

1 **Influence of Oxygen Minimum Zone on Macrobenthic Community Structure in the**  
2 **Northern Benguela Upwelling System: A Macro-Nematode Perspective**

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11 **Abstract**

12 Understanding the dynamics of marine ecosystems, particularly the benthic communities, is crucial  
13 for predicting the impacts of climate change and human activities on oceanic biodiversity. In this  
14 context, we conducted a study on the macrobenthic communities in the Benguela Upwelling  
15 System (BUS). Macrobenthic samples were collected offshore Namibia on board *R/V Mirabilis*  
16 during the 3<sup>rd</sup> RGNO training and the National Marine Information and Research Centre's  
17 (NatMIRC's) plankton survey from 13<sup>th</sup> May to 17<sup>th</sup> May 2016. Two transects, Terrace Bay (20°  
18 S) and Walvis Bay (23° S), hosted three stations each, while the third transect, Luderitz (26° S),  
19 hosted only one station. From the results, three oxygen zones were identified, namely Microxic  
20 (<0.1 ml l<sup>-1</sup>), Dysoxic (0.1-1.0 ml l<sup>-1</sup>), and Oxic (>1.0 ml l<sup>-1</sup>). A total of 20 macrobenthic taxa were  
21 identified; Nematoda, Polychaeta, Cumacea, and Oligochaeta were the most dominant taxa and  
22 were present in all oxygen zones. Eighteen genera of macro-nematoda were identified;  
23 *Desmolaimus* and *Paracomesoma* dominated in all oxygen zones, *Metoncholaimus* recorded  
24 higher abundance in the dysoxic stations, and were absent in the oxic stations, and the opposite  
25 was observed for *Halanonchus* and *Dorylaimopsis*. H' diversity values for both the general  
26 macrofauna and macro-nematoda were higher in the oxic stations and lower in both the dysoxic  
27 and microxic stations, while an opposite trend was observed for dominance values. Density values  
28 were lower in microxic stations and higher in dysoxic stations, while at the oxic stations, the  
29 density values fell in between the two hypoxic zones. In conclusion, this study provides an  
30 overview of the distribution, diversity, and response to varying oxygen conditions of macrobenthic  
31 communities and their importance in marine ecosystems.

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35 **Keywords:** Macrofauna, Macro-Nematoda, Oxygen minimum zone, Benguela Upwelling System  
36 (BUS), Dissolved oxygen, hypoxia

## 37 1.0 Introduction

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38 In recent decades, the concentration of dissolved oxygen (DO) in the ocean, specifically in the  
39 tropics, has been decreasing. This not only increases the size of areas under hypoxia but also their  
40 prevalence (Breitburg et al., 2018). Hypoxia is ranked among the major threats to the actualization  
41 of the blue economy and the achievement of the 14<sup>th</sup> pillar of the Sustainable Development Goals  
42 (SDGs), specifically target 14.7 which aims at increasing the economic benefits of marine resource  
43 utilization through its sustainable use by developing countries (Rossi, 2023; Liu, 2023). Although  
44 hypoxia can result from natural phenomena like upwelling and thermal stratification, the current  
45 expansion of hypoxic areas is mainly a result of accelerated nutrient inputs in coastal areas which  
46 increases algal production subsequently resulting in higher organic matter production which in  
47 turn results in increased aerobic microbial decomposition lowering the levels of DO in the water  
48 (Gobler & Baumann, 2016). It has been projected that such changes may affect different organisms  
49 differently depending on their tolerance and reactions to lower dissolved oxygen in their habitats  
50 (Rudolph et al., 2020). Studies on ecosystems with hypoxia as a natural phenomenon can assist in  
51 predicting and understanding how human-induced hypoxia might affect and shape marine  
52 ecosystems in the face of the expansion of marine areas under hypoxia.

53 The Benguela upwelling system (BUS) is located off the southwest coast of Africa. It extends from  
54 Cape Frio in Angola to the southern tip of the continent in Cape Agulhas, South Africa, and is one  
55 of the most productive regions of the world's ocean (Magalhães, 2018). The high productivity  
56 provides a huge source of carbon resulting from photosynthesis, which gradually sinks through the  
57 water column resulting in a rapid oxygen loss due to biochemical oxygen consumption. This  
58 consequently causes low dissolved oxygen concentrations and thus creates a permanent extensive

60 shallow oxygen minimum zone (Bohata & Koppelman, 2013; Emeis et al., 2018). When the  
61 oxygen minimum zone (OMZ) comes into contact with the seafloor, it creates a strong oxygen  
62 gradient at the benthic zone at depths ranging between 50 meters and 300 meters, resulting in a  
63 hypoxic (<0.5 ml l<sup>-1</sup>) inner shelf (Levin, 2003). The oxygen gradient created at the OMZ's benthic  
64 zone is believed to primarily regulate the benthic community distribution and diversity patterns  
65 (Zettler et al., 2013; Teuber et al., 2013). In these zones, microbial communities play a key role in  
66 the cycling of elements like carbon, nitrogen, and sulfur. The expansion of OMZs may increase  
67 nitrate use by nematode-associated microbes such as SAR11, a significant player in the marine  
68 nitrogen cycle. These nematodes and the communities they are part of influence biogeochemical  
69 processes and ecosystem dynamics within OMZs. For this reason, understanding these  
70 communities is crucial, especially with the ongoing expansion of OMZs due to climate change.

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71 The general trend observed in most OMZs in global oceans namely, Walvis Bay, Namibia (the  
72 location of this study), California, USA, and Oman margin (off the Arabian Peninsula) indicates  
73 that the densities of macrofauna (benthic organisms that are typically retained in a 0.5 mm sieve  
74 but pass through a 2.00 mm sieve (Bachelet, 1990) generally display a negative response to  
75 reducing oxygen levels within the OMZ with a 30% to 70% reduction in densities in regions with  
76 less than 0.15 ml l<sup>-1</sup> (Levin, 2003). Similarly, diversity reduces as oxygen levels reduce within the  
77 OMZ because of the loss of intolerant species and increased dominance of the tolerant species  
78 (Currie et al., 2018; Levin et al, 2009). Nematodes and some families from the annelid worms have  
79 been observed to be able to tolerate low oxygen, with Nematoda (in the meiofauna group) having  
80 95-99% abundance. Some polychaetes families like Spionidae, Dorvilleidae, and Lumbrineridae  
81 can also tolerate low oxygen in the OMZ by having a high gill surface area for increasing oxygen

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87 uptake. On the other hand, harpacticoid copepods are the most sensitive taxon to hypoxia (Levin  
88 et al., 2009; Zeppilli et al., 2015).

89 It has been observed that the macrofauna diversity is lowest off Walvis Bay, a city located on the  
90 western coast of Namibia, attributed to the perennial intense OMZ over the shelf (Currie et al.,  
91 2018). In contrast, the diversity increases significantly northwards off the Kunene River (Zettler  
92 et al., 2013), which flows from the highlands of Angola, along the border with Namibia, and into  
93 the Atlantic Ocean (See Figure 1). This increase in diversity is proposed to be a result of moving  
94 away from the intense OMZ cells off Walvis Bay and also the reflection of the biogeography where  
95 diversity increases with [lower](#) latitude (Zettler et al.,2009). Outside the OMZ, bathymetry, and  
96 latitude are said to be the factors affecting the infaunal communities at the Namibian shelf (Steffani  
97 et al., 2015).

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98 This study aims to identify the relationship between the levels of dissolved oxygen and the  
99 macrobenthic community distribution across the Northern Benguela Upwelling system as a guide  
100 on how the expansion of man-made hypoxia might influence the benthic fauna distribution on the  
101 seafloor.

## 102 **2.0 Material and Methods**

### 103 **2.1 Study Area**

104 The study area was located across the Northern Benguela Upwelling System (between 26° S and  
105 20° S) along the Namibian continental shelf, which hosts a deep continental shelf (around 300 m)  
106 (Figure 1). The intense upwelling in the study site has rendered the area highly productive,  
107 resulting in a characteristic shallow OMZ (50-200 m) with stronger productive cells around  
108 Luderitz and Walvis Bay (Bohata & Koppelman, 2013). The inner shelf is described to be

110 extremely oxygen-depleted, caused by in situ organic matter decomposition and warm poleward  
111 Angola current, which peaks in June-July while the continental slope below the OMZ is better  
112 oxygenated (Levin et al., 2009; Emeis et al., 2004).

113 The benthic zone in the OMZ in Northern BUS is characterized by extensive areas of diatomaceous  
114 mud, which are associated with high primary production at the ocean surface and low  
115 concentration of dissolved oxygen (Levin et al., 2009; Steffani et al., 2015). These low oxygen  
116 levels, as per the findings of Levin et al.(2009) were found to be less than 0.5 ml L<sup>-1</sup> in 55% of  
117 the total shelf. Furthermore, extreme anoxia, defined as oxygen concentrations less than 1 µM,  
118 was observed over an expansive area of almost 900 km<sup>2</sup>.

## 119 **2.2 Sample Collection**

120 Samples were collected from three transects (off Luderitz (OL) 26° S, off Walvis Bay (OWB) 23°  
121 S, and off Terrace Bay (OTB) 20° S) onboard *R/V Mirabilis* during the RGNO training National  
122 Marine Information and Research Centre's (NatMIRC's) plankton survey from 13<sup>th</sup> May to 17<sup>th</sup>  
123 May 2016. The sampling stations were located at 02 nm, 20 nm, 40 nm, or 70 nm [from the shore](#)  
124 at each transect, with the 26° S transect hosting only one station at 90 nm (Figure 1). However,  
125 benthic samples from these stations were dependent on the prevailing weather conditions and the  
126 ability to get good core samples (Table 1).

127 Sampling was done using a multi-corer from which individual cores were taken and sub-sampled  
128 for macrobenthos using a 6.4 cm diameter corer, and sediment samples for granulometry and  
129 organic matter analysis were taken simultaneously. Replicate samples were taken from a  
130 subsequent deployment of the multi-corer, where possible, to avoid pseudoreplication. The

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132 macrofauna cored samples were put in sampling bottles and immediately fixed with 5% buffered  
133 formalin, while samples for organic matter analysis were frozen to arrest microbial decomposition.  
134 Depth and temperature measurements were collected from a probe attached to the multi-corer,  
135 while dissolved oxygen concentrations in this study used the Winkler method from the overlying  
136 water (Montgomery et al., 1964).

### 137 **2.3 Laboratory analysis**

138 In the laboratory, macrobenthic samples were sieved between 2.00 mm and 0.45 mm sieves.  
139 Studies typically employ a sieve size of 0.5mm, our study however utilized a 0.45mm sieve due  
140 to its availability during the course of our research. This size, although slightly smaller, falls within  
141 the range of commonly used sieve sizes (300 to 500 microns) [for macrofaunal research](#). The use  
142 of a 0.45mm sieve is also not unprecedented in macrofaunal studies; studies by Li et al. (2018) and  
143 Zhang et al. (2022) have successfully employed this sieve size. Therefore, while acknowledging  
144 the slight deviation, results remain valid and comparable to those obtained using the more common  
145 sieve sizes.

146 The sieve fraction retained in the 0.45 mm mesh sieve was then preserved in [a](#) 5% buffered  
147 formalin solution, with 3-5 drops of Rose Bengal solution added to aid in sorting. After rinsing the  
148 samples with water, they were sorted under a dissecting microscope using the key provided by  
149 Thiel & Higgins (1988) to the class or order levels and then counted. Subsequently, the nematodes  
150 were pin-picked, fixed on permanent slides, and identified to the genus level using the key from  
151 Platt & Warwick (1988). Feeding types were determined for these genera following the  
152 methodology described by Wieser (1953), whereby nematodes are classified based on their buccal  
153 morphology, including shape, size, and presence or absence of buccal armature, resulting in four



154 groups. Those lacking buccal armature are categorized as deposit feeders (group 1), [they are](#) further  
155 subdivided into selective deposit feeders (1A) and non-selective deposit feeders (1B) based on the  
156 size of their buccal cavity, with the former possessing a very small buccal cavity, and the latter  
157 having a larger one (Moens and Vincx, 1997; Moens et al., 2013). Nematodes with buccal armature  
158 constitute group 2 and can be [selective epigrowth feeders and herbivores \(2A\)](#), using small teeth  
159 to graze diatoms, microalgae, and bacteria, or carnivores and omnivores (2B), characterized by the  
160 possession of teeth, mandibles, or onchia for piercing and consuming prey (Moens et al., 2013)

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161 For sediment granulometry, samples were wet-sieved using a 1.00 mm sieve, 0.5 mm, 0.25 mm,  
162 105  $\mu$ m, 0.063 mm, and those below 0.063 mm (<0.063 mm). The sediments collected at the  
163 different sieves were then put on pre-weighed Petri dishes and dried in an oven at 60 °C until a  
164 constant weight was reached, and these dried weights were used for sediment granulometry  
165 (Wentworth, 1922).

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166 The samples for total organic matter (TOM) were dried in an oven at 60 °C until a constant weight  
167 was achieved, [a known weight of each sample was then placed in labeled aluminum foil](#), and the  
168 loss of weight on ignition (LOI) technique was used to determine TOM from the study site (Thiel  
169 & Higgins, 1988).

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## 170 **2.4 Data analysis**

171 Out of the seven sampled stations, only three had replicates (Table 1), making direct statistical  
172 comparisons between stations impossible. To overcome this limitation, the sampling stations were  
173 grouped based on the measured environmental variable that provided the most significant  
174 relationship identified through step-wise regression analysis with the various biotic indices  
175 documented within the macrofauna community ([See supplementary material \(Tables S1-S4\)](#)). The

179 communities within these key 'habitat types' were then compared using the indices and analyses  
180 outlined below;

181 Grouping Stations; Following the approach of (Levin, 2003), stations with DO levels less than 0.1  
182 ml l<sup>-1</sup> were grouped as 'microxic,' those with DO levels between 0.1 and 1.0 ml l<sup>-1</sup> were grouped  
183 as 'dysoxic' and those with DO levels of 1.0 ml l<sup>-1</sup> and above grouped as 'oxic'. This approach was  
184 taken to investigate the role of DO levels in these communities. It was further justified by the  
185 application of SPSS, which indicated that DO was a key determinant in macrobenthic community  
186 structures ( $R^2=0.521$ ,  $P=.012$ ) (Table S2(a); S2(b)).

187 Abundance Analysis; The data recorded were analyzed for abundance (density and relative  
188 abundance) in Excel. Additionally, the Paleontological Statistics Software package (PAST v2.17c)  
189 (Hammer et al., 2001) was used to calculate the diversity and the diversity t-test between the  
190 stations. [For this study, H' diversity is calculated using the natural log \(loge\) while dominance was](#)  
191 [the complement of the Simpson Index \(1-Simpson index\).](#)

192 Community Similarity and Diversity; Bray Curtis similarity, Analysis of Similarity (ANOSIM),  
193 and Similarity percentages (SIMPER) were conducted using Plymouth Routines in Multivariate  
194 Ecological Research (PRIMER v5.2.9) (Clarke & Gorley, 2005).

### 195 3.0 Results

#### 196 3.1 Abiotic variables

197 Total Organic Matter (%TOM) demonstrated an inverse relationship with depth, with higher  
198 organic matter values recorded in the shallower stations. For example, the shallowest stations  
199 20020, 20002, and 23002 had the highest TOM of 38.6%±2.16, 27.5%±1.55 and 23.1%,

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205 respectively whilst the lowest TOM values were found in most offshore stations, stations 23070  
206 and 26090, with  $9.89 \pm 1.55$  and 4.4, respectively (Table 1). The lowest oxygen values were  
207 recorded on the OWB 23°S transect, with the two most onshore stations (23020 and 23002)  
208 recording the lowest levels of oxygen ( $0.02$  and  $0.06 \text{ ml l}^{-1}$ , respectively). The inshore station from  
209 transect OCF 20°S (20002) had a higher DO ( $1.8 \text{ ml l}^{-1}$ ) than stations 20020 and 20040 from the  
210 same transect, where the DO levels recorded were  $0.67$  and  $0.53 \text{ ml l}^{-1}$ , respectively, while the  
211 offshore stations 23070 in transect OWB 23°S and 26090 in transect OL 26°S recorded the highest  
212 dissolved oxygen ( $2.30$  and  $4.34 \text{ ml l}^{-1}$ , respectively) (Table 1).

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213 Fine sand was the most common sediment size ranging between 30-38.8% in all stations. Most  
214 stations from the sites also recorded higher proportions of medium sand and silt, except for station  
215 23020 in transect OWB 23°S, which recorded a lower proportion of silt and an increased abundance  
216 of coarse sand compared to the other stations (Table 2).

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### 217 3.2 Macrobenthic assemblages

218 Macrofaunal densities differed significantly across the various oxygen zones; In the microxic  
219 stations, very low densities were observed ( $4,661 \pm 4,834 \text{ ind. m}^{-2}$ ), and the lowest number of taxa  
220 was recorded, with only six taxa present. These included Nematoda, Polychaeta, Oligochaeta, and  
221 Cumacea, the only peracarid crustacean found across all oxygen zones. Despite the low number of  
222 taxa ( $S=6$ ), these stations recorded slightly higher  $H'$  diversity than the dysoxic stations, while the  
223 dominance index was intermediate ( $0.5 \pm 0.18$ ) (Figure 5).

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224 Dysoxic stations [in contrast](#) recorded the highest densities ( $74,108 \pm 134,126 \text{ ind. m}^{-2}$ ). One station  
225 within this Oxygen range, station (20040), recorded an extremely high density of  $274,991 \text{ ind. m}^{-2}$ ,  
226 primarily due to a high abundance of Nematoda and Oligochaeta. Although the taxa in the

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234 dysoxic stations were similar to those of the microxic stations, the composition and dominance  
235 differed significantly. The macro-nematoda was the most dominant taxon in this zone, with  
236 numbers reaching as high as 233,354 ind. m<sup>-2</sup> in one of the stations. Interestingly, Bivalvia and  
237 Ostracoda, absent in the microxic stations, were present here, albeit in low abundance (Figure 3).  
238 Dominance was highest in these stations (0.6±0.28) (Figure 5).

239 Oxic stations demonstrated the highest number of taxa (18) and the second-highest average density  
240 (14,345±6,726 ind. m<sup>-2</sup>) (Figure. 2). These stations recorded taxa like Echinodermata  
241 (Ophiuroidea), Holothuroidea, Aeolosomatidea, Isopoda, Aplacophora, and Amphipoda, which all  
242 had average abundances of more than 1%. Nemertina, Turbellaria, and Cnidaria were also recorded  
243 in these stations; however, their abundances were below 1%, and they were grouped as 'others'  
244 (Figure 3). These stations recorded the highest H' diversity (1.46±0.4) and the lowest dominance  
245 (0.31±0.18) (Figure 5).

246 All the oxygen zones were dissimilar to one another based on multivariate community analysis  
247 using Bray-Curtis analysis of dissimilarity. The highest dissimilarity was observed between the  
248 dysoxic and microxic zones which were 77.99% dissimilar despite both stations being  
249 characterized by low dissolved oxygen levels. The oxic stations were also highly dissimilar to the  
250 microxic and dysoxic zones, with values of 68.58% and 65.91%, respectively (Figure 4).

### 251 3.3 Macro-nematodes density and diversity

252 On average, macro-nematodes were the most dominant taxon in this study as a result of their  
253 dominance in the dysoxic station. Nematodes were abundant in all oxygen zones recording relative  
254 abundances of 8%, 74%, and 24% in microxic, dysoxic, and oxic zones respectively (See Figure  
255 3).

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259 In our study, eighteen different genera of macro-nematodes were identified. Notably,  
260 *Desmolaimus* and *Paracomesoma* were detected across all stations and were the only genera  
261 present in microxic stations. These two, along with *Metoncholaimus*, exhibited their highest  
262 abundance in dysoxic stations. Additionally, in these dysoxic environments,  
263 *Paralongicyatholaimus* and *Neochromadora* recorded high abundances, each constituting (>4%)  
264 of the total population (Figure 6), *Thalassolaimus*, *Paramesacanthion*, *Enoploides*, *Halanonchus*,  
265 *Rhabdodemia*, and *Dorylaimopsis* recorded high abundances in oxic stations but were absent in  
266 dysoxic stations except for *Thalassolaimus*. *Metoncholaimus* and *Paralongicyatholaimus*, were  
267 present in dysoxic stations but absent in oxic stations, while *Paramesacanthion*, *Enoploides*, and  
268 *Rhabdodemia* were present in the oxic station and absent in the dysoxic stations. For the  
269 purposes of graphing the relative abundance, *Thoracostomopsis*, *Anticoma*, *Cephalanticoma*,  
270 *Trileptium*, *Mesacanthoides*, *Terschellingia*, and *Marylinnia* were grouped as ‘others’ as they  
271 recorded low abundances (<4%) and were absent in dysoxic stations except for *Marylinnia* and  
272 *Terschellingia*, whereby, the former was absent in the oxic station while the latter was present in  
273 both oxygen zones (See Figure 6).

274 The feeding guild composition exhibited variation across distinct oxygen zones within the study  
275 site. Selective epigrowth feeders and herbivores, categorized as Type 2A, emerged as the  
276 predominant nematodes within dysoxic zones, comprising 40% of the population. This was closely  
277 followed by non-selective deposit feeders (Type 1B), contributing to 35% of nematodes within the  
278 same zone. Carnivores and omnivores, classified as Type 2B, constituted 28% of the nematode  
279 population, while selective deposit feeders and bacteriovores, categorized as Type 1A, accounted  
280 for 10%. Conversely, in the oxic zone, non-selective deposit feeders (Type 1B) and selective  
281 epigrowth feeders and herbivores (Type 2A) predominated, representing 35% and 32% of the

282 nematodes, respectively. Carnivores and omnivores (Type 2B) and selective deposit feeders (Type  
283 1A) accounted for 18% and 15%, respectively (Figure 7). Within the feeding modes, selective  
284 deposit feeders had the largest differences in composition between the two oxygen zones, with the  
285 highest concentration observed in the oxic zones. While 2A and 2B had slightly higher values in  
286 the dysoxic zone compared to the oxic zones.

287 The nematode diversity exhibited a pattern akin to the overall macrofaunal diversity, wherein one  
288 of the microoxic stations (23002) was devoid of nematodes, while its counterpart station (23020)  
289 harbored only two nematodes. Consequently, station 23002 was excluded from the diversity  
290 analysis. Across the remaining oxygen ranges (dysoxic and oxic), a parallel trend was observed as  
291 in the broader macrofaunal analysis within the study area. Specifically, oxic stations displayed  
292 higher H' diversity (1.38±0.5) compared to dysoxic stations (0.81±0.84). Conversely, dysoxic  
293 stations exhibited greater dominance (0.59±0.39) relative to oxic stations (0.32±0.18) (Fig. 8).

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#### 294 **4.0 Discussion**

295 Upwelling systems are known for their high surface productivity and Oxygen Minimum Zones  
296 (OMZ), which impinge on the benthic zone creating strong oxygen gradients on the seafloor and  
297 acting as the dominant driver for benthos diversity in these zones (Zettler et al., 2013). Despite the  
298 Benguela Upwelling System (BUS) being recognized as one of the major Eastern Boundary  
299 Upwelling systems, there is limited information on the structure and composition of the benthic  
300 communities. Information on macrofauna communities in the BUS will not only improve the  
301 existing database on benthic fauna but also provide insight into how increasing hypoxic areas in  
302 the ocean might structure benthic communities.

304 In this study, we adapted Levin's grouping system (Levin, 2003), classifying the different stations  
305 into three zones based on the oxygen levels recorded (microxic zone (<0.1 ml l<sup>-1</sup>); dysoxic zone  
306 (0.1-1.0 ml l<sup>-1</sup>); oxic zone (>1.0 ml l<sup>-1</sup>), which was supported by step-wise regression. The  
307 structure and composition of the macrofauna communities varied among the various oxygen zones.  
308 Most macrofaunal studies identify Polychaeta as the most abundant macrofauna taxon in both oxic  
309 and hypoxic areas (Eisenbarth & Zettler, 2016; Soto et al., 2017). However, in this study,  
310 Polychaeta only had the highest relative abundance in the oxic zones. In the microxic zone, the  
311 abundant taxon was Cumacea, followed by Polychaeta. It is essential to note that numerically  
312 Polychaeta was the most abundant taxon in this oxygen zone, but the presence of many individuals  
313 of other taxa in microxic stations reduced their proportional representation. The presence of  
314 cumaceans in high abundance in the core OMZ, comparable to the microxic zone in this study, has  
315 been reported by Zettler et al., (2013) and Eisenbarth & Zettler (2016), who described them as  
316 possible opportunistic species colonizing permanent hypoxic areas from adjacent areas, and thus  
317 their abundance may be season-specific. Currie et al. (2018) attributed the presence of Cumacea  
318 and other macrofauna taxa in the BUS to the Sulfur-oxidizing bacteria, possibly providing a  
319 detoxified condition in this area. In this case, the mobility of the cumaceans gives them an  
320 advantage over other tolerant taxa like polychaetes and nematodes at the core and hence their high  
321 relative abundances at the OMZs core.

322 Some polychaete families have physiological adaptations to tolerate the low oxygen concentrations  
323 found in OMZs (Hanz et al., 2019; Joydas & Damodaran, 2014; Levin et al., 2009). At the microxic  
324 zone, polychaetes had the numerical abundance in the microxic stations despite cumaceans leading  
325 in relative abundance. In station 23002, where Cumacea was located, only two taxa were present.  
326 As a result, cumaceans accounted for 75% of the population despite having a density of only 932

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342 [ind. m<sup>-2</sup>](#). In contrast, Polychaeta in [Station 23020](#) had a numerical high of 4350 [ind. m<sup>-2</sup>](#), which is  
343 relatively higher than the number recorded by cumaceans in [Station 23002](#). However, the diversity  
344 of other taxa in this station was much higher, and as a result, despite the high density, Polychaeta's  
345 proportion of the whole population was reduced to 53%, hence the lower relative abundance.

346 In the dysoxic zone, the macro-nematoda was the most dominant taxon, out-competing the  
347 polychaetes in all the dysoxic stations, recording more than 70% relative abundance. Oxygen can  
348 cause shifts in community structure and trophic transfer (Neira et al., 2018), as evidenced in this  
349 study. Nematoda as a taxon has not received significant attention in the macrofaunal size range in  
350 most studies despite evidence of their presence therein (Joydas & Damodaran, 2014; Sharma et  
351 al., 2011). An increase in the size of nematodes to macrobenthic class sizes has been reported in  
352 chemosynthetic environments that experience similar characteristics as OMZs, i.e., low oxygen  
353 and high sulfidic contents (Vanreusel et al., 2010). Apart from the increase in size, OMZs also  
354 tend to enhance the regional dominance of tolerant organisms such as nematodes with high  
355 biomass recorded in response to organic matter inputs. The high abundances are thought to reflect  
356 the availability of organic matter, a significant nutrient source for macrofauna, coupled with a  
357 reduction in predation by larger fauna that are affected adversely by the reduced oxygen  
358 concentrations (Moens et al., 2013).

359 The high nematode abundance in partnership with Oligochaeta occurred in the dysoxic zone;  
360 oxygen conditions were low enough to exclude some taxa but sufficient for tolerant species to  
361 survive and reproduce. Such conditions are referred to as the 'edge effect,' and such high densities  
362 are characteristics of the edge of the OMZs, where various species have been observed to have  
363 abnormally high densities. The reasons for these high abundances are not well understood, but  
364 Gutiérrez et al. (2008) [suggested](#) that the nematodes' population can multiply in low oxygen

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374 conditions, which experience high loads of organic matter input. Despite their tolerance to anoxia,  
375 nematodes cannot survive long-term exposure, as observed in the anoxic zone (Moens et al., 2013).  
376 It is not clearly understood then whether the high abundance of the nematodes in one of the stations  
377 is characteristic of the study site or just congregation to a food source. Thus, the patchiness and  
378 high variability in the dysoxic zone call for more studies (Buhl-Mortensen et al., 2010).

379 At dysoxic sites (DO 0.1-1.0 ml l<sup>-1</sup>), other taxa like Ostracoda and Bivalvia were observed, albeit  
380 in low numbers. Despite ostracodes flourishing better in well-oxygenated marine areas, various  
381 families (Platycopina) have been observed to tolerate and thrive in Oxygen Minimum Zones. At  
382 oxic sites, where DO levels were above 1.0 ml l<sup>-1</sup>, more taxa were recorded, and these numbers  
383 increased even more when DO levels surpassed 2.0 ml l<sup>-1</sup>.

384 Our findings were in line with the various studies in OMZs whereby a decrease in richness and  
385 diversity of macrofaunal species is observed towards the core and vice versa (McClain &  
386 Schlacher, 2015). In the core (microxic) area, the macrofauna taxa had the lowest density and  
387 diversity. Each square meter of core area contained 1243 individuals, indicating low species  
388 richness. A similar number was recorded by Zettler et al., (2009). In our study, we also observed  
389 an increase in the number of taxa recorded in sites with DO levels above 1 ml l<sup>-1</sup>. The high  
390 dominance of the tolerant taxa, as evidenced in the microxic and dysoxic areas, was reduced. Taxa  
391 such as Amphipoda, Isopoda, Echinodermata, Nemertina, Aeolosomatidae, Aplacophora,  
392 Holothuroidea, and Cnidaria were only recorded in the oxic zones indicating low tolerance to low  
393 oxygen levels. Of this fauna, crustaceans were the most abundant. This conforms to the  
394 observations of Soto et al. (2017) at oxic sites in an upwelling system in Chile. Conversely, Zettler  
395 et al. (2009) recorded amphipod species in low-oxygen areas. These contradictory results indicate  
396 that at least amongst the Amphipoda, tolerance/intolerance to hypoxia is species-specific.

397 Macro-nematoda abundance varied across the OMZ, with very low abundance in the microxic  
398 zones, extremely high numbers at the dysoxic zones, and a substantial amount at the oxic stations,  
399 accounting for only 28% of the total abundance. Nematodes are considered one of the most tolerant  
400 taxa in the marine environment, with the ability to tolerate low oxygen and high sulphidic  
401 environments characteristic of OMZs and may reach very high abundances in these environments  
402 (Neira et al., 2018; Gutierrez et al., 2008). This was the case at the dysoxic zone, where nematodes  
403 recorded high abnormal densities in one station, indicating the high tolerance of these taxa and  
404 their ability to dominate the macrofaunal component. Even with such high tolerance levels,  
405 nematode abundance can be impacted by microxic conditions, as observed in the microxic areas  
406 with a recording of only 621 nematodes m<sup>-2</sup> in this oxygen zone. This low value, however, may be  
407 at the macrofauna level, and the case may be different at the meiofauna level, where nematodes  
408 have recorded substantial densities in microxic environments (Neira et al., 2018; Steyaert et al.,  
409 2007).

410 Despite their high abundance in OMZs, not all nematodes are tolerant to low oxygen levels (Moens  
411 et al., 2013), as observed in this study. *Metoncholaimus*, *Paracomesoma*, and *Desmolaimus*  
412 dominated the dysoxic zone; these three genera are members of Oncholaimidae, Comesomatidae,  
413 and Linhomoeidae, respectively. Members of the family Oncholaimidae have large bodies that can  
414 disperse rapidly and colonize carcasses of macrofauna and even fish that may have succumbed to  
415 the low levels of oxygen found in the dysoxic zone (Moens et al., 2013). Nevertheless, their bodies  
416 are large enough to fit within the macrofauna size range, while their ability to swim ensures they  
417 actively locate their food source (Moens et al., 2013). Their high abundance in this study might  
418 reflect a congregation upon a food source that had attracted nearby members in large numbers.

419 Their ability to colonize the 'food source' in such numbers in a dysoxic environment indicates their  
420 tolerance to low oxygen levels.

421 On the other hand, Comesomatidae and Linhomoeidae members have been noted to have high  
422 abundances in enriched sediments with low oxygen, indicating tolerance to anoxic conditions  
423 (Steyaert et al., 2007). Their long and slender bodies might be the reason for their records at the  
424 macrofauna level. Despite this generalization at the family level and the assumptions that members  
425 of the same family may portray similar life strategies (Bongers et al., 1991), tolerance of nematodes  
426 to hypoxia is species-specific (Moens et al., 2013) as Steyaert et al. (2007) observed members of  
427 the same genera (*Sabatiera*) reacting differently to hypoxic and anoxic conditions. Thus, further  
428 analysis should be done to identify the species that are tolerant to hypoxia at these OMZs at  
429 macrofauna levels. Tolerance to hypoxia is indicated by both the presence and absence of taxa;  
430 most genera present in the oxic zone were absent in the dysoxic area and may be seen as genera  
431 intolerant to low oxygen levels.

432 Wieser's feeding types, as outlined in his study (Wieser, 1953), have long been used to assess the  
433 trophic structure of nematode communities. This study had a high abundance of non-selective  
434 feeders (1B) and selective epigrowth feeders and herbivores (2A), with the latter dominating the  
435 dysoxic zone, while the former dominating the oxic zones.

436 Non-selective deposit feeders are commonly observed to dominate substrates characterized by  
437 high levels of organic matter and smaller grain size, indicating their adaptation to environments  
438 with elevated organic matter input, as observed in this study (Singh and Ingole, 2016; Heip et al.,  
439 1985). In contrast to previous findings on meiofaunal nematodes from Oxygen Minimum Zones  
440 (OMZs) (Singh & Ingole, 2016; Neira et al., 2013) and macro-nematodes in deep-sea habitats

441 (Sharma & Bluhm, 2011), which reported lower abundances of selective epigrowth feeders, our  
442 study revealed a higher abundance of selective epigrowth feeders and herbivores. Within OMZs,  
443 such as our study site, the presence of giant bacteria such as *Thioploca*, *Beggiatoa*, and  
444 *Thiomargarita* forming extensive mats (Brüchert et al., 2009) may provide a substantial food  
445 source for nematodes (Giere, 2008).

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446 Adjacent to OMZs, the prevalence of diatomaceous mud and the settling of fresh phytodetritus  
447 contribute significantly to the diet of epistratum feeders, leading to their high abundance in dysoxic  
448 areas (Singh & Ingole, 2016; Sharma & Bluhm, 2011; Moens & Vincx, 1987; Heip et al., 1985).  
449 However, in regions offshore from the OMZ, where the OMZ no longer interacts with the benthic  
450 zone, reduced diatom production diminishes the abundance of epistratum feeders, resulting in the  
451 dominance of non-selective deposit feeders.

452 While non-selective deposit feeders typically dominate muddy and silty environments, selective  
453 deposit feeders and carnivores have been observed to exhibit lower abundances in these habitats  
454 (Heip et al., 1985). Our study aligns with this trend; however, other investigations have reported  
455 contradictory findings, with selective deposit feeders being identified among the most dominant  
456 nematodes in similar environments (Neira et al., 2013; Singh & Ingole, 2016). Additionally,  
457 carnivores and omnivores (2B) exhibited higher abundances in the dysoxic zone compared to the  
458 oxic zone, with their overall abundance surpassing that of selective deposit feeders. These  
459 observations suggest potential differences in nematode composition between the meiofaunal and  
460 macrofaunal levels. Sharma and Bluhm (2011) observed that category 2B feeders were the second  
461 most abundant feeding type in numerous stations within the Arctic deep-sea Canada Basin. They  
462 attributed these findings to the omission of larger nematodes in previous studies, as carnivores and  
463 omnivores are often large-bodied species.

464 **5.0 Conclusion**

465 In conclusion, this study provides insights into the intricate dynamics of macrofaunal communities  
466 in Oxygen Minimum Zones (OMZs). By classifying stations into microoxic, dysoxic, and oxic  
467 zones based on oxygen levels, we observed variations in macrofaunal composition and abundance  
468 across these zones. Contrary to typical findings, Polychaeta dominated only in oxic zones, while  
469 Cumacea and Nematoda were prevalent in microoxic and dysoxic zones, respectively. The presence  
470 of tolerant taxa like nematodes and Oligochaeta in dysoxic zones suggests their ability to thrive  
471 under low oxygen conditions. Conversely, taxa intolerant to low oxygen were absent in dysoxic  
472 zones, highlighting the importance of oxygen levels in shaping macrofaunal communities.

473 Macro-nematodes in general were the most abundant taxon. The high nematode densities suggest  
474 the ability of macro-nematodes to tolerate and thrive in low-oxygen environments. Our findings  
475 highlight the importance of considering macro-nematodes in studies of OMZ ecosystems and  
476 underscore the need for further research to elucidate their ecological roles and responses to  
477 environmental changes. Understanding the dynamics of macro-nematode populations coupled  
478 with their meiofaunal counterparts in OMZs is crucial for comprehensively assessing the impacts  
479 of oxygen depletion on benthic communities and ecosystem functioning.

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489 **6.0 Tables**

490 **Table 1:** Station information and abiotic factors information recorded from the stations in the Northern  
491 Benguela Upwelling System. NR stands for no replicate.

Transect	Station (No. Replicate)	Longitude	Latitude	Depth (m)	TOM(%)	DO (ml l <sup>-1</sup> )
Off Terrace Bay (OTB-20°S)	20002(2)	12.99905	-20	33	27.47±1.55	1.8
	20020(3)	12.67858	-20	125	38.64±2.16	0.67
	20040(N.R)	11.79321	-20	219	17.58 nr	0.53
Off Walvis Bay (OWB-23°S)	23002(NR)	14.3734	-23	39	23.08 nr	0.06
	23020(NR)	14.06986	-23	128	20.88 nr	0.02
	23070(3)	13.14	-23	318	9.89±1.55	2.3
Off Luderitz (OL 26°S)	26090(NR)	13.28	-26	1282	4.4 nr	4.34

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499 Table 2: Sediment size distribution in the Northern Benguela Upwelling System

Transect	Station	Very Coarse Sand	Coarse Sand	Medium sand	fine sand	very fine sand	Silt
Off Terrace Bay (20°S)	20002	0.65	0.67	13.81	34.82	21.29	28.76
	20020	0.93	0.62	28.17	30.89	12.98	26.42
	20040	2.71	4.37	18.55	33.33	17.04	23.98
Off Walvis Bay (23°S)	23002	0.00	0.28	18.74	36.17	17.25	27.57
	23020	0.98	6.85	14.91	38.88	18.83	19.56
	23070	0.00	0.00	13.92	33.73	20.29	32.06
Off Luderitz (26°S)	26090	0.00	0.29	23.99	30.46	19.25	26.01

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## 7.0 Figures

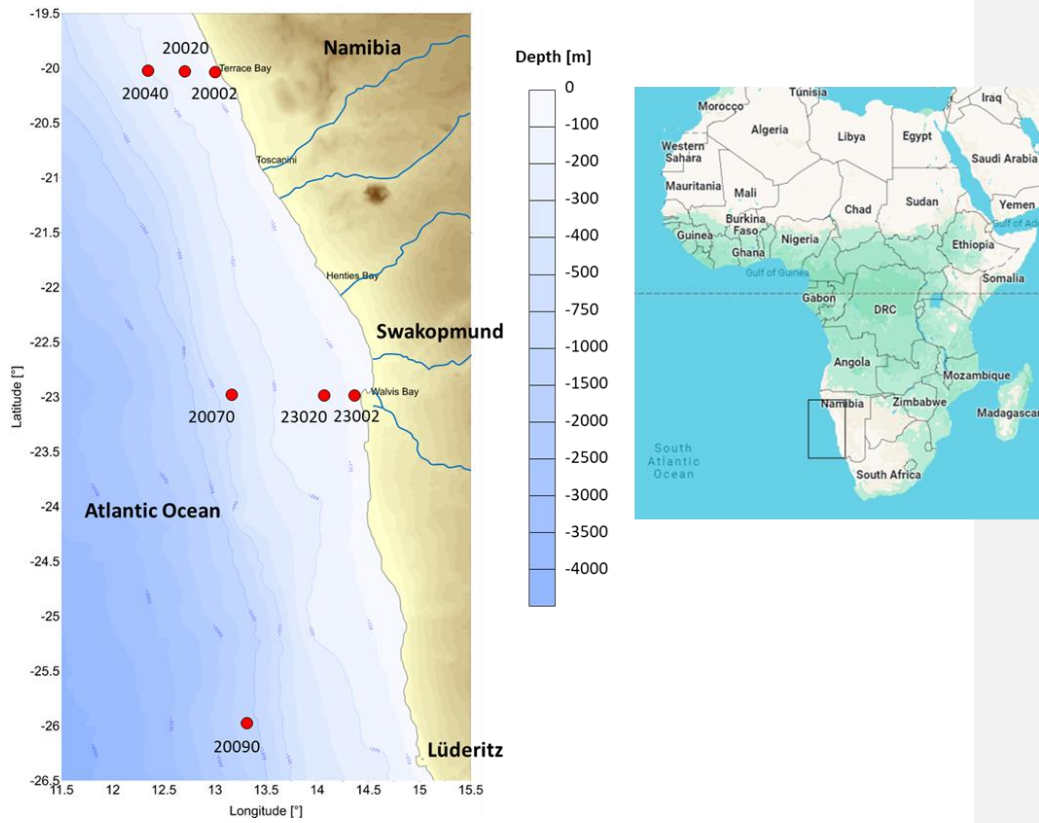


Figure 1: Sampling stations located across the Northern Benguela Upwelling System

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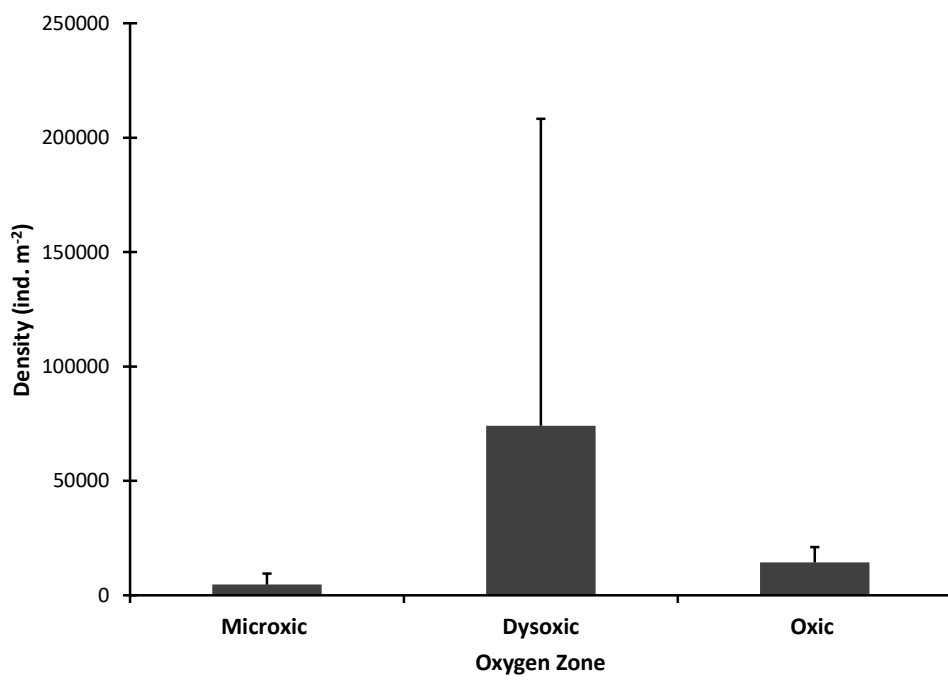


Figure 2: Macrobenthic densities in the different oxygen zones in the Northern Benguela Upwelling system. [Error bars represent standard deviation \(+1 SD\).](#)

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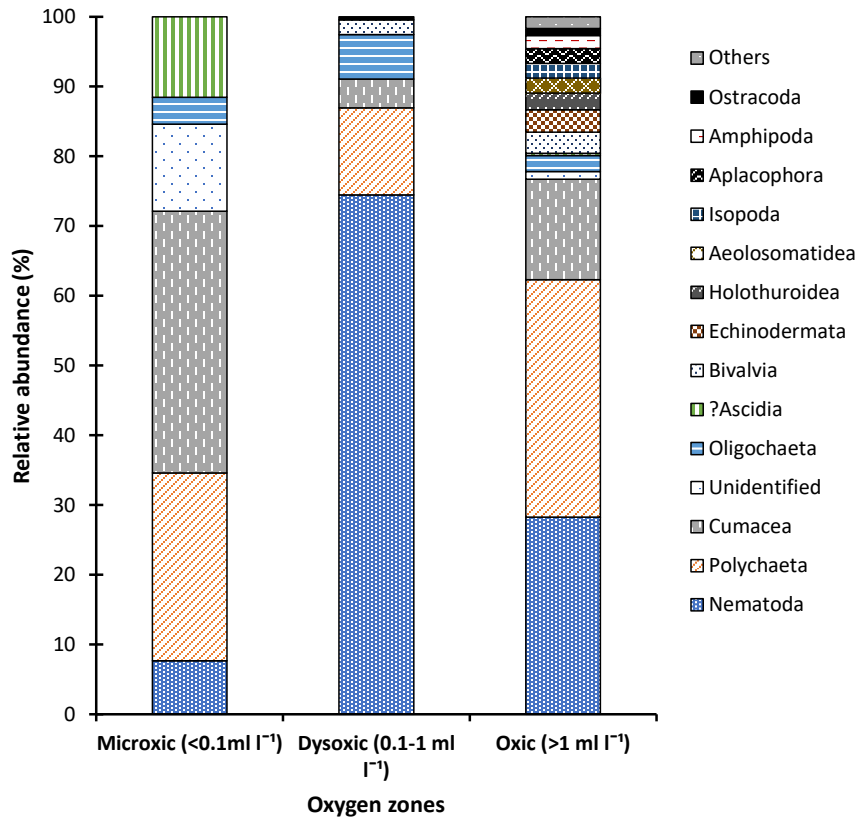


Figure 3: Macrofauna relative abundance in the Northern Benguela Upwelling System, based on the oxygen zones identified.

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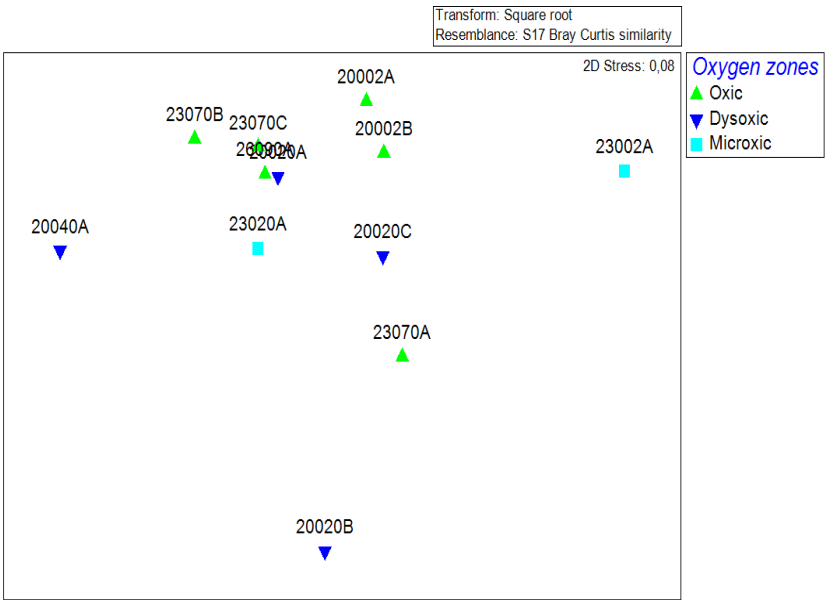


Figure 4: Non-metric multidimensional scaling (nMDS) plot based on Bray-Curtis Similarity index of macrobenthic fauna communities recorded in the Northern Benguela Upwelling System.

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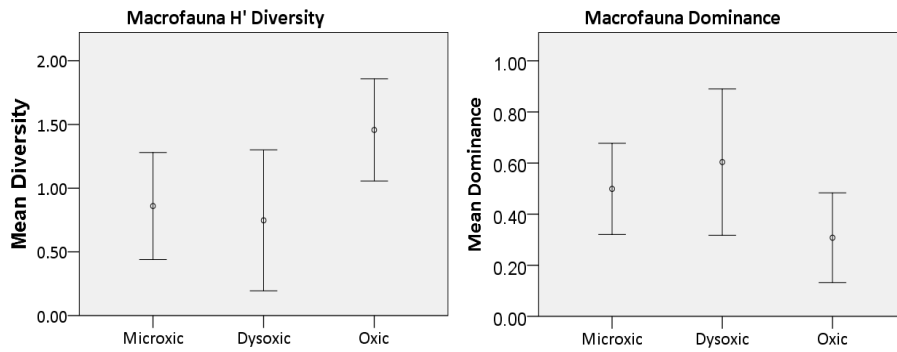


Figure 5: Macrofauna diversity indices recorded in the various oxygen zones in the Northern Benguela Upwelling system. H' diversity, calculated using the natural logarithm (loge), while dominance, represented the complement of the Simpson Index (1-Simpson index). Error bars represent standard deviation (+/-1 SD).

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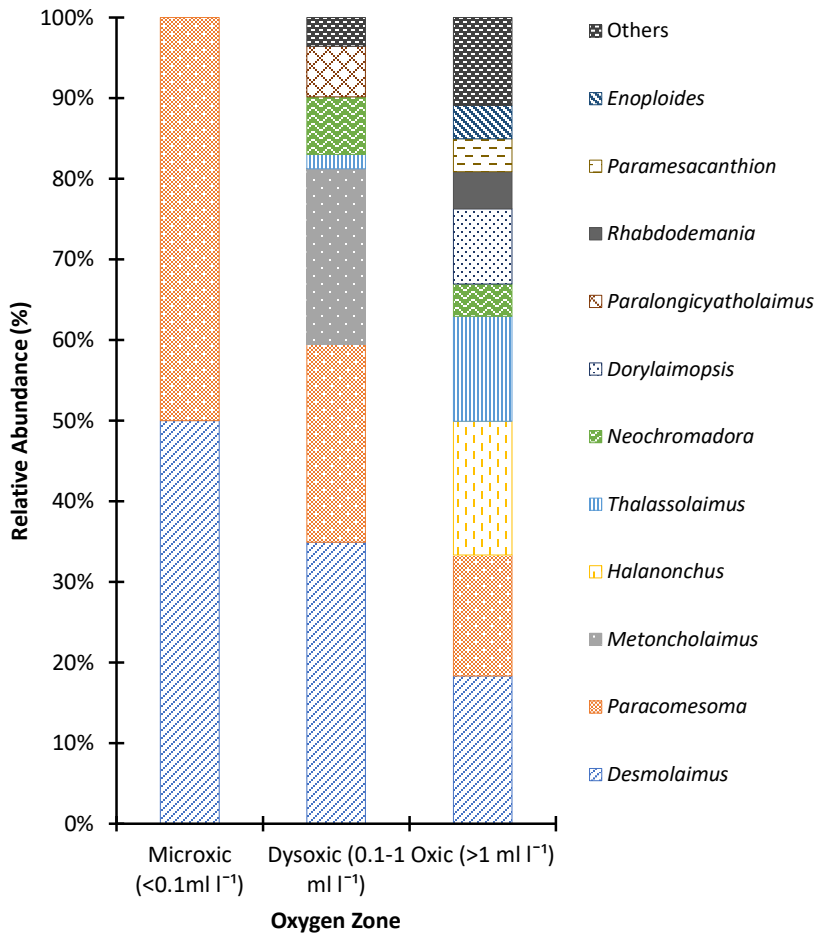


Figure 6: Macroneumatoda relative abundance in the Northern Benguela Upwelling System, based on the oxygen zones identified.

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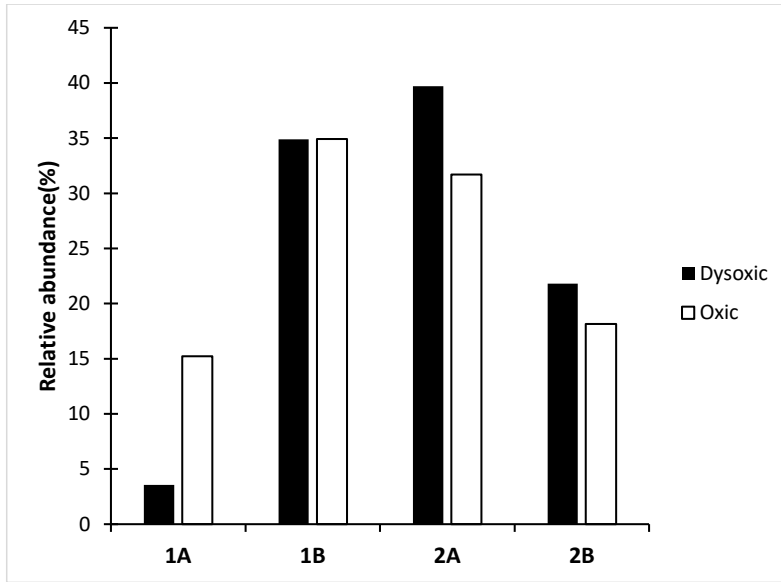


Figure 7: Feeding guilds based on Wieser (1953) of macro-Nematoda from the Northern Benguela Upwelling System. (1A= Selective deposit feeder, 1B- Non-Selective deposit feeder, 2A=Epigrowth feeders, 2B=Predators/Omnivores).

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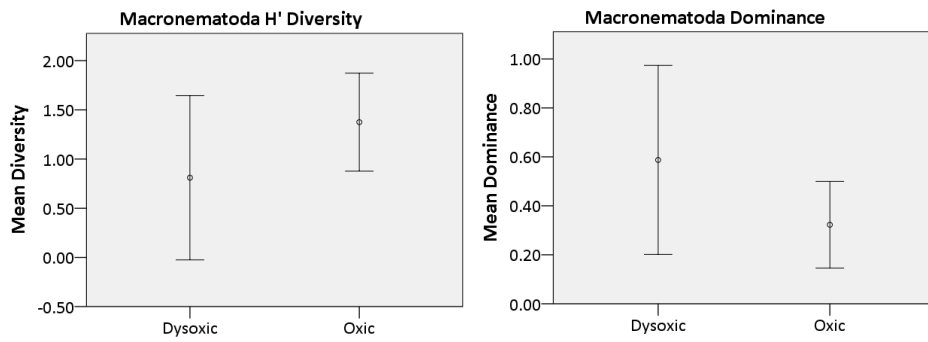


Figure 8: Macro-Nematoda diversity indices recorded in the various oxygen zones in the Northern Benguela Upwelling system. [H' diversity](#), calculated using the natural logarithm ( $\log_e$ ), while [dominance](#), represented the complement of the Simpson Index (1-Simpson index). Error bars represent standard deviation ( $\pm 1$  SD).

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## **8.0 Code/Data Availability**

Currently, the data is not available but can be released upon request to the authors.

## **9.0 Author contribution**

HS was responsible for the conceptualization of the study. HS and WB conducted the investigation, formal analysis, and drafting of the original draft. MA was responsible for supervision, review, and editing of the final draft.

## **10.0 Competing interests**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## **11.0 Special Issue Statement**

Part of this manuscript was presented as a poster presentation during the 53<sup>rd</sup> International Colloquium on Ocean Dynamics: 3rd GO2NE Oxygen Conference held in Liege, 2022. As a result, the corresponding author was invited to submit a manuscript for the special issue "*Low-oxygen environments and deoxygenation in open and coastal marine waters*" as part of the journal Biogeosciences.

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(NatMIRC) was instrumental in successfully sampling, processing, and transporting samples. The crew of the sampling vessel R/V Mirabilis ensured a smooth sampling experience, and hence we take this opportunity to appreciate their services.

We also extend our gratitude to the reviewers for their invaluable feedback and constructive criticism, which significantly enhanced the quality and clarity of this manuscript. We deeply appreciate their time, expertise, insights, suggestion, and dedication to improving the manuscript. Their contributions have undoubtedly strengthened the final version of this manuscript, and we are sincerely grateful for their inputs.

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