

1 **Rapidly increasing sulfate concentration: a hidden promoter of eutrophication in**
2 **shallow lakes**

3 Chuanqiao Zhou^{a,1}, Yu Peng^{a,1}, Li Chen^a, Miaotong Yu^a, Muchun Zhou^b, Runze Xu^a,
4 Lanqing Zhang^a, Siyuan Zhang^c, Xiaoguang Xu^{a,*}, Limin Zhang^a, Guoxiang Wang^a

5 ^a School of Environment, Nanjing Normal University, Jiangsu Center for Collaborative
6 Innovation in Geographical Information Resource Development and Application,
7 Jiangsu Key Laboratory of Environmental Change and Ecological Construction,
8 Nanjing 210023, China

9 ^b China Aerospace Science and Industry Nanjing Chenguang group, Nanjing 210022,
10 China

11 ^c School of Energy and Environment, Southeast University, Nanjing 210096, China

12 **Corresponding author: 1, Wenyuan Road, Xianlin University District, Nanjing,*
13 *210023, China*

14 *E-mail address: xxg05504118@163.com*

15 ¹ Both authors contributed equally

16 **Keywords:** Sulfate reduction; iron reduction; phosphorus release; eutrophication;
17 sulfate reduction bacteria

18 **Abstract:**

19 Except for excessive nutrient input and climate warming, the rapidly rising SO₄²⁻
20 concentration is considered as a crucial contributor to the eutrophication in shallow
21 lakes, however, the driving process and mechanism are still far from clear. In this study,
22 we constructed a series of microcosms with initial SO₄²⁻ concentrations of 0, 30, 60, 90,

23 120 and 150 mg/L to simulate the rapidly SO_4^{2-} increase of Lake Taihu subjected to
24 cyanobacteria blooms. Results showed that the sulfate reduction rate was stimulated by
25 the increase of initial SO_4^{2-} concentrations and cyanobacteria-derived organic matter,
26 with the maximal sulfate reduction rate of 39.68 mg/L·d in the treatment of 150 mg/L
27 SO_4^{2-} concentration. During the sulfate reduction, the produced maximal ΣS^{2-}
28 concentration in the overlying water and acid volatile sulfate (AVS) in the sediments
29 were 3.15 mg/L and 11.11 mg/kg, respectively, and both of them were positively
30 correlated with initial SO_4^{2-} concentrations ($R^2=0.97$; $R^2=0.92$). The increasing
31 abundance of sulfate reduction bacteria (SRB) was also linearly correlated with initial
32 SO_4^{2-} concentrations ($R^2=0.96$), ranging from 6.65×10^7 to 1.97×10^8 copies/g. However,
33 the Fe^{2+} concentrations displayed a negative correlation with initial SO_4^{2-}
34 concentrations, and the final Fe^{2+} concentrations were 9.68, 7.07, 6.5, 5.57, 4.42 and
35 3.46 mg/L, respectively. As a result, the released TP in the overlying water, to promote
36 the eutrophication, was up to 1.4 mg/L in the treatment of 150 mg/L SO_4^{2-} concentration.
37 Therefore, it is necessary to consider the effect of rapidly increasing SO_4^{2-}
38 concentrations on the release of endogenous phosphorus and the eutrophication in lakes.

39 **1.Introduction**

40 Nowadays, cyanobacteria bloom in eutrophic lakes has become one of the most
41 serious problems in freshwater lakes all over the world (Iwayama et al., 2017; Ho et al.,
42 2019). Phosphorus, as a necessary nutrient for biological growth, is considered to be
43 one of the main limiting factors of lake eutrophication (Ni et al., 2020). In recent years,
44 the input of exogenous phosphorus has been effectively controlled, while the release of

45 endogenous phosphorus is still an urgent problem in eutrophic lakes (Liu et al., 2018;
46 Guo et al., 2020). The release of endogenous phosphorus is affected by many factors,
47 such as wind and wave and the cyanobacteria decomposition (Xu et al., 2018; Zhao et
48 al., 2019). There are many forms of phosphorus in freshwater lake sediments, including
49 aluminum bound phosphorus (Al-P), iron bound phosphorus (Fe-P), etc. Among them,
50 Fe-P, formed under the condition of high dissolved oxygen (DO), is the most active
51 form of phosphorus in the sediments, which has a more obvious response to the change
52 of DO (Zhang et al., 2020). The accumulation and decay of cyanobacteria in eutrophic
53 lakes will change the physical and chemical environments of water body and form
54 anaerobic reduction conditions (Yan et al., 2017). This will facilitate the reduction of
55 iron oxides and lead to the desorption and release of Fe-P in sediments, resulting in the
56 increase of endogenous phosphorus release (Zhao et al., 2019).

57 Iron reduction plays an important role in natural ecosystems. It has been reported
58 that dissimilatory reduction of iron accounts for 22% of the total amount of organic
59 matter anaerobic mineralization in offshore areas (Thamdrup et al., 2004). According
60 to the classical theory, iron oxides or hydroxides can adsorb phosphorus in the water
61 and form Fe-P precipitation (Gunnars et al., 1997). In freshwater lakes, the lack of Fe(III)
62 content or the diagenesis of organic phosphorus may be the reason for the lack of
63 phosphorus in the overlying water. Therefore, the formation of iron oxides on the
64 surface of sediments is closely related to the phosphorus cycle process (Amirbahman
65 et al., 2003; Chen et al., 2014). The interaction between iron and phosphorus is reflected
66 in the effect of adsorption and desorption of Fe oxide on the P content in the overlying

67 water, since Fe-P is the main internal source of phosphorus (Wu et al., 2019). Iron
68 oxides can be used as both the source and destination of phosphorus in lake ecosystems
69 (Mort et al., 2010; Azam et al., 2014). In anaerobic reduction environments, iron
70 reduction can significantly promote the resolution of Fe-P. The Fe^{2+} generated by the
71 reaction can form FeS solid with soluble sulfide. In addition, free Fe^{3+} will combine
72 with humus to form stable complex, which further prevents the co-precipitation process
73 of phosphorus and iron oxides (Mort et al., 2010; Zhang et al., 2020). Therefore, iron
74 reduction process driven by cyanobacteria decomposition affects the circulation of
75 phosphorus in freshwater lakes.

76 Due to the SO_4^{2-} concentration in seawater reaching 28 mM, sulfate reduction
77 process with the participation of sulfate reduction bacteria (SRB) has received
78 considerable attention in the basic material cycle of marine biogeochemistry (Fike et
79 al., 2015; Pan et al., 2020). In freshwater lakes, the SO_4^{2-} concentration is less than 800
80 μM , which is generally considered insufficient for continuous sulfate reduction (Hansel
81 et al., 2015). However, in recent years, with the continuous input of exogenous sulfur,
82 the SO_4^{2-} concentration in freshwater lakes increases significantly and the degree of the
83 eutrophication and the SO_4^{2-} concentration show a positive correlation (Dierberg et al.,
84 2011; Yu et al., 2013). For instance, the SO_4^{2-} concentration in Lake Taihu, one of the
85 typical eutrophic lakes worldwide, has increased from 30 to 100 mg/L in the past 70
86 years and it will continue to rise in the future (Yu et al., 2013; Zhou et al., 2022). The
87 impact of sulfate reduction on the material cycle of lake ecosystems may be far beyond
88 our knowledge (Baldwin et al., 2012; Yu et al., 2013). On the other hand, it has been

89 reported that sulfate reduction process is one of the important ways of anaerobic
90 metabolism of organic matter in freshwater lakes, and $\sum S^{2-}$ produced by sulfate
91 reduction process can mediate the iron reduction process (Jorgensen et al., 2019; Zhang
92 et al., 2020). SRB mainly uses SO_4^{2-} as the electron acceptor to complete anaerobic
93 respiration, and the sulfur compounds produced by anaerobic metabolism are bound
94 with iron and so on, which are fixed in the sediments and form AVS on the surface of
95 sediments (Holmer et al., 2001; Chen et al., 2016). Therefore, with the input of
96 exogenous sulfur, sulfate reduction process produced $\sum S^{2-}$ will further promote iron
97 reduction in freshwater lakes.

98 In freshwater lakes, iron cycle affects the process of phosphorus cycle, and sulfur
99 cycle plays an important role in regulating iron cycle. Therefore, the cycle of iron, sulfur
100 and phosphorus in freshwater lakes is inseparable (Wu et al., 2019; Zhao et al., 2019).
101 Studies have shown that even when SO_4^{2-} content was as low as 20 mg/L, the anaerobic
102 metabolism of organic substrates was still dominated by sulfate reduction. Therefore,
103 sulfate reduction process plays an important role in the lacustrine biochemical cycle
104 (Hansel et al., 2015). In the absence of cyanobacteria, sulfate reduction doesn't occur
105 even if the SO_4^{2-} concentration is higher (Zhao et al., 2021). This is because the
106 accumulation and decomposition of cyanobacteria not only change the environment of
107 water body, but also release a large amount of organic matter, which provides the
108 necessary conditions for the circulation of iron, sulfur and phosphorus (Yan et al., 2017;
109 Melemdez-Pastor et al., 2019). Therefore, under the co-effect of the increase of SO_4^{2-}
110 and the cyanobacteria decomposition, the sulfate reduction process and the effect of

111 iron reduction process on endogenous phosphorus release from sediments need to be
112 further studied.

113 In this study, a series of different initial concentrations of SO_4^{2-} were set according
114 to the variation trend of SO_4^{2-} concentrations over the years and the possible rising trend
115 of eutrophic Lake Taihu. The effects of increased SO_4^{2-} concentration and cyanobacteria
116 bloom on sulfate reduction coupled with the microbial processes were investigated. The
117 dynamic changes of Fe^{2+} and Fe^{3+} concentrations during iron reduction were studied in
118 order to reveal the effect of sulfate reduction on iron reduction. In addition, the dynamic
119 changes of phosphorus in the overlying water and sediment were investigated. Finally,
120 the coupled sulfate, iron and phosphorus cyclic processes affected by the increasing
121 sulfate concentration and cyanobacteria bloom were also comprehensively analyzed for
122 elucidating the phosphorus release dynamics to tracking the hidden promoter of
123 cyanobacteria bloom in eutrophic lakes. The findings may be benefit for evaluating the
124 effect of sulfate reduction in freshwater lakes and its impact on the promotion of iron
125 reduction and the release of endogenous phosphorus.

126 **2. Materials and methods**

127 *2.1 Sample collection and preparation*

128 Lake Taihu (31°24' 40" N, 120°1' 3" E), one of the largest eutrophic shallow lakes
129 in China, with an average depth of 2.4 m and an area of 2340 m² (Mao et al., 2021).
130 Samples of sediments and cyanobacteria were collected in July 2020. Sediments from
131 the west shoreline of the lake (31°24'45"N, 120°0'42"E) were collected using a gravity
132 core sampler. Cyanobacteria was collected and concentrated by sieving water through

133 a fine-mesh plankton (250 mesh). All the sediment and cyanobacteria samples were
134 stored in an incubator with ice packs and delivered to the laboratory immediately. The
135 sediment samples were blended thoroughly, homogenized, and sieved (100 mesh) to
136 the polyethylene bag. The cyanobacteria samples were flushed and centrifuged at 1500
137 r/min for 5 min by a CT15RT versatile refrigerated centrifuge (China) and freeze-dried
138 by Biosafer-10A. Different gradient sulfate concentrations were prepared from
139 the high purity water and Na₂SO₄.

140 *2.2 Set-up of incubation microcosms*

141 To simulate the dramatical SO₄²⁻ increase and cyanobacteria blooms of eutrophic
142 Lake Taihu, a series of microcosms were constructed in this study. According to the
143 ratio of surface sediments and the average water depth and the cyanobacteria
144 accumulation density of 2500 g/m² during the breakout of cyanobacteria blooms of
145 Taihu Lake, 100 g of sediment, 200 ml of water and 0.11 g of cyanobacteria powder
146 were added into each bottle (Zhang et al., 2020). Meanwhile, according to the change
147 trend of SO₄²⁻ concentrations in Taihu Lake over the years and the possibility of further
148 increase in the future (Yu et al., 2013), the SO₄²⁻ concentrations in six microcosm
149 systems were configured as: 30, 60, 90, 120, 150 mg/L, and a control without SO₄²⁻,
150 respectively. The microcosm system adopted anaerobic bottles (Φ75 mm, length 180
151 mm, volume 500 ml) as the reaction device. There were three replicates in each SO₄²⁻
152 concentration experimental group. Each group was sampled 17 times on 1, 2, 3, 4, 5, 6,
153 7, 9, 11, 14, 18, 23, 28, 33, 38, 43 and 48 d. Totally, there were 306 anaerobic bottles,
154 and all the anaerobic bottles were placed in a biochemical incubator at a temperature of

155 25 °C. The water, gas and soil samples were collected by destructive sampling, that is,
156 at each sampling point, 18 anaerobic bottles were opened for testing, which ensured the
157 anaerobic environment and air pressure for other bottles. A part of sediment was used
158 for microbe determination and kept in a refrigerator at -80 °C, and the rest sediment and
159 other samples were kept at 0-4 °C for less than 24 h before analysis.

160 *2.3 Chemical analytical methods*

161 All water column and pore-water samples were filtered through 0.45 µm Nylon
162 filters prior. Dissolved total phosphorus (DTP) was determined by colorimetry after
163 digestion with $K_2S_2O_8+NaOH$, and the ammonium molybdate and ascorbic acid were
164 used as chromogenic agents (Ebina et al., 1983). Water DO, oxidation and reduction
165 potential (ORP) were measured using calibrated probes (MP525, China) during
166 destructive sampling. The SO_4^{2-} was detected using the turbidimetric method with the
167 stabilizer of $BaCl_2$ and gelatin (Tabatabai et al., 1974), and the ΣS^{2-} was detected by
168 methylene blue (Cline et al., 1969). Acid volatile sulfate (AVS), the ΣS^{2-} combined
169 with metal ions formed compounds in sediments, was determined by zinc cold diffusion
170 method (Hsieh et al., 1997). Fe^{2+} and Fe^{3+} was determined by colorimetric (Phillips
171 et al., 1987). The sediment total phosphorus (TP) was extracted and determined by
172 colorimetry (Ruban et al., 2001).

173 *2.4 Quantification of SRB in sediments*

174 In order to confirm the changes of sediment SRB in the microcosms, RT-QPCR
175 technologies were used to determine the cell copy numbers of MPA and SRB on 0,7
176 and 38 d in the sediments.

177 The sediment samples were collected and frozen at -80 °C in an ultra-low
178 temperature freezer. The E.Z.N.A. ®Soil DNA Kit (Omega Bio-Tek, Norcross, GA,
179 USA) was used to extract the total genomic DNA from each soil sample according to
180 the manufacturer's instructions. Nucleic acid quality and concentration were
181 determined by 1% agarose gel electrophoresis and NanoDrop 2000 UV
182 spectrophotometer (Thermo Scientific, USA), respectively.

183 SRB in sediments were quantified using the quantitative polymerase chain
184 reaction (qPCR) method. The qPCR with primer sets targeting DSR1F+ (5'-
185 ACSCACTGGAAGCACGGCGG-3') and DSR-R (5'-GTGGMRCCGTGCAKRTT
186 GG-3') were used for the SRB in this study. The q-PCR experiments were performed
187 on a ABI7300 q-PCR instrument (Applied Biosystems, USA) using ChamQ SYBR
188 Color qPCR Master Mix as the signal dye. Each 20 µL reaction mixture contained 2 µL
189 of the template DNA and 16.5 µL of ChamQ SYBR Color qPCR Master Mix. Standard
190 curves for each gene were obtained by the tenfold serial dilution of standard plasmids
191 containing the target functional gene. All operations were followed the MIQE
192 guidelines.

193 *2.5 Statistical analysis*

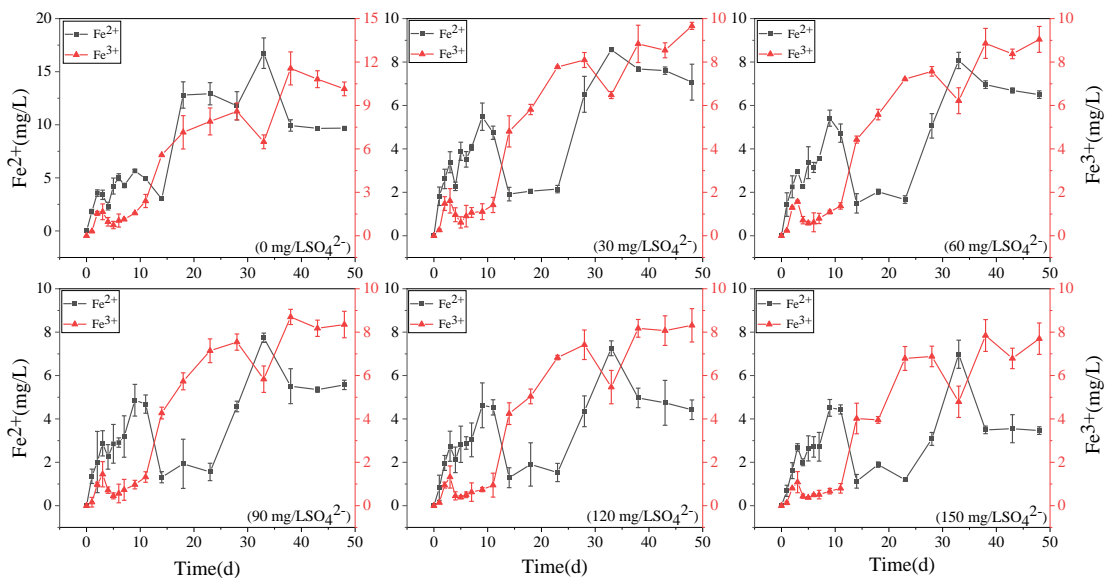
194 The Statistical Package of the Social Science 18.0 (SPSS 18.0) was used for
195 statistical analysis. The one-way analysis of variance (ANOVA) and correlation
196 analysis was carried out using bivariate correlations analysis.

197

198 **3.Results**

199 3.1 Fe^{2+} and Fe^{3+} dynamics in overlying water

200 The concentration variations of Fe^{2+} and Fe^{3+} in overlying water during the
201 incubation was presented in Fig.1. In the treatment without SO_4^{2-} , they increased
202 continuously to 9.68 mg/L and 10.15 mg/L, respectively. The concentration of Fe^{3+} in
203 the remaining five treatments decreased at the beginning and then increased to keep
204 stable. The higher the initial sulfate concentration was, the lower the final Fe^{3+}
205 concentration displayed. In the initial 150 mg/L SO_4^{2-} concentration treatment, the final
206 Fe^{3+} concentration was the lowest of 7.7 mg/L. The Fe^{2+} concentration in the five
207 treatments supplemented with SO_4^{2-} decreased significantly from 11 d to 23 d, and then
208 increased to a stable level. The final concentration of Fe^{2+} also showed a negative
209 correlation with the initial concentration of SO_4^{2-} . In the initial 30 mg/L SO_4^{2-}
210 concentration treatment, the final Fe^{2+} concentration was the highest of 7.07 mg/L.



211
212 Figure 1. The concentration variations of Fe^{2+} and Fe^{3+} in the water column during the
213 incubation

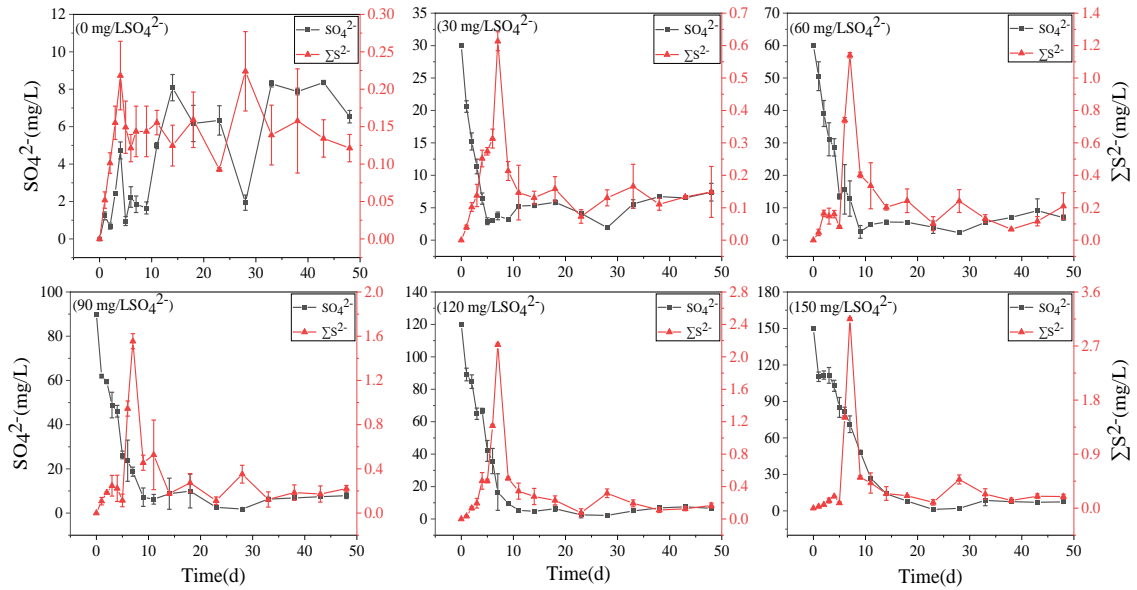
214 3.2 SO_4^{2-} and ΣS^{2-} dynamics in overlying water

215 All treatments had obvious sulfate reduction reaction, and the concentration of
 216 SO_4^{2-} decreased greatly except for the treatment without adding SO_4^{2-} (Fig.2). The
 217 higher the initial sulfate concentration was, the faster the sulfate reduction rate in the
 218 initial stage exhibited (Tab.1). In the treatment with initial SO_4^{2-} concentration of 150
 219 mg/L, the sulphate reduction rate was 39.68 mg/L·d, while it was only 9.39 mg/L·d in
 220 the 30 mg/L SO_4^{2-} treatment. The sulfate reduction rate at the beginning of other
 221 treatments was also positively correlated with the initial SO_4^{2-} concentration.

222 The higher the initial SO_4^{2-} concentration was, the higher the maximum
 223 concentration of $\sum\text{S}^{2-}$ was. In the treatment with initial SO_4^{2-} concentration of 30 mg/L,
 224 the lowest concentration was 2.93 mg/L on the 5th day. However, the lowest SO_4^{2-}
 225 concentration appeared on the 23rd day was 1.18 mg/L in the treatment with initial
 226 SO_4^{2-} concentration of 150 mg/L. The maximum concentration of $\sum\text{S}^{2-}$ was positively
 227 correlated with the initial SO_4^{2-} concentration. In the initial SO_4^{2-} concentrations of 30,
 228 60, 90, 120 and 150 mg/L SO_4^{2-} treatments, the highest $\sum\text{S}^{2-}$ concentrations at 7 d were
 229 0.14, 0.61, 1.14, 1.55, 2.15, and 3.15 mg/L, respectively.

230 Table 1. Sulphate reduction rate in the water column of microcosms (mg/L·d)

SO_4^{2-} (mg/L) \ Time(d)	0	7	38
0	-	-	-
30	9.39	0.74	0.05
60	9.44	2.84	0.07
90	28.02	4.98	0.11
120	30.89	19.45	0.11
150	39.68	10.42	0.21



231

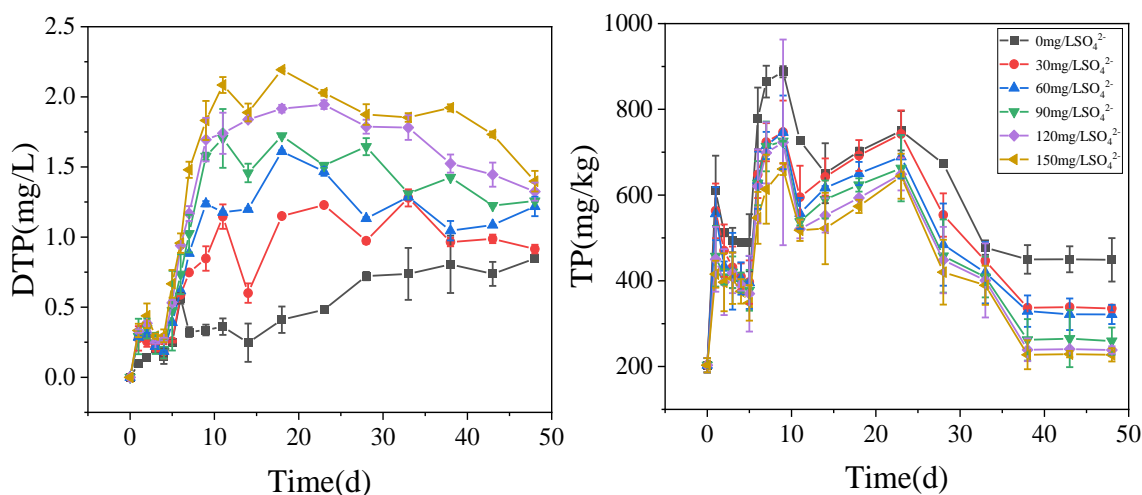
232 Figure 2. The concentration variations of SO_4^{2-} and ΣS^{2-} in the water column during
 233 the incubation

234 3.3 TP dynamics in overlying water and sediments

235 The dynamics of DTP concentrations in overlying water during the incubation was
 236 presented (Fig.3 left). The concentrations of DTP in overlying water were positively
 237 correlated with the initial SO_4^{2-} . The higher the initial concentrations of SO_4^{2-} were, the
 238 higher the concentrations of DTP in overlying water were. On 11 day, DTP in overlying
 239 water continued to rise and then kept stable. The highest DTP concentration was 2.08
 240 mg/L in the treatment with initial SO_4^{2-} concentration of 150 mg/L, while the highest
 241 DTP concentration was 0.36 mg/L in the treatment without SO_4^{2-} addition.

242 The concentrations of TP in the sediments increased significantly in all treatments
 243 with the cyanobacteria decomposition in the initial stage (Fig.3 right). Among of all
 244 treatments, on 9th day, the highest concentration of TP in the sediments was 887.69
 245 mg/kg in the treatment with initial SO_4^{2-} concentration of 0 mg/L. After 23 days, TP in
 246 the sediments decreased significantly and then stabilized. During cyanobacteria

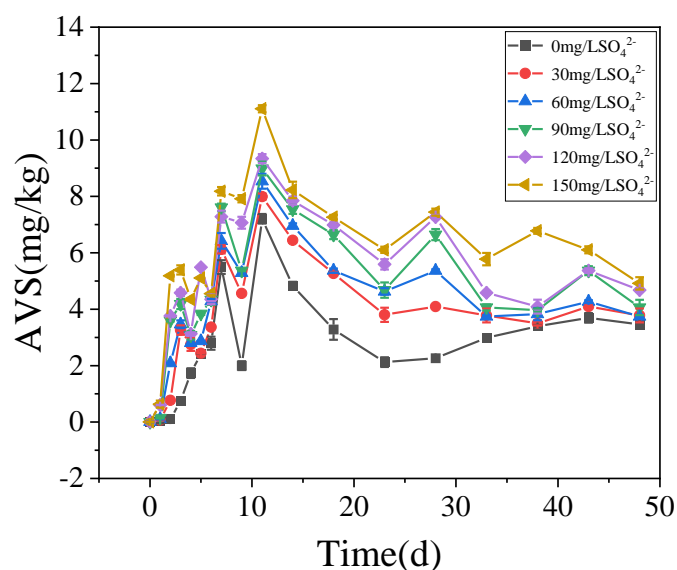
247 decomposition and sulfate reduction, the concentrations of TP in all treatments
 248 negatively correlated with the initial SO_4^{2-} concentration. The final TP concentration
 249 was 448.92, 335.32, 321.56, 259.32, 238.56 and 227.21 mg/kg, respectively in all
 250 treatments.



251
 252 Figure 3. The concentrations of TP in the overlying water (left) and sediments (right)
 253 during the incubation

254 3.4 AVS dynamics in the sediments

255 The concentrations of AVS in the sediments were positively correlated with the
 256 initial SO_4^{2-} concentrations. With the increase of TP in overlying water, the AVS in the
 257 sediments also increased steadily and reached the peak on the 11st days. In the treatment
 258 with initial SO_4^{2-} concentration of 0, 30, 60, 90, 120 and 150 mg/L, the highest
 259 concentration of AVS in the sediments were 7.21, 7.99, 8.54, 8.99, 9.34 and 11.11
 260 mg/kg, respectively.



261

262 Figure 4. The concentration of AVS in the sediments during the incubation

263 *3.5 SRB dynamics in the sediments*

264 During the decomposition of cyanobacteria, the SRB abundance significantly
 265 increased compared with the initial stage ($P < 0.01$). In the initial stage, the SRB
 266 abundance was 1.09×10^8 copies/g and the final value was positively correlated with the
 267 initial SO_4^{2-} . On 7 d, SRB of all treatments showed a downward trend compared with
 268 the initial value, and there was no significant difference in SRB values between each
 269 treatment. On 38 d, except for the initial SO_4^{2-} concentrations of 0 and 30 mg/L, SRB
 270 increased significantly in other treatments.

271 Table 2. Copy numbers of the *dsrB* gene of SRB in the sediments during the incubation

272 (copies/g)

Time SO_4^{2-} (mg/L)	0 d	7 d	38 d
0	1.09×10^8	5.81×10^7	6.65×10^7
30	1.09×10^8	6.13×10^7	7.71×10^7
60	1.09×10^8	7.61×10^7	1.15×10^8
90	1.09×10^8	7.87×10^7	1.31×10^8
120	1.09×10^8	7.99×10^7	1.49×10^8
150	1.09×10^8	8.23×10^7	1.91×10^8

273 4.Discussion

274 It is generally acknowledged that climate warming and exogenous nutrient input
275 are the important contributors to the occurrence of cyanobacteria blooms (Anneville et
276 al., 2015; Yan et al., 2017). However, in this study, we found that the dramatically
277 increasing SO_4^{2-} concentration in eutrophic lakes is also a non-negligible promoter for
278 the self-sustaining of cyanobacteria blooms. In eutrophic lakes, the decomposition of
279 cyanobacteria consumed DO in the water, and formed strong anaerobic reduction
280 conditions (Fig.S1). Fe-P was desorbed to from free Fe^{3+} , which was reduced to Fe^{2+} in
281 anaerobic environments (Fig.1). Free Fe^{2+} combined with $\sum\text{S}^{2-}$ which generated by
282 sulfate reduction and eventually formed AVS fixed in the sediments (Fig.4), and
283 phosphorus was released from the sediments (Fig.3). It has been reported that SRB and
284 iron reduction bacteria (IRB) are the main microorganisms that drive sulfate reduction
285 and iron reduction, respectively, and cyanobacteria decomposition promotes these
286 microorganisms' growth (Wu et al., 2018). Consistent with these results, our findings
287 also revealed that cyanobacteria released large amounts of organic matter to promote
288 microbial growth during their decay and decomposition (Fig.S2, Tab. 2) and ultimately
289 promoted anaerobic reduction of sulfur and iron (Holmer et al., 2001). Therefore, with
290 increasing SO_4^{2-} concentrations in eutrophic lakes, the influence of sulfate reduction on
291 phosphorus release is worth further investigation.

292 Sulfur and iron in eutrophic lake sediments are directly related to iron and
293 phosphorus, and sulfur and phosphorus are also closely linked to bridges under the
294 action of iron (Zhang et al., 2020). With the increase of SO_4^{2-} concentration in eutrophic

295 lakes, the effect of sulfate reduction on phosphorus release from sediments may be more
296 important than previously recognized (Pester et al., 2012). Sulfate reduction driven by
297 SRB is an important organic metabolism pathway in natural systems. During the sulfate
298 reduction process, SO_4^{2-} is an electron acceptor and its concentration variation can
299 significantly affect the sulfate reduction rate (Holmer et al., 2001; Nakagawa et al.,
300 2012). SO_4^{2-} is reduced to $\sum\text{S}^{2-}$ by acquiring the electrons supplied by SRB oxidation,
301 and thus SRB plays an important role in sulfate reduction (Sela-Adler et al., 2017). The
302 increase of SO_4^{2-} concentration promotes the SRB abundance, as evidenced by a
303 positive correlation (Wu et al., 2018). In the case of increased SRB abundance (Tab. 2)
304 and increased SO_4^{2-} concentration, the sulfate reduction reaction was enhanced. The
305 SO_4^{2-} concentration in the overlying water decreased significantly accompanied by a
306 temporary increase in $\sum\text{S}^{2-}$ (Fig.2). The highest concentrations of $\sum\text{S}^{2-}$ also increased
307 with the initial SO_4^{2-} concentrations (Fig.5a). Interestingly, the $\sum\text{S}^{2-}$ decreased rapidly
308 after day 10 to almost zero at the end (Fig.2). This may result from the two keys: (a)
309 hydrogen sulfide overflows from the incubator; (b) sulfide migrates downward, and
310 combines with other substances in the sediment and is immobilized (Zhang et al., 2020).
311 In this study, TP in the overlying water has a significant positive correlation with the
312 initial SO_4^{2-} concentrations ($R^2 = 0.96$; Fig.3). The classical theory presumes that iron
313 reduction by IRB leads to the release of iron-bound phosphorus in the anaerobic layer
314 of sediments, and when the formed Fe^{2+} enters the aerobic water layer, it is oxidized by
315 Fe^{3+} and bound to phosphorus again (Roden et al., 2006; Chen et al., 2016). When the
316 sulfate reduction process mediates the iron reduction process, the released Fe^{2+}

317 combines with the product $\sum S^{2-}$ of sulfate reduction to form Fe-S, thus weakening the
318 reoxidation process of Fe^{2+} , and increasing the release of phosphorus (Mort et al., 2010;
319 Zhao et al., 2019). Therefore, with the increase of SO_4^{2-} concentrations in eutrophic
320 lakes, it significantly promoted the release of endogenous phosphorus from the
321 sediments.

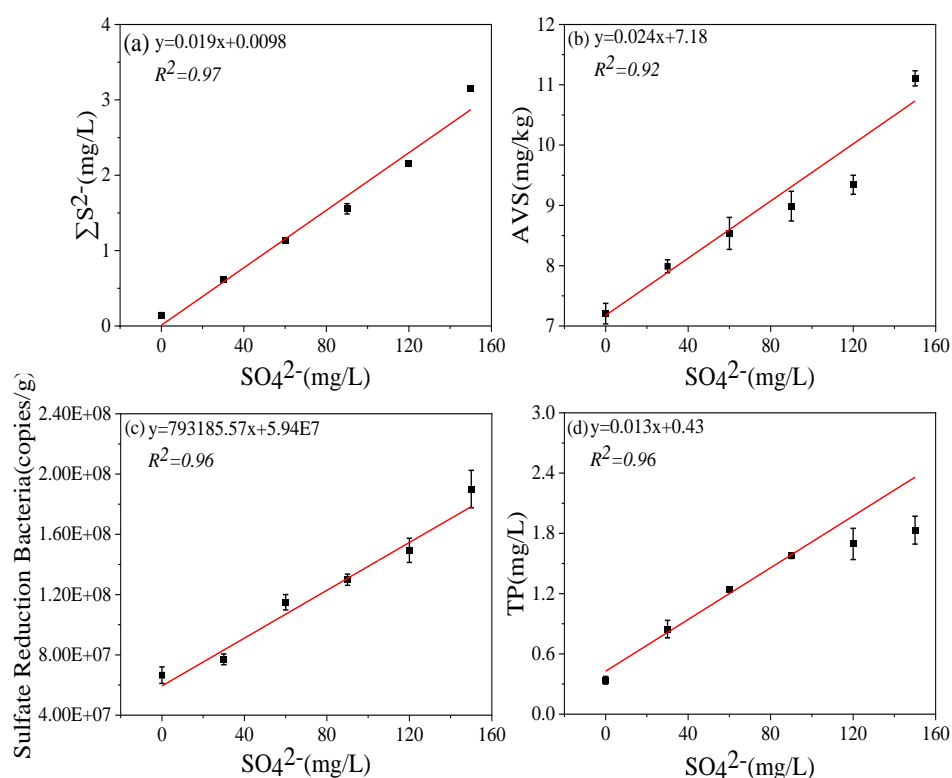
322 Although from a thermodynamic point of view, iron reduction should take
323 precedence over sulfur reduction (Han et al., 2015). However, due to chemical kinetics,
324 sulfur reduction occurs before iron reduction, resulting in the simultaneous appearance
325 of $\sum S^{2-}$ and iron oxides (Han et al., 2015; Hansel et al., 2015). This is consistent with
326 the concentration variation of iron and sulfur in this study (Fig.1-3). It has been reported
327 that iron cycles in the water body will produce an intense response to the accumulation
328 of sulfide, that is, sulfate reduction can promote iron reduction (Friedrich et al., 2014;
329 Zhang et al., 2020). $\sum S^{2-}$ is the final product of sulfate reduction, which is toxic to
330 microorganisms and easy to combine with heavy metals such as Fe^{2+} to form AVS in
331 lake sediments (Holmer et al., 2001). In this study, the concentration of AVS showed a
332 significant positive correlation with the initial concentration of SO_4^{2-} (Fig. 4, 5b), which
333 was consistent with the highest concentration of $\sum S^{2-}$ observed in the overlying water
334 (Fig. 2, 5c). The concentrations of Fe^{2+} and Fe^{3+} in the overlying water increased
335 significantly, and Fe^{2+} significantly decreased in the middle of the incubation (Fig. 1),
336 suggesting that Fe^{2+} reduced by sulfate can be combined with the product $\sum S^{2-}$ (Fig. 2).
337 These results consistent with the trend that AVS in the sediments reached a peak after
338 11 days and $\sum S^{2-}$ in the water decreased rapidly after 9 days and remained at a lower

339 concentration (Fig. 2, 3). The reason for this phenomenon may be the formation of Fe-
340 S compounds that is finally fixed in the sediments (Zhao et al., 2019).

341 The ΣS^{2-} mediated iron chemical reduction may lead to more environmental
342 effects, such as phosphorus mobilization (Zhang et al., 2020). For instance, a previous
343 investigation on the lakes along the Yangtze River demonstrates that the effects of
344 endogenous phosphorus release is probably related to the increase of SO_4^{2-}
345 concentration (Chen et al., 2016). In this study, the concentration of Fe^{2+} in the
346 treatment without SO_4^{2-} continued to rise, and was up to the highest concentration
347 among all treatments (Fig. 1). In contrast, the concentrations of TP in the treatment
348 without SO_4^{2-} showed the lowest concentration among all treatments (Fig. 1, 5a). This
349 is caused by Fe^{2+} and Fe^{3+} recombining with phosphorus and being immobilized in the
350 sediments (Wu et al., 2019). In general, iron combines with phosphorus to form siderite
351 ($FePO_4 \cdot 2H_2O$) and blue iron ($Fe_3(PO_4)_2 \cdot 8H_2O$) and is bound to the sediments (Taylor
352 et al., 2011). However, when precipitation or reduction separates iron from iron
353 phosphate minerals, phosphorus bound to iron is released (Gu et al., 2016).

354 In order to further elucidate whether the increasing SO_4^{2-} concentrations in
355 overlying water result in the self-sustaining of eutrophication in shallow lakes, a
356 conceptual diagram was put forward (Fig. 6). It has been accepted that exogenous
357 nutrient inputs and climate warming have positive effects on the breakout of
358 cyanobacteria blooms. With the continuous input of exogenous sulfur, the SO_4^{2-}
359 concentration in the lake water increases significantly. When cyanobacteria blooms
360 start to decay, the overlying water shifts from the aerobic state to the strong anaerobic

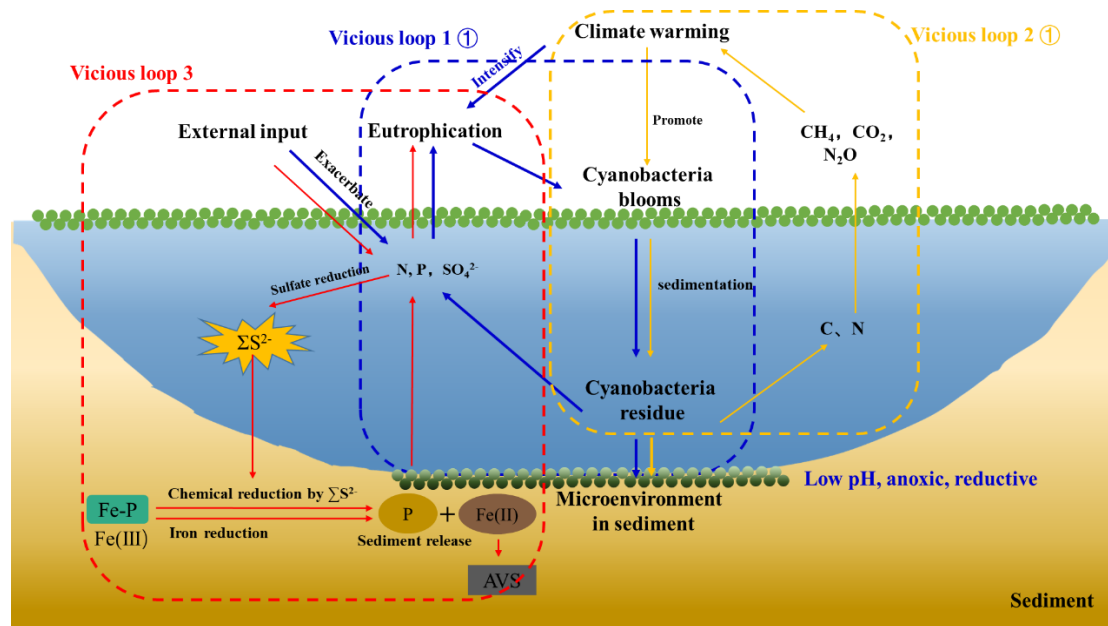
361 state, providing carbon source to promote the growth of microorganisms such as SRB.
 362 The increasing SO_4^{2-} concentrations provide the electron for the sulfate reduction
 363 process, resulting in the sulfate reduction and the release of a large amount of ΣS^{2-} . The
 364 Fe^{2+} released from the iron reduction process is captured by ΣS^{2-} , and finally the
 365 combination of iron and P was reduced, promoting the release of endogenous
 366 phosphorus. Therefore, it is necessary to pay attention to the effect of enhanced sulfate
 367 reduction on endogenous phosphorus release in eutrophic lakes.



368

369 Figure 5. Correlation of initial SO_4^{2-} concentrations with ΣS^{2-} (a), AVS(b), Sulfate-
 370 reducing bacteria (SRB) (c), TP (d) in the microcosm systems, respectively.

371



372

373 Figure 6. A simplified scheme of the relationship among climate warming, lake
 374 eutrophication and cyanobacteria blooms in eutrophic lakes. Under climate warming
 375 scenarios, extreme abiotic and biotic conditions facilitated the breakout of
 376 cyanobacteria blooms. After their collapse, the high amount of N, P, and C were
 377 released into the overlying water and reacted with the eutrophication. Furthermore, a
 378 large amount of CH_4 and CO_2 was produced and emitted to the atmosphere, contributing
 379 to global warming of freshwater lakes (Yan et al. 2017). With the external sulfur input,
 380 the concentration of SO_4^{2-} increased significantly and sulfate reduction was enhanced.
 381 The cyanobacteria decomposition created an anaerobic reduction environment, which
 382 will promote iron reduction and sulfate reduction. The free Fe^{3+} generated by Fe-P
 383 desorption was reduced to Fe^{2+} and combined with ΣS^{2-} which produced by sulfate
 384 reduction to form stable Fe-S in the sediments. Phosphorus was released from the
 385 sediment into the overlying water. Therefore, there are three vicious loops between
 386 cyanobacteria blooms occurrence, lake eutrophication and climate warming.

387

388 **5.Conclusion**

389 The dramatical increase of SO_4^{2-} concentration was up to more than 100 mg/L in
390 eutrophic lakes. There was a coupling relationship between sulfur, iron and phosphorus
391 cycles in lake ecosystems. Rapidly increasing sulfate concentration enhanced the
392 sulfate reduction to release of a large amount of $\sum\text{S}^{2-}$ mediated by the increasing
393 abundance of SRB with the adequate organic source from the decay processes of
394 cyanobacteria blooms. The iron reduction, in positive with initial sulfate concentration,
395 occurred with the cyanobacteria decomposition. The Fe^{2+} released from the iron
396 reduction process was captured by $\sum\text{S}^{2-}$, and finally the combination of iron and P was
397 reduced, promoting the release of endogenous phosphorus. Therefore, except for
398 climate warming and excessive nutrients, the increasing sulfate concentration is proved
399 to be another hidden promoter of eutrophication in shallow lakes.

400

401 **Author contributions**

402 Xu Xiaoguang: designed and led the study. Zhou Chuanqiao, Peng Yu, Chen Li,
403 Yu Miaotong, Muchun Zhou, Xu Runze, Lanqing Zhang, Siyuan Zhang: performed the
404 investigation and analysed the samples. Zhou Chuanqiao and Peng Yu: wrote the
405 original draft with major edits and inputs from Xu Xiaoguang, Zhang Limin and Wang
406 Guoxiang.

407

408 **Competing interests**

409 The authors declare that they have no known competing financial interests or

410 personal relationships that could have appeared to influence the work reported in this
411 paper.

412

413 **Acknowledgements**

414 This work was supported by the National Natural Science Foundation of China
415 (No.42077294, 41877336, 41971043), the Cooperation and Guidance Project of
416 Prospering Inner Mongolia through Science and Technology (No.2021CG0037), the
417 National Key Research and Development Program of China (No.2021YFC3200304),
418 the Guangxi Key Research and Development Program of China (No.2018AB36010).

419

420 **References**

421 Amirbahman, A., Pearce, A.R., Bouchard, R.J., Norton, S.A., Kahl, J.S.: Relationship
422 between hypolimnetic phosphorus and iron release from eleven lakes in Maine,
423 USA, *Biogeochemistry*, 65(3), 369-385, 10.1023/A:1026245914721, 2003.

424 Anneville, O., Domaizon, I., Kerimoglu, O., Rimet, F., Jacquet, S.: Blue-Green algae
425 in a “Greenhouse Century”? new insights from field data on climate change impacts
426 on cyanobacteria abundance, *Ecosystems*, 18(3), 441-458, 10.1007/s10021-014-
427 9837-6, 2015.

428 Azam, H.M., Finneran, K.T.: Fe(III) reduction-mediated phosphate removal as
429 vivianite ($\text{Fe}_3(\text{PO}_4)_2 \cdot 8\text{H}_2\text{O}$) in septic system wastewater, *Chemosphere*, 97, 1-9,
430 100.1016/j.chemosphere.2013.09.032, 2014.

431 Baldwin, D.S., Mitchell, A.: Impact of sulfate pollution on anaerobic biogeochemical

432 cycles in a wetland sediment, *Water Research*, 46(4), 965-974,
433 10.1016/j.watres.2011.11.065, 2012.

434 Chen, M., Li, X.H., He, Y.H., Song, N., Cai, H.Y., Wang, C.H., Li, Y.T., Chu, H.Y.,
435 Krumholz, L.R., Jing, H.L.: Increasing sulfate concentrations result in higher
436 sulfide production and phosphorous mobilization in a shallow eutrophic freshwater
437 lake, *Water Research*, 96, 94-104, 10.1016/j.watres.2016.03.030, 2016.

438 Chen, M., Ye, T.R., Krumholz, L.R., Jiang H.L.: Temperature and cyanobacteria bloom
439 biomass influence phosphorous cycling in eutrophic lake sediments, *Plos One*, 9(3),
440 e93130, 10.1371/journal.pone.0093130, 2014.

441 Cline, J.D.: Spectrophotometric determination of hydrogen sulfide in natural waters,
442 *Limnology and Oceanography*, 14, 454-458, 1969.

443 Dierberg, F.E., DeBusk, T.A., Larson, N.R., Kharbanda, M.D., Chan, N., Gabriel, M.C.:
444 Effect of sulfate amendments on mineralization and phosphorus release from South
445 Florida (USA) wetland soils under anaerobic conditions, *Soil Biology &*
446 *Biochemistry*, 43(1), 31-45, 10.1013/j.soilbio.2010.09.006, 2011.

447 Ebina, J., Tsutsui, T., Shirai, T.: Simultaneous determination of total nitrogen and total
448 phosphorus in water using peroxodisulfate oxidation, *Water Research*, 17(12),
449 1721-1726, 1983.

450 Fike, D.A., Bradley, A.S., Rose, C.V.: Rethinking the ancient sulfur cycle, *Annual*
451 *Review of Earth and Planetary Science*, 43, 593-622, 10.1146/annurev-warth-
452 060313-054802, 2015.

453 Friedrich, M.W., Finster, K.W.: How sulfur beats iron, *Science*, 344(6187), 974-975,

454 10.1126/science.1255442, 2014.

455 Gu, S., Qian, Y.G., Jiao, Y., Li, Q.M., Pinay, G., Gruau, G.: An innovative approach
456 for sequential extraction of phosphorus in sediments: Ferrous iron P as an
457 independent P fraction, *Water Research*, 103, 352-361,
458 10.1016/j.watres.2016.07.058, 2016.

459 Gunnars, A., Blomqvist, S.: Phosphate exchange across the sediment-water interface
460 when shifting from anoxic to oxic conditions an experimental comparison of
461 freshwater and brackish-marine systems, *Biogeochemistry*, 37(3), 203-226, 1997.

462 Guo, M.L., Li, X.L., Song, C.L., Liu, G.L., Zhou, Y.Y.: Photo-induced phosphate
463 release during sediment resuspension in shallow lakes: A potential positive
464 feedback mechanism of eutrophication, *Environmental Pollution*, 258, 113679,
465 10.1016/j.envpol.2019.113679, 2020.

466 Han, C., Ding, S.M., Yao, L., Shen, Q.S., Zhu, C.G., Wang, Y., Xu, D.: Dynamics of
467 phosphorus-iron-sulfur at the sediment-water interface influenced by algae blooms
468 decomposition, *Journal of Hazardous Materials*, 300, 329-337,
469 10.1016/j.jhazmat.2015.07.009, 2015.

470 Hansel, C.M., Lentini, C.J., Tang, Y.Z., Johnston, D.T., Wankel, S.D., Jardine, P.M.:
471 Dominance of sulfur-fueled iron oxide reduction in low-sulfate freshwater
472 sediments, *The ISME Journal*, 9(11), 2400-2412, 10.1038/ismej.2015.50, 2015.

473 Ho, J.C., Michalak, A.M., Pahlevan, N.: Widespread global increase in intense lake
474 phytoplankton blooms since the 1980s, *Nature* 574, 667-670, 10.1038/s41589-019-
475 1648-7, 2019.

476 Holmer, M., Storkholm, P.: Sulphate reduction and sulphur cycling in lake sediments:
477 a review, *Freshwater Biology*, 46, 431-451, 10.1046/j.1365-2427.2001.00687.x,
478 2001.

479 Hsieh, Y.P., Shieh, Y.N.: Analysis of reduced inorganic sulfur by diffusion methods:
480 improved apparatus and evaluation for sulfur isotopic studies, *Chemical Geology*,
481 137(3), 255-261, 1997.

482 Iwayama, A., Ogura, H., Hirama, Y., Chang, C.W., Hsieh, C.H., Kagami, M.:
483 Phytoplankton species abundance in Lake Inba (Japan) from 1986 to 2016,
484 *Ecological Research*, 32(6), 783-783, 10.1007/s11284-017-1482-z, 2017.

485 Jorgensen, B.B., Findlay, A.J., Pellerin, A.: The Biogeochemical sulfur cycle of Marine
486 sediments, *Frontiers in Microbiology*, 10, 849, 10.3389/fmicb.2019.00849, 2019.

487 Liu, Z.S., Zhang, Y., Han, F., Yan, P., Liu, B.Y., Zhou, Q.H., Min, F.L., He, F., Wu,
488 Z.B.: Investigation on the adsorption of phosphorus in all fractions from sediment
489 by modified maifanite, *Scientific Reports*, 8, 15619, 10.1038/s41598-018-34144-w,
490 2018.

491 Mao, Z.G., Gu, X.H., Cao, Y., Luo, J.H., Zeng, Q.F., Chen, H.H., Jeppesen, E.: How
492 does fish functional diversity respond to environmental changes in two large
493 shallow lakes? *Science of the total environment*, 753, 142158,
494 10.1016/j.scitotenv.2020.142158, 2021.

495 Mort, H.P., Slomp, C.P., Gustafsson, B.G., Andersen, T.J.: Phosphorus recycling and
496 burial in Baltic sea sediments with contrasting redox conditions, *Geochimica et*
497 *Cosmochimica Acta*, 74(4), 1350-1362, 10.1016/j.gca.2009.11.016, 2010.

498 Melemdez-Pastor, I., Isenstein, E.M., Navarro-Pedreno, J., Park, M.H.: Spatial
499 variability and temporal dynamics of cyanobacteria blooms and water quality
500 parameters in Missisquoi Bay (Lake Champlain), *Water Supply*, 19(5), 1500-1506,
501 10.2166/ws.2019.017, 2019.

502 Nakagawa, M., Ueno, Y., Hattori, S., Umemura, M., Yagi, A., Takai, K., Koba, K.,
503 Sasaki, Y., Makabe, A., Yoshida, N.: Seasonal change in microbial sulfur cycling
504 in monomictic Lake Fukami-ike, Japan, *Limnology and Oceanography*, 57(4), 974-
505 988, 10.4319/lo.2012.57.4.0974, 2012.

506 Ni, Z.K., Wang, S.R., Wu, Y., Pu, J.: Response of phosphorus fractionation in lake
507 sediments to anthropogenic activities in China, *Science of the Total Environment*,
508 699, 134242, 10.1016/j.scitotenv.2019.134242, 2020.

509 Pan, P., Guo, Z.R., Cai, Y., Liu, H.T., Wang, B., Wu, J.Y.: High-resolution imaging of
510 labile P&S in coastal sediment: Insight into the kinetics of P mobilization associated
511 with sulfate reduction, *Marine Chemistry*, 225, 103851, 10.1016/j.marchem.2020.
512 103851, 2020.

513 Pester, M., Knorr, K.H., Friedrich, M.W., Wagner, M., Loy, A.: Sulfate-reducing
514 microorganisms in wetlands-fameless actors in carbon cycling and climate change,
515 *Frontiers in Microbiology*, 3(72), 10.3389/fmicb.2012.00072, 2012.

516 Phillips, E.J.P., Lovley, D.R.: Determination of Fe(III) and Fe(II) in Oxalate Extracts
517 of Sediment, *Soil Science Society of America Journal*, 51: 938-941, 1987.

518 Roden, E.E.: Geochemical and microbiological controls on dissimilatory iron reduction,
519 *Comptes Rendus Geoscience*, 338(6-7), 456-467, 10.1016/j.crte.2006.04.009, 2006.

520 Ruban, V., Lopez-Sanchez, J.F., Pardo, P., Rauret, G., Muntau, H., Quevauviller, P.:
521 Harmonized protocol and certified reference material for the determination of
522 extractable contents of phosphorus in freshwater sediments-A synthesis of recent
523 works, *Fresenius J Anal Chem*, 370, 224-228, 10.1007/s002160100753, 2001.

524 Sela-Adler, M., Ronen, Z., Herut, B., Antler, G., Vigderovich, H., Eckert, W., Sivan,
525 O.: Co-existence of Methanogenesis and sulfate reduction with common substrates
526 in sulfate-rich estuarine sediments, *Frontiers in Microbiology*, 8(766),
527 10.3389/fmicb.2017.00766, 2017.

528 Tabatabai, M.: A rapid method for determination of sulfate in water samples,
529 *Environmental*, 7, 237-243, 1974.

530 Taylor, K.G., Konhauser, K.O.: Iron in Earth surface systems: a major player in
531 chemical and biological processes, *Elements*, 7(2), 83-87,
532 10.2113/gselements.7.2.83, 2011.

533 Thamdrup, B., Dalsgaard, T., Jensen, M.M., Petersen, J.: Anammox and the marine N
534 cycle, *Geochimica et cosmochimica acta*, 68(11), A325, 2004.

535 Wu, S.J., Zhao, Y.P., Chen, Y.Y., Dong, X.M., Wang, M.Y., Wang, G.X.: Sulfur
536 cycling in freshwater sediments: A cryptic driving force of iron deposition and
537 phosphorus mobilization, *Science of the total environment*, 657, 1294-1303,
538 10.1016/j.scitotenv.2018.12.161, 2019.

539 Xu, G.H., Sun, Z.H., Fang, W.Y., Liu, J.J., Xu, X.B., Lv, C.X.: Release of phosphorus
540 from sediments under wave-induced liquefaction, *Water Research*, 144, 503-511,
541 10.1016 /j.watres.2018.07.038, 2018.

542 Yan, X.C., Xu, X.G., Wang, M.Y., Wang, G.X., Wu, S.J., Li, Z.C., Sun, H., Shi, A.,
543 Yang, Y.H.: Climate warming and cyanobacteria blooms: Looks at their
544 relationships from a new perspective, *Water Research*, 125, 449-457,
545 10.1016/j.watres.2017.09.008, 2017.

546 Yu, T., Zhang, Y., Wu, F.C., Meng, W.: Six-Decade change in water chemistry of large
547 freshwater lake Taihu, China, *Environmental Science and Technology*, 47(16),
548 9093-9101, 10.1021/es401517h, 2013.

549 Zhang, S.Y., Zhao, Y.P., Zhou, C.Q., Duan, H.X., Wang, G.X.: Dynamic sulfur-iron
550 cycle promoted phosphorus mobilization in sediments driven by the algae
551 decomposition, *Ecotoxicology*, 30(8), 1662-1671, 10.1007/s10646-020-02316-y,
552 2020.

553 Zhao, Y.P., Wu, S.J., Yu, M.T., Zhang, Z.Q., Wang, X., Zhang, S.Y., Wang, G.X.:
554 Seasonal iron-sulfur interactions and the stimulated phosphorus mobilization in
555 freshwater lake sediments, *Science of the total environment*, 768, 144336,
556 10.1016/j.scitotenv.2020.144336, 2021.

557 Zhao, Y.P., Zhang, Z.Q., Wang, G.X., Li, X.J., Ma, J., Chen, S., Deng, H., Annalisa
558 O.H.: High sulfide production induced by algae decomposition and its potential
559 stimulation to phosphorus mobility in sediment, *Science of the total environment*,
560 650, 163-172, 10.1016/j.scitotenv.2018.09.010, 2019.

561 Zhou, C.Q., Peng, Y., Deng, Y., Yu, M.T., Chen, L., Zhang, L.Q., Xu, X.G., Zhao, F.J.,
562 Yan, Y., Wang, G.X.: Increasing sulfate concentration and sedimentary decaying
563 cyanobacteria co-affect organic carbon mineralization in eutrophic lakes sediments,

564 Science of the total environment, 2022, 806, 151260, 10.1016/j.scitotenv.2021.

565 151260, 2022.