

1 **Influence of Oxygen Minimum Zone on Macrobenthic Community Structure in the**  
2 **Northern Benguela Upwelling System: A Macro-Nematode Perspective**

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11 **Abstract**

12 Understanding the dynamics of marine ecosystems, particularly the benthic communities, is crucial  
13 for predicting the impacts of climate change and human activities on oceanic biodiversity. In this  
14 context, we conducted a study on the macrobenthic communities in the Benguela Upwelling  
15 System (BUS). Macrobenthic samples were collected offshore Namibia on board *R/V Mirabilis*  
16 during the 3<sup>rd</sup> RGNO training and the National Marine Information and Research Centre's  
17 (NatMIRC's) plankton survey from 13<sup>th</sup> May to 17<sup>th</sup> May 2016. Two transects, Terrace Bay (20°  
18 S) and Walvis Bay (23° S), hosted three stations each, while the third transect, Luderitz (26° S),  
19 hosted only one station. From the results, three oxygen zones were identified, namely Microxic  
20 (<0.1 ml l<sup>-1</sup>), Dysoxic (0.1-1.0 ml l<sup>-1</sup>), and Oxic (>1.0 ml l<sup>-1</sup>). A total of 20 macrobenthic taxa were  
21 identified; Nematoda, Polychaeta, Cumacea, and Oligochaeta were the most dominant taxa and  
22 were present in all oxygen zones. Eighteen genera of macro-nematoda were identified;  
23 *Desmolaimus* and *Paracomesoma* dominated in all oxygen zones, *Metoncholaimus* recorded  
24 higher abundance in the dysoxic stations, and were absent in the oxic stations, and the opposite  
25 was observed for *Halanonchus* and *Dorylaimopsis*. H' diversity values for both the general  
26 macrofauna and macro-nematoda were higher in the oxic stations and lower in both the dysoxic  
27 and microxic stations, while an opposite trend was observed for dominance values. Density values  
28 were lower in microxic stations and higher in dysoxic stations, while at the oxic stations, the  
29 density values fell in between the two hypoxic zones. In conclusion, this study provides an  
30 overview of the distribution, diversity, and response to varying oxygen conditions of macrobenthic  
31 communities and their importance in marine ecosystems.

- 32 **Keywords:** Macrofauna, Macro-Nematoda, Oxygen minimum zone, Benguela Upwelling System
- 33 (BUS), Dissolved oxygen, hypoxia

## 34 **1.0 Introduction**

35 In recent decades, the concentration of dissolved oxygen (DO) in the ocean, specifically in the  
36 tropics, has been decreasing. This not only increases the size of areas under hypoxia but also their  
37 prevalence (Breitburg et al., 2018). Hypoxia is ranked among the major threats to the actualization  
38 of the blue economy and the achievement of the 14<sup>th</sup> pillar of the Sustainable Development Goals  
39 (SDGs), specifically target 14.7 which aims at increasing the economic benefits of marine resource  
40 utilization through its sustainable use by developing countries (Rossi, 2023; Liu, 2023). Although  
41 hypoxia can result from natural phenomena like upwelling and thermal stratification, the current  
42 expansion of hypoxic areas is mainly a result of accelerated nutrient inputs in coastal areas, which  
43 increases algal production subsequently resulting in higher organic matter production which in  
44 turn results in increased aerobic microbial decomposition lowering the levels of DO in the water  
45 (Gobler & Baumann, 2016). It has been projected that such changes may affect different organisms  
46 differently depending on their tolerance and reactions to lower dissolved oxygen in their habitats  
47 (Rudolph et al., 2020). Studies on ecosystems with hypoxia as a natural phenomenon can assist in  
48 predicting and understanding how human-induced hypoxia might affect and shape marine  
49 ecosystems in the face of the expansion of marine areas under hypoxia.

50 The Benguela upwelling system (BUS) is located off the southwest coast of Africa. It extends from  
51 Cape Frio in Angola to the southern tip of the continent in Cape Agulhas, South Africa, and is one  
52 of the most productive regions of the world's ocean (Magalhães, 2018). The high productivity  
53 provides a huge source of carbon resulting from photosynthesis, which gradually sinks through the  
54 water column resulting in a rapid oxygen loss due to biochemical oxygen consumption. This  
55 consequently causes low dissolved oxygen concentrations and thus creates a permanent extensive

56 shallow oxygen minimum zone (Bohata & Koppelman, 2013; Emeis et al., 2018). When the  
57 oxygen minimum zone (OMZ) comes into contact with the seafloor, it creates a strong oxygen  
58 gradient at the benthic zone at depths ranging between 50 meters and 300 meters, resulting in a  
59 hypoxic ( $<0.5 \text{ ml l}^{-1}$ ) inner shelf (Levin, 2003). The oxygen gradient created at the OMZ's benthic  
60 zone is believed to primarily regulate the benthic community distribution and diversity patterns  
61 (Zettler et al., 2013; Teuber et al., 2013). In these zones, microbial communities play a key role in  
62 the cycling of elements like carbon, nitrogen, and sulfur. The expansion of OMZs may increase  
63 nitrate use by nematode-associated microbes such as SAR11, a significant player in the marine  
64 nitrogen cycle. These nematodes and the communities they are part of influence biogeochemical  
65 processes and ecosystem dynamics within OMZs. For this reason, understanding these  
66 communities is crucial, especially with the ongoing expansion of OMZs due to climate change.

67 The general trend observed in most OMZs in global oceans namely, Walvis Bay, Namibia (the  
68 location of this study), California, USA, and Oman margin (off the Arabian Peninsula) indicates  
69 that the densities of macrofauna (benthic organisms that are typically retained in a 0.5 mm sieve  
70 but pass through a 2.00 mm sieve (Bachelet, 1990)) generally display a negative response to  
71 reducing oxygen levels within the OMZ with a 30% to 70% reduction in densities in regions with  
72 less than  $0.15 \text{ ml l}^{-1}$  (Levin, 2003). Similarly, diversity reduces as oxygen levels reduce within the  
73 OMZ because of the loss of intolerant species and increased dominance of the tolerant species  
74 (Currie et al., 2018; Levin et al, 2009). Nematodes and some families from the annelid worms have  
75 been observed to be able to tolerate low oxygen, with Nematoda (in the meiofauna group) having  
76 95-99% abundance. Some polychaetes families like Spionidae, Dorvilleidae, and Lumbrineridae  
77 can also tolerate low oxygen in the OMZ by having a high gill surface area for increasing oxygen

78 uptake. On the other hand, harpacticoid copepods are the most sensitive taxon to hypoxia (Levin  
79 et al., 2009; Zeppilli et al., 2015).

80 It has been observed that the macrofauna diversity is lowest off Walvis Bay, a city located on the  
81 western coast of Namibia, attributed to the perennial intense OMZ over the shelf (Currie et al.,  
82 2018). In contrast, the diversity increases significantly northwards off the Kunene River (Zettler  
83 et al., 2013), which flows from the highlands of Angola, along the border with Namibia, and into  
84 the Atlantic Ocean (See Figure 1). This increase in diversity is proposed to be a result of moving  
85 away from the intense OMZ cells off Walvis Bay and also the reflection of the biogeography where  
86 diversity increases with lower latitude (Zettler et al., 2009). Outside the OMZ, bathymetry, and  
87 latitude are said to be the factors affecting the infaunal communities at the Namibian shelf (Steffani  
88 et al., 2015).

89 This study aims to identify the relationship between the levels of dissolved oxygen and the  
90 macrobenthic community distribution across the Northern Benguela Upwelling system as a guide  
91 on how the expansion of man-made hypoxia might influence the benthic fauna distribution on the  
92 seafloor.

## 93 **2.0 Material and Methods**

### 94 **2.1 Study Area**

95 The study area was located across the Northern Benguela Upwelling System (between 26° S and  
96 20° S) along the Namibian continental shelf, which hosts a deep continental shelf (around 300 m)  
97 (Figure 1). The intense upwelling in the study site has rendered the area highly productive,  
98 resulting in a characteristic shallow OMZ (50-200 m) with stronger productive cells around  
99 Luderitz and Walvis Bay (Bohata & Koppelman, 2013). The inner shelf is described to be

100 extremely oxygen-depleted, caused by in situ organic matter decomposition and warm poleward  
101 Angola current, which peaks in June-July while the continental slope below the OMZ is better  
102 oxygenated (Levin et al., 2009; Emeis et al., 2004).

103 The benthic zone in the OMZ in Northern BUS is characterized by extensive areas of diatomaceous  
104 mud, which are associated with high primary production at the ocean surface and low  
105 concentration of dissolved oxygen (Levin et al., 2009; Steffani et al., 2015). These low oxygen  
106 levels, as per the findings of Levin et al. (2009) were found to be less than  $0.5 \text{ ml l}^{-1}$  in 55% of the  
107 total shelf. Furthermore, extreme anoxia, defined as oxygen concentrations less than  $0.02 \text{ ml l}^{-1}$ ,  
108 was observed over an expansive area of almost  $900 \text{ km}^2$ .

## 109 **2.2 Sample Collection**

110 Samples were collected from three transects (off Luderitz (OL)  $26^\circ \text{ S}$ , off Walvis Bay (OWB)  $23^\circ$   
111  $\text{S}$ , and off Terrace Bay (OTB)  $20^\circ \text{ S}$ ) onboard *R/V Mirabilis* during the RGNO training National  
112 Marine Information and Research Centre's (NatMIRC's) plankton survey from 13<sup>th</sup> May to 17<sup>th</sup>  
113 May 2016. The sampling stations were located at 02 nm, 20 nm, 40 nm, or 70 nm from the shore  
114 at each transect, with the  $26^\circ \text{ S}$  transect hosting only one station at 90 nm (Figure 1). However,  
115 benthic samples from these stations were dependent on the prevailing weather conditions and the  
116 ability to get good core samples (Table 1).

117 Sampling was done using a multi-corer from which individual cores were taken and sub-sampled  
118 for macrobenthos using a 6.4 cm diameter corer, and sediment samples for granulometry and  
119 organic matter analysis were taken simultaneously. Replicate samples were taken from a  
120 subsequent deployment of the multi-corer, where possible, to avoid pseudoreplication. The

121 macrofauna cored samples were put in sampling bottles and immediately fixed with 5% buffered  
122 formalin, while samples for organic matter analysis were frozen to arrest microbial decomposition.  
123 Depth and temperature measurements were collected from a probe attached to the multi-corer,  
124 while dissolved oxygen concentrations in this study used the Winkler method from the overlying  
125 water (Montgomery et al., 1964).

### 126 **2.3 Laboratory analysis**

127 In the laboratory, macrobenthic samples were sieved between 2.00 mm and 0.45 mm sieves.  
128 Studies typically employ a sieve size of 0.5mm, our study however utilized a 0.45mm sieve due  
129 to its availability during the course of our research. This size, although slightly smaller, falls within  
130 the range of commonly used sieve sizes (300 to 500 microns) for macrofaunal research. The use  
131 of a 0.45mm sieve is also not unprecedented in macrofaunal studies; studies by Li et al. (2018) and  
132 Zhang et al. (2022) have successfully employed this sieve size. Therefore, while acknowledging  
133 the slight deviation, results remain valid and comparable to those obtained using the more common  
134 sieve sizes.

135 The sieve fraction retained in the 0.45 mm mesh sieve was then preserved in a 5% buffered  
136 formalin solution, with 3-5 drops of Rose Bengal solution added to aid in sorting. After rinsing the  
137 samples with water, they were sorted under a dissecting microscope using the key provided by  
138 Thiel & Higgins (1988) to the class or order levels and then counted. Subsequently, the nematodes  
139 were pin-picked, fixed on permanent slides, and identified to the genus level using the key from  
140 Platt & Warwick (1988). Feeding types were determined for these genera following the  
141 methodology described by Wieser (1953), whereby nematodes are classified based on their buccal  
142 morphology, including shape, size, and presence or absence of buccal armature, resulting in four



143 groups. Those lacking buccal armature are categorized as deposit feeders (group 1), they are further  
144 subdivided into selective deposit feeders (1A) and non-selective deposit feeders (1B) based on the  
145 size of their buccal cavity, with the former possessing a very small buccal cavity, and the latter  
146 having a larger one (Moens and Vincx, 1997; Moens et al., 2013). Nematodes with buccal armature  
147 constitute group 2 and can be selective epigrowth feeders and herbivores (2A), using small teeth  
148 to graze diatoms, microalgae, and bacteria, or carnivores and omnivores (2B), characterized by the  
149 possession of teeth, mandibles, or onchia for piercing and consuming prey (Moens et al., 2013)

150 For sediment granulometry, samples were wet-sieved using a 1.00 mm sieve, 0.5 mm, 0.25 mm,  
151 105  $\mu\text{m}$ , 0.063 mm, and those below 0.063 mm ( $<0.063 \text{ mm}$ ). The sediments collected at the  
152 different sieves were then put on pre-weighed Petri dishes and dried in an oven at 60 °C until a  
153 constant weight was reached, and these dried weights were used for sediment granulometry  
154 (Wentworth, 1922).

155 The samples for total organic matter (TOM) were dried in an oven at 60 °C until a constant weight  
156 was achieved, a known weight of each sample was then placed in labeled aluminum foil, and the  
157 loss of weight on ignition (LOI) technique was used to determine TOM from the study site (Thiel  
158 & Higgins, 1988).

## 159 **2.4 Data analysis**

160 Out of the seven sampled stations, only three had replicates (Table 1), making direct statistical  
161 comparisons between stations impossible. To overcome this limitation, the sampling stations were  
162 grouped based on the measured environmental variable that provided the most significant  
163 relationship identified through step-wise regression analysis with the various biotic indices  
164 documented within the macrofauna community (See supplementary material (Tables S1-S4)). The

165 communities within these key 'habitat types' were then compared using the indices and analyses  
166 outlined below;

167 Grouping Stations; Following the approach of (Levin, 2003), stations with DO levels less than 0.1  
168 ml l<sup>-1</sup> were grouped as 'microxic,' those with DO levels between 0.1 and 1.0 ml l<sup>-1</sup> were grouped  
169 as 'dysoxic' and those with DO levels of 1.0 ml l<sup>-1</sup> and above grouped as 'oxic'. This approach was  
170 taken to investigate the role of DO levels in these communities. It was further justified by the  
171 application of step-wise regression using SPSS, which indicated that DO was a key determinant in  
172 macrobenthic community structures ( $R^2=0.521$ ,  $P=.012$ ) (Table S2(a); S2(b)).

173 Abundance Analysis; The data recorded were analyzed for abundance (density and relative  
174 abundance) in Excel. Additionally, the Paleontological Statistics Software package (PAST v2.17c)  
175 (Hammer et al., 2001) was used to calculate the diversity and the diversity t-test between the  
176 stations. For this study, H' diversity is calculated using the natural log (log<sub>e</sub>) while dominance was  
177 the complement of the Simpson Index (1-Simpson index).

178 Community Similarity and Diversity; Bray Curtis similarity, Analysis of Similarity (ANOSIM),  
179 and Similarity percentages (SIMPER) were conducted using Plymouth Routines in Multivariate  
180 Ecological Research (PRIMER v5.2.9) (Clarke & Gorley, 2005).

## 181 **3.0 Results**

### 182 **3.1 Abiotic variables**

183 Total Organic Matter (%TOM) demonstrated an inverse relationship with depth, with higher  
184 organic matter values recorded in the shallower stations. For example, the shallowest stations  
185 20020, 20002, and 23002 had the highest TOM of 38.6%±2.16, 27.5%±1.55 and 23.1%,

186 respectively whilst the lowest TOM values were found in most offshore stations, stations 23070  
187 and 26090, with  $9.89 \pm 1.55$  and 4.4, respectively (Table 1). The lowest oxygen values were  
188 recorded on the OWB 23°S transect, with the two most onshore stations (23020 and 23002)  
189 recording the lowest levels of oxygen ( $0.02$  and  $0.06 \text{ ml l}^{-1}$ , respectively). The inshore station from  
190 transect OCF 20°S (20002) had a higher DO ( $1.8 \text{ ml l}^{-1}$ ) than stations 20020 and 20040 from the  
191 same transect, where the DO levels recorded were  $0.67$  and  $0.53 \text{ ml l}^{-1}$ , respectively, while the  
192 offshore stations 23070 in transect OWB 23°S and 26090 in transect OL 26°S recorded the highest  
193 dissolved oxygen ( $2.30$  and  $4.34 \text{ ml l}^{-1}$ , respectively) (Table 1).

194 Fine sand was the most common sediment size ranging between 30-38.8% in all stations. Most  
195 stations from the sites also recorded higher proportions of medium sand and silt, except for station  
196 23020 in transect OWB 23°S, which recorded a lower proportion of silt and an increased abundance  
197 of coarse sand compared to the other stations (Table 2).

### 198 **3.2 Macrobenthic assemblages**

199 Macrofaunal densities differed significantly across the various oxygen zones (ANOVA;  $P < 0.05$ ).  
200 In the microxic stations, very low densities were observed ( $4,661 \pm 4,834 \text{ ind. m}^{-2}$ ) (Figure 2), and  
201 the lowest number of taxa was recorded, with only six taxa present. These included Nematoda,  
202 Polychaeta, Oligochaeta, and Cumacea, the only peracarid crustacean found across all oxygen  
203 zones (Figure 3). Despite the low number of taxa ( $S=6$ ), these stations recorded slightly higher  $H'$   
204 diversity than the dysoxic stations, while the dominance index was intermediate ( $0.5 \pm 0.18$ ) (Figure  
205 5).

206 Dysoxic stations in contrast recorded the highest densities ( $74,108 \pm 134,126 \text{ ind. m}^{-2}$ ) (Figure 2).  
207 One station within this Oxygen range, station (20040), recorded an extremely high density of

208 274,991 ind. m<sup>-2</sup>, primarily due to a high abundance of Nematoda and Oligochaeta (Figure 3).  
209 Although the taxa in the dysoxic stations were similar to those of the microxic stations, the  
210 composition and dominance differed significantly. The macro-nematoda was the most dominant  
211 taxon in this zone, with numbers reaching as high as 233,354 ind. m<sup>-2</sup> in one of the stations.  
212 Interestingly, Bivalvia and Ostracoda, absent in the microxic stations, were present here, albeit in  
213 low abundance (Figure 3). Dominance was highest in these stations (0.6±0.28) (Figure 5).

214 Oxic stations demonstrated the highest number of taxa (18) and the second-highest average density  
215 (14,345±6,726 ind. m<sup>-2</sup>) (Figure. 2). These stations recorded taxa like Echinodermata  
216 (Ophiuroidea), Holothuroidea, Aeolosomatidea, Isopoda, Aplacophora, and Amphipoda, which all  
217 had average abundances of more than 1%. Nemertina, Turbellaria, and Cnidaria were also recorded  
218 in these stations; however, their abundances were below 1%, and they were grouped as 'others'  
219 (Figure 3). These stations recorded the highest H' diversity (1.46±0.4) and the lowest dominance  
220 (0.31±0.18) (Figure 5).

221 All the oxygen zones were dissimilar to one another based on multivariate community analysis  
222 using Bray-Curtis analysis of dissimilarity. The highest dissimilarity was observed between the  
223 dysoxic and microxic zones which were 77.99% dissimilar despite both stations being  
224 characterized by low dissolved oxygen levels. The oxic stations were also highly dissimilar to the  
225 microxic and dysoxic zones, with values of 68.58% and 65.91%, respectively (Figure 4).

### 226 **3.3 Macro-nematodes density and diversity**

227 On average, macro-nematodes were the most dominant taxon in this study as a result of their  
228 dominance in the dysoxic station. Nematodes were abundant in all oxygen zones recording relative

229 abundances of 8%, 74%, and 24% in microxic, dysoxic, and oxic zones respectively (See Figure  
230 3).

231 In this study, eighteen different genera of macro-nematodes were identified. Notably,  
232 *Desmolaimus* and *Paracomesoma* were detected across all stations and were the only genera  
233 present in microxic stations. These two, along with *Metoncholaimus*, exhibited their highest  
234 abundance in dysoxic stations. Additionally, in these dysoxic environments,  
235 *Paralongicyatholaimus* and *Neochromadora* recorded high abundances, each constituting (>4%)  
236 of the total population (Figure 6), *Thalassolaimus*, *Paramesacanthion*, *Enoploides*, *Halanonchus*,  
237 *Rhabdodemia*, and *Dorylaimopsis* recorded high abundances in oxic stations but were absent in  
238 dysoxic stations except for *Thalassolaimus*. *Metoncholaimus* and *Paralongicyatholaimus*, were  
239 present in dysoxic stations but absent in oxic stations, while *Paramesacanthion*, *Enoploides*, and  
240 *Rhabdodemia* were present in the oxic station and absent in the dysoxic stations. For the  
241 purposes of graphing the relative abundance, *Thoracostomopsis*, *Anticoma*, *Cephalanticoma*,  
242 *Trileptium*, *Mesacanthoides*, *Terschellingia*, and *Marylinnia* were grouped as ‘others’ as they  
243 recorded low abundances (<4%) and were absent in dysoxic stations except for *Marylinnia* and  
244 *Terschellingia*, whereby, the former was absent in the oxic station while the latter was present in  
245 both oxygen zones (See Figure 6).

246 The feeding guild composition exhibited variation across distinct oxygen zones within the study  
247 site. Selective epigrowth feeders and herbivores, categorized as Type 2A, emerged as the  
248 predominant nematodes within dysoxic zones, comprising 40% of the population. This was closely  
249 followed by non-selective deposit feeders (Type 1B), contributing to 35% of nematodes within the  
250 same zone. Carnivores and omnivores, classified as Type 2B, constituted 28% of the nematode  
251 population, while selective deposit feeders and bacteriovores, categorized as Type 1A, accounted

252 for 10%. Conversely, in the oxic zone, non-selective deposit feeders (Type 1B) and selective  
253 epigrowth feeders and herbivores (Type 2A) predominated, representing 35% and 32% of the  
254 nematodes, respectively. Carnivores and omnivores (Type 2B) and selective deposit feeders (Type  
255 1A) accounted for 18% and 15%, respectively (Figure 7). Within the feeding modes, selective  
256 deposit feeders had the largest differences in composition between the two oxygen zones, with the  
257 highest concentration observed in the oxic zones. While 2A and 2B had slightly higher values in  
258 the dysoxic zone compared to the oxic zones.

259 The nematode diversity exhibited a pattern akin to the overall macrofaunal diversity, wherein one  
260 of the microxic stations (23002) was devoid of nematodes, while its counterpart station (23020)  
261 harbored only two nematodes. Consequently, microxic stations were excluded from the diversity  
262 analysis. Across the remaining oxygen ranges (dysoxic and oxic), a parallel trend was observed as  
263 in the broader macrofaunal analysis within the study area. Specifically, oxic stations displayed  
264 higher H' diversity ( $1.38 \pm 0.5$ ) compared to dysoxic stations ( $0.81 \pm 0.84$ ). Conversely, dysoxic  
265 stations exhibited greater dominance ( $0.59 \pm 0.39$ ) relative to oxic stations ( $0.32 \pm 0.18$ ) (Fig. 8).

#### 266 **4.0 Discussion**

267 Upwelling systems are known for their high surface productivity and Oxygen Minimum Zones  
268 (OMZ), which impinge on the benthic zone creating strong oxygen gradients on the seafloor and  
269 acting as the dominant driver for benthos diversity in these zones (Zettler et al., 2013). Despite the  
270 Benguela Upwelling System (BUS) being recognized as one of the major Eastern Boundary  
271 Upwelling systems, there is limited information on the structure and composition of the benthic  
272 communities. Information on macrofauna communities in the BUS will not only improve the

273 existing database on benthic fauna but also provide insight into how increasing hypoxic areas in  
274 the ocean might structure benthic communities.

275 In this study, we adapted Levin's grouping system (Levin, 2003), classifying the different stations  
276 into three zones based on the oxygen levels recorded (microxic zone ( $<0.1 \text{ ml l}^{-1}$ ); dysoxic zone  
277 ( $0.1\text{-}1.0 \text{ ml l}^{-1}$ ); oxic zone ( $>1.0 \text{ ml l}^{-1}$ )), which was supported by step-wise regression. The  
278 structure and composition of the macrofauna communities varied among the various oxygen zones.  
279 Most macrofaunal studies identify Polychaeta as the most abundant macrofauna taxon in both oxic  
280 and hypoxic areas (Eisenbarth & Zettler, 2016; Soto et al., 2017). However, in this study,  
281 Polychaeta only had the highest relative abundance in the oxic zones. In the microxic zone, the  
282 abundant taxon was Cumacea, followed by Polychaeta. It is essential to note that numerically  
283 Polychaeta was the most abundant taxon in this oxygen zone, but the presence of many individuals  
284 of other taxa in one of the microxic stations reduced their proportional representation. The presence  
285 of cumaceans in high abundance in the core OMZ, comparable to the microxic zone in this study,  
286 has been reported by Zettler et al., (2013) and Eisenbarth & Zettler (2016), who described them as  
287 possible opportunistic species colonizing permanent hypoxic areas from adjacent areas, and thus  
288 their abundance may be season-specific. Currie et al. (2018) attributed the presence of Cumacea  
289 and other macrofauna taxa in the BUS to the Sulfur-oxidizing bacteria, possibly providing a  
290 detoxified condition in this area. In this case, the mobility of the cumaceans gives them an  
291 advantage over other tolerant taxa like polychaetes and nematodes at the core and hence their high  
292 relative abundances at the OMZs core.

293 Some polychaete families have physiological adaptations to tolerate the low oxygen concentrations  
294 found in OMZs (Hanz et al., 2019; Joydas & Damodaran, 2014; Levin et al., 2009). At the microxic  
295 zone, polychaetes had the numerical abundance in the microxic stations despite cumaceans leading

296 in relative abundance. In station 23002, where Cumacea was located, only two taxa were present.  
297 As a result, cumaceans accounted for 75% of the population despite having a density of only 932  
298 ind. m<sup>-2</sup>. In contrast, Polychaeta in Station 23020 had a numerical high of 4350 ind. m<sup>-2</sup>, which is  
299 relatively higher than the number recorded by cumaceans in Station 23002. However, the diversity  
300 of other taxa in this station was much higher, and as a result, despite the high density, Polychaeta's  
301 proportion of the whole population was reduced to 53%, hence the lower relative abundance.

302 In the dysoxic zone, the macro-nematoda was the most dominant taxon, out-competing the  
303 polychaetes in all the dysoxic stations, recording more than 70% relative abundance. Oxygen can  
304 cause shifts in community structure and trophic transfer (Neira et al., 2018), as evidenced in this  
305 study. Nematoda as a taxon has not received significant attention in the macrofaunal size range in  
306 most studies despite evidence of their presence therein (Joydas & Damodaran, 2014; Sharma et  
307 al., 2011). An increase in the size of nematodes to macrobenthic class sizes has been reported in  
308 chemosynthetic environments that experience similar characteristics as OMZs, i.e., low oxygen  
309 and high sulfidic contents (Vanreusel et al., 2010). Apart from the increase in size, OMZs also  
310 tend to enhance the regional dominance of tolerant organisms such as nematodes with high  
311 biomass recorded in response to organic matter inputs. The high abundances are thought to reflect  
312 the availability of organic matter, a significant nutrient source for macrofauna, coupled with a  
313 reduction in predation by larger fauna that are affected adversely by the reduced oxygen  
314 concentrations (Moens et al., 2013).

315 The high nematode abundance in partnership with Oligochaeta occurred in the dysoxic zone;  
316 oxygen conditions were low enough to exclude some taxa but sufficient for tolerant species to  
317 survive and reproduce. Such conditions are referred to as the 'edge effect,' and such high densities  
318 are characteristics of the edge of the OMZs, where various species have been observed to have



319 abnormally high densities. The reasons for these high abundances are not well understood, but  
320 Gutiérrez et al. (2008) suggested that the nematodes' population can multiply in low oxygen  
321 conditions, which experience high loads of organic matter input. Despite their tolerance to anoxia,  
322 nematodes cannot survive long-term exposure, as observed in the microxic zone (Moens et al.,  
323 2013). It is not clearly understood then whether the high abundance of the nematodes in one of the  
324 stations is characteristic of the study site or just congregation to a food source. Thus, the patchiness  
325 and high variability in the dysoxic zone call for more studies (Buhl-Mortensen et al., 2010).

326 At dysoxic sites (DO 0.1-1.0 ml l<sup>-1</sup>), other taxa like Ostracoda and Bivalvia were observed, albeit  
327 in low numbers. Despite ostracodes flourishing better in well-oxygenated marine areas, various  
328 families (Platycopina) have been observed to tolerate and thrive in Oxygen Minimum Zones. At  
329 oxic sites, where DO levels were above 1.0 ml l<sup>-1</sup>, more taxa were recorded, and these numbers  
330 increased even more when DO levels surpassed 2.0 ml l<sup>-1</sup>.

331 Our findings were in line with the various studies in OMZs whereby a decrease in richness and  
332 diversity of macrofaunal species is observed towards the core and vice versa (McClain &  
333 Schlacher, 2015). In the core (microxic) area, the macrofauna taxa had the lowest density and  
334 diversity. Each square meter of core area contained 1243 individuals, indicating low species  
335 density. A similar number was recorded by Zettler et al., (2009). In this study, an increase in the  
336 number of taxa recorded in sites with DO levels above 1 ml l<sup>-1</sup> was also observed. The high  
337 dominance of the tolerant taxa, as evidenced in the microxic and dysoxic areas, was reduced. Taxa  
338 such as Amphipoda, Isopoda, Echinodermata, Nemertina, Aeolosomatidae, Aplacophora,  
339 Holothuroidea, and Cnidaria were only recorded in the oxic zones indicating low tolerance to low  
340 oxygen levels. Of this fauna, crustaceans were the most abundant. This conforms to the  
341 observations of Soto et al. (2017) at oxic sites in an upwelling system in Chile. Conversely, Zettler

342 et al. (2009) recorded amphipod species in low-oxygen areas. These contradictory results indicate  
343 that at least amongst the Amphipoda, tolerance/intolerance to hypoxia is species-specific.

344 Macro-nematoda abundance varied across the OMZ, with very low abundance in the microxic  
345 zones, extremely high numbers at the dysoxic zones, and a substantial amount at the oxic stations,  
346 accounting for only 28% of the total abundance. Nematodes are considered one of the most tolerant  
347 taxa in the marine environment, with the ability to tolerate low oxygen and high sulphidic  
348 environments characteristic of OMZs and may reach very high abundances in these environments  
349 (Neira et al., 2018; Gutierrez et al., 2008). This was the case at the dysoxic zone, where nematodes  
350 recorded high abnormal densities in one station, indicating the high tolerance of these taxa and  
351 their ability to dominate the macrofaunal component. Even with such high tolerance levels,  
352 nematode abundance can be impacted by microxic conditions, as observed in the microxic areas  
353 with a recording of only 621 nematodes m<sup>-2</sup> in this oxygen zone. This low value, however, may be  
354 at the macrofauna level, and the case may be different at the meiofauna level, where nematodes  
355 have recorded substantial densities in microxic environments (Neira et al., 2018; Steyaert et al.,  
356 2007).

357 Despite their high abundance in OMZs, not all nematodes are tolerant to low oxygen levels (Moens  
358 et al., 2013), as observed in this study. *Metoncholaimus*, *Paracomesoma*, and *Desmolaimus*  
359 dominated the dysoxic zone; these three genera are members of Oncholaimidae, Comesomatidae,  
360 and Linhomoeidae, respectively. Members of the family Oncholaimidae have large bodies that can  
361 disperse rapidly and colonize carcasses of macrofauna and even fish that may have succumbed to  
362 the low levels of oxygen found in the dysoxic zone (Moens et al., 2013). Nevertheless, their bodies  
363 are large enough to fit within the macrofauna size range, while their ability to swim ensures they  
364 actively locate their food source (Moens et al., 2013). Their high abundance in this study might

365 reflect a congregation upon a food source that had attracted nearby members in large numbers.  
366 Their ability to colonize the 'food source' in such numbers in a dysoxic environment indicates their  
367 tolerance to low oxygen levels.

368 On the other hand, Comesomatidae and Linhomoeidae members have been noted to have high  
369 abundances in enriched sediments with low oxygen, indicating tolerance to anoxic conditions  
370 (Steyaert et al., 2007). Their long and slender bodies might be the reason for their records at the  
371 macrofauna level. Despite this generalization at the family level and the assumptions that members  
372 of the same family may portray similar life strategies (Bongers et al., 1991), tolerance of nematodes  
373 to hypoxia is species-specific (Moens et al., 2013) as Steyaert et al. (2007) observed members of  
374 the same genera (*Sabatiera*) reacting differently to hypoxic and anoxic conditions. Thus, further  
375 analysis should be done to identify the species that are tolerant to hypoxia at these OMZs at  
376 macrofauna levels. Tolerance to hypoxia is indicated by both the presence and absence of taxa;  
377 most genera present in the oxic zone were absent in the dysoxic area and may be seen as genera  
378 intolerant to low oxygen levels.

379 Wieser's feeding types, as outlined in his study (Wieser, 1953), have long been used to assess the  
380 trophic structure of nematode communities. This study had a high abundance of non-selective  
381 feeders (1B) and selective epigrowth feeders and herbivores (2A), with the latter dominating the  
382 dysoxic zone, while the former dominating the oxic zones. Non-selective deposit feeders are  
383 commonly observed to dominate substrates characterized by high levels of organic matter and  
384 smaller grain size, indicating their adaptation to environments with elevated organic matter input,  
385 as observed in this study (Singh and Ingole, 2016; Heip et al., 1985). In contrast to previous  
386 findings on meiofaunal nematodes from Oxygen Minimum Zones (OMZs) (Singh & Ingole, 2016;  
387 Neira et al., 2013) and macro-nematodes in deep-sea habitats (Sharma & Bluhm, 2011), which

388 reported lower abundances of selective epigrowth feeders, our study revealed a higher abundance  
389 of selective epigrowth feeders and herbivores (2A). Within OMZs, such as our study site, the  
390 presence of giant bacteria such as *Thioploca*, *Beggiatoa*, and *Thiomargarita* forming extensive  
391 mats (Brüchert et al., 2009) may provide a substantial food source for nematodes (Giere, 2008).

392 Adjacent to OMZs, the prevalence of diatomaceous mud and the settling of fresh phytodetritus  
393 contribute significantly to the diet of epistratum feeders, leading to their high abundance in dysoxic  
394 areas (Singh & Ingole, 2016; Sharma & Bluhm, 2011; Moens & Vincx, 1987; Heip et al., 1985).  
395 However, in regions offshore from the OMZ, where the OMZ no longer interacts with the benthic  
396 zone, reduced diatom production diminishes the abundance of epistratum feeders, resulting in the  
397 dominance of non-selective deposit feeders.

398 While non-selective deposit feeders typically dominate muddy and silty environments, selective  
399 deposit feeders and carnivores have been observed to exhibit lower abundances in these habitats  
400 (Heip et al., 1985). Our study aligns with this trend; however, other investigations have reported  
401 contradictory findings, with selective deposit feeders being identified among the most dominant  
402 nematodes in similar environments (Neira et al., 2013; Singh & Ingole, 2016). Additionally,  
403 carnivores and omnivores (2B) exhibited higher abundances in the dysoxic zone compared to the  
404 oxic zone, with their overall abundance surpassing that of selective deposit feeders. These  
405 observations suggest potential differences in nematode composition between the meiofaunal and  
406 macrofaunal levels. Sharma and Bluhm (2011) observed that category 2B feeders were the second  
407 most abundant feeding type in numerous stations within the Arctic deep-sea Canada Basin. They  
408 attributed these findings to the omission of larger nematodes in previous studies, as carnivores and  
409 omnivores are often large-bodied species.

## 410 **5.0 Conclusion**

411 In conclusion, this study provides insights into the intricate dynamics of macrofaunal communities  
412 in Oxygen Minimum Zones (OMZs). By classifying stations into microxic, dysoxic, and oxic  
413 zones based on oxygen levels, we observed variations in macrofaunal composition and abundance  
414 across these zones. Contrary to typical findings, Polychaeta dominated only in oxic zones, while  
415 Cumacea and Nematoda were prevalent in microxic and dysoxic zones, respectively. The presence  
416 of tolerant taxa like nematodes and Oligochaeta in dysoxic zones suggests their ability to thrive  
417 under low oxygen conditions. Conversely, taxa intolerant to low oxygen were absent in dysoxic  
418 zones, highlighting the importance of oxygen levels in shaping macrofaunal communities.

419 Macro-nematodes in general were the most abundant taxon. The high nematode densities suggest  
420 the ability of macro-nematodes to tolerate and thrive in low-oxygen environments. Our findings  
421 highlight the importance of considering macro-nematodes in studies of OMZ ecosystems and  
422 underscore the need for further research to elucidate their ecological roles and responses to  
423 environmental changes. Understanding the dynamics of macro-nematode populations coupled  
424 with their meiofaunal counterparts in OMZs is crucial for comprehensively assessing the impacts  
425 of oxygen depletion on benthic communities and ecosystem functioning.

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435 **6.0 Tables**

436 Table 1: Station information and abiotic factors information recorded from the stations in the Northern  
 437 Benguela Upwelling System. NR stands for no replicate.

Transect	Station (No. Replicate)	Longitude	Latitude	Depth (m)	TOM(%)	DO (ml l <sup>-1</sup> )
Off Terrace Bay (OTB-20°S)	20002(2)	12.99905	-20	33	27.47±1.55	1.8
	20020(3)	12.67858	-20	125	38.64±2.16	0.67
	20040(N.R)	11.79321	-20	219	17.58 nr	0.53
Off Walvis Bay (OWB-23°S)	23002(NR)	14.3734	-23	39	23.08 nr	0.06
	23020(NR)	14.06986	-23	128	20.88 nr	0.02
	23070(3)	13.14	-23	318	9.89±1.55	2.3
Off Luderitz (OL 26°S)	26090(NR)	13.28	-26	1282	4.4 nr	4.34

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440 Table 2: Sediment size distribution in the Northern Benguela Upwelling System

Transect	Station	Very Coarse Sand	Coarse Sand	Medium sand	fine sand	very fine sand	Silt
Off Terrace Bay (20°S)	20002	0.65	0.67	13.81	34.82	21.29	28.76
	20020	0.93	0.62	28.17	30.89	12.98	26.42
	20040	2.71	4.37	18.55	33.33	17.04	23.98
Off Walvis Bay (23°S)	23002	0.00	0.28	18.74	36.17	17.25	27.57
	23020	0.98	6.85	14.91	38.88	18.83	19.56
	23070	0.00	0.00	13.92	33.73	20.29	32.06
Off Luderitz (26°S)	26090	0.00	0.29	23.99	30.46	19.25	26.01

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## 7.0 Figures

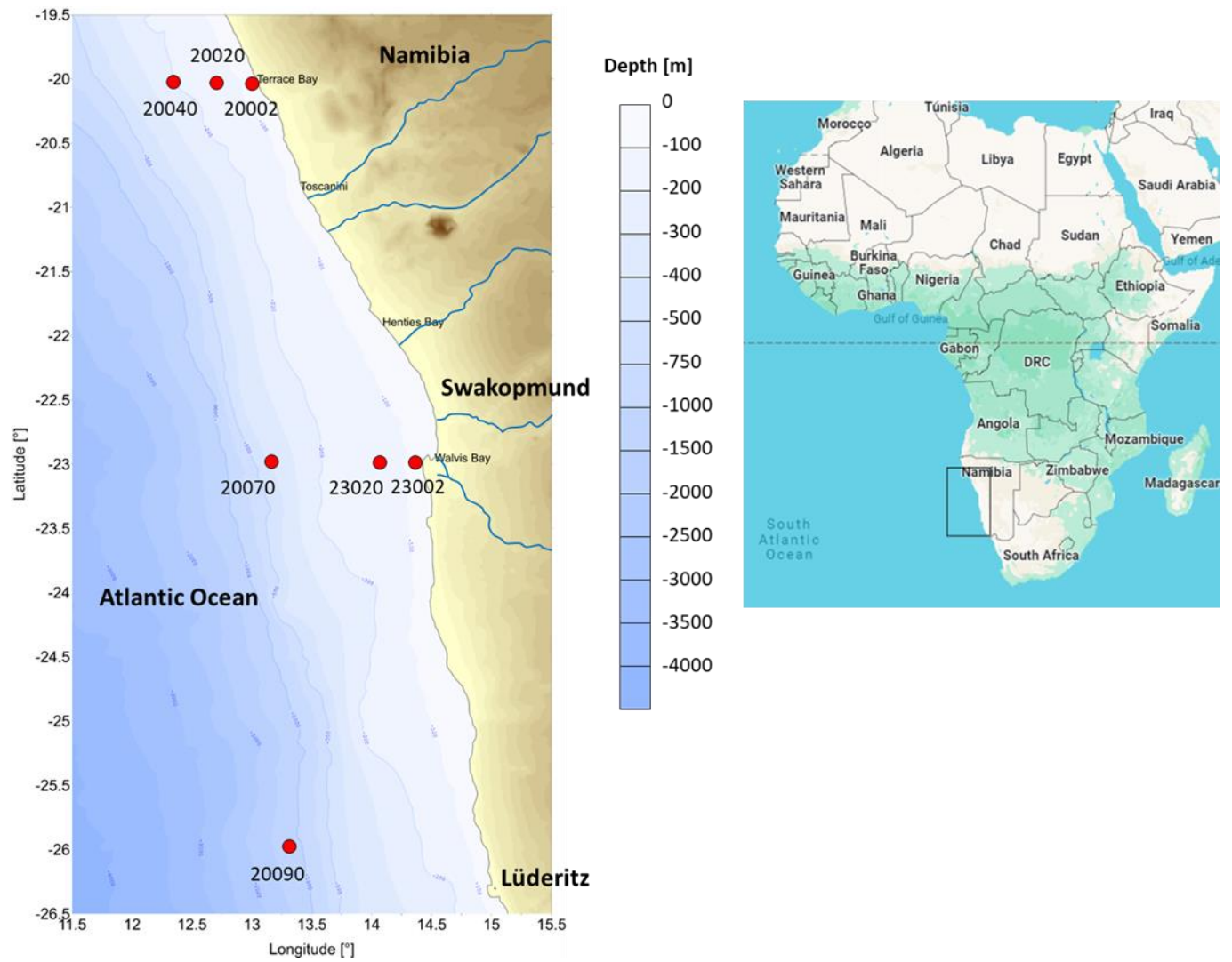


Figure 1: Sampling stations located across the Northern Benguela Upwelling System



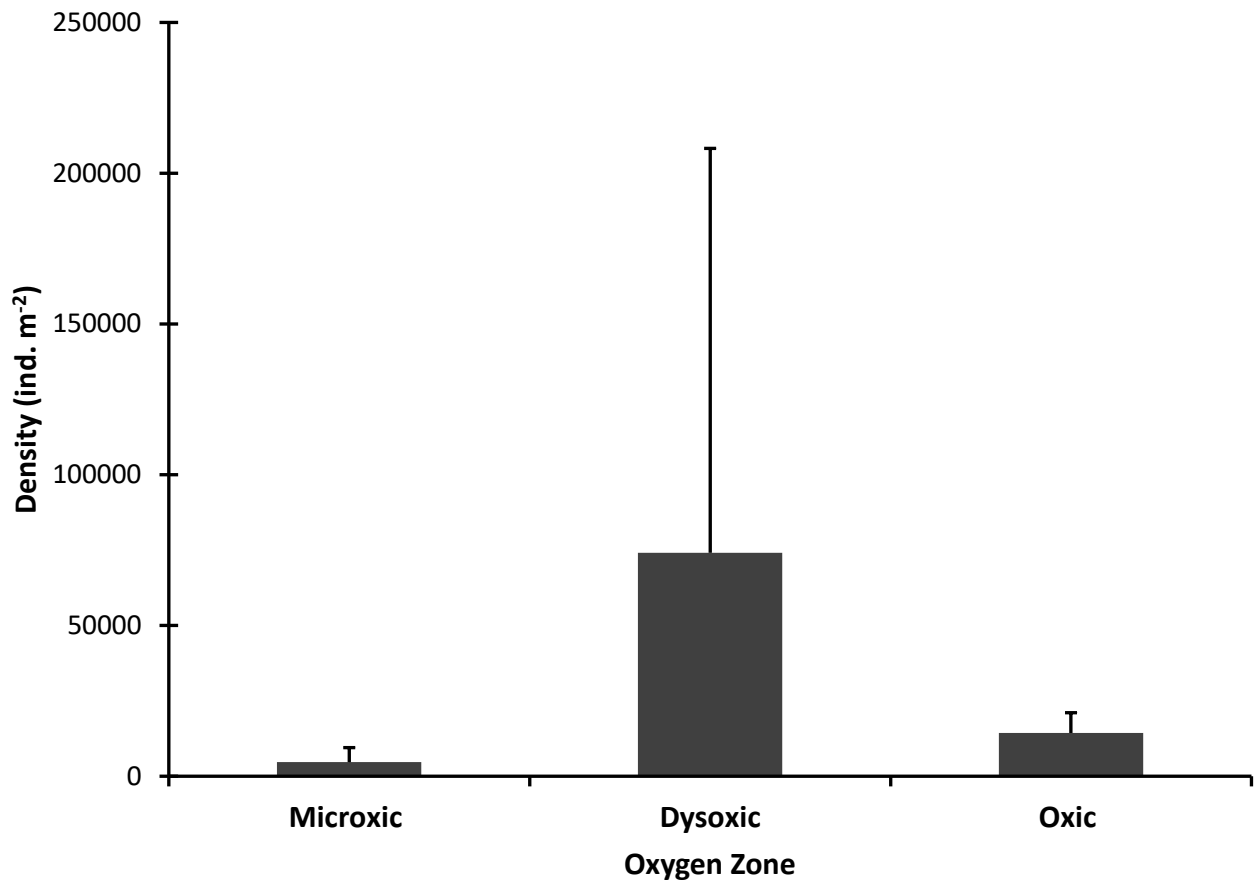


Figure 2: Macrobenthic densities in the different oxygen zones in the Northern Benguela Upwelling system. Error bars represent standard deviation (+1 SD).

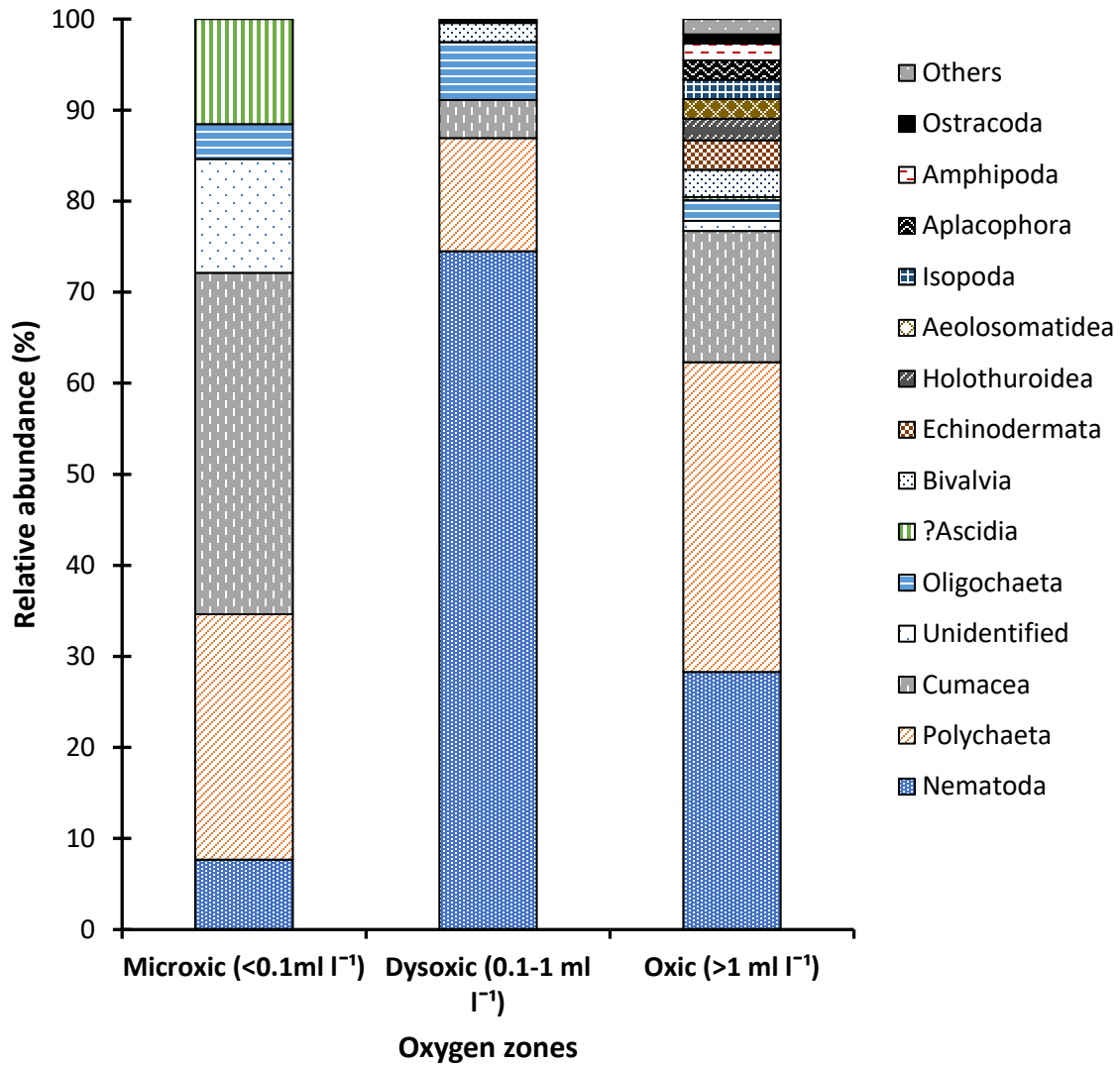


Figure 3: Macrofauna relative abundance in the Northern Benguela Upwelling System, based on the oxygen zones identified.

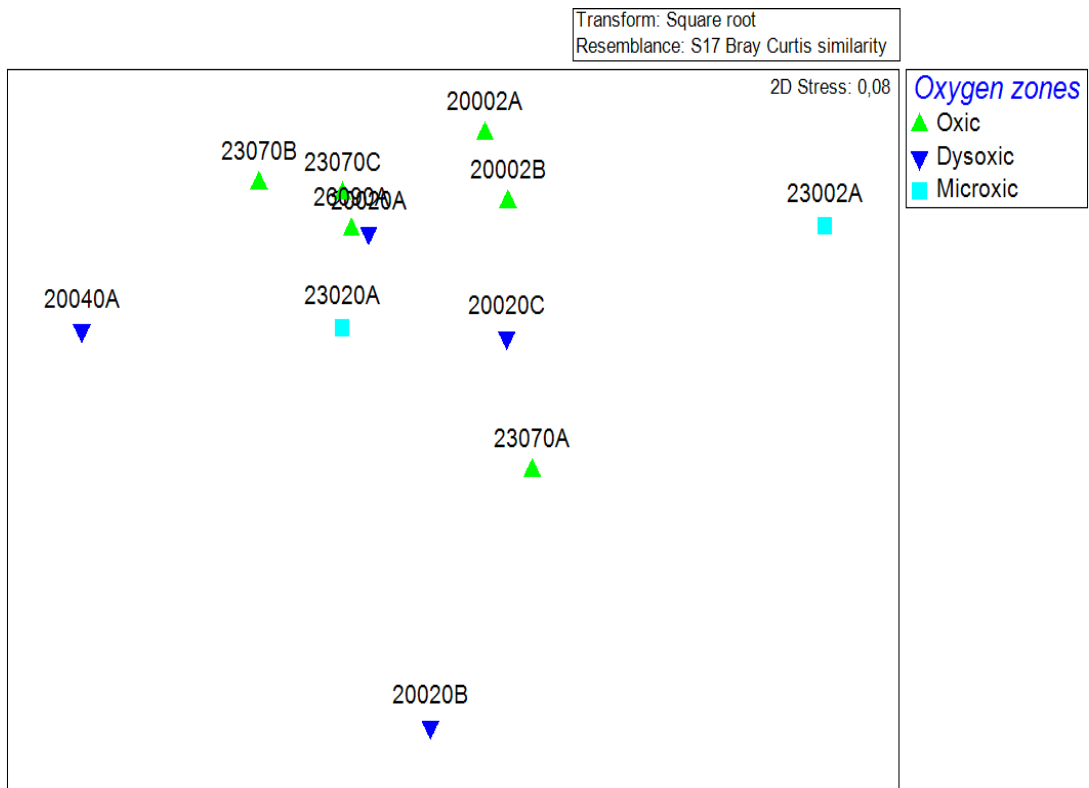


Figure 4: Non-metric multidimensional scaling (nMDS) plot based on Bray-Curtis Similarity index of macrobenthic fauna communities recorded in the Northern Benguela Upwelling System.

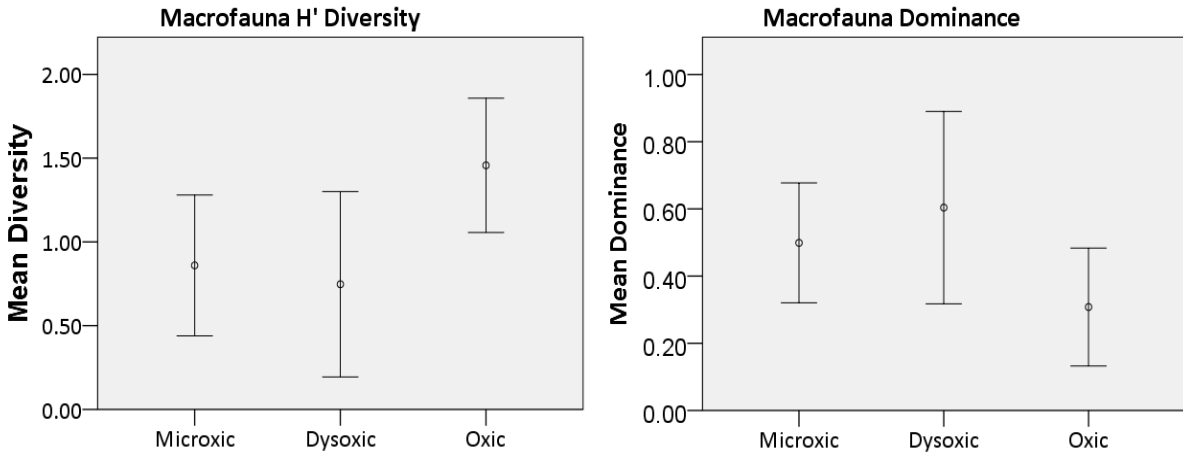


Figure 5: Macrofauna diversity indices recorded in the various oxygen zones in the Northern Benguela Upwelling system. H' diversity, calculated using the natural logarithm ( $\log_e$ ), while dominance, represented the complement of the Simpson Index ( $1 - \text{Simpson index}$ ). Error bars represent standard deviation ( $\pm 1 \text{ SD}$ ).

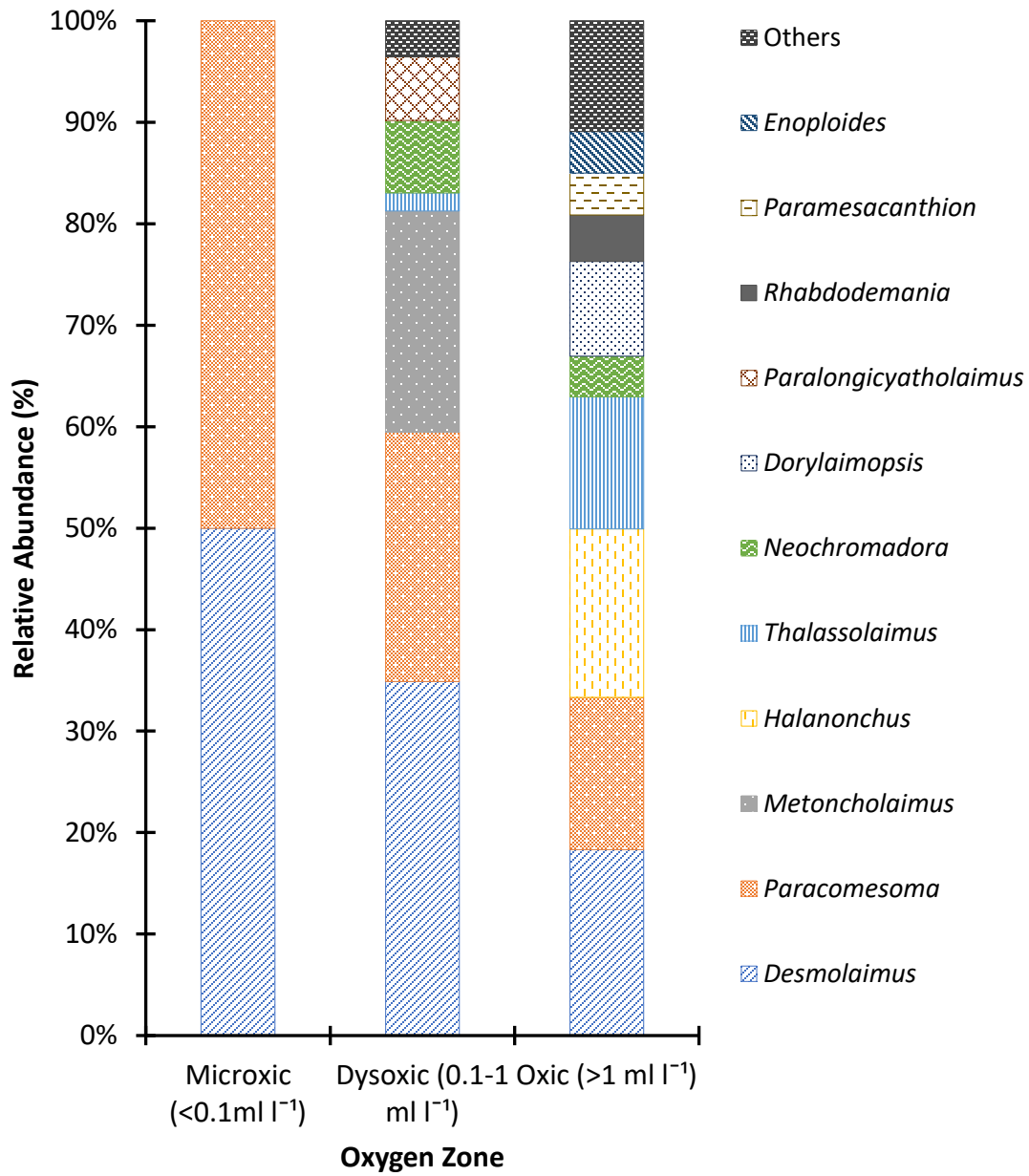


Figure 6: Macroneumatoda relative abundance in the Northern Benguela Upwelling System, based on the oxygen zones identified.

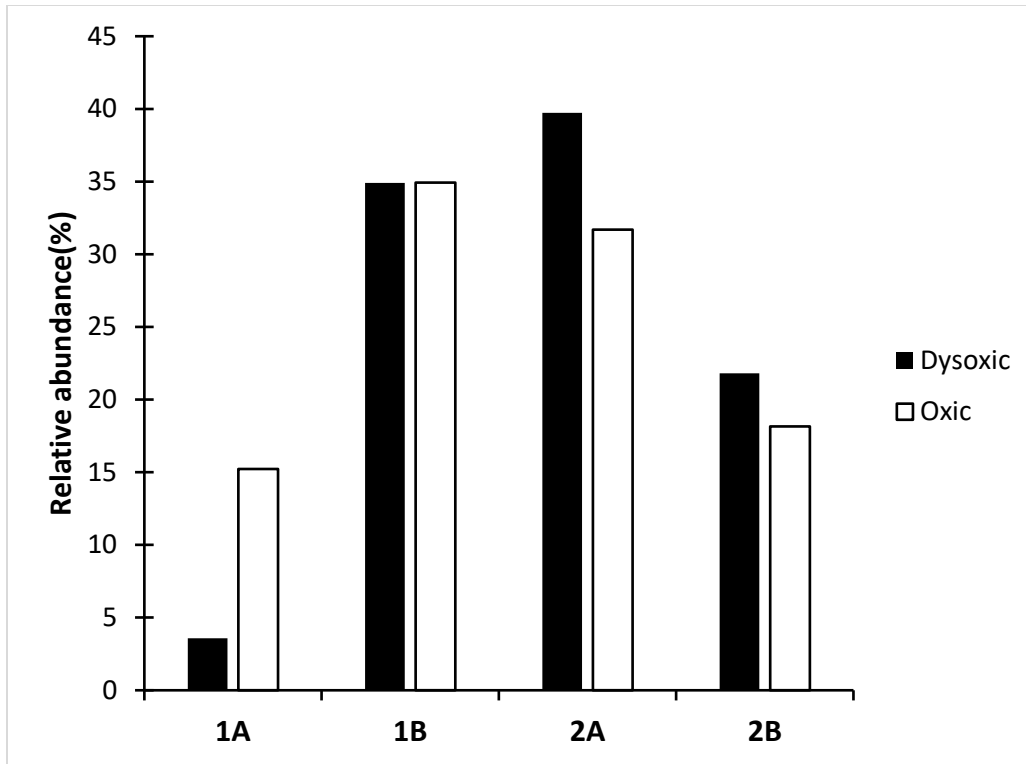


Figure 7: Feeding guilds based on Wieser (1953) of macro-Nematoda from the Northern Benguela Upwelling System. (1A= Selective deposit feeder, 1B- Non-Selective deposit feeder, 2A=Epigrwoth feeders, 2B=Predators/Omnivores).

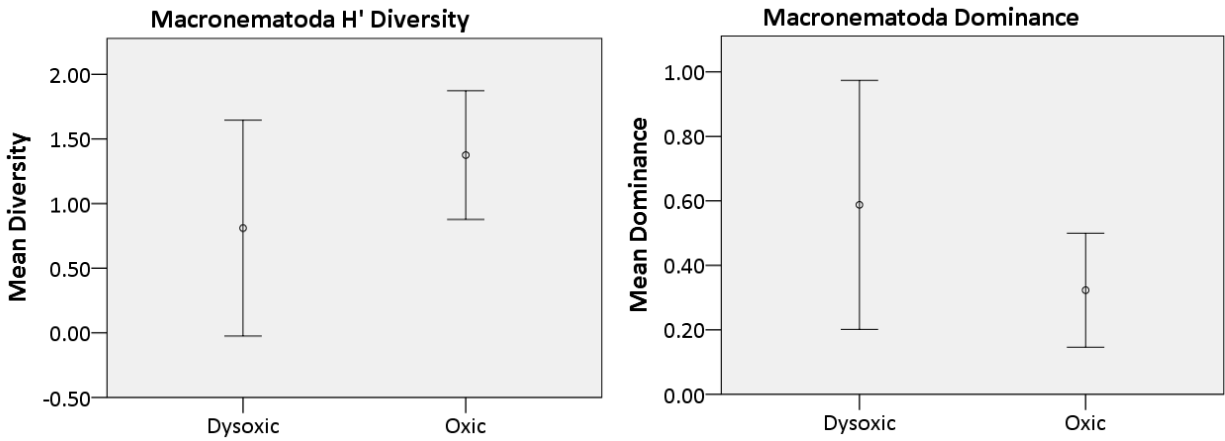


Figure 8: Macro-Nematoda diversity indices recorded in the various oxygen zones in the Northern Benguela Upwelling system. H' diversity, calculated using the natural logarithm (loge), while dominance, represented the complement of the Simpson Index (1-Simpson index). Error bars represent standard deviation (+/-1 SD).

## **8.0 Code/Data Availability**

Currently, the data is not available but can be released upon request to the authors.

## **9.0 Author contribution**

HS was responsible for the conceptualization of the study. HS and WB conducted the investigation, formal analysis, and drafting of the original draft. MA was responsible for supervision, review, and editing of the final draft.

## **10.0 Competing interests**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## **11.0 Special Issue Statement**

Part of this manuscript was presented as a poster presentation during the 53<sup>rd</sup> International Colloquium on Ocean Dynamics: 3rd GO2NE Oxygen Conference held in Liege, 2022. As a result, the corresponding author was invited to submit a manuscript for the special issue "*Low-oxygen environments and deoxygenation in open and coastal marine waters*" as part of the journal Biogeosciences.

## **12.0 Acknowledgements**

The authors would like to thank the 3rd Regional Research Graduate Network in Oceanography (RGNO) organizers, all the sponsors, and participants for technically and financially supporting the authors in the participation and subsequent sampling that resulted in the successful completion of this study. The assistance from the Namibian National Marine Information and Research Centre



(NatMIRC) was instrumental in successfully sampling, processing, and transporting samples. The crew of the sampling vessel R/V Mirabilis ensured a smooth sampling experience, and hence we take this opportunity to appreciate their services.

We also extend our gratitude to the reviewers for their invaluable feedback and constructive criticism, which significantly enhanced the quality and clarity of this manuscript. We deeply appreciate their time, expertise, insights, suggestion, and dedication to improving the manuscript. Their contributions have undoubtedly strengthened the final version of this manuscript, and we are sincerely grateful for their inputs.

### 13.0 References

- Bachelet, G. (1990). The choice of a sieving mesh size in the quantitative assessment of marine macrobenthos: a necessary compromise between aims and constraints. *Marine Environmental Research*, 30(1), 21–35.
- Bohata, K., & Koppelman, R. (2013). Chaetognatha of the Namibian upwelling region: taxonomy, distribution and trophic position. *PloS One*, 8(1), e53839.
- Bongers, T., Alkemade, R., & Yeates, G. W. (1991). Interpretation of disturbance-induced maturity decrease in marine nematode assemblages by means of the Maturity Index. *Marine Ecology Progress Series*, 135–142.
- Breitburg, D., Levin, L. A., Oschlies, A., Grégoire, M., Chavez, F. P., Conley, D. J., Garçon, V., Gilbert, D., Gutiérrez, D., & Isensee, K. (2018). Declining oxygen in the global ocean and coastal waters. *Science*, 359(6371), eaam7240.
- Brodie Rudolph, T., Ruckelshaus, M., Swilling, M., Allison, E. H., Österblom, H., Gelcich, S., & Mbatha, P. (2020). A transition to sustainable ocean governance. *Nature Communications*, 11(1), 3600.
- Brüchert, V., Currie, B., & Peard, K. R. (2009). Hydrogen sulphide and methane emissions on the central Namibian shelf. *Progress in Oceanography*, 83(1-4), 169-179.
- Buhl-Mortensen, L., Vanreusel, A., Gooday, A. J., Levin, L. A., Priede, I. G., Buhl-Mortensen, P., Gheerardyn, H., King, N. J., & Raes, M. (2010). Biological structures as a source of habitat heterogeneity and biodiversity on the deep ocean margins. *Marine Ecology*, 31(1), 21–50.
- Clarke, K. R., & Gorley, R. N. (2005). PRIMER: plymouth routines in multivariate ecological research. *PRIMER-E Ltd., Plymouth*.
- Currie, B., Utne-Palm, A. C., & Salvanes, A. G. V. (2018). Winning ways with hydrogen sulphide on the Namibian shelf. *Frontiers in Marine Science*, 5, 341.
- Dietrich, P., Cesarz, S., Liu, T., Roscher, C., & Eisenhauer, N. (2021). Effects of plant species diversity on nematode community composition and diversity in a long-term biodiversity experiment. *Oecologia*, 197(2), 297–311.
- Eisenbarth, S., & Zettler, M. L. (2016). Diversity of the benthic macrofauna off northern Namibia from the shelf to the deep sea. *Journal of Marine Systems*, 155, 1–10.

- Emeis, K., Eggert, A., Flohr, A., Lahajnar, N., Nausch, G., Neumann, A., Rixen, T., Schmidt, M., Van der Plas, A., & Wasmund, N. (2018). Biogeochemical processes and turnover rates in the Northern Benguela Upwelling System. *Journal of Marine Systems*, 188, 63–80.
- Emeis, K.-C., Brüchert, V., Currie, B., Endler, R., Ferdelman, T., Kiessling, A., Leipe, T., Noli-Peard, K., Struck, U., & Vogt, T. (2004). Shallow gas in shelf sediments of the Namibian coastal upwelling ecosystem. *Continental Shelf Research*, 24(6), 627–642.  
<https://doi.org/10.1016/j.csr.2004.01.007>
- Giere, O. (2008). *Meiobenthology: the microscopic motile fauna of aquatic sediments*. Springer Science & Business Media.
- Gobler, C. J., & Baumann, H. (2016). Hypoxia and acidification in ocean ecosystems: Coupled dynamics and effects on marine life. *Biology Letters*, 12(5).  
<https://doi.org/10.1098/rsbl.2015.0976>
- Gutiérrez, D., Enríquez, E., Purca, S., Quipúzcoa, L., Marquina, R., Flores, G., & Graco, M. (2008). Oxygenation episodes on the continental shelf of central Peru: Remote forcing and benthic ecosystem response. *Progress in Oceanography*, 79(2–4), 177–189.
- Hammer, Ø., Harper, D. A. T., & Ryan, P. D. (2001). PAST: paleontological statistics software package for education and data analysis. *Palaeontologia Electronica*, 4(1), 9.
- Hanz, U., Wienberg, C., Hebbeln, D., Duineveld, G., Lavaleye, M., Juva, K., Dullo, W.-C., Freiwald, A., Tamborrino, L., & Reichart, G.-J. (2019). Environmental factors influencing benthic communities in the oxygen minimum zones on the Angolan and Namibian margins. *Biogeosciences*, 16(22), 4337–4356.
- Ingels, J., Zeppilli, D., & Giere, O. (2023). Meiofauna—Adapted to Life at the Limits. In *New Horizons in Meiobenthos Research: Profiles, Patterns and Potentials* (pp. 363–400). Springer.
- Joydas, T. V., & Damodaran, R. (2014). Infaunal macrobenthos of the oxygen minimum zone on the Indian western continental shelf. *Marine Ecology*, 35(1), 22–35.
- Levin, L. A., Ekau, W., Gooday, A. J., Jorissen, F., Middelburg, J. J., Naqvi, S. W. A., Neira, C., Rabalais, N. N., & Zhang, J. (2009). Effects of natural and human-induced hypoxia on coastal benthos. *Biogeosciences*, 6(10), 2063–2098.
- Levin Lisa. (2003). Oxygen minimum zone benthos: Adaptation and community response to hypoxia. In Gibson R. N & Atkinson R.J.A (Eds.), *Oceanography and Marine Biology: An Annual Review* (Vol. 41, pp. 1–45). CRC Press.

- Li, S., Cui, B., Xie, T., Bai, J., Wang, Q., & Shi, W. (2018). What drives the distribution of crab burrows in different habitats of intertidal salt marshes, Yellow River Delta, China. *Ecological indicators*, 92, 99-106.
- Liu, S. (2023). Interactions within Sustainable Development Goals (SDGs): the economic and environmental dimensions of SDG Indicator 14.7. In Swain, R. B., & Min, Y. (Eds.), (2023). *Interlinkages Between the Sustainable Development Goals* (pp. 37-51). Edward Elgar Publishing.
- Magalhães, A. V. (2018). Comparison of zooplankton communities between cold and hot seasons in Lobito-Angola. (Doctoral dissertation).
- McClain, C. R., & Schlacher, T. A. (2015). On some hypotheses of diversity of animal life at great depths on the sea floor. *Marine Ecology*, 36(4), 849–872.
- Moens, T., Braeckman, U., Derycke, S., Fonseca, G., Gallucci, F., Gingold, R., Guilini, K., Ingels, J., Leduc, D., & Vanaverbeke, J. (2013). Ecology of free-living marine nematodes. *Nematoda*, 2, 109–152.
- Moens, T., & Vincx, M. (1997). Observations on the feeding ecology of estuarine nematodes. *Journal of the Marine Biological Association of the United Kingdom*, 77(1), 211–227.
- Montgomery, H., Thom, N. S., & Cockburn, A. (1964). Determination of dissolved oxygen by the Winkler method and the solubility of oxygen in pure water and sea water. *Journal of Applied Chemistry*, 14(7), 280–296.
- Neira, C., Ingels, J., Mendoza, G., Hernandez-Lopez, E., & Levin, L. A. (2018). Distribution of meiofauna in bathyal sediments influenced by the oxygen minimum zone off Costa Rica. *Frontiers in Marine Science*, 5, 448.
- Neira, C., King, I., Mendoza, G., Sellanes, J., De Ley, P., & Levin, L. A. (2013). Nematode community structure along a central Chile margin transect influenced by the oxygen minimum zone. *Deep Sea Research Part I: Oceanographic Research Papers*, 78, 1–15.
- Platt, H. M., & Warwick, R. M. (1988). *Freeliving marine nematodes: Part II. British Chromadorida. Synopses of the British Fauna No. 38*. EJ Brill, Dr. W. Backhuys for the Linnean Society of London and the ....
- Ridall, A., & Ingels, J. (2021). Suitability of free-living marine nematodes as bioindicators: Status and future considerations. *Frontiers in Marine Science*, 8, 685327.
- Rossi, S. (2023). A Comprehensive Overview of SDG 14: Life Below Water\_Final. In: SDG 14: Life Below Water. Springer, Cham. [https://doi.org/10.1007/978-3-031-19467-2\\_1](https://doi.org/10.1007/978-3-031-19467-2_1)

- Sharma, J., Baguley, J., Bluhm, B. A., & Rowe, G. (2011). Do meio-and macrobenthic nematodes differ in community composition and body weight trends with depth? *PLoS One*, 6(1), e14491.
- Sharma, J., & Bluhm, B. A. (2011). Diversity of larger free-living nematodes from macrobenthos (> 250 µm) in the Arctic deep-sea Canada Basin. *Marine Biodiversity*, 41, 455-465.
- Soto, E., Quiroga, E., Ganga, B., & Alarcón, G. (2017). Influence of organic matter inputs and grain size on soft-bottom macrobenthic biodiversity in the upwelling ecosystem of central Chile. *Marine Biodiversity*, 47, 433–450.
- Steffani, N., Sedick, S., Rogers, J., & Gibbons, M. J. (2015). Infaunal benthic communities from the inner shelf off Southwestern Africa are characterised by generalist species. *Plos One*, 10(11), e0143637.
- Steyaert, M., Moodley, L., Nadong, T., Moens, T., Soetaert, K., & Vincx, M. (2007). Responses of intertidal nematodes to short-term anoxic events. *Journal of Experimental Marine Biology and Ecology*, 345(2), 175–184.
- Teuber, L., Schukat, A., Hagen, W., & Auel, H. (2013). Distribution and ecophysiology of calanoid copepods in relation to the oxygen minimum zone in the eastern tropical Atlantic. *PloS One*, 8(11), e77590.
- Thiel, H., & Higgins, R. P. (1988). Introduction to the study of meiofauna. *Smithsonian*.
- Vanreusel, A., De Groote, A., Gollner, S., & Bright, M. (2010). Ecology and biogeography of free-living nematodes associated with chemosynthetic environments in the deep sea: a review. *PLoS One*, 5(8), e12449.
- Wieser, W. (1953). Die Beziehung zwischen Mundhohlengestalt, Ernährungsweise und Vorkommen bei freilebenden marinen Nematoden. Eine skologischen-morphologische studie. *Arkiv fiir Zoologie*, 4, 439-484.
- Wentworth, C. K. (1922). A scale of grade and class terms for clastic sediments. *The Journal of Geology*, 30(5), 377–392.
- Zeppilli, D., Sarrazin, J., Leduc, D., Arbizu, P. M., Fontaneto, D., Fontanier, C., Gooday, A. J., Kristensen, R. M., Ivanenko, V. N., & Sørensen, M. V. (2015). Is the meiofauna a good indicator for climate change and anthropogenic impacts? *Marine Biodiversity*, 45(3), 505–535.
- Zettler, M. L., Bochert, R., & Pollehne, F. (2009). Macrozoobenthos diversity in an oxygen minimum zone off northern Namibia. *Marine Biology*, 156, 1949–1961.

Zettler, M. L., Bochert, R., & Pollehne, F. (2013). Macrozoobenthic biodiversity patterns in the northern province of the Benguela upwelling system. *African Journal of Marine Science*, 35(2), 283–290.

Zhang, Z., Xia, S., Sun, X., Zhou L. (2022). Effects of Macrobenthos Relative to Floating-Leaved Plants on the Wintering Shorebird Assemblages at Shengjin Lake, China. *Diversity*; 14(12):1072. <https://doi.org/10.3390/d14121072>

