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Biogeosciences Discussions

Unusual *Hemiaulus* Bloom Influences Ocean Productivity in Northeast U.S. Shelf Waters

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Abstract. Ocean production and trophic transfer rates, including Net Community Production (NCP), Net Primary Production (NPP), Gross Oxygen Production (GOP), and microzooplankton grazing rates are key metrics for understanding marine ecosystem dynamics and impacts on biogeochemical cycles. Because of its temperate location and high dynamic range of environmental conditions and long-term human utilization, the long-term ecological research site in the coastal Northeastern U.S. Shelf (NES) of the Northwestern Atlantic Ocean offers an ideal opportunity to understand how productivity shifts in response to changes in planktonic community composition. While small phytoplankton usually dominate in the NES waters during the summer, a bloom of the large diatom genus *Hemiaulus*, with N_2 fixing symbionts, was observed in the mid-shelf region during the summer of 2019. NCP was 2.5 to 9 times higher when *Hemiaulus* dominated compared to NCP throughout the same geographic area during the summers of 2020–2022. The *Hemiaulus* bloom in summer 2019 also coincided with higher trophic transfer efficiency from phytoplankton to microzooplankton, higher GOP and NPP, and higher sea surface temperatures than summers 2020-2022. This study shows that the presence of an atypical phytoplankton community that alters the typical size distribution of the primary producers can greatly influence productivity and trophic transfer, highlighting the dynamic nature of the coastal ocean. Notably, summer 2018 NCP levels were also high although no atypical phytoplankton community was present. A better understanding of the dynamics of the NES in terms of biological productivity is of primary importance, especially in the context of changing environmental conditions due to climate processes.

1 Introduction

Oceans regulate atmospheric carbon dioxide (CO_2) concentrations and support life on Earth via several mechanisms (Friedlingstein et al., 2022). One of these mechanisms is the biological pump, which involves biological, physical, and chemical processes that aid in transporting and sequestering organic carbon from CO_2 (Boyd et al., 2019). As the main primary producers in the ocean, phytoplankton play a major role in the biological pump (Field et al., 1998). Diatoms, a type of photosynthetic algae, are believed to account for nearly half of net marine primary productivity globally and are important contributors to the biological pump (Jin et al., 2006). Diatoms characteristically thrive in nutrient-rich surface layers and turbulent conditions, and are thus typically found at high latitudes and in coastal upwelling regions (Armbrust, 2009). However, new technology (e.g., molecular biology and imaging) has revealed that diatoms may be more prevalent in low nutrient, oligotrophic systems than traditionally considered (Malviya et al., 2016), likely due to unique metabolic capabilities involving nutrient acquisition strategies that enable their survival in low nutrient regimes.

One specific metabolic capability within diatoms is the ability to form a symbiosis with nitrogen-fixing cyanobacteria. This symbiosis, known as a diatom-diazotroph association has been observed around the globe, mostly in oligotrophic regions (Foster and Zehr, 2019), but also in temperate continental shelf waters (Wang et al., 2021). Furthermore, some diatom-diazotroph association have the capability to grow very quickly, forming localized blooms (Villareal et al., 2011). Diatom-diazotroph blooms, specifically involving the diatom genus

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Hemiaulus and the symbiont *Richelia*, have been found in warm, stratified waters in various regions around the globe and have been associated with high carbon export observed via a combination of modern oceanographic measurements and paleo-flux case studies (Kemp and Villareal, 2013, Malviya et al., 2016). Examples include blooms in the eastern Equatorial Atlantic (Foster and Zehr, 2006), tropical North Atlantic (Carpenter et al., 1999, Subramaniam et al., 2008), North Pacific Subtropical Gyre (Dore et al., 2008, Villareal et al., 2011), and South China Sea (Grosse et al., 2010). Furthermore, at the ALOHA site in the Pacific Ocean north of Hawaii, blooms of the *Hemiaulus-Richelia* association can last as long as 30 days and contribute significantly (20%) to annual carbon flux in this region (Karl et al., 2012, Kemp and Villareal, 2018). As described by these examples, diatoms with nitrogen-fixing symbionts are thus important contributors to primary productivity and carbon export, especially at times when surface waters are depleted of dissolved inorganic nitrogen (Pyle et al., 2020, Tang et al., 2020).

An intense bloom of *Hemiaulus* and its symbiont *Richelia* was observed in summer 2019 in temperate Northeast U.S. Shelf (NES) surface waters. The NES region in the Northwestern Atlantic Ocean is particularly productive, favoring enhanced inorganic carbon sequestration by the biological pump, and supports an ecologically and economically important ecosystem (Townsend et al., 2006). Like other marine regions, the NES ecosystem is fueled by phytoplankton which are the main primary producers and fundamental components to ecosystem function (e.g. Mouw and Yoder, 2005, O'Reilly and Zetlin, 1998, Yoder et al., 2002). Productivity is heavily influenced by abiotic factors in the NES region. For instance, strong seasonal variations in water temperature, stratification and cross-shelf advection on the NES affect nutrient supply and lead to seasonal shifts in phytoplankton productivity and species composition (Li et al., 2015, Oliver et al., 2022, Zhang et al., 2023). Furthermore, the water temperature of the NES is rising faster than the global average (Chen et al., 2020, Karmalkar and Horton, 2021, Shearman and Lentz, 2010), leading to unknown consequences for phytoplankton community composition and productivity within this important and dynamic coastal region.

To further understand phytoplankton population dynamics and their influence on the ocean's biological pump, the NES Long-Term Ecological Research (NES-LTER, https://nes-lter.whoi.edu/) project investigates primary productivity, food web structure and ecosystem dynamics with a focus on southern New England coastal waters. As part of the NES-LTER project, phytoplankton and zooplankton community composition, phytoplankton growth rates, microzooplankton grazing rates, and productivity rates are determined on week-long research cruises which have occurred quarter-annually since 2018. To quantify productivity, several different rates are estimated from data collected on these cruises, including Gross Oxygen Production (GOP), Net Primary Production (NPP), Net Community Production (NCP) and export efficiency ratios (NCP/GOP). GOP is similar to Gross Primary Production; it represents total photosynthesis in oxygen units and also includes photoprocesses that produce oxygen (Juranek and Quay, 2013). NPP is photosynthetic production minus autotrophic respiration and thus represents the net production activity of the phytoplankton community. NCP is the balance of photosynthesis and community respiration (autotrophic plus heterotrophic) and is equal, on long enough spatial and temporal scales, to the amount of carbon exported out of the surface of the ocean (Emerson, 2014). The NCP/GOP ratio, analogous to the f-ratio (Dugdale and Goering, 1967), is indicative of export efficiency, with a high ratio implying that the community is exporting most of the carbon (organic matter) produced and recycling only a little (Juranek and Quay, 2013).

The composition and size structure of the phytoplankton community in the NES-LTER study are investigated concurrently from automated imaging and size-fractionated chlorophyll-a (Chl-a). In winter, the NES waters tend to be nutrient-rich due to enhanced vertical mixing and input of river and estuary waters that promote high levels of surface Chl-a, with a dominance of large phytoplankton cells (>10 μm) that are growing slowly (Marrec et al., 2021). Conversely, during a typical summer, nutrients become depleted in the surface mixed layer, leading to low Chl-a concentrations dominated by fast-growing small phytoplankton cells (<10 μm) (Marrec et al., 2021, O'Reilly and Zetlin, 1998).

To complement production estimates and phytoplankton community structure observations, the flow of carbon from primary producers to higher trophic levels is investigated. Microzooplankton, protists smaller than 200 μm, are a crucial link between primary producers and higher trophic levels because they often consume 60–70% of daily primary production (Landry and Calbet, 2004, Schmoker et al., 2013). In the NES, while phytoplankton grow faster during the summer than in winter, microzooplankton grazing rates tend to stay relatively constant across seasons (Marrec et al., 2021). Thus, during winter, phytoplankton growth rates and microzooplankton grazing rates are typically well coupled and show a close 1:1 ratio, with microzooplankton consuming most of the primary production (Marrec et al., 2021). During the summer, the phytoplankton growth and microzooplankton grazing rates are typically decoupled, with higher growth rates than in winter, but grazing rates in the same range, leading to less than 50% of the primary production consumed by microzooplankton. The degree of coupling between microzooplankton grazing and phytoplankton growth rates is associated with phytoplankton size structure (Marrec et

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al., 2021) and likely composition, and is an important indicator of the trophic transfer efficiency from phytoplankton to microzooplankton, at the basis of the planktonic food web.

Here, we examined the association between productivity, phytoplankton composition and microzooplankton grazing, key components of trophic transfer efficiency and thus ecosystem function. During a NES-LTER cruise in summer 2019, we observed an anomalous relationship between growth and grazing rates, as well as dramatically different productivity rates and community composition compared to other summer cruises in the NES region. We thus investigated how a diatom bloom of *Hemiaulus* with diazotrophic symbionts affected metrics of productivity and grazing on the NES during the summer of 2019. Our results provide insights into the effects of community composition on productivity rates.

2 Methods

Measurements of environmental conditions, chemical and biological stocks, and productivity and grazing rates were conducted on multiple cruises within the framework of the NES-LTER program (Table 1). Measurements from three other cruises from different projects on the NES were also included in this analysis for comparison (project names in Table 1). From this time series, we were able to better understand an event observed on the 2019 NES-LTER summer cruise (EN644) which occurred from August 19 to 25 (Table 1). Some data during that event, such as surface seawater temperature (SST), salinity (SSS), NCP rates, and phytoplankton composition were collected continuously from the underway system (i.e., km-scale resolution), while other parameters (e.g., NPP, grazing rates, Chl-a, nutrients) were measured discretely at the NES-LTER stations. Main stations were located with ~ 19 km spacing on a north-to-south transect primarily along 70º 53' W (Fig. 1). In particular, the mid-shelf region, which is where the *Hemiaulus* bloom primarily occurred, corresponds to 50 – 100 m water depth (Fig. 1) and contains 4 stations. At each station, water was collected via Niskin bottles mounted on a CTD-rosette (conductivity-temperature-depth, Seabird SBE32 Carousel Water Sampler). The CTD-rosette system consisted of a 24-bottle rosette frame with 10-L Niskin bottles. Depth, temperature, and salinity were collected with a SBE911 CTD (Seabird Electronics) equipped with additional sensors for chlorophyll fluorescence (WET Labs ECO-AFL/FL), photosynthetically active radiation (PAR, Biospherical Instruments® QSP2000), and beam attenuation (WET Labs C-Star 25-cm transmissometer). The Niskin bottles were closed at various depths ranging from surface to near bottom, based on the depths of the mixed layer, euphotic zone, and Chl-a maximum. Water from the Niskins was used to quantify a number of parameters as described in Sections 2.2 through 2.5.
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Table 1. Dates of the summer cruises, as well as project and ship names and cruise numbers, that are presented in this paper. Project name abbreviations are as follows: OTZ– Ocean Twilight Zone, SPIROPA–Shelfbreak Productivity Interdisciplinary Research Operation at the Pioneer Array (Oliver et al., 2021), and EcoMon–Ecosystem Monitoring program run by the National Oceanic and Atmospheric Administration.

Cruise Name	Start date/End date	Project name	Ship name
EN617	20 July 2018 – 25 July 2018	NES-LTER	*R/V Endeavor*
TN368	05 July 2019 – 18 July 2019	SPIROPA	*R/V Thomas G. Thompson*
HB1907	25 July 2019 – 08 Aug 2019	OTZ	*NOAA Ship Henry B Bigelow*
GU1902	16 Aug 2019 – 29 Aug 2019	EcoMon	*NOAA Ship Gordon Gunter*
EN644	20 Aug 2019 – 25 Aug 2019	NES-LTER	*R/V Endeavor*
EN655	25 July 2020 – 28 July 2020	NES-LTER	*R/V Endeavor*
EN668	16 July 2021 – 21 July 2021	NES-LTER	*R/V Endeavor*
EN687	29 July 2022 – 03 Aug 2022	NES-LTER	*R/V Endeavor*

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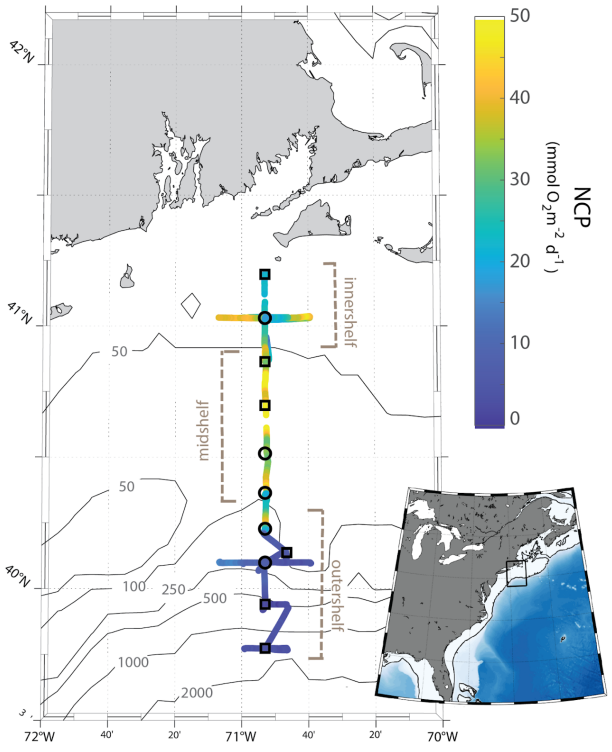

Fig 1. Map of the NES-LTER August 2019 cruise track colored according to rates of NCP as measured continuously by an at-sea mass spectrometer for the first half of the cruise. Station locations are marked with circles or squares and are ~19 km apart. Rates of NCP, GOP, and phytoplankton community composition were quantified at all stations. Grazing rates and NPP were calculated only at stations marked by squares. Other NES-LTER cruises have a similar track although issues such as weather sometimes change the track slightly. The inset shows the location of the cruise (rectangular box) in the context of the east coast of the United States.

The underway system consisted of continuous surface seawater pumped throughout the ship by an impeller pump and a diaphragm pump located near the ship's bow. Using water from the impeller pump, continuous measurements of surface temperature and salinity were obtained from a Seabird SBE38 (temperature) sensor installed at the water intake and by a Seabird SBE45 sensor (temperature and salinity) located further away in the underway system. Because the diaphragm pump is less likely to damage plankton, its underway flow was used for measurements to quantify NCP (Section 2.1), GOP (Section 2.2) and phytoplankton community composition (Section 2.6

2.1 Net Community Production

Net community production rates were calculated from O_2/Ar ratios measured by an at-sea Equilibrator Inlet Mass Spectrometer (EIMS) (Cassar et al., 2009) analyzing water from the ship's underway system and from discrete samples collected from both CTD Niskin bottles and from the underway system. The EIMS was used to collect continuous data on O_2/Ar ratios via the diaphragm pump of the underway system that, on the *R/V Endeavor*, pumps seawater from a depth of 5 m. The underway system seawater flows through a debubbler into a bucket at a constant rate that allows for continuous overflow for consistent head pressure. Water is then pumped from the bucket at ~1.1 L min^{-1} by a gear pump through two filters: a bag with a 25-µm pore size, and a 2-layered sock with a 5-µm inner and 100-µm outer pore size. The gear pump then pushes the water through an equilibrator membrane contactor

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cartridge (Liqui-Cel Extra-Flow 2.5x8 model G540). The equilibrated headspace gas from the cartridge is then dried by flowing through the dessicants Nafion and Drierite and then passed via a fused silica capillary into a Hiden Residual Gas Analyzer (RGA) (HAL 7) quadrupole mass spectrometer. Details of the equilibration method can be found in (2016), but in this instance were modified to not use SAES getters as they would have removed the O_2. The EIMS was operated throughout the whole cruise (starting one hour after the ship left port and ending a few hours before return to port). To calibrate the mass spectrometer, the capillary was switched to an air inlet for twenty minutes approximately every six hours as the ratio of O_2/Ar in air is stable and well-known. Additionally, bottle samples were collected from the underway system at least once per day and were subsequently measured on an isotope ratio mass spectrometer at Woods Hole Oceanographic Institution (see Section 2.2). These bottle samples were used to provide additional calibration as necessary–such additional corrections changed the O_2/Ar ratios by at most 0.67%.

The O_2/Ar ratios were then used to calculate NCP (Hendricks et al., 2004, Juranek and Quay, 2005, Stanley et al., 2010). With data from the EIMS and the bottle samples, the biological oxygen saturation $\Delta(O_2/Ar)$ was calculated via the equation below:

$$\Delta\left(\frac{O_2}{Ar}\right) = \frac{\left(\frac{O_2}{Ar}\right)_{smpl}}{\left(\frac{O_2}{Ar}\right)_{eq}} - 1 \quad (1)$$

where $(O_2/Ar)_{smpl}$ represents the ratio of O_2 to Ar ion currents detected by the EIMS after being calibrated with bottle data, and $(O_2/Ar)_{eq}$ represents the ratio of equilibrium concentrations of the gases determined from the gases' solubility (Garcia and Gordon, 1992, Hamme and Emerson, 2004) at the seawater temperature and salinity.

The NCP integrated over the mixed layer, in units of mmol O_2 m^{-2} d^{-1}, is calculated as

$$NCP = \Delta\left(\frac{O_2}{Ar}\right) \lfloor O_2 \rfloor_{eq} k\rho \quad (2)$$

where $[O_2]_{eq}$ represents the equilibrium concentration of O_2 at the relevant temperature and salinity (mmol kg^{-1}), k is the weighted gas transfer velocity (m d^{-1}), and ρ is the density of seawater (kg m^{-3}) (Millero and Poisson, 1981). The weighted gas transfer velocity is a time-weighted average from over the past 30 days calculated as described in Reuer et al. (2007), with the gas exchange parameterization of Stanley et al., (2009) and wind speeds from NCEP Reanalysis (Kalnay et al., 1996, Kistler et al., 2001). Many physical considerations altering O_2 saturations, such as changes in temperature and bubble injection, do not need to be considered due to the inclusion of Ar which has similar solubility and diffusivity as O_2; however, a few assumptions were made for these calculations. Firstly, this equation assumes steady state within the mixed layer, i.e. no change in O_2/Ar in the ocean with time. While O_2/Ar was likely changing in actuality, assuming steady state simply means that the rates calculated reflect an exponentially weighted average of NCP over the past few residence times of oxygen (residence time equals a few days in these conditions) (Teeter et al., 2018). Thus, the assumption of steady state does not majorly impact our conclusions. We were not able to calculate the time rate of change term in O_2/Ar (Manning et al., 2017b) because the cruise was not Lagrangian, and even though the ship returned to the same geographic location, the water at that location changed due to ocean currents. To check the assumption that there is negligible respiration within the ship's lines (Juranek et al., 2010), bottle samples were collected in duplicate from Niskins at the same time as samples were collected from the underway system several times during every cruise; gas concentrations in the bottle samples from the underway and Niskin were identical within measurement errors, confirming there was no detectable respiration in the ship's line.

2.2 Gross Oxygen Production

Discrete samples of triple oxygen isotopes (TOI) were collected from the surface Niskin bottles on the CTD-rosette system at all stations as well as from the underway system between stations. Samples from the CTD-rosette system were also collected from bottles fired at ~ 5 m below the mixed layer and often one greater depth to provide information for assessing whether vertical corrections to O_2/Ar ratios were significant. Samples were collected in custom-made ~500-mL sample bottles which were pre-poisoned with 100 µl of saturated mercuric chloride solution and filled with around 300 mL of seawater from the underway system or from the Niskin at each station (Stanley et al., 2015). Samples were brought to Woods Hole Oceanographic Institution where they were analyzed for TOI with a custom-made processing line and a Thermofisher MAT 253 isotope ratio mass spectrometer as detailed in Stanley et al (2015). The same samples were also analyzed for O_2/Ar which yielded rates of NCP from discrete data as well as an independent method for calibrating the EIMS (see above). Corrections for the effect of argon on the triple oxygen isotope ratio and the effect of varying sizes of the sample vs. reference standard were made for every

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sample. Reproducibility from duplicate samples collected on these cruises ranged from 4 to 8 per meg for $^{17}\Delta$, 0.008 to 0.03 per mil for $\delta^{17}O$, and 0.008 to 0.05 per mil for $\delta^{18}O$ depending on the cruise.

From these samples, GOP is calculated in units of mmol O_2 m^{-2} d^{-1} following Prokopenko et al., (2011) according to:

$$GOP = kO_{eq}\frac{\frac{x_{dis}^{17}-x_{eq}^{17}}{x_{dis}^{17}}-\lambda\frac{x_{dis}^{18}-x_{eq}^{18}}{x_{dis}^{18}}}{\frac{x_{P}^{17}-x_{eq}^{17}}{x_{dis}^{17}}-\lambda\frac{x_{P}^{18}-x_{eq}^{18}}{x_{dis}^{18}}} \quad (3)$$

where k again represents the time-weighted gas transfer velocity (m d^{-1}), O_{eq} represents the equilibrium concentration of oxygen, λ represents the respiration slope factor = 0.5179, X_{dis}* represents the ratio of isotopes (*O/^{16}O) dissolved in the sample, X_{eq}* represents the ratio of isotopes (*O/^{16}O) dissolved in seawater equilibrated with the atmosphere, and X_P* stands for the ratio of isotopes (*O/^{16}O) in oxygen that was produced via photosynthesis. The photosynthetic end member used was the average of the phytoplankton value determined by Barkan and Luz (2011), and Vienna Standard Mean Ocean Water (VSMOW) was used for the isotopic composition of oxygen in H_2O. The actual isotopic composition of H_2O was measured in a subset of samples to see if corrections needed to be made (Manning et al., 2017a). It was found to be very similar to VSMOW, leading to an error of less than 10% in GOP due to isotopic water variations.

Confirmation that the water from the underway system was representative of the oceanic TOI signature of dissolved oxygen was obtained by comparing samples collected from the underway system to those collected concurrently from the surface Niskin bottle. All cruises, other than 2019, showed that there was statistically no difference in TOI between water from the underway system and the CTD and thus that the water from the underway system was representative of the mixed layer at that location and time. During the summer of 2019, the water from the underway system had TOI values 4.1 per meg lower than that from the CTD – this is within measurement errors but since it might have led to systematic biases, we corrected for this offset before calculating GOP from the data. The GOP rates, along with the NCP rates, represent productivity integrated throughout the mixed layer.

2.3 Net Primary Productivity

Water samples for NPP were collected at 4-7 stations (cruise dependent) from 3-4 depths (station dependent) from the Niskins on the CTD-rosette system. During collection, water was pre-filtered through 200-µm mesh (to remove mesozooplankton) into acid-washed 2-L polycarbonate bottles. Water collection and associated incubation occurred in triplicate for surface samples at each station. Bottles were spiked with a solution of 99% $NaH^{13}CO_3$ (Cambridge Isotope Lab, Tewksbury, MA) for a final 10% enrichment of the dissolved inorganic carbon (DIC) pool and placed in various mesh bags to simulate *in situ* light levels. Bottles were incubated for 24 h in clear deck-board incubators with flowthrough seawater and Onset HOBO data loggers monitored tank water temperature. At each station, the natural ^{13}C in the water was determined from an un-spiked sample and dark carbon assimilation was determined from a spiked dark bottle sample. Dark carbon assimilation was negligible (<1%) so no correction for dark carbon assimilation was applied to this dataset.

The corresponding light levels at collection depths were determined using either PAR or beam attenuation from the CTD cast for each station. When PAR data were not available (e.g., night-time casts), a relationship was established (eq. 4) with previous daytime cast information between beam attenuation (c, measured by transmissometer, m^{-1}) and the light extinction coefficient (Kd, m^{-1}) for each cruise. During night-time casts, Kd was estimated from the average c in the upper 10 m during the cast with the slope (m) and intercept (b) from the daytime plot, according to equation 4:

$$K_d = (m * At) + b \quad (4)$$

The appropriate shading in incubations (%PAR) for each depth of sample collection (z) was estimated as:

$$\%PAR = 100e^{-K_d \times z} \quad (5)$$

At the end of each incubation, bottles were filtered under low vacuum (5-10 in. Hg) over pre-combusted Whatman GF/F filters (450˚C; 6h). Filters were stored at -20˚C until further analysis on shore. NPP rates were quantified by measuring the incorporation of isotopically heavy carbon into phytoplankton biomass. Prior to measuring ^{13}C in the samples, filters were acid fumigated with concentrated HCl in a desiccator overnight to remove inorganic carbon. They were dried in an oven at 60˚C for 24 h, individually wrapped in tin capsules and analyzed on a Carlo Erba NC2500 elemental analyzer interfaced with a Thermo Delta V+ isotope ratio mass spectrometer. The δ

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^{13}C values were reported relative to the international standard Vienna PeeDee Belemnite (Coplen, 1995) and converted to atom percent values.

NPP rates were calculated from atom percent values with the equation from Hama et al. (1983)

$$NPP = \frac{POC \cdot (a_{is} - a_{ns})}{t \cdot (a_{ic} - a_{ns})} \quad (6)$$

where NPP is the net primary production rate ($\mu g \cdot L^{-1} \cdot day^{-1}$), POC is the particulate organic carbon; ($\mu g\ L^{-1}$), t is the incubation time (h), a_{is} is the atom % of ^{13}C in the incubated sample, a_{ns} is the atom % of ^{13}C in the natural sample (un-spiked sample described above) and a_{ic} is the atom % of ^{13}C in the total DIC pool. POC measurements were blank corrected with the mean value of triplicate combusted filter blanks. The DIC concentration was determined from salinity (S) according to the following equation from Parsons et al. (1984):

$$DIC = \left((S * 0.067) - 0.05\right) * 0.96 \quad (7)$$

NPP rates were integrated to the depth of the mixed layer (Table S1) to align with NCP and GOP integrated rate calculations. The mixed layer depths were calculated from the temperature and salinity data from the CTD with the threshold method where the mixed layer was declared to be the depth where the density difference between the surface density and the mixed layer was greater than $\Delta\sigma_\theta = 0.125$ kg m^{-3} (de Boyer Montegut et al., 2004). Mixed layer depths were confirmed to be similar when a gradient criterion with a difference of 0.0125 kg m^{-3} was used instead (Kara et al., 2000). Primary production rates for 2018 were estimated from the growth/grazing rates. The surface values of phytoplankton growth rates were converted from Chl-a to carbon (mg C m^{-3} d^{-1}) with a constant ratio of 50 which was then multiplied by the mixed layer depth to get values in mg C m^{-2} d^{-1}.

2.4 Autotrophic and Heterotrophic Respiration

Assuming a photosynthetic quotient (O:C ratio) of 1.4, respiration rates were calculated from the productivity values (GOP, NPP, and NCP) and following the relationships below:

$$NPP = GOP - R_A \quad (8)$$

$$NCP = NPP - R_H \quad (9)$$

where R_A is autotrophic respiration and R_H is heterotrophic respiration.

2.5 Growth Rates and Grazing rates

Rates of phytoplankton growth and protistan grazing were quantified with a 2-point modification of the dilution method (Chen, 2015, Landry et al., 2008, Morison et al., 2020) following methods in Marrec et al. (2021). Briefly, surface samples were collected at 4 to 7 stations throughout the cruise. For each sample, whole seawater (WSW) from the Niskin bottles was transferred into a 10-L polycarbonate carboy through a 200-µm mesh filter to remove mesozooplankton predators. Diluent was prepared by gravity filtration through a 0.2 µm membrane filter capsule (PALL®) from the Niskin to the carboys and mixed with WSW to obtain a 20% WSW dilution. A total of 6 bottles per experiment were prepared: 2 bottles with nutrient amended 20% WSW, 2 bottles with nutrient-amended WSW, and 2 bottles with unamended WSW to assess nutrient limitation. Incubations took place for 24 h in a clear, 1m^3 deck-board incubator. Paired bottles were placed into mesh bags that simulated the effective light availability in the surface mixed layer, which corresponded to 65% of sea surface irradiance. Phytoplankton growth and grazing mortality rates were then estimated from changes in Chl-a over the 24 h incubation. For dilution experiments, Chl-a concentrations were obtained from triplicate 150-mL subsamples filtered on GF/F filters, after a 12-h dark extraction period at room temperature in 95% ethanol and measured on a calibrated Turner 10 AU fluorometer. The full extraction method is detailed in Marrec et al. (2021).

2.6 Discrete Chlorophyll-a sample collection and processing

Samples for Chl-a analysis were collected into brown amber bottles from Niskins on the CTD Rosette system. A known sample volume (250-500 mL) was filtered at low pressure (5-10 in. Hg) through either a GF/F filter or a 20 µm polycarbonate Sterlitech filter. Filters were transferred to either tissue capsules (GF/F) or cryogenic vials (20 µm) and then flash frozen in liquid nitrogen until extraction. Later, filters were extracted in 5 mL of 90% acetone for 24 hours in a dark refrigerator, then tubes were vortexed and centrifuged (only GF/F filters), and the solution was

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measured on a calibrated Turner Designs Handheld Aquaflor fluorometer, acidified with 2 drops of 10% hydrochloric acid and measured again. Chl-a concentrations for different size fractions were calculated by difference.

2.7 Satellite and radar data

To look at variability in SST and surface Chl-a, a proxy for phytoplankton biomass, throughout the summers over multiple years, on a wider spatial and temporal scale than the at-sea chlorophyll data permitted, SST and surface Chl-a concentrations from remote sensing sources were retrieved and analyzed. In particular, both snapshots and monthly averages of MODIS (Moderate Resolution Imaging Spectroradiometer) SST and chlorophyll data with a horizontal resolution of 1 km were used to examine the spatial coverage of the *Hemiaulus* bloom in summer 2019 (when it dominated phytoplankton biomass) and compare the surface temperature and chlorophyll in the NES region in summers 2018-2022.

To examine possible origins of the bloom water, backward particle trajectory simulations were carried out with the OceanParcels Python package https://oceanparcels.org/index.html (Lange and van Sebille, 2017). High frequency (HF) radar-measured sea surface velocity data in the NES region in Jul-Aug 2019 with 6-km spatial resolution and hourly temporal resolution were used as the background flow. Particles were released at mid-shelf sites along the NES-LTER transect on Aug 21, 2019 and advected backward for 30 days until Jul 22, 2019.

2.8 Imaging FlowCytobot

Composition of the phytoplankton community was assessed with Imaging FlowCytobots (IFCB; McLane Research Laboratories, Inc.). IFCB uses a combination of video and flow cytometry technology to capture images of plankton and other particles in the size range ~5-150 µm (Olson and Sosik, 2007). During the cruises reported here, IFCB instruments were configured to record images of particles with laser-based chlorophyll fluorescence or light scattering signals above trigger thresholds and samples were pre-screened with 150 µm Nitex. IFCB instruments were operated two ways. First, on all cruises, an IFCB was configured to sample 5 mL automatically from the ship's underway system every 25 minutes. Second, at stations occupied on the NES-LTER and SPIROPA cruises, IFCB instruments were used to analyze depth profiles from discrete samples collected with Niskin bottles. Typically, three 5-mL subsamples were measured for each depth. The fraction of each 5-mL sample imaged by IFCB decreases with increasing trigger rate but is recorded precisely during sample acquisition enabling calculation of concentrations. IFCB image data were automatically analyzed following approaches developed for the IFCB time series at the Martha's Vineyard Coastal Observatory (MVCO) (Brownlee et al., 2016). In particular, cell biovolume was estimated from IFCB images (Moberg and Sosik, 2012) and converted to cell carbon following the relationships described by Menden-Deuer and Lessard (2000). IFCB images were classified with a convolutional neural network (CNN) trained to separate 155 categories of plankton and other particles observed at MVCO and across the NES region. We used the Inception v3 (Szegedy et al., 2016) CNN architecture as implemented in PyTorch, pre-trained with ImageNet (Russakovsky et al., 2015) and fine-tuned with an NES IFCB training set (97026 images, 155 classes, 80-20 split for training and validation). In addition, an independent test set of manually annotated images in 51 IFCB samples from EcoMon cruises was used to evaluate *Hemiaulus* quantification as a function of classifier score threshold. From this independent analysis, classifier predictions with scores above 0.9 performed very well for *Hemiaulus* (class-specific F1-score = 0.936; CNN-count vs. manual-count: $r^2 = 0.999$, slope = 0.915; intercept = 0.005).

2.9 Nutrients

Dissolved inorganic nutrient concentrations (ammonium, phosphate, silicate, and nitrate + nitrite) were obtained from CTD bottle samples with duplicates. Seawater was passed through an EMD Millipore sterile Sterivex 0.22 µm filter with filtrate collected into acid-washed 20-ml scintillation vials (after triplicate rinses), which were then stored at -20 °C until analysis. Samples were processed at Woods Hole Oceanographic Institution's Nutrient Analytical Facility with a four-channel segmented flow SEAL AA3 HR Autoanalyzer. Detection levels are as follows: 0.01 µmol L^{-1} for silicate, 0.03 µmol L^{-1} for phosphate, 0.04 µmol L^{-1} for nitrate + nitrite, and 0.03 µmol L^{-1} for ammonium.

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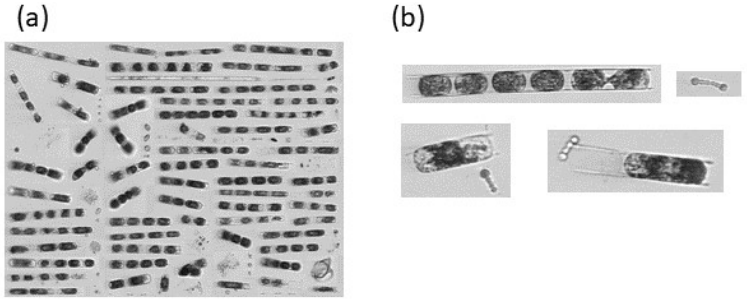

Fig 2. IFCB images of the **(a)** phytoplankton community during the summer 2019 NES-LTER cruise and **(b)** closer up individual *Hemiaulus* cells found with the nitrogen-fixing symbiont *Richelia*.

Results

3.1 *Hemiaulus* distribution and Chlorophyll

During the NES-LTER summer 2019 cruise, through automated image classification and analysis and through visual microscopic confirmation, a bloom of the diatom genus *Hemiaulus* was observed in the surface waters of the mid-shelf region (Fig. 2a). These images also showed N_2 fixing symbionts, namely *Richelia,* inside or next to the *Hemiaulus* cells (Fig. 2b). Additionally, *Hemiaulus* carbon was highest in the mid-shelf waters between latitudes of 40.1° N and 41.1° N, a span of 111 km (Fig. 3a), with concentrations ranging from 6.8 to 68.3 µg L^{-1}. This bloom was only observed in the surface waters of the mid-shelf region, as can be seen by discrete IFCB measurements from Niskin samples (Fig. 3b). *Hemiaulus* carbon concentrations observed in other years on NES-LTER transect cruises never reached values above 0.30 µg L^{-1}.

The presence of the diatom bloom was consistent with the size-fractionated Chl-a data. Surface Chl-a concentrations in the mid-shelf region in summer are typically low (< 0.50 µg L^{-1}, Fig. 4a) and progressively decrease with decreasing latitude.. However, during the NES-LTER summer 2019, Chl-a concentrations were observed up to 3.50 µg L^{-1} in the surface waters of the mid-shelf (mean Chl-a of 1.96 µg L^{-1}, Table 2) with up to 80% of the Chl-a associated with the > 20 µm fraction (Fig. 4c). This is in contrast to other summers when most of the Chl-a was

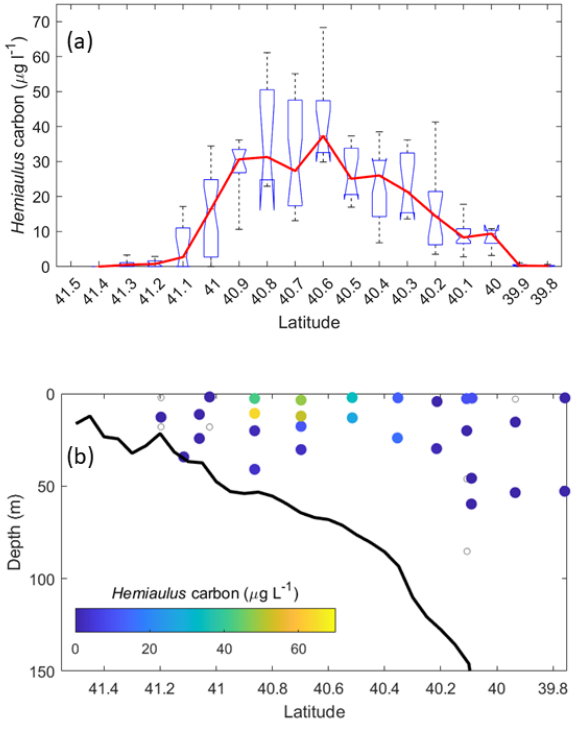

Fig 3. Cross-shelf distribution of *Hemiaulus* carbon concentration in August 2019 showing the mixed-layer bloom in the mid-shelf region, corresponding to water column depths ranging from approximately 50 to 100 m. Results are derived from IFCB observations in **(a)** surface waters and **(b)** discrete samples from depth profiles with symbols color-coded by *Hemiaulus* carbon concentration (open symbols indicate samples where *Hemiaulus* was not detected).

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associated with the < 20 µm fraction (Fig. 4b, d-f). Concentrations of Chl-a in the > 20 µm size fraction and concentrations of *Hemiaulus* carbon in the NES-LTER summer 2019 cruise were larger at co-located sampling locations in the beginning of the cruise than at the end, suggesting that the bloom may have peaked before the cruise started and thus was in decline during the cruise period.

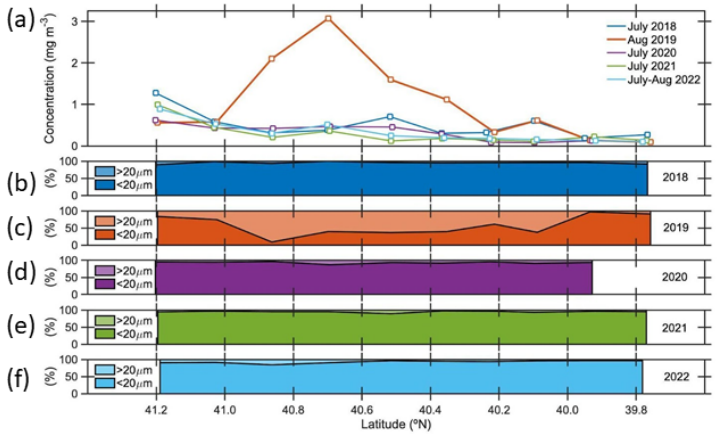

Fig 4. (a) Surface Chl-a concentration versus latitude for NES-LTER summer cruises 2018-2022. **(b-f)** Percentages of surface Chl-a associated with the > 20 µm phytoplankton (lighter shade) and < 20 µm phytoplankton (darker shade) versus latitude for each year. Note that the August 2019 cruise had the largest surface Chl-a concentrations and also the largest fractions associated with the > 20 µm fraction.

Monthly mean surface Chl-a concentrations from remote sensing were used to investigate if the observed differences in Chl-a and productivity between the summers were related to differences in the timing of the cruise as opposed to differences in in community composition (Fig. S1). In many of the summers (2018, 2021, and 2022), Chl-a in July was actually higher than in August, suggesting that the timing of the 2019 cruise (end of August instead of end of July) was not a factor in explaining the anomalous results. In 2019, in situ observations from both July and August confirm the conclusion from the satellite data that the transect area had larger Chl-a concentrations in August than in July. In particular, a patch of high chlorophyll in the mid-shelf between 40.2°N and 40.7°N to the immediate west of the transect was present in August 2019, likely resulting from the observed *Hemiaulus* bloom.

3.2 Physical properties

During the NES-LTER summer 2019 cruise, SST was higher than during most of the other summer cruises in the mid-shelf region (Table 2). Furthermore, SST in summer 2019 on the outer-shelf region in particular was substantially higher than on any other summer cruise (Fig. 5a). Notably, the 2019 cruise occurred later in the summer season (August) than the NES-LTER cruises in other years (July). Along the NES-LTER transect specifically, SST in July 2019 was lower than in August 2019 and was similar to other years. In general, monthly-averaged satellite SST data in the broader NES region usually show lower SST values in July compared to August (2018, 2020-2022) (Fig. S2). Interestingly, however, in summer 2019, the monthly averaged satellite data actually show higher SST in July, because of impingement of a Gulf Stream warm-core ring on the shelf edge (Zhang et al., 2023) and the subsequent onshore intrusion of the ring water in July 2019. The fact that monthly averaged satellite SST was higher in July than August but the local NES LTER transect data shows temperature higher in August than July suggests that the high SST observed during late August 2019 reflected an ephemeral event and not a mean condition during that month. During the NES-LTER August 2019 cruise, surface salinity was lower than on 2018, 2021, and 2022 summer cruises, but similar to surface salinity during the July 2020 cruise (Fig. 5b) and to salinities observed in July 2019 along the NES-LTER transect. Despite the NES-LTER summer 2019 cruise being in a specific week of August and the conditions suggesting an ephemeral event, for simplicity, we will refer to it as August 2019 in this paper.

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Table 2. Averages (+/- standard error) of surface mixed layer characteristics (productivity values integrated through the surface mixed layer; physical conditions, nutrients, and Chl-a concentrations from the surface) in the mid-shelf region measured during NES-LTER summer cruises for each year and overall averages from NES-LTER winter cruises 2018-2022 in the mid-shelf waters of the transect. The NPP average in 2018 (*) was calculated based on the phytoplankton growth rate since direct NPP measurements were not available for this year. The standard error for 2020 NPP reflects the triplicate values from one station rather than multiple mid-shelf stations. The winter averages reflects mean of data collected in winters of 2018-2022.

		NCP (mmol O_2 $m^{-2}d^{-1}$)	GOP (mmol O_2 $m^{-2}d^{-1}$)	NCP/ GOP	NPP (mg C $m^{-2}d^{-1}$)	Phyto-plankton growth rate (d^{-1})	Micro-zoop grazing (d^{-1})	Temp-erature (ºC)	Salinity	*Hemiau-lus* Carbon (µg L^{-1})	Chl-a (mg m^{-3})	% Chl-a > 20 µm	Silicate (µmol L^{-1})	Phosphate (µmol L^{-1})
Summer	2018	28.8	160	0.18	* 324	1.12	0.19	21.44	32.71	0.002	0.43	0.02	1.9	0.11
		+/0.2	+/- 10	+/- 0.01	+/- 20	+/- 0.11	+/- 0.03	+/- 0.01	+/- 0.01	+/- 0.002	+/- 0.10	+/- 0.006	+/- 0.3	+/- 0.01
	2019	**28.8**	**110**	**0.24**	**398**	**0.20**	**0.17**	**24.29**	**31.83**	**28.4**	**1.97**	**1.37**	**0.27**	**0.025**
		+/- 0.2	**+/- 9**	**+/- 0.09**	**+/- 213**	**+/- 0.17**	**+/- 0.04**	**+/- 0.01**	**+/- 0.001**	**+/- 1.3**	**+/- 0.59**	**+/- 0.43**	**+/- 0.5**	**+/- 0.02**
	2020	11.3	100	0.13	225	0.80	0.22	23.88	31.80	0.00	0.40	0.03	1.4	0.085
		+/- 0.1	+/- 7	+/- 0.01	+/- 4	+/- 0.10	+/- 0.09	+/- 0.02	+/- 0.002	+/- 0.00	+/- 0.06	+/- 0.01	+/- 0.5	+/- 0.10
	2021	8.4	104	0.09	81	0.83	0.51	22.61	33.17	0.001	0.22	0.01	0.75	0.00
		+/- 0.05	+/- 26	+/- 0.01	+/- 10	+/- 0.64	+/- 0.25	+/- 0.02	+/- 0.01	+/- 0.001	+/- 0.07	+/- 0.004	+/- 0.4	+/- 0.00
	2022	3.8	40	0.07	191	0.31	0.11	24.46	32.53	0.003	0.32	0.03	0.42	0.06
		+/- 0.1	+/- 5	+/- 0.01	+/- 10	+/- 0.1	+/- 0.05	+/- 0.01	+/- 0.01	+/- 0.003	+/- 0.10	+/- 0.02	+/- 0.18	+/- 0.06
Winter	All	5.1	101	0.11	464	0.30	0.24	6.131	32.786	0.018	2.17	1.56	1.7	0.50
	yrs	+/- 0.1	+/- 20	+/- 0.04	+/- 63	+/- 0.04	+/- 0.05	+/- 0.007	+/- 0.002	+/- 0.006	+/- 0.16	+/- 0.14	+/- 0.5	+/- 0.30

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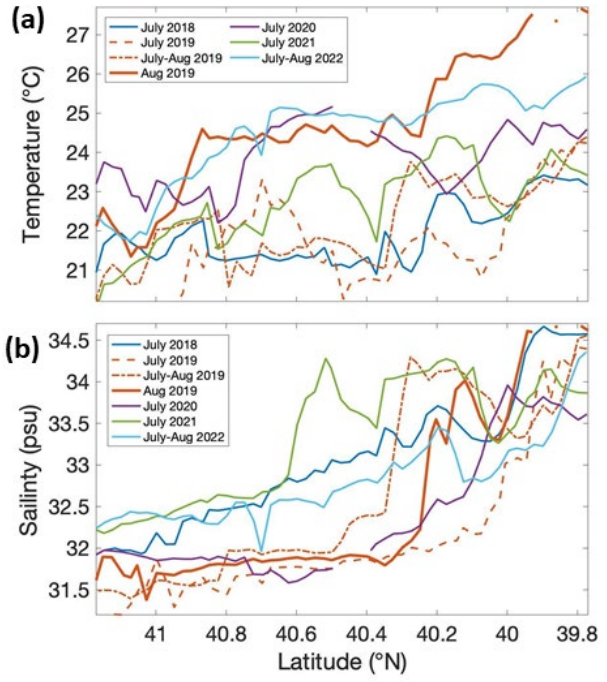

Fig 5. (a) Temperature and **(b)** salinity 5 m below the surface versus latitude for NES-LTER summer cruises (2018-2022) and the SPIROPA and OTZ summer 2019 cruises. For clarity, the values are averaged in 0.025 degree latitude bands when there were multiple occupations of the same region.

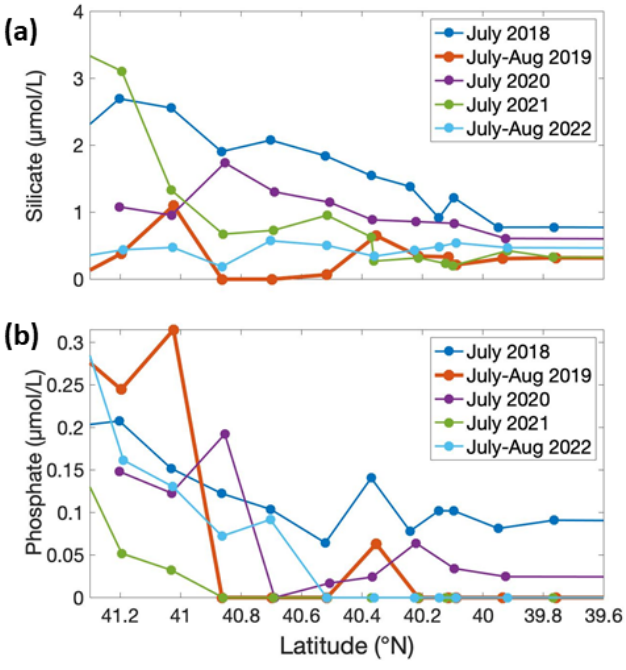

Fig 6. (a) Silicate and **(b)** phosphate concentrations (in µmol/L) in the upper 12 m of the water column for NES-LTER summer cruises (2018-2022).

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3.3 Nutrients

Nutrient concentrations differed between the August 2019 cruise and other summer cruises. Specifically, phosphate and silicate concentrations in surface waters were lower in August 2019 compared to other summers (Table 2). In other summers, silicate decreased with distance from shore, but in 2019, silicate was depleted between 41ºN and 40.4 ºN (Fig. 6a) coincident with the location of the *Hemiaulus* bloom. Additionally, higher levels of silicate were found around depths of 50 m to 140 m in August 2019 than during other summer NES-LTER cruises (Fig. S3), which may be associated with diatoms that had sunk and were starting to be remineralized, releasing silicate back into the water column. Surface water phosphate concentrations in August 2019 were depleted south of 41º N (Fig. 6b). However, low concentrations of phosphate were also found in summers of 2020 and 2021. Lastly, nitrate + nitrite concentrations were close to the detection level in the surface samples for all summer cruises except a few stations in 2018. Ammonium levels are not discussed because the samples were frozen at sea and thus may not be reliable; additionally, ammonium levels showed no clear relationship over the transect cruises.

3.4 Productivity and grazing rates

In August 2018 NCP was elevated in the mid-shelf waters, coincident with the location of the *Hemiaulus* bloom (Fig. 1). NCP peaked in the first half of the cruise and decreased during the second half, supporting the earlier supposition that the *Hemiaulus* bloom was likely in decline. Additionally the area of maximum NCP moved shoreward in the second half of the cruise. The high NCP was primarily constrained to the main longitude sampling line and usually did not extend, at least at those points in time, spatially off the main transect.

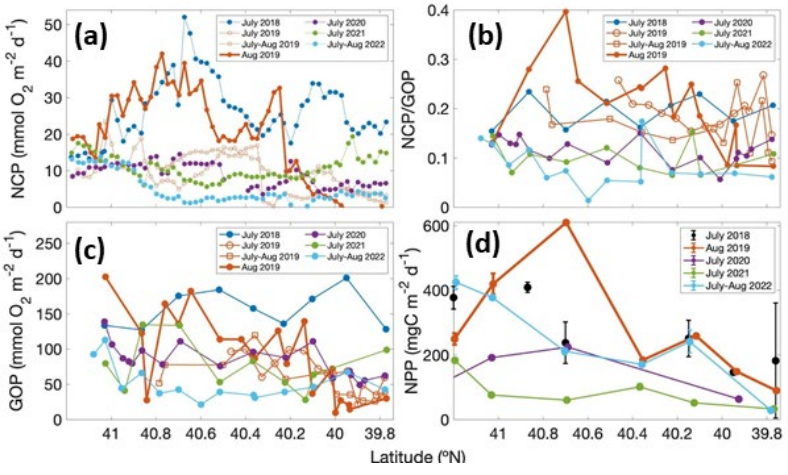

Fig 7. (a) NCP, **(b)** NCP/GOP, and **(c)** GOP rates integrated throughout the mixed layer for NES-LTER summer cruises (2018-2022) and SPIROPA and OTZ summer 2019 cruises. Values are averaged in 0.025 degree latitude bands to average multiple occupations of the same region. The same overall patterns are seen with and without the averaging within these latitude bands. **(d)** Average NPP values integrated to the bottom of the mixed layer for NES-LTER summer cruises (2019-2021) with error bars reflecting the standard deviation of triplicate surface water incubations. NPP values were not directly measured for summer 2018 but were instead estimated from phytoplankton growth rate in the grazing incubation experiments (black circles).

During August 2019, waters with high carbon concentrations of *Hemiaulus* showed higher rates of NCP (Fig. 7a), NCP/GOP (Fig. 7b), GOP (Fig. 7c), and NPP (Fig. 7d) compared to these rates at mid-shelf waters in most other years. More specifically, the mid-shelf waters where *Hemiaulus* was present in Aug 2019 displayed NCP values approximately 2.5 to 9 times larger than in the same mid-shelf latitudes in summers of 2020-22 (Table 2). Furthermore, we observed a correlation between NCP and *Hemiaulus* carbon between Aug 21 and Aug 23 (Fig. 8; $R^2 = 0.54$, $p < 0.001$). The patchiness of the diatom bloom corresponded to the patchiness in NCP. Additionally, one can see that the cooler shelf water was associated with higher abundances of *Hemiaulus* than the warmer slope water (Fig. 8), suggesting a water mass dependence on the location of the *Hemiaulus* bloom. Thus the patchiness in the bloom and NCP is likely a result of the ship crossing different water masses.

GOP rates were higher by a factor of 1.1 in waters with the *Hemiaulus* bloom in 2019 than during the summers of 2020-21; GOP rates were much higher in August 2019 than in summer of 2022 by a factor of 2.75.

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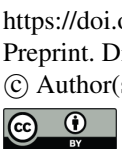

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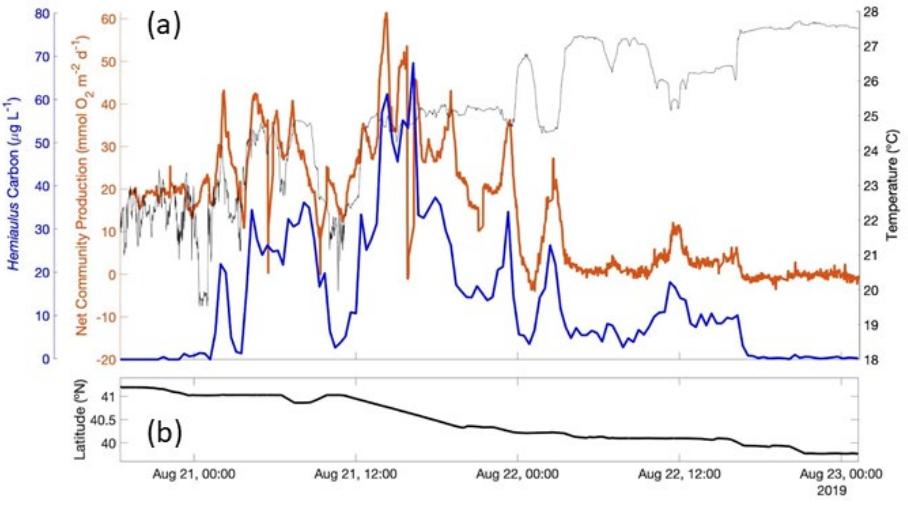

Fig 8. (a) *Hemiaulus* carbon concentration (blue), as estimated from IFCB data, showing a strong positive correspondence with NCP (orange) (R^2 = 0.54, p < 0.001) and a weak negative correlation with SST (black) (R^2=0.07, p=0.002) during one of the times during the August 2019 NES-LTER cruise that the ship was transiting the mid-shelf region, and **(b)** the latitude of the ship during the time period reflected in panel a.

Additionally, during August 2019, NPP rates were generally higher throughout the cruise compared to rates from July 2020–2022 and estimates for July 2018. Within the region that corresponds directly with the location of the *Hemiaulus* bloom, NPP rates in 2019 were ~1.5 - 2.5 times higher than NPP rates during other summer cruises. (Fig. 7d; Table 2). More specifically, NPP at 40.7 °N was approximately double the NPP measured in 2020 and more than double the rate measured in 2021. Furthermore, at 40.4 °N, NPP in 2019 was about 40% higher than in 2021 (no data for this station in 2020) (Fig. 7d).

A larger difference between NCP in the various summers than between GOP in the summers suggests that the increase in NCP in August 2019 was due to both increased photosynthesis and decreased community respiration. As a first approach, we calculated autotrophic respiration and heterotrophic respiration, to show that autotrophic respiration was lower than average in August 2019 (R_A = 308 mgC $m^{-2}d^{-1}$ in August 2019 versus 496 mgC $m^{-2}d^{-1}$ average for the other summers). This approach also showed that heterotrophic respiration was higher than average in August 2019 (431 mgC $m^{-2}d^{-1}$ in August 2019 versus 247 mgC $m^{-2}d^{-1}$ average for the other summers). NCP, GOP, and NCP/GOP rates in summer 2018 were comparable to these rates in August 2019 (discussed below in section 4.1).

Since the summer 2019 NES-LTER cruise occurred in the middle of August rather than in mid to late July as was typical for most other summers, the physical conditions were inherently different in 2019. We compared NCP and GOP data (NPP not available) from two earlier cruises in summer 2019 (cruise details in Table 1) whose stations overlapped with those of the LTER cruise (Fig. 7). These cruises occurred before the *Hemiaulus* bloom and their IFCB records did not show high abundance of *Hemiaulus* (< 1 µgC L^{-1}), although it was detected at that time. These two July 2019 cruises had much lower NCP rates compared to August 2019 NCP rates, specifically with rates similar to those observed in summer 2020-22 NES-LTER transect cruises (Fig. 7a & b). Together, these data suggest that higher production rates were uniquely tied to the presence of *Hemiaulus* rather than representing deviations in timing or environmental conditions.

The ratio of protistan grazing to phytoplankton growth rates provides an estimate of the percent of primary production (%PP) consumed by microzooplankton (Fig. 9). In contrast to typical summer conditions (> 0.6 d^{-1}, Table 2), during August 2019, phytoplankton growth rates during the *Hemiaulus* bloom were low (< 0.2 d^{-1}, Table 2), likely because the bloom was near its end, with most of the primary production consumed by microzooplankton (%PP = 84%, Table 2). Thus in August 2019, the phytoplankton growth and microzooplankton grazing were well coupled (though only in the part of the transect where *Hemiaulus* bloomed), like typical winter conditions, when the

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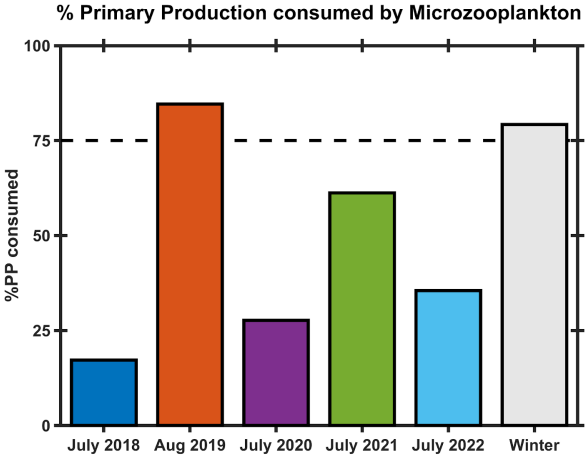

Fig 9. Percentage of primary production (%PP) consumed by microzooplankton in surface waters in the mid-shelf region during NES-LTER cruises for each summer and the overall average from NES-LTER winter cruises 2018-2022. %PP consumed by microzooplankton is calculated as the ratio of microzooplankton grazing rate (d^{-1}) to phytoplankton growth rate (d^{-1}).

phytoplankton community structure is dominated by large cells, instead of the decoupling typically observed in most summer conditions dominated by picoplankton (Marrec et al., 2021). We note that coupling between phytoplankton growth and microzooplankton grazing was occasionally observed during other summer cruises, but mostly in inner-shelf waters (except one mid-shelf station in July 2021). Overall, most of the primary production during the *Hemiaulus* bloom was grazed by microzooplankton indicating high trophic transfer efficiency from phytoplankton to microzooplankton.

4 Discussion

4.1 Change in community composition altering biological rates

A bloom of *Hemiaulus* has not been observed on any previous NES-LTER cruise and to our knowledge, has not been reported in the broader NES region before. Additionally, only in August 2019, compared to summers 2018 and 2020-22, was most of the Chl-a associated with the > 20 µm size fraction. Thus, the presence of the diatom bloom found in August 2019 is a major change in phytoplankton composition observed in this region of the NES that led to large changes in productivity rates and to the coupling between phytoplankton growth and grazing.

It is likely that the nitrogen-fixing symbionts in *Hemiaulus* allowed the diatom to thrive in the stratified, low nutrient surface waters of the summer shelf. This is supported by phosphate being drawn down to levels below detection only in August 2019; the nitrogen-fixing symbionts in the *Hemiaulus* likely made phosphate a limiting factor for growth (Tang et al., 2020) in August 2019 whereas nitrate limitation is typical for NES summer conditions. Additionally, silicate is especially important for diatoms because it is required for formation of their cell frustules. Moreover, previous studies show that the availability of dissolved silica seems to be an important control for many diatom-diazotroph blooms by affecting the growth rate and size of the diatom's frustules (Kemp and Villareal, 2013, Spitzer, 2015). The observed depletion of silicate and phosphate in the surface water during the August 2019 cruise suggests that, at the time of the cruise, the *Hemiaulus* bloom might have been on the decline. The very low phytoplankton growth rates might also support the idea that the bloom had peaked but also might be attributed to the inverse relationship between phytoplankton cell size and growth rate (e.g. Finkel et al., 2010). The higher levels of silicate observed at depth in August 2019 are likely due to the *Hemiaulus* sinking out of the euphotic zone and their frustules re-mineralizing at depth, which would release the silicate–and other nutrients–back into the

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water (Twining et al., 2014). In any case, it appears the *Hemiaulus* population could have been limited by phosphate, silicate, or both.

The strong coherence between the high spatial resolution data on *Hemiaulus* carbon concentrations and NCP (Fig. 8), as well as the other data presented here and a clear potential mechanism, strongly support the idea that the high productivity rates observed in August 2019 are directly due to the presence of *Hemiaulus*. In particular, the high NCP rates observed during the August 2019 NES-LTER cruise and their overlap with the location of the diatom bloom, suggest a high export ecosystem developed due to *Hemiaulus*' influence on productivity and biological rates. Here we are defining export as a flux away from the local biological production compartment, which can include losses of carbon (or oxygen) to depth or transfer to higher trophic levels. While the *Hemiaulus* bloom slightly increased total photosynthesis, as seen from the GOP rates, the bloom presence affected NCP, and thus by extension, export production, even more, potentially due to the large size of *Hemiaulus* cells and chains. The NCP/GOP ratio in August 2019 was double the ratio observed in the summers 2020-22 (Table 2). Other studies have shown links between variations in NCP/GOP and changes in planktonic community composition (Palevsky et al., 2016). Bigger phytoplankton cells sink faster than small ones, making them less likely to be grazed before sinking out of the euphotic zone, allowing for a higher export efficiency. Hence, the NES-LTER summer of 2019 cruise appears to represent a high carbon export efficiency system.

Not only did NCP and GOP rates change because of the *Hemiaulus* bloom, but so did NPP, phytoplankton growth rates, chl-a concentrations, and the trophic transfer efficiency within the planktonic food web. The presence of *Hemiaulus* in the mid-shelf region led to higher NPP rates during August 2019 compared to all other observed summers in the mid-shelf region of the NES (Fig. 7). High NPP rates associated with diatom blooms have been observed in other systems such as on the Eastern Bering Shelf (Lomas et al., 2012) and in the Gulf of California (Puigcorbe et al., 2015). In particular, diatom-diazotroph associations such as *Hemiaulus-Richelia* are known to significantly increase NPP (Gaysina et al., 2019). For example, Tang et al. (2020) reported a high contribution of nitrogen fixation to NPP off the coast of New Jersey during their 2015-2016 survey in the Western North Atlantic. Even though high NPP was associated with the location of the *Hemiaulus* bloom in our study, phytoplankton growth rates were low (<0.2 d^{-1}). This decoupling between NPP and growth was likely due to the order of magnitude higher chl-a concentrations observed during August 2019 (1.37 µg L^{-1}) compared to other summers (0.01 - 0.03 µg L^{-1}; Table 2) since NPP is roughly the product of phytoplankton growth and biomass (Marchetti et al. 2009). Thus, although growth rate was low, biomass was so high that NPP was also high. Furthermore, most of the primary production was directly consumed by microzooplankton, which we have not observed during any other summer NES-LTER cruise, suggesting the presence of *Hemiaulus* led to more efficient trophic transfer during August 2019. While conditions with high NCP (i.e. low community respiration) and high grazing pressure as observed in August 2019 may seem counterintuitive, they are not contradictory since grazing cannot be equated with respiration. First, much of respiration is bacterial and therefore not reflected by the grazing rates (Robinson and Williams, 2005). Second, it has been observed that after starvation, protozoan grazers increase their organic matter production by accumulating lipids and increasing their cell size (Anderson and Menden-Deuer, 2017, Morison et al., 2020). Thus, high grazing could suggest a buildup of organic matter through secondary production, which is consistent with the higher than average microzooplankton biomass and would be reflected as large NCP. Third, microzooplankton can produce fecal pellets (Buck and Newton, 1995), removing carbon from the system without respiration and thus lead to high NCP. The dominant presence, and slow growth, of large *Hemiaulus* cells within the phytoplankton community was likely a main factor promoting the higher trophic transfer efficiency from phytoplankton to microzooplankton, as is typical during winter (Marrec et al., 2021).

Interestingly, NCP and GOP values in summer 2018 were similar to those in August 2019 (Table 2, Fig. 7) and also much higher than during subsequent summers (2020-2022), in spite of no *Hemiaulus* being present in summer 2018. Additionally, the ratio of NCP/GOP in summer of 2018 was also significantly larger than in 2020-22 (Fig. 7). Remote sensing shows an elevated Chl-a patch (less concentrated than the patch in August 2019) in summer 2018 west of the transect that could be the driving factor behind the high NCP and GOP values (Fig. S1). The summer of 2018 was dominated by small phytoplankton similar to observations in summers of 2020 and 2021, although the summer of 2018 had a particularly high concentration of dinoflagellates over parts of the shelf. The summer 2018 data did not show an increase in trophic transfer efficiency due to coupled microzooplankton grazing and phytoplankton growth nor did it show increased NPP. The high NCP in summer 2018 could be due to a variety of environmental (biotic and abiotic) factors that were different from other cruises. For example, in the summer of 2018, saline waters from offshore intruded much farther inshore than during most of the other summers and these high-salinity mid-shelf waters were particularly productive (Mehta, 2022). Additionally, correspondence was seen between NCP and dinoflagellate biomass in summer 2018, although not as significantly as was seen with the *Hemiaulus* and NCP in 2019 (Aldrett, 2021). Thus, this study shows that a change in community composition,

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such as the *Hemiaulus* bloom in August 2019, can dramatically change the productivity rates of the ecosystem and yet productivity rates can also be similarly high with a very different phytoplankton community structure.

4.2 Aggregate vs Compositional Variability

The changes in community composition, productivity rates, and chlorophyll in August 2019 compared to the other summers shed interesting light on the question of synchrony or compensation between aggregate and compositional variability at the NES-LTER site (Micheli et al., 1999, Shoemaker et al., 2022). The resilience of an ecosystem may be related to the compensation or synchrony between different types of variability (Lindegren et al., 2016). During August 2019, the phytoplankton composition in the NES changed dramatically due to the bloom of the diatom *Hemiaulus*. This change was associated with increases in Chl-a, higher productivity rates, tighter coupling between microzooplankton grazing and phytoplankton growth, and increases in export efficiency. These latter terms are all metrics of aggregate properties and thus this bloom event exhibited high compositional and high aggregate variability compared to the ecosystem in July of 2020-22. Thus, during this event, a change in compositional variability (e.g., the change in phytoplankton community composition) was synchronous with a change in aggregate variability. However, when NCP rates are compared from summer 2018 to summer 2019, the compositional variability is still quite different (*Hemiaulus* in 2019, mostly small phytoplankton in 2018) but the aggregate variability in terms of NCP is the same in both years – thus comparing those two summers is a case where compositional variability changed and yet aggregate variability (at least for NCP) did not. Why does community composition sometimes affect NCP dramatically (i.e., August 2019 vs July of 2020-22) and sometimes it does not (July 2018)? Perhaps NCP is affected more by the abundance of certain species in the community rather than size distribution of the community. This concurrent investigation of plankton community composition and production rates within a well-studied ecosystem highlights how shifts in community size distribution can greatly affect productivity. However, it also shows that multiple factors change from year to year, leading to different effects.

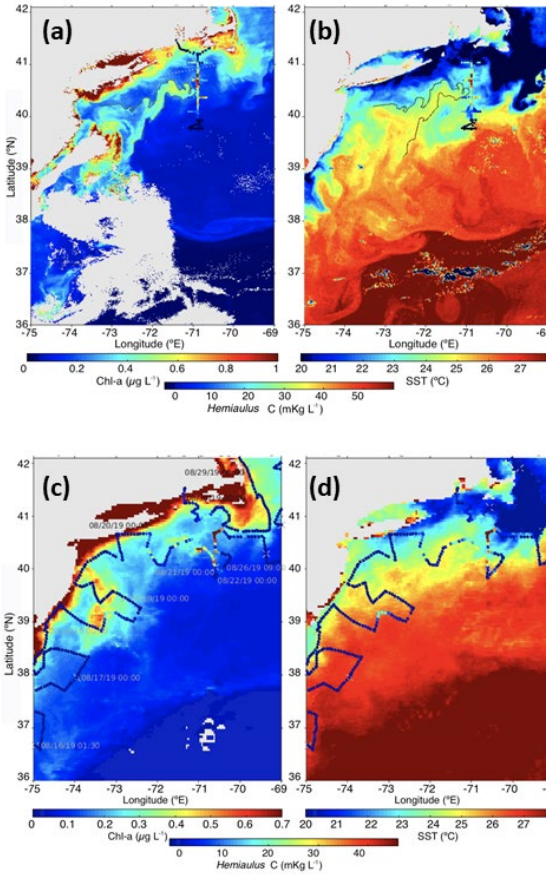

Fig 10. Snapshots of satellite-measured **(a)** Chl-a and **(b)** SST on 11 August 2019. The 0.5 μg L^{-1} chlorophyll contour is plotted as a solid line. *Hemiaulus* carbon from underway surface samples during the NES-LTER Aug 2019 cruise is overlaid with colored dots. The monthly composite **(c)** Chl a and **(d)** SST for August 2019. *Hemiaulus* carbon from underway surface samples during ECOMON cruise GU1902 is overlaid with colored dots. Daily ship positions are indicated in the left panel. Animated versions of individual Chl a and SST images are available at: http://science.whoi.edu/users/olga/outgoing/Aug_2019_chl/NEW_2019_Hemiaulus/

4.3 Origin of Bloom

The *Hemiaulus* bloom was likely more widespread than what was observed in the NES-LTER 2019 summer cruise. For

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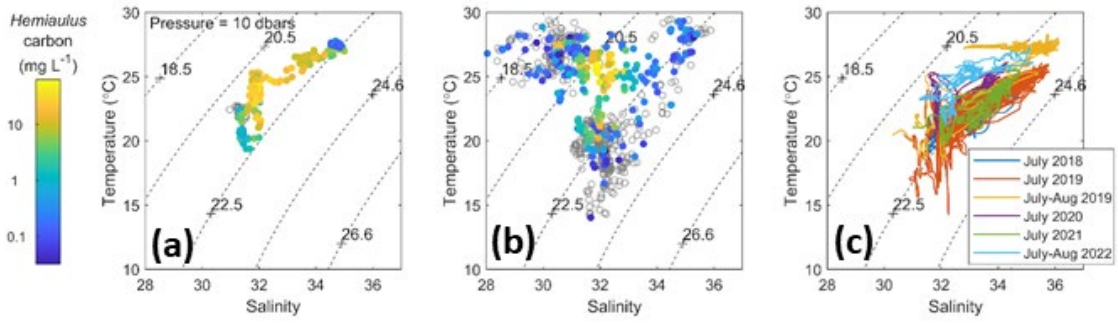

Fig 11. T-S plots, colored according to *Hemiaulus* carbon, suggesting that the highest *Hemiaulus* abundances were associated with a "sweet spot" in T-S space that was found during **(a)** the 2019 summer NES-LTER cruise in August and **(b)** the Ecomon August 2019 cruise. Empty circles represent locations where *Hemiaulus* was not detected. **(c)** TS plots from other years, colored according to cruise, show that a few of the other summer cruises from 2018 to 2022 have the same sweet spot in physical conditions even though they did not have detectable *Hemiaulus*.

example, satellite imagery from August 11 shows a filament of warm, high Chl-a waters oriented southwest-northeast and ending in the region where *Hemiaulus* was abundant (Fig. 10a & b); the advective continuity of the filament with the *Hemiaulus* patch suggests the filament may have had high *Hemiaulus* as well. Direct support for a widespread bloom comes from IFCB data collected on the NOAA EcoMon Cruise (GU1902) that occurred at a similar time as the August 2019 NES-LTER transect cruise. The IFCB data shows that *Hemiaulus* was present both farther east as well as to the southwest of where it was observed on the LTER transect cruise and that some of the points in the high chlorophyll filament observed from satellite chlorophyll contained *Hemiaulus* (Fig. 10c).

Backward particle trajectory analysis based on HF radar-measured sea surface velocities show that the water with high *Hemiaulus* biomass during the August 2019 transect cruise could have been advected from the inner-shelf around Narragansett Bay and Georges Bank rather than from the mid-shelf further south (Fig. S4). In particular, coastal upwelling probably brought the inner-shelf water into the mid-shelf transect area where it was observed to have high *Hemiaulus*. The salinity of the water with the high *Hemiaulus* biomass is consistent with the water having originated from the shelf. The water with high amounts of *Hemiaulus* carbon was associated with salinity ranging from 31.6 to 34 psu and temperatures of 22° C to 27° C (Fig. 11a & b). T-S plots of data from other years (Fig. 11c) suggest that several other summers also had similarly warm, low salinity water (in particular July of 2020 and especially 2022) but interestingly *Hemiaulus* were not observed on those cruises.

Although multiple lines of evidence suggest that the water containing the high biomass of *Hemiaulus* initially originated from the inner-shelf, *Hemiaulus* is typically found in warm, low nutrient water – characteristics that are not present on the inner-shelf, where water is instead colder and often richer in nutrients. In this case, the inner-shelf water warmed as it was transported offshore and thus it reached temperatures warm enough for *Hemiaulus* to thrive by the time it reached the mid-shelf (the timing of warming is not known). But how did this inner-shelf water acquire *Hemiaulus* as it was transported offshore in August 2019? One possibility is that it was seeded by the warmer low nutrient surface slope and ring waters; in particular, these slope and ring waters were observed earlier in the summer of 2019 to have a small population of *Hemiaulus* that could have served as a seed population (Oliver et al., 2021). However, there is no evidence of surface transport from slope-water to the *Hemiaulus* patch. Another possibility is that *Hemiaulus* were already present in the deeper coastal water and then thrived as the deep water was mixed upward, warmed and reached the higher light surface waters. However, the vertical distributions of *Hemiaulus* (Fig. 3) do not support this hypothesis, since a deeper population was not observed. Thus, the reason *Hemiaulus* bloomed in 2019, and not in other years, remains a topic for future research and continued speculation. Future years of the NES-LTER program may shed light on the variable effects of disturbances, such as this *Hemiaulus* bloom, as more factors that lead to high or low export in summer are determined and explored.

5 Conclusions

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An unusual bloom of the diatom genus *Hemiaulus* with nitrogen-fixing symbionts in the mid-shelf region of the Northeast U.S. shelf in August 2019 led to increases in NCP, GOP, NPP, higher export efficiency, and higher trophic transfer efficiency from phytoplankton to microzooplankton. Very tight coupling observed between kilometer-scale changes in NCP and the carbon biomass of *Hemiaulus* showed how substantial an effect the *Hemiaulus* bloom had on the aggregate variability of the Northeast U.S. shelf. While the source of the *Hemiaulus* on the inner-shelf remains unknown, the bloom was associated with warmer temperatures than usually observed on the shelf which may have been an important factor that facilitated the bloom when it was transported from the inner-shelf.

The *Hemiaulus* bloom, which was associated with warmer temperatures, was intriguing in that it led to unusually high productivity rates, increases in Chl-a concentrations, and tighter food-web coupling. While the warm SST may have contributed to the *Hemiaulus* bloom, the summer cruises of 2020 and 2022 also had anomalously high water temperatures, but relatively low (i.e., average summer) productivity rates and Chl-a were observed during those times. So, these summers of 2020 and 2022 had similar physical conditions to that of 2019, but no significant bloom was observed, and no high-carbon export system was present. Thus, higher temperatures are not enough to explain higher productivity rates, a shift in community composition is also necessary. A mixture of the right physical conditions and community composition, like this special case of 2019, are needed for a high-carbon export system to be supported on the mid-shelf during summer.

With climate change, the oceans are warming at a rapid rate, and are likely moving towards warmer more stratified conditions (e.g., lower nitrate stock in surface waters) (Li et al., 2020) which may lead to less productivity and thus lower export efficiencies. However, these conditions may also lead to unusual phytoplankton composition as species shift and the work presented here shows that those unusual events can lead to large locally and episodically enhanced productivity and export; despite the ecosystem commonly becoming nitrate-limited during the summer season, an intense phytoplankton bloom in summer occurred due to a symbiotic diatom-diazotroph relationships. These observations lead to further questions about how the NES ecosystem is responding to the effects of climate change such as enhanced stratification. Monitoring future disturbances and their effects will provide new insights into relationships, mechanisms, and patterns of composition and productivity that may be only occasionally occurring now but are likely more prevalent in the future.

6 Data Availability

All in situ data are available at the EDI data repository. In particular, the raw gas tracer data used for calculating NCP and GOP is available at https://portal.edirepository.org/nis/mapbrowse?packageid=knb-lter-nes.6.2. The calculated rates of NCP data is accessible at https://portal.edirepository.org/nis/mapbrowse?packageid=knb-lter-nes.7.2 and https://portal.edirepository.org/nis/mapbrowse?packageid=knb-lter-nes.15.2. NPP data is available at https://portal.edirepository.org/nis/metadataviewer?packageid=knb-lter-nes.16.4. Grazing rate data is available at https://portal.edirepository.org/nis/mapbrowse?packageid=knb-lter-nes.5.1. Chlorophyll data is available at https://portal.edirepository.org/nis/mapbrowse?packageid=knb-lter-nes.8.1. IFCB data is available at https://portal.edirepository.org/nis/mapbrowse?packageid=knb-lter-nes.9.1 and on the IFCB dashboard at https://ifcb-data.whoi.edu/timeline?dataset=NESLTER_transect.

The MODIS SST and chlorophyll snapshot data were produced by NASA Goddard Space Flight Center, Ocean Ecology Laboratory, Ocean Biology Processing Group, and the data are publicly available at https://oceancolor.gsfc.nasa.gov/. The 8-day composite data were retrieved from the public-accessible University of Delaware ERDDAP server (https://basin.ceoe.udel.edu//erddap/index.html) maintained by the Ocean Exploration, Remote Sensing and Biogeography Laboratory led by Dr. Matthew Oliver at University of Delaware. The HF radar-measured sea surface velocity data in July-August 2019 was obtained from the public-accessible Rutgers University Center for Ocean Observing Leadership ERDDAP server (http://hfr.marine.rutgers.edu/erddap/griddap/).

Author Contribution

SAC, RHRS, ZOS, and DA measured and calculated rates of productivity from gas tracers. SMD and PM measured grazing rates. TAR and DNF measured and calculated rates of net primary productivity from bottle incubations. HMS, ETC and EEP imaged and quantified phytoplankton abundances. DJM and WGZ analyzed remote sensing

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data. Everyone participated in study design. SAC and RHRS prepared the manuscript with contributions from all co-authors.

Competing Interests

The authors declare that they have no conflict of interest.

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