



# Biomarker characterization of the North Water Polynya, Baffin Bay: Implications for local sea ice and temperature proxies

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**Abstract.** The North Water Polynya (NOW, Greenlandic Inuit: *Pikialasorsuaq*), Baffin Bay, is the largest polynya and one of the most productive regions in the Arctic. This area of thin to absent sea ice is a critical moisture source for local ice sheet sustenance and coupled with the inflow of nutrient-rich Arctic Surface Water, supports a diverse community of Arctic fauna and indigenous people. Although paleoceanographic records can provide critical insight into the NOW's past behavior, it is critical that we better understand the modern functionality of paleoceanographic proxies. In this study, we analyzed lipid biomarkers, including algal highly-branched isoprenoids and sterols for sea ice extent and pelagic productivity, and archaeal GDGTs for ocean temperature, in a suite of modern surface sediment samples from within and around the NOW. Our data show that all highly-branched isoprenoids exhibit strong correlations with each other and show highest concentrations within the NOW, which suggests a spring/autumn sea ice diatom source rather than a combination of sea ice and open water diatoms as seen elsewhere in the Arctic. Sterols are also highly concentrated in the NOW and exhibit an order of magnitude higher concentration here compared to sites south of the NOW, consistent with the order of magnitude higher primary productivity observed within the NOW relative to surrounding waters in spring/summer months. Finally, our temperature calibrations for GDGTs and OH-GDGTs reduce the uncertainty present in global temperature calibrations, but also identify some additional variables that may be important in controlling their local distribution, such as nutrient availability and dissolved oxygen. Collectively, our datasets provide new insight into the utility of these lipid biomarker proxies in high-latitude settings and will help provide a refined perspective on the Holocene development of the NOW with their application in downcore reconstructions.

## 1 Introduction

Arctic and Antarctic polynyas are key sites for deep water formation (Kuhlbrodt et al., 2017), moisture sources for adjacent ice sheets (Smith et al., 2010), and enhanced productivity that can sequester atmospheric CO<sub>2</sub> (Arrigo and van Dijken, 2004). As global temperatures continue to climb, further reductions in sea ice are projected along Arctic coastlines (Barnhart et al., 2015; Onarheim et al., 2018), calling the future status of polynyas into question. Polynyas will cease to exist where seasonal sea ice vanishes or transitions to marginal sea ice zones, which will both result in cascading negative effects on regional and

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global environments (Meredith et al., 2019; Moore et al., 2021). One way to alleviate some uncertainty about the future status of polynyas is by reconstructing changes in sea ice extent and productivity in the recent geologic past to understand how polynyas have responded to past climate change. In this light, the Holocene instability of coastal polynyas has recently been shown in the Barents Sea (Knies et al., 2018), East Greenland (Syring et al., 2020) and northern Baffin Bay (Jackson et al., 2021; Ribeiro et al., 2021) using lipid biomarker climate proxies, such as IP<sub>25</sub> (sea ice) and sterols (open water productivity). However, understanding the modern distributions of biomarkers and the influencing environmental factors within dynamic Arctic regions is critical before making down-core interpretations (e.g., Smik and Belt, 2017; Belt et al., 2019; Kolling et al., 2020).

Here, we focus on the North Water Polynya (NOW, Greenlandic Inuit: *Pikialasorsuaq*), which is the largest polynya (85,000 km²) and one of the most productive regions in the Arctic (Barber et al., 2001; Klein et al., 2002). The latent heat NOW forms when an ice bridge consolidates at the head of Smith Sound, which restricts the passage of Arctic ice floes but allows throughflow of nutrient-rich Arctic Surface Water (ASW) into Baffin Bay through Nares Strait (Fig. 1, Melling et al., 2001). This throughflow and open water fuels high local productivity, which supports a diverse community of Arctic fauna, including higher-trophic level seabirds and marine mammals (Stirling, 1980; Tremblay et al., 2002a). Due to the rich biodiversity, the region has also supported intermittent human occupation for at least 4000 years (Dorset, 500 BCE to 1500 CE; Thule, 1000 to 1600 CE, Schlederman, 1980), with modern Inuit inhabitants continuing to rely on the NOW for their food security and subsistence economy today (Hastrup et al., 2018). Given that stable ice arches fail to form as reliably as they once did (Moore et al., 2021), the NOW is becoming geographically and seasonally less defined (Ryan and Münchow, 2017).

In this study, we aim to characterize the NOW through the distribution of lipid biomarkers archived in marine seafloor surface sediments taken in 2008 and 2017 that encompass its modern area in Baffin Bay. We focus on different lipid classes that inform us about seasonal sea ice extent, surface productivity, and ocean temperature. Our assessment of these biomarker proxies against modern instrumental data provides a key baseline for interpreting the presence and extent of the NOW in the geologic past (Georgiadis et al., 2020; Jackson et al., 2021). On a broader scale, our work is also critical for the community's general understanding of these lipids' environmental relationships at high northern latitudes where some proxy datasets are currently sparse (e.g., Tierney and Tingley, 2014, 2018). For example, existing biomarker temperature calibrations are often global in scale and feature high uncertainties at the low end of the temperature spectrum (Kim et al., 2010, 2012). However, uncertainty can be substantially reduced by filling in high latitude regions and isolating their distinct characteristics with local calibrations (Tierney and Tingley, 2014, 2018; Harning et al., 2019).

## 2. Study Area

Ocean circulation in Baffin Bay is cyclonic, involving the northward flowing West Greenland Current (WGC) and the southward flowing Baffin Current (BC) (Fig. 1). The warm and saline WGC carries a mixture of Atlantic Water from the Irminger Current and Polar Water from the East Greenland Current, whereas the BC is comprised of low salinity Arctic surface water (ASW) that enters Baffin Bay from the Arctic Ocean through the Canadian Arctic Archipelago (CAA) channels. The



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ASW is modified by mixing with terrestrial-derived freshwater and by sea-ice processes en route to Baffin Bay (Tang et al., 2004; Münchow et al., 2006, 2015, Azetsu-Scott et al., 2010). The present-day depths of the CAA channels govern the composition of inflowing ASW (Jones et al., 2003); Nares Strait has a sill depth of 220 m that allows passage of both the Polar Mixed Layer (containing high-nutrient Pacific Water from Bering Strait) and some of the halocline layer that has been mixed with the underlying Atlantic layer of the Arctic Ocean (Azetsu-Scott et al., 2010). Lancaster and Jones Sounds have shallower sill depths that exclude all but the most carbonate-undersaturated Polar Mixed Layer (Azetsu-Scott et al., 2010). These Arctic outflows join the BC and form the upper 100 to 300 m of surface water in Baffin Bay, except where the WGC dominates in the southeast (Tang et al., 2004).

Sea ice covers nearly all of Baffin Bay in winter, except in the southeast due to the warmth and salinity of the WGC (Fig. 1). Sea ice begins to form in September and reaches maximum coverage in March and is thickest along the Baffin Island coast where the ASW flow is concentrated (Fig. 1, Tang et al., 2004). In contrast, the NOW has low concentrations of thin sea ice, even during winter months. Consolidation of an ice arch at the head of Smith Sound initiates the formation of the polynya, which is further stimulated by northerly winds and currents that remove newly formed sea ice (Ingram et al., 2002; Bi et al., 2019), and sensible heat from WGC upwelling on the Greenland side (Melling et al., 2001; Ingram et al., 2002). Baffin Bay sea ice concentrations decrease between April and August; beginning in the NOW region before propagating southward and creating a generally ice-free Baffin Bay by June (Bi et al., 2019). The Pacific Water, a major component of the ASW, has twice the nitrogen and phosphorus and seven times the silica of Atlantic Water (Jones et al., 2003). The high nutrient content of incoming ASW, along with higher light levels and stratification in the NOW, fuels high seasonal phytoplankton productivity (Lewis et al., 1996; Ingram et al., 2002; Tremblay et al., 2002a). Productivity is an order of magnitude higher in the NOW than in adjacent areas of Baffin Bay, making it one of the most important areas for new production in the Arctic (Tremblay et al., 2002a).

## 3. Background on lipid biomarkers

Highly branched isoprenoids (HBIs) are unsaturated hydrocarbons biosynthesized by a narrow range of marine diatoms (see review by Belt, 2018). The mono-unsaturated HBI termed IP<sub>25</sub>, first discovered in Canadian Arctic sea ice (Belt et al., 2007), has developed into an important seasonal sea ice proxy due to its production during spring blooms (Brown et al., 2011; Limoges et al., 2018; Amiraux et al., 2019) of Arctic sea ice diatoms (Brown et al., 2014). Based on a distinctly heavy stable carbon isotopic composition, in addition to similar concentration profiles to IP<sub>25</sub> across Arctic marine surface sediment, the diunsaturated HBI II also likely has an Arctic sea ice diatom source (Belt et al., 2008; Cabedo-Sanz et al., 2013; Brown et al., 2014; Limoges et al., 2018). IP<sub>25</sub> and HBI II below the limit of detection have often been interpreted as either reflecting a lack of seasonal sea ice cover or permanent and thick sea ice that blocks sunlight penetration needed for sea ice diatom photosynthesis. However, this is likely an oversimplification of a broader range of scenarios that result in absent IP<sub>25</sub> (Belt, 2018). Other HBIs, such as the tri-unsaturated isomers HBI III and IV, have been attributed to biosynthesis by open-water phytoplankton (Belt et al., 2000, 2008, 2015, 2017; Rowland et al., 2001), which may help in differentiating between



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open water or thick sea ice conditions inferred from IP<sub>25</sub> and HBI II in the Arctic (Cabedo-Sanz et al., 2013; Smik et al., 2016; Köseoğlu et al., 2018). Along with certain sterols (see following paragraph), the PIP<sub>25</sub> index has been developed for semi-quantitative sea-ice reconstructions (e.g., Müller et al., 2011):

$$PIP_{25} = \frac{IP_{25}}{IP_{25} + (phytoplankton\ biomarker\ x\ c)},\tag{1}$$

where the balance factor c is a ratio of mean IP<sub>25</sub> and mean phytoplankton biomarker concentrations. Although in some regions high concentrations of HBI III have also been associated with highly productive marginal ice zones (Barents Sea, Belt et al., 2019), marine fronts (North Iceland Shelf, Harning et al., 2021) and sea ice (Amiraux et al., 2019, 2020; Koch et al., 2020) that may obscure PIP<sub>25</sub>-derived indices, recent compilations of Arctic surface sediments show that PIP<sub>25</sub>-based indices broadly correlate with spring and autumn sea ice concentrations (Xiao et al., 2015; Kolling et al., 2020).

Sterols are ubiquitous components in eukaryotic organisms (Volkman,1986), and similar to HBI III and IV, have become common complimentary biomarkers in IP<sub>25</sub> and PIP<sub>25</sub> datasets. Although these biomarkers have often been attributed to specific sources, such as pelagic phytoplankton (brassicasterol, e.g., Navarro-Rodriguez et al., 2013), dinoflagellates (dinosterol, e.g., Boon et al., 1979) and vascular plants (campesterol and β-sitosterol, e.g., Huang and Meinschein, 1976), they are now known to derive from variable sources that complicate their source specificity. For example, brassicasterol and dinosterol are also found in sea ice (Nichols et al., 1990; Belt et al., 2013, 2018) and pennate diatoms (e.g., Volkman et al., 1993; Rampen et al., 2010), and campesterol and β-sitosterol can be produced by diatoms as well (Belt et al., 2013, 2018; Rampen et al., 2010). Hence, the abundance of these sterols within marine sedimentary records are more broadly reflective of general marine productivity (e.g., Köseoğlu et al., 2019). In the CAA where terrestrial biomass is low (Gould et al., 2003), we further assume that the contribution of terrestrial-derived campesterol and β-sitosterol is minimal compared to that produced in the ocean.

Isoprenoid GDGTs are cell membrane-spanning lipids biosynthesized by archaea (Pearson and Ingalls, 2013), including ammonia oxidizing Thaumarchaeota (Schouten et al., 2002; Könneke et al., 2005; Pitcher et al., 2011; Besseling et al., 2020). Thaumarchaeota can modify the degree of cyclization in GDGTs in response to *in situ* temperature variability, a process known as homeoviscous adaptation (e.g., Elling et al., 2015). Thus, the correlation between the degree of cyclization in global surface sediment datasets and upper ocean temperature results in various versions of the tetraether index of tetraethers consisting of 86 carbons (TEX<sub>86</sub>) paleothermometer (Schouten et al., 2002, 2013; Kim et al., 2010, 2012). Evidence from a latitudinal transect in the western Atlantic Ocean demonstrates that GDGTs are most likely produced and exported to the seafloor from 80–250 m water depth (Hurley et al., 2018), which compares well to archaea abundance maxima at 200 m water depths in the Pacific Ocean (Karner et al., 2001). Considering that Thaumarchaeota are chemolithoautotrophs that perform ammonia oxidation (conversion of ammonia to nitrite), they are typically more abundant around the primary nitrite maximum near the base of the photic zone (Church et al., 2010; Francis et al., 2005; Hurley et al., 2018) and are most productive when there is minimized phytoplanktic competition over ammonia (Schouten et al., 2013). In the higher latitudes, the latter occurs during the less productive dark winter months when photosynthesis for sea surface species is inhibited, which may explain the



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seasonal winter temperature bias of GDGTs observed in this latitudinal band (Herfort et al., 2006; Rueda et al., 2009; Rodrigo-Gámiz et al., 2015; Harning et al., 2019). Although the temperature relationship of TEX<sub>86</sub>-based indices deviates from the linear global temperature calibrations and features higher uncertainty at the cold end of the spectrum (Schouten et al., 2002; Kim et al., 2010, 2012), regional calibrations have proven useful for reducing estimate uncertainty (Harning et al., 2019) that may be at least partially attributed to changes in community composition (e.g., Elling et al., 2017). Moreover, new indices based on hydroxylated isoprenoid GDGTs (OH-GDGTs, e.g., RI-OH and RI-OH' indices, Lü et al., 2015) also produced by planktic Thaumarchaeota (Elling et al., 2017; Bale et al., 2019) have been suggested to improve upon TEX<sub>86</sub>-based proxies in the polar oceans (Fietz et al., 2013, 2020; Huguet et al., 2013). This is because the addition of one hydroxyl group may further reduce membrane rigidity at lower temperatures (Huguet et al., 2017). In addition to temperature, recent studies have shown that several other environmental and geochemical factors can influence the degree of GDGT cyclization, such as growth phase (Elling et al., 2014), ammonia oxidation rates (Hurley et al., 2016), and oxygen concentrations (Qin et al., 2015), but not likely salinity (Wuchter et al., 2004, 2005; Elling et al., 2015). Although the effects of these environmental parameters on OH-GDGT cyclization have not been rigorously tested, emerging evidence suggests that salinity, sea ice, seasonality, and terrestrial input may be complicating factors in some oceanic settings (Fietz et al., 2013; Kang et al., 2017; Lü et al., 2019; Park et al., 2019; Wei et al., 2020).

## 4. Materials and Methods

# 145 4.1. Marine surface sediment samples

We studied marine surface sediment samples (*n*=13) collected from box core and trigger core tops collected during the 2008 CSS *Hudson* and 2017 CSS *Amundsen* research cruises, and range in water depths from 267 to 2373 m bsl (Table 1). As each sample integrates the upper 1 to 2 centimeters of sediment, they may reflect several decades to centuries of time depending upon site-specific sedimentation rates and mixing by bioturbation. Therefore, each sample reflects time averages of biomarker production in and out of the polynya. Marine surface sediment samples collected on these cruises as well as other cruises over the past decades have previously been analyzed for HBIs and select sterols (i.e., brassicasterol and dinosterol) as part of larger pan-Arctic datasets (Stoynova et al., 2013; Kolling et al., 2020). We compare our new datasets with a subset of samples (n=70) previously published by Kolling et al. (2020). To avoid environmental settings that may be disproportionately impacted by terrestrial and/or glacier runoff, we exclude samples collected from within fjords (e.g., Kangiqtugaapik, formerly Clyde Inlet) and bays (e.g., Disko Bugt). Moreover, while the International Hydrographic Organization defines the southern limit of Baffin Bay as 70 °N (Limits of Oceans and Seas, 1953), we extend our range into Davis Strait to 67 °N to provide greater breadth in the environmental gradients.

## 4.2. Bulk geochemistry

At the University of Colorado Boulder's Earth Systems Stable Isotope Laboratory, freeze dried marine surface sediment subsamples were analyzed for bulk elemental (%CaCO<sub>3</sub>, %TC, %TN) and stable isotope ( $\delta^{13}$ C relative and  $\delta^{15}$ N) geochemistry



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on a Thermo Delta V elemental analyzer (EA) interfaced to an isotope ratio mass spectrometer (IRMS), and computed for elemental C/N values.  $\delta^{13}$ C values are expressed relative to Vienna Pee Dee Belemnite (VPDB) and  $\delta^{15}$ N relative to Air.

#### 4.3. Lipid biomarkers

At the University of Colorado Boulder's Organic Geochemistry Laboratory, freeze dried marine surface sediment subsamples (~1-4 g) were extracted two times on a Dionex accelerated solvent extractor (ASE 200) using dichloromethane (DCM):methanol (9:1, v/v) at 100 °C and 2,000 psi. A 25 percent aliquot of the total lipid extract (TLE) was taken for GDGT analysis. The remaining 75 percent of TLE was then separated into five fractions (F1-F5) using silica column chromatography, after elution with ½ dead volume (DV) hexane (F1), 2 DV hexane:DCM (8:2, v/v) (F2), 2 DV DCM (F3), 2 DV DCM:ethyl acetate (EtOAc) (1:1, v/v) (F4) and 2 DV EtOAc (F5). Each of these extractions contained 500 ng of the following internal standards: 3-methylheneicosane (F1), p-Terphenyl-d14 (F2), docosanoic acid (F3), 1-nonadecanol (F4), 2-Me octadecanoic acid (F5).

From F1, we focus on highly branched isoprenoid (HBI) IP<sub>25</sub> (C<sub>25:1</sub>), HBI II (C<sub>25:2</sub>), HBI III (C<sub>25:3</sub>) and HBI IV (C<sub>25:3</sub>) biomarkers. HBIs were analyzed via gas chromatography-mass spectrometry (GC-MS) on a Thermo Trace 1310 Gas Chromatograph interfaced to a TSQ Evo 8000 triple quadrupole mass spectrometer and fitted with an Agilent DB-1MS GC column (60 m x 250 μm x 250 μm) following modified methods and operating conditions of Belt et al. (2012). We used an ion source temperature of 250 °C rather than 300 °C to prevent excessive fragmentation during ionization, which yields larger molecular ions that facilitated compound identification both in full scan (FS) and selected reaction monitoring (SRM) modes (e.g., Boudinot et al., 2020). The identification and quantification of IP<sub>25</sub> (*m/z* 350, Belt et al., 2007), HBI II (*m/z* 348, Belt et al., 2007), and HBI III and IV (*m/z* 346, Belt et al., 2000) was based on their respective mass spectra compared to that of the internal standard (3-methylheneicosane, *m/z* 310.6). To account for the varying response factors of different lipid classes and to make our sample set comparable with other published HBI datasets, we corrected calculated concentrations by the response factor of a 5-point external HBI dilution series comprised of 7-hexylnonadecane (7-HND) and 9-octylheptadec-8-ene (9-OHD) (Belt et al., 2012).

From F4, we focus on a series of diagnostic sterols, namely brassicasterol (24-Methylcholesta-5,22*E*-dien-3β-ol), dinosterol (4α,23,24-Trimethyl-5α-cholesta-22*E*-en-3β-ol), campesterol (24-Methylcholesta-5-en-3β-ol) and β-sitosterol (24-Ethylcholesta-5-en-3β-ol). Before analyses, each sample was derivatized with N,O-bis(trimethylsilyl)trifluoroacetamide (BSTFA; 25 μl) and pyridine (catalyst, 25 μl) at 70 °C for 20 min.). Sterols were analyzed using the same GC-MS system described above but with an Agilent DB-5MS GC column (60 m x 250 μm x 250 μm) and under the following operating conditions: initial temperature of 80 °C (held for 2 min.), ramp 20 °C/min. (2.5 min.), ramp 5 °C/min. (68 min., held for 30 min.). Mass spectrometric analyses were carried out in full scan (FS) and selected reaction monitoring (SRM) modes. Quantification of individual sterols was achieved by comparison of their respective mass spectra (Boon et al., 1979) to that of the internal standard (1-nonadecanol, *m/z* 284.5). To account for the varying response factors of different lipid classes and to



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195 make our sample set directly comparable with Kolling et al. (2020), we corrected calculated concentrations by the response factor of a 5-point external dilution series of cholesterol.

For GDGTs, we focus on isoprenoid and hydroxylated isoprenoid GDGTs. A 25 percent aliquot of dry TLE samples was resuspended in hexane:isopropanol (99:1, v/v), sonicated, vortexed, and then filtered using a 0.45 µm polytetrafluoroethylene (PTFE) syringe filter. Prior to analysis samples were spiked with 10 ng of the C<sub>46</sub> GDGT internal standard (Huguet et al., 2006). GDGTs were identified and quantified via high performance liquid chromatography-mass spectrometry (HPLC-MS) following modified methods of Hopmans et al. (2016) on a Thermo Scientific Ultimate 3000 HPLC interfaced to a Q Exactive Focus Orbitrap-Quadrupole MS (Harning et al., 2019). GDGTs were identified based on their characteristic masses and elution patterns. For isoprenoid GDGTs, we explore the original TEX86 index (Schouten et al., 2002) and the more recent TEX<sub>86</sub><sup>L</sup> index, which is a modification of the former for temperatures <15 °C (Kim et al., 2010, 2012), to reflect relative changes in temperature:

$$TEX_{86} = \frac{[GDGT-2] + [GDGT-3] + [cren.i]}{[GDGT-1] + [GDGT-2] + [GDGT-3] + [cren.i]},$$
(2)

$$TEX_{86}^{L} = \log(\frac{[GDGT-2]}{[GDGT-1] + [GDGT-2] + [GDGT-3]}),$$
 (3)

To evaluate the degree of influence from non-thermal factors on GDGT-derived indices, we compute the Ring Index for each sample (RI, Zhang et al., 2016):

$$210 \quad RI = 0x[GDGT - 0] + 1x[GDGT - 1] + 2x[GDGT - 2] + 3x[GDGT - 3] + 4x[cren.] + 4x[cren.'], \tag{4}$$

and compare with the global core tope polynomial regression for TEX<sub>86</sub> values (Zhang et al., 2016):

$$RI_{TEX} = -0.77(\pm 0.38)xTEX_{86} + 3.32(\pm 0.34)x(TEX_{86})^2 + 1.59(\pm 0.10),$$
(5)

Finally, for hydroxylated isoprenoid GDGTs, we explore two different relative temperature indices, RI-OH and RI-OH' 215 developed for regions over and under 15 °C, respectively (Lü et al., 2015):

$$RI - OH = \frac{[OH - GDGT - 1] + 2x[OH - GDGT - 2]}{[OH - GDGT - 1] + [OH - GDGT - 2]},\tag{6}$$

$$RI - OH = \frac{[OH - GDGT - 1] + 2x[OH - GDGT - 2]}{[OH - GDGT - 1] + [OH - GDGT - 2]},$$

$$RI - OH' = \frac{[OH - GDGT - 1] + 2x[OH - GDGT - 2]}{[OH - GDGT - 0] + [OH - GDGT - 1] + [OH - GDGT - 2]},$$
(7)

## 4.4. WOA18 datasets

220 To assess and calibrate our GDGT-based proxies against modern climatological fields, we use World Ocean Atlas 2018 (WOA18) decadal mean datasets from 2007 to 2017 CE. We compare GDGTs against various depth integrations (0-200 mbsl) of the following variables: temperature (Locarnini et al., 2018), salinity (Zweng et al., 2018), dissolved oxygen (Garcia et al., 2018a), and nitrate (Garcia et al., 2018b) (Fig. 2). For each of these variables, we compared proxy data against the mean annual values as well as mean seasonal values where complete datasets were available. Since winter and spring seasonal data were 225 either fragmentary or not available, the mean annual datasets used in this study likely represent the ice-free season, integrating summer and shoulder season months.

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## 5. Results

## 5.1. Bulk geochemistry

Although only 9 out of the 13 total surface sediment samples were analyzed for bulk geochemistry, those that were represent the full geographical range of sample types throughout Baffin Bay (Fig. 1). Bi-plots of δ<sup>13</sup>C and C/N tightly cluster within the most typical range of marine algae (Fig. 3, Meyers, 1994).

#### **5.2. HBIs**

HBIs are present above the detection limit in all sediment samples. IP<sub>25</sub> is the most dominant HBI in the dataset, followed by HBI II, HBI III, and HBI IV (Fig. 4a). HBIs are generally more abundant at NOW sites compared to sites outside this region, although there is some overlap when the standard deviation of mean concentrations is considered for IP<sub>25</sub>, HBI II and HBI IV, but not for HBI III (Figs. 4a and S1). Based on Pearson correlation analysis, each of the HBIs strongly correlates with the others (>0.74, Fig. 5).

## 240 **5.3. Sterols**

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Sterols are present above the detection limit in all sediment samples. β-sitosterol is the most abundant sterol in the dataset, followed by campesterol, brassicasterol, and dinosterol (Fig. 4c). Sterols are generally more abundant at NOW sites compared to sites outside this region (Fig. 4c and S2). Although the standard deviations of mean dinosterol and campesterol concentrations overlaps between NOW and non-NOW sites, the standard deviations of β-sitosterol and brassicasterol for the two regions is statistically different (Fig. 4c and S2). Based on Pearson correlation analysis, the sterols all strongly correlate with each other (>0.72), apart from dinosterol and β-sitosterol whose correlation is insignificant (Fig. 5).

## 5.4. PIP<sub>25</sub>

We calculated new Baffin Bay balance factors (c) for P<sub>III</sub>IP<sub>25</sub> (3.19), P<sub>IV</sub>IP<sub>25</sub> (1.02), P<sub>B</sub>IP<sub>25</sub> (0.54), and P<sub>D</sub>IP<sub>25</sub> (0.63) based on a combination of the 70 previously published samples from Baffin Bay (Kolling et al., 2020) and this study's sample set to reflect the combined datasets. Although HBI IV was quantified by Kolling et al. (2020), we note that the authors did not explore its potential as part of a PIP<sub>25</sub> index. For all four PIP<sub>25</sub> indices in our dataset, we observe slightly higher mean values in non-NOW sites compared to NOW sites, although the standard deviation of mean values between these regions is not statistically different (Fig. 4e and S3).

## 5.5. GDGTs and OH-GDGTs

GDGTs and OH-GDGTs are present above the detection limit in all sediment samples. In terms of RI values, all samples plot below the global core top polynomial regression for TEX<sub>86</sub> (Fig. 6a). In terms of total fractional abundance, GDGTs (0.88  $\pm$  0.03) dominate over their OH-GDGT counterparts (0.12  $\pm$  0.03) (Fig. 6b). GDGTs and OH-GDGTs with no cyclopentane



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moieties are the most dominant (e.g., GDGT-0 and OH-GDGT-0), followed by crenarchaeol and then the GDGTs and OH-GDGTs with 1, 2, and 3 cyclopentane moieties in the case of GDGTs (Fig. 6c-d).

In terms of regressions against different environmental variables (e.g., temperature, salinity, dissolved oxygen, and nitrate), we find that the tested GDGT- (TEX<sub>86</sub> and TEX<sub>86</sub><sup>L</sup>) and OH-GDGT-based indices (RI-OH and RI-OH') range in the strength of the correlation coefficients (R<sup>2</sup>) and significance (p values) (Fig. 7). Moreover, differences in the season and depth integration of the environmental variables also appear to influence the strength of correlations (Figs. 7, S4, S5, and S6). For TEX<sub>86</sub>, the strongest significant temperature correlation is achieved with summer SST above 20 m bsl ( $R^2 = 0.21$  to 0.35, Fig. 7a). Although salinity and dissolved oxygen seem to have little confounding influence ( $R^2 < 0.22$ ), we find a moderately strong but significant correlation between annual TEX<sub>86</sub> and nitrate concentrations above 5 m bsl (R<sup>2</sup> = 0.40, Fig. S6). For TEX<sub>86</sub><sup>L</sup>, the strongest significant temperature correlations ( $R^2 > 0.40$ ) are found with annual (>30 m bsl, Fig. 7b). The influence of other environmental variables (e.g., salinity, dissolved oxygen, and nitrate) on  $TEX_{86}^{L}$  appear to be generally weak ( $R^2 < 0.27$ ), although summer nitrate concentration correlations at 0-200 m bsl are slightly higher and significant ( $R^2 = 0.37$ , Fig. S6). For RI-OH, the strongest significant temperature correlations are achieved with annual SST between 10 and 20 m bsl ( $R^2 = 0.44$ to 0.46), although autumn subsurface temperature between 60 and 80 m bsl also feature similarly significant correlations (R<sup>2</sup> = 0.41 to 0.42) (Fig. 7c). The influence of the other tested environmental variables on RI-OH appears to be generally weak ( $R^2$ < 0.27), although summer and annual 0-20 m dissolved oxygen concentration correlations are slightly higher ( $R^2 = 0.39$  and  $R^2 = 0.36$ , respectively, Fig. S5). Finally, for RI-OH', the strongest significant temperature correlations are achieved with annual 0-20, annual 0-30 m SST, and autumn 0-100 m SST, although correlation coefficients for all are relatively low (R<sup>2</sup>= 0.23) (Fig. 7d). Based on our R<sup>2</sup> values, the influence of other environmental variables, namely annual and summer subsurface dissolved oxygen and nitrate below 100 m depth, appears to exert relatively more influence than temperature on RI-OH' (Figs. S5 and S6).

For the two indices that feature the strongest significant correlations with temperature ( $R^2 > 0.40$ ), we derived individual linear temperature calibrations for northern Baffin Bay. The TEX<sub>86</sub><sup>L</sup> calibration ( $R^2 = 0.45$ ) encompasses the surface and subsurface waters from 0-90 m bsl and features a standard error (S.E.) of 0.13 °C (Fig. 9a):

$$subT = 10.573x + 6.636, (10)$$

The RI-OH calibration ( $R^2 = 0.46$ ) encompasses the surface waters from 0-20 m bsl and features a S.E. of 0.26 °C (Fig. 9b): SST = 27.982x - 31.524 (11)

#### 6. Discussion

## 6.1. Spatial variability in surface productivity

A recent and expanded Arctic study by Kolling et al. (2020) explored the efficacy of using a variety of sedimentary HBIs (i.e., 290 IP<sub>25</sub>, HBI II, and HBI III), sterols (i.e., brassicasterol and dinosterol) and their PIP<sub>25</sub>-derived indices to reconstruct sea ice extent/concentration and pelagic productivity by comparison to satellite data. For Baffin Bay, the main conclusions were that P<sub>B</sub>IP<sub>25</sub>, P<sub>D</sub>IP<sub>25</sub> and P<sub>III</sub>IP<sub>25</sub> all exhibited strong correlations with modern spring and autumn seasonal sea ice concentration



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(Kolling et al., 2020). On the other hand, TR<sub>25</sub>, the recently proposed HBI proxy for spring phytoplankton bloom in the Barents Sea (Belt et al., 2019), did not exhibit a clear relationship with chlorophyll *a*, but instead paralleled spring/autumn/winter sea ice extent (Kolling et al., 2020). For this reason, we did not further explore the controls of chlorophyll *a*, which is a limited and discontinuous dataset in the Baffin Bay region (NASA MODIS), on the distribution of HBI III and HBI IV compounds that comprise the TR<sub>25</sub> proxy (e.g., Belt et al., 2019). However, we do further explore the relationship of HBIs, sterols and PIP<sub>25</sub> indices in their ability to track sea ice extent, and more importantly, the elevated surface productivity associated with the NOW. Moreover, we include several additional sterols (β-sitosterol and campesterol) and introduce another PIP<sub>25</sub> index (P<sub>IV</sub>IP<sub>25</sub>) that have not previously been tested in Baffin Bay. The following discussion considers the datasets from this study and that of Kolling et al. (2020) (Fig. 1).

In terms of HBI concentrations, all compounds were considerably higher in sites within the NOW compared to sites outside the NOW (Fig. 4a-b and S1). For IP<sub>25</sub> and HBI II, the detection of each in all samples is consistent with the spring and autumn sea ice limits in Baffin Bay (Fig. S1, Bi et al., 2019). We also observe a strong positive correlation between IP<sub>25</sub> and HBI II (Fig. 5a-b), which likely supports a common sympagic diatom source as observed elsewhere in the Arctic (Belt et al., 2008; Cabedo-Sanz et al., 2013; Brown et al., 2014; Limoges et al., 2018). Given the distinct oceanographic seasonal characteristics of the NOW (open water) and central Baffin Bay (seasonal sea ice), we expected higher IP25 and HBI II concentration in the sites south of the NOW compared to sites within the NOW itself. One possibility is that these HBIs are produced earlier in the spring or later in the autumn when sea ice covers both the NOW and east coast of Baffin Island (Bi et al., 2019). Another possibility, although seemingly counterintuitive, may be that the higher concentrations of IP25 and HBI II within the NOW reflect the southward transport of drift ice through the open NOW and the production and deposition of IP<sub>25</sub> and HBI II en route. While HBI III and IV are found in relatively lower concentrations than IP25 and HBI II (Fig. 4a-b), all four HBI's spatial distributions are similar (Fig. S1), which presents several possibilities. First, as observed in other regions of the Arctic such as the Barents Sea, increased concentrations of HBI III and IV are associated with the open water of Marginal Ice Zones (Smik et al., 2016; Belt et al., 2015, 2019). In this sense, the concentrations we observe in Baffin Bay are consistent with increased production of these HBIs during the spring transition from seasonal sea ice to open water in the NOW compared to sites outside that maintain seasonal sea ice cover during the summer. Alternatively, recent work that has monitored the production of various HBIs through the spring sea ice melt season in southwest Baffin Bay shows that HBI III is produced under the sea ice before and concurrent with IP25, and therefore, is likely biosynthesized by sympagic diatoms in this region (Amiraux et al., 2019, 2021). Given that all HBIs show limited significant correlations with sterols, which reflect open water productivity, we argue that the latter scenario where HBI III and IV are produced by sympagic diatoms is most likely (Fig. 5ab).

In terms of sterol concentrations, all compounds were considerably higher in sites within the NOW compared to sites outside the NOW (Fig. 4c-d and S2). We do note that the non-NOW data for brassicasterol and dinosterol from Kolling et al. (2020) are proportionally higher than ours, which may relate to the fact that some of theirs sites are situated in front of large Baffin Island fjords where terrestrial detritus may stimulate localized increases in surface productivity. While a recent



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Holocene marine record from Petermann Fjord (Northwest Greenland) interprets campesterol and β-sitosterol as indicators of terrestrial input (Detlef et al., 2021), we note that all sterols are highly correlated in our dataset (Fig. 5a), which suggests a common marine source consistent. This is consistent with our bulk geochemistry data (Fig. 3), their known production by a variety of marine diatoms (Belt et al., 2013, 2018; Rampen et al., 2010), and the low terrestrial biomass of surrounding landmasses (Gould et al., 2003). Although the sterols do not feature the same degree of source specificity that the HBIs do, the spatial variability of our results is broadly consistent with the high seasonal biological productivity that the NOW supports. Moreover, studies based on nutrient concentrations of particulate matter in Baffin Bay show that primary productivity in the NOW is an order of magnitude higher than adjacent areas in Baffin Bay during spring/summer (Tremblay et al., 2002a). The localized and high concentrations of sterols in the NOW compared to non-NOW sites is broadly consistent with these trends (Fig. 4c-d and S2). Given the southward propagation of sea ice, and that the development of the NOW occurs at the transition between spring and summer (Bi et al., 2019), the sterols are likely recording a spring/summer season signal of pelagic productivity. The one site within the NOW that does not exhibit high sterol concentrations is at the northern limit, and therefore either may not receive the full effects of nutrient delivery and warmth from the WGC and/or is still under the influence of residual sea ice drift or Smith Sound blockage during the summer months. We also observe generally higher concentrations of sterols in the central NOW compared to sites on the NOW periphery abutting Jones and Lancaster Sounds that may feature more persistent seasonal sea ice presence than those in the center (Fig. S2).

Finally, in terms of PIP<sub>25</sub>, all four indices (P<sub>III</sub>IP<sub>25</sub>, P<sub>IV</sub>IP<sub>25</sub>, P<sub>B</sub>IP<sub>25</sub>, and P<sub>D</sub>IP<sub>25</sub>) for our samples were lower in sites within the NOW compared to sites outside the NOW (Fig. 4e and S3). On the other hand, P<sub>B</sub>IP<sub>25</sub> and P<sub>D</sub>IP<sub>25</sub> from Kolling et al. (2020) is higher in the NOW compared to non-NOW sites, while P<sub>III</sub>IP<sub>25</sub> and P<sub>IV</sub>IP<sub>25</sub> is consistent with ours (Fig. 4f). The difference in P<sub>B</sub>IP<sub>25</sub> and P<sub>D</sub>IP<sub>25</sub> between datasets likely originates from the different types of studied sample sites, where Kolling et al. (2020) has sites in front of fjords that contain proportionally higher sterol concentrations. While the HBI-derived indices (P<sub>III</sub>IP<sub>25</sub> and P<sub>IV</sub>IP<sub>25</sub>) feature overall higher values compared to those derived from sterols (P<sub>B</sub>IP<sub>25</sub>, and P<sub>D</sub>IP<sub>25</sub>) in both datasets, we caution against the application of the HBI-derived indices in Baffin Bay for sea ice concentration since these biomarkers may all originate from sea ice diatoms (e.g., Amiraux et al., 2019, 2021), rather than a combination of sea ice and open water diatoms. Looking at the sterol-derived indices (P<sub>B</sub>IP<sub>25</sub>, and P<sub>D</sub>IP<sub>25</sub>) for this study's dataset, values indicate that the NOW sites feature reduced sea ice cover (0.3 to 0.5) whereas non-NOW sites feature seasonal (0.5 to 0.7) sea ice cover (Fig. 4e). The spatial variability of our data is consistent with late spring and summer sea ice cover in Baffin Bay, where the highest PIP<sub>25</sub> index values are found off eastern and northeastern Baffin Island where sea ice persists the longest (Bi et al., 2019).

In summary, we have several recommendations for future paleoenvironmental reconstructions in Arctic oceanographic settings. First, for complex oceanographic settings like Baffin Bay, we recommend the analysis of both sterols and HBIs to test the performance of various sea ice cover proxies. If possible, this would be best achieved through the analysis of modern surface sediments and sediment traps over an environmental gradient and through the seasons to capture proxy response to known variable changes (e.g., Navarro et al., 2013; Smik and Belt, 2017; Koch et al., 2020). Second, continued research on sterol and HBI sources and seasonality of production is critical for the development of more refined sea ice and

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marine productivity reconstructions (e.g., Limoges et al., 2018; Amiraux et al., 2019, 2021). This is particularly necessary when combining IP<sub>25</sub>, a known sea ice diatom proxy, with presumably more complicated open water proxies such as HBI III and sterols. Based on our dataset, which shows an order of magnitude higher sterol concentrations within the NOW relative to sites outside of its modern limits, compared to HBIs that show only slighter higher concentrations within the NOW, we suggest that sterols, rather than HBIs, may be a more appropriate tool to characterize the presence/absence of the NOW in the recent geologic past.

## **6.2.** Temperature calibrations

Temperature correlations for GDGTs first relied on empirical correlations between global surface sediments and the variability in biomarker structure (e.g., Schouten et al., 2002). Subsequent iterations and developments of TEX<sub>86</sub>-based indices removed some compounds (e.g., crenarchaeol regioisomer for TEX<sub>86</sub><sup>L</sup>) for improved performance in certain latitudinal bands (Kim et al., 2012). Most recently, Bayesian statistics have been employed to generate spatially varying calibrations (BAYSPAR) that reach a compromise between data-constrained global and regional calibrations to capture regional oceanographic variability and site-specific uncertainty more accurately (Tierney and Tingley, 2014). However, BAYSPAR does not include surface sediment for sensitive oceanographic regions like Baffin Bay (Tierney and Tingley, 2014). For this reason, developing local correlation-constrained calibrations can be particularly useful to capture the nuances of oceanographic variability in regions where global dataset coverage is lacking or where the temperature relationship deviates from global linear calibrations (e.g., Harning et al., 2019; Park et al., 2019; Fietz et al., 2020). Moreover, exploring the potential influence of additional environmental variables (e.g., salinity, DO, nutrients) will help us better understand the mechanisms behind and functionality of this commonly applied paleotemperature proxy. Given the relatively small sample size and narrow temperature range of our dataset compared to global compilations (e.g., Kim et al., 2012; Lü et al., 2015), we acknowledge that our conclusions in this study are relatively limited. However, as additional data is added from Baffin Bay and other high-latitude regions, we will be better poised to test these observations.

#### 385 6.2.1. GDGTs and OH-GDGTs

While a more detailed analysis of intact polar lipid production and genetic diversity in Baffin Bay is lacking, the distribution of GDGT and OH-GDGT in our study area (Fig. 6) and understanding of their production in cultures (Elling et al., 2017) indicate that planktic group 1.1a Thaumarchaeota are likely the dominant producers. Therefore, the global relationship between TEX<sub>86</sub> and the RI (Fig. 6a, black polynomial line) serves as a simple means to evaluate whether the TEX<sub>86</sub>-based indices are influenced by additional nonthermal factors (Zhang et al., 2016). Even though our data exhibit a correlation between TEX<sub>86</sub> and RI values ( $R^2 = 0.46$ ) with a slope like the global polynomial regression, all samples plot below the lower 95% uncertainty limit (Fig. 6a). A recent study from the South China Sea found a similar relationship to ours, in that TEX<sub>86</sub> values were well correlated with RI values, but did not conform to the global polynomial trend's uncertainty (Wei et al., 2020). Wei et al. (2020) posited that the shallow shelf environment (neritic zone) of the South China Sea may result in the observed deviation from the



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395 global polynomial TEX<sub>86</sub>-RI relationship, as shallow water Thaumarchaeota respond differently to temperature than the deep-dwelling communities (e.g., Kim et al., 2015, 2016; Villanueva et al., 2015; Zhu et al., 2016; Jia et al., 2017). While the depths of our sites are deeper than the neritic zone (>200 m bsl, Table 1), many are much shallower than open ocean sites used in the global calibrations (Kim et al., 2010, 2012). One possibility is that the shallower environment of northern Baffin Bay may result in a different response of Thaumarchaeota to temperature and different distribution of GDGTs than in the open ocean.

400 In any case, temperature seems to remain the dominant control on GDGT cyclization in this region.

Our regression analysis against temperature, salinity, DO and nitrate further supports temperature as the dominant environmental control on GDGT distributions in Baffin Bay for the seasons available in the WOA18 dataset. The lack of WOA18 winter temperatures, in addition to the fragmentary dataset for spring temperatures in Baffin Bay (Locarnini et al., 2018), prevents us from assessing the impact of these individual seasons, which is unfortunate given that cold season temperatures in other high-latitude settings exhibit a stronger correlation with GDGT distributions (Herfort et al., 2006; Rueda et al., 2009; Rodrigo-Gámiz et al., 2015; Harning et al., 2019). However, the higher correlation between TEX<sub>86</sub><sup>L</sup> and annual temperatures compared to summer/autumn seasons (Fig. 7b) suggests a bias for Baffin Bay GDGT production towards the cooler seasons. Although we explored the relationship of the original TEX<sub>86</sub> index, we prefer to rely on the low-temperature TEX<sub>86</sub><sup>L</sup> modification to assess GDGT depth and season and temperature relationships. This decision is supported by 1) generally low correlation coefficients for temperature and TEX<sub>86</sub>, except for summer SST (Fig. 7a), and 2) the better correlations of TEX<sub>86</sub><sup>L</sup> and lower water depths where ammonia-oxidation likely occurs (Fig. 6b, e.g., Hurley et al., 2018, Park et al., 2019). The moderate correlation between TEX<sub>86</sub> and summer SST, and the lack of correlation between TEX<sub>86</sub><sup>L</sup> and SST from any season, may indicate that the crenarchaeol regioisomer, which is included in the TEX<sub>86</sub> index but not in TEX<sub>86</sub><sup>L</sup>, may be physiologically advantageous for surface dwelling Thaumarchaeota during the warmer summer months in Baffin Bay. We also cannot rule out the possibility of a yet unknown alternative biological source.

In terms of other tested environmental variables, correlations between TEX<sub>86</sub><sup>L</sup> and salinity and dissolved oxygen are generally poor (R<sup>2</sup> < 0.3, Figs. S4 and S5), consistent with previous reports on the lack of relationship between GDGT production and salinity (Wuchter et al., 2004, 2005; Elling et al., 2015). The presence of a well-oxygenated water column in Baffin Bay (Fig. 2c) likely has little influence on GDGT cyclization, which has been reported to only occur in oxygen-limited environments (e.g., Qin et al., 2015). The correlation between TEX<sub>86</sub><sup>L</sup> and nitrate, while weak, is more pronounced in the lowermost integrated depth (200 m bsl) (Fig. S6). The main product of ammonia-oxidation is nitrite, for which we have no data for in this region. However, since nitrite can be subsequently oxidized to nitrate by bacteria (Kuypers et al., 2018), we assume that nitrate provides indirect evidence for both reactions. This allows us to confirm the subsurface depth habitat of ammonia oxidizing Thaumarchaeota in Baffin Bay, which supports our observations that TEX<sub>86</sub><sup>L</sup> correlates best with annual subT (Fig. 9b).

Our complimentary analysis of OH-GDGTs and environmental variables reveals several differences with the conclusions drawn for GDGTs in Baffin Bay and with OH-GDGTs elsewhere. First, the RI-OH' index, which was developed as a modified version of RI-OH for low-temperature environments (Lü et al., 2015), does not perform as well as the RI-OH



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index in our dataset (Fig. 7c-d). Second, we find that RI-OH is best correlated with annual SST in the top 40 m of the water column (R<sup>2</sup> = 0.46), and with autumn in the subsurface (>50 m depth, Fig. 7c). The slightly stronger correlation between RI-OH and SST compared to subT is supported by Lü et al. (2019), who observed higher concentrations of OH-GDGTs in the upper portion of the water column compared to GDGTs in the East China Sea. Additional studies have found that the ring composition of OH-GDGT often differs from GDGTs, which may suggest that these two lipid classes are sourced from different Thaumarchaeota subgroups or produced in different niches of the water column characterized by different environmental conditions (Liu et al., 2012).

In terms of additional environmental variables, only surface dissolved oxygen appears to exert a partial influence on OH-GDGT cyclization in the RI-OH index (Fig. S5c). However, when OH-GDGT-0 is considered in the RI-OH' index, moderate correlations with dissolved oxygen (Fig. S5d) and nitrate appear within the subsurface waters (Fig. S6d). These latter observations conflict with our RI-OH temperature correlations that suggest a surface-dwelling producer of OH-GDGTs. Instead, this may suggest that OH-GDGT-0 is important for membrane functionality at these depths and/or for these environmental variables. Alternatively, OH-GDGT-0 may also have a different biological source in Baffin Bay, as at least for GDGTs, GDGT-0 can be produced by other groups of archaea (e.g., Pearson and Ingalls, 2013). Given the relatively low correlations between RI-OH', which includes OH-GDGT-0, and temperature, our data further supports the potential role of OH-GDGT-0 for membrane response to these non-thermal environmental variables in Baffin Bay. Although our data suggests that OH-GDGTs, reflected by the RI-OH index, are predominately a proxy for annual SST, more detailed studies on the biological producers, depth habitat, and response to environmental variables of OH-GDGTs will undoubtedly benefit future applications and interpretations of downcore OH-GDGT proxy records.

Following our evaluation of GDGT and OH-GDGT indices in terms of their ability to capture temperature, amongst other environmental variables, we present two temperature calibrations that can benefit future paleoceanographic reconstructions from Baffin Bay (Fig. 8). Like other regional TEX<sub>86</sub><sup>L</sup> temperature calibrations in the greater North Atlantic region (e.g., Harning et al., 2019), our Baffin Bay TEX<sub>86</sub><sup>L</sup> temperature calibration captures a subsurface signal (0-90 m bsl) with a considerably lower S.E. (0.13 °C) compared to the latest global calibration (4.0 °C, Kim et al., 2012) despite the lower R<sup>2</sup> (0.45 vs. 0.87, respectively). The strength of our local TEX<sub>86</sub><sup>L</sup> subT calibration is also supported by the fact that other regional (Harning et al., 2019) and global calibrations (Kim et al., 2012) overestimate the observed WOA18 temperature in Baffin Bay (Fig. 8c), with residuals as high as 5.8 °C for the Iceland calibration and 5.0 °C for the global calibration (Fig. 8e). In terms of the RI-OH SST calibration, to the best of our knowledge there are no regional calibrations to compare with. However, comparison with the global calibration that features a S.E. of 6.0 °C (Lü et al., 2015) again reveals the power of reducing uncertainty with regional calibrations as ours from Baffin Bay features a smaller S.E. of 0.26 °C. While the global TEX<sub>86</sub><sup>L</sup> calibration overestimates Baffin Bay temperatures by up to 5.0 °C, the RI-OH calibration overestimates local temperatures by less than 3.3 °C (Fig. 8d-f). In future studies, we suggest that these two calibrations may bolster the quantification of paleotemperature change in Baffin Bay surface and subsurface waters. For other high-latitude locations that do not yet have local GDGT-temperature calibrations, our comparison between instrumental temperature and global calibration





estimates caution against quantitative interpretations due to large over estimations. However, as the trends between regional and global calibrations are similar (Fig. 8c-d), qualitative interpretations remain valid.

## 465 7. Conclusions

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As global climate change continues in the current century, the sustainability of Arctic polynyas is in jeopardy. While proxy reconstructions of polynyas prior to the instrumental period can shed light on key climate and environmental mechanisms that lead to their presence/absence, a detailed understanding of those proxies is needed. Therefore, we evaluated a series of lipid biomarkers (HBIs, sterols, GDGTs, and OH-GDGTs) in surface sediment samples from Baffin Bay to characterize how these biomarkers capture sea ice and productivity conditions in the North Water Polynya (NOW) as well as inform the utility of commonly applied paleotemperature proxies in Baffin Bay.

- All studied HBIs (IP25, HBI II, HBI III, and HBI IV) exhibit strong correlations with each other and are found in
  highest concentrations within the modern limits of the NOW. We suggest all HBIs are, at least partially, produced by
  sympagic diatoms under the sea ice that is formed in the NOW during spring and autumn, and subsequently advected
  southward along the coast of eastern Baffin Island.
- All studied sterols (dinosterol, campesterol, β-sitosterol, and brassicasterol) exhibit an order of magnitude higher concentrations within the NOW compared to sites further south in Baffin Bay, consistent with the order of magnitude higher spring/summer primary productivity that is observed within the NOW today relative to surrounding waters. Hence, we suggest that sterols, rather than HBIs, are more suitable for open water paleoproductivity reconstructions in this region.
- Application of the GDGT-based TEX<sub>86</sub><sup>L</sup> index is optimal over TEX<sub>86</sub> in Baffin Bay and captures an integration of annual subT consistent with the depth habitat of Thaumarchaeota observed elsewhere around the globe. Other tested environmental variables, such as salinity, dissolved oxygen, and nitrate reveal low correlations, which we infer to reflect negligible influence on GDGT cyclization. We present a local annual subT calibration that provides a considerably lower S.E. (0.13 °C) compared to the latest global calibration (4.0 °C, Kim et al., 2012).
- Application of the OH-GDGT-based RI-OH index is optimal over RI-OH', even though the latter was originally developed for low temperature environments. The RI-OH index captures an integration of annual SST, which may suggest that OH-GDGTs in Baffin Bay are biosynthesized by different microbes than those that produce GDGTs and/or are sensitive to different environmental stresses. Our local annual SST calibration provides a considerably lower S.E. (0.26 °C) compared to the latest global calibration (6.0 °C, Lü et al., 2015).

## **Data Availability**

After acceptance, all data will be stored on the PANGAEA repository.





#### **Author Contributions**

495 JS and DJH designed the study and JS and AEJ funded the study. DJH led the analyses of samples and developed GC-MS methods under the supervision of JS. BH and LW assisted with the extraction and purification of samples. DJH wrote the manuscript with discussion and contribution from all co-authors.

## **Competing Interests**

The authors declare they have no conflicts of interest.

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#### References

505

- Amiraux, R., Smik, L., Köseoğlu, D., Rontani, J.-F., Galindo, V., Grondin, P.-L., Babin, M., and Belt, S. T.: Temporal evolution of IP<sub>25</sub> and other highly branched isoprenoid lipids in sea ice and the underlying water column during an Arctic melting season. Elementa-Sci. Anthrop., 7, 1-23, 2019.
- Amiraux, R., Archambault, P., Moriceau, B., Lemire, M., Babin, M., Memery, L., Massé, G., and Tremblay, J.-E., 2021: Efficiency of the sympagic-benthic coupling revealed by n-3 fatty acids, IP<sub>25</sub> and other highly branched isoprenoid analyses of two filter-feeding Arctic benthic molluscs: *Mya truncate* and *Serripes groenlandicus*. Org. Geochem., 151, 104160.
- Arrigo, K. R., and van Dijken, G. L.: Annual cycles of sea ice and phytoplankton in Cape Bathurst polynya, southeastern Beaufort Sea, Canadian Arctic. Geophys. Res. Lett., 31, L08304, 2004.
- Azetsu-Scott, K., Clarke, A., Falkner, K., Hamilton, J., Jones, P. E., Lee, C., Petrie, B., Prinsenberg, S., Starr, M., and Yeats, P.: Calcium carbonate saturation states in the waters of the Canadian Arctic Archipelago and the Labrador Sea. J. Geophys. 520 Res., 115, 1-18, 2010.
  - Bale, N. J., Palatinszky, M., Rijpstra, W. I. C., Herbold, C. W., Wagner, M., and Sinninghe Damsté, J. S.: Membrane lipid composition of the moderately thermophilic ammonia-oxidizing archaeon "*Candidatus* Nitrosotenuis uzonensis" at different growth temperatures. Appl. Environ. Microbiol., 85, e01332-19, 2019.
- Barber, D., Mardsen, R., Minnett, P., Ingram, G., and Fortier, L.: Physical processes within the North Water (NOW) polynya. Atmosphere-Ocean, 39, 163-166, 2001.
- Barnhart, K. R., Miller, C. R., Overeem, I., and Kay, J. E.: Mapping the future expansion of Arctic open water. Nat. Clim. 530 Change, 6, 280-285, 2015.





565

- Bauch, D., van der Loeff, M. R., Andersen, N., Torres-Valdes, S., Bakker, K., and Abrahamsen, E. P.: Origin of freshwater and polynya water in the Arctic Ocean halocline in summer 2007. Prog. Oceanogr., 91, 482-495, 2011.
- Belt, S. T.: Source-specific biomarkers as proxies for Arctic and Antarctic sea ice. Org. Geochem., 125, 277-298, 2018.
  - Belt, S. T., Allard, W. G., Massé, G., Robert, J.-M., and Rowland, S. J.: Highly branched isoprenoids (HBIs): Identification of the most common and abundant sedimentary isomers. Geochim. Cosmochim. Acta, 64, 3839–3851, 2000.
- Belt, S. T., Brown, T. A., Navarro Rodriguez, A., Cabedo-Sanz, P., Tonkin, A., and Ingle, R.: A reproducible method for the extraction, identification and quantification of the Arctic sea ice proxy IP<sub>25</sub> from marine sediments. Anal. Methods, 4, 705-713, 2012.
- Belt, S. T., Brown, T. A., Ringrose, A. E., Cabedo-Sanz, P., Mundy, C. J., Gosselin, M., and Poulin, M.: Quantitative measurement of the sea ice diatom biomarker IP25 and sterols in Arctic sea ice and underlying sediments: further considerations for palaeo sea ice reconstruction. Org. Geochem., 62, 33-45, 2013.
  - Belt, S. T., Brown, T. A., Smik, L., Assmy, P., and Mundy, C. J.: Sterol identification in floating Arctic sea ice algal aggregates and the Antarctic sea ice diatom *Berkeleva adeliensis*. Org. Geochem., 118, 1-3, 2018.
  - Belt, S. T., Brown, T. A., Smik, L., Tatarek, A., Wiktor, J., Stowasser, G., Assmy, P., Allen, C. S., and Husum, K.: Identification of C<sub>25</sub> highly branched isoprenoid (HBI) alkenes in diatoms of the genus *Rhizosolenia* in polar and sub-polar marine phytoplankton. Org. Geochem. 110, 65–72, 2017.
- Belt, S. T., Cabedo-Sanz, P., Smik, L., Rodriguez-Navarro, A., Berben, S. M. P., Knies, J., and Husum, K.: Identification of paleo Arctic winter sea ice limits and the marginal ice zone: Optimised biomarker-based reconstructions of late Quaternary Arctic sea ice. Earth Planet. Sci. Lett., 431, 127-139, 2015.
- Belt, S. T., Massé, G., Rowland, S. J., Poulin, M., Michel, C., and LeBlanc, B.: A novel chemical fossil of palaeo sea ice: IP<sub>25</sub>. Org. Geochem., 38, 16–27, 2007.
  - Belt, S. T., Massé, G., Vare, L. L., Rowland, S. J., Poulin, M., Sicre, M.-A., Sampei, M., and Fortier, L.: Distinctive <sup>13</sup>C isotopic signature distinguishes a novel sea ice biomarker in Arctic sediments and sediment traps. Mar. Chem., 112, 158–167, 2008.
- Belt, S. T., Smik, L., Köseoğlu, D., Knies, J., and Husum, K.: A novel biomarker-based proxy for the spring phytoplankton bloom in Arctic and sub-arctic settings HBI T<sub>25</sub>. Earth and Planet. Sci. Lett., 523, 115703, 2019.
- Besseling, M. A., Hopmans, E. C., Bale, N. J., Schouten, S., Sinninghe Damsté, J. S., and Villanueva, L.: The absence of intact polar lipid-derived GDGTs in marine waters dominated by Marine Group II: Implications for lipid biosynthesis in Archaea. Sci. Rep., 10, 1-10.
  - Bi, H., Zhang, Z., Wang, Y., Xu, X., Liang, Y., Huang, J., Liu, Y., and Fu, M.: 2019. Baffin Bay sea ice inflow and outflow: 1978-1979 to 2016-2017. Cryosphere, 13, 1025-1042, 2019.
- Boon, J. J., Rijpstra, W. I. C., De Lange, F., De Leeuw, J. W., Yoshioka, M., and Shimizu, Y.: Black Sea sterol a molecular fossil for dinoflagellate blooms. Nature, 277, 125-127, 1976.
- Boudinot, F. G., Dildar, N., Leckie, R. M., Parker, A., Jones, M. M., Sageman, B. B., Bralower, T. J., and Sepúlveda, J.: Neritic ecosystem response to Oceanic Anoxic Event 2 in the Cretaceous Western Interior Seaway, USA. Palaeogr. Palaeoclimatol. Palaeoecol., 546, 109673.





- Brown, T. A., Belt, S. T., Philippe, B., Mundy, C. J., Massé, G., Poulin, M., and Gosselin, M.: Temporal and vertical variations of lipid biomarkers during a bottom ice diatom bloom in the Canadian Beaufort Sea: further evidence for the use of the IP<sub>25</sub> biomarker as a proxy for spring Arctic sea ice. Polar Biol., 34, 1857-1868, 2011.
  - Brown, T. A., Belt, S. T., Tatarek, A., and Mundy, C. J.: Source identification of the Arctic sea ice proxy IP<sub>25</sub>. Nat. Comm. 5, 1-7, 2014.
- 590 Cabedo-Sanz, P., Belt, S. T., Knies, J., and Husum, K.: Identification of contrasting seasonal sea ice conditions during the Younger Dryas. Quat. Sci. Rev., 79, 74–86, 2013.
  - Cabedo-Sanz, P., Smik, L., and Belt, S.T.: On the stability of various highly branched isoprenoids (HBI) lipids in stored sediments and sediment extracts: Org. Geochem., 97, 74-77. 2016.
- Cavalieri, D. J., Parkinson, C. L., Gloersen, P., Zwally, H. J.: Sea Ice Concentrations from Nimbus-7 SMMR and DMSP SSM/I-SSMIS Passive Microwave Data, Version 1. updated yearly. Boulder, Colorado USA. NASA National Snow and Ice Data Center Distributed Active Archive Center, 1996.
- 600 Church, M. J., Wai, B., Karl, D. M., and DeLong, E. F.: Abundances of crenarchaeal amoA genes and transcripts in the Pacific Ocean. Environ. Microbiol., 12, 679–688, 2010.
  - Detlef, H., Reilly, B., Jennings, A., Jensen, M. M., O'Regan, M., Glasius, M., Olsen, J., Jakobsson, M., Pearce, C.: Holocene sea-ice dynamics in Petermann Fjord. The Cryosphere, 15, 4357-4380.
- Dunbar, M.: The geophysical position of the North Water. Arctic, 22, 438–441, 1969.
- Elling, F. J., Könneke, M., Lipp, J. S., Becker, K. W., Gagen, E. J., and Hinrichs, K.-U.: Effects of growth phase on the membrane lipid composition of the thaumarchaeon Nitrosopumilus maritimus and their implications for archaeal lipid distributions in the marine environment. Geochim. Cosmochim. Acta, 141, 579–597, 2014.
  - Elling, F. J., Könneke, M., Mußmann, M., Greve, A., and Hinrichs, K.-U.: Influence of temperature, pH, and salinity on membrane lipid composition and TEX<sub>86</sub> of marine planktonic thaumarchaeal isolates. Geochim. Cosmochim. Acta, 171, 238–255, 2015.
- Elling, F. J., Könneke, M., Nicol, G. W., Stieglmeier, M., Bayer, B., Spieck, E., de la Torre, J. R., Becker, K. W., Thomm, M., Prosser, J. I., Herndl, G. J., Schleper, C., and Hinrichs, K.-U.: Chemotaxonomic characterisation of the thaumarchaeal lipidome. Environ. Microbiol., 19, 681–2,700, 2017.
- 620 Fietz, S., Ho, S. L., and Huguet, C.: Archaeal membrane lipid-based paleothermometry for application in polar oceans. Oceanography 33, 2020.
  - Fietz, S., Huguet, C., Rueda, G., Hambach, B., and Rosell-Melé, A.: Hydroxylated isoprenoidal GDGTs in the Nordic Seas. Mar. Chem., 152, 1-10, 2013.
- Francis, C. A., Roberts, K. J., Beman, J. M., Santoro, A. E., and Oakley, B. B.: Ubiquity and diversity of ammonia-oxidizing archaea in water columns and sediments of the ocean. Proc. Natl. Acad. Sci., 102, 14,683–14,688, 2005.
- Garcia, H. E., Weathers, K., Paver, C. R., Smolyar, I., Boyer, T. P., Locarnini, R. A., Zweng, M. M., Mishonov, A. V., Baranova, O. K., Seidov, D., and Reagan, J. R.: World Ocean Atlas 2018, Volume 3: Dissolved Oxygen, Apparent Oxygen Utilization, and Oxygen Saturation. A. Mishonov Technical Ed.; NOAA Atlas NESDIS 83, 38 pp., 2018a.





- Garcia, H. E., Weathers, K., Paver, C. R., Smolyar, I., Boyer, T. P., Locarnini, R. A., Zweng, M. M., Mishonov, A. V., Baranova, O. K., Seidov, D., and Reagan, J. R.: *World Ocean Atlas 2018, Volume 4: Dissolved Inorganic Nutrients (phosphate, nitrate and nitrate+nitrite, silicate).* A. Mishonov Technical Ed.; NOAA Atlas NESDIS 84, 35 pp., 2018b.
  - Georgiadis, E., Giraudeau, J., Jennings, A., Limoges, A., Jackson, R., Ribeiro, S., and Massé, G.: Local and regional controls on Holocene sea ice dynamics and oceanography in Nares Strait, Northwest Greenland. Mar. Geol., 442, 106115, 2020.
- 640 Gould, W. A., Raynolds, M., and Walker, D. A.: Vegetation, plant biomass, and net primary productivity patterns in the Canadian Arctic. J. Geophys. Res. Atmos., 108, 1-14, 2003.
- Harning, D. J., Andrews, J. T., Belt, S. T., Cabedo-Sanz, P., Geirsdóttir, Á, Dildar, N., Miller, G. H., and Sepúlveda, J.: Sea ice control on winter subsurface temperatures of the North Iceland Shelf during the Little Ice Age: A TEX<sub>86</sub> calibration case study. Paleoceanogr. Paleoclimatol., 34, 1006-1021, 2019.
  - Harning, D. J., Jennings, A. E., Köseoğlu, D., Belt, S. T., Geirsdóttir, Á., and Sepúlveda, J.: Response of biological productivity to North Atlantic marine front migration during the Holocene. Clim. Past., 17, 379-396, 2021.
- Hastrup, K., Mosbech, A., and Grønnow, B.: Introducing the North Water: Histories of exploration, ice dynamics, living resources, and human settlement in the Thule Region. Ambio 47, 162-174, 2018.
  - Herfort, L., Schouten, S., Boon, J. P., and Sinninghe Damsté, J. S.: Application of TEX<sub>86</sub> temperature proxy to the southern North Sea. Org. Geochem., 37, 1715–1726, 2006.
- Hopmans, E. C., Schouten, S., and Sinninghe Damsté, J. S.: The effect of improved chromatography on GDGT-based palaeoproxies. Org. Geochem., 93, 1-6, 2016.
- Huang, W. Y., and Meinschein, W. G.: Sterols as source indicators of organic materials in sediments. Geochem. Cosmochim. Acta 40, 323-330, 1976.
  - Huguet, C., Fietz, S., and Rosell-Melé, A.: Global distribution patterns of hydroxy glycerol dialkyl glycerol tetraethers. Org. Geochem. 57, 107-118, 2013.
- 665 Huguet, C., Fietz, S., Rosell-Melé, A., Daura, X., and Costenaro, L.: Molecular dynamics simulation study of the effect of glycerol dialkyl glycerol tetraether hydroxylation on membrane thermostability. Biochim. Biophysi. Acta (BBA) Biomembranes, 1859, 966–974, 2017.
- Huguet, C., Hopmans, E. C., Febo-Ayala, W., Thompson, D. H., Sinninghe Damsté, J. S., and Schouten, S.: An improved method to determine the absolute abundance of glycerol dibiphytanyl glycerol tetraether lipids. Org. Geochem., 37, 1036–1041, 2006.
- Hurley, S. J., Elling, F. J., Könneke, M., Buchwald, C., Wankel, S. D., Santoro, A. E., Lipp, J. S., Hinrichs, K.-U., and Pearson, A.: Influence of ammonia oxidation rate on thaumarchaeal lipid composition and the TEX<sub>86</sub> temperature proxy. Proc. Natl. Acad. Sci., 113, 7762-7767, 2016.
  - Hurley, S. J., Lipp, J. S., Close, H. G., Hinrichs, K.-U., and Pearson, A.: Distribution and export of isoprenoid tetraether lipids in suspended particulate matter from the water column of the Western Atlantic Ocean. Org. Geochem., 116, 90–102, 2018.
- Ingram, R. G., Bacle, J., Barber, D. G., Gratton, Y., and Melling, H.: An overview of physical processes in the North Water. Deep-Sea Res. II, 49, 4893–4906, 2002.



690

710



- Jackson, R., Kvorning, A. B., Limoges, A., Georgiadis, E., Olsen, S. M., Tallberg, P., Andersen, T. J., Mikkelsen, N., Giraudeau, J., Massé, G., Wacker, L., and Ribeiro, S.: Holocene polynya dynamics and their interaction with oceanic heat transport in northernmost Baffin Bay. Sci. Rep., 11, 1-17, 2021.
  - Jia, G., Wang, X., Guo, W., and Dong, L.: Seasonal distribution of archaeal lipids in surface water and its constraint on their sources and the TEX86 temperature proxy in sediments of the South China Sea, J. Geophys. Res.-Biogeo., 122, 592–606, 2017.
- Jones, E. P., Swift, J. H. Anderson, L. G., Lipizer, M., Civitarese, G., Falkner, K. K., Kattner, G., and McLaughlin, F.: Tracing Pacific water in the North Atlantic Ocean. J. Geophys. Res. 108, 3116, 2003.
- Kang, S., Shin, K. H., and Kim, J. H.: Occurrence and distribution of hydroxylated isoprenoid glycerol dialkyl glycerol tetraethers (OH-GDGTs) in the Han River system, South Korea. Acta Geochimica 36, 367-369, 2017.
  - Karner, M. B., DeLong, E. F., and Karl, D. M.: Archaeal dominance in the mesopelagic zone of the Pacific Ocean. Nature, 409, 507–510, 2001.
- Kim, J.-H., van der Meer, J., Schouten, S., Helmke, P., Willmott, V., Sangiorgi, F., Koç, N., Hopmans, E. C., and Sinninghe Damsté, J. S.: New indices and calibrations derived from the distribution of crenarchaeal isoprenoid tetraether lipids: Implications for past sea surface temperature reconstructions. Geochim. Cosmochim. Acta 74, 4639–4654, 2010.
- Kim, J.-H., Crosta, X., Willmott, V., Renssen, H., Bonnin, J., Helmke, P., Schouten, S., and Sinninghe Damsté, J. S.: Holocene subsurface temperature variability in the eastern Antarctic continental margin. Geophys. Res. Lett., 39, 3-8, 2012.
  - Kim, J. H., Schouten, S., Rodrigo-Gámiz, M., Rampen, S., Marino, G., Huguet, C., Helmke, P., Buscail, R., Hopmans, E. C., Pross, J., Sangiorgi, F., Middelburg, J. B. M., and Sinninghe Damsté, J. S.: Influence of deep-water derived isoprenoid tetraether lipids on the TEX<sup>H</sup><sub>86</sub> paleothermometer in the Mediterranean Sea. Geochim. Cosmochim. Ac., 150, 125–141, 2015.
- Kim, J. H., Villanueva, L., Zell, C., and Sinninghe Damsté, J. S.: Biological source and provenance of deep-water derived isoprenoid tetraether lipids along the Portuguese continental margin. Geochim. Cosmochim. Ac., 172, 177–204, 2016.
- Klein, B., LeBlanc, B., Mei, Z.-P., Beret, R., Michaud, J., Mundy, C. J., von Quillfeldt, C. H., Ève Garneau, M., Roy, S., Gratton, Y., Cochran, J. K., Bélanger, S., Larouche, P., Pakulski, J. D., Rivkin, R. B., and Legendre, L.: Phytoplankton biomass, production and potential export in the North Water. Deep Sea Res. II, 49, 4983–5002, 2002.
- Knies, J., Köseoğlu, D., Rise, L., Baeten, N., Bellec, V.K., Bøe, R., Klug, M., Panieri, G., Jernas, P. E., and Belt, S. T.: Nordic Seas polynyas and their role in preconditioning marine productivity during the Last Glacial Maximum. Nat. Comm., 9, 3959,
  2018.
  - Koch, C. W., Cooper, L. W., Lalande, C., Brown, T. A., Frey, K. E., and Grebmeler, J. M.: Seasonal and latitudinal variations in sea ice algae deposition in the Northern Bering and Chukchi Seas determined by algal biomarkers. PLoS ONE, 15, e0231178, 2020.
- Kolling, H. M., Stein, R., Fahl, K., Sadatzki, H., de Vernal, A., and Xiao, X.: Biomarker distributions in (sub)-Arctic surface sediments and their potential for sea-ice reconstructions. Geochem., Geophys., Geosyst., 21, e2019GC008629, 2020.
- Könneke, M., Bernhard, A. E., de la Torre, J. R., Walker, C. B., Waterbury, J. B., and Stahl, D. A.: Isolation of an autotrophic ammonia-oxidizing marine archaeon. Nature, 437, 543-546, 2005.



735



- Köseoğlu, D., Belt, S. T., Smik, L., Yao, H., Panieri, G., and Knies, J.: Complementary biomarker-based methods for characterizing Arctic sea ice conditions: A case study comparison between multivariate analysis and the PIP25 index. Geochim. Cosmochim. Acta, 222, 406–420, 2018.
- Köseoğlu, D., Belt., S. T., and Knies, J.: Abrupt shifts of productivity and sea ice regimes at the western Barents Sea slope from the Last Glacial Maximum to the Bølling-Allerød interstadial. Quat. Sci. Rev., 222, 105903, 2019.
- Kuhlbrodt, T., Griesel, A., Montoya, M., Levermann, A., Hofmann, M., and Rahmstorf, S.: On the driving processes of the Atlantic meridional overturning circulation. Rev. Geophys., 45, RG2001, 2007.
  - Kuypers, M. M. M., Marchant, H. K., and Kartal, B.: The microbial nitrogen-cycling network. Nat. Rev. Microbiol., 16, 263-276, 2018.
- 745 Lewis, L., Ponton, D., Legendre, L., and Leblanc, B.: Springtime sensible heat, nutrients and phytoplankton in the Northwater Polynya, Canadian Arctic. Cont. Shelf Res., 16, 1775-1792, 1996.
  - Limits of Oceans and Seas, 3<sup>rd</sup> edition. International Hydrographic Organization, 1953.
- Limoges, A., Massé, G., Weckström, K., Poulin, M., Ellegaard, M., Heikkilä, M., Geilfus, N.-X., Sejr, M. K., Rysgaard, S., and Ribeiro, S.: Spring succession and vertical export of diatoms and IP<sub>25</sub> in a seasonally ice-covered high arctic fjord. Frontiers in Earth Science, 6, 1-15, 2018.
- Liu, X.-L., Lipp, J. S., Simpson, J. H., Lin, Y.-S., Summons, R. E., and Hinrichs, K.-U.: Mono- and dihydroxyl dibiphytanyl glycerol tetraethers in marine sediments: Identification of both core and intact polar lipid forms. Geochim. Cosmochim. Acta, 89, 102-115, 2012.
- Locarnini, R. A., Mishonov, A. V., Baranova, O. K., Boyer, T. P., Zweng, M. M., Garcia, H. E., Reagan, J. R., Seidov, D., Weathers, K., Paver, C. R., and Smolyar, I.: *World Ocean Atlas 2018, Volume 1: Temperature.* A. Mishonov Technical Ed.; NOAA Atlas NESDIS 81, 52 pp, 2018.
  - Lü, X., Chen, J., Han, T., Yang, H., Wu, W., Ding, W., and Hinrichs, K. U.: Origin of hydroxyl GDGTs and regular isoprenoid GDGTs in suspended particulate matter of Yangtze River Estuary. Org. Geochem., 128, 78–85, 2019.
- Lü, X., Liu, X.-L., Elling F. J., Yang, H., Xie, S., Song, J., Li, X., Yuan, H., Li, N., and Hinrichs, K. U.: Hydroxylated isoprenoid GDGTs in Chinese coastal seas and their potential as a paleo-temperature proxy for mid-to-low latitude marginal seas. Org. Geochem., 89–90, 31–43, 2015.
- Melling, H., Gratton, Y., and Ingram, G.: Ocean circulation within the North Water polynya of Baffin Bay. Atmosphere-770 Ocean, 39, 301–325, 2001.
  - Meredith, M., Sommerkorn, M., Cassotta, S., Derksen, C., Ekaykin, A., Hollowed, A., Kofinas, G., Mackintosh, A., Melbourne-Thomas, J., Muelbert, M. M. C., Ottersen, G., Pritchard, H., and Schuur, E. A. G.: Polar Regions. In: IPCC Special Report on the Ocean and Cryosphere in a Changing Climate [Pörtner, H.-O., Roberts, D. C., Masson-Delmotte, V., Zhai, P.,
- 775 Tignor, M., Poloczanska, E., Mintenbeck, K., Alegría, A., Nicolai, M., Okem, A., Petzold, J., Rama, B., Weyer, N. M. (eds.)], 2019.
  - Meyers, P. A.: Preservation of elemental and isotopic source identification of sedimentary organic matter. Chem. Geol., 114, 289-302, 1994.



800

810



- Mezgec, K., Stenni, B., Crosta, X., Masson-Delmotte, V., Baroni, C., Braida, M., Ciardini, V., Colizza, E., Melis, R., Salvatore, M.C., Severi, M., Scarchilli, C., Traversi, R., Udisti, R., and Frezzotti, M.: Holocene sea ice variability driven by wind and polynya efficiency in the Ross Sea. Nat. Comm., 8, 1-12, 2017.
- 785 Moore, G. W. K., Howell, S. E. L., Brady, M., Xu, X., and McNeil, K.: Anomalous collapses of Nares Strait ice arches leads to enhanced export of Arctic sea ice. Nat. Comm., 12, 1-8, 2021.
- Müller, J., Wagner, A., Fahl, K., Stein, R., Prange, M., and Lohmann, G.: Towards quantitative sea ice reconstructions in the northern North Atlantic: A combined biomarker and numerical modelling approach. Earth Planet. Sci. Lett., 306, 137-148, 2011.
  - Münchow, A., Falkner, K. K., and Melling, H.: Spatial continuity of measured seawater and tracer fluxes through Nares Strait, a dynamically wide channel bordering the Canadian Archipelago. J. Marine Res., 65, 759–788, 2006.
- 795 Münchow, A., Kelly, K. Falkner, K. K., and Melling, H.: Baffin Island and West Greenland Current Systems in northern Baffin Bay. Prog. Oceanogr., 132, 305-337, 2015.
  - Navarro-Rodriguez, A., Belt, S. T., Knies, J., and Brown, T. A.: Mapping recent sea ice conditions in the Barents Sea using the proxy biomarker IP25: implications for palaeo sea ice reconstructions. Quat. Sci. Rev., 79, 26-39, 2013.
- Nichols, P. D., Palmisano, A. C., Rayner, M. S., Smith, G. A., and White, D. C.: Occurrence of novel C30 sterols in Antarctic sea-ice diatom communities during a spring bloom. Org. Geochem., 15, 503-508, 1990.
- Onarheim, I. H., Eldevik, T., Smedsrud, L. H., and Stroeve, J. C.: Seasonal and regional manifestation of Arctic sea ice loss. J. Clim., 31, 4917-4932, 2018.
  - Park, E., Hefter, J., Fischer, G., Iversen, M. H., Ramondenc, S., Nöthig, E.-M., and Mollenhauer, G.: Seasonality of archaeal lipid flux and GDGT-based thermometry in sinking particles of high-latitude oceans: Fram Strait (79°N) and Antarctic Polar Front (50°S). Biogeosciences, 16, 2247-2268, 2019.
  - Pearson, A., and Ingalls, A. E.: Assessing the use of archaeal lipids in marine environmental proxies. Annu. Rev. Earth Planet. Sci., 41, 359-384, 2013.
- Pitcher, A., Hopmans, E. C., Mosier, A.C., Park, S.-J., Rhee, S.-K., Francis, C. A., Schouten, S., and Sinninghe Damsté, J. S.:
  Core and intact polar glycerol dibiphytanyl glycerol tetraether lipids of ammonia-oxidizing archaea enriched from marine and estuarine sediments. Appl. Environ. Microbiol., 3468–3477, 2011.
  - Qin, W., Carlson, L. T., Armbrust, E. V., Devol, A. H., Moffett, J. W., Stahl, D. A., and Ingalls, A. E.: Confounding effects of oxygen and temperature on the TEX86 signature of marine Thaumarchaeota. Proc. Natl. Acad. Sci., 112, 10,979–10,984.
- Rampen, S. W., Abbas, B. A., Schouten, S., and Sinninghe Damste, J. S.: A comprehensive study of sterols in marine diatoms (Bacillariophyta): implications for their use as tracers for diatom productivity. Limnol. Oceanogr., 55, 91-105, 2010.
- Ribeiro, S., Limoges, A., Massé, G., Johansen, K. L., Colgan, W., Weckström, K., Jackson, R., Georgiadis, E., Mikkelsen, N., Kuijpers, A., Olsen, J., Olsen, S. M., Nissen, M., Andersen, T. J., Strunk, A., Wetterich, S., Syväranta, J., Henderson, A. C. G., Mackay, H., Taipale, S., Jeppesen, E., Larsen, N. K., Crosta, X., Giraudeau, J., Wengrat, S., Nuttal, N., Grønnow, B., Mosbech, A., and Davidson, T.A.: Vulnerability of the North Water ecosystem to climate change. Nat. Comm., 12, 1-12, 2021.





- Rodrigo-Gámiz, M., Rampen, S. W., de Haas, H., Baas, M., Schouten, S., and Sinninghe-Damsté, J. S.: Constraints on the applicability of the organic temperature proxies Uk37′, TEX<sub>86</sub> and LDI in the subpolar region around Iceland. Biogeosciences, 12, 6573-6590, 2015.
- Rowland, S. J., Allard, W. G., Belt, S. T., Massé, G., Robert, J.-M., Blackburn, S., Frampton, D., Revill, A. T., and Volkman, J. K.: Factors influencing the distributions of polyunsaturated terpenoids in the diatom, Rhizosolenia setigera. Phytochemistry, 58, 717–728, 2001.
  - Rueda, G., Rosell-Melé, A., Escala, M., Gyllencreutz, R., and Backman, J.: Comparison of instrumental and GDGT-based estimates of sea surface and air temperatures from the Skagerrak. Org. Geochem., 40, 287–291, 2009.
- Ryan, P. A., and Münchow, A.: Sea ice draft observations in Nares Strait from 2003 to 2012. J. Geophys. Res.: Oceans, 122, 3057-3080, 2017.
  - Schlitzer, R.: Ocean Data View, odv.awi.de, 2020.
- Schouten, S., Hopmans, E. C., Schefuss, E., and Sinninghe Damsté, J. S.: Distributional variations in marine crenarchaeotal membrane lipids: a new tool for reconstructing ancient sea water temperatures? Earth Planet. Sci. Lett., 204, 265-274, 2002.
  - Schouten, S., Hopmans, E. C., and Sinninghe Damsté, J. S.: The organic geochemistry of glycerol dialkyl glycerol tetraether lipids: A review. Org. Geochem., 54, 19-61, 2013.
- Schlederman, P.: Polynyas and prehistoric settlement patterns. Arctic 33, 292–302, 1980.
- Sinninghe Damsté, J. S., Rijpatra, W. I. C., Hopmans, E. C., Jung, M.-Y., Kim, J.-G., Rhee, S.-K., Stieglmeier, M., and Schleper, C.: Intact polar and core glycerol dibiphytanyl glycerol tetraether lipids of Group 1.1a and 1.1b Thaumarchaeota in soil. Appl. Environ. Microbiol., 78, 6866–6874, 2012.
  - Smik, L., Cabedo-Sanz, P., and Belt, S. T.: Semi-quantitative estimates of paleo Arctic sea ice concentration based on source-specific highly branched isoprenoid alkenes: A further development of the PIP25 index. Org. Geochem., 92, 63-69, 2016.
- 860 Smik, L., and Belt, S. T.: Distributions of the Arctic sea ice biomarker proxy IP<sub>25</sub> and two phytoplanktonic biomarkers in surface sediments from West Svalbard. Org. Geochem., 105, 39-41, 2017.
- Smith, J. A., Hillenbrand, C. D., Pudsey, C. J., Allen, C. S., and Graham, A. G. C.: The presence of polynyas in the Weddell Sea during the Last Glacial Period with implications for the reconstruction of sea-ice limits and ice sheet history. Earth Planet. Sci. Lett., 296, 287–298, 2010.
  - Stirling, I.: The biological importance of polynyas in the Canadian Arctic. Arctic 33, 303-315, 1980.
- Stoynova, V., Shanahan, T. M., Hughen, K. A., and de Vernal, A.: Insights into Circum-Arctic sea ice variability from molecular geochemistry. Quat. Sci. Rev., 79, 63-73, 2013.
  - Syring, N., Stein, R., Fahl, K., Vahlenkamp, M., Zehnich, M., Spielhagen, R. F., and Niessen, F.: Holocene changes in sea-ice cover and polynya formation along the eastern North Greenland shelf: New insights from biomarker records. Quat. Sci. Rev., 231, 106173, 2020.
- Tang, C. C. L., Ross, C. K., Yao, T., Petrie, B., DeTracey, B. M., and Dunlap, E.: The circulation, water masses and sea-ice of Baffin Bay. Prog. Oceanogr., 63, 183-228, 2004.



890

905



- Tierney, J. E., and Tingley, M. P.: A Bayesian, spatially-varying calibration model for the TEX<sub>86</sub> proxy. Geochim. Cosmochim. Acta, 127, 83-106, 2014.
  - Tremblay, J.-E., Gratton, Y., Carmack, E. C., Payne, C. D., and Price, N. M.: Impact of the large-scale Arctic circulation and the North Water Polynya on nutrient inventories in Baffin Bay. J. Geophys. Res. 107, 261-26-14, 2002b.
- Tremblay, J.-E., Gratton, Y., Fauchot, J., and Price, N. M.: Climatic and oceanic forcing of new, net, and diatom production in the North Water. Deep-Sea Res. II, 49, 4927-4946, 2002a.
  - Xiao, X., Fahl, K., Müller, J., and Stein, R.: Sea-ice distribution in the modern Arctic Ocean: Biomarker records from trans-Arctic Ocean surface sediments. Geochim. Cosmochim. Acta, 155, 16–29, 2015.
  - Villanueva, L., Schouten, S., and Sinninghe Damsté, J. S.: Depth- related distribution of a key gene of the tetraether lipid biosynthetic pathway in marine Thaumarchaeota. Environ. Microbiol., 17, 3527–3539, 2015.
  - Volkman, J. K.: A review of sterol markers for marine and terrigenous organic matter. Org. Geochem., 9, 83-99, 1986.
- Volkman, J. K., Barrett, S. M., Dunstan, G. A., and Jeffrey, S. W.: Geochemical significance of the occurrence of dinosterol and other 4-methyl sterols in a marine diatom. Org. Geochem., 20, 7-15, 1993.
- Wei, B., Jia, G., Hefter, J., Kang, M., Park, E., and Mollenhauer, G.: Comparison of the U<sub>37</sub><sup>K'</sup>, LDI, TEX<sub>86</sub><sup>H</sup> and RI-OH temperature proxies in the northern shelf of the South China Sea. Biogeosciences, 17, 4489-4508, 2020.
  - Wuchter, C., Schouten, S., Coolen, M. J. L., and Sinninghe Damsté, J. S.: Temperature-dependent variation in the distribution of tetraether membrane lipids of marine Crenarchaeota: Implications for TEX86 paleothermometry. Paleoceanography, 19, PA4028, 2004.
- Wuchter, C., Schouten, S., Wakeham, S. G., and Sinninghe Damsté, J. S.: Temporal and spatial variation in tetraether membrane lipids of marine Crenarchaeota in particulate organic matter: Implications for TEX<sub>86</sub> paleothermometry. Paleoceanography, 20, PA3013, 2005.
- Plang, Y. G., Pagani, M., Wang, Z.: Ring index: A new strategy to evaluate the integrity of TEX<sub>86</sub> paleothermometry. Paleoceanography, 32, 220-232, 2016.
  - Zhu, C., Wakeham, S. G., Elling, F. J., Basse, A., Mollenhauer, G., Versteegh, G. J. M., Könneke, M., and Hinrichs, K. U.: Stratification of archaeal membrane lipids in the ocean and implications for adaptation and chemotaxonomy of planktonic archaea. Environ. Microbiol., 18, 4324–4336, 2016.
  - Zweng, M. M., Reagan, J. R., Seidov, D., Boyer, T. P., Locarnini, R. A., Garcia, H. E., Mishonov, A. V., Baranova, O. K., Weathers, K., Paver, C. R., and Smolyar, I.: *World Ocean Atlas 2018, Volume 2: Salinity.* A. Mishonov Technical Ed.; NOAA Atlas NESDIS 82, 50 pp, 2018.





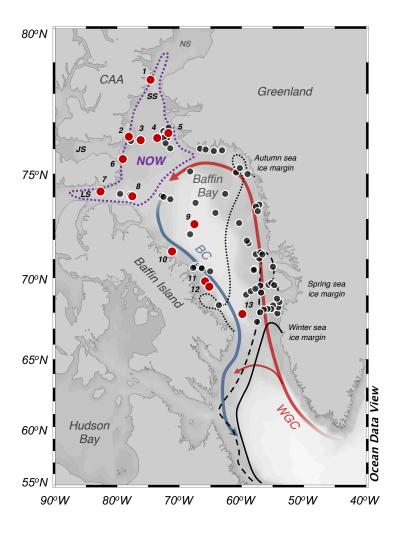


Figure 1: Overview map of Baffin Bay. Simplified ocean surface currents shown in bold lines where red reflects warm, Atlantic Water (West Greenland Current, WGC) and blue reflects cool, Arctic Water (Baffin Current, BC). To the north is the June limit of the NOW (purple dotted line). Seasonal sea ice limits shown with black dotted (autumn), dashed (spring) and solid lines (winter) (Cavalieri et al., 1996). The numbered locations of this study's modern sites are shown with red circles, and those of Kolling et al. (2020) in black. CAA = Canadian Arctic Archipelago, NS = Nares Strait, SS = Smith Sound, JS = Jones Sound, and LS = Lancaster Sound. See Table 1 for further sample site information. Ocean Data View base map after Schlitzer (2019).





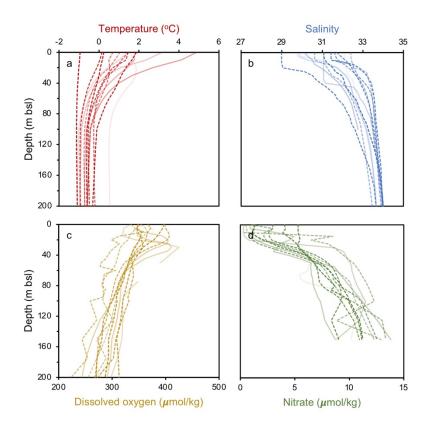


Figure 2: WOA18 Annual 2007-2017 oceanographic variables from Baffin Bay against depth (m bsl). Individual profiles are from each study site, where darker (lighter) colors reflect sites farther north (south) and dashed (solid) lines denote those within (outside) the modern limits of the NOW. Data from Garcia et al. (2018a, 2018b), Locarnini et al. (2018), and Zweng et al. (2018).

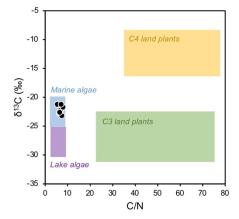


Figure 3: Organic matter provenance based on bulk C/N and  $\delta^{13}$ C values. Reference values for four endmembers after Meyers (1994).





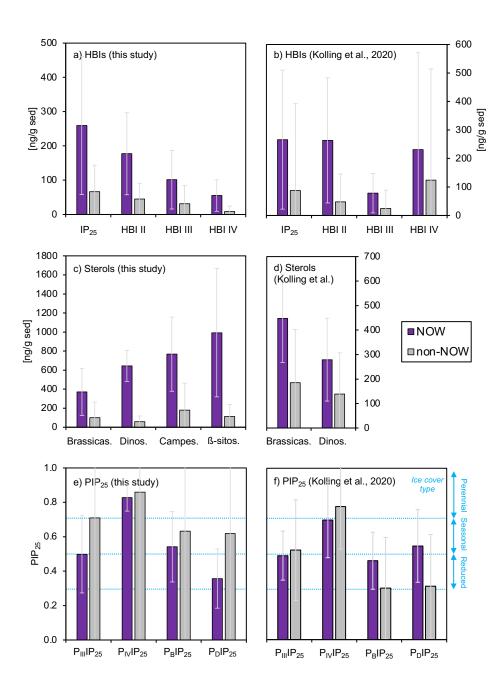


Fig. 4: Average concentrations of a-b) HBIs, c-d) sterols, and e-f) PIP<sub>25</sub> indices for sample sets from this study and Kolling et al. (2020). Standard deviations for each shown in light gray. Sites within the NOW are colored purple whereas sites outside the NOW are gray. Qualitative sea ice concentration limits in panel e-f after Müller et al. (2011) for no sea ice (0 to 0.3), reduced sea ice (0.3 to 0.5), seasonal sea ice (0.5 to 0.7), and perennial sea ice (0.7 to 1).





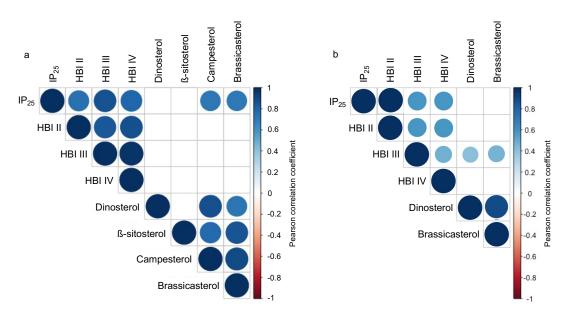


Fig. 5: Pearson correlation coefficients between HBIs and sterols for a) this study's samples and b) Kolling et al. (2020). Positive correlations are displayed in blue and negative correlations in red. Both color and the size of the circle are proportional to the correlation coefficients. Insignificant correlations (*p*-values >0.01) are left blank.



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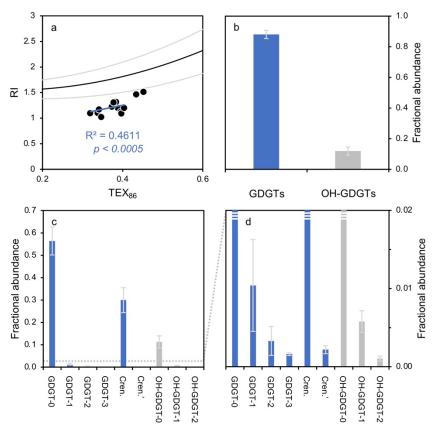


Figure 6: GDGT- and OH-GDGT distributions and fractional abundances. a) TEX<sub>86</sub> versus RI showing the global polynomial equation and 95% uncertainty envelope (black and gray lines, Zhang et al., 2016) and Baffin Bay sediments (black dots), b) fractional abundance of total GDGTs (blue) and OH-GDGTs (gray), c) and d) fractional abundance of individual GDGTs (blue) and OH-GDGTs (gray) at two different scales.



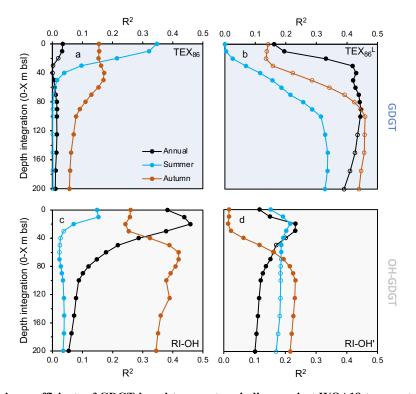


Figure 7: Regression coefficients of GDGT-based temperature indices against WOA18 temperature at various depth integrations and seasons. a)  $TEX_{86}$ , b)  $TEX_{86}^{L}$ , c) RI-OH, and d) RI-OH'. WOA18 data from Locarnini et al. (2018). Data points not filled indicate that p values are > 0.05.





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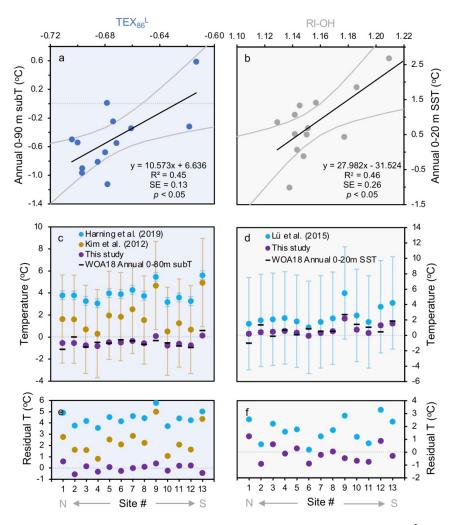


Figure 8: GDGT- and OH-GDGT-temperature calibrations. Shown are a) TEX<sub>86</sub><sup>L</sup> vs annual 0-90 m subT, b) RI-OH (OH-GDGTs) vs annual 0-20 m SST, c) TEX<sub>86</sub><sup>L</sup> subT estimates based on calibrations from Iceland (blue, Harning et al., 2019), global (yellow, Kim et al., 2012), and Baffin Bay (purple, this study) against WOA18 annual 0-90m subT (black dashes, Locarnini et al., 2018), d) RI-OH SST estimates based on calibrations from global (blue, Lü et al., 2015) and Baffin Bay (purple, this study) against WOA18 annual 0-20m SST (black dashes, Locarnini et al., 2018). Panels e and f show residuals of calibration estimates and WOA18 data for TEX<sub>86</sub><sup>L</sup> and RI-OH, respectively. The x-axis refers to site # as shown in Fig. 1 and Table 1.





Table 1: Marine surface sediment site information for this study.

Site #	Core Site Name	Lat	Long	Water depth (m bsl)
1	AMD17-129-BC	78.42	-74.24	521
2	AMD17-101-BC	76.48	-77.77	378
3	AMD17-108-BC	76.47	-74.70	449
4	AMD17-111-BC	76.44	-73.32	593
5	AMD17-115-BC	76.57	-71.33	668
6	HU2008 029-040 BX	75.58	-78.63	580
7	HU2008 029-59 TC	74.26	-82.23	791
8	HU2008 029-49 BX	74.03	-77.13	868
9	AMD17-BB2-BC	72.77	-67.25	2373
10	AMD15-CASQ1-BC4	71.41	-70.89	702
11	AMD17-176-BC	69.82	-65.46	267
12	AMD17-Site 8.1-BC	69.52	-64.97	1054
13	AMD17-Disko Fan-BC	67.99	-59.60	1012