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Reconstructing the ocean's mesopelagic zone carbon budget: sensitivity and estimation of parameters associated with prokaryotic remineralization

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Abstract

Through the constant rain of sinking marine particles in the ocean, carbon (C) trapped within is exported into the water column and sequestered when reaching depths below the mesopelagic zone. Atmospheric CO_2 levels are thereby strongly related to the magnitude of carbon export fluxes in the mesopelagic zone. Sinking particles represent the main source of carbon and energy for mesopelagic organisms, attenuating the C export flux along the water column. Attempts to quantify the amount of C exported versus consumed by heterotrophic organisms have increased in recent decades. Yet, most of the conducted estimations have led to estimated C demands several times higher than the measured C export fluxes. The choice of parameters such as growth efficiencies or various conversion factors is known to greatly impact the resulting C budget. In parallel, field or experimental data are sorely lacking to obtain accurate values of these crucial overlooked parameters. In this study, we identify the most influential of these parameters and perform inversion of a mechanistic model. Further, we determine the optimal parameter values as the ones that best explain the observed prokaryotic respiration, the prokaryotic production, and the zooplankton respirations. The consistency of the resulting C-budget suggests that such budgets can be adequately balanced when using appropriate parameters.

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1. Introduction

The biological carbon pump (BCP) is the main mechanism by which CO_2 is exported and stored in the deep ocean in the long term. This ecosystem service is defined as the sum of the biological processes that lead to carbon export from the euphotic zone into the deep ocean (Eppley and Peterson 1979). This process exports from 5 to 20 Gt C yr^{-1} in the form of particulate organic carbon (POC) gravitationally sinking from the sunlit ocean to the mesopelagic zone roughly located between 200 and 1000 m (Henson et al. 2011). Therefore, atmospheric CO_2 levels are strongly related to any change in carbon export into the mesopelagic zone (Kwon et al. 2009). Five downward pathways of organic matter export to the mesopelagic zone are defined: through phytoplankton (senescent cells, colonies, spores, cysts), zooplankton (carcasses or fecal pellets), aggregates (marine snow of different compositions including the two latter categories), vertical migration of zooplankton and mixing/diffusion/advection (Siegel et al. 2016; Le Moigne 2019).

Gravitational sinking POC supply, known as the dominant pathway, constitutes the main organic carbon input to the mesopelagic zone (Boyd et al. 2019). Consequently, the downward flux of organic carbon is attenuated with increasing depth as it is fragmented, metabolized and remineralized by different biological processes until only the refractory material remains. The majority of POC flux attenuation occurs in the mesopelagic zone (Martin et al. 1987; Marsay et al. 2015; Fuchs et al. 2022). The remineralization of exported carbon is mainly performed by two types of organisms: micro-organisms (mostly heterotrophic prokaryotes i.e. Bacteria and Archaea) and zooplankton. Heterotrophic prokaryotes primarily use dissolved organic carbon (DOC) as a source of carbon. However, some prokaryotes, colonizing particles upon formation, undergo changes in environmental conditions during their descent, such as the increase of the hydrostatic pressure and the variations of temperature (Tamburini et al. 2003, 2021; Baumas et al. 2021). Such particle-attached prokaryotes primarily use POC as a carbon source. Only organic matter of size below 600 Da diffuses directly through prokaryotic membranes, therefore attached prokaryotes produce ectoenzymes required to solubilize larger molecules (Weiss et al. 1991). Smith et al. (1992) observed that the amount of DOC produced by ecto-enzymatic solubilization of POC may be 10 to 100 times greater than the absorption capacity of a cell. DOC is thereby released into the surrounding water (the so-called

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65 solubilization). This increases the amount of DOC available for free-living prokaryotes. In
66 addition, several types of zooplankton are involved in marine particles: POC-feeding
67 detritivores (e.g. copepods), prokaryotes consumers (e.g. flagellates), and carnivores (e.g.
68 chaetognaths). Besides, zooplankton lose POC through excretion (moult, mucilage, urine),
69 fecal pellets (decomposed organic matter), and sloppy feeding. Giering et al. (2014) specify
70 that 30% of a particle supplied by the downward flux is fragmented by the action of the
71 detritivores and is transformed into suspended matter.

72 Given their importance regarding the BCP, all the processes described above were extensively
73 studied in the last decades (e.g. Alldredge and Silver 1988; Smith et al. 1992; Kiørboe et al.
74 2002, 2003; Kiørboe 2003; Lampitt et al. 2008; Steinberg et al. 2008; Iversen et al. 2010;
75 Giering et al. 2014; Koski et al. 2020 and references therein). However, the scientific
76 community has struggled to reconcile the mesopelagic carbon budget with measurements and
77 estimates showing a biological carbon demand often greater than the amount of known organic
78 carbon sources (Reinthaler et al. 2006; Steinberg et al. 2008; Burd et al. 2010; Collins et al.
79 2015; Boyd et al. 2019). In other words, the measured export flux cannot sustain measured
80 metabolic demands of prokaryotes and zooplankton altogether in the mesopelagic zone, leading
81 to a discrepancy in C budgets.

82 A first explanation may lie in the choices of the boundaries of the mesopelagic zone used to
83 integrate fluxes and to estimate the carbon budget as investigated in Fuchs et al. (2022). Indeed
84 they specifically designed a method to determine from CTD-cast variables (fluorescence, O2
85 concentration, potential temperature, salinity, and density) accurate boundaries. With their
86 method named RUBALIZ, they show that 90% of the POC flux attenuation occurs within new
87 determined boundaries which is not the case of the fixed 200-1000m often used. Besides,
88 integrating prokaryotic C demand within RUBALIZ boundaries helps to reduce the
89 discrepancy. Other response elements may be found focusing on the carbon demand of
90 prokaryotes (which are responsible for the final step of the remineralization), whose estimation
91 is usually provided by adding rates of prokaryotic heterotrophic production (PHP) to that of
92 prokaryotic respiration (PR) (Burd et al. 2010). PHP rates are often measured from tritiated
93 leucine incorporation rates in incubations which are then multiplied by a conversion factor
94 Leu/Carbon (CF) (Kirchman et al. 1985). The PR is more challenging to measure (especially
95 in the dark ocean, (Nagata et al. 2010)) and, therefore, often estimated from measurements of
96 PHP and a prokaryotic growth efficiency (PGE) taken from the literature (as PR = PHP x (1-
97 PGE)/PGE, del Giorgio and Cole 1998). Unfortunately, *in-situ* measurements of both CF and

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PGE are time-consuming and operationally complex to perform (especially for the mesopelagic zone). In addition, such data for attached to sinking particles prokaryotic communities are scarce since the adequate sampling devices (to specifically sample biologically intact sinking particles) were only recently validated (Baumas et al. 2021). Besides, PHP and PR data are usually obtained after decompression or carried out from experiments at atmospheric pressure, being a source of misevaluation (Tamburini et al. 2013). As a result, values from the mean of global literature compilation or theoretical values are often used as references for both CF or PGE (Burd et al. 2010; Giering and Evans 2022) and may be far from the actual *in situ* values.

In parallel, model predictions help to estimate unmeasurable processes along with the comparison and validation of data. The biological processes occurring in the mesopelagic zone are not yet well constrained (see sections above). Consequently, only a few models specifically designed to assess the fluxes governing the BCP in the mesopelagic zone exist (e.g. Tian et al. 2000; Anderson and Ryabchenko 2009; Anderson and Tang 2010; Fennel et al. 2022). For instance, the model developed by Anderson and Tang (2010) enables the evaluation of the remineralization of different compartments such as attached prokaryotes to sinking and suspended particles, free-living prokaryotes and up to six trophic levels of zooplankton. This model describes the various known biological processes involved in the BCP system. However, the model also requires to be set up with parameters such as the PGE. For example, Anderson's model requires 24 parameters which often present large uncertainties.

Giering et al. (2014) attempted to reconcile carbon input and biological carbon demand in the mesopelagic zone using the Anderson and Tang (2010) model and measurements carried out in the North Atlantic (Porcupine Abyssal Plain site, 49.0°N 16.5°W, summer 2009). They found that prokaryotes were responsible for 70-92% of the remineralization of organic carbon. In this study, the model results were consistent with the measurements performed *in situ*, both showing a reconciliation of the carbon budget between 50 and 1000 m depths. Giering et al. (2014) balanced their C-budget by using a rather low CF (CF = 0.44 kg C mol^{-1}) compared to the one generally used in the literature (CF = 1.55 kg C mol^{-1}) and a PGE of 8% for free-living prokaryotes and 24% for prokaryotes attached to the particles. All these values were chosen as medians of literature values compiled from various measurement methods. Wisely choosing these parameter is therefore crucial to determine the reconciliation or the imbalance of carbon budget.

In this respect, we rely on model inversion methods (Tarantola 2005) to provide meaningful estimations of parameters of interest. For a given phenomenon, inversion methods rely on a

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model taking as input the parameters to be estimated and whose outputs can be compared with *in situ* measurements. The inversion procedure thus gives the value of the parameters that best replicate the *in situ* measurements. This type of procedure has already been used in oceanography modeling. For instance, Saint-Béat et al. (2018) studied phytoplankton marine food web in the Arctic and Saint-Béat et al. (2020) examined pelagic ecosystems of two different zones in the Arctic Baffin Bay using inversion method and sensitivity analyses to identify which biological processes impact the most the planktonic ecosystem functioning.

Here, we investigate the impact of overlooked but widely used parameters associated with the prokaryotic remineralization (e.g. CF, PGEs) on the magnitude of the discrepancy. Our aims are: 1) to highlight the most sensitive parameters for which the determination of an accurate value is critical in the context of balancing of mesopelagic carbon budget; 2) to perform a mathematical inversion method to estimate the most plausible *in situ* values of the most sensitive parameters from a limited field dataset; 3) to discuss our results in the context of mesopelagic carbon budget and carbon sequestration by the BCP.

2. Material & methods

2.1 *In situ* Data

Most of the data used in this study originated from the DY032 (June-July 2015) cruise at the PAP (Porcupine Abyssal Plain) site in the North Atlantic onboard the RRS Discovery. Some data unavailable for DY032 were estimated from a previous PAP cruise, D341 (July-August 2009). Most of the *in situ* data were compiled from already published cruise data (e.g. Giering et al. 2014; Belcher et al. 2016; Baumas et al. 2021; Fuchs et al. 2022). Their post-treatments to suit our study framework are described below. Additionally, we used data (ecto-enzymatic activities along with total hydrolysable amino acids and carbohydrates, depth profile of heterotrophic prokaryotic production and respiration under *in situ* pressure versus atmospheric) from the PEACETIME cruise (Guieu et al. 2020) that occurred in May 2017 in the Mediterranean Sea to illustrate some points in our discussions (see supp data).

2.1.1 Carbon fluxes

a) Determination of the Active Mesopelagic zone boundaries

Fuchs et al. (2022) introduced the “RUBALIZ” method, using CTD data, which allows the estimation of vertical boundaries targeting the zone of the dark ocean where most of the POC

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fluxes attenuation occurs. At station PAP during cruise DY032, this so-called "Active Mesopelagic Zone" was located between 127 and 751 m.

b) Carbon inputs

The POC inputs to the active mesopelagic zone mainly involve the gravitational export of POC. Gravitational input was taken from Fuchs et al. (2022) who fitted a power law Martin curve (b of 0.84) on data obtained from 30 to 500m using Marine Snow Catcher (Belcher et al. 2016). However, gravitational input is not the only POC input known in the literature. Recently, Boyd et al. (2019), provided an estimation of other particle-injection pumps (PIPs) such as the mixed layer pump, physical pump, the seasonal lipid pump or the active transport related to metazoans migrations. At the PAP site during summer, only the eddy subduction pump, metazoans migrations, and large-scale physical pumps were relevant to take into account. Other PIPs do not correspond to the location and season considered in our study. From Boyd et al. (2019) review, these three particle-injection pumps seem to represent altogether around 52% of the gravitational export of POC. We therefore add up this proportion of POC to the purely gravitational inputs. This yields an overall POC flux of 134 mg C m^{-2} d^{-1} exported into the active mesopelagic zone. The corresponding net POC input is 117 mg C m^{-2} d^{-1} (that is POC fluxes at the end - 751 m - of the active mesopelagic zone subtracted to the one at the start - 127 m - for PAP DY032).

DOC inputs are taken from Giering et al. (2014) and are considered as the sum of direct DOC export via physical processes (advection-diffusion) and active flux from zooplankton migrations. We estimated from their extended Data Fig. 2 that the DOC gradient below 100m is hardly visible meaning that physical vertical DOC export is insignificant for the active mesopelagic zone which is studied here. As a result, we set the DOC export at 3 mg C m^{-2} d^{-1}, which corresponds only to the active flux from zooplankton migrations from Giering et al. (2014).

c) Carbon demands

As explained above, prokaryotic carbon demand is generally assessed by adding rates of prokaryotic heterotrophic production (PHP) to that of prokaryotic respiration (PR). PHP of non-sinking prokaryotes (that is free-living and attached to suspended particles prokaryotes) are derived from leucine incorporation measurements on seawater samples and are taken from Fuchs et al. (2022). These data did not permit the separation of the free-living from attached to suspended particles (Baumas et al. 2021). Hence, in the sequel, we no longer make this

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distinction and group both types under the term "non-sinking prokaryotes". During DY032, Marine Snow Catchers (MSC) were deployed to separate slow and fast-sinking particles from 100L of samples (Riley et al. 2012; Baumas et al. 2021). PHP rates associated with prokaryotic communities of fast-sinking particles were taken from Baumas et al. (2021) and slow-sinking particles are presented here. Briefly, slow-sinking particle fractions were sampled in the 7L base of the MSC. Samples were incubated and leucine incorporation rates were measured as for fast-sinking particles in Baumas et al. (2021). The formula described in Baumas et al. (2021) was then applied to normalize to 100L as particles were concentrated in 7L after 2h of decantation and to remove the contribution of non-sinking prokaryotes which were primarily in this compartment around slow-sinking particles of interest. Total sinking prokaryotes PHP rates were obtained by adding both fast-sinking and slow-sinking prokaryotes PHP rates. In addition, we were able to use the respiration rates of prokaryotes attached to fast-sinking particles obtained by Belcher et al. (2016). For each depth (30-500m) the mean total O_2 consumption per particle in nmol $agg^{-1}d^{-1}$ was converted to mg C m^{-3} d^{-1} (assuming a respiration quotient $RQ(CO_2/O_2) = 1$) by multiplying by the total number of particles (i.e. fecal pellets + phytoplanktonic aggregates) and dividing by 95L which is the volume of the MSC used (Riley et al. 2012). It is also important to note that PR for slow-sinking particles is missing. Thus, when we mention the respiration of sinking prokaryotes, only attached to fast-sinking prokaryotes are taken into account which certainly underestimates the respiration used. All prokaryotic carbon demand (PHPs and PRs) estimates were integrated within RUBALIZ boundaries (i.e. 127m - 751m). Non-sinking prokaryotes PHP rates were integrated using a piecewise model with a single node on the log-data as described in Fuchs et al. (2022). Sinking prokaryotes PHP rates were integrated using power law. Sinking PR were integrated using trapeze because data are only available until 500m and without any *a priori* on the curve shape, this method is certainly the most conservative.

Zooplankton activities are known to be related to POC concentration (Steinberg et al. 2008). Zooplankton respiration data were available only for the cruise D341 when the net POC input into the active mesopelagic layer was 59 mg C m^{-2} d^{-1} (including PIPs) instead of 134 mg C m^{-2} d^{-1} for DY032 (see above). For D341, zooplankton respiration integrated within the active mesopelagic zone (135-726m, Fuchs et al. 2022) was 9 mg C m^{-2} d^{-1}. Zooplankton respiration was integrated using a power law as in Giering et al. (2014). Zooplankton respiration data are missing for DY032, thus we consider this quantity as a percentage of the POC input that we calculate from the D341 data set, i.e. 14.67%. The zooplankton respiration value used here is therefore 17 mg C m^{-2} d^{-1}.

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Table 1: Fluxes and their associated values used in this study. Anderson & Tang model's terms (Anderson and Tang 2010) corresponding to these fluxes are also shown. Values are integrated between 127 and 751m which are boundaries of the active mesopelagic zone defined by Fuchs et al. (2022). *POC and DOC refer respectively to Particulate and Dissolved Organic Carbon, PHP to Prokaryotic Heterotrophic Production, and PR to Prokaryotic Respiration.*

Name	Anderson and Tang's Model term correspondence	Values	units	sources
Net POC input	$D1ex$	117	mg C m^{-2} d^{-1}	Belcher et al. (2016); Boyd et al. (2019)
DOC input	$DOCex$	3	mg C m^{-2} d^{-1}	Giering et al. (2014)
Non-sinking prokaryotes PHP	$F_{BFL}+ F_{BAD2}$	1.10E+07	pmol Leu m^{-2} d^{-1}	Baumas et al. (2021)
Sinking prokaryotes PHP	F_{BAD1}	1.02E+06	pmol Leu m^{-2} d^{-1}	Baumas et al. (2021)
Sinking prokaryotes PR	R_{BAD1}	19	mg C m^{-2} d^{-1}	Adapted from Belcher et al. (2016)
Zooplankton respiration	$R_{VA}+R_{VFL}+R_H+R_{Z1:6}$	17	mg C m^{-2} d^{-1}	Adapted from Giering et al. (2014)

2.2 Mathematical methods

2.2.1 Parameter estimation

The scope of our study is to estimate *in situ* parameters by inverting the model introduced by Anderson and Tang (2010), adapted by Giering et al. (2014). We do not intend to present the model in details here. The details of the equations constituting the version of the model used can be found in the original paper (Anderson and Tang 2010), in the R code available at https://github.com/RobeeF/InverseCarbonBudgetEstim and the specific terms related to variables used are reported in Table 1. The model is calibrated by choosing the set of input

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parameters that yields the best fit between the model output and the data. As the model outputs 85 outfluxes, we used a subset of four measurable outfluxes to calibrate the model: the PHP of non-sinking prokaryotes, the PHP of sinking prokaryotes, the PR of sinking prokaryotes and the respiration of zooplankton. These fluxes have been chosen because of their near direct correspondence with outputs of the model linked to the C demand of all groups (sinking prokaryotes, non-sinking prokaryotes, detritivores, bacterivores, and carnivores).

Similarly, the model relies on 24 input parameters (Table S1), which makes the parameter space of significant size and therefore challenging to explore. As such, we first determine the set of parameters that have the largest impact on the output of the model. Then for these parameters, the values that give the best fit between the data and the solution given by the model are determined.

a) Sensitivity of the model to its inputs

In order to reduce the size of the input parameter space, Sobol Indices (Sobol 1993) were used to determine the most influential parameters. These indices enable quantification of the share of the variation of the output that can be imputed to each input parameter.

In essence, the first-order Sobol indices account for the direct influence of an input variable on the output. However, first-order Sobol indices neglect the interactions existing between this input variable and the other input variables. As such, in addition to the first-order Sobol Indices, we used the total Sobol indices introduced by Homma and Saltelli (1996) which encompass both the direct effect of a parameter and also its interactions with the other parameters.

First-order and total Sobol indices were computed to quantify the influence of each parameter over each of the four outfluxes. Only the parameters which had significant Sobol indices (i.e. Sobol indices > 0.20) for at least one outflux were kept.

b) Estimation of the parameters

The parameters which had no substantial effects on the output of the model were set to the values indicated by Anderson and Tang (2010) and Giering et al. (2014) and given in Appendix (Table S1). The other parameters were estimated by minimizing the distance existing between the four outfluxes predicted by the model and their *in situ* measured counterpart. The distance chosen here is a standardized Euclidean distance:

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$$\sum_{i=1}^{4}\left(\frac{outflux_{obs,i}-outflux_{model,i}}{outflux_{obs,i}}\right)^2 \quad (1)$$

where $outflux_{obs,i}$ is the i-th measured flux and $outflux_{model,i}$ its modeled counterpart. The optimization method used is the Nelder-Mead algorithm (Nelder and Mead 1965): if the function to minimize depends on N variables (the number of input parameters here), a simplex constituted by N + 1 points is defined. The coordinates of the simplex are updated in turn so that the simplex vertices get closer to the local minimum. Even if this method gives little theoretical guarantees of convergence, it has proven to work well in practice (Lagarias et al. 1998) and has the advantage that it does not require computing the gradient of each outflux with respect to each input parameter.

As the model takes 24 inputs and outputs 85 fluxes, concerns might be raised about the uniqueness of the solution found to minimize the term (1). To make the model identifiable, the number of input parameters to estimate is limited to the number of output fluxes available, here four. In this respect, the CFs have been fixed to 0.5 kg C mol Leu^{-1}. This value, contrary to the previously classically used value of 1.55 kg C mol Leu^{-1} (Simon and Azam 1989; Nagata et al. 2010), was determined by Giering and Evans (2022) as the median value of 15 studies conducted in the mesopelagic zone. Doing so, we limit the number of free parameters to be estimated to four so that the model remains identifiable. The model is mostly linear and our experiments have shown the solution to be unique and independent of the initial values taken.

The codes and data to reproduce the results are available at https://github.com/RobeeF/InverseCarbonBudgetEstim

3.Results

3.1 Most sensitive parameters

Using Sobol indices, we identified the most sensitive parameters from the 24 of the Anderson and Tang (2010) model on the 4 fluxes outputs of the model for which we have the measured counterpart (i.e. PHP and PR of sinking prokaryotes, PHP of non-sinking prokaryotes and respiration of zooplankton). All parameter definitions are given in Table S1. For the outflux "PHP of non-sinking prokaryotes", only the $PGE_{non\text{-}sinking}$ appears to be sensitive with a Sobol index of 0.68 meaning that it explains 68% of the variance (Table 2). Fluxes related to sinking prokaryotes, i.e. their PHP and their PR, appear to be highly influenced both by Ψ, $\boldsymbol{\alpha}$, and

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$PGE_{sinking}$ with indices of 0.22 and 0.23 for Ψ, 0.24 and 0.24 for **α** and 0.27, 0.25 for $PGE_{sinking}$ respectively. Surprisingly, zooplankton respiration is more impacted by the $PGE_{non\text{-}sinking}$ (Sobol index of 0.52) than proper zooplankton parameters. All other parameters exhibit Sobol indices below 1%. Total Sobol indices, indicating the part of the variance of fluxes due to the parameter alone and in interaction with the others, were similar to the first-order indices, suggesting no interactions of parameters regarding the variance of fluxes. This sensitivity analysis enabled the identification of Ψ, **α**, and both PGEs as the most influential parameters, suggesting that their values should be set with particular care. Especially for the $PGE_{non\text{-}sinking}$ which can be responsible for more than 50% of the variance of $PHP_{non\text{-}sinking}$ and zooplankton respiration. PGEs are growth efficiencies defined as the amount of new prokaryotic biomass produced per unit of organic C substrate assimilated and is a way to relate PHP and PR (del Giorgio and Cole 1998). Ψ corresponds to the percentage of POC consumed by prokaryotes and **α** to the fraction of hydrolyzed POC which is lost into the surrounding water, i.e. not assimilated by sinking prokaryotes that hydrolyzed it.

Table 2: First-order Sobol indices for the parameters of the model by Anderson and Tang (2010). The definition of each parameter can be found in Table S1. Significant Sobol indices (>0.2) are shown in red. PHP and PR respectively refer to Prokaryotic Heterotrophic Production and to Prokaryotic Respiration.

	ψ	PGE sinking	PGE non-sinking	α	Φv	βv	Kv	Φv	βv	Kv	Φz	βz	λz	Kz	Φh	βh	λh	Kh	ζ	ζ2
Non-sinking prokaryotes PHP	<0.01	0.021	0.681	0.01	<0.01	<0.01	<0.01	0.014	<0.01	0.011	-0.012	<0.01	<0.01	0.015	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
Sinking prokaryotes PHP	0.222	0.24	<0.01	0.265	<0.01	<0.01	<0.01	<0.01	0.011	<0.01	<0.01	<0.01	<0.01	<0.01	0.011	<0.01	<0.01	-0.012	<0.01	-0.011
Sinking prokaryotes PR	0.225	0.243	<0.01	0.252	-0.019	<0.01	<0.01	<0.01	-0.011	<0.01	<0.01	<0.01	0.012	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
Zooplankton respiration	<0.01	0.023	0.507	<0.01	<0.01	0.014	<0.01	0.064	0.027	0.041	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	0.028	<0.01	<0.01	<0.01

3.2 Model inversion

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The optimization method, described in the material and method section, enabled the determination of the 4 parameters identified as sensitive above: Ψ, **α**, $PGE_{sinking}$, and $PGE_{non-sinking}$ in the case study of PAP DY032. Table 3 reports the combination found by model inversion. By construction of the procedure (e.g. same number of input and output), the solution is unique, explaining why no confidence intervals are reported. The errors between the four fluxes generated by the model and their measured counterparts were less than 1%, far lower than potential measurement errors. The zooplankton flux was the best matched, followed by the PR of the sinking prokaryotes, the PHP of the non-sinking prokaryotes, and of the sinking prokaryotes.

*Table 3: Estimation of the parameters Ψ, **α**, $PGE_{sinking}$ and $PGE_{non-sinking}$ obtained by inversion of the model by Anderson and Tang (2010). As the model was made identifiable, the solutions are unique, explaining the absence of confidence intervals. The remaining differences between the model outfluxes deriving from the estimated input values and the actual in situ measurements are referred to as "Errors" and are expressed in percentage. PHP, PR, and ZR respectively stand for Prokaryotic Heterotrophic Production, to Prokaryotic Respiration and to Zooplankton Respiration.*

Estimations				**Errors**			
Ψ	**α**	**$PGE_{sinking}$**	**$PGE_{non-sinking}$**	**$PHP_{non-sinking}$**	**$PHP_{sinking}$**	**$PR_{sinking}$**	**ZR**
0.675	0.777	0.026	0.087	-0.487%	0.524%	0.184%	-0.05%

3.3 C budget

The two PGEs presented above along with CF of 0.5 kg C mol Leu^{-1} were applied to leucine-incorporation rates measurements to build the corresponding active mesopelagic C budget. The resulting C budget was compared with two other C budgets calculated with different sets of parameters. The three active mesopelagic zone C budgets resulting from DY032 measurements or estimation are represented in Fig. 1 with the budget (1) obtained with the classical CF value of 1.55 kg C mol Leu^{-1} and median literature values for PGEs, i.e. 0.07 for $PGE_{non-sinking}$ (Arístegui et al. 2005; Reinthaler et al. 2006; Baltar et al. 2010; Collins et al. 2015) and 0.02 for $PGE_{sinking}$ (Collins et al. 2015); the budget (2) obtained with the parameter values from Giering et al. (2014) who reconcile C budget, i.e. CF of 0.44 kg C mol Leu^{-1}, $PGE_{non-sinking}$ of

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0.07, and $PGE_{sinking}$ of 0.24 and the budget (3) obtained with a CF, $PGE_{sinking}$ and $PGE_{non\text{-}sinking}$ of 0.5, 0.026 and 0.087, respectively, determined in this study. The combination yielding to the largest discrepancy is the budget (1) (Fig. 1) (discrepancy of -194 mg C m^{-2} d^{-1}). The C input seems to support the zooplankton respiration and total C demand of sinking prokaryotes but not the one of non-sinking prokaryotes especially due to their PR of 218 mg C m^{-2} d^{-1}. Combination of budget (2) and (3) presented both an excess of C (60 and 40 mg C m^{-2} d^{-1} respectively) compared to the biological C demand. These two differ mainly on the PR of sinking prokaryotes which is negligible in combination (2) but which is the second largest flux in the C demand in our study. In all cases, the C demand of non-sinking prokaryotes accounts for most of the total C demand.

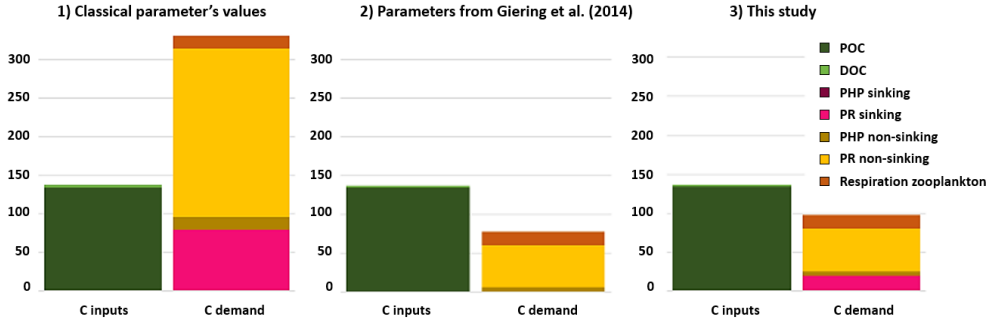

Figure 1: Carbon budget for the active mesopelagic zone estimation resulting from DY032 measurements or estimation and on which different combination of CF (1.55, 0.44 and 0.5 respectively for budget 1) 2) and 3)) , $PGE_{sinking}$ (0.02, 0.24 and 0.026 respectively for budget 1) 2) and 3)) and $PGE_{non\text{-}sinking}$ (0.07, 0.08 and 0.087 respectively for budget 1) 2) and 3)) were applied on leucine incorporation rates of sinking and non-sinking prokaryotes. See Fig. S1 for value details.

4.Discussion:

As stated in the introduction, the scientific community has struggled to reconcile the mesopelagic carbon budget with measurements and estimates showing a carbon demand often greater than the amount of known organic C sources (e.g. Reinthaler et al. 2006; Steinberg et al. 2008; Burd et al. 2010; Collins et al. 2015; Boyd et al. 2019). Building C budget involves a plethora of parameters whose impacts are overlooked and often neglected, mainly because neither their ideal values nor their underlying mechanism in the water column across space and time are clearly understood. The scientific community is concerned about this issue (e.g. Burd et al. 2010; Giering and Evans 2022), but in the absence of a better option and in an attempt to encourage comparisons, the same parameter values are universally used. A first step towards

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this direction was conducted thanks to the RUBALIZ method (Fuchs et al. 2022) which precisely determines the vertical location of the "active mesopelagic zone" and thereby estimates the boundaries between which to integrate C fluxes. In the current study, we pursue this investigation and combine measurements with modeling approaches to investigate the role of sensitive parameters related to the remineralization of POC in the mesopelagic zone.

4.1 Optimization method: Consistency of parameters estimated

The Anderson and Tang (2010) model takes as inputs the measured C inputs as well as 24 parameters related to the activity of organisms such as sinking prokaryotes, non-sinking prokaryotes, zooplankton detritivores, bacterivores, and carnivores. Among the 24 parameters, four have been found to be particularly sensitive in assessing the carbon demands of the various groups: Ψ (percentage of particle consumption by prokaryotes), $\boldsymbol{\alpha}$ (percentage of C hydrolyzed released in surrounding water), $PGE_{non\text{-}sinking}$ and $PGE_{sinking}$ (growth efficiencies of sinking and non-sinking prokaryotes). It is interesting to note that zooplankton respiration (which is the sum of detritivores, bacterivores and carnivores respiration) is mostly sensitive to one parameter: $PGE_{non\text{-}sinking}$ but not to a parameter specific to zooplankton. This counter-intuitive result suggests a strong synergy between the two model compartments. At this point, it is challenging to establish whether this is the outcome of a complex ecological process or a model artifact.

In the model, the consumption of particles is done by two groups: prokaryotes (Ψ) and detritivores (1-Ψ). It can be estimated by taking the average ratio between PHP and ZR. Anderson and Ryabchenko (2009) estimated Ψ using calculations of POC consumption by prokaryotes and zooplanktons between 150 and 1000m performed by Steinberg et al. (2008) in the Pacific. Following this, they set Ψ at 0.76. The inversion of the Anderson and Tang model (2010) leads to a well-identified solution of Ψ, i.e. 0.67 in the case of PAP DY032 cruise. This value is in line with the one used by Anderson and Tang (2010). However, data are lacking to compare and explore variations of Ψ value across seasons, locations or depths. In the model, Ψ participates in the repartition of POC input between prokaryotes and detritivores. Whether for modeling purposes to determine Ψ or to build a C-budget without a model, PHP and ZR are required. It remains too rare to have both together and more future efforts should be devoted to get PHP and ZR concomitantly.

Beyond Ψ, according to Sobol indices, $\boldsymbol{\alpha}$ is the second parameter of interest. Amino acids and sugar are major components of POC, constituting between 40 to 70% of POC in the

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411 mesopelagic zone (Wakeham et al. 1997). When prokaryotes consume POC using hydrolytic
412 enzymes, a major fraction of the hydrolyzed C is lost to the surrounding environment as DOC
413 (Smith et al. 1992; Vetter et al. 1998). This loss is represented by **α** and is very difficult to
414 quantify accurately. Two major experiments, focused on amino acid hydrolysis, aimed to
415 determine such losses: Smith et al. (1992) and Grossart and Ploug (2001). Smith et al. (1992)
416 sampled particles at 25m and showed that 97% of particulate combined amino acids are
417 released in the surrounding water. Later, Grossart and Ploug (2001) using aggregates from
418 phytoplankton cultures show a loss of POC of 74%. Relying on these two studies, Anderson
419 and Tang (2010) followed by Giering et al. (2014) consider that the value should be lower than
420 that of a fresh detritus and choose a conservative value of 0.5. In the case of these two
421 experiments, only the amino acids are considered and the experiments were conducted under
422 laboratory-controlled settings. Conversely, we used unpublished data from PEACETIME
423 cruise (see methods details in supp. data) of *in situ* hydrolysis rates of aminopeptidase and β-
424 glucosydase from sinking prokaryotes (which hydrolyze amino acids and sugar, respectively)
425 that we were able to convert into hydrolyzed carbon fluxes (see measurements and calculation
426 details in supp. data). Unfortunately, total hydrolyzed C fluxes were most of the time below
427 the C demand of the sinking prokaryotes which is unrealistic and probably due to the low
428 amount of POC (sinking POC concentration of <1 mg L^{-1} in the sinking fraction) resulting in
429 insufficient sinking prokaryotes abundance to detect their activity by volume. However, when
430 total hydrolyzed C fluxes were superior to $PHP_{sinking}$ (indicating that some hydrolyzed C is not
431 assimilated and is released), **α** was estimated between 0.19 and 0.79 with a mean of 0.41±0.24
432 and seems to decrease with depth (see calculations details in supp data). This could confirm
433 Grossart and Ploug's (2001) work showing that the older a detritus is, the less enzymatic
434 activity there is and therefore the less amino acid loss. Even if **α** is not measurable easily, this
435 parameter is identified at 0.78 by the inversion method during a post-bloom period at the PAP
436 site. This value is consistent with Smith et al. (1992) and Grossart and Ploug (2001) evidencing
437 high **α** for surface aggregates (0.97) with laboratory-made phytoplankton aggregates (0.74), or
438 with our calculations for the Mediterranean Sea (0.41±0.24), an oligotrophic region. This
439 suggests that the optimization method is a relevant alternative to determine **α**. In addition, **α**
440 corresponds to a release of C in the surrounding water. Regarding the model, the C demand of
441 free-living prokaryotes matches the hydrolyzed C released which constitutes their main C
442 sources. The relationship between enzymatic activities and heterotrophic production of free-
443 living prokaryotes is well documented in the deep-sea ocean (Cho and Azam 1988; Smith et
444 al. 1992; Hoppe and Ullrich 1999; Tamburini et al. 2002, 2003; Nagata et al. 2010). Total C
445 demand of non-sinking prokaryotes is challenging to measure due to the diversity of existing

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446 methods, especially the PR (e.g. Table S2), which leads to an incredibly wide range of
447 estimated values. Subsequently, identifying **α** via the optimization method could help to avoid
448 these conflicting PR measurements.
449
450 The last two sensitive parameters according to Sobol indices were $PGE_{non\text{-}sinking}$ and $PGE_{sinking}$.
451 A wide range of $PGE_{non\text{-}sinking}$ has been estimated using $PHP_{non\text{-}sinking}$ and $PR_{non\text{-}sinking}$ in the open
452 ocean (e.g. Sherry et al. 1999; Lemée et al. 2002; Carlson et al. 2004; Arístegui et al. 2005;
453 Reinthaler et al. 2006; Baltar et al. 2009, 2010; Collins et al. 2015). Overall it varies from 0.001
454 to 0.64 (Collins et al. (2015) and Sherry et al. (1999), respectively). However, these values
455 were produced from different protocols for the PHP (changes in biomass, thymidine or leucine
456 incorporation, each with its own conversion factors and/or constants) and for the PR methods
457 (by ETS measurements, micro-winkler titration, changes in dissolved O_2, or using optodes
458 sensors spots, see Table S2) and correspond to various locations, seasons and depths. These are
459 all valid reasons that can potentially explain the stark contrast in the values reported. If one
460 focuses only on the mesopelagic zone in the North Atlantic, the median is 0.07 (Arístegui et al.
461 2005; Reinthaler et al. 2006; Baltar et al. 2010; Collins et al. 2015). The optimization method
462 yielded to a value of 0.087 and therefore produces very consistent results for a post-bloom
463 period at the PAP site. Concerning $PGE_{sinking}$, too few values are available. To our knowledge,
464 only Collins et al. (2015) provided *in situ* values associated with sinking prokaryotes (from
465 0.01 to 0.03) at 150m. This is the only comparison we have, and our value of 0.026 matches
466 this order of magnitude. To provide further comparison, the DY032 data before integration
467 allows us to calculate a $PGE_{sinking}$ (using $PGE_{sinking} = PHP_{sinking}/(PHP_{sinking}+PR_{sinking})$ from del
468 Giorgio and Cole 1998) per $PR_{sinking}$ and $PHP_{sinking}$ of sinking prokaryotes points performed at
469 the same depth. This led to a variation from 0.033 at 70m to 0.0013 at 500m. Although the lack
470 of datapoints deeper than 500m and the low number of points forces us to stay cautious about
471 these estimates, it may indicate that $PGE_{sinking}$ is not constant throughout the mesopelagic zone
472 and decreases with depth. Constraining conditions due to the increase of hydrostatic pressure
473 and decrease in temperature experienced by prokaryotes attached to sinking particles could
474 explain this decrease in $PGE_{sinking}$ (Stief et al. 2021; Tamburini et al. 2021). Under highly
475 constrained conditions, Russell and Cook (1995) explained that maintaining respiration at the
476 highest possible rate would allow the supply of active membrane transporters which are vital
477 to the cell. This implies a low but optimal PGE (Westerhoff et al. 1983) which could thus
478 decrease with depth and time as the POC becomes less labile (Grossart and Ploug 2000). On
479 the contrary, the Anderson and Tang (2010) model, and the associated model inversion
480 presented here, is built so that the mesopelagic zone is considered as one homogeneous entity.

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Explicitly, specifying depth-dependent $PGE_{sinking}$ in the mesopelagic zone could lead to more realistic modeling, but would entail a non-negligible additional model complexity.

It is worth noting that the $PGE_{sinking}$ and $PGE_{non\text{-}sinking}$ estimated here rely on a leucine-to-carbon Conversion Factor (CF) of 0.5 kg C mol Leu^{-1}. This value comes from the median of 15 values obtained on the free-living prokaryotes of the mesopelagic zone (between 300 to 1000m), which do not sink and are adapted to their place in the water column (Giering and Evans 2022). However, to our knowledge, there are no such values measured for the specific case of sinking prokaryotes. The latter are surface prokaryotes that have attached to the particles and will experience changes in conditions (e.g. pressure, temperature) linked to their sink (Baumas et al. 2021; Tamburini et al. 2021). The CF depends, among other things, on the leucine fraction in the proteins and the cellular carbon/protein ratio (Kirchman and Ducklow 1993). It is known that stresses can affect the incorporation of leucine into proteins and general protein production (e.g. Young 1968; Welch et al. 1993) and that these parameters can vary with prokaryotic diversity, especially between bacteria and archaea (Bogatyreva et al. 2006). Stresses occur during the descent throughout the water column and sinking prokaryotes experienced a drastic decrease in diversity following the sink at PAP DY032 (Baumas et al. 2021; Tamburini et al. 2021). We can therefore easily imagine that the CF for sinking prokaryotes could be impacted. Despite this, without having further data, we applied the same CF on sinking as the 0.5 recommended by Giering and Evans (2022) for non-sinking prokaryotes and the results were consistent.

4.2 Influence on mesopelagic C Budget

As stated in the introduction, mesopelagic C budgets are constructed by applying a CF and a PGE on leucine incorporation rates data to assess prokaryotic C demand. In Fig. 1, we applied three different combinations of CFs and PGEs to the same data. The combination using conventional CF of 1.55 kg C mol Leu^{-1}, $PGE_{non\text{-}sinking}$ of 0.07, and $PGE_{sinking}$ of 0.02 led to an aberrant discrepancy such that more than the entire C pool would be remineralized in the active mesopelagic zone and that there would be no source of C to sustain deeper zone life nor sequestration by the BCP. As stated above, this was a recurrent issue in the field (Reinthaler et al. 2006; Steinberg et al. 2008; Burd et al. 2010; Collins et al. 2015; Boyd et al. 2019) with the exception of Giering et al. (2014) who reconcile the C budget of the mesopelagic zone. Although Giering et al. (2014) did not take into account sinking prokaryotes from *in situ* data point of view, their results were mainly due to the difference in CF used, i.e. 0.44 kg C mol Leu^{-}

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[1]. However, from a model point of view, the main difference between C budgets estimated using Giering et al. (2014) parameters and those determined by our optimization method is due to the 10-fold difference between $PGE_{sinking}$ used. Giering et al. (2014) used 0.24 which is the mean of a 14 days incubation experiment during which PGE varied from 0.45 in the first 3 days to 0.04 at the end for riverine aggregates (Grossart and Ploug 2000). Despite the fact that $PGE_{sinking}$ data are very scarce, riverine values of 0.24 seem highly unlikely and inappropriate to mesopelagic sinking prokaryotes compared to what is known in marine environments (e.g. Collins et al. 2015). Indeed, if we consider that enzymes account for a large proportion of the proteins produced by cells (see above) the $PGE_{sinking}$ must be low due to the high metabolic cost of their production (Grossart and Ploug 2000). Finally, the C budget built from a combination of CFs of 0.5 kg C mol Leu^{-1} and PGEs revealed by our optimization method seems the most reasonable option (from the three budgets built, Fig. 1) with an excess of C input of 40 mg C m^{-2} d^{-1}. In this case, PGEs were determined by the model, which in addition to PHP and PR of sinking and non-sinking prokaryotes and zooplankton respiration, also accounts for the production of zooplankton biomass into calculations. We do not have measurements or estimates for the production of zooplankton biomass but based on the model, this biomass production is 11 mg C m^{-2} d^{-1}. Adding this value to the C demand implies a leftover of 29 mg C m^{-2} d^{-1} that is not used and is exported under the active mesopelagic zone via gravitational sinking POC. This value is in accordance with the POC flux estimated from measures at 751m (thus at the exit of our zone): 17 mg C m^{-2} d^{-1}. Being aware of the biases that may exist in the fluxes used as well as in the construction of the model itself, our optimization method enables the determination of realistic values of parameters and thus constructing robust C budgets. As far as we know, the combination of field measurements (using consistently defined integration depths, such as RUBALIZ (Fuchs et al. 2022) with the use of optimization method on the Anderson & Tang model has led to the most complete and realistic mesopelagic carbon budget.

4.3 Model: reliability and potential biases

The Anderson and Tang model (Anderson and Tang 2010) was originally parametrized with 24 input parameters and 85 output fluxes, and is hence by definition an underdetermined model as the number of outputs is higher than the number of inputs. To make the model identifiable, i.e. obtaining unique solutions for each parameter value, the number of parameters allowed to vary, namely: Ψ, $\boldsymbol{\alpha}$, $PGE_{non\text{-}sinking}$, and $PGE_{sinking}$, was restricted to the number of measurable outputs (here four, $PHP_{sinking}$, $PR_{sinking}$, $PHP_{non\text{-}sinking}$, and zooplankton respiration). Measurement errors (e.g. measurement device errors, *in situ* variabilities, errors due to

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integration methods) are typically challenging to characterize. Furthermore, even if these four outfluxes well describe the prokaryotic and zooplankton compartment fluxes, one may wonder about the sensitivity of the results to the fact that a given outflux is not available or estimated with error.

As a result, we have tested two settings: a model inversion without the zooplankton respiration flux (using only three fluxes) and a second setting where the PGEs were estimated from the leucine incorporation rate using freely varying CFs, i.e. with CFs no more fixed at 0.5 as a value. The results are reported in Table S3 and S4. Not using the zooplankton flux to inverse the model mechanically adds some variability to the estimation results, especially concerning Ψ, $\boldsymbol{\alpha}$, and $PGE_{non\text{-}sinking}$, in decreasing order of variability (Table S3). The $PGE_{sinking}$ was not affected as its confidence interval length was inferior to 10^{-7}: this underlines the very limited influence between the zooplankton and sinking prokaryote compartments in the model (contrary to the zooplankton and non-sinking prokaryote compartments). Yet, the difference between the four-flux and three-flux parameter estimations was negligible (<1% variation for each estimate), highlighting the robustness of the estimates to the potential unavailability of the zooplankton respiration. On the contrary, as made visible in Table S4, not fixing the CFs to estimate the PGEs created more variations in the PGE estimations, while the estimations of Ψ and $\boldsymbol{\alpha}$ changed by less than 5% with respect to Table 2 estimations. The PGEs of the attached and free-living parameters get significantly closer to their fixed boundaries (10%), while the CFs rise, especially the CF of the attached particles (=1.865). Similarly, if PGEs are no longer bounded, the estimates of PGEs (0.173 for attached prokaryotes and 0.226 for free-living prokaryotes) and CFs (3.927 for attached prokaryotes and 1.526 for free-living prokaryotes) become unrealistic. This can be explained by the fact that the PGEs and CFs play similar roles in the current formulation of the model. Hence, without additional fluxes ensuring full model identifiability, one of these two types of quantities needs to be fixed to estimate the other.

In addition to these sensitivity analyses, an uncertainty analysis has been run by simulating errors in the measurements of the POC, DOC and the four output fluxes (see Table S5 in supp. data). Simulating errors from -10% to 10% for each flux, the estimation of the four parameters of interest were lowly affected: 1%, 2%, 3% and 1% on average for the Ψ, $PGE_{sinking}$, $PGE_{non\text{-}sinking}$ and $\boldsymbol{\alpha}$, respectively. The $PGE_{non\text{-}sinking}$ was mostly sensitive to measurement errors of POC flux, DOC flux and $PHP_{non\text{-}sinking}$ (generating variations of 6%, 5% and 5%, respectively). Similarly, the $PGE_{sinking}$ was logically mostly sensitive to errors in the $PHP_{sinking}$ and $PR_{sinking}$

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583 (generating variations of 6% for both). For the measurement errors, the generated variations all
584 remained under 3% which is reassuring concerning the stability of the estimation.
585
586 Finally, the last potential source of estimation bias results from the assumed stationarity
587 hypothesis of the mesopelagic system. For logistical and technical reasons, measurements and
588 sampling between the upper and lower boundary of the mesopelagic zone are typically
589 performed simultaneously. The stationarity assumption is thus a natural foundation ground
590 upon interpretations and models. However, there is a temporal delay in flux variations between
591 the upper layer and lower measurements (Giering et al. 2017; Stange et al. 2017). This delay
592 depends on the particles sinking speed typically ranging from 2 to 1500 m d^{-1} (Alldredge and
593 Silver 1988; Armstrong et al. 2002; Trull et al. 2008; Turner 2015), their morphotype, density
594 and porosity as well as the timing of their production. Strong meteorological events can also
595 perturbate C fluxes from the water column with an increasing time lag over depth (e.g. Pedrosa-
596 Pàmies et al. 2019). Admittedly, C budgets suffer from lack of time integration into the
597 analysis. Our study regarding PAP site is also concerned as it undergoes a substantial
598 seasonality (Cole et al. 2012; Giering et al. 2017). Although, we do not have enough
599 understanding of vertical time lag to change the model and to avoid such bias yet. Some long-
600 term observatories such as BATS in the Bermuda Atlantic or HOT in Hawaii provide
601 biogeochemical flux time series but monthly sampling focuses mostly on the euphotic zone
602 and does not investigate the mesopelagic zone enough. Sampling at discrete times following
603 the sink of a bloom (e.g. Le Moigne et al. 2016) could be a solution, which would nevertheless
604 entail a significant cruise planning effort.
605

606 ## 4.4 Grounds for improvements

607 Anderson & Tang model allowed us to have a comprehensive vision of the remineralization
608 processes in the mesopelagic zone by including the interactions between various
609 compartments, completing *in situ* measurements with a comprehensive vision of the
610 mechanisms at stake. The described inversion of the Anderson & Tang model provided
611 meaningful estimations of the parameters of interest. However, as most models represent
612 complex phenomena, some processes are not fully and properly captured by the model. Below,
613 we provide a list of processes that may help refining mesopelagic C budget estimations.
614

615 ### 4.4.1 Other microorganisms

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Are not included in the model, the role of microbial eukaryotes, viruses, and the input of C by chemolithotrophs whose potential role has gained in importance (Herndl and Reinthaler 2013; Lara et al. 2017; Kuhlisch et al. 2021; Luo et al. 2022). For instance, eukaryotes can dominate microbial biomass on bathypelagic particles (Bochdansky et al. 2017), and have the potential to promote the aggregation of particles (Jain et al. 2005; Chang et al. 2014; Hamamoto and Honda 2019; Xie et al. 2022). Viruses could be the main cause of prokaryotic and phytoplanktonic mortality. Thus, DOC fluxes could be attributed to them, in particular with the cell lyses they provoke (Fuhrman 2000 and ref within, Lara et al. 2017; Kuhlisch et al. 2021). In the North Atlantic, 9 to 12% of cells could be infected by viruses which would cause a DOC production of 0.1 mg C m^{-3} d^{-1} (Wilhem and Suttle 1999). For comparison, PHP results on PAP before integration (with a conversion factor of 0.5 kg C mol^{-1} Leu) were mostly below this value. In addition, inorganic C fixation by chemoautotrophy would be of the same order of magnitude as $PHP_{non\text{-}sinking}$ rates (Herndl et al. 2005; Reinthaler et al. 2010). It would be important to verify what microbial eukaryotes, chemolithotrophs or viruses contributions are, even if the poor understanding of these processes currently prevents properly integrating them into models.

4.4.2 Lifestyles

More surprisingly, sinking prokaryotes are poorly considered as they are not sampled with the Niskin bottles classically used in oceanography (Planquette and Sherrell 2012; Baumas et al. 2021). However, the use of the MSC at PAP DY032 allows us to access fractions of particulate organic carbon that will allow us to evaluate the importance of sinking prokaryotes. We have seen that their C demand is not negligible and represents 18% of total C demand. Anderson & Tang model distinguishes sinking particles from neutrally buoyant particles, each with distinct attached communities. Since sampling with MSC only allows us to separate what is sinking from what is not, we merged free-living prokaryotes with those attached to neutrally buoyant particles without distinction. However, unlike free-living prokaryotes, prokaryotes attached to neutrally buoyant particles have access to POC and must produce enzyme activity with different metabolisms than their free-living counterparts. On the other hand, prokaryotes attached to neutrally buoyant particles are also different from prokaryotes attached to sinking particles since they do not undergo changes in temperature and pressure related to the sink. They must therefore surely have intrinsically different PGE and associated remineralization rates. It would therefore be valuable to consider them as a third distinct group in laboratory experiments and sampling. Contrary to the sinking or ascending particles which are naturally split by their sinking/ascending velocity (e.g. respectively Smith et al. 1989; Cowen et al. 2001;

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McDonnell et al. 2015), no means allow the selective and exclusive sampling of neutrally buoyant particles. The only valid way is to use the MSC to let the sinking particles fall into the lower compartments and to filter the "non-sinking" part to retain the particulate fraction. However, it is known that filtration affects the activities of prokaryotes and generates biases (Edgcomb et al. 2016). This makes investigations of prokaryotes associated with neutrally buoyant particles particularly challenging and future endeavors should urgently attempt to target them.

4.4.3 OC inputs

Continuing in the same line, the inputs of C that the model takes into account are only the gravitational POC and the DOC. We chose to artificially increase the gravitational POC flux to add sources of neutrally buoyant particles in the form of PIPs (eddy subduction pump, metazoans migrations and large-scale physical pumps). Indeed, Boyd et al. (2019) clearly showed that these PIPs can be of paramount importance (here we have estimated them at 51.6% of the gravitational flux). Accounting for these neutrally buoyant particles through the POC flux was performed due to the model structure. Yet, explicitly describing them in a dedicated compartment of the model could be an improvement for future research, as these neutrally buoyant particles have an effect on the whole system, including the prokaryotes linked to various types of particles and their predators or on particle fragmentation. Given the existence of the neutrally buoyant particle compartment, it is feasible to adapt the model to account for these C inputs. This is even more relevant as new optical instruments have flourished (e.g. Briggs et al. 2013; Giering et al. 2020; Picheral et al. 2022) and would make it easier to better quantify these neutrally buoyant particle fluxes.

4.4.4 *In situ* pressure effect

Our last major concern deals with the fact that neither Niskin nor MSC avoid disruption introduced through the process of depressurization when samples are collected at depth (Tamburini et al. 2013; Garel et al. 2019). Heterotrophic activities associated to non-sinking prokaryotes are known to decrease with depth but were mostly sampled without taking care of the *in situ* pressure (e.g. Turley and Mackie 1994; Arístegui et al. 2009). From our knowledge, some devices such as the $IODA_{6000}$ (Robert 2012) were specifically designed to measure *in situ* PR of non-sinking prokaryotes. However, enigmatically high PR values (2-3 orders of magnitude higher than PHP) are measured by $IODA_{6000}$, making it difficult to have confidence in these *in situ* measured PR rates. During the PEACETIME cruise, we use a pressure-retaining

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685 sampler (methods presented in supp data), allowing for the first time to access both $PHP_{non-sinking}$ and $PR_{non-sinking}$ rates and to compare it with classical depressurization procedures (Fig. S1). We observed that activity rates of non-sinking prokaryotes kept under pressure were always higher when kept at *in situ* hydrostatic pressure than their decompressed counterparts and, surprisingly, seem to increase with depth rather than decrease typically depicted and found when the samples are decompressed (Fig. S1). Focusing on $PR_{non-sinking}$ rates, obtained values are also several orders of magnitude too high to be realistic in regard to C-Budgets and prevent us from calculating PGEs. As PHP and PR are linked, it is very likely that the pressure effect (here, an increase) is reflected on both and thus in the associated $PGE_{non-sinking}$. Taking hydrostatic pressure into account could thus drastically affect C-budgets and even for zooplankton respiration as we saw in the model that they are really sensitive to $PGE_{non-sinking}$. We highly recommend using either direct *in situ* measurements or pressure retaining systems for future research. This advice should be followed carefully, especially from 500m where the pressure effect starts to be very important (Fig. S1), while the piezosphere was previously considered below 1000m depth (Jannasch and Taylor 1984; Yayanos 1986). Furthermore, this shows the crucial interest to measure points below 500m in order to get a global trend of the profile, which could not have been done here for sinking prokaryotes (the MSC were deployed only up to 500m during the cruise DY032). From a C-budget point of view, taking *in situ* pressure into account will increase C demand of free-living prokaryotes well adapted to their living depth.

The effect of pressure acts inversely on sinking prokaryotes, as they are surface prokaryotes (unadapted to high-hydrostatic pressure) that undergo a dynamic pressure increase as the particle sinks (Baumas et al. 2021; Tamburini et al. 2021). Besides, repeated results (Tamburini et al. 2006, 2009, 2021; Riou et al. 2018) have shown that, while performing a sinking simulation experiment the activities of sinking prokaryotes are affected during the sink. For instance, they noticed that the aminopeptidase activity was always lower with increasing pressure over time than at atmospheric pressure on diatom aggregates (Tamburini et al. 2006). This may reflect the stress endured by the sinking prokaryotes as they experience the sink. This could also be another explanation of why the fraction of hydrolyzed C released (**α**) tends to decrease with depth as it is directly linked with aminopeptidase activity. In view of these statements, it is not surprising that the PHP and PR, and therefore a PGE, are impacted by increasing pressure (e.g. Stief et al. 2021; Tamburini et al. 2021). Only the RESPIRE from Boyd et al. (2015) provides *in situ* measurements of the PR of sinking prokaryotes. However, in line with the previous comments, it gives unrealistically rather high values. Thus, sinking

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720 simulation experiments remains, in the present, the best alternative to understand the mechanics
721 of sinking prokaryotes during the sink of their associated particle. Several systems exist to
722 simulate the sink (e.g. de Jesus Mendes et al. 2007; Grossart and Gust 2009; Tamburini et al.
723 2009; Mendes and Thomsen 2012; Dong et al. 2018; Stief et al. 2021; Liu et al. 2022), all
724 showing a general tendency that hydrostatic pressure affects activities (and diversity) of
725 surface-originated prokaryotes, decreasing the integrated C-demand when taking into account.
726 Handling high-pressure sampling or experiments requires much more effort and material than
727 usual methods. However, it seems highly worthy when investigating both, sinking and non-
728 sinking prokaryotes activities, in regard to C-budget purposes.
729

730 # 5. Conclusion

731 By combining *in situ* data from the DY032 cruise at the PAP site with inversion of the
732 Anderson & Tang model which includes known processes from the biological C pump, we
733 provide robust and ecologically realistic estimates of key parameters and to better characterize
734 the patterns at stake.

735 1) We showed that the most sensitive parameters in the model are the ones related to
736 prokaryotes such as prokaryotic growth efficiencies, leucine-to-carbon conversion
737 factor, and C hydrolyzed by sinking prokaryotes released to the surrounding water.

738 2) By inversion of Anderson and Tang's model, we determined consistent values of the
739 parameters listed above.

740 3) We showed that using these values instead of the classical mean from literature or
741 inadequate theoretical values resulted in a more consistent and realistic C-budget than
742 previously considered.

743 4) Additional measurements are needed to better understand both prokaryotic growth
744 efficiencies and Leucine-to-Carbon conversion factors in the mesopelagic zone.
745 However, we recommend measuring fewer fluxes for which we are confident
746 associated with inversion model procedures in order to access parameter values
747 challenging to measure in other places, cruises, or seasons.

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Fig. 2 summarizes processes involved in mesopelagic C budgets estimations and highlights missing knowledges. We attempt to classify the processes according to their degree of understanding (well known, insufficient data or unknown) and point out that majority of these processes require a better understanding. Among others, it is crucial to quantify the roles of microbial eukaryotes, viruses, and chemoautotrophs in the entire process of C budgets. Suspended particles should have a dedicated well-identified compartment in future studies instead of being neglected and drowned into others. Finally, accounting for *in situ* hydrostatic pressure when studying prokaryotic C demand is key. This is because: 1) it may reduce PCD for sinking prokaryotes unadapted to increasing pressure and 2) it may increase PCD for free-living prokaryotes well-adapted to their living depth.

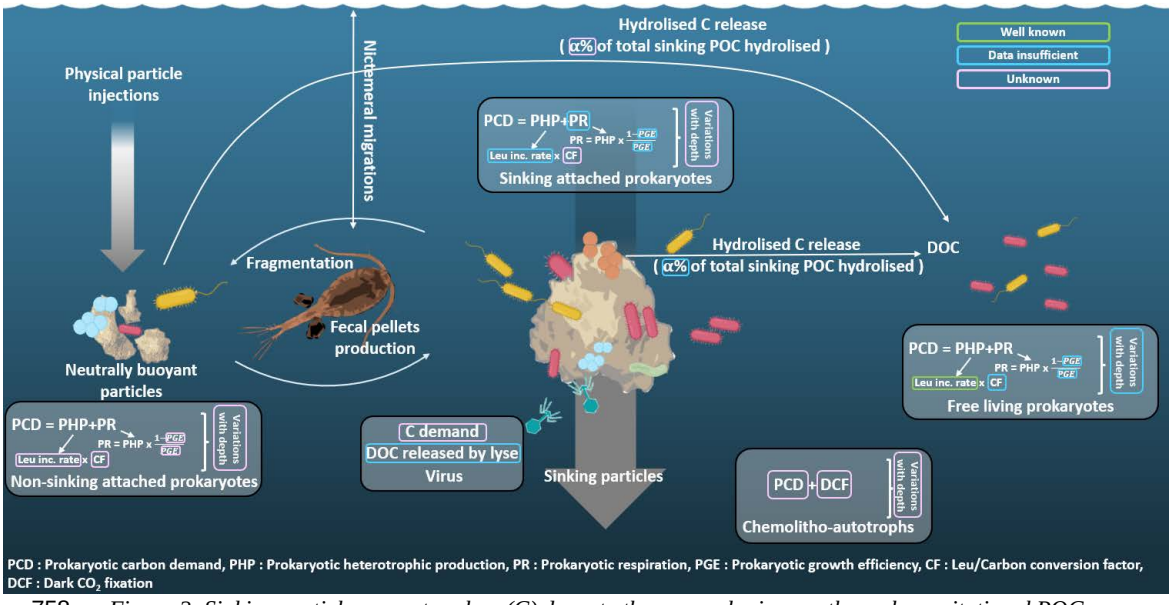

Figure 2: Sinking particles export carbon (C) down to the mesopelagic zone through gravitational POC fluxes where this latter is attenuated to satisfy C demand of different groups of organisms such as prokaryotes living attached to sinking particles, attached to non-sinking particles, or free-living prokaryotes. In turn, viruses and chemoautotrophs can increase the amount of usable labile C. Quantifying C demand and role on POC fluxes of these different groups is crucial to truly assess C sequestration in the deeper layer of the water column. However, a multitude of uncertainties remains for each group. The quantities enclosed in green are well known, in blue lack data and in pink are unknown. C demand is the sum of heterotrophic production (PHP) and respiration (PR). The understanding of these two quantities is currently better for the free-living prokaryotes whereas data are still insufficient for sinking prokaryotes and even absent for prokaryotes attached to non-sinking particles. Moreover, to build C budgets, these variables are integrated over a few hundred meters of water column and the relationship between in situ pressure and C demand remains often neglected even if this relationship highly depends on the prokaryote type considered (not constant for sinking

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prokaryotes unadapted to the increased pressure, constant for free-living prokaryotes well adapted to their living depth and constant for prokaryotes attached to non-sinking particles which can be adapted or not if the particle was sinking before being stopped in its sink).

Code/Data availability

The codes and data to reproduce the results are available at https://github.com/RobeeF/InverseCarbonBudgetEstim

Author contribution

The idea was conceived by CB, CT and JCP. Sampling and experiments onboard PEACETIME cruise were conducted by CT and MG. The data processing of PAP DY032 data was conducted by CB with advices from FLM, and the one from PEACETIME data by CB and MG. RF designed the inversion detection methodology and performed the estimation with advices from LM. CB and RF led the writing with significant contributions from all authors.

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Competing interests

The authors declare that they have no conflict of interest.

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