/10.1002/2015GB005115





624 Mopper, K., Kieber, D.J., 2002. Photochemistry and the cycling of carbon, sulfur, nitrogen and 625 phosphorus. In: Hansell DA, Carlson CA (eds) Biogeochemistry of dissolved organic matter. 626 Academic Press, San Diego, pp. 455-507. 627 Momzikoff, A., Brinis, A., Dallot, S., Gondry, G., Saliot, A., Lebaron, P., 2004. Field study of the 628 chemical characterization of the upper ocean surface using various samplers. Limnol. Oceanogr. Methods 2(11), 374-384. https://doi.org/10.4319/lom.2004.2.374 629 630 Nichols, C.M., Lardière, S.G., Bowman, J.P., Nichols, P.D., Gibson, J.A.E., Guézennec, J., 2005. 631 Chemical Characterization of Exopolysaccharides from Antarctic Marine Bacteria. Microb. Ecol. 632 49(4), 578-589. https://doi.org/10.1007/s00248-004-0093-8 633 Obernosterer, I., Catala, P., Reinthaler, T., Herndl, G.J., Lebaron, P., 2005. Enhanced heterotrophic 634 activity in the surface microlayer of the Mediterranean Sea. Aquat. Microb. Ecol. 39(3), 293-302. 635 https://doi.org/10.3354/ame039293 Ogawa, H., Amagai, Y., Koike, I., Kaiser, K., Benner, R., 2001. Production of refractory dissolved 636 637 292(5518), 917-920. organic matter by bacteria. Science 638 https://doi.org/10.1126/science.1057627 639 Orellana, M.V., Matrai, P.A., Leck, C., Rauschenberg, C.D., Lee, A.M., Coz, E., 2011. Marine 640 microgels as a source of cloud condensation nuclei in the high Arctic. Proc. Natl. Acad. Sci. 641 108(33), 13612-13617. https://doi.org/10.1073/pnas.1102457108 642 Ortega-Retuerta, E., Passow, U., Duarte, C.M., Reche, I., 2009. Effects of ultraviolet B radiation on 643 (not so) transparent exopolymer particles. Biogeosciences 6(12),3071-3080. 644 https://doi.org/10.5194/bg-6-3071-2009 645 Osburn, C.L., Handsel, L.T., Mikan, M.P., Paerl, H.W., Montgomery, M.T., 2012. Fluorescence





646	tracking of dissolved and particulate organic matter quality in a river-dominated estuary. Environ.
647	Sci. Technol. 46(16), 8628–8636. https://doi.org/10.1021/es3007723
648	Parsons, T.R., Matia, Y., Lalli, C.M., 1984. A Manual of Chemical and Biological Methods for
649	Seawater Analysis. Pergamon Press, Oxford.
650	Reinthaler, T., Sintes, E., Herndl., G.J., 2008. Dissolved organic matter and bacterial production and
651	respiration in the sea-surface microlayer of the open Atlantic and the western Mediterranean sea.
652	Limnol. Oceanogr. 53(1), 122-136. https://doi.org/10.4319/lo.2008.53.1.0122
653	Romera-castillo, C., Sarmento, H., Alvarezsalgado, X.A., Gasol, J.M., Marrase, C., 2010. Production
654	of chromophoric dissolved organic matter by marine phytoplankton. Limnol. Oceanogr. 55(1),
655	446–454. https://doi.org/10.4319/lo.2010.55.1.0446
656	Siegel, D.A., 2005. Colored dissolved organic matter and its influence on the satellite - based
657	characterization of the ocean biosphere. Geophys. Res. Lett. 32(20), 469-496.
658	https://doi.org/10.1029/2005GL024310
659	Singh, S., D'Sa, E., Swenson, E., 2010. Seasonal variability in CDOM absorption and fluorescence
660	properties in the Barataria Basin, Louisiana, USA. J. Environ. Sci. 22(10), 1481-1490.
661	https://doi.org/10.1016/S1001-0742(09)60279-5
662	State Bureau of Technical Supervision Bureau, 1992. Specifications for Oceanographic Survey-Survey
663	of Biology in Sea Water. Standard Press of China, Beijing, pp. 17-20.
664	Stedmon, C.A., Bro, R., 2008. Characterizing dissolved organic matter fluorescence with parallel
665	factor analysis: a tutorial. Limnol. Oceanogrmethods 6(11), 572–579.
666	https://doi.org/10.4319/lom.2008.6.572b
667	Stedmon, C.A., Markager, S., 2005. Resolving the variability in dissolved organic matter fluorescence
668	in a temperate estuary and its catchment using PARAFAC analysis. Limnol. Oceanogr. 50(2),
669	686–697. https://doi.org/10.4319/lo.2005.50.2.0686





670 Stedmon, C.A., Markager, S., Bro, R., 2003. Tracing dissolved organic matter in aquatic environments 671 using a new approach to fluorescence spectroscopy. Mar. Chem. 82(3-4), 239-254. https://doi.org/10.1016/s0304-4203(03)00072-0 672 673 Stedmon, C.A., Markager, S., Tranvik, L., Kronberg, L., Slätis, T., Martinsen, W., 2007. 674 Photochemical production of ammonium and transformation of dissolved organic matter in the Baltic Sea. Mar. Chem. 104(3-4), 227-240. https://doi.org/10.1016/j.marchem.2006.11.005 675 676 Su, Y., Hu, E., Feng, M., Zhang, Y., Chen, F., Liu, Z., 2017. Comparison of bacterial growth in 677 response to photodegraded terrestrial chromophoric dissolved organic matter in two lakes. Sci. 678 Total. Environ. 579(Feb. 1), 1203-1214. https://doi.org/10.1016/j.scitotenv.2016.11.104 679 Sun, H., Zhang, Y.H., Tan, S., Zheng, Y.F., Zhou, S., Ma, Q.Y., Yang, G.P., Todd, J., Zhang, X.H., 2020. 680 DMSP-Producing Bacteria Are More Abundant in the Surface Microlayer than Subsurface 681 Seawater of the East China Sea. Microb. Ecol. 80(1), 350-365. https://doi.org/10.1007/s00248-020-01507-8 682 683 Wang, F., Feng, T., Guo, Z., Li, Y., Lin, T., Rose, N.L., 2019. Sources and dry deposition of 684 carbonaceous aerosols over the coastal East China Sea: Implications for anthropogenic pollutant 685 pathways and deposition. Environ. Pollut. 245(Feb.), 771-779. 686 https://doi.org/10.1016/j.envpol.2018.11.059 687 Weishaar, J.L., Aiken, G.R., Bergamaschi, B.A., Fram, M.S., Fujii, R., Mopper, K., 2003. Evaluation 688 of specific ultraviolet absorbance as an indicator of the chemical composition and reactivity of 689 dissolved organic carbon. Environ. Sci. Technol. 37(20), 4702-4708. https://doi.org/10.1021/es030360x 690 691 Weng, H., Tian, R., Ji, Z., Yu, X., 2011. Potential relationships between atmospheric particulate matter





692	transported by winter monsoons and red tides in the East China Sea. Sci. Bull. 56(3), 297-305.
693	https://doi.org/10.1007/s11434-010-4209-x
694	Wotton, R.S., Preston, T.M., 2005. Surface Films: Areas of Water Bodies That Are Often Overlooked.
695	Bioscience 55(2), 137-145. https://doi.org/10.1641/0006-3568(2005)055[0137:SFAOWB]2.0.CO
696	Wurl, O., Wurl, E., Miller, L.A., Johnson, K., Vagle, S., 2011. Formation and global distribution of
697	sea-surface microlayers. Biogeosciences 8(1), 121–135. https://doi.org/10.5194/bg-8-121-2011
698	Wurl, O., Holmes, M., 2008. The gelatinous nature of the sea-surface microlayer. Mar. Chem.
699	110(1–2), 89–97. https://doi.org/10.1016/j.marchem.2008.02.009
700	Yamashita, Y., Hashihama, F., Saito, H., Fukuda, H., Ogawa, H., 2017. Factors controlling the
701	geographical distribution of fluorescent dissolved organic matter in the surface waters of the
702	Pacific Ocean. Limnol. Oceanogr. 62(6), 2360–2374. https://doi.org/10.1002/lno.10570
703	Yamashita, Y., 2004. In situ production of chromophoric dissolved organic matter in coastal
704	environments. Geophys. Res. Lett. 31(14), 189-207. https://doi.org/10.1029/2004GL019734
705	Yang, L., Zhang, J., Yang, G.P., 2020. Mixing behavior, biological and photolytic degradation of
706	dissolved organic matter in the East China Sea and the Yellow Sea. Sci. Total Environ. 762(6),
707	143164. https://doi.org/10.1016/j.scitotenv.2020.143164
708	Yang, L., Zhuang, W., Chen, C.A., Wang, B., Kuo, F., 2017. Unveiling the transformation and
709	bioavailability of dissolved organic matter in contrasting hydrothermal vents using fluorescence
710	EEM-PARAFAC. Water Res. 111(Mar. 15), 195–203.
711	https://doi.org/10.1016/j.watres.2017.01.001
712	Zepp, R.G., Sheldon, W.M., Moran, M.A., 2004. Dissolved organic fluorophores in southeastern US
713	coastal waters: correction method for eliminating Rayleigh and Raman scattering peaks in





714	excitation-emission	matrices.	Mar.	Chem.	89(1),	15–36.
715	https://doi.org/10.1016/j.a	marchem.2004.02.	006			
716	Zhu, W.Z., Yang, G., Zhang,	H., 2017. Photoc	hemical bel	havior of dissol	ved and colloi	dal organic
717	matter in estuarine and	l oceanic waters.	Sci. Total	Environ. s607-	-608(Dec. 31)	, 214–224.
718	https://doi.org/10.1016/j.s	scitotenv.2017.06.1	163			





 Table 1 Spectral characteristics of the four fluorescent components identified by the PARAFAC

modal in this study, compared with those preciously identified.

Component	Ex <sub>max</sub> (nm)	Em <sub>max</sub> (nm)	Coble (1996)	Comparison with other studies using PARAFAC	Description and probable source
C1	345	455	peak C 320-360/42 0-480	Osburn et al. (2012)	Terrestrial-like humic substances
C2	255	375 (310)	peak A 230-260/380 -460	Stedmon et al. (2003)	Tyrosine-like substances
СЗ	315	385	peak M 290-310/370 -420	Stedmon and Markager (2005)	Marine humic-like substances (biological degradation)
C4	280	335	peak T 270-280/340 -350	Coble (1996)	Tryptophan-like; Non-Humic-like; Biological production in the water column





**Table 2** Average temperature, salinity, wind speed, CDOM a(254), DOC, Chlorophyll-*a* (Chl-*a*), dissolved oxygen (DO), S<sub>275-295</sub>, S<sub>R</sub>, and SUVA<sub>254</sub> of the SSW and SML in the BS, YS and ECS during spring, summer, autumn, and winter.

		Spring		Sun	nmer	Aut	umn	Win	nter	
	Water layer	mean	SD	mean	SD	mean	SD	mean	SD	
Temperature (°C)	SSW	14.0	4.91	24.0	3.66	13.7	2.69	14.0	5.23	
Salinity	SSW	32.5	1.92	31.7	2.17	30.7	1.10	32.7	1.41	
Wind Speed (m s <sup>-1</sup> )	SSW	5.98	2.86	5.47	2.51	7.30	3.82	6.09	2.52	
DO (mg L <sup>-1</sup> )	SSW	6.44	0.85	7.57	1.07	7.49	0.51	8.32	0.99	
Chl-a (μg L <sup>-1</sup> )	SSW	1.26	2.38	1.13	1.48	0.74	0.36	0.42	0.25	
Спі-и (µg L )	SML	1.63	3.66	1.28	1.13	0.61 0.29	0.29	no data		
DOC (µmol L <sup>-1</sup> )	SSW	91.3	25.7	109.4	33.55	151.2	83.8	88.4	88.4 22.51	
DOC (µmor L )	SML	132.9	77.4	145.7	49.8	146	33.5	131.3	91.1	
a(254) (m <sup>-1</sup> )	SSW	3.20	2.49	3.10	1.34	5.21	1.84	2.52	1.26	
a(234) (III )	SML	3.70	1.98	4.05	1.66	5.51	1.82	4.74	2.50	
S <sub>275-295</sub> (nm <sup>-1</sup> )	SSW	0.0201	0.0049	0.0188	0.0035	0.0214	0.0075	0.0207	0.0068	
32/5-295 (IIIII )	SML	0.0222	0.0073	0.0178	0.0021	0.0207	0.0067	0.021	0.0055	
<b>C</b> -	SSW	1.723	1.026	1.731	1.557	1.493	1.312	1.521	0.52	
$S_R$	SML	1.095	0.218	1.361	0.296	1.357	0.772	1.416	0.214	
SUVA <sub>254</sub>	SSW	2.067	0.664	2.244	0.671	3.008	0.949	3.008	0.949	
(L mg-C <sup>-1</sup> m <sup>-1</sup> )	SML	1.911	0.768	1.951	0.359	3.196	1.126	2.992	1.034	





**Table 3** Correlation coefficients between EF of DOM optical properties, Chl-a, DOC, PO<sub>4</sub><sup>3</sup>, NO<sub>3</sub>,

NO<sub>2</sub>-, SiO<sub>3</sub><sup>2</sup>-, Cyanobacteria, Picophytoplankton, Bacterial abundance.

	EF of	EF of	EF of	EF of	EF of	EF of							
	a(254)	DOC	Chl-a	C1	C2	C3	C4	PO <sub>4</sub> 3-	NO <sub>3</sub> -	$NO_2$	SiO <sub>3</sub> <sup>2-</sup>	Cyanobacteria	picophytoplankton
EF of DOC	0.185										•		
EF of Chl-a	0.092	0.021											
Ef of C1	.336**	0.047	-0.119										
EF of C2	0.163	0.073	-0.017	.635**									
EF of C3	.413**	0.179	-0.096	.907**	.557**								
EF of C4	.319**	0.021	0.011	.574**	.368**	.628**							
EF of PO <sub>4</sub> <sup>3-</sup>	0.129	0.267	.319*	0.131	-0.037	0.139	0.087						
EF of NO <sub>3</sub>	-0.065	-0.054	0.235	-0.037	-0.044	-0.027	-0.053	0.26					
EF of NO <sub>2</sub> -	0.15	0.208	.307*	0.142	-0.017	0.192	0.035	.571**	0.271				
EF of SiO <sub>3</sub> <sup>2-</sup>	.634*	0.004	0.074	0.122	-0.101	0.305	0.151	0.205	-0.118	-0.141			
EF of													
Cyanobacteria	0.091	-0.017	0.028	-0.027	-0.105	-0.047	-0.052	0.218	0.027	.755**	-0.286		
EF of													
Picophytoplankton	.347**	0.0281	0.252	-0.067	-0.082	-0.077	0.025	0.218	-0.08	0.113	-0.327	0.081	
EF of													
Bacterial													
abundance	-0.036	-0.069	-0.063	-0.061	0.004	0.014	-0.093	-0.12	-0.026	-0.073	-0.099	.730**	-0.064

<sup>\*\*</sup> Correlation is significant at the 0.01 level (two-tailed)

<sup>\*</sup> Correlation is significant at the 0.05 level (two-tailed)