

# Growth and actual leaf temperature modulate CO<sub>2</sub>-responsiveness of monoterpene emissions from Holm oak in opposite ways

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**Abstract.** Climate change can profoundly alter VOC emissions from vegetation and thus influence climate evolution. Yet, the short and long-term effects of elevated CO<sub>2</sub> concentrations on emissions in interaction with temperature are not enough understood, especially for VOCs other than isoprene. To gain additional insight, we conducted a study on holm oak, which is known for its strong foliar monoterpene emissions that are directly linked to their synthesis. We measured CO<sub>2</sub>-response curves of ~~leaf monoterpene emissions, CO<sub>2</sub>/H<sub>2</sub>O gas exchanges and chlorophyll fluorescence and photosynthetic parameters~~ at two assay temperatures (30 and 35°C) on saplings of four populations of holm oak saplings grown under normal and double CO<sub>2</sub> concentrations combined with two temperature growth regimes differing by 5 °C (day/night: 25/15 and 30/20 °C). A stepwise reduction in CO<sub>2</sub> resulted in a decrease in emissions, occasionally preceded by an increase, with the overall decrease in emissions being greater at 35 °C than at 30 °C assay temperature. During ramping to high CO<sub>2</sub>, emissions remained mostly unchanged at 35 °C, whereas at 30 °C they often dropped, especially at the highest CO<sub>2</sub> levels (≥ 1200 ppm). In addition to the actual leaf temperature, the high CO<sub>2</sub>-responsiveness of emissions was modulated by the plant's growth temperature with warm-grown plants being more sensitive than cool-crown plants. In contrast, growth CO<sub>2</sub> had no significant effect on the emission-CO<sub>2</sub> sensitivity of emissions, although it promoted plant growth and the leaf's emission factor. Correlation analyses suggest that the emission response to CO<sub>2</sub> depended primarily on the availability of energetic-cofactors produced by photosynthetic electron transport. This availability was likely limited by different processes that occurred during CO<sub>2</sub>-ramping including photooxidative stress and induction of protective and repair mechanisms as well as -competition with CO<sub>2</sub>-fixation and photorespiration. In addition, feedback inhibition of photosynthesis may have played a role, especially in leaves, whose emissions were inhibited only at very high CO<sub>2</sub> levels. Correlation analyses suggest that the emission response to low CO<sub>2</sub> depended on the leaf's initial carbon balance and its actual energy status, whereas the response to high CO<sub>2</sub> depended only on the leaf's actual energy status, which was affected by the occurrence of photooxidative stress and feedback limitation of photosynthesis. Overall, our results confirm an isoprene-analogous behavior of monoterpene emissions from holm oak. Emissions exhibit a nonlinear response curve to CO<sub>2</sub> similar to that currently used for isoprene emission in the MEGAN model, with no difference between major individual monoterpene species and plant chemotype. Simulations estimating the annual VOC releases from holm oak leaves at double atmospheric CO<sub>2</sub> indicate that the observed high-CO<sub>2</sub> inhibition is unlikely to offset the increase in emissions due to the expected-predicted warming.

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## 1 Introduction

Terrestrial vegetation has been identified as the main source of biogenic volatile organic compounds (BVOCs). Besides their multiple biological functions, BVOCs influence several climate forcing components in the atmosphere, notably the concentrations of the greenhouse gases methane and ozone, and in pristine environments the formation of secondary organic aerosols (Fuentes et al., 2001; Arneth et al., 2010). Aerosols can have a cooling effect on the earth climate by increasing the diffusive fraction of radiation and by changing cloud properties in the atmosphere (Zhu et al., 2019; Yli-Juuti et al., 2021). Furthermore, increased diffusive light can favor photosynthesis possibly enhancing carbon sequestration in forest ecosystems (Ezhova et al., 2018; Rap et al., 2018). Consequently, large-scale alteration of BVOC emissions due to global change could feedback on future climate evolution (Scott et al., 2018; Sporre et al., 2019). Yet, the interacting effects of climate change factors on emissions are currently not enough understood. Among these, increasing atmospheric concentrations of carbon dioxide [CO<sub>2</sub>] and temperature are major factors with no or relative weak regional differences compared to other factors such as precipitation.

Globally, volatile isoprenoids constitute the largest fraction of BVOCs (Guenther et al., 2012). Isoprene alone accounts about the half, which is produced at high rates in the photosynthesizing tissues of about 30% of vascular plant species with a higher presence in woody species than in herbs (Monson et al., 2012; Fineschi et al., 2013; Sharkey et al., 2013; Dani et al., 2014a). Accordingly, most studies have focused on isoprene emissions (for recent reviews see Sharkey and Monson, 2014; Lantz et al., 2019a; Monson et al. 2021). Isoprene is synthesized in chloroplasts from dimethylallyl diphosphate (DMADP) by isoprene synthase, an enzyme with relatively low affinity for its substrate (Lehning et al., 1999; Sharkey et al., 2013). Isoprene synthesis and emissions are strongly regulated by temperature and light, which is mainly related to changes in the pool size of DMADP. DMADP is built in the chloroplastic methyl-D-erythritol-4-phosphate (MEP) pathway from the C<sub>3</sub>-substrates glyceraldehyde-3-phosphate (triose-phosphate) and pyruvate, plus reduction power (NADPH or equivalents) and phosphorylation power (ATP or equivalents) as energetic cofactors. Glyceraldehyde-3-phosphate and energetic cofactors come directly from ongoing photosynthesis, while a large fraction of pyruvate is formed from phosphoenolpyruvate (PEP), which is imported from the cytosol in exchange of inorganic phosphate (Pi) and thus can stem from “older” carbon sources (Lantz et al. 2019a and references therein). Regarding CO<sub>2</sub>-effects, many studies observed that isoprene emissions decrease rapidly at high [CO<sub>2</sub>] (e.g., Monson and Fall, 1989; Loreto and Sharkey, 1990; Monson et al., 1991; Rasulov et al., 2009; Wilkinson et al., 2009; Possell and Hewitt, 2011; Morfopoulos et al., 2014), potentially counteracting the expected increase in emissions from rising temperatures and CO<sub>2</sub> fertilization on plant growth in a future warmer, high-CO<sub>2</sub> world (Pacifico et al., 2012; Bauwens et al. 2018). This CO<sub>2</sub>-responsiveness however can be variable from one species to another (e.g. Sharkey et al., 1991; Lantz et al., 2019b; Niinemets et al., 2021), and be modulated by the atmospheric CO<sub>2</sub>-regimes, in which the plants were grown or acclimated (e.g. Wilkinson et al., 2009; Possell and Hewitt, 2011; Sun et al., 2013). Further, it depends on the actual leaf temperature; high temperatures generally suppress the high-CO<sub>2</sub> sensitivity of isoprene emissions (e.g. Affek and Yakir, 2002; Rasulov et al., 2010; Sun et al., 2013; Potosnak et al., 2014, Dani et al., 2014b; Monson et al., 2016).

Monoterpenes (MTs) are less emitted globally (<15%; Guenther et al., 2012). However, in some vegetation types such as boreal, temperate mountainous and Mediterranean forests, MTs can largely dominate the total BVOC release (e.g. Rantala et al., 2015; Seco et al., 2017; Tani and Mochizuki, 2021 and references therein), and due to their bigger size and high reactivity, these emissions might be particularly relevant for local and regional SOA formation (Jokinen et al., 2015; Zhang et al., 2018; McFiggans et al., 2019). MTs are essentially produced in the same pathway as isoprene (but see Pazouki and Niinemets, 2016 for exceptions). However, the responses of MT emissions to elevated [CO<sub>2</sub>] are less understood and show more contrasting results ranging from no effect, increases and decreases (Arneeth et al., 2008; Peñuelas and Staudt, 2010; Feng et al., 2019; Daussy and Staudt, 2020). There are several reasons why MT emissions may behave differently than isoprene. First, in many plants MTs are synthesized outside photosynthetic source tissues in glandular organs (trichomes, resin ducts), where there can be accumulated in high concentrations (e.g. Huang et al., 2018; Dehimeche et al., 2021). As a result, emission rates vary independent from their biosynthesis rates, which furthermore, is less coupled to ongoing photosynthesis due to additional regulatory processes associated with the partitioning and transport of photosynthates. Second, the biosynthesis of MTs involves at least two other enzymes (i.e. one MT synthase and one geranyl diphosphate synthase) whose in-plant catalytic rates may be less substrate regulated than isoprene due to their high substrate affinities (Harrison et al., 2013; Rasulov et al., 2014 and references therein). In addition, MT emitters typically produce several MTs formed by several MT synthases thus introducing further complexity in their responses to [CO<sub>2</sub>].

One of the best studied MT emitter is the Mediterranean evergreen oak *Quercus ilex* L. (QI, holm oak). Its strong MT emissions show much analogy to isoprene in terms of quantity and responses to environmental factors (Loreto et al., 1996a; Staudt and Bertin, 1998). Regarding CO<sub>2</sub> effects on emissions, two studies reported that emissions become significantly inhibited at high ~~very~~[CO<sub>2</sub>] but not at moderately increased [CO<sub>2</sub>] (Loreto et al., 1996b; Staudt et al., 2001). Yet, Loreto et al., (2001) compared emissions from QI trees growing in open top chambers with normal and double [CO<sub>2</sub>] and concluded that 700 ppm [CO<sub>2</sub>] significantly inhibits the emissions some of the major MTs while enhancing others ~~in-consistence with differences in the tree's activity of MT synthases~~. However, because in that study emissions at different CO<sub>2</sub> levels were determined on different trees, the seeming compound specificity could be confounded with the chemotype of the tree, a possible misinterpretation mentioned by the authors. Later, Rapparini et al. (2004) investigated emissions from QI trees growing near natural CO<sub>2</sub> springs. Switching from 350 ppm to 1000 ppm [CO<sub>2</sub>] reduced all emissions in the control site but not in the elevated CO<sub>2</sub> site. They also found unexplained seasonal differences in CO<sub>2</sub> responsiveness possibly associated with water stress. Long-term, seasonal CO<sub>2</sub> effects were also reported by Staudt et al. (2001) who observed that the emission factor (EF, i.e. the foliar emission rate under standard temperature, light and [CO<sub>2</sub>]) of elevated CO<sub>2</sub>-grown plants was significantly increased during the winter season but not ~~before~~ during the warm season. Thus, while previous studies provide evidence that MT emissions from QI can be inhibited by elevated [CO<sub>2</sub>], the exact CO<sub>2</sub>-response, its compound specificity, and its dependence on actual temperature and growth conditions are unclear or not known. To gain additional insight, we conducted a study in which we measured CO<sub>2</sub> response curves of foliar MT emissions, CO<sub>2</sub>/H<sub>2</sub>O gas exchanges, and chlorophyll fluorescence at two assay temperatures on four QI populations that were grown under two [CO<sub>2</sub>] in combination with two temperature regimes. In particular, we addressed the following

questions: Do MT emissions respond to low and/or high [CO<sub>2</sub>] and how are these responses related to the leaf's emission factor and primary metabolism? Does the CO<sub>2</sub>-responsiveness differ between individual MT compounds? Is the CO<sub>2</sub>-responsiveness affected by the actual leaf temperature? Do seedlings grown under warmer and/or higher CO<sub>2</sub> regimes differ in their leaf emission factors and/or responsiveness to CO<sub>2</sub>? Are the observed effects relevant for estimating MT emissions in a future warmer and CO<sub>2</sub>-enriched world?

## 2 Material and methods

### 2.1 Plants and growth conditions

105 QI acorns were sampled in fall from various adult trees growing around Montpellier. They were stored in boxes with humidified paper tissues in a cold chamber at 5°C, where they started to germinate during winter and early spring. The germinating acorns were subsequently potted in PVC-pipes (diameter 16 cm, height 100 cm) containing a mix of sand, clay and peat, and were immediately transferred in four identical, controlled-environment greenhouse compartments. Temperature regimes and atmospheric [CO<sub>2</sub>] of the compartments were set to 400 ppm CO<sub>2</sub> and 15/25 °C night/day temperature (400/20),  
110 800 ppm CO<sub>2</sub> and 15/25 °C (800/20), 400 ppm CO<sub>2</sub> and 20/30 °C, (400/25) and 800 ppm CO<sub>2</sub> and 20/30 °C (800/25). Currently in the Montpellier region, a temperature range of 15/25 °C is common in late spring when new leaf growth is at its peak, whereas the 20/30 °C is typical for the summer (see table S1 in supplement 1). However, in a future atmosphere with 800 ppm [CO<sub>2</sub>], such high temperatures are expected to occur more frequently outside the summer season especially in the Mediterranean, where climate warming proceeds faster than global average (Seneviratne et al., 2016). Plants were grown in  
115 these compartments under the same temperature and CO<sub>2</sub> regimes for 4-5 months until measurements started. The greenhouse facility of our institute consists of eight serial compartments in about east-west direction with the southern facades exposed to a large open field (grassland). To avoid any edge effect and uneven light exposure, the four inner compartments were used for the experiment. In addition, plants were regularly moved within the greenhouse compartments as well as between greenhouse compartments after having interchanged the growth temperature and CO<sub>2</sub> regimes.

### 120 2.2 CO<sub>2</sub>-response curve measurements

Leaf MT emissions (E), CO<sub>2</sub>/H<sub>2</sub>O gas exchange and chlorophyll fluorescence were measured using two LI-6400 Portable Photosynthesis Systems (LI-COR Biosciences, Lincoln, NE, USA). The large majority of measurements were made with the small 2 cm<sup>2</sup> Li6400-40 leaf chamber equipped with a blue/red LED light source and an integrated [chlorophyll](#) fluorometer. At the beginning of the experiment, few additional measurements were conducted with a 6 cm<sup>2</sup> broadleaf chamber equipped with a LED source but without fluorometer. For measuring the CO<sub>2</sub> response of VOC emission, a mature, healthy leaf of a sapling was gently clamped into the chamber and subsequently exposed to seven [ambient-\[CO<sub>2</sub>\]](#)CO<sub>2</sub> levels in the following order: 400, 200, 100, 800, 1200, 1600, 2000 ppm. The flow rate of air was set at 300 μmol s<sup>-1</sup> (ca. 450 ml min<sup>-1</sup>), [the](#) photosynthetic photon flux density (PPFD) at 1000 μmol m<sup>-2</sup> s<sup>-1</sup> (10% blue, 90% red LEDs) and [the chamber](#) block temperature at either 30

or 35 °C. Leaves were ~~equilibrated-acclimatized~~ at every ~~[CO<sub>2</sub>]~~~~CO<sub>2</sub>-level~~ for at least 30 min before starting data recording and  
130 VOC sampling. We applied a relative long waiting periods compared to analog studies on isoprene (e.g. Monson et al., 2016;  
Lantz et al., 2019b), because MT emissions need longer to come to a new steady state due to their lower volatility (Niinemets  
et al., 2002a; Staudt et al., 2003). BVOCs were sampled with a programmable air sampler (Gillian GilAir Plus, Sensidyne LP,  
USA) passing 2.4 L of chamber air at 150 ml min<sup>-1</sup> through adsorption cartridges packed with about 180 mg Tenax TA and  
135 130 mg Carbotrap B. The chamber air was taken from the air hose connecting the chamber and match valve via a 3-way Teflon  
valve (Bola, Bohlender GmbH, Germany). CO<sub>2</sub>/H<sub>2</sub>O gas exchange and chlorophyll fluorescence data were recorded before  
and after VOC sampling surveying each time that gas exchange was stable. The mean values of both records were used for  
further data evaluation. All photosynthetic variables (net CO<sub>2</sub>-assimilation (A), transpiration, conductance to water vapor (G),  
substomatal [CO<sub>2</sub>] (Ci)) were calculated by the LiCOR software including corrections for diffusion leaks as recommended for  
small leaf chambers. ~~In a few occasions when the 6 cm<sup>2</sup> leaf chamber was used, the enclosed leaf did not completely cover the  
140 chamber surface. In this case, we measured the projected surface area of the enclosed leaf part and recalculated the gas  
exchange variables accordingly, and if necessary for leaf area. In fact a few times, when the 6 cm<sup>2</sup>-leaf chamber was used, the  
enclosed leaf did not completely cover the chamber surface. In that case, we marked the enclosed leaf part and measured  
separately its projected surface.~~

The integrated leaf chamber fluorimeter determined the actual quantum efficiency of photosystem II (PSII) electron transport  
145 in the light (ΦPSII) by measuring first the steady-state fluorescence (Fs) of the light adapted leaf and then the maximum  
fluorescence (Fm') by applying a saturating light pulse of ca. 10000 μmol m<sup>-2</sup> s<sup>-1</sup> PPFD (ΦPSII = (Fm'-Fs) Fm<sup>-1</sup>) (Murchie  
and Lawson, 2013). ΦPSII is proportional to the flow of electrons in PSII (electron transport rate, ETR), which was calculated  
by multiplying ΦPSII with PPFD assuming that 87\_% of the incident PPFD was absorbed by QI leaves, of which half is  
attributed to PSII (i.e. ETR = ΦPSII \* PPFD \* 0.87 \* 0.5). It should be noted that these correction factors may somewhat have  
150 varied among individual leaves and measurements, for example due to differences in the leaf's structure and light acclimation  
(Laisk and Loreto, 1996; Niinemets et al., 2006; McClain and Sharkey, 2019). However, leaf-to-leaf variation of calculated  
ETRs ~~showed no significant relation to the variation of~~~~was not related to~~ leaf structural variables (leaf ~~dry~~ mass per leaf area  
(LMA), chlorophyll concentration (R<sup>2</sup> < 0.1, P > 0.15)). In addition to measurements under light, foliar dark respiration and  
maximum quantum efficiency of PSII photochemistry (Fv/Fm) were measured at the beginning and at the end of CO<sub>2</sub>-ramping  
155 after having leaves adapted to dark and 400 ppm [CO<sub>2</sub>] for 30 min. Fv/Fm is given as (Fm-Fo) Fm<sup>-1</sup>, where Fo and Fm are,  
respectively, the steady-state and maximum fluorescence of the dark-adapted leaf. Fm and Fm' data were further used to  
calculate the non-photochemical quenching (NPQ) as (Fm'-Fm) Fm<sup>-1</sup>. NPQ reflects the fraction of absorbed light energy  
dissipated as heat from PSII. ~~NPQ reflects protective mechanisms against the absorption of excessive light energy, which  
otherwise leads to photodamage, as evidenced by a decrease in Fv/Fm. Photodamage is caused by the over-reduction of PSII  
160 along with the formation of reactive oxygen species (ROS: mainly singlet oxygen, superoxide, hydroxyl radicals and hydrogen  
peroxide; Asada, 2006). NPQ englobes protective mechanisms against excessive light, which otherwise leads to the over-~~

reduction of PSII along with the formation of Reactive Oxygen Species (ROS: mainly singlet oxygen, superoxide, hydroxyl radicals and hydrogen peroxide; Asada, 2006) and ultimately to the persistent photoinhibition of the photosystem as indicated by a drop in Fv/Fm.

NPQ processes are regulated by the acidification of the chloroplast thylakoid lumen leading to the activation of the integral membrane protein PsbS and the xanthophyll cycle, both triggering conformational changes in PSII antenna (Ruban, 2016).

Other derived variables considered in the data evaluation were the ETR/A, E/A and E/ETR ratios. ETR/A is the amount of electrons per net-assimilated CO<sub>2</sub>. Variation in ETR/A reflects the excess of photochemical energy produced via PSII not used for CO<sub>2</sub> reduction in the Calvin-Benson-Bassham (CBB) cycle, hence the amount of NADPH and ATP available for other metabolic pathways inside chloroplasts such as photorespiration, starch synthesis, nitrite reduction, Mehler reaction (oxygen reduction), xanthophyll cycle and isoprenoid biosynthesis. Under physiological normal conditions, about half of the ETR is used for CO<sub>2</sub> reduction (Dani et al., 2014b), with about four to five moles of electrons required per assimilated mole of CO<sub>2</sub>. The E/A and E/ETR ratios are respectively the percentage losses of assimilated carbon (C-loss) and PSII photosynthetic electron transport (é-loss) by MT emissions assuming that one mole emitted MT consumes 10 moles of assimilated carbon and 56 moles of electrons (28 moles NADPH or equivalents and 2 moles electrons per NADPH; Niinemets et al., 2002b).

All response curves were run in the greenhouse compartments. The air for the LicOR instrument was always taken from outside the greenhouse and filtered with charcoal to minimize [CO<sub>2</sub>] fluctuations and contamination with ambient VOC. The response curves at 30 and 35 °C were always measured on different mature leaves of a given sapling. Due to logistic constrains, the number of replicates per growth treatment varied between 5 and 8 at 30 °C and between 4 and 6 at 35 °C assay temperature (totally 26 CO<sub>2</sub>-response curves at 30 °C and 20 CO<sub>2</sub>-response curves at 35 °C on 26 saplings). We favored running more replicates at 30°C than at 35°C due to the stronger and more variable CO<sub>2</sub>-responsiveness at this assay temperature. On the whole, the measurement of a CO<sub>2</sub>-response curve took about 6 hours and was usually carried out between 10 am and 4 pm.

In order to check whether BVOC emissions from QI leaves changed during the day independently of external factors, we repeatedly measured emissions from QI leaves according the same protocol but without changing in the same time frame at constant assay [CO<sub>2</sub>], temperature or PPFD.

Additional ancillary measurements were made after each experiment: Relative chlorophyll contents of the measurement leaves were assessed using a SPAD-502 instrument (Minolta, Ltd, Japan). SPAD data were converted to foliar Chlorophyll concentration ([Chloro]) based on the calibration realized in a previous study on QI seedlings (Staudt et al., 2017). Further, projected leaf area were determined by means of a scanner plus image software (Epson perfection V800; Image J5 software, National Institutes of Health, Bethesda, MD, USA) and dry weights on a microbalance after oven drying at 60 °C for 48 hours. Plant growth was assessed by measuring the number of leaves and ramifications and total plant height.

### 2.3 BVOC analyses

Adsorption Cartridges were analyzed using a gas chromatograph coupled with mass spectrometer (GC-MS) Shimadzu QP2010 Plus equipped with a Shimadzu TD-20 thermodesorber (Shimadzu, Kyoto, Japan). Prior to analysis, cartridges were purged

195 for 1 min with dry N<sub>2</sub> at room temperature to remove excess water. BVOCs were thermally desorbed from cartridges at 250 °C in a 30 ml min<sup>-1</sup> He flow for 10 min on a cold trap filled with Tenax TA and maintained at - 10 °C. The focused VOCs were then thermally injected into the GC-column with a split ratio of 4 by flash heating the cold trap to 240 °C for 5 min. BVOCs were separated on a DB5 column (30 m x 0.25 mm, 0.25 μm film thickness) with helium as carrier gas (constant flow 1 ml min<sup>-1</sup>) using the following oven temperature program: 2 min at 40 °C, 5 °C min<sup>-1</sup> to 200°C, 10 °C min<sup>-1</sup> to 270 °C held for 6 min. Eluting BVOCs were identified by comparison of mass spectra and arithmetic retention indices with commercial databases (NIST 2005; Wiley 2009; Adams 2005) as well as with commercial pure standards (Fluka, Sigma) dissolved in methanol to achieve realistic concentrations. Liquid standards stepwise dissolved in methanol were also used to calibrate the GC-MS system. The present study was focused on the five predominantly emitted MTs that were α-pinene, sabinene, β-pinene, myrcene and limonene. The emission rates were calculated by multiplying the chamber net BVOC concentration (i.e. chamber BVOC concentration with plant minus BVOC concentration of empty chamber) with the chamber flow rate divided by the enclosed leaf area, which in most cases was equal the chamber area (see above). Empty chamber was either measured before or after CO<sub>2</sub>-ramping. The emission rates per leaf dry weight were calculated using the LMA of the measured leaf.

#### 2.4 Data treatments and statistical tests

During CO<sub>2</sub>-ramping stomatal conductance and transpiration frequently increased at low CO<sub>2</sub> and diminished at high CO<sub>2</sub>. As a result, leaf temperature slightly changed during CO<sub>2</sub>-ramping, sometimes by much as 1°C owing to changes in evaporative cooling of the leaf by transpiration. To avoid that potential CO<sub>2</sub> effects on emissions were biased by leaf temperature changes, we normalized the emissions rate to the same standard temperature of 30 °C (hereafter referred to as the emission factor (EF)) using the temperature algorithm for light dependent isoprenoid emission (Guenther et al., 1993) with coefficients adjusted for QI emissions according to Staudt and Bertin (1998). ~~The 30°C-normalized emission rate measured on a leaf at the beginning of CO<sub>2</sub>-ramping (EF<sub>30</sub>) is defined as:  $EF_{30} = \frac{E_{CO_2} \cdot V_{CO_2}}{V_{leaf} \cdot V_{400}} \cdot \left( \frac{30 - T_{leaf}}{10} \right)^{-2}$  (Staudt and Bertin, 1998).~~

215 To examine the relative changes of BVOC emissions and photosynthetic variables in response to CO<sub>2</sub>-ramping, the data of each CO<sub>2</sub>-response curve were normalized in two ways: i) By dividing the individual values of a measurement series by the mean of the series ( $\frac{X_{V_{CO_2}}}{X_{mean} \cdot V_{mean}^{-1}}$ ); ii) by dividing the individual values of a measurement series by the initial value, i.e. the measurement made at 400 ppm [CO<sub>2</sub>] ( $\frac{X_{V_{CO_2}}}{X_{400} \cdot V_{400}^{-1}}$ ). The first normalization is relatively insensitive to outliers and served to assess and illustrate the overall CO<sub>2</sub> responsiveness of a measured variable. However, it is less suitable to differentiate the responsiveness to low CO<sub>2</sub> from that to high CO<sub>2</sub>, because the response of one will affect the relative response of the other (see also Fig. S1 in Supplement 2 for illustration of the potential biases generated by these data normalizations). Hence, the second normalization was specifically used to analyze separately the responsiveness to low CO<sub>2</sub> (2 measurements at [CO<sub>2</sub>] < 400 ppm) and to high CO<sub>2</sub> (4 measurements at [CO<sub>2</sub>] > 400 ppm) of a measured variable. To describe the global responsiveness of a variable to low and high CO<sub>2</sub>, we used the mean values of the individual 400-CO<sub>2</sub>-normalized data for the measurements in a response curve below 400 ppm CO<sub>2</sub> ( $\mu V_{<400} V_{400}^{-1} = \frac{1}{n} \sum_{i=1}^n V_{CO_2} V_{400}^{-1}, n=2$ ) and above 400 ppm CO<sub>2</sub> ( $\mu V_{>400} V_{400}^{-1} =$

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$\frac{1}{n} \sum_{i=1}^n V_{CO_2} V_{400}^{-1}$ ,  $n=4$ ), respectively. The relative change in Fv/Fm and dark respiration (R) was expressed as the difference between the value before and after CO<sub>2</sub>-ramping divided by its initial value (e.g.  $\delta Fv/Fm = (Fv/Fm_{ini} - Fv/Fm_{end}) / Fv/Fm_{ini}$ ).

The individual 400-CO<sub>2</sub>-normalized emission rates ( $E_{CO_2} E_{400}^{-1}$ ) were fitted to the algorithm described in Wilkinson et al. (2009) that is used in the MEGAN model (Guenther et al., 2012) to account for the CO<sub>2</sub> response of isoprene emissions:

$$C_{Ci} = E_{max} - \frac{E_{max} C_i^h}{C_i^h + C_i^h} \quad (1)$$

where  $C_{Ci}$  is the CO<sub>2</sub> activity factor,  $C_i$  the leaf internal CO<sub>2</sub> concentration, and  $E_{max}$ ,  $C^*$  and  $h$  are empirical coefficients. The algorithm simulates an inverse sigmoidal relationship between emissions and  $C_i$ , where  $C_{Ci}$  scales the emission rate at standard [CO<sub>2</sub>] (400 ppm) to the progressive inhibitory effects of increasing  $C_i$ .

The influence of growth conditions were assessed using Analysis of Variance on each data set of the two assay temperatures after having tested for normality (Shapiro-Wilk test) and equal variance (Levene test). If tests failed, the non-parametric Kruskal-Wallis test was applied. Post-hoc Tukey HSD and Dunn's tests were used for pairwise comparison. The influence of assay temperature on pooled data of growth conditions was examined using Student or Mann-Whitney rank sum test. Paired Student tests or Wilcoxon signed-rank tests were used to compare data of two assay [CO<sub>2</sub>] (400 vs 800 ppm) measured during CO<sub>2</sub>-ramping on a same leaf. The differences between groups of measured variables were considered to be significant at the level  $\alpha = 0.05$ . Pearson correlation analyses were performed in order to test the degree of linear covariation relationships among variables. Consistency of correlations (linearity, outliers) was visually checked by scatter plots. In general, the data distributions seen on scatter plots provided no clear evidence for non-linear relationships with little exception: the relation between  $G_{400}$  and  $A_{400}$  was slightly curved at highest values ( $A_{400}$  changed somewhat less with increasing  $G_{400}$ ). All statistical analyses were done with addinssoft (2021) XLSTAT statistical and data analysis solution except the non-linear regression analyses (curve fitting), which was done-carried out with the SigmaStat 2.0 (Jandel Scientific Software).

### 3 Results

#### 3.1 BVOC emission pattern and chemotypes

Foliar VOC emission of all oak saplings were mainly composed of five MTs  $\alpha$ -pinene, sabinene,  $\beta$ -pinene, myrcene and limonene accounting to  $95 \pm 5$  %. The remainder was composed of  $\alpha$ -tujene, camphene, 1,8-cineol and  $\beta$ -ocimene. Individual trees released the five major MTs in different proportions, roughly according two type of emission profiles: About two thirds of the trees (17 of 26) produced  $\alpha$ -,  $\beta$ -pinene and sabinene in high proportions (60-90 %), while one third (9 of 26) high proportions of limonene and myrcene (60-90 %). All replicate measurements made on a same or different leaf of individual trees showed that the relative proportions of these 5 VOCs were not different between leaves and not influenced by assay temperature or [CO<sub>2</sub>], and hence tree specific (chemotype) (Fig. S2 in Supplement 1). Apart from the emission composition,



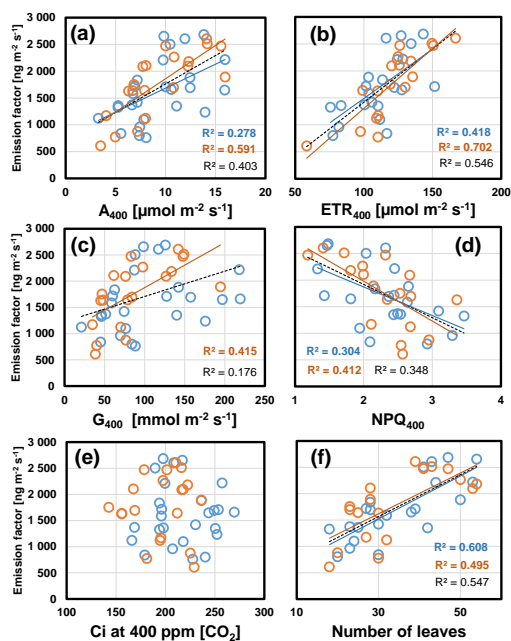
there was no apparent difference between the two chemotypes in any of the measured variables including the total VOC emission rate and responses to CO<sub>2</sub> (Figs. S3 and S4 in Supplement 1). These observations allowed us to restrict our data analyses on the sum of the major compounds.

### 260 3.2 Intraspecific variability of the ~~foliar~~ emission factor

The mean emission rates across all growth treatments of the sum of the 5 major emitted MTs measured at the beginning of the CO<sub>2</sub>-ramping were 1491 ± 537 and 2456 ± 865 ng m<sup>-2</sup> s<sup>-1</sup> (11.0 ± 4.0 and 18.1 ± 6.4 nmol m<sup>-2</sup> s<sup>-1</sup>, 32.6 ± 11.4 and 55.8 ± 21.4 μg g<sup>-1</sup> h<sup>-1</sup>) for 30 °C and 35 °C assay temperature respectively. The deduced temperature-normalized EF ~~ranged by a factor 4~~ varied more than fourfold between 610 and 2686 ng m<sup>-2</sup> s<sup>-1</sup> (4.5-19.7 nmol m<sup>-2</sup> s<sup>-1</sup>, 13.1-60.8 μg g<sup>-1</sup> h<sup>-1</sup>) and averaged ~~on~~ 1694 ± 589 ng m<sup>-2</sup> s<sup>-1</sup> (37.6 ± 13.6 μg g<sup>-1</sup> h<sup>-1</sup>). There was no significant difference between the mean EF deduced from the 30°C and that from the 35 °C measurements (1626 ± 575 vs 1781 ± 605 ng m<sup>-2</sup> s<sup>-1</sup> (P=0.38, t-test on merged data of the growth treatments/regimes). Regarding the effect of growth conditions, there was no significant difference between the four populations in terms of EF or any other variable except plant leaf mass (i.e. the number of leaves per plant; ANOVA, P=0.030), with the lowest mass observed in plants grown under the 400/25 regime (Table S2 in Supplement 1). Pooling the 30°-data of the two growth temperature regimes suggest that growth under elevated CO<sub>2</sub> increased the emission factor. However, this effect was only significant for EF per leaf area (t-test, P=0.012; Table S2) and not for EF per leaf dry weight (t-test, P=0.087), as leaves grown under elevated CO<sub>2</sub> had increased LMA values (t-test, P=0.022). In addition, growth under double CO<sub>2</sub> promoted the leaf mass of the saplings (t-test, P=0.008). Pooling the data of the two growth temperature regimes revealed that growth under elevated CO<sub>2</sub> significantly increased seedling growth (t test, P=0.008) and LMA (t test, P=0.022), though it had no effect on estimated foliar chlorophyll content (t test, P=0.296). Growth at double CO<sub>2</sub> had also a general positive effect on the mean EF per leaf surface determined at 30° assay temperature (t test, P=0.012; Table S2). This difference was however not significant for EF per leaf dry mass (t test, P=0.087). The EFs deduced from the 35°C measurements were not different between the two growth [CO<sub>2</sub>]. However, the initial C loss at 35 °C (E<sub>400</sub>/A<sub>400</sub>) and the initial respiration rate (R<sub>ini</sub>) were respectively lower and higher in leaves grown under elevated CO<sub>2</sub> than under normal CO<sub>2</sub> (t tests, P=0.029 and 0.015). In contrast, Pooling the data of the two growth CO<sub>2</sub> regimes did not show/reveal any significant effect of growth temperature on leaf growth, leaf structure (LMA, [Chloro]), EFs or any photosynthetic variables measured at the beginning of a series at either assay temperature.

Overall, the foliar EF results showed a large variability of EF varied largely within all and across growth regimes. Pearson C correlation analyses revealed revealed that the leaf-to-leaf variation in EF measured at 30 °C assay temperature correlated most strongly with the plant's leaf growth rate (i.e. number of leaves per plant; Fig. 1 (f)). In addition, EF scaled positively with the actual CO<sub>2</sub>-assimilation (A<sub>400</sub>) and electron transport rate (ETR<sub>400</sub>), and negatively with the non-photochemical quenching NPQ<sub>400</sub> (Figs. 1(a), (b), (d)). Leaf growth, A<sub>400</sub>, ETR<sub>400</sub> and stomatal conductance (G<sub>400</sub>) were all positively correlated with each other (see Table S3a in Supplement 2). The same relationships held for the data gained at an assay temperature of 35 °C. However, the correlation of the deduced EFs with leaf growth rate was less strong than at 30 °C, while

290 those with ongoing photosynthetic processes  $A_{400}$ ,  $ETR_{400}$  and  $NPQ_{400}$  were strengthened including a positive correlation between EF and  $G_{400}$  (Fig. 1(c)). By contrast, the variability of EF was not correlated with  $C_i$  at either experimental temperature, which ranged from 150 to 260 ppm (Fig. 1(e)) at neither assay temperature, EF variability was correlated to that of the initial  $C_i$ , which ranged between 150 and 260 ppm (Fig. 1(e)).

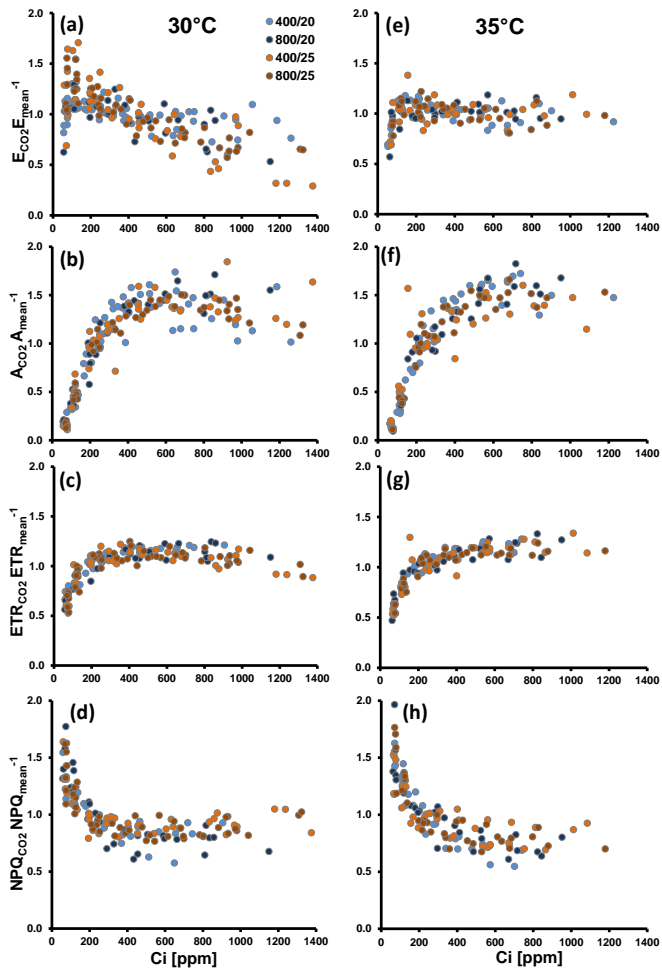


295 **Figure 1.** Scatter plots of the foliar emission factors measured at the beginning of  $\text{CO}_2$ -ramping (at 400 ppm [ $\text{CO}_2$ ]) and assay temperatures of 30 °C (blue) and 35 °C (red) against simultaneously measured photosynthetic variables ((a)-(e)) and the number of leaves per plant (f). Lines with determination coefficients  $R^2$  show best-fit results from Pearson correlation analyses with  $P < 0.05$ . Broken lines and  $R^2$  in black are from pooled data. See Table S3a in Supplement 2 for more information.

### 3.3 Response pattern of VOC emissions and photosynthetic variables during $\text{CO}_2$ -ramping

300 Control runs, in which repeated measurements were taken during the course of the day while temperature, PPFD, and [ $\text{CO}_2$ ] were held at standard conditions showed that leaf emission rates changed very little during the day (Fig. S5 in Supplement 1), ruling out a possible major bias in detecting  $\text{CO}_2$  effects due to endogenous diel variation in leaf emissions.

The relative changes of MT emissions and photosynthetic variables during CO<sub>2</sub>-ramping exhibited different pattern according to the assay temperature, regardless of whether the emission data were normalized to the means per series (Fig. 2) or to the initial measurements at 400 ppm CO<sub>2</sub> (Fig. S6 in Supplement 1).



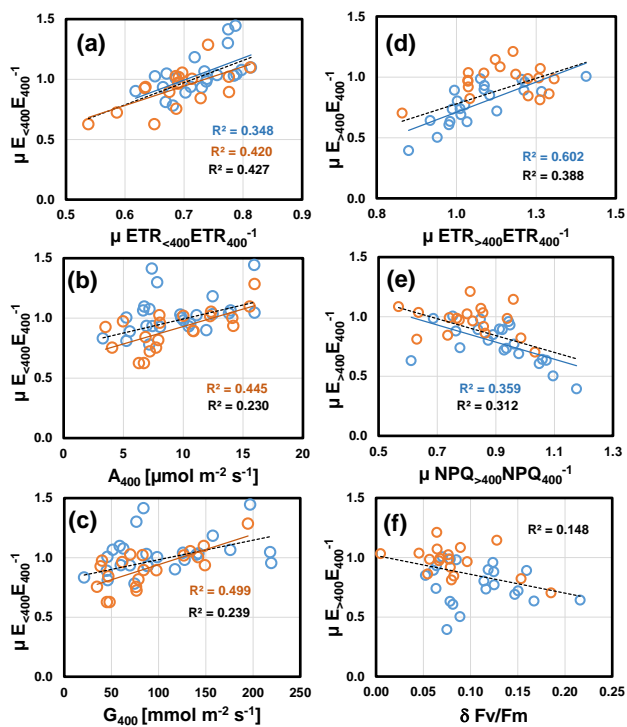
310 **Figure 2. Relative-Normalized** MT emission ((a), (e)), CO<sub>2</sub>-assimilation ((b), (f)), electron transport rate (ETR; (c), (g)) and non-  
photochemical quenching (NPQ; (d), (g)) against ~~calculated~~ leaf internal CO<sub>2</sub> concentration (Ci) measured during CO<sub>2</sub> ramping at assay  
temperatures of 30 °C (left panels) and 35 °C (right panels). Altogether, 26 and 20 CO<sub>2</sub>-response curves were run at 30 and 35 °C,  
respectively. Colors of the dots denote the temperature and CO<sub>2</sub> regimes, in which plants have been grown. Data were normalized by devising  
the individual data of a CO<sub>2</sub>-ramping-response curve by its mean. To compare the overall amplitude of responses, all y-axis were set to the  
same scale.

315 Indeed, comparing the averages of E<sub>400</sub> normalized emissions across all populations, relative emission changes at both low or  
high CO<sub>2</sub> were significantly different between the two assay temperatures (t-tests;  $\mu_{E_{400}E_{400}^{-1}}$ : 1.03 ± 0.16 at 30 °C vs 0.91  
± 0.16 at 35 °C, P=0.021 (t-test);  $\mu_{E_{400}E_{400}^{-1}}$ : 0.78 ± 0.16 at 30 °C vs 0.97 ± 0.12 at 35 °C, P<0.001 (t-test); Table S2 in  
Supplement 1 P<0.001). At 30 °C (Fig. 2(a)-(d), Fig. S6(a)-(d)), emissions frequently decreased under high CO<sub>2</sub> ([CO<sub>2</sub>] > 400)  
320 and showed variable responses to low CO<sub>2</sub> ([CO<sub>2</sub>] < 400) (Fig. 2(a)-(d), Fig. S6(a)-(d)). The variable response of emissions to  
low CO<sub>2</sub> included both emission increases, which occurred more frequently at 200 ppm CO<sub>2</sub>, and emission decreases, which  
occurred more frequently at the subsequent exposure to 100 ppm including increases and decreases CO<sub>2</sub>. CO<sub>2</sub>-assimilation (A,  
Fig. 2(b), Fig. S6(b)) Photosynthesis-continuously increased until 400 to 600 ppm Ci, leveled off beyond with occasional  
decreases at highest Ci. The amplitude of change in ETR (Fig. 2(c)-(d), Fig. S6(c)-(d)) was smaller than in photosynthesis.  
325 Nevertheless, it generally dropped at Ci lower than 200 ppm and tended to decrease at highest Ci. The pattern of NPQ changes  
somewhat mirrored that of ETR. NPQ increased at Ci below 400 ppm and mostly remained unchanged or slightly decreased  
at higher Ci (Fig. 2(d), Fig. S6(d)). Fv/Fm values were significantly lowered after CO<sub>2</sub>-response curves (from 0.79 ± 0.02 to  
0.71 ± 0.04 (δFv/Fm, t-test: p<0.001; Table S2 in Supplement 1) indicating that leaves did not fully recover from  
photoinhibition and were subject to oxidative stress that occurred during CO<sub>2</sub>-ramping (δFv/Fm, Table S1 in Supplement 1).  
330 At 35 °C assay temperature (Fig. 2(e)-(h), Fig. S6(e)-(h) in Supplement 1), the relative emission rates expressed a less variable  
responsiveness to CO<sub>2</sub> than at 30 °C. It more frequently decreased at low Ci than at 30 °C but remained largely insensitive to  
high Ci. Photosynthesis leveled off later at higher Ci than at 30 °C with no or less inhibition at highest Ci. Similarly, ETR never  
decreased during high CO<sub>2</sub> exposure compared to 30 °C but rather slightly increased with increasing Ci. Analogously, during  
the ramping to high CO<sub>2</sub>, relative NPQ decreased more and leveled off later at 35 °C than at 30 °C. The decrease-loss in Fv/Fm  
335 was also significant lower at 35 °C (0.79 ± 0.03 vs 0.73 ± 0.05, t-test: p<0.001; Table S2 in Supplement 1), though smaller  
than at 30 °C. However, though the difference between the two assay temperatures was not significant (δFv/Fm: 8 ± 4% vs: 11 ± 4%,  
t-test: P=0.06).

340 Correlation analyses (for overview see Table S3b and Fig. S7 in Supplement 2) revealed that at both assay temperatures, mean  
relative emissions at low CO<sub>2</sub> ( $\mu_{E_{400}E_{400}^{-1}}$ ) scaled positively with those of ETR ( $\mu_{ETR_{400}ETR_{400}^{-1}}$ , P=0.004; Fig. 3(a)) and  
negatively with the leaf's initial C-losses (E<sub>400</sub>/A<sub>400</sub>) measured at the beginning at normal [CO<sub>2</sub>] (30 °C: R=-0.52, P=0.006; 35

345 °C:  $R=-0.51$ ,  $P=0.021$ ; data not shown). ~~The latter correlation should be viewed with caution because  $\mu$   $ETR_{<400}ETR_{400}^{-1}$  and  $E_{400}/A_{400}$  contain  $E_{400}$  as a common variable. As a result, random variations of  $E_{400}$  due to limited precision of BVOC measurements will produce negative correlations.~~ However at 35 °C,  $\mu E_{<400}E_{400}^{-1}$  was also strongly correlated with the leaf's initial photosynthesis  $A_{400}$  ( $P=0.001$ ; Fig. 3(b)) and stomatal conductance rate  $G_{400}$  ( $P<0.001$ ; Fig. 3(c)). These correlations were not significant at 30 °C, mainly because two leaves exhibited increased emissions at reduced  $[CO_2]$  along with a relatively high ETR, while their initial photosynthetic and stomatal conductance rates were rather low. At either assay temperature,  $\mu ETR_{<400}ETR_{400}^{-1}$  was unrelated to  $A_{400}$ ,  $G_{400}$  and  $C-loss_{400}$  (Table S3b in Supplement 2). During subsequent ramping to high  $CO_2$ , the emission reductions observed at 30 °C ( $\mu E_{>400}E_{400}^{-1}$ ) were best explained by concomitant reductions in ETR ( $\mu ETR_{>400}ETR_{400}^{-1}$ ,  $P<0.001$ ; Fig. 3(d)) and, anti-correlated with ETR, by increases in NPQ ( $\mu NPQ_{<400}NPQ_{400}^{-1}$ ,  $P=-0.003$ ; Fig. 3(e)).  $\mu E_{>400}E_{400}^{-1}$  was not related to any other variable except a weak negative correlation with the relative emissions before to low  $CO_2$   $\mu E_{>400}E_{400}^{-1}$  ( $R=-0.44$ ,  $P=0.026$ , data not shown). By contrast at 35 °C, the ~~small-variable-minor~~ emission changes ~~observed~~ under high  $CO_2$  ( $\mu E_{>400}E_{400}^{-1}$ ) ~~were at 35 °C showed a strong~~ positively correlation ~~ed~~ with  $\mu E_{<400}E_{400}^{-1}$  ~~the relative emission rates measured before under low  $CO_2$  ( $R=-0.71$ ,  $P<0.001$ ), which is explained by~~. This correlation should be considered with caution. ~~Due to their mathematical interdependency (common denominator), random variation in the absolute emission rates associated with limited precision in BVOC measurements generate positive correlations without any  $CO_2$  effect~~ (Fig. S1 in supplement 2). Finally, when data of both assay temperatures were pooled,  $\mu E_{>400}E_{400}^{-1}$  was negatively correlated with the leaf loss in  $Fv/Fm$  ( $\delta Fv/Fm$ :  $P=0.014$ ; Fig. 3(f)). Interestingly,  $\delta Fv/Fm$  was also negatively correlated with plant growth at both assay temperatures (30 °C:  $R=-0.51$ ,  $P=0.015$ ; 35 °C:  $R=-0.67$ ,  $P=0.003$ ).

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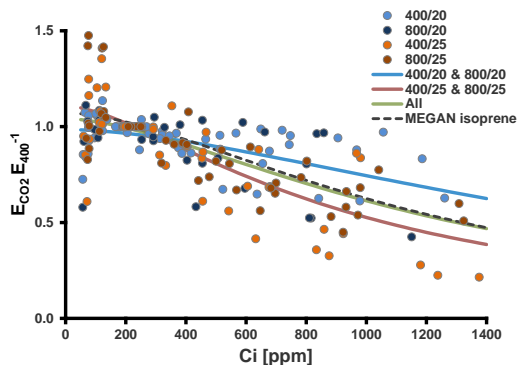
**Figure 3.** Key correlations of relative emissions to low  $\text{CO}_2$  ( $\mu E_{<400} E_{400}^{-1}$  left panels) and high  $\text{CO}_2$  ( $\mu E_{>400} E_{400}^{-1}$ , right panels) measured at assay temperatures of 30 °C (blue) and 35 °C (red). Relative rates of a measured variable are calculated as the mean of  $V_{<400}/V_{400}$  and  $V_{>400}/V_{400}$  where  $V_{400}$  is the value measured at the beginning of  $\text{CO}_2$  ramping at 400 ppm  $\text{CO}_2$ , and  $V_{<400}$  and  $V_{>400}$  are the values subsequently measured at lower and higher  $[\text{CO}_2]$ . Lines with determination coefficients  $R^2$  show best-fit results from Pearson correlation analyses with  $P < 0.05$ . Lines with  $R^2$  given in black are from pooled data. For a complete overview, see Table S3b and Fig. S7 in Supplement 2.

$\text{CO}_2$ -responses of emissions and photosynthetic variables did not differ significantly among the four growth populations with one exception (Table S2 in Supplement 1): At 30 °C, there was a significant difference in relative NPQ at high  $\text{CO}_2$  ( $\mu NPQ_{>400} NPQ_{400}^{-1}$ ) with 800/25 and 400/25 grown plants showing less reduction in NPQ than 800/20 grown plants (ANOVA,  $P = 0.006$ ). When the data of the two  $\text{CO}_2$  growth regimes were pooled, growth temperature significantly affected the high- $\text{CO}_2$ -responses of emissions ( $\mu E_{>400} E_{400}^{-1}$ : t-test,  $P = 0.008$ ), ETR ( $\mu ETR_{>400} ETR_{400}^{-1}$ : t-test,  $P = 0.005$ ) and NPQ (at 30 °C ( $E_{>400}$

375  $E_{400}^{-1}$ : t-test,  $P=0.008$ ;  $ETR_{>400}-ETR_{400}^{-1}$ : t-test,  $P=0.005$ ;  $\mu_{NPQ_{>400}-NPQ_{400}^{-1}}$ : t-test,  $P=0.001$ ). In fact, at 30 °C assay  
temperature, warm grown plants mostly often continued to non-photochemically dissipate light energy to the expense of ETR  
and MT emissions, whereas NPQ of cool grown plants frequently relaxed during high-CO<sub>2</sub>-ramping along with keeping higher  
ETR and emission rates. Accordingly, Fv/Fm was significantly more reduced in warm grown plants compared to cool grown  
plants ( $\delta Fv/Fm$ : t-test,  $P=0.016$ ). Growth temperature also affected the CO<sub>2</sub>-responsiveness of some photosynthetic variables  
at 35 °C assay temperature (table-Table S2 Supplement 1): Leaves grown at elevated temperature opened stomata less at low  
380 [CO<sub>2</sub>] and closed them more at high [CO<sub>2</sub>] than leaves grown at low temperature~~Leaves grown under elevated temperature~~  
~~opened less at low CO<sub>2</sub> and closed more stomata at high CO<sub>2</sub> than in leaves grown under low temperature~~ (t-tests:  $\mu_{G_{<400}-G_{400}^{-1}}$   
 $P=0.009$ ,  $\mu_{G_{>400}-G_{400}^{-1}}$ :  $P=0.003$ ). Furthermore at high CO<sub>2</sub>, warm grown leaves had lower CO<sub>2</sub>-assimilation rates ( $\mu_{A_{>400}}$   
 $A_{400}^{-1}$ :  $P=0.017$ ) and higher NPQ (~~t-test:  $\mu_{NPQ_{>400}-NPQ_{400}^{-1}}$~~  t-test,  $P=0.001$ ) and ETR/A ratios (~~t-test:  $\mu_{ETR/A_{>400}-ETR/A_{400}^{-1}}$~~   
385  $t$ -test,  $P=0.014$ ) compared to leaves grown under low temperature. However, growth temperature had no significant effect  
on emission responses to low and high CO<sub>2</sub> at 35 °C. Pooling the data of the two growth temperature regimes did not reveal  
any effect of growth CO<sub>2</sub> on CO<sub>2</sub>-responsiveness of emissions or photosynthetic variables

### 3.4 Implications for predicting future MT emissions from Holm oak

We tested whether the MEGAN algorithm (eq. (1)) can be used to simulate the CO<sub>2</sub>-response of MT emissions at 30 °C (Fig.  
390 4). Using the whole data set for the fit resulted in a response curve with coefficients close to that currently applied to predict  
the CO<sub>2</sub>-response of isoprene emissions under current CO<sub>2</sub>-level (Table S4 in Supplement 3). However, as indicated by the  
statistics (see above) and can be seen from Fig. 4, relative emissions considerably varied with data from the low and high  
temperature grown plants predominately scattering above and below the total fit, respectively. Consequently, separate fits  
resulted in two distinct curves, which differed mainly in the coefficient C\* determining the amplitude of the emission reduction  
395 and less in the coefficient h, which sets the C<sub>i</sub> level, at which emissions decline.



**Figure 4.** Relative MT emission rates (normalized to the initial measurement at 400 ppm CO<sub>2</sub>) measured during CO<sub>2</sub>-ramping at the assay temperatures of 30 °C (26 response curves). Colours of the dots denote the CO<sub>2</sub> and temperature regimes, in which plants have been grown.

Solid lines present best fits to the algorithm of the MEGAN modelling framework accounting for the short-term effect of CO<sub>2</sub> on isoprene emissions (equation (1)):
$$C_{Ci} = E_{max} - ((E_{max} C_i^h) / (C_i^{*h} + C_i^h))$$
where  $C_{Ci}$  is the CO<sub>2</sub>-scaling factor,  $C_i$  the leaf internal CO<sub>2</sub> concentration, and  $E_{max}$ ,  $C_i^*$  and  $h$  are empirical coefficients. The algorithm simulates an inverse sigmoidal relationship between emissions and  $C_i$ , where  $C_{Ci}$  scales the emission rate at standard [CO<sub>2</sub>] (400 ppm) to the progressive inhibitory effects of increasing  $C_i$ . The green line shows the fit from all data and the red and blue lines from the warm and cool grown plants, respectively. Black broken line depicts the CO<sub>2</sub>-scaling currently used in MEGAN (Guenther et al, 2012). All coefficients values and additional information are given in Table S4 of supplement [1].

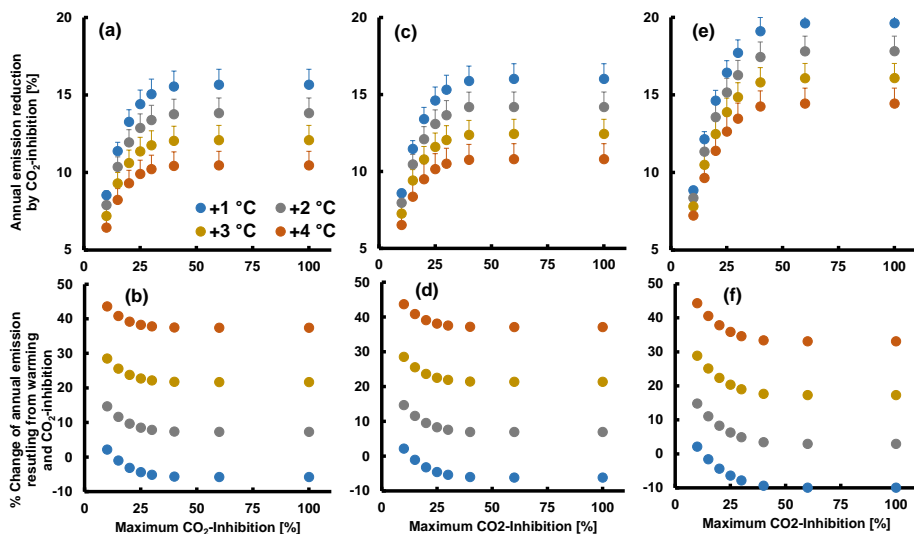
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We further analyzed to what extent the observed emission inhibition by double CO<sub>2</sub> could compensate the increase in emissions due to global warming. Doubling assay [CO<sub>2</sub>] from 400 to 800 ppm reduced temperature normalized emission rates on average by 8 % at 30 °C (1626 ± 575 vs 1499 ± 538 ng m<sup>-2</sup> s<sup>-1</sup>; P=0.001; paired t-test; n=26) and had no effect at 35 °C (1781 ± 605 vs 1800 ± 719; P=0.755; paired t-test; n=20). However, the highest and significant decrease in 30°-emissions by double CO<sub>2</sub> was observed for plants grown at elevated CO<sub>2</sub> and temperature (800/25) amounting to ca. 10 % (1808 ± 661 vs 1625 ± 591; P=0.033; paired t-test; n=7). Compared to the short-term effect of temperature on emissions, this decrease might compensate the emission increase by one degree warming, which is about 13 % in the range between 10 and 30 °C and then gradually decreases until the temperature optimum around 41 °C (Staudt and Bertin, 1998). Yet, it is very likely that the high-CO<sub>2</sub> inhibition we observed becomes stronger at temperatures lower than 30 °C. Several studies on isoprene emissions suggest that the high-CO<sub>2</sub> inhibition on emissions increases with temperature linearly and can suppress emissions by 50 % or more at lowest temperatures (Rasulov et al., 2010; Potosnak et al., 2014; Sharkey and Monson, 2014; and references therein). Accordingly, the high-CO<sub>2</sub> inhibition of MT emissions would be most relevant during the cooler daytime hours, and, because QI is an evergreen species, all along the cooler seasons, while having less effect during the warm season and hours. There are additional factors that shape the seasonal course of emissions and hence will change the impact of CO<sub>2</sub>-inhibition on annual emissions. MT emissions from QI are strictly light dependent (Staudt and Bertin, 1998). Low temperatures are commonly



425 associated with lower light levels and shorter day lengths, resulting in lower emissions during these periods. Moreover, the EF  
of QI leaves usually diminishes towards the winter season (Peñuelas and Llusà, 1999; Staudt et al., 2002; Ciccioli et al., 2003)  
along with the down-regulation of the activity of MT synthases (Fischbach et al., 2002; Grote et al., 2006), further reducing  
its contribution to the annual VOC budget. However, in summer the physiological activity of QI leaves is frequently restrained  
430 by drought, potentially reducing their emissions (Staudt et al., 2002; Lavoit et al., 2009). To summarize, differences in the  
high-CO<sub>2</sub> responsiveness of emissions were modulated by both actual leaf and the plant's growth temperature, albeit in  
opposite manner. The observed correlations suggest that the emission response to low CO<sub>2</sub> depended on the instantaneous  
energy status and the leaf's initial carbon balance, whereas the response to high CO<sub>2</sub> depended mostly on the instantaneous  
435 energy status, which was affected by photooxidative stress. Figure 5 provides a schematic overview on the key correlations  
regarding the initial emission factor and the relative emissions under low and high [CO<sub>2</sub>].

To better understand the relative importance of CO<sub>2</sub> inhibition in interaction with other determinants of leaf MT-emissions  
from QI, we computed annual emissions by combining eight high-CO<sub>2</sub>-inhibition scenarios differing in their maximum high-  
CO<sub>2</sub> inhibition with four warming scenarios (1-4 °C warming) and three scenarios of EF seasonality (seasonality without and  
440 with summer drought, no seasonality). The details of the procedure are described in Supplement 3. Briefly, simulations were  
based on climate data recorded over 2019-2021 by a flux tower in a nearby QI forest station. The temperature data were  
stepwise increased by 1 to 4°C encompassing the range predicted to occur with doubling [CO<sub>2</sub>] by the end of the century (2.8-  
4.6 °C relative to the 1850-1900 period, scenario SSP3-7.0; IPCC 2021). The seasonal courses of EF were deduced from the  
results of a former rain exclusion experiment (Staudt et al., 2002; Fig. S9 in Supplement 3). The CO<sub>2</sub> inhibition scenarios  
445 assume that emission inhibition is zero at an air temperature of 35 °C or higher and increases by 2 % per 1 °C-decrease to  
reach eight different maximum inhibitions varying between 10 % and 100 %. Hence, in all scenarios the inhibition was the  
same between 30 °C (10 % as observed on elevated CO<sub>2</sub> and warm-grown plants) and higher temperatures. Overall, 96  
simulations were run, whose outputs are summarized in Fig. 5 and Table S5 in Supplement 3. Fig. S10 in Supplement 3 shows  
an example of the diurnal and annual emission courses resulting from the simulations.



**Figure 5.** Assessment of the potential effect of high- $\text{CO}_2$ -inhibition on total annual MT emission from QI leaves under a future warmer climate with a doubled atmospheric  $[\text{CO}_2]$  combining different scenarios of maximum  $\text{CO}_2$ -inhibition (x-axes), warming (colour of dots) and seasonality of the emission factor (left panels: seasonality without summer drought, middle panels: seasonality with summer drought, right panels: no seasonality (constant EF)). The lower graphs **To account for the high- $\text{CO}_2$ -inhibition of emissions at double  $\text{CO}_2$ , we assumed that inhibition is 0% at  $\geq 35^\circ\text{C}$  and progressively increases at lower temperatures by 2% per  $1^\circ\text{C}$  decrease to reach maximum inhibitions of either 10, 15, 20, 25, 30, 40 or 100%. For example for maximum inhibitions of 25% and 40%, emissions become reduced by 25% at  $\leq 22.5^\circ\text{C}$ , and by 40% at  $\leq 15^\circ\text{C}$ , respectively.** (b), (d), (e) show the percentage change of annual emission with respect to today values  $((\text{future annual } E_{+T+\text{CO}_2} - \text{today annual } E) * \text{today annual } E^{-1} * 100)$ , and the upper graphs ((a), (c), (d)) the percentage reduction of the future annual emissions by  $\text{CO}_2$ -inhibition  $((\text{annual } E_{+T} - \text{annual } E_{+T+\text{CO}_2}) * \text{annual } E_{+T}^{-1} * 100)$ . The simulations were run with temperature and PPFD data of the years 2019, 2020 and 2021. Data points and error bars (not visible in (b), (d), (f)) are means and standard errors of  $n = 3$  years. The seasonal variations of the emission factor were deduced from Staudt et al. (2002). A detailed explanation of the simulations and additional illustrations of the results are given in Supplement 3.

As expected, the relative importance of  $\text{CO}_2$  inhibition decreases with the level of climate warming. In absence of summer drought (Fig. 5(a), (b)), warming from  $1^\circ$  to  $4^\circ$  decreases the annual emission reduction by  $\text{CO}_2$ -inhibition from  $8.5 \pm 0.3$  to  $6.5 \pm 0.5$  % at 10 % maximal  $\text{CO}_2$ -inhibition and from  $15.7 \pm 1.0$  to  $10.5 \pm 0.9$  at 100 % maximal  $\text{CO}_2$ -inhibition. Thus, even when  $\text{CO}_2$ -inhibition of emissions would continue to increase with decreasing temperature, its impact on the annual emission budget is moderate due to the non-linear effects of temperature and light on emissions, and due to the strong seasonality of EF. In fact, the calculated amounts of MT emitted from a leaf per day are about two orders of magnitude lower during the winter

470 season than during the hot summer season (see examples in Fig. S10(e) and (g) in Supplement 3). As a result, assuming higher maximum inhibitions than 40 % at temperatures below 15 °C has no additional effect on the annual VOC budget (Fig. 5 (a), (c) and (e)). Drought or any event that significantly curtails summer emissions is expected to enhance the annual impact of CO<sub>2</sub>-inhibition. Indeed, running simulations with EF seasonality from drought-exposed QI trees instead of irrigated trees increased the annual CO<sub>2</sub>-inhibition by 0.1-0.5 % (Fig. 5 (c), (d)), which is attributable to the lower annual EF maximum and its delayed occurrence in the summer season when decreasing day length, light intensity, and temperature already constrain the daily VOC release. Accordingly, the complete suppression of EF seasonality (constant mean EF throughout the year) increased the annual CO<sub>2</sub>-effect by 0.5-4 % (Figs. 5 (e), (f)). However, even in this very unlikely seasonality scenario, CO<sub>2</sub>-inhibition did not reduce annual emissions by more than 20 %. A decrease in future annual VOC emissions relative to current levels was observed only in the simulations with 1°C warming (negative values in Figs. 5 (b), (d), (f) and Table S5 in Supplement 3), while warming of 2 °C or more always resulted in a net increase in annual emissions. Additional simulations with intermediate temperatures suggest that the emission inhibition by double CO<sub>2</sub> could offset the emissions increase by 1.5 °C warming. This is at the low end of the 2.8-4.6 °C warming range predicted by the IPCC SSP3-7.0 scenario, considering 480 that the temperatures recorded in 2019-2021 were already about 1 °C higher than in the pre-industrial period.

## 4 Discussion

### 4.1 Variability in of the foliar emission capacity factor – what makes the difference?

485 Growth at double CO<sub>2</sub> enhanced the foliar emission factor while a 5-degree difference in growth temperature had no significant effect on the EF. The enhancement of EF by elevated growth CO<sub>2</sub> was little affected by growth conditions, apart from that was partly due to its positive effect on elevated growth CO<sub>2</sub> on plant growth and LMA, as confirming observed in previous CO<sub>2</sub> greenhouse study on QI (Staudt et al., 2001). On the other hand, the non-effect of growth temperature on EF the lack of a significant effect of growth temperature on EF observed in the present study contrasts to diverges from the results of several field studies (Peñuelas and Llusà, 1999; Staudt et al., 2002; Ciccioli et al., 2003; Lavoit et al., 2009) as well as a growth chamber study (Staudt et al., 2003) and several field studies (Peñuelas and Llusà, 1999; Staudt et al., 2002; Ciccioli et al., 2003; Lavoit et al., 2009), which showed suggesting that the EF of QI leaves undergoes is subject to strong seasonal cycles that are likely related to prevailing meteorological conditions related to prevailing meteorological conditions. We explain this apparent contradiction as indicating that the long-term regulation of the leaf's emission capacity by temperature is non-linear and that persistent lower temperatures than that applied here in the present study are necessary for its down regulation, presumably associated with the expression and turnover of MT synthases (Fischbach et al., 2002; Lavoit et al., 2009). Furthermore, it is conceivable that the exposure to temperature changes in the temperature regimes rather than constant lifelong temperature differences trigger induce acclimation processes in EF (Staudt et al., 2003; Wiberley et al., 2008). Finally, moderate effects of growth temperature on EF may have been overlooked in our study due to its large variability of the EF within the four populations.

The EF variability scaled positively with the leaf's actual photosynthetic activity and the individual plant's leaf growth-rate, which was negatively related to the subsequent loss in Fv/Fm that occurred during subsequent CO<sub>2</sub>-ramping. This suggests that the plant's overall capacity to use photosynthates for growth overall (sink capacity) of individual plants was associated with the capacity of its individual leaves to fix carbon, and to produce MTs and to avoid persistent photooxidative inhibition stress. Positive relations between isoprenoid emissions, photosynthesis, growth performance and resistance to harsh environmental conditions within and across natural or genetically manipulated populations have already reported (e.g. Monson and Fall, 1989; Staudt et al., 2001; Possell et al., 2004; Eller et al., 2012; Lantz et al., 2019a; Zuo et al. 2019; Niinemets et al., 2021; Dani et al. 2022), though also opposite-negative or no relations have been observed (e.g., Guidolotti et al., 2011; Behnke et al., 2012; Zuo et al., 2019; Monson et al., 2020; Sun et al., 2020). The positive association we observed on QI saplings may be due to the beneficial effects of chloroplastic volatile isoprenoid production on plant growth and stress resistance their photosynthetic and growth performances by modulating cellular signaling networks optimizing plant growth and stress resistance (Frank et al., 2021; Monson et al., 2021; Dani et al., 2022). However, because photosynthetic processes supply directly chloroplastic isoprenoid biosynthesis with carbon substrates and energetic cofactors, it is also conceivable that oppositely higher foliar emission capacity of QI saplings resulted from their higher foliar photosynthetic activity constrained by the plant's sink capacity to use photosynthates (Ainsworth and Bush, 2011). Covariation of leaf emission and photosynthesis might also reflect variation in leaf anatomy as for example the per leaf surface quantity of photosynthetically active tissues and density of chloroplasts ((Sun et al., 2012; Rasulov et al., 2015). We found no consistent correlation between estimated chlorophyll content or LMA and initial photosynthesis or emission rates not supporting leaf anatomical differences being the major cause of the observed correlation pattern. Instead, a portion of variability in initial A<sub>400</sub> was simply related to leaf to leaf variability in stomatal conductance G<sub>400</sub>. Especially at 35°C assay temperature, when terpene production was almost double as high as at 30°C, EF scaled positively with G<sub>400</sub> along with A<sub>400</sub> and ETR<sub>400</sub>, while at 30°C it was more related to the plant's growth performance (Fig. 5). Considering that stomatal opening does not directly control MT emissions (Niinemets et al., 2014), this shift in the correlation pattern indicates that at 35 °C the chloroplastic MT production was frequently limited by ongoing CO<sub>2</sub> assimilation via the supply of basic carbon substrates and energetic cofactors. Instead at 30°C, the leaf to leaf variability in the more moderate MT production might have more frequently mirrored the variability in the foliar amount of active MT synthases (see e.g. Loreto et al., 2001; Fischbach et al., 2002) and/or other rate limiting enzymes of the MEP pathway (Lantz et al., 2019a), which seemingly was associated with the plant's growth performance perhaps due to a congruent variability in growth hormones produced within the same metabolic pathway (Dani et al., 2022). By contrast, CO<sub>2</sub>-responsiveness was not the reason for the leaf to leaf variability of EF (for an example see Guidolotti et al., 2011), since it was unrelated to initial Ci at both assay temperatures (Fig. 1(e)). However, it is also known that the sink capacity of a plant can constrain the photosynthetic activity of its leaves (Ainsworth and Bush, 2011). Therefore, QI saplings with lower growth could have lower EFs if their leaf monoterpene biosynthesis was limited by carbon substrates and/or energetic cofactors coming from photosynthesis. At 35°C assay temperature, when terpene production was almost twice that at 30°C, the correlations between the deduced EFs and photosynthetic variables, including stomatal conductance, were stronger than at 30 °C (Fig. 1).

535 Given that stomatal opening has no direct control on the emissions of the MTs considered in the present study (Niinemets et al., 2014), this shift in the correlation pattern with assay temperature indicates that the leaf-to-leaf variability of EF was indeed partly due to limitation by photosynthetic substrates. In addition, variations in leaf anatomy could explain the correlation of leaf EF and photosynthesis, such as the quantity of photosynthetically active tissues per leaf surface and density of chloroplasts (Sun et al., 2012; Rasulov et al., 2015). However, we found no consistent correlation between estimated chlorophyll content or LMA and photosynthesis or emission rates, suggesting that leaf anatomical differences were of minor importance. CO<sub>2</sub>-responsiveness of emissions was also not the main reason for the leaf-to-leaf variability of EF (for an example see Guidolotti et al., 2011), since EF was not related to Ci at either assay temperatures (Fig. 1(e)).

#### 540 4.2 What are the drivers of the CO<sub>2</sub>-responsiveness? of emissions

If leaf-MT biosynthesis in leaves was constrained by its low-G and A-photosynthetic activity already at the beginning of CO<sub>2</sub>-ramping at 400 ppm CO<sub>2</sub>, it is likely that emissions decline under low CO<sub>2</sub>-ramping when photosynthetic processes carbon fixation became rapidly reduced. Indeed at 35°C all leaves that had low initial assimilation and stomatal conductance rates (A<sub>400</sub>, G<sub>400</sub>) showed the most pronounced emission decrease under low CO<sub>2</sub> ( $\mu E_{<400}E_{400}^{-1}$ ) (Fig. 3 (b), (c)), whereas at 30°C only leaves having low assimilation together with high emission rates (hence high C-loss<sub>400</sub>). In addition and unrelated to the leaf's initial photosynthetic and MT production status, relative emission rates under low CO<sub>2</sub> ( $E_{<400}E_{400}^{-1}$ ) scaled positively with relative ETR ( $\mu ETR_{<400}ETR_{400}^{-1}$ ) at both assay temperatures (Fig. 3 (a)). We interpret these observations that MT production during low CO<sub>2</sub>-ramping was curbed by two rather independent constraints, one associated with the availability of basic C3-substrates entering in the MEP pathway and one with the availability of energetic co-factors necessary to reduce them further downstream. The latter predominated initially at moderate low CiO<sub>2</sub> when energetic cofactors were still primarily used in the CBB-cycle for CO<sub>2</sub>-reduction and photorespiration, whereas the former when Ci approached the CO<sub>2</sub> compensation point (i.e., when A = 0) and the more and earlier the leaf-initial emission rates were high and assimilation low. Labelling studies have shown that the fraction of 'older' carbon incorporated in the biosynthesis of isoprene increases during exposure to low [CO<sub>2</sub>] and/or high temperatures (Funk et al., 2004; Trowbridge et al., 2012; de Souza et al., 2018; Guidolotti et al., 2019; Yanez-Serrano et al., 2019) as well as including the leaf-internally re-cycled fixation of respired CO<sub>2</sub> (Garcia et al., 2019).

555 Relative ETR was also by far the best predictor of emission changes to high CO<sub>2</sub> at 30°C (Fig. 3 (d)), suggesting that the same mechanisms contributed to modulate emissions at moderate low and at high CO<sub>2</sub>. Earlier studies on isoprene emissions suggested that this high-CO<sub>2</sub> inhibition results from an activation of the cytosolic PEP-carboxylase under high [CO<sub>2</sub>] (but see Abadie and Tcherkez, 2019) leading to a reduction of PEP available for import into chloroplasts and in turn less pyruvate for isoprenoid biosynthesis (Rosenstiel et al., 2003). This hypothesis however does not explain its temperature dependency (Sun et al., 2013; Monson et al., 2016) and was not confirmed by studies experiments using competitive PEP-carboxylase inhibitors (Rasulov et al., 2018). An alternative hypothesis links the emission reduction at high CO<sub>2</sub> to the occurrence of feed-back inhibition of photosynthesis processes (Sharkey and Monson, 2014; and references therein). A lack of increase or decreases of A and ETR at high CO<sub>2</sub> are typically observed when the production of triose phosphate from CO<sub>2</sub>-fixation in the CBB-

cycle largely exceeds its utilization for starch and sucrose synthesis (Triose-Phosphate-Utilization-limitation; McClain and Sharkey, 2019). As a result, ~~the~~ accumulation of sugar phosphates will lead to causes the depletion of inorganic phosphate (Pi) necessary to sustain ATP synthesis, ~~and~~ This ultimately leads to an inhibiting on of photosynthetic light and dark reactions, and also the availability of pyruvate inside the chloroplasts by compromising the exchange rates ~~to~~ from the cytosol via Pi transporters (Sharkey and Monson, 2014; de Souza et al., 2018). TPU-limitation of photosynthesis occurs less under high temperature, mostly because sucrose synthesis and consumption in sink tissues is are enhanced thereby restoring Pi levels. Beside its direct effect on metabolic rates, high temperature decreases CO<sub>2</sub>-solubility (in pure water approx. -10 % from 30 to 35 °C), which possibly may lowering photosynthetic reduction of CO<sub>2</sub>-fixation (Potosnak et al., 2014) and hence alleviate TPU-limitation coming from both external (ambient air) and internal (respiration) sources (Potosnak et al., 2014). Growth conditions can affect TPU. Plants acclimatized to low temperature or elevated CO<sub>2</sub> tend to have an increased Pi regeneration capacity thus being less vulnerable for TPU limitation (McClain and Sharkey, 2019). Consistent with this, we found little evidence of TPU limitation in our study at 35 °C assay temperature. Moreover, the greatest decrease in emission and photosynthetic processes was observed in warm-grown plants, particularly in the 400/25 regime. These plants had the lowest leaf growth and thus a low capacity to utilize photosynthates, which is expected to favor TPU limitation. Indeed in this study, signs of TPU limitation were virtually absent at 35 °C assay temperature, and at 30 °C, the strongest decline in emission and photosynthetic processes was observed on 400/25 grown plants showing the lowest growth, indicating that a limited capacity to use photosynthates for growth favored TPU limitation of photosynthesis and inhibition of emissions. On the other hand, there was no overall correlation between the plant's growth rates and their emission responsiveness to high CO<sub>2</sub> (Table S3b and Fig. S7 in Supplement 2, Fig. 5). Furthermore, comparing the shape of response curves of individual normalized E and ETR data rather than their means, suggests that their evolutions during high and low CO<sub>2</sub>-ramping were partly disconnected in opposite ways during high and low CO<sub>2</sub>-ramping (Figs. 2 and S6 in Supplement 1): During low CO<sub>2</sub>-ramping, emissions often increased ~~occasionally~~ while ETR always ~~dropped~~ decreased, and during high CO<sub>2</sub>-ramping, emissions frequently decreased earlier and more than ETR. Similarly, Monson et al. (2016) and Lantz et al. (2019b) reported that isoprene emissions decreased in response to high CO<sub>2</sub> before TPU-limitation appeared. In ~~for~~ our study, ETR rarely decreased at a C<sub>i</sub> lower than 800 ppm, which is consistent with the global medianimum C<sub>i</sub> value ~~for~~ TPU-limitations deduced by Kumarathunge et al. (2019). Similarly, Monson et al. (2016) and Lantz et al. (2019b) reported that isoprene emissions decreased in response to high CO<sub>2</sub> before TPU-limitation appeared. Thus, with the exception of a few cases, unless TPU-limitations occurred before being detectable by gas exchange and fluorescence measurements (Sharkey, 2019), MT emission started began to decrease line when ETR was mostly insensitive to increasing CO<sub>2</sub>, i.e. at a stage when CO<sub>2</sub>-fixation by the CBB-cycle is typically limited by the production of energetic cofactors from ETR to regenerate the primary CO<sub>2</sub>/O<sub>2</sub>-acceptor Ribulose-1,5-bisphosphate. At this stage, A is expected to should still slightly increase with increasing C<sub>i</sub>, because photorespiration, the second most important electron sink, is progressively inhibited, as evidenced by a decrease in the ETR/A ratio. As a result, energetic cofactors might be less available for the MEP pathway, notably reduction power, which is consumed more during CO<sub>2</sub>-reduction than during photorespiration with respect to ATP (Niinemets et al., 2021; and references therein). This hypothesis has been used as the

600 basis for photosynthesis-linked modelling of isoprene emissions named ('excess energy' or 'energy status' model<sup>2</sup>; (e.g.,  
Morfopoulos et al., 2014; Grote et al., 2014). The temperature dependence of the high-CO<sub>2</sub>-inhibition is explained by the fact  
that at high temperatures a high ETR is maintained while the fraction of electrons consumed in the CBB-cycle is reduced. In  
the same context, it was suggested that the emissions of isoprene emissions were assumed to are be positively related to NPQ  
as an ~~indicating or an~~ excess of reduction ~~power~~ available for the isoprenoid biosynthesis of isoprenoids (Peñuelas et al.,  
605 2013; Filella et al., 2018). The lack of high-CO<sub>2</sub>-inhibition of emissions at higher temperatures is explained by the fact that a  
higher ETR is usually maintained without being consumed in the CBB-cycle. Our results provide partial support for the excess  
energy hypothesis: The ETR/A ratios were indeed generally higher at 35 °C than at 30 °C due to higher ETR at 35 °C (Table  
S2 in Supplement 1). The difference was only significant for warm grown plants consistent with their stronger emission  
responsiveness to high-CO<sub>2</sub>: (t-test, P=0.025). Furthermore, during initial CO<sub>2</sub>-ramping to low CO<sub>2</sub> the ETR/A ratios strongly  
610 increased (Fig. S78(a) in Supplement 1), which would explain why emissions sometimes-frequently increased under moderate  
low CO<sub>2</sub> at 30 °C before presumably being constrained by the lack of basic C3-substrates (see discussion above). Yet, in  
response to high CO<sub>2</sub>, ETR/A-ratios decreased little, and the variations were less related to the emission changes than ETR  
(Table S3 in supplement 2 and Fig. S78(b), (c) in Supplement 1). Also, our study showed not a positive but a clear-negative  
correlation between emission responses to high CO<sub>2</sub> and NPQ and emission responses to high CO<sub>2</sub> (Fig. 3 (c)) indicating that  
615 the maintenance of NPQ processes during high CO<sub>2</sub> co-constrained ETR, A and MT biosynthesis. Given that Since our plants  
were adapted to greenhouse light conditions, the continuous exposure to a relatively-high PPFD level at various [CO<sub>2</sub>] caused  
some persistent photoinhibition oxidative stress as evidenced by the loss of (Fv/Fm reduction). Especially during exposure to  
lowest [CO<sub>2</sub>] when ETR and A rapidly declined, electrons or excitation energy from excessive light were likely transferred to  
O<sub>2</sub> generating ROS. NPQ and ROS formation efficiently-reduces the availability of reduction power in two ways: First by  
620 reducing its formation during PSII electron transport by diverting the absorption or the absorbed light energy from PS (thus  
lowering ETR), and second, by enhancing its consumption for ROS detoxification and NPQ mechanisms inside chloroplasts,  
notably in redox reactions associated with the xanthophyll cycle, the water-water cycle starting with the Mehler reaction, the  
glutathione-ascorbate cycle and the ferredoxin thioredoxin system (for overviews see e.g. Asada, 2006; Foyer and Noctor,  
2014; Choudhury et al., 2016, Ruban, 2016; Kang et al. 2019). We speculate that Thus during initial high CO<sub>2</sub>-ramping, when  
625 ETR and A frequently co-evolved, and were still not feedback-inhibited, a variable portion of PSII electrons was diverted from  
MT-biosynthesis and the CBB-cycle for repair and protective mechanisms in addition to that for the CBB cycle. This initial  
decrease depending on NPQ relaxation was essentially caused by the lack of reduction equivalents while ATP synthesis was  
still maintained by linear and cyclic electron flow, which subsequently became rapidly compromised with the onset of TPU  
limitation thus explaining the uneven, non-linear character of high-CO<sub>2</sub>-inhibition. We are unable to quantify the losses and  
630 altered allocation of photochemical energy and carbon precursors linked to the photooxidative stress that occurred in our  
experiments. Based on the measured Fv/Fm values, the total loss of the leaf's capacity for PSII electron transport during CO<sub>2</sub>  
ramping was about 10%, to which would be added the reduction in availability use of reduction equivalents due to their use in  
photostress-related redox systems. For comparison, the calculated amount of electrons spent for MT emissions (é-losses) rarely

exceeded 1 % ( $0.57 \pm 0.17$  and  $0.80 \pm 0.21$  at 30 °C and 35 °C respectively), of which less than the half ~~was~~ used in the MEP-  
635 pathway (12 of the total 28 moles reduction equivalents for 1 mole MT; Sharkey and Monson (2014)). Hence, the fraction of  
excess electrons used for MT synthesis was very small compared to the total stress-related ETR reduction and to other  
alternative electron sinks in general (Dani et al., 2014b). Furthermore, photooxidative stress also occurred at 35 °C (albeit to a  
lower extent than at 30 °C), when emissions were much higher but hardly affected by high-CO<sub>2</sub> exposure. ~~These facts~~  
~~collectively shed doubts that the loss of ETR by photooxidative stress was the sole cause of the initial high CO<sub>2</sub> emission~~  
640 ~~inhibition.~~ These facts suggest that additional processes must have determined the availability of energetic cofactors and carbon  
intermediates for MT synthesis during high CO<sub>2</sub> exposure. Given the relative long exposure times applied in our study, these  
have likely included regulation of enzymes activities at transcriptional level. For example, photooxidative stress induces the  
biosynthesis of downstream higher isoprenoids such as carotenoids and tocopherols via retrograde signals of ROS, MEP-  
pathway precursors or carotenoid degradation products (Xiao et al. 2012; Ramel et al., 2013; Foyer, 2018; Jiang and Dehesh,  
645 2021). This might have curtailed the synthesis of MTs through competition for the same precursors, or oppositely enhanced it  
by relieving feedback inhibitions in the MEP-pathway thus keeping precursors at higher levels (Behnke et al. 2009; Banerjee  
et al., 2013; Ghirardo et al., 2014; Rasulov et al., 2014; Zuo et al. 2019; Sun et al., 2020 and references therein).  
In summary, while the data of our study clearly indicate a link between photosynthetic electron transport and CO<sub>2</sub>-  
responsiveness of MT emissions from QI leaves, they do not allow us to confirm or refute current hypotheses about the CO<sub>2</sub>-  
650 sensitivity of isoprenoid emissions, which might not be mutually exclusive but have interacted during the course of CO<sub>2</sub>-  
ramping. Shifts in the availability of limiting metabolites are complex and possibly did not really reach a steady state within  
the applied time steps of our protocol, but continuously adjusted and fluctuated among sources and sinks of energetic cofactors  
constantly interacting with the availability of carbon intermediates including feedback and feedforward controls within the  
MEP pathway (for overview see Sharkey and Monson, 2014; and de Souza et al., 2018). At timescales over hours, these shifts  
655 have likely included regulations of enzymes activities at transcriptional level (e.g. Hartikainen et al., 2012; Kanagendran et al.  
2018). Photooxidative stress for example induces the biosynthesis of downstream higher isoprenoids such as carotenoids and  
tocopherols, possibly via retrograde signals of ROS, MEP pathway precursors or carotenoid degradation products (Xiao et al.  
2012; Ramel et al., 2013; Foyer, 2018; Jiang and Dehesh, 2021), which in turn could curtail the production of MTs through  
competition for the same precursors, or oppositely enhancing it by relieving feedback inhibitions in the MEP pathway and  
660 keeping a higher level of MT precursors (Behnke et al. 2009; Banerjee et al., 2013; Ghirardo et al., 2014; Rasulov et al., 2014;  
Zuo et al. 2019; Sun et al., 2020 and references therein).  
The results of our simulations suggest that at annual scale, the observed emission inhibition under double CO<sub>2</sub> is unlikely to  
compensate for the emission increase from projected warming. However, even though we combined a wide range of scenarios  
in these simulations, the general validity of these results and their extrapolation to other BVOC-emitting species should be  
665 viewed with caution for several reasons: An important determinant of the annual weight of CO<sub>2</sub> inhibition was the degree of  
its temperature dependence. The slope (2% °C<sup>-1</sup>, see Supplement 3) we derived from the measurements on plants grown at  
elevated CO<sub>2</sub> and elevated temperature is only about half that reported in Potosnak et al. (2014) and Sharkey and Monson



(2014). This may indicate that our simulations underestimated CO<sub>2</sub> inhibition at temperatures below 30 °C. However, the results shown in those studies were obtained for isoprene with other tree species at more than double [CO<sub>2</sub>], limiting their validity for our study. Another issue that questions the transferability of our results concerns the Ci values. Because the sclerophyllous leaves of QI assimilate CO<sub>2</sub> at relative high rates with respect to their stomatal conductance, the Ci will be lower in QI than in many other deciduous tree species at same ambient [CO<sub>2</sub>]. For example, in our study, the ratio of Ci to 400 ppm ambient [CO<sub>2</sub>] was 0.55, whereas global emission and vegetation models typically assume a ratio of 0.7 (e.g., Guenther et al., 2012; Grote et al., 2014). Therefore, QI emissions may be less inhibited under elevated [CO<sub>2</sub>] than other species. Changes in Ci may also play a role in emission responses to drought, which were not accounted for in our simulations. Under moderate water deficit, when photosynthetic processes are still fully active, stomata partially close to conserve water. As a result, Ci decreases and leaf temperature increases (due to lower evaporative cooling), reducing the inhibition of emissions by high CO<sub>2</sub>. This phenomenon may explain why emissions sometimes increased during the initial phase of water stress (e.g., Pegoraro et al., 2007; Staudt et al., 2008). However, during severe drought events, which are expected to increase in the future (Gao and Giorgi, 2008), emissions decrease, as assumed in our simulations. There are other factors that could change the annual VOC budget of leaves in a warmer and CO<sub>2</sub>-rich world, such as earlier leaf development, longer leaf lifespan and associated duration of emissions (Staudt et al., 2017; Mochizuki et al., 2020). Further studies are needed to gain in-depth knowledge of the variation in CO<sub>2</sub> sensitivity of QI emissions, especially in mature trees under real field conditions. In particular, it would be interesting to know whether the temperature sensitivity is stable throughout the day and year, or whether it changes with actual light conditions and photoperiod, plant water and phenological status due to changes in the production and allocation of energetic cofactors and precursors of the MEP pathway within the chloroplasts (see, e.g., Sun et al., 2012; Grote et al., 2014; Monson et al., 2016). Generally, the extrapolation of the emission responses to high CO<sub>2</sub> we observed should be viewed with caution, since plants under natural conditions do not undergo CO<sub>2</sub> response curves like those in our study, in which emission reductions were partially related to simultaneous inhibition of photosynthetic processes and oxidative stress. On the other hand, we found little evidence that emission inhibition by high CO<sub>2</sub> was influenced by the preceding exposure to low CO<sub>2</sub>. Furthermore, the occurrence of oxidative stress associated with photosynthetic feedback limitation is more likely in a high CO<sub>2</sub> world. Keeping this in mind, we attempted to estimate the extent to which the observed emission inhibition by double [CO<sub>2</sub>] in a future warmer world could offset the emission increase by the congruent increase in temperature. Doubling assay [CO<sub>2</sub>] from 400 to 800 ppm reduced temperature normalized emission rates on average by 8 % at 30 °C (1626 ± 575 vs. 1499 ± 538; P = 0.001; paired t-test; n = 26) and had no effect at 35 °C (1781 ± 605 vs. 1800 ± 719; P = 0.755; paired t-test; n = 20). Regarding the individual growth populations, double [CO<sub>2</sub>] significantly decreased 30° emissions only in the 800/25 and 400/20 grown plants by respectively 10 % and 5 %. Yet, it is very likely that the high-CO<sub>2</sub> inhibition becomes stronger at temperatures lower than 30°C. Several studies on isoprene emissions reported a negative linear relationship between high CO<sub>2</sub> inhibition and leaf temperature perhaps dumping emissions at 800 ppm CO<sub>2</sub> by 40 % or more under lowest temperatures (Rasulov et al., 2010; Sharkey and Monson, 2014; Zuo et al., 2019; Niinemets et al., 2021; and references therein). In turn, the short term increase in emissions per one degree temperature increase is about 13 % in the range between 10 and 30 °C and then gradually decreases

## 5 Conclusions

The results from CO<sub>2</sub> response curves measured at 30°C assay temperature showed that MT emissions from QI become essentially inhibited under very high [CO<sub>2</sub>] ( $\geq 1200$  ppm), whereas [CO<sub>2</sub>] lower than 400 ppm both increased and decreased emissions. This CO<sub>2</sub>-responsiveness was clearly temperature dependent. High assay temperature (here 35 °C) neutralized the high-CO<sub>2</sub> inhibition of emissions and accentuated the emission decrease at low [CO<sub>2</sub>]. In addition, growth temperature influenced CO<sub>2</sub>-responsiveness and this in the opposite way than did assay temperature. Emissions of plants grown under an elevated temperature regime were more inhibited by high [CO<sub>2</sub>] than plants grown under lower temperatures. Growth under elevated CO<sub>2</sub> had no significant effect on the CO<sub>2</sub>-response of emissions though it enhanced the leaf growth of the plants. The CO<sub>2</sub>-responsiveness of emissions was not also not different between chemotypes and was similar for all individual major MTs. Correlation analyses suggest that the changes in MT emissions in response to CO<sub>2</sub> changes were mainly driven by concurrent changes in the availability of energetic cofactors from photosynthetic electron transport, which are required to maintain monoterpene synthesis in chloroplasts. However, at lowest [CO<sub>2</sub>] MT production was likely co-constrained by the availability of basic carbon substrates as indicated by a relationship between the drop in emissions and the leaf's initial CO<sub>2</sub>-assimilation rate. We hypothesize that several processes, whose magnitudes changed during the different phases of CO<sub>2</sub>-ramping and which differed between leaf and plant replicates, determined the availability of energetic cofactors. These included, on the one hand, changes in their production due to photoinhibition and photooxidative damage or feedback inhibition of photosynthesis (TPU limitation) and, on the other hand, changes in their distribution between MT synthesis, CO<sub>2</sub> fixation and photorespiration, non-photochemical quenching, and repair and detoxification mechanisms associated with oxidative stress. The results of the correlation analyses also suggest that the growth performance of the plants (leaf mass) was related to the ability of their leaves to produce photosynthates and MTs.

Overall, our results confirm an isoprene-analogous behavior of MT emissions from QI. Fitting the algorithm used in MEGAN to account for CO<sub>2</sub> effects to our emission data obtained at 30° resulted in a nonlinear response curve that is very similar to the curve currently used for isoprene emissions. In addition, we performed several simulations to estimate the annual BVOC release from QI leaves under a warmer climate at double atmospheric [CO<sub>2</sub>]. The results showed that the observed emission inhibition at 800 ppm CO<sub>2</sub> would be insufficient to offset the increase in foliar emissions due to the projected warming. Our results confirm an isoprene-analogous behavior commonly attributed to MT emissions from QI and clarify and provide more depth on several aspects regarding their CO<sub>2</sub>-responsiveness: First, they corroborate earlier studies (Loreto et al., 1996b, Staudt et al., 2001; Rapparini et al., 2004) showing that MT emissions become essentially inhibited under very high [CO<sub>2</sub>] (C<sub>i</sub> > 500 ppm), whereas smaller CO<sub>2</sub> variations (C<sub>i</sub>: 200-500) affect little emissions. Second, contrary to the conclusions of Loreto et al. (2001), CO<sub>2</sub>-responsiveness is the same for all major MTs, regardless of tree chemotype. Third, the CO<sub>2</sub>-responsiveness is clearly temperature dependent. High leaf temperatures reduce the high-CO<sub>2</sub> inhibition of emissions, which can explain seasonal



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