



Biogeochemistry of climate driven shifts in Southern Ocean primary producers

Ben J. Fisher¹, Alex J. Poulton², Michael P. Meredith³, Kimberlee Baldry⁴, Oscar Schofield⁵, Sian F. Henley¹.

5 ¹ School of GeoSciences, University of Edinburgh, Edinburgh, United Kingdom

² The Lyell Centre for Earth and Marine Science, Heriot-Watt University, Edinburgh, United Kingdom

³ British Antarctic Survey, Cambridge, United Kingdom

⁴ Institute for Marine and Antarctic Studies, College of Sciences and Engineering, University of Tasmania, Hobart, TAS, Australia

10 ⁵ Center of Ocean Observing Leadership, School of Environmental and Biological Sciences, Rutgers University, New Brunswick, NJ 08901, USA

Correspondence to: Ben J. Fisher (ben.fisher@ed.ac.uk)

Abstract. As a net source of nutrients fuelling global primary production, changes in Southern Ocean productivity are expected to influence biological carbon storage across the global ocean. Following a high emissions, low mitigation pathway, primary productivity in the Southern Ocean is predicted to increase by up to 40% over the 21st century. The ecophysiological response of marine phytoplankton experiencing climate change will be a key determinant in understanding the impact of Southern Ocean productivity shifts on the carbon cycle. Yet, phytoplankton ecophysiology is poorly represented in CMIP6 climate models, leading to substantial uncertainty in the representation of their role in carbon sequestration. Here we synthesise the existing spatial and temporal projections of Southern Ocean productivity from CMIP6 models, separated by phytoplankton class and identify key processes where greater observational data coverage can help to improve future model performance. We find bidirectional changes in iron and light limitation of phytoplankton, while the greatest changes in productivity occur in the coastal zone of the Southern Ocean. Different phytoplankton groups are responsible for driving productivity increases at different latitudes, yet we observe that models disagree on the ecological mechanism behind these productivity changes. We propose that an evidence-based sampling approach targeting climate-driven changes in ocean biogeochemistry and community assemblages in the regions of rapid projected productivity changes could help to resolve the empirical principles underlying phytoplankton community structure in the Southern Ocean.

1. Vulnerability of Southern Ocean ecosystems to climate change

The biological uptake of carbon by marine phytoplankton represents an increasingly important process in the Earth system, with ocean carbon storage mediating atmospheric CO₂ concentrations, including CO₂ of anthropogenic origin (Riebesell et al., 2007, Friedlingstein et al., 2022). The Southern Ocean is a disproportionately large carbon and heat sink relative to its size (Frölicher et al., 2015). Across the global ocean, uptake of carbon accounts for ~25% of CO₂ released by human activities



(Friedlingstein et al., 2022), with the Southern Ocean accounting for 30-40% of this global anthropogenic CO₂ uptake (e.g. Caldeira and Duffy, 2000, DeVries, 2014), predominantly due to enhanced atmosphere-ocean exchange at increased atmospheric CO₂ concentrations (Friedlingstein et al., 2022). While biological uptake is considered to play a minor role in total CO₂ uptake (Landschützer et al., 2015, Gruber et al., 2019), variability in pCO₂ has been associated with summertime blooms in the Southern Ocean (Gregor et al., 2018). Under a future climate scenario with longer growth seasons (Moreau et al., 2015), increased seasonal productivity (Leung et al., 2015, Fu et al., 2016) and a higher Revelle factor (Hauck et al., 2015), pelagic ecosystems are expected to become increasingly important in Southern Ocean carbon uptake (Henley et al., 2020).

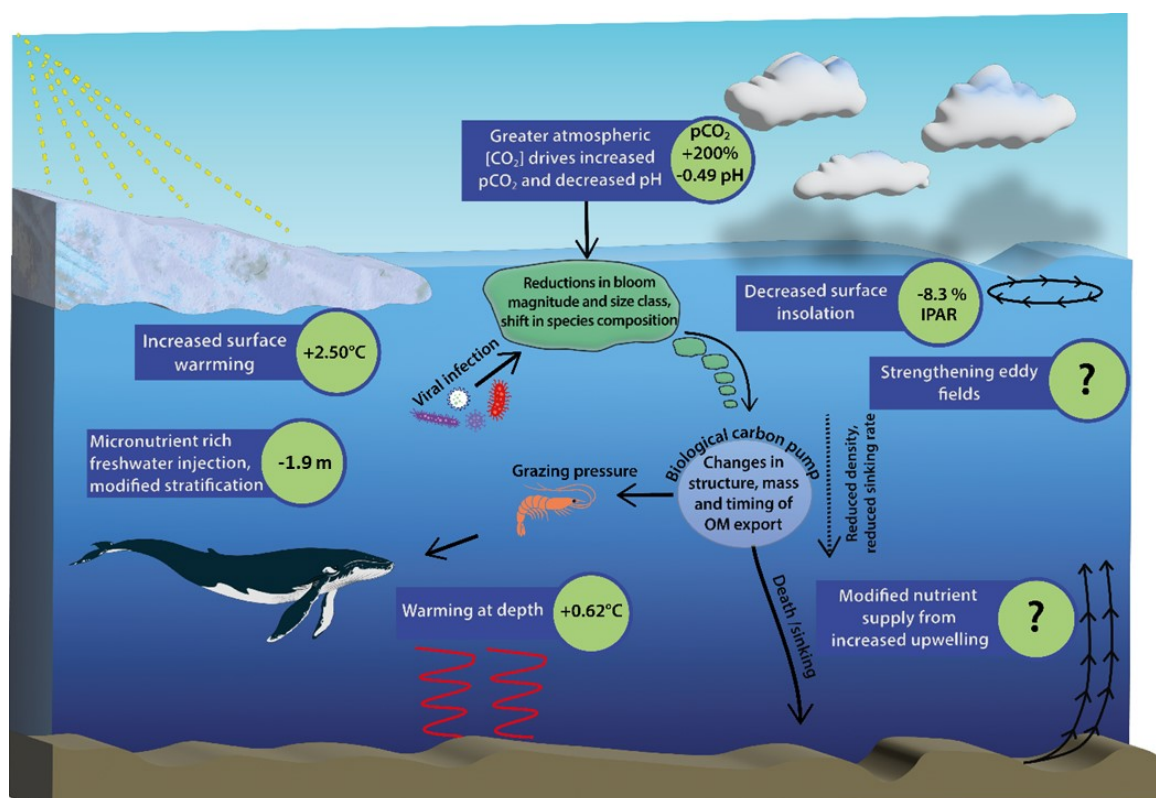


Figure 1: Schematic diagram of Southern Ocean pressures associated with climate change and the downstream biogeochemical consequences for ecosystem productivity. Values shown are 100 year mean changes to 2100 under a SSP5-8.5 scenario south of 65° and are taken from CMIP6 models and existing literature (see Table S2 for a full description). Question marks indicate processes where estimations of change do not exist.

Biological carbon cycling is especially vulnerable to climate change because of the small cell size of marine phytoplankton (0.002-0.2 mm), coupled with their specific requirements for light and iron, which are the preliminary factors limiting their growth in high nutrient low chlorophyll (HNLC) zones of the Southern Ocean (Moore et al., 2013). Primary production by Southern Ocean phytoplankton represents ~25% of global marine primary production, equivalent to 11.4 Pg C yr⁻¹ (Okin et al., 2011). Climate impacts on Southern Ocean phytoplankton are likely to manifest in increased productivity (Leung et al., 2015, Fu et al., 2016), ecological shifts towards smaller cell sizes (Venables et al., 2013, Saba et al., 2014, Schofield et al., 2018, Mascioni et al., 2019, Biggs et al., 2019) and changes in seasonal phenology (Moreau et al., 2015). Increases in overall



productivity can be most closely associated with reduced sea ice duration, increasing ocean surface area and light supply to the light and iron co-limited productivity system. Strengthened upwelling is also likely to increase sedimentary iron supply to the surface, however, the extent to which changes in ocean mixing can be expected to impact nutrient supplies remains largely unknown (Figure 1).

Shifts in community composition from diatoms to smaller cryptophytes have already been documented along the west Antarctic Peninsula (Montes-Hugo et al., 2008, Moline et al., 2008, Ducklow et al., 2007, Rozema et al., 2017, Moline et al., 2004), and are thought to be due to tolerance of cryptophytes to low-salinity waters induced by increased sea ice melt (Moline et al., 2004). Meanwhile, in culture-based competition experiments, diatoms are more successful in simulated future ocean conditions over prevalent haptophytes such as *Phaeocystis antarctica*, albeit with reduced diatom cell sizes (Xu et al., 2014). This difference is potentially driven by reduced iron limitation of diatoms and their greater tolerance to temperature change (Zhu et al., 2016). These varied responses between manipulation experiments and in situ observations suggest that physiological as well as ecological changes are important in understanding the net biogeochemical implications of phytoplankton community change.

In the sea-ice zone, grazing by zooplankton accounts for ~90% of phytoplankton losses (Moreau et al., 2020). Shifts in phytoplankton size class could rapidly cascade through the ecosystem as the dominant Southern Ocean zooplankton, krill are unable to graze the small cryptophytes (Haberman et al., 2003), instead promoting the dominance of carbon-poor salps (*Salpa thompsoni*), which reduces the overall efficiency of the marine food web (Ballerini et al., 2014) and potentially weakens the biological carbon pump (Biggs et al., 2021, Quéguiner, 2013). Additionally, water temperature, alongside changes to zooplankton abundance and diversity, has been shown to drive zooplankton metabolism (Mayzaud and Pakhomov, 2014), which can in turn be expected to modulate the grazing pressure and therefore the amount of POC exported. The Southern Ocean supplies $62 \pm 5\%$ of pycnocline nitrate and phosphate (Fripiat et al., 2021), therefore increased Southern Ocean productivity and regional nutrient utilization can be expected to reduce global nutrient export (Moore et al., 2018). Models of the Southern Ocean with complete nutrient trapping (no nutrient export) or no productivity (all nutrients available for export) suggest that regional nutrient supplies fuel 17 to 75% of productivity outside the Southern Ocean (Hauck et al., 2018, Primeau et al., 2013, Palter et al., 2010, Sarmiento et al., 2004).

Projections of productivity in the Southern Ocean under future climate scenarios from the Coupled Model Intercomparison Project Phase 6 (CMIP6) class Earth System Models (ESMs) are actively informing research directions, carbon budgets (Friedlingstein et al., 2022) and governmental policy (Touzé-Peiffer et al., 2020). Yet, between CMIP5 and CMIP6 the spread of model projections has increased as different models incorporate more complexity and additional processes (e.g. varying elemental stoichiometry, phytoplankton class diversity, complex elemental cycling) (Seferian et al., 2020). While representation of ocean physical drivers and nutrient fields compared to observations has improved between CMIP5 and



CMIP6, surface chlorophyll is one of three key parameters which did not show improvement in CMIP6 (Canadell et al., 2021, Fu et al., 2022). Deficiencies in model projections of phytoplankton and ocean biogeochemistry have been linked to the use of fixed C:N:P elemental stoichiometry (Kwiatkowski et al., 2018), and an inability to reflect physiological adaptations, e.g. the ability of diatoms to maintain growth under iron limitation (Person et al., 2018).

As the main source of organic carbon in the Southern Ocean, uncertainty in projections of phytoplankton composition compounds existing model uncertainty in the biological carbon flux to the ocean's interior and seafloor (Henson et al., 2022), where a fraction is available for long term storage. Simulations of phytoplankton productivity exert a particular bias on the Southern Ocean due to data scarcity resulting from weather conditions which impede chlorophyll remote sensing and sea ice preventing observation or sampling throughout the winter and early spring (Holm-Hansen et al., 2004, Laufkötter et al., 2018). Therefore, we argue that a targeted approach to establish phytoplankton-environment response interactions within the regions of the most rapid projected changes is essential to accelerate the improvement of phytoplankton representation in future generations of ESMs.

2. Physical climate driven changes in Southern Ocean water masses

Climate change is driving substantial changes in Southern Ocean water masses (Bindoff et al., 2019). The strengthening of Southern Ocean winds (Figure 2a) and increased buoyancy fluxes (including freshwater inputs) act as opposing drivers of stratification, modifying mixed layer depth (Figure 2b) and influencing iron and light availability to the upper ocean, thereby determining the abundance and composition of primary producers. Despite the importance of changes in Southern Ocean circulation for global ocean nutrient supply, the cumulative influence of microscale physical processes results in poor overall performance of CMIP-class models in this region when their historical runs are compared with observations (Meredith et al., 2019).

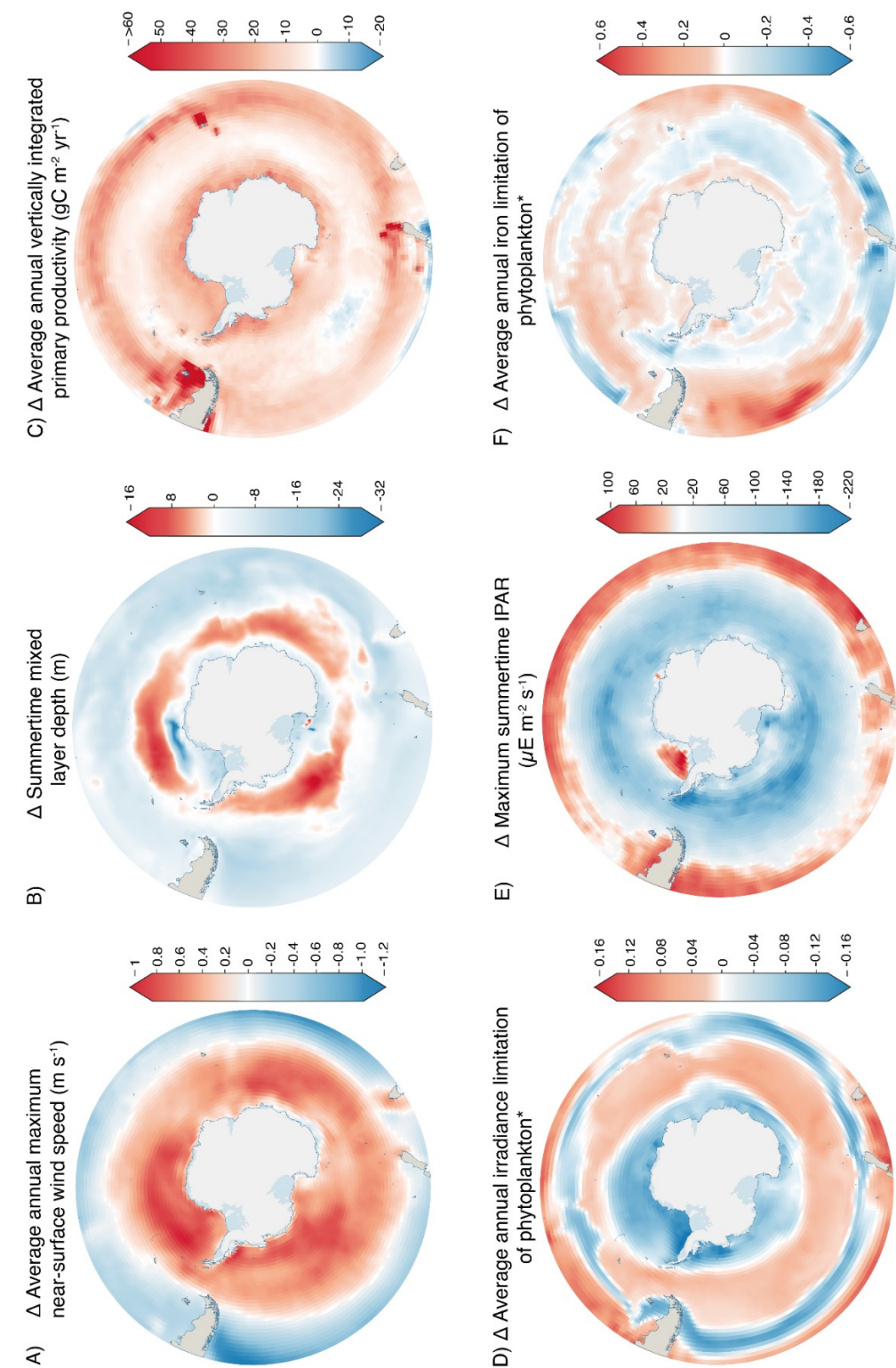
Across the Southern Ocean, the timing of the springtime onset of net primary production and the magnitude of summer biomass accumulation are controlled by light availability, as dictated by sea ice extent, cloud cover and water column structure (Henley et al., 2017). CMIP6 models project the greatest increase in productivity to occur across the coastal zone of the Southern Ocean (65-90°S) (Figure 2c), where irradiance limitation is reduced (Figure 2d) due to a shallowing of the mixed layer (Figure 2b) and increased light delivery in the Weddell Sea (Figure 2e), before experiencing stronger iron limitation (Figure 2f). Conversely, across the Transitional zone (40-50°S) the mixed layer deepens, incidental photosynthetically active radiation (IPAR) reduces, irradiance limitation increases and productivity increases are restricted compared to the rest of the Southern Ocean. Increased iron limitation (Figure 2f) likely manifests from greater competition for iron driven by increased productivity (Figure 2c) and a deepening of the mixed layer in the Subantarctic (50-65°S) (Figure 2b). Iron supply to the surface is subject to changes to the forcing and properties of water masses, resulting in variable circulation strengths, depth boundaries, heat content and carbon sequestration resulting from climate-driven perturbations to the ice-ocean-atmosphere system (Meredith et al., 2019, Bindoff et al., 2019). Upwelling of nutrients and light availability for phytoplankton are both strongly influenced by



120 mixed layer depth, which in turn varies seasonally with increased solar warming and ice melt driving deeper Southern Ocean
 pycnocline stratification through the summer (Sallée et al., 2021).

Additionally, a substantial proportion of the extra energy resulting from zonal wind strengthening (Figure 2d) is imparted to
 the mesoscale eddy field (Meredith and Hogg, 2006, Hogg and Munday, 2014, Hogg et al., 2015). This intensification of the
 125 eddies may lead to regional enhancements in vertical mixing and nutrient supply, especially where it is co-located with regions
 of rugged bathymetry (Sheen et al., 2014) and sedimentary sources of iron (Ellwood et al., 2020, Uchida et al., 2020, Kahru et
 al., 2007). Increased near-surface wind strength has also been linked to reduced upper-ocean stratification and deeper mixed
 layers around the Southern Ocean (Carranza and Gille, 2015, Sallée et al., 2021), as models predict south of 55° (Figure 2b).
 However this is counteracted near to the coast by the stabilising influences of buoyancy forcing (Shi et al., 2020). Alongside
 130 wind strength, buoyancy forcing determines the strength of the meridional overturning circulation, responsible for atmosphere-
 ocean exchange of heat and carbon (Pellichero et al., 2018). Wind activity can drive increased outgassing of CO₂ from
 upwelling of deep waters (Mikaloff Fletcher et al., 2007, Lovenduski et al., 2008), resulting in a weakening of the Southern
 Ocean CO₂ sink (Le Quéré et al., 2007) and atmospheric CO₂ increases of up to 20 µatm (Lauderdale et al., 2013). This
 outgassing can be expected to increase with more intense storm events (Nicholson et al., 2022) and in line with sea ice retreat
 135 leaving a greater amount of ocean surface available for gas exchange (Shadwick et al., 2021).

Despite the importance of buoyancy in influencing atmosphere-ocean exchange, the sea ice changes that drive buoyancy
 forcing are poorly reproduced in CMIP6 models (Roach et al., 2020, Shu et al., 2020). Currently, CMIP6 models project a
 decrease in sea ice extent during the course of this century as warming continues, albeit with further work required to reduce
 140 uncertainties surrounding the magnitudes and timings (Roach et al., 2020). Stronger westerly winds enhance the northward
 export of sea ice in the Southern Ocean, promoting freshwater injection at more northerly latitudes, and enhancing stratification
 (Haumann et al., 2016), which will increase light availability. This northward flux also increases salinity at high latitudes
 (Pellichero et al., 2018), but this is compensated and exceeded regionally by increased discharge of glacial meltwater from the
 Antarctic Ice Sheet (Fogwill et al., 2015). This glacial discharge is likely to accelerate under continued warming in both West
 145 and East Antarctica, with the Amundsen and Bellingshausen Seas likely to be impacted most rapidly, and accelerating
 freshwater discharge reaching as far west as the Ross Sea (Levermann et al., 2014, Meredith et al., 2019). Recent large and
 unexpected changes in sea ice around Antarctica emphasise that greater knowledge of the key drivers and controls is required,
 in order to improve predictive skill in models (Turner and Comiso, 2017).



150 Figure 2: CMIP6 anomaly representing 100 year change in A) near-surface wind speed, B) mixed layer depth, C) primary productivity, D) irradiance limitation of phytoplankton, E) incidental photosynthetically active radiation (IPAR), F) iron limitation of phytoplankton. Changes are calculated from an ensemble of CMIP6 models, comparing a historical (1985-2015) average against 2100 under the SSP5-8.5 climate scenario. Details of ensemble members are given in Table S1. *Units in panels E and F are arbitrary ratios of growth under environmental irradiance or iron concentrations against potential growth under unlimited irradiance or iron concentrations.



155 3 Changing biogeochemistry of the Southern Ocean

3.1 Micronutrient supply and uptake

Iron acts as the primary limiting nutrient across the Southern Ocean (Watson et al., 2000, de Baar et al., 1995), due to supply limitation from low atmospheric inputs and distance from terrigenous sources (Boyd and Ellwood, 2010). Around the Antarctic coast, iron concentrations are set by processes including the resuspension of shelf sediments (Blain et al., 2001), melting of
 160 sea ice (Lannuzel et al., 2016) and potential transformation of iron into more labile forms by glacial retreat, as seen in the Arctic (Laufer-Meiser et al., 2021). While supplies of ice-rafter iron in the open ocean (Hopwood et al., 2019, Raiswell et al., 2008) may be expected to increase with warming, this increase in supply appears to be offset by an increased total iron demand to support enhanced phytoplankton growth, stimulated by increased light availability (Figure 2a). Much of the Southern Ocean relies on regenerated iron (i.e. that released from remineralisation), denoted by the fe ratio (uptake of new iron divided by
 165 uptake of new and recycled iron), with a value of ~ 0.1 in HNLC waters beyond the coastal zone (Bowie et al., 2009, Boyd et al., 2005). Therefore, the class of phytoplankton present in any region will act as a determinant on iron resupply since physiological differences between phytoplankton species will influence sinking rate, cellular iron content, and the rate of remineralisation. While iron is known to be an essential micronutrient for phytoplankton, there are large uncertainties in our understanding of iron cycling, particularly in parameterising rates of ligand adsorption and desorption (Tagliabue et al., 2016).
 170 Shifts in primary producers may influence rates of iron ligand sorption and desorption by modulating the size and molecular composition of the organic ligand binding pool.

Iron is utilised by phytoplankton in key metabolic processes, including the production of electron transfer proteins used within photosystems, for the reduction of carbon dioxide, sulphate and nitrate, and synthesis of chlorophyll (Street and Paytan, 2005).
 175 Other micronutrients beyond iron, predominantly manganese, have been shown to play an important role in controlling oxidative stress by catalysing antioxidant production in some diatom species (McCain et al., 2021), explaining the observed phenomena of iron-manganese co-limitation in the Southern Ocean (Browning et al., 2021, Pausch et al., 2019, Balaguer et al., 2022). Despite the importance of micronutrients such as manganese in controlling phytoplankton growth, particularly during seasonal transitions (Browning et al., 2021), only iron is considered in ESMs, due at least partially to the lack of
 180 observational data to underpin distribution modelling of other micronutrients. Future work should continue to develop our understanding of the metabolic role of other micronutrients and additionally consider the extent to which diversity exists in micronutrient demand between Southern Ocean phytoplankton species.



3.2 Macronutrient supply and uptake

185 Nitrogen species, silicic acid (DSi) and phosphate are essential for the growth and survival of diatoms, with nitrate and phosphate also being required by all other phytoplankton classes for cellular metabolism. The ratio of utilisation between nitrogen (N) and phosphorus (P) deviates from the Redfield (1958) ratio of 16:1 across the Southern Ocean according to changes in community composition (Henley et al., 2020, Weber and Deutsch, 2010). Unlike much of the global ocean (Moore et al., 2013), high rates of macronutrient supply from the Circumpolar Deep Water (CDW) prevent widespread N or P
 190 limitation in the Southern Ocean except in periods of intense summer growth in high-productivity coastal regions (Henley et al., 2017). Although projections indicate an increase in biomass across such regions (Figure 3), models do not show any increases in nitrate limitation over the remainder of the century, suggesting that iron and light will continue to be the primary constraints on productivity.

195 While macronutrients are not usually limiting to Southern Ocean phytoplankton, growth of diatom communities, particularly around high productivity coastal and island zones (supported by lateral iron advection) (Robinson et al., 2016), is likely to place an increased demand on DSi availability (Table 1). The relationship between Si and N is denoted as Si^* ($[Si(OH)_4] - [NO_3^-]$) (Sarmiento et al., 2004), with high Si^* values (> 25) indicating plentiful DSi availability that supports diatom growth, while low values (< 10) suggest conditions which favour non-silicifying phytoplankton, such as the smaller cryptophytes and
 200 haptophytes. Si^* is highest in the Antarctic zone (Henley et al., 2020) because of silica input from upwelling of CDW, but remains spatially heterogeneous within this region (Table 1). Si^* is consistently high in the Weddell Sea, while across the WAP and Ross, Amundsen and Bellingshausen Seas there is a moderate mean Si^* with large variability, and the Indian Sector has a substantially lower DSi availability. The relative availability of DSi is generally reflected in projected patterns of chlorophyll over the current century; large increases in chlorophyll are projected in the Weddell Sea with much less growth in
 205 the Indian sector (Figure 3), suggesting that Si limitation of diatoms will continue to constrain overall patterns of phytoplankton productivity.

The impact of climate change on DSi supply to the surface is difficult to evaluate because it is dependent on the competing stratification effects from strengthening upwelling and an increase in freshening. Export of DSi from the surface and
 210 remineralisation at depth additionally act as important controls on supply; more rapid remineralisation of organic matter in the upper ocean as warming increases the microbial respiration rate may increase demand for OM and release nutrients at shallower depths. Such an effect would also reduce both the rain rate and the quality of OM available at the seafloor for long-term carbon sequestration. Increases in viral lysis under warmer ocean conditions will also influence nutrient dynamics either by retaining nutrients at the surface through shallow cell lysis and remineralisation (Biggs et al., 2021), or by shuttling nutrients to depth
 215 when infection of iron-limited cells delays death and increases sinking (Kranzler et al., 2021). Efforts to better define the nutrient budgets, particularly in low sea ice years, across different sectors of the Southern Ocean, as well as understanding the



changing nutrient demands of phytoplankton will be essential for determining future trends in nutrient limitation (Henley et al., 2019).

3.3 Ocean acidification

220 Across all regions of the Southern Ocean, continued uptake of anthropogenic CO₂ is expected to elicit a decrease in pH of
 ~0.45 units south of 55°S (Table 1). While ocean acidification (OA) is typically considered to have the greatest effect on
 marine calcifiers through impacts on the production and dissolution of calcium carbonate (Figuerola et al., 2021), OA is also
 likely to impact diatom (Petrou et al., 2019), picophytoplankton (Tortell et al., 2008) and krill (Kawaguchi et al., 2013)
 populations which form the base of Southern Ocean ecosystems. Unravelling the specific impact of OA on phytoplankton
 225 species is complex due to the fact that OA often acts on phytoplankton indirectly, for example Petrou et al. (2019) showed that
 acidification reduces silicification of diatoms, likely reducing sinking capacity and increasing rates of remineralisation in the
 upper ocean, in turn weakening ocean carbon drawdown and acting as a positive feedback on the carbon cycle. Additionally,
 in their experiment, Petrou et al. (2019)'s modelled OA conditions resulted in reduced photosynthetic capability showing a
 direct effect of OA on diatom physiology. OA has further been shown to elicit change in biogeochemical parameters such as
 230 carbon uptake, with reduced carbon uptake by Ross Sea assemblages under elevated pCO₂ concentrations (Tortell et al., 2008).
 However, this effect may be offset or reversed by overall increases in productivity linked to shifts towards larger diatom
 species under increasing OA (Tortell et al., 2008). Alternatively, some studies have shown that smaller size classes (<20 µm)
 are more successful at higher pCO₂ values (Hancock et al., 2018) or that total productivity may decrease under OA (Westwood
 et al., 2018).

235 The ability for OA to promote increased productivity has been linked to the availability of Fe and species shift effects (Hoppe
 et al., 2013), whereby species with the greatest capacity to tolerate variability in pH (e.g. *Fragilariopsis cylindrus*, *Chaetoceros*
sp.) outcompete less tolerant, typically faster growing species such as *Pseudo-nitzschia* (Hoppe et al., 2013, Tortell et al.,
 2008). At an ecosystem level, OA has been shown to reduce the lipid content, and therefore nutritional value of large
 240 phytoplankton to zooplankton grazers (Duncan et al., 2022). While protein and lipid content of small taxa may increase, their
 poor utilisation due to size will likely lead to rapid remineralisation in the surface layer, further reducing energy transfer to
 higher trophic levels. The impacts of OA on phytoplankton ecology and physiology are not currently reflected in CMIP6-class
 ESMs (Seferian et al., 2020), but are likely to become increasingly important in the future as the rate of OA increases. To
 support this development, empirical research will need to focus on further disentangling the indirect influences of OA on
 245 phytoplankton on a species-specific basis, particularly for picophytoplankton which are less well studied in incubation
 experiments but are expected to become more abundant. The interspecies divergence of responses to OA, and differential
 responses at different pCO₂ thresholds suggests that OA-induced ecophysiological change is likely to remain too complex to
 model accurately at the basin or global scale.

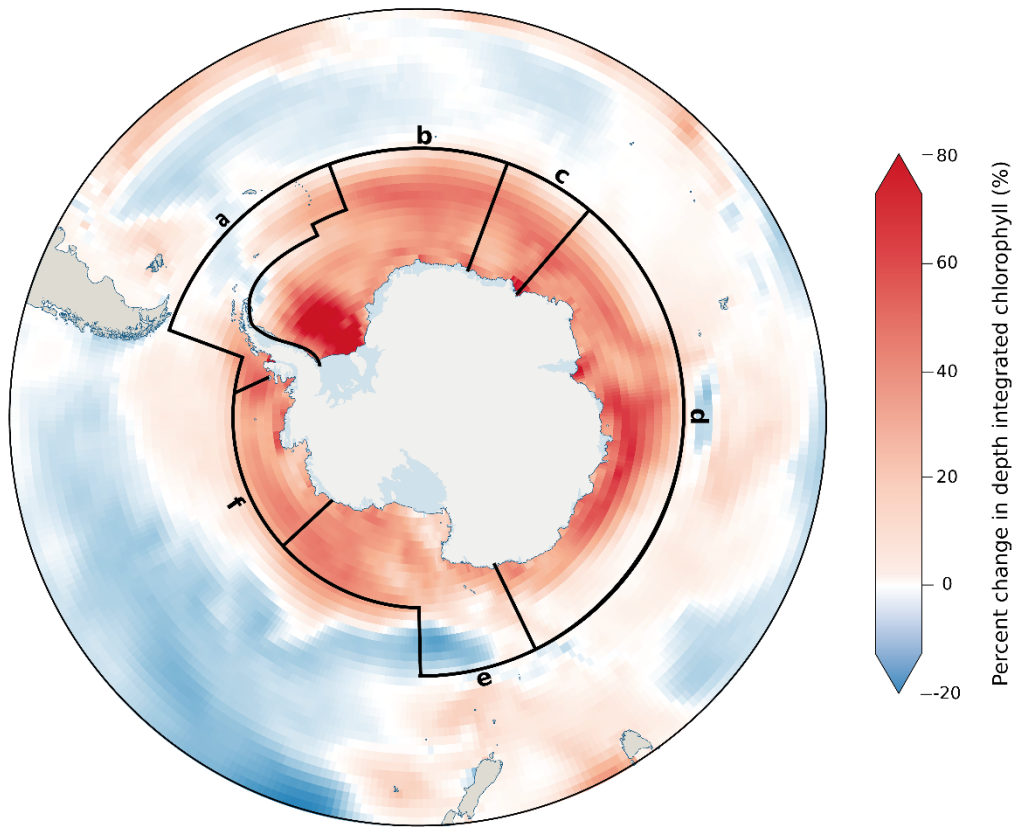


Figure 3: Change in depth-integrated chlorophyll (0-500 m) from all phytoplankton, displayed as the percentage change between the annual historical average (1985-2015) and projected values for 2100. Values shown are multi-model means of the models listed in Table S1. Spatial boundaries show the Southern Ocean Observing System (SOOS) regions south of 55°S, which are defined in Table 1.

Table 1: Biogeochemical parameters of the Southern Ocean Observing System regions. SOOS regional working groups (as defined at: www.soos.aq/activities/rwg) indicated on Figure 3; section C is an overlap section of sections B and D. Data shown are: Si* ([Si(OH)₄]–[NO₃[–]]) values determined from World Ocean Atlas 2018 data, multi-model means of pH and temperature anomalies based on comparisons between the mean annual historical value (1985-2015) and projected values for 2100 under SSP5-8.5 for a CMIP6 ensemble (detailed in Table S1). Values in brackets are standard deviations.

Section	SOOS Region	Si*	Δ pH	Δ Temperature (°C)
A	West Antarctic Peninsula & Scotia Arc	17.24 (17.82)	-0.45 (0.01)	1.99 (0.40)
B	Weddell Sea & Dronning Maud Land (WSDML)	37.37 (9.70)	-0.43 (0.01)	1.61 (0.49)



C	SOIS/WSDML	23.16 (6.67)	-0.44 (0.01)	2.09 (0.37)
D	Southern Ocean Indian Sector (SOIS)	4.71 (3.72)	-0.44 (0.01)	1.99 (0.52)
E	Ross Sea	19.82 (18.49)	-0.42 (0.02)	1.16 (0.47)
F	Amundsen and Bellingshausen Seas	17.59 (14.02)	-0.45 (0.01)	1.97 (0.32)

4 Primary production and representation in CMIP6

4.1 Productivity parameters

Across CMIP6 models, primary productivity is expressed as the depth-integrated production of particulate organic carbon (POC, as intpp) and the biomass of phytoplankton expressed as chlorophyll (Chl). While chlorophyll and POC production both represent phytoplankton related parameters, they are independent variables, with the relationship between the two (Chl/C ratio) being spatially and seasonally variable across the Southern Ocean (Thomalla et al., 2017). Additionally, the biological origin of POC in CMIP6 differs based on the ecosystem complexity of different models, for example between those models which include bacteria and those which do not (Table 1 in Seferian et al. (2020)). Although Chl/C ratios are fundamental in coupling marine ecosystems to the biological carbon pump, estimates of Chl:C carry substantial uncertainties, particularly when sensed remotely (Baldry et al., 2020). Resolving these uncertainties, stemming from non-photochemical quenching, interference from non-phytoplankton POC, and differences in intracellular chlorophyll contents between phytoplankton species (Baldry et al., 2020) will improve our ability to utilise satellite- and Argo-derived ocean chlorophyll data and improve ecosystem-carbon coupling across ocean biogeochemical models.

4.2 Phytoplankton classes

A major difference in the representation of productivity between CMIP6 models is the extent to which they consider different classes of phytoplankton. Diatoms (>20 µm) and pico/nano phytoplankton (predominantly cryptophytes and haptophytes) represent the vast majority of productivity across all latitudes of the Southern Ocean. Diatoms are a significant contributor to primary production and carbon export, accounting for ~40% of global marine primary production and POC exported to depth in the ocean (Treguer and De La Rocha, 2013, Jin et al., 2006). Diazotrophs (nitrogen-fixing phytoplankton) are present in small numbers, usually only in environmental niches, due to the excess supply of nitrogen across the Southern Ocean (Nissen



et al., 2018). Calcifiers, mostly coccolithophores, inhabit waters north of 60°S where there is a strong supply of light but low Si, high Fe conditions, preventing the growth of diatoms (Charalampopoulou et al., 2016). Of ~90 distinct models (excluding multiple resolutions of the same model) included in CMIP6, 11 specifically include diatoms under future warming conditions and only three of these additionally consider picophytoplankton. Two CMIP6 models (GFDL-ESM4 and CESM2-WACCM) showed substantial differences in productivity projections south of 65° (Figure 3). While GFDL-ESM4 projects that in this region diatoms account for the majority (55%) of the change in productivity under SSP5-8.5 (Figure 4a,b), diatoms represent only 26% of productivity in CESM2-WACCM, while picophytoplankton form the major (74%) phytoplankton group (Figure 4e,d). Additionally, the GFDL model indicates that increased productivity is driven by increases in both diatoms and picophytoplankton, representing a simultaneous growth scenario while CESM2-WACCM favours a replacement mechanism with diatoms decreasing as picophytoplankton populations grow (Figure 4c,f. This demonstrates that further parameterisation is itself not a solution to integrating phytoplankton ecophysiology while fundamental gaps remain in the empirical understanding of mutualism and resource limitation between phytoplankton classes. Here, we argue for an expansion in the biological understanding of phytoplankton responses to climate, moving beyond single species to community based experiments alongside a shift towards holistic marine ecosystem models which account for an expanded range of biological interactions including phytoplankton-zooplankton predation and bacterially-driven mixotrophic effects (Seferian et al., 2020). Trait-based approaches have been explored as a means of modelling phytoplankton community composition, distinguishing functional groups based on life histories, morphology and physiology (Litchman and Klausmeier, 2008). Ocean biological sampling has some of the lowest coverage in the Southern Ocean (Sunagawa et al., 2020). The lack of coverage compounds uncertainty associate with the unique phytoplankton traits in the region, for example the lack of nitrogen fixation and cryobiological traits such as ice-binding, resulting in a weak overall understanding of Southern Ocean phytoplankton biology. Expansion of ecosystem observing at the metagenomics level offers a promising opportunity to expand our knowledge of traits and trade-offs in Southern Ocean phytoplankton communities, facilitating their integration into ocean climate models.

4.3 Ecological dynamics and ecophysiology

In a changing ocean, phytoplankton will succeed where they have the greatest biological plasticity, for example the ability to photo-acclimate rapidly (Arrigo et al., 2010) or scavenge and utilise a diverse range of micronutrients. The physiological properties of any individual species ultimately determines their ability to survive in a particular region at a particular time under ever changing climate-driven conditions. Subsequently, species ecology determines the abundance and temporal extent with which a species can exist or compete in a particular region. As the warming of the climate continues to bring about an earlier retreat of sea ice, growth seasons are expected to lengthen, altering the temporal dynamics of species progression (Moreau et al., 2015). In the coastal zone of the Southern Ocean, changes in light appear to be the main influence on productivity with decreased irradiance limitation stimulating picophytoplankton growth to a greater extent than diatoms



(Figure 4 j,k,m,n); meanwhile iron limitation shows little correlation with productivity changes in this region (Figure 4 g,h),
325 likely because of replete iron supplies from coastal upwelling.

While few CMIP6 models account for multiple phytoplankton groups, among the models that do contain different classes there is a vast variability between the magnitudes of change they project over the century (Figure 5). Accurate representation of the seasonal progression of phytoplankton in models is important because pre-bloom community composition is closely linked to
330 the eventual extent of the summer bloom (Biggs et al., 2019). In addition, the magnitude of early spring and late autumn productivity is likely to constrain the length of the grazing season for zooplankton and food supply to benthic communities (Constable et al., 2014). Early season productivity is known to occur in under-ice algal communities (Lizotte, 2001), and potentially by phytoplankton (Ardyna et al., 2020), which makes both in situ and satellite measurements of this productivity even more complex due to the physical and reflective barrier of sea ice. Sensing early season under-ice productivity represents
335 a strategic priority because of the likely effect of climate change in producing shifts in seasonal timing related to earlier sea ice retreat, potentially shifting the summer bloom timing or amplitude. The Intergovernmental Panel on Climate Change (IPCC) notes that lack of data relating to snow cover on Antarctic sea ice limits mass balance estimations, and therefore results in a “very limited predictive skill” for ice-dependent ecosystems (Meredith et al., 2019). Subsequently, expanded understanding of productivity onset and early season physiology and phenology will allow for better integration of
340 phytoplankton ecophysiology in ESMs, which has been identified as a step change required to improve biogeochemical processes and climate projections in future generations of CMIP (Seferian et al., 2020).

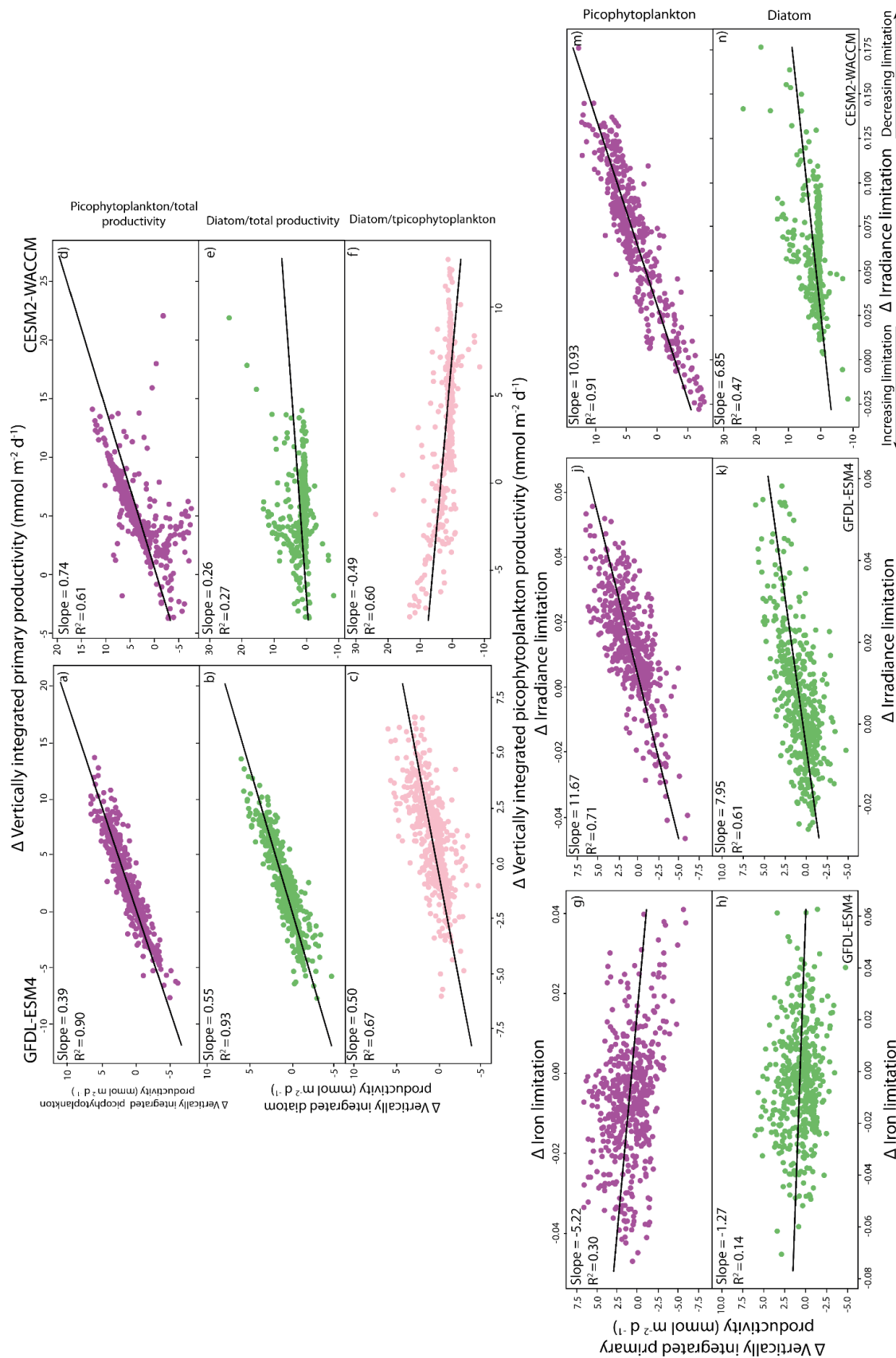


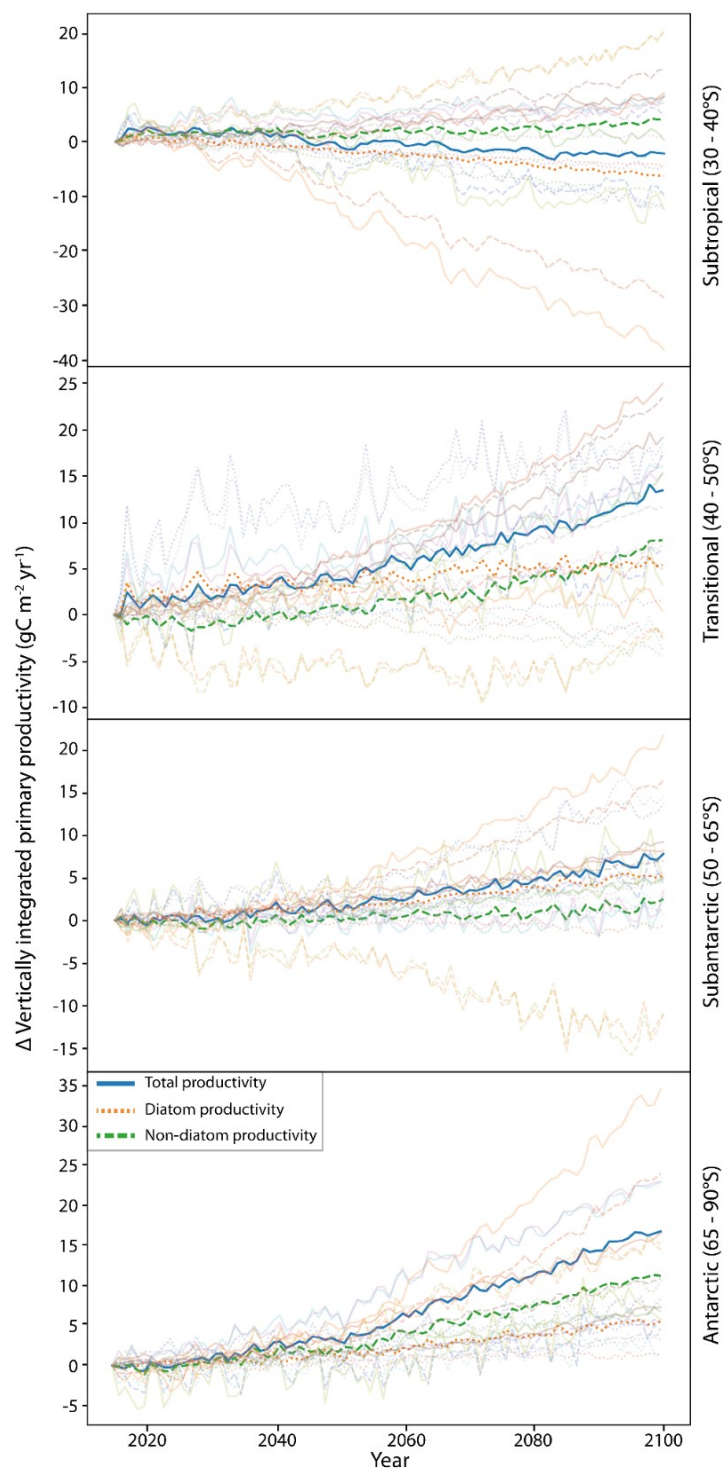
Figure 4: Evaluation of GFDL-ESM4 and CESM2-WACCM models using an anomaly between 2100 (SSP5-8.5) and a historical average (1985-2015). Linear regression between change in total productivity and picophytoplankton productivity for GFDL-ESM4 (A) and CESM2-WACCM (D). Linear regression between change in total productivity and diatom productivity for GFDL-ESM4 (B) and CESM2-WACCM (E). Linear regression between change in picophytoplankton productivity and diatom productivity for GFDL-ESM4 (C) and CESM2-WACCM (F). Change in iron limitation with picophytoplankton (G) and diatom (H) productivity for GFDL-ESM4. Change in irradiance limitation with picophytoplankton (I) and diatom (N) productivity for CESM2-WACCM.



348 5. CMIP6 Projections

349 From those models that do distinguish between at least diatoms and other phytoplankton (Table S1) we are able
 350 to examine projected changes in community composition over the 21st century under a continued warming
 351 scenario (SSP5-8.5) (Figure 5). In CMIP5, Leung et al. (2015) found a latitudinally banded response of
 352 phytoplankton to continued warming, driven by the bottom-up dynamics of nitrate, iron and light limitation. From
 353 this, we applied the same latitudinal bands to our analysis of the changes in whole community, diatom and non-
 354 diatom productivity across CMIP6. Our whole community projections agree with the trends shown by Leung et
 355 al. (2015), showing a southward movement of phytoplankton productivity, increasing average total productivity
 356 south of 40°S, with the strongest increases in the Transitional (40-50°S; 14 g C m⁻² yr⁻¹) and Antarctic (or coastal)
 357 zones (65-90°S; 16 g C m⁻² yr⁻¹). In relative terms, this reflects a ~10% increase in total productivity over the
 358 SSP5-8.5 run (2015-2100) for both the Transitional and Subantarctic zones, with a ~40% increase in productivity
 359 for the Antarctic zone (Figure S4). An ensemble mean shows no overall change in productivity across the
 360 Subtropics, however individual models show the widest degree of divergence in this region, indicating a large
 361 amount of uncertainty in the directionality of productivity change.

362
 363 In the Transitional zone, we initially see growth in the proportion of productivity linked to diatoms at a rate greater
 364 than that of non-diatoms until 2085, likely stimulated by increased iron availability (Figure 2f). Between 2085
 365 and 2100, this trend is inverse with non-diatom productivity overtaking diatoms, potentially reflecting the increase
 366 in light limitation across the region (Figure 2d) which favours smaller, less light-dependent, phytoplankton. In the
 367 Subantarctic (50-65°S), despite a large projected increase in light availability (Figure 2e), models project only a
 368 minor increase in productivity driven by a small amount of diatom growth, suggesting that growth of both diatom
 369 and non-diatom species remains largely iron-limited in this region. The coastal zone shows the greatest degree of
 370 change in phytoplankton growth; by 2100 the non-diatom population will have increased at twice the rate of
 371 diatoms. The continued increase in all phytoplankton classes can be attributed to the increased iron availability
 372 (Figure 2f), with the success of non-diatoms reflecting the increase in light limitation (Figure 2e). While CMIP6
 373 models do not explicitly consider phytoplankton size, the shift from diatoms to, typically smaller, non-diatom
 374 species is consistent with more advanced ecological models such as DARWIN which predict a decrease in the
 375 slope of the phytoplankton size spectrum, albeit over a greater area of the Southern Ocean than shown in CMIP6
 376 (Henson et al., 2021). Despite the clear differences between latitudinal bands, spatial heterogeneity continues to
 377 exist within these zones, particularly for the coastal zone where the majority of change occurs on the WAP and
 378 Weddell Sea (Figure 3), reflecting the disproportionately high DSi supply in these regions (Table 1). Resolving
 379 spatial heterogeneity of phytoplankton in global-scale models such as those in CMIP6 is likely to require an
 380 increased reliance on, and integration with, regional-scale modelling (Person et al., 2018). The rapid increase of
 381 non-diatom species around the coast is in agreement with studies describing declining large diatom (>20 µm)
 382 abundances (Wright et al., 2010, Kang et al., 2001, Pearce et al., 2011); however, while it is true that diatoms are
 383 projected to decrease as a proportion of the community, diatom-derived carbon production is still projected to
 384 increase under continued warming, suggesting that the coastal biological carbon pump may be less threatened by
 385 this shift in community composition than previously thought.



386

387 **Figure 5: Changes in productivity ($\text{g C m}^{-2} \text{yr}^{-1}$) and the contribution of different phytoplankton classes to productivity, 2015–2100.**
 388 **The anomaly in CMIP6 model productivity projections (as POC production) compared to 2015 for SSP5-8.5 conditions across 4**
 389 **latitudinal bands of the Southern Ocean, per Leung et al. (2015). Thick lines represent multi-model means of total productivity**
 390 **(intpp), diatom productivity (intppdiat) and non-diatom productivity (intpp-intppdiat). Light lines represent the individual model**
 391 **projections. Details of the models assessed are given in Table S1.**



6. Global implications of Southern Ocean productivity shifts

The impact of climate change on phytoplankton has the potential to restructure ecosystems of the Southern Ocean, with wider consequences for global productivity and climate. Earth System Models are essential for understanding the impact of regional scale processes on global climate, and CMIP6 represents our best understanding of future climate change under continued warming. Despite this, key processes that will impact phytoplankton are absent from most models, resulting in large uncertainty for all phytoplankton classes across all zones of the Southern Ocean. While computational limits prevent ecological parameterisation on the scale of purely biological models, the lack of empirical understanding prevents inclusion of some key physical and biological processes in these models.

We have identified changes in nutrient upwelling (both upwelling strength and concentration of nutrients at the surface), directionality of mixed layer depth change, mutualism and resource competition between phytoplankton classes, and eddy strengthening as key processes which require improved representation in future generations of climate models. In some cases this is because of lack of monitoring (nutrients, eddies), while the relatively recent reversal in Antarctic sea ice trends means it is not yet possible to determine the scale of coastal MLD change from excess freshwater injection. Additionally, there is a need for ongoing monitoring of phytoplankton functional group composition, particularly in areas of the greatest projected change (WAP and Weddell Sea), ideally capturing temporally-resolved new productivity in the early spring. Crucially, to then link ecological shifts to global climate, the diversity in fate of phytoplankton species represents a substantial knowledge gap required for improving the predictive ability of ESMs. Finally, there is a need for process-based studies including a diverse set of phytoplankton species to further our understanding of fundamental life cycle processes and ecological-biogeochemical coupling. A focus towards defining variability in phytoplankton buoyancy, carbon uptake and release as DOM, grazing by zooplankton, interactions with viruses, macronutrient stoichiometry and micronutrient utilisation will enhance our core understanding of Southern Ocean phytoplankton dynamics in a changing climate.

7. Methods

Model and observational data for the Southern Ocean were collected and visualised to determine a) the physical and biogeochemical changes which force or result from shifts in productivity, b) the extent of primary productivity shifts in CMIP6.

7.1 CMIP projections

Model output was obtained from the Climate Model Intercomparison Project Phase 6 (CMIP6) data server via pangeo.io using the XMIP package in Python on JASMIN (Lawrence et al., 2013). Ensemble members for each parameter were chosen based on their availability for historical and SSP5-8.5 (ScenarioMIP) data (O'Neill et al., 2016). Some models that were available were not included in the analysis due to compatibility issues where significant differences in data structure existed. All



variables were initially selected with monthly frequency except for surface winds where data were initially obtained daily; subsequently, annual means were generated for all parameters.

425 Model data were processed in python to apply the desired analysis (e.g. annual average, annual maximum) and then further averaged over residual variables (e.g. member_id). Subsequently, all models were re-gridded to a common grid using XESMF before being averaged to create multi model means.

For spatial plotting, data were exported from python and a common area of the grid selected in Climate Data Operator. The
430 subset data were projected to the Antarctic Polar Stereographic (3031) coordinate reference system in ArcGIS pro. Finally, spatial plots were visualised in QGIS using the Quantarctica (Matsuoka et al., 2021) package, with post processing using SAGA and GDAL tools to remove imperfections in grid alignment through interpolation. The full range of CMIP6 parameters accessed, models evaluated and transformations applied are detailed in Table S1. All code to extract, analyse and visualise CMIP6 data is available open access.

435

7.2 Regional data

Si* across the Southern Ocean was mapped from the World Ocean Atlas 2018 data product (Garcia et al., 2019). Annually averaged data for nitrate and silicic acid were exported at a 1x1 degree resolution and subtracted from one another to produce Si*. The output was visualised in QGIS using Quantarctica base layers (Matsuoka et al., 2021). For Si*, pH and temperature
440 values by SOOS area, SOOS regions south of 55° were drawn as mask layers and subset using the zonal statistics function in QGIS.

Acknowledgements

BJF is supported by a NERC Doctoral Training Partnership Grant (NE/S007407/1). This work used JASMIN, the UK's collaborative data analysis environment (<https://jasmin.ac.uk>). SFH would like to acknowledge support from NERC
445 (NE/K010034/1).

Data availability

CMIP6 data were obtained through, and are freely available at panego.io. Specific models and parameters extracted for each analysis are listed in Supplementary Table 1. Code for CMIP6 data analysis are available at: *(zenodo link to be later inserted,*
450 *available to reviewers as attached zip file).*



Author contributions

BJF, SFH, MPM and AJP devised the concept for the paper and contributed towards initial drafting and editing. BJF performed model analysis and produced the figures. OS and KB provided input on conceptual design and edited previous versions of the manuscript.

455

Competing interests

The authors declare no competing interests.

References

- 460 Ardyna, M., Mundy, C. J., Mayot, N., Matthes, L. C., Oziel, L., Horvat, C., Leu, E., Assmy, P., Hill, V., Matrai, P. A., Gale, M., Melnikov, I. A. & Arrigo, K. R. 2020. Under-Ice Phytoplankton Blooms: Shedding Light on the “Invisible” Part of Arctic Primary Production. *Frontiers in Marine Science*, 7.
- Arrigo, K. R., Mills, M. M., Kropuenske, L. R., Van Dijken, G. L., Alderkamp, A.-C. & Robinson, D. H. 2010. Photophysiology in Two Major Southern Ocean Phytoplankton Taxa: Photosynthesis and Growth of *Phaeocystis* antarctica and *Fragilariopsis cylindrus* under Different Irradiance Levels. *Integrative and Comparative Biology*, 50, 465 950-966.
- Balaguer, J., Koch, F., Hassler, C. & Trimborn, S. 2022. Iron and manganese co-limit the growth of two phytoplankton groups dominant at two locations of the Drake Passage. *Communications Biology*, 5, 207.
- Baldry, K., Strutton, P. G., Hill, N. A. & Boyd, P. W. 2020. Subsurface Chlorophyll-a Maxima in the Southern Ocean. *Frontiers in Marine Science*, 7.
- 470 Ballerini, T., Hofmann, E. E., Ainley, D. G., Daly, K., Marrari, M., Ribic, C. A., Smith Jr, W. O. & Steele, J. H. 2014. Productivity and linkages of the food web of the southern region of the western Antarctic Peninsula continental shelf. *Progress in Oceanography*, 122, 10-29.
- Biggs, T. E. G., Alvarez-Fernandez, S., Evans, C., Mojica, K. D. A., Rozema, P. D., Venables, H. J., Pond, D. W. & Brussaard, C. P. D. 2019. Antarctic phytoplankton community composition and size structure: importance of ice 475 type and temperature as regulatory factors. *Polar Biology*, 42, 1997-2015.
- Biggs, T. E. G., Huisman, J. & Brussaard, C. P. D. 2021. Viral lysis modifies seasonal phytoplankton dynamics and carbon flow in the Southern Ocean. *The ISME Journal*.
- Bindoff, N. L., Cheung, W. W., Kairo, J. G., Arístegui, J., Guinder, V. A., Hallberg, R., Hilmi, N. J. M., Jiao, N., Karim, M. S. & Levin, L. 2019. Changing ocean, marine ecosystems, and dependent communities. *IPCC special report on the 480 ocean and cryosphere in a changing climate*, 477-587.



- Blain, S., Tréguer, P., Belviso, S., Bucciarelli, E., Denis, M., Desabre, S., Fiala, M., Jézéquel, V. M., Le Fèvre, J. & Mayzaud, P. 2001. A biogeochemical study of the island mass effect in the context of the iron hypothesis: Kerguelen Islands, Southern Ocean. *Deep Sea Research Part I: Oceanographic Research Papers*, 48, 163-187.
- Bowie, A. R., Lannuzel, D., Remenyi, T. A., Wagener, T., Lam, P. J., Boyd, P. W., Guieu, C., Townsend, A. T. & Trull, T. W. 2009. Biogeochemical iron budgets of the Southern Ocean south of Australia: Decoupling of iron and nutrient cycles in the subantarctic zone by the summertime supply. *Global Biogeochemical Cycles*, 23.
- Boyd, P. W. & Ellwood, M. J. 2010. The biogeochemical cycle of iron in the ocean. *Nature Geoscience*, 3, 675-682.
- Boyd, P. W., Law, C. S., Hutchins, D. A., Abraham, E. R., Croot, P. L., Ellwood, M., Frew, R. D., Hadfield, M., Hall, J., Handy, S., Hare, C., Higgins, J., Hill, P., Hunter, K. A., Leblanc, K., Maldonado, M. T., McKay, R. M., Mioni, C., Oliver, M., Pickmere, S., Pinkerton, M., Safi, K., Sander, S., Sanudo-Wilhelmy, S. A., Smith, M., Strzepek, R., Tovar-Sanchez, A. & Wilhelm, S. W. 2005. FeCycle: Attempting an iron biogeochemical budget from a mesoscale SF6 tracer experiment in unperturbed low iron waters. *Global Biogeochemical Cycles*, 19.
- Browning, T. J., Achterberg, E. P., Engel, A. & Mawji, E. 2021. Manganese co-limitation of phytoplankton growth and major nutrient drawdown in the Southern Ocean. *Nature communications*, 12, 1-9.
- Caldeira, K. & Duffy, P. B. 2000. The Role of the Southern Ocean in Uptake and Storage of Anthropogenic Carbon Dioxide. *Science*, 287, 620-622.
- Canadell, J. G., Monteiro, P. M., Costa, M. H., Da Cunha, L. C., Cox, P. M., Alexey, V., Henson, S., Ishii, M., Jaccard, S., Koven, C., Lohila, A., Patra, P. K., Piao, S., Rogelj, J., Syampungani, S., Zaehle, S. & Zickfeld, K. 2021. Global carbon and other Biogeochemical Cycles and Feedbacks. In: MASSON-DELMOTTE, V., P. ZHAI, A. PIRANI, S.L. CONNORS, C. PÉAN, S. BERGER, N. CAUD, Y. CHEN, L. GOLDFARB, M.I. GOMIS, M. HUANG, K. LEITZELL, E. LONNOY, J.B.R. MATTHEWS, T.K. MAYCOCK, T. WATERFIELD, O. YELEKÇİ, R. YU & ZHOU, B. (eds.) *Climate Change 2021: The Physical Science Basis. Contribution of Working Group I to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change* In Press: Cambridge University Press.
- Carranza, M. M. & Gille, S. T. 2015. Southern Ocean wind-driven entrainment enhances satellite chlorophyll-a through the summer. *Journal of Geophysical Research: Oceans*, 120, 304-323.
- Charalampopoulou, A., Poulton, A. J., Bakker, D. C. E., Lucas, M. I., Stinchcombe, M. C. & Tyrrell, T. 2016. Environmental drivers of coccolithophore abundance and calcification across Drake Passage (Southern Ocean). *Biogeosciences*, 13, 5917-5935.
- Constable, A. J., Melbourne-Thomas, J., Corney, S. P., Arrigo, K. R., Barbraud, C., Barnes, D. K. A., Bindoff, N. L., Boyd, P. W., Brandt, A., Costa, D. P., Davidson, A. T., Ducklow, H. W., Emmerson, L., Fukuchi, M., Gutt, J., Hindell, M. A., Hofmann, E. E., Hosie, G. W., Iida, T., Jacob, S., Johnston, N. M., Kawaguchi, S., Kokubun, N., Koubbi, P., Lea, M.-A., Makhado, A., Massom, R. A., Meiners, K., Meredith, M. P., Murphy, E. J., Nicol, S., Reid, K., Richerson, K., Riddle, M. J., Rintoul, S. R., Smith Jr, W. O., Southwell, C., Stark, J. S., Sumner, M., Swadling, K. M., Takahashi, K. T., Trathan, P. N., Welsford, D. C., Weimerskirch, H., Westwood, K. J., Wienecke, B. C., Wolf-



- 515 Gladrow, D., Wright, S. W., Xavier, J. C. & Ziegler, P. 2014. Climate change and Southern Ocean ecosystems I: how changes in physical habitats directly affect marine biota. *Global Change Biology*, 20, 3004-3025.
- De Baar, H. J. W., De Jong, J. T. M., Bakker, D. C. E., Löscher, B. M., Veth, C., Bathmann, U. & Smetacek, V. 1995. Importance of iron for plankton blooms and carbon dioxide drawdown in the Southern Ocean. *Nature*, 373, 412-415.
- 520 Devries, T. 2014. The oceanic anthropogenic CO₂ sink: Storage, air-sea fluxes, and transports over the industrial era. *Global Biogeochemical Cycles*, 28, 631-647.
- Ducklow, H. W., Baker, K., Martinson, D. G., Quetin, L. B., Ross, R. M., Smith, R. C., Stammerjohn, S. E., Vernet, M. & Fraser, W. 2007. Marine pelagic ecosystems: the West Antarctic Peninsula. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 362, 67-94.
- 525 Duncan, R. J., Nielsen, D. A., Sheehan, C. E., Deppeler, S., Hancock, A. M., Schulz, K. G., Davidson, A. T. & Petrou, K. 2022. Ocean acidification alters the nutritional value of Antarctic diatoms. *New Phytologist*, 233, 1813-1827.
- Ellwood, M. J., Strzepek, R. F., Strutton, P. G., Trull, T. W., Fourquez, M. & Boyd, P. W. 2020. Distinct iron cycling in a Southern Ocean eddy. *Nature Communications*, 11, 825.
- Figuerola, B., Hancock, A. M., Bax, N., Cummings, V. J., Downey, R., Griffiths, H. J., Smith, J. & Stark, J. S. 2021. A
 530 Review and Meta-Analysis of Potential Impacts of Ocean Acidification on Marine Calcifiers From the Southern Ocean. *Frontiers in Marine Science*, 8.
- Fogwill, C. J., Phipps, S. J., Turney, C. S. M. & Golledge, N. R. 2015. Sensitivity of the Southern Ocean to enhanced regional Antarctic ice sheet meltwater input. *Earth's Future*, 3, 317-329.
- Friedlingstein, P., Jones, M. W., O'sullivan, M., Andrew, R. M., Bakker, D. C. E., Hauck, J., Le Quéré, C., Peters, G. P.,
 535 Peters, W., Pongratz, J., Sitch, S., Canadell, J. G., Ciais, P., Jackson, R. B., Alin, S. R., Anthoni, P., Bates, N. R., Becker, M., Bellouin, N., Bopp, L., Chau, T. T. T., Chevallier, F., Chini, L. P., Cronin, M., Currie, K. I., Decharme, B., Djeutchouang, L. M., Dou, X., Evans, W., Feely, R. A., Feng, L., Gasser, T., Gilfillan, D., Gkritzalis, T., Grassi, G., Gregor, L., Gruber, N., Gürses, Ö., Harris, I., Houghton, R. A., Hurtt, G. C., Iida, Y., Ilyina, T., Luijkx, I. T., Jain, A., Jones, S. D., Kato, E., Kennedy, D., Klein Goldewijk, K., Knauer, J., Korsbakken, J. I., Körtzinger, A.,
 540 Landschützer, P., Lauvset, S. K., Lefèvre, N., Lienert, S., Liu, J., Marland, G., McGuire, P. C., Melton, J. R., Munro, D. R., Nabel, J. E. M. S., Nakaoka, S. I., Niwa, Y., Ono, T., Pierrot, D., Poulter, B., Rehder, G., Resplandy, L., Robertson, E., Rödenbeck, C., Rosan, T. M., Schwinger, J., Schwingshackl, C., Séférian, R., Sutton, A. J., Sweeney, C., Tanhua, T., Tans, P. P., Tian, H., Tilbrook, B., Tubiello, F., Van Der Werf, G. R., Vuichard, N., Wada, C., Wanninkhof, R., Watson, A. J., Willis, D., Wiltshire, A. J., Yuan, W., Yue, C., Yue, X., Zaehle, S. &
 545 Zeng, J. 2022. Global Carbon Budget 2021. *Earth Syst. Sci. Data*, 14, 1917-2005.
- Fripiat, F., Martínez-García, A., Marconi, D., Fawcett, S. E., Kopf, S. H., Luu, V. H., Rafter, P. A., Zhang, R., Sigman, D. M. & Haug, G. H. 2021. Nitrogen isotopic constraints on nutrient transport to the upper ocean. *Nature Geoscience*, 14, 855-861.



- Frölicher, T. L., Sarmiento, J. L., Paynter, D. J., Dunne, J. P., Krasting, J. P. & Winton, M. 2015. Dominance of the Southern
 550 Ocean in Anthropogenic Carbon and Heat Uptake in CMIP5 Models. *Journal of Climate*, 28, 862-886.
- Fu, W., Moore, J. K., Primeau, F., Collier, N., Ogunro, O. O., Hoffman, F. M. & Randerson, J. T. 2022. Evaluation of CMIP
 models with IOMB: Rates of contemporary ocean carbon uptake linked with vertical temperature gradients and
 transport to the ocean interior. *arXiv preprint arXiv:2202.12933*.
- Fu, W., Randerson, J. T. & Moore, J. K. 2016. Climate change impacts on net primary production (NPP) and export
 555 production (EP) regulated by increasing stratification and phytoplankton community structure in the CMIP5
 models. *Biogeosciences*, 13, 5151-5170.
- Garcia, H., Weathers, K., Paver, C., Smolyar, I., Boyer, T., Locarnini, M., Zweng, M., Mishonov, A., Baranova, O. &
 Seidov, D. 2019. World ocean atlas 2018. Vol. 4: Dissolved inorganic nutrients (phosphate, nitrate and nitrate+
 nitrite, silicate).
- 560 Gregor, L., Kok, S. & Monteiro, P. M. S. 2018. Interannual drivers of the seasonal cycle of CO₂ in the Southern Ocean.
Biogeosciences, 15, 2361-2378.
- Gruber, N., Landschützer, P. & Lovenduski, N. S. 2019. The variable Southern Ocean carbon sink. *Annual review of marine
 science*, 11, 159-186.
- Haberman, K. L., Ross, R. M. & Quetin, L. B. 2003. Diet of the Antarctic krill (*Euphausia superba* Dana): II. Selective
 565 grazing in mixed phytoplankton assemblages. *Journal of Experimental Marine Biology and Ecology*, 283, 97-113.
- Hancock, A. M., Davidson, A. T., Mckinlay, J., McMinn, A., Schulz, K. G. & Van Den Enden, R. L. 2018. Ocean
 acidification changes the structure of an Antarctic coastal protistan community. *Biogeosciences*, 15, 2393-2410.
- Hauck, J., Lenton, A., Langlais, C. & Mearns, R. 2018. The Fate of Carbon and Nutrients Exported Out of the Southern
 Ocean. *Global Biogeochemical Cycles*, 32, 1556-1573.
- 570 Hauck, J., Völker, C., Wolf-Gladrow, D. A., Laufkötter, C., Vogt, M., Aumont, O., Bopp, L., Buitenhuis, E. T., Doney, S.
 C., Dunne, J., Gruber, N., Hashioka, T., John, J., Quéré, C. L., Lima, I. D., Nakano, H., Séférian, R. & Totterdell, I.
 2015. On the Southern Ocean CO₂ uptake and the role of the biological carbon pump in the 21st century. *Global
 Biogeochemical Cycles*, 29, 1451-1470.
- Haumann, F. A., Gruber, N., Münnich, M., Frenger, I. & Kern, S. 2016. Sea-ice transport driving Southern Ocean salinity
 575 and its recent trends. *Nature*, 537, 89-92.
- Henley, S. F., Cavan, E. L., Fawcett, S. E., Kerr, R., Monteiro, T., Sherrell, R. M., Bowie, A. R., Boyd, P. W., Barnes, D. K.
 A., Schloss, I. R., Marshall, T., Flynn, R. & Smith, S. 2020. Changing Biogeochemistry of the Southern Ocean and
 Its Ecosystem Implications. *Frontiers in Marine Science*, 7.
- Henley, S. F., Schofield, O. M., Hendry, K. R., Schloss, I. R., Steinberg, D. K., Moffat, C., Peck, L. S., Costa, D. P., Bakker,
 580 D. C. E., Hughes, C., Rozema, P. D., Ducklow, H. W., Abele, D., Stefels, J., Van Leeuwe, M. A., Brussaard, C. P.
 D., Buma, A. G. J., Kohut, J., Sahade, R., Friedlaender, A. S., Stammerjohn, S. E., Venables, H. J. & Meredith, M.



- P. 2019. Variability and change in the west Antarctic Peninsula marine system: Research priorities and opportunities. *Progress in Oceanography*, 173, 208-237.
- 585 Henley, S. F., Tuerena, R. E., Annett, A. L., Fallick, A. E., Meredith, M. P., Venables, H. J., Clarke, A. & Ganeshram, R. S. 2017. Macronutrient supply, uptake and recycling in the coastal ocean of the west Antarctic Peninsula. *Deep-Sea Research Part II-Topical Studies in Oceanography*, 139, 58-76.
- Henson, S. A., Cael, B. B., Allen, S. R. & Dutkiewicz, S. 2021. Future phytoplankton diversity in a changing climate. *Nat Commun*, 12, 5372.
- 590 Henson, S. A., Laufkötter, C., Leung, S., Giering, S. L. C., Palevsky, H. I. & Cavan, E. L. 2022. Uncertain response of ocean biological carbon export in a changing world. *Nature Geoscience*, 15, 248-254.
- Hogg, A. M., Meredith, M. P., Chambers, D. P., Abrahamsen, E. P., Hughes, C. W. & Morrison, A. K. 2015. Recent trends in the Southern Ocean eddy field. *Journal of Geophysical Research: Oceans*, 120, 257-267.
- Hogg, A. M. & Munday, D. R. 2014. Does the sensitivity of Southern Ocean circulation depend upon bathymetric details? *Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences*, 372, 20130050.
- 595 Holm-Hansen, O., Kahru, M., Hewes, C. D., Kawaguchi, S., Kameda, T., Sushin, V. A., Krasovski, I., Priddle, J., Korb, R., Hewitt, R. P. & Mitchell, B. G. 2004. Temporal and spatial distribution of chlorophyll-a in surface waters of the Scotia Sea as determined by both shipboard measurements and satellite data. *Deep Sea Research Part II: Topical Studies in Oceanography*, 51, 1323-1331.
- 600 Hoppe, C. J. M., Hassler, C. S., Payne, C. D., Tortell, P. D., Rost, B. & Trimborn, S. 2013. Iron Limitation Modulates Ocean Acidification Effects on Southern Ocean Phytoplankton Communities. *PLOS ONE*, 8, e79890.
- Hopwood, M. J., Carroll, D., Höfer, J., Achterberg, E. P., Meire, L., Le Moigne, F. a. C., Bach, L. T., Eich, C., Sutherland, D. A. & González, H. E. 2019. Highly variable iron content modulates iceberg-ocean fertilisation and potential carbon export. *Nature Communications*, 10, 5261.
- 605 Jin, X., Gruber, N., Dunne, J., Sarmiento, J. L. & Armstrong, R. 2006. Diagnosing the contribution of phytoplankton functional groups to the production and export of particulate organic carbon, CaCO₃, and opal from global nutrient and alkalinity distributions. *Global Biogeochemical Cycles*, 20.
- Kahru, M., Mitchell, B. G., Gille, S. T., Hewes, C. D. & Holm-Hansen, O. 2007. Eddies enhance biological production in the Weddell-Scotia Confluence of the Southern Ocean. *Geophysical Research Letters*, 34.
- 610 Kang, S.-H., Kang, J.-S., Lee, S., Chung, K. H., Kim, D. & Park, M. G. 2001. Antarctic phytoplankton assemblages in the marginal ice zone of the northwestern Weddell Sea. *Journal of Plankton Research*, 23, 333-352.
- Kawaguchi, S., Ishida, A., King, R., Raymond, B., Waller, N., Constable, A., Nicol, S., Wakita, M. & Ishimatsu, A. 2013. Risk maps for Antarctic krill under projected Southern Ocean acidification. *Nature Climate Change*, 3, 843-847.



- Kranzler, C. F., Brzezinski, M. A., Cohen, N. R., Lampe, R. H., Maniscalco, M., Till, C. P., Mack, J., Latham, J. R.,
 615 Bruland, K. W., Twining, B. S., Marchetti, A. & Thamtracoln, K. 2021. Impaired viral infection and reduced
 mortality of diatoms in iron-limited oceanic regions. *Nature Geoscience*, 14, 231-237.
- Kwiatkowski, L., Aumont, O., Bopp, L. & Ciais, P. 2018. The Impact of Variable Phytoplankton Stoichiometry on
 Projections of Primary Production, Food Quality, and Carbon Uptake in the Global Ocean. *Global Biogeochemical
 Cycles*, 32, 516-528.
- 620 Landschützer, P., Gruber, N., Haumann, F. A., Rödenbeck, C., Bakker, D. C. E., Heuven, S. V., Hoppema, M., Metzl, N.,
 Sweeney, C., Takahashi, T., Tilbrook, B. & Wanninkhof, R. 2015. The reinvigoration of the Southern Ocean carbon
 sink. *Science*, 349, 1221-1224.
- Lannuzel, D., Chever, F., Van Der Merwe, P. C., Janssens, J., Roukaerts, A., Cavagna, A.-J., Townsend, A. T., Bowie, A. R.
 & Meiners, K. M. 2016. Iron biogeochemistry in Antarctic pack ice during SIPEX-2. *Deep Sea Research Part II:
 625 Topical Studies in Oceanography*, 131, 111-122.
- Lauderdale, J. M., Garabato, A. C. N., Oliver, K. I. C., Follows, M. J. & Williams, R. G. 2013. Wind-driven changes in
 Southern Ocean residual circulation, ocean carbon reservoirs and atmospheric CO₂. *Climate Dynamics*, 41, 2145-
 2164.
- Laufer-Meiser, K., Michaud, A. B., Maisch, M., Byrne, J. M., Kappler, A., Patterson, M. O., Røy, H. & Jørgensen, B. B.
 630 2021. Potentially bioavailable iron produced through benthic cycling in glaciated Arctic fjords of Svalbard. *Nature
 Communications*, 12, 1349.
- Laufkötter, C., Stern, A. A., John, J. G., Stock, C. A. & Dunne, J. P. 2018. Glacial Iron Sources Stimulate the Southern
 Ocean Carbon Cycle. *Geophysical Research Letters*, 45, 13,377-13,385.
- Lawrence, B. N., Bennett, V. L., Churchill, J., Jukes, M., Kershaw, P., Pascoe, S., Pepler, S., Pritchard, M. & Stephens, A.
 635 Storing and manipulating environmental big data with JASMIN. 2013 IEEE International Conference on Big Data,
 6-9 Oct. 2013 2013. 68-75.
- Leung, S., Cabré, A. & Marinov, I. 2015. A latitudinally banded phytoplankton response to 21st century climate change in
 the Southern Ocean across the CMIP5 model suite. *Biogeosciences*, 12, 5715-5734.
- Levermann, A., Winkelmann, R., Nowicki, S., Fastook, J. L., Frieler, K., Greve, R., Hellmer, H. H., Martin, M. A.,
 640 Meinshausen, M., Mengel, M., Payne, A. J., Pollard, D., Sato, T., Timmermann, R., Wang, W. L. & Bindschadler,
 R. A. 2014. Projecting Antarctic ice discharge using response functions from SeaRISE ice-sheet models. *Earth Syst.
 Dynam.*, 5, 271-293.
- Litchman, E. & Klausmeier, C. A. 2008. Trait-Based Community Ecology of Phytoplankton. *Annual Review of Ecology,
 Evolution, and Systematics*, 39, 615-639.
- 645 Lizotte, M. P. 2001. The contributions of sea ice algae to Antarctic marine primary production. *American zoologist*, 41, 57-
 73.



- Lovenduski, N. S., Gruber, N. & Doney, S. C. 2008. Toward a mechanistic understanding of the decadal trends in the Southern Ocean carbon sink. *Global Biogeochemical Cycles*, 22.
- 650 Mascioni, M., Almandoz, G. O., Cefarelli, A. O., Cusick, A., Ferrario, M. E. & Vernet, M. 2019. Phytoplankton composition and bloom formation in unexplored nearshore waters of the western Antarctic Peninsula. *Polar Biology*, 42, 1859-1872.
- Matsuoka, K., Skoglund, A., Roth, G., De Pomereu, J., Griffiths, H., Headland, R., Herried, B., Katsumata, K., Le Brocq, A., Licht, K., Morgan, F., Neff, P. D., Ritz, C., Scheinert, M., Tamura, T., Van De Putte, A., Van Den Broeke, M., Von Deschanden, A., Deschamps-Berger, C., Van Liefferinge, B., Tronstad, S. & Melvær, Y. 2021. Quantarctica, an
 655 integrated mapping environment for Antarctica, the Southern Ocean, and sub-Antarctic islands. *Environmental Modelling & Software*, 140, 105015.
- Mayzaud, P. & Pakhomov, E. A. 2014. The role of zooplankton communities in carbon recycling in the Ocean: the case of the Southern Ocean. *Journal of Plankton Research*, 36, 1543-1556.
- 660 McCain, J. S. P., Tagliabue, A., Susko, E., Achterberg, E. P., Allen, A. E. & Bertrand, E. M. 2021. Cellular costs underpin micronutrient limitation in phytoplankton. *Science Advances*, 7, eabg6501.
- Meredith, M., Sommerkorn, M., Cassotta, S., Derksen, C., Ekaykin, A., Hollowed, A., Kofinas, G., Mackintosh, A., Melbourne-Thomas, J. & Muelbert, M. 2019. Chapter 3: polar regions. *IPCC special report on the ocean and cryosphere in a changing climate*, 5.
- 665 Meredith, M. P. & Hogg, A. M. 2006. Circumpolar response of Southern Ocean eddy activity to a change in the Southern Annular Mode. *Geophysical Research Letters*, 33.
- Mikaloff Fletcher, S., Gruber, N., Jacobson, A. R., Gloor, M., Doney, S., Dutkiewicz, S., Gerber, M., Follows, M., Joos, F. & Lindsay, K. 2007. Inverse estimates of the oceanic sources and sinks of natural CO₂ and the implied oceanic carbon transport. *Global Biogeochemical Cycles*, 21.
- Moline, M. A., Claustre, H., Frazer, T. K., Schofield, O. & Vernet, M. 2004. Alteration of the food web along the Antarctic
 670 Peninsula in response to a regional warming trend. *Global Change Biology*, 10, 1973-1980.
- Moline, M. A., Karnovsky, N. J., Brown, Z., Divoky, G. J., Frazer, T. K., Jacoby, C. A., Torres, J. J. & Fraser, W. R. 2008. High latitude changes in ice dynamics and their impact on polar marine ecosystems. *Annals of the New York Academy of Sciences*, 1134, 267.
- 675 Montes-Hugo, M., Vernet, M., Martinson, D., Smith, R. & Iannuzzi, R. 2008. Variability on phytoplankton size structure in the western Antarctic Peninsula (1997–2006). *Deep Sea Research Part II: Topical Studies in Oceanography*, 55, 2106-2117.
- Moore, J. K., Fu, W., Primeau, F., Britten, G. L., Lindsay, K., Long, M., Doney, S. C., Mahowald, N., Hoffman, F. & Randerson, J. T. 2018. Sustained climate warming drives declining marine biological productivity. *Science*, 359, 1139-1143.



- 680 Moreau, S., Boyd, P. W. & Strutton, P. G. 2020. Remote assessment of the fate of phytoplankton in the Southern Ocean sea-
 ice zone. *Nature Communications*, 11, 3108.
- Moreau, S., Mostajir, B., Bélanger, S., Schloss, I. R., Vancoppenolle, M., Demers, S. & Ferreyra, G. A. 2015. Climate
 change enhances primary production in the western Antarctic Peninsula. *Global Change Biology*, 21, 2191-2205.
- Nicholson, S.-A., Whitt, D. B., Fer, I., Du Plessis, M. D., Lebéhot, A. D., Swart, S., Sutton, A. J. & Monteiro, P. M. S. 2022.
 685 Storms drive outgassing of CO₂ in the subpolar Southern Ocean. *Nature Communications*, 13, 158.
- Nissen, C., Vogt, M., Münnich, M., Gruber, N. & Haumann, F. A. 2018. Factors controlling coccolithophore biogeography
 in the Southern Ocean. *Biogeosciences*, 15, 6997-7024.
- O'Neill, B. C., Tebaldi, C., Van Vuuren, D. P., Eyring, V., Friedlingstein, P., Hurtt, G., Knutti, R., Kriegler, E., Lamarque, J.
 F., Lowe, J., Meehl, G. A., Moss, R., Riahi, K. & Sanderson, B. M. 2016. The Scenario Model Intercomparison
 690 Project (ScenarioMIP) for CMIP6. *Geosci. Model Dev.*, 9, 3461-3482.
- Okin, G. S., Baker, A. R., Tegen, I., Mahowald, N. M., Dentener, F. J., Duce, R. A., Galloway, J. N., Hunter, K., Kanakidou,
 M., Kubilay, N., Prospero, J. M., Sarin, M., Surapipith, V., Uematsu, M. & Zhu, T. 2011. Impacts of atmospheric
 nutrient deposition on marine productivity: Roles of nitrogen, phosphorus, and iron. *Global Biogeochemical Cycles*,
 25.
- 695 Palter, J. B., Sarmiento, J. L., Gnanadesikan, A., Simeon, J. & Slater, R. D. 2010. Fueling export production: nutrient return
 pathways from the deep ocean and their dependence on the Meridional Overturning Circulation. *Biogeosciences*, 7,
 3549-3568.
- Pausch, F., Bischof, K. & Trimborn, S. 2019. Iron and manganese co-limit growth of the Southern Ocean diatom
Chaetoceros debilis. *PLoS One*, 14, e0221959.
- 700 Pearce, I., Davidson, A. T., Thomson, P. G., Wright, S. & Van Den Enden, R. 2011. Marine microbial ecology in the sub-
 Antarctic Zone: Rates of bacterial and phytoplankton growth and grazing by heterotrophic protists. *Deep Sea
 Research Part II: Topical Studies in Oceanography*, 58, 2248-2259.
- Pellichero, V., Sallée, J.-B., Chapman, C. C. & Downes, S. M. 2018. The southern ocean meridional overturning in the sea-
 ice sector is driven by freshwater fluxes. *Nature Communications*, 9, 1789.
- 705 Person, R., Aumont, O. & Lévy, M. 2018. The Biological Pump and Seasonal Variability of pCO₂ in the Southern Ocean:
 Exploring the Role of Diatom Adaptation to Low Iron. *Journal of Geophysical Research: Oceans*, 123, 3204-3226.
- Petrou, K., Baker, K. G., Nielsen, D. A., Hancock, A. M., Schulz, K. G. & Davidson, A. T. 2019. Acidification diminishes
 diatom silica production in the Southern Ocean. *Nature Climate Change*, 9, 781-786.
- Primeau, F. W., Holzer, M. & Devries, T. 2013. Southern Ocean nutrient trapping and the efficiency of the biological pump.
 710 *Journal of Geophysical Research: Oceans*, 118, 2547-2564.
- Quéguiner, B. 2013. Iron fertilization and the structure of planktonic communities in high nutrient regions of the Southern
 Ocean. *Deep Sea Research Part II: Topical Studies in Oceanography*, 90, 43-54.



- Raiswell, R., Benning, L. G., Tranter, M. & Tulaczyk, S. 2008. Bioavailable iron in the Southern Ocean: the significance of the iceberg conveyor belt. *Geochemical Transactions*, 9, 7.
- 715 Redfield, A. C. 1958. The biological control of chemical factors in the environment. *American scientist*, 46, 230A-221.
- Riebesell, U., Schulz, K. G., Bellerby, R. G. J., Botros, M., Fritsche, P., Meyerhöfer, M., Neill, C., Nondal, G., Oeschies, A., Wohlers, J. & Zöllner, E. 2007. Enhanced biological carbon consumption in a high CO₂ ocean. *Nature*, 450, 545-548.
- Roach, L. A., Dörr, J., Holmes, C. R., Massonnet, F., Blockley, E. W., Notz, D., Rackow, T., Raphael, M. N., O'farrell, S. P.,
 720 Bailey, D. A. & Bitz, C. M. 2020. Antarctic Sea Ice Area in CMIP6. *Geophysical Research Letters*, 47, e2019GL086729.
- Robinson, J., Popova, E. E., Srokosz, M. A. & Yool, A. 2016. A tale of three islands: Downstream natural iron fertilization in the Southern Ocean. *Journal of Geophysical Research: Oceans*, 121, 3350-3371.
- Rozema, P., Venables, H., Van De Poll, W., Clarke, A., Meredith, M. & Buma, A. 2017. Interannual variability in
 725 phytoplankton biomass and species composition in northern Marguerite Bay (West Antarctic Peninsula) is governed by both winter sea ice cover and summer stratification. *Limnology and Oceanography*, 62, 235-252.
- Saba, G. K., Fraser, W. R., Saba, V. S., Iannuzzi, R. A., Coleman, K. E., Doney, S. C., Ducklow, H. W., Martinson, D. G., Miles, T. N., Patterson-Fraser, D. L., Stammerjohn, S. E., Steinberg, D. K. & Schofield, O. M. 2014. Winter and spring controls on the summer food web of the coastal West Antarctic Peninsula. *Nature Communications*, 5, 4318.
- 730 Sallée, J.-B., Pellichero, V., Akhondas, C., Pauthenet, E., Vignes, L., Schmidtko, S., Garabato, A. N., Sutherland, P. & Kuusela, M. 2021. Summertime increases in upper-ocean stratification and mixed-layer depth. *Nature*, 591, 592-598.
- Sarmiento, J. L., Gruber, N., Brzezinski, M. A. & Dunne, J. P. 2004. High-latitude controls of thermocline nutrients and low latitude biological productivity. *Nature*, 427, 56-60.
- 735 Schofield, O., Brown, M., Kohut, J., Nardelli, S., Saba, G., Waite, N. & Ducklow, H. 2018. Changes in the upper ocean mixed layer and phytoplankton productivity along the West Antarctic Peninsula. *Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences*, 376, 20170173.
- Seferian, R., Berthet, S., Yool, A., Palmieri, J., Bopp, L., Tagliabue, A., Kwiatkowski, L., Aumont, O., Christian, J., Dunne, J., Gehlen, M., Ilyina, T., John, J. G., Li, H., Long, M. C., Luo, J. Y., Nakano, H., Romanou, A., Schwinger, J.,
 740 Stock, C., Santana-Falcon, Y., Takano, Y., Tjiputra, J., Tsujino, H., Watanabe, M., Wu, T., Wu, F. & Yamamoto, A. 2020. Tracking Improvement in Simulated Marine Biogeochemistry Between CMIP5 and CMIP6. *Curr Clim Change Rep*, 6, 1-25.
- Shadwick, E. H., De Meo, O. A., Schroeter, S., Arroyo, M. C., Martinson, D. G. & Ducklow, H. 2021. Sea Ice Suppression of CO₂ Outgassing in the West Antarctic Peninsula: Implications For The Evolving Southern Ocean Carbon Sink.
 745 *Geophysical Research Letters*, 48, e2020GL091835.



- Sheen, K. L., Naveira Garabato, A. C., Brearley, J. A., Meredith, M. P., Polzin, K. L., Smeed, D. A., Forryan, A., King, B. A., Sallée, J. B., St. Laurent, L., Thurnherr, A. M., Toole, J. M., Waterman, S. N. & Watson, A. J. 2014. Eddy-induced variability in Southern Ocean abyssal mixing on climatic timescales. *Nature Geoscience*, 7, 577-582.
- Shi, J.-R., Talley, L. D., Xie, S.-P., Liu, W. & Gille, S. T. 2020. Effects of Buoyancy and Wind Forcing on Southern Ocean Climate Change. *Journal of Climate*, 33, 10003-10020.
- Shu, Q., Wang, Q., Song, Z., Qiao, F., Zhao, J., Chu, M. & Li, X. 2020. Assessment of Sea Ice Extent in CMIP6 With Comparison to Observations and CMIP5. *Geophysical Research Letters*, 47, e2020GL087965.
- Street, J. H. & Paytan, A. 2005. Iron, phytoplankton growth, and the carbon cycle. *Met Ions Biol Syst*, 43, 153-93.
- Sunagawa, S., Acinas, S. G., Bork, P., Bowler, C., Acinas, S. G., Babin, M., Bork, P., Boss, E., Bowler, C., Cochrane, G., De Vargas, C., Follows, M., Gorsky, G., Grimsley, N., Guidi, L., Hingamp, P., Iudicone, D., Jaillon, O., Kandels, S., Karp-Boss, L., Karsenti, E., Lescot, M., Not, F., Ogata, H., Pesant, S., Poulton, N., Raes, J., Sardet, C., Sieracki, M., Speich, S., Stemmann, L., Sullivan, M. B., Sunagawa, S., Wincker, P., Eveillard, D., Gorsky, G., Guidi, L., Iudicone, D., Karsenti, E., Lombard, F., Ogata, H., Pesant, S., Sullivan, M. B., Wincker, P., De Vargas, C. & Tara Oceans, C. 2020. Tara Oceans: towards global ocean ecosystems biology. *Nature Reviews Microbiology*, 18, 428-445.
- Tagliabue, A., Aumont, O., Death, R., Dunne, J. P., Dutkiewicz, S., Galbraith, E., Misumi, K., Moore, J. K., Ridgwell, A., Sherman, E., Stock, C., Vichi, M., Völker, C. & Yool, A. 2016. How well do global ocean biogeochemistry models simulate dissolved iron distributions? *Global Biogeochemical Cycles*, 30, 149-174.
- Thomalla, S. J., Ogunkoya, A. G., Vichi, M. & Swart, S. 2017. Using Optical Sensors on Gliders to Estimate Phytoplankton Carbon Concentrations and Chlorophyll-to-Carbon Ratios in the Southern Ocean. *Frontiers in Marine Science*, 4.
- Tortell, P. D., Payne, C. D., Li, Y., Trimborn, S., Rost, B., Smith, W. O., Riesselman, C., Dunbar, R. B., Sedwick, P. & Ditullio, G. R. 2008. CO₂ sensitivity of Southern Ocean phytoplankton. *Geophysical Research Letters*, 35.
- Touzé-Peiffer, L., Barberousse, A. & Le Treut, H. 2020. The Coupled Model Intercomparison Project: History, uses, and structural effects on climate research. *Wiley Interdisciplinary Reviews: Climate Change*, 11, e648.
- Treguer, P. J. & De La Rocha, C. L. 2013. The world ocean silica cycle. *Ann Rev Mar Sci*, 5, 477-501.
- Turner, J. & Comiso, J. 2017. Solve Antarctica's sea-ice puzzle. *Nature*, 547, 275-277.
- Uchida, T., Balwada, D., P. Abernathey, R., A. McKinley, G., K. Smith, S. & Lévy, M. 2020. Vertical eddy iron fluxes support primary production in the open Southern Ocean. *Nature Communications*, 11, 1125.
- Venables, H. J., Clarke, A. & Meredith, M. P. 2013. Wintertime controls on summer stratification and productivity at the western Antarctic Peninsula. *Limnology and Oceanography*, 58, 1035-1047.
- Watson, A. J., Bakker, D. C. E., Ridgwell, A. J., Boyd, P. W. & Law, C. S. 2000. Effect of iron supply on Southern Ocean CO₂ uptake and implications for glacial atmospheric CO₂. *Nature*, 407, 730-733.
- Weber, T. S. & Deutsch, C. 2010. Ocean nutrient ratios governed by plankton biogeography. *Nature*, 467, 550-554.



- Westwood, K. J., Thomson, P. G., Van Den Enden, R. L., Maher, L. E., Wright, S. W. & Davidson, A. T. 2018. Ocean
 780 acidification impacts primary and bacterial production in Antarctic coastal waters during austral summer. *Journal of
 Experimental Marine Biology and Ecology*, 498, 46-60.
- Wright, S. W., Van Den Enden, R. L., Pearce, I., Davidson, A. T., Scott, F. J. & Westwood, K. J. 2010. Phytoplankton
 community structure and stocks in the Southern Ocean (30–80 E) determined by CHEMTAX analysis of HPLC
 pigment signatures. *Deep Sea Research Part II: Topical Studies in Oceanography*, 57, 758-778.
- 785 Xu, K., Fu, F.-X. & Hutchins, D. A. 2014. Comparative responses of two dominant Antarctic phytoplankton taxa to
 interactions between ocean acidification, warming, irradiance, and iron availability. *Limnology and Oceanography*,
 59, 1919-1931.
- Zhu, Z., Xu, K., Fu, F., Spackeen, J. L., Bronk, D. A. & Hutchins, D. A. 2016. A comparative study of iron and temperature
 interactive effects on diatoms and *Phaeocystis antarctica* from the Ross Sea, Antarctica. *Marine Ecology Progress
 790 Series*, 550, 39-51.